

## Review

# An Observatory for the *MET* Oncogene: A Guide for Targeted Therapies

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**Simple Summary:** The *MET* gene encodes a receptor critical for cell growth and repair. It plays diverse roles in processes like organ development, wound healing, and blood vessel formation. Genetic alterations in *MET* contribute to cancer progression, enabling tumor spread and resistance to treatment. Scientists are studying how to target *MET* to treat cancer. Despite progress, the complexity of *MET*'s functions in cancer challenges our understanding. This review explores recent discoveries about *MET* in cancer, its effects, potential therapies, and future directions.

**Abstract:** The *MET* proto-oncogene encodes a pivotal tyrosine kinase receptor, binding the hepatocyte growth factor (HGF, also known as scatter factor, SF) and governing essential biological processes such as organogenesis, tissue repair, and angiogenesis. The pleiotropic physiological functions of *MET* explain its diverse role in cancer progression in a broad range of tumors; genetic/epigenetic alterations of *MET* drive tumor cell dissemination, metastasis, and acquired resistance to conventional and targeted therapies. Therefore, targeting *MET* emerged as a promising strategy, and many efforts were devoted to identifying the optimal way of hampering *MET* signaling. Despite encouraging results, however, the complexity of *MET*'s functions in oncogenesis yields intriguing observations, fostering a humbler stance on our comprehension. This review explores recent discoveries concerning *MET* alterations in cancer, elucidating their biological repercussions, discussing therapeutic avenues, and outlining future directions. By contextualizing the research question and articulating the study's purpose, this work navigates *MET* biology's intricacies in cancer, offering a comprehensive perspective.

**Keywords:** *MET* alterations; cancer progression; invasive growth; therapeutic targeting



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## 1. Introduction

The human *MET* protooncogene encodes the tyrosine kinase receptor for the hepatocyte growth factor/scatter factor (HGF/SF) [1–3]. It is expressed in a broad range of epithelial cells, and the resulting receptor is a 170 kDa transmembrane protein organized in two disulfide bond-linked  $\alpha$  and  $\beta$  subunits of, respectively, 50 kDa and 145 kDa [4,5]. The ligand, SF, is produced and secreted in physiologic conditions by mesenchymal cells close to *MET*-expressing epithelial cells [6,7]. This paracrine effect leads to *MET* activation by autophosphorylation of the cytoplasmic catalytic domain and recruitment of adaptor proteins, promoting signal transduction. The mitogen-activated protein kinase (MAPK) pathway, Phosphoinositide 3-kinase/Protein kinase B (PI3K/AKT) signaling, and Signal Transducer and Activator of Transcription 3 (STAT3) constitute the primary signal transducers [8–10]. *MET* plays an essential role during embryogenesis (e.g., epithelial to mesenchymal transition, organ development) and the postnatal period (e.g., angiogenesis, organ regeneration, and wound healing, as reviewed in [11]).

The broad range of critical biological responses induced by *MET* awards this oncogene as a crucial oncogene in tumor progression, enabling cancer cells to survive and escape the hostile primary tumor microenvironment and form distal metastases [12]. *MET* is altered in multiple cancer types and behaves as a pivotal regulator of invasive growth, a complex

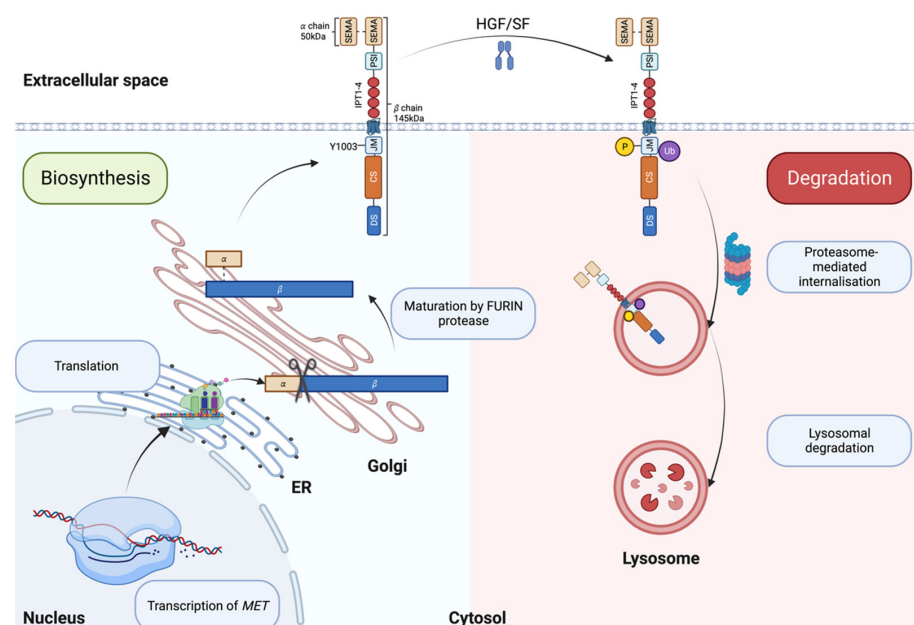
and intertwined sequence of events including epithelial-to-mesenchymal transition (EMT), scattering, migration, and growth [3,13–17]. Tumor cells harboring such alterations become ‘addicted’ to MET (described in [3]), therefore instituting a tumor cell-specific vulnerability point and justifying targeted therapies. Accordingly, MET inhibition reduces tumor size and impedes metastases in rodent models [12,18,19]. Recently, MET has been suggested as one of the top five proteins to focus on in targeted cancer therapies [20].

However, it should be noted that the path to efficient MET-targeted therapy for patients is long and covered with complications. Despite the presumed ‘addiction’ to MET, many patients do not respond to therapy [12,21–23]. Moreover, acquired resistance to MET inhibitors might arise [24–27].

This review reminds the overall structure of MET and summarizes the observed *MET* alterations in cancer, their impact on invasive growth, and their therapeutic potential. The lessons from the disappointing results of targeted therapies will be investigated to propose more accurate strategies to extend disease-free survival time.

## 2. Structure and Function of the MET Kinase

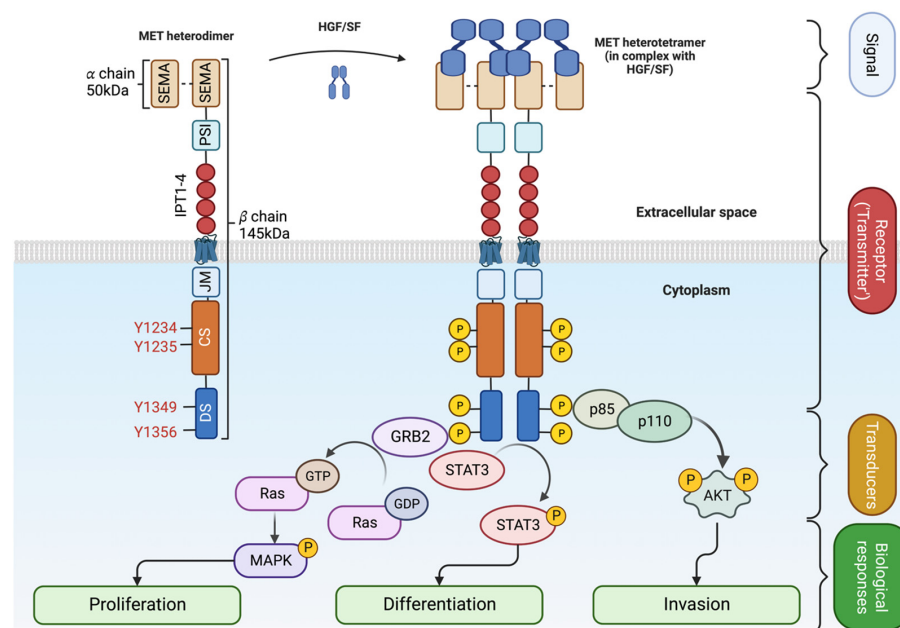
MET is the receptor tyrosine kinase (RTK) for SF. The translated precursor protein of 175kDa matures through proteolytic cleavage by FURIN protease in the Golgi apparatus (Figure 1, left panel and [28]). The mature protein forms a heterodimer of an extracellular  $\alpha$  subunit (50 kDa) linked via disulfide bonds to the 145 kDa transmembrane  $\beta$  subunit. The extracellular portion of the  $\beta$  chain comprises a semaphorin (SEMA) domain, a Plexin–Semaphorin–Integrin (PSI) homology domain, and four immunoglobulin-like IPT domains [29,30]. SEMA and IPT domains are crucial for ligand binding and receptor dimerization, while the PSI domain is essential for the proper maturation of the receptor through its recently described disulfide isomerase activity [31]. The intracellular part of MET is composed of a short juxtamembrane domain (JM), followed by the catalytic site and the docking site for signal transducers.



**Figure 1.** Biosynthesis (left, blue box) and degradation (right, pink box) of MET. ER: Endoplasmic reticulum; SEMA: semaphorin domain; PSI: plexin–semaphoring–integrin homology domain; IPT: immunoglobulin-like domain; JM: juxtamembrane domain; CS: catalytic site; DS: docking site. Created with [BioRender.com](https://www.biorender.com) (accessed on 5 August 2023).

Upon SF binding, the catalytic domain of MET becomes auto-phosphorylated on Tyrosine 1234 and Tyrosine 1235 [32]. The activation of the catalytic site triggers the phosphorylation

of two tyrosine on the docking site (Y1349 and Y1356), priming the interaction with Src homology 2 (SH2) domain-containing proteins [10,33,34]. The MET docking site is multifunctional; Y1349 phosphorylation leads to the activation of the PI3K/AKT pathway (migration/survival) while Y1356 phosphorylation activates the RAS/MAPK pathway (proliferation/cell cycle progression) [8,10]. The STAT3 transcription factor is also activated by MET [9]. Like other RTKs, MET transmits the information from the extracellular space to the cytoplasm, generating a multilayered network that activates various biological processes, including migration, growth, and differentiation/stemness (Figure 2 and [35]).



**Figure 2.** SF-dependent MET activation. Upon ligand binding (HGF/SF), MET heterodimer forms a tetramer [5]. The extracellular signal is transmitted into the cytoplasm, leading to the recruitment of adaptor proteins (GRB2 for MAPK pathway and p85/p110 for AKT) and signal transduction. Created with [BioRender.com](https://www.biorender.com) (accessed on 5 August 2023).

SF/MET interaction induces the phosphorylation of Y1003 within the juxtamembrane domain, allowing the recruitment of the E3 ubiquitin ligase casitas B-lineage lymphoma (CBL), promoting MET monoubiquitylation [36], receptor internalization and lysosomal degradation (Figure 1, right panel and [37]). Accordingly, experimental evidence has shown that Y1003F mutation stabilizes the receptor [38]. The juxtamembrane domain is therefore accepted as a negative regulator of the MET/SF axis [36,39–42].

Proteomics-based studies have been invaluable in understanding conventional RTKs like EGFR, where signaling typically shows rapid activation and deactivation in response to ligand binding [43]. In contrast, MET diverges from this pattern, making it an ‘unconventional RTK’. Unlike conventional RTKs, which generally experience transient phosphorylation followed by rapid deactivation, MET activation remains persistent after ligand binding [44]. In the broader context of RTK signaling, conventional RTKs often follow a swift sequence of activation events involving various cellular components such as MAPK, adaptor proteins, and guanine nucleotide exchange factors [45]. This unique activation profile of MET opens new avenues for future research, particularly to explore the implications of its long-lasting response in both physiology and tumor biology [17].

### 3. MET Alterations in Tumors and Their Biological Significance

Recent advances in next-generation sequencing technologies allowed a significant reduction in their cost, ultimately leading to an inflation of publicly available ‘OMICS’ data. Additionally, improvements in the standardization of data curation, analyses, and

presentation offered researchers an unprecedented quantity of comprehensive information regarding genetic alterations during cancer onset and progression. In line with this, we developed an auto-updatable ‘MET observatory’ to catalogue genetic alterations of the *MET* gene in cancer. Here, we present some of the features of this observatory; notably, the catalog of alterations results from data collection from The Cancer Genome Atlas (TCGA), Catalogue of Somatic Mutations in Cancer (COSMIC), and ClinVar datasets.

#### 4. MET Amplification

An oncogene can be defined as an entity that can transform cells by conferring some attributes of a cancer cell. *MET* is an excellent example of an oncogene because of its critical involvement in cell migration, metastasis, and cell survival. Unsurprisingly, *MET* is altered in many cancers, including but not limited to non-small cell lung cancer (NSCLC), lung squamous carcinoma, gastric cancer, colorectal adenocarcinoma, melanoma, gliomas, and renal cancer [46–51]. One of the most frequently observed molecular aberrations involving *MET* is gene amplification, exhibiting a prevalence rate of approximately 4% across various tumor types, as illustrated in Figure 3A and corroborated by multiple studies [52–57]. Notably, the incidence of *MET* amplification surges to nearly 20% in kidney papillary cell carcinomas (KIRP), a data point presented in Figure 3B. Moreover, *MET* amplification has been shown to confer resistance to therapies targeting the epidermal growth factor receptor (EGFR) in malignancies such as NSCLC and colorectal cancer, underlining its role in both cell survival and acquired resistance to EGFR-targeting therapies [56,58,59].

While interpreting these findings, it is crucial to place *MET* amplification in the broader context of common chromosomal aberrations in cancer, such as cellular aneuploidy. Specifically, the trisomy of chromosome 7 is often observed across multiple cancer types and serves as a pan-cancer genetic marker [60–62]. This trisomy could confound the assessment of *MET* amplification because both are related, but have distinct genomic alterations affecting cellular phenotype and treatment response. Unlike chromosome 7 trisomy, which is not a primary cancer driver, *MET* amplification acts as a driver and represents a true biological selection [3]. In vitro and preclinical studies suggest that a ‘threshold’ of five copies of the *MET* gene drives addiction, thus justifying targeted therapies [63]. Although no clinical consensus exists for such a cut-off, it is critical for effective patient stratification in *MET*-targeted therapies. Fluorescence in situ hybridization (FISH) techniques can distinguish between chromosome 7 polysomy and true *MET* amplification. In the case of polysomy, the *MET*-to-centromere of the chromosome 7 ratio (*MET*/*CEN7*) remains constant, whereas it increases for biologically selected true *MET* amplification, identifying a patient subgroup that could benefit from targeted therapies. Current advances in next-generation sequencing (NGS) techniques can also offer invaluable information for better patient stratification. Indeed, recent studies have demonstrated the efficiency of *MET* inhibition when patients were classified using either the *MET*/*CEN7* ratio or NGS-based detection of the *MET* copy number, as shown in Table 1 and supported by studies [64,65].

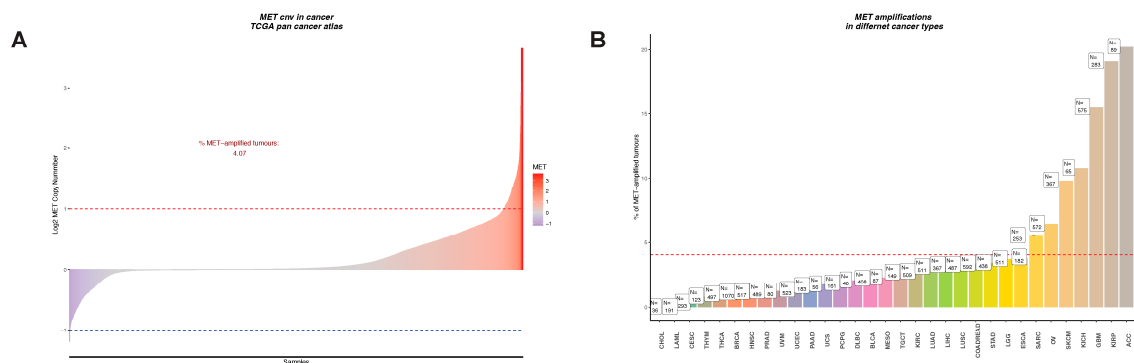
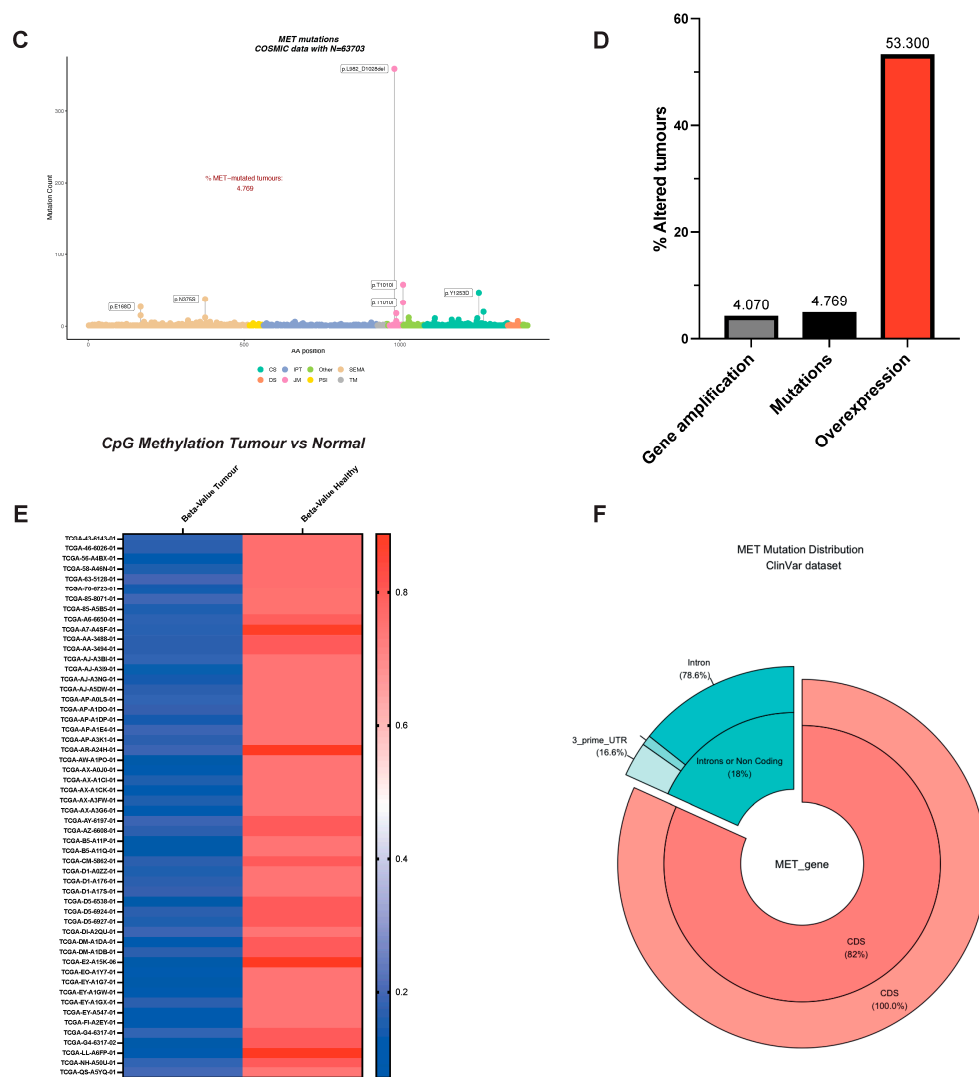


Figure 3. Cont.



**Figure 3.** The MET Observatory. **(A)** Copy number variation (CNV) in human tumors (TCGA pan cancer atlas). Dashed lines correspond to a copy number variation equal to 2 in absolute values. **(B)** *MET* CNV across tumor types. The dashed line corresponds to the average percentage of CNV in all types of cancer using TCGA abbreviations: ACC (Adrenocortical carcinoma), BLCA (Bladder Urothelial Carcinoma), BRCA (Breast invasive carcinoma), CESC (Cervical squamous cell carcinoma and endocervical adenocarcinoma), CHOL (Cholangiocarcinoma), COAD (Colon adenocarcinoma), DLBC (Lymphoid Neoplasm Diffuse Large B-cell Lymphoma), ESCA (Esophageal carcinoma), GBM (Glioblastoma multiforme), HNSC (Head and Neck squamous cell carcinoma), KICH (Kidney Chromophobe), KIRC (Kidney renal clear cell carcinoma), KIRP (Kidney renal papillary cell carcinoma), LAML (Acute Myeloid Leukemia), LGG (Brain Lower Grade Glioma), LIHC (Liver hepatocellular carcinoma), LUAD (Lung adenocarcinoma), LUSC (Lung squamous cell carcinoma), MESO (Mesothelioma), OV (Ovarian serous cystadenocarcinoma), PAAD (Pancreatic adenocarcinoma), PCPG (Pheochromocytoma and Paraganglioma), PRAD (Prostate adenocarcinoma), READ (Rectum adenocarcinoma), SARC (Sarcoma), SKCM (Skin Cutaneous Melanoma), STAD (Stomach adenocarcinoma), TGCT (Testicular Germ Cell Tumors), THCA (Thyroid carcinoma), THYM (Thymoma), UCEC (Uterine Corpus Endometrial Carcinoma), UCS (Uterine Carcinosarcoma), UVM (Uveal Melanoma). **(C)** *MET* mutations in the protein-coding region and distribution over the *MET* protein domains. **(D)** Comparison between the incidence of *MET* amplification, mutations, and overexpression. **(E)** B-value of CpG methylation of the *MET* promoter using the cg22116492 probe in cancer and in non-tumoral tissues. **(F)** Percentage of mutations affecting protein-coding and regulatory regions of *MET* mRNA (ClinVar).



**Table 1.** \* As described in August 2022 in clinicaltrials.gov. Early phase I: exploratory trials before phase I. NA: trials without FDA-defined phases. \*\* Advanced cancers of various origins; ADCC: antibody-dependent cell-mediated cytotoxicity; AML: acute myeloid lymphoma; HCC: hepatocellular carcinoma; NSCLC: non-small cell lung cancer; PRCC: papillary renal cell carcinoma; SCLC: small cell lung cancer.

Drug	Number of Trials (Phase) *	Cancer Types	Principal Outcome	Notes
Multitarget tyrosine kinase inhibitors				
PF02341066 (Crizotinib)	51 (early I/I); 62 (II); 18 (III); 5 (IV); 7 (NA)	Breast cancer, renal clear cell cancer, glioblastoma, inflammatory myofibroblastic tumors, lymphoma, papillary renal cancers, MET+ gastric adenocarcinoma, MET+ or RON+ metastatic urothelial cancer and NSCLC	Substantial antitumor activity in patients with <i>MET</i> amplification and/or <i>MET</i> Δ14 [66–68]. Crizotinib overcomes resistance to selpercatinib in <i>RET</i> -fusion positive NSCLC patients [69].	<ul style="list-style-type: none"> <li>Targets: <i>MET</i>, <i>ROS1</i>, and <i>ALK</i></li> <li>Approved for treating NSCLC with <i>EML4</i>–<i>ALK</i> in 2011 and NSCLC with <i>CD74</i>–<i>ROS1</i> in 2016 [67].</li> </ul>
XL184 (Cabozantinib)	54 (early I/I); 157 (II); 19 (III); 2 (IV); 7 (NA)	Breast cancer, glioblastoma, HCC, kidney cancer, medullary thyroid cancer, melanoma, NSCLC, ovarian cancer, and prostate cancer	Cabozantinib significantly improved progression-free survival in patients with metastatic PRCC and melanoma [70,71]. Complete response was reported in one patient with <i>MET</i> Δ14 [72].	<ul style="list-style-type: none"> <li>Targets: <i>MET</i>, <i>RET</i>, and others.</li> <li>Approved for the treatment of medullary thyroid cancer [73].</li> </ul>
GSK1363089 (Foretinib)	6 (early I/I); 7 (II)	Mixed cancer, breast cancer, gastric cancer, head and neck cancer, liver cancer, NSCLC, and PRCC	No activity in unselected patients [74].	Targets: <i>MET</i> , <i>RET</i> , and others.
MGCD265 (Glesatinib)	4 (early I/I); 2 (II)	Mixed cancer and NSCLC	Results are pending. The safety profile is acceptable in non-genetically selected patients with advanced solid tumors [75].	Targets: <i>MET</i> and <i>AXL</i>
MP470 (Amuvatinib)	2 (early I/I); 1 (II); 1 (NA)	Mixed cancer, gastric cancer, glioblastoma, pancreatic cancer, and SCLC	Results are pending.	Targets: <i>MET</i> , <i>RET</i> , <i>FLT3</i> , and <i>PDGFRA</i>
E7050 (Golvatinib)	8 (early I/I)	Mixed cancer, gastric cancer, head and neck cancer, and HCC	Results are pending.	Targets: <i>MET</i> and <i>VEGFR2</i>
Specific <i>MET</i> inhibitors (small molecules)				
ARQ197 (Tivantinib)	25 (early I/I); 21 (II); 4 (III)	Mixed cancer **, colorectal cancer, HCC, liver cancer, mesothelioma, NSCLC, SCLC, and stomach cancer	Tivantinib treatment did not demonstrate efficacy in a Phase III trial for HCC patients with high <i>MET</i> levels (based on staining intensity) [76]. Capmatinib showed substantial antitumor activity in patients with <i>MET</i> amplification or <i>MET</i> Δ14 [79–81]. Capmatinib induces potentially similar resistance mechanisms as Crizotinib [82] but is a promising option in <i>MET</i> -amplified, <i>EGFR</i> -inhibitor-resistant tumors [80].	Specificity to <i>MET</i> is controversial [77,78].
INCB28060 (Capmatinib)	18 (early I/I); 27 (II); 3 (III); 1 (IV)	Mixed cancer, colorectal cancer, glioblastoma, head and neck cancer, HCC, NSCLC, and PRCC	Encouraging results in <i>EGFR</i> -mutated, <i>MET</i> -amplified tumors [64,65].	Approved to treat adult patients with metastatic NSCLC with <i>MET</i> Δ14 [83].
AZD6094 (Savolitinib or Volitinib)	7 (early I/I); 8 (II); 3 (III)	Mixed cancer, colorectal cancer, gastric cancer, NSCLC, kidney cancer, and PRCC	Results are pending.	NA
AMG337	3 (early I/I); 5 (II)	Mixed cancer, renal clear cell cancer, esophageal cancer, and stomach cancer	Results are pending.	NA
MSC2156119J (Tepotinib)	5 (early I/I); 5 (II)	Mixed cancer, lung cancer, and NSCLC	Partial response in NSCLC patients with <i>MET</i> Δ14 [84]. Promising results in patients with <i>MET</i> amplification [85,86].	NA
OMO-1 (JNJ-38877618)	1 (early I/I)	Mixed cancer, lung cancer, and NSCLC	Results are pending.	NA

Table 1. Cont.

Drug	Number of Trials (Phase) *	Cancer Types	Principal Outcome	Notes
MET antibodies				
MetMab (Onartuzumab)	6 (early I/I); 8 (II); 5 (III)	Mixed cancer, breast cancer, colorectal cancer, glioblastoma, HER2- and MET+ gastric cancer, HCC, and MET+ NSCLC	Some clinical trials were inconclusive due to poor patient selection or the premature termination of the study [87–89]. Other results are pending. Cytostatic antitumor activity [91]. Significant increase in median progression-free survival for patients with the highest MET expression [92]. It cannot reverse acquired resistance to Erlotinib, an EGFR inhibitor [93].	One-armed monoclonal antibody [90].
LY2875358 (Emibetuzumab)	1 (early I/I); 2 (II)	Mixed cancer, gastric cancer, and NSCLC		Humanized IgG4 bivalent monoclonal antibody [94].
ARGX-111	1 (early I/I)	Mixed cancer, gastric cancer, glioblastoma, liver cancer, and renal cancer	Results are pending.	Bivalent monoclonal antibody with the property to activate ADCC [95].
SAIT301	1 (early I/I)	Mixed cancer	Phase I completed, and the recommended dose for phase II was established [96].	Bivalent monoclonal antibodies targeted against the MET $\alpha$ chain, inducing CBL-independent degradation of MET to circumvent the detrimental agonist effect of other bivalent antibodies [97,98].
SF antibodies				
AMG 102 (Rilotumumab)	6 (early I/I); 13 (II); 3 (III)	Mixed cancer, gastric cancer, glioblastoma, lung cancer, mesothelioma, and prostate cancer	No improvement in the clinical outcome in patients with MET+ gastric cancer [99]. Toxicity issue [100].	Humanized IgG2 monoclonal antibody [101].
AV-299 (Ficlatuzumab)	6 (early I/I); 4 (II); 3 (III)	AML, head and neck cancer, liver cancer, and NSCLC	Low benefit compared to other drugs [102,103].	Humanized IgG1 monoclonal antibody [104].

### 5. Exon14 ‘Skipping’, the Predominant MET Alteration

It was thus proposed that other mutations might drive ligand-independent activation of MET. Surprisingly, for an RTK (e.g., *EGFR*, fibroblast growth factor receptor (*FGFR*), for review, see [105]), very few patients exhibited *MET* mutations in the kinase domain or regulatory regions (Figure 3C and [17,106]). Splice site mutations spanning the *Exon14* were by far the most common *MET* mutations. These mutations (complex or simple) are a consequence of the loss of acceptor or donor sites, resulting in *Exon14* ‘skipping’. Indeed, the latter was described in a significant number of patients: 13% in pulmonary sarcomatoid carcinoma, 6% in adenosquamous carcinoma, 3% in lung adenocarcinomas, 2% in lung squamous cell carcinomas, 0.4% in gliomas, and 0.4% in cancers of unknown primary origin (CUP) [16,107]. *MET Exon14* encodes for the juxtamembrane domain. Because of the regulatory role of the JM described above, it was thought that its loss would lead to increased receptors, and the subsequent ligand-independent uncontrolled activation of MET, thus driving invasive growth. Accordingly, the re-insertion of *Exon14* into the oncogenic gene fusion (*TPR*)–*MET*, which consists of the *MET* sequence downstream from the juxtamembrane domain fused to the dimerization motif of *TPR*, resulted in decreased oncogenic potential [108]. Therefore, targeted therapies against MET constituted appealing strategies for cancer patients carrying *MET Exon14* deletion (*METΔ14*) [21,23,72,79,84,109–115]. However, only half of the patients harboring *METΔ14* benefited from MET-targeted therapies, suggesting that the critical aspects of *METΔ14* remain to be elucidated [116,117]. Recently, two independent studies demonstrated that the deletion of *Exon14* does not result in constitutive activation of the kinase. *METΔ14* activity requires SF and drives a robust and selective AKT activation, rendering cancer cells more prone to survival and migration [17,118]. Therefore, it is proposed that cancer cells expressing *METΔ14* choose the astute strategy to ‘fly’ the local hostile micro milieu to form distal metastases instead of ‘fighting’ to proliferate locally. The absence of SF in the tumor microenvironment or PI3K/AKT axis mutations may explain the insensitivity to targeted therapies. Fittingly, PI3K/AKT activating mutations co-occur

with *MET Exon14* ‘skipping’ in 14% of cancer patients [119,120]. Based on these results, a better stratification of patients might lead to a better response to MET-targeted therapies.

## 6. Point Mutations within the *MET* Coding Sequence

As shown in Figure 3C, mutations affecting the catalytic site or regulatory sites of *MET* are sporadic but do exist. The first activating mutations of *MET* were identified in hereditary papillary renal carcinoma (HPRC), and the authors suggested that the mutations affecting the kinase domain of *MET* (M1149T, V1206L, V1238L, D1246N, and Y1248C) were causal in HPRC [121]. Similar *MET* mutations (D1246H, Y1228C, and M1268T) spanning the critical Y1234 and Y1235 were described in the sporadic renal carcinoma [122]. Moreover, cytogenetic analyses showed non-random chromosome 7 trisomy, which affected the mutated *MET* allele [123]. All these mutations have the common feature of inducing the constitutive activation of the kinase [122,124,125], leading to oncogene ‘addiction’ [3]. Experiments in transgenic murine demonstrated the oncogenic potential of these activating mutants.

Interestingly, tumors formed in mice were not restricted to the kidney; animals developed lymphomas, carcinomas, or aggressive mammary tumors [126,127]. Importantly, these independent studies demonstrate that activating mutations affecting the *MET* catalytic site drive tumorigenesis in multiple tissues. Researchers attempted to identify activating mutations in other human cancers in the following years. Somatic or germline mutations were described in hepatocellular carcinoma, head and neck cancers, oropharynx squamous cell cancer, gastric cancer, cancers of unknown primary origin (CUP), and colorectal cancer [128–134].

A growing number of point mutations were described in the SEMA domain (both in  $\alpha$  and  $\beta$  chains, Figure 3C) responsible for the ligand binding [135]. Six non-small cell lung cancer patients out of 127 harbored mutations (L229F, N375S, E168D, N375S, S323G, N375S) within the *Exon2* encoding for the SEMA domain [136]. SEMA domain mutations are not restricted to lung cancer. In an independent study, *MET* mutations were detected in 9% of advanced breast cancer (8/88 patients). Six of eight *MET* mutations affected the SEMA domain (N375S in five patients, M362T in one patient, [137]). Despite some evidence showing that SEMA domain mutations are oncogenic [133,138], our knowledge of the biological significance of *MET* mutations affecting the SEMA domain remains poor. They likely affect the ligand-binding domain’s structure, promoting a constitutively active or ligand-hypersensitive kinase. Accordingly, in CUP—owning the unique ability of self-renewal in the absence of any exogenous growth factor [139]—*MET* mutations were clustered within the SEMA domain; in a total of 23 CUP patients, five out of seven *MET* alterations were localized in the SEMA domain (H150Y, Q142X, C385Y, and two patients with E168D [133]).

Point mutations, although rare, should not be overlooked. With recent advancements in genomic screening technologies, we might find a growing number of these mutations. By combining *in silico*, *in vitro*, and *in vivo* findings, we aim to have a sharper picture of *MET* alterations in cancer and their biological significance, opening new avenues for targeted therapies.

For RTKs, it is generally accepted that gene amplification leads to a higher number of receptors at the cell surface, priming kinase activation in the presence of small amounts of ligands. *MET* challenges this concept; an overexpression of human *MET* in mouse liver induces hepatocellular carcinomas [140]. Since human *MET* cannot interact with murine SF [141], these results demonstrate the ligand-independent activation of the receptor when overexpressed.

## 7. Fusion Partners of *MET* Drive Oncogene ‘Addiction’

For decades, the only known *MET* gene rearrangement in human tumors has been *TPR-MET*, mostly occurring in gastric cancers [142]. Recently, the thorough analyses of the vast TCGA tumor collection uncovered new hybrid proteins [143]: the *MET* intracellular domain fused at the N-terminus with several partners, some of them encompass the dimerization

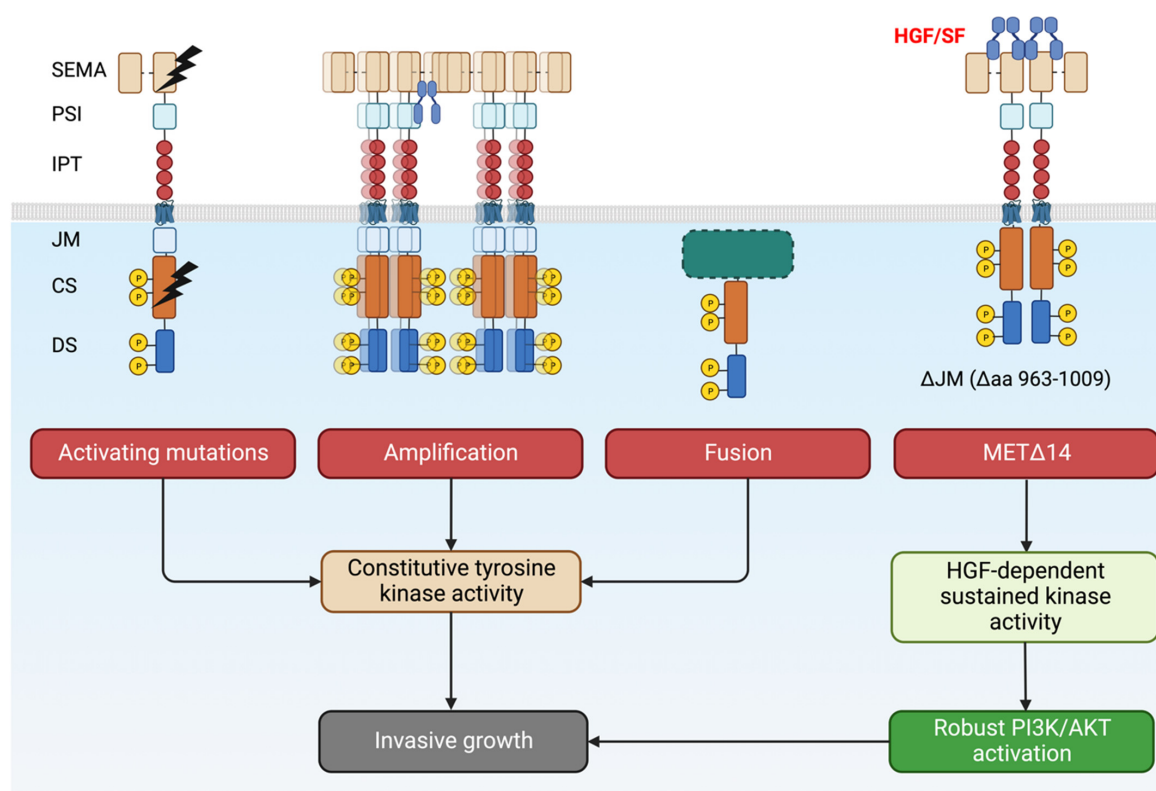


‘coiled-coil’ (CC) motif (i.e., C8orf34, BAIAP2L1, TFG, and KIF5B). Consequently, the chimeric MET dimerizes in a ligand-independent fashion, driving constitutive kinase activity and tumorigenesis. Although occurring at low frequencies, these fusions have been found in lung adenocarcinomas, hepatocellular carcinomas, papillary renal carcinomas, and thyroid carcinomas; thus, they cannot be ignored [143].

Another recurrent gene rearrangement involves *MET* and the *PTPRZ1* gene, encoding a tyrosine phosphatase [144]. *PTPRZ1*–*MET* fusions include almost the entire *MET* sequence fused at its 5′ end with a variable number of *exons* of the *PTPRZ1* gene [145]. *PTPRZ1*–*MET* fusions have been found in brain tumors, such as low-grade gliomas, secondary glioblastomas arising in adults from the progression of lower-grade gliomas, and pediatric glioblastomas at a remarkably high frequency (~10%, [145]). Notably, the chromosomal rearrangement between *PTPRZ1* and *MET* leads to fusion protein overexpression and enhanced kinase activation [146]. The mechanism explaining enhanced *MET* activity in tumors expressing the fusion protein remains to be determined; the highly active *PTPRZ1* promoter fused to the *MET* gene [145] and the coiled-coil domain of *PTPRZ1* fused to *MET* [147] are two mutually non-exclusive hypotheses.

Experiments show that *MET* fusion proteins respond to anti-*MET* monotherapy: *PTPRZ1*–*MET* in a pediatric glioma [145] and *KIF5B*–*MET* in lung cancers [148]. *MET* gene fusions also happen in melanomas, where six different N-terminal partners fused in-frame with the intracellular *MET* domain have been described [23].

Different *MET* genetic alterations can induce either ligand-independent (or hypersensitive) or SF-dependent (*MET*Δ14) activation of the kinase (Figure 4). They have a common denominator for driving invasive growth. Tumor cells thus become addicted to *MET* and become vulnerable to targeted therapies.



**Figure 4.** Overview of *MET* alterations in cancer that drives invasive growth. Tumors become ‘addicted’ to *MET* and become a candidate for targeted therapies. Created with [BioRender.com](https://www.biorender.com) (accessed on 5 August 2023).

## 8. Genetic Alterations of *MET*: The Peak of the Iceberg?

*MET*-amplified tumors represent 4%, where *MET* overexpression is observed in more than 50% of cancers (Figure 3D). The discrepancy is too large and deserves the attention of the scientific community. It should be noted that a large dataset often restricts their analyses to protein-coding regions, overlooking important regulatory regions crucial for gene expression. The inducible nature of the *MET* promoter was previously described and its importance in tumor biology was established [149–151]. Thus, alterations affecting the *MET* promoter should not be disregarded: re-analyses of TCGA dataset showed a remarkable decrease in promoter methylation in cancer patients, a synonym for transcription activation and *MET* overexpression (Figure 3E). This observation suggests that central regulatory mechanisms remain to be elucidated. Gene expression is not only regulated at the transcription level. Post-transcription control mechanisms affecting the translation efficiency and the messenger's stability are hubs for deregulations observed in cancer. Indeed, we observed a small but significant fraction of patients harboring mutations within untranslated regions of the mRNA (Figure 3F). However, despite their biological significance being unknown, their exploration might reveal an unsuspected 'dark energy' for tumor cells. Additionally, we have recently revealed *MET* translational regulation by the PI3K/AKT/mTOR axis and its relevance in therapy resistance. This aspect is detailed below.

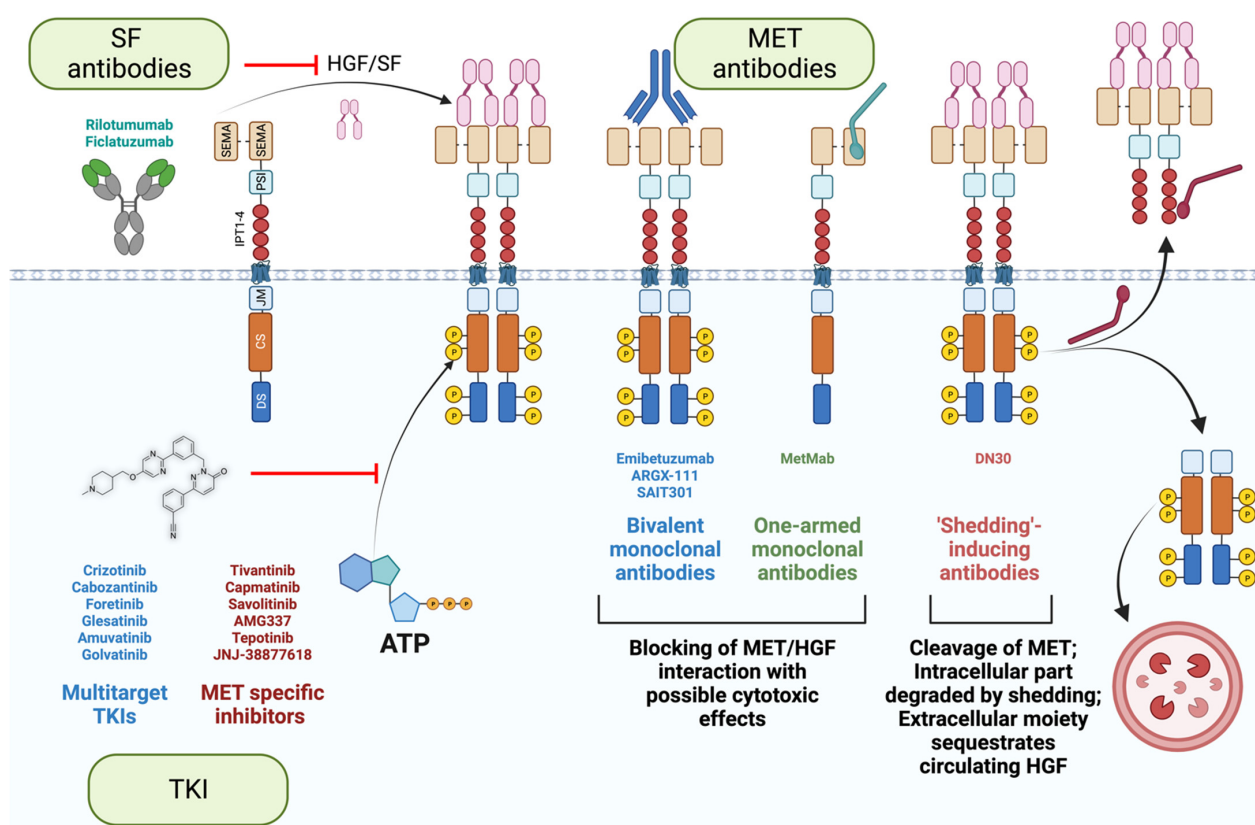
## 9. *MET*-Targeted Therapies

Several *MET*-targeting agents have been developed in line with the statements above (summarized in Table 1). As with other targeted therapies, several questions must be addressed before trying to quench *MET* signaling and tumor growth in patients. We can encapsulate these matters in three main topics: (1) how to quench *MET* signaling in patients; (2) who should benefit from *MET*-targeting agents; and (3) how to face the inevitable problem in the targeted therapy: drug resistance.

## 10. Different *MET*-Blocking Agents: Advantages and Pitfalls

Three main strategies were employed to extinguish the *MET* signaling (schematized in Figure 5). Firstly, small kinase inhibitors and monoclonal antibodies targeting SF or its receptor [3,12,152]. Small kinase inhibitors are chemical compounds that pass through the plasma membrane and interact with the receptor kinase domain. They can target a large panel of receptors (multitarget tyrosine kinase inhibitors), specifically *MET*. The latter has the advantage of reducing off-target effects. On the other hand, because of the crosstalk of receptors [44,153–155], targeting a broader number of RTKs might induce a better clinical outcome. Tyrosine kinase inhibitors act as ATP mimetics, hampering receptor phosphorylation and subsequent kinase activity [156]. However, as discussed in the first chapter, *MET* phosphorylation is indispensable for its degradation [36,37]. In the chronic treatment setting, small molecules can thus potentially increase the number of receptors at the cell surface, suffocating the treatment efficacy. Additionally, an acquired, or existing mutation in the ATP binding pocket can engender resistance, as previously observed for small molecule inhibitors of EGFR, KIT, and BCR-ABL [157–160].

Secondly, an alternative approach is to target the extracellular moiety of *MET* using antibodies. They are more specific by nature than chemical inhibitors. Importantly, antibodies are insensitive to multidrug resistance, a feature of aggressive cancer cells that augments the drug efflux, reduces its influx, or increases the drug catabolism [56,161–164]. Furthermore, antibodies recognize *MET*, even if the intracellular part is mutated in cancer cells (*MET*Δ14 or activating mutations in the catalytic site). However, antibodies can induce receptor dimerization and activation. Different strategies have been employed to circumvent this issue. One-armed monoclonal antibodies (MetMab, also known as Onartuzumab, [90]) and antibodies inducing CBL-independent degradation of *MET* (SAIT301, [97,98]) have been developed and tested in clinics (Table 1).



**Figure 5.** Targeting MET/SF axis in cancer. Tyrosine kinase inhibitors (TKI) act as ATP mimetics to impede receptor phosphorylation. MET/SF interaction using can be prevented using either SF antibodies or MET antibodies. Some MET antibodies can induce receptor degradation in CBL-independent manner (SAIT301) or by shedding (DN30). Additionally, in the case of DN30, the extracellular part of the trimmed receptor has the potential to sequester circulating SF. Created with BioRender.com (accessed on 5 August 2023).

Lastly, another innovative strategy is taking advantage of the shedding of the plasma membrane to maintain homeostasis [165]; DN30 antibodies interact with the IPT domain of MET with subnanomolar affinities and induce shedding through the cleavage of the extracellular moiety of MET [166–168]. The proteasome subsequently degrades the intracellular moiety [169,170]. Additionally, since the extracellular part of MET is trimmed, it can potentially sequester the circulating SF, further hampering the SF/MET axis. Furthermore, based on the straightforward elimination of MET from the cell surface, independently of receptor activation (phosphorylated vs. unphosphorylated state), DN30 has a substantial advantage over other MET antibodies, as it is effective in the full spectrum of MET activation mechanisms, whether SF-dependent or independent (that is, induced by mutations, gene fusions, or amplification). Multiple studies demonstrated its remarkable potential; DN30 hampers cell growth and induces apoptosis in multiple MET-addicted cell lines in vitro, and induces an impressive reduction of tumor mass in vivo [168,171]. Importantly, DN30 displayed a favorable pharmacokinetics and safety profile in non-human primates [171], encouraging the design of a clinical trial in MET-addicted cancer patients.

Targeting the ligand is an alternative option to impede MET signaling. The benefit of the SF antibodies AMG 102 (Rilotumumab, [101]) and AV-299 (Ficlatuzumab) was assessed in many clinical trials with disappointing results (Table 1). In one clinical trial, SF antibodies significantly increased mortality, causing the premature ending of the study [99]. As discussed previously, *MET* alterations in tumors typically promote ligand-independent kinase activity (gene amplification, activating mutations, fusion proteins), partially explaining these frustrating results. Nevertheless, targeting SF should not be dismissed; their ability

to sequester the ligand could be exploited to inhibit SF-dependent invasion and survival in tumors without *MET* amplification (e.g., *MET* $\Delta$ 14). Furthermore, it should be reminded that SF acts on the tumor micro milieu on cancer-associated fibroblasts and macrophages to foster angiogenesis [172–175]; therefore, a strategy is valid only if employed toward the right target.

### 11. Patient Stratification: A Key for Success in Targeted Therapies

Identifying the target population for treatment might look self-evident, but it is laborious and critical for setting the scene for success in a clinical trial. More than three decades of basic research guided drawing an overall picture of patients that can benefit from *MET* inhibition. Studies using cell lines or patient-derived xenografts have shown that only tumors harboring *MET* alterations (mostly amplification) respond to the *MET* blockade. The cell cycle is arrested, and/or apoptosis is induced in vitro [176], and complete inhibition of tumor growth (and even tumor shrinkage) is observed in vivo [59]. *MET* alterations must be assessed in patients before assigning them to the group receiving targeted therapies. This is indeed a golden rule to follow, as targeted therapies can only benefit patients presenting *MET* alterations. Nevertheless, their unambiguous identification is challenging. In many clinical trials, *MET* levels were assessed by immunohistochemistry [83,92], a strategy being far from objective and hardly reproducible. Moreover, the increased protein intensity is not necessarily a synonym for *MET* amplification and ‘addiction’. Because of the inducible nature of the *MET* promoter [149–151], high-*MET*-protein levels may be transient due to changes in the tumor microenvironment (e.g., hypoxia, ionizing radiation, cytotoxic reagents, described as ‘expedience’ in [3]). Indeed, post hoc analyses of some clinical trials illustrate this issue; patients with a gene copy number gain of *MET* > 4 exhibited the uppermost progression-free survival [92]. Another study testing the efficiency of MetMab failed to demonstrate any benefit of the treatment, although 88% of patients were *MET*-negative [87], demonstrating abruptly the validity of the golden rule stated above.

For tumors expressing wt *MET* (the vast majority of patients), a priori ineligible for *MET*-targeted therapies, it should be noted that hampering the *MET* signaling reduces migration and metastatic dissemination drastically without affecting the growth of cancer cells [63,177,178]. Using *MET*-targeted therapy might constitute a promising adjuvant therapy after tumor resection with curative intent, an ideal setting to eradicate the persistence and dissemination of subclinical tumor foci.

With the progress in next-generation sequencing technologies (lesser quantity of material required for acceptable coverage), genomic interrogations of liquid biopsies (circulating tumor DNA and circulating tumor cells) are largely feasible [179,180] to adequately and objectively stratify patients. Ideally, *MET* alterations must be assessed before the initiation of the clinical trial. Liquid biopsies also enable longitudinal evaluation of tumor evolution in a non-invasive manner, crucial for understanding mechanisms hidebound to drug resistance, the major challenge in targeted therapy.

### 12. Understanding and Overcoming Drug Resistance

Targeted therapies were a breakthrough in cancer research, emphasizing our cutting-edge understanding of cancer cells’ vulnerabilities. The design of appropriate treatment strategies was thus achievable. However, they suffer a significant limitation: resistance frequently occurs [181–183] after an initial response. Understanding how cancer cells evade targeted therapies constitutes a substantial challenge in the clinic.

Tumors can be defined as pseudo-organs constituted of heterogeneous clones presenting highly diverse genotypes and phenotypes. Some might prosper while others regress depending on their ability to face microenvironmental selection pressures [183]. *MET* genetic alterations might dominate the majority of tumor cells and dictate drug sensitivity; however, minor subclones harboring other mutations that confer resistance to *MET* blockade may coexist and be positively selected under drug pressure. Resistant subclones must be promptly detected (e.g., liquid biopsies at regular intervals). In line with this idea,



many efforts have been made to characterize the molecular profile of emerging resistant clones [184–186]. These forerunner studies have undeniable value for discovering new targetable biomarkers in persistent clones that limit their propagation with proper and timely therapeutic interventions.

### 13. The Flare Effect: mTOR Pathway Comes to Scene

In current clinical practice, the line of treatment is dismissed when resistance arises. In many patients, discontinuation of kinase inhibitors results in rapid tumor regrowth: this phenomenon is known as disease ‘flare’ or tumor ‘rebound’ [187–190], characterized by an unknown incidence and a cumbersome prognosis [191]. The occurrence of tumor ‘rebound’ complicates disease management and has led to the idea of continuing the therapy beyond the progression [191]. Notably, treatment with a MET therapeutic antibody that induces ‘shedding’ (proteolytic cleavage of the receptor at the cell surface) substantially prevents this effect, providing a rationale to combine, or alternate, MET-targeted drugs with different mechanisms of action [192].

We recently presented a mechanistic elucidation of the ‘flare’ effect [193]. Within MET-amplified cells, halting the administration of the small molecule JNJ-38877618 (MET inhibitor) triggered activation within the protein kinase B/mechanistic target of the rapamycin (AKT/mTOR) pathway. Notably, mTOR orchestrates cell growth by governing mRNA translation, ribosome biogenesis, and metabolic processes [194–197]. Noteworthy transformations have been observed due to the overexpression of mTOR’s target, eIF4E, in rodent fibroblasts [198], consequently prompting extensive investigations into mTOR’s role in cancer [194]. Efforts towards inhibiting mTOR with rapalogs (including rapamycin and analogs) have been pursued in cancer therapeutics; however, the exclusive utilization of rapalogs displayed suboptimal efficacy [199].

A captivating observation emerged as mTOR swiftly augmented *MET* translation upon MET inhibitor withdrawal [193], uncovering a novel aspect of the AKT/mTOR axis in conferring resistance against therapies. Beyond de novo protein synthesis, the influence of the AKT/mTOR pathway on MET extends further. This pathway’s Ser/Thr kinase activity phosphorylates and incapacitates the protein phosphatase PTP1B, resulting in elevated phosphorylated (active) MET at the cell surface [193]. Consequently, the AKT/mTOR pathway emerges as a promising candidate for targeted interventions due to its paramount role in driving tumor rebound. These mechanistic insights hold the potential to guide the formulation of metronomic treatment strategies, incorporating alternating MET inhibitors and mTOR inhibitors with minimal washout periods.

In a broader context, the intricate interplay between the mTOR axis and MET biology underscores the pivotal role of translational regulation in governing MET expression within the realm of cancer. Translational control, distinct from the transcriptional regulations, operates on pre-existing mRNA, facilitating swift adaptability to the dynamic shifts within the tumor microenvironment. While a comprehensive evaluation of the relevance of *MET*’s translational control in the context of cancer demands further exploration, it might provide insight into the apparent paradox between sporadic MET amplifications (accounting for 3–5% of cases) and the nearly ubiquitous MET overexpression observed in cancer [200–203].

### 14. Conclusions

MET is a potent oncogene driving invasive growth [3,13–17]. Decades of fundamental research led to understanding MET’s different roles in human tumors. New studies are ongoing to fine-tune our understanding of the role of varying *MET* mutations in cancer [17,118,138]. The biological knowledge of MET needs to be translated into clinical applications. Recent failures of clinical trials can be primarily explained by the lack of consideration of *MET* genetic alterations before patient selection. On the contrary, when patient stratification was appropriately performed a priori or during post hoc analyses, a general picture of the target patients’ genomic profile can be pictured: *MET* alterations are an absolute prerequisite for the success of targeted therapies.



These lessons bear witness to crucial importance of the functional preclinical insights in guiding clinical practice. Indeed, it was already clear from studies in cell lines and animal models that substantial reduction of cancer cell viability in vitro and tumor shrinkage in vivo occur only in settings where stable and heritable genetic alterations of *MET* sustain oncogene ‘addiction’. For the large majority of cancer patients, where *MET* is not mutated, preclinical comprehension of the role of *MET* can be directly translated to the clinic: *MET* has anti-apoptotic and pro-invasive functions [177,178]. Adjuvant treatment with *MET* inhibitors after tumor resection with curative intent could be a viable strategy.

Another unexplained phenomenon is the relatively poor frequency of *MET*-amplified tumors [16,52–57,107], compared to many *MET*-overexpressing tumors [200–203]. This apparent paradox can be partially explained by the inducible nature of the *MET* promoter: ionizing radiations, hypoxia, and cytotoxic reagents enhance *MET* expression [149–151], previously described as oncogene-mediated ‘expedience’ [3]. Translational control mechanisms (e.g., mTOR) might add another layer of regulation of the receptor expression [193]. Another possibility is the existence of other mutations, either in the coding sequence or regulatory regions of the *MET* gene. The number of previously unidentified *MET* mutations is constantly increasing thanks to the advances in next-generation sequencing techniques and comprehensive publicly available databases (TCGA, cBioPortal, ClinVar). Thus, efforts aiming at the identification of new variants in cancer must be encouraged. The primary task should be to functionally characterize these mutations to broaden the panel of *MET* ‘addicted’ tumors and increase the number of patients eligible for next-generation precision medicine.

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## References

1. Weidner, K.M.; Hartmann, G.; Naldini, L.; Comoglio, P.M.; Sachs, M.; Fonatsch, C.; Rieder, H.; Birchmeier, W. Molecular characteristics of HGF-SF and its role in cell motility and invasion. *Exs* **1993**, *65*, 311–328.
2. Graveel, C.R.; Tolbert, D.; Vande Woude, G.F. *MET*: A critical player in tumorigenesis and therapeutic target. *Cold Spring Harb. Perspect. Biol.* **2013**, *5*, a009209. [[CrossRef](#)]
3. Comoglio, P.M.; Trusolino, L.; Boccaccio, C. Known and novel roles of the *MET* oncogene in cancer: A coherent approach to targeted therapy. *Nat. Rev. Cancer* **2018**, *18*, 341–358. [[CrossRef](#)]
4. Giordano, S.; Di Renzo, M.F.; Narsimhan, R.P.; Cooper, C.S.; Rosa, C.; Comoglio, P.M. Biosynthesis of the protein encoded by the c-met proto-oncogene. *Oncogene* **1989**, *4*, 1383–1388. [[PubMed](#)]
5. Uchikawa, E.; Chen, Z.; Xiao, G.Y.; Zhang, X.; Bai, X.C. Structural basis of the activation of c-MET receptor. *Nat. Commun.* **2021**, *12*, 4074. [[CrossRef](#)] [[PubMed](#)]
6. Nakamura, T. Structure and function of hepatocyte growth factor. *Prog. Growth Factor. Res.* **1991**, *3*, 67–85. [[CrossRef](#)] [[PubMed](#)]
7. Tajima, H.; Matsumoto, K.; Nakamura, T. Regulation of cell growth and motility by hepatocyte growth factor and receptor expression in various cell species. *Exp. Cell Res.* **1992**, *202*, 423–431. [[CrossRef](#)]
8. Ponzetto, C.; Bardelli, A.; Maina, F.; Longati, P.; Panayotou, G.; Dhand, R.; Waterfield, M.D.; Comoglio, P.M. A novel recognition motif for phosphatidylinositol 3-kinase binding mediates its association with the hepatocyte growth factor/scatter factor receptor. *Mol. Cell Biol.* **1993**, *13*, 4600–4608.
9. Boccaccio, C.; Ando, M.; Tamagnone, L.; Bardelli, A.; Michieli, P.; Battistini, C.; Comoglio, P.M. Induction of epithelial tubules by growth factor HGF depends on the STAT pathway. *Nature* **1998**, *391*, 285–288. [[CrossRef](#)]

10. Ponzetto, C.; Bardelli, A.; Zhen, Z.; Maina, F.; dalla Zonca, P.; Giordano, S.; Graziani, A.; Panayotou, G.; Comoglio, P.M. A multifunctional docking site mediates signaling and transformation by the hepatocyte growth factor/scatter factor receptor family. *Cell* **1994**, *77*, 261–271. [\[CrossRef\]](#)
11. Trusolino, L.; Bertotti, A.; Comoglio, P.M. MET signalling: Principles and functions in development, organ regeneration and cancer. *Nat. Rev. Mol. Cell Biol.* **2010**, *11*, 834–848. [\[CrossRef\]](#)
12. Gherardi, E.; Birchmeier, W.; Birchmeier, C.; Vande Woude, G. Targeting MET in cancer: Rationale and progress. *Nat. Rev. Cancer* **2012**, *12*, 89–103. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Boccaccio, C.; Comoglio, P.M. Invasive growth: A MET-driven genetic programme for cancer and stem cells. *Nat. Rev. Cancer* **2006**, *6*, 637–645. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Trusolino, L.; Comoglio, P.M. Scatter-factor and semaphorin receptors: Cell signalling for invasive growth. *Nat. Rev. Cancer* **2002**, *2*, 289–300. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Christofori, G. New signals from the invasive front. *Nature* **2006**, *441*, 444–450. [\[CrossRef\]](#)
16. Frampton, G.M.; Ali, S.M.; Rosenzweig, M.; Chmielecki, J.; Lu, X.; Bauer, T.M.; Akimov, M.; Bufill, J.A.; Lee, C.; Jentz, D.; et al. Activation of MET via diverse exon 14 splicing alterations occurs in multiple tumor types and confers clinical sensitivity to MET inhibitors. *Cancer Discov.* **2015**, *5*, 850–859. [\[CrossRef\]](#)
17. Cerqua, M.; Botti, O.; Arigoni, M.; Gioelli, N.; Serini, G.; Calogero, R.; Boccaccio, C.; Comoglio, P.M.; Altintas, D.M. MET14 promotes a ligand-dependent, AKT-driven invasive growth. *Life Sci. Alliance* **2022**, *5*, e202201409. [\[CrossRef\]](#)
18. Comoglio, P.M.; Giordano, S.; Trusolino, L. Drug development of MET inhibitors: Targeting oncogene addiction and expedience. *Nat. Rev. Drug Discov.* **2008**, *7*, 504–516. [\[CrossRef\]](#)
19. Corso, S.; Migliore, C.; Ghiso, E.; De Rosa, G.; Comoglio, P.M.; Giordano, S. Silencing the MET oncogene leads to regression of experimental tumors and metastases. *Oncogene* **2008**, *27*, 684–693. [\[CrossRef\]](#)
20. Behan, F.M.; Iorio, F.; Picco, G.; Goncalves, E.; Beaver, C.M.; Migliardi, G.; Santos, R.; Rao, Y.; Sassi, F.; Pinnelli, M.; et al. Prioritization of cancer therapeutic targets using CRISPR-Cas9 screens. *Nature* **2019**, *568*, 511–516. [\[CrossRef\]](#)
21. Drilon, A.; Clark, J.W.; Weiss, J.; Ou, S.I.; Camidge, D.R.; Solomon, B.J.; Otterson, G.A.; Villaruz, L.C.; Riely, G.J.; Heist, R.S.; et al. Antitumor activity of crizotinib in lung cancers harboring a MET exon 14 alteration. *Nat. Med.* **2020**, *26*, 47–51. [\[CrossRef\]](#)
22. Wolf, J.; Seto, T.; Han, J.-Y.; Reguart, N.; Garon, E.B.; Groen, H.J.M.; Tan, D.S.-W.; Hida, T.; Jonge, M.J.D.; Orlov, S.V.; et al. Capmatinib (INC280) in MET $\Delta$ ex14-mutated advanced non-small cell lung cancer (NSCLC): Efficacy data from the phase II GEOMETRY mono-1 study. *J. Clin. Oncol.* **2019**, *37*, 9004. [\[CrossRef\]](#)
23. Landi, L.; Chiari, R.; Tiseo, M.; D’Inca, F.; Dazzi, C.; Chella, A.; Delmonte, A.; Bonanno, L.; Giannarelli, D.; Cortinovis, D.L.; et al. Crizotinib in MET-Deregulated or ROS1-Rearranged Pretreated Non-Small Cell Lung Cancer (METROS): A Phase II, Prospective, Multicenter, Two-Arms Trial. *Clin. Cancer Res.* **2019**, *25*, 7312–7319. [\[CrossRef\]](#)
24. Cruickshanks, N.; Zhang, Y.; Hine, S.; Gibert, M.; Yuan, F.; Oxford, M.; Grello, C.; Patuski, M.; Dube, C.; Guessous, F.; et al. Discovery and Therapeutic Exploitation of Mechanisms of Resistance to MET Inhibitors in Glioblastoma. *Clin. Cancer Res.* **2019**, *25*, 663–673. [\[CrossRef\]](#)
25. Kim, S.; Kim, T.M.; Kim, D.W.; Kim, S.; Kim, M.; Ahn, Y.O.; Keam, B.; Heo, D.S. Acquired Resistance of MET-Amplified Non-small Cell Lung Cancer Cells to the MET Inhibitor Capmatinib. *Cancer Res. Treat.* **2019**, *51*, 951–962. [\[CrossRef\]](#)
26. Suzawa, K.; Offin, M.; Lu, D.; Kurzatkowski, C.; Vojnic, M.; Smith, R.S.; Sabari, J.K.; Tai, H.; Mattar, M.; Khodos, I.; et al. Activation of KRAS Mediates Resistance to Targeted Therapy in MET Exon 14-mutant Non-small Cell Lung Cancer. *Clin. Cancer Res.* **2019**, *25*, 1248–1260. [\[CrossRef\]](#)
27. Virzi, A.R.; Gentile, A.; Benvenuti, S.; Comoglio, P.M. Reviving oncogenic addiction to MET bypassed by BRAF (G469A) mutation. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 10058–10063. [\[CrossRef\]](#) [\[PubMed\]](#)
28. Komada, M.; Hatsuzawa, K.; Shibamoto, S.; Ito, F.; Nakayama, K.; Kitamura, N. Proteolytic processing of the hepatocyte growth factor/scatter factor receptor by furin. *FEBS Lett.* **1993**, *328*, 25–29. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Gherardi, E.; Youles, M.E.; Miguel, R.N.; Blundell, T.L.; Iamelle, L.; Gough, J.; Bandyopadhyay, A.; Hartmann, G.; Butler, P.J. Functional map and domain structure of MET, the product of the c-met protooncogene and receptor for hepatocyte growth factor/scatter factor. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 12039–12044. [\[CrossRef\]](#) [\[PubMed\]](#)
30. Zhen, Z.; Giordano, S.; Longati, P.; Medico, E.; Campiglio, M.; Comoglio, P. Structural and functional domains critical for constitutive activation of the HGF-receptor (Met). *Oncogene* **1994**, *9*, 1691–1697.
31. Altintas, D.M.; Gallo, S.; Basilico, C.; Cerqua, M.; Bocedi, A.; Vitacolonna, A.; Botti, O.; Casanova, E.; Rancati, I.; Milanese, C.; et al. The PSI Domain of the MET Oncogene Encodes a Functional Disulfide Isomerase Essential for the Maturation of the Receptor Precursor. *Int. J. Mol. Sci.* **2022**, *23*, 12427. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Longati, P.; Bardelli, A.; Ponzetto, C.; Naldini, L.; Comoglio, P.M. Tyrosines1234-1235 are critical for activation of the tyrosine kinase encoded by the MET proto-oncogene (HGF receptor). *Oncogene* **1994**, *9*, 49–57. [\[PubMed\]](#)
33. Giordano, S.; Bardelli, A.; Zhen, Z.; Menard, S.; Ponzetto, C.; Comoglio, P.M. A point mutation in the MET oncogene abrogates metastasis without affecting transformation. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 13868–13872. [\[CrossRef\]](#) [\[PubMed\]](#)
34. Ponzetto, C.; Zhen, Z.; Audero, E.; Maina, F.; Bardelli, A.; Basile, M.L.; Giordano, S.; Narsimhan, R.; Comoglio, P. Specific uncoupling of GRB2 from the Met receptor. Differential effects on transformation and motility. *J. Biol. Chem.* **1996**, *271*, 14119–14123. [\[CrossRef\]](#) [\[PubMed\]](#)
35. Yarden, Y.; Sliwkowski, M.X. Untangling the ErbB signalling network. *Nat. Rev. Mol. Cell Biol.* **2001**, *2*, 127–137. [\[CrossRef\]](#)

36. Peschard, P.; Fournier, T.M.; Lamorte, L.; Naujokas, M.A.; Band, H.; Langdon, W.Y.; Park, M. Mutation of the c-Cbl TKB domain binding site on the Met receptor tyrosine kinase converts it into a transforming protein. *Mol. Cell* **2001**, *8*, 995–1004. [[CrossRef](#)] [[PubMed](#)]
37. Lefebvre, J.; Ancot, F.; Leroy, C.; Muharram, G.; Lemiere, A.; Tulasne, D. Met degradation: More than one stone to shoot a receptor down. *FASEB J.* **2012**, *26*, 1387–1399. [[CrossRef](#)] [[PubMed](#)]
38. Abella, J.V.; Peschard, P.; Naujokas, M.A.; Lin, T.; Saucier, C.; Urbe, S.; Park, M. Met/Hepatocyte growth factor receptor ubiquitination suppresses transformation and is required for Hrs phosphorylation. *Mol. Cell Biol.* **2005**, *25*, 9632–9645. [[CrossRef](#)]
39. Tan, Y.H.; Krishnaswamy, S.; Nandi, S.; Kanteti, R.; Vora, S.; Onel, K.; Hasina, R.; Lo, F.Y.; El-Hashani, E.; Cervantes, G.; et al. CBL is frequently altered in lung cancers: Its relationship to mutations in MET and EGFR tyrosine kinases. *PLoS ONE* **2010**, *5*, e8972. [[CrossRef](#)]
40. Villa-Moruzzi, E.; Puntoni, F.; Bardelli, A.; Vigna, E.; De Rosa, S.; Comoglio, P.M. Protein tyrosine phosphatase PTP-S binds to the juxtamembrane region of the hepatocyte growth factor receptor Met. *Biochem. J.* **1998**, *336 Pt 1*, 235–239. [[CrossRef](#)]
41. Lee, C.C.; Yamada, K.M. Identification of a novel type of alternative splicing of a tyrosine kinase receptor. Juxtamembrane deletion of the c-met protein kinase C serine phosphorylation regulatory site. *J. Biol. Chem.* **1994**, *269*, 19457–19461. [[CrossRef](#)]
42. Lee, J.H.; Gao, C.F.; Lee, C.C.; Kim, M.D.; Vande Woude, G.F. An alternatively spliced form of Met receptor is tumorigenic. *Exp. Mol. Med.* **2006**, *38*, 565–573. [[CrossRef](#)]
43. Blagoev, B.; Ong, S.E.; Kratchmarova, I.; Mann, M. Temporal analysis of phosphotyrosine-dependent signaling networks by quantitative proteomics. *Nat. Biotechnol.* **2004**, *22*, 1139–1145. [[CrossRef](#)] [[PubMed](#)]
44. Trusolino, L.; Bertotti, A.; Comoglio, P.M. A signaling adapter function for alpha6beta4 integrin in the control of HGF-dependent invasive growth. *Cell* **2001**, *107*, 643–654. [[CrossRef](#)]
45. Schlessinger, J. Receptor tyrosine kinases: Legacy of the first two decades. *Cold Spring Harb. Perspect. Biol.* **2014**, *6*, a008912. [[CrossRef](#)]
46. Go, H.; Jeon, Y.K.; Park, H.J.; Sung, S.W.; Seo, J.W.; Chung, D.H. High MET gene copy number leads to shorter survival in patients with non-small cell lung cancer. *J. Thorac. Oncol.* **2010**, *5*, 305–313. [[CrossRef](#)]
47. Graziano, F.; Galluccio, N.; Lorenzini, P.; Ruzzo, A.; Canestrari, E.; D’Emidio, S.; Catalano, V.; Sisti, V.; Ligorio, C.; Andreoni, F.; et al. Genetic activation of the MET pathway and prognosis of patients with high-risk, radically resected gastric cancer. *J. Clin. Oncol.* **2011**, *29*, 4789–4795. [[CrossRef](#)]
48. Zhang, M.; Li, G.; Sun, X.; Ni, S.; Tan, C.; Xu, M.; Huang, D.; Ren, F.; Li, D.; Wei, P.; et al. MET amplification, expression, and exon 14 mutations in colorectal adenocarcinoma. *Hum. Pathol.* **2018**, *77*, 108–115. [[CrossRef](#)]
49. Pal, S.K.; Ali, S.M.; Yakirevich, E.; Geynisman, D.M.; Karam, J.A.; Elvin, J.A.; Frampton, G.M.; Huang, X.; Lin, D.I.; Rosenzweig, M.; et al. Characterization of Clinical Cases of Advanced Papillary Renal Cell Carcinoma via Comprehensive Genomic Profiling. *Eur. Urol.* **2018**, *73*, 71–78. [[CrossRef](#)] [[PubMed](#)]
50. Cheng, F.; Guo, D. MET in glioma: Signaling pathways and targeted therapies. *J. Exp. Clin. Cancer Res.* **2019**, *38*, 270. [[CrossRef](#)] [[PubMed](#)]
51. Zhou, Y.; Song, K.Y.; Giubellino, A. The Role of MET in Melanoma and Melanocytic Lesions. *Am. J. Pathol.* **2019**, *189*, 2138–2148. [[CrossRef](#)]
52. Houldsworth, J.; Cordon-Cardo, C.; Ladanyi, M.; Kelsen, D.P.; Chaganti, R.S. Gene amplification in gastric and esophageal adenocarcinomas. *Cancer Res.* **1990**, *50*, 6417–6422.
53. Kuniyasu, H.; Yasui, W.; Kitadai, Y.; Yokozaki, H.; Ito, H.; Tahara, E. Frequent amplification of the c-met gene in scirrhous type stomach cancer. *Biochem. Biophys. Res. Commun.* **1992**, *189*, 227–232. [[CrossRef](#)] [[PubMed](#)]
54. Hara, T.; Ooi, A.; Kobayashi, M.; Mai, M.; Yanagihara, K.; Nakanishi, I. Amplification of c-myc, K-sam, and c-met in gastric cancers: Detection by fluorescence in situ hybridization. *Lab. Invest.* **1998**, *78*, 1143–1153.
55. Miller, C.T.; Lin, L.; Casper, A.M.; Lim, J.; Thomas, D.G.; Orringer, M.B.; Chang, A.C.; Chambers, A.F.; Giordano, T.J.; Glover, T.W.; et al. Genomic amplification of MET with boundaries within fragile site FRA7G and upregulation of MET pathways in esophageal adenocarcinoma. *Oncogene* **2006**, *25*, 409–418. [[CrossRef](#)] [[PubMed](#)]
56. Engelman, J.A.; Zejnullahu, K.; Mitsudomi, T.; Song, Y.; Hyland, C.; Park, J.O.; Lindeman, N.; Gale, C.M.; Zhao, X.; Christensen, J.; et al. MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. *Science* **2007**, *316*, 1039–1043. [[CrossRef](#)] [[PubMed](#)]
57. Bubendorf, L.; Dafni, U.; Schobel, M.; Finn, S.P.; Tischler, V.; Sejda, A.; Marchetti, A.; Thunnissen, E.; Verbeken, E.K.; Warth, A.; et al. Prevalence and clinical association of MET gene overexpression and amplification in patients with NSCLC: Results from the European Thoracic Oncology Platform (ETOP) Lungscape project. *Lung Cancer* **2017**, *111*, 143–149. [[CrossRef](#)]
58. Bean, J.; Brennan, C.; Shih, J.Y.; Riely, G.; Viale, A.; Wang, L.; Chitale, D.; Motoi, N.; Szoke, J.; Broderick, S.; et al. MET amplification occurs with or without T790M mutations in EGFR mutant lung tumors with acquired resistance to gefitinib or erlotinib. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 20932–20937. [[CrossRef](#)]
59. Bardelli, A.; Corso, S.; Bertotti, A.; Hobor, S.; Valtorta, E.; Siravegna, G.; Sartore-Bianchi, A.; Scala, E.; Cassingena, A.; Zecchin, D.; et al. Amplification of the MET receptor drives resistance to anti-EGFR therapies in colorectal cancer. *Cancer Discov.* **2013**, *3*, 658–673. [[CrossRef](#)]

60. Bandyk, M.G.; Zhao, L.; Troncoso, P.; Pisters, L.L.; Palmer, J.L.; von Eschenbach, A.C.; Chung, L.W.; Liang, J.C. Trisomy 7: A potential cytogenetic marker of human prostate cancer progression. *Genes. Chromosomes Cancer* **1994**, *9*, 19–27. [[CrossRef](#)] [[PubMed](#)]
61. Broberg, K.; Toksvig-Larsen, S.; Lindstrand, A.; Mertens, F. Trisomy 7 accumulates with age in solid tumors and non-neoplastic synovia. *Genes. Chromosomes Cancer* **2001**, *30*, 310–315. [[CrossRef](#)]
62. Ried, T.; Knutzen, R.; Steinbeck, R.; Blegen, H.; Schröck, E.; Heselmeyer, K.; du Manoir, S.; Auer, G. Comparative genomic hybridization reveals a specific pattern of chromosomal gains and losses during the genesis of colorectal tumors. *Genes Chromosomes Cancer* **1996**, *15*, 234–245. [[CrossRef](#)]
63. Benvenuti, S.; Gentile, A.; Lazzari, L.; Arnesano, A.; Trusolino, L.; Comoglio, P.M. An ‘in-cell trial’ to assess the efficacy of a monovalent anti-MET antibody as monotherapy and in association with standard cytotoxics. *Mol. Oncol.* **2014**, *8*, 378–388. [[CrossRef](#)]
64. Sequist, L.V.; Han, J.Y.; Ahn, M.J.; Cho, B.C.; Yu, H.; Kim, S.W.; Yang, J.C.; Lee, J.S.; Su, W.C.; Kowalski, D.; et al. Osimertinib plus savolitinib in patients with EGFR mutation-positive, MET-amplified, non-small-cell lung cancer after progression on EGFR tyrosine kinase inhibitors: Interim results from a multicentre, open-label, phase 1b study. *Lancet Oncol.* **2020**, *21*, 373–386. [[CrossRef](#)] [[PubMed](#)]
65. Choueiri, T.K.; Heng, D.Y.C.; Lee, J.L.; Cancel, M.; Verheijen, R.B.; Mellemgaard, A.; Ottesen, L.H.; Frigault, M.M.; L’Hernault, A.; Szigyarto, Z.; et al. Efficacy of Savolitinib vs Sunitinib in Patients With MET-Driven Papillary Renal Cell Carcinoma: The SAVOIR Phase 3 Randomized Clinical Trial. *JAMA Oncol.* **2020**, *6*, 1247–1255. [[CrossRef](#)]
66. Awad, M.M.; Oxnard, G.R.; Jackman, D.M.; Savukoski, D.O.; Hall, D.; Shivdasani, P.; Heng, J.C.; Dahlberg, S.E.; Janne, P.A.; Verma, S.; et al. MET Exon 14 Mutations in Non-Small-Cell Lung Cancer Are Associated With Advanced Age and Stage-Dependent MET Genomic Amplification and c-Met Overexpression. *J. Clin. Oncol.* **2016**, *34*, 721–730. [[CrossRef](#)]
67. Lennerz, J.K.; Kwak, E.L.; Ackerman, A.; Michael, M.; Fox, S.B.; Bergethon, K.; Lauwers, G.Y.; Christensen, J.G.; Wilner, K.D.; Haber, D.A. MET amplification identifies a small and aggressive subgroup of esophagogastric adenocarcinoma with evidence of responsiveness to crizotinib. *J. Clin. Oncol.* **2011**, *29*, 4803. [[CrossRef](#)]
68. Mo, H.N.; Liu, P. Targeting MET in cancer therapy. *Chronic Dis. Transl. Med.* **2017**, *3*, 148–153. [[CrossRef](#)] [[PubMed](#)]
69. Rosen, E.Y.; Johnson, M.L.; Clifford, S.E.; Somwar, R.; Kherani, J.F.; Son, J.; Bertram, A.A.; Davare, M.A.; Gladstone, E.; Ivanova, E.V.; et al. Overcoming MET-Dependent Resistance to Selective RET Inhibition in Patients with RET Fusion-Positive Lung Cancer by Combining Selpercatinib with Crizotinib. *Clin. Cancer Res.* **2021**, *27*, 34–42. [[CrossRef](#)] [[PubMed](#)]
70. Pal, S.K.; Tangen, C.; Thompson, I.M., Jr.; Balzer-Haas, N.; George, D.J.; Heng, D.Y.C.; Shuch, B.; Stein, M.; Tretiakova, M.; Humphrey, P.; et al. A comparison of sunitinib with cabozantinib, crizotinib, and savolitinib for treatment of advanced papillary renal cell carcinoma: A randomised, open-label, phase 2 trial. *Lancet* **2021**, *397*, 695–703. [[CrossRef](#)]
71. Daud, A.; Kluger, H.M.; Kurzrock, R.; Schimmoller, F.; Weitzman, A.L.; Samuel, T.A.; Moussa, A.H.; Gordon, M.S.; Shapiro, G.I. Phase II randomised discontinuation trial of the MET/VEGF receptor inhibitor cabozantinib in metastatic melanoma. *Br. J. Cancer* **2017**, *116*, 432–440. [[CrossRef](#)]
72. Paik, P.K.; Drilon, A.; Fan, P.-D.; Yu, H.; Rekhtman, N.; Ginsberg, M.S.; Borsu, L.; Schultz, N.; Berger, M.F.; Rudin, C.M. Response to MET Inhibitors in Patients with Stage IV Lung Adenocarcinomas Harboring MET Mutations Causing Exon 14 Skipping MET Inhibitors in MET Exon 14 Splice Variant Lung Cancer. *Cancer Discov.* **2015**, *5*, 842–849. [[CrossRef](#)]
73. Elisei, R.; Schlumberger, M.J.; Muller, S.P.; Schoffski, P.; Brose, M.S.; Shah, M.H.; Licitra, L.; Jarzab, B.; Medvedev, V.; Kreissl, M.C.; et al. Cabozantinib in progressive medullary thyroid cancer. *J. Clin. Oncol.* **2013**, *31*, 3639–3646. [[CrossRef](#)]
74. Shah, M.A.; Wainberg, Z.A.; Catenacci, D.V.; Hochster, H.S.; Ford, J.; Kunz, P.; Lee, F.C.; Kallender, H.; Cecchi, F.; Rabe, D.C.; et al. Phase II study evaluating 2 dosing schedules of oral foretinib (GSK1363089), cMET/VEGFR2 inhibitor, in patients with metastatic gastric cancer. *PLoS ONE* **2013**, *8*, e54014. [[CrossRef](#)] [[PubMed](#)]
75. Patnaik, A.; Gadgeel, S.; Papadopoulos, K.P.; Rasco, D.W.; Haas, N.B.; Der-Torossian, H.; Faltaos, D.; Potvin, D.; Tassell, V.; Tawashi, M.; et al. Phase I Study of Glesatinib (MGCD265) in Combination with Erlotinib or Docetaxel in Patients with Advanced Solid Tumors. *Target. Oncol.* **2022**, *17*, 125–138. [[CrossRef](#)] [[PubMed](#)]
76. Kudo, M.; Morimoto, M.; Moriguchi, M.; Izumi, N.; Takayama, T.; Yoshiji, H.; Hino, K.; Oikawa, T.; Chiba, T.; Motomura, K.; et al. A randomized, double-blind, placebo-controlled, phase 3 study of tivantinib in Japanese patients with MET-high hepatocellular carcinoma. *Cancer Sci.* **2020**, *111*, 3759–3769. [[CrossRef](#)] [[PubMed](#)]
77. Basilico, C.; Pennacchietti, S.; Vigna, E.; Chiriaco, C.; Arena, S.; Bardelli, A.; Valdembri, D.; Serini, G.; Michieli, P. Tivantinib (ARQ197) displays cytotoxic activity that is independent of its ability to bind MET. *Clin. Cancer Res.* **2013**, *19*, 2381–2392. [[CrossRef](#)] [[PubMed](#)]
78. Calles, A.; Kwiatkowski, N.; Cammarata, B.K.; Ercan, D.; Gray, N.S.; Janne, P.A. Tivantinib (ARQ 197) efficacy is independent of MET inhibition in non-small-cell lung cancer cell lines. *Mol. Oncol.* **2015**, *9*, 260–269. [[CrossRef](#)] [[PubMed](#)]
79. Wolf, J.; Seto, T.; Han, J.-Y.; Reguart, N.; Garon, E.B.; Groen, H.J.; Tan, D.S.; Hida, T.; de Jonge, M.; Orlov, S.V. Capmatinib in MET exon 14-mutated or MET-amplified non-small-cell lung cancer. *N. Engl. J. Med.* **2020**, *383*, 944–957. [[CrossRef](#)]
80. Wu, Y.L.; Zhang, L.; Kim, D.W.; Liu, X.; Lee, D.H.; Yang, J.C.; Ahn, M.J.; Vansteenkiste, J.F.; Su, W.C.; Felip, E.; et al. Phase Ib/II Study of Capmatinib (INC280) Plus Gefitinib After Failure of Epidermal Growth Factor Receptor (EGFR) Inhibitor Therapy in Patients With EGFR-Mutated, MET Factor-Dysregulated Non-Small-Cell Lung Cancer. *J. Clin. Oncol.* **2018**, *36*, 3101–3109. [[CrossRef](#)]



81. Schuler, M.; Berardi, R.; Lim, W.T.; de Jonge, M.; Bauer, T.M.; Azaro, A.; Gottfried, M.; Han, J.Y.; Lee, D.H.; Wollner, M.; et al. Molecular correlates of response to capmatinib in advanced non-small-cell lung cancer: Clinical and biomarker results from a phase I trial. *Ann. Oncol.* **2020**, *31*, 789–797. [[CrossRef](#)] [[PubMed](#)]
82. Dagogo-Jack, I.; Moonsamy, P.; Gainor, J.F.; Lennerz, J.K.; Piotrowska, Z.; Lin, J.J.; Lennes, I.T.; Sequist, L.V.; Shaw, A.T.; Goodwin, K.; et al. A Phase 2 Study of Capmatinib in Patients With MET-Altered Lung Cancer Previously Treated With a MET Inhibitor. *J. Thorac. Oncol.* **2021**, *16*, 850–859. [[CrossRef](#)]
83. Grande, E.; Giovannini, M.; Marriere, E.; Pultar, P.; Quinlan, M.; Chen, X.; Rahmanzadeh, G.; Curigliano, G.; Cui, X. Effect of capmatinib on the pharmacokinetics of digoxin and rosuvastatin administered as a 2-drug cocktail in patients with MET-dysregulated advanced solid tumours: A phase I, multicentre, open-label, single-sequence drug-drug interaction study. *Br. J. Clin. Pharmacol.* **2021**, *87*, 2867–2878. [[CrossRef](#)] [[PubMed](#)]
84. Paik, P.K.; Felip, E.; Veillon, R.; Sakai, H.; Cortot, A.B.; Garassino, M.C.; Mazieres, J.; Viteri, S.; Senellart, H.; Van Meerbeeck, J.; et al. Tepotinib in Non-Small-Cell Lung Cancer with MET Exon 14 Skipping Mutations. *N. Engl. J. Med.* **2020**, *383*, 931–943. [[CrossRef](#)]
85. Wu, Y.L.; Cheng, Y.; Zhou, J.; Lu, S.; Zhang, Y.; Zhao, J.; Kim, D.W.; Soo, R.A.; Kim, S.W.; Pan, H.; et al. Tepotinib plus gefitinib in patients with EGFR-mutant non-small-cell lung cancer with MET overexpression or MET amplification and acquired resistance to previous EGFR inhibitor (INSIGHT study): An open-label, phase 1b/2, multicentre, randomised trial. *Lancet Respir. Med.* **2020**, *8*, 1132–1143. [[CrossRef](#)] [[PubMed](#)]
86. Decaens, T.; Barone, C.; Assenat, E.; Wermke, M.; Fasolo, A.; Merle, P.; Blanc, J.F.; Grando, V.; Iacobellis, A.; Villa, E.; et al. Phase 1b/2 trial of tepotinib in sorafenib pretreated advanced hepatocellular carcinoma with MET overexpression. *Br. J. Cancer* **2021**, *125*, 190–199. [[CrossRef](#)] [[PubMed](#)]
87. Dieras, V.; Campone, M.; Yardley, D.A.; Romieu, G.; Valero, V.; Isakoff, S.J.; Koeppen, H.; Wilson, T.R.; Xiao, Y.; Shames, D.S.; et al. Randomized, phase II, placebo-controlled trial of onartuzumab and/or bevacizumab in combination with weekly paclitaxel in patients with metastatic triple-negative breast cancer. *Ann. Oncol.* **2015**, *26*, 1904–1910. [[CrossRef](#)]
88. Shah, M.A.; Bang, Y.J.; Lordick, F.; Alsina, M.; Chen, M.; Hack, S.P.; Bruey, J.M.; Smith, D.; McCaffery, I.; Shames, D.S.; et al. Effect of Fluorouracil, Leucovorin, and Oxaliplatin With or Without Onartuzumab in HER2-Negative, MET-Positive Gastroesophageal Adenocarcinoma: The METGastric Randomized Clinical Trial. *JAMA Oncol.* **2017**, *3*, 620–627. [[CrossRef](#)]
89. Kishi, K.; Sakai, H.; Seto, T.; Kozuki, T.; Nishio, M.; Imamura, F.; Nokihara, H.; Satouchi, M.; Nakagawa, S.; Tahata, T.; et al. First-line onartuzumab plus erlotinib treatment for patients with MET-positive and EGFR mutation-positive non-small-cell lung cancer. *Cancer Treat. Res. Commun.* **2019**, *18*, 100113. [[CrossRef](#)]
90. Martens, T.; Schmidt, N.-O.; Eckerich, C.; Fillbrandt, R.; Merchant, M.; Schwall, R.; Westphal, M.; Lamszus, K. A novel one-armed anti-c-Met antibody inhibits glioblastoma growth in vivo. *Clin. Cancer Res.* **2006**, *12*, 6144–6152. [[CrossRef](#)]
91. Harding, J.J.; Zhu, A.X.; Bauer, T.M.; Choueiri, T.K.; Drlon, A.; Voss, M.H.; Fuchs, C.S.; Abou-Alfa, G.K.; Wijayawardana, S.R.; Wang, X.A.; et al. A Phase Ib/II Study of Ramucirumab in Combination with Emibetuzumab in Patients with Advanced Cancer. *Clin. Cancer Res.* **2019**, *25*, 5202–5211. [[CrossRef](#)]
92. Scagliotti, G.; Moro-Sibilot, D.; Kollmeier, J.; Favaretto, A.; Cho, E.K.; Grosch, H.; Kimmich, M.; Girard, N.; Tsai, C.M.; Hsia, T.C.; et al. A Randomized-Controlled Phase 2 Study of the MET Antibody Emibetuzumab in Combination with Erlotinib as First-Line Treatment for EGFR Mutation-Positive NSCLC Patients. *J. Thorac. Oncol.* **2020**, *15*, 80–90. [[CrossRef](#)] [[PubMed](#)]
93. Camidge, D.R.; Moran, T.; Demedts, I.; Grosch, H.; Mileham, K.; Molina, J.; Juan-Vidal, O.; Bepler, G.; Goldman, J.W.; Park, K.; et al. A Randomized, Open-Label Phase II Study Evaluating Emibetuzumab Plus Erlotinib and Emibetuzumab Monotherapy in MET Immunohistochemistry Positive NSCLC Patients with Acquired Resistance to Erlotinib. *Clin. Lung Cancer* **2022**, *23*, 300–310. [[CrossRef](#)] [[PubMed](#)]
94. Liu, L.; Zeng, W.; Wortinger, M.A.; Yan, S.B.; Cornwell, P.; Peek, V.L.; Stephens, J.R.; Tetreault, J.W.; Xia, J.; Manro, J.R. LY2875358, a neutralizing and internalizing anti-MET bivalent antibody, inhibits HGF-dependent and HGF-independent MET activation and tumor growth. *Clin. Cancer Res.* **2014**, *20*, 6059–6070. [[CrossRef](#)]
95. Hultberg, A.; Morello, V.; Huyghe, L.; De Jonge, N.; Blanchetot, C.; Hanssens, V.; De Boeck, G.; Silence, K.; Festjens, E.; Heukens, R. Depleting MET-Expressing Tumor Cells by ADCC Provides a Therapeutic Advantage over Inhibiting HGF/MET Signaling. *ARGX-111 Kills MET-Expressing Cancer Cells by ADCC. Cancer Res.* **2015**, *75*, 3373–3383. [[CrossRef](#)]
96. Lee, J.; Kim, S.T.; Park, S.; Lee, S.; Park, S.H.; Park, J.O.; Lim, H.Y.; Ahn, H.; Bok, H.; Kim, K.M.; et al. Phase I Trial of Anti-MET Monoclonal Antibody in MET-Overexpressed Refractory Cancer. *Clin. Color. Cancer* **2018**, *17*, 140–146. [[CrossRef](#)]
97. Lee, B.-S.; Kang, S.; Kim, K.-A.; Song, Y.-J.; Cheong, K.H.; Cha, H.-Y.; Kim, C.H. Met degradation by SAIT301, a Met monoclonal antibody, reduces the invasion and migration of nasopharyngeal cancer cells via inhibition of EGR-1 expression. *Cell Death Dis.* **2014**, *5*, e1159. [[CrossRef](#)]
98. Lee, J.M.; Kim, B.; Lee, S.; Jeong, Y.; Oh, Y.; Song, Y.; Jung, S.; Choi, J.; Lee, S.; Cheong, K. Cbl-independent degradation of Met: Ways to avoid agonism of bivalent Met-targeting antibody. *Oncogene* **2014**, *33*, 34–43. [[CrossRef](#)]
99. Catenacci, D.V.T.; Tebbutt, N.C.; Davidenko, I.; Murad, A.M.; Al-Batran, S.E.; Ilson, D.H.; Tjulandin, S.; Gotovkin, E.; Karaszewska, B.; Bondarenko, I.; et al. Rilotumumab plus epirubicin, cisplatin, and capecitabine as first-line therapy in advanced MET-positive gastric or gastro-oesophageal junction cancer (RILOMET-1): A randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol.* **2017**, *18*, 1467–1482. [[CrossRef](#)]



100. Affronti, M.L.; Jackman, J.G.; McSherry, F.; Herndon, J.E., 2nd; Massey, E.C., Jr.; Lipp, E.; Desjardins, A.; Friedman, H.S.; Vlahovic, G.; Vredenburgh, J.; et al. Phase II Study to Evaluate the Efficacy and Safety of Rilotumumab and Bevacizumab in Subjects with Recurrent Malignant Glioma. *Oncologist* **2018**, *23*, 889–e898. [\[CrossRef\]](#) [\[PubMed\]](#)
101. Burgess, T.L.; Sun, J.; Meyer, S.; Tsuruda, T.S.; Sun, J.; Elliott, G.; Chen, Q.; Haniu, M.; Barron, W.F.; Juan, T. Biochemical Characterization of AMG 102: A Neutralizing, Fully Human Monoclonal Antibody to Human and Nonhuman Primate Hepatocyte Growth Factor/AMG 102 Neutralizes Hepatocyte Growth Factor. *Mol. Cancer Ther.* **2010**, *9*, 400–409. [\[CrossRef\]](#)
102. Mok, T.S.; Geater, S.L.; Su, W.C.; Tan, E.H.; Yang, J.C.; Chang, G.C.; Han, M.; Komarnitsky, P.; Payumo, F.; Garrus, J.E.; et al. A Randomized Phase 2 Study Comparing the Combination of Ficlatusumab and Gefitinib with Gefitinib Alone in Asian Patients with Advanced Stage Pulmonary Adenocarcinoma. *J. Thorac. Oncol.* **2016**, *11*, 1736–1744. [\[CrossRef\]](#) [\[PubMed\]](#)
103. Patnaik, A.; Weiss, G.J.; Papadopoulos, K.P.; Hofmeister, C.C.; Tibes, R.; Tolcher, A.; Isaacs, R.; Jac, J.; Han, M.; Payumo, F.C.; et al. Phase I ficlatuzumab monotherapy or with erlotinib for refractory advanced solid tumours and multiple myeloma. *Br. J. Cancer* **2014**, *111*, 272–280. [\[CrossRef\]](#) [\[PubMed\]](#)
104. D’Arcangelo, M.; Cappuzzo, F. Focus on the potential role of ficlatuzumab in the treatment of non-small cell lung cancer. *Biol. Targets Ther.* **2013**, *7*, 61. [\[CrossRef\]](#) [\[PubMed\]](#)
105. Du, Z.; Lovly, C.M. Mechanisms of receptor tyrosine kinase activation in cancer. *Mol. Cancer* **2018**, *17*, 58. [\[CrossRef\]](#) [\[PubMed\]](#)
106. Zehir, A.; Benayed, R.; Shah, R.H.; Syed, A.; Middha, S.; Kim, H.R.; Srinivasan, P.; Gao, J.; Chakravarty, D.; Devlin, S.M.; et al. Mutational landscape of metastatic cancer revealed from prospective clinical sequencing of 10,000 patients. *Nat. Med.* **2017**, *23*, 703–713. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Vuong, H.G.; Ho, A.T.N.; Altibi, A.M.A.; Nakazawa, T.; Katoh, R.; Kondo, T. Clinicopathological implications of MET exon 14 mutations in non-small cell lung cancer—A systematic review and meta-analysis. *Lung Cancer* **2018**, *123*, 76–82. [\[CrossRef\]](#)
108. Vigna, E.; Gramaglia, D.; Longati, P.; Bardelli, A.; Comoglio, P.M. Loss of the exon encoding the juxtamembrane domain is essential for the oncogenic activation of TPR-MET. *Oncogene* **1999**, *18*, 4275–4281. [\[CrossRef\]](#)
109. US Food and Drug Administration. *TABRECTA (Capmatinib) Prescribing Information*; US Food and Drug Administration: Silver Spring, MD, USA, 2022.
110. Awad, M.M.; Leonardi, G.C.; Kravets, S.; Dahlberg, S.E.; Drilon, A.; Noonan, S.A.; Camidge, D.R.; Ou, S.-H.I.; Costa, D.B.; Gadgeel, S.M. Impact of MET inhibitors on survival among patients with non-small cell lung cancer harboring MET exon 14 mutations: A retrospective analysis. *Lung Cancer* **2019**, *133*, 96–102. [\[CrossRef\]](#)
111. Lu, S.; Fang, J.; Li, X.; Cao, L.; Zhou, J.; Guo, Q.; Liang, Z.; Cheng, Y.; Jiang, L.; Yang, N.; et al. Phase II study of savolitinib in patients (pts) with pulmonary sarcomatoid carcinoma (PSC) and other types of non-small cell lung cancer (NSCLC) harboring MET exon 14 skipping mutations (METex14+). *J. Clin. Oncol.* **2020**, *38*, 9519. [\[CrossRef\]](#)
112. Engstrom, L.D.; Aranda, R.; Lee, M.; Tovar, E.A.; Essenburg, C.J.; Madaj, Z.; Chiang, H.; Briere, D.; Hallin, J.; Lopez-Casas, P.P.; et al. Glesatinib Exhibits Antitumor Activity in Lung Cancer Models and Patients Harboring MET Exon 14 Mutations and Overcomes Mutation-mediated Resistance to Type I MET Inhibitors in Nonclinical Models. *Clin. Cancer Res.* **2017**, *23*, 6661–6672. [\[CrossRef\]](#)
113. Hu, H.; Mu, Q.; Bao, Z.; Chen, Y.; Liu, Y.; Chen, J.; Wang, K.; Wang, Z.; Nam, Y.; Jiang, B.; et al. Mutational Landscape of Secondary Glioblastoma Guides MET-Targeted Trial in Brain Tumor. *Cell* **2018**, *175*, 1665–1678.e18. [\[CrossRef\]](#)
114. Klempner, S.J.; Borghei, A.; Hakimian, B.; Ali, S.M.; Ou, S.I. Intracranial Activity of Cabozantinib in MET Exon 14-Positive NSCLC with Brain Metastases. *J. Thorac. Oncol.* **2017**, *12*, 152–156. [\[CrossRef\]](#) [\[PubMed\]](#)
115. Moro-Sibilot, D.; Cozic, N.; Perol, M.; Mazieres, J.; Otto, J.; Souquet, P.J.; Bahleda, R.; Wislez, M.; Zalcman, G.; Guibert, S.D.; et al. Crizotinib in c-MET- or ROS1-positive NSCLC: Results of the AcSe phase II trial. *Ann. Oncol.* **2019**, *30*, 1985–1991. [\[CrossRef\]](#) [\[PubMed\]](#)
116. Salgia, R.; Sattler, M.; Scheele, J.; Stroh, C.; Felip, E. The promise of selective MET inhibitors in non-small cell lung cancer with MET exon 14 skipping. *Cancer Treat. Rev.* **2020**, *87*, 102022. [\[CrossRef\]](#)
117. Fujino, T.; Suda, K.; Mitsudomi, T. Lung Cancer with MET exon 14 Skipping Mutation: Genetic Feature, Current Treatments, and Future Challenges. *Lung Cancer* **2021**, *12*, 35–50. [\[CrossRef\]](#) [\[PubMed\]](#)
118. Wang, F.; Liu, Y.; Qiu, W.; Shum, E.; Feng, M.; Zhao, D.; Zheng, D.; Borczuk, A.; Cheng, H.; Halmos, B. Functional Analysis of MET Exon 14 Skipping Alteration in Cancer Invasion and Metastatic Dissemination. *Cancer Res.* **2022**, *82*, 1365–1379. [\[CrossRef\]](#)
119. Rotow, J.K.; Gui, P.; Wu, W.; Raymond, V.M.; Lanman, R.B.; Kaye, F.J.; Peled, N.; Fece de la Cruz, F.; Nades, B.; Corcoran, R.B.; et al. Co-occurring Alterations in the RAS-MAPK Pathway Limit Response to MET Inhibitor Treatment in MET Exon 14 Skipping Mutation-Positive Lung Cancer. *Clin. Cancer Res.* **2020**, *26*, 439–449. [\[CrossRef\]](#)
120. Jamme, P.; Fernandes, M.; Copin, M.C.; Descarpentries, C.; Escande, F.; Morabito, A.; Gregoire, V.; Jamme, M.; Baldacci, S.; Tulasne, D.; et al. Alterations in the PI3K Pathway Drive Resistance to MET Inhibitors in NSCLC Harboring MET Exon 14 Skipping Mutations. *J. Thorac. Oncol.* **2020**, *15*, 741–751. [\[CrossRef\]](#)
121. Schmidt, L.; Duh, F.M.; Chen, F.; Kishida, T.; Glenn, G.; Choyke, P.; Scherer, S.W.; Zhuang, Z.; Lubensky, I.; Dean, M.; et al. Germline and somatic mutations in the tyrosine kinase domain of the MET proto-oncogene in papillary renal carcinomas. *Nat. Genet.* **1997**, *16*, 68–73. [\[CrossRef\]](#)
122. Schmidt, L.; Junker, K.; Nakaigawa, N.; Kinjerski, T.; Weirich, G.; Miller, M.; Lubensky, I.; Neumann, H.P.; Brauch, H.; Decker, J. Novel mutations of the MET proto-oncogene in papillary renal carcinomas. *Oncogene* **1999**, *18*, 2343–2350. [\[CrossRef\]](#) [\[PubMed\]](#)

123. Zhuang, Z.; Park, W.S.; Pack, S.; Schmidt, L.; Vortmeyer, A.O.; Pak, E.; Pham, T.; Weil, R.J.; Candidus, S.; Lubensky, I.A.; et al. Trisomy 7-harboring non-random duplication of the mutant MET allele in hereditary papillary renal carcinomas. *Nat. Genet.* **1998**, *20*, 66–69. [\[CrossRef\]](#)
124. Jeffers, M.; Schmidt, L.; Nakaigawa, N.; Webb, C.P.; Weirich, G.; Kishida, T.; Zbar, B.; Vande Woude, G.F. Activating mutations for the met tyrosine kinase receptor in human cancer. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 11445–11450. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Jeffers, M.; Fiscella, M.; Webb, C.P.; Anver, M.; Koochekpour, S.; Vande Woude, G.F. The mutationally activated Met receptor mediates motility and metastasis. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 14417–14422. [\[CrossRef\]](#) [\[PubMed\]](#)
126. Graveel, C.; Su, Y.; Koeman, J.; Wang, L.-M.; Tessarollo, L.; Fiscella, M.; Birchmeier, C.; Swiatek, P.; Bronson, R.; Vande Woude, G. Activating Met mutations produce unique tumor profiles in mice with selective duplication of the mutant allele. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 17198–17203. [\[CrossRef\]](#)
127. Graveel, C.R.; London, C.A.; Woude, G.F.V. A mouse model of activating Met mutations. *Cell Cycle* **2005**, *4*, 518–520. [\[CrossRef\]](#)
128. Park, W.S.; Dong, S.M.; Kim, S.Y.; Na, E.Y.; Shin, M.S.; Pi, J.H.; Kim, B.J.; Bae, J.H.; Hong, Y.K.; Lee, K.S. Somatic mutations in the kinase domain of the Met/hepatocyte growth factor receptor gene in childhood hepatocellular carcinomas. *Cancer Res.* **1999**, *59*, 307–310.
129. Di Renzo, M.F.; Olivero, M.; Martone, T.; Maffe, A.; Maggiora, P.; De Stefani, A.; Valente, G.; Giordano, S.; Cortesina, G.; Comoglio, P. Somatic mutations of the MET oncogene are selected during metastatic spread of human HNSC carcinomas. *Oncogene* **2000**, *19*, 1547–1555. [\[CrossRef\]](#)
130. Aebbersold, D.M.; Landt, O.; Berthou, S.; Gruber, G.; Beer, K.T.; Greiner, R.H.; Zimmer, Y. Prevalence and clinical impact of Met Y1253D-activating point mutation in radiotherapy-treated squamous cell cancer of the oropharynx. *Oncogene* **2003**, *22*, 8519–8523. [\[CrossRef\]](#)
131. Lee, J.-H.; Han, S.-U.; Cho, H.; Jennings, B.; Gerrard, B.; Dean, M.; Schmidt, L.; Zbar, B.; Vande Woude, G.F. A novel germ line juxtamembrane Met mutation in human gastric cancer. *Oncogene* **2000**, *19*, 4947–4953. [\[CrossRef\]](#)
132. Ghadjari, P.; Blank-Liss, W.; Simcock, M.; Hegyi, I.; Beer, K.T.; Moch, H.; Aebbersold, D.M.; Zimmer, Y. MET Y1253D-activating point mutation and development of distant metastasis in advanced head and neck cancers. *Clin. Exp. Metastasis* **2009**, *26*, 809–815. [\[CrossRef\]](#)
133. Stella, G.M.; Benvenuti, S.; Gramaglia, D.; Scarpa, A.; Tomezzoli, A.; Cassoni, P.; Senetta, R.; Venesio, T.; Pozzi, E.; Bardelli, A. MET mutations in cancers of unknown primary origin (CUPs). *Hum. Mutat.* **2011**, *32*, 44–50. [\[CrossRef\]](#)
134. Neklason, D.W.; Done, M.W.; Sargent, N.R.; Schwartz, A.G.; Anton-Culver, H.; Griffin, C.A.; Ahnen, D.J.; Schildkraut, J.M.; Tomlinson, G.E.; Strong, L.C. Activating mutation in MET oncogene in familial colorectal cancer. *BMC Cancer* **2011**, *11*, 424. [\[CrossRef\]](#)
135. Kong-Beltran, M.; Stamos, J.; Wickramasinghe, D. The Sema domain of Met is necessary for receptor dimerization and activation. *Cancer Cell* **2004**, *6*, 75–84. [\[CrossRef\]](#) [\[PubMed\]](#)
136. Ma, P.C.; Jagadeeswaran, R.; Jagadeesh, S.; Tretiakova, M.S.; Nallasura, V.; Fox, E.A.; Hansen, M.; Schaefer, E.; Naoki, K.; Lader, A. Functional expression and mutations of c-Met and its therapeutic inhibition with SU11274 and small interfering RNA in non-small cell lung cancer. *Cancer Res.* **2005**, *65*, 1479–1488. [\[CrossRef\]](#) [\[PubMed\]](#)
137. de Melo Gagliato, D.; Jardim, D.L.F.; Falchook, G.; Tang, C.; Zinner, R.; Wheler, J.J.; Janku, F.; Subbiah, V.; Piha-Paul, S.A.; Fu, S. Analysis of MET genetic aberrations in patients with breast cancer at MD Anderson Phase I unit. *Clin. Breast Cancer* **2014**, *14*, 468–474. [\[CrossRef\]](#) [\[PubMed\]](#)
138. Sebai, M.; Tulasne, D.; Caputo, S.M.; Verkarre, V.; Fernandes, M.; Guerin, C.; Reinhart, F.; Adams, S.; Maugard, C.; Caron, O.; et al. Novel germline MET pathogenic variants in French patients with papillary renal cell carcinomas type I. *Hum. Mutat.* **2022**, *43*, 316–327. [\[CrossRef\]](#)
139. Verginelli, F.; Pisacane, A.; Gambardella, G.; D'Ambrosio, A.; Candiello, E.; Ferrio, M.; Panero, M.; Casorzo, L.; Benvenuti, S.; Cascardi, E.; et al. Cancer of unknown primary stem-like cells model multi-organ metastasis and unveil liability to MEK inhibition. *Nat. Commun.* **2021**, *12*, 2498. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Wang, R.; Ferrell, L.D.; Faouzi, S.; Maher, J.J.; Bishop, J.M. Activation of the Met receptor by cell attachment induces and sustains hepatocellular carcinomas in transgenic mice. *J. Cell Biol.* **2001**, *153*, 1023–1034. [\[CrossRef\]](#)
141. Jeffers, M.; Rong, S.; Vande Woude, G.F. Hepatocyte growth factor/scatter factor-Met signaling in tumorigenicity and invasion/metastasis. *J. Mol. Med.* **1996**, *74*, 505–513. [\[CrossRef\]](#)
142. Yu, J.; Miehlik, S.; Ebert, M.P.; Hoffmann, J.; Breidert, M.; Alpen, B.; Starzynska, T.; Stolte Prof, M.; Malfertheiner, P.; Bayerdorffer, E. Frequency of TPR-MET rearrangement in patients with gastric carcinoma and in first-degree relatives. *Cancer* **2000**, *88*, 1801–1806. [\[CrossRef\]](#)
143. Stransky, N.; Cerami, E.; Schalm, S.; Kim, J.L.; Lengauer, C. The landscape of kinase fusions in cancer. *Nat. Commun.* **2014**, *5*, 4846. [\[CrossRef\]](#)
144. Bao, Z.S.; Chen, H.M.; Yang, M.Y.; Zhang, C.B.; Yu, K.; Ye, W.L.; Hu, B.Q.; Yan, W.; Zhang, W.; Akers, J.; et al. RNA-seq of 272 gliomas revealed a novel, recurrent PTPRZ1-MET fusion transcript in secondary glioblastomas. *Genome Res.* **2014**, *24*, 1765–1773. [\[CrossRef\]](#) [\[PubMed\]](#)
145. International Cancer Genome Consortium PedBrain Tumor Project. Recurrent MET fusion genes represent a drug target in pediatric glioblastoma. *Nat. Med.* **2016**, *22*, 1314–1320. [\[CrossRef\]](#) [\[PubMed\]](#)

146. Chen, H.M.; Yu, K.; Tang, X.Y.; Bao, Z.S.; Jiang, T.; Fan, X.L.; Chen, X.W.; Su, X.D. Enhanced expression and phosphorylation of the MET oncoprotein by glioma-specific PTPRZ1-MET fusions. *FEBS Lett.* **2015**, *589*, 1437–1443. [\[CrossRef\]](#)
147. Huang, R.; Liu, Y.; Wang, K.; Wang, Z.; Zhang, C.; Zhang, W.; Zhao, Z.; Li, G.; Huang, L.; Chang, Y.; et al. High-sensitive clinical diagnostic method for PTPRZ1-MET and the characteristic protein structure contributing to ligand-independent MET activation. *CNS Neurosci. Ther.* **2021**, *27*, 617–628. [\[CrossRef\]](#) [\[PubMed\]](#)
148. Plenker, D.; Bertrand, M.; de Langen, A.J.; Riedel, R.; Lorenz, C.; Scheel, A.H.; Muller, J.; Bragelmann, J.; Dassler-Plenker, J.; Kobe, C.; et al. Structural Alterations of MET Trigger Response to MET Kinase Inhibition in Lung Adenocarcinoma Patients. *Clin. Cancer Res.* **2018**, *24*, 1337–1343. [\[CrossRef\]](#)
149. Pennacchietti, S.; Michieli, P.; Galluzzo, M.; Mazzone, M.; Giordano, S.; Comoglio, P.M. Hypoxia promotes invasive growth by transcriptional activation of the met protooncogene. *Cancer Cell* **2003**, *3*, 347–361. [\[CrossRef\]](#)
150. De Bacco, F.; Luraghi, P.; Medico, E.; Reato, G.; Girolami, F.; Perera, T.; Gabriele, P.; Comoglio, P.M.; Boccaccio, C. Induction of MET by ionizing radiation and its role in radioresistance and invasive growth of cancer. *JNCI J. Natl. Cancer Inst.* **2011**, *103*, 645–661. [\[CrossRef\]](#)
151. Gambarotta, G.; Pisto, S.; Giordano, S.; Comoglio, P.M.; Santoro, C. Structure and inducible regulation of the human MET promoter. *J. Biol. Chem.* **1994**, *269*, 12852–12857. [\[CrossRef\]](#)
152. Peters, S.; Adjei, A.A. MET: A promising anticancer therapeutic target. *Nat. Rev. Clin. Oncol.* **2012**, *9*, 314–326. [\[CrossRef\]](#)
153. Linklater, E.S.; Tovar, E.A.; Essenburg, C.J.; Turner, L.; Madaj, Z.; Winn, M.E.; Melnik, M.K.; Korkaya, H.; Maroun, C.R.; Christensen, J.G.; et al. Targeting MET and EGFR crosstalk signaling in triple-negative breast cancers. *Oncotarget* **2016**, *7*, 69903–69915. [\[CrossRef\]](#) [\[PubMed\]](#)
154. Jo, M.; Stolz, D.B.; Esplen, J.E.; Dorko, K.; Michalopoulos, G.K.; Strom, S.C. Cross-talk between epidermal growth factor receptor and c-Met signal pathways in transformed cells. *J. Biol. Chem.* **2000**, *275*, 8806–8811. [\[CrossRef\]](#)
155. Bhanumathy, K.K.; Balagopal, A.; Vizeacoumar, F.S.; Vizeacoumar, F.J.; Freywald, A.; Giambra, V. Protein Tyrosine Kinases: Their Roles and Their Targeting in Leukemia. *Cancers* **2021**, *13*, 184. [\[CrossRef\]](#) [\[PubMed\]](#)
156. Toschi, L.; Janne, P.A. Single-agent and combination therapeutic strategies to inhibit hepatocyte growth factor/MET signaling in cancer. *Clin. Cancer Res.* **2008**, *14*, 5941–5946. [\[CrossRef\]](#)
157. Bell, D.W.; Gore, I.; Okimoto, R.A.; Godin-Heymann, N.; Sordella, R.; Mulloy, R.; Sharma, S.V.; Brannigan, B.W.; Mohapatra, G.; Settleman, J. Inherited susceptibility to lung cancer may be associated with the T790M drug resistance mutation in EGFR. *Nat. Genet.* **2005**, *37*, 1315–1316. [\[CrossRef\]](#)
158. Kobayashi, S.; Boggon, T.J.; Dayaram, T.; Janne, P.A.; Kocher, O.; Meyerson, M.; Johnson, B.E.; Eck, M.J.; Tenen, D.G.; Halmos, B. EGFR mutation and resistance of non-small-cell lung cancer to gefitinib. *N. Engl. J. Med.* **2005**, *352*, 786–792. [\[CrossRef\]](#)
159. Chen, L.L.; Trent, J.C.; Wu, E.F.; Fuller, G.N.; Ramdas, L.; Zhang, W.; Raymond, A.K.; Prieto, V.G.; Oyedele, C.O.; Hunt, K.K. A missense mutation in KIT kinase domain 1 correlates with imatinib resistance in gastrointestinal stromal tumors. *Cancer Res.* **2004**, *64*, 5913–5919. [\[CrossRef\]](#) [\[PubMed\]](#)
160. Gorre, M.E.; Mohammed, M.; Ellwood, K.; Hsu, N.; Paquette, R.; Rao, P.N.; Sawyers, C.L. Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. *Science* **2001**, *293*, 876–880. [\[CrossRef\]](#)
161. Chun, S.-Y.; Kwon, Y.-S.; Nam, K.-S.; Kim, S. Lapatinib enhances the cytotoxic effects of doxorubicin in MCF-7 tumorspheres by inhibiting the drug efflux function of ABC transporters. *Biomed. Pharmacother.* **2015**, *72*, 37–43. [\[CrossRef\]](#)
162. Brouwer, K.; Chu, X.; Dahlin, A.; Evers, R.; Fischer, V.; Hillgren, K.; Hoffmaster, K.; Ishikawa, T.; Keppler, D.; Kim, R. Membrane transporters in drug development. *Nature* **2010**, *9*, 215–236.
163. Filomeni, G.; Turella, P.; Dupuis, M.L.; Forini, O.; Ciriolo, M.R.; Cianfriglia, M.; Pezzola, S.; Federici, G.; Caccuri, A.M. 6-(7-Nitro-2, 1, 3-benzoxadiazol-4-ylthio) hexanol, a specific glutathione S-transferase inhibitor, overcomes the multidrug resistance (MDR)-associated protein 1-mediated MDR in small cell lung cancer. *Mol. Cancer Ther.* **2008**, *7*, 371–379. [\[CrossRef\]](#)
164. Rodriguez-Antona, C.; Ingelman-Sundberg, M. Cytochrome P450 pharmacogenetics and cancer. *Oncogene* **2006**, *25*, 1679–1691. [\[CrossRef\]](#) [\[PubMed\]](#)
165. Edwards, D.R.; Handsley, M.M.; Pennington, C.J. The ADAM metalloproteinases. *Mol. Asp. Med.* **2008**, *29*, 258–289. [\[CrossRef\]](#) [\[PubMed\]](#)
166. Vigna, E.; Pacchiana, G.; Mazzone, M.; Chiriaco, C.; Fontani, L.; Basilico, C.; Pennacchietti, S.; Comoglio, P.M. “Active” cancer immunotherapy by anti-Met antibody gene transfer. *Cancer Res.* **2008**, *68*, 9176–9183. [\[CrossRef\]](#) [\[PubMed\]](#)
167. Prat, M.; Crepaldi, T.; Pennacchietti, S.; Bussolino, F.; Comoglio, P.M. Agonistic monoclonal antibodies against the Met receptor dissect the biological responses to HGF. *J. Cell Sci.* **1998**, *111*, 237–247. [\[CrossRef\]](#)
168. Petrelli, A.; Circosta, P.; Granziero, L.; Mazzone, M.; Pisacane, A.; Fenoglio, S.; Comoglio, P.M.; Giordano, S. Ab-induced ectodomain shedding mediates hepatocyte growth factor receptor down-regulation and hampers biological activity. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 5090–5095. [\[CrossRef\]](#)
169. Schelter, F.; Kobuch, J.; Moss, M.L.; Becherer, J.D.; Comoglio, P.M.; Boccaccio, C.; Krüger, A. A disintegrin and metalloproteinase-10 (ADAM-10) mediates DN30 antibody-induced shedding of the met surface receptor. *J. Biol. Chem.* **2010**, *285*, 26335–26340. [\[CrossRef\]](#)
170. Foveau, B.; Ancot, F.; Leroy, C.; Petrelli, A.; Reiss, K.; Vingtdoux, V.; Giordano, S.; Fafeur, V.; Tulasne, D. Down-regulation of the met receptor tyrosine kinase by presenilin-dependent regulated intramembrane proteolysis. *Mol. Biol. Cell* **2009**, *20*, 2495–2507. [\[CrossRef\]](#)



171. Martinelli, I.; Modica, C.; Chiriaco, C.; Basilico, C.; Hughes, J.M.; Corso, S.; Giordano, S.; Comoglio, P.M.; Vigna, E. hOA-DN30: A highly effective humanized single-arm MET antibody inducing remission of 'MET-addicted' cancers. *J. Exp. Clin. Cancer Res.* **2022**, *41*, 112. [[CrossRef](#)] [[PubMed](#)]
172. Bussolino, F.; Di Renzo, M.F.; Ziche, M.; Bocchietto, E.; Olivero, M.; Naldini, L.; Gaudino, G.; Tamagnone, L.; Coffer, A.; Comoglio, P. Hepatocyte growth factor is a potent angiogenic factor which stimulates endothelial cell motility and growth. *J. Cell Biol.* **1992**, *119*, 629–641. [[CrossRef](#)] [[PubMed](#)]
173. Wu, X.; Chen, X.; Zhou, Q.; Li, P.; Yu, B.; Li, J.; Qu, Y.; Yan, J.; Yu, Y.; Yan, M. Hepatocyte growth factor activates tumor stromal fibroblasts to promote tumorigenesis in gastric cancer. *Cancer Lett.* **2013**, *335*, 128–135. [[CrossRef](#)] [[PubMed](#)]
174. Galimi, F.; Cottone, E.; Vigna, E.; Arena, N.; Boccaccio, C.; Giordano, S.; Naldini, L.; Comoglio, P.M. Hepatocyte growth factor is a regulator of monocyte-macrophage function. *J. Immunol.* **2001**, *166*, 1241–1247. [[CrossRef](#)] [[PubMed](#)]
175. Wang, R.; Zhang, J.; Chen, S.; Lu, M.; Luo, X.; Yao, S.; Liu, S.; Qin, Y.; Chen, H. Tumor-associated macrophages provide a suitable microenvironment for non-small lung cancer invasion and progression. *Lung Cancer* **2011**, *74*, 188–196. [[CrossRef](#)]
176. McDermott, U.; Sharma, S.V.; Dowell, L.; Greninger, P.; Montagut, C.; Lamb, J.; Archibald, H.; Raudales, R.; Tam, A.; Lee, D.; et al. Identification of genotype-correlated sensitivity to selective kinase inhibitors by using high-throughput tumor cell line profiling. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 19936–19941. [[CrossRef](#)]
177. Michieli, P.; Mazzone, M.; Basilico, C.; Cavassa, S.; Sottile, A.; Naldini, L.; Comoglio, P.M. Targeting the tumor and its microenvironment by a dual-function decoy Met receptor. *Cancer Cell* **2004**, *6*, 61–73. [[CrossRef](#)]
178. Mazzone, M.; Basilico, C.; Cavassa, S.; Pennacchietti, S.; Risio, M.; Naldini, L.; Comoglio, P.M.; Michieli, P. An uncleavable form of pro-scatter factor suppresses tumor growth and dissemination in mice. *J. Clin. Investig.* **2004**, *114*, 1418–1432. [[CrossRef](#)]
179. Siravegna, G.; Marsoni, S.; Siena, S.; Bardelli, A. Integrating liquid biopsies into the management of cancer. *Nat. Rev. Clin. Oncol.* **2017**, *14*, 531–548. [[CrossRef](#)]
180. Wan, J.; Massie, C.; Garcia-Corbacho, J.; Mouliere, F.; Brenton, J.D.; Caldas, C.; Pacey, S.; Baird, R.; Rosenfeld, N. Liquid biopsies come of age: Towards implementation of circulating tumour DNA. *Nat. Rev. Cancer* **2017**, *17*, 223–238. [[CrossRef](#)]
181. Wagle, N.; Emery, C.; Berger, M.F.; Davis, M.J.; Sawyer, A.; Pochanard, P.; Kehoe, S.M.; Johannessen, C.M.; MacConaill, L.E.; Hahn, W.C. Dissecting therapeutic resistance to RAF inhibition in melanoma by tumor genomic profiling. *J. Clin. Oncol.* **2011**, *29*, 3085. [[CrossRef](#)]
182. Russo, M.; Siravegna, G.; Blaszkowsky, L.S.; Corti, G.; Crisafulli, G.; Ahronian, L.G.; Mussolin, B.; Kwak, E.L.; Buscarino, M.; Lazzari, L. Tumor Heterogeneity and Lesion-Specific Response to Targeted Therapy in Colorectal Cancer Lesion-Specific Response to Therapy in Colorectal Cancer. *Cancer Discov.* **2016**, *6*, 147–153. [[CrossRef](#)]
183. Tabassum, D.P.; Polyak, K. Tumorigenesis: It takes a village. *Nat. Rev. Cancer* **2015**, *15*, 473–483. [[CrossRef](#)]
184. Russo, M.; Pompei, S.; Sogari, A.; Corigliano, M.; Crisafulli, G.; Puliafito, A.; Lamba, S.; Erriquez, J.; Bertotti, A.; Gherardi, M.; et al. A modified fluctuation-test framework characterizes the population dynamics and mutation rate of colorectal cancer persister cells. *Nat. Genet.* **2022**, *54*, 976–984. [[CrossRef](#)] [[PubMed](#)]
185. Cipponi, A.; Goode, D.L.; Bedo, J.; McCabe, M.J.; Pajic, M.; Croucher, D.R.; Rajal, A.G.; Junankar, S.R.; Saunders, D.N.; Lobachevsky, P. MTOR signaling orchestrates stress-induced mutagenesis, facilitating adaptive evolution in cancer. *Science* **2020**, *368*, 1127–1131. [[CrossRef](#)] [[PubMed](#)]
186. Russo, M.; Crisafulli, G.; Sogari, A.; Reilly, N.M.; Arena, S.; Lamba, S.; Bartolini, A.; Amodio, V.; Magri, A.; Novara, L. Adaptive mutability of colorectal cancers in response to targeted therapies. *Science* **2019**, *366*, 1473–1480. [[CrossRef](#)]
187. Chaft, J.E.; Oxnard, G.R.; Sima, C.S.; Kris, M.G.; Miller, V.A.; Riely, G.J. Disease flare after tyrosine kinase inhibitor discontinuation in patients with EGFR-mutant lung cancer and acquired resistance to erlotinib or gefitinib: Implications for clinical trial design. *Clin. Cancer Res.* **2011**, *17*, 6298–6303. [[CrossRef](#)] [[PubMed](#)]
188. Zuniga, R.M.; Torcuator, R.; Jain, R.; Anderson, J.; Doyle, T.; Schultz, L.; Mikkelsen, T. Rebound tumour progression after the cessation of bevacizumab therapy in patients with recurrent high-grade glioma. *J. Neurooncol* **2010**, *99*, 237–242. [[CrossRef](#)]
189. West, H.; Oxnard, G.R.; Doebele, R.C. Acquired resistance to targeted therapies in advanced non-small cell lung cancer: New strategies and new agents. *Am. Soc. Clin. Oncol. Educ. Book* **2013**, *33*, e272–e278. [[CrossRef](#)]
190. Pop, O.; Pirvu, A.; Toffart, A.C.; Moro-Sibilot, D. Disease flare after treatment discontinuation in a patient with EML4-ALK lung cancer and acquired resistance to crizotinib. *J. Thorac. Oncol.* **2012**, *7*, e1–e2. [[CrossRef](#)]
191. Yap, T.A.; Macklin-Doherty, A.; Popat, S. Continuing EGFR inhibition beyond progression in advanced non-small cell lung cancer. *Eur. J. Cancer* **2017**, *70*, 12–21. [[CrossRef](#)]
192. Pupo, E.; Ducano, N.; Lupo, B.; Vigna, E.; Avanzato, D.; Perera, T.; Trusolino, L.; Lanzetti, L.; Comoglio, P.M. Rebound Effects Caused by Withdrawal of MET Kinase Inhibitor Are Quenched by a MET Therapeutic Antibody. *Cancer Res.* **2016**, *76*, 5019–5029. [[CrossRef](#)] [[PubMed](#)]
193. Altintas, D.M.; Cerqua, M.; De Laurentiis, A.; Trusolino, L.; Boccaccio, C.; Comoglio, P.M. An mTOR feedback loop mediates the 'flare' ('rebound') response to MET tyrosine kinase inhibition. *Sci. Rep.* **2023**, *13*, 1378. [[CrossRef](#)]
194. Guertin, D.A.; Sabatini, D.M. Defining the Role of mTOR in Cancer. *Cancer Cell* **2007**, *12*, 9–22. [[CrossRef](#)] [[PubMed](#)]
195. Mamane, Y.; Petroulakis, E.; LeBacquer, O.; Sonenberg, N. mTOR, translation initiation and cancer. *Oncogene* **2006**, *25*, 6416–6422. [[CrossRef](#)]

196. Huber, A.; Bodenmiller, B.; Uotila, A.; Stahl, M.; Wanka, S.; Gerrits, B.; Aebersold, R.; Loewith, R. Characterization of the rapamycin-sensitive phosphoproteome reveals that Sch9 is a central coordinator of protein synthesis. *Genes. Dev.* **2009**, *23*, 1929–1943. [[CrossRef](#)]
197. Thoreen, C.C.; Chantranupong, L.; Keys, H.R.; Wang, T.; Gray, N.S.; Sabatini, D.M. A unifying model for mTORC1-mediated regulation of mRNA translation. *Nature* **2012**, *485*, 109–113. [[CrossRef](#)]
198. Lazaris-Karatzas, A.; Montine, K.S.; Sonenberg, N. Malignant transformation by a eukaryotic initiation factor subunit that binds to mRNA 5' cap. *Nature* **1990**, *345*, 544–547. [[CrossRef](#)]
199. Blagosklonny, M.V. Rapalogs in cancer prevention: Anti-aging or anticancer? *Cancer Biol. Ther.* **2012**, *13*, 1349–1354. [[CrossRef](#)] [[PubMed](#)]
200. Wei, K.; Li, M.; Zöller, M.; Wang, M.; Mehrabi, A.; Hoffmann, K. Targeting c-MET by Tivantinib through synergistic activation of JNK/c-jun pathway in cholangiocarcinoma. *Cell Death Dis.* **2019**, *10*, 231. [[CrossRef](#)]
201. Lee, Y.J.; Han, J.-Y.; Lee, G.K.; Shin, J.; Yun, S.A.; Oh, J.Y.; Lee, S.; Kim, H.T.; Lee, J.S. C-MET overexpression as a resistance biomarker to epidermal growth factor receptor tyrosine kinase inhibitors in EGFR-mutant non-small cell lung cancer. *J. Clin. Oncol.* **2016**, *34*, e20660. [[CrossRef](#)]
202. Danilkovitch-Miagkova, A.; Zbar, B. Dysregulation of Met receptor tyrosine kinase activity in invasive tumors. *J. Clin. Investig.* **2002**, *109*, 863–867. [[CrossRef](#)] [[PubMed](#)]
203. Blumenschein, G.R., Jr.; Mills, G.B.; Gonzalez-Angulo, A.M. Targeting the hepatocyte growth factor–cMET axis in cancer therapy. *J. Clin. Oncol.* **2012**, *30*, 3287. [[CrossRef](#)] [[PubMed](#)]

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