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Designing oligonucleotides to disrupt lacZ gene in E coli DH5 alpha

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Goal: Knock-out the lacZ gene.

Steps:

Order two oligonucleotides (instructions will be below).

**Anneal them**  $\rightarrow$  mix forward and reverse oligos, heat and cool so they form a double-stranded insert.

Clone into Cas9 plasmid using Golden Gate (BbsI) or similar cloning strategy.

Transform the plasmid into E. coli DH5α.

Inside the cells, the plasmid will express **Cas9 + our gRNA**, which together target lacZ and cut it at the chosen site.

The cut will usually inactivate lacZ via small indels.

## Oligonucleotide design:

**Step 1**: Obtain the genbank/FASTA link for the E. Coli strand we're targeting. This is the link.

**Step 2**: Control-F to find the lacZ gene's coordinates. Note the CDS, or coding DNA sequence, maps to  $\beta$ -galactosidase, which is what we want to make non-functional.

- **Step 3**: <u>Using a python script</u>, I created a new FASTA file that only contained nucleotides in the range of 1239256..1242168.
- **Step 4**: Go to CHOPCHOP, and find the best protospacer. <u>Here's a link</u> to the CHOPCHOP output with the new FASTA file from Step 3.
- **Step 5**: According to CHOPCHOP, our best protospacer is 'CTGACAATGGCAGATCCCAG'. So our forward oligo will be:

## CACC CTGACAATGGCAGATCCCAG

And our reverse oligo will be:

## AAAC CTGGGATCTGCCATTGTCAG

after we add the overhangs.

Step 6: Order the oligonucleotides on idtdna's website.

