"Simple and multiple regression are two of the most-used statistical procedures in biology. Statistical results from both procedures are commonly interpreted as metrics of the degree of relationship between (sometimes multiple) explanatory and response variables. This rough interpretation may generally be satisfactory for simple regressions, i.e., models involving only one explanatory variable. However, this interpretation can lead to confusion for multiple regression, where the coefficients of a multiple regression model measure something subtly but crucially different..."

Morrissey & Ruxton (2018) Multiple Regression Is Not Multiple Regressions: The Meaning of Multiple Regression and the Non-Problem of Collinearity. PTPBIO 10(3)

Timothée Bonnet

May 16, 2019

BDSI / RSB

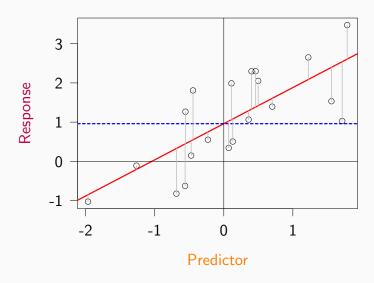
Linear model, reminder

Multiple regression

Interaction

A simple linear model

 $Response = Intercept + Slope \times Predictor + Error$



A multiple linear model

```
\label{eq:Response} \textbf{Response} = \textbf{Intercept} + \textbf{Slope1} \times \textbf{Predictor1} + \textbf{Slope2} \times \textbf{Predictor2} + \\ \textbf{Error}
```

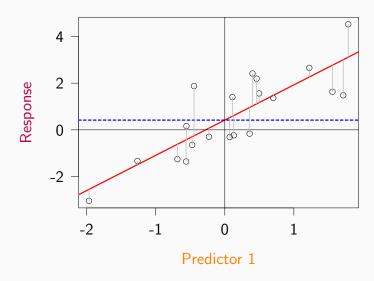
In R:

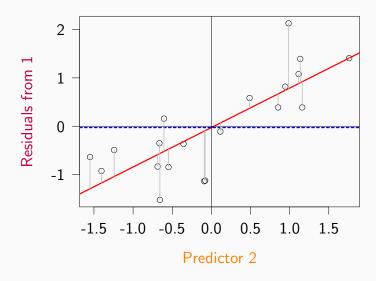
```
lm(response ~ 1 + predictor1 + predictor2, data=data)
```

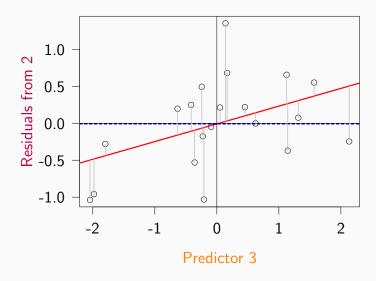
Linear model, reminder

Multiple regression

Interaction







```
m1 <- lm(y ~ x1)
m2 <- lm(m1$residuals ~ x2)
m3 <- lm(m2$residuals ~ x3)
```

But estimates in

are different from

Also what happens with classical ANOVA (aov in R)

```
summary(aov(y ~x1 + x2 + x3))
          Df Sum Sq Mean Sq F value Pr(>F)
x1
          1 40.88 40.88 251.862 3.27e-11 ***
       1 16.40 16.40 101.051 2.55e-08 ***
x2.
     1 0.02 0.02 0.135 0.719
x.3
Residuals 16 2.60 0.16
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
 summary(aov(y ~x2 + x3 + x1))
          Df Sum Sq Mean Sq F value Pr(>F)
x2
          1 46.20 46.20 284.62 1.30e-11 ***
          1 2.46 2.46 15.16 0.00129 **
x3
       1 8.65 8.65 53.27 1.79e-06 ***
x1
Residuals 16 2.60
                    0.16
```

10

In contrast Im() optimizes relationships simultaneously Order does **not** matter:

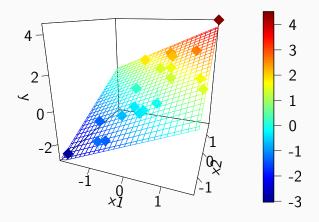
BUT estimates may change with extra covariates

BUT estimates may change with extra covariates

??

- That is a good thing
- Estimates are independent effects, conditional on the other parameters

```
library("plot3D")
fit <-lm(y ~x1 + x2)
# predict values on regular xy grid
grid.lines = 26
x1.pred <- seq(min(x1), max(x1), length.out = grid.lines)
x2.pred <- seq(min(x2), max(x2), length.out = grid.lines)</pre>
x1x2 \leftarrow expand.grid(x1 = x1.pred, x2 = x2.pred)
v.pred <- matrix(predict(fit, newdata = x1x2),</pre>
                 nrow = grid.lines, ncol = grid.lines)
fitpoints <- predict(fit)</pre>
scatter3D(x1, x3, y, pch = 18, cex = 2,
    theta =18, phi = -18, ticktype = "detailed",
    xlab = "x1", ylab = "x2", zlab = "y",
    surf = list(x = x1.pred, y = x2.pred, z = y.pred,
    facets = NA, fit = fitpoints), main = "")
```



```
library("plot3Drg1")
plotrgl(lighting = FALSE, new=TRUE)
```

Exercise

- 1. load jumpingdistance.csv
- 2. Use plots and Im() to test whether mass increases jumping distance

```
jumping <- read.csv(file = "jumpingdistance.csv")</pre>
```

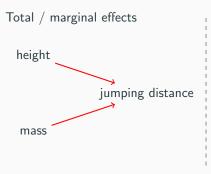
A first approach suggests mass increases jumping distance:

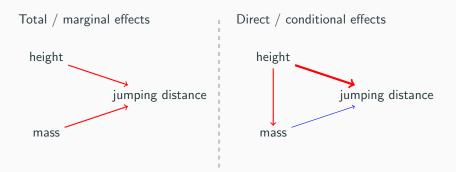
```
summary(lm(jump ~ mass, data=jumping))
plot(mass, jump)
```

But that is incorrect and due to the correlation between mass and height:

```
summary(lm(jump ~ mass + height, data=jumping))
```

The direct (causal) effect of mass is negative, as revealed by a multiple regression





- ullet Marginal effects pprox raw correlations, sum of direct and indirect effects
- Multiple regression estimates direct effects (conditional on other predictors)
 → may reveal causal relationships

Exercise

- 1. Load babies.csv
- 2. What drives change in number of babies born?

```
babies <- read.csv("babies.csv")</pre>
```

Marginal (=total) effects of storks and temperature

```
plot(babies)
summary(lm(babies_born ~ number_of_storks, data=babies))
summary(lm(babies_born ~ mean_temperature, data=babies))
```

But this is a spurious effect due to time covariation. Adding year as a covariate shows that there is no evidence of an effect of storks and temperatures on number of births.

```
summary(lm(babies_born ~ mean_temperature + year, data=babies))
summary(lm(babies_born ~ number_of_storks + year, data=babies))
```

Are more innovative papers less rigorous?

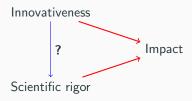
Research question



Should you correct for publication impact?

Are more innovative papers less rigorous?

Research question



Should you correct for publication impact?

Should you include publication impact?

```
Summary(lm(rigor ~ innovativeness + impact))$coefficients

Estimate Std. Error t value Pr(>|t|)

(Intercept) 0.0301366 0.02188752 1.376885 1.688569e-01

innovativeness -0.3150363 0.03051417 -10.324262 8.238502e-24

impact 0.5135830 0.01538756 33.376503 1.361378e-164
```

Apparent negative effect of innovativeness?

```
Summary(lm(rigor ~ innovativeness))$coefficients

Estimate Std. Error t value Pr(>|t|)

(Intercept) 0.04104524 0.03182923 1.289545 1.975073e-01
innovativeness 0.38804729 0.03210760 12.085841 1.758144e-31
```

Apparent positive effect of innovativeness?

Should you include publication impact?

Should you include publication impact?

Data simulated with positive effect of innovativeness on rigor (simulated slope 0.3)

Should you include publication impact?

Data simulated with positive effect of innovativeness on rigor (simulated slope 0.3)

You should NOT correct for impact

Should you include publication impact?

Data simulated with positive effect of innovativeness on rigor (simulated slope 0.3)

You should NOT correct for impact

Rule of Thumb: Do not correct for variables influenced by your predictor outside the causal path of interest

Want more on multiple regression?

Morrissey & Ruxton (2018) Multiple Regression Is Not Multiple Regressions: The Meaning of Multiple Regression and the Non-Problem of Collinearity. PTPBIO 10(3)

dx.doi.org/10.3998/ptpbio.16039257.0010.003

Simple regression, and multiple regression, typically correspond to very different biological questions. The former use regression lines to describe univariate associations. The latter describe the partial, or direct, effects of multiple variables, conditioned on one another. We suspect that the superficial similarity of simple and multiple regression leads to confusion in their interpretation. [...] There is no general sense in which collinearity is a problem. [...] Purported solutions to the perceived problems of collinearity are detrimental to most biological analyses.

Linear model, reminder

Multiple regression

Interaction

Vocabulary warning!

• **correlation**: linear association between two variables "how well does x explain y ?"

Vocabulary warning!

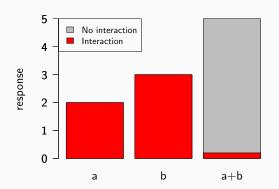
- correlation: linear association between two variables "how well does x explain y?"
- **interaction**: non-additive effect of two or more variables "does the effect of x_1 on y change as a function of x_2 ?". Adds a predictor (or several) to a model.

Vocabulary warning!

- correlation: linear association between two variables "how well does x explain y?"
- **interaction**: non-additive effect of two or more variables "does the effect of x_1 on y change as a function of x_2 ?". Adds a predictor (or several) to a model.

Vocabulary warning!

- correlation: linear association between two variables "how well does x explain y ?"
- interaction: non-additive effect of two or more variables "does the effect
 of x₁ on y change as a function of x₂?". Adds a predictor (or several) to a
 model.



```
lm(y ~ 1 + x1 * x2)
lm(y ~ 1 + x1 + x2 + x1:x2)
```

```
lm(y ~1 + x1 * x2)
 lm(y ~1 + x1 + x2 + x1:x2)
 summary(lm(y^{-1} + x1*x2))
Call:
lm(formula = y ~1 + x1 * x2)
Residuals:
   Min 1Q Median 3Q Max
-1.8719 -0.6777 -0.1086 0.5897 2.3166
Coefficients:
          Estimate Std. Error t value Pr(>|t|)
(Intercept) 1.14098 0.09578 11.913 < 2e-16 ***
x1 -0.49281 0.10834 -4.549 1.58e-05 ***
       0.53434 0.09881 5.408 4.67e-07 ***
x2
```

Why the multiplication sign?

Why the multiplication sign?

x1Xx2 <- x1*x2

Why the multiplication sign?

```
x1Xx2 <- x1*x2
```

```
summary(lm(y~ 1 + x1 + x2 + x1Xx2))

Call:
lm(formula = y~ 1 + x1 + x2 + x1Xx2)
```

Residuals:

```
Min 1Q Median 3Q Max -1.8719 -0.6777 -0.1086 0.5897 2.3166
```

Coefficients:

```
Estimate Std. Error t value Pr(>|t|)

(Intercept) 1.14098 0.09578 11.913 < 2e-16 ***

x1 -0.49281 0.10834 -4.549 1.58e-05 ***

x2 0.53434 0.09881 5.408 4.67e-07 ***
```

Modeling warning!

 DO NOT COMPARE P-VALUES OF TWO MODELS TO TEST FOR AN INTERACTION

- 1. Load the data masssex.csv
- 2. Fit a simple regression explaining movement by mass for each sex separately. Is the relationship different between sexes?
- 3. Fit the multiple regression explaining movement by mass, sex, and mass:sex, using the full dataset. Is the relationship different between sexes?
- 4. Try to understand the discreapancy by plotting the data

1.

```
masssex <- read.csv(file="masssex.csv")</pre>
```

1.

```
masssex <- read.csv(file="masssex.csv")
```

2

```
summary(lm(movement ~ mass, data=masssex[masssex$sex==0,]))
summary(lm(movement ~ mass, data=masssex[masssex$sex==1,]))
```

1.

```
masssex <- read.csv(file="masssex.csv")
```

2

```
summary(lm(movement ~ mass, data=masssex[masssex$sex==0,]))
summary(lm(movement ~ mass, data=masssex[masssex$sex==1,]))
```

3.

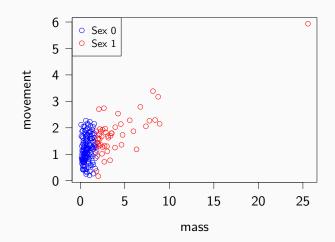
```
summary(lm(movement ~ mass*sex, data=masssex))
```

4. Visualize the problem (result on next slide):

```
plot(masssex[masssex$sex==0,"mass"],masssex[masssex$sex==0,"movement"],
    col="blue", xlim=range(masssex$mass), ylim=range(masssex$movement),
        xlab="mass", ylab="movement")
points(masssex[masssex$sex==1,"mass"],
        masssex[masssex$sex==1,"movement"], col="red")
legend(x="topleft", col=c("blue", "red"),
        legend = c("Sex 0", "Sex 1"), pch=1)
```

The slope of movement on mass is the same for both sexes, but the range of values is much smaller for sex 0, so that there is no power to detect a significant effect. Analysing sexes separately is unsound. You must fit an interaction in a model with both sexes to test for an interaction.

4.



- 1. Load plantsize.csv and plot the data
- 2. Fit an additive model explaining plant size by x and y coordinates

```
plantsize <- read.csv("plantsize.csv")
m0 <- lm(plantsize ~ x_location + y_location, data=plantsize)</pre>
```

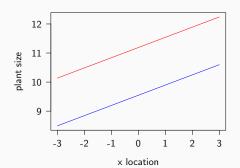
- 1. Load plantsize.csv and plot the data
- 2. Fit an additive model explaining plant size by x and y coordinates
- 3. Create a prediction for plant size as a function of x for two values of y

```
plantsize <- read.csv("plantsize.csv")
m0 <- lm(plantsize ~ x_location + y_location, data=plantsize)</pre>
```

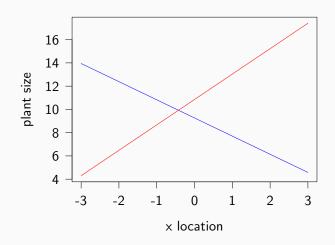
3.1. Predict

3.2 Visualize

```
plot(newdata$x_location[newdata$y_location==-3],
    newdata$prediction[newdata$y_location==-3],
    xlab="x location", ylab="plant size", type="1",
    ylim = range(newdata$prediction), col="blue")
lines(newdata$x_location[newdata$y_location==4],
    newdata$prediction[newdata$y_location==4], col="red")
```



- 1. Load plantsize.csv and plot the data
- 2. Fit an additive model explaining plant size by x and y coordinates
- 3. Create a prediction for plant size as a function of \boldsymbol{x} for two values of \boldsymbol{y} and plot it
- 4. Fit an interaction between x and y coordinates
- 5. Create a new prediction with interaction, and plot it



- 1. Load plantsize.csv and plot the data
- 2. Fit an additive model explaining plant size by x and y coordinates
- 3. Create a prediction for plant size as a function of x for two values of y and plot it
- 4. Fit an interaction between x and y coordinates
- 5. Create a new prediction with interaction, and plot it
- 6. Compare estimates and p-values across models. Do you think x location has an effect or not?

```
library(reshape2)
 matpred <- acast(newdata, x_location~y_location, value.var="prediction")</pre>
 layout (mat = matrix (data = c(1,2), nrow = 1), widths = c(3,1))
 image(t(matpred), col = topo.colors(10), xlab = "x location",
       vlab = "v location")
 par(mar=c(5, 0, 4, 6)+0.1)
 image(matrix(data = seq(min(newdata$prediction),
              max(newdata$prediction), length.out = 10), nrow= 1 ),
        col=topo.colors(10), xaxt = "n", yaxt="n", main="legend")
 axis(side = 4, at = c(0,0.2,0.4,0.6,0.8,1),
labels = round(seq(min(newdata$prediction), max(newdata$prediction),
                   length.out =6),1)
```

