

Statistical inference and linear models

December 6, 2018

- 1 Statistical inference
- 2 Little R-Studio tricks
- 3 t-test, ANOVA, regression: all is one, one is all
- 4 Linear models in details

General approach

1. Scientific question

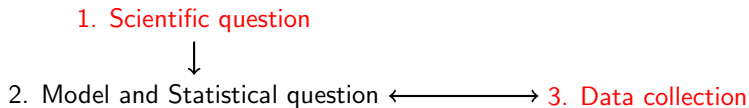
General approach

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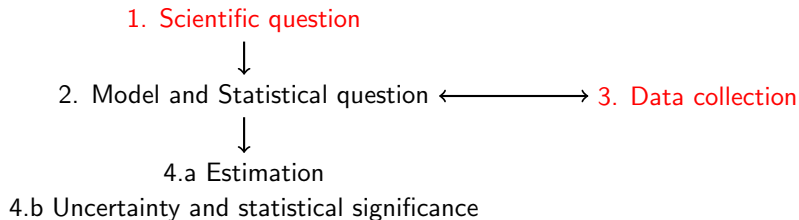


2. Model and Statistical question

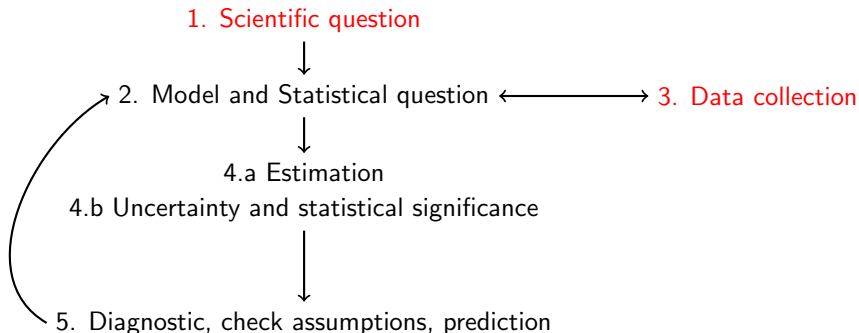
General approach



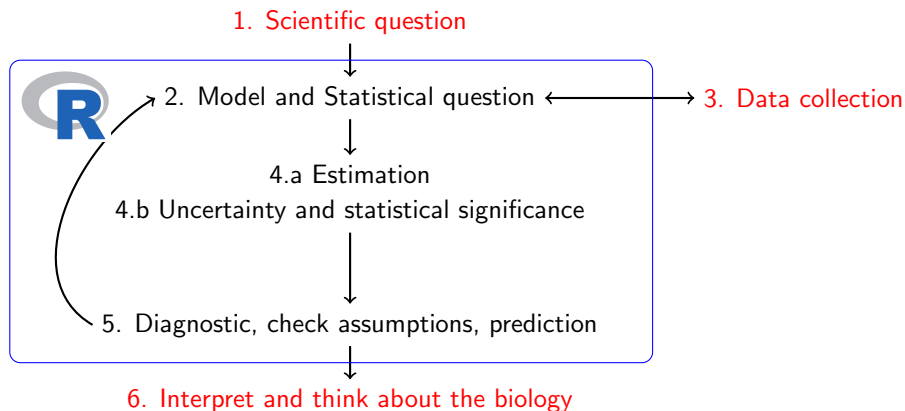
General approach



General approach



General approach



Iris t.test

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```
data("iris")
```

```
str(iris)  
plot(iris)
```

- 1 Scientific question: Are the taxa "setosa" and "versicolor" different species?

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- ① Scientific question: Are the taxa "setosa" and "versicolor" different species?
- ② Model and stat question:
 - ▶ Model:
 - ★ There is an intrinsic/expected sepal length value for a species; an individual value is the sum of this expectation and a random Gaussian deviation.
 - ★ $y_i = \mu_{species_i} + \epsilon_i$ with $\epsilon \sim N(0, \sigma^2)$
 - ★ t-test

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► Model:

- ★ There is an intrinsic/expected sepal length value for a species; an individual value is the sum of this expectation and a random Gaussian deviation.
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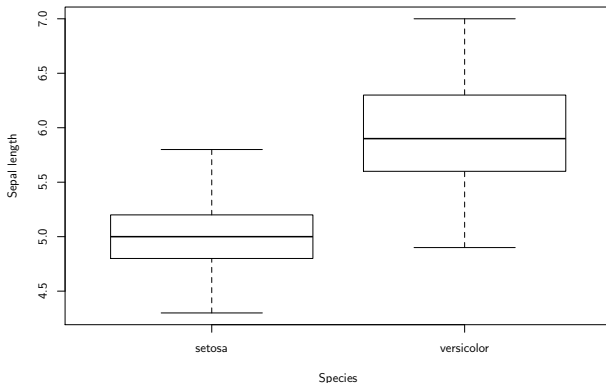
► Statistical question:

- ★ Does sepal length **differ significantly** between the two taxa **in our sample**?
- ★ Is the observed difference between taxa likely if both taxa have the same intrinsic/expected value?

③ Data collection

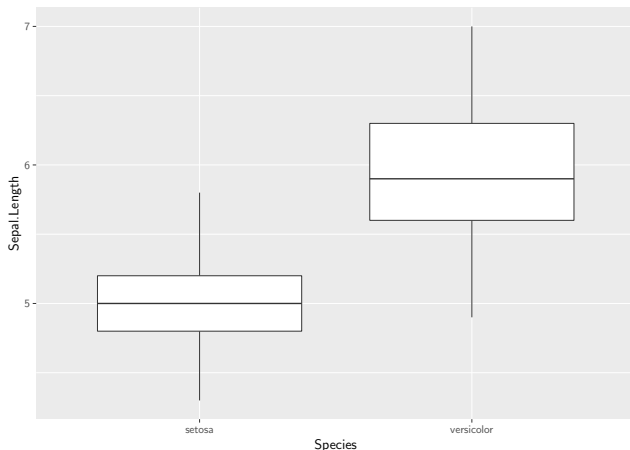
Iris t.test, visualization

```
boxplot(Sepal.Length ~ Species,  
  data = iris[iris$Species %in% c("setosa","versicolor"),],  
  drop = TRUE, ylab="Sepal length", xlab="Species")
```



Iris t.test, visualization

```
library(ggplot2)
ggplot( iris[iris$Species %in% c("setosa","versicolor"),],
  aes(x=Species, y=Sepal.Length)) + geom_boxplot()
```



Iris t.test

One t-test for sepal length between *setosa* and *versicolor*:

```
t.test(x = iris$Sepal.Length[iris$Species == "setosa"],  
       y = iris$Sepal.Length[iris$Species == "versicolor"])
```

Welch Two Sample t-test

data: iris\$Sepal.Length[iris\$Species == "setosa"] and iris\$Sepal.Length[iris\$Species == "versicolor"]

t = -10.521, df = 86.538, p-value < 2.2e-16

alternative hypothesis: true difference in means is not equal to 0

95 percent confidence interval:

-1.1057074 -0.7542926

sample estimates:

mean of x mean of y

5.006 5.936

When do we know it is different?

④ Statistical estimation

► a Estimation

- ★ Cannot know true difference $\mu_{species_1} - \mu_{species_2}$
- ★ Estimated difference = **Mean₁ - Mean₂**
- ★ Estimated difference contains random variation

► b Quantify uncertainty / Statistical significance

- ★ $t = \frac{\text{Mean}_1 - \text{Mean}_2}{\text{Variation}} \frac{\sqrt{\text{Sample Size}}}{\sqrt{2}}$
- ★ We know exactly how t is distributed when $\mu_{species_1} - \mu_{species_2} = 0$
- ★ Hence we know probability of $\geq t$ if $\mu_{species_1} - \mu_{species_2} = 0$ (p -value)
- ★ Can derive confidence interval and standard error

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Less uncertainty with

- **Larger absolute difference**
- **Smaller variability**
- **Larger sample size**

When do we know it is different? Simulations

Simulations in R

```
rnorm()
```

When do we know it is different? Simulations

When do we know it is different? Simulations

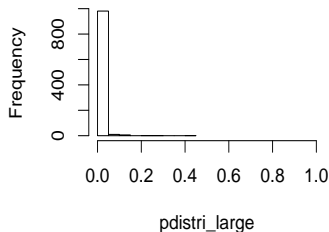
1. Larger absolute difference

```
nbsim <- 1000
pdistri_large <- vector(length = nbsim)
pdistri_small <- vector(length = nbsim)
for (i in 1:nbsim)
{
  x1 <- rnorm(n = 10, mean = 2, sd = 1)
  x2 <- rnorm(n = 10, mean = 4, sd = 1) #large diff
  x3 <- rnorm(n = 10, mean = 2.5, sd = 1) #small diff
  out_large <- t.test(x1, x2)
  out_small <- t.test(x1, x3)
  pdistri_large[i] <- out_large$p.value
  pdistri_small[i] <- out_small$p.value
}
```

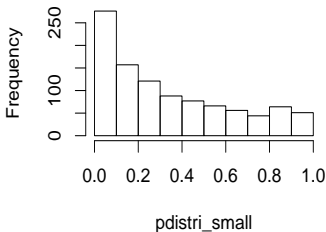
When do we know it is different?

```
par(mfrow=c(1,2), cex=2)
hist(pdistrib_large, xlim=c(0,1),
     main=paste("Prop signif=",mean(pdistrib_large<0.05)))
hist(pdistrib_small, xlim=c(0,1),
     main=paste("Prop signif=",mean(pdistrib_small<0.05)))
```

Prop signif= 0.981



Prop signif= 0.173



```
par(mfrow=c(1,1))
```

When do we know it is different? Try it!

Exercise

Check the effect of **smaller variability** and/or **larger sample size**.

By the way, what are these p-values?

*Probability for a summary statistic to be greater or equal to the observed summary statistic, **when the null-hypothesis of a given statistical model is true.***

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Properties

- Depends on the null-hypothesis (H_0) of a given model with assumptions
- Uniform distribution under H_0 ...
- ... hence $\text{proportion}(\text{significance under } H_0) = \text{significance threshold}$

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- Depends on the null-hypothesis (H_0) of a given model with assumptions
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- ... hence $\text{proportion}(\text{significance under } H_0) = \text{significance threshold}$

NB: Focus on p -value criticized, but common and they are no more evil than other misused statistics!

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Column selection

```
lm1  <- lm(y ~ x1 + x2)
lm2  <- lm(y ~ 1)
lm3  <- lm(y ~ x1*x2)
lm3  <- lm(y ~ x2)
lm4  <- lm(y ~ x1)
...

plot(lm1)
plot(lm2)
...
```

Column selection

You changed the name to be more explicit

```
lmadd <- lm(y ~ x1 + x2)
lmnull <- lm(y ~ 1)
lmff <- lm(y ~ x1*x2)
lmx2 <- lm(y ~ x2)
lmx1 <- lm(y ~ x1)
...

plot(lm1)
plot(lm2)
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```

What a pain to repeat the changes in `plot()`!

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Alt + click

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What if my code was not well aligned??

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...

plot(lm1)
plot(lm2)
...
```

Ctrl + Alt + clicks creates multiple cursors
then Shift + Home and Ctrl + C

Shortcuts

Alt + Shift + K

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A small example

Animal behavior in response to weather

Load data:

```
getwd()  
setwd()
```

```
dat.behav <- read.csv(file = "Data/datbehav.csv") # path to file
```

A small example

Animal behavior in response to weather

Load data:

```
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setwd()
```

```
dat.behav <- read.csv(file = "Data/datbehav.csv") # path to file
```

STEP 1: have a look at your data

```
str(dat.behav)  
summary(dat.behav)  
plot(dat.behav)
```

t-test

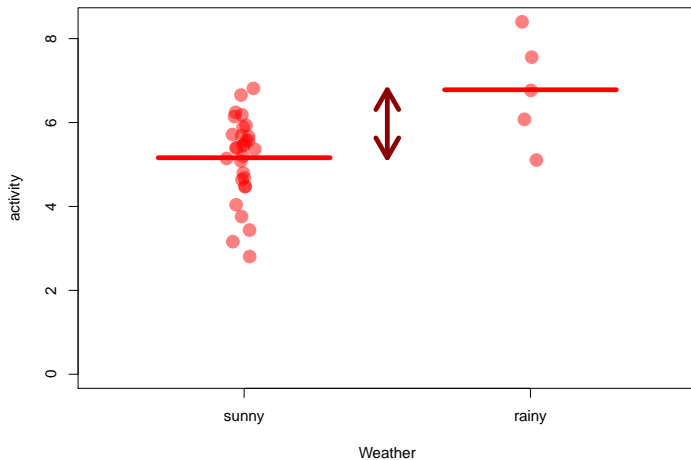
```
fitstudent <- t.test(x = dat.behav$activity[dat.behav$weather=="rainy"],  
                    y = dat.behav$activity[dat.behav$weather=="sunny"],  
                    var.equal = TRUE)  
print(fitstudent)
```

Two Sample t-test

```
data: dat.behav$activity[dat.behav$weather == "rainy"] and dat.behav$activity[dat.behav$weather == "sunny"]  
t = 3.2752, df = 33, p-value = 0.002485  
alternative hypothesis: true difference in means is not equal to 0  
95 percent confidence interval:  
 0.6138373 2.6270325  
sample estimates:  
mean of x mean of y  
6.781476 5.161041
```

t-test, graphically

Difference between means



ANOVA

- 1 What is the fastest way to get row averages in a data-frame?
- 2 Create a function called colVars, like colMeans but for variance
- 3 Create nice plots to visualize iris data (ideally journal-quality)

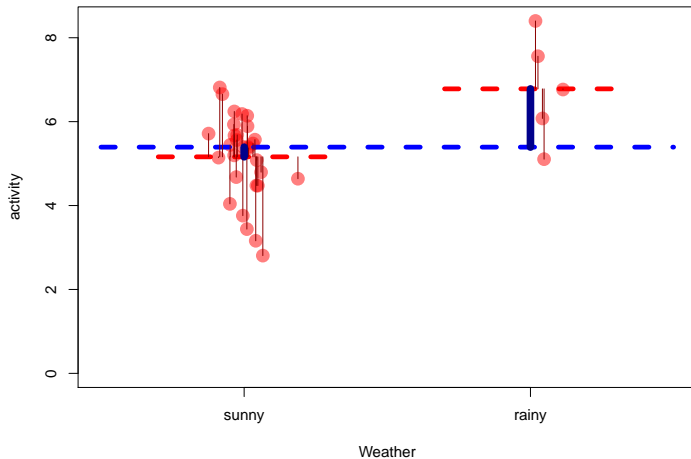
```
fitanova <- aov(data = dat.behav, formula = activity ~ weather)
summary(fitanova)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
weather	1	11.25	11.253	10.73	0.00248 **
Residuals	33	34.62	1.049		

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

ANOVA, graphically

Variance decomposition



Linear regression

```
fitlm <- lm(data = dat.behav, formula = activity ~ weather)
summary(fitlm)
```

Call:

```
lm(formula = activity ~ weather, data = dat.behav)
```

Residuals:

	Min	1Q	Median	3Q	Max
	-2.3547	-0.6028	0.2346	0.6419	1.6534

Coefficients:

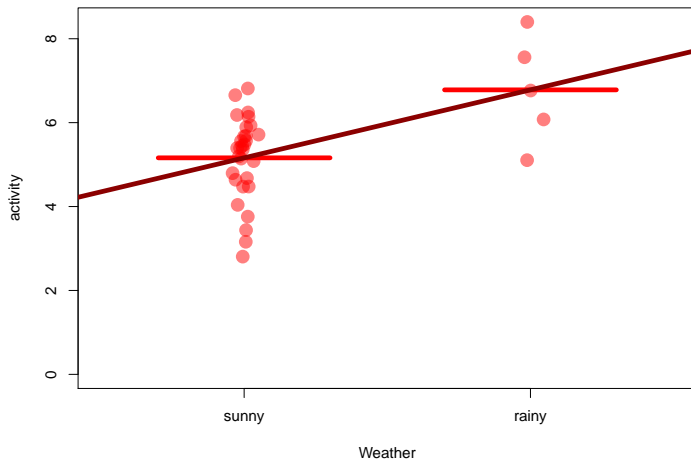
	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	6.7815	0.4581	14.805	3.94e-16 ***
weathersunny	-1.6204	0.4948	-3.275	0.00248 **

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 1.024 on 33 degrees of freedom

Regression, graphically

Rate of change



NB: aov() vs. anova()

```
aov(data = dat.behav, formula = activity ~ weather)
anova(fitlm)
```

All is one. . .

All is one. . .

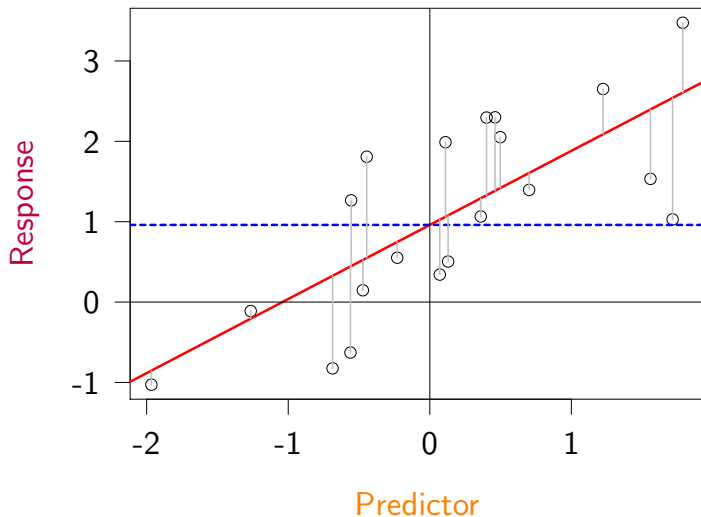
...but `lm()` rules!

- t-test, ANOVA, regression and others can be mathematically equivalent
- In R, `lm()` and related functions can do them all. . .
- ...and much more!

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A simple linear model

$$\text{Response} = \text{Intercept} + \text{Slope} \times \text{Predictor} + \text{Error}$$



A simple linear model

$$\text{Response} = \text{Intercept} + \text{Slope} \times \text{Predictor} + \text{Error}$$

In R:

```
lm(response ~ 1 + predictor1 + predictor2, data=data)
# equivalent to
lm(response ~ predictor1 + predictor2, data=data)
```

- Intercept can be explicit or implicit
- Can remove intercept with $\dots \sim 0 + \dots$
- Error is implicit
- Feed the option `data=` to keep code short, reliable and flexible
- Order of predictors do not matter

Interpretation

```
Ans <- read.csv(file = "Anscombe.csv")
```

```
Warning in file(file, "rt"): cannot open file 'Anscombe.csv':  
No such file or directory  
Error in file(file, "rt"): cannot open the connection
```

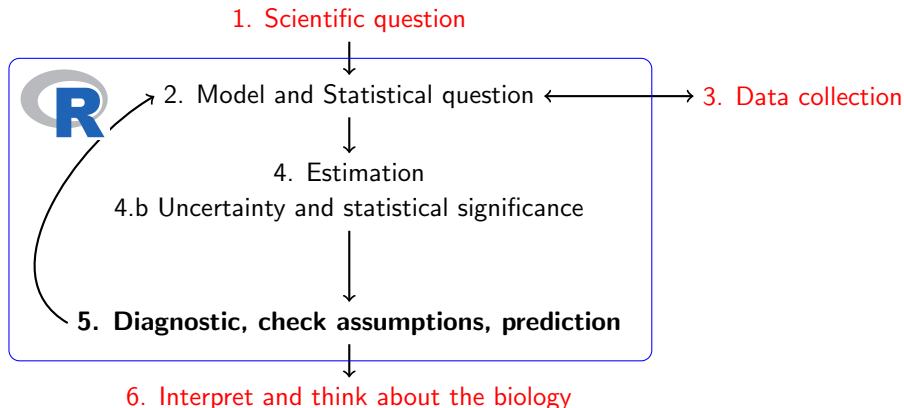
```
lm1 <- lm(y ~ x , data=Ans[Ans$distri==1,])  
summary(lm1)  
plot(Ans$x[Ans$distri==1], Ans$y[Ans$distri==1],  
      xlim=c(0,15), ylim=c(0,12))  
abline(lm1)
```

Interpretation

lm vs. plot

- Fit a linear model $y \sim x$ for each of the four “distri”
- Plot the relationship $y \sim x$ for each of the four “distri”
- Can we trust these models? For what? *I expect more than “it’s all bullshit”*

General approach



Linear model basic assumptions

Not necessarily wrong, but typical interpretation assumes:

- Linear combination of parameters (including transformation, polynoms, interactions. . .)
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- Homoscedasticity (constant error variance)
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Risk: Over-optimistic uncertainty, unreliable predictions
- Independence of error
Risk: Bias and over-optimistic uncertainty

Diagnostic: summary and plot

```
lm1 <- lm(y ~ x , data=Ans[Ans$distri==1,])
```

Error in is.data.frame(data): object 'Ans' not found

```
lm2 <- lm(y ~ x , data=Ans[Ans$distri==2,])
```

Error in is.data.frame(data): object 'Ans' not found

```
summary(lm1)
par(mfrow=c(2,2))
plot(lm1)
```

```
summary(lm2)
plot(lm2)
par(mfrow=c(1,1))
```

Diagnostic: prediction

```
pred2 <- predict(lm2, se.fit = TRUE, interval = "confidence")
```

```
Error in predict(lm2, se.fit = TRUE, interval = "confidence"):
object 'lm2' not found
```

```
pred2 <- cbind(Ans[Ans$distri==2,], pred2)
```

```
Error in cbind(Ans[Ans$distri == 2, ], pred2): object 'Ans' not
found
```

Diagnostic: prediction

```
pred2 <- predict(lm2, se.fit = TRUE, interval = "confidence")
```

```
Error in predict(lm2, se.fit = TRUE, interval = "confidence"):
object 'lm2' not found
```

```
pred2 <- cbind(Ans[Ans$distri==2,], pred2)
```

```
Error in cbind(Ans[Ans$distri == 2, ], pred2): object 'Ans' not
found
```

```
Error in plot(pred2$x, pred2$y, ylim = c(0, 12), col = "red", pch
= 16, : object 'pred2' not found
```

```
Error in lines(pred2$x, pred2$fit.fit): object 'pred2' not found
```

```
Error in arrows(x0 = pred2$x, y0 = pred2$fit.lwr, y1 =
pred2$fit.upr, : object 'pred2' not found
```

```
Error in strwidth(legend, units = "user", cex = cex, font =
text.font): plot.new has not been called yet
```

Practice lm() with parasites

What explains variation in parasitic load?

You collected ecto-parasites on some furry large mammals at three locations. Parasites break easily when we collect them and are impossible to count, so we decide to measure parasitic load as their mass. **Why do some mammals have larger parasitic load?**

Practice `lm()` with parasites

What explains variation in parasitic load?

You collected ecto-parasites on some furry large mammals at three locations. Parasites break easily when we collect them and are impossible to count, so we decide to measure parasitic load as their mass. **Why do some mammals have larger parasitic load?**

- Load the `Para.csv` data (don't forget: `str()`, `summary()`, `plot()`...)
- Model `Parasite_Mass` using `lm()`
- Find what variables predict `Parasite_Mass`
- How good are your models? Assumptions? Prediction?
- What biological interpretation can you imagine?