

Bayesian linear mixed models using Stan: A tutorial for psychologists, linguists, and
cognitive scientists

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Abstract

With the arrival of the R packages `nlme` and `lme4`, linear mixed models (LMMs) have come to be widely used in experimentally-driven areas like psychology, linguistics, and cognitive science. This tutorial provides a practical introduction to fitting LMMs in a Bayesian framework using the probabilistic programming language Stan. We choose Stan (rather than WinBUGS or JAGS) because it provides an elegant and scalable framework for fitting models in most of the standard applications of LMMs. We ease the reader into fitting increasingly complex LMMs, first using a two-condition repeated measures self-paced reading study, followed by a more complex 2×2 repeated measures factorial design that can be generalized to much more complex designs.

Keywords: Bayesian data analysis, linear mixed models, Stan

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Introduction

Linear mixed models, or hierarchical/multilevel linear models, have become the main workhorse of experimental research in psychology, linguistics, and cognitive science, where repeated measures designs are the norm. Within the programming environment R (R Development Core Team, 2006), the `nlme` package (Pinheiro & Bates, 2000) and its successor, `lme4` (D. M. Bates, Mächler, Bolker, & Walker, 2015) have revolutionized the use of linear mixed models (LMMs) due to their simplicity and speed: one can fit fairly complicated models relatively quickly, often with a single line of code. A great advantage of LMMs over traditional approaches such as repeated measures ANOVA and paired t-tests is that there is no need to aggregate over subjects and items to compute two sets of F-scores (or several t-scores) separately; a single model can take all sources of variance into account simultaneously. Furthermore, comparisons between conditions can easily be implemented in a single model through appropriate contrast coding.

Other important developments related to LMMs have been unfolding in computational statistics. Specifically, probabilistic programming languages like WinBUGS (Lunn, Thomas, Best, & Spiegelhalter, 2000), JAGS (Plummer, 2012) and Stan (Stan Development Team, 2014), among others, have made it possible to fit Bayesian LMMs quite easily. However, one prerequisite for using these programming languages is that some background statistical knowledge is needed before one can define the model. This difficulty is well-known; for example, Spiegelhalter, Abrams, and Myles (2004, 4) write: “Bayesian statistics has a (largely deserved) reputation for being mathematically challenging and difficult to put into practice...”.

The purpose of this paper is to facilitate a first encounter with model specification in one of these programming languages, Stan. The tutorial is aimed primarily at psychologists, linguists, and cognitive scientists who have used `lme4` to fit models to their

data, but may have only a basic knowledge of the underlying LMM machinery. A diagnostic test is that they may not be able to answer some or all of these questions: what is a design matrix; what is contrast coding; what is a random effects variance-covariance matrix in a linear mixed model? Our tutorial is not intended for statisticians or psychology researchers who could, for example, write their own Markov Chain Monte Carlo samplers in R or C++ or the like; for them, the Stan manual is the optimal starting point. The present tutorial attempts to ease the beginner into their first steps towards fitting Bayesian linear mixed models. More detailed presentations about linear mixed models are available in several textbooks; references are provided at the end of this tutorial.

We have chosen Stan as the programming language of choice (over JAGS and WinBUGS) because it is possible to fit arbitrarily complex models with Stan. For example, it is possible (if time consuming) to fit a model with 14 fixed effects predictors and two crossed random effects by subject and item, each involving a 14×14 variance-covariance matrix (D. Bates, Kliegl, Vasishth, & Baayen, 2015); as far as we are aware, such models cannot, as far as we know, be fit in JAGS or WinBUGS.¹

In this tutorial, we take it as a given that the reader is interested in learning how to fit Bayesian linear mixed models. The tutorial is structured as follows. After a short introduction to Bayesian modeling, we begin by successively building up increasingly complex LMMs using the data-set reported by Gibson and Wu (2013), which has a simple two-condition design. At each step, we explain the structure of the model. The next section takes up inference for this two-condition design. Then we demonstrate how one can fit models using the matrix formulation of the design.

This paper was written using a literate programming tool, **knitr** (Xie, 2015); this integrates documentation for the accompanying code with the paper. The **knitr** file that generated this paper, as well as all the code and data used in this tutorial, can be downloaded from our website:

¹Whether it makes sense in general to fit such a complex model is a different issue; see Gelman et al. (2014), and D. Bates et al. (2015) for recent discussion.

<http://www.ling.uni-potsdam.de/~vasishth/statistics/BayesLMMs.html>

In addition, the source code for the paper, all R code, and data are available on github at:

<https://github.com/vasishth/BayesLMMTutorial>

We start with the two-condition repeated measures data-set (Gibson & Wu, 2013) as a concrete running example. This simple example serves as a starter kit for fitting commonly used LMMs in the Bayesian setting. We assume that the reader has the relevant software installed; specifically, `rstan` in R. For detailed instructions, see

<https://github.com/stan-dev/rstan/wiki/RStan-Getting-Started>

Bayesian modeling

Bayesian modeling has two major advantages over frequentist analysis with linear mixed models. First, information based on preceding results can be incorporated using different priors. Second, complex models with a large number of random variance components can be fit. In the following, we will provide a short introduction to Bayesian statistics.

Bayesian modeling is based on Bayes' Theorem. It can be seen as a way of understanding how the probability that a hypothesis is true is affected by new data. In mathematical notation,

$$P(H|D) = \frac{P(D|H)P(H)}{P(D)},$$

H is the hypothesis we are interested in, and D represents new data. Since D is fixed for a given data-set, the theorem can be rephrased as,

$$P(H|D) \propto P(D|H)P(H).$$

The *posterior* probability that the hypothesis is true given new data, $P(H|D)$, is proportional to the product of the *likelihood* of the new data given the hypothesis,

$P(D|H)$, and the *prior* probability of the hypothesis, $P(H)$.

For the purposes of this paper, the goal of a Bayesian analysis is simply to derive the posterior distribution of each parameter of interest, given some data and prior beliefs about the distributions of the parameters. The following example illustrates how the posterior belief depends on the properties of the likelihood and the prior. Before collecting data, a researcher has some hypothesis concerning the distribution of the response variable in an experiment. The researcher's belief can be expressed via prior distributions. A normal distribution with a mean value of $\mu_0 = 80$ represents the prior distribution. The left-hand panel of Figure 1 displays the prior distribution with a relatively large variance of $\sigma_0^2 = 1000$ (solid line). The large variance reflects the researcher's uncertainty concerning the true mean of the distribution. Alternatively, the researcher may be very certain concerning μ_0 , as represented by the prior distribution in the right-hand panel of Figure 1 (solid line). This prior has the same mean but a much lower variance of $\sigma_0^2 = 100$.

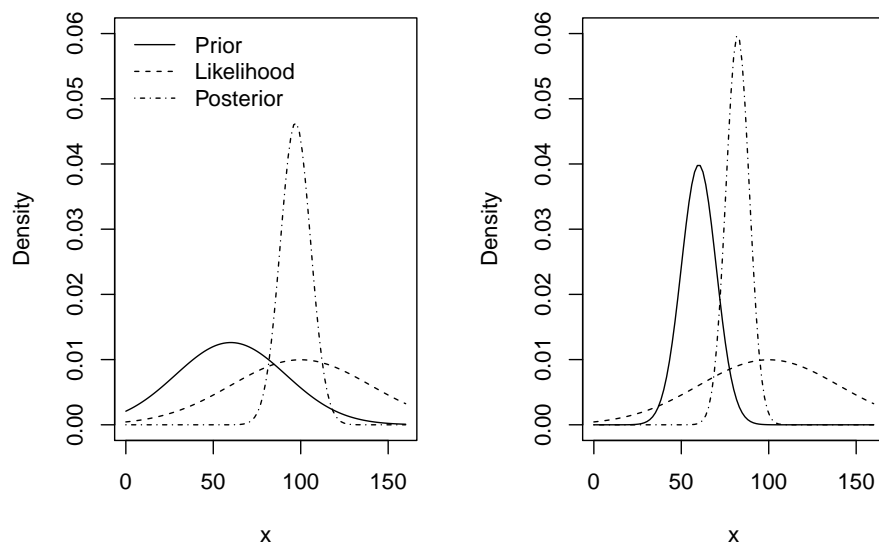


Figure 1. Prior, likelihood, and posterior normal distributions. The likelihood is based on $n = 20$ observations with sample mean $\mu = 100$ and standard deviation $\sigma = 40$. The prior has mean $\mu_0 = 60$ and variance $\sigma_0^2 = 1000$ (left-hand panel) or $\sigma_0^2 = 100$ (right-hand panel), respectively.

The researcher starts to collect data. In our example, there are $n = 20$ values with a sample mean of $\bar{x} = 100$ and a standard deviation of $\sigma = 40$. The corresponding likelihood distribution is displayed in Figure 1 (dashed line). The resulting posterior distribution (dash-dot line) is a result of prior and likelihood. Given the prior with the large variance (left-hand panel), the posterior is largely influenced by the data. If the prior has a lower variance (right-hand panel), its influence on the posterior is much stronger resulting in a stronger shift towards the prior's mean.

This toy example illustrates the central idea of Bayesian modeling. The prior reflects our knowledge of past results. In most cases, we will use so-called vague flat priors such that the posterior distribution is mainly affected by the data. The resulting posterior distribution allows for calculating credible intervals of true parameter values (see below).

For further explanation of the advantages this approach affords beyond the classical frequentist approach, the reader is directed to the rich literature relating to a comparison between Bayesian versus frequentist statistics (such as the provocatively titled paper by Lavine 1999; and the highly accessible textbook by Kruschke 2014).

The second advantage of Bayesian modeling concerns variance components (random effects). Fitting a large number of random effects in non-Bayesian settings requires a large amount of data. Often, the data-set is too small to fit reliable distributions of random effects (D. Bates et al., 2015). However, if a researcher is interested in differences between individual subjects or items (random intercepts and random slopes) or relationships between differences (correlations between variance components), Bayesian modeling can be used even if there is not enough data for inferential statistics. The resulting posterior distributions might have high variance but they still allow for calculating probabilities of true parameter values of variance components. Note that we do not intend to criticise classical LMMs but highlight possibilities of Bayesian modeling concerning random effects.

Example 1: A two-condition repeated measures design

This section motivates the LMM with the self-paced reading data-set of Gibson and Wu (2013). We introduce the data-set, state our modeling goals here, and proceed to build up increasingly complex LMMs.

The scientific question. Subject and object relative clauses have been widely used in reading studies to investigate sentence comprehension processes. A subject relative is a sentence like *The senator who interrogated the journalist resigned* where a noun (*senator*) is modified by a relative clause (*who interrogated the journalist*), and the modified noun is the grammatical subject of the relative clause. In an object relative, the noun modified by the relative clause is the grammatical object of the relative clause (e.g., *The senator who the journalist interrogated resigned*). In both cases, the noun that is modified (*senator*) is called the head noun.

A typical finding for English is that subject relatives are easier to process than object relatives (Just & Carpenter, 1992). Natural languages generally have relative clauses, and the subject relative advantage has until recently been considered to be true cross-linguistically. However, Chinese relative clauses apparently represent an interesting counter-example to this generalization; recent work by Hsiao and Gibson (2003) has suggested that in Chinese, *object* relatives are easier to process than subject relatives at a particular point in the sentence (the head noun of the relative clause). We now present an analysis of a subsequently published data-set (Gibson & Wu, 2013) that evaluates this claim.

The data. The dependent variable of the experiment of Gibson and Wu (2013) was the reading time `rt` of the head noun of the relative clause. This was recorded in two conditions (subject relative and object relative), with 37 subjects and 15 items, presented in a standard Latin square design. There were originally 16 items, but one item was removed, resulting in $37 \times 15 = 555$ data points. However, eight data points from one subject (id 27) were missing. As a consequence, we have a total of $555 - 8 = 547$ data

points. The first few lines from the data frame are shown in Table 1; “o” refers to object relative and “s” to subject relative.

row	subj	item	so	rt
1	1	13	o	1561
2	1	6	s	959
3	1	5	o	582
4	1	9	o	294
5	1	14	s	438
6	1	4	s	286
\vdots	\vdots	\vdots	\vdots	
547	9	11	o	350

Table 1

First six rows, and the last row, of the data-set of Gibson and Wu (2013), as they appear in the data frame.

We build up the Bayesian LMM from a fixed effects simple linear model to a varying intercepts model and finally to a varying intercepts, varying slopes model (the “maximal model” of Barr, Levy, Scheepers, and Tily 2013). The result is a probability model that expresses how the dependent variable, the reading time labeled `rt`, was generated in the experiment of Gibson and Wu (2013).

As mentioned above, the goal of Bayesian modeling is to derive the *posterior probability distribution* of the model parameters from a *prior probability distribution* and a *likelihood function*. Stan makes it easy to compute this posterior distribution of each parameter of interest. The posterior distribution reflects what we should believe, given the data, regarding the value of that parameter.

Fixed Effects Model (Simple Linear Model)

We begin by making the working assumption that the dependent variable of reading time `rt` on the head noun is approximately log-normally distributed (Rouder, 2005). This assumes that the logarithm of `rt` is approximately normally distributed. The logarithm of the reading times, $\log \text{rt}$, has some unknown grand mean β_0 . The mean of the log-normal distribution of `rt` is the sum of β_0 and an adjustment $\beta_1 \times \text{so}$ whose magnitude depends on

the categorical predictor `so`, which has the value -1 when `rt` is from the subject relative condition, and 1 when `rt` is from the object relative condition. One way to write the model in terms of the logarithm of the reading times is as follows:

$$\log \mathbf{rt}_i = \beta_0 + \beta_1 \mathbf{so}_i + \varepsilon_i \quad (1)$$

The index i represents the i -th row in the data-frame (in this case, $i \in \{1, \dots, 547\}$); the term ε_i represents the error in the i -th row. With the above ± 1 contrast coding, β_0 represents the grand mean of $\log \mathbf{rt}$, regardless of relative clause type. It can be estimated by simply taking the grand mean of $\log \mathbf{rt}$. The parameter β_1 is an adjustment to β_0 so that the mean of $\log \mathbf{rt}$ is $\beta_0 + 1\beta_1$ when $\log \mathbf{rt}$ is from the object relative condition, and $\beta_0 - 1\beta_1$ when $\log \mathbf{rt}$ is from the subject relative condition. Notice that $2 \times \beta_1$ will be the difference in the means between the object and subject relative clause conditions. Together, β_0 and β_1 make up the part of the model which characterizes the effect of the experimental manipulation, relative clause type (`so`), on the dependent variable `rt`. We call this a fixed effects model because we estimate the β parameters, which are unvarying from subject to subject and from item to item. In R, this would correspond to fitting a simple linear model using the `lm` function, with `so` as predictor and `log rt` as dependent variable.

The error ε_i is positive when $\log \mathbf{rt}_i$ is greater than the expected value $\mu_i = \beta_0 + \beta_1 \mathbf{so}_i$ and negative when $\log \mathbf{rt}_i$ is less than the expected value μ_i . Thus, the error is the amount by which the expected value differs from actually observed value. It is standardly assumed that the ε_i are independently and identically distributed as a normal distribution with mean zero and unknown standard deviation σ_e . Stan parameterizes the normal distribution by the mean and standard deviation, and we follow that convention here, by writing the distribution of ε as $\mathcal{N}(0, \sigma_e)$ (the standard notation in statistics is in terms of mean and variance). A consequence of the assumption that the errors are identically distributed is that the distribution of ε should, at least approximately, have the same shape as the normal distribution. Independence implies that there should be no

```

1 # read in data:
2 rDat <- read.table("gibsonwu2012data.txt", header = TRUE)
3 # subset critical region:
4 rDat <- subset(rDat, region == "headnoun")
5
6 # convert subjects and items to factors
7 rDat$subj <- factor(rDat$subj)
8 rDat$item <- factor(rDat$item)
9 # contrast coding of type (-1 vs. 1)
10 rDat$so <- ifelse(rDat$type == "subj-ext", -1, 1)
11
12 # create data as list for Stan, and fit model:
13 stanDat <- list(rt = rDat$rt, so = rDat$so, N = nrow(rDat))
14 library(rstan)
15 fixEfFit <- stan(file = "fixEf.stan", data = stanDat,
16                 iter = 2000, chains = 4)
17
18 # plot traceplot, excluding warm-up:
19 traceplot(fixEfFit, pars = c("beta", "sigma_e"),
20          inc_warmup = FALSE)
21
22 # examine quantiles of posterior distributions:
23 print(fixEfFit, pars = c("beta", "sigma_e"),
24       probs = c(0.025, 0.5, 0.975))
25
26 # examine quantiles of parameter of interest:
27 betal <- unlist(extract(fixEfFit, pars = "beta[2]"))
28 print(quantile(betal, probs = c(0.025, 0.5, 0.975)))

```

Listing 1: Code for the fixed effects Model 1.

correlation between the errors—this is not the case in the data, since we have multiple measurements from each subject, and from each item.

Setting up the data. We now fit the fixed effects Model 1. For the following discussion, please refer to the code in Listings 1 (R code) and 2 (Stan code). First, we read the Gibson and Wu (2013) data into a data frame `rDat` in R, and then subset the critical region (Listing 1, lines 2 and 4). Next, we create a data list `stanDat` for Stan, which contains the data (line 7). Stan requires the data to be of type list; this is different from the `lm` and `lmer` functions, which assume that the data are of type data-frame.

Defining the model. The next step is to write the Stan model in a text file with extension `.stan`. A Stan model consists of several *blocks*. A block is a set of statements

```

1 data {
2   int<lower=1> N;           //number of data points
3   real rt[N];              //reading time
4   real<lower=-1,upper=1> so[N]; //predictor
5 }
6 parameters {
7   vector[2] beta;          //intercept and slope
8   real<lower=0> sigma_e;    //error sd
9 }
10 model {
11   real mu;
12   for (i in 1:N){          // likelihood
13     mu <- beta[1] + beta[2] * so[i];
14     rt[i] ~ lognormal(mu, sigma_e);
15   }
16 }

```

Listing 2: Stan code for the fixed effects Model 1.

surrounded by brackets and preceded by the block name. We open up a file `fixEf.stan` in a text editor and write down the first block, the *data block*, which contains the declaration of the variables in the data object `stanDat` (Listing 2, lines 1–5). The strings `real` and `int` specify the data type for each variable. A `real` variable is a real number (\mathbb{R}), and an `int` variable is an integer (\mathbb{Z}). For instance, `N` is the integer number of data points. The variables `so` and `rt` are arrays of length `N` whose entries are `real`. We constrain a variable to take only a subset of the values allowed by its type (e.g. `int` or `real`) by specifying in brackets lower and upper bounds (e.g. `<lower=-1,upper=1>`). The variables in the data block, `N`, `rt`, and `so`, correspond to the values of the list `stanDat` in R.

Next, we turn to the *parameters block*, where the parameters are defined (Listing 2, lines 6–9). These are the parameters for which posterior distributions are of interest. The fixed effects Model 1 has three parameters: the fixed intercept β_0 , the fixed slope β_1 , and the standard deviation σ_e of the error. The fixed effects β_0 and β_1 are in the vector `beta` of length two; note that although we called our parameters β_0 and β_1 in Model 1, in Stan, these are contained in a vector with indices 1 and 2, so β_0 is in `beta[1]` and β_1 in `beta[2]`. The third parameter, the standard deviation σ_e of the error (`sigma_e`), is also defined here,

and is constrained to have lower bound 0 (Listing 2, line 8).

Finally, the *model block* specifies the prior distribution and the likelihood (Listing 2, lines 10–15). To understand the Stan syntax, compare the Stan code above to the specification of Model 1. The Stan code literally writes out this model. The block begins with a local variable declaration for `mu`, which is the mean of `rt` conditional on whether `so` is `-1` for the subject relative condition or `1` for the object relative condition.

The prior distributions on the parameters `beta` and `sigma_e` would ordinarily be declared in the model block. If we don't declare any prior, it is assumed that they have a uniform prior distribution. Note that the distribution of `sigma_e` is truncated at zero because `sigma_e` is constrained to be positive (see the declaration `real<lower=0> sigma_e;` in the parameters block). So, this means that the error has a uniform prior with lower bound 0.

In the model block, the for-loop assigns to `mu` the mean for the log-normal distribution of `rt[i]`, conditional on the value of the predictor `so[i]` for relative clause type. The statement `rt[i] ~ lognormal(mu, sigma_e)` means that the logarithm of each value in the vector `rt` is normally distributed with mean `mu` and standard deviation `sigma_e`. One could have equally well log-transformed the reading time and assumed a normal distribution instead of the lognormal.

Running the model. We save the file `fixEf.stan` which we just wrote and fit the model in R with the function `stan` from the package `rstan` (Listing 1, lines 9 and 10). This call to the function `stan` will compile a C++ program which produces samples from the joint posterior distribution of the fixed intercept β_0 , the fixed slope β_1 , and the standard deviation σ_e of the error. Here, the function generates four *chains* of samples, each of which contains 2000 samples of each parameter. Samples 1 to 1000 are part of the *warmup*, where the chains settle into the posterior distribution. We analyze samples 1001 to 2000. The result is saved to an object `fixEfFit` of class `stanFit`.

The warmup samples, also known as the *burn-in* period, are intended to allow the

MCMC sampling process to converge to the equilibrium distribution, the desired joint distribution over the variables. This is necessary since the initial values of the parameters might be very unlikely under the equilibrium distribution and hence bias the result. Once a chain has converged, the samples remain quite stable. Before the MCMC sampling process, the number of iterations necessary for convergence is unknown. Therefore, all warmup iterations are discarded.

The number of iterations necessary for convergence to the equilibrium distribution depends on the number of parameters. The probability to reach convergence increases with the number of iterations. Hence, we generally recommend using a large number of iterations although the process might converge after a smaller number of iterations. In the examples in the present paper, we use 1000 iterations for warmup and another 1000 iterations for analyzing the posterior distribution. For more complex models, more iterations might be necessary before the MCMC sampling process converges to the equilibrium distribution. Although there are ways to determine how long the simulation needs to be run and the number of warmup iterations given the type of posterior distribution (Raftery & Lewis, 1992), we illustrate below practical convergence diagnostics for the evaluation of convergence in the samples.

Besides the number of iterations, we specified sampling in four different chains. Each chain is independent from the others and starts with different random initial values. Running multiple chains has two advantages over a single chain. First, the independent chains are helpful for diagnostics. If all chains have converged to the same region of the parameter space, there is a high probability that they converged to the equilibrium distribution. Second, running multiple chains generally allows for parallel simulations on multiple cores.

Evaluating model convergence and summarizing results. The first step after running the above function should be to look at the *trace plot* of each chain after warmup, using the command shown in Listing 1, lines 13 and 14 (function `traceplot`). We

choose the parameters β_i and σ_e (`pars = c("beta", "sigma_e")`) and omit the warmup samples (`inc_warmup = FALSE`). A trace plot has the chains plotted against the sample ID. In Figure 2, we see three different chains plotted against sample number going from 1001 to 2000. If the trace plot looks like a “fat, hairy caterpillar” (Lunn, Jackson, Spiegelhalter, Best, & Thomas, 2012) which does not bend, this suggests that the chains have converged to the posterior distribution.

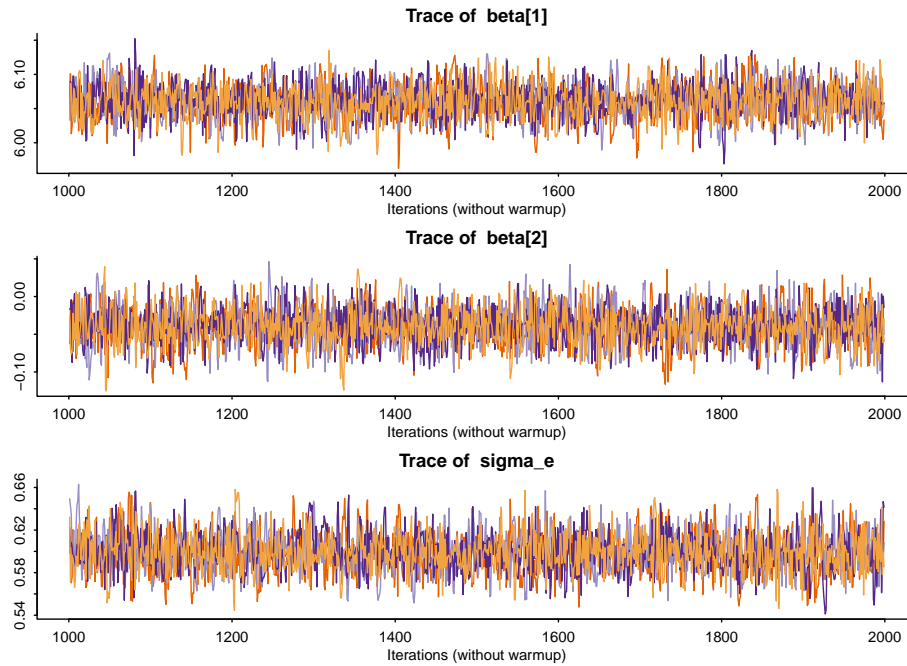


Figure 2. Trace plots of the fixed intercept β_0 (`beta[1]`), the fixed slope β_1 (`beta[2]`), and the standard deviation σ_e (`sigma_e`) of the error for the fixed effects Model 1.

Different colours denote different chains.

The second diagnostic which we use to assess whether the chains have converged to the posterior distribution is the statistic `Rhat`. Each parameter has the `Rhat` statistic associated with it (Gelman & Rubin, 1992); this is essentially the ratio of between-chain variance to within-chain variance (analogous to ANOVA). The `Rhat` statistic should be approximately 1 ± 0.1 if the chain has converged. This is shown in the rightmost column of the model summary, see Table 2. The information can be obtained with `print(fixEffFit)`, where `fixEffFit` is the `stan.model`.

Having satisfied ourselves that the chains have converged, we turn to examine this

posterior distribution. (If there is an indication that convergence has not happened, then, assuming that the model has no errors in it, increasing the number of samples usually resolves the issue.)

parameter	mean	2.5%	97.5%	\hat{R}
$\hat{\beta}_0$	6.06	0.03	6.11	1
$\hat{\beta}_1$	-0.04	-0.09	0.01	1
$\hat{\sigma}_e$	0.60	0.56	0.64	1

Table 2

Examining the credible intervals and the R -hat statistic in the Gibson and Wu data.

The result of fitting the fixed effects Model 1 is the *joint posterior probability distribution* of the parameters β_0 , β_1 , and σ_e . The distribution is joint because each of the 4000 (4 chains \cdot 1000 post-warmup iterations) posterior samples which the call to `stan` generates is a vector $\theta = (\beta_0, \beta_1, \sigma_e)^T$ of three model parameters. Thus, the object `fixEffFit` contains 4000 parameter vectors θ which occupy a three dimensional space. Already in three dimensions, the posterior distribution becomes difficult to view in one graph. Figure 3 displays the joint posterior probability distribution of the elements of θ by projecting it down onto planes. In each of the three planes (lower triangular scattergrams) we see how one parameter varies with respect to the other. In the diagonal histograms, we visualize the marginal probability distribution of each parameter separately from the other parameters.

Of immediate interest is the marginal distribution of the slope β_1 . Figure 3 suggests that the posterior probability density of β_1 is mainly spread over the interval $(-\infty, 0)$. One quantitative way to assess the posterior probability distribution is to examine its quantiles; see Table 2. Here, it is useful to define the concept of the *credible interval*. The $(1 - \alpha)\%$ credible interval contains $(1 - \alpha)\%$ of the posterior probability density. Unlike the $(1 - \alpha)\%$ confidence interval from the frequentist setting, the $(1 - \alpha)\%$ credible interval represents the range within which we are $(1 - \alpha)\%$ certain that the true value of the parameter lies, given the prior and the data (see Morey, Hoekstra, Rouder, Lee, and

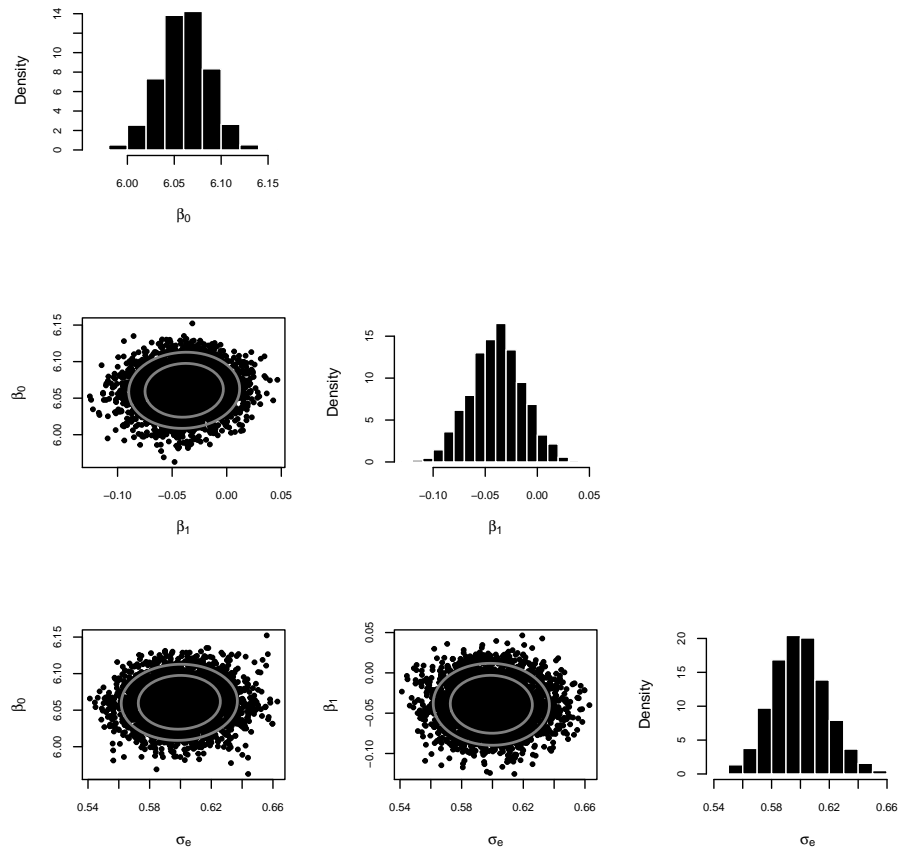


Figure 3. Bivariate joint posterior probability distribution of each element of θ with each other element (lower diagonal) and marginal posterior probability distribution of each element of θ separately (diagonal). All parameters are on the log scale, but note the difference in length scale between β_1 on the one hand and β_0 and σ_e on the other.

Wagenmakers 2015 for further discussion on CIs vs credible intervals). A common convention is to use the interval ranging from the 2.5th to 97.5th percentiles. We follow this convention and 95% credible intervals in Table 2. The last lines of 1 illustrate how these quantiles of the posterior distribution of β_1 (`beta[2]`) can be computed.

The samples of β_1 suggests that approximately 94% of the posterior probability density is below zero, suggesting that there is some evidence that object relatives are easier to process than subject relatives in Chinese, given the Gibson and Wu data. However, since the 95% credible interval includes 0, we may be reluctant to draw this conclusion. We will say more about the evaluation of research hypotheses further on.

Varying Intercepts Mixed Effects Model

The fixed effects Model 1 is inappropriate for the Gibson and Wu data because it does not take into account the fact that we have multiple measurements for each subject and item. As mentioned above, these multiple measurements lead to a violation of the independence of errors assumption. Moreover, the fixed effects coefficients β_0 and β_1 represent means over all subjects and items, ignoring the fact that some subjects will be faster and some slower than average; similarly, some items will be read faster than average, and some slower.

In linear mixed models, we take this by-subject and by-item variability into account by adding adjustment terms u_{0j} and w_{0k} , which adjust β_0 for subject j and item k . This partially decomposes ε_i into a sum of the terms u_{0j} and w_{0k} , which are adjustments to the intercept β_0 for the subject j and item k associated with \mathbf{rt}_i . If subject j is slower than the average of all the subjects, u_j would be some positive number, and if item k is read faster than the average reading time of all the items, then w_k would be some negative number. Each subject j has their own adjustment u_{0j} , and each item its own w_{0k} . These adjustments u_{0j} and w_{0k} are called *random intercepts* by Pinheiro and Bates (2000) and *varying intercepts* by Gelman and Hill (2007), and by adjusting β_0 by these we account for the variability between speakers, and between items.

It is standardly assumed that these adjustments are normally distributed around zero with unknown standard deviation: $u_0 \sim \mathcal{N}(0, \sigma_u)$ and $w_0 \sim \mathcal{N}(0, \sigma_w)$; the subject and item adjustments are also assumed to be mutually independent. We now have three sources of variance in this model: the standard deviation of the errors σ_e , the standard deviation of the by-subject random intercepts σ_u , and the standard deviation of the by-item varying intercepts σ_w . We will refer to these as variance components.

We now express the logarithm of reading time, which was produced by subjects $j \in \{1, \dots, 37\}$ reading items $k \in \{1, \dots, 15\}$, in conditions $i \in \{1, 2\}$ (1 refers to subject relatives, 2 to object relatives), as the following sum. Notice that we are now using a

slightly different way to describe the model, compared to the fixed effects model. We are using indices for subject, item, and condition to identify unique rows. Also, instead of writing $\beta_1\mathbf{so}$, we index β_1 by the condition i . This follows the notation used in the textbook on linear mixed models, written by the authors of `nlme` (Pinheiro & Bates, 2000), the precursor to `lme4`.

$$\log \mathbf{rt}_{ijk} = \beta_0 + \beta_{1i} + u_{0j} + w_{0k} + \varepsilon_{ijk} \quad (2)$$

Model 2 is an LMM, and more specifically a *varying intercepts model*. The coefficient β_{1i} is the one of primary interest; it will have some mean value $-\beta_1$ for subject relatives and β_1 for object relatives due to the contrast coding. So, if our posterior mean for β_1 is negative, this would suggest that object relatives are read faster than subject relatives.

We fit the varying intercepts Model 2 in Stan in much the same way as the fixed effects Model 1. For the following discussion, please consult Listing 3 for the R code used to run the model, and Listing 4 for the Stan code.

Setting up the data. The data which we prepare for passing on to the function `stan` now includes subject and item information (Listing 3, lines 2–8). The data block in the Stan code accordingly includes the number J , K of subjects and items, respectively; and the variable N records the number of rows in the data frame.

Defining the model. Model 2, shown in Listing 4, still has the fixed intercept β_0 , the fixed slope β_1 , and the standard deviation σ_e of the error, and we specify these in the same way as we did for the fixed effects Model 1. In addition, the varying intercepts Model 2 has by-subject varying intercepts u_{0j} for $j \in \{1, \dots, J\}$ and by-item varying intercepts w_{0k} for $k \in \{1, \dots, K\}$. The standard deviation of u_0 is σ_u and the standard deviation of w_0 is σ_w . We again constrain the standard deviations to be positive.

The model block places normal distribution priors on the varying intercepts u_0 and w_0 . We implicitly place uniform priors on `sigma_u`, `sigma_w`, and `sigma_e` by omitting

```

1 # format data for Stan:
2 stanDat <- list(subj = as.integer(rDat$subj),
3               item = as.integer(rDat$item),
4               rt = rDat$rt,
5               so = rDat$so,
6               N = nrow(rDat),
7               J = nlevels(rDat$subj),
8               K = nlevels(rDat$item))
9
10 # Sample from posterior distribution:
11 ranIntFit <- stan(file = "ranInt.stan", data = stanDat,
12                 iter = 2000, chains = 4)
13 # Summarize results:
14 print(ranIntFit, pars = c("beta", "sigma_e", "sigma_u", "sigma_w"),
15       probs = c(0.025, 0.5, 0.975))
16
17 betal <- unlist(extract(ranIntFit, pars = "beta[2]"))
18 print(quantile(betal, probs = c(0.025, 0.5, 0.975)))
19
20 # Posterior probability of betal being less than 0:
21 mean(betal < 0)

```

Listing 3: Code for running Model 2, the varying intercepts model.

them from the model block. As pointed out earlier for `sigma_e`, these prior distributions have lower bound zero because of the constraint `<lower=0>` in the variable declarations.

The statement about how each row in the data is generated is shown in Listing 4, lines 26–29; here, both the fixed effects and the varying intercepts for subjects and items determine the expected value `mu`. The vector `u` has varying intercepts for subjects. Likewise, the vector `w` has varying intercepts for items. The for-loop in lines 26–29 now adds `u[subj[i]] + w[item[i]]` to the mean `beta[1]` of the distribution of `rt[i]`. These are subject- and item-specific adjustments to the fixed-effects intercept `beta[1]`. The term `u[subj[i]]` identifies the id of the subject for row i in the data-frame; thus, if $i = 1$, then `subj[1] = 1`, and `item[1] = 13` (see Table 1).

Running the model. In R, we pass the list `stanDat` of data to `stan`, which compiles a C++ program to sample from the posterior distribution of Model 2. Stan samples from the posterior distribution of the model parameters, including the varying intercepts u_{0j} and w_{0k} for each subject $j \in \{1, \dots, J\}$ and item $k \in \{1, \dots, K\}$.

```

1  data {
2    int<lower=1> N;                //number of data points
3    real rt[N];                   //reading time
4    real<lower=-1, upper=1> so[N]; //predictor
5    int<lower=1> J;                //number of subjects
6    int<lower=1> K;                //number of items
7    int<lower=1, upper=J> subj[N]; //subject id
8    int<lower=1, upper=K> item[N]; //item id
9  }
10
11 parameters {
12   vector[2] beta;                //fixed intercept and slope
13   vector[J] u;                   //subject intercepts
14   vector[K] w;                   //item intercepts
15   real<lower=0> sigma_e;         //error sd
16   real<lower=0> sigma_u;         //subj sd
17   real<lower=0> sigma_w;         //item sd
18 }
19
20 model {
21   real mu;
22   //priors
23   u ~ normal(0, sigma_u);        //subj random effects
24   w ~ normal(0, sigma_w);        //item random effects
25   // likelihood
26   for (i in 1:N){
27     mu <- beta[1] + u[subj[i]] + w[item[i]] + beta[2] * so[i];
28     rt[i] ~ lognormal(mu, sigma_e);
29   }
30 }

```

Listing 4: Stan code for running Model 2, the varying intercepts model.

It may be helpful to rewrite the model in mathematical form following the Stan syntax (Gelman and Hill 2007 use a similar notation); the Stan statements are slightly different from the way that we expressed Model 2. Defining i as the row id in the data, i.e., $i \in \{1, \dots, 547\}$, we can write:

Likelihood :

$$\mu_i = \beta_0 + u_{[subj[i]]} + w_{[item[i]]} + \beta_1 \cdot \mathbf{so}_i$$

$$\mathbf{rt}_i \sim \text{LogNormal}(\mu_i, \sigma_e)$$

Priors : (3)

$$u \sim \text{Normal}(0, \sigma_u) \quad w \sim \text{Normal}(0, \sigma_w)$$

$$\sigma_e, \sigma_u, \sigma_w \sim \text{Uniform}(0, \infty)$$

$$\beta \sim \text{Uniform}(-\infty, \infty)$$

Here, notice that the i -th row in the statement for μ identifies the subject id (k) ranging from 1 to 37, and the item id (k) ranging from 1 to 15.

Summarizing the results. The posterior distributions of each of the parameters is summarized in Table 3. The \hat{R} values suggest that model has converged. Note also that compared to Model 1, the estimate of σ_e is smaller; this is because the other two variance components are now being estimated as well. Note that the 95% credible interval for the estimate $\hat{\beta}_1$ includes 0; thus, there is some evidence that object relatives are easier than subject relatives, but we cannot exclude the possibility that there is no difference in the reading times between the two relative clause types.

parameter	mean	2.5%	97.5%	\hat{R}
$\hat{\beta}_0$	6.06	5.92	6.20	1
$\hat{\beta}_1$	-0.04	-0.08	0.01	1
$\hat{\sigma}_e$	0.52	0.49	0.55	1
$\hat{\sigma}_u$	0.25	0.19	0.34	1
$\hat{\sigma}_w$	0.20	0.12	0.32	1

Table 3

The quantiles and the \hat{R} statistic in the Gibson and Wu data, the varying intercepts model.

Varying Intercepts, Varying Slopes Mixed Effects Model

Consider now that subjects who are faster than average (i.e., who have a negative varying intercept) may exhibit greater slowdowns when they read subject relatives compared to object relatives. Similarly, it is in principle possible that items which are read faster (i.e., which have a large negative varying intercept) may show a greater slowdown in subject relatives than object relatives. The opposite situation could also hold: faster subjects may show smaller SR-OR effects, or items read faster may show smaller SR-OR effects. Although such individual-level variability was not of interest in the original paper by Gibson and Wu, it could be of theoretical interest (see, for example, Kliegl, Wei, Dambacher, Yan, and Zhou 2010). Furthermore, as Barr et al. (2013) point out, it is in principle desirable to include a fixed effect factor in the random effects as a varying slope if the experiment design is such that subjects see both levels of the factor (cf. D. Bates et al. 2015).

In order to express this structure in the LMM, we must make two changes in the varying intercepts Model 2.

Adding varying slopes. The first change is to let the size of the effect for the predictor `so` vary by subject and by item. The goal here is to express that some subjects exhibit greater slowdowns in the object relative condition than others. We let effect size vary by subject and by item by including in the model by-subject and by-item varying slopes which adjust the fixed slope β_1 in the same way that the by-subject and by-item varying intercepts adjust the fixed intercept β_0 . This adjustment of the slope by subject and by item is expressed by adjusting β_1 by adding two terms u_{1j} and w_{1k} . These are *random* or *varying slopes*, and by adding them we account for how the effect of relative clause type varies by subject j and by item k . We now express the logarithm of reading time, which was produced by subject j reading item k , as the following sum. The subscript i indexes the conditions.

$$\log \mathbf{rt}_{ijk} = \underbrace{\beta_0 + u_{0j} + w_{0k}}_{\text{varying intercepts}} + \underbrace{\beta_1 + u_{1ij} + w_{1ik}}_{\text{varying slopes}} + \varepsilon_{ijk} \quad (4)$$

Defining a variance-covariance matrix for the random effects. The second change which we make to Model 2 is to define a covariance relationship between by-subject varying intercepts and slopes, and between by-items intercepts and slopes. This amounts to adding an assumption that the by-subject slopes u_1 could in principle have some correlation with the by-subject intercepts u_0 ; and by-item slopes w_1 with by-item intercept w_0 . We explain this in detail below.

Let us assume that the adjustments u_0 and u_1 are normally distributed with mean zero and some variances σ_{u0}^2 and σ_{u1}^2 , respectively; also assume that u_0 and u_1 have correlation ρ_u . It is standard to express this situation by defining a variance-covariance matrix Σ_u (sometime this is simply called a variance matrix). This matrix has the variances of u_0 and u_1 respectively along the diagonals, and the covariances on the off-diagonals. (The covariance $\text{Cov}(X, Y)$ between two variables X and Y is defined as the product of their correlation ρ and their standard deviations σ_X and σ_Y : $\text{Cov}(X, Y) = \rho\sigma_X\sigma_Y$.)

$$\Sigma_u = \begin{pmatrix} \sigma_{u0}^2 & \rho_u\sigma_{u0}\sigma_{u1} \\ \rho_u\sigma_{u0}\sigma_{u1} & \sigma_{u1}^2 \end{pmatrix} \quad (5)$$

Similarly, we can define a variance-covariance matrix Σ_w for items, using the standard deviations σ_{w0} , σ_{w1} , and the correlation ρ_w .

$$\Sigma_w = \begin{pmatrix} \sigma_{w0}^2 & \rho_w\sigma_{w0}\sigma_{w1} \\ \rho_w\sigma_{w0}\sigma_{w1} & \sigma_{w1}^2 \end{pmatrix} \quad (6)$$

The standard way to express this relationship between the subject intercepts u_0 and slopes u_1 , and the item intercepts w_0 and slopes w_1 , is to define a bivariate normal distribution as follows:

$$\begin{pmatrix} u_0 \\ u_1 \end{pmatrix} \sim \mathcal{N} \left(\begin{pmatrix} 0 \\ 0 \end{pmatrix}, \Sigma_u \right), \quad \begin{pmatrix} w_0 \\ w_1 \end{pmatrix} \sim \mathcal{N} \left(\begin{pmatrix} 0 \\ 0 \end{pmatrix}, \Sigma_w \right) \quad (7)$$

An important point to notice here is that any $n \times n$ variance-covariance matrix has associated with it an $n \times n$ correlation matrix. In the subject variance-covariance matrix Σ_u , the correlation matrix is

$$\begin{pmatrix} 1 & \rho_{01} \\ \rho_{01} & 1 \end{pmatrix} \quad (8)$$

In a correlation matrix, the diagonal elements will always be 1, because a variable always has a correlation of 1 with itself. The off-diagonals will have the correlations between the variables. Note also that, given the variances σ_{u0}^2 and σ_{u1}^2 , we can always recover the variance-covariance matrix, if we know the correlation matrix. This is because of the above-mentioned definition of covariance.

A correlation matrix can be decomposed into a square root of the matrix, using the Cholesky decomposition. Thus, given a correlation matrix C , we can obtain its square root L ; an obvious consequence is that we can square L to get the correlation matrix C back. This is easy to illustrate with a simple example. Suppose we have a correlation matrix:

$$C = \begin{pmatrix} 1 & -0.5 \\ -0.5 & 1 \end{pmatrix} \quad (9)$$

We can use the Cholesky decomposition function in R, `chol`, to derive the lower triangular square root L of this matrix. This gives us:

$$L = \begin{pmatrix} 1 & 0 \\ -0.5 & 0.8660254 \end{pmatrix} \quad (10)$$

We can confirm that this is a square root by multiplying L with itself to get the correlation matrix back (squaring a matrix is done by multiplying the matrix by its

transpose):

$$LL^T = \begin{pmatrix} 1 & 0 \\ -0.5 & 0.8660254 \end{pmatrix} \begin{pmatrix} 1 & -0.5 \\ 0 & 0.8660254 \end{pmatrix} = \begin{pmatrix} 1 & -0.5 \\ -0.5 & 1 \end{pmatrix} \quad (11)$$

The reason that we bring up the Cholesky decomposition here is that we will use it to generate the by-subject and by-item adjustments to the intercept and slope fixed-effects parameters.

Generating correlated random variables using the Cholesky decomposition. The by-subject and by-item adjustments are generated using the following standard procedure for generating correlated random variables $\mathbf{x} = (x_1, x_2)$:

1. Given a vector of standard deviances (e.g., σ_{u0}, σ_{u1}), create a diagonal matrix:

$$\tau = \begin{pmatrix} \sigma_{u0} & 0 \\ 0 & \sigma_{u1} \end{pmatrix} \quad (12)$$

2. Premultiply the diagonalized matrix τ with the Cholesky decomposition L of the correlation matrix C to get a matrix Λ .
3. Generate values from a random variable $\mathbf{z} = (z_1, z_2)^T$, where z_1 and z_2 each have independent $\mathcal{N}(0, 1)$ distributions (left panel of Figure 4).
4. Multiply Λ with \mathbf{z} ; this generates the correlated random variables \mathbf{x} (right panel of Figure 4).

This digression into Cholesky decomposition and the generation of correlated random variables is important to understand for building the Stan model. We will define a vague prior distribution on L — the square root of the correlation matrix — and a vague prior on the standard deviations. This allows us to generate the by-subject and by-item adjustments to the fixed effects intercepts and slopes.

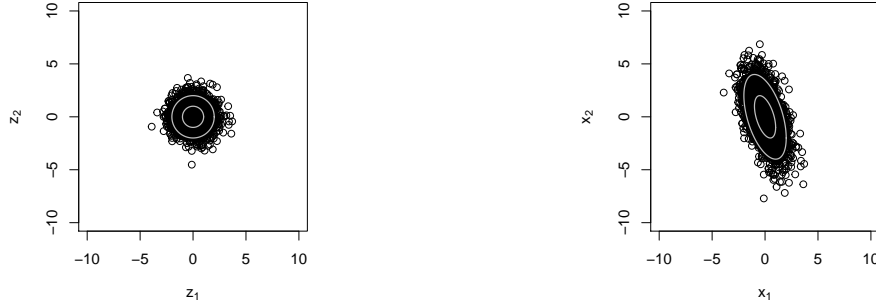


Figure 4. Uncorrelated random variables $\mathbf{z} = (z_1, z_2)^\top$ (left) and correlated random variables $\mathbf{x} = (x_1, x_2)^\top$ (right).

Defining the model. With this background, implementing the varying intercepts, varying slope Model 4 is straightforward; see Listing 5 for the R code and Listing 6 for the Stan code. The R list `stanDat` is identical to the one of Model 3, and therefore we will focus on the Stan code. The data block is the same as before. The parameters block contains several new parameters. This time, we have vectors `sigma_u` and `sigma_w` which are $(\sigma_{u0}, \sigma_{u1})^\top$ and $(\sigma_{w0}, \sigma_{w1})^\top$, instead of scalar values as in Model 2. The variables `L_u`, `L_w`, `z_u`, and `z_w`, which have been declared in the parameters block, play a role in the *transformed parameters block*, a block which we did not use in the earlier models. The transformed parameters block generates the by-subject and by-item varying intercepts and slopes using the parameters `sigma_u`, `sigma_w`, `L_u`, `L_w`, `z_u`, and `z_w`. The J pairs of by-subject varying intercepts and slopes are in the rows of the $J \times 2$ matrix `u`, and the K pairs of by-item varying intercepts and slopes are in the rows of the $K \times 2$ matrix `w`.

These varying intercepts and slopes are obtained through the statements `diag_pre_multiply(sigma_u, L_u) * z_u` and `diag_pre_multiply(sigma_w, L_w) * z_w`. This statement generates varying intercepts and slopes from the joint probability distribution of equation 7. The parameters `L_u`, `L_w` are the Cholesky decompositions of the subject and item correlation matrices, respectively, and `z_u`, and `z_w` are $\mathcal{N}(0, 1)$ random variables.

It is helpful to walk through steps 1 to 4 involved in generating the varying intercepts and slopes using the procedure described above for generating correlated random variables.

```

1 ranIntSlpFit <- stan(file = "ranIntSlp.stan", data = stanDat,
2                     iter = 2000, chains = 4)
3
4 # posterior probability of beta 1 being less
5 # than 0:
6 betal <- unlist(extract(ranIntSlpFit, pars = "beta[2]"))
7 print(quantile(betal, probs = c(0.025, 0.5, 0.975)))
8 mean(betal < 0)

```

Listing 5: Code for running Model 3, the varying intercepts, varying slopes model.

The statement `diag_pre_multiply(sigma_u, L_u) * z_u` computes the transpose matrix product (steps 1 and 2). The right multiplication of this product by `z_u`, a matrix of normally distributed random variables (step 3), yields the varying intercepts and slopes (step 4).

$$\begin{aligned}
 \begin{pmatrix} u_{01} & u_{11} \\ u_{02} & u_{12} \\ \vdots & \vdots \\ u_{0J} & u_{1J} \end{pmatrix} &= \left(\text{diag}(\sigma_{u0}, \sigma_{u1}) \mathbf{L}_u \mathbf{z}_u \right)^\top \\
 &= \left(\begin{pmatrix} \sigma_{u0} & 0 \\ 0 & \sigma_{u1} \end{pmatrix} \begin{pmatrix} \ell_{11} & 0 \\ \ell_{21} & \ell_{22} \end{pmatrix} \begin{pmatrix} z_{11} & z_{12} & \dots & z_{1J} \\ z_{21} & z_{22} & \dots & z_{2J} \end{pmatrix} \right)^\top
 \end{aligned} \tag{13}$$

Turning to the model block, here, we place priors on the parameters declared in the parameters block, and define how these parameters generate `log rt` (Listing 6, lines 30–42). The definition of the prior `L_u ~ lkj_corr_cholesky(2.0)` implicitly places a so-called `lkj` prior with shape parameter $\eta = 2.0$ on the correlation matrices

$$\begin{pmatrix} 1 & \rho_u \\ \rho_u & 1 \end{pmatrix} \text{ and } \begin{pmatrix} 1 & \rho_w \\ \rho_w & 1 \end{pmatrix}, \tag{14}$$

where ρ_u is the correlation between the by-subject varying intercept σ_{u0} and slope σ_{u1} (cf.

```

1 data {
2   int<lower=1> N;           //number of data points
3   real rt[N];              //reading time
4   real<lower=-1, upper=1> so[N]; //predictor
5   int<lower=1> J;           //number of subjects
6   int<lower=1> K;           //number of items
7   int<lower=1, upper=J> subj[N]; //subject id
8   int<lower=1, upper=K> item[N]; //item id
9 }
10
11 parameters {
12   vector[2] beta;           //intercept and slope
13   real<lower=0> sigma_e;     //error sd
14   vector<lower=0>[2] sigma_u; //subj sd
15   vector<lower=0>[2] sigma_w; //item sd
16   cholesky_factor_corr[2] L_u;
17   cholesky_factor_corr[2] L_w;
18   matrix[2,J] z_u;
19   matrix[2,K] z_w;
20 }
21
22 transformed parameters{
23   matrix[2,J] u;
24   matrix[2,K] w;
25
26   u <- diag_pre_multiply(sigma_u, L_u) * z_u; //subj random effects
27   w <- diag_pre_multiply(sigma_w, L_w) * z_w; //item random effects
28 }
29
30 model {
31   real mu;
32   //priors
33   L_u ~ lkj_corr_cholesky(2.0);
34   L_w ~ lkj_corr_cholesky(2.0);
35   to_vector(z_u) ~ normal(0,1);
36   to_vector(z_w) ~ normal(0,1);
37   //likelihood
38   for (i in 1:N){
39     mu <- beta[1] + u[1,subj[i]] + w[1,item[i]]
40       + (beta[2] + u[2,subj[i]] + w[2,item[i]]) * so[i];
41     rt[i] ~ lognormal(mu, sigma_e);
42   }
43 }

```

Listing 6: The Stan code for Model 3, the varying intercepts, varying slopes model.

the covariance matrix of Equation 5) and ρ_w is the correlation between the by-item varying intercept σ_{w0} and slope σ_{w1} . The lkj distribution with shape parameter $\eta = 1.0$ is a uniform prior over all 2×2 correlation matrices; it scales up to larger correlation matrices. The parameter η has an effect on the shape of the lkj distribution. Our choice of $\eta = 2.0$ implies that the correlations in the off-diagonals are near zero, reflecting the fact that we have no prior information about the correlation between intercepts and slopes.

The statement `to_vector(z_u) ~ normal(0,1)` places a normal distribution with mean zero and standard deviation one on `z_u`. The same goes for `z_w`. The for-loop assigns to `mu` the mean of the log-normal distribution from which we draw `rt[i]`, conditional on the value of the predictor `so[i]` for relative clause type and the subject and item identity.

We can now fit the varying intercepts, varying slopes Model 3; see Listing 5 for the code. We see in the model summary in Table 4 that the model has converged, and that the credible intervals of the parameter of interest, β_1 , still include 0. In fact, the posterior probability of the parameter being less than 0 is now 90% (this information can be extracted as shown in Listing 5, lines 6–8).

parameter	mean	2.5%	97.5%	\hat{R}
$\hat{\beta}_0$	6.06	5.92	6.21	1
$\hat{\beta}_1$	−0.04	−0.09	0.02	1
$\hat{\sigma}_e$	0.51	0.48	0.55	1
$\hat{\sigma}_{u0}$	0.25	0.19	0.34	1
$\hat{\sigma}_{u1}$	0.07	0.01	0.14	1
$\hat{\sigma}_{w0}$	0.20	0.13	0.32	1
$\hat{\sigma}_{w1}$	0.04	0.0	0.10	1

Table 4

The quantiles and the \hat{R} statistic in the Gibson and Wu data, the varying intercepts, varying slopes model.

Figure 5 plots the varying slope’s posterior distribution against the varying intercept’s posterior distribution for each subject. The correlation between u_0 and u_1 is negative, as captured by the marginal posterior distributions of the correlation ρ_u between

u_0 and u_1 . Thus, Figure 5 suggests that the slower a subject's reading time is on average, the slower they read object relatives. In contrast, Figure 5 shows no clear pattern for the by-item varying intercepts and slopes. The broader distribution of the correlation parameter for items compared to slopes illustrates the greater uncertainty concerning the true value of the parameter. We briefly discuss inference next.

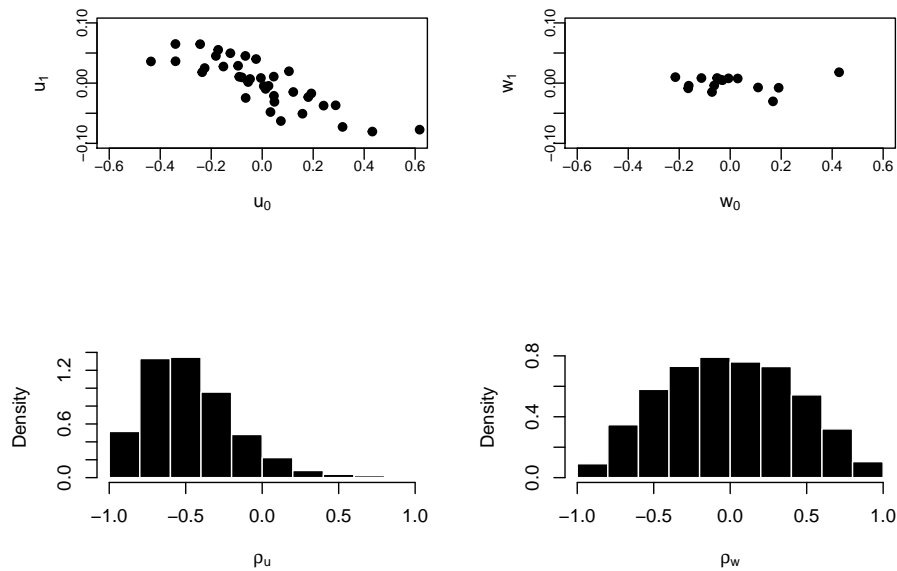


Figure 5. The top row shows the relationship between the varying slopes (y-axis) and intercepts (x-axis) for each subject (left panel) and item (right panel). The bottom row shows the posterior distribution of the parameter of correlation between the varying slopes and intercepts for each subject (left panel) and item (right panel).

Random effects in a non-Bayesian LMM. We fit the same model also as a classical non-Bayesian LMM with the `lmer` function from the `lme4` package. This allows for comparing the results to the Stan results. Here, we focus on random effects. As illustrated in Figure 6, the estimates of the random-effect standard deviations of the classical LMM are in agreement with the modes of the posterior distributions. However, the classical LMM is overparameterized due to an insufficient number of data points. Hence, correlations between random effects could not be estimated, as indicated by perfect correlations of -1 and 1 . In contrast, Stan can still estimate posterior distributions for

these parameters (Figure 5).

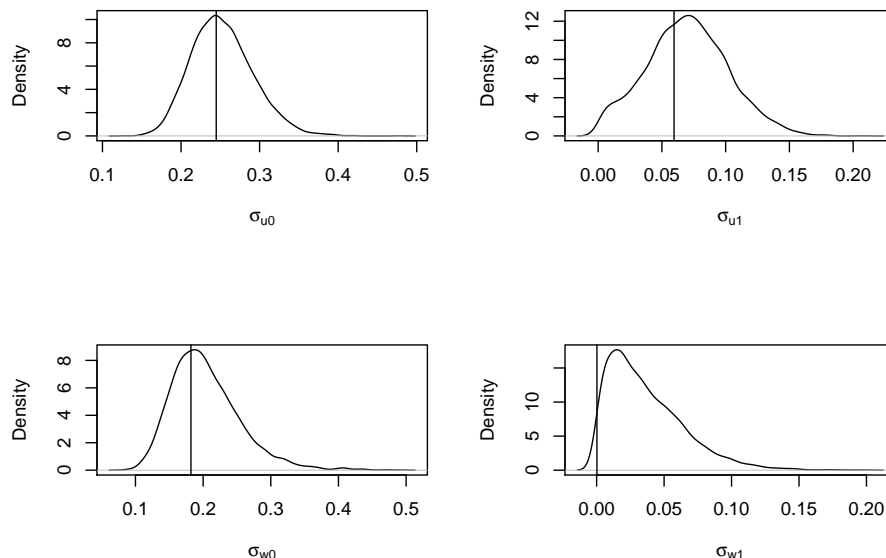


Figure 6. The curves show the density of the posterior distributions of the random-effect standard deviations. The vertical bars indicate the corresponding `lmer` estimates. The top row shows the random effects for subjects, the bottom row shows the random effects for items. Left-hand panels correspond to random intercepts, right-hand panels correspond to random slopes.

Inference

Having fit a varying intercepts, varying slopes Model 3, we now explain one way to carry out statistical inference, using credible intervals. We have used this approach to draw inferences from data in previously published work (e.g., Frank, Trompenaars, and Vasishth 2015, Hofmeister and Vasishth 2014). There are of course other approaches possible for carrying out inference. Bayes Factors are an example; see Lee and Wagenmakers (2013) and Rouder and Morey (2012). Another is to define a Region of Practical Equivalence (Kruschke, 2014). The reader can choose the approach they find the most appealing.

The result of fitting the varying intercepts, varying slopes Model 4 is the posterior distribution of the model parameters. As mentioned above in connection with Models 1-3,

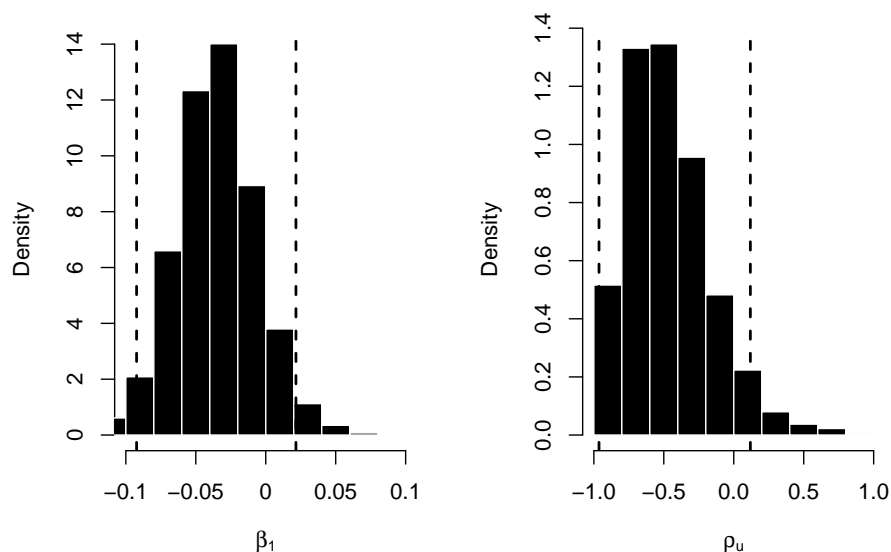


Figure 7. Upper and lower bounds on the credible intervals (dashed lines) plotted over the marginal posterior distribution of the fixed slope β_1 (left) and of the correlation ρ_u between the by-subject varying intercepts and varying slopes (right).

direct inference from the posterior distributions is possible. For instance, we can find the posterior probability with which the fixed intercept β_1 or the correlation ρ_u between by-subject varying intercepts and slopes take on any given value by consulting the marginal posterior distributions whose histograms are shown in Figure 7. The information conveyed by such graphs can be sharpened by using the 95% credible interval, mentioned earlier. Approximately 95% of the posterior density of β_1 lies between the 2.5 percentile -0.09 and the 97.5 percentile 0.02 . This leads us to conclude that the slope β_1 for relative clause type `so` is less than zero with probability 90% (see line 8 in Listing 5). Since 0 is included in the credible interval, it is difficult to draw the inference that object relative clauses are read faster than subject relative clauses. However, one could perhaps still make a weak claim to that effect, especially if a lot of evidence has accumulated in other experiments that supports such a conclusion (see Vasishth, Chen, Li, and Guo 2013 for a more detailed discussion).

What about the correlations between varying intercepts and varying slopes for

subject and for item? What can we infer from the analysis about these relationships? The 95% credible interval for ρ_u is $(-1, 0.1)$. Our belief that ρ_u is less than zero is rather uncertain, although we can conclude that ρ_u is less than zero with probability 90%. There is only weak evidence that subjects who read faster than average exhibit greater slowdowns at the head noun of object relative clauses than subjects who read slower than average. For the by-item varying intercepts and slopes, it is pretty clear that we do not have enough data (15 items) to draw any conclusions. For these data, it probably makes sense to fit a simpler model (D. Bates et al., 2015), with only varying intercepts and slopes for subject, and only varying intercepts for items; although there is no harm done in a Bayesian setting if we fit a model with a full variance-covariance matrix for both subjects and items.

In sum, regarding our main research question, our conclusion here is that we cannot say that object relatives are harder to process than subject relatives, because the credible interval for β_1 includes 0. However, one could argue that there is *some* weak evidence in favor of the hypothesis, since the posterior probability of the parameter being negative is approximately 90%.

Matrix formulation of the linear mixed model

We fit three models with increasing complexity to the data-set. In all specifications, there was an explicit vector `so` for the predictor variable in Stan. However, if we want to fit more complex models with a multitude of categorical and continuous predictors and interactions, this approach requires increasingly complex specifications in Stan code. Alternatively, we can use the matrix formulation of the linear mixed model that allows for using the same code for models of different complexity. In the following, we will apply this approach for an alternative version of Model 3 including random intercepts and slopes for subjects and items (Equation 4).

Again, we fit a varying intercepts, varying slopes model. The grand mean β_0 of `log rt` is adjusted by subject and by item through the varying intercepts u_0 and w_0 , which are

unique values for each subject and item respectively. Likewise, the fixed effect β_1 , which is associated with the predictor `so`, is adjusted by the by-subject varying slope u_1 and by-item varying slope w_1 .

It is more convenient to represent this model in matrix form. We build up the model specification by first noting that, for each subject, the by-subject varying intercept u_0 and slope u_1 have a multivariate normal prior distribution with mean zero and covariance matrix Σ_u . Similarly, for each item, the by-item varying intercept w_0 and slope w_1 have a multivariate normal prior distribution with mean zero and covariance matrix Σ_w . The error ε is assumed to have a normal distribution with mean zero and standard deviation σ_e .

We proceed to implement the model in Stan. Instead of passing the predictor `so` to `stan` as vector, as we did earlier, we make `so` into a design matrix `X` using the function `model.matrix` available in R (see Listing 7, line 2).² The command `model.matrix(~ 1 + so, rDat)` creates a model matrix with two fixed effects, the intercept (1) and a factor (`so`), based on the data frame `rDat`. The first column of the design matrix `X` consists of all ones; this column represents the intercept. The second column is the predictor `so` and consists of values in $\{-1, 1\}$. The model matrix thus consists of a two-level factorial design, with blocks of this design repeated for each subject. For the full data-set, we could write it very compactly in matrix form as follows:

$$\log \mathbf{rt} = \mathbf{X}\beta + \mathbf{Z}_u\mathbf{u} + \mathbf{Z}_w\mathbf{w} + \varepsilon \quad (15)$$

Here, \mathbf{X} is the $N \times P$ model matrix (with $N = 547$, since we have 547 data points; and $P = 2$ since we have the intercept plus another fixed effect), β is a vector of length P including fixed effects parameters, \mathbf{Z}_u and \mathbf{Z}_w are the subject and item model matrices ($N \times P$), and \mathbf{u} and \mathbf{w} are the by-subject and by-item adjustments to the fixed effects estimates; these are identical to the design matrix \mathbf{X} in the model with varying intercepts

²Here, we would like to acknowledge the contribution of Douglas Bates in specifying the model in this general matrix form.

```

1 # Make design matrix
2 X <- unname(model.matrix(~ 1 + so, rDat))
3 attr(X, "assign") <- NULL
4 # Make Stan data
5 stanDat <- list(N = nrow(X),
6               P = ncol(X),
7               n_u = ncol(X),
8               n_w = ncol(X),
9               X = X,
10              Z_u = X,
11              Z_w = X,
12              J = nlevels(rDat$subj),
13              K = nlevels(rDat$item),
14              rt = rDat$rt,
15              subj = as.integer(rDat$subj),
16              item = as.integer(rDat$item))
17 # Fit the model
18 matrixFit <- stan(file = "matrixModel.stan", data = stanDat,
19                  iter = 2000, chains = 4)

```

Listing 7: Matrix formulation code for running the varying intercepts, varying slopes model.

and varying slopes included. For more examples of similar model specifications in Stan, see the R package `RePsychLing` on github (<https://github.com/dmbates/RePsychLing>).

Note that we remove the column names and the attributes of the model matrix `X` in order to use it for Stan; please refer to Listing 7. Having defined the model, we proceed to assemble the list `stanDat` of data, relying on the above matrix formulation. The number `N` of observations, the number `J` of subjects and `K` of items, the reading times `rt`, and the subject and item indicator variables `subj` and `item` are familiar from the previous models presented. The integer `P` is the number of fixed effects (two including the intercept). Model 4 includes a varying intercept u_0 and a varying slope u_1 for each subject, and so the number `n_u` of by-subject random effects equals `P`. Likewise, Model 4 includes a varying intercept w_0 and a varying slope w_1 for each item, and so the number `n_w` of by-item random effects also equals `P`.

We also have to adapt the Stan code to the model formulation (see Listing 8). The data block contains the corresponding variables. Using the command

```

1  data {
2    int<lower=0> N;                //no trials
3    int<lower=1> P;                //no fixefs
4    int<lower=0> J;                //no subjects
5    int<lower=1> n_u;              //no subj ranefs
6    int<lower=0> K;                //no items
7    int<lower=1> n_w;              //no item ranefs
8    int<lower=1,upper=J> subj[N]; //subject indicator
9    int<lower=1,upper=K> item[N]; //item indicator
10   row_vector[P] X[N];           //fixef design matrix
11   row_vector[n_u] Z_u[N];       //subj ranef design matrix
12   row_vector[n_w] Z_w[N];       //item ranef design matrix
13   vector[N] rt;                 //reading time
14 }
15 parameters {
16   vector[P] beta;                //fixef coefs
17   cholesky_factor_corr[n_u] L_u; //cholesky factor of subj ranef corr matrix
18   cholesky_factor_corr[n_w] L_w; //cholesky factor of item ranef corr matrix
19   vector<lower=0>[n_u] sigma_u; //subj ranef std
20   vector<lower=0>[n_w] sigma_w; //item ranef std
21   real<lower=0> sigma_e;         //residual std
22   vector[n_u] z_u[J];           //subj ranef
23   vector[n_w] z_w[K];           //item ranef
24 }
25 transformed parameters {
26   vector[n_u] u[J];             //subj ranefs
27   vector[n_w] w[K];             //item ranefs
28   {
29     matrix[n_u,n_u] Sigma_u;    //subj ranef cov matrix
30     matrix[n_w,n_w] Sigma_w;    //item ranef cov matrix
31     Sigma_u <- diag_pre_multiply(sigma_u, L_u);
32     Sigma_w <- diag_pre_multiply(sigma_w, L_w);
33     for(j in 1:J)
34       u[j] <- Sigma_u * z_u[j];
35     for(k in 1:K)
36       w[k] <- Sigma_w * z_w[k];
37   }
38 }
39 model {
40   //priors
41   L_u ~ lkj_corr_cholesky(2.0);
42   L_w ~ lkj_corr_cholesky(2.0);
43   for (j in 1:J)
44     z_u[j] ~ normal(0,1);
45   for (k in 1:K)
46     z_w[k] ~ normal(0,1);
47   //likelihood
48   for (i in 1:N)
49     rt[i] ~ lognormal(X[i] * beta +
50                       Z_u[i] * u[subj[i]] +
51                       Z_w[i] * w[item[i]],
52                       sigma_e);
53 }

```

Listing 8: Stan code for the matrix formulation of the varying intercepts, varying slopes model.

`row_vector[P] X[N]`, we declare the fixed effects design matrix \mathbf{X} as an array of N row vectors of length P whose components are the predictors associated with the N reading times. Likewise for the subject and item random effects design matrices \mathbf{Z}_u and \mathbf{Z}_w , which correspond to \mathbf{Z}_u and \mathbf{Z}_w respectively in Equation 15. The vector `beta` contains the fixed effects β_0 and β_1 . The matrices `L_u`, `L_w` and the arrays `z_u`, `z_w` of vectors (not to be confused with the design matrices \mathbf{Z}_u and \mathbf{Z}_w) will generate the varying intercepts and slopes u_0, u_1 and w_0, w_1 , using the procedure described for Model 3. For example, the command `vector[n_u] u[J]` specifies `u` as an array of J vectors of length `n_u`; hence, there is one vector per subject. The vector `sigma_u` contains the standard deviations of the by-subject varying intercepts and slopes u_0, u_1 , and the vector `sigma_w` contains the standard deviations of the by-item varying intercepts and slopes w_0, w_1 . The variable `sigma_e` is the standard deviation σ_e of the error ε . The transformed parameters block generates the by-subject intercepts and slopes u_0, u_1 and the by-item intercepts and slopes w_0, w_1 .

We place `lkj` priors on the random effects correlation matrices through the `lkj_corr_cholesky(2.0)` priors on their Cholesky factors `L_u` and `L_w`. We implicitly place uniform priors on the fixed effects β_0, β_1 , the random effects standard deviations σ_{u0}, σ_{u1} , and σ_{w0}, σ_{w1} and the error standard deviation σ_e by omitting any prior specifications for them in the model block. We specify the likelihood with the probability statement that `rt[i]` is distributed log-normally with mean `X[i] * beta + Z_u[i] * u[subj[i]] + Z_w[i] * w[item[i]]` and standard deviation `sigma_e`. The next step towards model-fitting is to pass the list `stanDat` to `stan`, which compiles a C++ program to sample from the posterior distribution of the model parameters.

A major advantage of the above matrix formulation is that we do not need to write a new Stan model for a future repeated measures design. All we have to do now is define the design matrix \mathbf{X} appropriately, and include it (along with appropriately defined \mathbf{Z}_u and \mathbf{Z}_w for the subjects and items random effects) as part of the data specification that is passed to

Stan.

Concluding remarks, and further reading

We hope that this tutorial has given the reader a flavor of what it would be like to fit Bayesian linear mixed models. There is of course much more to say on the topic, and we hope that the interested reader will take a look at some of the excellent books that have recently come out. We suggest below a sequence of reading that we found helpful. A good first general textbook is by Gelman and Hill (2007); it begins with the frequentist approach and only later transitions to Bayesian models. The book by McElreath (2016) is also excellent. For those looking for a psychology-specific introduction, the books by Kruschke (2014) and Lee and Wagenmakers (2013) are to be recommended, although for the latter the going might be easier if the reader has already looked at Gelman and Hill (2007). As a second book, Lunn et al. (2012) is recommended; it provides many interesting and useful examples using the BUGS language, which are discussed in exceptionally clear language. Many of these books use the BUGS syntax (Lunn et al., 2000), which the probabilistic programming language JAGS (Plummer, 2012) also adopts; however, Stan code for these books is slowly becoming available on the Stan home page (<https://github.com/stan-dev/example-models/wiki>). For those with introductory calculus, a slightly more technical introduction to Bayesian methods by Lynch (2007) is an excellent choice. Finally, the textbook by Gelman et al. (2014) is the definitive modern guide, and provides a more advanced treatment.

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