Expression Analysis of Breast Cancer Cell-lines (E-GEOD-18494, GSE47533 and GSE41491)

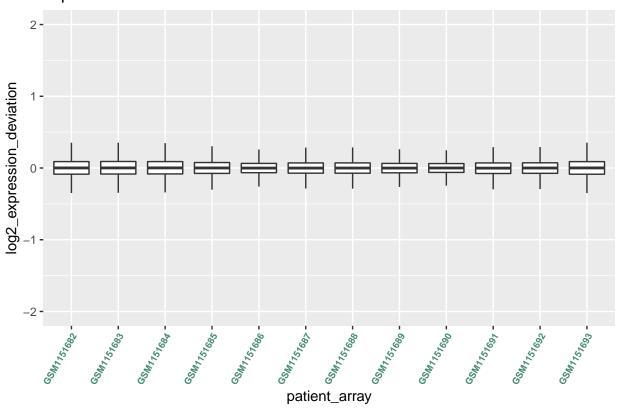
```
Rede Alexandre (reunião 27-11-2020)
HIF1a, !VHL & !O2
p53, !Mdm2
Mdm2, p53 & !VHL
VHL, HIF1a & !p53
p300, ((p53 & HIF1a) & !VHL) | (!(p53 & HIF1a) & VHL)
BIM, !MCL 1 & !BCLXL & !BCL2
BAD, p53
BID, (!HIF1a & (p53 & VHL)) | (!MCL 1 & !BCLXL & !BCL2)
BIK, !MCL 1 & !BCLXL & !BCL2
MCL 1, HIF1a
BCLXL, HIF1a & !(p53 & VHL) & ((!Casp3 & !BAD) | (!Casp3 & BCL2))
BCL2, HIF1a & !(p53 & VHL) & ((MCL_1 & !BIM & !BIK & !BAD) | (!BIM & !BIK & BCLXL & !BAD))
IAPs, !DIABLO
BAX, (BIM \& !BCLXL) \mid (BIK \& !BCLXL \& !BCL2) \mid (BID \& !BCLXL \& !BCL2) \mid (BIM \& BID) 
BIK) | (BIM & !BCL2) | (!MCL_1 & BIM)
BAK, (!MCL_1 & BIM & !BCLXL) | (BID & !BCL2) | (BID & !BCLXL) | (!MCL_1 & BID) | (!MCL_1 &
BIK & !BCLXL) | (BIM & BID) | (BIK & BID)
DIABLO, BAX | BAK
Cyto_C, BAX | BAK
Casp9, Casp3 | (!IAPs & Cyto_C)
Casp3, !IAPs & Casp9
BCLXL?
No VHL in GSE41491
# Selected genes from HIF Axis
#hif.symbols <- c("HIF1A", "TP53", "MDM2", "VHL", "EP300", "TMBIM1", "TMBIM4", "TMBIM6", "BAD", "BIK", "MCL1
hif.symbols <- c("HIF1A", "TP53", "MDM2", "VHL", "EP300")
hif.probes <- anno.EGEOD18494$probes[anno.EGEOD18494$symbol %in% hif.symbols]
# Select the probes and genes
# EGEOD18494
expr.EGEOD18494.hif <- as.data.frame(expr.EGEOD18494) %>%
    rownames to column('probes') %>%
    filter(probes %in% hif.probes) %>%
    merge(anno.EGEOD18494[anno.EGEOD18494$symbol %in% hif.symbols, c("probes", "symbol")], by = "probes")
    mutate(., symbol=ifelse(symbol %in% c("TMBIM1", "TMBIM4", "TMBIM6"), "BIM", symbol)) %>%
    mutate(., symbol=ifelse(symbol %in% c("BIRC2", "BIRC3", "BIRC5", "BIRC6", "BIRC7"), "IAPs", symbol))
    group_by(symbol) %>%
    summarise_at(vars(-probes), funs(mean(., na.rm=TRUE))) %>%
    column_to_rownames(var = "symbol") %>%
    dplyr::select(c(data.EGEOD18494$codes[data.EGEOD18494$cell_line == "MDA-MB231 breast cancer"]))
```

```
## Warning: `funs()` is deprecated as of dplyr 0.8.0.
## Please use a list of either functions or lambdas:
##
##
    # Simple named list:
##
    list(mean = mean, median = median)
##
    # Auto named with `tibble::lst()`:
##
    tibble::lst(mean, median)
##
##
##
    # Using lambdas
    list(~ mean(., trim = .2), ~ median(., na.rm = TRUE))
## This warning is displayed once every 8 hours.
## Call `lifecycle::last_warnings()` to see where this warning was generated.
hif.probes <- anno.GSE47533$probes[anno.GSE47533$symbol %in% hif.symbols]
# GSE47533
expr.GSE47533.hif <- as.data.frame(expr.GSE47533) %>%
 rownames_to_column('probes') %>%
 filter(probes %in% hif.probes) %>%
 merge(anno.GSE47533[anno.GSE47533$symbol %in% hif.symbols, c("probes", "symbol")], by = "probes") %>%
 mutate(., symbol=ifelse(symbol %in% c("TMBIM1","TMBIM4", "TMBIM6"), "BIM", symbol)) %>%
 mutate(., symbol=ifelse(symbol %in% c("BIRC2", "BIRC3", "BIRC5", "BIRC6", "BIRC7"), "IAPs", symbol))
 group_by(symbol) %>%
 column to rownames(var = "symbol")
hif.probes <- anno.GSE41491$probes[anno.GSE41491$symbol %in% hif.symbols]
# GSE41491
expr.GSE41491.hif <- as.data.frame(expr.GSE41491) %>%
 rownames_to_column('probes') %>%
 filter(probes %in% hif.probes) %>%
 merge(anno.GSE41491[anno.GSE41491$symbol %in% hif.symbols, c("probes", "symbol")], by = "probes") %>%
 mutate(., symbol=ifelse(symbol %in% c("TMBIM1","TMBIM4", "TMBIM6"), "BIM", symbol)) %>%
 mutate(., symbol=ifelse(symbol %in% c("BIRC2", "BIRC3", "BIRC5", "BIRC6", "BIRC7"), "IAPs", symbol))
 group_by(symbol) %>%
 summarise_at(vars(-probes), funs(mean(., na.rm=TRUE))) %>%
 column_to_rownames(var = "symbol")
write.table(expr.GSE47533.hif,"expr.GSE47533.txt", sep ="\t")
write.table(expr.EGEOD18494.hif, "expr.EGEOD18494.txt", sep ="\t")
write.table(expr.GSE41491.hif, "expr.GSE41491.txt", sep ="\t")
expr.EGEOD18494.tdm <- TDM::tdm_transform(ref_file = "expr.GSE47533.txt", file = "expr.EGEOD18494.txt")
##
## Attaching package: 'data.table'
## The following objects are masked from 'package:dplyr':
##
##
      between, first, last
## The following object is masked from 'package:purrr':
##
```

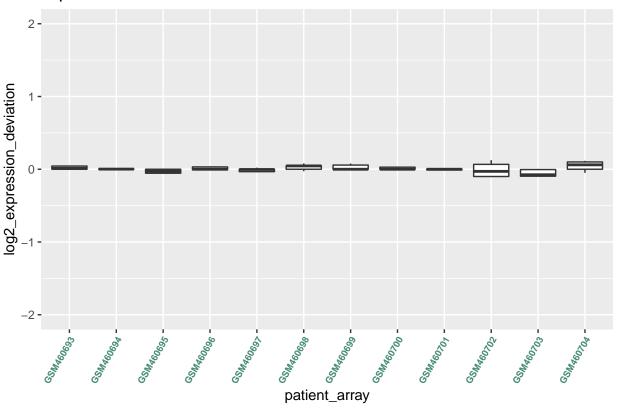
```
##
       transpose
##
## Attaching package: 'scales'
## The following object is masked from 'package:purrr':
##
##
       discard
## The following object is masked from 'package:readr':
##
##
       col_factor
expr.GSE41491.tdm <- TDM::tdm_transform(ref_file = "expr.GSE47533.txt", file = "expr.GSE41491.txt")
symbols <- expr.EGEOD18494.tdm$gene
expr.EGEOD18494.tdm$gene <- NULL
expr.EGEOD18494.tdm <- as.data.frame(matrix(as.numeric(unlist(expr.EGEOD18494.tdm)),</pre>
                                             nrow = dim(expr.EGEOD18494.tdm)[1],
                                             ncol = dim(expr.EGEOD18494.tdm)[2]))
colnames(expr.EGEOD18494.tdm) <- colnames(expr.EGEOD18494.hif)</pre>
rownames(expr.EGEOD18494.tdm) <- symbols</pre>
row medians assayData <-
 Biobase::rowMedians(as.matrix(expr.GSE47533))
RLE_data <- sweep(expr.GSE47533, 1, row_medians_assayData)</pre>
RLE_data <- as.data.frame(RLE_data)</pre>
RLE data gathered <-
 tidyr::gather(RLE_data, patient_array, log2_expression_deviation)
ggplot2::ggplot(RLE_data_gathered, aes(patient_array,
                                        log2_expression_deviation)) +
  geom_boxplot(outlier.shape = NA) +
 ylim(c(-2, 2)) +
  ggtitle("expr.GSE47533") +
  theme(axis.text.x = element_text(colour = "aquamarine4",
                                   angle = 60, size = 6.5, hjust = 1,
                                   face = "bold"))
```

Warning: Removed 142 rows containing non-finite values (stat_boxplot).

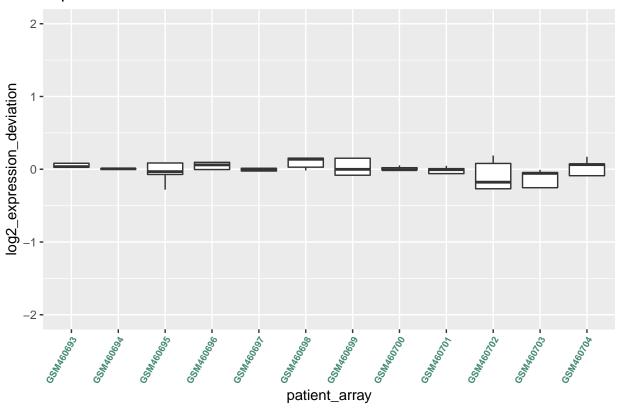
expr.GSE47533



expr.EGEOD18494.tdm

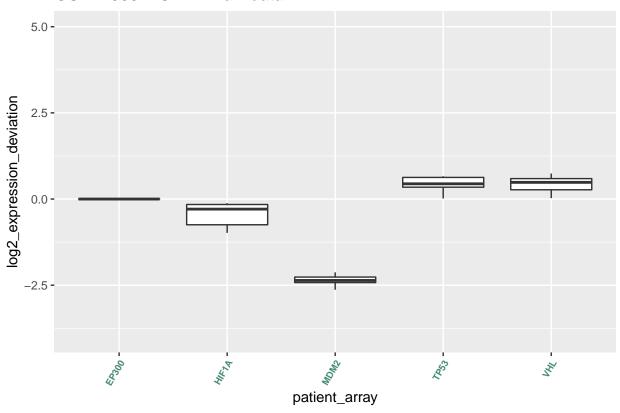


expr.EGEOD18494.hif

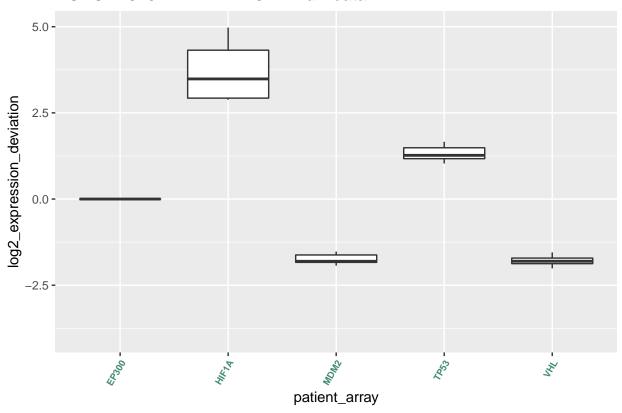


```
rm(RLE_data, RLE_data_gathered, row_medians_assayData)
```

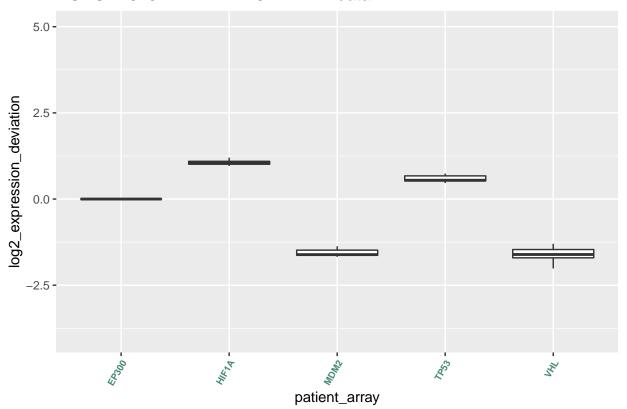
GSE47533 MCF7 - Raw data



EGEOD18494 MDA-MB231 - Raw data



EGEOD18494 MDA-MB231 - TDM data



rm(RLE_data, RLE_data_gathered, row_medians_assayData)

require(BiTrinA)

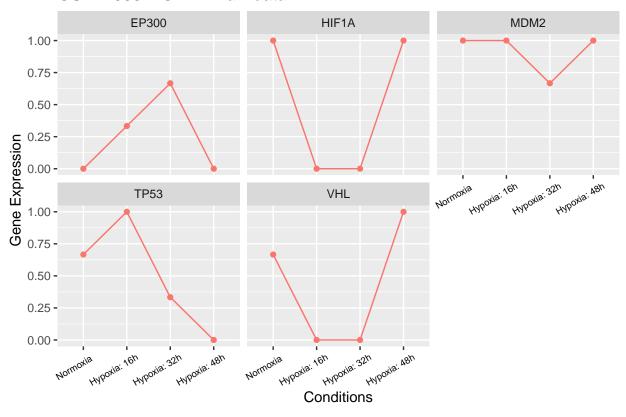
Loading required package: BiTrinA
Loading required package: diptest

Norm	.0 N orm.	0 № orm.	оњуро.	1 6Н у.фо.	1 6l y. p o.	.1 Ы у. β о.	3 21 y.plo	.3 24 y. p o	.3 21 у. β о	.4 81 y. p o.	.4 8h y. p o	.481hr3shopdvalueymbol
EP300 0	0	0	1	0	0	0	1	1	0	0	0	9.13376 6 .386 EP300
HIF1A 1	1	1	0	0	0	0	0	0	1	1	1	8.699330.001 HIF1A

Norm.0Norm.0Norm.0Hypo.16Hypo.16Hypo.16Hypo.16Hypo.3Hypo.3Hypo.3Hypo.3Hypo.4HHypo.4HHypo.4Hhr6shopdvalueymbol													
MDM2	2 1	1	1	1	1	1	1	0	1	1	1	1	6.619755000 MDM2
TP53	0	1	1	1	1	1	0	0	1	0	0	0	9.531254.001 TP53
VHL	1	1	0	0	0	0	0	0	0	1	1	1	9.55876 6 .806 VHL

```
expr.GSE47533.hif.mean <- expr.GSE47533.hif.bin %>%
  mutate(Norm = rowMeans(dplyr::select(., starts_with("Norm"))),
                  Hypo.16h = rowMeans(dplyr::select(., starts_with("Hypo.16h"))),
                  Hypo.32h = rowMeans(dplyr::select(., starts_with("Hypo.32h"))),
                  Hypo.48h = rowMeans(dplyr::select(., starts_with("Hypo.48h")))) %>%
    dplyr::select(., -ends_with(c(".1",".2", ".3")))
expr.GSE47533.hif.pivot <- expr.GSE47533.hif.mean %>%
    group_by(symbol) %>%
    pivot_longer(cols = starts_with(c("Norm","Hypo")), names_to = "codes", values_to = "value")
expr.GSE47533.hif.pivot$codes <- factor(expr.GSE47533.hif.pivot$codes, levels = c("Norm", "Hypo.16h"
expr.GSE47533.hif.pivot$time <- as.numeric(expr.GSE47533.hif.pivot$codes)
# hif.symbols <- c("HIF1A", "TP53", "MDM2", "VHL", "EP300", "TMBIM1", "TMBIM4", "TMBIM6", "BAD", "BIK", "MCL
p.MCF7 <- ggplot(aes(x = factor(time), y = value, group = symbol, color="red"),</pre>
                          \#data = expr.GSE47533.hif.pivot[expr.GSE47533.hif.pivot$symbol %in% c("HIF1A", "TP53", "MDM2") and the substitution of the s
                         data = expr.GSE47533.hif.pivot[expr.GSE47533.hif.pivot$symbol %in% c("HIF1A", "TP53", "MDM2"
    geom_point() +
    geom line() +
    scale_x_discrete(breaks = c(1, 2, 3, 4),
                                       labels = c("Normoxia", "Hypoxia: 16h", "Hypoxia: 32h", "Hypoxia: 48h")) +
    xlab("Conditions") + ylab("Gene Expression") +
    ggtitle("GSE47533 MCF7 - Raw data") +
    theme(legend.position = "none", axis.text.x=element text(color = "black", size=7, angle=30, vjust=.8,
    #geom_line(aes(linetype=Symbol, color=Symbol)) +
    facet wrap(~ symbol)
p.MCF7
```

GSE47533 MCF7 - Raw data

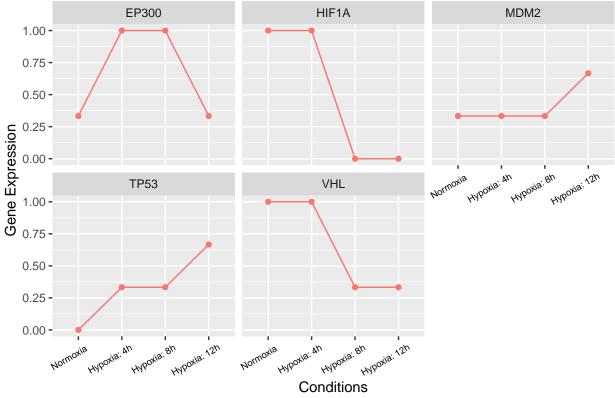


EGEOD18494

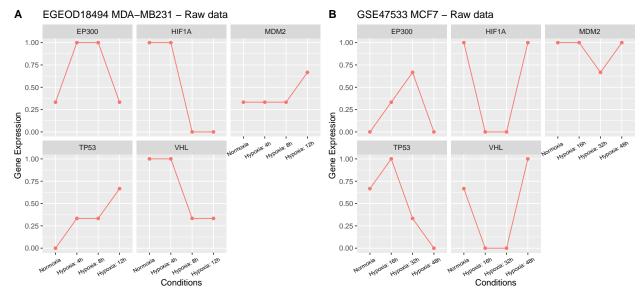
```
# expr.EGEOD18494.hif.bin <- binarizeMatrix(expr.EGEOD18494.tdm,
                   method = c("BASCA"),
#
#
                    tau = 0.15,
#
                    \#sigma = 0.9,
                    adjustment = "none")
expr.EGEOD18494.hif.bin <- binarizeMatrix(expr.EGEOD18494.hif)</pre>
#expr.EGEOD18494.hif.bin <- expr.EGEOD18494.tdm</pre>
expr.EGEOD18494.hif.bin$symbol <- row.names(expr.EGEOD18494.hif.bin)</pre>
row <- data.EGEOD18494$cell_line == "MDA-MB231 breast cancer"</pre>
expr.EGEOD18494.hif.bin <- expr.EGEOD18494.hif.bin[, c(as.character(data.EGEOD18494$codes[row]), c("thr
names(expr.EGEOD18494.hif.bin) <- c(paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD18</pre>
\#names(expr.EGEOD18494.hif.bin) \leftarrow c(pasteO(substr(data.EGEOD18494\$condition[row],1,4),".", data.EGEOD18494\$condition[row],1,4),".", data.EGEOD18494\$condition[row],1,4),"."
# head(expr.EGEOD18494.hif.bin) %>%
    knitr::kable(.)
```

```
expr.EGEOD18494.hif.mean <- expr.EGEOD18494.hif.bin %>%
 mutate(norm = rowMeans(dplyr::select(., starts_with("norm"))),
       hypo.4h = rowMeans(dplyr::select(., starts_with("hypo.4h"))),
       hypo.8h = rowMeans(dplyr::select(., starts_with("hypo.8h"))),
       hypo.12h = rowMeans(dplyr::select(., starts_with("hypo.12h")))) %>%
  dplyr::select(., -ends_with(c(".1",".2", ".3")))
expr.EGEOD18494.hif.pivot <- expr.EGEOD18494.hif.mean %>%
  group_by(symbol) %>%
 pivot_longer(cols = starts_with(c("Norm", "Hypo")), names_to = "codes", values_to = "value")
expr.EGEOD18494.hif.pivot$codes <- factor(expr.EGEOD18494.hif.pivot$codes, levels = c("norm", "hypo.4
expr.EGEOD18494.hif.pivot$time <- as.numeric(expr.EGEOD18494.hif.pivot$codes)
# hif.symbols <- c("HIF1A", "TP53", "MDM2", "VHL", "EP300", "TMBIM1", "TMBIM4", "TMBIM6", "BAD", "BIK", "MCL
p.MDA <- ggplot(aes(x = factor(time), y = value, group = symbol, color="red"),</pre>
           #data = expr.EGEOD18494.hif.pivot[expr.EGEOD18494.hif.pivot$symbol %in% c("HIF1A", "TP53", "
           data = expr.EGEOD18494.hif.pivot[expr.EGEOD18494.hif.pivot$symbol %in% c("HIF1A", "TP53", "M
  geom_point() +
  geom_line() +
  scale_x_discrete(breaks = c(1, 2, 3, 4),
                 labels = c("Normoxia", "Hypoxia: 4h", "Hypoxia: 8h", "Hypoxia: 12h")) +
  xlab("Conditions") + ylab("Gene Expression") +
  ggtitle("EGEOD18494 MDA-MB231 - Raw data") +
  theme(legend.position = "none", axis.text.x=element_text(color = "black", size=7, angle=30, vjust=.8,
  #geom_line(aes(linetype=Symbol, color=Symbol)) +
  facet_wrap(~ symbol)
p.MDA
```

EGEOD18494 MDA-MB231 - Raw data



library(cowplot)
plot_grid(p.MDA, p.MCF7, labels = c('A', 'B'))



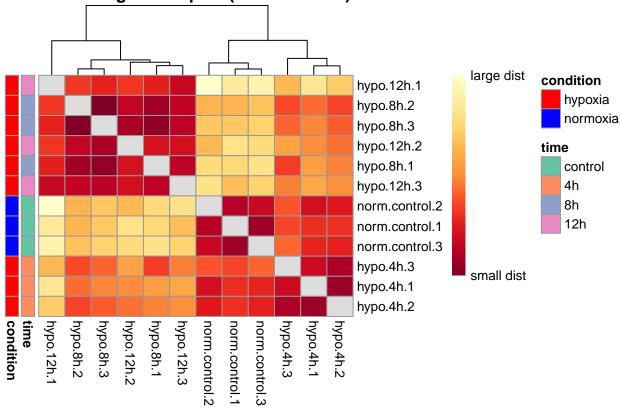
Heatmaps - EGEOD18494

Multivariate Shapiro-Wilk normality test

From the output, the p-value > 0.05 implying that the distribution of the data are not significantly different from normal distribution. In other words, we can assume the normality.

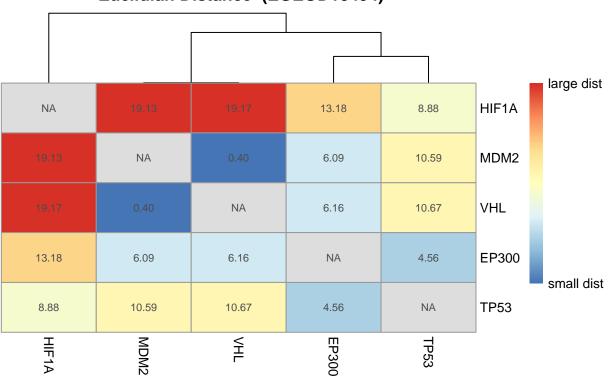
```
#library(rstatix)
#rstatix::mshapiro_test(expr.EGEOD18494.hif)
library("pheatmap")
library("ComplexHeatmap")
## =============
## ComplexHeatmap version 2.4.3
## Bioconductor page: http://bioconductor.org/packages/ComplexHeatmap/
## Github page: https://github.com/jokergoo/ComplexHeatmap
## Documentation: http://jokergoo.github.io/ComplexHeatmap-reference
##
## If you use it in published research, please cite:
## Gu, Z. Complex heatmaps reveal patterns and correlations in multidimensional
##
     genomic data. Bioinformatics 2016.
##
## This message can be suppressed by:
     suppressPackageStartupMessages(library(ComplexHeatmap))
data.EGEOD18494$time <- factor(data.EGEOD18494$time, levels = c("control", "4h", "8h", "12h"))
row <- data.EGEOD18494$cell_line == "MDA-MB231 breast cancer"</pre>
annotation_for_heatmap <- droplevels(data.frame(time = data.EGEOD18494\stime[row], condition = data.EGEOD
row.names(annotation_for_heatmap) <- paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD1
dists <- as.matrix(dist(t(expr.EGEOD18494.hif), method = "manhattan"))</pre>
rownames(dists) <- c(paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD18494$time[row],
colnames(dists) <- c(paste0(substr(data.EGEOD18494$condition[row],1,4),".", data.EGEOD18494$time[row],</pre>
hmcol <- rev(colorRampPalette(RColorBrewer::brewer.pal(9, "Y10rRd"))(255))</pre>
diag(dists) <- NA</pre>
ann_colors <- list(</pre>
 time = RColorBrewer::brewer.pal(length(levels(data.EGEOD18494$time)), "Set2"),
  condition = c("red", "blue")
ann_colors
## $time
## [1] "#66C2A5" "#FC8D62" "#8DA0CB" "#E78AC3"
##
```

Clustering of Samples (EGEOD18494)



```
max(dists, na.rm = TRUE)),
legend_labels = (c("small dist", "large dist")),
main = "Clustering of Gene Expression \n Euclidian Distance (EGEOD18494)")
```

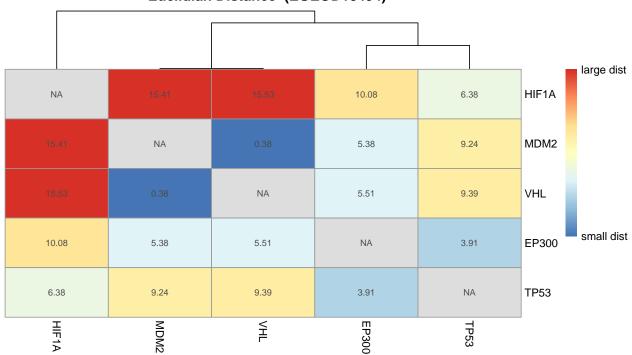
Clustering of Gene Expression Euclidian Distance (EGEOD18494)



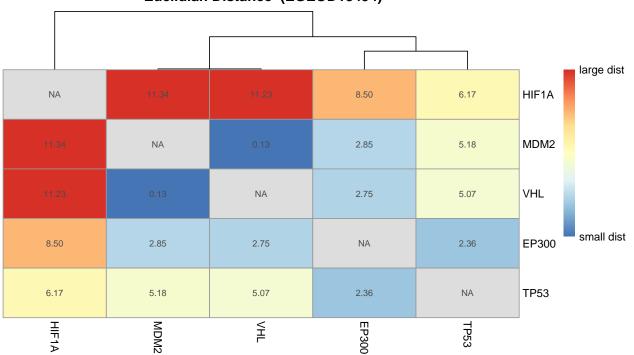
```
expr.row <- (colnames(expr.EGEOD18494.hif) %in% data.EGEOD18494$codes[data.EGEOD18494$cell_line == "MDA
dists <- as.matrix(dist(expr.EGEOD18494.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.EGEOD18494.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.EGEOD18494.hif[expr.row])</pre>
diag(dists) <- NA</pre>
p1 <- pheatmap(dists,
         legend = TRUE,
         display_numbers = T,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                           max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Hypoxia \n Euclidian Distance (EGEOD18494)",
         silent=T)
row <- data.EGEOD18494$cell_line == "MDA-MB231 breast cancer" & data.EGEOD18494$condition == "normoxia
annotation_for_heatmap <- droplevels(data.frame(time = data.EGEOD18494$time[row], condition = data.EGEOD
```

```
expr.row <- (colnames(expr.EGEOD18494.hif) %in% data.EGEOD18494$codes[data.EGEOD18494$cell_line == "MDA
row.names(annotation for heatmap) <- colnames(expr.EGEOD18494.hif[expr.row])
dists <- as.matrix(dist(expr.EGEOD18494.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.EGEOD18494.hif[expr.row])</pre>
hmcol <- rev(colorRampPalette(RColorBrewer::brewer.pal(9, "Y10rRd"))(255))</pre>
colnames(dists) <- rownames(expr.EGEOD18494.hif[expr.row])</pre>
diag(dists) <- NA</pre>
ann_colors <- list(</pre>
 time = RColorBrewer::brewer.pal(length(levels(data.EGEOD18494$time)), "Set2"),
  condition = c("#EF8A62", "#67A9CF")
ann_colors
## $time
## [1] "#66C2A5" "#FC8D62" "#8DA0CB" "#E78AC3"
## $condition
## [1] "#EF8A62" "#67A9CF"
names(ann_colors$time) <- levels(data.EGEOD18494$time)</pre>
names(ann_colors$condition) <- levels(data.EGEOD18494$condition)</pre>
p2 <- pheatmap(dists, #row = (hmcol),
         #annotation_col = annotation_for_heatmap,
         #annotation_colors = ann_colors,
         legend = TRUE,
         display_numbers = T,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Normoxia \n Euclidian Distance (EGEOD18494)",
         silent=T)
gridExtra::grid.arrange(grobs=list(p1$gtable, p2$gtable),
                         nrow = 2 , labels=c('A', 'B'))
```

Clustering of Gene Expression on Hypoxia Euclidian Distance (EGEOD18494)

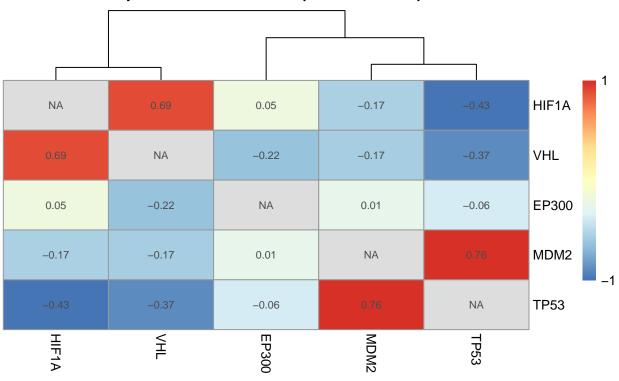


Clustering of Gene Expression on Normoxia Euclidian Distance (EGEOD18494)

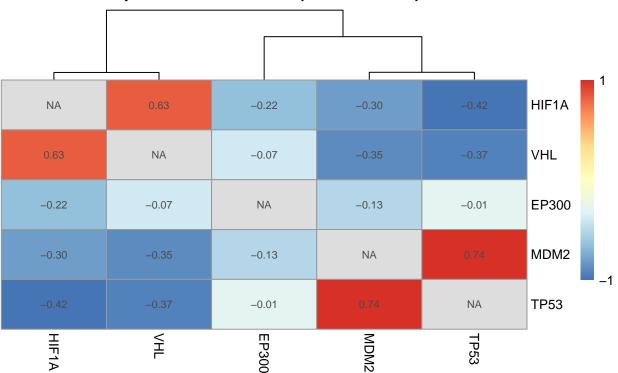


data.EGEOD18494\$time <- factor(data.EGEOD18494\$time, levels = c("control", "4h", "8h", "12h 4h"))
dists <- cor(t(expr.EGEOD18494.hif), use = "pairwise.complete.obs", method = "spearman")
rownames(dists) <- rownames(expr.EGEOD18494.hif)</pre>

Clustering of Gene Expression Spearman Correlation (EGEOD18494)



Clustering of Gene Expression Spearman Correlation (EGEOD18494)



Heatmaps - GSE47533

Multivariate Shapiro-Wilk normality test

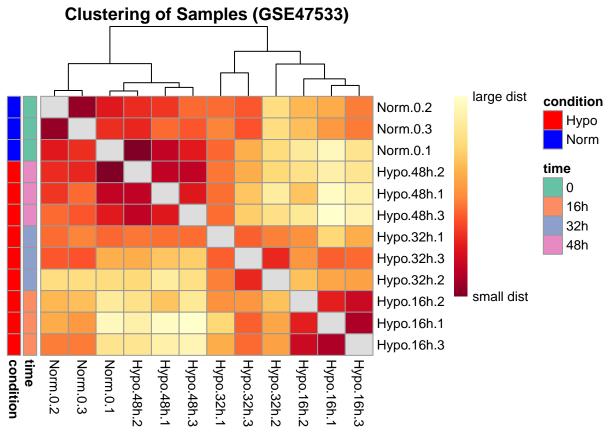
From the output, the p-value > 0.05 implying that the distribution of the data are not significantly different from normal distribution. In other words, we can assume the normality.

```
# library(rstatix)
#
# rstatix::mshapiro_test(expr.GSE47533.hif) %>%
# knitr::kable(.)

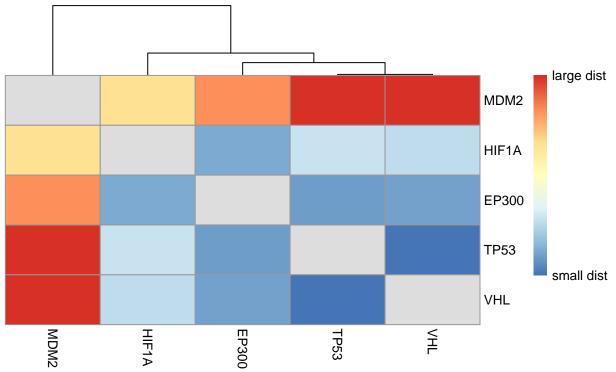
library("pheatmap")
library("ComplexHeatmap")

data.GSE47533$time <- factor(data.GSE47533$time, levels = c("0", "16h", "32h", "48h"))
annotation_for_heatmap <- droplevels(data.frame(time = data.GSE47533$time, condition = data.GSE47533$condition_for_heatmap) <- pasteO(substr(data.GSE47533$condition,1,4),".", data.GSE47533$time</pre>
```

```
dists <- as.matrix(dist(t(expr.GSE47533.hif), method = "manhattan"))</pre>
rownames(dists) <- c(paste0(substr(data.GSE47533$condition,1,4),".", data.GSE47533$time, ".", data.GSE4
colnames(dists) <- c(paste0(substr(data.GSE47533$condition,1,4),".", data.GSE47533$time, ".", data.GSE4
hmcol <- rev(colorRampPalette(RColorBrewer::brewer.pal(9, "Y10rRd"))(255))</pre>
diag(dists) <- NA</pre>
ann_colors <- list(</pre>
 time = RColorBrewer::brewer.pal(length(levels(data.GSE47533$time)), "Set2"),
 condition = c("red", "blue")
ann_colors
## $time
## [1] "#66C2A5" "#FC8D62" "#8DA0CB" "#E78AC3"
##
## $condition
## [1] "red" "blue"
names(ann_colors$time) <- levels(data.GSE47533$time)</pre>
names(ann_colors$condition) <- levels(data.GSE47533$condition)</pre>
pheatmap(dists, col = (hmcol),
         annotation_row = annotation_for_heatmap,
         annotation_colors = ann_colors,
         legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Samples (GSE47533)")
```

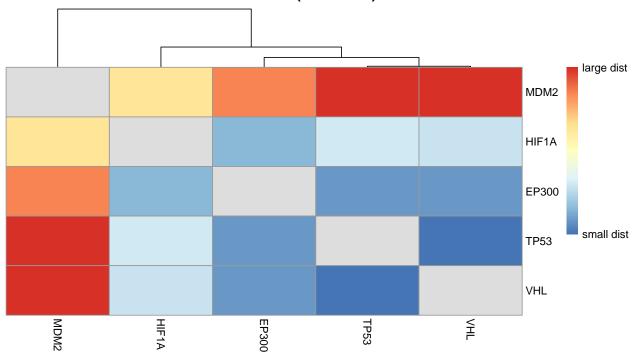


Clustering of Gene Expression Euclidian Distance (GSE47533)

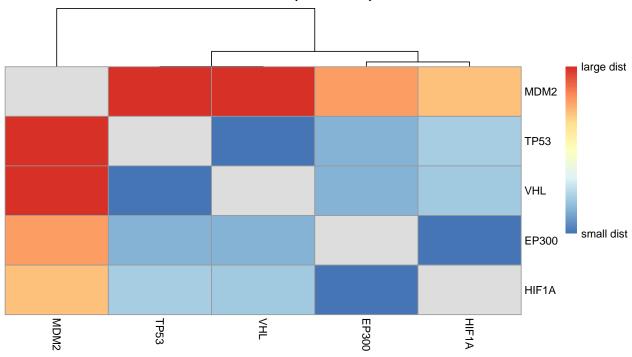


```
expr.row <- (colnames(expr.GSE47533.hif) %in% data.GSE47533$codes[data.GSE47533$condition == "Hypo"])
dists <- as.matrix(dist(expr.GSE47533.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.GSE47533.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.GSE47533.hif[expr.row])</pre>
diag(dists) <- NA</pre>
p1 <- pheatmap(dists,
         legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Hypoxia \n Euclidian Distance (GSE47533)",
         silent=T)
expr.row <- (colnames(expr.GSE47533.hif) %in% data.GSE47533$codes[data.GSE47533$condition == "Norm"])
dists <- as.matrix(dist(expr.GSE47533.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.GSE47533.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.GSE47533.hif[expr.row])</pre>
diag(dists) <- NA</pre>
```

Clustering of Gene Expression on Hypoxia Euclidian Distance (GSE47533)

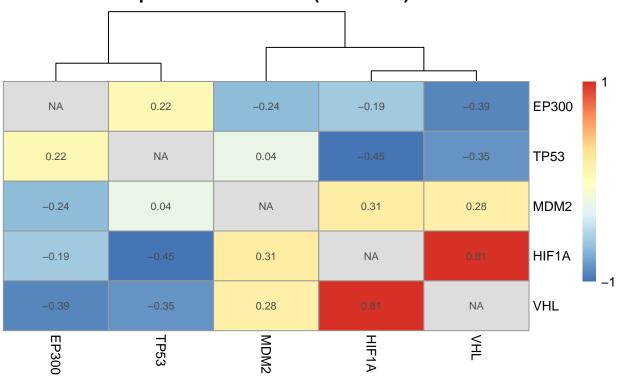


Clustering of Gene Expression on Normoxia Euclidian Distance (GSE47533)

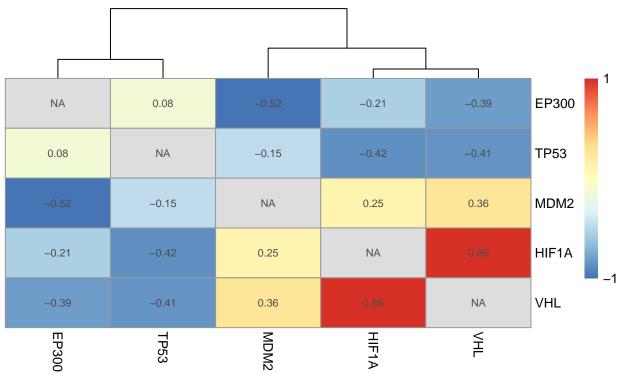


```
dists <- cor(t(expr.GSE47533.hif), use = "pairwise.complete.obs", method = "spearman")
rownames(dists) <- rownames(expr.GSE47533.hif)
colnames(dists) <- rownames(expr.GSE47533.hif)
diag(dists) <- NA</pre>
```

Clustering of Gene Expression Spearman Correlation (GSE47533)



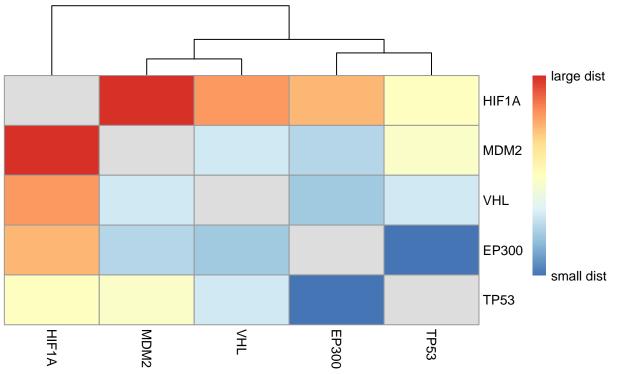
Clustering of Gene Expression Pearson Correlation (GSE47533)



Heatmaps - All datasets Breast Cell-lines (E-GEOD-18494, GSE47533, and GSE41491)

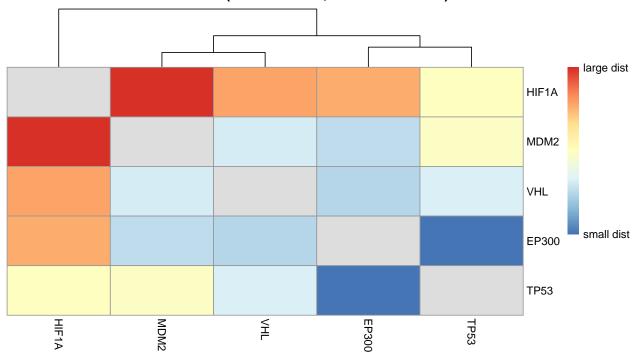
- E-GEOD-18494 2012 / MDA-MB231 / breast / 4h, 8h, 12h / microarray
- GSE41491 2012 / MCF7 / breast / 1h, 2h, 4h, 8h, 12h, 16h, 24h / microarray
- GSE47534 2014 / MCF7 / breast / normoxia, 16h, 32h, 48h / mRNA

Clustering of Gene Expression Euclidian Distance (All 3 datasets, breast cell-lines)

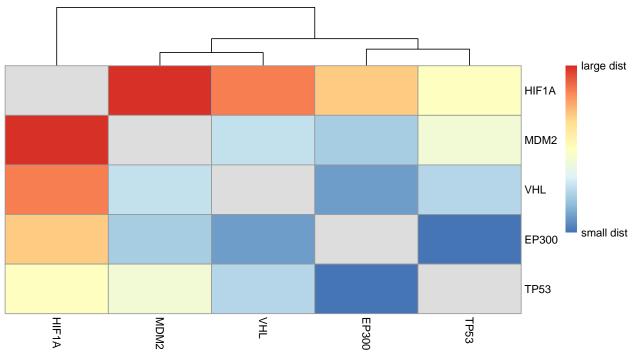


```
main = "Clustering of Gene Expression on Hypoxia \n Euclidian Distance (All 3 datasets, brea
         silent=T)
col_norm <- union(data.GSE47533$codes[data.GSE47533$condition == "Norm"],</pre>
                  union(data.EGEOD18494$codes[data.EGEOD18494$condition == "normoxia"],
                         data.GSE41491$codes[data.GSE41491$condition == "no"]))
expr.row <- (colnames(expr.all.hif) %in% col_norm)</pre>
dists <- as.matrix(dist(expr.all.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.all.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.all.hif[expr.row])</pre>
diag(dists) <- NA</pre>
p2 <- pheatmap(dists,</pre>
         legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Normoxia \n Euclidian Distance (All 3 datasets, breas
         silent=T)
gridExtra::grid.arrange(grobs=list(p1$gtable, p2$gtable),
                        nrow = 2 , labels=c('A', 'B'))
```

Clustering of Gene Expression on Hypoxia Euclidian Distance (All 3 datasets, breast cell-lines)

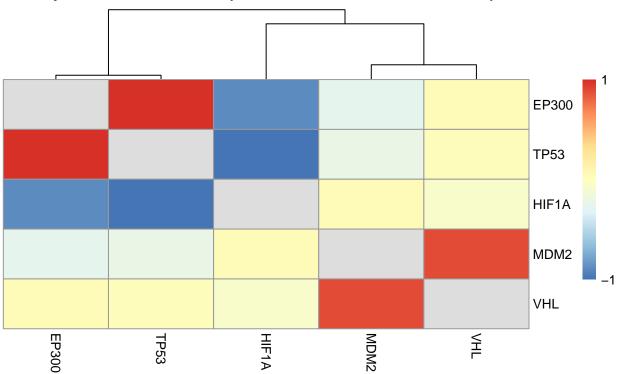


Clustering of Gene Expression on Normoxia Euclidian Distance (All 3 datasets, breast cell-lines)



```
dists <- cor(t(expr.all.hif), use = "pairwise.complete.obs", method = "spearman")
rownames(dists) <- rownames(expr.all.hif)
colnames(dists) <- rownames(expr.all.hif)
diag(dists) <- NA</pre>
```

Clustering of Gene Expression Spearman Correlation (All 3 datasets, breast cell-lines)



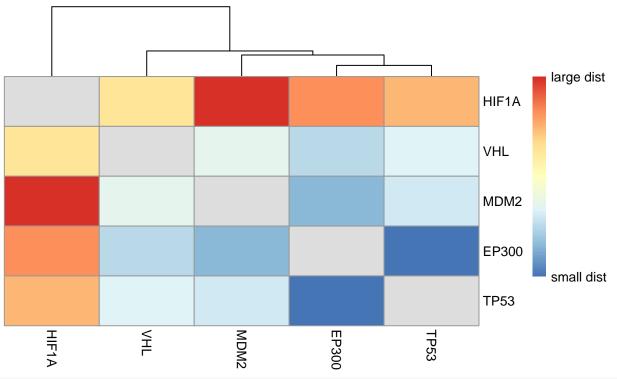
Heatmaps - All datasets All Cell-lines (E-GEOD-18494, GSE47533, and GSE41491)

- E-GEOD-18494 2012 / HepG2, U87, MDA-MB231 / hepatoma, glioma, breast / 4h, 8h, 12h / microarray
- GSE41491 2012 / DU145, HT29, MCF7 / prostate, colon, breast / 1h, 2h, 4h, 8h, 12h, 16h, 24h / microarray
- GSE47534 2014 / MCF7 / breast / normoxia, 16h, 32h, 48h / mRNA

```
# Imput the mean of all VHL values
mean.vhl <- mean(unlist(expr.GSE47533.hif["VHL",], expr.EGE0D18494.hif["VHL",]))
expr.GSE41491.hif["VHL",] <- rep(mean.vhl, 24)

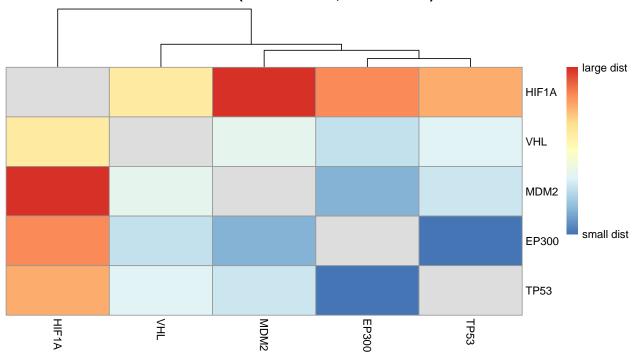
expr.all.hif <- cbind(expr.GSE47533.hif, expr.EGE0D18494.hif, expr.GSE41491.hif)</pre>
```

Clustering of Gene Expression Euclidian Distance (All 3 datasets, all cell-lines)

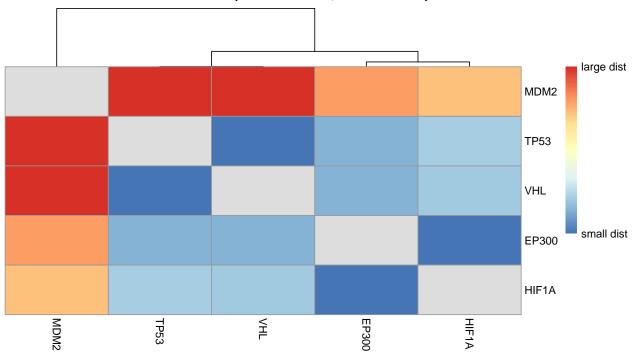


```
legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                           max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Hypoxia \n Euclidian Distance (All 3 datasets, all
         silent=T)
expr.row <- (colnames(expr.all.hif) %in% data.GSE47533$codes[data.GSE47533$condition == "Norm"])
dists <- as.matrix(dist(expr.all.hif[expr.row], method = "euclidean"))</pre>
rownames(dists) <- rownames(expr.all.hif[expr.row])</pre>
colnames(dists) <- rownames(expr.all.hif[expr.row])</pre>
diag(dists) <- NA</pre>
p2 <- pheatmap(dists,</pre>
         legend = TRUE,
         treeheight_row = 0,
         legend_breaks = c(min(dists, na.rm = TRUE),
                            max(dists, na.rm = TRUE)),
         legend_labels = (c("small dist", "large dist")),
         main = "Clustering of Gene Expression on Normoxia \n Euclidian Distance (All 3 datasets, all c
         silent=T)
gridExtra::grid.arrange(grobs=list(p1$gtable, p2$gtable),
                        nrow = 2 , labels=c('A', 'B'))
```

Clustering of Gene Expression on Hypoxia Euclidian Distance (All 3 datasets, all cell-lines)



Clustering of Gene Expression on Normoxia Euclidian Distance (All 3 datasets, all cell-lines)



```
dists <- cor(t(expr.all.hif), use = "pairwise.complete.obs", method = "spearman")
rownames(dists) <- rownames(expr.all.hif)
colnames(dists) <- rownames(expr.all.hif)
diag(dists) <- NA</pre>
```

Clustering of Gene Expression Spearman Correlation (All 3 datasets, all cell-lines)

