

Simulating with Parameter Uncertainty

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1 Purpose

This script shows how to conduct a simulation that considers uncertainty in the parameter estimates. See also http://www.page-meeting.org/page/page2006/P2006III_11.pdf.

2 Data

Here we load metrumrg and read in the data to be used for simulations.

Listing 1:

```
> library(metrumrg)
> data <- read.csv("../data/derived/phase1.csv")
> head(data)
```

	C	ID	TIME	SEQ	EVID	AMT	DV	SUBJ	HOUR	HEIGHT	WEIGHT	SEX	AGE	DOSE	FED	SMK
1	C	1	0.00	0	0	.	0	1	0.00	174	74.2	0	29.1	1000	1	0
2	.	1	0.00	1	1	1000	.	1	0.00	174	74.2	0	29.1	1000	1	0
3	.	1	0.25	0	0	.	0.363	1	0.25	174	74.2	0	29.1	1000	1	0
4	.	1	0.50	0	0	.	0.914	1	0.50	174	74.2	0	29.1	1000	1	0
5	.	1	1.00	0	0	.	1.12	1	1.00	174	74.2	0	29.1	1000	1	0
6	.	1	2.00	0	0	.	2.28	1	2.00	174	74.2	0	29.1	1000	1	0

	DS	CRCN	TAFD	TAD	LDOS	MDV	predose	zerodv
1	0	83.5	0.00	.	.	0	1	0
2	0	83.5	0.00	0	1000	1	0	0
3	0	83.5	0.25	0.25	1000	0	0	0
4	0	83.5	0.50	0.5	1000	0	0	0
5	0	83.5	1.00	1	1000	0	0	0
6	0	83.5	2.00	2	1000	0	0	0

We use NONMEM output from a simple two compartment model to generate parameters. We use 1005.lst and 1005.cov output from NM7 to populate a call to metrumrg::simpar().

Listing 2:

```
> cov <- read.table("../nonmem/1005/1005.cov", skip=1, header=T)
> head(cov)
```

	NAME	THETA1	THETA2	THETA3	THETA4	THETA5
1	THETA1	0.85947800	0.7848260	1.05073e-03	0.06297000	-1.6425100
2	THETA2	0.78482600	4.7421000	6.67920e-03	0.89652600	5.3176400
3	THETA3	0.00105073	0.0066792	2.75922e-05	0.00222269	-0.0304355
4	THETA4	0.06297000	0.8965260	2.22269e-03	0.28707800	0.1958110
5	THETA5	-1.64251000	5.3176400	-3.04355e-02	0.19581100	563.8350000
6	THETA6	-0.04113180	-0.0252131	-1.04883e-04	-0.01065710	0.7701760

	THETA6	THETA7	SIGMA.1.1.	SIGMA.2.1.	SIGMA.2.2.	OMEGA.1.1.
1	-0.041131800	-0.176199000	-5.18961e-04	0	2.06030e-02	6.09321e-03
2	-0.025213100	0.068704500	-3.12567e-03	0	1.90856e-02	5.73980e-03
3	-0.000104883	-0.000135683	-1.02658e-05	0	5.89818e-05	3.21218e-06

```

4 -0.010657100  0.015500000 -6.28838e-04      0  2.53788e-03  4.30468e-03
5  0.770176000 -0.633694000  4.55841e-02      0 -4.24311e-01  2.73913e-01
6  0.013008700  0.000572209  1.20518e-04      0 -1.04929e-03  1.65243e-03
      OMEGA.2.1.  OMEGA.2.2.  OMEGA.3.1.  OMEGA.3.2.  OMEGA.3.3.
1 -2.40140e-04 -4.36000e-03 -5.36432e-03 -2.57895e-03 -3.33572e-03
2 -2.19992e-02 -2.44779e-02 -1.95821e-02 -1.12216e-02  4.78859e-03
3 -6.51294e-05 -7.81938e-05 -6.76593e-05 -2.76252e-05  2.83097e-05
4 -6.21638e-03 -7.79632e-03 -4.55928e-03 -2.25358e-03  3.07430e-03
5  1.60551e-01  2.73617e-02 -5.50834e-03  7.46183e-02 -3.41613e-02
6  3.04227e-04  5.99259e-04 -5.34503e-04 -5.46264e-05 -3.36875e-04

```

We are interested in theta covariance, so we remove extra columns and rows.

Listing 3:

```
> cov<- cov[1:7,c(2:8)]
```

3 Parameters

Now we generate 10 sets of population parameters based on the 1005.lst results.

Listing 4:

```

> set.seed(10)
> PKparms <- simpar(
+   nsim=10,
+   theta=c(8.58,21.6, 0.0684, 3.78, 107, 0.999, 1.67),
+   covar=cov,
+   omega=list(0.196, 0.129, 0.107),
+   odf=c(40,40,40),
+   sigma=list(0.0671),
+   sdf=c(200)
+ )
> PKparms

```

	TH.1	TH.2	TH.3	TH.4	TH.5	TH.6	TH.7	OM1.1	OM2.2	OM3.3	SG1.1
1	7.565	19.23	0.06670	3.882	107.50	1.1020	1.340	0.1847	0.15400	0.13630	0.06894
2	6.531	20.18	0.06637	3.861	102.60	1.0680	2.325	0.2862	0.12000	0.16400	0.06099
3	8.257	21.93	0.06598	3.722	74.43	0.8294	2.140	0.1647	0.12770	0.11300	0.06041
4	6.394	19.65	0.06679	3.521	92.78	0.9400	2.011	0.1886	0.11460	0.08460	0.07700
5	7.266	20.13	0.07281	4.136	114.00	0.9471	1.937	0.1526	0.08448	0.13140	0.06269
6	8.205	21.46	0.07480	4.221	116.30	0.9340	1.544	0.2462	0.17640	0.08805	0.07274
7	8.495	23.50	0.07476	4.147	78.29	1.0610	1.906	0.2221	0.14440	0.09957	0.06160
8	7.988	21.95	0.07318	4.524	98.36	0.9228	1.700	0.2287	0.13820	0.06118	0.06692
9	8.268	19.21	0.07017	3.554	68.39	0.9785	1.814	0.1765	0.12310	0.08504	0.06092
10	8.144	20.51	0.06545	3.754	100.90	1.0090	1.511	0.2116	0.11940	0.09954	0.06269

4 Control Streams

We read in a control stream and clean out extra xml markup.

Listing 5:

```
> ctl <- as.nmctl(readLines("../nonmem/ctl/1005.ctl"))
> ctl[] <- lapply(ctl,function(rec)sub("<.*","",rec))
```

Now we iterate across the rows of PKparms, writing out a separate ctl for each.

Listing 6:

```
> dir.create('../nonmem/sim')
> set <- lapply(
+   rownames(PKparms),
+   function(row,params,ctl){
+     params <- as.character(PKparms[row,])
+     ctl$prob <- sub(1005,row,ctl$prob)
+     ctl$theta <- params[1:7]
+     ctl$omega <- params[8:10]
+     ctl$sigma <- params[11]
+     names(ctl)[names(ctl)=='estimation'] <- 'simulation'
+     ctl$simulation <- paste(
+       '(',
+       as.numeric(row) + 7995,
+       'NEW) (' ,
+       as.numeric(row) + 8996,
+       'UNIFORM) ONLYSIMULATION'
+     )
+     ctl$cov <- NULL
+     ctl$table <- NULL
+     ctl$stable <- NULL
+     ctl$stable <- 'ID TIME DV WT SEX LDOS NOPRINT NOAPPEND FILE=sim.tab
+   ,
+     write.nmctl(ctl,file=file.path('../nonmem/sim',paste(sep='.',row,'
+   ctl'))))
+     return(ctl)
+   },
+   params=PKparms,
+   ctl=ctl
+ )
```

5 Simulation

Finally, we run NONMEM simulations using NONR.

Listing 7:

```
> NONR (
```

```
+      run=1:10,  
+      command="/opt/NONMEM/nm72/nmqual/autolog.pl",  
+      project="../nonmem/sim",  
+      diag=FALSE,  
+      checkrunno=FALSE,  
+      grid=TRUE  
+ )
```