avocado: A Variant Caller, Distributed

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ABSTRACT

In this paper, we present avocado, a distributed variant caller built on top of ADAM and Spark. avocado's goal is to provide both high performance and high accuracy in an open source variant calling framework. To achieve this, we implement both local assembly and pileup-based single nucleotide polymorphism (SNP) calling. A key innovation presented in our work involves the development of heuristics for when to choose more expensive assembly-based methods instead of pileup-based methods. Additionally, we introduce the concept of "significant statistics," a tool for performing incremental joint variant calling.

Categories and Subject Descriptors

Applied Computing [Life and Medical Sciences]: Computational Biology—Sequencing and Genotyping Technologies; Applied Computing [Genomics]: Computational Genomics; Computing Methodologies [Distributed Computing Methodologies]: Distributed Algorithms—MapReduce Algorithms

General Terms

Algorithms, Performance

Keywords

Variant Calling, Genotyping, Genomics Pipeline, Local Assembly, Distributed Computing, MapReduce

1. INTRODUCTION

Modern genomics processing pipelines can be divided into four primary ordered stages:

- 1. Sequencing: Gathering of read data from DNA
- 2. **Alignment:** Alignment of read data against reference genome

- 3. Variant Calling: Statistical determination of differences against reference genome
- 4. Variant Annotation: Annotation of impact of variation

Currently, to run a genomics pipeline end-to-end for a single high coverage genome¹ consumes approximately 100 hours [?]. Of this 100 hour figure, both alignment and variant calling consume approximately 50 hours each.

Although some applications that use genomic data are latency insensitive (for example, population genomics), many medical applications like genomic medicine, or genomic classification of viral outbreaks [?] are latency sensitive. However, it is unacceptable to sacrifice accuracy in the pursuit of speed. Recent work has focused on the problem of accelerating alignment [?]; in this paper, we address accelerating variant calling.

As noted above, it is unacceptable to sacrifice accuracy for performance. To achieve improved performance, we implement several enhancements:

- Current pipelines are penalized by I/O performance, we address this by using an in-memory MapReduce framework [?] to reduce I/O pressure
- Additionally, we leverage the new ADAM data format [?], a high performance file format for distributed genomics
- Finally, we achieve high accuracy at a low performance cost by using high fidelity assembly-based methods only on complex segments of the genome

In this paper, we discuss this system, related work, and perform a performance analysis. We start with a discussion of the related work in $\S 2$. We then describe our architecture and algorithms in $\S 3$. Finally, we analyze the performance of our system in $\S 4$, and propose future research directions in $\S 5$.

 $^{^1{\}rm High}$ coverage refers to having on average >30× bases aligned to each location in the reference genome.

2. RELATED WORK

There has been significant work related to variant calling, and towards accelerating the genomic processing pipeline. In this section, we discuss other variant calling pipelines, and tools that we use in our evaluation.

2.1 ADAM

ADAM [?] is a new data format for genomics meant to replace the Sequence/Binary Alignment/Map (SAM/BAM) formats for read data [?], and the Variant Call Format (VCF) for variant/genotype data [?]. The original SAM/BAM/VCF formats were designed for single-node processing, and do not easily distribute across several machines. Although a library was designed for processing BAM/VCF data in Hadoop [?], this API does not scale well past 8 nodes. ADAM achieves scalability beyond 100 machines by eliminating the central file header, and by using the Parquet data store which is optimized for parallel data access [?].

In the process of developing avocado, we contributed 2,500 lines of code (LOC) to the ADAM project. This contribution comprised the variant and genotype format, code for calculating normalized variant data from genotypes, and converters to/from the VCF format.

2.2 The Genome Analysis Toolkit

[?, ?]

2.3 Samtools Mpileup

[?]

2.4 FreeBayes

[?]

- **2.5** SNAP
- 2.6 SMaSH
- 3. ARCHITECTURE

3.1 Local Assembly

From the assembled haplotypes, we order them according to the haplotype likelihood:

$$\mathcal{L}(H_j) \equiv \mathbf{P}(\{r_i\}|H_j) \tag{1}$$

$$= \prod_{i} \mathbf{P}(r_i|H_j) \tag{2}$$

where $\mathbf{P}(r_i|H_j)$ is obtained from aligning the read and the candidate haplotype by a pairwise HMM alignment. Among the ordered haplotypes, we pick the top scoring haplotypes and ignore the low scoring ones. The likelihood of observing the reads $\{r_i\}$, given a pair of haplotypes H_j and $H_{j'}$, is defined to be:

$$\mathcal{L}(H_j, H_{j'}) \equiv \mathbf{P}(\{r_i\} | H_j, H_{j'}) \tag{3}$$

$$= \prod_{i} \left[\frac{\mathbf{P}(r_i|H_j)}{2} + \frac{\mathbf{P}(r_i|H_{j'})}{2} \right]. \tag{4}$$

We compute the posterior probability of observing the pair of haplotypes H_j and $H_{j'}$ from the haplotype pair likelihood and a haplotype pair prior probability:

$$\mathbf{P}(H_j, H_{j'} | \{r_i\}) = \frac{1}{Z} \mathcal{L}(H_j, H_{j'}) \mathbf{P}(H_j, H_{j'})$$
 (5)

where Z is a normalization:

$$Z = \sum_{j} \sum_{j'} \mathcal{L}(H_j, H_{j'}) \mathbf{P}(H_j, H_{j'})$$

and where we obtain the prior $\mathbf{P}(H_j,H_{j'})$ by aligning the haplotype pair with the same pairwise HMM alignment as above, and taking the product of the prior probabilities for each SNP and indel event.

We choose the maximum a priori estimate among haplotypes with any variants as the called non-reference maternal and paternal haplotype pair:

$$(H_{\text{mat}}^{\text{nonref}}, H_{\text{pat}}^{\text{nonref}}) = \arg \max_{H_j, H_{j'}: n_{\text{var}}(H_j, H_{j'}) > 0} \mathbf{P}(H_j, H_{j'} | \{r_i\}).$$

$$(6)$$

Similarly, we may define the reference haplotype pair as $(H_{\text{mat}}^{\text{ref}}, H_{\text{pat}}^{\text{ref}})$. The error probability of calling the non-reference haplotype pair is:

$$\mathbf{P}_{\text{error}}(H_{\text{mat}}^{\text{nonref}}, H_{\text{pat}}^{\text{nonref}}) = \frac{\mathbf{P}(H_{\text{mat}}^{\text{ref}}, H_{\text{pat}}^{\text{ref}})}{\mathbf{P}(H_{\text{mat}}^{\text{nonref}}, H_{\text{pat}}^{\text{nonref}}) + \mathbf{P}(H_{\text{mat}}^{\text{ref}}, H_{\text{pat}}^{\text{ref}})}.$$
(7)

The quality score of all variants present in the nonreference haplotype pair is defined as the Phred scaling of $\mathbf{P}_{\text{error}}$.

- 3.2 SNP Calling
- 3.3 Joint Variant Calling
- 3.4 Algorithm Selection
- 4. EVALUATION
- 4.1 Accuracy
- 4.2 Performance
- 5. FUTURE WORK
- 6. CONCLUSION

APPENDIX

A. AVAILABILITY

 ${\tt avocado}$ is open source and is licensed under the Apache 2 license. The source code is available at:

http://www.github.com/bigdatagenomics/avocado