# avocado: A Variant Caller, Distributed

Frank Austin Nothaft, Peter Jin, Brielin Brown {fnothaft, phj, brielin}@berkeley.edu



### Background

Three stages in modern DNA processing pipelines:

- 1. Sequencing: Generate 100-250 base pair reads
- 2. Alignment: Align these reads to the reference genome
- 3. Variant Calling: Determine gene variants & genotypes Variant calling is an interesting area: "Accurate" algorithms are slow and don't scale (60 hrs/genome), and are inaccurate for high complexity regions (error is > 75%).

#### Goals:

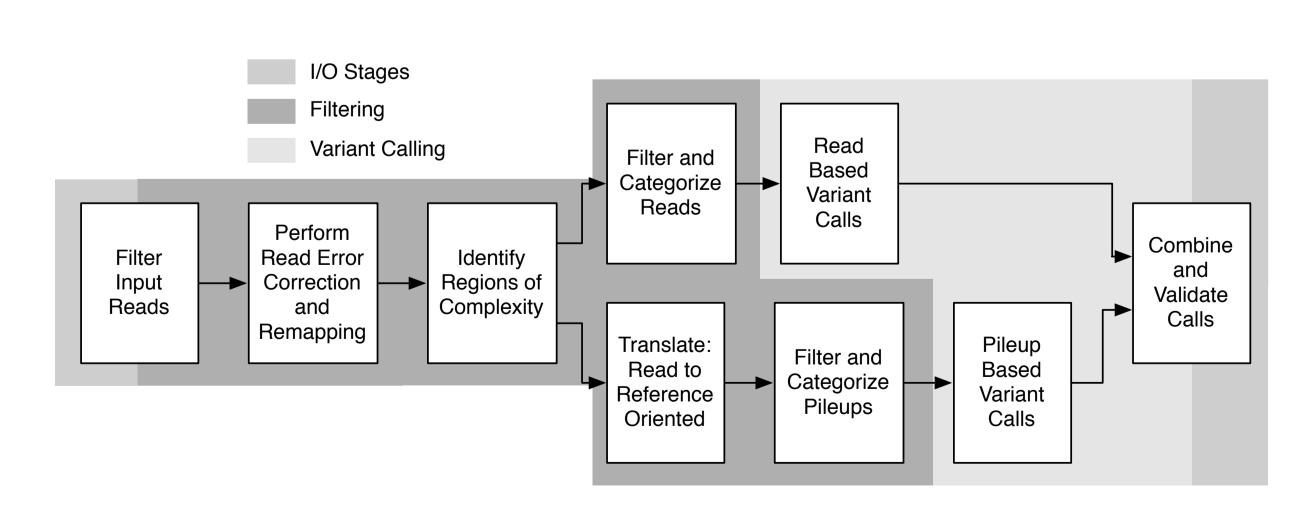
- 1. Build a variant caller designed for distributed computing
- 2. Develop an open-source alternative to the GATK

### Pipeline

### **Tech Specs:**

- Built in Scala on top of Parquet and BDAS Spark
- Leverages new ADAM read/pileup/variant call format
- Scalability well past 30+ nodes; other pipelines are limited to 26 (1/chromosome)

### Pipeline:



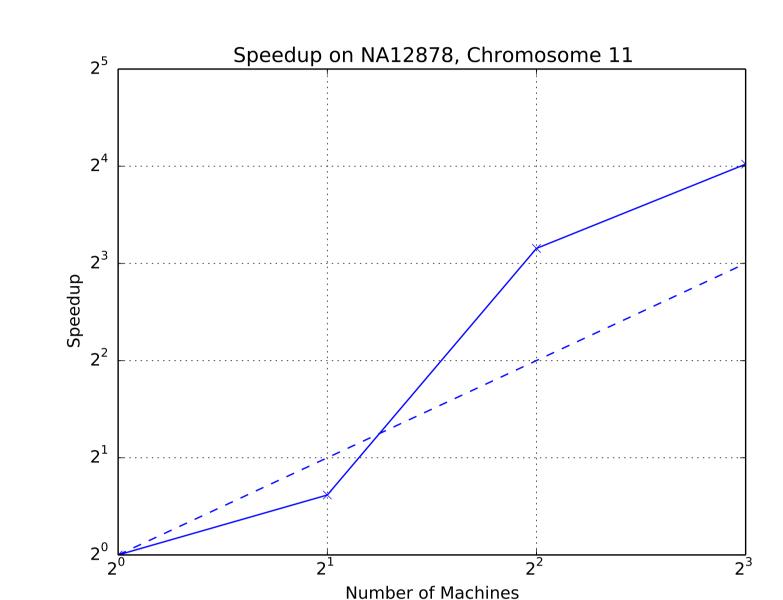
### Design Principles:

- Use mapping quality/coverage as filtering heuristic
- Use assembly methods on high complexity regions
- Design is modular: easy to add new calling algorithms

### Performance

#### **Notes:**

- Algorithm is currently disk bound due to shuffles: performance bug in pileup creation due to partitioning
- Plan to fix performance bug by doing interval-based rod conversion:
- Lump reads by reference position group to maintain locality
- Fewer objects created than reads  $\rightarrow$  pileups  $\rightarrow$  rods



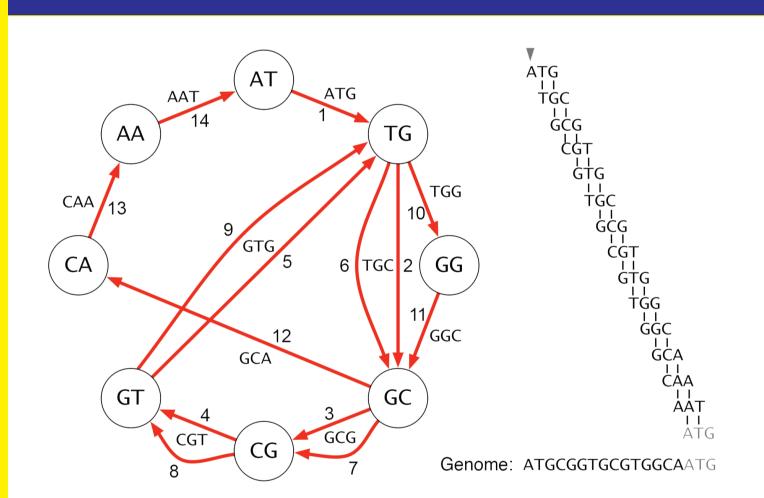
MapQ,Cov	0.2	0.4	0.6
40	88%	54%	20%
<b>50</b>	90%	56%	22%
60	98%	91%	88%

% Reads in High Complexity Region

Sample	Time	Size	Machines
Venter	6hr	99 GB	$20 \times \text{cr1.8xlarge}$
NA12878, chr11		673 MB	$4 \times \text{ m2.4xlarge}$
$\mathbf{NA12878,chr20}$	$0.25~\mathrm{Hr}$	297  MB	$4 \times \text{m2.4xlarge}$

Performance Over Different Datasets

## Local Assembly



We partition the high-complexity locations into regions and use local k-mer assembly to discover the most likely haplotype pair per region. The likelihood of a pair of haplotypes  $H_j$  and  $H_{j'}$  is given by:

$$\mathcal{L}(H_j, H_{j'}) = \prod_i \left[ \frac{P(r_i|H_j)}{2} + \frac{P(r_i|H_{j'})}{2} \right]$$

where  $r_i$  is a read. We obtain P(r|H) by a pairwise HMM alignment model.

Figure credit: P.E.C. Compeau, P.A. Pevner, G. Tesler, "How to apply de Bruijn graphs to genome assembly," Nature Biotech. 29(11), 2011.

### Base SNP Calling

For calling SNPs on a single sample, we look at genome loci that show evidence of a SNP (at least one non-reference base). Genotype likelihoods are calculated by:

$$\mathcal{L}(g) = \frac{1}{m^k} \prod_{j=1}^l (m-g)\epsilon + g(1-\epsilon) \prod_{j=l+1}^k (m-g)(1-\epsilon) + g\epsilon$$

m= ploidy, g= genotype state,  $\epsilon=$  likelihood of error, l= bases matching reference, k= bases at locus

Genotyping is biased towards the reference. We compensate by the allele frequency and call a non-reference genotype if  $g \in (1,2)$  has the highest probability.

## Sufficient Statistics/Joint Calling

For a few samples, one may look-up the MAF  $\phi$  in a reference and compensate the the single sample likelihood

$$\hat{g} = \arg \max_{g} \mathcal{L}(g) \mathbf{P}(g|\phi)$$

When many samples are collected it can be desirable to compute a population MAF while performing genotype calling. For each SNP a, this is done via EM:

$$\phi_{a,t+1} = \frac{1}{M} \sum_{i=1}^{N} \frac{\sum_{g_i} g_i \mathcal{L}(g_i) \mathbf{P}(g_i | \phi_{a,t})}{\sum_{g_i} \mathcal{L}(g) \mathbf{P}(g | \phi_{a,t})}$$

 $M = \sum_i m_i = ext{total number of chromosomes } N = ext{number of individuals}$