Index Selection

Sweetpotato breeders Meeting, Malawi

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Outline

- Two selection indices
- 2 Computations
- 3 Example
- 4 Exercise
- Conclusions

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Why to use a selection index?

- In breeding usually several traits have to be improved simultaneously.
- Very often intuitive procedures are used.
- Intuition usually fails when several correlated traits are involved:
 - Positive correlations: Improving one trait improves the other.
 - Negative correlations: Improving one trait can diminish the other.
- And do not forget the variability:
 - High variability: A lot of room to improve.
 - Low variability: No so much room to improve.

Two selection indices

- Elston, R. C. (1963). A weight-free index for the purpose of ranking or selection with respect to several traits at a time. *Biometrics*. 19(1): 85-97.
- Pesek, J. and R.J. Baker.(1969). Desired improvement in relation to selection indices. *Can. J. Plant. Sci.* 9:803-804.

Elston

The Elston index is a weight free index. Given p traits, for each genotype the Elston index is computed as

$$I_E = \prod_{i=1}^p (x_i - k_i)$$

where x_i is the value of the genotype for trait i and k_i is some lower bound. Two options for k:

- $k_i = \min x_i$
- $k_i = \frac{n \min x_i \max x_i}{n-1}$.

Pesek-Baker

The Pesek Baker is an index where weights are given in the form of desired gains. Given p traits, this index is defined by

$$I_{PB} = \sum_{i=1}^{p} b_i x_i$$

where x_i is the value of the genotype for trait i. The coefficients, b_i , are computed from:

$$\mathbf{b} = \mathbf{V}^{-1}\mathbf{g}$$

with

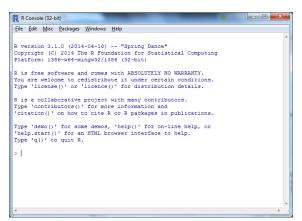
- **b** the vector of index coefficients.
- V the genetic variance-covariance matrix, and
- g the vector of desired genetic gains to be specified by the breeder.

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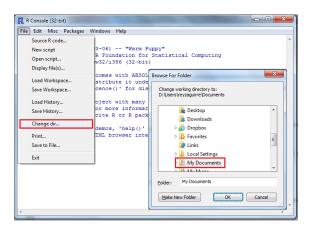
Open R

We will use R for the example. If you have Clone Selector, you have R. Open R.



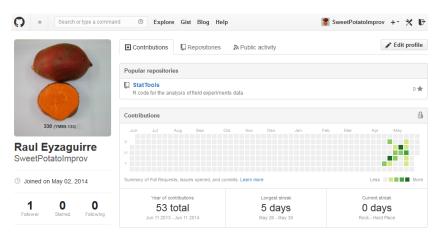
Define your working directory

Define your working directory, My Documents by default.



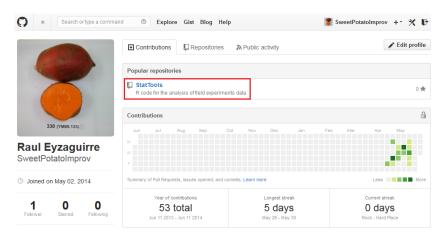
GitHub account

GitHub is a good place to put code. https://github.com/SweetPotatoImprov



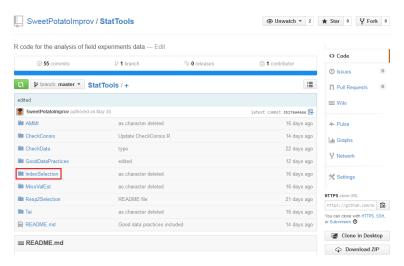
GitHub repo

There is a GitHub repository for the R functions and some other documents: https://github.com/SweetPotatoImprov/StatTools

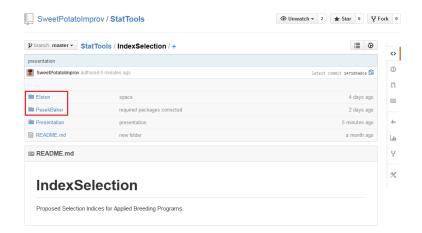


GitHub repo

Go into the IndexSelection folder:



Two selection indices



Download the files

Download the R functions (Elston.R and PesekBaker.R) into your working directory.

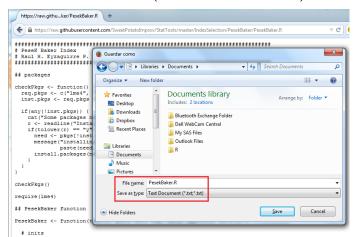
Click on the function.



Click on Raw.

Download the files

Right click on the mouse and Save as ... or go to Save web page as ... Beware of browsers, some are not so smart. Make sure to save the file with the real name (Elston.R or PesekBaker.R)



Load the functions directly from GitHub

You can also load the R functions directly from GitHub into R. First define the URL of the files:

This works well if you use RStudio:

```
> source(urlfile1)
> source(urlfile2)
```

This works well directly from R:

```
> require(RCurl)
> file1 <- getURL(urlfile1, ssl.verifypeer = FALSE)
> file2 <- getURL(urlfile2, ssl.verifypeer = FALSE)
> eval(parse(text = file1))
> eval(parse(text = file2))
```

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Data

Data is in the file SI_example.csv.

This file contains data for:

- 2 locations: La Molina and Satipo.
- 2 replications in each location.
- 1041 genotypes.
- 25 traits.
- 3 plants per plot.

Small data

Let us start with a small subset in the file SI_example_small.csv.

This file contains data for:

- 2 locations: La Molina and Satipo.
- 2 replications in each location.
- 8 genotypes.
- 5 traits: RYTHA, BC, DM, STAR, NOCR.
- 3 plants per plot.

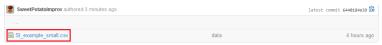
Download the data

Data are on GitHub, so you can proceed in the same way as we did to download the functions

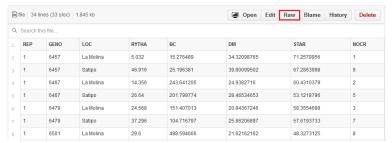


Download the data

Click on the file.

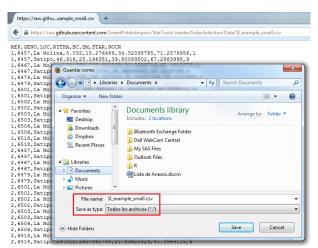


Click on Raw.



Download the data

Right click on the mouse and Save as . . . or go to Save web page as . . . Make sure to save the file with the real name and format.



Load the data directly from GitHub

You can also load the data directly from GitHub into R. First define the URL of the file:

```
> urlfile <- "https://raw.githubusercontent.com/SweetPotatoImprov/StatTools/
+ master/IndexSelection/Data/SI_example_small.csv"</pre>
```

This works well if you use RStudio:

```
> mydata <- read.csv(urlfile)
```

This works well directly from R:

```
> require(RCurl)
> mydata <- getURL(urlfile, ssl.verifypeer = FALSE)
> mydata <- read.csv(textConnection(mydata))</pre>
```

Load the data and functions

If you have the files (SI_example_small.csv, Elston.R, PesekBaker.R) in your working directory, proceed like this:

```
> mydata <- read.csv("SI_example_small.csv")
> mydata$REP <- factor(mydata$REP)
> mydata$GENO <- factor(mydata$GENO)
> str(mvdata)
                    32 obs. of 8 variables:
'data.frame':
 $ REP : Factor w/ 2 levels "1", "2": 1 1 1 1 1 1 1 1 1 1 ...
 $ GENO : Factor w/ 8 levels "6457", "6467", ...: 1 1 2 2 3 3 4 4 5 5 ...
 $ LOC : Factor w/ 2 levels "La Molina", "Satipo": 1 2 1 2 1 2 1 2 1 2 ...
 $ RYTHA: num 5.03 46.92 14.36 26.64 24.57 ...
 $ BC : num 15.3 25.2 243.6 201.8 151.4 ...
 $ DM : num 34.3 39.8 24.9 28.5 20.8 ...
 $ STAR : num 71.3 67.3 60.4 53.1 58.4 ...
 $ NOCR: int 19253781167...
> source('Elston.R')
> source('PesekBaker.R')
```

Elston index - Syntaxes

The syntaxes and arguments for this function are:

Elston(traits, geno, data, lb = 1)

- traits is a list of traits,
- geno is the index for genotypes,
- data is the data frame containing the data,
- 1b is the lower bound, 1 for $k_i = \min x_i$ and 2 for $k_i = \frac{n \min x_i \max x_i}{n-1}$ 1 by default.

Elston index - Computation

```
> Elston(c('RYTHA', 'BC', 'DM', 'STAR', 'NOCR'), 'GENO', mydata)
$Elston.Index
   6457
            6467
                    6479
                             6501
                                  6502
                                              6503
                                                      6506
                                                               6518
0.09313 0.00000 1.18126 4.82440 33.84316 0.28160 0.00000 0.00000
$Sorted.Elston.Index
   6502
            6501
                    6479
                             6503
                                     6457
                                              6467
                                                      6506
                                                               6518
33.84316 4.82440
                1.18126 0.28160 0.09313 0.00000
                                                   0.00000 0.00000
```

Remember that by default it uses $k_i = \min x_i$ for the index

$$I_E = \prod_{i=1}^p (x_i - k_i)$$

Elston index - Computation

```
We can use the second lower bound, k_i = \frac{n \min x_i - \max x_i}{n-1}, writing 1b=2.
> Elston(c('RYTHA', 'BC', 'DM', 'STAR', 'NOCR'), 'GENO', lb=2, mydata)
$Elston.Index
 6457 6467
             6479
                      6501
                             6502
                                    6503
                                           6506
                                                  6518
 4.862 0.710 6.790 19.822 84.154 11.965 18.425
                                                 3.439
$Sorted.Elston.Index
 6502
        6501
               6506
                      6503 6479
                                    6457 6518
                                                  6467
84.154 19.822 18.425 11.965 6.790 4.862 3.439 0.710
```

Pesek Baker index

Let us compute the Pesek Baker index using these traits (means between brackets):

- RYTHA (31.3 tons/ha)
- BC (188 ppm)
- DM (30.05%)
- STAR (60.3%)
- NOCR (6.66)

Which are your desired genetic gains?

My guess: 5, 100, 2, 2, 5.

Pesek Baker index - Syntaxes

The syntaxes and arguments for this function are:

```
PesekBaker(traits, geno, env, rep, data, dgg = NULL, units =
"sdu", sf = 0.1)
```

- traits is a list of traits,
- geno is the index for genotypes,
- env is the index for environments,
- rep is the index for replications,
- data is the data frame containing the data,
- dgg is the vector of desired genetic gains, one standard deviation by default.
- units are the units for dgg, "actual" for actual units and "sdu" for standard deviations, "sdu" by default, and
- sf is the selected fraction, 0.1 by default.

Pesek Baker index - Output

The PesekBaker function returns the following elements:

- \$Desired.Genetic.Gains: The desired genetic gains in actual units.
- \$Standard.Deviations: The estimated standard deviations.
- \$Genetic.Variance: The estimated genetic variances.
- \$Correlation.Matrix: The estimated correlation matrix.
- \$Index.Coefficients: The index coefficients.
- \$Response.to.Selection: The response to selection.
- \$Std.Response.to.Selection: The standardized response to selection.
- \$Pesek.Baker.Index: The Pesek-Baker index value.
- \$Sorted.Pesek.Baker: The Pesek-Baker index value sorted in descending order.

Pesek Baker Index - Computation

Why is the RYTHA coefficient negative?

- It does not mean that we are going to select to diminish root yield.
- We must see the correlations.
- We must see the variances.

See the correlations

We want to improve all the traits, but some have a negative correlation.

There are also traits with high positive correlations.

It is difficult to have this kind of correlation structures on mind.

> output\$Correlation.Matrix

	RYTHA	BC	DM	STAR	NOCR
RYTHA	1.0000	0.26732	-0.231076	-0.15266	0.615558
BC	0.2673	1.00000	-0.775152	-0.85198	0.091521
DM	-0.2311	-0.77515	1.000000	0.89555	0.004778
STAR	-0.1527	-0.85198	0.895548	1.00000	0.077349
NOCR	0.6156	0.09152	0.004778	0.07735	1.000000

See the standard deviations

Maybe we want to improve too much a trait with a very low variability. If a trait has high variability, then there is a lot of room to improve. Let us remember my desired gains: 5, 100, 2, 2, 5.

```
> output$Standard.Deviations
[1] 5.068 151.583 5.859 8.231 1.663
> output$Response.to.Selection
[1] 1.765 35.298 0.706 0.706 1.765
> output$Std.Response.to.Selection
[1] 0.34827 0.23286 0.12049 0.08577 1.06140
```

Compute the index - second try

I will relax a little bit my ambition about NOCR.

```
> output <- PesekBaker(c('RYTHA', 'BC', 'DM', 'STAR', 'NOCR'), 'GENO', 'LOC', 'REP',
                      mydata, c(5, 100, 2, 2, 2.5), "actual")
> output$Index.Coefficients
         coef
BYTHA 0.01853
BC.
   0.01669
DM
     0.20259
STAR 0.15350
NOCR.
    0.66803
> output$Response.to.Selection
    3,605 72,102 1,442 1,442 1,803
[1]
> output$Std.Response.to.Selection
[1] 0.7114 0.4757 0.2461 0.1752 1.0841
```

Using the defaults

By default the function gives weights so that the desired genetic gains are one standard deviation for each trait. Same relative weight for each trait.

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Load the data

Load the complete data set.

```
> mydata <- read.csv("SI_example.csv")
> mydata$REP <- factor(mydata$REP)
> mydata$GENO <- factor(mydata$GENO)
> names(mydata)
```

```
[1] "REP"
             "GENO"
                      "LOC"
                              "PROT"
                                       "BC"
                                               "FE"
                                                        "ZN"
                                                                "CA"
                                                                         "MG"
[10] "STAR"
             "FRUC"
                      "GLUC"
                              "SUCR"
                                               "DM"
                                                        "VV"
                                                                "NOPS"
                                                                        "NOCR."
                                       "MALT"
[19] "NONC"
             "CRW"
                      "NCRW"
                              "RYTHA" "TRW"
                                               "BIOM"
                                                        "HT"
                                                                "CT"
                                                                        "FYTHA"
```

[28] "CYTHA"

Choose a set of traits to improve

Choose a set of traits to improve and run Pesek-Baker using defaults. See the correlations, standard deviations and response to selection.

```
> output <- PesekBaker(c('RYTHA', 'BC', 'DM', 'STAR', 'NOCR'), 'GENO', 'LOC', 'REP',
                      mvdata)
> output$Correlation.Matrix
        RYTHA
                    BC.
                            DM
                                  STAR
                                          NOCR.
RYTHA 1.00000 0.06816 -0.1677 -0.0496 0.8300
BC.
      0.06816 1.00000 -0.5271 -0.6703 0.1144
DM
     -0.16766 -0.52711 1.0000 0.8277 -0.1710
STAR -0.04960 -0.67032 0.8277 1.0000 -0.1026
NOCR.
    0.83003 0.11437 -0.1710 -0.1026 1.0000
> output$Standard.Deviations
[1]
     5.626 145.822 4.631 7.170 1.394
> output$Response.to.Selection
[1] 3.1484 81.5980 2.5916 4.0124 0.7798
> output$Std.Response.to.Selection
[1] 0.5596 0.5596 0.5596 0.5596 0.5596
```

Get your index

With an eye on the correlations and standard deviations, define your own desired genetic gains. Run the index again.

See the response to selection. Not satisfied? Modify your desired genetic gains and run the index again.

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Conclusions for the Pesek Baker index

- Keep an eye in the correlations.
- Keep both eyes in the standard deviations.
- Specify your desired genetic gains in relative terms,
- and even better, specify your desired genetic gains in relative terms and in standard deviation units.
- If a trait is important give it a weight a little bit higher than one standard deviation (somewhere between 1 and 2). If a trait is not so important give it a weight lower than 1 standard deviation (between 0 and 1).
- Do not ever try to improve a trait in more than 2 standard deviations (or maybe 1.5 could be a better and more conservative upper bound).
- Play with different values for the desired genetic gains and compare the response to selection that you get with each group of values.