**MetaCHIP User Manual**

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**Introduction**

MetaCHIP  is  implemented  in  Python,  a list of dependencies needs to be installed before running. Details of these dependencies can be found at: <https://github.com/songweizhi/MetaCHIP>.

To  install  MetaCHIP  simply  download  the  package  and  run  the  programs  from  the  command  line  interface. **Full path to a list of dependencies needs to be specified in the config.txt file if they are not in environment variables**; otherwise, keep the config.txt file as it is.

MetaCHIP’s input is the sequence file of a set of genome bins derived from metagenomic data.

The MetaCHIP pipeline contains three scripts: Get\_clusters.py, Best-match.py and Phylogenetic.py.

1. **Get\_clusters.py** clusters input genome bins into sub-groups according to their phylogenetic relationships.
2. **Best-match.py** performs the best-match approach.
3. **Phylogenetic.py** performs the phylogenetic approach.

**Get\_clusters.py**

-i input genome folder

-x file extension

-p output prefix

-dc distance cutoff

-fs leaf name font size

Get\_clusters.pywill cluster input genome bins into sub-groups based on the phylogenetic tree derived from the protein sequences of 43 universal single-copy genes (SCG) used by CheckM [1].

Clustering profile generated in this step **should be manually curated** by comparing it with the SCG tree or taxonomic classifications of the input genomes (if available) prior to the HGT identification step. You can do this by changing the group assignment of input genome bins specified in the first column of [prefix]\_grouping\_g[num].txt file. Or, you can modify clustering sensitivity by re-run this step with a customized distance cutoff (-dc) after had a look at the [prefix]\_grouping\_g[num].png file.

**Example command:**

# default setting

$ python Get\_clusters.py -i human\_gut\_bins -x fna -p human\_gut

# with modified distance cutoff

$ python Get\_clusters.py -i human\_gut\_bins -x fna -p human\_gut -dc 1.2

**Output files:**

1. Clustering results were exported to [prefix]\_grouping\_g[num].txt.
2. SCG trees ([prefix]\_grouping\_g[num]\_tree.jpg) of the input genome bins.
3. A dendrogram ([prefix]\_grouping\_g[num].png) showing the hierarchical clustering of input genome bins.

The **[prefix]\_grouping\_g[num].txt** file will be used as input for running Best-match.py and Phylogenetic.py.

**Best-match.py**

-p output prefix

-g grouping file

-blastall all vs all blast results

-cov coverage cutoff

-al alignment length cutoff

-flk the length of flanking sequences to plot

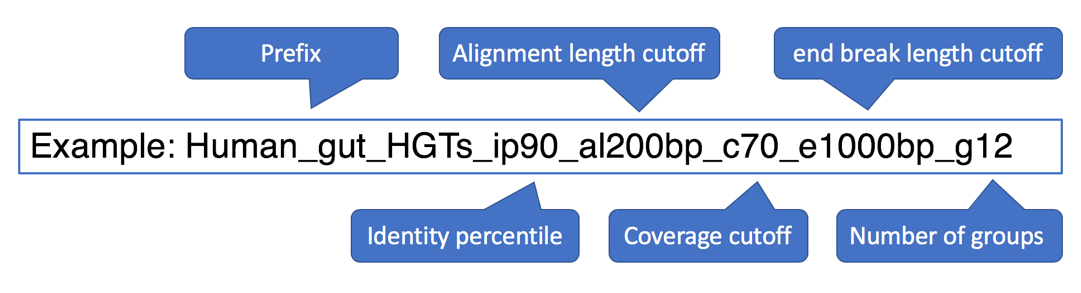
-ip identity percentile cutoff

-eb minimal length to be considered as end break

-tmp keep temporary files

-num\_threads number of threads for running blastn

HGT candidates predicted by the best-match approach, as well as the plots of their flanking regions are exported to a folder with name in the following format:

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**Example command:**

# default setting

$ python Best-match.py -p human\_gut -num\_threads 9

**Output files:**

A list of HGT candidates identified by best-match approach are exported to **HGT\_candidates\_BM.txt**. Their nucleotide and amino acid sequences are exported to HGT\_candidates\_BM\_nc.fasta and HGT\_candidates\_BM\_aa.fasta.

**Phylogenetic.py**

-p output prefix

-g grouping file

-cov coverage cutoff

-al alignment length cutoff

-ip identity percentile

-eb the minimal length to be considered as end break

-a Prokka output

-o orthologs folder

All protein orthologs within the input genomes need to be obtained for the phylogenetic approach, you can get it with GET\_HOMOLOGUES [2]. The input is a bunch of annotation files for input genome bins in Genbank format, which have been generated by Get\_clusters.py (**[prefix]\_gbk\_files**).

**Example command:**

# run GET\_HOMOLOGUES

$ cd human\_gut\_MetaCHIP\_wd

$ get\_homologues.pl -f 70 -t 3 -S 70 -E 1e-05 -C 70 -G -n 16 -d human\_gut\_gbk\_files

# default setting

$ python Phylogenetic.py -p human\_gut

**Output files:**

HGT candidates validated by phylogenetic approach are exported to the same folder as best-match approach.

1. **HGT\_candidates\_PG.txt**: Best-match approach predicted HGTs, with additional information provided by phylogenetic approach.
2. **HGT\_candidates\_PG\_validated.txt**: Phylogenetic approach validated HGTs only.
3. **HGT\_candidates\_PG\_aa.fasta**: Nucleotide sequences of phylogenetic approach validated HGTs.
4. **HGT\_candidates\_PG\_nc.fasta**: Amino acid sequences of phylogenetic approach validated HGTs.

**References**

1. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW: CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome research 2015, 25:1043-1055.
2. Contreras-Moreira B, Vinuesa P: GET\_HOMOLOGUES, a versatile software package for scalable and robust microbial pangenome analysis. Applied & Environmental Microbiology 2013, 79:7696-7701.