## Friendly guide for performing electron neutrino analysis with WatChMaL framework.

First of all, let's describe the pipeline that we will follow.

 $WCSim.root \rightarrow WCSim.npz \rightarrow WCSim.h5$ ;  $split\_WCSim.npz \rightarrow (model evaluation) \rightarrow softmax\_WCSim.npy \rightarrow softmax\_WCSim.root$ .

So, we have to convert initial WCSim files to h5 format, prepare split\_files for the model evaluation, run model, achive softmax files and convert them back to root format. All steps have corresponding runner scripts.

- Step1: WCSim.root → WCSim.npz
  python convert\_wcsim\_to\_npz.py
  In first step we have to prepare npz files ("npzfiles") for initial
  root files from WCSim simulation and initialize the structure of
  data directory and copy fiTQun files in the neighbour directory
  "fitqunfiles".
- Step2: WCSim.npz → WCSim.h5
   python convert\_npz\_to\_h5.py
   In this step we create corresponding directories and convert "npz-data" files to "h5data".
- Step3: prepare  $split\_WCSim.npz$  module load scipy-stack/2020b python make\_split\_idxs\_path.py
  In this step we prepare data.split\_path files ("splitfiles") for further evaluation and files.txt with h5 and fitqun listed in them.
- Step4: run model evaluation python eval\_files.py #TODO In this step we should one-by-one evaluate model on h5 files data listed in txt format. Output is gathered in rundir.

• Step5:  $softmax\_WCSim.npy \rightarrow softmax\_WCSim.root$  python softmax\_to\_root.py
This step was done locally. We have to convert softmax files to root format and list them. Yet to list cedar locations of softmax.root files in "softmax.txt".

For running Charlie's analysis you need to have corresponding fitqun and softmax files, where softmax file is an output of evaluation with WatChMaL model on wesim data. Then you need to list files locations row-by-row in fitqun\_files.txt and softmax\_files.txt.

Usage of this files together required changes in **RecoInterface**. Also, Charlie's script *analysis\_1ringe.cc*, was modified for next purposes:

- new\_first\_cut\_analysis\_1ringe\_pmu, new\_first\_cut\_analysis\_1ringe\_pe, new\_first\_cut\_analysis\_1ringe\_pgamma for each class probability histos.
- log\_first\_cut\_analysis\_1ringe\_pmu for log scale 1D probability histo.
- analysis\_energy and analysis\_momentum for energy and momentum 1D histos.
- logmomentum\_pmu for reco momentum with log scale probability mu 2D histo.

For running the code you have to perform next steps:

module load StdEnv/2016 source Source\_At\_Start\_nuPRISM.sh source \$DataTools/cedar\_scripts/sourceme.sh

Then for single job run  $run\_single\_job.sh$ . Here you can change the scripts you need, e.g.:

./bin/logmomentum\_pmu \$1 \$2 ./outputs/histo\_\$3.root. Here arguments correspond to single fitqun\_files\_N.txt, softmax\_files\_N.txt and N respectively.

For multiple jobs in parallel just run  $run\_all\_jobs$ . It will produce multiple histos in root files, so you need split your softmax and fitqun files (as it was done in np\\_splited directory).

For converting softmax files to root format run  $softmax\_to\_root.py$ . Modify it for proper files locations which would be listed. If you need to split list of all files, use  $files\_split.py$ .

Jupyter notebook contains plots for this analysis.