

Personalized Phased Diploid Genomes of the EN-TEx Samples

Michael Schatz, Fritz Sedlazeck, Han Fang, Maria Nattestad, Ruibang Luo, Srividya Ramakrishnan, Charlotte Darby, Philipp Rescheneder, Alex Dobin, Carrie Davis, Ashwin Prakash, Anna Vlasova, Alessandra Breschi, Roderic Guigo, Tom Gingeras

Oct 2, 2017

ENTex Update



NGMLR and Sniffles

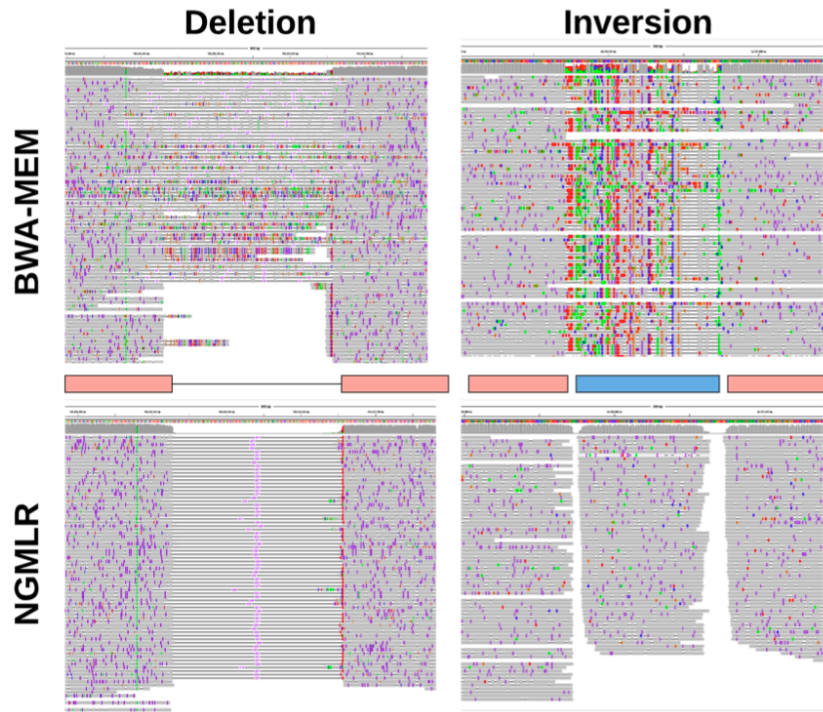
“NGMLR + Sniffles paper”: *Accurate detection of complex structural variations using single molecule sequencing*

Sedlazeck, FJ et al. (2017) *bioRxiv* <https://doi.org/10.1101/169557>

“SKBR3 paper”: *Complex rearrangements and oncogene amplifications revealed by long-read DNA and RNA sequencing of a breast cancer cell line*

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NGMLR and Sniffles



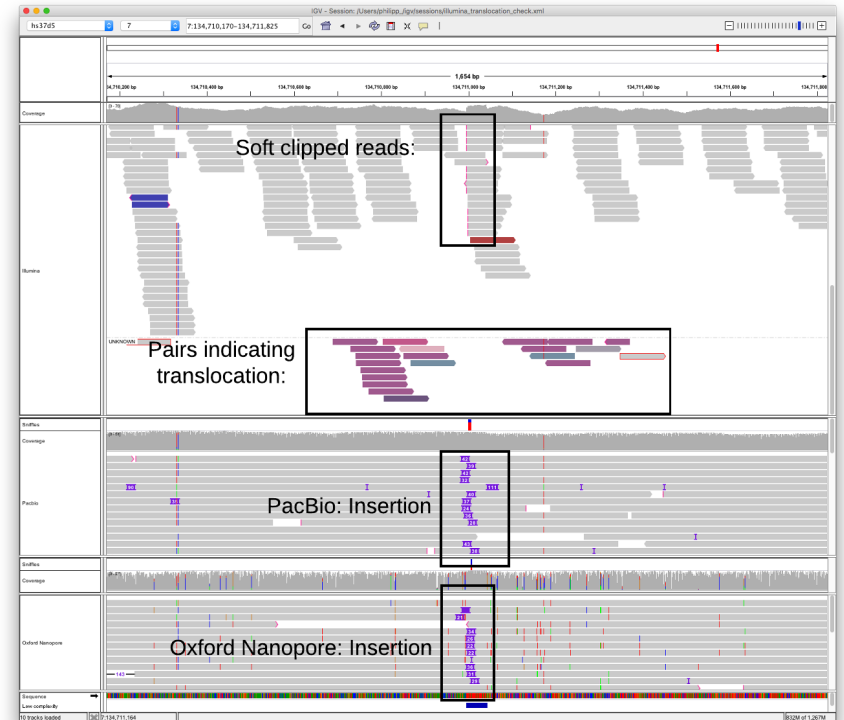
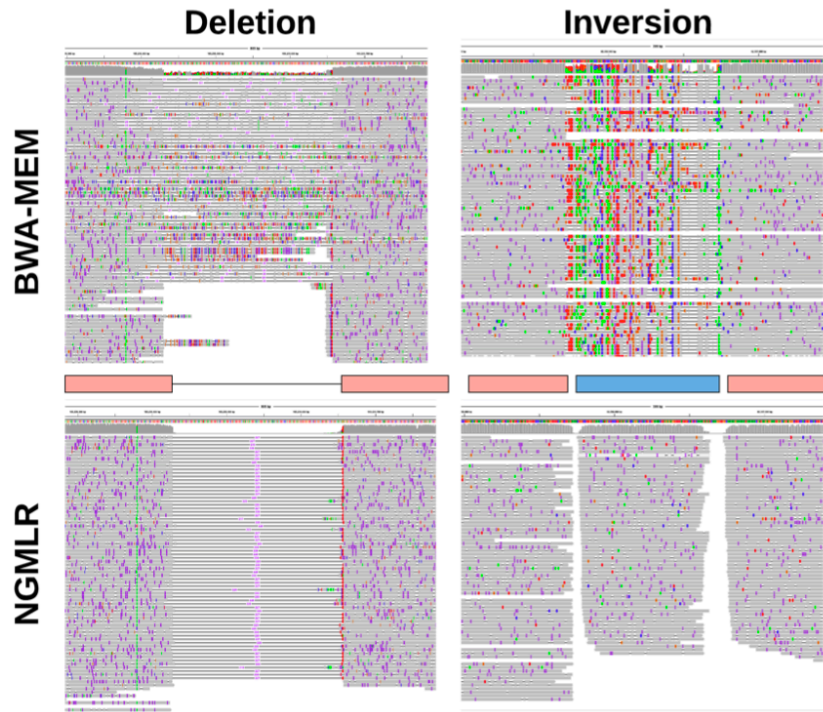
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NGMLR and Sniffles



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NGMLR and Sniffles

Data Set	Tech.	Cov.	Avg. read length(bp)	Total SVs	DEL	DUP	INS	INV	TRA
Arabidopsis Col-0	PacBio	127x	6,482	355	67	63	106	68	51
Arabidopsis CVI	PacBio	123x	6,073	9,652	3,822	904	1,823	478	2,625
Arabidopsis Col-0 x CVI F1	PacBio	155x	11,206	11,935	4,974	582	4,049	567	1,763
Arabidopsis Col-0 X CVI F1	Illumina	40x	250	10,950	4,324	643	0	671	5,312
Giab HG002 (son)	PacBio	69x	8,540	19,131	7,957	1,084	9,656	232	202
Giab HG002 (son)	Illumina	80x	148	10,822	5,018	863	0	823	4,118
Giab HG003 (father)	PacBio	32x	6,284	11,964	5,296	408	6,048	99	113
Giab HG003 (father)	Illumina	80x	148	11,395	5,553	869	0	818	4,155
Giab HG004 (mother)	PacBio	30x	7,285	10,463	4,590	276	5,436	93	68
Giab HG004 (mother)	Illumina	80x	148	8,901	5,000	868	0	829	2,204
NA12878 (healthy female)	PacBio	55x	4,334	15,499	6,734	606	7,880	160	119
NA12878 (healthy female)	Oxford Nanopore	28x	6,432	17,155	12,301	323	4,401	87	43
NA12878 (healthy female)	Illumina	50x	101	7,275	3,744	553	0	731	2,247
SKBR3 (Breast Cancer)	PacBio	69x	9,872	19,165	7,268	1,019	10,391	328	159
SKBR3 (Breast Cancer)	Illumina	25x	101	5,046	2,776	483	0	627	1,160

Table 1: Summary of detected SVs across 15 different data sets. SVs were reported with a min. size of 50bp using SURVIVOR based on Delly, Lumpy and Manta for Illumina or Sniffles for PacBio or Oxford Nanopore requiring at least 10 reads. Supplementary Table 5 shows all the data sets used.

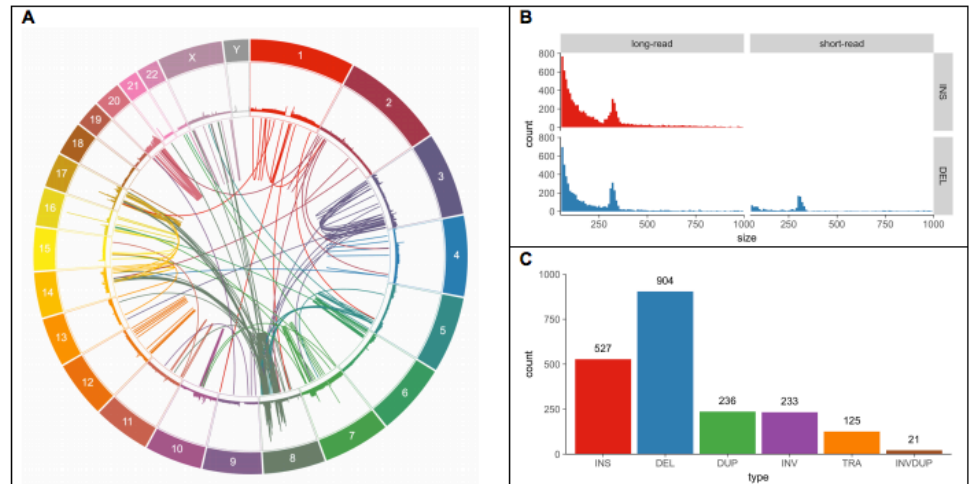


Figure 1 | Variants found in SK-BR-3 with PacBio long-read sequencing. (A) Circos plot showing long-range (larger than 10 kbp or interchromosomal) variants found by Sniffles from split-read alignments, with read coverage shown in the outer track. **(B)** Variant size histogram of deletions and insertions from size 50 bp up to 1 kbp found by log-read (Sniffles) and short-read (Survivor 2-caller consensus) variant-calling, showing similar size distributions for insertions and deletions from long reads but not for short reads where insertions are entirely missing. **(C)** Sniffles variant counts by type for variants above 1 kbp in size, including translocations and inverted duplications.

“NGMLR + Sniffles paper”: Accurate detection of complex structural variations using single molecule sequencing

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Structural Variations Concordance (ENC-002)

Sniffles	17,107	PacBio				
Falcon	7,857	12,241				
LongRanger	2,823	1,946	3,785	10X Genomics		
SuperNova	3,394	2,837	1,486	18,862		
SURVIVOR2	3,291	2,163	2,274	1,646	6,631	Illumina
MegaHit	1,858	1,529	569	1,378	687	3,855
	Sniffles	Falcon	LongRanger	SuperNova	SURVIVOR2	MegaHit

Main Diagonal

- Calls per tool

Outer triplets

- Concordance by Technology

Inner triplets

- Concordance by Assembly
- Concordance by Mappers

Overall:

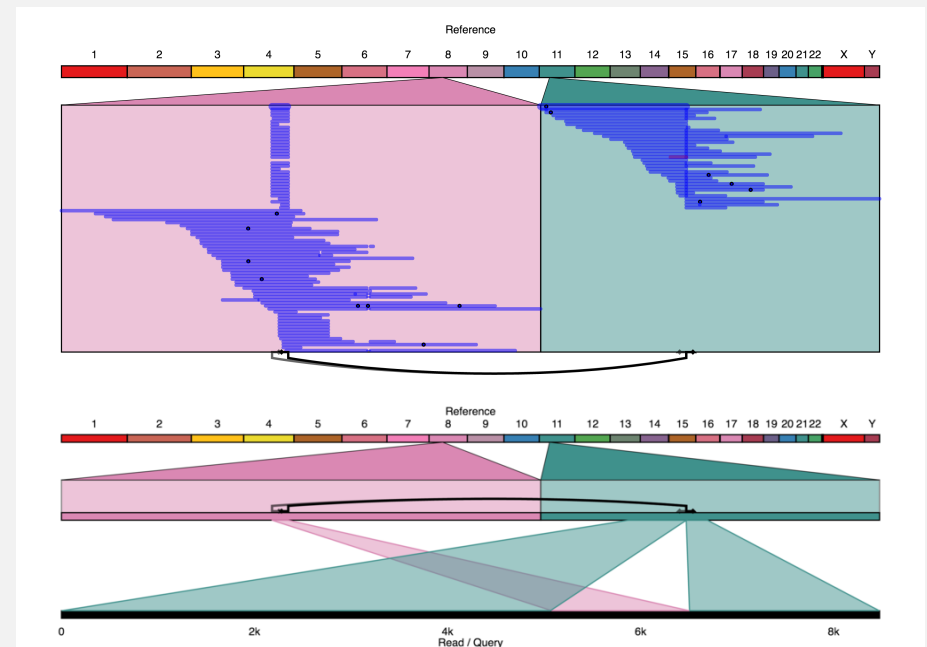
- We need multiple technologies and approaches

Sniffles PacBio Variant Calls (ENC-002)

Sniffles calls

	All SVs (50bp+)	Large SVs (10kbp+)
Deletions	7,124	163
Duplications	1,642	153
Insertions	7,904	0
Inversions	275	144
Translocations	162	162
All	17,107	622

Translocation in Ribbon



Ribbon: Visualizing complex genome alignments and structural variation

Nattestad et al. (2016) *bioRxiv* doi: <http://dx.doi.org/10.1101/082123>

Sniffles PacBio Variant Calls

ENC-002

	All SVs (50bp+)	Large SVs (10kbp+)
Deletions	7,124	163
Duplications	1,642	153
Insertions	7,904	0
Inversions	275	144
Translocations	162	162
All	17,107	622

ENC-003

	All SVs (50bp+)	Large SVs (10kbp+)
Deletions	7,747	128
Duplications	1,511	116
Insertions	9,528	0
Inversions	224	106
Translocations	101	101
All	19,111	451

Current Calls: 50bp+ & supported by 10+ reads (High Confidence)

Sensitive Analysis: 10bp+ event & supported by 5+ reads

Variants Per Chromosome

ENC-002

Chr	DEL	DUP	INV	INS	TRA
1	459	106	13	606	19
2	550	116	10	576	21
3	376	50	7	453	16
4	526	50	9	451	7
5	400	69	13	383	4
6	443	88	5	502	7
7	441	103	10	445	16
8	372	75	2	388	7
9	242	50	15	340	13
10	372	84	10	408	10
11	285	72	16	397	4
12	315	64	16	405	7
13	252	56	3	319	8
14	179	26	1	210	1
15	168	27	1	212	1
16	358	226	115	250	2
17	271	63	2	273	2
18	204	46	3	219	0
19	215	42	6	255	0
20	199	88	7	244	8
21	161	44	3	167	7
22	124	44	5	156	0
X	197	41	2	201	2
Y	15	11	0	29	0
M	0	1	1	15	0

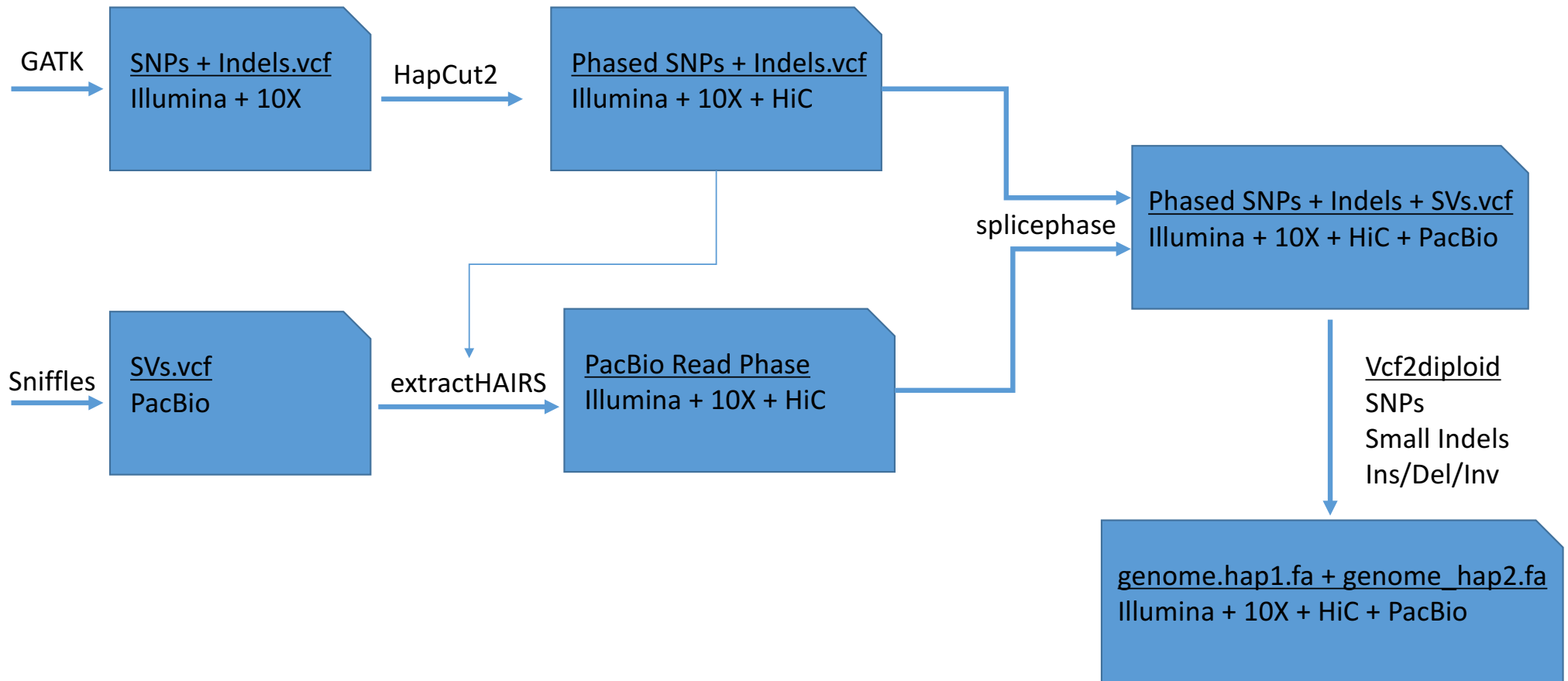
Raw Coverage: 165,114,151,916bp / 3Gb = 55.0x
 Raw Length: 7538 +/- 5610 bp
 Mean alignment length: 5257 +/- 5457 bp

ENC-003

Chr	DEL	DUP	INV	INS	TRA
1	480	91	11	727	10
2	609	125	9	686	7
3	390	46	6	560	9
4	525	47	6	557	2
5	439	62	13	461	4
6	490	75	7	594	3
7	507	105	9	546	9
8	372	73	7	482	4
9	286	49	19	397	12
10	409	78	9	450	1
11	316	71	11	439	4
12	344	49	9	451	1
13	294	57	3	389	11
14	182	31	3	244	1
15	189	21	0	235	1
16	347	163	67	266	2
17	261	71	5	352	1
18	232	41	2	283	0
19	262	54	3	302	0
20	206	78	12	306	8
21	157	45	4	188	8
22	168	37	1	188	1
X	273	39	8	403	2
Y	9	2	0	18	0
M	0	1	0	4	0

Raw coverage: 171,860,416,260bp / 3Gb = 57.2x
 Raw length: 6974 +/- 5854 bp
 Mean alignment length: 5682 +/- 4673 bp

Diploid Construction

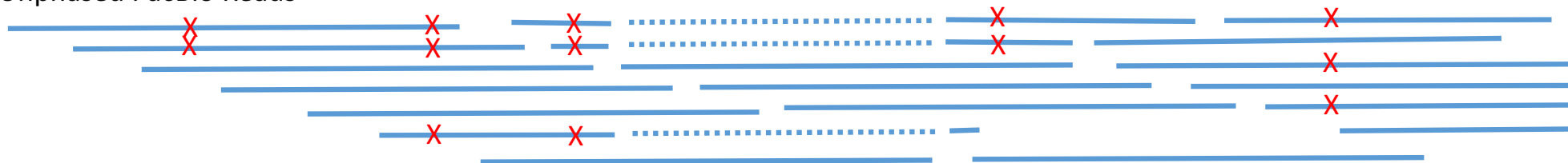


SVPhaser

Unphased Illumina Variants



Unphased PacBio Reads

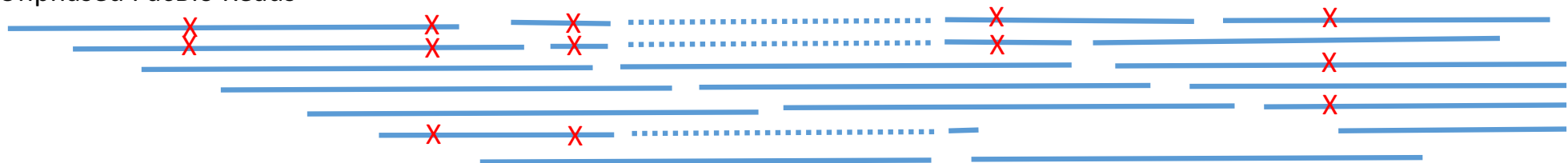


SVPhaser

Phased Illumina Variants



Unphased PacBio Reads

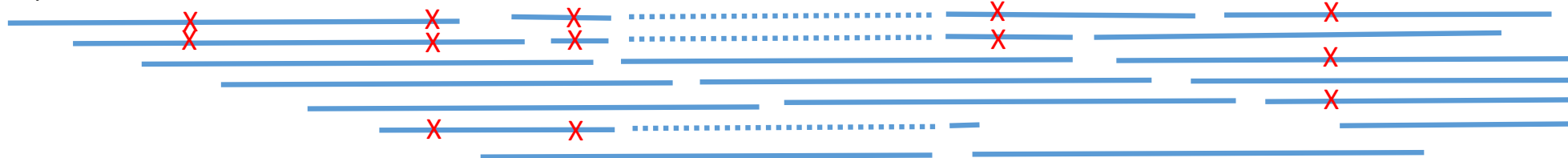


SVPhaser

Phased Illumina Variants



Unphased PacBio Reads

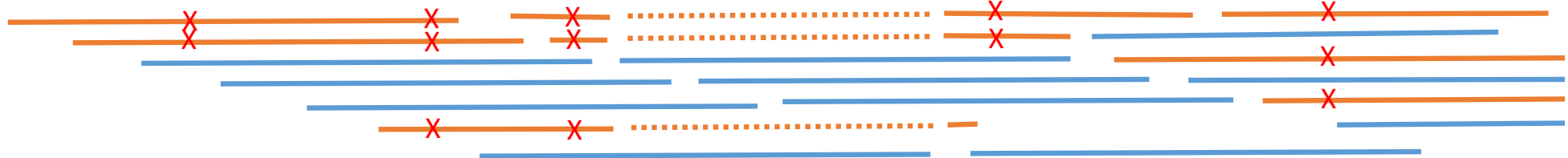


SVPhaser

Phased Illumina Variants



Phased PacBio Reads

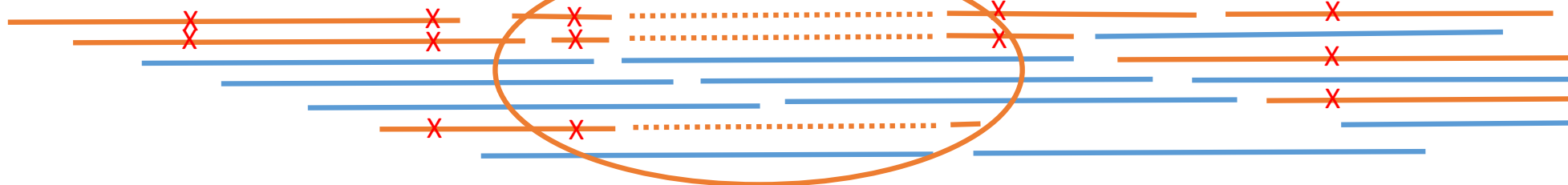


SVPhaser

Phased Illumina Variants

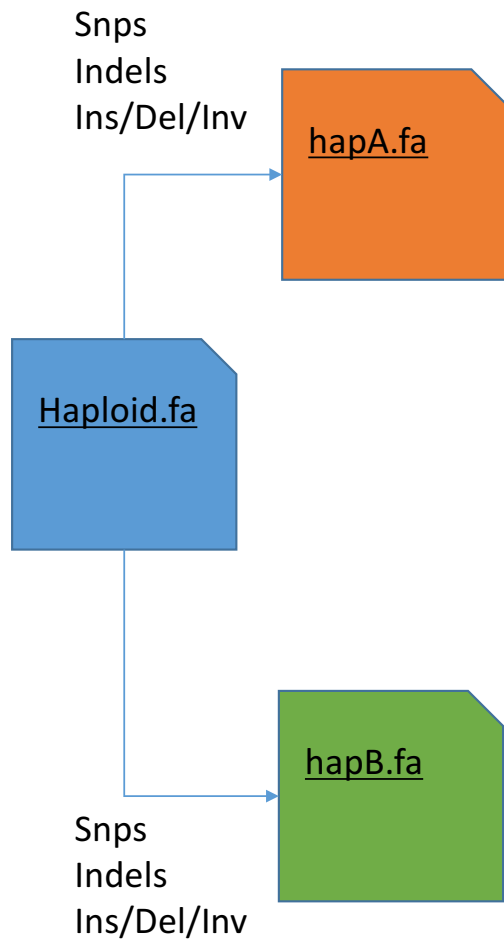


Phased PacBio Variants

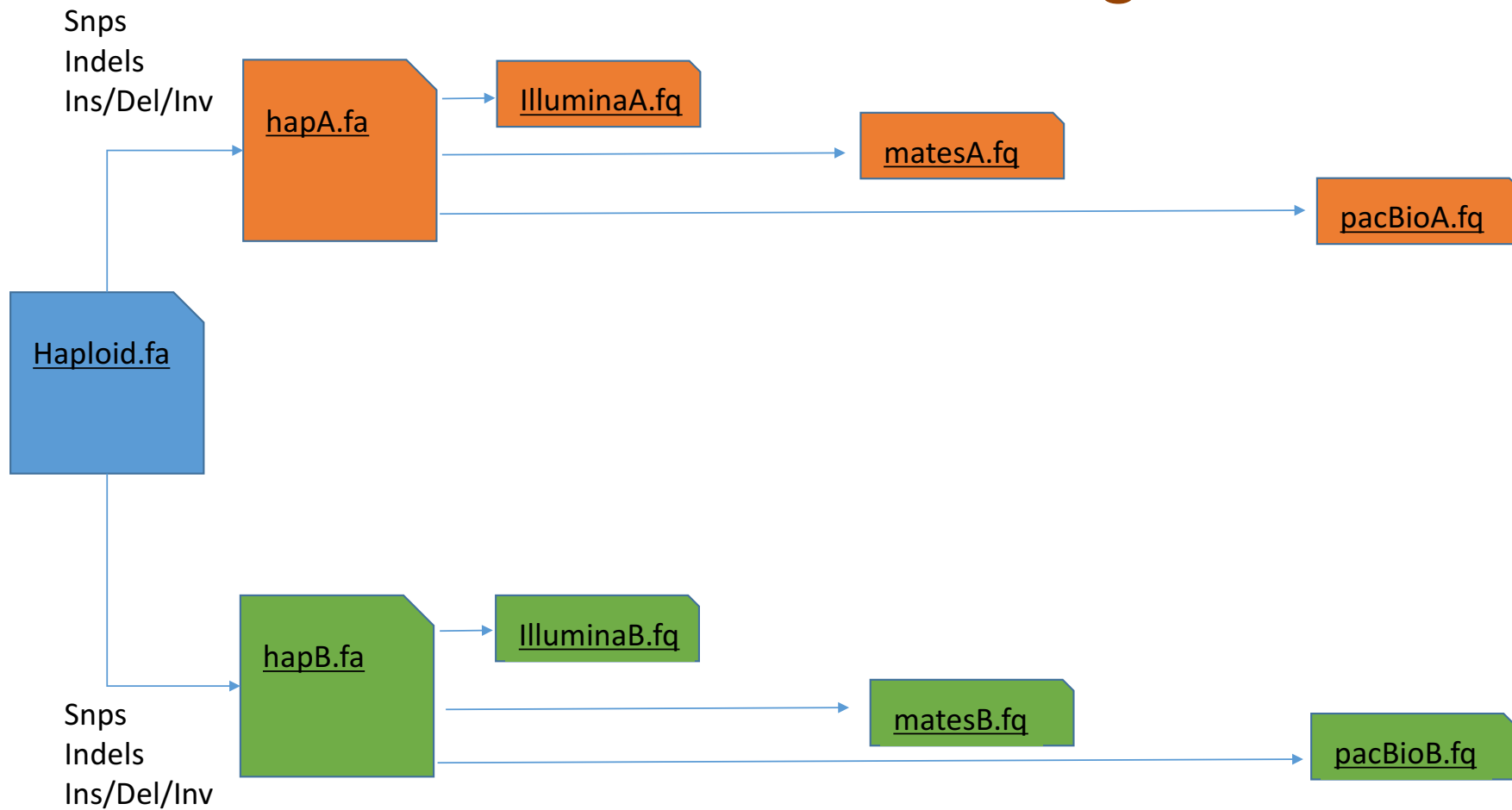


Deletion must be on the orange haplotype!

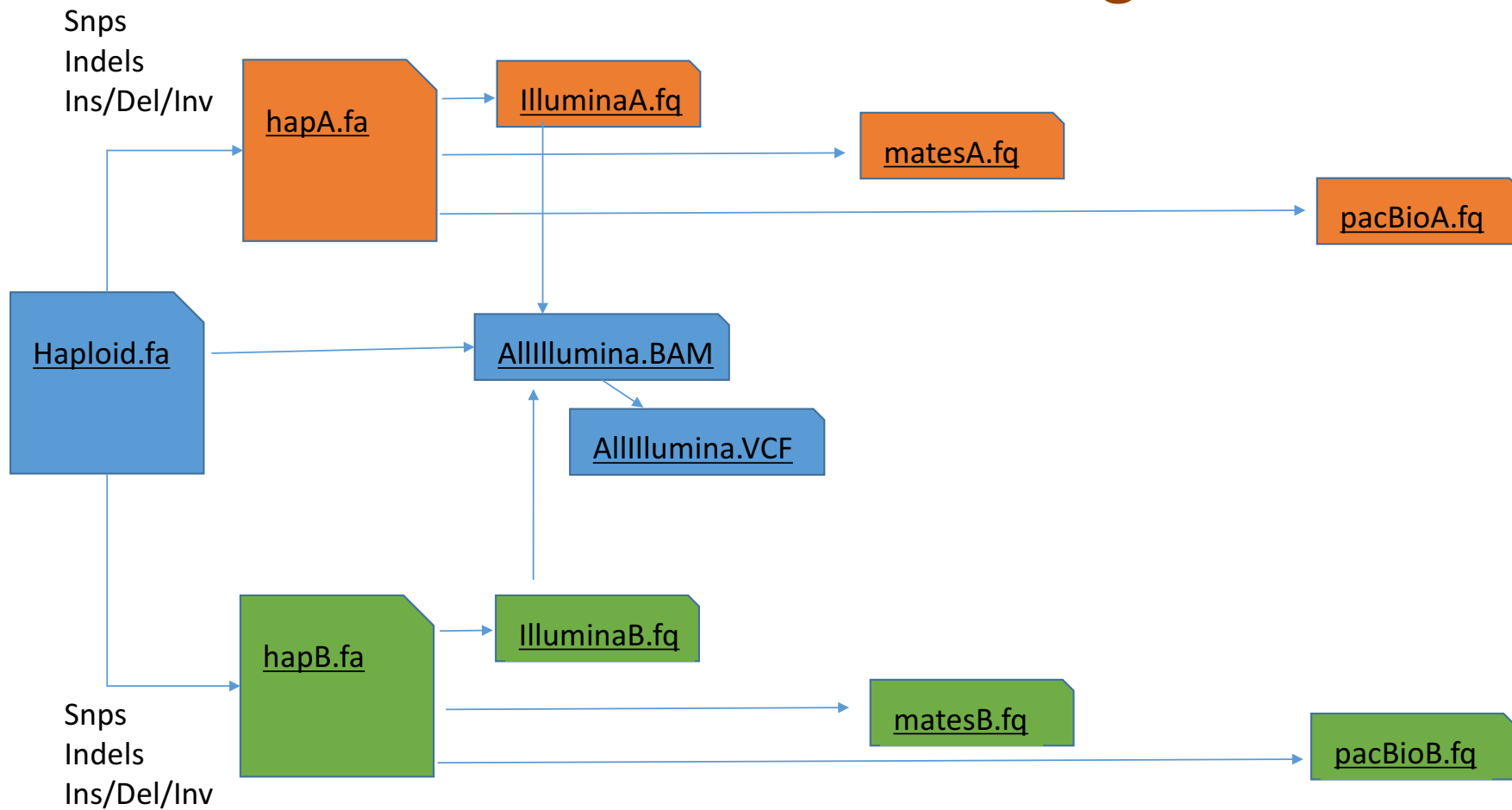
SVPhaser Testing



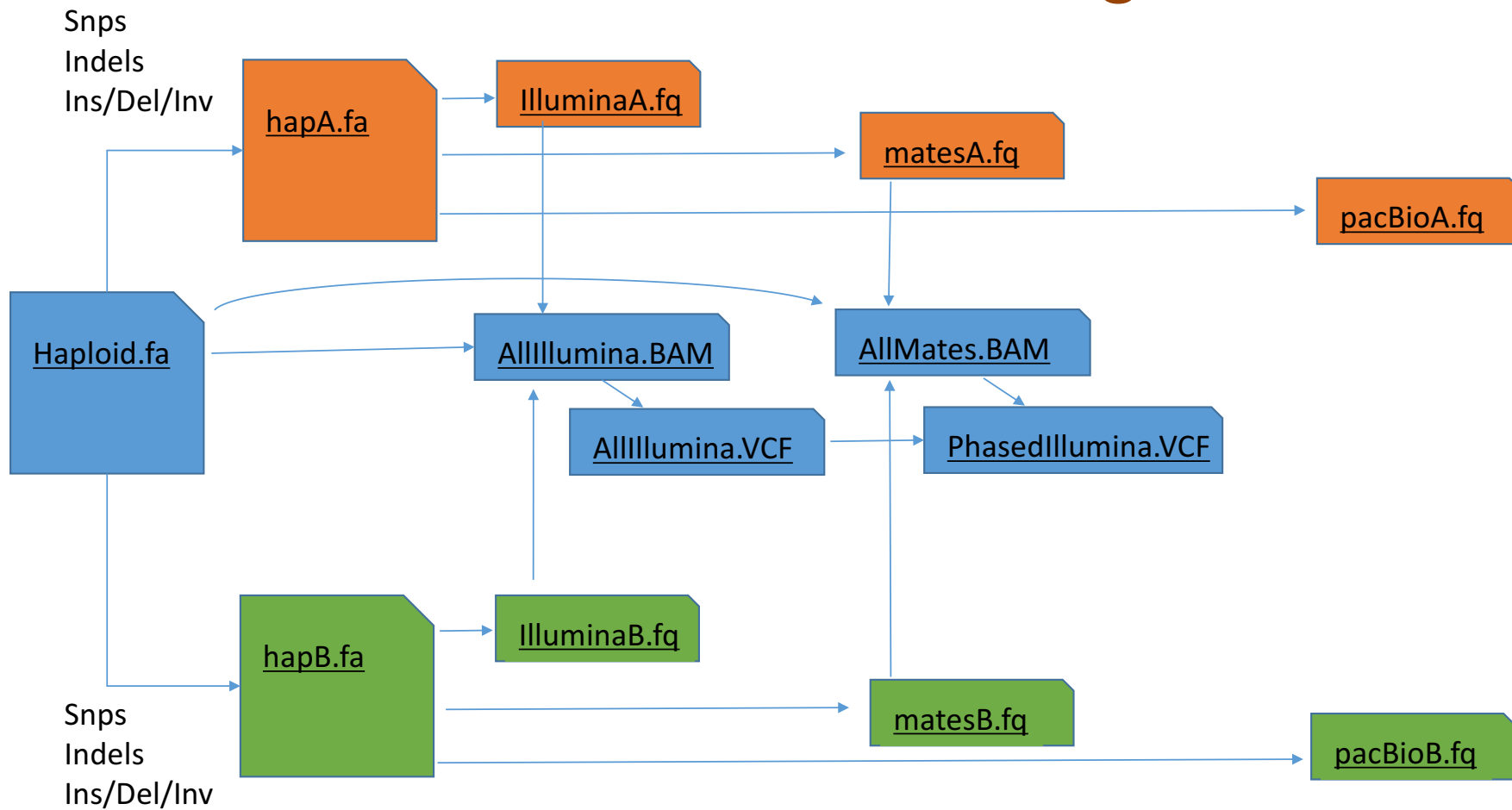
SVPhaser Testing



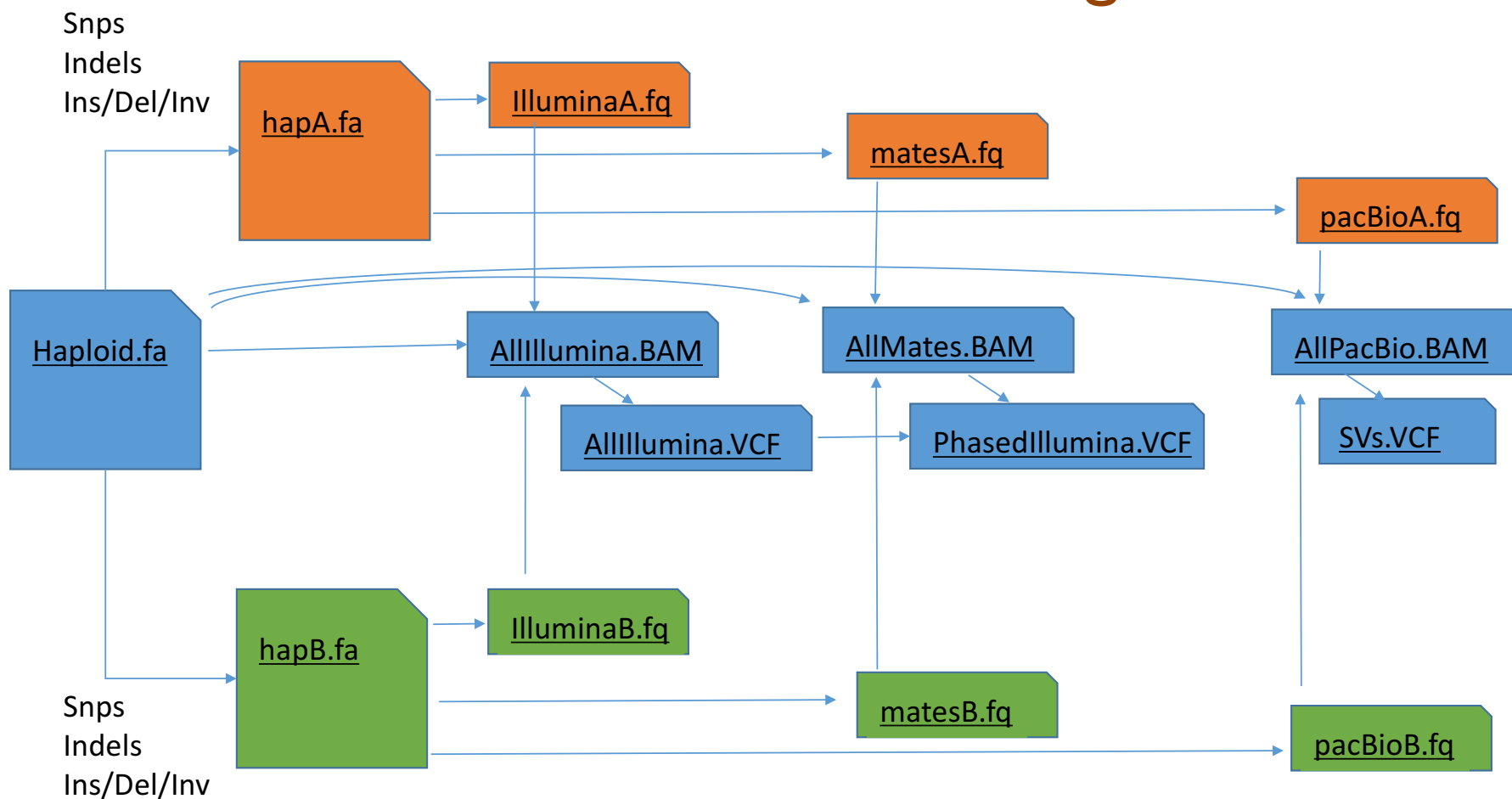
SVPhaser Testing



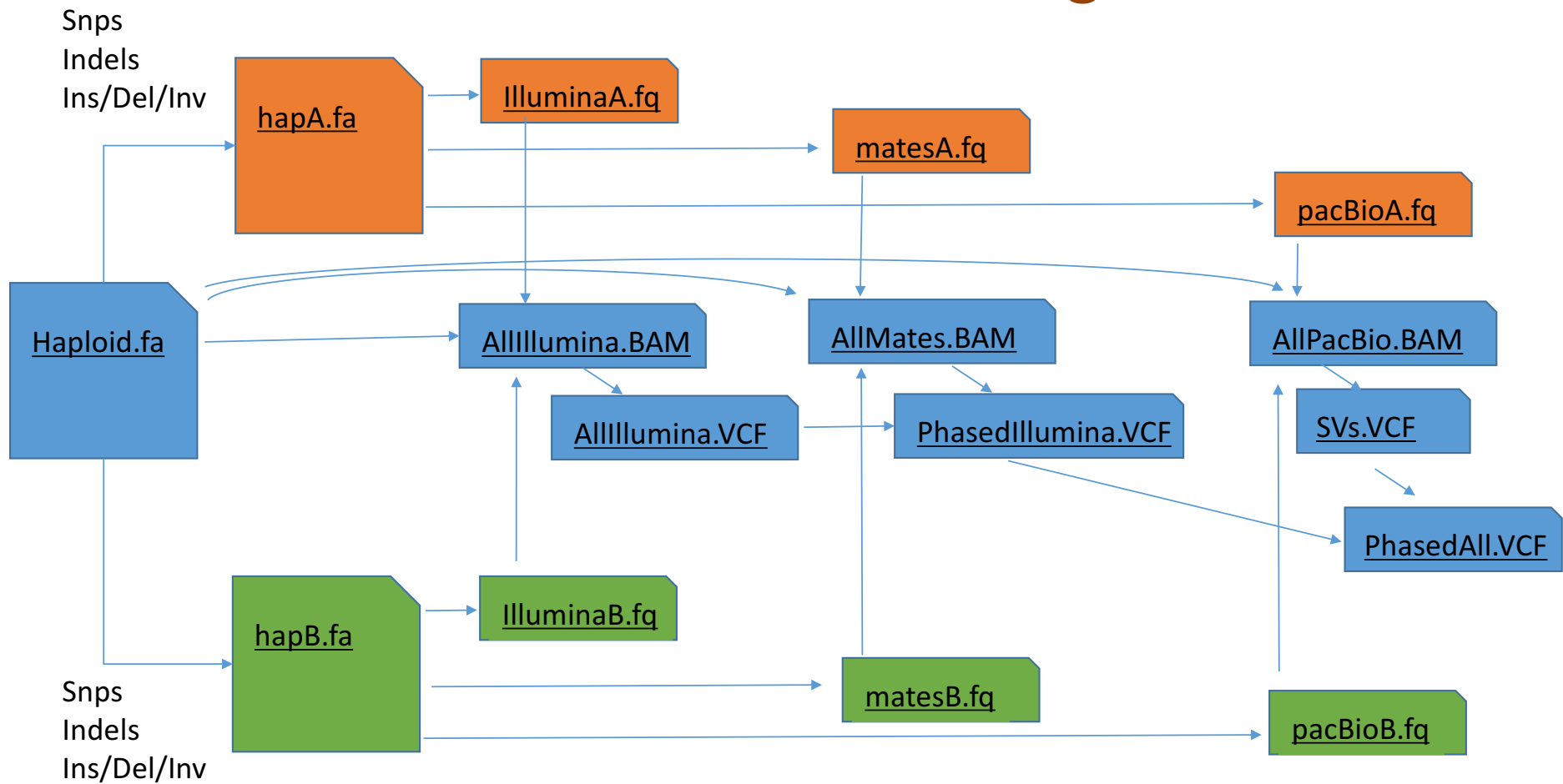
SVPhaser Testing



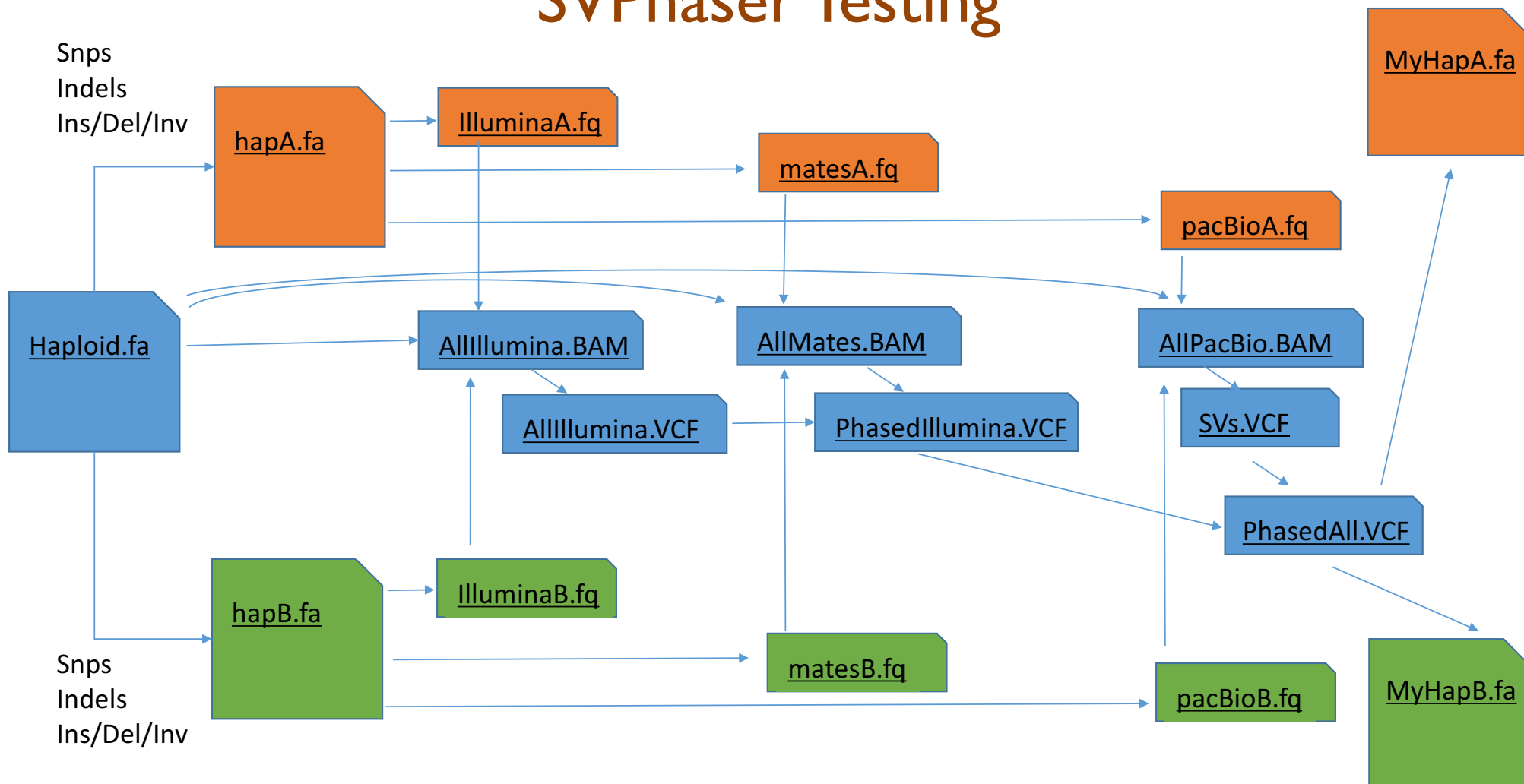
SVPhaser Testing



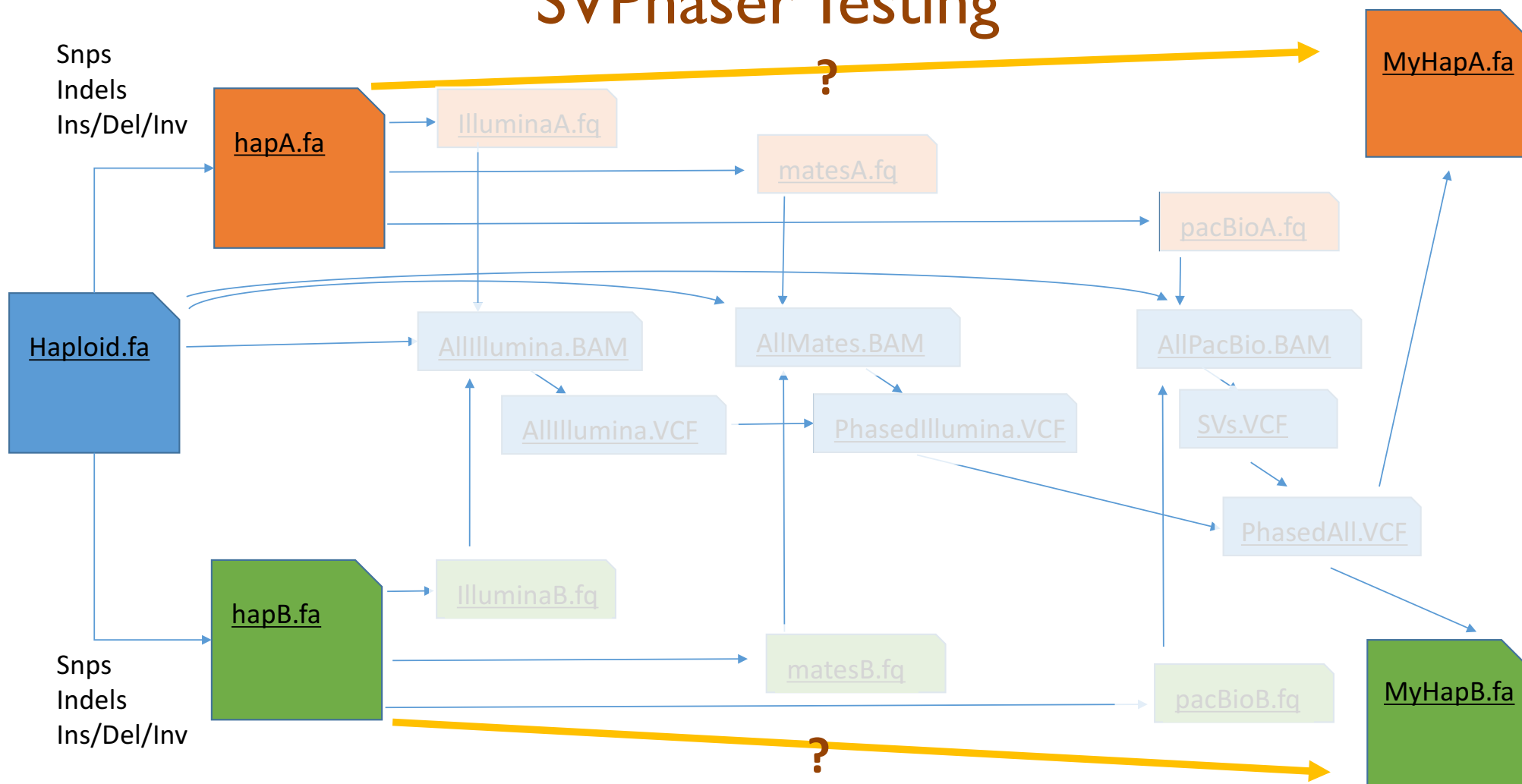
SVPhaser Testing



SVPhaser Testing



SVPhaser Testing



SVPhaser Testing (standard AlleleSeq)

```
$ nucmer mutA.fasta chr1_hapA.fa
```

[S1]	[E1]	[S2]	[E2]	[LEN 1]	[LEN 2]	[% IDY]	[LEN R]	[LEN Q]	[COV R]	[COV Q]	[TAGS]	
=====	=====	=====	=====	=====	=====	=====	=====	=====	=====	=====	=====	=====
1	6093	1	6093	6093	6093	100.00	705447	699930	0.86	0.87	chr1	chr1_maternal
7339	21137	6094	19892	13799	13799	99.99	705447	699930	1.96	1.97	chr1	chr1_maternal
22158	28972	19892	26706	6815	6815	100.00	705447	699930	0.97	0.97	chr1	chr1_maternal
28973	66028	28648	65703	37056	37056	100.00	705447	699930	5.25	5.29	chr1	chr1_maternal
66833	71776	65699	70641	4944	4943	99.94	705447	699930	0.70	0.71	chr1	chr1_maternal
71768	75348	71610	75190	3581	3581	100.00	705447	699930	0.51	0.51	chr1	chr1_maternal
75348	103011	76044	103707	27664	27664	99.98	705447	699930	3.92	3.95	chr1	chr1_maternal
103011	123468	104645	125102	20458	20458	100.00	705447	699930	2.90	2.92	chr1	chr1_maternal
124039	141409	125103	142473	17371	17371	100.00	705447	699930	2.46	2.48	chr1	chr1_maternal
142112	145387	142474	145749	3276	3276	100.00	705447	699930	0.46	0.47	chr1	chr1_maternal
146107	178148	145750	177791	32042	32042	99.99	705447	699930	4.54	4.58	chr1	chr1_maternal
178826	189042	177792	188008	10217	10217	99.97	705447	699930	1.45	1.46	chr1	chr1_maternal
189665	195023	188009	193367	5359	5359	100.00	705447	699930	0.76	0.77	chr1	chr1_maternal
195024	200386	195238	200600	5363	5363	100.00	705447	699930	0.76	0.77	chr1	chr1_maternal
201520	231233	200601	230315	29714	29715	99.99	705447	699930	4.21	4.25	chr1	chr1_maternal
231222	268940	232037	269755	37719	37719	100.00	705447	699930	5.35	5.39	chr1	chr1_maternal
268941	273282	271210	275551	4342	4342	99.98	705447	699930	0.62	0.62	chr1	chr1_maternal
273282	275274	276293	278285	1993	1993	100.00	705447	699930	0.28	0.28	chr1	chr1_maternal
276698	292589	278286	294177	15892	15892	100.00	705447	699930	2.25	2.27	chr1	chr1_maternal
294512	306412	294178	306078	11901	11901	100.00	705447	699930	1.69	1.70	chr1	chr1_maternal
307636	313944	306078	312386	6309	6309	100.00	705447	699930	0.89	0.90	chr1	chr1_maternal
313942	323954	313941	323953	10013	10013	100.00	705447	699930	1.42	1.43	chr1	chr1_maternal
325156	327294	323952	326090	2139	2139	100.00	705447	699930	0.30	0.31	chr1	chr1_maternal
327987	340849	326090	338952	12863	12863	100.00	705447	699930	1.82	1.84	chr1	chr1_maternal
340843	345937	340468	345562	5095	5095	100.00	705447	699930	0.72	0.73	chr1	chr1_maternal
347851	361244	345563	358956	13394	13394	100.00	705447	699930	1.90	1.91	chr1	chr1_maternal
361838	378128	358957	375247	16291	16291	100.00	705447	699930	2.31	2.33	chr1	chr1_maternal
379230	427511	375248	423529	48282	48282	99.99	705447	699930	6.84	6.90	chr1	chr1_maternal
427511	447306	424063	443858	19796	19796	100.00	705447	699930	2.81	2.83	chr1	chr1_maternal
447307	448221	445585	446499	915	915	100.00	705447	699930	0.13	0.13	chr1	chr1_maternal
449845	480347	446500	477002	30503	30503	99.99	705447	699930	4.32	4.36	chr1	chr1_maternal
480348	506343	478764	504759	25996	25996	100.00	705447	699930	3.69	3.71	chr1	chr1_maternal
507867	509684	504760	506577	1818	1818	100.00	705447	699930	0.26	0.26	chr1	chr1_maternal
509684	528583	507296	526195	18900	18900	100.00	705447	699930	2.68	2.70	chr1	chr1_maternal

SVPhaser Testing (enhanced AlleleSeq)

```
$ nucmer mutA.fasta chr1_hapA.fa
```

[S1]	[E1]		[S2]	[E2]		[LEN 1]	[LEN 2]		[% IDY]		[LEN R]	[LEN Q]		[COV R]	[COV Q]		[TAGS]
1	705447		1	706650		705447	706650		99.52		705447	706650		100.00	100.00		chr1 chr1_maternal

```
$ nucmer mutB.fasta chr1_hapB.fa
```

[S1]	[E1]		[S2]	[E2]		[LEN 1]	[LEN 2]		[% IDY]		[LEN R]	[LEN Q]		[COV R]	[COV Q]		[TAGS]
1	706496		1	707791		706496	707791		99.49		706496	707791		100.00	100.00		chr1 chr1_paternal

SVPhaser Testing (after)

```
$ nucmer mutA.fasta chr1_hapA.fa
```

[S1]	[E1]		[S2]	[E2]		[LEN 1]	[LEN 2]		[% IDY]		[LEN R]	[LEN Q]		[COV R]	[COV Q]		[TAGS]
1	705447		1	706650		705447	706650		99.52		705447	706650		100.00	100.00		chr1 chr1_maternal

```
$ nucmer mutB.fasta chr1_hapB.fa
```

[S1]	[E1]		[S2]	[E2]		[LEN 1]	[LEN 2]		[% IDY]		[LEN R]	[LEN Q]		[COV R]	[COV Q]		[TAGS]
1	706496		1	707791		706496	707791		99.49		706496	707791		100.00	100.00		chr1 chr1_paternal



One end-to-end alignment per haplotype
All SVs correctly phased and inserted



SVPhaser Testing (after)

```
$ nucmer mutA.fasta chr1_hapA.fa
```

[S1]	[E1]		[S2]	[E2]		[LEN 1]	[LEN 2]		[% IDY]		[LEN R]	[LEN Q]		[COV R]	[COV Q]		[TAGS]
1	705447		1	706650		705447	706650		99.52		705447	706650		100.00	100.00		chr1 chr1_maternal

```
$ nucmer mutB.fasta chr1_hapB.fa
```

[S1]	[E1]		[S2]	[E2]		[LEN 1]	[LEN 2]		[% IDY]		[LEN R]	[LEN Q]		[COV R]	[COV Q]		[TAGS]
1	706496		1	707791		706496	707791		99.49		706496	707791		100.00	100.00		chr1 chr1_paternal

Small amount of residual differences (0.5%)

- Nearly impossible to have 100% perfect SNP calling
- A small percent of insertions have misreported sequence
 - -> Fix is in progress

One end-to-end alignment per haplotype

All SVs correctly phased and inserted

