

# blood

2011 117: 3770-3779  
Prepublished online February 1, 2011;  
doi:10.1182/blood-2010-08-301119

## **In vitro requirement for periostin in B lymphopoiesis**

Basile T. Siewe, Susan L. Kalis, Phong T. Le, Pamela L. Witte, Sangdun Choi, Simon J. Conway, Laurel Druschitz and Katherine L. Knight

---

Updated information and services can be found at:  
<http://bloodjournal.hematologylibrary.org/content/117/14/3770.full.html>

Articles on similar topics can be found in the following Blood collections  
[Hematopoiesis and Stem Cells](#) (2814 articles)  
[Immunobiology](#) (4412 articles)

---

Information about reproducing this article in parts or in its entirety may be found online at:  
[http://bloodjournal.hematologylibrary.org/site/misc/rights.xhtml#repub\\_requests](http://bloodjournal.hematologylibrary.org/site/misc/rights.xhtml#repub_requests)

Information about ordering reprints may be found online at:  
<http://bloodjournal.hematologylibrary.org/site/misc/rights.xhtml#reprints>

Information about subscriptions and ASH membership may be found online at:  
<http://bloodjournal.hematologylibrary.org/site/subscriptions/index.xhtml>



## In vitro requirement for periostin in B lymphopoiesis

\*Basile T. Siewe,<sup>1</sup> \*Susan L. Kalis,<sup>1</sup> Phong T. Le,<sup>1</sup> Pamela L. Witte,<sup>1</sup> Sangdun Choi,<sup>2</sup> Simon J. Conway,<sup>3</sup> Laurel Druschitz,<sup>1</sup> and Katherine L. Knight<sup>1</sup>

<sup>1</sup>Department of Microbiology and Immunology, Loyola University, Chicago, IL; <sup>2</sup>Department of Molecular Science and Technology, Ajou University, Suwon, Korea; and <sup>3</sup>Developmental Biology and Neonatal Medicine Program, Herman B Wells Center for Pediatric Research, Riley Hospital for Children, Indiana University School of Medicine, Indianapolis, IN

**B lymphopoiesis arrests in rabbits by 4 months of age. To identify molecules that contribute to this arrest, cDNA-representational difference analysis on BM stromal cells from young and adult rabbits showed that expression of *Postn* that encodes for the extracellular matrix protein periostin dramatically reduced with age. *Postn*-small interfering RNA OP9 cells lost their capacity to support B-cell development from rabbit or murine BM cells, and reexpression of periostin restored this potential, indicat-**

**ing an in vitro requirement for periostin in B lymphopoiesis. In our system, we determined that periostin deficiency leads to increased cell death and decreased proliferation of B-lineage progenitors. Further, RGD peptide inhibition of periostin/ $\alpha_v\beta_3$  interaction resulted in a marked decrease in B lymphopoiesis in vitro. Microarray analysis of the *Postn*-small interfering RNA OP9 cells showed decreased expression of key B-lymphopoietic factors, including IL-7 and CXCL12. In vivo, unidentified mol-**

**ecule(s) probably compensate periostin loss because *Postn*<sup>-/-</sup> mice had normal numbers of B-cell progenitors in BM. We conclude that the decline in periostin expression in adult rabbit BM does not solely explain the arrest of B lymphopoiesis. However, the interaction of periostin with  $\alpha_v\beta_3$  on lymphoid progenitors probably provides both proliferative and survival signals for cells in the B-cell development pathway. (*Blood*. 2011;117(14):3770-3779)**

### Introduction

The differentiation and proliferation of B-lineage cells in the BM is orchestrated internally by a set of nonredundant transcription factors that are crucial for B-cell development.<sup>1</sup> B-cell progenitors also require fine-tuned interactions with the supportive BM stroma and extracellular matrix (ECM) molecules that create specialized microenvironments, or niches, within the BM. These niches provide both nurturing cell contacts and cytokines that guide all of the hematopoietic cell differentiation programs.<sup>2</sup> The ECM molecules, including periostin, fibronectin, laminin, and proteoglycans, are ligands of integrins that not only control anchorage, spreading, and migration of hematopoietic cells but also induce signal transduction pathways in these cells.<sup>3-5</sup>

Unlike mice and humans, rabbits generate high levels of new B-cell precursors only until a few weeks of age, after which time B lymphopoiesis gradually wanes; by 4 months of age, essentially no new pro-B or pre-B-cell precursors are found in the BM.<sup>6</sup> Lymphoid progenitors are present in the BM of adult rabbits as evidenced by the finding that total BM cells from adults differentiated into B-cell precursors after adoptive transfer into young recipients and also that rabbit lymphoid progenitors differentiated into B- and T-cell precursors when cocultured with OP9 or OP9-DL1 stromal cells.<sup>7</sup> These data suggested to us that the suppression of B lymphopoiesis in the BM of adult rabbits might be mediated through age-related changes in the BM environment. To investigate this hypothesis, a cDNA representational difference analysis (cDNA-RDA) was performed to identify genes down-regulated in BM stromal cells from adults compared with BM

stromal cells from young rabbits. We found several ECM molecules, including periostin, whose expression was down-regulated in adult rabbit stroma. Periostin, a secreted ECM protein of poorly defined function, was originally identified in cells of the mesenchymal lineage, leading to suggestions that it contributed to bone development. Periostin has also been implicated in heart development (reviewed in Hoersch and Andrade-Navarro<sup>8</sup>). Using the murine OP9 stromal cell system, we show in vitro that periostin is required for B lymphopoiesis in mouse and rabbit.

### Methods

#### Animals

*Periostin* (*peri*<sup>lacZ</sup> knockin) knockout mice were as described<sup>9</sup>; all animal experimentation was performed in accordance with guidelines from the National Institutes of Health, and protocols were approved by the respective Institutional Animal Care and Use Committees of all participating institutions.

#### Antibodies and reagents

Antibodies used in this study are listed in Table 1. Reagents used are LIVE/DEAD cell stain Kit (Invitrogen); 7-amino-actinomycin D (7-AAD; BD Bioscience); Annexin V (BD Bioscience); CFSE (Invitrogen); RGD-peptide cyclo (Arg-Gly-Asp-DPhe-Lys; PCI-3661-PI; Peptide International); RGD-peptide cyclo (Arg-Gly-Asp-DPhe-Lys; PCI-3661-PI; Peptide International).

Submitted August 11, 2010; accepted January 15, 2011. Prepublished online as *Blood* First Edition paper, February 1, 2011; DOI 10.1182/blood-2010-08-301119.

\*B.T.S. and S.L.K. contributed equally to this study.

The online version of this article contains a data supplement.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

© 2011 by The American Society of Hematology

**Table 1. Antibodies used in this study**

Anti-CD79a (BD clone HM47)	Anti-ms-Ter119 (clone Ter-119)
Biotin-labeled Anti-rb-IgG (clone C101-167; BD Bioscience)	Anti-ms-CD49b (clone DX5) Anti-ms-B220 (clone RA3-6B2) Anti-ms-CD3 (clone RA3-6B2) Anti-ms-CD8 (H35-17.2) Anti-ms-Gr-1 (clone RB6-8C5) Anti-ms-CD11b (clone M1/70) Anti-ms-Sca1 (clone D7) Anti-ms-c-Kit (clone ACK2) Anti-ms-CD51/CD61 (clone LM609) Anti-ms-IgM (clone R6-8Cs) Anti-ms-CD43 (clone R2/60) Anti-ms-Cd19 (clone 1D3) Anti-ms-periostin (gift of Dr LanBo Chen, Harvard University) Anti-6xHis (clone G-18) Anti-actin (clone AC-15) Rabbit IgG anti-ms-Cxcl12 (polyclonal) Anti-ms-CD29 (clone KM16) Anti-ms-MHC-I (clone 24-1-4-8)

All antibodies were purchased from eBioscience, Inc unless otherwise stated.  
rb indicates rabbit; and ms = mouse.

**cDNA-RDA**

Tissue culture flasks were seeded with  $25 \times 10^6$  nucleated BM cells from newborn and 1-year-old rabbits in 5 mL of  $\alpha$ -MEM. Nonadherent cells were removed after 24-48 hours, and stromal cells were passaged after reaching ~ 90%-100% confluence. On the third passage, RNA was isolated with the use of CsCl (Sigma-Aldrich) gradient centrifugation, and cDNA-RDA was performed.<sup>10</sup> First difference products were cloned into pGEMT (Promega) and identified by National Center for Biotechnology Information BLASTn.

**End point, semiquantitative, and quantitative RT-PCR**

Primers used are listed in Table 2. Total cellular RNA was isolated with an RNeasy kit (QIAGEN) and reverse-transcribed with SuperScript III reverse

**Table 2. PCR Primers used in this study**

	Primers
<b>End point PCR</b>	
Mouse <i>Postn</i>	5'-ATTGGTACCTCCATCCAAGACT-3' 5'-TGTGTTCAGGAGATCTTTCAAAAT-3'
<i>Gapdh</i> control	5'-TGAGTATGTCGTGGAGTCTACTGG-3' 5'-GTGGCAGTGATGGCATGGAC-3'
<b>sqRT-PCR</b>	
Rabbit <i>Postn</i>	5'-CATGAACAGATCTCTTGACACAA-3' 5'-TCCAATGGGTATGTCTGCT-3'
<i>Gapdh</i> control	5'-CATCACTGCCACCCAGAAGA-3' 5'-CCTGCTTCACCACCTTCTTGA-3'
<b>qRT-PCR</b>	
Mouse <i>IL-7</i>	5'-GGCAGGTTGTGCAAAACAGTAACA-3' 5'-AAAGGAAGCCTCTGTTCACATGTGT-3'
Mouse <i>CXCL12</i>	5'-ACACTCCAAGCTGTGCCCTTCAGA-3' 5'-AAGAGGCTCAAGATGTGAGAGGCT-3'
Mouse <i>Postn</i>	5'-AGGCAACAGCTCAGAGTCTTCGT-3' 5'-ATGGAGGGATTCTCTGCTGGCTT-3'
<i>Hgprt</i> control	5'-GTCAACGGGGACATAAAAG-3' 5'-TGGGGCTGTACTGCTTAACC-3'
<i>Postn</i> mutagenesis primers*	5'-GCTGCCCCGAGT <u>CATGCTATA</u> GATCATGTTTATGGCAC-3' 5'-GTGCCATAAACATGATCTATAGG CATGACTGCGGGGCAGC-3'

\*Inserted mutations are underlined.

transcriptase (Invitrogen). For quantitative (q) PCR a total volume of 20  $\mu$ L with ~ 6 ng of cDNA template and 0.5  $\mu$ L of each primer and SYBR Green PCR Master Mix (Applied Biosystems) was analyzed in triplicate. Gene expression was analyzed with an ABI Biosystems 7300 Real-Time PCR Sequence Detector and ABI Prism Sequence Detection Software (Applied Biosystems). Results were normalized to a housekeeping gene control as described in each experiment. For semiquantitative (sq) RT-PCR, the thermocycler was paused after 28, 30, 32, and 34 cycles, and at each time point a 15- $\mu$ L aliquot of the PCR product was removed and resolved on a 5% polyacrylamide gel. Visualization was performed on a Typhoon imaging system (GE Healthcare).

**CFSE labeling**

Cells ( $10^6$ ) were washed, resuspended in 1 mL of  $1 \times$  PBS, and CFSE was added to a final concentration of 5  $\mu$ M. The cell suspension was then mixed and incubated for 10 minutes at 37°C. Cold cell culture medium (1 mL) was added and incubated for 1 minute at room temperature. Cells were washed twice with PBS and counted.

**Stromal cell cultures**

Murine OP9-DL1, OP9-GFP stromal cells<sup>11,12</sup> and OP9-GFP variants transfected with small interfering RNA (siRNA) *GFP* or *Postn* constructs were seeded at ~ 10 000 OP9 cells/well in 96-well plates. After 24 hours, the OP9 cells were overlaid with ~ 10 000 total rabbit or mouse BM cells. Initially, most of the BM cells died; however, after 1 week, stromal-associated cell clusters became visible. Growth of nonadherent cells (B-lineage cells) in the cocultures was monitored by harvesting the cells, straining them (70- $\mu$ m cell strainer; BD Biosciences) to remove loose feeder cells, and quantifying them using trypan blue exclusion. Cells arising from the cocultures were fixed and permeabilized (Cytofix/Perm; BD PharMingen); rabbit cells were stained with anti-CD79a to identify B-lineage cells, and murine cells were stained with anti-B220 and anti-IgM to identify B-lineage cells, and with anti-CD3 to identify T-lineage cells. Stained cells were examined by flow cytometry with the use of FACSCanto (BD Biosciences); only cells that fell within a lymphocyte forward scatter versus side scatter gate were analyzed by FlowJo software (Version 7.2.2; TreeStar Inc). Cultures were routinely supplemented with IL-7 (10 ng/mL).

**RGD-peptide inhibition**

BM cocultured on OP9 cells were supplemented with RGD peptides at the concentrations stated. After 7 days in culture, inhibition of lymphopoiesis was assayed by cell enumeration with the use of trypan blue and by flow cytometry for analysis of differentiation.

**Murine hematopoietic stem cells (Lin<sup>-</sup>Sca1<sup>+</sup>c-Kit<sup>+</sup>) and B-cell progenitors**

BM cells were stained with a lineage cocktail (anti-Ter119, anti-CD49b, anti-B220, anti-CD3, anti-CD8; anti-Gr-1, and anti-CD11b), anti-Sca1 and anti-c-Kit to identify Lin<sup>-</sup>Sca1<sup>+</sup>c-Kit<sup>+</sup> (LSK) cells. B-cell progenitors were identified as follows: CLPs were LSK-IL-7R<sup>+</sup>, pre-pro-B cells were B220<sup>+</sup>c-Kit<sup>-</sup>CD19<sup>-</sup>, pro-B cells were B220<sup>+</sup>CD43<sup>+</sup>CD19<sup>+</sup>, and pre-B cells were B220<sup>+</sup>CD43<sup>-</sup>CD19<sup>+</sup>. To identify cells expressing the  $\alpha_v\beta_3$  receptor, cells were stained as above and also with anti-CD51/CD61 Ab. This monoclonal Ab (mAb) binds to a conformational epitope, resulting from the posttranslational association of the  $\alpha_v$  and  $\beta_3$  subunits. For coculture experiments, LSK  $\alpha_v\beta_3$ <sup>+</sup> cells (3000-4000 cells/well) underwent FACS directly onto OP9 or OP9-DL1 cells.

**siRNA transfection of OP9 stromal cells**

OP9-GFP cells were transfected with *GFP* or murine *Postn* siRNA constructs. Two *Postn* siRNAs were designed (siRNAwiz; InvivoGen) with sequences 309 (GATGCCTATTGACCATGTTTA) and 2292 (GGTCTCCAAGGTCAAAAGTT), corresponding to the starting nucleotides, 309 and 2292, respectively. The siRNAs were cloned into psiRNA-h7Skzeo (InvivoGen), and stable siRNA transfectants of OP9-GFP cells

**Table 3. Genes identified by cDNA-RDA of BM stromal cells from young and adult rabbits**

Clone no.	Encoding for	Function
1	Periostin	ECM protein
2	Fibronectin	ECM protein
3	Collagen type I	ECM protein
4	Thrombospondin	ECM protein
5	Ubiquitin C <sub>1</sub> hydrolase	Protein ubiquitination
6	FAT	Tumor suppressor (cell-cell interaction)
7	Intersectin	Clathrin mediated endocytosis
8	Frizzled 4	Wnt signaling

Driver was cDNA from adult rabbit (2 years old); tester was cDNA from newborn rabbit.

were selected in zeocin. The siRNA for GFP was psiRNA-h7SKze (InvivoGen). OP9 transfectants derived from *Postn* and *GFP* siRNAs were cloned by limiting dilution. DNA microarray analysis from one OP9-GFP siRNA clone and one OP9-OSF2 siRNA clone was performed in duplicate with Affymetrix Mouse Genome 430 2.0 Array, which included 45 K gene elements. Data were analyzed by the Affymetrix Microarray Suite Software (Version 5.0). All microarray data are deposited in ArrayExpress under accession no. E-MTAB-519.

#### Site-directed mutagenesis of *Postn* and transfection of OP9-309 cells

Silent mutations were introduced into murine His-tagged-*Postn* by PCR amplification of the pcDNA-*Postn* construct with the use of PCR primers shown in Table 2 (mutations are underlined). The PCR product was digested with *DpnI* restriction enzyme, gel-purified, and chemically transformed into XL1 Blue cells (Stratagene). Insertion of the desired mutations was confirmed by sequence analysis, and the mutated pcDNA-*Postn* construct was used to transfect OP9-309 cells. Stable clones were selected on neomycin (2 mg/mL). Reexpression of periostin was confirmed by staining with anti-ms-periostin and also by both sqRT-PCR and qRT-PCR.

#### Western blot analysis

Total cell lysates from 10<sup>6</sup> OP9-309 and OP9-309 cells transfected with mutated *Postn* were resolved by 7% SDS-PAGE and then transferred to an Immuno-Blot polyvinylidene difluoride membrane (Millipore). The blot was probed with anti-6xHis Ab and anti-actin Ab as a control.

## Results

#### Age-related changes in gene expression in rabbit BM stromal cells

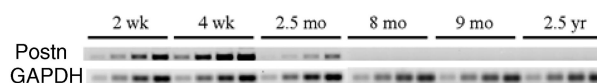
Throughout the lives of mice and humans, B cells are continuously generated in the BM.<sup>13</sup> In contrast, B cells are produced in rabbits only during gestation and until ~ 4 months after birth.<sup>6</sup> Investigating the mechanism by which the observed B lymphopoiesis arrests in adult rabbits, Kalis et al<sup>7</sup> showed that BM stroma from adult rabbits loses its potential to support proliferation and differentiation of HSCs into B-lineage cells. As a result of this finding, we searched for age-related differences in young and adult rabbit BM stromal cells. Stromal cells were cultured from BM of young and adult rabbits in  $\alpha$ -MEM, and RDA was performed with cDNA from adult-derived stromal cells as the driver. We recovered 13 different cDNA clones and identified 8 by nucleotide sequence analysis and National Center for Biotechnology Information BLASTn search (Table 3). By sqRT-PCR we found that one of these molecules, *Postn*, which encodes for the ECM protein periostin, was robustly expressed in the BM of 2- and 4-week-old rabbits (Figure 1). Its

expression began to wane soon thereafter, and by 8 months of age *Postn* expression was undetectable. The decline in expression of *Postn* after ~ 4 weeks of age correlated with the timing of decreased B-cell precursor production in the BM. This initial decline in B lymphopoiesis is followed by a gradual waning of B-cell precursors that continues until 4 months of age, after which time, B lymphopoiesis is arrested.<sup>6</sup> We therefore investigated whether reduced *Postn* expression plays a role in the decline and ultimate arrest of B lymphopoiesis.

#### In vitro inhibition of B lymphopoiesis by *Postn* siRNA

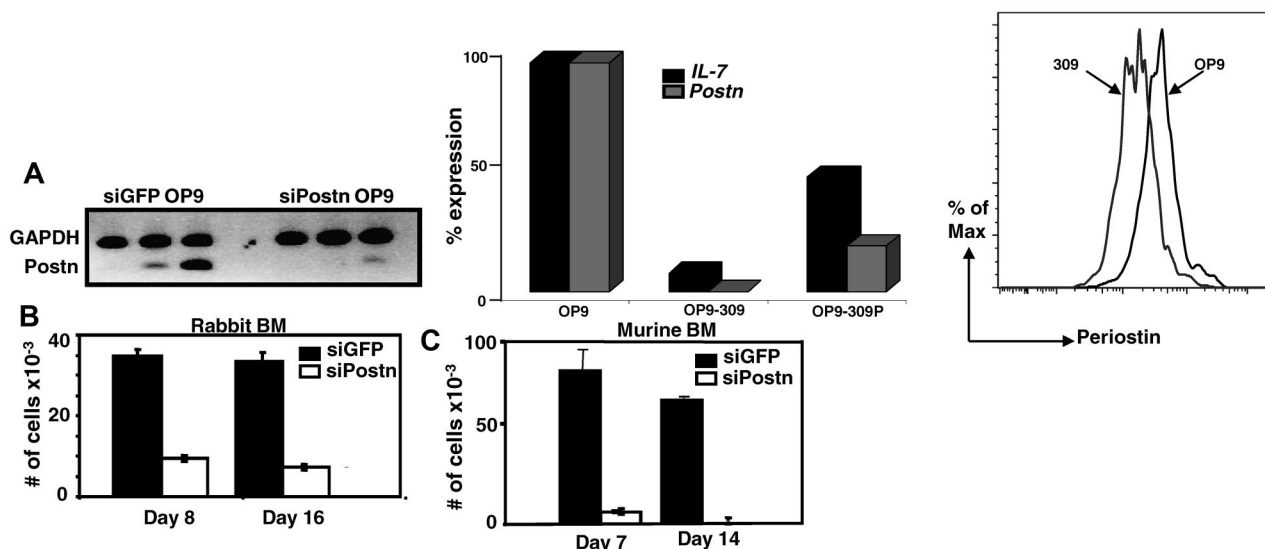
Previous in vitro studies have shown that the ECM protein collagen type I is required for hematopoiesis,<sup>14,15</sup> and, similarly, fibronectin has been shown both in vitro and in vivo to promote proliferation of HSCs.<sup>16</sup> Whether the ECM protein periostin is also important for hematopoiesis, specifically B lymphopoiesis, has not been investigated. We tested this possibility using the OP9 stromal cell coculture system that preferentially promotes B lymphopoiesis from BM cells.<sup>11</sup> We stably transfected OP9-GFP cells with a *Postn* siRNA construct (no. 309) or a control GFP siRNA construct and found considerable reduction in the expression of *Postn* as determined by sqRT-PCR, qRT-PCR, and flow cytometry (Figure 2A left, middle, and right panels, respectively). To determine whether the *Postn* siRNA OP9 (OP9-309) cells retained the capacity to support B-cell development, we overlaid 10 000 total BM cells from adult rabbits on *Postn* siRNA OP9 and on control *GFP* siRNA OP9 cells and found that 1 and 2 weeks later the number of cells that grew out on the *Postn* siRNA OP9 cells was decreased 4- to 5-fold compared with those that grew out on the control *GFP* siRNA OP9 cells (Figure 2B). Similar results were obtained with a second OP9 variant transfected with the *Postn* siRNA construct no. 2292 (data not shown). As shown previously, the cells that grew out from rabbit BM were confirmed as B lineage on the basis of their expression of CD79a (Figure 5C in Kalis et al<sup>7</sup>). We repeated these experiments with the use of murine BM cells and found an 18-fold reduction in the number of cells arising from *Postn* siRNA OP9 cells compared with cells arising from the *GFP* siRNA OP9 cells (Figure 2C). The murine cells that developed on the *GFP* siRNA OP9 cells were confirmed by flow cytometry to be early B-lineage cells (B220<sup>+</sup>CD43<sup>+</sup>; data not shown). These data showed that enforced reduction of *Postn* expression in OP9 cells greatly inhibited their capacity to promote B-cell development from both rabbit and murine BM progenitors, suggesting that periostin is required for B lymphopoiesis in vitro.

To confirm that the siRNA-mediated knockdown of *Postn* in the OP9-309 cells was responsible for the marked reduction of differentiation and proliferation of BM cells into B-lineage cells, we tested if the re-expression of periostin would restore their capacity to support differentiation and proliferation of BM lymphoid progenitors into cells of the B lineage. We generated a construct of *Postn* containing silent mutations in the hybridizing region of the *Postn* siRNA (see "Site-directed mutagenesis of *Postn* and transfection of OP9-309 cells"). We reasoned that transfection of OP9-309 cells with the mutated construct would reestablish



**Figure 1. Semiquantitative RT-PCR of expressed *Postn* in BM from young and adult rabbits.** Aliquots were removed 4 times at 2-cycle intervals during the PCR and analyzed by PAGE. Age of rabbits was from 2 weeks to 2.5 years. *Postn* indicates periostin; *Gapdh* was used as a control. Results are representative of 2 experiments.





**Figure 2.** Rabbit and murine BM cells cocultured with *Postn* siRNA OP9 or GFP siRNA OP9 cells. (A) SqRT-PCR for *Postn* in stable transfectants of GFP siRNA OP9 and *Postn* siRNA cells (left). QRT-PCR for *Postn* and *IL-7* in OP9, OP9-309, and OP9-309P cells; expression levels were normalized to Hgprt housekeeping gene (similar results were obtained in 2 independent experiments; middle). Flow cytometric analysis of OP9 and OP9-309 cells stained with anti-periostin Ab (right). (B-C) Total number of nonadherent cells harvested 1 and 2 weeks after coculture of 10 000 total rabbit (B) and murine (C) BM cells with GFP siRNA OP9 and *Postn* siRNA OP9 (OP9-309) cells. Error bars are SEM from triplicate wells. Results are representative of 3 experiments.

expression of periostin because the mutated construct is impervious to the silencing effects of the *Postn* siRNA. After transfection and selection of cells on neomycin, a stable transfectant was obtained (OP9-309P), which by Western blot analysis (Figure 3A) and qRT-PCR (Figure 2A middle) had restored expression of periostin. To determine whether reexpression of periostin restored the capacity of *Postn* siRNA OP9 cells to support B lymphopoiesis, we cocultured rabbit and mouse BM cells on the OP9-309 and OP9-309P cells, and after 7 days, the cells arising from the cocultures were quantified (Figure 3B) and analyzed by flow cytometry (Figure 3C). We found that reexpression of periostin restored the capacity of OP9-309 cells to support differentiation of BM cells to B-lineage cells. Five-fold (mouse) or 2-fold (rabbit) more cell growth was found in cocultures with the OP9-309P cells than with the OP9-309 cells (Figure 3B), and nearly all cells (88.9%) arising in the mouse BM cocultures with OP9-309P were B220<sup>+</sup> (Figure 3C left). Greater than 60% of the cocultures of rabbit BM cells with OP9-309P were CD79a<sup>+</sup> B-lineage cells; in contrast < 5% of cells growing on OP9-309 were B-lineage cells (Figure 3C). These data indicate that with the use of the OP9 stromal cell system, periostin is required for B lymphopoiesis in vitro.

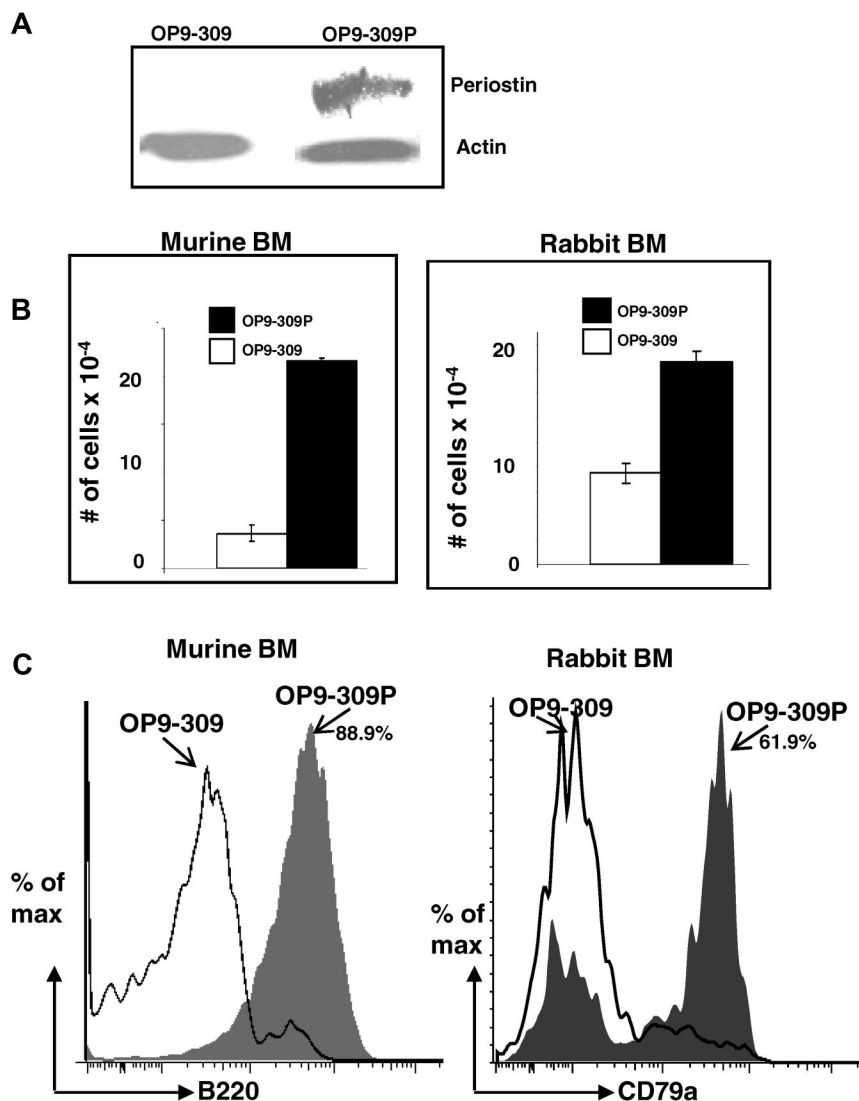
#### Proliferation and survival of BM cells in periostin-deficient OP9-309 cultures

Periostin can act as a ligand for  $\alpha_v\beta_3$  and  $\alpha_v\beta_5$  cellular integrins<sup>17</sup> and  $\alpha_v\beta_3$ -ligand interactions have been shown to activate the Akt/protein kinase B signaling pathway, promoting cell survival, proliferation, and cell motility in vitro.<sup>17-20</sup> We investigated if the reduction in B lymphopoiesis observed in cocultures with the periostin-deficient OP9-309 cells was because of a decrease in proliferation or survival or both of B-lineage cells. Murine CFSE-labeled total BM cells were cultured on OP9 or OP9-309 cells, and cell proliferation, survival, and differentiation were assayed 2-7 days later. After 2 days in culture, the percentage of cells in the lymphocyte gate was similar in both cultures, whereas by day 4 a noticeable reduction in the population from the OP9-309

cultures was observed (Figure 4A) with 43% of the cells being dead and only 22% dead in the OP9 cultures (data not shown). By day 7 we found a dramatic reduction of lymphocyte-sized cells in 309-OP9 cultures (~ 10-fold less) compared with the cells arising in the OP9 cultures (Figure 4A). Further, most of the lymphocytes from the OP9-309 cultures were dead (53%) compared with 6% from the OP9 cultures, showing that most lymphocyte-sized cells from the periostin-deficient OP9-309 cultures were either dead or dying.

Integrin signaling activates the Akt pathway, which promotes cell survival by inhibiting apoptosis.<sup>20,21</sup> We predicted that the observed cell death in the periostin-deficient 309 cells was because of increased apoptosis. By staining with 7-AAD and Annexin V, we found > 60% of the lymphocytes from the OP9-309 cultures were apoptotic (7-AAD<sup>+/+</sup>-Annexin V<sup>+</sup>), compared with only < 25% from the OP9 cultures (Figure 4B). These data indicate that lymphoid cells in the periostin-deficient OP9-309 cultures are dying primarily by apoptosis.

To determine the effect of periostin deficiency on cell proliferation, we analyzed the distribution of CFSE in B220<sup>+</sup> cells arising from both OP9 and OP9-309 cultures by flow cytometry and found that at day 4 essentially all B220<sup>+</sup> cells from both OP9 and OP9-309 cultures had proliferated (Figure 4C left). By day 7, consistent with the high level of apoptosis (Figure 4B), we found almost no B220<sup>+</sup> cells that had proliferated in the OP9-309 cultures (Figure 4C right), indicating that the B220<sup>+</sup> cells seen at day 4 had died. Interestingly, the few B220<sup>+</sup> cells from the OP9-309 cultures had proliferated albeit poorly, signifying that periostin is probably required for the development/expansion of B220<sup>+</sup> cells as well as B220<sup>+</sup> cells. We repeated these experiments with the use of FACS of LSK cells (1000 cells/well) instead of total BM cells to investigate the requirement of periostin in B lymphopoiesis with the use of a homogenous progenitor population. After 7 days in culture, almost no B220<sup>+</sup> cells could be identified in the OP9-309 cultures compared with 23% from the OP9 cultures (Figure 4D arrowheads). However, unlike cultures with total BM cells in



**Figure 3. Restoration of capacity to support B lymphopoiesis by OP9-309 cells transfected with periostin.** (A) Western blot analysis of OP9-309 and OP9-309P cells reexpressing his-tagged periostin. Blot probed with anti-his Ab and anti-actin Ab (control). (B) Cocultures of murine and rabbit BM with OP9-309 and OP9-309P cells; number of total cells in cocultures is indicated; error bars shown as SEM from a range of 2 samples. (C) Flow cytometric analysis of B220<sup>+</sup> (murine) and CD79a<sup>+</sup> (rabbit) cells arising from cocultures (gray histograms indicate cells from OP9-309P cultures; and open histograms, cells from OP9-309 cultures). Similar results were obtained in 3 independent experiments.

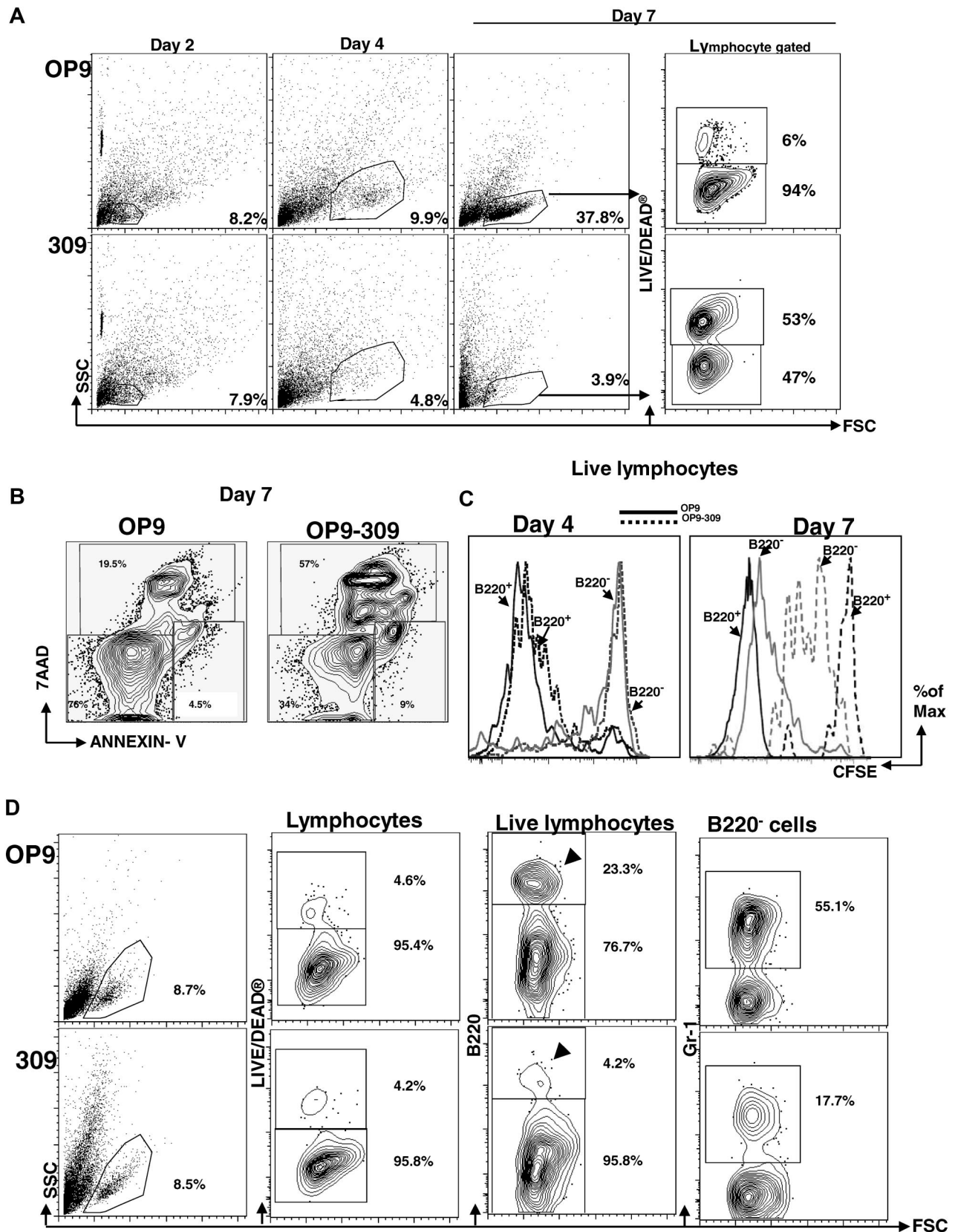
which most cells are dead after 7 days, most of the lymphocyte-sized cells were alive in these cultures. We think that this is because of the self-renewal potential of LSK cells.<sup>22,23</sup> This self-renewal effect is largely undetectable when total BM cells are cultured on OP9-309, because LSK cells make up only 0.05% of total BM cells.<sup>24,25</sup> Most of the cells arising from the OP9-309 cultures were B220<sup>+</sup> and some of these were Gr-1<sup>+</sup> monocytes, although the number of monocytes was 3-fold less (55% vs 18%) than from the OP9 cultures. This finding confirms our previous observation that periostin is probably required for the development/expansion of non-B-lineage cells (Figure 4C). Taken together, the data from the experiments that used LSK cells confirm the data obtained with total BM cells. We conclude that the periostin-deficient OP9-309 cells do not support the proliferation of B-lineage progenitors, indicating a requirement for periostin in B-lineage development in vitro.

#### Receptor for periostin on mouse BM precursors

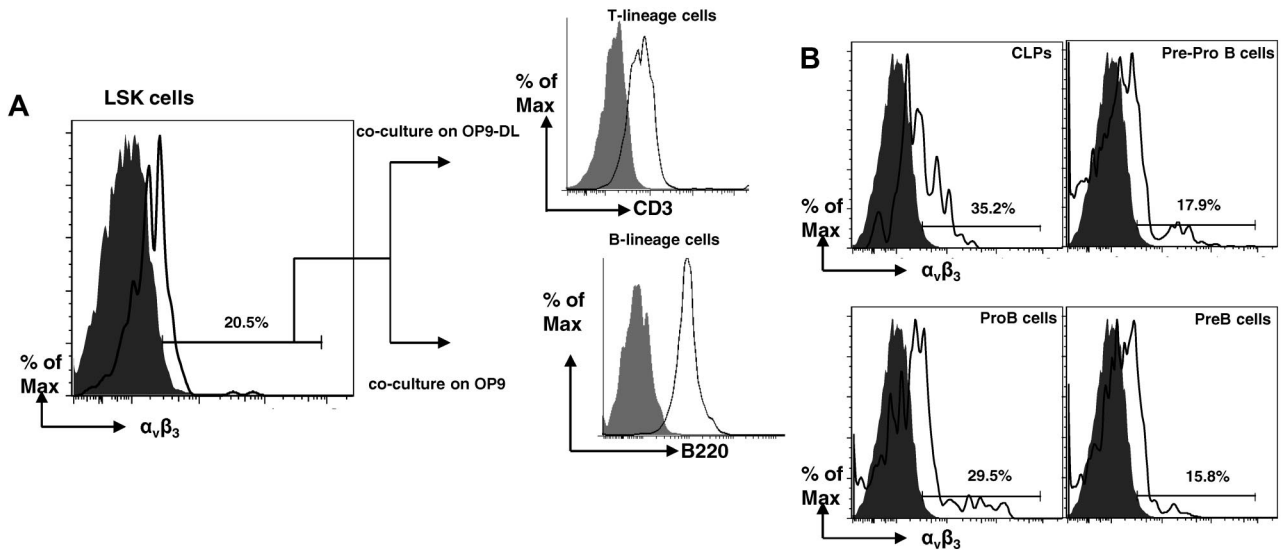
Our results show that periostin is required for B lymphopoiesis in vitro most probably by promoting cell proliferation and survival. Umemoto et al<sup>26</sup> reported that murine HSCs express the  $\beta_3$  chain (CD61) common to multiple integrin receptors,<sup>27</sup> and we investi-

gated if murine LSK and other early progenitors express this  $\beta_3$  complexed with  $\alpha_v$  (CD51), forming a functional periostin receptor  $\alpha_v\beta_3$ .<sup>17</sup> With the use of a mAb against an  $\alpha_v\beta_3$  conformational epitope resulting from the posttranslational association of the  $\alpha_v$  and  $\beta_3$  subunits,<sup>28</sup> we found a population of LSK cells expressing the  $\alpha_v\beta_3$  heterodimer (Figure 5A). To confirm that this LSK  $\alpha_v\beta_3^+$  population contained HSCs, the cells underwent FACS and were cocultured on either OP9 or OP9-DL1 stromal cells. After 7 days, the cells arising from the cocultures were analyzed by flow cytometry, and we identified B- and T-lineage cells arising from the OP9 and OP9-DL1 cultures, respectively (Figure 5A), indicating that the LSK  $\alpha_v\beta_3^+$  cells include HSCs or early lymphoid progenitors or both. We also found that some common lymphoid progenitors (CLPs), pre-pro-B, pro-B, and pre-B cells express  $\alpha_v\beta_3$  (Figure 5B). These results corroborate and extend the findings of Umemoto et al,<sup>26</sup> demonstrating that on murine HSCs, the  $\beta_3$  chain (CD61) is expressed in a heterodimer with an  $\alpha_v$  chain (CD51) forming the receptor for periostin.

The presence of the periostin receptor  $\alpha_v\beta_3$  on lymphoid progenitor cells in BM (Figure 5) suggests that an  $\alpha_v\beta_3$ -mediated interaction between BM progenitor cells and periostin molecules on the OP9 cells is probably required to facilitate proliferation,



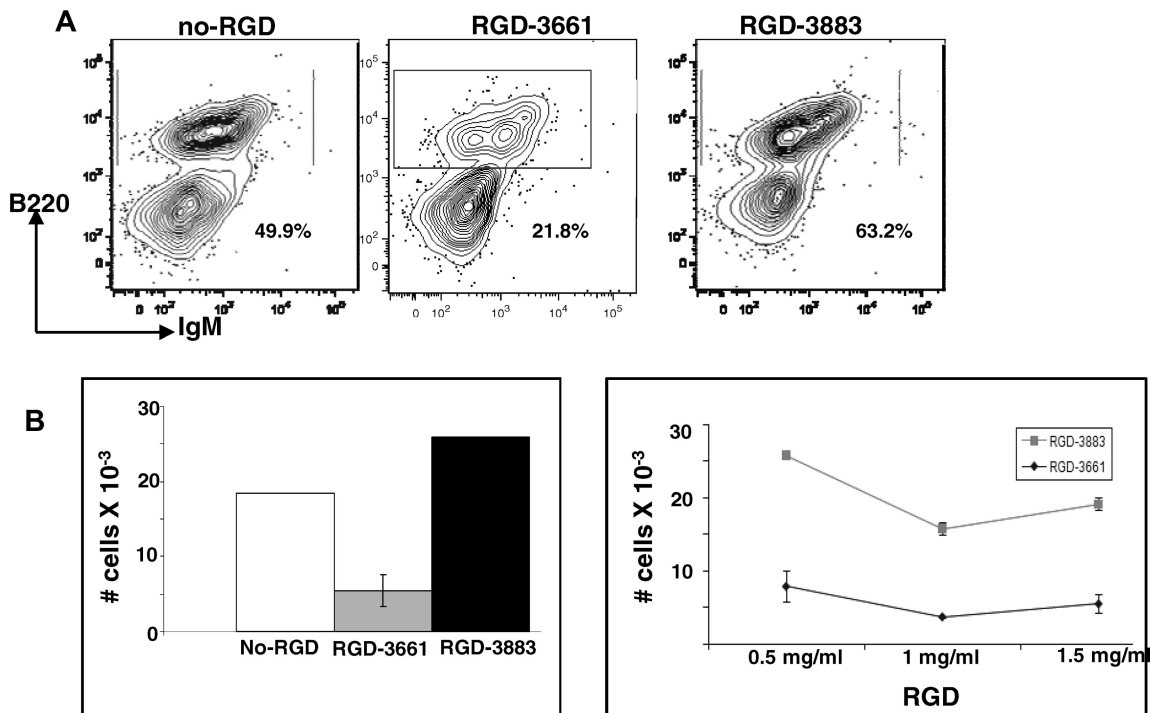
**Figure 4.** Cell death, proliferation, and differentiation of BM cells with periostin-deficient OP9-309 cells. (A) Flow cytometric analysis of cells arising from cocultures of total BM and OP9 or OP9-309 cells after 2, 4, and 7 days. The lymphocyte gates are indicated in percentages; cell death was determined at day 7 (LIVE/DEAD staining). (B) After 7 days in cultures, apoptosis in the lymphocyte populations from OP9 and OP9-309 cultures was assayed by staining cells with 7-AAD and Annexin V and analyzing the cells by flow cytometry. (C) Flow cytometric analysis of CFSE distribution in BM cells after 4 and 7 days in culture (solid gray and black lines indicate cells from OP9 cultures; and dotted black and gray lines, cells from OP9-309 cultures). (D) Flow cytometry of LSK cultured 7 days on either OP9 or OP9-309 cells. The data are representative of 2 experiments.



**Figure 5. Identification and characterization of BM B-cell progenitors expressing the periostin receptor  $\alpha_v\beta_3$ .** (A) Flow cytometry of Lin<sup>−</sup>Sca1<sup>+</sup>c-Kit<sup>+</sup>  $\alpha_v\beta_3$ <sup>+</sup> murine BM cells after 7 days of coculture with OP9 or OP9-DL cells. T-lineage cells identified with anti-CD3; B-lineage cells identified with anti-B220. (B) Flow cytometry of B-cell progenitors for  $\alpha_v\beta_3$ ; gray histograms represent isotype controls. The data are representative of 2 experiments.

survival, and differentiation. Cell-surface integrins, including  $\alpha_v\beta_3$ , interact with ECM proteins by recognizing a specific RGD protein sequence shared by several ECM proteins, including periostin, collagen, and fibronectin.<sup>29–31</sup> We reasoned that, if periostin interacts with  $\alpha_v\beta_3$  on LSK and B-cell progenitors to drive B lymphopoiesis, then inhibiting this interaction would prevent B lymphopoiesis. Because it has been shown that millimolar concentrations of synthetic RGD peptides can inhibit this receptor-ligand interaction in a variety of cells in vitro,<sup>30–34</sup> we cultured mouse BM cells on

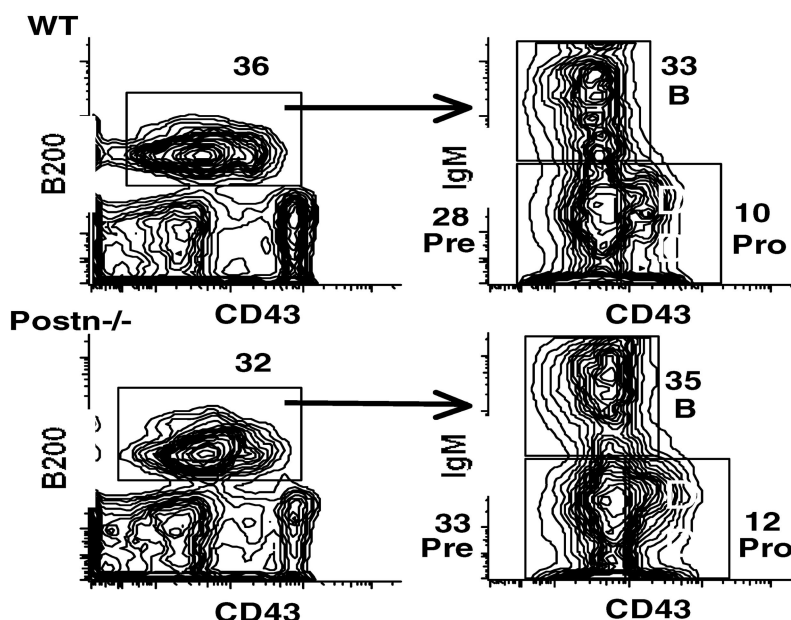
OP9 stromal cells in the presence of an RGD inhibitory peptide (RGD-3661) or a control peptide (RGD-3883) and determined whether B lymphopoiesis was inhibited in vitro. After 7 days, the cells arising from the cocultures were examined for evidence of proliferation and differentiation. The addition of inhibitory RGD-3661 exhibited a ~2-fold reduction in differentiation of BM progenitors to B-lineage cells compared with cells cocultured with control peptide (Figure 6A). This reduction in differentiation was reflected by marked reduction in cell growth at 3 different



**Figure 6. In vitro inhibition of B lymphopoiesis by synthetic RGD peptides.** (A) Flow cytometric analysis of cells arising from cocultures of murine BM and OP9 cells (left) supplemented with inhibitory RGD-3661 peptide (1 mg/mL; middle) or control RGD-3883 peptide (1 mg/mL; right). (B) Quantification of B-lineage cells arising from the cocultures. Error bars are SEM of 3 independent experiments. Left panel depicts cell numbers from experiments with 1 mg/mL RGD peptide; right panel depicts cell numbers from RGD dose-dependent experiments.



**Figure 7. Flow cytometric analysis of BM cells from wild-type (WT) and *Postn*<sup>-/-</sup> mice.** BM cells from 16-week-old mice were analyzed for the expression of B220 and for CD43 (left) and for surface IgM and CD43 within the B220<sup>+</sup> population (right). Numbers indicate the percentages of various lymphocyte stages: pre indicates pre-B stage; and pro, pro-B stage. Similar results were obtained in  $\geq 3$  separate experiments and also with BM of 3-week-old mice; a total of 10 mice were examined.



concentrations of the RGD peptide (Figures 6B). However, supplementation of cocultures with the control RGD-3883 had no inhibitory effect, neither on differentiation (Figure 6A right) nor on the proliferation of B-lineage cells (Figure 6B). We conclude that early BM progenitors need stimulation through their  $\alpha_v\beta_3$  receptor, probably through periostin and perhaps other stromal ligands, for optimal proliferation and differentiation into B-lineage cells.

#### B and T lymphopoiesis in *Postn*<sup>-/-</sup> mice

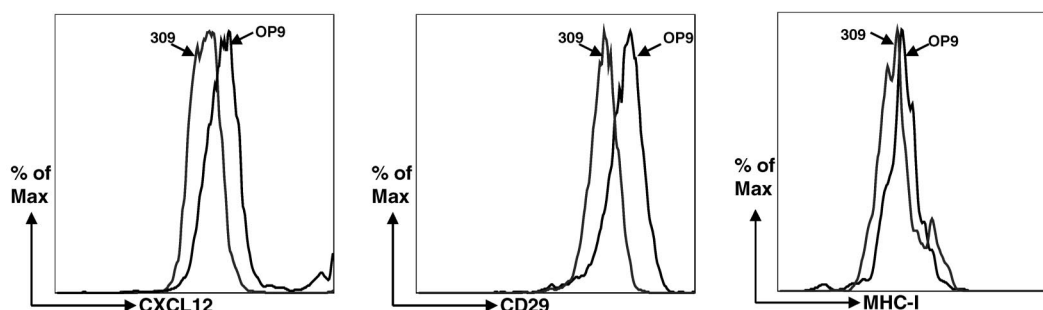
After determining that periostin is required for B lymphopoiesis in vitro, we tested if periostin is required for B or T lymphopoiesis in vivo by examining B- and T-cell populations in *Postn*<sup>-/-</sup> mice.<sup>9</sup> BM, spleen, thymus, and peripheral blood cells from a total of 10 mice aged 3-16 weeks were examined by flow cytometric analysis to determine whether the number and distribution of lymphocytes and lymphoid progenitors were altered. Flow cytometric analysis of these cells from both 3- and 16-week-old mice showed that the percentages of BM B-cell precursor subsets (Figure 7) and the number of splenic B cells (data not shown) was similar to age-matched controls. Although *Postn*<sup>-/-</sup> mice had slightly lower levels of splenic CD4 T cells (73% wild type vs 57% *Postn*<sup>-/-</sup>) and slightly increased levels of splenic CD8 T cells (20% wild type vs. 37% *Postn*<sup>-/-</sup>), the development of thymocyte subsets was also not altered (data not shown). These results indicate

that periostin is not required in vivo for murine B and T lymphopoiesis.

#### Microarray analysis of *Postn* siRNA and *GFP* siRNA OP9 cells

To begin elucidating changes in OP9 cells that occur in response to reduced periostin expression, we compared expression profiles of the *Postn* siRNA OP9 cell line derived from siRNA no.309 and the *GFP* siRNA OP9 cell line by microarray analysis. We found that several critical lymphoid-supporting factors were down-regulated in the *Postn* siRNA OP9 cells, including *IL-7* and *CXCL12* (supplemental Table 1, available on the *Blood* Web site; see the Supplemental Materials link at the top of the online article). Expression of *IL-7* and *CXCL12* were reduced by  $\sim 8$ -fold. Down-regulation of *IL-7* was confirmed by qRT-PCR (Figure 2 middle) and down-regulation of *CXCL12* was confirmed by flow cytometry (Figure 8). By flow cytometry, we also found that CD29 (integrin  $\beta 1$ ) was down-regulated (Figure 8). This was expected from the microarray data because anti-CD29 mAb recognizes the  $\beta 1$  common chain associated with integrin  $\alpha 11$  (ITga11),<sup>35</sup> the expression of which is down-regulated in OP9-309 cells.

The down-regulation of *CXCL12* in the periostin-deficient OP9-309 cells may in part account for the decreased capacity of the OP9-309 cells to support B lymphopoiesis. The reduction in *CXCL12* expression would limit its interaction with CCR4 on



**Figure 8. Flow cytometric analysis of OP9-309 cells for down-regulation of CXCL12 and integrin  $\alpha 11$  (ITga11).** Anti-MHC-1 serves as a control for a molecule not found by microarray analysis to have dysregulated expression.

B-cell progenitors. This interaction is important for adhesion, migration, and retention of the B-cell progenitors in the BM hematopoietic niche as well as for signaling via the Akt pathway. The addition of exogenous IL-7 and CXCL12 to BM:OP9-309 cocultures did not restore B lymphopoiesis (data not shown), indicating that still other molecules are needed to restore the OP9-309 cells. The microarray data provide the basis for continued studies to delineate the role of periostin in B lymphopoiesis.

## Discussion

Throughout the life of mice and humans, HSCs in the BM continuously proliferate and differentiate into B-lineage cells.<sup>13</sup> In other species, including chickens, sheep, and rabbits, B lymphopoiesis occurs primarily during gestation,<sup>36,37</sup> and in rabbits it ceases a few months after birth.<sup>6</sup> The proliferation and differentiation of HSCs to B-lineage cells requires vital signals from the BM microenvironment, including cytokines, growth factors, and adhesion molecules secreted by stromal cells and the ECM.<sup>38-40</sup> Previous investigation of the mechanism responsible for the observed arrest of B lymphopoiesis in rabbits led Kalis et al<sup>7</sup> to suggest that the BM stroma in adult rabbits loses its potential to support B lymphopoiesis. Surprisingly, IL-7, a cytokine critical for B lymphopoiesis, was found to be up-regulated in adult rabbits.<sup>7</sup> A cDNA-RDA assay to compare cultured stromal cells from adult and young rabbits identified the ECM molecule periostin as a molecule that appeared to be down-regulated in adult BM stromal cells. Periostin is a secreted ECM protein of poorly defined function but has been associated with development of bone, teeth, and heart.<sup>41-45</sup> Periostin is also deposited on osteoblasts,<sup>46</sup> which have been shown to support B-lymphocyte commitment and differentiation from HSCs.<sup>47</sup> This finding in combination with a recent report showing the importance of another ECM protein, fibronectin, in lymphopoiesis<sup>48,49</sup> led us to test if periostin could similarly be important for B lymphopoiesis. Our results indicate that enforced reduction of *Postn* expression in OP9 cells (OP9-309) led to the near ablation of proliferation and differentiation of mouse and rabbit BM progenitors to B-lineage cells. We confirmed that the decline in B lymphopoiesis in the OP9-309 cells was the result of reduction of periostin by reexpressing periostin in the OP9-309 cells and showing that the capacity to support differentiation and proliferation of mouse and rabbit BM progenitors into B-lineage cells was regained. These results indicate a novel role for periostin in B lymphopoiesis.

Periostin is the ligand for integrin  $\alpha_v\beta_3$ , and  $\alpha_v\beta_3$ -ligand interactions activate the Akt signaling pathway resulting in cell survival and proliferation.<sup>16,18,19,21</sup> We postulate that in vitro periostin deficiency prevents signaling via the Akt pathway, leading to loss of proliferation and cell death, thereby inhibiting differentiation of BM progenitors into B-lineage cells. Consistent with this idea, we found pronounced apoptosis and almost no proliferation or differentiation of BM progenitors cultured on periostin-deficient OP9 cells. This requirement for periostin in B lymphopoiesis was confirmed with the use of purified LSK cells in coculture experiments. Although cell death was minimal in these cultures, probably because of the self-renewal potential of LSK,<sup>22,23</sup> almost no differentiation of LSK into the B lineage occurred. Further, although cell viability was extremely low, we found almost no live cells in purified pre-pro-B cells from OP9-309 cultures (data not shown).

Additional evidence that BM progenitors interact with periostin was the finding that  $\alpha_v\beta_3$  is expressed on LSK, CLP, pre-pro-B, pro-B, and pre-B cell subsets and also that inhibiting the periostin- $\alpha_v\beta_3$  interactions with RGD peptide inhibited the proliferation and differentiation of BM progenitors into B-lineage cells. On the basis of these data, we conclude that in vitro, periostin- $\alpha_v\beta_3$  interactions are required to drive proliferation and differentiation of BM progenitors into B-lineage cells, most probably by activating the Akt signaling pathway. Because early B-cell precursors reside in the subendosteal region of the BM cavity,<sup>50</sup> we hypothesize that, during aging, disruption of the ECM in the subendosteal region could diminish attachments and survival signals between B-lineage progenitors and the stroma and lead to a decline in the levels of B lymphopoiesis.

We were surprised to find no decline in either B or T lymphopoiesis in young or adult *Postn*<sup>-/-</sup> mice, and, in addition,  $\beta_3$ -deficient mice appear to have normal lymphocyte compartments.<sup>51</sup> It is probable that in vivo other stimuli compensate for the loss of periostin in activating the Akt signaling pathway. Another possibility is that other ECM molecules present in vivo compensate for the loss of periostin expression, because, for example, a possible redundancy of integrin ( $\alpha_4\beta_5$  and  $\alpha_4\beta_7$ ) interactions with fibronectin has been reported (discussed in Bungartz et al<sup>52</sup>). However, we found that the exogenous addition of fibronectin to cocultures of BM cells and *Postn* siRNA OP9 cells was not sufficient to compensate for the loss of periostin (data not shown). Further studies are warranted to explore these possibilities as a means of delineating this novel role of periostin in B lymphopoiesis.

Even though we found a decline in expression of periostin in the BM of adult rabbits, we think this decline is isoform specific. Periostin isoforms have been identified in other species,<sup>8</sup> and using PCR primers for domains 5' of those used in the present study, we found periostin expressed in the BM throughout life (data not shown). Isoforms of rabbit periostin will need to be identified and examined before conclusions can be made about the potential role of specific periostin isoforms in rabbit B lymphopoiesis, in vivo. Taken together the data presented herein highlight the importance of the ECM in B lymphopoiesis, with the identification of a novel in vitro requirement for periostin. This interaction between periostin and its receptor  $\alpha_v\beta_3$  on early BM progenitors is required for proliferation and differentiation of these progenitors into B-lineage cells in vitro.

## Acknowledgments

We thank Haiyan Wang for animal husbandry and Shi-Kang Zhai and Pi-Chen Yam for excellent technical assistance and Patricia Simms of the Loyola University Medical Center Flow Cytometry Facility.

This work was supported by from the National Institutes of Health (grant AI 054508 to K.L.K.), NRF of Korea (grant 2010-0016256 to S.C.), and grants from the Riley Foundation and IU Department of Pediatrics/Cardiology (S.J.C.).

## Authorship

Contribution: B.T.S. and S.L.K. designed research, performed research, analyzed data, and wrote the paper; P.T.L. and P.L.W. designed research and analyzed data; S.C. performed microarray research; S.J.C. provided vital mice and scientific expertise; L.D.

performed research; and K.L.K. designed research, analyzed data, and wrote the paper.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Katherine L. Knight, Department of Microbiology and Immunology, Stritch School of Medicine, Loyola University Chicago, 2160 S First Ave, Maywood, IL 60153; e-mail: [kknight@lumc.edu](mailto:kknight@lumc.edu).

## References

- Northrup DL, Allman D. Transcriptional regulation of early B cell development. *Immunol Res*. 2008; 42(1-3):106-117.
- Bertrand FE, Eckfeldt CE, Fink JR, et al. Microenvironmental influences on human B-cell development. *Immunol Rev*. 2000;175:175-186.
- Adams JC, Watt FM. Regulation of development and differentiation by the extracellular matrix. *Development*. 1993;117(4):1183-1198.
- Hynes RO. Integrins: versatility, modulation, and signaling in cell adhesion. *Cell*. 1992;69(1):11-25.
- Ruoslahti E, Yamaguchi Y. Proteoglycans as modulators of growth factor activities. *Cell*. 1991; 64(5):867-869.
- Jasper PJ, Zhai SK, Kalis SL, Kingzette M, Knight KL. B lymphocyte development in rabbit: progenitor B cells and waning of B lymphopoiesis. *J Immunol*. 2003;171(12):6372-6380.
- Kalis SL, Zhai SK, Yam PC, Witte PL, Knight KL. Suppression of B lymphopoiesis at a lymphoid progenitor stage in adult rabbits. *Int Immunol*. 2007;19(6):801-811.
- Hoersch S, Andrade-Navarro MA. Periostin shows increased evolutionary plasticity in its alternatively spliced region. *BMC Evol Biol*. 2010; 10:30.
- Rios H, Koushik SV, Wang H, et al. periostin null mice exhibit dwarfism, incisor enamel defects, and an early-onset periodontal disease-like phenotype. *Mol Cell Biol*. 2005;25(24):11131-11144.
- Hubank M, Schatz DG. cDNA representational difference analysis: a sensitive and flexible method for identification of differentially expressed genes. *Methods Enzymol*. 1999;303: 325-349.
- Carlyle JR, Michie AM, Furlonger C, et al. Identification of a novel developmental stage marking lineage commitment of progenitor thymocytes. *J Exp Med*. 1997;186(2):173-182.
- Schmitt TM, Zuniga-Pflucker JC. Induction of T cell development from hematopoietic progenitor cells by delta-like-1 in vitro. *Immunity*. 2002;17(6): 749-756.
- Stephan RP, Sanders VM, Witte PL. Stage-specific alterations in murine B lymphopoiesis with age. *Int Immunol*. 1996;8(4):509-518.
- Zuckerman KS, Rhodes RK, Goodrum DD, et al. Inhibition of collagen deposition in the extracellular matrix prevents the establishment of a stroma supportive of hematopoiesis in long-term murine bone marrow cultures. *J Clin Invest*. 1985;75(3): 970-975.
- Sweeney E, Roberts D, Corbo T, Jacenko O. Congenic mice confirm that collagen X is required for proper hematopoietic development. *PLoS One*. 2010;5(3):e9518.
- Yokota T, Oritani K, Mitsui H, et al. Growth-supporting activities of fibronectin on hematopoietic stem/progenitor cells in vitro and in vivo: structural requirement for fibronectin activities of CS1 and cell-binding domains. *Blood*. 1998; 91(9):3263-3272.
- Gillan L, Matei D, Fishman DA, Gerbin CS, Karlan BY, Chang DD. Periostin secreted by epithelial ovarian carcinoma is a ligand for alpha(V) beta(3) and alpha(V)beta(5) integrins and promotes cell motility. *Cancer Res*. 2002;62(18): 5358-5364.
- Yamada KM, Even-Ram S. Integrin regulation of growth factor receptors. *Nat Cell Biol*. 2002;4(4): E75-E76.
- Su CC, Lin YP, Cheng YJ, et al. Phosphatidylinositol 3-kinase/Akt activation by integrin-tumor matrix interaction suppresses Fas-mediated apoptosis in T cells. *J Immunol*. 2007;179(7):4589-4597.
- Byzova TV, Goldman CK, Pampori N, et al. A mechanism for modulation of cellular responses to VEGF: activation of the integrins. *Mol Cell*. 2000;6(4):851-860.
- Wu M, Wang B, Fei J, Santanam N, Blough ER. Important roles of Akt/PKB signaling in the aging process. *Front Biosci (Schol Ed)*. 2010;2:1169-1188.
- Oostendorp RA, Audet J, Miller C, Eaves CJ. Cell division tracking and expansion of hematopoietic long-term repopulating cells. *Leukemia*. 1999; 13(4):499-501.
- Renstrom J, Kroger M, Peschel C, Oostendorp RA. How the niche regulates hematopoietic stem cells. *Chem Biol Interact*. 2010;184(1-2):7-15.
- Ikuta K, Uchida N, Friedman J, Weissman IL. Lymphocyte development from stem cells. *Annu Rev Immunol*. 1992;10:759-783.
- Ikuta K, Weissman IL. Evidence that hematopoietic stem cells express mouse c-kit but do not depend on steel factor for their generation. *Proc Natl Acad Sci U S A*. 1992;89(4):1502-1506.
- Umamoto T, Yamato M, Shiratsuchi Y, et al. CD61 enriches long-term repopulating hematopoietic stem cells. *Biochem Biophys Res Commun*. 2008;365(1):176-182.
- Luo BH, Carman CV, Springer TA. Structural basis of integrin regulation and signaling. *Annu Rev Immunol*. 2007;25:619-647.
- Lee DY, Yeh CR, Chang SF, et al. Integrin-mediated expression of bone formation-related genes in osteoblast-like cells in response to fluid shear stress: roles of extracellular matrix, Shc, and mitogen-activated protein kinase. *J Bone Miner Res*. 2008;23(7):1140-1149.
- Ruoslahti E, Pierschbacher MD. New perspectives in cell adhesion: RGD and integrins. *Science*. 1987;238(4826):491-497.
- Pierschbacher MD, Ruoslahti E. Variants of the cell recognition site of fibronectin that retain attachment-promoting activity. *Proc Natl Acad Sci U S A*. 1984;81(19):5985-5988.
- Pierschbacher MD, Ruoslahti E. Cell attachment activity of fibronectin can be duplicated by small synthetic fragments of the molecule. *Nature*. 1984;309(5963):30-33.
- Gehlsen KR, Argraves WS, Pierschbacher MD, Ruoslahti E. Inhibition of in vitro tumor cell invasion by Arg-Gly-Asp-containing synthetic peptides. *J Cell Biol*. 1988;106(3):925-930.
- Dedhar S, Ruoslahti E, Pierschbacher MD. A cell surface receptor complex for collagen type I recognizes the Arg-Gly-Asp sequence. *J Cell Biol*. 1987;104(3):585-593.
- Hayman EG, Pierschbacher MD, Ruoslahti E. Detachment of cells from culture substrate by soluble fibronectin peptides. *J Cell Biol*. 1985; 100(6):1948-1954.
- Hemler ME. VLA proteins in the integrin family: structures, functions, and their role on leukocytes. *Annu Rev Immunol*. 1990;8:365-400.
- Reynaud CA, Bertocchi B, Dahan A, Weill JC. Formation of the chicken B-cell repertoire: ontogenesis, regulation of Ig gene rearrangement, and diversification by gene conversion. *Adv Immunol*. 1994;57:353-378.
- Weill JC, Reynaud CA. Early B-cell development in chickens, sheep and rabbits. *Curr Opin Immunol*. 1992;4(2):177-180.
- Zuckerman KS, Rhodes RK. The hematopoietic extracellular matrix. *Prog Clin Biol Res*. 1985; 184:257-266.
- Campbell A, Wicha MS, Long M. Extracellular matrix promotes the growth and differentiation of murine hematopoietic cells in vitro. *J Clin Invest*. 1985;75(6):2085-2090.
- Long MW, Briddell R, Walter AW, Bruno E, Hoffman R. Human hematopoietic stem cell adherence to cytokines and matrix molecules. *J Clin Invest*. 1992;90(1):251-255.
- Oshima A, Tanabe H, Yan T, Lowe GN, Glackin CA, Kudo A. A novel mechanism for the regulation of osteoblast differentiation: transcription of periostin, a member of the fasciclin I family, is regulated by the bHLH transcription factor, twist. *J Cell Biochem*. 2002;86(4):792-804.
- Kruzyńska-Freitag A, Machnicki M, Rogers R, Markwald RR, Conway SJ. Periostin (an osteoblast-specific factor) is expressed within the embryonic mouse heart during valve formation. *Mech Dev*. 2001;103(1-2):183-188.
- Kruzyńska-Freitag A, Wang J, Maeda M, et al. Periostin is expressed within the developing teeth at the sites of epithelial-mesenchymal interaction. *Dev Dyn*. 2004;229(4):857-868.
- Suzuki H, Amizuka N, Kii I, et al. Immunohistochemical localization of periostin in tooth and its surrounding tissues in mouse mandibles during development. *Anat Rec A Discov Mol Cell Evol Biol*. 2004;281(2):1264-1275.
- Hamilton DW. Functional role of periostin in development and wound repair: implications for connective tissue disease. *J Cell Commun Signal*. 2008;2(1-2):9-17.
- Takeshita S, Kikuno R, Tezuka K, Amann E. Osteoblast-specific factor 2: cloning of a putative bone adhesion protein with homology with the insect protein fasciclin I. *Biochem J*. 1993;294(Pt 1):271-278.
- Zhu J, Garrett R, Jung Y, et al. Osteoblasts support B-lymphocyte commitment and differentiation from hematopoietic stem cells. *Blood*. 2007; 109(9):3706-3712.
- Oritani K, Kincade PW. Identification of stromal cell products that interact with pre-B cells. *J Cell Biol*. 1996;134(3):771-782.
- van der Loo JC, Xiao X, McMillin D, Hashino K, Kato I, Williams DA. VLA-5 is expressed by mouse and human long-term repopulating hematopoietic cells and mediates adhesion to extracellular matrix protein fibronectin. *J Clin Invest*. 1998;102(5):1051-1061.
- Balduino A, Hurtado SP, Frazao P, et al. Bone marrow subendosteal microenvironment harbours functionally distinct haemosupportive stromal cell populations. *Cell Tissue Res*. 2005; 319(2):255-266.
- Hodivala-Dilke KM, McHugh KP, Tsakiris DA, et al. Beta3-integrin-deficient mice are a model for Glanzmann thrombasthenia showing placental defects and reduced survival. *J Clin Invest*. 1999; 103(2):229-238.
- Bungartz G, Stiller S, Bauer M, et al. Adult murine hematopoiesis can proceed without beta1 and beta7 integrins. *Blood*. 2006;108(6):1857-1864.