

Package ‘semTools’

February 26, 2014

Type Package

Title Useful tools for structural equation modeling.

Version 0.4-3

Date 2014-2-24

Author Sunthud Pornprasertmanit [aut, cre], Patrick Miller [aut], Alex Schoemann [aut], Yves Rosseel [aut], Corbin Quick [ctb], Mauricio Garnier-Villarreal [ctb], James Selig [ctb], Aaron Boulton [ctb], Kristopher Preacher [ctb], Donna Coffman [ctb], Mijke Rhemtulla [ctb], Alexander Robitzsch [ctb], Craig Enders [ctb], Ruber Arslan [ctb], Bell Clinton [ctb], Pavel Panko [ctb], Ed Merkle [ctb]

Maintainer Sunthud Pornprasertmanit <psunthud@ku.edu>

Depends R(>= 3.0), lavaan, methods

Suggests MASS, parallel, Amelia, mice, foreign, OpenMx, GPArotation

Description This package provide useful tools for structural equation modeling analysis.

License GPL (>= 2)

LazyLoad yes

LazyData yes

URL <https://github.com/simsem/semTools/wiki>

R topics documented:

auxiliary	2
BootMiss-class	4
bsBootMiss	5
clipboard_saveFile	7
compareFit	8
dat2way	10
dat3way	10
EFA-class	11
efaUnrotate	12
exLong	13
findRMSEApower	14

findRMSEApowernested	15
findRMSEAsamplesize	16
findRMSEAsamplesizenested	17
FitDiff-class	18
fitMeasuresMx	19
fmi	20
impliedFactorStat	22
imposeStart	23
indProd	25
kd	27
kurtosis	29
lavaanStar-class	30
lisrel2lavaan	31
loadingFromAlpha	33
longInvariance	33
mardiaKurtosis	36
mardiaSkew	37
measurementInvariance	38
miPowerFit	39
monteCarloMed	42
moreFitIndices	44
net	46
Net-class	47
nullMx	48
nullRMSEA	49
parcelAllocation	50
plotProbe	52
plotRMSEAdist	54
plotRMSEApower	55
plotRMSEApowernested	57
probe2WayMC	58
probe2WayRC	60
probe3WayMC	63
probe3WayRC	66
reliability	69
reliabilityL2	71
residualCovariate	72
rotate	73
runMI	74
saturateMx	77
simParcel	78
skew	79
splitSample	80
standardizeMx	81
tukeySEM	83
wald	84

auxiliary

*Analyzing data with full-information maximum likelihood with auxiliary variables***Description**

Analyzing data with full-information maximum likelihood with auxiliary variables. The techniques used to account for auxiliary variables are both extra-dependent-variables and saturated-correlates approaches (Enders, 2008). The extra-dependent-variables approach is used for all single variables in the model (such as covariates or single-indicator dependent variable) For variables that are belong to a multiple-indicator factor, the saturated-correlates approach is used. Note that all covariates are treated as endogenous variables in this model (`fixed.x = FALSE`) so multivariate normality is assumed for the covariates. **CAUTION:** (1) this function will automatically change the missing data handling method to full-information maximum likelihood and (2) this function is still not applicable for categorical variables (because the maximum likelihood method is not available in lavaan for estimating models with categorical variables currently).

Usage

```
auxiliary(model, aux, fun, ...)
cfa.auxiliary(model, aux, ...)
sem.auxiliary(model, aux, ...)
growth.auxiliary(model, aux, ...)
lavaan.auxiliary(model, aux, ...)
```

Arguments

<code>model</code>	The lavaan object, the parameter table, or lavaan script. If the lavaan object is provided, the lavaan object must be evaluated with mean structure.
<code>aux</code>	The list of auxiliary variable
<code>fun</code>	The character of the function name used in running lavaan model ("cfa", "sem", "growth", "lavaan").
<code>...</code>	The additional arguments in the lavaan function.

Value

The `lavaanStar` object which contains the original `lavaan` object and the additional values of the null model, which need to be adjusted to account for auxiliary variables.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

Enders, C. K. (2008). A note of the use of missing auxiliary variables in full information maximum likelihood-based structural equation models. *Structural Equation Modeling*, 15, 434-448.

See Also

`lavaanStar`

Examples

```
# Example of confirmatory factor analysis

HS.model <- ' visual  =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed   =~ x7 + x8 + x9 '

dat <- data.frame(HolzingerSwineford1939, z=rnorm(nrow(HolzingerSwineford1939), 0, 1))

fit <- cfa(HS.model, data=dat, meanstructure=TRUE)
fitaux <- auxiliary(HS.model, aux="z", data=dat, fun="cfa") # Use lavaan script
fitaux <- cfa.auxiliary(fit, aux="z", data=dat) # Use lavaan output

# Example of multiple groups confirmatory factor analysis

fitgroup <- cfa(HS.model, data=dat, group="school", meanstructure=TRUE)
fitgroupaux <- cfa.auxiliary(fitgroup, aux="z", data=dat, group="school")

# Example of path analysis

mod <- ' x5 ~ x4
         x4 ~ x3
         x3 ~ x1 + x2 '

fitpath <- sem(mod, data=dat, fixed.x=FALSE, meanstructure=TRUE) # fixed.x must be FALSE
fitpathaux <- sem.auxiliary(fitpath, aux="z", data=dat)

# Example of full structural equation modeling

dat2 <- data.frame(PoliticalDemocracy, z=rnorm(nrow(PoliticalDemocracy), 0, 1))
model <- '
  ind60 =~ x1 + x2 + x3
  dem60 =~ y1 + a*y2 + b*y3 + c*y4
  dem65 =~ y5 + a*y6 + b*y7 + c*y8

  dem60 ~ ind60
  dem65 ~ ind60 + dem60

  y1 ~~ y5
  y2 ~~ y4 + y6
  y3 ~~ y7
  y4 ~~ y8
  y6 ~~ y8
,
fitsem <- sem(model, data=dat2, meanstructure=TRUE)
fitsemaux <- sem.auxiliary(fitsem, aux="z", data=dat2, meanstructure=TRUE)

# Example of covariate at the factor level

HS.model.cov <- ' visual  =~ x1 + x2 + x3
                  textual =~ x4 + x5 + x6
                  speed   =~ x7 + x8 + x9
  visual ~ sex
  textual ~ sex
  speed ~ sex'
```

```

fitcov <- cfa(HS.model.cov, data=dat, fixed.x=FALSE, meanstructure=TRUE)
fitcovaux <- cfa.auxiliary(fitcov, aux="z", data=dat)

# Example of Endogenous variable with single indicator
HS.model.cov2 <- ' visual  =~ x1 + x2 + x3
                  textual =~ x4 + x5 + x6
                  x7 ~ visual + textual'

fitcov2 <- sem(HS.model.cov2, data=dat, fixed.x=FALSE, meanstructure=TRUE)
fitcov2aux <- sem.auxiliary(fitcov2, aux="z", data=dat)

# Multiple auxiliary variables
HS.model2 <- ' visual  =~ x1 + x2 + x3
              speed   =~ x7 + x8 + x9'
fit <- cfa(HS.model2, data=HolzingerSwineford1939)
fitaux <- cfa.auxiliary(HS.model2, data=HolzingerSwineford1939, aux=c("x4", "x5"))

```

BootMiss-class

Class For the Results of Bollen-Stine Bootstrap with Incomplete Data

Description

This class contains the results of Bollen-Stine bootstrap with missing data.

Objects from the Class

Objects can be created via the `bsBootMiss` function.

Slots

timeTrans: Time (in seconds) using for data transformation
timeFit: Time (in seconds) using for fitting Bootstrap data sets
transData: Transformed data
bootDist: The vector of chi-square values from Bootstrap data sets fitted by the target model
origChi: The chi-square value from the original data set
df: The degree of freedom of the model
bootP: The p-value comparing the original chi-square with the bootstrap distribution

methods

- **summary** The summary function is used to provide the detailed results of the Bollen-Stine bootstrap.
- **hist** The hist function is used to provided the bootstrap distribution compared with the original chi-square value.

Author(s)

Terrence D. Jorgensen (University of Kansas; <TJorgensen314@gmail.com>)

See Also

`bsBootMiss`

Examples

```
# See the example from the bsBootMiss function
```

bsBootMiss

Bollen-Stine Bootstrap with the Existence of Missing Data

Description

Implement the Bollen and Stine's (1992) Bootstrap when missing observations exist. The implemented method is proposed by Savalei and Yuan (2009). This can be used in two ways. The first and easiest option is to fit the model to incomplete data in `lavaan` using the FIML estimator, then pass that `lavaan` object to `bsBootMiss`.

The second is designed for users of other software packages (e.g., LISREL, EQS, Amos, or Mplus). Users can import their data, chi-squared value, and model-implied moments from another package, and they have the option of saving (or writing to a file) either the transformed data or bootstrapped samples of that data, which can be analyzed in other programs. In order to analyze the bootstrapped samples and return a p value, users of other programs must still specify their model using `lavaan` syntax.

Usage

```
bsBootMiss(x, transformation = 2, nBoot = 500, model, rawData,
Sigma, Mu, group, ChiSquared, EMcov,
writeTransData = FALSE, transDataOnly = FALSE,
writeBootData = FALSE, bootSamplesOnly = FALSE,
writeArgs, seed = NULL, suppressWarn = TRUE, ...)
```

Arguments

<code>x</code>	A target <code>lavaan</code> object used in the Bollen-Stine bootstrap
<code>transformation</code>	The transformation methods in Savalei and Yuan (2009). There are three methods in the article, but only the first two are currently implemented here. Use <code>transformation = 1</code> when there are few missing data patterns, each of which has a large size, such as in a planned-missing-data design. Use <code>transformation = 2</code> when there are more missing data patterns. The currently unavailable <code>transformation = 3</code> would be used when several missing data patterns have $n = 1$.
<code>nBoot</code>	The number of bootstrap samples.
<code>model</code>	Optional. The target model if <code>x</code> is not provided.
<code>rawData</code>	Optional. The target raw data set if <code>x</code> is not provided.
<code>Sigma</code>	Optional. The model-implied covariance matrix if <code>x</code> is not provided.
<code>Mu</code>	Optional. The model-implied mean vector if <code>x</code> is not provided.
<code>group</code>	Optional character string specifying the name of the grouping variable in <code>rawData</code> if <code>x</code> is not provided.
<code>ChiSquared</code>	Optional. The model-implied mean vector if <code>x</code> is not provided.
<code>EMcov</code>	Optional, if <code>x</code> is not provided. The EM (or Two-Stage ML) estimated covariance matrix used to speed up Transformation 2 algorithm.

<code>transDataOnly</code>	Logical. If TRUE, the result will provide the transformed data only.
<code>writeTransData</code>	Logical. If TRUE, the transformed data set is written to a text file, <code>transDataOnly</code> is set to TRUE, and the transformed data is returned invisibly.
<code>bootSamplesOnly</code>	Logical. If TRUE, the result will provide bootstrap data sets only.
<code>writeBootData</code>	Logical. If TRUE, the stacked bootstrap data sets are written to a text file, <code>bootSamplesOnly</code> is set to TRUE, and the list of bootstrap data sets are returned invisibly.
<code>writeArgs</code>	Optional. If TRUE or TRUE are TRUE, user can pass arguments to the <code>write.table</code> function as a list. Some default values are provided: <code>file = "bootstrappedSamples.dat"</code> , <code>row.names = FALSE</code> , and <code>na = "-999"</code> , but the user can override all of these by providing other values for those arguments in the <code>writeArgs</code> list.
<code>seed</code>	The seed number used in randomly drawing bootstrap samples.
<code>suppressWarn</code>	Logical. If TRUE, warnings from <code>lavaan</code> function will be suppressed when fitting the model to each bootstrap sample.
<code>...</code>	The additional arguments in the <code>lavaan</code> function.

Value

As a default, this function returns `BootMiss` object containing the results of the bootstrap samples. Users can use `print`, `summary`, or `hist` functions to examine the results. The transformed data is returned instead if `transDataOnly` is TRUE. The bootstrap data sets are returned if `bootSamplesOnly` is TRUE.

Author(s)

Terrence D. Jorgensen (University of Kansas; <TJorgensen314@gmail.com>)

References

- Bollen, K. A., & Stine, R. A. (1992). Bootstrapping goodness-of-fit measures in structural equation models. *Sociological Methods & Research*, 21, 205-229. doi:10.1177/0049124192021002004
- Savalei, V., & Yuan, K.-H. (2009). On the model-based bootstrap with missing data: obtaining a p-value for a test of exact fit. *Multivariate Behavioral Research*, 44, 741-763. doi:10.1080/00273170903333590

See Also

`BootMiss`

Examples

```
dat1 <- HolzingerSwineford1939
dat1$x5 <- ifelse(dat1$x1 <= quantile(dat1$x1, .3), NA, dat1$x5)
dat1$x9 <- ifelse(is.na(dat1$x5), NA, dat1$x9)

targetModel <- "
visual  =~ x1 + x2 + x3
textual =~ x4 + x5 + x6
speed   =~ x7 + x8 + x9
```

```
"
targetFit <- sem(targetModel, dat1, meanstructure = TRUE, std.lv = TRUE,
               missing = "fiml", group = "school")
summary(targetFit, fit = TRUE, standardized = TRUE)

# The number of bootstrap samples should be much higher.
temp <- bsBootMiss(targetFit, transformation = 1, nBoot = 10, seed = 31415)

temp
summary(temp)
hist(temp)
## user can specify confidence level (default = 0.95), the location of
## the legend ("none", "right", or "left"), and pass other arguments to hist()
hist(temp, conf = .90, legend = "none", xlab = "something else", breaks = 25)
```

clipboard_saveFile *Copy or save the result of lavaan or FitDiff objects into a clipboard or a file*

Description

Copy or save the result of lavaan or FitDiff object into a clipboard or a file. From the clipboard, users may paste the result into the Microsoft Excel or spreadsheet application to create a table of the output.

Usage

```
clipboard(object, what="summary", ...)
saveFile(object, file, what="summary", tableFormat=FALSE, ...)
```

Arguments

object	The lavaan or FitDiff object
what	The attributes of the lavaan object to be copied in the clipboard. "summary" is to copy the screen provided from the summary function. "mifit" is to copy the result from the miPowerFit function. Other attributes listed in the inspect method in the lavaan-class could also be used, such as "coef", "se", "fit", "samp", and so on. For the The FitDiff object, this argument is not active yet.
file	A file name used for saving the result
tableFormat	If TRUE, save the result in the table format using tabs for separation. Otherwise, save the result as the output screen printed in the R console.
...	Additional argument listed in the miPowerFit function (for lavaan object only).

Value

The resulting output will be saved into a clipboard or a file. If using the clipboard function, users may paste it in the other applications.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

Examples

```
## Not run:
library(lavaan)
HW.model <- ' visual  =~ x1 + c1*x2 + x3
              textual =~ x4 + c1*x5 + x6
              speed   =~ x7 + x8 + x9 '

fit <- cfa(HW.model, data=HolzingerSwineford1939, group="school", meanstructure=TRUE)

# Copy the summary of the lavaan object
clipboard(fit)

# Copy the modification indices and the model fit from the miPowerFit function
clipboard(fit, "mifit")

# Copy the parameter estimates
clipboard(fit, "coef")

# Copy the standard errors
clipboard(fit, "se")

# Copy the sample statistics
clipboard(fit, "samp")

# Copy the fit measures
clipboard(fit, "fit")

# Save the summary of the lavaan object
saveFile(fit, "out.txt")

# Save the modification indices and the model fit from the miPowerFit function
saveFile(fit, "out.txt", "mifit")

# Save the parameter estimates
saveFile(fit, "out.txt", "coef")

# Save the standard errors
saveFile(fit, "out.txt", "se")

# Save the sample statistics
saveFile(fit, "out.txt", "samp")

# Save the fit measures
saveFile(fit, "out.txt", "fit")

## End(Not run)
```

Description

This function will create the template that compare fit indices across multiple lavaan outputs. The results can be exported to a clipboard or a file later.

Usage

```
compareFit(..., nested = TRUE)
```

Arguments

<code>...</code>	lavaan outputs or lists of lavaan outputs
<code>nested</code>	Logical whether the specified models are nested

Value

A `FitDiff` object that saves model fit comparisons across multiple models. If the output is not assigned as an object, the output is printed in two parts: 1) nested model comparison (if models are nested) and 2) fit indices summaries. In the fit indices summaries, daggers are tagged to the model with the best fit for each fit index.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

`FitDiff`, `clipboard`

Examples

```
m1 <- ' visual  =~ x1 + x2 + x3
      textual  =~ x4 + x5 + x6
      speed    =~ x7 + x8 + x9 '

fit1 <- cfa(m1, data=HolzingerSwineford1939)

m2 <- ' f1  =~ x1 + x2 + x3 + x4
      f2  =~ x5 + x6 + x7 + x8 + x9 '
fit2 <- cfa(m2, data=HolzingerSwineford1939)
compareFit(fit1, fit2, nested=FALSE)

HW.model <- ' visual =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed  =~ x7 + x8 + x9 '

out <- measurementInvariance(HW.model, data=HolzingerSwineford1939, group="school", quiet=TRUE)
compareFit(out)
```

`dat2way`*Simulated Dataset to Demonstrate Two-way Latent Interaction*

Description

A simulated data set with 2 independent factors and 1 dependent factor where each factor has three indicators

Usage

```
data(dat2way)
```

Format

A data frame with 500 observations of 9 variables.

- x1** The first indicator of the first independent factor
- x2** The second indicator of the first independent factor
- x3** The third indicator of the first independent factor
- x4** The first indicator of the second independent factor
- x5** The second indicator of the second independent factor
- x6** The third indicator of the second independent factor
- x7** The first indicator of the dependent factor
- x8** The second indicator of the dependent factor
- x9** The third indicator of the dependent factor

Source

Data was generated by the `mvrnorm` function in the `MASS` package.

Examples

```
head(dat2way)
```

`dat3way`*Simulated Dataset to Demonstrate Three-way Latent Interaction*

Description

A simulated data set with 3 independent factors and 1 dependent factor where each factor has three indicators

Usage

```
data(dat3way)
```

Format

A data frame with 500 observations of 12 variables.

x1 The first indicator of the first independent factor

x2 The second indicator of the first independent factor

x3 The third indicator of the first independent factor

x4 The first indicator of the second independent factor

x5 The second indicator of the second independent factor

x6 The third indicator of the second independent factor

x7 The first indicator of the third independent factor

x8 The second indicator of the third independent factor

x9 The third indicator of the third independent factor

x10 The first indicator of the dependent factor

x11 The second indicator of the dependent factor

x12 The third indicator of the dependent factor

Source

Data was generated by the `mvrnorm` function in the `MASS` package.

Examples

```
head(dat3way)
```

EFA-class

Class For Rotated Results from EFA

Description

This class contains the results of rotated exploratory factor analysis

Objects from the Class

Objects can be created via the `orthRotate` or `oblqRotate` function.

Slots

loading: Rotated standardized factor loading matrix

rotate: Rotation matrix

gradRotate: The gradient of the objective function at the rotated loadings

convergence: Convergence status

phi: Factor correlation. Will be an identity matrix if orthogonal rotation is used.

se: Standard errors of the rotated standardized factor loading matrix

method: Method of rotation

call: The command used to generate this object

methods

- `summary` The `summary` function shows the detailed results of the rotated solution. This function has two arguments: `suppress` and `sort`. The `suppress` argument is used to not show the standardized loading values that less than the specified value. The default is 0.1. The `sort` is used to sort the factor loadings by the sizes of factor loadings in each factor. The default is `TRUE`.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

`efaUnrotate`; `orthRotate`; `oblqRotate`

Examples

```
unrotated <- efaUnrotate(HolzingerSwineford1939, nf=3, varList=paste0("x", 1:9), estimator="ml",
  summary(unrotated, std=TRUE)
inspect(unrotated, "standardized")

# Rotated by Quartimin
rotated <- oblqRotate(unrotated, method="quartimin")
summary(rotated)
```

efaUnrotate	Analyze Unrotated Exploratory Factor Analysis Model
-------------	---

Description

This function will analyze unrotated exploratory factor analysis model. The unrotated solution can be rotated by the `orthRotate` and `oblqRotate` functions.

Usage

```
efaUnrotate(data, nf, varList=NULL, start=TRUE, aux=NULL, ...)
```

Arguments

<code>data</code>	A target data frame.
<code>nf</code>	The desired number of factors
<code>varList</code>	Target observed variables. If not specified, all variables in the target data frame will be used.
<code>start</code>	Use starting values in the analysis from the <code>factanal</code> function. If <code>FALSE</code> , the starting values from the <code>lavaan</code> package will be used.
<code>aux</code>	The list of auxiliary variables. These variables will be included in the model by the saturated-correlates approach to account for missing information.
<code>...</code>	Other arguments in the <code>cfa</code> function in the <code>lavaan</code> package, such as <code>ordered</code> , <code>se</code> , or <code>estimator</code>

Details

This function will generate a lavaan script for unrotated exploratory factor analysis model such that 1) all factor loadings are estimated, 2) factor variances are fixed to 1, 3) factor covariances are fixed to 0, and 4) the dot products of any pairs of columns in the factor loading matrix are fixed to zero (Johnson and Wichern, 2002). The reason for creating this function in addition to the `factanal` function is that users can enjoy some advanced features from the `lavaan` package such as scaled chi-square, diagonal weighted least square for ordinal indicators, or full-information maximum likelihood.

Value

A lavaan output of unrotated exploratory factor analysis solution.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

Examples

```
unrotated <- efaUnrotate(HolzingerSwineford1939, nf=3, varList=paste0("x", 1:9), estimator="ML",
summary(unrotated, std=TRUE)
inspect(unrotated, "standardized")

dat <- data.frame(HolzingerSwineford1939, z=rnorm(nrow(HolzingerSwineford1939), 0, 1))
unrotated2 <- efaUnrotate(dat, nf=2, varList=paste0("x", 1:9), aux="z")
```

exLong

Simulated Data set to Demonstrate Longitudinal Measurement Invariance

Description

A simulated data set with 1 factors with 3 indicators in three timepoints

Usage

```
data(exLong)
```

Format

A data frame with 200 observations of 10 variables.

sex Sex of respondents

y1t1 Indicator 1 in Time 1

y2t1 Indicator 2 in Time 1

y3t1 Indicator 3 in Time 1

y1t2 Indicator 1 in Time 2

y2t2 Indicator 2 in Time 2

y3t2 Indicator 3 in Time 2

y1t3 Indicator 1 in Time 3

y2t3 Indicator 2 in Time 3

y3t3 Indicator 3 in Time 3

Source

Data was generated using the `simsem` package.

Examples

```
head(exLong)
```

<code>findRMSEApower</code>	<i>Find the statistical power based on population RMSEA</i>
-----------------------------	---

Description

Find the proportion of the samples from the sampling distribution of RMSEA in the alternative hypothesis rejected by the cutoff derived from the sampling distribution of RMSEA in the null hypothesis. This function can be applied for both test of close fit and test of not-close fit (MacCallum, Browne, & Suguwara, 1996)

Usage

```
findRMSEApower(rmseao, rmseaA, df, n, alpha=.05, group=1)
```

Arguments

<code>rmseao</code>	Null RMSEA
<code>rmseaA</code>	Alternative RMSEA
<code>df</code>	Model degrees of freedom
<code>n</code>	Sample size of a dataset
<code>alpha</code>	Alpha level used in power calculations
<code>group</code>	The number of group that is used to calculate RMSEA.

Details

This function find the proportion of sampling distribution derived from the alternative RMSEA that is in the critical region derived from the sampling distribution of the null RMSEA. If `rmseaA` is greater than `rmseao`, the test of close fit is used and the critical region is in the right hand side of the null sampling distribution. On the other hand, if `rmseaA` is less than `rmseao`, the test of not-close fit is used and the critical region is in the left hand side of the null sampling distribution (MacCallum, Browne, & Suguwara, 1996).

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

MacCallum, R. C., Browne, M. W., & Sugawara, H. M. (1996). Power analysis and determination of sample size for covariance structure modeling. *Psychological Methods*, 1, 130-149.

See Also

- `plotRMSEApower` to plot the statistical power based on population RMSEA given the sample size
- `plotRMSEAdist` to visualize the RMSEA distributions
- `findRMSEAsamplesize` to find the minimum sample size for a given statistical power based on population RMSEA

Examples

```
findRMSEApower(rmse0=.05, rmseA=.08, df=20, n=200)
```

```
findRMSEApowernested
```

Find power given a sample size in nested model comparison

Description

Find the sample size that the power in rejection the samples from the alternative pair of RMSEA is just over the specified power.

Usage

```
findRMSEApowernested(rmse0A = NULL, rmse0B = NULL,
  rmse1A, rmse1B = NULL, dfA, dfB, n, alpha=.05,
  group=1)
```

Arguments

<code>rmse0A</code>	The H0 baseline RMSEA.
<code>rmse0B</code>	The H0 alternative RMSEA (trivial misfit).
<code>rmse1A</code>	The H1 baseline RMSEA.
<code>rmse1B</code>	The H1 alternative RMSEA (target misfit to be rejected).
<code>dfA</code>	degree of freedom of the more-restricted model.
<code>dfB</code>	degree of freedom of the less-restricted model.
<code>n</code>	Sample size.
<code>alpha</code>	The alpha level.
<code>group</code>	The number of group in calculating RMSEA.

Author(s)

Bell Clinton (University of Kansas; <clintonbell@ku.edu>); Pavel Panko (University of Kansas; <pavel@ku.edu>); Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

MacCallum, R. C., Browne, M. W., & Cai, L. (2006). Testing differences between nested covariance structure models: Power analysis and null hypotheses. *Psychological Methods*, 11, 19-35.

See Also

- `plotRMSEApowernested` to plot the statistical power for nested model comparison based on population RMSEA given the sample size
- `findRMSEAsamplesizenested` to find the minimum sample size for a given statistical power in nested model comparison based on population RMSEA

Examples

```
findRMSEApowernested(rmse0A = 0.06, rmse0B = 0.05, rmse1A = 0.08,
rmse1B = 0.05, dfA = 22, dfB = 20, n = 200, alpha = 0.05, group = 1)
```

```
findRMSEAsamplesize
```

Find the minimum sample size for a given statistical power based on population RMSEA

Description

Find the minimum sample size for a specified statistical power based on population RMSEA. This function can be applied for both test of close fit and test of not-close fit (MacCallum, Browne, & Suguwara, 1996)

Usage

```
findRMSEAsamplesize(rmse0, rmseA, df, power=0.80, alpha=.05, group=1)
```

Arguments

<code>rmse0</code>	Null RMSEA
<code>rmseA</code>	Alternative RMSEA
<code>df</code>	Model degrees of freedom
<code>power</code>	Desired statistical power to reject misspecified model (test of close fit) or retain good model (test of not-close fit)
<code>alpha</code>	Alpha level used in power calculations
<code>group</code>	The number of group that is used to calculate RMSEA.

Details

This function find the minimum sample size for a specified power based on an iterative routine. The sample size keep increasing until the calculated power from `findRMSEApower` function is just over the specified power. If `group` is greater than 1, the resulting sample size is the sample size per group.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

MacCallum, R. C., Browne, M. W., & Sugawara, H. M. (1996). Power analysis and determination of sample size for covariance structure modeling. *Psychological Methods*, 1, 130-149.

See Also

- `plotRMSEApower` to plot the statistical power based on population RMSEA given the sample size
- `plotRMSEAdist` to visualize the RMSEA distributions
- `findRMSEApower` to find the statistical power based on population RMSEA given a sample size

Examples

```
findRMSEAsamplesize(rmse0=.05, rmseA=.08, df=20, power=0.80)
```

```
findRMSEAsamplesizenested
```

Find sample size given a power in nested model comparison

Description

Find the sample size that the power in rejection the samples from the alternative pair of RMSEA is just over the specified power.

Usage

```
findRMSEAsamplesizenested(rmse0A = NULL, rmse0B = NULL, rmse1A,
rmse1B = NULL, dfA, dfB, power=0.80, alpha=.05, group=1)
```

Arguments

<code>rmse0A</code>	The H0 baseline RMSEA.
<code>rmse0B</code>	The H0 alternative RMSEA (trivial misfit).
<code>rmse1A</code>	The H1 baseline RMSEA.
<code>rmse1B</code>	The H1 alternative RMSEA (target misfit to be rejected).
<code>dfA</code>	degree of freedom of the more-restricted model.
<code>dfB</code>	degree of freedom of the less-restricted model.
<code>power</code>	The desired statistical power.
<code>alpha</code>	The alpha level.
<code>group</code>	The number of group in calculating RMSEA.

Author(s)

Bell Clinton (University of Kansas; <clintonbell@ku.edu>); Pavel Panko (University of Kansas; <pavel@ku.edu>); Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

MacCallum, R. C., Browne, M. W., & Cai, L. (2006). Testing differences between nested covariance structure models: Power analysis and null hypotheses. *Psychological Methods*, 11, 19-35.

See Also

- `plotRMSEApowernested` to plot the statistical power for nested model comparison based on population RMSEA given the sample size
- `findRMSEApowernested` to find the power for a given sample size in nested model comparison based on population RMSEA

Examples

```
findRMSEAsamplesizenested(rmse0A = 0, rmse0B = 0, rmse1A = 0.06,
rmse1B = 0.05, dfA = 22, dfB = 20, power=0.80, alpha=.05, group=1)
```

FitDiff-class

*Class For Representing A Template of Model Fit Comparisons***Description**

This class contains model fit measures and model fit comparisons among multiple models

Objects from the Class

Objects can be created via the `compareFit` function.

Slots

name: The name of each model

nested: Model fit comparisons between adjacent nested models that are ordered based on their degrees of freedom

ordernested: The order of nested models regarding to their degrees of freedom

fit: Fit measures of all models specified in the `name` slot

methods

- **summary** The summary function is used to provide the nested model comparison results and the summary of the fit indices across models. This function has one argument: `fit.measures`. If "default" is specified, chi-square values, degree of freedom, *p* value, CFI, TLI, RMSEA, SRMR, AIC, and BIC are provided. If "all" is specified, all information given in the `fitMeasures` function is provided. Users may specify a vector of the name of fit indices that they wish.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

`compareFit`; `clipboard`

Examples

```
HW.model <- ' visual =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed =~ x7 + x8 + x9 '

out <- measurementInvariance(HW.model, data=HolzingerSwineford1939, group="school", quiet=TRUE)
modelDiff <- compareFit(out)
summary(modelDiff)
summary(modelDiff, fit.measures="all")
summary(modelDiff, fit.measures=c("aic", "bic"))

## Not run:
# Save results to a file
saveFile(modelDiff, file="modelDiff.txt")

# Copy to a clipboard
clipboard(modelDiff)

## End(Not run)
```

fitMeasuresMx

Find fit measures from an MxModel result

Description

Find fit measures from MxModel result. The saturate and null models are analyzed in the function and fit measures are calculated based on the comparison with the null and saturate models. The function is adjusted from the `fitMeasures` function in the `lavaan` package.

Usage

```
fitMeasuresMx(object, fit.measures="all")
```

Arguments

`object` The target MxModel object
`fit.measures` Target fit measures

Value

A vector of fit measures

Author(s)

The original function is the `fitMeasures` function written by Yves Rosseel in the `lavaan` package. The function is adjusted for an MxModel object by Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

`nullMx`, `saturateMx`, `standardizeMx`

Examples

```
## Not run:
library(OpenMx)
data(demoOneFactor)
manifests <- names(demoOneFactor)
latents <- c("G")
factorModel <- mxModel("One Factor",
  type="RAM",
  manifestVars=manifests,
  latentVars=latents,
  mxPath(from=latents, to=manifests),
  mxPath(from=manifests, arrows=2),
  mxPath(from=latents, arrows=2, free=FALSE, values=1.0),
  mxData(observed=cov(demoOneFactor), type="cov", numObs=500)
)
factorFit <- mxRun(factorModel)
round(fitMeasuresMx(factorFit), 3)

# Compare with lavaan
library(lavaan)
script <- "f1 =~ x1 + x2 + x3 + x4 + x5"
fitMeasures(cfa(script, sample.cov = cov(demoOneFactor), sample.nobs = 500, std.lv = TRUE))

## End(Not run)
```

fmi

Fraction of Missing Information.

Description

This function takes a list of imputed data sets and estimates the Fraction of Missing Information of the Variances and Means for each variable.

Usage

```
fmi(dat.imp, method="saturated", varnames=NULL, group=NULL, exclude=NULL,
    digits=3)
```

Arguments

<code>dat.imp</code>	List of imputed data sets, the function only accept a list of data frames.
<code>method</code>	Specified the model used to estimated the variances and means. Can be one of the following: "saturated" ("sat") or "null", the default is "saturated". See Details for more information.
<code>varnames</code>	A vector of variables names. This argument allow the user to get the fmi of a subset of variables. The function by default will estimate the fmi for all the variables.
<code>group</code>	A variable name defining the groups. This will give the fmi for each group.
<code>exclude</code>	A vector of variables names. These variables will be excluded from the analysis.
<code>digits</code>	Number of decimals to print in the results.

Details

The function estimates a variance/covariance model for each data set using lavaan. If method = "saturated" the function will estimate all the variances and covariances, if method = "null" the function will only estimate the variances. The saturated model gives more reliable estimates. With big data sets using the saturated model could take a lot of time. In the case of having problems with big data sets it is helpful to select a subset of variables with varnames and/or use the "null" model. The function does not accept character variables.

Value

fmi returns a list with the Fraction of Missing Information of the Variances and Means for each variable in the data set.

Variances	The estimated variance for each variable, and the respective standard error. Two estimates Fraction of Missing Information of the Variances. The first estimate of fmi (fmi.1) is asymptotic fmi and the second estimate of fmi (fmi.2) is corrected for small numbers of imputations
Means	The estimated mean for each variable, and the respective standard error. Two estimates Fraction of Missing Information of the Means. The first estimate of fmi (fmi.1) is asymptotic fmi and the second estimate of fmi (fmi.2) is corrected for small numbers of imputations

Author(s)

Mauricio Garnier Villarreal (University of Kansas; <mgv@ku.edu>)

References

- Rubin, D.B. (1987) *Multiple Imputation for Nonresponse in Surveys*. J. Wiley & Sons, New York.
- Savalei, V. & Rhemtulla, M. (2012) On Obtaining Estimates of the Fraction of Missing Information From Full Information Maximum Likelihood, *Structural Equation Modeling: A Multidisciplinary Journal*, 19:3, 477-494.
- Wagner, J. (2010) The Fraction of Missing Information as a Tool for Monitoring the Quality of Survey Data, *Public Opinion Quarterly*, 74:2, 223-243.

Examples

```
library(Amelia)
library(lavaan)

modsim <- '
f1 =~ 0.7*y1+0.7*y2+0.7*y3
f2 =~ 0.7*y4+0.7*y5+0.7*y6
f3 =~ 0.7*y7+0.7*y8+0.7*y9'

datSIM <- simulateData(modsim,model.type="cfa", meanstructure=TRUE,
                        std.lv=TRUE, sample.nobs=c(200,200))
randomMiss2 <- rbinom(prod(dim(datSIM)), 1, 0.1)
randomMiss2 <- matrix(as.logical(randomMiss2), nrow=nrow(datSIM))
randomMiss2[,10] <- FALSE
datSIM[randomMiss2] <- NA
datSIMMI <- amelia(datSIM,m=3,idvars="group")

out1 <- fmi(datSIMMI$imputations, exclude="group")
```

```

out1

out2 <- fmi(datsimMI$imputations, exclude="group", method="null")
out2

out3 <- fmi(datsimMI$imputations, varnames=c("y1", "y2", "y3", "y4"))
out3

out4 <- fmi(datsimMI$imputations, group="group")
out4

```

`impliedFactorStat` *Calculate the model-implied factor means and covariance matrix.*

Description

Calculate reliability values of factors by coefficient omega

Usage

```

impliedFactorStat(object)
impliedFactorMean(object)
impliedFactorCov(object)

```

Arguments

`object` The lavaan model object provided after running the `cfa`, `sem`, `growth`, or `lavaan` functions.

Details

The `impliedFactorMean` function is used to calculated model-implied factor means:

$$\mu = (\mathbf{I} - \mathbf{B})^{-1} \alpha,$$

where μ is the model-implied factor mean, \mathbf{I} is an identity matrix, \mathbf{B} is an regression coefficient matrix, and α is a vector of factor intercepts.

The `impliedFactorCov` function is used to calculated model-implied covariance matrix:

$$\Phi = (\mathbf{I} - \mathbf{B})^{-1} \Psi (\mathbf{I} - \mathbf{B})^{-1'}$$

where Φ is the model-implied factor covariance matrix, Ψ is the residual factor covariance matrix.

The `impliedFactorStat` function is used to provide both model-implied means (if the mean structure is estimated) and covariance matrix.

Value

Model-implied factor means or model-implied factor covariance matrix, or both

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

Examples

```
HS.model <- ' visual  =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed   =~ x7 + x8 + x9 '

fit <- cfa(HS.model, data=HolzingerSwineford1939, group="school")
impliedFactorStat(fit)
```

imposeStart	<i>Specify starting values from a lavaan output</i>
-------------	---

Description

This function will save the parameter estimates of a lavaan output and impose those parameter estimates as starting values for another analysis model. The free parameters with the same names or the same labels across two models will be imposed the new starting values. This function may help to increase the chance of convergence in a complex model (e.g., multitrait-multimethod model or complex longitudinal invariance model).

Usage

```
imposeStart(out, expr, silent = TRUE)
```

Arguments

out	The lavaan output that users wish to use the parameter estimates as starting values for an analysis model
expr	The original code that users use to run a lavaan model
silent	Logical to print the parameter table with new starting values

Value

A fitted lavaan model

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

Examples

```
# The following example show that the longitudinal weak invariance model
# using effect coding was not convergent with three time points but convergent
# with two time points. Thus, the parameter estimates from the model with
# two time points are used as starting values of the three time points.
# The model with new starting values is convergent properly.

weak2time <- '
# Loadings
```



```

f1t1 =~ LOAD1*y1t1 + LOAD2*y2t1 + LOAD3*y3t1
f1t2 =~ LOAD1*y1t2 + LOAD2*y2t2 + LOAD3*y3t2

# Factor Variances
f1t1 ~~ f1t1
f1t2 ~~ f1t2

# Factor Covariances
f1t1 ~~ f1t2

# Error Variances
y1t1 ~~ y1t1
y2t1 ~~ y2t1
y3t1 ~~ y3t1
y1t2 ~~ y1t2
y2t2 ~~ y2t2
y3t2 ~~ y3t2

# Error Covariances
y1t1 ~~ y1t2
y2t1 ~~ y2t2
y3t1 ~~ y3t2

# Factor Means
f1t1 ~ NA*1
f1t2 ~ NA*1

# Measurement Intercepts
y1t1 ~ INT1*1
y2t1 ~ INT2*1
y3t1 ~ INT3*1
y1t2 ~ INT4*1
y2t2 ~ INT5*1
y3t2 ~ INT6*1

# Constraints for Effect-coding Identification
LOAD1 == 3 - LOAD2 - LOAD3
INT1 == 0 - INT2 - INT3
INT4 == 0 - INT5 - INT6
'

model2time <- lavaan(weak2time, data = exLong)

weak3time <- '
# Loadings
f1t1 =~ LOAD1*y1t1 + LOAD2*y2t1 + LOAD3*y3t1
f1t2 =~ LOAD1*y1t2 + LOAD2*y2t2 + LOAD3*y3t2
f1t3 =~ LOAD1*y1t3 + LOAD2*y2t3 + LOAD3*y3t3

# Factor Variances
f1t1 ~~ f1t1
f1t2 ~~ f1t2
f1t3 ~~ f1t3

# Factor Covariances
f1t1 ~~ f1t2 + f1t3
f1t2 ~~ f1t3

```

```

# Error Variances
y1t1 ~~ y1t1
y2t1 ~~ y2t1
y3t1 ~~ y3t1
y1t2 ~~ y1t2
y2t2 ~~ y2t2
y3t2 ~~ y3t2
y1t3 ~~ y1t3
y2t3 ~~ y2t3
y3t3 ~~ y3t3

# Error Covariances
y1t1 ~~ y1t2
y2t1 ~~ y2t2
y3t1 ~~ y3t2
y1t1 ~~ y1t3
y2t1 ~~ y2t3
y3t1 ~~ y3t3
y1t2 ~~ y1t3
y2t2 ~~ y2t3
y3t2 ~~ y3t3

# Factor Means
f1t1 ~ NA*1
f1t2 ~ NA*1
f1t3 ~ NA*1

# Measurement Intercepts
y1t1 ~ INT1*1
y2t1 ~ INT2*1
y3t1 ~ INT3*1
y1t2 ~ INT4*1
y2t2 ~ INT5*1
y3t2 ~ INT6*1
y1t3 ~ INT7*1
y2t3 ~ INT8*1
y3t3 ~ INT9*1

# Constraints for Effect-coding Identification
LOAD1 == 3 - LOAD2 - LOAD3
INT1 == 0 - INT2 - INT3
INT4 == 0 - INT5 - INT6
INT7 == 0 - INT8 - INT9
'

### The following command does not provide convergent result
# model3time <- lavaan(weak3time, data = exLong)

### Use starting values from the model with two time points
model3time <- imposeStart(model2time, lavaan(weak3time, data = exLong))
summary(model3time)

```

Description

The `indProd` function will make products of indicators using no centering, mean centering, double-mean centering, or residual centering. The `orthogonalize` function is the shortcut of the `indProd` function to make the residual-centered indicators products.

Usage

```
indProd(data, var1, var2, var3=NULL, match = TRUE, meanC = TRUE,
        residualC = FALSE, doubleMC = TRUE, namesProd = NULL)
orthogonalize(data, var1, var2, var3=NULL, match=TRUE, namesProd=NULL)
```

Arguments

<code>data</code>	The desired data to be transformed.
<code>var1</code>	Names or indices of the variables loaded on the first factor
<code>var2</code>	Names or indices of the variables loaded on the second factor
<code>var3</code>	Names or indices of the variables loaded on the third factor (for three-way interaction)
<code>match</code>	Specify <code>TRUE</code> to use match-paired approach (Marsh, Wen, & Hau, 2004). If <code>FALSE</code> , the resulting products are all possible products.
<code>meanC</code>	Specify <code>TRUE</code> for mean centering the main effect indicator before making the products
<code>residualC</code>	Specify <code>TRUE</code> for residual centering the products by the main effect indicators (Little, Bovaird, & Widaman, 2006).
<code>doubleMC</code>	Specify <code>TRUE</code> for centering the resulting products (Lin et. al., 2010)
<code>namesProd</code>	The names of resulting products

Value

The original data attached with the products.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>) Alexander Schoemann (East Carolina University; <schoemanna@ecu.edu>)

References

- Marsh, H. W., Wen, Z. & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, 9, 275-300.
- Lin, G. C., Wen, Z., Marsh, H. W., & Lin, H. S. (2010). Structural equation models of latent interactions: Clarification of orthogonalizing and double-mean-centering strategies. *Structural Equation Modeling*, 17, 374-391.
- Little, T. D., Bovaird, J. A., & Widaman, K. F. (2006). On the merits of orthogonalizing powered and product terms: Implications for modeling interactions among latent variables. *Structural Equation Modeling*, 13, 497-519.

See Also

- `probe2WayMC` For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe3WayMC` For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe2WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `probe3WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `plotProbe` Plot the simple intercepts and slopes of the latent interaction.

Examples

```
# Mean centering / two-way interaction / match-paired
dat <- indProd(attitude[, -1], var1=1:3, var2=4:6)

# Residual centering / two-way interaction / match-paired
dat2 <- indProd(attitude[, -1], var1=1:3, var2=4:6, match=FALSE, meanC=FALSE,
residualC=TRUE, doubleMC=FALSE)

# Double-mean centering / two-way interaction / match-paired
dat3 <- indProd(attitude[, -1], var1=1:3, var2=4:6, match=FALSE, meanC=TRUE,
residualC=FALSE, doubleMC=TRUE)

# Mean centering / three-way interaction / match-paired
dat4 <- indProd(attitude[, -1], var1=1:2, var2=3:4, var3=5:6)

# Residual centering / three-way interaction / match-paired
dat5 <- indProd(attitude[, -1], var1=1:2, var2=3:4, var3=5:6, match=FALSE, meanC=FALSE,
residualC=TRUE, doubleMC=FALSE)

# Double-mean centering / three-way interaction / match-paired
dat6 <- indProd(attitude[, -1], var1=1:2, var2=3:4, var3=5:6, match=FALSE, meanC=TRUE,
residualC=TRUE, doubleMC=TRUE)
```

kd

Generate data via the Kaiser-Dickman (1962) algorithm.

Description

Given a covariance matrix and sample size, generate raw data that correspond to the covariance matrix. Data can be generated to match the covariance matrix exactly, or to be a sample from the population covariance matrix.

Usage

```
kd(covmat, n, type = c("exact", "sample"))
```

Arguments

<code>covmat</code>	a symmetric, positive definite covariance matrix
<code>n</code>	the sample size for the data that will be generated
<code>type</code>	type of data generation. <code>exact</code> generates data that exactly correspond to <code>covmat</code> . <code>sample</code> treats <code>covmat</code> as a population covariance matrix, generating a sample of size <code>n</code> .

Details

By default, R's `cov()` function divides by $n-1$. The data generated by this algorithm result in a covariance matrix that matches `covmat`, but you must divide by n instead of $n-1$.

Value

`kd` returns a data matrix of dimension `n` by `nrow(covmat)`.

Author(s)

Ed Merkle (University of Missouri; <merkle@missouri.edu>)

References

Kaiser, H. F. and Dickman, K. (1962). Sample and population score matrices and sample correlation matrices from an arbitrary population correlation matrix. *Psychometrika*, 27, 179-182.

Examples

```
#### First Example

## Get data
dat <- HolzingerSwineford1939[,7:15]
hs.n <- nrow(dat)

## Covariance matrix divided by n
hscov <- ((hs.n-1)/hs.n) * cov(dat)

## Generate new, raw data corresponding to hscov
newdat <- kd(hscov, hs.n)

## Difference between new covariance matrix and hscov is minimal
newcov <- (hs.n-1)/hs.n * cov(newdat)
summary(as.numeric(hscov - newcov))

## Generate sample data, treating hscov as population matrix
newdat2 <- kd(hscov, hs.n, type="sample")

#### Another example

## Define a covariance matrix
covmat <- matrix(0, 3, 3); diag(covmat) <- 1.5; covmat[2:3,1] <- c(1.3, 1.7); covmat[3,2]
covmat <- covmat + t(covmat)

## Generate data of size 300 that have this covariance matrix
rawdat <- kd(covmat, 300)
```

```
## Covariances are exact if we compute sample covariance matrix by
## dividing by n (vs by n-1)
summary(as.numeric((299/300)*cov(rawdat) - covmat))

## Generate data of size 300 where covmat is the population covariance matrix
rawdat2 <- kd(covmat, 300)
```

kurtosis

Finding excessive kurtosis

Description

Finding excessive kurtosis (g_2) of an object

Usage

```
kurtosis(object, population=FALSE)
```

Arguments

object	A vector used to find a excessive kurtosis
population	TRUE to compute the parameter formula. FALSE to compute the sample statistic formula.

Details

The excessive kurtosis computed is g_2 . The parameter excessive kurtosis γ_2 formula is

$$\gamma_2 = \frac{\mu_4}{\mu_2^2} - 3,$$

where μ_i denotes the i order central moment.

The excessive kurtosis formula for sample statistic g_2 is

$$g_2 = \frac{k_4}{k_2^2},$$

where k_i are the i order k -statistic.

The standard error of the excessive kurtosis is

$$Var(\hat{g}_2) = \frac{24}{N}$$

where N is the sample size.

Value

A value of an excessive kurtosis with a test statistic if the population is specified as FALSE

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

Weisstein, Eric W. (n.d.). *Kurtosis*. Retrived from MathWorld—A Wolfram Web Resource <http://mathworld.wolfram.com/Kurtosis.html>

See Also

- `skew` Find the univariate skewness of a variable
- `mardiaSkew` Find the Mardia's multivariate skewness of a set of variables
- `mardiaKurtosis` Find the Mardia's multivariate kurtosis of a set of variables

Examples

```
kurtosis(1:5)
```

lavaanStar-class	<i>Class For Representing A (Fitted) Latent Variable Model with Additional Elements</i>
------------------	---

Description

This is the `lavaan` class that contains additional information about the fit values from the null model. Some functions are adjusted according to the change.

Objects from the Class

Objects can be created via the `auxiliary` function or `runMI`.

Slots

call: The function call as returned by `match.called()`.

timing: The elapsed time (user+system) for various parts of the program as a list, including the total time.

Options: Named list of options that were provided by the user, or filled-in automatically.

ParTable: Named list describing the model parameters. Can be coerced to a `data.frame`. In the documentation, this is called the 'parameter table'.

Data: Object of internal class "Data": information about the data.

SampleStats: Object of internal class "SampleStats": sample statistics

Model: Object of internal class "Model": the internal (matrix) representation of the model

Fit: Object of internal class "Fit": the results of fitting the model

nullfit: The fit-indices information from the null model

imputed: The list of information from running multiple imputation. The first element is the convergence rate of the target and null models. The second element is the fraction missing information. The first estimate of FMI (FMI.1) is asymptotic FMI and the second estimate of FMI (FMI.2) is corrected for small numbers of imputation. The third element is the fit values of the target model by the specified chi-squared methods. The fourth element is the fit values of the null model by the specified chi-square methods.

auxNames: The list of auxiliary variables in the analysis.

References

see lavaan

See Also

auxiliary; runMI

Examples

```
HS.model <- ' visual  =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed   =~ x7 + x8 + x9 '

dat <- data.frame(HolzingerSwineford1939, z=rnorm(nrow(HolzingerSwineford1939), 0, 1))

fit <- cfa(HS.model, data=dat)
fitaux <- auxiliary(HS.model, aux="z", data=dat, fun="cfa")
```

lisrel2lavaan

Latent variable modeling in lavaan using LISREL syntax

Description

This function can be used to estimate a structural equation model in lavaan using LISREL syntax. Data are automatically imported from the LISREL syntax file, or, if data files names are provided within LISREL syntax, from the same directory as the syntax itself, as per standard LISREL data importation.

Usage

```
lisrel2lavaan(filename = NULL, analyze = TRUE, silent = FALSE, ...)
```

Arguments

filename	Filename of the LISREL syntax file. If the filename argument is not specified, the user will be prompted with a file browser with which LISREL syntax file can be selected (recommended).
analyze	Logical. If analyze==TRUE (default), data will be automatically imported and analyzed; lavaan summary output displayed and fit object will be returned silently. If analyze==FALSE, data will not be imported or analyzed; instead, a lavaan parameter table containing the model specifications will be returned.
silent	Logical. If false (default) the data will be analyzed and output displayed. If true, a fit object will be returned and summary output will not be displayed.
...	Additional arguments to be passed to lavaan.

Value

Output summary is printed to screen and lavaan fit object is returned.

Note

lisrel2lavaan is still in development, and not all LISREL commands are currently functional. A number of known limitations are outlined below. If an error is encountered that is not listed, please contact <corbing@ku.edu>.

1. data importation `lisrel2lavaan` currently supports `.csv`, `.dat`, and most other delimited data formats. However, formats that are specific to LISREL or PRELIS (e.g., the `.PSF` file format) cannot be imported. `lisrel2lavaan` supports raw data, covariance matrices, and correlation matrices (accompanied by a variance vector). Symmetric matrices can either contain lower triangle or full matrix. For MACS structure models, either raw data or summary statistics (that include a mean vector) are supported.
2. variable labels Certain variable labels that are permitted in LISREL cannot be supported in `lisrel2lavaan`. duplicate labels Most importantly, no two variables of any kind (including phantom variables) should be given the same label when using `lisrel2lavaan`. If multiple variables are given the same label, `lavaan` will estimate an incorrect model.
numeric character labels All variable labels are recommended to include non-numeric characters. In addition, the first character in each variable label is recommended to be non-numeric.
labels not specified If variable labels are not provided by the user, names will be generated reflecting variable assignment (e.g. 'eta1', 'ksi1'); manifest variables will be in lower case and latent variables in upper case.
3. OU paragraph Not all commands in the OU paragraph are presently supported in `lisrel2lavaan`. The ME command can be used to specify estimation method; however, not all estimations available in LISREL are currently supported by `lavaan`. If the specified ME is unsupported, `lisrel2lavaan` will revert to default estimation. The AD, EP, IT, ND and NP keywords will be ignored. Requests for text files containing starting values (e.g., OU BE) will also be ignored.
4. starting values Certain functionalities related to starting values in LISREL are not yet operational in `lisrel2lavaan`. Note that due to differences in estimation, starting values are not as important in `lavaan` model estimation as in LISREL. text file output Requests for text files containing starting values for individual matrices in the in the OU command (e.g., OU BE) are not currently supported. These requests will be ignored.
MA paragraph Specification of matrix starting values using the MA command is permitted by providing starting values within syntax directly. However, `lisrel2lavaan` has sometimes encountered problems with importation when files are specified following the MA paragraph.

Author(s)

Corbin Quick (University of Kansas; <corbing@ku.edu>)

Examples

```
## Not run:
## calling lisrel2lavaan without specifying the filename argument will
## open a file browser window with which LISREL syntax can be selected.

## any additional arguments to be passed to lavaan for data analysis can
## be specified normally.

lisrel2lavaan(se="standard")
## lavaan output summary printed to screen
## lavaan fit object returned silently
```

```
## manual file specification

lisrel2lavaan(filename="myFile.LS8", se="standard")
## lavaan output summary printed to screen
## lavaan fit object returned silently

## End(Not run)
```

loadingFromAlpha	<i>Find standardized factor loading from coefficient alpha</i>
------------------	--

Description

Find standardized factor loading from coefficient alpha assuming that all items have equal loadings.

Usage

```
loadingFromAlpha(alpha, ni)
```

Arguments

alpha	A desired coefficient alpha value.
ni	A desired number of items.

Value

result	The standardized factor loadings that make desired coefficient alpha with specified number of items.
--------	--

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

Examples

```
loadingFromAlpha(0.8, 4)
```

longInvariance	<i>Measurement Invariance Tests Within Person</i>
----------------	---

Description

Testing measurement invariance across timepoints (longitudinal) or any context involving the use of the same scale in one case (e.g., a dyad case with husband and wife answering the same scale). The measurement invariance uses a typical sequence of model comparison tests. This function currently works with only one scale.

Usage

```
longInvariance(model, varList, auto = "all", constrainAuto = FALSE,
fixed.x = TRUE, std.lv = FALSE, group=NULL, group.equal="",
group.partial="", warn=TRUE, debug=FALSE, strict = FALSE, quiet = FALSE,
...)
```

Arguments

model	lavaan syntax or parameter table
varList	A list containing indicator names of factors used in the invariance testing, such as the list that the first element is the vector of indicator names in the first time-point and the second element is the vector of indicator names in the second timepoint. The order of indicator names should be the same (but measured in different times or different units).
auto	The order of autocorrelation on the measurement errors on the similar items across factor (e.g., Item 1 in Time 1 and Time 2). If 0 is specified, the autocorrelation will be not imposed. If 1 is specified, the autocorrelation will imposed for the adjacent factor listed in <code>varList</code> . The maximum number can be specified is the number of factors specified minus 1. If "all" is specified, the maximum number of order will be used.
constrainAuto	If TRUE, the function will equate the auto-covariance to be equal within the same item across factors. For example, the covariance of item 1 in time 1 and time 2 is equal to the covariance of item 1 in time 2 and time 3.
fixed.x	See lavaan.
std.lv	See lavaan.
group	See lavaan.
group.equal	See lavaan.
group.partial	See lavaan.
warn	See lavaan.
debug	See lavaan.
strict	If TRUE, the sequence requires 'strict' invariance. See details for more information.
quiet	If TRUE, a summary is printed out containing an overview of the different models that are fitted, together with some model comparison tests.
...	Additional arguments in the lavaan function.

Details

If `strict = FALSE`, the following four models are tested in order:

1. Model 1: configural invariance. The same factor structure is imposed on all units.
2. Model 2: weak invariance. The factor loadings are constrained to be equal across units.
3. Model 3: strong invariance. The factor loadings and intercepts are constrained to be equal across units.
4. Model 4: The factor loadings, intercepts and means are constrained to be equal across units.

Each time a more restricted model is fitted, a chi-square difference test is reported, comparing the current model with the previous one, and comparing the current model to the baseline model (Model 1). In addition, the difference in cfi is also reported (delta.cfi).

If `strict = TRUE`, the following five models are tested in order:

1. Model 1: configural invariance. The same factor structure is imposed on all units.
2. Model 2: weak invariance. The factor loadings are constrained to be equal across units.
3. Model 3: strong invariance. The factor loadings and intercepts are constrained to be equal across units.
4. Model 4: strict invariance. The factor loadings, intercepts and residual variances are constrained to be equal across units.
5. Model 5: The factor loadings, intercepts, residual variances and means are constrained to be equal across units.

Note that if the chi-square test statistic is scaled (eg. a Satorra-Bentler or Yuan-Bentler test statistic), a special version of the chi-square difference test is used as described in <http://www.statmodel.com/chidiff.shtml>

Value

Invisibly, all model fits in the sequence are returned as a list.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>); Yves Rosseel (Ghent University; <Yves.Rosseel@UGent.be>)

References

Vandenberg, R. J., and Lance, C. E. (2000). A review and synthesis of the measurement invariance literature: Suggestions, practices, and recommendations for organizational research. *Organizational Research Methods*, 3, 4-70.

See Also

`measurementinvariance` For the measurement invariance test between groups

Examples

```
model <- ' f1t1 =~ y1t1 + y2t1 + y3t1
          f1t2 =~ y1t2 + y2t2 + y3t2
          f1t3 =~ y1t3 + y2t3 + y3t3'

# Create list of variables
var1 <- c("y1t1", "y2t1", "y3t1")
var2 <- c("y1t2", "y2t2", "y3t2")
var3 <- c("y1t3", "y2t3", "y3t3")
constrainedVar <- list(var1, var2, var3)

# Invariance of the same factor across timepoints
longInvariance(model, auto=1, constrainAuto=TRUE, varList=constrainedVar, data=exLong)

# Invariance of the same factor across timepoints and groups
longInvariance(model, auto=1, constrainAuto=TRUE, varList=constrainedVar, data=exLong, group.equal=c("loadings", "intercepts"))
```

mardiaKurtosis	<i>Finding Mardia's multivariate kurtosis</i>
----------------	---

Description

Finding Mardia's multivariate kurtosis of multiple variables

Usage

```
mardiaKurtosis(dat)
```

Arguments

dat The target matrix or data frame with multiple variables

Details

The Mardia's multivariate kurtosis formula (Mardia, 1970) is

$$b_{2,d} = \frac{1}{n} \sum_{i=1}^n \left[(\mathbf{X}_i - \bar{\mathbf{X}})' \mathbf{S}^{-1} (\mathbf{X}_i - \bar{\mathbf{X}}) \right]^2,$$

where d is the number of variables, X is the target dataset with multiple variables, n is the sample size, S is the sample covariance matrix of the target dataset, and \bar{X} is the mean vectors of the target dataset binded in n rows. When the population multivariate kurtosis is normal, the $b_{2,d}$ is asymptotically distributed as normal distribution with the mean of $d(d+2)$ and variance of $8d(d+2)/n$.

Value

A value of a Mardia's multivariate kurtosis with a test statistic

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

Mardia, K. V. (1970). Measures of multivariate skewness and kurtosis with applications. *Biometrika*, 57, 519-530.

See Also

- `skew` Find the univariate skewness of a variable
- `kurtosis` Find the univariate excessive kurtosis of a variable
- `mardiaSkew` Find the Mardia's multivariate skewness of a set of variables

Examples

```
library(lavaan)
mardiaKurtosis(HolzingerSwineford1939[,paste("x", 1:9, sep="")])
```

mardiaSkew

*Finding Mardia's multivariate skewness***Description**

Finding Mardia's multivariate skewness of multiple variables

Usage

```
mardiaSkew(dat)
```

Arguments

`dat` The target matrix or data frame with multiple variables

Details

The Mardia's multivariate skewness formula (Mardia, 1970) is

$$b_{1,d} = \frac{1}{n^2} \sum_{i=1}^n \sum_{j=1}^n \left[(\mathbf{X}_i - \bar{\mathbf{X}})' \mathbf{S}^{-1} (\mathbf{X}_j - \bar{\mathbf{X}}) \right]^3,$$

where d is the number of variables, \mathbf{X} is the target dataset with multiple variables, n is the sample size, \mathbf{S} is the sample covariance matrix of the target dataset, and $\bar{\mathbf{X}}$ is the mean vectors of the target dataset binded in n rows. When the population multivariate skewness is normal, the $\frac{n}{6}b_{1,d}$ is asymptotically distributed as chi-square distribution with $d(d+1)(d+2)/6$ degrees of freedom.

Value

A value of a Mardia's multivariate skewness with a test statistic

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

Mardia, K. V. (1970). Measures of multivariate skewness and kurtosis with applications. *Biometrika*, 57, 519-530.

See Also

- `skew` Find the univariate skewness of a variable
- `kurtosis` Find the univariate excessive kurtosis of a variable
- `mardiaKurtosis` Find the Mardia's multivariate kurtosis of a set of variables

Examples

```
library(lavaan)
mardiaSkew(HolzingerSwineford1939[,paste("x", 1:9, sep="")])
```

measurementInvariance

Measurement Invariance Tests

Description

Testing measurement invariance across groups using a typical sequence of model comparison tests.

Usage

```
measurementInvariance(..., strict = FALSE, quiet = FALSE)
```

Arguments

<code>...</code>	The same arguments as for any lavaan model. See <code>cfa</code> for more information.
<code>strict</code>	If <code>TRUE</code> , the sequence requires ‘strict’ invariance. See details for more information.
<code>quiet</code>	If <code>TRUE</code> , a summary is printed out containing an overview of the different models that are fitted, together with some model comparison tests.

Details

If `strict = FALSE`, the following four models are tested in order:

1. Model 1: configural invariance. The same factor structure is imposed on all groups.
2. Model 2: weak invariance. The factor loadings are constrained to be equal across groups.
3. Model 3: strong invariance. The factor loadings and intercepts are constrained to be equal across groups.
4. Model 4: The factor loadings, intercepts and means are constrained to be equal across groups.

Each time a more restricted model is fitted, a chi-square difference test is reported, comparing the current model with the previous one, and comparing the current model to the baseline model (Model 1). In addition, the difference in cfi is also reported (`delta.cfi`).

If `strict = TRUE`, the following five models are tested in order:

1. Model 1: configural invariance. The same factor structure is imposed on all groups.
2. Model 2: weak invariance. The factor loadings are constrained to be equal across groups.
3. Model 3: strong invariance. The factor loadings and intercepts are constrained to be equal across groups.
4. Model 4: strict invariance. The factor loadings, intercepts and residual variances are constrained to be equal across groups.
5. Model 5: The factor loadings, intercepts, residual variances and means are constrained to be equal across groups.

Note that if the chi-square test statistic is scaled (eg. a Satorra-Bentler or Yuan-Bentler test statistic), a special version of the chi-square difference test is used as described in <http://www.statmodel.com/chidiff.shtml>

Value

Invisibly, all model fits in the sequence are returned as a list.

Author(s)

Yves Rosseel <Yves.Rosseel@UGent.be>

References

Vandenberg, R. J., and Lance, C. E. (2000). A review and synthesis of the measurement invariance literature: Suggestions, practices, and recommendations for organizational research. *Organizational Research Methods*, 3, 4-70.

See Also

longInvariance For the measurement invariance test within person

Examples

```
HW.model <- ' visual =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed =~ x7 + x8 + x9 '

measurementInvariance(HW.model, data=HolzingerSwineford1939, group="school")
```

miPowerFit

Modification indices and their power approach for model fit evaluation

Description

The model fit evaluation approach using modification indices and their power proposed by Saris, Satorra, and van der Veld (2009, pp. 570-573).

Usage

```
miPowerFit(lavaanObj, stdLoad=0.4, cor=0.1, stdBeta=0.1, intcept=0.2, stdDelta=NULL,
delta=NULL)
```

Arguments

lavaanObj	The lavaan model object used to evaluate model fit
stdLoad	The amount of standardized factor loading that one would like to be detected (rejected). The default value is 0.4, which is suggested by Saris and colleagues (2009, p. 571).
cor	The amount of factor or error correlations that one would like to be detected (rejected). The default value is 0.1, which is suggested by Saris and colleagues (2009, p. 571).
stdBeta	The amount of standardized regression coefficients that one would like to be detected (rejected). The default value is 0.1, which is suggested by Saris and colleagues (2009, p. 571).

intcept	The amount of standardized intercept (similar to Cohen's d that one would like to be detected (rejected). The default value is 0.2, which is equivalent to a low effect size proposed by Cohen (1988, 1992).
stdDelta	The vector of the standardized parameters that one would like to be detected (rejected). If this argument is specified, the value here will overwrite the other arguments above. The order of the vector must be the same as the row order from modification indices from the <code>lavaan</code> object. If a single value is specified, the value will be applied to all parameters.
delta	The vector of the unstandardized parameters that one would like to be detected (rejected). If this argument is specified, the value here will overwrite the other arguments above. The order of the vector must be the same as the row order from modification indices from the <code>lavaan</code> object. If a single value is specified, the value will be applied to all parameters.

Details

In the `lavaan` object, one can inspect the modification indices and expected parameter changes. Those values can be used to evaluate model fit by the method proposed by Saris and colleagues (2009). First, one should evaluate whether the modification index of each parameter is significant. Second, one should evaluate whether the power to detect a target expected parameter change is high enough. If the modification index is not significant and the power is high, there is no misspecification. If the modification index is significant and the power is low, the fixed parameter is misspecified. If the modification index is significant and the power is high, the expected parameter change is investigated. If the expected parameter change is large (greater than the target expected parameter change), the parameter is misspecified. If the expected parameter change is low (lower than the target expected parameter change), the parameter is not misspecified. If the modification index is not significant and the power is low, the decision is inconclusive.

Value

A data frame with these variables:

1. lhs The left-hand side variable (with respect to the `lavaan` operator)
2. op The `lavaan` syntax operator: "`~~`" represents covariance, "`=~`" represents factor loading, "`~`" represents regression, and "`~1`" represents intercept.
3. rhs The right-hand side variable (with respect to the `lavaan` operator)
4. group The group of the parameter
5. mi The modification index of the fixed parameter
6. epc The expected parameter change if the parameter is freely estimated
7. target.epc The target expected parameter change that represents the minimum size of misspecification that one would like to be detected by the test with a high power
8. std.epc The standardized expected parameter change if the parameter is freely estimated
9. std.target.epc The standardized target expected parameter change
10. significant.mi Represents whether the modification index value is significant
11. high.power Represents whether the power is enough to detect the target expected parameter change
12. decision The decision whether the parameter is misspecified or not: "`M`" represents the parameter is misspecified, "`NM`" represents the parameter is not misspecified, "`EPC:M`" represents the parameter is misspecified decided by checking the expected parameter change value, "`EPC:NM`" represents the parameter is not misspecified decided by checking the expected parameter change value, and "`I`" represents the decision is inconclusive.

The row numbers matches with the results obtained from the `inspect(object, "mi")` function.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

Cohen, J. (1988). *Statistical power analysis for the behavioral sciences* (2nd ed.). Hillsdale, NJ: Erlbaum.

Cohen, J. (1992). A power primer. *Psychological Bulletin*, 112, 155-159.

Saris, W. E., Satorra, A., & van der Veld, W. M. (2009). Testing structural equation models or detection of misspecifications? *Structural Equation Modeling*, 16, 561-582.

See Also

`moreFitIndices` For the additional fit indices information

Examples

```
library(lavaan)

HS.model <- ' visual  =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed   =~ x7 + x8 + x9 '

fit <- cfa(HS.model, data=HolzingerSwineford1939, group="sex", meanstructure=TRUE)
miPowerFit(fit)

model <- '
  # latent variable definitions
  ind60 =~ x1 + x2 + x3
  dem60 =~ y1 + a*y2 + b*y3 + c*y4
  dem65 =~ y5 + a*y6 + b*y7 + c*y8

  # regressions
  dem60 ~ ind60
  dem65 ~ ind60 + dem60

  # residual correlations
  y1 ~~ y5
  y2 ~~ y4 + y6
  y3 ~~ y7
  y4 ~~ y8
  y6 ~~ y8
  '

fit2 <- sem(model, data=PoliticalDemocracy, meanstructure=TRUE)
miPowerFit(fit2, stdLoad=0.3, cor=0.2, stdBeta=0.2, intcept=0.5)
```

Description

This function takes an expression for an indirect effect, the parameters and standard errors associated with the expression and returns a confidence interval based on a Monte Carlo test of mediation (MacKinnon, Lockwood, & Williams, 2004).

Usage

```
monteCarloMed(expression, ..., ACM=NULL, rep=20000, CI=95, plot=FALSE, outputVal
```

Arguments

<code>expression</code>	A character scalar representing the computation of an indirect effect. Different parameters in the expression should have different alphanumeric values. Expressions can use either addition (+) or multiplication (*) operators.
<code>...</code>	Parameter estimates for all parameters named in <code>expression</code> . The order of parameters should follow from <code>expression</code> (the first parameter named in <code>expression</code> should be the first parameter listed in <code>...</code>). Alternatively <code>...</code> can be a vector of parameter estimates.
<code>ACM</code>	A matrix representing the asymptotic covariance matrix of the parameters described in <code>expression</code> . This matrix should be a symmetric matrix with dimensions equal to the number of parameters names in <code>expression</code> . Information on finding the ACOV in popular SEM software is described below.)
<code>rep</code>	The number of replications to compute. Many thousand are recommended.
<code>CI</code>	Width of the confidence interval computed.
<code>plot</code>	Should the function output a plot of simulated values of the indirect effect?
<code>outputValues</code>	Should the function output all simulated values of the indirect effect?

Details

This function implements the Monte Carlo test of mediation first described in MacKinnon, Lockwood, & Williams (2004) and extends it to complex cases where the indirect effect is more than a function of two parameters. The function takes an expression for the indirect effect, randomly simulated values of the indirect effect based on the values of the parameters (and the associated standard errors) comprising the indirect effect, and outputs a confidence interval of the indirect effect based on the simulated values. For further information on the Monte Carlo test of mediation see MacKinnon, Lockwood, & Williams (2004), Preacher & Selig (in press), and Selig & Preacher (2008). For a Monte Carlo test of mediation with a random effects model see Selig & Preacher (2010).

The asymptotic covariance matrix can be easily found in many popular SEM software applications.

- **LISREL** Including the EC option on the OU line will print the ACM to a separate file. The file contains the lower triangular elements of the ACM in free format and scientific notation
- **Mplus** Include the command TECH3; in the OUTPUT section. The ACM will be printed in the output.
- **lavaan** Use the command `vcov` on the fitted lavaan object to print the ACM to the screen

Value

A list with two elements. The first element is the point estimate for the indirect effect. The second element is a matrix with values for the upper and lower limits of the confidence interval generated from the Monte Carlo test of mediation. If `outputValues=TRUE`, output will be a list with a list with the point estimate and values for the upper and lower limits of the confidence interval as the first element and a vector of simulated values of the indirect effect as the second element.

Author(s)

Corbin Quick (University of Kansas; <corbinq@ku.edu>) Alexander M. Schoemann (East Carolina University; <schoemanna@ecu.edu>) James P. Selig (University of New Mexico; <selig@unm.edu>)

References

Preacher, K. J., & Selig, J. P. (2010, July). Monte Carlo method for assessing multilevel mediation: An interactive tool for creating confidence intervals for indirect effects in 1-1-1 multilevel models [Computer software]. Available from <http://quantpsy.org/>.

Preacher, K. J., & Selig, J. P. (in press). Advantages of Monte Carlo confidence intervals for indirect effects. *Communication Methods and Measures*.

Selig, J. P., & Preacher, K. J. (2008, June). Monte Carlo method for assessing mediation: An interactive tool for creating confidence intervals for indirect effects [Computer software]. Available from <http://quantpsy.org/>.

Examples

```
#Simple two path mediation
#Write expression of indirect effect
med <- 'a*b'
#Parameter values from analyses
aparam <- 1
bparam<-2
#Asymptotic covariance matrix from analyses
AC <- matrix(c(.01,.00002,
               .00002,.02), nrow=2, byrow=TRUE)
#Compute CI, include a plot
monteCarloMed(med, coef1=aparam, coef2=bparam, outputValues=FALSE, plot=TRUE, ACM=AC)

#Use a matrix of parameter estimates as input
aparam<-c(1,2)
monteCarloMed(med, coef1=aparam, outputValues=FALSE, plot=TRUE, ACM=AC)

#complex mediation with two paths for the indirect effect
#Write expression of indirect effect
med <- 'a1*b1 + a1*b2'
#Parameter values and standard errors from analyses
aparam <- 1
b1param<-2
b2param<-1
#Asymptotic covariance matrix from analyses
AC <- matrix(c(1,.00002, .00003,
               .00002,1, .00002,
```

```
.00003, .00002, 1), nrow=3, byrow=TRUE)
#Compute CI do not include a plot
monteCarloMed(med, coef1=aparam, coef2=b1param, coef3=b2param, ACM=AC)
```

moreFitIndices	<i>Calculate more fit indices</i>
----------------	-----------------------------------

Description

Calculate more fit indices that are not already provided in lavaan.

Usage

```
moreFitIndices(object, nPrior = 1)
```

Arguments

object	The lavaan model object provided after running the <code>cfa</code> , <code>sem</code> , <code>growth</code> , or <code>lavaan</code> functions.
nPrior	The sample size on which prior is based. This argument is used to compute BIC*.

Details

Gamma Hat (`gammaHat`; West, Taylor, & Wu, 2012) is a global fit index which can be computed by

$$gammaHat = \frac{p}{p + 2 \times \frac{\chi_k^2 - df_k}{N-1}},$$

where p is the number of variables in the model, χ_k^2 is the chi-square test statistic value of the target model, df_k is the degree of freedom when fitting the target model, and N is the sample size. This formula assumes equal number of indicators across groups.

Adjusted Gamma Hat (`adjGammaHat`; West, Taylor, & Wu, 2012) is a global fit index which can be computed by

$$adjGammaHat = \left(1 - \frac{K \times p \times (p + 1)}{2 \times df_k}\right) \times (1 - gammaHat),$$

where K is the number of groups (please refer to Dudgeon, 2004 for the multiple-group adjustment for `agfi*`).

Corrected Akaike Information Criterion (`aic.smallN`; Burnham & Anderson, 2003) is the corrected version of `aic` for small sample size:

$$aic.smallN = f + \frac{2k(k + 1)}{N - k - 1},$$

where f is the minimized discrepancy function, which is the product of the log likelihood and -2, and k is the number of parameters in the target model.

Corrected Bayesian Information Criterion (`bic.priorN`; Kuha, 2004) is similar to `bic` but explicitly specifying the sample size on which the prior is based (N_{prior}).

$$bic.priorN = f + k \log(1 + N/N_{prior}),$$

Stochastic information criterion (`sic`; Preacher, 2006) is similar to `aic` or `bic`. This index will account for model complexity in the model's function form, in addition to the number of free parameters. This index will be provided only when the chi-squared value is not scaled. `sic` can be computed by

$$sic = \frac{1}{2} \left(f - \log \det I(\hat{\theta}) \right),$$

where $I(\hat{\theta})$ is the information matrix of the parameters.

Hannan-Quinn Information Criterion (`hqc`; Hannan & Quinn, 1979) is used for model selection similar to `aic` or `bic`.

$$hqc = f + 2k \log(\log N),$$

Note that if Satorra-Bentler or Yuan-Bentler's method is used, the fit indices using the scaled chi-square values are also provided.

See `nullRMSEA` for the further details of the computation of RMSEA of the null model.

Value

1. `gammaHat` Gamma Hat
2. `adjGammaHat` Adjusted Gamma Hat
3. `baseline.rmsea` RMSEA of the Baseline (Null) Model
4. `aic.smallN` Corrected (for small sample size) Akaike Information Criterion
5. `bic.priorN` Bayesian Information Criterion with specifying the prior sample size
6. `sic` Stochastic Information Criterion
7. `hqc` Hannan-Quinn Information Criterion
8. `gammaHat.scaled` Gamma Hat using Scaled Chi-square
9. `adjGammaHat.scaled` Adjusted Gamma Hat using Scaled Chi-square
10. `baseline.rmsea.scaled` RMSEA of the Baseline (Null) Model using Scaled Chi-square

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>) Aaron Boulton (University of Kansas; <aboulton@ku.edu>) Ruben Arslan (Humboldt-University of Berlin, <rubenarslan@gmail.com>) Terrence Jorgensen (University of Kansas; <tdj@ku.edu>)

References

- Burnham, K., & Anderson, D. (2003). *Model selection and multimodel inference: A practical-theoretic approach*. New York, NY: Springer-Verlag.
- Dudgeon, P. (2004). A note on extending Steiger's (1998) multiple sample RMSEA adjustment to other noncentrality parameter-based statistic. *Structural Equation Modeling*, 11, 305-319.
- Kuha, J. (2004). AIC and BIC: Comparisons of assumptions and performance. *Sociological Methods Research*, 33, 188-229.

Preacher, K. J. (2006). Quantifying parsimony in structural equation modeling. *Multivariate Behavioral Research*, 43, 227-259.

West, S. G., Taylor, A. B., & Wu, W. (2012). Model fit and model selection in structural equation modeling. In R. H. Hoyle (Ed.), *Handbook of Structural Equation Modeling*. New York: Guilford.

See Also

- `miPowerFit` For the modification indices and their power approach for model fit evaluation
- `nullRMSEA` For RMSEA of the null model

Examples

```
HS.model <- ' visual  =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed   =~ x7 + x8 + x9 '
```

```
fit <- cfa(HS.model, data=HolzingerSwineford1939)
moreFitIndices(fit)
```

```
fit2 <- cfa(HS.model, data=HolzingerSwineford1939, estimator="mlr")
moreFitIndices(fit2)
```

 net

Nesting and Equivalence Testing

Description

This test examines whether models are nested or equivalent based on Bentler and Satorra's (2010) procedure.

Usage

```
net(..., crit = .0001)
```

Arguments

<code>...</code>	The lavaan objects used for test of nesting and equivalence
<code>crit</code>	The upper-bound criterion for testing the equivalence of models. Models are considered nested (or equivalent) if the difference between their chi-squared fit statistics is less than this criterion.

Value

The Net object representing the outputs for nesting and equivalent testing, including a logical matrix of test results and a vector of degrees of freedom for each model.

Author(s)

Terrence D. Jorgensen (University of Kansas; <TJorgensen314@gmail.com>)

References

Bentler, P. M., & Satorra, A. (2010). Testing model nesting and equivalence. *Psychological Methods*, 15, 111-123. doi:10.1037/a0019625

Examples

```
m1 <- ' visual  =~ x1 + x2 + x3
textual =~ x4 + x5 + x6
speed   =~ x7 + x8 + x9 '

m2 <- ' f1  =~ x1 + x2 + x3 + x4
f2 =~ x5 + x6 + x7 + x8 + x9 '

m3 <- ' visual  =~ x1 + x2 + x3
textual =~ eq*x4 + eq*x5 + eq*x6
speed   =~ x7 + x8 + x9 '

fit1 <- cfa(m1, data = HolzingerSwineford1939)
fit1a <- cfa(m1, data = HolzingerSwineford1939, std.lv = TRUE) # Equivalent to fit1
fit2 <- cfa(m2, data = HolzingerSwineford1939) # Not equivalent to or nested in fit1
fit3 <- cfa(m3, data = HolzingerSwineford1939) # Nested in fit1 and fit1a

tests <- net(fit1, fit1a, fit2, fit3)
tests
summary(tests)
```

Net-class

Class For the Result of Nesting and Equivalence Testing

Description

This class contains the results of nesting and equivalence testing among multiple models

Objects from the Class

Objects can be created via the `net` function.

Slots

test: Logical matrix of results of nesting and equivalence testing across models
df: The degrees of freedom of tested models

methods

- `summary` The summary function is used to provide the results in narrative.

Author(s)

Terrence D. Jorgensen (University of Kansas; <TJorgensen314@gmail.com>)

See Also

net

Examples

```
# See the example in the net function.
```

nullMx

Analyzing data using a null model

Description

Analyzing data using a null model by full-information maximum likelihood. In the null model, all means and covariances are free if items are continuous. All covariances are fixed to 0. For ordinal variables, their means are fixed as 0 and their variances are fixed as 1 where their thresholds are estimated. In multiple-group model, all means and variances are separately estimated.

Usage

```
nullMx(data, groupLab = NULL)
```

Arguments

data	The target data frame
groupLab	The name of grouping variable

Value

The `MxModel` object which contains the analysis result of the null model.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

saturateMx, fitMeasuresMx, standardizeMx

Examples

```
## Not run:
library(OpenMx)
data(demoOneFactor)
nullModel <- nullMx(demoOneFactor)

## End(Not run)
```

nullRMSEA

Calculate the RMSEA of the null model

Description

Calculate the RMSEA of the null (baseline) model

Usage

```
nullRMSEA(object, scaled = FALSE, silent=FALSE)
```

Arguments

<code>object</code>	The lavaan model object provided after running the <code>cfa</code> , <code>sem</code> , <code>growth</code> , or <code>lavaan</code> functions.
<code>scaled</code>	If TRUE, calculate the null model from the scaled test.
<code>silent</code>	If TRUE, do not print anything on the screen.

Details

RMSEA of the null model is calculated similar to the formula provided in the `lavaan` package. The standard formula of RMSEA is

$$RMSEA = \sqrt{\frac{\chi^2}{N \times df} - \frac{1}{N}} \times \sqrt{G}$$

where χ^2 is the chi-square test statistic value of the target model, N is the total sample size, df is the degree of freedom of the hypothesized model, G is the number of groups. Kenny proposed in his website that

"A reasonable rule of thumb is to examine the RMSEA for the null model and make sure that is no smaller than 0.158. An RMSEA for the model of 0.05 and a TLI of .90, implies that the RMSEA of the null model is 0.158. If the RMSEA for the null model is less than 0.158, an incremental measure of fit may not be that informative."

See <http://davidakenny.net/cm/fit.htm>.

Value

A value of RMSEA of the null model. This value is hidden. Users may be assigned the output of this function to any object for further usage.

Author(s)

Ruben Arslan (Humboldt-University of Berlin, <rubenarslan@gmail.com>)

References

Kenny, D. A., Kaniskan, B., & McCoach, D. B. (2011). *The performance of RMSEA in models with small degrees of freedom*. Unpublished paper, University of Connecticut.

See Also

- `miPowerFit` For the modification indices and their power approach for model fit evaluation
- `moreFitIndices` For other fit indices

Examples

```
HS.model <- ' visual  =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed   =~ x7 + x8 + x9 '

fit <- cfa(HS.model, data=HolzingerSwineford1939)
nullRMSEA(fit)
```

parcelAllocation *Random Allocation of Items to Parcels in a Structural Equation Model*

Description

This function generates a given number of randomly generated item-to-parcel allocations, fits a model to each allocation, and provides averaged results over all allocations.

Usage

```
parcelAllocation(nPerPar, facPlc, nAlloc=100, syntax, dataset, names='default',
  leaveout=0, ...)
```

Arguments

<code>nPerPar</code>	A list in which each element is a vector corresponding to each factor indicating sizes of parcels. If variables are left out of parceling, they should not be accounted for here (there should NOT be parcels of size "1").
<code>facPlc</code>	A list of vectors, each corresponding to a factor, specifying the variables in that factor (whether included in parceling or not). Either variable names or column numbers. Variables not listed will not be modeled or included in output datasets.
<code>nAlloc</code>	The number of random allocations of items to parcels to generate.
<code>syntax</code>	lavaan syntax. If substituted with a file name, <code>parcelAllocation</code> will print output data sets to a specified folder rather than analyzing using lavaan (note for Windows users: file path must be specified using forward slashes).
<code>dataset</code>	Data set. Can be file path or R object (matrix or dataframe). If the data has missing values multiple imputation before parceling is recommended.
<code>names</code>	(Optional) A character vector containing the names of parceled variables.
<code>leaveout</code>	A vector of variables to be left out of randomized parceling. Either variable names or column numbers are allowed.
<code>...</code>	Additional arguments to be passed to lavaan

Details

This function implements the random item to parcel allocation procedure described in Sterba (2011) and Sterba and MccCallum (2010). The function takes a single data set with item level data, randomly assigns items to parcels, fits a structural equation model to the parceled data (using lavaan), and repeats this process for a user specified number of random allocations. Results from all fitted models are summarized and output. For further details on the benefits of the random allocation of items to parcels see Sterba (2011) and Sterba and MccCallum (2010).

Value

Estimates	A data frame containing results related to parameter estimates with columns corresponding to parameter names, average parameter estimates across allocations, the standard deviation of parameter estimates across allocations, the minimum parameter estimate across allocations, the maximum parameter estimate across allocations, the range of parameter estimates across allocations, and the proportions of allocations in which the parameter estimate is significant.
SE	A data frame containing results related to standard errors with columns corresponding to parameter names, average standard errors across allocations, the standard deviation of standard errors across allocations, the minimum standard error across allocations, the maximum standard error across allocations, and the range of standard errors across allocations.
Fit	A data frame containing results related to model fit with columns corresponding to fit index names, the average of each index across allocations, the standard deviation of each fit index across allocations, the minimum of each fit index across allocations, the maximum of each fit index across allocations, and the range of each fit index across allocations.

Author(s)

Corbin Quick (University of Kansas; <corbinq@ku.edu>) Alexander M. Schoemann (East Carolina University; <schoemanna@ecu.edu>)

References

- Sterba, S.K. (2011). Implications of parcel-allocation variability for comparing fit of item-solutions and parcel-solutions. *Structural Equation Modeling*, 18, 554-577.
- Sterba, S.K. & MacCallum, R.C. (2010). Variability in parameter estimates and model fit across random allocations of items to parcels. *Multivariate Behavioral Research*, 45, 322-358.

Examples

```
#Fit 3 factor CFA to simulated data.
#Each factor has 9 indicators that are randomly parceled into 3 parcels
#Lavaan syntax for the model to be fit to parceled data
syntax <- 'La =~ V1 + V2 + V3
          Lb =~ V4 + V5 + V6
          '

#Parcel and fit data 20 times. The actual parcel number should be higher than 20 times.
name1 <- colnames(simParcel)[1:9]
name2 <- colnames(simParcel)[10:18]
parcelAllocation(list(c(3,3,3),c(3,3,3)), list(name1, name2), nAlloc=20, syntax=syntax,
dataset=simParcel)
```

plotProbe

Plot the graphs for probing latent interaction

Description

This function will plot the line graphs representing the simple effect of the independent variable given the values of the moderator.

Usage

```
plotProbe(object, xlim, xlab="Indepedent Variable", ylab="Dependent Variable", .
```

Arguments

object	The result of probing latent interaction obtained from <code>probe2WayMC</code> , <code>probe2WayRC</code> , <code>probe3WayMC</code> , or <code>probe3WayRC</code> function.
xlim	The vector of two numbers: the minimum and maximum values of the independent variable
xlab	The label of the x-axis
ylab	The label of the y-axis
...	Any addition argument for the <code>plot</code> function

Value

None. This function will plot the simple main effect only.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

- `indProd` For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- `probe2WayMC` For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe3WayMC` For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe2WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `probe3WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.

Examples

```

library(lavaan)

dat2wayMC <- indProd(dat2way, 1:3, 4:6)

modell1 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f12 =~ x1.x4 + x2.x5 + x3.x6
f3 =~ x7 + x8 + x9
f3 ~ f1 + f2 + f12
f12 ~~ 0*f1
f12 ~~ 0*f2
x1 ~ 0*1
x4 ~ 0*1
x1.x4 ~ 0*1
x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f12 ~ NA*1
f3 ~ NA*1
"

fitMC2way <- sem(modell1, data=dat2wayMC, meanstructure=TRUE, std.lv=FALSE)
result2wayMC <- probe2WayMC(fitMC2way, c("f1", "f2", "f12"), "f3", "f2", c(-1, 0, 1))
plotProbe(result2wayMC, xlim=c(-2, 2))

dat3wayMC <- indProd(dat3way, 1:3, 4:6, 7:9)

modell3 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f3 =~ x7 + x8 + x9
f12 =~ x1.x4 + x2.x5 + x3.x6
f13 =~ x1.x7 + x2.x8 + x3.x9
f23 =~ x4.x7 + x5.x8 + x6.x9
f123 =~ x1.x4.x7 + x2.x5.x8 + x3.x6.x9
f4 =~ x10 + x11 + x12
f4 ~ f1 + f2 + f3 + f12 + f13 + f23 + f123
f1 ~~ 0*f12
f1 ~~ 0*f13
f1 ~~ 0*f123
f2 ~~ 0*f12
f2 ~~ 0*f23
f2 ~~ 0*f123
f3 ~~ 0*f13
f3 ~~ 0*f23
f3 ~~ 0*f123
f12 ~~ 0*f123
f13 ~~ 0*f123
f23 ~~ 0*f123
x1 ~ 0*1
x4 ~ 0*1
x7 ~ 0*1
x10 ~ 0*1

```

```

x1.x4 ~ 0*1
x1.x7 ~ 0*1
x4.x7 ~ 0*1
x1.x4.x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f3 ~ NA*1
f12 ~ NA*1
f13 ~ NA*1
f23 ~ NA*1
f123 ~ NA*1
f4 ~ NA*1
"

fitMC3way <- sem(model3, data=dat3wayMC, meanstructure=TRUE, std.lv=FALSE)
result3wayMC <- probe3WayMC(fitMC3way, c("f1", "f2", "f3", "f12", "f13", "f23", "f123"),
"f4", c("f1", "f2"), c(-1, 0, 1), c(-1, 0, 1))
plotProbe(result3wayMC, xlim=c(-2, 2))

```

plotRMSEAdist

Plot the sampling distributions of RMSEA

Description

Plots the sampling distributions of RMSEA based on the noncentral chi-square distributions

Usage

```
plotRMSEAdist(rmseas, n, df, ptile=NULL, caption=NULL, rmseaScale = TRUE, group=1)
```

Arguments

rmseas	The vector of RMSEA values to be plotted
n	Sample size of a dataset
df	Model degrees of freedom
ptile	The percentile rank of the distribution of the first RMSEA that users wish to plot a vertical line in the resulting graph
caption	The name vector of each element of rmseas
rmseaScale	If TRUE, the RMSEA scale is used in the x-axis. If FALSE, the chi-square scale is used in the x-axis.
group	The number of group that is used to calculate RMSEA.

Details

This function creates overlapping plots of the sampling distribution of RMSEA based on non-central chi-square distribution (MacCallum, Browne, & Suguwara, 1996). First, the noncentrality parameter (λ) is calculated from RMSEA (Steiger, 1998; Dudgeon, 2004) by

$$\lambda = (N - 1)d\varepsilon^2/K,$$

where N is sample size, d is the model degree of freedom, K is the number of group and ε is the population RMSEA. Next, the noncentral chi-square distribution with a specified degree of freedom

and noncentrality parameter is plotted. Thus, the x-axis represent the sample chi-square value. The sample chi-square value can be transformed to the sample RMSEA scale ($\hat{\varepsilon}$) by

$$\hat{\varepsilon} = \sqrt{K} \sqrt{\frac{\chi^2 - d}{(N - 1)d}},$$

where χ^2 is the chi-square value obtained from the noncentral chi-square distribution.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

Dudgeon, P. (2004). A note on extending Steiger's (1998) multiple sample RMSEA adjustment to other noncentrality parameter-based statistic. *Structural Equation Modeling*, 11, 305-319.

MacCallum, R. C., Browne, M. W., & Sugawara, H. M. (1996). Power analysis and determination of sample size for covariance structure modeling. *Psychological Methods*, 1, 130-149.

Steiger, J. H. (1998). A note on multiple sample extensions of the RMSEA fit index. *Structural Equation Modeling*, 5, 411-419.

See Also

- `plotRMSEApower` to plot the statistical power based on population RMSEA given the sample size
- `findRMSEApower` to find the statistical power based on population RMSEA given a sample size
- `findRMSEAsamplesize` to find the minium sample size for a given statistical power based on population RMSEA

Examples

```
plotRMSEAdist(rmse=c(.05, .08), n=200, df=20, ptile=0.95, rmseaScale = TRUE)
plotRMSEAdist(rmse=c(.05, .01), n=200, df=20, ptile=0.05, rmseaScale = FALSE)
```

<code>plotRMSEApower</code>	<i>Plot power curves for RMSEA</i>
-----------------------------	------------------------------------

Description

Plots power of RMSEA over a range of sample sizes

Usage

```
plotRMSEApower(rmse0, rmseaA, df, nlow, nhigh, steps=1, alpha=.05, group=1, ...)
```


Arguments

rmsea0	Null RMSEA
rmseaA	Alternative RMSEA
df	Model degrees of freedom
nlow	Lower sample size
nhigh	Upper sample size
steps	Increase in sample size for each iteration. Smaller values of steps will lead to more precise plots. However, smaller step sizes means a longer run time.
alpha	Alpha level used in power calculations
group	The number of group that is used to calculate RMSEA.
...	The additional arguments for the plot function.

Details

This function creates plot of power for RMSEA against a range of sample sizes. The plot places sample size on the horizontal axis and power on the vertical axis. The user should indicate the lower and upper values for sample size and the sample size between each estimate ("step size") We strongly urge the user to read the sources below (see References) before proceeding. A web version of this function is available at: <http://quantpsy.org/rmsea/rmseaplot.htm>.

Value

1. plot Plot of power for RMSEA against a range of sample sizes

Author(s)

Alexander M. Schoemann (East Carolina University; <schoemanna@ecu.edu>) Kristopher J. Preacher (Vanderbilt University; <kris.preacher@vanderbilt.edu>) Donna L. Coffman (Pennsylvania State University; <dlc30@psu.edu.>)

References

- MacCallum, R. C., Browne, M. W., & Cai, L. (2006). Testing differences between nested covariance structure models: Power analysis and null hypotheses. *Psychological Methods*, 11, 19-35.
- MacCallum, R. C., Browne, M. W., & Sugawara, H. M. (1996). Power analysis and determination of sample size for covariance structure modeling. *Psychological Methods*, 1, 130-149.
- MacCallum, R. C., Lee, T., & Browne, M. W. (2010). The issue of isopower in power analysis for tests of structural equation models. *Structural Equation Modeling*, 17, 23-41.
- Preacher, K. J., Cai, L., & MacCallum, R. C. (2007). Alternatives to traditional model comparison strategies for covariance structure models. In T. D. Little, J. A. Bovaird, & N. A. Card (Eds.), *Modeling contextual effects in longitudinal studies* (pp. 33-62). Mahwah, NJ: Lawrence Erlbaum Associates.
- Steiger, J. H. (1998). A note on multiple sample extensions of the RMSEA fit index. *Structural Equation Modeling*, 5, 411-419.
- Steiger, J. H., & Lind, J. C. (1980, June). *Statistically based tests for the number of factors*. Paper presented at the annual meeting of the Psychometric Society, Iowa City, IA.

See Also

- `plotRMSEAdist` to visualize the RMSEA distributions
- `findRMSEApower` to find the statistical power based on population RMSEA given a sample size
- `findRMSEAsamplesize` to find the minium sample size for a given statistical power based on population RMSEA

Examples

```
plotRMSEApower(.025, .075, 23, 100, 500, 10)
```

```
plotRMSEApowernested
```

Plot power of nested model RMSEA

Description

Plot power of nested model RMSEA over a range of possible sample sizes.

Usage

```
plotRMSEApowernested(rmse0A = NULL, rmse0B = NULL, rmse1A, rmse1B = NULL,
  dfA, dfB, nlow, nhigh, steps=1, alpha=.05, group=1, ...)
```

Arguments

<code>rmse0A</code>	The H0 baseline RMSEA.
<code>rmse0B</code>	The H0 alternative RMSEA (trivial misfit).
<code>rmse1A</code>	The H1 baseline RMSEA.
<code>rmse1B</code>	The H1 alternative RMSEA (target misfit to be rejected).
<code>dfA</code>	degree of freedom of the more-restricted model.
<code>dfB</code>	degree of freedom of the less-restricted model.
<code>nlow</code>	Lower bound of sample size.
<code>nhigh</code>	Upper bound of sample size.
<code>steps</code>	Step size.
<code>alpha</code>	The alpha level.
<code>group</code>	The number of group in calculating RMSEA.
<code>...</code>	The additional arguments for the plot function.

Author(s)

Bell Clinton (University of Kansas; <clintonbell@ku.edu>); Pavel Panko (University of Kansas; <pavel@ku.edu>); Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

MacCallum, R. C., Browne, M. W., & Cai, L. (2006). Testing differences between nested covariance structure models: Power analysis and null hypotheses. *Psychological Methods*, 11, 19-35.

See Also

- `findRMSEApowernested` to find the power for a given sample size in nested model comparison based on population RMSEA
- `findRMSEAsamplesizenested` to find the minimum sample size for a given statistical power in nested model comparison based on population RMSEA

Examples

```
plotRMSEApowernested(rmse0A = 0, rmse0B = 0, rmse1A = 0.06, rmse1B = 0.05,
  dfA=22, dfB=20, nlow=50, nhigh=500, steps=1, alpha=.05, group=1)
```

probe2WayMC	<i>Probing two-way interaction on the residual-centered latent interaction</i>
-------------	--

Description

Probing interaction for simple intercept and simple slope for the no-centered or mean-centered latent two-way interaction

Usage

```
probe2WayMC(fit, nameX, nameY, modVar, valProbe)
```

Arguments

<code>fit</code>	The lavaan model object used to evaluate model fit
<code>nameX</code>	The vector of the factor names used as the predictors. The first-order factor will be listed first. The last name must be the name representing the interaction term.
<code>nameY</code>	The name of factor that is used as the dependent variable.
<code>modVar</code>	The name of factor that is used as a moderator. The effect of the other independent factor on each moderator variable value will be probed.
<code>valProbe</code>	The values of the moderator that will be used to probe the effect of the other independent factor.

Details

Before using this function, researchers need to make the products of the indicators between the first-order factors using mean centering (Marsh, Wen, & Hau, 2004). Note that the double-mean centering may not be appropriate for probing interaction if researchers are interested in simple intercepts. The mean or double-mean centering can be done by the `indProd` function. The indicator products can be made for all possible combination or matched-pair approach (Marsh et al., 2004). Next, the hypothesized model with the regression with latent interaction will be used to fit all original indicators and the product terms. See the example for how to fit the product term below. Once the lavaan result is obtained, this function will be used to probe the interaction.

Let that the latent interaction model regressing the dependent variable (Y) on the independent variable (X) and the moderator (Z) be

$$Y = b_0 + b_1X + b_2Z + b_3XZ + r,$$

where b_0 is the estimated intercept or the expected value of Y when both X and Z are 0, b_1 is the effect of X when Z is 0, b_2 is the effect of Z when X is 0, b_3 is the interaction effect between X and Z , and r is the residual term.

For probing two-way interaction, the simple intercept of the independent variable at each value of the moderator (Aiken & West, 1991; Cohen, Cohen, West, & Aiken, 2003; Preacher, Curran, & Bauer, 2006) can be obtained by

$$b_{0|X=0,Z} = b_0 + b_2Z.$$

The simple slope of the independent variable at each value of the moderator can be obtained by

$$b_{X|Z} = b_1 + b_3Z.$$

The variance of the simple intercept formula is

$$Var(b_{0|X=0,Z}) = Var(b_0) + 2ZCov(b_0, b_2) + Z^2Var(b_2)$$

where Var denotes the variance of a parameter estimate and Cov denotes the covariance of two parameter estimates.

The variance of the simple slope formula is

$$Var(b_{X|Z}) = Var(b_1) + 2ZCov(b_1, b_3) + Z^2Var(b_3)$$

Wald statistic is used for test statistic.

Value

A list with two elements:

1. SimpleIntercept The intercepts given each value of the moderator. This element will be shown only if the factor intercept is estimated (e.g., not fixed as 0).
2. SimpleSlope The slopes given each value of the moderator.

In each element, the first column represents the values of the moderators specified in the `valProbe` argument. The second column is the simple intercept or simple slope. The third column is the standard error of the simple intercept or simple slope. The fourth column is the Wald (z) statistic. The fifth column is the p -value testing whether the simple intercepts or slopes are different from 0.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

- Aiken, L. S., & West, S. G. (1991). Multiple regression: Testing and interpreting interactions. Newbury Park, CA: Sage.
- Cohen, J., Cohen, P., West, S. G., & Aiken, L. S. (2003). Applied multiple regression/correlation analysis for the behavioral sciences (3rd ed.). New York: Routledge.
- Marsh, H. W., Wen, Z., & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, 9, 275-300.
- Preacher, K. J., Curran, P. J., & Bauer, D. J. (2006). Computational tools for probing interactions in multiple linear regression, multilevel modeling, and latent curve analysis. *Journal of Educational and Behavioral Statistics*, 31, 437-448.

See Also

- `indProd` For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- `probe3WayMC` For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe2WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `probe3WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `plotProbe` Plot the simple intercepts and slopes of the latent interaction.

Examples

```
library(lavaan)

dat2wayMC <- indProd(dat2way, 1:3, 4:6)

modell <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f12 =~ x1.x4 + x2.x5 + x3.x6
f3 =~ x7 + x8 + x9
f3 ~ f1 + f2 + f12
f12 ~~ 0*f1
f12 ~~ 0*f2
x1 ~ 0*1
x4 ~ 0*1
x1.x4 ~ 0*1
x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f12 ~ NA*1
f3 ~ NA*1
"

fitMC2way <- sem(modell, data=dat2wayMC, meanstructure=TRUE, std.lv=FALSE)
summary(fitMC2way)

result2wayMC <- probe2WayMC(fitMC2way, c("f1", "f2", "f12"), "f3", "f2", c(-1, 0, 1))
result2wayMC
```

probe2WayRC

Probing two-way interaction on the residual-centered latent interaction

Description

Probing interaction for simple intercept and simple slope for the residual-centered latent two-way interaction (Pornprasertmanit, Schoemann, Geldhof, & Little, submitted)

Usage

```
probe2WayRC(fit, nameX, nameY, modVar, valProbe)
```

Arguments

<code>fit</code>	The lavaan model object used to evaluate model fit
<code>nameX</code>	The vector of the factor names used as the predictors. The first-order factor will be listed first. The last name must be the name representing the interaction term.
<code>nameY</code>	The name of factor that is used as the dependent variable.
<code>modVar</code>	The name of factor that is used as a moderator. The effect of the other independent factor on each moderator variable value will be probed.
<code>valProbe</code>	The values of the moderator that will be used to probe the effect of the other independent factor.

Details

Before using this function, researchers need to make the products of the indicators between the first-order factors and residualize the products by the original indicators (Lance, 1988; Little, Bovaird, & Widaman, 2006). The process can be automated by the `indProd` function. Note that the indicator products can be made for all possible combination or matched-pair approach (Marsh et al., 2004). Next, the hypothesized model with the regression with latent interaction will be used to fit all original indicators and the product terms. To use this function the model must be fit with a mean structure. See the example for how to fit the product term below. Once the lavaan result is obtained, this function will be used to probe the interaction.

The probing process on residual-centered latent interaction is based on transforming the residual-centered result into the no-centered result. See Pornprasertmanit, Schoemann, Geldhof, and Little (submitted) for further details. Note that this approach based on a strong assumption that the first-order latent variables are normally distributed. The probing process is applied after the no-centered result (parameter estimates and their covariance matrix among parameter estimates) has been computed. See the `probe2WayMC` for further details.

Value

A list with two elements:

1. `SimpleIntercept` The intercepts given each value of the moderator. This element will be shown only if the factor intercept is estimated (e.g., not fixed as 0).
2. `SimpleSlope` The slopes given each value of the moderator.

In each element, the first column represents the values of the moderators specified in the `valProbe` argument. The second column is the simple intercept or simple slope. The third column is the standard error of the simple intercept or simple slope. The fourth column is the Wald (z) statistic. The fifth column is the p -value testing whether the simple intercepts or slopes are different from 0.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

- Lance, C. E. (1988). Residual centering, exploratory and confirmatory moderator analysis, and decomposition of effects in path models containing interactions. *Applied Psychological Measurement*, 12, 163-175.
- Little, T. D., Bovaird, J. A., & Widaman, K. F. (2006). On the merits of orthogonalizing powered and product terms: Implications for modeling interactions. *Structural Equation Modeling*, 13, 497-519.

Marsh, H. W., Wen, Z., & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, 9, 275-300.

Pornprasertmanit, S., Schoemann, A. M., Geldhof, G. J., & Little, T. D. (submitted). *Probing latent interaction estimated with a residual centering approach.*

See Also

- `indProd` For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- `probe2WayMC` For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe3WayMC` For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe3WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `plotProbe` Plot the simple intercepts and slopes of the latent interaction.

Examples

```
library(lavaan)

dat2wayRC <- orthogonalize(dat2way, 1:3, 4:6)

modell <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f12 =~ x1.x4 + x2.x5 + x3.x6
f3 =~ x7 + x8 + x9
f3 ~ f1 + f2 + f12
f12 ~~ 0*f1
f12 ~~ 0*f2
x1 ~ 0*1
x4 ~ 0*1
x1.x4 ~ 0*1
x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f12 ~ NA*1
f3 ~ NA*1
"

fitRC2way <- sem(modell, data=dat2wayRC, meanstructure=TRUE, std.lv=FALSE)
summary(fitRC2way)

result2wayRC <- probe2WayRC(fitRC2way, c("f1", "f2", "f12"), "f3", "f2", c(-1, 0, 1))
result2wayRC
```

probe3WayMC	<i>Probing two-way interaction on the residual-centered latent interaction</i>
-------------	--

Description

Probing interaction for simple intercept and simple slope for the no-centered or mean-centered latent two-way interaction

Usage

```
probe3WayMC(fit, nameX, nameY, modVar, valProbe1, valProbe2)
```

Arguments

fit	The lavaan model object used to evaluate model fit
nameX	The vector of the factor names used as the predictors. The three first-order factors will be listed first. Then the second-order factors will be listed. The last element of the name will represent the three-way interaction. Note that the fourth element must be the interaction between the first and the second variables. The fifth element must be the interaction between the first and the third variables. The sixth element must be the interaction between the second and the third variables.
nameY	The name of factor that is used as the dependent variable.
modVar	The name of two factors that are used as the moderators. The effect of the independent factor on each combination of the moderator variable values will be probed.
valProbe1	The values of the first moderator that will be used to probe the effect of the independent factor.
valProbe2	The values of the second moderator that will be used to probe the effect of the independent factor.

Details

Before using this function, researchers need to make the products of the indicators between the first-order factors using mean centering (Marsh, Wen, & Hau, 2004). Note that the double-mean centering may not be appropriate for probing interaction if researchers are interested in simple intercepts. The mean or double-mean centering can be done by the `indProd` function. The indicator products can be made for all possible combination or matched-pair approach (Marsh et al., 2004). Next, the hypothesized model with the regression with latent interaction will be used to fit all original indicators and the product terms. See the example for how to fit the product term below. Once the lavaan result is obtained, this function will be used to probe the interaction.

Let that the latent interaction model regressing the dependent variable (Y) on the independent variable (X) and two moderators (Z and W) be

$$Y = b_0 + b_1X + b_2Z + b_3W + b_4XZ + b_5XW + b_6ZW + b_7XZW + r,$$

where b_0 is the estimated intercept or the expected value of Y when X , Z , and W are 0, b_1 is the effect of X when Z and W are 0, b_2 is the effect of Z when X and W is 0, b_3 is the effect of W when X and Z are 0, b_4 is the interaction effect between X and Z when W is 0, b_5 is the interaction

effect between X and W when Z is 0, b_6 is the interaction effect between Z and W when X is 0, b_7 is the three-way interaction effect between X , Z , and W , and r is the residual term.

For probing three-way interaction, the simple intercept of the independent variable at the specific values of the moderators (Aiken & West, 1991) can be obtained by

$$b_{0|X=0,Z,W} = b_0 + b_2Z + b_3W + b_6ZW.$$

The simple slope of the independent variable at the specific values of the moderators can be obtained by

$$b_{X|Z,W} = b_1 + b_3Z + b_4W + b_7ZW.$$

The variance of the simple intercept formula is

$$Var(b_{0|X=0,Z,W}) = Var(b_0) + Z^2Var(b_2) + W^2Var(b_3) + Z^2W^2Var(b_6) + 2ZCov(b_0, b_2) + 2WCov(b_0, b_3) + 2ZW$$

where Var denotes the variance of a parameter estimate and Cov denotes the covariance of two parameter estimates.

The variance of the simple slope formula is

$$Var(b_{X|Z,W}) = Var(b_1) + Z^2Var(b_4) + W^2Var(b_5) + Z^2W^2Var(b_7) + 2ZCov(b_1, b_4) + 2WCov(b_1, b_5) + 2ZW$$

Wald statistic is used for test statistic.

Value

A list with two elements:

1. SimpleIntercept The intercepts given each value of the moderator. This element will be shown only if the factor intercept is estimated (e.g., not fixed as 0).
2. SimpleSlope The slopes given each value of the moderator.

In each element, the first column represents the values of the first moderator specified in the `valProbe1` argument. The second column represents the values of the second moderator specified in the `valProbe2` argument. The third column is the simple intercept or simple slope. The fourth column is the standard error of the simple intercept or simple slope. The fifth column is the Wald (z) statistic. The sixth column is the p -value testing whether the simple intercepts or slopes are different from 0.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

- Aiken, L. S., & West, S. G. (1991). Multiple regression: Testing and interpreting interactions. Newbury Park, CA: Sage.
- Marsh, H. W., Wen, Z., & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, 9, 275-300.

See Also

- `indProd` For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- `probe2WayMC` For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe2WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `probe3WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `plotProbe` Plot the simple intercepts and slopes of the latent interaction.

Examples

```
library(lavaan)

dat3wayMC <- indProd(dat3way, 1:3, 4:6, 7:9)

model3 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f3 =~ x7 + x8 + x9
f12 =~ x1.x4 + x2.x5 + x3.x6
f13 =~ x1.x7 + x2.x8 + x3.x9
f23 =~ x4.x7 + x5.x8 + x6.x9
f123 =~ x1.x4.x7 + x2.x5.x8 + x3.x6.x9
f4 =~ x10 + x11 + x12
f4 ~ f1 + f2 + f3 + f12 + f13 + f23 + f123
f1 ~~ 0*f12
f1 ~~ 0*f13
f1 ~~ 0*f123
f2 ~~ 0*f12
f2 ~~ 0*f23
f2 ~~ 0*f123
f3 ~~ 0*f13
f3 ~~ 0*f23
f3 ~~ 0*f123
f12 ~~ 0*f123
f13 ~~ 0*f123
f23 ~~ 0*f123
x1 ~ 0*1
x4 ~ 0*1
x7 ~ 0*1
x10 ~ 0*1
x1.x4 ~ 0*1
x1.x7 ~ 0*1
x4.x7 ~ 0*1
x1.x4.x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f3 ~ NA*1
f12 ~ NA*1
f13 ~ NA*1
f23 ~ NA*1
f123 ~ NA*1
```

```
f4 ~ NA*1
"

fitMC3way <- sem(model3, data=dat3wayMC, meanstructure=TRUE, std.lv=FALSE)
summary(fitMC3way)

result3wayMC <- probe3WayMC(fitMC3way, c("f1", "f2", "f3", "f12", "f13", "f23", "f123"),
"f4", c("f1", "f2"), c(-1, 0, 1), c(-1, 0, 1))
result3wayMC
```

probe3WayRC	<i>Probing three-way interaction on the residual-centered latent interaction</i>
-------------	--

Description

Probing interaction for simple intercept and simple slope for the residual-centered latent three-way interaction (Pornprasertmanit, Schoemann, Geldhof, & Little, submitted)

Usage

```
probe3WayRC(fit, nameX, nameY, modVar, valProbe1, valProbe2)
```

Arguments

fit	The lavaan model object used to evaluate model fit
nameX	The vector of the factor names used as the predictors. The three first-order factors will be listed first. Then the second-order factors will be listed. The last element of the name will represent the three-way interaction. Note that the fourth element must be the interaction between the first and the second variables. The fifth element must be the interaction between the first and the third variables. The sixth element must be the interaction between the second and the third variables.
nameY	The name of factor that is used as the dependent variable.
modVar	The name of two factors that are used as the moderators. The effect of the independent factor on each combination of the moderator variable values will be probed.
valProbe1	The values of the first moderator that will be used to probe the effect of the independent factor.
valProbe2	The values of the second moderator that will be used to probe the effect of the independent factor.

Details

Before using this function, researchers need to make the products of the indicators between the first-order factors and residualize the products by the original indicators (Lance, 1988; Little, Bovaird, & Widaman, 2006). The process can be automated by the `indProd` function. Note that the indicator products can be made for all possible combination or matched-pair approach (Marsh et al., 2004). Next, the hypothesized model with the regression with latent interaction will be used to fit all original indicators and the product terms (Geldhof, Pornprasertmanit, Schoemann, & Little, in press). To use this function the model must be fit with a mean structure. See the example for how

to fit the product term below. Once the lavaan result is obtained, this function will be used to probe the interaction.

The probing process on residual-centered latent interaction is based on transforming the residual-centered result into the no-centered result. See Pornprasertmanit, Schoemann, Geldhof, and Little (submitted) for further details. Note that this approach based on a strong assumption that the first-order latent variables are normally distributed. The probing process is applied after the no-centered result (parameter estimates and their covariance matrix among parameter estimates) has been computed. See the `probe3WayMC` for further details.

Value

A list with two elements:

1. `SimpleIntercept` The intercepts given each value of the moderator. This element will be shown only if the factor intercept is estimated (e.g., not fixed as 0).
2. `SimpleSlope` The slopes given each value of the moderator.

In each element, the first column represents the values of the first moderator specified in the `valProbe1` argument. The second column represents the values of the second moderator specified in the `valProbe2` argument. The third column is the simple intercept or simple slope. The fourth column is the standard error of the simple intercept or simple slope. The fifth column is the Wald (z) statistic. The sixth column is the *p*-value testing whether the simple intercepts or slopes are different from 0.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

- Geldhof, G. J., Pornprasertmanit, S., Schoemann, A., & Little, T. D. (in press). Orthogonalizing through residual centering: Applications and caveats. *Educational and Psychological Measurement*.
- Lance, C. E. (1988). Residual centering, exploratory and confirmatory moderator analysis, and decomposition of effects in path models containing interactions. *Applied Psychological Measurement*, 12, 163-175.
- Little, T. D., Bovaird, J. A., & Widaman, K. F. (2006). On the merits of orthogonalizing powered and product terms: Implications for modeling interactions. *Structural Equation Modeling*, 13, 497-519.
- Marsh, H. W., Wen, Z., & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, 9, 275-300.
- Pornprasertmanit, S., Schoemann, A. M., Geldhof, G. J., & Little, T. D. (submitted). *Probing latent interaction estimated with a residual centering approach*.

See Also

- `indProd` For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- `probe2WayMC` For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering.

- `probe3WayMC` For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- `probe2WayRC` For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- `plotProbe` Plot the simple intercepts and slopes of the latent interaction.

Examples

```
library(lavaan)

dat3wayRC <- orthogonalize(dat3way, 1:3, 4:6, 7:9)

model3 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f3 =~ x7 + x8 + x9
f12 =~ x1.x4 + x2.x5 + x3.x6
f13 =~ x1.x7 + x2.x8 + x3.x9
f23 =~ x4.x7 + x5.x8 + x6.x9
f123 =~ x1.x4.x7 + x2.x5.x8 + x3.x6.x9
f4 =~ x10 + x11 + x12
f4 ~ f1 + f2 + f3 + f12 + f13 + f23 + f123
f1 ~~ 0*f12
f1 ~~ 0*f13
f1 ~~ 0*f123
f2 ~~ 0*f12
f2 ~~ 0*f23
f2 ~~ 0*f123
f3 ~~ 0*f13
f3 ~~ 0*f23
f3 ~~ 0*f123
f12 ~~ 0*f123
f13 ~~ 0*f123
f23 ~~ 0*f123
x1 ~ 0*1
x4 ~ 0*1
x7 ~ 0*1
x10 ~ 0*1
x1.x4 ~ 0*1
x1.x7 ~ 0*1
x4.x7 ~ 0*1
x1.x4.x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f3 ~ NA*1
f12 ~ NA*1
f13 ~ NA*1
f23 ~ NA*1
f123 ~ NA*1
f4 ~ NA*1
"

fitRC3way <- sem(model3, data=dat3wayRC, meanstructure=TRUE, std.lv=FALSE)
summary(fitRC3way)

result3wayRC <- probe3WayRC(fitRC3way, c("f1", "f2", "f3", "f12", "f13", "f23", "f123"),
```

```
"f4", c("f1", "f2"), c(-1, 0, 1), c(-1, 0, 1))
result3wayRC
```

reliability

Calculate reliability values of factors

Description

Calculate reliability values of factors by coefficient omega

Usage

```
reliability(object)
```

Arguments

object The lavaan model object provided after running the `cfa`, `sem`, `growth`, or `lavaan` functions.

Details

The coefficient alpha (Cronbach, 1951) can be calculated by

$$\alpha = \frac{k}{k-1} \left[1 - \frac{\sum_{i=1}^k \sigma_{ii}}{\sum_{i=1}^k \sigma_{ii} + 2 \sum_{i < j} \sigma_{ij}} \right],$$

where k is the number of items in a factor, σ_{ii} is the item i observed variances, σ_{ij} is the observed covariance of items i and j .

The coefficient omega (Raykov, 2001) can be calculated by

$$\omega = \frac{\left(\sum_{i=1}^k \lambda_i \right)^2 \text{Var}(\psi)}{\left(\sum_{i=1}^k \lambda_i \right)^2 \text{Var}(\psi) + \sum_{i=1}^k \theta_{ii} + 2 \sum_{i < j} \theta_{ij}},$$

where λ_i is the factor loading of item i , ψ is the factor variance, θ_{ii} is the variance of measurement errors of item i , and θ_{ij} is the covariance of measurement errors from item i and j .

The second coefficient omega (Bentler, 1972, 2009) can be calculated by

$$\omega_2 = 1 - \frac{\mathbf{1}'\Theta\mathbf{1}}{\mathbf{1}'\hat{\Sigma}\mathbf{1}},$$

where Θ is the measurement error covariance matrix, $\hat{\Sigma}$ is the model-implied covariance matrix, and $\mathbf{1}$ is the k -dimensional vector of 1. The first and the second coefficients omega will have different values if there are dual loadings (or the existence of method factors). The first coefficient omega can be viewed as the reliability controlling for the other factors. The second coefficient omega can be viewed as the unconditional reliability.

The third coefficient omega (McDonald, 1999) can be calculated by

$$\omega_3 = 1 - \frac{\mathbf{1}'\Theta\mathbf{1}}{\mathbf{1}'\Sigma\mathbf{1}},$$

where Σ is the observed covariance matrix. If the model fits the data well, the third coefficient omega will be similar to the other two. Note that if there is a directional effect in the model, all coefficients omega will use the total factor variances, which is calculated by the `impliedFactorCov` function.

If measurement errors are not correlated, the last term in the denominator of the middle expression is dropped. For the categorical items, the reliability indices are calculated from the latent variable underlying the categorical items (i.e., using polychoric/polyserial correlations). Therefore, the coefficient alpha from this function may be not the same as the standard alpha calculation for categorical items. Researchers may check the `alpha` function in the `psych` package for the standard coefficient alpha calculation.

Value

Reliability values (coefficient alpha, coefficients omega) of each factor in each group

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>); Yves Rosseel (Ghent University; <Yves.Rosseel@UGent.be>)

References

- Bentler, P. M. (1972). A lower-bound method for the dimension-free measurement of internal consistency. *Social Science Research*, 1, 343-357.
- Bentler, P. M. (2009). Alpha, dimension-free, and model-based internal consistency reliability. *Psychometrika*, 74, 137-143.
- Cronbach, L. J. (1951). Coefficient alpha and the internal structure of tests. *Psychometrika*, 16, 297-334.
- McDonald, R. P. (1999). Test theory: A unified treatment. Mahwah, NJ: Erlbaum.
- Raykov, T. (2001). Estimation of congeneric scale reliability using covariance structure analysis with nonlinear constraints *British Journal of Mathematical and Statistical Psychology*, 54, 315-323.

See Also

`reliabilityL2` for reliability value of a desired second-order factor

Examples

```
HS.model <- ' visual  =~ x1 + x2 + x3
               textual =~ x4 + x5 + x6
               speed   =~ x7 + x8 + x9 '

fit <- cfa(HS.model, data=HolzingerSwineford1939)
reliability(fit)
```

reliabilityL2

*Calculate the reliability values of a second-order factor***Description**

Calculate the reliability values (coefficient omega) of a second-order factor

Usage

```
reliabilityL2(object, secondFactor)
```

Arguments

object The lavaan model object provided after running the `cfa`, `sem`, `growth`, or `lavaan` functions that has a second-order factor

secondFactor The name of the second-order factor

Details

The first formula of the coefficient omega (in the `reliability`) will be mainly used in the calculation. The model-implied covariance matrix of a second-order factor model can be separated into three sources: the second-order factor, the uniqueness of the first-order factor, and the measurement error of indicators:

$$\hat{\Sigma} = \Lambda B \Phi_2 B' \Lambda' + \Lambda \Psi_u \Lambda' + \Theta,$$

where $\hat{\Sigma}$ is the model-implied covariance matrix, Λ is the first-order factor loading, B is the second-order factor loading, Φ_2 is the covariance matrix of the second-order factors, Ψ_u is the covariance matrix of the unique scores from first-order factors, and Θ is the covariance matrix of the measurement errors from indicators. Thus, the proportion of the second-order factor explaining the total score, or the coefficient omega at Level 1, can be calculated:

$$\omega_{L1} = \frac{\mathbf{1}' \Lambda B \Phi_2 B' \Lambda' \mathbf{1}}{\mathbf{1}' \Lambda B \Phi_2 B' \Lambda' \mathbf{1} + \mathbf{1}' \Lambda \Psi_u \Lambda' \mathbf{1} + \mathbf{1}' \Theta \mathbf{1}},$$

where $\mathbf{1}$ is the k -dimensional vector of 1 and k is the number of observed variables. When model-implied covariance matrix among first-order factors (Φ_1) can be calculated:

$$\Phi_1 = B \Phi_2 B' + \Psi_u,$$

Thus, the proportion of the second-order factor explaining the variance at first-order factor level, or the coefficient omega at Level 2, can be calculated:

$$\omega_{L2} = \frac{\mathbf{1}'_F B \Phi_2 B' \mathbf{1}_F}{\mathbf{1}'_F B \Phi_2 B' \mathbf{1}_F + \mathbf{1}'_F \Psi_u \mathbf{1}_F},$$

where $\mathbf{1}_F$ is the F -dimensional vector of 1 and F is the number of first-order factors.

The partial coefficient omega at Level 1, or the proportion of observed variance explained by the second-order factor after partialling the uniqueness from the first-order factor, can be calculated:

$$\omega_{L1} = \frac{\mathbf{1}' \Lambda B \Phi_2 B' \Lambda' \mathbf{1}}{\mathbf{1}' \Lambda B \Phi_2 B' \Lambda' \mathbf{1} + \mathbf{1}' \Theta \mathbf{1}},$$

Note that if the second-order factor has a direct factor loading on some observed variables, the observed variables will be counted as first-order factors.

Value

Reliability values at Levels 1 and 2 of the second-order factor, as well as the partial reliability value at Level 1

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

`reliability` for the reliability of the first-order factors.

Examples

```
HS.model3 <- ' visual  =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed   =~ x7 + x8 + x9
              higher  =~ visual + textual + speed'

fit6 <- cfa(HS.model3, data=HolzingerSwineford1939)
reliability(fit6) # Should provide a warning for the endogenous variable
reliabilityL2(fit6, "higher")
```

`residualCovariate` *Residual centered all target indicators by covariates*

Description

This function will regress target variables on the covariate and replace the target variables by the residual of the regression analysis. This procedure is useful to control the covariate from the analysis model (Geldhof, Pornprasertmanit, Schoemann, & Little, in press).

Usage

```
residualCovariate(data, targetVar, covVar)
```

Arguments

<code>data</code>	The desired data to be transformed.
<code>targetVar</code>	Variable names or the position of indicators that users wish to be residual centered (as dependent variables)
<code>covVar</code>	Covariate names or the position of the covariates using for residual centering (as independent variables) onto target variables

Value

The data that the target variables replaced by the residuals

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

Geldhof, G. J., Pornprasertmanit, S., Schoemann, A. M., & Little, T. D. (in press). Orthogonalizing through residual centering: Applications and caveats. *Educational and Psychological Measurement*.

See Also

`indProd` For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.

Examples

```
dat <- residualCovariate(attitude, 2:7, 1)
```

rotate	<i>Implement orthogonal or oblique rotation</i>
--------	---

Description

These functions will implement orthogonal or oblique rotation on standardized factor loadings from a lavaan output.

Usage

```
orthRotate(object, method="varimax", ...)
oblqRotate(object, method="quartimin", ...)
funRotate(object, fun, ...)
```

Arguments

object	A lavaan output
method	The method of rotations, such as "varimax", "quartimax", "geomin", "oblimin", or any gradient projection algorithms listed in the GPA function in the GPArotation package.
fun	The name of the function that users wish to rotate the standardized solution. The functions must take the first argument as the standardized loading matrix and return the GPArotation object. Check this page for available functions: rotations .
...	Additional arguments for the GPForth function (for orthRotate), the GPFoblq function (for oblqRotate), or the function that users provide in the fun argument.

Details

These functions will rotate the unrotated standardized factor loadings by orthogonal rotation using the GPForth function or oblique rotation using the GPFoblq function the GPArotation package. The resulting rotation matrix will be used to calculate standard errors of the rotated standardized factor loading by delta method by numerically computing the Jacobian matrix by the lavJacobianD function in the lavaan package.

Value

An `linkS4class{EFA}` object that saves the rotated EFA solution.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

Examples

```
unrotated <- efaUnrotate(HolzingerSwineford1939, nf=3, varList=paste0("x", 1:9), estimator="ml")

# Orthogonal varimax
out.varimax <- orthRotate(unrotated, method="varimax")
summary(out.varimax, sort=FALSE, suppress=0.3)

# Orthogonal Quartimin
orthRotate(unrotated, method="quartimin")

# Oblique Quartimin
oblqRotate(unrotated, method="quartimin")

# Geomin
oblqRotate(unrotated, method="geomin")

## Not run:
# Target rotation
library(GPARotation)
target <- matrix(0, 9, 3)
target[1:3, 1] <- NA
target[4:6, 2] <- NA
target[7:9, 3] <- NA
colnames(target) <- c("factor1", "factor2", "factor3")
# This function works with GPARotation version 2012.3-1
funRotate(unrotated, fun="targetQ", Target=target)

## End(Not run)
```

runMI

Multiply impute and analyze data using lavaan

Description

This function takes data with missing observations, multiple imputes the data, runs a SEM using lavaan and combines the results using Rubin's rules. Note that parameter estimates and standard errors are pooled by the Rubin's (1987) rule. The chi-square statistics and the related fit indices are pooled by the method described in "chi" argument. SRMR is calculated based on the average model-implied means and covariance matrices across imputations.

Usage

```
runMI(model, data, m, miArgs=list(), chi="all", miPackage="Amelia",
seed=12345, fun, ...)
cfa.mi(model, data, m, miArgs=list(), miPackage="Amelia", chi="all",
```

```

seed=12345, ...)
sem.mi(model, data, m, miArgs=list(), miPackage="Amelia", chi="all",
seed=12345, ...)
growth.mi(model, data, m, miArgs=list(), miPackage="Amelia", chi="all",
seed=12345, ...)
lavaan.mi(model, data, m, miArgs=list(), miPackage="Amelia", chi="all",
seed=12345, ...)

```

Arguments

<code>model</code>	lavaan syntax for the the model to be analyzed.
<code>data</code>	Data frame with missing observations or a list of data frames where each data frame is one imputed data set (for imputed data generated outside of the function). If a list of data frames is supplied, then other options can be left at the default.
<code>m</code>	Number of imputations wanted.
<code>miArgs</code>	Addition arguments for the multiple-imputation function. The arguments should be put in a list (see example below).
<code>miPackage</code>	Package to be used for imputation. Currently these functions only support "Amelia" or "mice" for imputation.
<code>chi</code>	The method to combine the chi-square. Can be one of the following: "mr" for the method proposed for Meng & Rubin (1992), "mplus" for the method used in Mplus (Asparouhov & Muthen, 2010), "lmrr" for the method proposed by Li, Meng, Raghunathan, & Rubin (1991), "all" to show the three methods in the output, and "none" to not pool any chi-square values. The default is "all".
<code>seed</code>	Random number seed to be used in imputations.
<code>fun</code>	The character of the function name used in running lavaan model ("cfa", "sem", "growth", "lavaan").
<code>...</code>	Other arguments to be passed to the specified lavaan function ("cfa", "sem", "growth", "lavaan").

Value

The `lavaanStar` object which contains the original lavaan object (where the appropriate parameter estimates, appropriate standard errors, and chi-squares are filled), the additional fit-index values of the null model, which need to be adjusted to multiple datasets, and the information from pooling multiple results.

Author(s)

Alexander M. Schoemann (University of Kansas; <schoemann@ku.edu>) Patrick Miller (University of Kansas; <patrickm@ku.edu>) Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>) Mijke Rhemtulla (University of Kansas; <mijke@ku.edu>) Alexander Robitzsch (Federal Institute for Education Research, Innovation, and Development of the Austrian School System, Salzburg, Austria; <a.robitzsch@bifie.at>) Craig Enders (Arizona State University; <Craig.Enders@asu.edu>) Mauricio Garnier Villarreal (University of Kansas; <mgv@ku.edu>) Yves Rosseel (Ghent University; <Yves.Rosseel@UGent.be>)

References

- Asparouhov T. & Muthen B. (2010). *Chi-Square Statistics with Multiple Imputation*. Technical Report. www.statmodel.com.
- Li, K.H., Meng, X.-L., Raghunathan, T.E. and Rubin, D.B. (1991). Significance Levels From Repeated p-values with Multiply-Imputed Data. *Statistica Sinica*, 1, 65-92.
- Meng, X.L. & Rubin, D.B. (1992). Performing likelihood ratio tests with multiply-imputed data sets. *Biometrika*, 79, 103 - 111.
- Rubin, D.B. (1987) *Multiple Imputation for Nonresponse in Surveys*. J. Wiley & Sons, New York.

Examples

```
library(lavaan)

HS.model <- ' visual   =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6
              speed    =~ x7 + x8 + x9 '

HSMiss <- HolzingerSwineford1939[,paste("x", 1:9, sep="")]
randomMiss <- rbinom(prod(dim(HSMiss)), 1, 0.1)
randomMiss <- matrix(as.logical(randomMiss), nrow=nrow(HSMiss))
HSMiss[randomMiss] <- NA

out <- cfa.mi(HS.model, data=HSMiss, m = 3, chi="all")
summary(out)
inspect(out, "fit")
inspect(out, "impute")

## Not run:
##Multiple group example
HSMiss2 <- cbind(HSMiss, school = HolzingerSwineford1939[, "school"])
out2 <- cfa.mi(HS.model, data=HSMiss2, m = 3, miArgs=list(noms="school"), chi="MR", group)
summary(out2)
inspect(out2, "fit")
inspect(out2, "impute")

##Example using previously imputed data with runMI
library(Amelia)

modsim <- '
f1 =~ 0.7*y1+0.7*y2+0.7*y3
f2 =~ 0.7*y4+0.7*y5+0.7*y6
f3 =~ 0.7*y7+0.7*y8+0.7*y9'

mod <- '
f1 =~ y1+y2+y3
f2 =~ y4+y5+y6
f3 =~ y7+y8+y9'

datSIM <- simulateData(modsim,model.type="cfa", meanstructure=TRUE,
std.lv=TRUE, sample.nobs=c(200,200))
randomMiss2 <- rbinom(prod(dim(datSIM)), 1, 0.1)
randomMiss2 <- matrix(as.logical(randomMiss2), nrow=nrow(datSIM))
datSIM[randomMiss2] <- NA
datSIMMI <- amelia(datSIM,m=3, noms="group")
```

```

out3 <- runMI(mod, data=datsimMI$imputations, chi="LMRR", group="group", fun="cfa")
summary(out3)
inspect(out3, "fit")
inspect(out3, "impute")

# Categorical variables
dat <- simulateData(popModel, sample.nobs = 200L)
miss.pat <- matrix(as.logical(rbinom(prod(dim(dat)), 1, 0.2)), nrow(dat), ncol(dat))
dat[miss.pat] <- NA
out5 <- cfa.mi(analyzeModel, data=dat, ordered=paste0("y", 1:4), m = 3,
miArgs=list(ords = c("y1", "y2", "y3", "y4")))
summary(out5)
inspect(out5, "fit")
inspect(out5, "impute")

## End(Not run)

```

saturateMx

*Analyzing data using a saturate model***Description**

Analyzing data using a saturate model by full-information maximum likelihood. In the saturate model, all means and covariances are free if items are continuous. For ordinal variables, their means are fixed as 0 and their variances are fixed as 1—their covariances and thresholds are estimated. In multiple-group model, all means and variances are separately estimated.

Usage

```
saturateMx(data, groupLab = NULL)
```

Arguments

data	The target data frame
groupLab	The name of grouping variable

Value

The `MxModel` object which contains the analysis result of the saturate model.

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

`nullMx`, `fitMeasuresMx`, `standardizeMx`

Examples

```
## Not run:
library(OpenMx)
data(demoOneFactor)
satModel <- saturateMx(demoOneFactor)

## End(Not run)
```

simParcel

*Simulated Data set to Demonstrate Random Allocations of Parcels***Description**

A simulated data set with 2 factors with 9 indicators for each factor

Usage

```
data(simParcel)
```

Format

A data frame with 800 observations of 18 variables.

f1item1 Item 1 loading on factor 1
f1item2 Item 2 loading on factor 1
f1item3 Item 3 loading on factor 1
f1item4 Item 4 loading on factor 1
f1item5 Item 5 loading on factor 1
f1item6 Item 6 loading on factor 1
f1item7 Item 7 loading on factor 1
f1item8 Item 8 loading on factor 1
f1item9 Item 9 loading on factor 1
f2item1 Item 1 loading on factor 2
f2item2 Item 2 loading on factor 2
f2item3 Item 3 loading on factor 2
f2item4 Item 4 loading on factor 2
f2item5 Item 5 loading on factor 2
f2item6 Item 6 loading on factor 2
f2item7 Item 7 loading on factor 2
f2item8 Item 8 loading on factor 2
f2item9 Item 9 loading on factor 2

Source

Data was generated using the `simsem` package.

Examples

```
head(simParcel)
```

skew

*Finding skewness***Description**

Finding skewness (g1) of an object

Usage

```
skew(object, population=FALSE)
```

Arguments

object	A vector used to find a skewness
population	TRUE to compute the parameter formula. FALSE to compute the sample statistic formula.

Details

The skewness computed is g1. The parameter skewness γ_2 formula is

$$\gamma_2 = \frac{\mu_3}{\mu_2^{3/2}},$$

where μ_i denotes the i order central moment.

The excessive kurtosis formula for sample statistic g_2 is

$$g_2 = \frac{k_3}{k_2^2},$$

where k_i are the i order k -statistic.

The standard error of the skewness is

$$Var(\hat{g}_2) = \frac{6}{N}$$

where N is the sample size.

Value

A value of a skewness with a test statistic if the population is specified as FALSE

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

References

Weisstein, Eric W. (n.d.). *Skewness*. Retrived from MathWorld—A Wolfram Web Resource <http://mathworld.wolfram.com/Skewness.html>

See Also

- `kurtosis` Find the univariate excessive kurtosis of a variable
- `mardiaSkew` Find the Mardia's multivariate skewness of a set of variables
- `mardiaKurtosis` Find the Mardia's multivariate kurtosis of a set of variables

Examples

```
skew(1:5)
```

splitSample	<i>Randomly Split a Data Set into Halves</i>
-------------	--

Description

This function randomly splits a data set into two halves, and saves the resulting data sets to the same folder as the original.

Usage

```
splitSample(dataset, path="default", div=2, type="default", name="splitSample")
```

Arguments

dataset	The original data set to be divided. Can be a file path to a .csv or .dat file (headers will automatically be detected) or an R object (matrix or dataframe). (Windows users: file path must be specified using FORWARD SLASHES ONLY.)
path	File path to folder for output data sets. NOT REQUIRED if dataset is a filename. Specify ONLY if dataset is an R object, or desired output folder is not that of original data set. If path is specified as "object", output data sets will be returned as a list, and not saved to hard drive.
div	Number of output data sets. NOT REQUIRED if default, 2 halves.
type	Output file format ("dat" or "csv"). NOT REQUIRED unless desired output formatting differs from that of input, or dataset is an R object and csv formatting is desired.
name	Output file name. NOT REQUIRED unless desired output name differs from that of input, or input dataset is an R object. (If input is an R object and name is not specified, name will be "splitSample".)

Details

This function randomly orders the rows of a data set, divides the data set into two halves, and saves the halves to the same folder as the original data set, preserving the original formatting. Data set type (.csv or .dat) and formatting (headers) are automatically detected, and output data sets will preserve input type and formatting unless specified otherwise. Input can be in the form of a file path (.dat or .csv), or an R object (matrix or dataframe). If input is an R object and path is default, output data sets will be returned as a list object.

Value

dataL List of output data sets. ONLY IF dataset is an R object and path is default. Otherwise, output will saved to hard drive with the same formatting as input.

Author(s)

Corbin Quick (University of Kansas; <corbing@ku.edu>)

Examples

```
#### Input is .dat file
#splitSample("C:/Users/Default/Desktop/MYDATA.dat")
#### Output saved to "C:/Users/Default/Desktop/" in .dat format
#### Names are "MYDATA_s1.dat" and "MYDATA_s2.dat"

#### Input is R object
##Split C02 dataset from the datasets package
library(datasets)
splitMyData <- splitSample(CO2, path="object")
summary(splitMyData[[1]])
summary(splitMyData[[2]])
#### Output object splitMyData becomes list of output data sets

#### Input is .dat file in "C:/" folder
#splitSample("C:/testdata.dat", path = "C:/Users/Default/Desktop/", type = "csv")
#### Output saved to "C:/Users/Default/Desktop/" in .csv format
#### Names are "testdata_s1.csv" and "testdata_s2.csv"

#### Input is R object
#splitSample(myData, path = "C:/Users/Default/Desktop/", name = "splitdata")
#### Output saved to "C:/Users/Default/Desktop/" in .dat format
#### Names are "splitdata_s1.dat" and "splitdata_s2.dat"
```

standardizeMx

Find standardized estimates for OpenMx output

Description

Find standardized estimates for OpenMx output. This function is applicable for the MxRAMObjective only.

Usage

```
standardizeMx(object, free = TRUE)
```

Arguments

object Target OpenMx output using MxRAMObjective

free If TRUE, the function will show only standardized values of free parameters. If FALSE, the function will show the results for fixed and free parameters.

Value

A vector of standardized estimates

Author(s)

Sunthud Pornprasertmanit (University of Kansas; <psunthud@ku.edu>)

See Also

saturateMx, nullMx, fitMeasuresMx

Examples

```
## Not run:
library(OpenMx)
data(myFADataRaw)
myFADataRaw <- myFADataRaw[,c("x1", "x2", "x3", "x4", "x5", "x6")]
oneFactorModel <- mxModel("Common Factor Model Path Specification",
  type="RAM",
  mxData(
    observed=myFADataRaw,
    type="raw"
  ),
  manifestVars=c("x1", "x2", "x3", "x4", "x5", "x6"),
  latentVars="F1",
  mxPath(from=c("x1", "x2", "x3", "x4", "x5", "x6"),
    arrows=2,
    free=TRUE,
    values=c(1, 1, 1, 1, 1, 1),
    labels=c("e1", "e2", "e3", "e4", "e5", "e6")
  ),
  # residual variances
  # -----
  mxPath(from="F1",
    arrows=2,
    free=TRUE,
    values=1,
    labels="varF1"
  ),
  # latent variance
  # -----
  mxPath(from="F1",
    to=c("x1", "x2", "x3", "x4", "x5", "x6"),
    arrows=1,
    free=c(FALSE, TRUE, TRUE, TRUE, TRUE, TRUE),
    values=c(1, 1, 1, 1, 1, 1),
    labels=c("l1", "l2", "l3", "l4", "l5", "l6")
  ),
  # factor loadings
  # -----
  mxPath(from="one",
    to=c("x1", "x2", "x3", "x4", "x5", "x6", "F1"),
    arrows=1,
    free=c(TRUE, TRUE, TRUE, TRUE, TRUE, TRUE, FALSE),
    values=c(1, 1, 1, 1, 1, 1, 0),
    labels=c("meanx1", "meanx2", "meanx3", "meanx4", "meanx5", "meanx6", NA)
```

```

)
# means
# -----
) # close model
# Create an MxModel object
# -----
oneFactorFit <- mxRun(oneFactorModel)
standardizeMx(oneFactorFit)

# Compare with lavaan
library(lavaan)
script <- "f1 =~ x1 + x2 + x3 + x4 + x5 + x6"
fit <- cfa(script, data=myFADataRaw, meanstructure=TRUE)
standardize(fit)

## End(Not run)

```

tukeySEM

Tukey's WSD post-hoc test of means for unequal variance and sample size

Description

This function computes Tukey's WSD post-hoc test of means when variances and sample sizes are not equal across groups. It can be used as a post-hoc test when comparing latent means in multiple group SEM.

Usage

```
tukeySEM(m1, m2, var1, var2, n1, n2, ng)
```

Arguments

m1	Mean of group 1.
m2	Mean of group 2.
var1	Variance of group 1.
var2	Variance of group 2.
n1	Sample size of group 1.
n2	Sample size of group 2.
ng	Total number of groups to be compared (i.e., the number of groups compared in the omnibus test).

Details

After conducting an omnibus test of means across three or more groups, researchers often wish to know which sets of means differ at a particular Type I error rate. Tukey's WSD test holds the error rate stable across multiple comparisons of means. This function implements an adaptation of Tukey's WSD test from Maxwell & Delaney (2004), that allows variances and sample sizes to differ across groups.

Value

A vector with three elements:

1. q The q statistic
2. df The degrees of freedom for the q statistic
3. p A p value based on the q statistic, degrees of freedom and the total number of groups to be compared

Author(s)

Alexander M. Schoemann (East Carolina University; <schoemanna@ecu.edu>)

References

Maxwell, S. E., & Delaney, H. D. (2004). *Designing experiments and analyzing data: A model comparison perspective* (2nd ed.). Mahwah, NJ.: Lawrence Erlbaum Associates.

Examples

```
##For a case where three groups have been compared:
##Group 1: mean = 3.91, var = 0.46, n = 246
##Group 2: mean = 3.96, var = 0.62, n = 465
##Group 3: mean = 2.94, var = 1.07, n = 64

#compare group 1 and group 2
tukeySEM(3.91, 3.96, 0.46, 0.62, 246, 425, 3)

#compare group 1 and group 3
tukeySEM(3.91, 2.94, 0.46, 1.07, 246, 64, 3)

#compare group 2 and group 3
tukeySEM(3.96, 2.94, 0.62, 1.07, 465, 64, 3)
```

wald

Calculate multivariate Wald statistics

Description

Calculate multivariate Wald statistics based on linear combinations of model parameters

Usage

```
wald(object, syntax)
```

Arguments

object	An output from lavaan
syntax	Syntax that each line represents one linear constraint. A plus or minus sign is used to separate between each coefficient. An asterisk is used to separate between coefficients and parameters. The coefficient can have a forward slash to represent a division. The parameter names must be matched with the names of lavaan parameters investigated by running the <code>coef</code> function on a lavaan

output. Lines can be separated by semi-colon. A pound sign is allowed for comments. Note that the defined parameters (created by ":=") do not work with this function.

Details

The formula for multivariate Wald test is

$$\chi^2 = (C\hat{b})' [C\hat{V}C']^{-1} (C\hat{b}),$$

where C is the contrast matrix, \hat{b} is the estimated fixed effect, \hat{V} is the asymptotic covariance matrix among fixed effects.

Value

Chi-square value with p value.

Author(s)

Sunthud Pornprasertmanit (<psunthud@ku.edu>)

Examples

```
# Test the difference in factor loadings
HS.model <- ' visual  =~ x1 + con1*x2 + con1*x3
              textual =~ x4 + x5 + x6
              speed   =~ x7 + con2*x8 + con2*x9 '

fit <- cfa(HS.model, data=HolzingerSwineford1939)
wald(fit, "con2 - con1")

# Simultaneously test the difference in the influences
# of x1 and x2 on intercept and slope
model.syntax <- '
  i =~ 1*t1 + 1*t2 + 1*t3 + 1*t4
  s =~ 0*t1 + 1*t2 + 2*t3 + 3*t4
  i ~ x1 + x2
  s ~ x1 + x2
  t1 ~ c1
  t2 ~ c2
  t3 ~ c3
  t4 ~ c4
'

fit2 <- growth(model.syntax, data=Demo.growth)
wald.syntax <- '
i~x1 - i~x2
1/2*s~x1 - 1/2*s~x2
'
wald(fit2, wald.syntax)

# Mplus example of MODEL TEST
model3 <- ' f1  =~ x1 + p2*x2 + p3*x3 + p4*x4 + p5*x5 + p6*x6
p4 == 2*p2 '
```

```
fit3 <- cfa(model3, data=HolzingerSwineford1939)
wald(fit3, "p3; p6 - 0.5*p5")
```