# Expression Analysis of Breast Cancer Cell-lines (E-GEOD-18494, GSE47533 and GSE41491)

```
Rede Alexandre (reunião 27-11-2020)
HIF1a, !VHL & !O2
p53, !Mdm2
Mdm2, p53 & !VHL
VHL, HIF1a & !p53
p300, ((p53 & HIF1a) & !VHL) | (!(p53 & HIF1a) & VHL)
BIM, !MCL 1 & !BCLXL & !BCL2
BAD, p53
BID, (!HIF1a & (p53 & VHL)) | (!MCL 1 & !BCLXL & !BCL2)
BIK, !MCL 1 & !BCLXL & !BCL2
MCL 1, HIF1a
BCLXL, HIF1a & !(p53 & VHL) & ((!Casp3 & !BAD) | (!Casp3 & BCL2))
BCL2, HIF1a & !(p53 & VHL) & ((MCL_1 & !BIM & !BIK & !BAD) | (!BIM & !BIK & BCLXL & !BAD))
IAPs, !DIABLO
BAX, (BIM \& !BCLXL) \mid (BIK \& !BCLXL \& !BCL2) \mid (BID \& !BCLXL \& !BCL2) \mid (BIM \& BID) 
BIK) | (BIM & !BCL2) | (!MCL_1 & BIM)
BAK, (!MCL_1 & BIM & !BCLXL) | (BID & !BCL2) | (BID & !BCLXL) | (!MCL_1 & BID) | (!MCL_1 &
BIK & !BCLXL) | (BIM & BID) | (BIK & BID)
DIABLO, BAX | BAK
Cyto_C, BAX | BAK
Casp9, Casp3 | (!IAPs & Cyto_C)
Casp3, !IAPs & Casp9
BCLXL?
No VHL in GSE41491
# Selected genes from HIF Axis
#hif.symbols <- c("HIF1A", "TP53", "MDM2", "VHL", "EP300", "TMBIM1", "TMBIM4", "TMBIM6", "BAD", "BIK", "MCL1
hif.symbols <- c("HIF1A", "TP53", "MDM2", "VHL", "EP300")
hif.probes <- anno.EGEOD18494$probes[anno.EGEOD18494$symbol %in% hif.symbols]
# Select the probes and genes
# EGEOD18494
expr.EGEOD18494.hif <- as.data.frame(expr.EGEOD18494) %>%
    rownames to column('probes') %>%
    filter(probes %in% hif.probes) %>%
    merge(anno.EGEOD18494[anno.EGEOD18494$symbol %in% hif.symbols, c("probes", "symbol")], by = "probes")
    mutate(., symbol=ifelse(symbol %in% c("TMBIM1", "TMBIM4", "TMBIM6"), "BIM", symbol)) %>%
    mutate(., symbol=ifelse(symbol %in% c("BIRC2", "BIRC3", "BIRC5", "BIRC6", "BIRC7"), "IAPs", symbol))
    group_by(symbol) %>%
    summarise_at(vars(-probes), funs(mean(., na.rm=TRUE))) %>%
    column_to_rownames(var = "symbol") %>%
    dplyr::select(c(data.EGEOD18494$codes[data.EGEOD18494$cell_line == "MDA-MB231 breast cancer"]))
```

```
## Warning: `funs()` is deprecated as of dplyr 0.8.0.
## Please use a list of either functions or lambdas:
##
##
    # Simple named list:
##
    list(mean = mean, median = median)
##
    # Auto named with `tibble::lst()`:
##
    tibble::1st(mean, median)
##
##
##
    # Using lambdas
    list(~ mean(., trim = .2), ~ median(., na.rm = TRUE))
## This warning is displayed once every 8 hours.
## Call `lifecycle::last_warnings()` to see where this warning was generated.
expr.EGEOD18494.hif <- expr.EGEOD18494.hif[c("HIF1A", "TP53", "MDM2", "VHL", "EP300"),]
hif.probes <- anno.GSE47533$probes[anno.GSE47533$symbol %in% hif.symbols]
# GSE47533
expr.GSE47533.hif <- as.data.frame(expr.GSE47533) %>%
 rownames_to_column('probes') %>%
 filter(probes %in% hif.probes) %>%
 merge(anno.GSE47533[anno.GSE47533$symbol %in% hif.symbols, c("probes", "symbol")], by = "probes") %>%
 mutate(., symbol=ifelse(symbol %in% c("TMBIM1","TMBIM4", "TMBIM6"), "BIM", symbol)) %>%
 mutate(., symbol=ifelse(symbol %in% c("BIRC2", "BIRC3", "BIRC5", "BIRC6", "BIRC7"), "IAPs", symbol))
 group by(symbol) %>%
 summarise_at(vars(-probes), funs(mean(., na.rm=TRUE))) %>%
 column to rownames(var = "symbol")
expr.GSE47533.hif <- expr.GSE47533.hif[c("HIF1A", "TP53", "MDM2", "VHL", "EP300"),]
hif.probes <- anno.GSE41491$probes[anno.GSE41491$symbol %in% hif.symbols]
# GSE41491
expr.GSE41491.hif <- as.data.frame(expr.GSE41491) %>%
 rownames_to_column('probes') %>%
 filter(probes %in% hif.probes) %>%
 merge(anno.GSE41491[anno.GSE41491$symbol %in% hif.symbols, c("probes", "symbol")], by = "probes") %>%
 mutate(., symbol=ifelse(symbol %in% c("TMBIM1","TMBIM4", "TMBIM6"), "BIM", symbol)) %>%
 mutate(., symbol=ifelse(symbol %in% c("BIRC2", "BIRC3", "BIRC5", "BIRC6", "BIRC7"), "IAPs", symbol))
 group_by(symbol) %>%
 column_to_rownames(var = "symbol")
write.table(expr.GSE47533.hif,"expr.GSE47533.txt", sep ="\t")
write.table(expr.EGEOD18494.hif, "expr.EGEOD18494.txt", sep ="\t")
write.table(expr.GSE41491.hif, "expr.GSE41491.txt", sep ="\t")
expr.EGEOD18494.tdm <- TDM::tdm_transform(ref_file = "expr.GSE47533.txt", file = "expr.EGEOD18494.txt")
## Attaching package: 'data.table'
## The following objects are masked from 'package:dplyr':
##
```

```
## The following object is masked from 'package:purrr':
##
##
       transpose
##
## Attaching package: 'scales'
##
  The following objects are masked from 'package:psych':
##
##
       alpha, rescale
## The following object is masked from 'package:purrr':
##
##
       discard
## The following object is masked from 'package:readr':
##
       col_factor
expr.GSE41491.tdm <- TDM::tdm_transform(ref_file = "expr.GSE47533.txt", file = "expr.GSE41491.txt")
symbols <- expr.EGEOD18494.tdm$gene
expr.EGEOD18494.tdm$gene <- NULL
expr.EGEOD18494.tdm <- as.data.frame(matrix(as.numeric(unlist(expr.EGEOD18494.tdm)),</pre>
                                              nrow = dim(expr.EGEOD18494.tdm)[1],
                                             ncol = dim(expr.EGEOD18494.tdm)[2]))
colnames(expr.EGEOD18494.tdm) <- colnames(expr.EGEOD18494.hif)</pre>
rownames(expr.EGEOD18494.tdm) <- symbols</pre>
require(BiTrinA)
## Loading required package: BiTrinA
## Loading required package: diptest
expr.GSE47533.hif.bin <- binarizeMatrix(expr.GSE47533.hif,</pre>
               method = c("BASCA"),
```

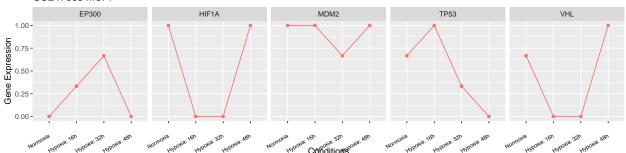
##

between, first, last

	Nor	m.0N	lorm	.0 <b>N</b> orm	.0Њуро	.1 <b>61</b> y.plo	.1 <b>6l</b> y. <b>p</b> o	.1 <b>6</b> ly. <b>β</b> о	.3 <b>21</b> y.plc	.3 <b>21</b> y.pc	.3 <b>2Н</b> у. <b>β</b> с	.4 <b>8</b> ly.фо	.4 <b>8h</b> y. <b>p</b> o	.48hr3shopdvalusymbol
TP53	3 (	0	1	1	1	1	1	0	0	1	0	0	0	9.531254.001 TP53
MDN	12	1	1	1	1	1	1	1	0	1	1	1	1	6.619755000  MDM2
VHL		1	1	0	0	0	0	0	0	0	1	1	1	9.558 <b>766</b> .780 VHL
EP30	00	0	0	0	1	0	0	0	1	1	0	0	0	9.13376 <b>6</b> .374 EP300

```
expr.GSE47533.hif.mean <- expr.GSE47533.hif.bin %>%
  mutate(Norm = rowMeans(dplyr::select(., starts_with("Norm"))),
                  Hypo.16h = rowMeans(dplyr::select(., starts_with("Hypo.16h"))),
                  Hypo.32h = rowMeans(dplyr::select(., starts_with("Hypo.32h"))),
                  Hypo.48h = rowMeans(dplyr::select(., starts_with("Hypo.48h")))) %>%
    dplyr::select(., -ends_with(c(".1",".2", ".3")))
expr.GSE47533.hif.pivot <- expr.GSE47533.hif.mean %>%
    group by(symbol) %>%
    pivot_longer(cols = starts_with(c("Norm","Hypo")), names_to = "codes", values_to = "value")
expr.GSE47533.hif.pivot$codes <- factor(expr.GSE47533.hif.pivot$codes, levels = c("Norm", "Hypo.16h"
expr.GSE47533.hif.pivot$time <- as.numeric(expr.GSE47533.hif.pivot$codes)
# hif.symbols <- c("HIF1A", "TP53", "MDM2", "VHL", "EP300", "TMBIM1", "TMBIM4", "TMBIM6", "BAD", "BIK", "MCL
p.MCF7 <- ggplot(aes(x = factor(time), y = value, group = symbol, color="red"),</pre>
                          \#data = expr. GSE47533.hif.pivot[expr. GSE47533.hif.pivot$symbol %in% c("HIF1A", "TP53", "MDM2") and the symbol %in% c("HIF1A", "TP53", "MDM2") and "HIF1A", "TP53", "MDM2", "MDM2
                         data = expr.GSE47533.hif.pivot[expr.GSE47533.hif.pivot$symbol %in% c("HIF1A", "TP53", "MDM2"
    geom point() +
    geom line() +
    scale_x_discrete(breaks = c(1, 2, 3, 4),
                                       labels = c("Normoxia", "Hypoxia: 16h", "Hypoxia: 32h", "Hypoxia: 48h")) +
    xlab("Conditions") + ylab("Gene Expression") +
    ggtitle("GSE47533 MCF7") +
    theme(legend.position = "none", axis.text.x=element_text(color = "black", size=7, angle=30, vjust=0.5
    #geom line(aes(linetype=Symbol, color=Symbol)) +
    facet_wrap(~ symbol, nrow = 1)
p.MCF7
```

#### GSE47533 MCF7



#### **EGEOD18494**

```
# expr.EGEOD18494.hif.bin <- binarizeMatrix(expr.EGEOD18494.tdm,
                 method = c("BASCA"),
#
#
                 tau = 0.15,
#
                 \#sigma = 0.9,
#
                 adjustment = "none")
expr.EGEOD18494.hif.bin <- binarizeMatrix(expr.EGEOD18494.hif)</pre>
#expr.EGEOD18494.hif.bin <- expr.EGEOD18494.tdm
expr.EGEOD18494.hif.bin$symbol <- row.names(expr.EGEOD18494.hif.bin)
row <- data.EGEOD18494$cell_line == "MDA-MB231 breast cancer"</pre>
expr.EGEOD18494.hif.bin <- expr.EGEOD18494.hif.bin[, c(as.character(data.EGEOD18494$codes[row]), c("thr
names(expr.EGEOD18494.hif.bin) <- c(paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD18
\#names(expr.EGEOD18494.hif.bin) < -c(paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD18494
# head(expr.EGEOD18494.hif.bin) %>%
# knitr::kable(.)
expr.EGEOD18494.hif.mean <- expr.EGEOD18494.hif.bin %>%
mutate(norm = rowMeans(dplyr::select(., starts_with("norm"))),
        hypo.4h = rowMeans(dplyr::select(., starts_with("hypo.4h"))),
        hypo.8h = rowMeans(dplyr::select(., starts_with("hypo.8h"))),
        hypo.12h = rowMeans(dplyr::select(., starts_with("hypo.12h")))) %>%
  dplyr::select(., -ends_with(c(".1",".2", ".3")))
expr.EGEOD18494.hif.pivot <- expr.EGEOD18494.hif.mean %>%
  group_by(symbol) %>%
  pivot_longer(cols = starts_with(c("Norm","Hypo")), names_to = "codes", values_to = "value")
expr.EGEOD18494.hif.pivot$codes <- factor(expr.EGEOD18494.hif.pivot$codes, levels = c("norm", "hypo.4
expr.EGEOD18494.hif.pivot$time <- as.numeric(expr.EGEOD18494.hif.pivot$codes)
# hif.symbols <- c("HIF1A", "TP53", "MDM2", "VHL", "EP300", "TMBIM1", "TMBIM4", "TMBIM6", "BAD", "BIK", "MCL
p.MDA <- ggplot(aes(x = factor(time), y = value, group = symbol, color="red"),</pre>
           #data = expr.EGEOD18494.hif.pivot[expr.EGEOD18494.hif.pivot$symbol %in% c("HIF1A", "TP53", "
           data = expr.EGEOD18494.hif.pivot[expr.EGEOD18494.hif.pivot$symbol %in% c("HIF1A", "TP53", "M
  geom_point() +
  geom_line() +
  scale_x_discrete(breaks = c(1, 2, 3, 4),
                 labels = c("Normoxia", "Hypoxia: 4h", "Hypoxia: 8h", "Hypoxia: 12h")) +
  xlab("Conditions") + ylab("Gene Expression") +
  ggtitle("EGEOD18494 MDA-MB231") +
```

```
theme(legend.position = "none", axis.text.x=element_text(color = "black", size=7, angle=30, vjust=.8,
              #geom_line(aes(linetype=Symbol, color=Symbol)) +
              facet_wrap(~ symbol, nrow = 1)
p.MDA
                               EGEOD18494 MDA-MB231
                                                                           EP300
                                                                                                                                                                                              HIF1A
                                                                                                                                                                                                                                                                                                                MDM2
                                                                                                                                                                                                                                                                                                                                                                                                                                    TP53
             1.00 -
0.75 - 0.50 - 0.25 - 0.25
             0.00
                                                                                                                                                                                                                                                                                                      Conditions
library(cowplot)
plot_grid(p.MDA, p.MCF7, labels = c('A', 'B'), ncol = 1)
                              EGEOD18494 MDA-MB231
                                                                            EP300
                                                                                                                                                                                                                                                                                                                MDM2
                                                                                                                                                                                                                                                                                                                                                                                                                                    TP53
              1.00
0.75 - 0.50 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.25 - 0.
             0.00
                                                                                                                                                                                                                                                                                                      Conditions
                               GSE47533 MCF7
   В
                                                                                                                                                                                              HIF1A
              1.00 -
0.75 - 0.50 - 0.25 - 0.25 -
              0.00
```

#### ${\bf Heatmaps\ \textbf{-}\ EGEOD18494}$

#### Multivariate Shapiro-Wilk normality test

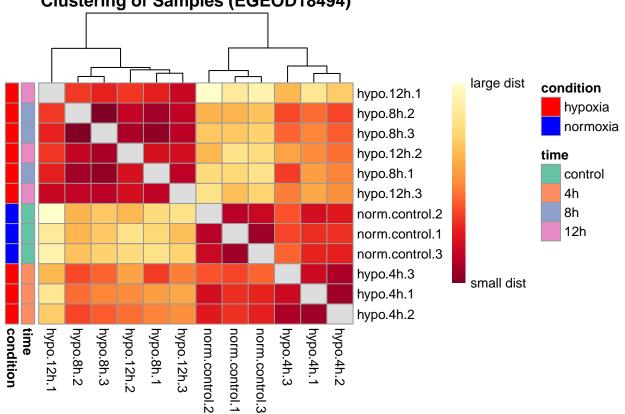
From the output, the p-value > 0.05 implying that the distribution of the data are not significantly different from normal distribution. In other words, we can assume the normality.

```
#library(rstatix)
#rstatix::mshapiro_test(expr.EGEOD18494.hif)
```

```
library("pheatmap")
library("ComplexHeatmap")
## ComplexHeatmap version 2.4.3
## Bioconductor page: http://bioconductor.org/packages/ComplexHeatmap/
## Github page: https://github.com/jokergoo/ComplexHeatmap
## Documentation: http://jokergoo.github.io/ComplexHeatmap-reference
##
## If you use it in published research, please cite:
## Gu, Z. Complex heatmaps reveal patterns and correlations in multidimensional
    genomic data. Bioinformatics 2016.
##
## This message can be suppressed by:
   suppressPackageStartupMessages(library(ComplexHeatmap))
data.EGEOD18494$time <- factor(data.EGEOD18494$time, levels = c("control", "4h", "8h", "12h"))
row <- data.EGEOD18494$cell_line == "MDA-MB231 breast cancer"</pre>
annotation_for_heatmap <- droplevels(data.frame(time = data.EGEOD18494\stime[row], condition = data.EGEOD
row.names(annotation_for_heatmap) <- paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD1
dists <- as.matrix(dist(t(expr.EGEOD18494.hif), method = "manhattan"))</pre>
rownames(dists) <- c(paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD18494$time[row],
colnames(dists) <- c(paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD18494$time[row],
hmcol <- rev(colorRampPalette(RColorBrewer::brewer.pal(9, "Y10rRd"))(255))</pre>
diag(dists) <- NA</pre>
ann colors <- list(</pre>
 time = RColorBrewer::brewer.pal(length(levels(data.EGEOD18494$time)), "Set2"),
  condition = c("red", "blue")
ann_colors
## $time
## [1] "#66C2A5" "#FC8D62" "#8DA0CB" "#E78AC3"
##
## $condition
## [1] "red" "blue"
names(ann_colors$time) <- levels(data.EGEOD18494$time)</pre>
names(ann_colors$condition) <- levels(data.EGEOD18494$condition)</pre>
pheatmap(dists, col = (hmcol),
         annotation_row = annotation_for_heatmap,
         annotation_colors = ann_colors,
         legend = TRUE,
         treeheight_row = 0,
```

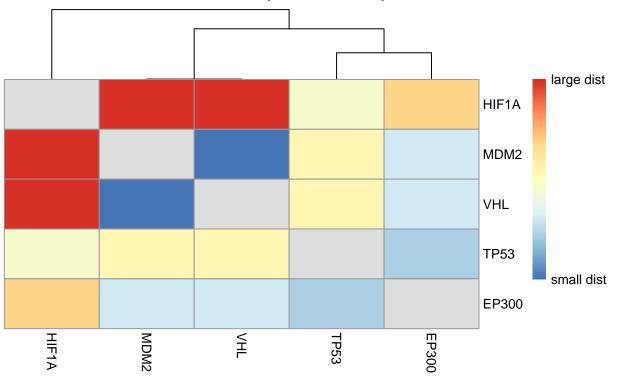
```
legend_breaks = c(min(dists, na.rm = TRUE),
                  max(dists, na.rm = TRUE)),
legend_labels = (c("small dist", "large dist")),
main = "Clustering of Samples (EGEOD18494)")
```

#### **Clustering of Samples (EGEOD18494)**



```
dists <- as.matrix(dist(expr.EGEOD18494.hif, method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.EGEOD18494.hif)</pre>
colnames(dists) <- rownames(expr.EGEOD18494.hif)</pre>
diag(dists) <- NA</pre>
pheatmap(dists, #row = (hmcol),
         #annotation_col = annotation_for_heatmap,
         #annotation_colors = ann_colors,
         legend = TRUE,
         #display_numbers = T,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression \n Euclidian Distance (EGEOD18494)")
```

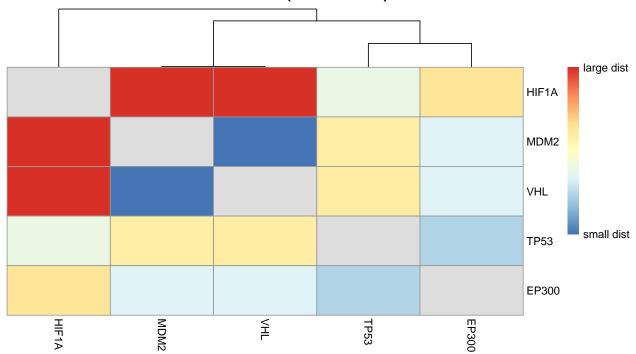
# Clustering of Gene Expression Euclidian Distance (EGEOD18494)



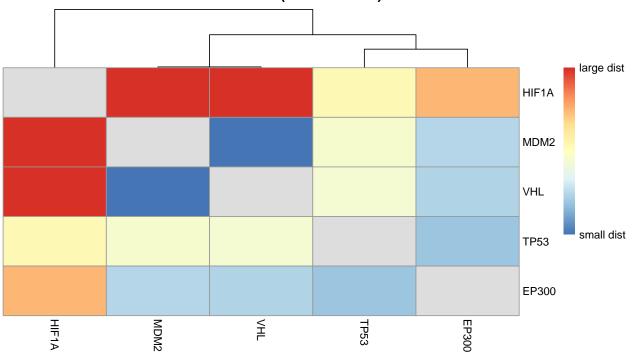
```
expr.row <- (colnames(expr.EGEOD18494.hif) %in% data.EGEOD18494$codes[data.EGEOD18494$cell_line == "MDA
dists <- as.matrix(dist(expr.EGEOD18494.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.EGEOD18494.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.EGEOD18494.hif[expr.row])</pre>
diag(dists) <- NA</pre>
p1 <- pheatmap(dists,
         legend = TRUE,
         #display_numbers = T,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                           max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Hypoxia \n Euclidian Distance (EGEOD18494)",
         silent=T)
row <- data.EGEOD18494$cell_line == "MDA-MB231 breast cancer" & data.EGEOD18494$condition == "normoxia
annotation_for_heatmap <- droplevels(data.frame(time = data.EGEOD18494\stime[row], condition = data.EGEOD
expr.row <- (colnames(expr.EGEOD18494.hif) %in% data.EGEOD18494$codes[data.EGEOD18494$cell_line == "MDA
row.names(annotation_for_heatmap) <- colnames(expr.EGEOD18494.hif[expr.row])</pre>
```

```
dists <- as.matrix(dist(expr.EGEOD18494.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.EGEOD18494.hif[expr.row])</pre>
hmcol <- rev(colorRampPalette(RColorBrewer::brewer.pal(9, "Y10rRd"))(255))</pre>
colnames(dists) <- rownames(expr.EGEOD18494.hif[expr.row])</pre>
diag(dists) <- NA</pre>
ann_colors <- list(</pre>
 time = RColorBrewer::brewer.pal(length(levels(data.EGEOD18494$time)), "Set2"),
  condition = c("#EF8A62", "#67A9CF")
ann_colors
## $time
## [1] "#66C2A5" "#FC8D62" "#8DA0CB" "#E78AC3"
##
## $condition
## [1] "#EF8A62" "#67A9CF"
names(ann_colors$time) <- levels(data.EGEOD18494$time)</pre>
names(ann_colors$condition) <- levels(data.EGEOD18494$condition)</pre>
p2 <- pheatmap(dists, #row = (hmcol),
         #annotation_col = annotation_for_heatmap,
         #annotation_colors = ann_colors,
         legend = TRUE,
         #display_numbers = T,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Normoxia \n Euclidian Distance (EGEOD18494)",
         silent=T)
gridExtra::grid.arrange(grobs=list(p1$gtable, p2$gtable),
                         nrow = 2 , labels=c('A', 'B'))
```

### Clustering of Gene Expression on Hypoxia Euclidian Distance (EGEOD18494)

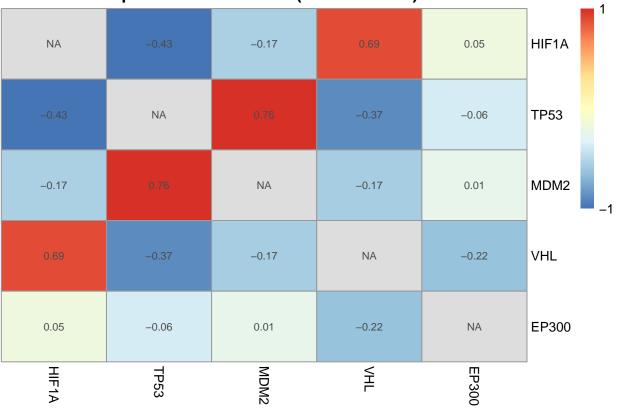


## Clustering of Gene Expression on Normoxia Euclidian Distance (EGEOD18494)

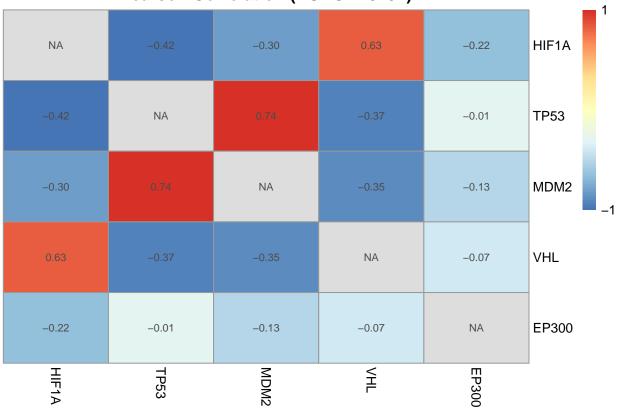


data.EGEOD18494\$time <- factor(data.EGEOD18494\$time, levels = c("control", "4h", "8h", "12h 4h"))
dists.EGEOD18494\_spearman <- cor(t(expr.EGEOD18494.hif), use = "pairwise.complete.obs", method = "spear"
rownames(dists.EGEOD18494\_spearman) <- rownames(expr.EGEOD18494.hif)</pre>

#### **Spearman Correlation (EGEOD18494)**



#### Pearson Correlation (EGEOD18494)



#### Heatmaps - GSE47533

#### Multivariate Shapiro-Wilk normality test

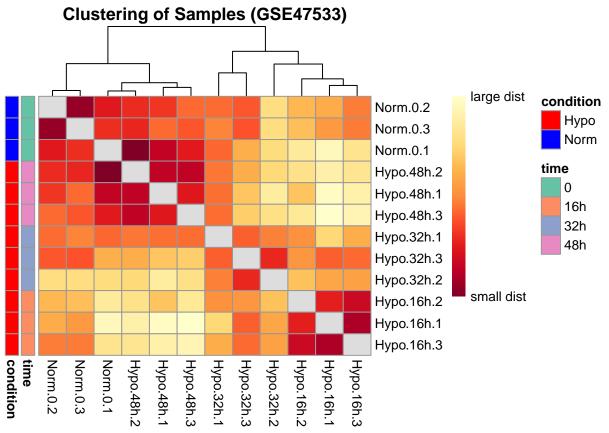
From the output, the p-value > 0.05 implying that the distribution of the data are not significantly different from normal distribution. In other words, we can assume the normality.

```
# library(rstatix)
#
# rstatix::mshapiro_test(expr.GSE47533.hif) %>%
# knitr::kable(.)

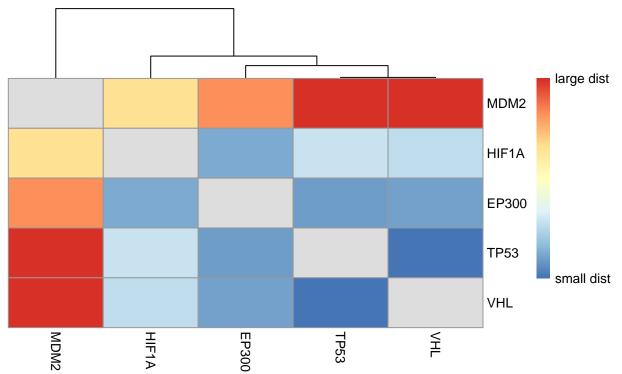
library("pheatmap")
library("ComplexHeatmap")

data.GSE47533$time <- factor(data.GSE47533$time, levels = c("0", "16h", "32h", "48h"))</pre>
```

```
annotation_for_heatmap <- droplevels(data.frame(time = data.GSE47533$time, condition = data.GSE47533$co.
row.names(annotation_for_heatmap) <- paste0(substr(data.GSE47533$condition,1,4),".", data.GSE47533$time
dists <- as.matrix(dist(t(expr.GSE47533.hif), method = "manhattan"))</pre>
rownames(dists) <- c(paste0(substr(data.GSE47533$condition,1,4),".", data.GSE47533$time, ".", data.GSE4
colnames(dists) <- c(paste0(substr(data.GSE47533$condition,1,4),".", data.GSE47533$time, ".", data.GSE4
hmcol <- rev(colorRampPalette(RColorBrewer::brewer.pal(9, "Y10rRd"))(255))</pre>
diag(dists) <- NA</pre>
ann_colors <- list(</pre>
 time = RColorBrewer::brewer.pal(length(levels(data.GSE47533$time)), "Set2"),
  condition = c("red", "blue")
ann_colors
## $time
## [1] "#66C2A5" "#FC8D62" "#8DA0CB" "#E78AC3"
## $condition
## [1] "red" "blue"
names(ann_colors$time) <- levels(data.GSE47533$time)</pre>
names(ann_colors$condition) <- levels(data.GSE47533$condition)</pre>
pheatmap(dists, col = (hmcol),
         annotation_row = annotation_for_heatmap,
         annotation_colors = ann_colors,
         legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Samples (GSE47533)")
```

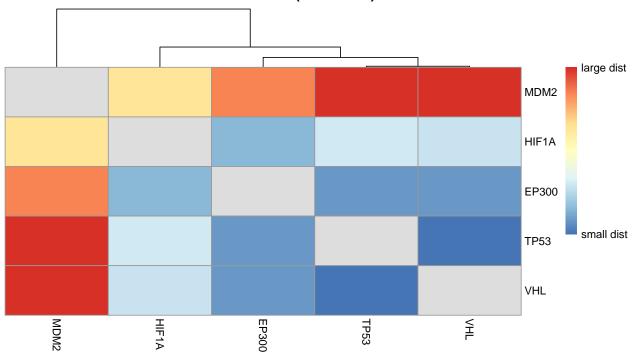


# Clustering of Gene Expression Euclidian Distance (GSE47533)

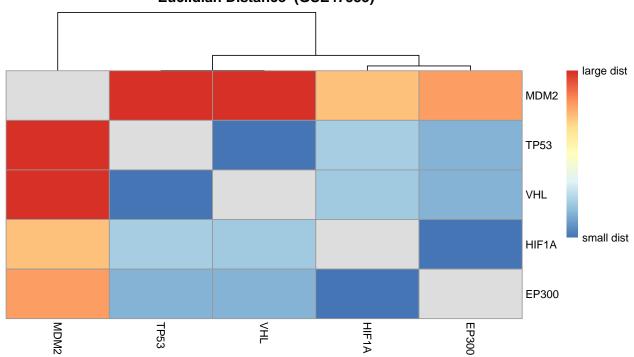


```
expr.row <- (colnames(expr.GSE47533.hif) %in% data.GSE47533$codes[data.GSE47533$condition == "Hypo"])
dists <- as.matrix(dist(expr.GSE47533.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.GSE47533.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.GSE47533.hif[expr.row])</pre>
diag(dists) <- NA</pre>
p1 <- pheatmap(dists,
         legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Hypoxia \n Euclidian Distance (GSE47533)",
         silent=T)
expr.row <- (colnames(expr.GSE47533.hif) %in% data.GSE47533$codes[data.GSE47533$condition == "Norm"])
dists <- as.matrix(dist(expr.GSE47533.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.GSE47533.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.GSE47533.hif[expr.row])</pre>
diag(dists) <- NA</pre>
```

### Clustering of Gene Expression on Hypoxia Euclidian Distance (GSE47533)

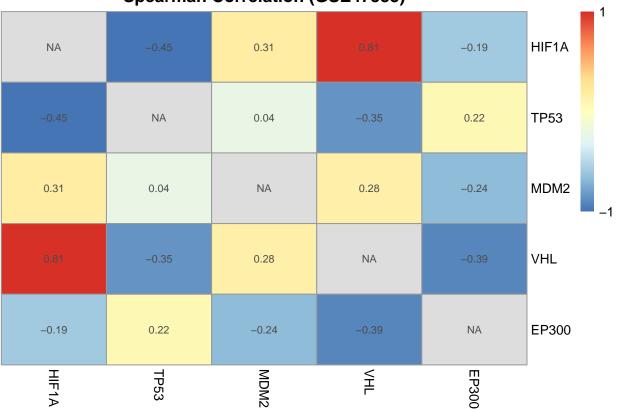


## Clustering of Gene Expression on Normoxia Euclidian Distance (GSE47533)

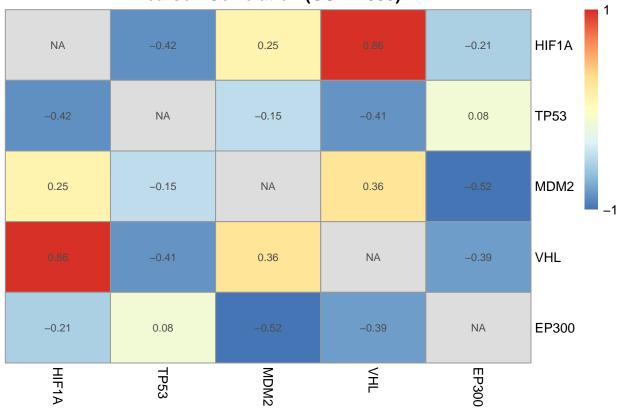


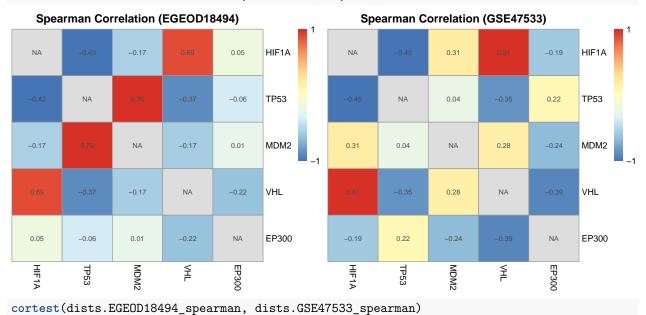
dists.GSE47533\_spearman <- cor(t(expr.GSE47533.hif), use = "pairwise.complete.obs", method = "spearman"
rownames(dists.GSE47533\_spearman) <- rownames(expr.GSE47533.hif)
colnames(dists.GSE47533\_spearman) <- rownames(expr.GSE47533.hif)
diag(dists.GSE47533\_spearman) <- NA</pre>

#### **Spearman Correlation (GSE47533)**



#### **Pearson Correlation (GSE47533)**



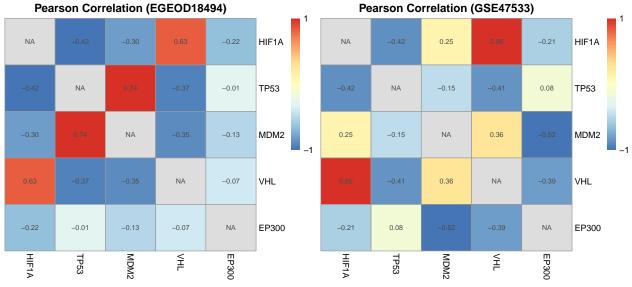


## Warning in cortest(dists.EGEOD18494\_spearman, dists.GSE47533\_spearman): n not

## specified, 100 used

## Tests of correlation matrices

## Call:cortest(R1 = dists.EGEOD18494\_spearman, R2 = dists.GSE47533\_spearman)



```
cortest(dists.EGEOD18494_pearson, dists.GSE47533_pearson)
```

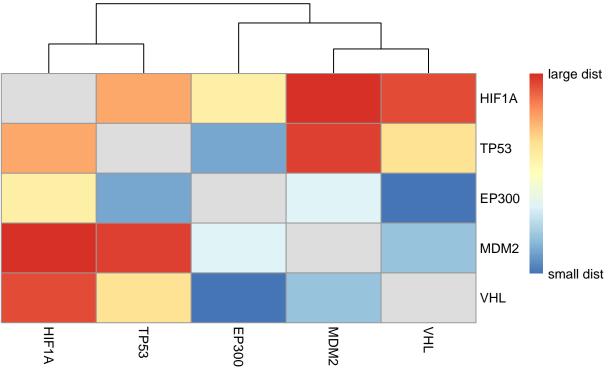
```
## Warning in cortest(dists.EGEOD18494_pearson, dists.GSE47533_pearson): n not
## specified, 100 used

## Tests of correlation matrices
## Call:cortest(R1 = dists.EGEOD18494_pearson, R2 = dists.GSE47533_pearson)
## Chi Square value 127.9 with df = 10 with probability < 1.3e-22
## z of differences = 0.4</pre>
```

## Heatmaps - All datasets Breast Cell-lines (E-GEOD-18494, GSE47533, and GSE41491)

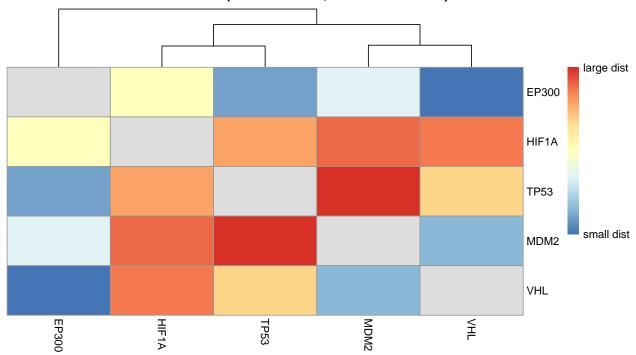
- E-GEOD-18494 2012 / MDA-MB231 / breast / 4h, 8h, 12h / microarray
- GSE41491 2012 / MCF7 / breast / 1h, 2h, 4h, 8h, 12h, 16h, 24h / microarray
- GSE47534 2014 / MCF7 / breast / normoxia, 16h, 32h, 48h / mRNA

## Clustering of Gene Expression Euclidian Distance (All 3 datasets, breast cell-lines)

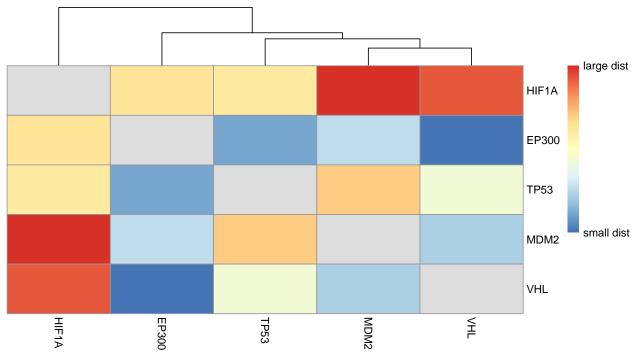


```
legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                           max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Hypoxia \n Euclidian Distance (All 3 datasets, brea
         silent=T)
col_norm <- union(data.GSE47533$codes[data.GSE47533$condition == "Norm"],</pre>
                  union(data.EGEOD18494$codes[data.EGEOD18494$condition == "normoxia"],
                         data.GSE41491$codes[data.GSE41491$condition == "no"]))
expr.row <- (colnames(expr.all.hif) %in% col_norm)</pre>
dists <- as.matrix(dist(expr.all.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.all.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.all.hif[expr.row])</pre>
diag(dists) <- NA</pre>
p2 <- pheatmap(dists,
         legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Normoxia \n Euclidian Distance (All 3 datasets, breas
         silent=T)
gridExtra::grid.arrange(grobs=list(p1$gtable, p2$gtable),
                        nrow = 2 , labels=c('A', 'B'))
```

### Clustering of Gene Expression on Hypoxia Euclidian Distance (All 3 datasets, breast cell-lines)

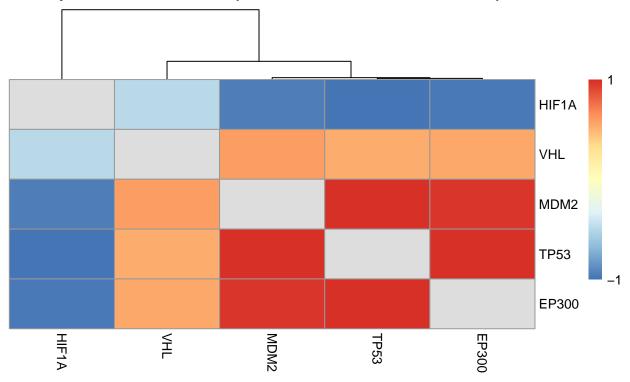


## Clustering of Gene Expression on Normoxia Euclidian Distance (All 3 datasets, breast cell-lines)



```
dists <- cor(t(expr.all.hif), use = "pairwise.complete.obs", method = "spearman")
rownames(dists) <- rownames(expr.all.hif)
colnames(dists) <- rownames(expr.all.hif)
diag(dists) <- NA</pre>
```

# Clustering of Gene Expression Spearman Correlation (All 3 datasets, breast cell-lines)



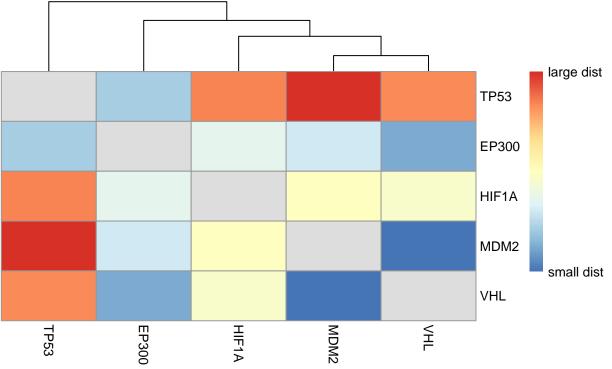
# Heatmaps - All datasets All Cell-lines (E-GEOD-18494, GSE47533, and GSE41491)

- E-GEOD-18494 2012 / HepG2, U87, MDA-MB231 / hepatoma, glioma, breast / 4h, 8h, 12h / microarray
- GSE41491 2012 / DU145, HT29, MCF7 / prostate, colon, breast / 1h, 2h, 4h, 8h, 12h, 16h, 24h / microarray
- GSE47534 2014 / MCF7 / breast / normoxia, 16h, 32h, 48h / mRNA

```
# Imput the mean of all VHL values
mean.vhl <- mean(unlist(expr.GSE47533.hif["VHL",], expr.EGEOD18494.hif["VHL",]))
expr.GSE41491.hif["VHL",] <- rep(mean.vhl, 24)

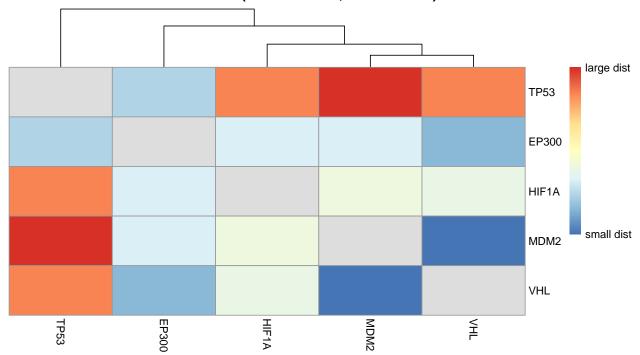
expr.all.hif <- cbind(expr.GSE47533.hif, expr.EGEOD18494.hif, expr.GSE41491.hif)</pre>
```

## Clustering of Gene Expression Euclidian Distance (All 3 datasets, all cell-lines)

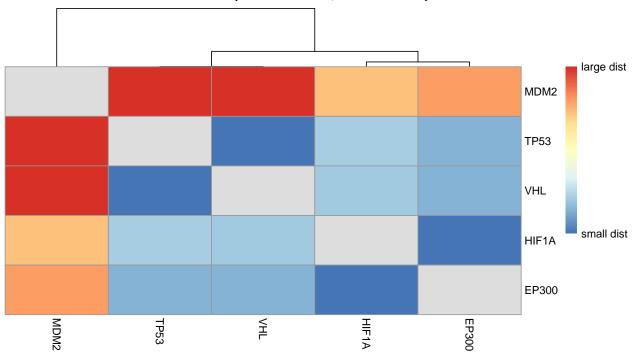


```
legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                           max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Hypoxia \n Euclidian Distance (All 3 datasets, all
         silent=T)
expr.row <- (colnames(expr.all.hif) %in% data.GSE47533$codes[data.GSE47533$condition == "Norm"])
dists <- as.matrix(dist(expr.all.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.all.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.all.hif[expr.row])</pre>
diag(dists) <- NA</pre>
p2 <- pheatmap(dists,</pre>
         legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                           max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Normoxia \n Euclidian Distance (All 3 datasets, all c
         silent=T)
gridExtra::grid.arrange(grobs=list(p1$gtable, p2$gtable),
                        nrow = 2 , labels=c('A', 'B'))
```

### Clustering of Gene Expression on Hypoxia Euclidian Distance (All 3 datasets, all cell-lines)



### Clustering of Gene Expression on Normoxia Euclidian Distance (All 3 datasets, all cell-lines)



```
dists <- cor(t(expr.all.hif), use = "pairwise.complete.obs", method = "spearman")
rownames(dists) <- rownames(expr.all.hif)
colnames(dists) <- rownames(expr.all.hif)
diag(dists) <- NA</pre>
```

# Clustering of Gene Expression Spearman Correlation (All 3 datasets, all cell-lines)

