Assignment 3: Data Exploration

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## OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

## Directions

1. Change “Student Name” on line 3 (above) with your name.
2. Work through the steps, **creating code and output** that fulfill each instruction.
3. Be sure to **answer the questions** in this assignment document.
4. When you have completed the assignment, **Knit** the text and code into a single PDF file.
5. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai. Add your last name into the file name (e.g., “Salk\_A03\_DataExploration.Rmd”) prior to submission.

The completed exercise is due on <>.

## Set up your R session

1. Check your working directory, load necessary packages (tidyverse), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX\_Neonicotinoids\_Insects\_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON\_NIWO\_Litter\_massdata\_2018-08\_raw.csv). Name these datasets “Neonics” and “Litter”, respectively.

#checking working directory  
getwd()

## [1] "/Users/amritasood/Desktop/CLASSES MEM/Spring 2021/EDA - ENV872/Environmental\_Data\_Analytics\_2021/Assignments"

#set absolute working directory  
setwd("/Users/amritasood/Desktop/CLASSES MEM/Spring 2021/EDA - ENV872/Environmental\_Data\_Analytics\_2021")  
  
#install packages  
library(tidyverse)

#upload datasets  
Neonics <- read.csv("../Data/Raw/ECOTOX\_Neonicotinoids\_Insects\_raw.csv")  
Litter <- read.csv("../Data/Raw/NEON\_NIWO\_Litter\_massdata\_2018-08\_raw.csv")

## Learn about your system

1. The neonicotinoid dataset was collected from the Environmental Protection Agency’s ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicologoy of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: The impact of this class of insecticides on pollinating insects such as honey bees and native bees is a cause for concern. Because they are systemic chemicals absorbed into the plant, neonicotinoids can be present in pollen and nectar, making them toxic to pollinators that feed on them. Neonicotinoids can kill beneficial insects such as honey bees, hoverflies, and parasitic wasps. Thus, we are interested in the ecotoxicology of neonicotinoids on insects such as bees.

1. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: We are interested in studying the litter and woody debris that falls to the ground in forests because it may provide useful insight about the nutrient cycle; help us understand the impact of different pesticides or insecticides and there leftover remanants. It is highly useful to gain a better understanding of nutrient availability and habitat information for different microbes and organisms.

1. How is litter and woody debris sampled as part of the NEON network? Read the NEON\_Litterfall\_UserGuide.pdf document to learn more. List three pieces of salient information about the sampling methods here:

Answer: Litter and fine woody debris sampling is conducted at terrestrial NEON sites that contain woody vegetation >2m tall. *Spatial Design : One litter trap pair (one elevated trap and one ground trap) is deployed for every 400 m2 plot area, resultng in 1-4 trap pairs per plot. Trap placement within plots may be either targeted or randomized, depending on the vegetation. Locations of tower plots are selected randomly within the 90% flux footprint of the primary and secondary airsheds (and additional areas in close proximity to the airshed, as necessary to accommodate sufficient spacing between plots). So the sampling spatial design was both randomized and targeted.* Temporal Sampling Design: Ground traps are sampled once per year. Target sampling frequency for elevated traps varies by vegetation present at the site, with frequent sampling 1x every 2 weeks) in deciduous forest sites during senescence, and year-round sampling (1x every 1-2 months) at evergreen sites. Sampling occured at different frequencies for different plants/trees. \*Location of sampling was selected randomly within the 90% flux footprint of the primary and secondary airsheds. The available space, plot spacing requirements, and/or the tower airshed size restricts the number of plots that can be sampled for litter.

## Obtain basic summaries of your data (Neonics)

1. What are the dimensions of the dataset?

view(Neonics)  
view(Litter)  
#dimensions of datasets  
  
dim(Neonics)

## [1] 4623 30

dim(Litter)

## [1] 188 19

Length of neonics dataset is 30 and width is 4623. Length of Litter dataset is 19 and width is 188.

1. Using the summary function on the “Effects” column, determine the most common effects that are studied. Why might these effects specifically be of interest?

#summary of the effects column  
common\_effects<-as.factor(Neonics$Effect)  
summary(common\_effects)

## Accumulation Avoidance Behavior Biochemistry   
## 12 102 360 11   
## Cell(s) Development Enzyme(s) Feeding behavior   
## 9 136 62 255   
## Genetics Growth Histology Hormone(s)   
## 82 38 5 1   
## Immunological Intoxication Morphology Mortality   
## 16 12 22 1493   
## Physiology Population Reproduction   
## 7 1803 197

Answer: We are studying the ecotoxicological effect of neonictonoids on insects and these effects such as immnology or genetics or intoxication may be useful in deepening our understanding about how the effects of this insecticide on different insects such as bees.

1. Using the summary function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.

commonly\_studied\_species <- as.factor(Neonics$Species.Common.Name)  
summary(commonly\_studied\_species)

## Honey Bee Parasitic Wasp   
## 667 285   
## Buff Tailed Bumblebee Carniolan Honey Bee   
## 183 152   
## Bumble Bee Italian Honeybee   
## 140 113   
## Japanese Beetle Asian Lady Beetle   
## 94 76   
## Euonymus Scale Wireworm   
## 75 69   
## European Dark Bee Minute Pirate Bug   
## 66 62   
## Asian Citrus Psyllid Parastic Wasp   
## 60 58   
## Colorado Potato Beetle Parasitoid Wasp   
## 57 51   
## Erythrina Gall Wasp Beetle Order   
## 49 47   
## Snout Beetle Family, Weevil Sevenspotted Lady Beetle   
## 47 46   
## True Bug Order Buff-tailed Bumblebee   
## 45 39   
## Aphid Family Cabbage Looper   
## 38 38   
## Sweetpotato Whitefly Braconid Wasp   
## 37 33   
## Cotton Aphid Predatory Mite   
## 33 33   
## Ladybird Beetle Family Parasitoid   
## 30 30   
## Scarab Beetle Spring Tiphia   
## 29 29   
## Thrip Order Ground Beetle Family   
## 29 27   
## Rove Beetle Family Tobacco Aphid   
## 27 27   
## Chalcid Wasp Convergent Lady Beetle   
## 25 25   
## Stingless Bee Spider/Mite Class   
## 25 24   
## Tobacco Flea Beetle Citrus Leafminer   
## 24 23   
## Ladybird Beetle Mason Bee   
## 23 22   
## Mosquito Argentine Ant   
## 22 21   
## Beetle Flatheaded Appletree Borer   
## 21 20   
## Horned Oak Gall Wasp Leaf Beetle Family   
## 20 20   
## Potato Leafhopper Tooth-necked Fungus Beetle   
## 20 20   
## Codling Moth Black-spotted Lady Beetle   
## 19 18   
## Calico Scale Fairyfly Parasitoid   
## 18 18   
## Lady Beetle Minute Parasitic Wasps   
## 18 18   
## Mirid Bug Mulberry Pyralid   
## 18 18   
## Silkworm Vedalia Beetle   
## 18 18   
## Araneoid Spider Order Bee Order   
## 17 17   
## Egg Parasitoid Insect Class   
## 17 17   
## Moth And Butterfly Order Oystershell Scale Parasitoid   
## 17 17   
## Hemlock Woolly Adelgid Lady Beetle Hemlock Wooly Adelgid   
## 16 16   
## Mite Onion Thrip   
## 16 16   
## Western Flower Thrips Corn Earworm   
## 15 14   
## Green Peach Aphid House Fly   
## 14 14   
## Ox Beetle Red Scale Parasite   
## 14 14   
## Spined Soldier Bug Armoured Scale Family   
## 14 13   
## Diamondback Moth Eulophid Wasp   
## 13 13   
## Monarch Butterfly Predatory Bug   
## 13 13   
## Yellow Fever Mosquito Braconid Parasitoid   
## 13 12   
## Common Thrip Eastern Subterranean Termite   
## 12 12   
## Jassid Mite Order   
## 12 12   
## Pea Aphid Pond Wolf Spider   
## 12 12   
## Spotless Ladybird Beetle Glasshouse Potato Wasp   
## 11 10   
## Lacewing Southern House Mosquito   
## 10 10   
## Two Spotted Lady Beetle Ant Family   
## 10 9   
## Apple Maggot (Other)   
## 9 670

Answer: These species all belong to same clas/family. They are all flower visiting insects. Neonicotinoid has a a toxicological effect on bees specifically and maybe some of these other insects listed above.

1. Concentrations are always a numeric value. What is the class of Conc.1..Author. in the dataset, and why is it not numeric?

class(Neonics$Conc.1..Author.)

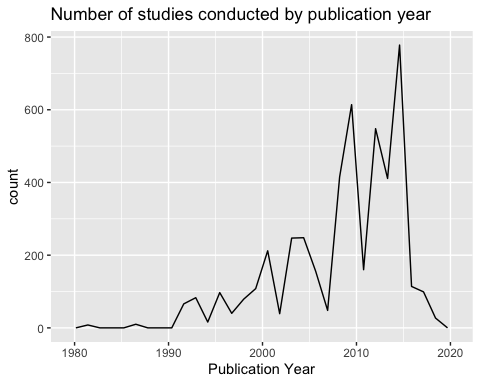
## [1] "character"

Answer: Concnentrations are always numeric however, in this dataset Conc.1.Author has other non numeric charcaters mentioneds such as “~” which makes R read it as a character instead of numeric.

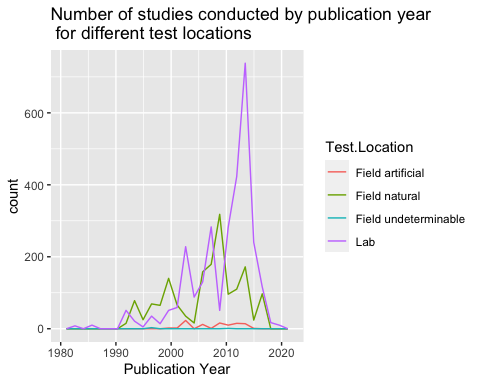
## Explore your data graphically (Neonics)

1. Using geom\_freqpoly, generate a plot of the number of studies conducted by publication year.

ggplot(Neonics) +  
 geom\_freqpoly(aes(x = Publication.Year), bins = 30) + ggtitle("Number of studies conducted by publication year") + labs(x = "Publication Year")

 10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

ggplot(Neonics) +  
 geom\_freqpoly(aes(x = Publication.Year, color= Test.Location), bins = 25)+ ggtitle("Number of studies conducted by publication year \n for different test locations") + labs(x = "Publication Year")

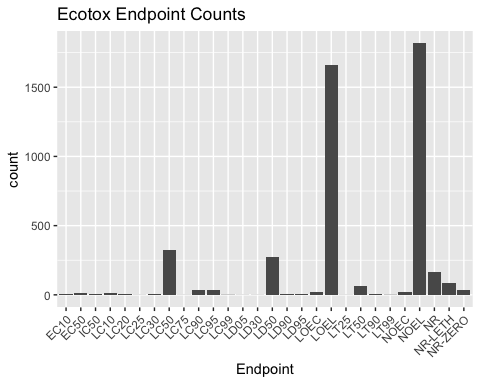


Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: Lab is the most common test location. The second most common is Field Natural.

1. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX\_CodeAppendix for more information.

ggplot(Neonics, aes(x = Endpoint)) +  
 geom\_bar()+ theme(axis.text.x = element\_text(angle = 45, hjust =1) )+ ggtitle("Ecotox Endpoint Counts")



Answer: NOEL (No-observable-effect-level) and LOEL (Lowest-observable-effect-level)are the 2 most common endpoints. LOEL is defined as Lowest-observable-effect-level: lowest dose (concentration) producing effects that were significantly different (as reported by authors) from responses of controls. NOEL is defined as No-observable-effect-level: highest dose (concentration) producing effects not significantly different from responses of controls according to author’s reported statistical test.

## Explore your data (Litter)

1. Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the unique function, determine which dates litter was sampled in August 2018.

class(Litter$collectDate)

## [1] "character"

Litter$collectDate <- as.Date(Litter$collectDate, format = "%Y-%m-%d")  
class(Litter$collectDate)

## [1] "Date"

August2018 <- unique(Litter$collectDate)   
August2018

## [1] "2018-08-02" "2018-08-30"

Collect date was a character so I converted it into date. 13. Using the unique function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from unique different from that obtained from summary?

plots\_sampled <- unique(Litter$plotID)   
length(plots\_sampled)

## [1] 12

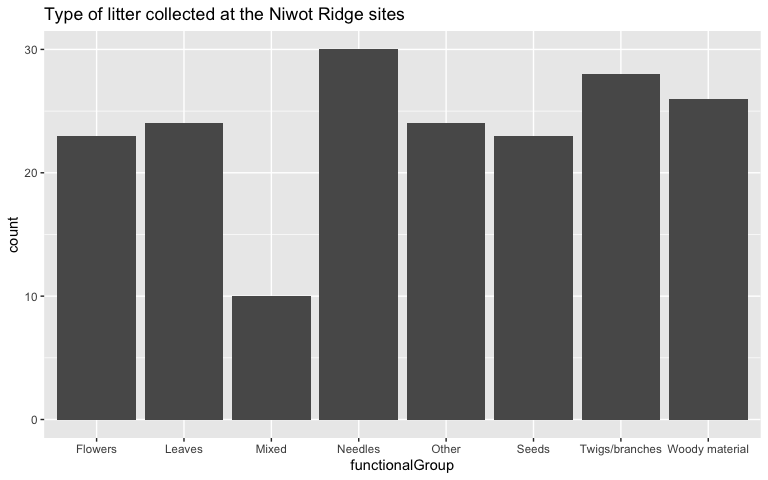
plots <- as.factor(Litter$plotID)  
summary(plots)

## NIWO\_040 NIWO\_041 NIWO\_046 NIWO\_047 NIWO\_051 NIWO\_057 NIWO\_058 NIWO\_061   
## 20 19 18 15 14 8 16 17   
## NIWO\_062 NIWO\_063 NIWO\_064 NIWO\_067   
## 14 14 16 17

Answer: 12 plots were sampled. According to summary there are also 12 plots. The information obtained is same.

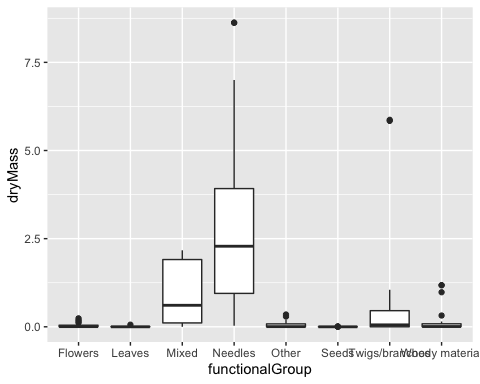
1. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

ggplot(Litter, aes(x = functionalGroup)) +  
 geom\_bar()+ ggtitle("Type of litter collected at the Niwot Ridge sites")



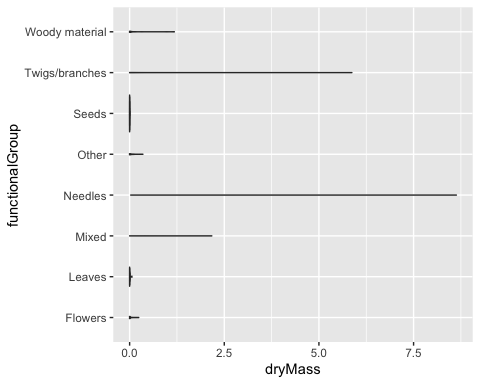
1. Using geom\_boxplot and geom\_violin, create a boxplot and a violin plot of dryMass by functionalGroup.

ggplot(Litter) +  
 geom\_boxplot(aes(x = functionalGroup, y = dryMass))



ggplot(Litter) +  
 geom\_violin(aes(x = dryMass, y = functionalGroup),   
 draw\_quantiles = c(0.25, 0.5, 0.75))

## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):  
## collapsing to unique 'x' values  
  
## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):  
## collapsing to unique 'x' values  
  
## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):  
## collapsing to unique 'x' values



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: Violin plots show probability density of a rotated kernel on each side. In this case boxplot is more effective because the data distribution is more clearly visible and easy to read. The violin plot doesnt provide any useful information. A violin plot is more useful when the data is multimodal but in our case we can visualize the data better with just a boxplot.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles