UNIVERSITATEA "ALEXANDRU-IOAN CUZA" DIN IAȘI

FACULTATEA DE INFORMATICĂ



LUCRARE DE LICENȚĂ

Aspecte computaționale în biologie

propusă de

Andi Munteanu

Sesiunea: februarie, 2019

Coordonator științific

Conf. Dr. Liviu Ciortuz

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Conf. Dr. Liviu Ciortuz

	Avizat
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Motivație

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Chapter 1

Description of methods

This chapter contains informations about the methods used for graph clustering, the sequencing and processing the biological data and eventually mentions of other works / papers that were focusing on assessing the robustness on changing the seed.

1.1 Graph Clustering

- 1.1.1 Short intro about what graph clustering is
- 1.1.2 Why graph clustering instead other traditional methods such as k-means, density based techniques etc
- 1.1.3 Types of graph clustering
- 1.1.4 Community detection Optimizing the quality function

1.1.5 Louvain

The Louvain algorithm [1] is the state-of-the art community detection algorithm that uses an iterative greedy approach. The method can be used to optimize a partition provided by the user, but the default behaviour is to initially assign each node to its own cluster. Each iteration is described by two repeating steps. The first step (called local moving heuristic) is to change the partition structure in a greedy manner. For each node the algorithm evaluates whether the change of its label could improve the overall quality or not. If so, the node moves to the cluster that provides the greatest increase of quality. These operations are repeated until no change is longer possible. The second

step is shrinking the graph, meaning each community with a super-node. Thus, the number of nodes of the resulting graph will be the same as the number of clusters that were identified in the first step. The weight of the edges are also recalculated using the sum of inter-cluster weights of the original graph. These two steps are repeated until the partition doesn't change after the first phase.

The algorithm can use multiple runs, when the current iterations takes as starting point the partition that is obtained in the previous one. The algorithm is said to reach convergence if no change is noticed at two consecutive iterations.

Although it is a greedy algorithm, Louvain proved it can obtain qualitative clusters. Another advantage of the method is computational efficiency: the average time complexity is $O(n \log n)$, where n is the number of nodes.

1.1.6 Louvain with multilevel refinement

Rotta and Noack [2] proposed an extension of the Louvain algorithm that introduces an extra layer of refinement. The problem the authors were trying to overcome was escaping the local optima that the first step was reaching.

The solution they provided is to exploit a consequence of the second step of the algorithm: after the graph is shrunk and re-clustered, the clusters' composition can suffer some changes. The idea of the Louvain with multi-level refinement is to add another step after the graph is expanded back to its original structure, which is reapplying the local moving heuristic. That ensures that, at the end of each iteration, a local optima is always reached.

1.1.7 SLM

The Louvain algorithm (and its refined version) ensure local optimal node movements, but it doesn't optimize the movement of a set of nodes or the splitting of a community. Waltman and van Eck [3] proposed the Smart Local Moving algorithm (SLM). In this method, for each community that is obtained in the first step of the Louvain algorithm, the associated subgraph is generated and clustered using the local moving heuristic. If the subgraph will contain more than one cluster, the initial community will be replaced with its subgroups. This procedure enables the algorithm to move subsets of points in a local optimal way.

One of the main differences between Louvain and SLM relates to the convergence

of the results. Louvain (and its refined variant) quickly converge after a few iterations, as no community merging or node moving leads to better results. However, SLM, by splitting the communities into subclusters and moving a group of nodes, can improve its results after multiple runs. The downside is that the lack of convergence will lead to more execution time.

1.1.8 Leiden

1.2 PhenoGraph pipeline

PhenoGraph [4] is a pipeline proposed by Levine et al. to process biological data and obtain a clustering that is interpreted as different cell types. The pipeline is consists of the following steps:

- 1. dimensionality reduction
- 2. graph construction
- 3. graph clustering

Each step will be described in detail in the following sections.

1.2.1 Dimensionality reduction

Given that the human genome contains approximately 25-30000 genes, it is expected that the input data (that is, cells extracted from a tissue from multiple donors) will be highly dimensional. Clustering techinques are highly reliant on calculating distances between points, thus an increased number of dimension will lead to expensive computations. The solution for this is to reduce the input space such that no information is lost.

One of the most used approach is Principal Component Analysis (PCA) [5], which is a method that uses linear combinations (called principal components) of the initial features to reduce the number of dimensions. This algorithm relies on computing the singular values decomposition, which is a heavy computational task, but several methods of truncating the calculation were developed in order to increase the algorithm's efficiency [6]. To prevent the loss of the original information, the common practice is to use somewhere between 30 and 50 principal components.

Dimensionality reduction can also be performed in a non-linear fashion. Here we mention UMAP [7], an graph-based method that tries to optimize a cross-entropy function in order to create a reduced space that preserves the topology of the original data: the similar points are kept in close proximity, while maintaining the separation between distinct well-defined groups. Compared to the linear methods, UMAP manages to preserve the structure of the original data in only two or three dimensions. This characteristic makes UMAP a more suitable choice when it comes to visualising the data. The downside of the non-linear method that, given its stochastic nature, it can be affected by the value of the random seeds. Usually the effect is presented as slight changes of the topology of the groups or rotations of the representation.

1.2.2 Graph construction

As mentioned before, the relationship between nodes is better described in a graph structure, where the existence of the edges between nodes indicates a specific degree of similarity. The conversion is performed using the kNN alogrithm. This explains the need of performing dimensionality reduction prior to the graph construction, as the distance calculation becomes computationally expensive on large number of features.

The first step is to calculate the k nearest neighbours for each node using a distance metric (usually Euclidian or cosine). The graph will be created by drawing edges between a node and its k neighbours. The neighbourhood relationship is not symmetrical, thus the graph will be directed.

Given the curse of dimensionality [8], distance cannot be trusted to be used for the weight of the edges. Thus, the authors of the PhenoGraph proposed to calculate the weights using the Jaccard Similarity Index (JSI) of the neighbourhoods. The weights can take values between 0 and 1 using the formula

$$W_{ij} = \frac{|v(i) \cap v(j)|}{|v(i) \cup v(j)|},$$

where v(i) indicates the neighbourhood of the node i. Thus, weights closer to zero indicate that the two points have significantly different neighbourhoods, whereas values closer to one mark a good overlap between the sets of neighbours.

The weighted graph is also referred in literature as Shared Nearest Neighbour graph (SNN).

It should be mentioned that the node its included in his own neighbourhood. Otherwise, this would lead to misclassification of weights, especially in the case where the two nodes are direct neighbours, but the rest of their neighbourhoods are disjoint. If v(i) doesn't include i, the intersection will not contain any element, thus the edge will get a weight of zero, altough the points are direct neighbours.

[[Add the two photos with the examples from the supplementary]]
Comp

1.2.3 Graph clustering

The last step consists in identifying the clusters using a community detection algorithm. As presented in the previous section, this method is computationally efficient and manages to group points into dense subgraphs by optimizing a quality function, without being dependant on the cluster size or shape.

The original PhenoGraph article proposed using the Louvain article, but in our experiments we will use its improvements aswell, namely Louvain with multilevel refinement, SLM and Leiden.

The authors' approach on getting results that are not affected by the seed was to run the community detection 100 times and choose the most qualitative partition.

1.2.4 Describing the pipeline

Present the steps that describe the pipeline. (Dimensionality reduction, graph building and graph clustering)

About dimensionality reduction

- 1.2.5 How to convert matrix data into a graph using kNN
- 1.2.6 SNN providing weights using Jaccard Similarity Index

1.3 Element-Centric Similarity

1.3.1 Introductory terms

Cluster affiliation graph: a bipartite graph that represents the relationship between points and clusters. One vertex set will contain the points, while for the other one

node is associated with a cluster. An edge indicates that a point belongs to a cluster (or, viceversa, that a cluster contains that point).

Cluster-induced element graph: a directed graph where the node set is represented by the original points. Two nodes share an edge if they belong to the same cluster.

Affinity matrix The affinity matrix, or the personalized PageRank (PPR) affinity is determined by calculating the paths between elements of the cluster-induced element graph. The PPR of a node i to the others is computed as follows:

$$p_i = (1 - \alpha)v_i + \alpha p_i W$$

where v_i is a one-hot encoded vector with 1 on the i^{th} position, W the weighted adjacency matrix of the cluster-induced element graph and α is a parameter that determines the importance of overlaps in overlapping or hierarchical communities.

For disjoint partitions, the formula is simplified to the following form:

$$p_{ij} = \begin{cases} 0, \text{if i and j don't belong to the same cluster} \\ \frac{\alpha}{|C_{\beta}|}, \text{if i and j belong to the cluster } \beta \\ 1 - \alpha + \frac{\alpha}{|C_{\beta}|}, \text{if } i = j \end{cases}$$

1.3.2 Description about how it works

Element-Centric similarity (ECS) is a new clustering comparison measure proposed by Gates et al [9]. This score is based on calculating the L1 distance between the affinity matrices that are associated with the two clusterings:

$$S_i(\mathcal{A}, \mathcal{B}) = 1 - \frac{1}{2\alpha} \sum_{j=1}^{N} |p_{ij}^{\mathcal{A}} - p_{ij}^{\mathcal{B}}|$$

A major aspect that differentiate ECS from the other metrics is that it is elementcentric and not cluster-centric, that is the score is calculated individually for each element of the partition. The overall score can be calculated as the mean.

Describe the intuition behind ECS: the idea of the bipartite graph between points and clusters.

More details about how to calculate ECS. Talk about the affinity matrix and the L1 distance.

1.3.3 Properties, comparison with other clustering metrics

Present some limitation of other clustering metrics such as bias toward cluster sizes, shapes and so on. Perhaps present some comparison figures from the main article.

Present some properties of ECS:

- 1. the fact that it can be used not only for flat disjoint clusterings, but also for overlapping or hierarchical partitions
- 2. it overcomes the biases present in the other clustering metrics
- 3. ECS illustrates the overall similarity between two partitions but also can help in identifying the points where the clustering are not similar

1.3.4 Element-Centric Consistency

Gates et. al also introduce in the same paper the concept of *frustration*, which is used when a list of clusterings is provided. The frustration is calculated as the average of the pairwise similarity scores:

$$\frac{2}{T(T-1)} \sum_{i=1}^{T} \sum_{k=1}^{j-1} S_i(\mathcal{R}_k, \mathcal{R}_j),$$

where *T* denotes the number of clusterings. We will refer to this metric as *Element-Centric Consistency* (ECC), as higher values of the score indicate a more consistent labeling of the points in the same cluster.

1.4 Intro info about biological data and sequencing techniques

Tell about sequencing techniques, how the initial data looks, about cells, genes, what they mean, what is the role and the purpose of the clusters in the biological interpretation.

Chapter 2

The importance of parameter values in the clustering output

2.1 Monocle and Seurat

As stated in the previous chapter, the PhenoGraph pipeline was widely adopted in the task of processing biological data, due to its efficiency and quality of results. The pipeline was incorporated and implemented in various programming languages.

The most frequently used R packages that perform single-cell data processing are Seurat (currently on the third version) [10] and Monocle (currently at the third version) [11]. Both packages use the PhenoGraph method for the cell clustering. The goal of this chapter is to evaluate the output of the two packages and compare the results side by side.

2.2 Data used for experiments

To perform this comparison, we used the Cuomo data [12], an in-vitro SMART-seq dataset of human endoderm differentiation. The dataset contains 1880 cells extracted from six donors (hayt, naah, vils, pahc, melw and qunz) at four different time-points. The cells are in expressed in approximately 30 000 genes. The dataset was preprocessed by removing the cells that are expressed in less than 200 features and the genes that are expressed in less than three cells. MT (mitochondrial) and RP (ribosomal protein) genes were also excluded from the feature set.

The purpose of this thesis is to analyze the clustering pipeline, therefore the pre-

	Dim reduction	Graph construction	Graph clustering	
Seurat	PCA followed	kNN based	Louvain, Louvain refined,	
Seurat	by tSNE or UMAP	support for SNN	SLM, Leiden	
Monocle	PCA followed	kNN based	Louvain, Leiden	
Monocie	by tSNE or UMAP	support for SNN	Louvaiii, Leideii	

Table 2.1: The algorithms used by Monocle and Seurat inside the Phenograph pipeline

processing parameters (normalization and scaling) are done in the same manner for both packages.

2.3 Algorithms used in the pipeline

In chapter one we presented the steps that are involved in the PhenoGraph pipeline, namely the dimensionality reduction, the graph construction and the community detection. Each step can be performed by a varying number of algorithms. Therefore, in order to compare the two R packages, we must identify the algorithms that they are using in the clustering pipeline (see Table 2.1).

For the dimensionality reduction, both packages allow the usage of either linear methods (PCA) or non-linear ones (UMAP and tSNE). The developers of the both packages recommend performing the PCA reduction on the raw data, followed by tSNE or UMAP. The graph construction is performed using the kNN based method. For the community detection, Monocle uses the Louvain and Leiden alogrithms, whereas Seurat can cluster the cells using Louvain with multi-level refinement and SLM, too.

We note that all the algorithms involved in the clustering pipeline contain at least one stochastic component, therefore

2.4 Comparing the results

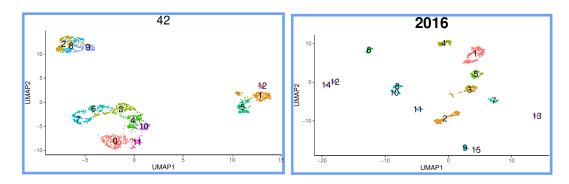


Figure 2.1: Clustering distribution with default parameters for Monocle (left) and Seurat (right). The title indicates the default random seed that is used.

If the Cuomo [12] dataset is clustered using the Monocle and Seurat packages with default parameters, significantly different outputs are produced, as it can be observed in Figure 2.1. Firstly, there are differences with respect to the dimensionality reduction. The cells are scattered in multiple islands in the Monocle package, whereas Seurat obtains a more compact grouping. Also, the two packages disagree regarding the number of clusters: Seurat outputs 13 clusters, Monocle 15.

The biological interpretation of the data heavily relies on the clusters' structure. Thus, is expected for the discrepancies between the two packages to extend over the marker identification and the biological interpretation. The question we pose is whether the divergence between Monocle and Seurat has technical (such as the implementation of the clustering pipeline) or biological (such as additional pre- or post-processing steps that rely on the sequencing information).

2.5 Aligning the results

Chapter 3

ClustAssess

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Chapter 4

Experiments and results

Conclusions, Future Work

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Title: ClustAssess: Tools for Assessing Clustering

Student name: Munteanu Andi

Coordinator: Conf dr. Liviu Ciortuz

The thesis is based on the ClustAssess paper [13], where I contributed as an author and developer of the R package.

Clustering is an unsupervised method that is used to classify and label points based on different similarity metrics. Comparing to the supervised classifiers, clustering algorithms do not require training and using a model and are most suitable when the label of the points are not priorly known and are inferred based on the features that describe them.

Depending on the approach, clustering methods can be divided in multiple categories, such as centroid-based (k-means), density-based (DBSCAN), hierarchical, distribution-based (EM). One approach that gained popularity in the last decades are the graph-based clustering. Some of its advantages is providing flexibility when it comes to the cluster shapes and sizes, or the scalability.

Our thesis focus is set on the community detection techniques, that is graph-based clustering approach that rely on optimizing an objective function (also known in literature as quality function or quality metric, as it tries to evaluate the quality of the clustering by encouraging high density of edges inside clusters and as few intercluster links as possible). This method is intensively used, as it manages to obtain close to optimal results in an efficient time.

Given that many datasets are provided as points displayed on a high-dimensional space, a methodology that enables the use of community detection method on this data is required. Our reference is the PhenoGraph algorithm presented by Levine et. al [4], that establishes a pipeline that firstly applies a dimensionality reduction technique (a linear one such as PCA or non-linear one such as UMAP), then generates a graph using the Nearest Neighbour algorithm. The resulting graph is then used as input for the community detection algorithm.

The PhenoGraph pipeline has been used frequently in several domains. One of them is the downstream analysis of biological data, where the goal is to cluster the cells in multiple groups that will be used to infer some biological conclusions. The input data is usually provided as a matrix that has cells sampled from different donors at different timepoints on the rows. The columns represent the genes that can be expressed in the cells. Given that the human genome contains approximately 30.000 genes, the input space is a high dimensional one, which makes the PhenoGraph pipeline a suitable choice for processing the data. Also, using graphs to represent data is a more appropriate way to describe the cell-cell interaction.

The current state-of the art of processing the biological data is thus using the PhenoGraph pipeline: for dimensionality reduction, approximate PCA using the Lanczos bidiagonalization method [6] or UMAP [7]; for graph building, the Nearest Neighbour algorithm [14]; for community detection, Louvain [1], Louvain with multilevel refinement [2], Smart Local Moving Algorithm [3] and Leiden [15].

Currently, some of the most popular tools used for analyzing single-cell data are Seurat [10], Monocle [11] and SCANPY [16]. Our thesis makes a comparison between Seurat and Monocle regarding the implementation of the PhenoGraph pipeline. This was motivated by the significant differences between the results of the two packages and the subsequent divergent biological interpretation of the obtained partitions. The question we wanted to answer is whether this differences are caused by computational or biological factors (such as sequencing depth or how the data was pre-processed).

In our thesis we showcase how the divergence was caused by using parameters values that do not match. The conclusion we draw is that tuning the algorithm's parameters is essential in obtaining reproducable results. Given the stochastic nature of the algorithms that are involved in the pipeline, we also noticed how changing the random seed value is a direct factor that affects the clustering output.

The instability caused by random seed was previously identified in several papers that pursued the algorithm modification in order to achieve stability. Such example is kmeans++ [17], where the authors replaced the random initialization of the centroids with assigning probabilities of selection based on the distance to the existing center points. Another example is provided in the clust-perturb algorithm proposed by Stacey et al. [18], where the robustness is evaluated by introducing random noise in the graph.

Our work is focused on providing a pipeline that follows the algorithms involved in the PhenoGraph. The package that we developed its purposed to provide informative plots that would give the user insight about how different parameters impact the number of clusters and the partitioning. Another purpose that we try to achieve is to evaluate and provide insight about parameters configuration that are robust to the

change of seeds. The robustness is determined by using Element Centric Similarity (ECS) [9], a measurement that determines how similar are two clustering of the same data.

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