Logistic Regression Homework, STAT5120, Allen Baumgarten

Load the faraway package and then type "wbca" at the prompt...but without the quotes. Read about this dataset here: https://cran.r-project.org/web/packages/faraway/faraway.pdf. There are 681 cases of potentially cancerous tumors of which 238 are actually malignant. Determining whether a tumor is really malignant is traditionally determined by an invasive surgical procedure. The purpose of this study was to determine whether a new procedure called fine needle aspiration which draws only a small sample of tissue could be effective in determining tumor status.

(a) Fit a binomial regression with Class as the response and the other nine variables as predictors. Just do model1 <- glm(Class ~ ., data = wbca, family=binomial) summary(model1)

A scatterplot matrix was constructed to examine these variables. Output for an all-inclusive model shows that five of the nine variables are statistically significant.

wbca data 8 10 6 8 10 wbca.Adhes wbca.BNucl wbca.Chrom wbca.Epith wbca.Mitos wbca.NNucl wbca\Thick 6 8 10 6 8 10 6 8 10 4 6 8 10

Deviance Residuals:

```
Min
          1Q
                 Median
                           3Q
                                    Max
-2.48282 -0.01179 0.04739 0.09678 3.06425
```

Coefficients:

```
Estimate
                    Std. Error z value Pr(>|z|)
                              7.892 2.97e-15 ***
(Intercept) 11.16678 1.41491
Adhes
           -0.39681 0.13384
                              -2.965 0.00303 **
BNucl
          -0.41478 0.10230
                              -4.055
                                      5.02e-05 ***
Chrom
           -0.56456 0.18728
                              -3.014 0.00257 **
           -0.06440 0.16595
                             -0.388 0.69795
Epith
           -0.65713 0.36764
                             -1.787 0.07387.
Mitos
                              -2.271 0.02315 *
NNucl
           -0.28659 0.12620
Thick
           -0.62675 0.15890
                             -3.944 8.01e-05 ***
UShap
           -0.28011 0.25235
                              -1.110 0.26699
USize
                              0.246
            0.05718 0.23271
                                     0.80589
```

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 881.388 on 680 degrees of freedom Residual deviance: 89.464 on 671 degrees of freedom

AIC: 109.46

Number of Fisher Scoring iterations: 8

Report the residual deviance and associated degrees of freedom. Can this information be used to determine if this model fits the data well (think in terms of sample sizes)? If so, what does the chi-squared test for the residual deviance indicate?

Joseph Hilbe comments on the Null and Residual deviances by saying that, "...an intercept model only, that is, a model with no predictors, is the called the null deviance."1

The residual deviance and df for this model iteration are 89.464 on 671 df, respectively. Df for the null deviance are the number of observations minus 1 (n-1). The null deviance and residual deviance can be used to calculate the Likelihood Ratio Test (G²) which is basically a difference in the model deviances, i.e., for our model of interest vs. a null model with an intercept only. In this case, we get 881.388 - 89.464 = 791.9 which compared to a χ^2 provides strong evidence for us to reject the null hypothesis that our betas = 0 and provide no information on our predicted variable.

(b) Use AIC as the criterion in best-subsets method to determine the best subset of variables. This is easy-just do reduced <- step(model1) summary(reduced)

Alan Agresti remarks that, "The AIC judges a model by how close its fitted values tend to be from the true mean values, in terms of expected value. Even though a simple model farther from the true relationship than is a more

¹ Hilbe, Joseph M., Practical Guide to Logistic Regression, (CRC Press: 2016), 54.

complex model, it may be preferred because it tends to provide better estimates of certain characteristics, such as cell probabilities. Thus, the optimal model is the one that tends to have fit closest to the true values. Akaike defined closeness in terms of a Kullback-Leibler measure of distance...With a sample, this criterion selects the model minimizes

```
AIC = -2(maximized log likelihood - number of parameters in model)^2
```

Dr. Hilbe's comments on the AIC, however, suggest care in using it: "The AIC is perhaps the most well-known and well used information statistic in current research. What may seem surprising to many readers is that there are a plethora of journal articles detailing studies proving how poor the AIC test is in assessing which of two models is the better fitted. Even Akaike himself later developed another criterion which he preferred to the original. However, it is his original 1973 version that is used by most researchers and that is found in most journals to assess comparative model fit. The traditional AIC statistic is found in two versions:

```
AIC = -2L + 2k \text{ or } -2(L - k) or AIC = (-2L + 2k)/n \text{ or } 2(L - k)/n
```

where L is the log-likelihood model, k is the number of parameter estimates in the model, and n is the number of observations in the model."³

Models were split into subsets as follows. The lowest AIC statistic was shown to be around 105 in the select stepwise model below under part (c).

```
Start: AIC=109.46
Class ~ Adhes + BNucl + Chrom + Epith + Mitos + NNucl + Thick + UShap + USize
```

```
Df Deviance AIC
- USize 1 89.523
                   107.52
- Epith 1 89.613
                   107.61
- UShap 1 90.627
                   108.63
<none>
           89.464
                    109.46
- Mitos 1 93.551
                    111.55
- NNucl 1 95.204
                    113.20
- Adhes 1 98.844
                    116.84
- Chrom 1 99.841
                    117.84
- BNucl 1 109.000
                    127.00
- Thick 1 110.239
                    128.24
```

Step: AIC=107.52

Class ~ Adhes + BNucl + Chrom + Epith + Mitos + NNucl + Thick + UShap

```
Df Deviance AIC
- Epith 1 89.662 105.66
- UShap 1 91.355 107.36
```

² Agresti, Alan, Categorical Data Analysis, 3rd ed., (New Jersey: John Wiley & Sons, 2013), 212.

³ Hilbe, 58.

```
<none>
           89.523
                   107.52
           93.552
                   109.55
- Mitos 1
- NNucl 1
           95.231
                   111.23
- Adhes 1 99.042
                   115.04
- Chrom 1 100.153
                   116.15
- BNucl 1 109.064
                   125.06
- Thick 1 110.465
                    126.47
```

Step: AIC=105.66

Class ~ Adhes + BNucl + Chrom + Mitos + NNucl + Thick + UShap

```
      Df
      Deviance
      AIC

      <none>
      89.662
      105.66

      - UShap 1
      91.884
      105.88

      - Mitos 1
      93.714
      107.71

      - NNucl 1
      95.853
      109.85

      - Adhes 1
      100.126
      114.13

      - Chrom 1
      100.844
      114.84

      - BNucl 1
      109.762
      123.76

      - Thick 1
      110.632
      124.63
```

(c) Use the reduced model (the one from part (b)) to predict the outcome (malignant or not) for a patient with Adhes = 1, BNucl = 1, Chrom = 3, Epith = 2, Mitos = 1, NNucl = 1, Thick = 4, UShap = 1, USize = 1. Also, give a 95% confidence interval for your prediction. Our reduced model is seen to be:

Deviance Residuals:

```
Min 1Q Median 3Q Max -2.44161 -0.01119 0.04962 0.09741 3.08205
```

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	11.0333	1.3632	8.094	5.79e-16 ***
Adhes	-0.3984	0.1294	-3.080	0.00207 **
BNucl	-0.4192	0.1020	-4.111	3.93e-05 ***
Chrom	-0.5679	0.1840	-3.085	0.00203 **
Mitos	-0.6456	0.3634	-1.777	0.07561.
NNucl	-0.2915	0.1236	-2.358	0.01837 *
Thick	-0.6216	0.1579	-3.937	8.27e-05 ***
UShap	-0.2541	0.1785	-1.423	0.15461

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 881.388 on 680 degrees of freedom Residual deviance: 89.662 on 673 degrees of freedom

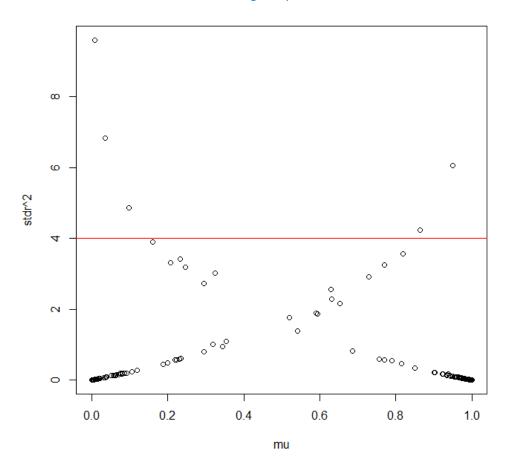
AIC: 105.66

Number of Fisher Scoring iterations: 8

BIC: 141.8503

Hilbe advises that, "The Schwartz Bayesian information criterion (BIC) is the most used BIC test found in the literature...My recommendation is to test models with both [the AIC and BIC]. If the values substantially differ, it is likely the model is mis-specified." Here we see that the AIC and BIC so seem to differ somewhat. Residuals analysis would be in order but we will forgo that for the time being.

There are some residuals that seem to greatly skewed as seen in the standardized residuals plot:



We will use this reduced model to predict the outcome of a patient with the values above. What we will have is the log-odds of having (or not having) a cancerous tumor, given these values. Our model is:

Y = 11.033 + -0.398(Adhes) + -0.419(BNucl) + -0.568(Chrom) + -0.646(Mitos) + -0.292(NNucl) + -0.622(Thick) + -0.254(Ushap)

Our proposed values above suggest the likelihood of cancerous tumor expression to be:

$$Y = 11.033 + -0.398(1) + -0.419(1) + -0.568(3) + -0.646(1) + -0.292(1) + -0.622(4) + -0.254(1)$$

$$Y = 11.033 - 0.398 - 0.419 - 1.704 - 0.646 - 0.292 - 2.488 - 0.254$$

$$Y = rt$$

⁴ Hilbe, 60.

- (d) Suppose a tumor is classified as benign if p > 0.5 and classified as malignant if p < 0.5 (remember "1" means benign and "0" indicates malignant for the Class variable). Compute the number of errors of both types that will be made if this method is applied to the current data with the reduced model. Also, compute the percentage of classifications that result in each kind of error (that is, compute the error rates- false positive and false negative).
- (e) Suppose we move the cut-off point to 0.9 so that p < 0.9 indicates malignant and p > 0.9 indicates benign. What are the new error counts and rates?
- (f) It can be misleading to use the same data to fit a model and test its predictive ability. So split the original dataset into two parts: assign every third record to the test set and all the others to the training set. Use the training set to determine a good model (repeat parts (a) and (b)). Then use the test set to assess predictive performance (repeat parts (d) and (e)).
- (g) Discuss how you could search for the "best" cut-off point to use for classifying tumors. Then write an R program to carry it out. What is the "best" cut-off value? What do you mean by "best"?

APPENDIX: R SCRIPTS

Part (a):

>scatterplotMatrix(~wbca\$Adhes+wbca\$BNucl+wbca\$Chrom+wbca\$Epith+wbca\$Mitos+wbca\$NNucl+wbca\$Thic k, data=wbca, main="wbca data")

> model1 <- glm(Class ~ ., data = wbca, family=binomial)

> summary(model1)

Call:

glm(formula = Class ~ ., family = binomial, data = wbca)

Deviance Residuals:

Min 1Q Median 3Q Max -2.48282 -0.01179 0.04739 0.09678 3.06425

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	11.16678	1.41491	7.892	2.97e-15 ***
Adhes	-0.39681	0.13384	-2.965	0.00303 **
BNucl	-0.41478	0.10230	-4.055	5.02e-05 ***
Chrom	-0.56456	0.18728	-3.014	0.00257 **
Epith	-0.06440	0.16595	-0.388	0.69795
Mitos	-0.65713	0.36764	-1.787	0.07387.
NNucl	-0.28659	0.12620	-2.271	0.02315 *
Thick	-0.62675	0.15890	-3.944	8.01e-05 ***
UShap	-0.28011	0.25235	-1.110	0.26699
USize	0.05718	0.23271	0.246	0.80589

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 881.388 on 680 degrees of freedom

Residual deviance: 89.464 on 671 degrees of freedom

AIC: 109.46

Number of Fisher Scoring iterations: 8

Part (b):

> reduced <- step(model1)

Start: AIC=109.46

Class ~ Adhes + BNucl + Chrom + Epith + Mitos + NNucl + Thick + UShap + USize

```
        Df
        Deviance
        AIC

        - USize
        1
        89.523
        107.52

        - Epith
        1
        89.613
        107.61

        - UShap
        1
        90.627
        108.63

        <none>
        89.464
        109.46

        - Mitos
        1
        93.551
        111.55

        - NNucl
        1
        95.204
        113.20

        - Adhes
        1
        98.844
        116.84

        - Chrom
        1
        99.841
        117.84

        - BNucl
        1
        109.000
        127.00

        - Thick
        1
        110.239
        128.24
```

Step: AIC=107.52

Class ~ Adhes + BNucl + Chrom + Epith + Mitos + NNucl + Thick + UShap

```
Df Deviance AIC
- Epith 1 89.662 105.66
- UShap 1 91.355 107.36
<none> 89.523 107.52
- Mitos 1 93.552 109.55
- NNucl 1 95.231 111.23
- Adhes 1 99.042 115.04
- Chrom 1 100.153 116.15
- BNucl 1 109.064 125.06
- Thick 1 110.465 126.47
```

Step: AIC=105.66

Class ~ Adhes + BNucl + Chrom + Mitos + NNucl + Thick + UShap

```
Df Deviance AIC
<none> 89.662 105.66
- UShap 1 91.884 105.88
- Mitos 1 93.714 107.71
- NNucl 1 95.853 109.85
- Adhes 1 100.126 114.13
- Chrom 1 100.844 114.84
- BNucl 1 109.762 123.76
- Thick 1 110.632 124.63
```

```
Part (c):
```

```
> summary(reduced)
```

Call:

glm(formula = Class ~ Adhes + BNucl + Chrom + Mitos + NNucl + Thick + UShap, family = binomial, data = wbca)

Deviance Residuals:

```
Min 1Q Median 3Q Max -2.44161 -0.01119 0.04962 0.09741 3.08205
```

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	11.0333	1.3632	8.094	5.79e-16 ***
Adhes	-0.3984	0.1294	-3.080	0.00207 **
BNucl	-0.4192	0.1020	-4.111	3.93e-05 ***
Chrom	-0.5679	0.1840	-3.085	0.00203 **
Mitos	-0.6456	0.3634	-1.777	0.07561.
NNucl	-0.2915	0.1236	-2.358	0.01837 *
Thick	-0.6216	0.1579	-3.937	8.27e-05 ***
UShap	-0.2541	0.1785	-1.423	0.15461

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 881.388 on 680 degrees of freedom Residual deviance: 89.662 on 673 degrees of freedom

AIC: 105.66

Number of Fisher Scoring iterations: 8

```
> wbca_regmod <- glm(wbca$Class ~ wbca$Adhes + wbca$BNucl + wbca$Chrom + wbca$Mitos + wbca$NNucl + wbca$Thick + wbca$UShap, data=wbca, family = binomial)
```

> summary(wbca_regmod)

current_model <- wbca_regmod #Run glm(), assign to this object; also update lines 96, 103 and 114 current_data <- wbca #Be sure this points to the current data source

```
G2 <- current_model$null.deviance - current_model$deviance
EL50 <- -(-12.3508)/0.4972 # Input the EL50 = -a/B (negated intercept / Beta)
```

```
# Print summary of results and coefficients
print(summary(current_model))
writeLines("Deviance Statistic:")
print(G2)
writeLines("")
writeLines("Coefficients Exponentiated:")
print(exp(coef(current_model)))
writeLines("")
```

```
writeLines("")
writeLines("95% CI's (Wald):")
print(confint.default(current model))
writeLines("")
writeLines("")
writeLines("95% CI's (Wald) Exponentiated:")
print(exp(confint.default(current model)))
writeLines("")
writeLines("")
writeLines("95% CI's (Profile/Likelihood Ratio):")
print(confint(current model))
writeLines("")
writeLines("")
writeLines("95% CI's (Profile/Likelihood Ratio) Exponentiated:")
print(exp(confint(current model)))
writeLines("")
writeLines("")
# Runs a Pearson GOF test. If p>.05 then model is probably well-fitted though check for overdispersion, AIC, etc.
pr <- sum(residuals(current model, type = "pearson")^2)</pre>
df <- current model$df.residual
p_value <- pchisq(pr, current_model$df.residual, lower=F)</pre>
print(matrix(c("Pearson Chi GOF", "Chi2", "df", "p-value", "Parameters", round(pr,4),df,round(p_value,4)),
ncol=2))
writeLines("")
writeLines("")
# Print standardized residuals (full model)
mu <- current_model$fitted.value</pre>
dr <- resid(current model, type = "deviance")</pre>
hat <- hatvalues(current_model)</pre>
stdr <- dr/sqrt(1-hat)
windows()
plot(mu, stdr^2)
abline(h = 4, col = "red")
# Predicted values (odds) of y given a continuous predictor
predict <- predict(current_model)</pre>
fit <- current model$fitted.values
# Calculate standard errors of the linear predictor
lpred <- predict(current_model, newdata = current_data, type = "link", se.fit = TRUE)</pre>
up <- lpred$fit + (qnorm(.975) * lpred$se.fit)
low <- lpred$fit - (qnorm(.975) * lpred$se.fit)</pre>
eta <- Ipred$fit
upci <- current model$family$linkinv(up)</pre>
mu <- current model$family$linkinv(eta)</pre>
loci <- current model$family$linkinv(low)</pre>
writeLines("Lower CI:")
print(summary(loci))
```

```
writeLines("")
writeLines("Mean CI:")
print(summary(mu))
writeLines("")
writeLines("Upper CI:")
print(summary(upci))
writeLines("")
# Bayseian Information Criterion
library(COUNT)
writeLines("AIC and BIC Statistics:")
print(modelfit(current model))
# Graph of probabilities of continuous predictor. Remove #s and > and write in continuous variable name to the
right of $s:
# > layout(1)
# > plot(current_data$BMI, mu, col = 1)
# > lines(current data$BMI, loci, col = 2, type = 'p')
# > lines(current data$BMI, upci, col = 3, type = 'p')
# To check for overdispersion in the model: scaled and robust/sandwhich se's:
coef <- current_model$coefficients</pre>
se <- sqrt(diag(vcov(current model)))</pre>
coefse <- data.frame(coef, se)</pre>
writeLines("")
pr <- resid(current model, type = "pearson")</pre>
pchi2 <- sum(residuals(current model, type = "pearson")^2)</pre>
disp <- pchi2/current_model$df.residual</pre>
scse <- se*sqrt(disp)</pre>
library(sandwich)
rmodel <- glm(current_data$Class ~ current_data$Adhes + current_data$BNucl + current_data$Chrom +
current_data$Mitos + current_data$NNucl + current_data$Thick + current_data$UShap, family = binomial, data =
current data) #Update variables!!
# WITH FACTOR: rmodel <- glm(current data$y ~ + current data$weight+ current data$los +
factor(current data$type), family = binomial, data = current data) #Update variables!!
rse <- sqrt(diag(vcovHC(rmodel, type = "HC0")))
newcoefse <- data.frame( coef, se, scse, rse)</pre>
print(newcoefse)
# Quasibinomial model: check that coefficients are identical or nearly so. If so, then good fit:
quasibinomialmod <- glm(current_data$Class ~ current_data$Adhes + current_data$BNucl + current_data$Chrom
+ current_data$Mitos + current_data$NNucl + current_data$Thick + current_data$UShap, family = quasibinomial,
data = current data) #Update variables!!
# WITH FACTOR: quasibinomialmod <- glm(current_data$died ~ + current_data$white + current_data$hmo +
current data$los + factor(current data$type), family = quasibinomial, data = current data) #Update variables!!
writeLines("")
writeLines("")
writeLines("Quasibinomial model:")
print(summary(quasibinomialmod))
```

```
> source("Log_REG_working.r")

Call:
glm(formula = wbca$Class ~ wbca$Adhes + wbca$BNucl + wbca$Chrom +
    wbca$Mitos + wbca$NNucl + wbca$Thick + wbca$UShap, family = binomial,
    data = wbca)

Deviance Residuals:
    Min 1Q Median 3Q Max
-2.44161 -0.01119 0.04962 0.09741 3.08205

Coefficients:
    Estimate Std. Error z value Pr(>|z|)
```

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 881.388 on 680 degrees of freedom Residual deviance: 89.662 on 673 degrees of freedom

AIC: 105.66

Number of Fisher Scoring iterations: 8

Deviance Statistic:

[1] 791.7264

Coefficients Exponentiated:

(Intercept) wbca\$Adhes wbca\$BNucl wbca\$Chrom wbca\$Mitos wbca\$NNucl wbca\$Thick wbca\$UShap 6.190351e+04 6.713727e-01 6.575746e-01 5.667417e-01 5.243413e-01 7.471176e-01 5.370696e-01 7.756406e-01

95% CI's (Wald):

	2.5 %	97.5 %
(Intercept)	8.3615066	13.70515770
wbca\$Adhes	-0.6519565	-0.14490519
wbca\$BNucl	-0.6190357	-0.21935844
wbca\$Chrom	-0.9285735	-0.20712955
wbca\$Mitos	-1.3578036	0.06657864
wbca\$NNucl	-0.5338499	-0.04921541
wbca\$Thick	-0.9311288	-0.31212640

95% CI's (Wald) Exponentiated:

2.5 % 97.5 %

(Intercept) 4279.1369847 8.955181e+05 wbca\$Adhes 0.5210254 8.651043e-01 wbca\$BNucl 0.5384634 8.030338e-01 wbca\$Chrom 0.3951169 8.129143e-01 wbca\$NNucl 0.5863432 9.519760e-01 wbca\$Thick 0.3941086 7.318890e-01 wbca\$UShap 0.5466757 1.100503e+00

95% CI's (Profile/Likelihood Ratio):

Waiting for profiling to be done...

2.5 % 97.5 %

(Intercept) 8.7277704 14.15772490 wbca\$Adhes -0.6722270 -0.15465473 wbca\$BNucl -0.6339696 -0.22932801 wbca\$Chrom -0.9534347 -0.22625284 wbca\$Mitos -1.2507890 -0.01303244 wbca\$NNucl -0.5466091 -0.05972893 wbca\$Thick -0.9635139 -0.33585675 wbca\$UShap -0.6291944 0.07595514

95% CI's (Profile/Likelihood Ratio) Exponentiated:

Waiting for profiling to be done...

2.5 % 97.5 %

(Intercept) 6171.9518030 1.408062e+06 wbca\$Adhes 0.5105703 8.567109e-01 wbca\$BNucl 0.5304818 7.950677e-01 wbca\$Chrom 0.3854150 7.975164e-01 wbca\$Mitos 0.2862788 9.870521e-01 wbca\$NNucl 0.5789095 9.420199e-01 wbca\$UShap 0.5330210 1.078914e+00

- [,2] [,1]
- [1,] "Pearson Chi GOF" "Parameters"
- [2,] "Chi2" "227.9781"
- [3,] "df" "673"
- "1" [4,] "p-value"

Lower CI:

Min. 1st Qu. Median Mean 3rd Qu. Max.

$0.000000\ 0.000915\ 0.966333\ 0.621669\ 0.990807\ 0.997395$

Mean CI:

Min. 1st Qu. Median Mean 3rd Qu. Max. 0.000000 0.007241 0.989560 0.650514 0.997888 0.999605

Upper CI:

Min. 1st Qu. Median Mean 3rd Qu. Max. 0.0000022 0.0732503 0.9970059 0.6850585 0.9995259 0.9999401

AIC and BIC Statistics:

\$AIC

[1] 105.6618

\$AICn

[1] 0.1551568

\$BIC

[1] 141.8503

\$BICqh

[1] 0.1805181

	coef	se	scse	rse
(Intercept)	11.0333322	1.3632013	0.79341251	1.1988868
wbca\$Adhes	-0.3984309	0.1293522	0.07528577	0.1510720
wbca\$BNucl	-0.4191971	0.1019604	0.05934312	0.1216056
wbca\$Chrom	-0.5678516	0.1840452	0.10711828	0.1603045
wbca\$Mitos	-0.6456125	0.3633695	0.21148886	0.3238224
wbca\$NNucl	-0.2915327	0.1236335	0.07195737	0.1128367
wbca\$Thick	-0.6216276	0.1579117	0.09190800	0.1314029
wbca\$UShap	-0.2540660	0.1784898	0.10388488	0.2047329

Quasibinomial model:

Call:

```
glm(formula = current_data$Class ~ current_data$Adhes + current_data$BNucl +
    current_data$Chrom + current_data$Mitos + current_data$NNucl +
    current_data$Thick + current_data$UShap, family = quasibinomial,
    data = current_data)
```

Deviance Residuals:

```
Min 1Q Median 3Q Max -2.44161 -0.01119 0.04962 0.09741 3.08205
```

Coefficients:

```
Estimate Std. Error t value Pr(>|t|)
(Intercept) 11.03333 0.79351 13.904 < 2e-16 ***
```

(Dispersion parameter for quasibinomial family taken to be 0.338836)

Null deviance: 881.388 on 680 degrees of freedom Residual deviance: 89.662 on 673 degrees of freedom

AIC: NA

Number of Fisher Scoring iterations: 8