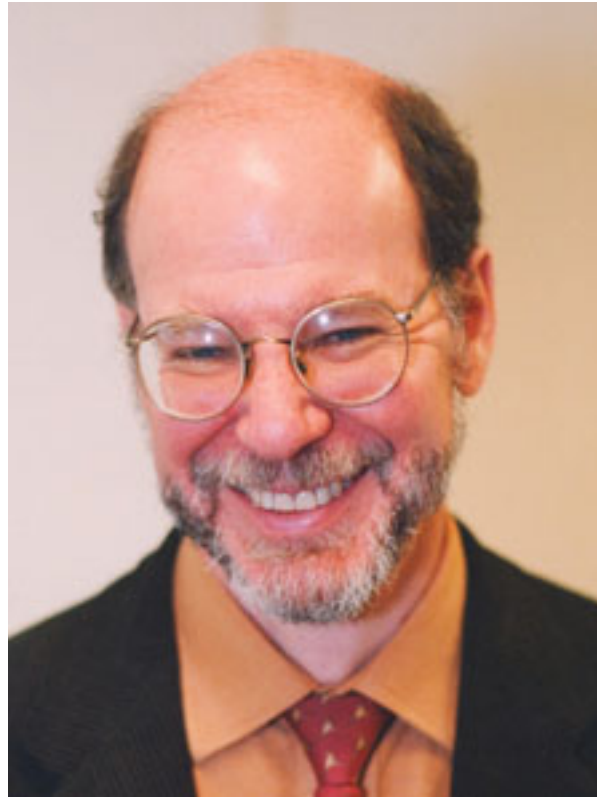


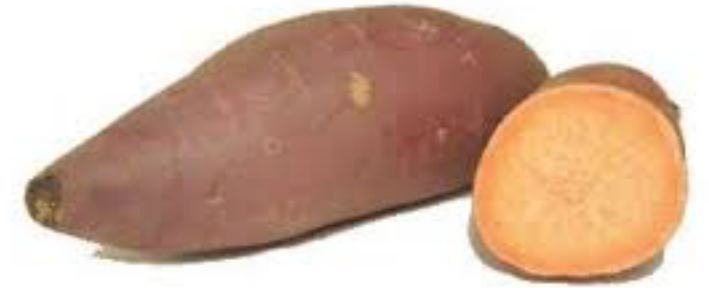
# Bio393: Genetic Analysis

Step-wise genetic analysis



**Bob Horvitz**

# “Model organisms” are everywhere now



# 1. Define the problem



Let the question influence the choice of organism  
(not the other way around)

## 2. Choose an organism

Organism	Time to $10^6$	Space
Bacteriophage	1 hour	10 nL

$10^6$  individuals to study  $10^{-6}$  mutation rate

$$2^{20} \approx 10^6 \text{ individuals}$$

## 2. Choose an organism

Organism	Time to $10^6$	Space
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Mouse	3 years	Half Pancoe

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## 2. Choose an organism

Organism	Time to $10^6$	Space
Bacteriophage	1 hour	10 nL
Bacteria	15 hours	1 $\mu$ L
Yeast	1 day	0.1 mL
Worm	10 days	6 cm cube
Fly	6 weeks	0.5 m cube
Mouse	3 years	Half Pancoe
Human	750 years	Chicago suburbs

$10^6$  individuals to study  $10^{-6}$  mutation rate

$$2^{20} \approx 10^6 \text{ individuals}$$

### 3. Perform a mutant hunt

To mutagenize?

Yes	No	
$10^{-3}$	$10^{-6}$	LoF mutation
$10^{-5}$ - $10^{-6}$	$10^{-8}$ - $10^{-9}$	Specific mutation

*C. elegans*



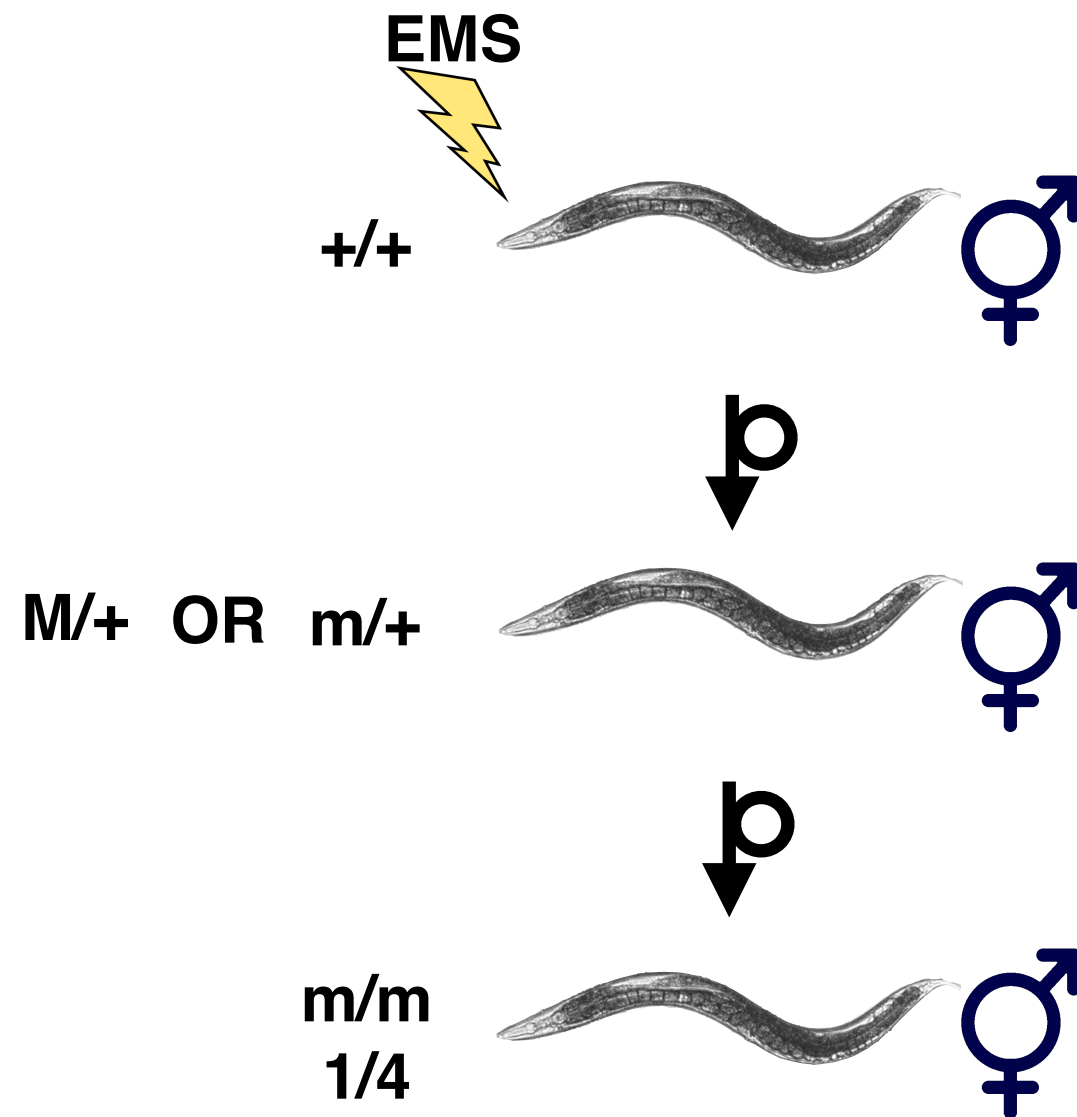
~20,000 genes  
20 LoF mutations

*D. melanogaster*



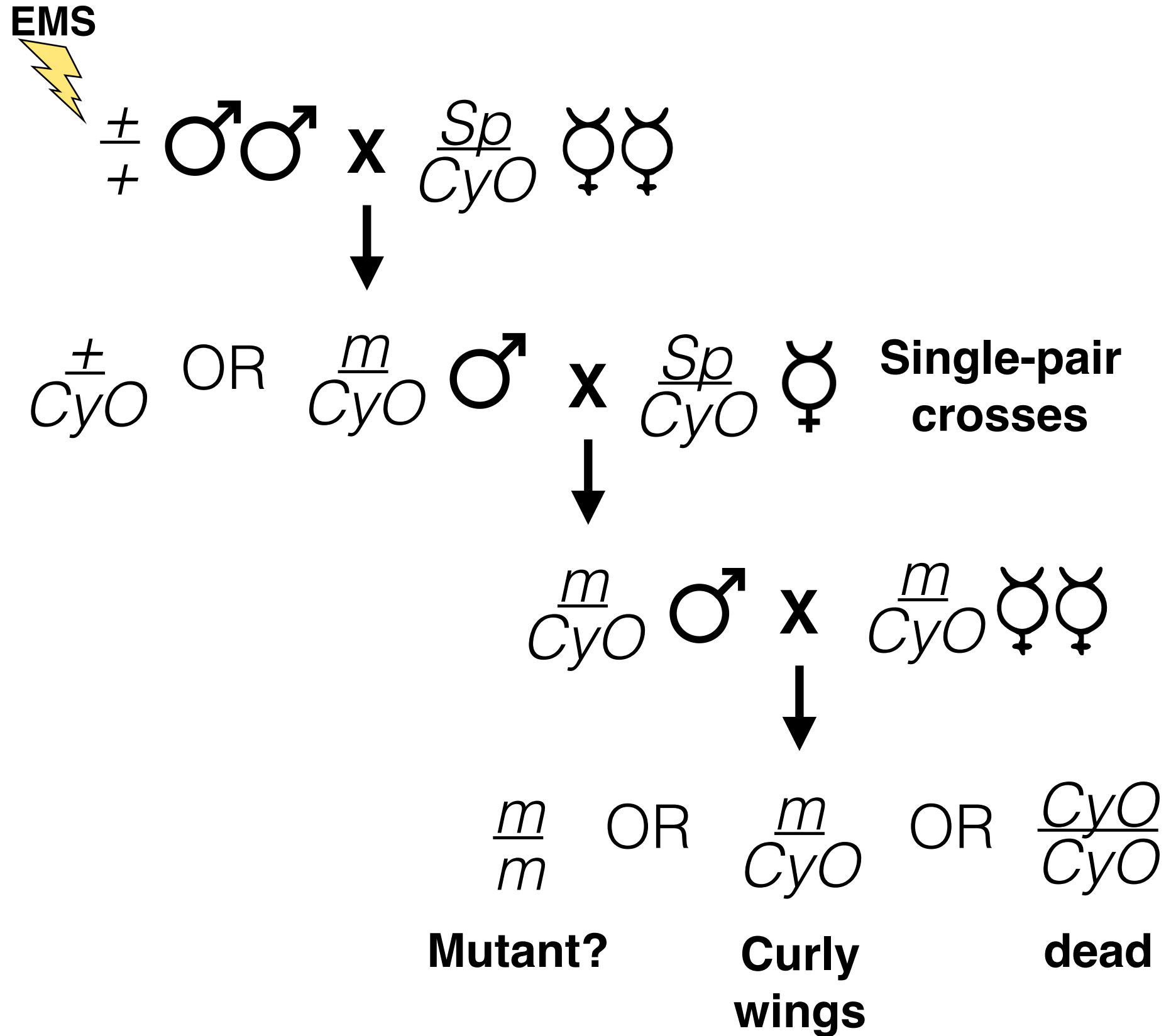
~12,000 genes  
12 LoF mutations

# Screen or selection?

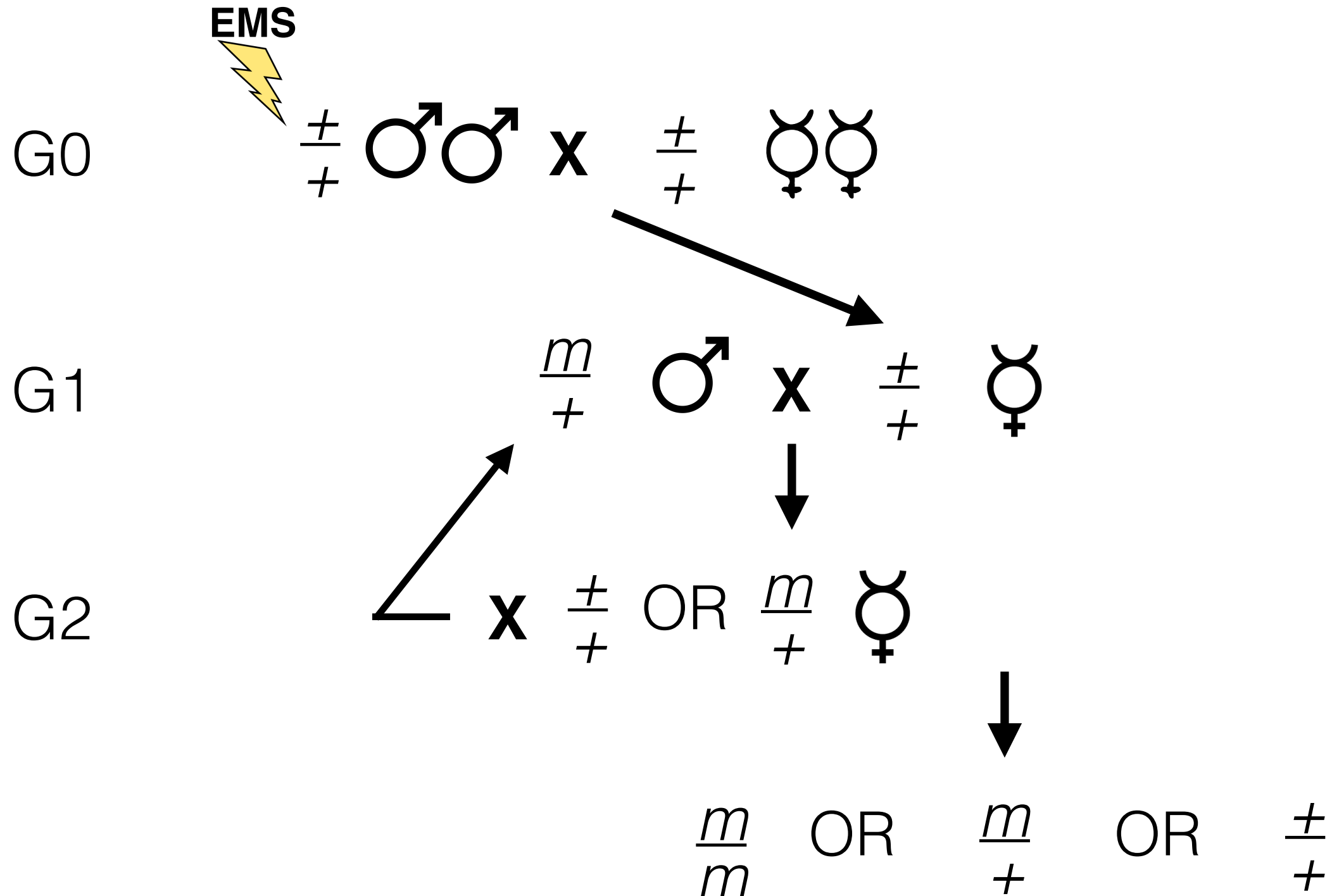


*C. elegans* screens for dominant or recessive phenotypes

# Screen or selection?



# Screen or selection?



Mouse screens for dominant or recessive phenotypes

Remember hemizygous screens too

## **4. Screen until saturation?**

Use Poisson sampling and common sense

Saturation of the investigator's patience

Change mutagens

### **Why might we miss genes?**

Numbers are too small

Pleiotropy (sterility or lethality)

Redundancy



## **5. Establish a strain**

True-breeding stocks

Balancers, balanced stocks

Freeze organisms

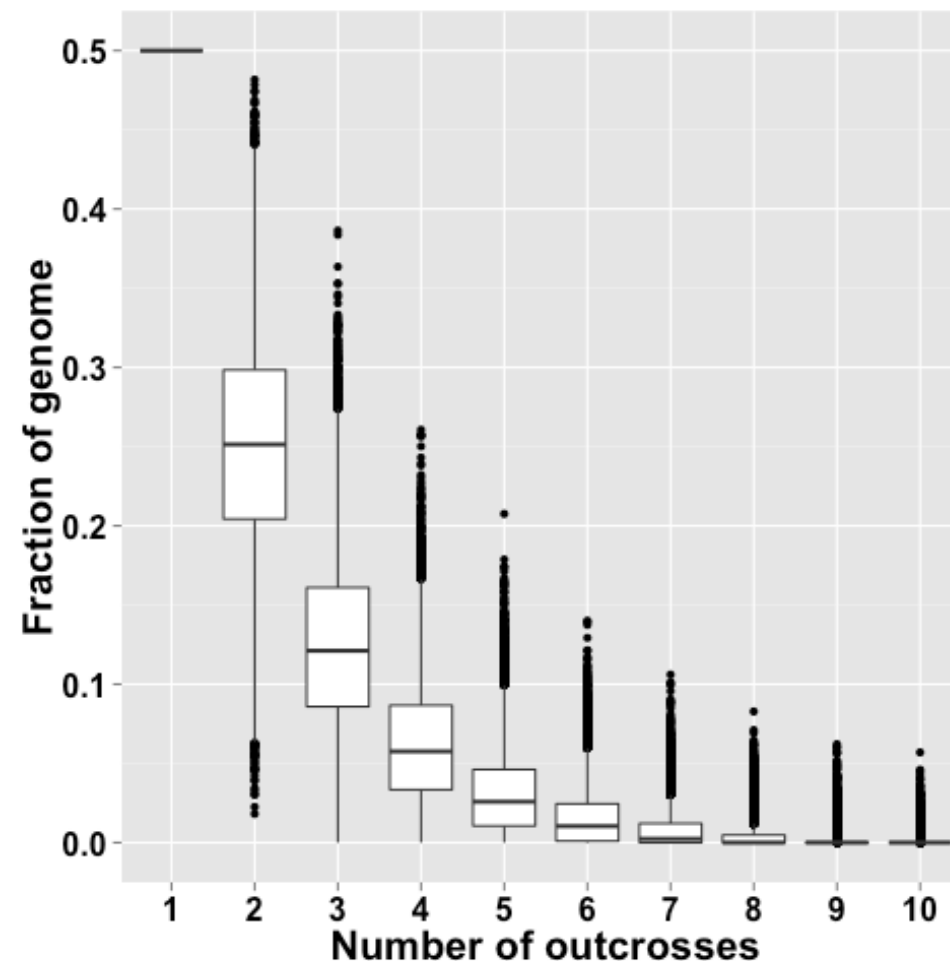
**The most common phenotypes are sterile or dead!**

## 6. Backcross and/or outcross

Mutagenesis adds hundreds of mutations randomly throughout the genome.

Backcross = cross to parent used in the screen/selection

Outcross = cross to a wild-type strain



**7. Test for dominance**

**8. Single-gene phenotype?**

**9. Mapping and complementation**

What have we discovered so far?

# 10. Characterize the phenotype

Look at the wild-type and mutant organisms *in detail*



Let's say you  
screened for mutants  
that failed to lay eggs

What could be  
mutated?

No embryos

No vulva

No vulval muscles

No neurons

Or malfunction of any vulva, muscle, or neuron

# Pleiotropy

A single mutation causes many different mutant traits

Mutation in gene X



Mutant with...

long hairs  
disrupted sleep patterns  
slow growth  
enhanced metabolism of high fat diet

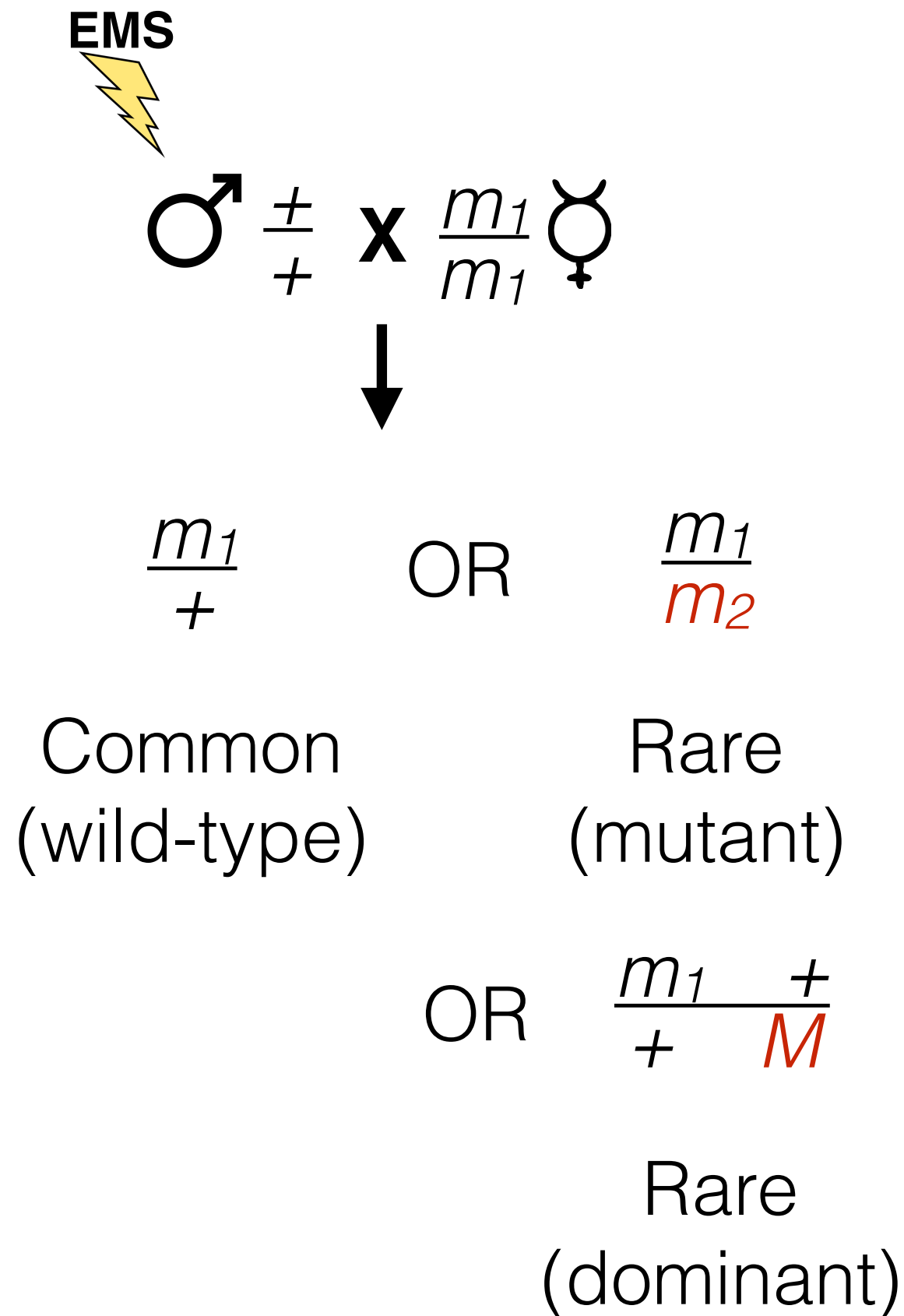
gene X  $\longrightarrow$  short hair  
gene X  $\longrightarrow$  normal sleep  
gene X  $\longrightarrow$  normal growth  
gene X  $\longrightarrow$  fat metabolism

# **11. Define the nature of the mutant allele(s): gene dosage**

1. Dominant or recessive?
2. Frequency of mutant?
3. Where is the mutant allele in allelic series?
4. Look at deficiency heterozygotes for haploinsufficiency
5. Antagonism by wild-type copies of gene

What if you only have one mutant?

# 12. Perform non-complementation screens



# **13. Define the null phenotype**

What happens with a complete loss of gene function?

Dosage studies, non-complementation screens, and  
characterization of the mutant phenotype  
tell you about the null phenotype

What if you have a mutant  
with a dominant gain-of-function phenotype?



# Cis-dominant suppressor screen

EMS



♂  $\frac{D}{D}$  ×  $\frac{+}{+}$  ♀



$\frac{D}{+}$

OR

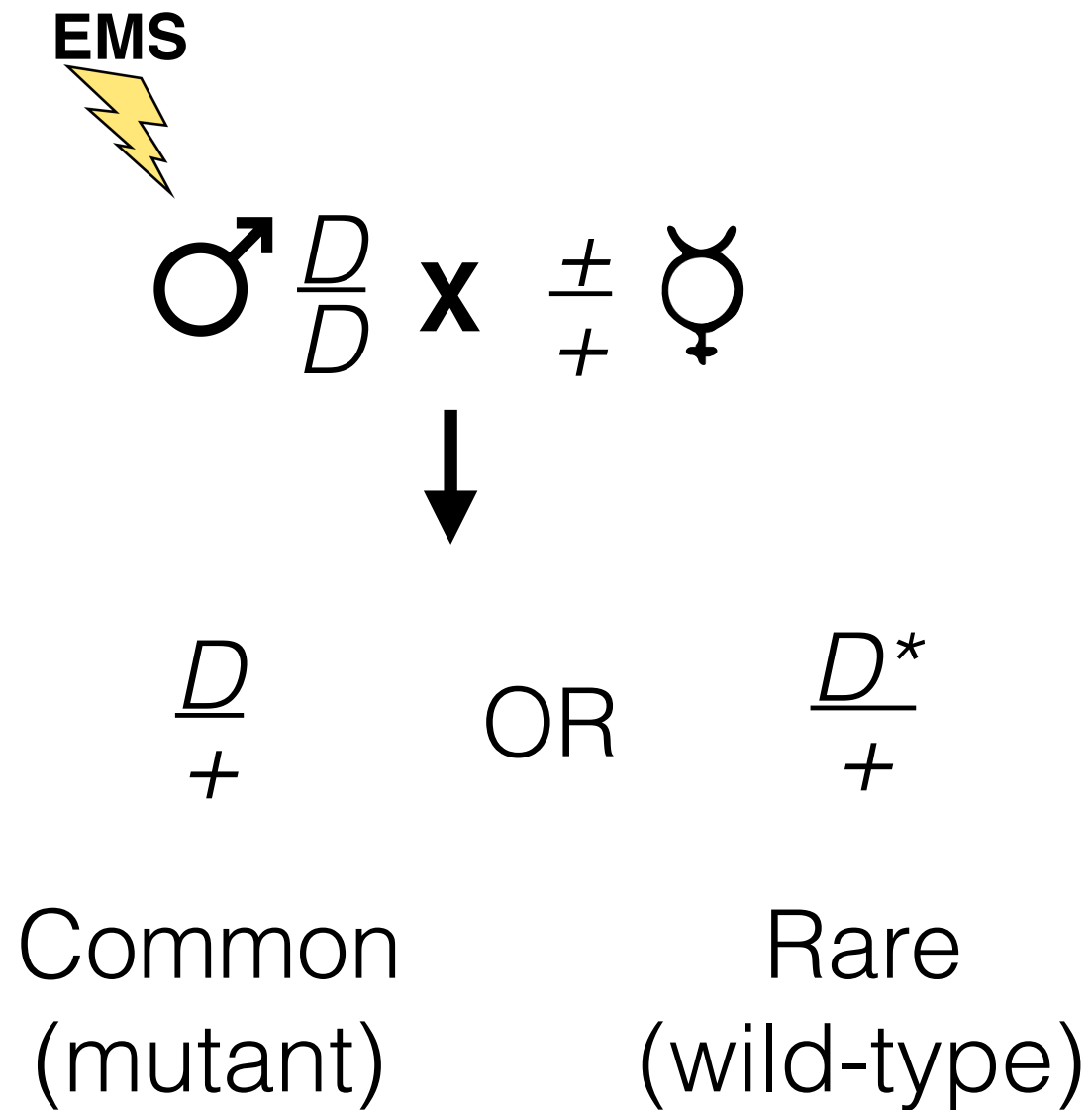
$\frac{D^*}{+}$

Common  
(mutant)

Rare  
(wild-type)

What could  $D^*$  be?

# Cis-dominant suppressor screen



**Revertant**  $\frac{D^*}{+} \rightarrow \frac{\pm}{+}$

**Null mutant**  $\frac{D^*}{+} \rightarrow \frac{\Delta D^*}{+}$

**Intragenic supp.**  $\frac{D^*}{+} \rightarrow \frac{sup D^*}{+}$

**Dominant extragenic suppressor**  $\frac{D^*}{+} \rightarrow \frac{D^*}{+} ; \frac{Sup}{+}$

How can we tell what we got?