

7393 Exam 2 R Code Andrei Matveev

```
#library(data.table)
library(truncnorm)
library(coda)
#devtools::install_github("stan-dev/loo")
library(loo)

## This is loo version 2.4.0
## - Online documentation and vignettes at mc-stan.org/loo
## - As of v2.0.0 loo defaults to 1 core but we recommend using as many as possible. Use the 'cores' arg
library(rstan)

## Loading required package: StanHeaders
## Loading required package: ggplot2
## rstan (Version 2.21.2, GitRev: 2e1f913d3ca3)
## For execution on a local, multicore CPU with excess RAM we recommend calling
## options(mc.cores = parallel::detectCores()).
## To avoid recompilation of unchanged Stan programs, we recommend calling
## rstan_options(auto_write = TRUE)

##
## Attaching package: 'rstan'
## The following object is masked from 'package:coda':
##
##   traceplot
library(shinystan)

## Loading required package: shiny
##
## This is shinystan version 2.5.0
library(invgamma)
library(bayesplot)

## This is bayesplot version 1.7.2
## - Online documentation and vignettes at mc-stan.org/bayesplot
## - bayesplot theme set to bayesplot::theme_default()
##   * Does _not_ affect other ggplot2 plots
##   * See ?bayesplot_theme_set for details on theme setting
library(posterior)
library(rstanarm)

## Loading required package: Rcpp
```

```

## This is rstanarm version 2.21.1
## - See https://mc-stan.org/rstanarm/articles/priors for changes to default priors!
## - Default priors may change, so it's safest to specify priors, even if equivalent to the defaults.
## - For execution on a local, multicore CPU with excess RAM we recommend calling
##   options(mc.cores = parallel::detectCores())
##
## Attaching package: 'rstanarm'
## The following object is masked from 'package:rstan':
##
##   loo
devtools::install_github("jfrench/bayesutils")

## Skipping install of 'bayesutils' from a github remote, the SHA1 (17473e8d) has not changed since last
##   Use `force = TRUE` to force installation
setwd('/Users/AM/Documents/_CU Masters/2020 fall Bayesian_7393/code/Bayesian_Statistics_Class_Code/Exam')
#df1 = load("diamonds_simple.rda")
#rm(df1, diamonds_simple)
df = bayesutils::diamonds_simple

df$lprice = log(df$price)
df$lcarat = log(df$carat)

df_covid = bayesutils::covid_dec4

lq_theta_y = function(sigmaSQ, beta0, beta1, lpr = df$lprice, lcar = df$lcarat) {
  ld = dnorm(beta0, 0, 100, log = TRUE) +
    dnorm(beta1, 0, 100, log = TRUE) +
    dinvgamma(sigmaSQ, shape = 0.01, rate = 0.01, log = TRUE) +
    sum(dnorm(lpr, mean = (beta0 + beta1 * lcar), sd = sqrt(sigmaSQ), log = TRUE))
  return(ld)
}

mh = function(B, theta_start) {
  theta = array(0, c((B+1), 3), dimnames = list(c(), c("sigmaSQ", "beta0", "beta1")))

  theta[1,1] = theta_start[1]
  theta[1,2] = theta_start[2]
  theta[1,3] = theta_start[3]

  for (i in 2:dim(theta)[1]) {

    ### step for sigmaSQ
    beta0_star = theta[(i-1),2]
    beta1_star = theta[(i-1),3]

    sigmaSQ_star = rtruncnorm(n = 1, a=0, b=Inf, mean = theta[(i-1),1], sd = 0.1)

    num_logr = lq_theta_y(sigmaSQ = sigmaSQ_star, beta0 = beta0_star, beta1 = beta1_star) -
      log(dtruncnorm(x = sigmaSQ_star, a=0, b=Inf, mean = theta[(i-1),1], sd = 0.1))
    den_logr = lq_theta_y(sigmaSQ = theta[i-1, 1], beta0 = beta0_star, beta1 = beta1_star) -
      log(dtruncnorm(x = theta[i-1, 1], a=0, b=Inf, mean = sigmaSQ_star, sd = 0.1))
  }
}

```

```

logr = num_logr - den_logr
if (log(runif(1)) <= min(logr, 0)) {
  theta[i,1] = sigmaSQ_star
} else {
  theta[i,1] = theta[(i - 1), 1]
}

### step for beta0
beta1_star = theta[(i-1),3] # it is the repeated code, but I need it to keep the interpretability
sigmaSQ_star = theta[i,1] # update sigmaSQ_star after the Gibbs step for sigmaSQ

beta0_star = rnorm(1, theta[(i-1),2], 0.1) # !!! check the parametrization (0.1 or 0.1^2)

num_logr = lq_theta_y(sigmaSQ = sigmaSQ_star, beta0 = beta0_star, beta1 = beta1_star) -
  dnorm(x = beta0_star, mean = theta[(i-1),2], sd = 0.1, log = TRUE)
den_logr = lq_theta_y(sigmaSQ = sigmaSQ_star, beta0 = theta[i-1, 2], beta1 = beta1_star) -
  dnorm(x = theta[(i-1),2], mean = beta0_star, sd = 0.1, log = TRUE)

logr = num_logr - den_logr
if (log(runif(1)) <= min(logr, 0)) {
  theta[i,2] = beta0_star
} else {
  theta[i,2] = theta[(i - 1), 2]
}

### step for beta1
sigmaSQ_star = theta[i,1] # it is the repeated code, but I need it to keep the interpretability
beta0_star = theta[i,2] # update beta0_star after the Gibbs step for beta0

beta1_star = rnorm(1, theta[(i-1),3], 0.1) #

num_logr = lq_theta_y(sigmaSQ = sigmaSQ_star, beta0 = beta0_star, beta1 = beta1_star) -
  dnorm(x = beta1_star, mean = theta[(i-1),3], sd = 0.1, log = TRUE)
den_logr = lq_theta_y(sigmaSQ = sigmaSQ_star, beta0 = beta0_star, beta1 = theta[i-1, 3]) -
  dnorm(x = theta[(i-1),3], mean = beta1_star, sd = 0.1, log = TRUE)

logr = num_logr - den_logr
if (log(runif(1)) <= min(logr, 0)) {
  theta[i,3] = beta1_star
} else {
  theta[i,3] = theta[(i - 1), 3]
}
}
return(theta)
}

B = 10^5
keep = (B/2 + 1):(B + 1)
chain1 = mh(B, theta_start = c(0.1, -1, -1))

```

```

chain2 = mh(B, theta_start = c(0.3, 0, 0))
chain3 = mh(B, theta_start = c(0.5, -1, 1))
chain4 = mh(B, theta_start = c(0.2, 1, 1))

mc = mcmc.list(mcmc(chain1[keep,]), mcmc(chain2[keep,]),
               mcmc(chain3[keep,]), mcmc(chain4[keep,]))
summary(mc)

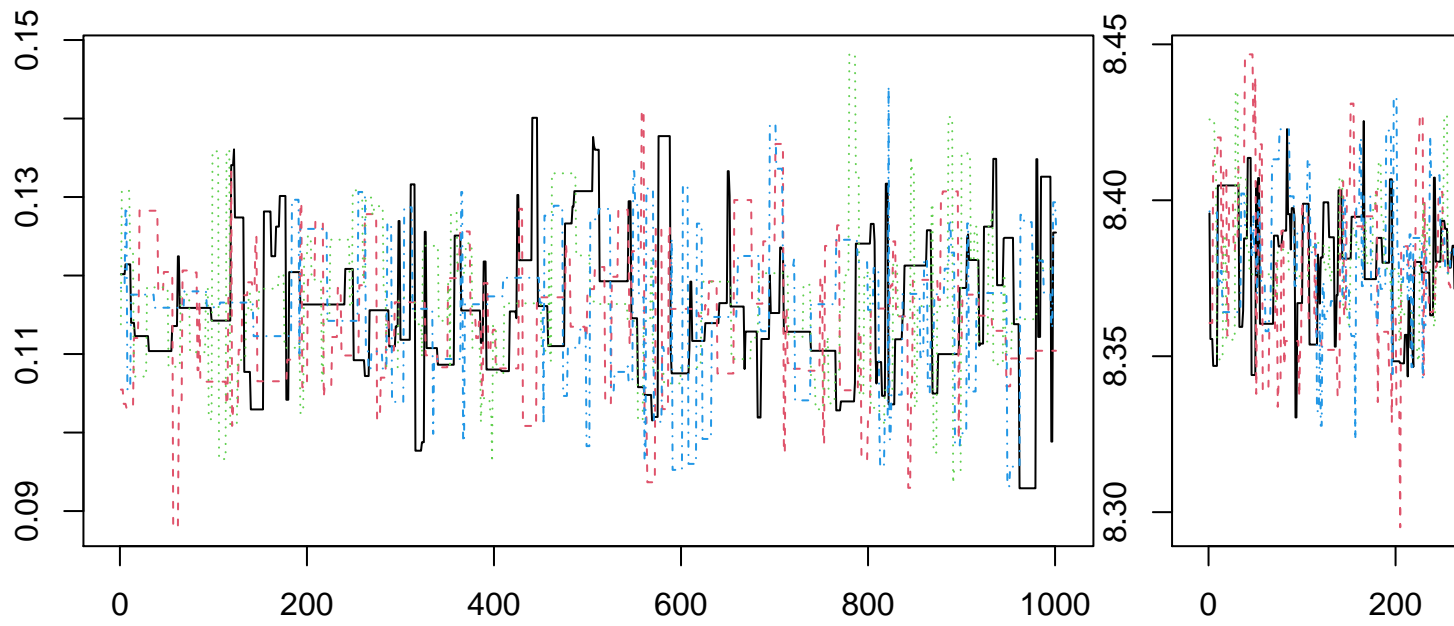
##
## Iterations = 1:50001
## Thinning interval = 1
## Number of chains = 4
## Sample size per chain = 50001
##
## 1. Empirical mean and standard deviation for each variable,
##    plus standard error of the mean:
##
##           Mean          SD Naive SE Time-series SE
## sigmaSQ 0.1151 0.009198 2.057e-05    7.053e-05
## beta0    8.3795 0.022435 5.017e-05    1.643e-04
## beta1    1.5061 0.031364 7.013e-05    2.142e-04
##
## 2. Quantiles for each variable:
##
##           2.5%    25%    50%    75%    97.5%
## sigmaSQ 0.09875 0.1086 0.1146 0.1209 0.1346
## beta0    8.33571 8.3643 8.3796 8.3947 8.4234
## beta1    1.44474 1.4851 1.5060 1.5274 1.5677

keep = (B/2 + 17001):(B/2 + 18001)

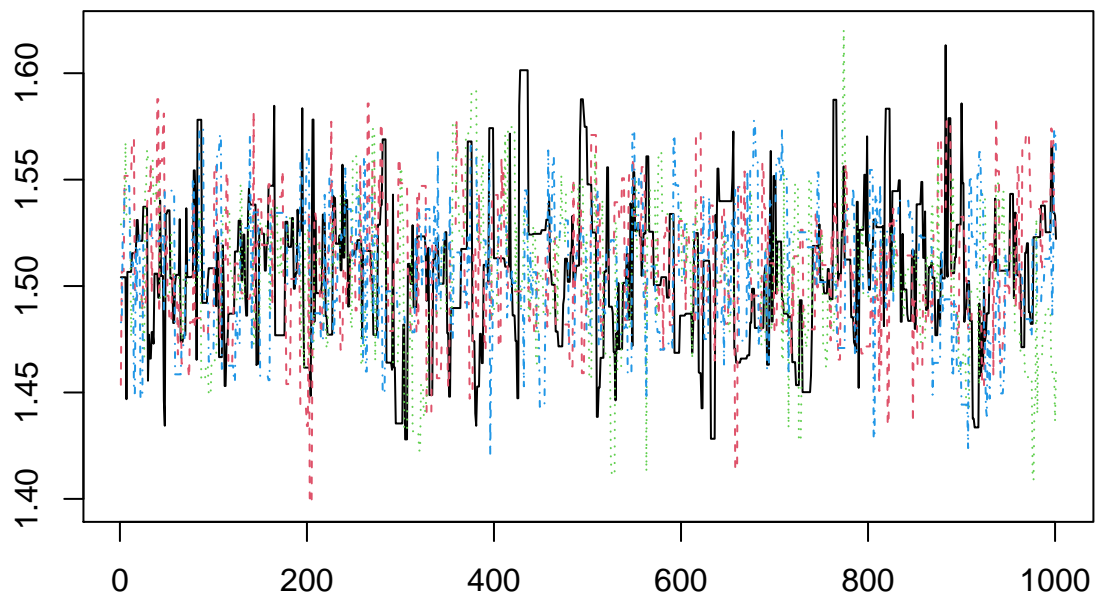
mc = mcmc.list(mcmc(chain1[keep,]), mcmc(chain2[keep,]),
               mcmc(chain3[keep,]), mcmc(chain4[keep,]))
coda::traceplot(mc)

```

Trace of sigmaSQ

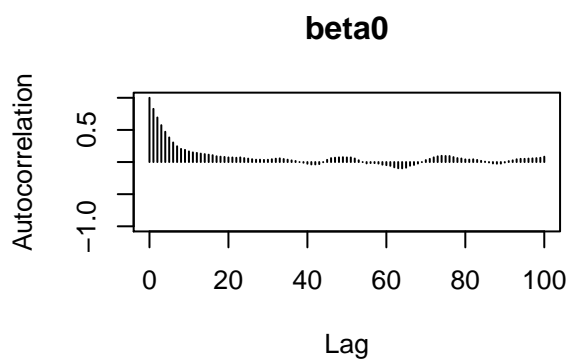
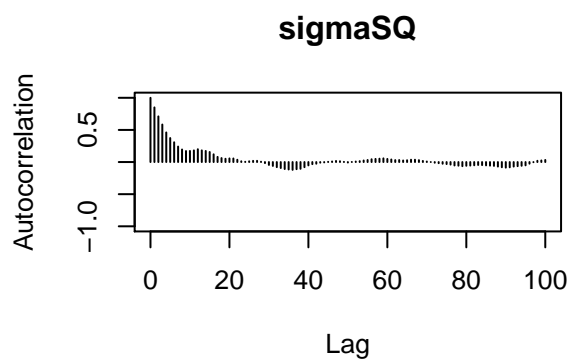
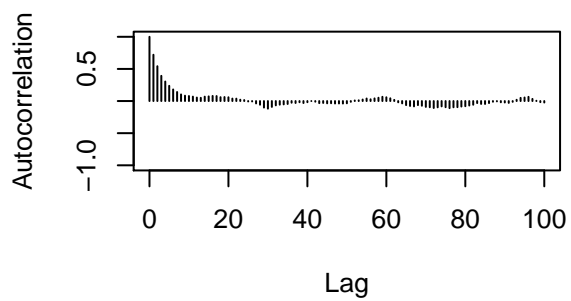
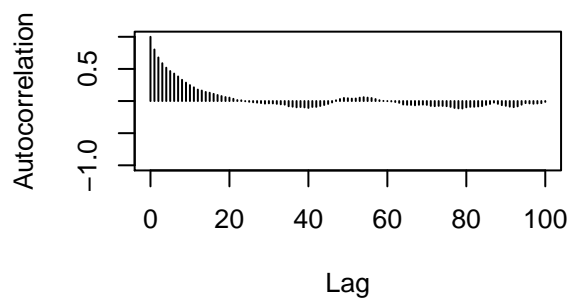
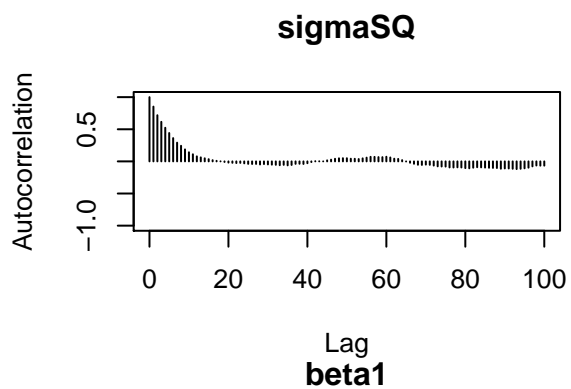
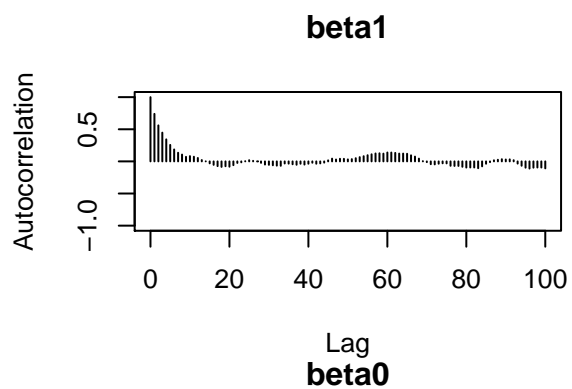
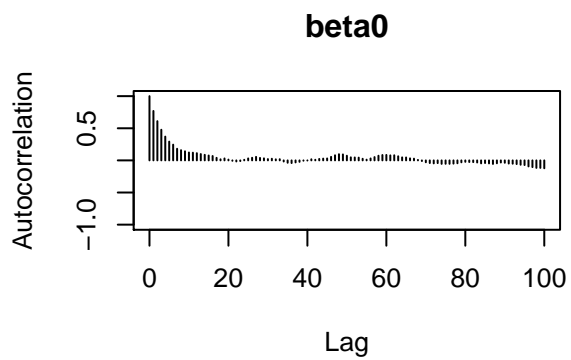
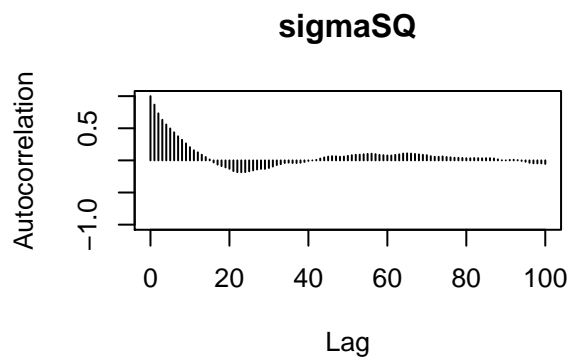


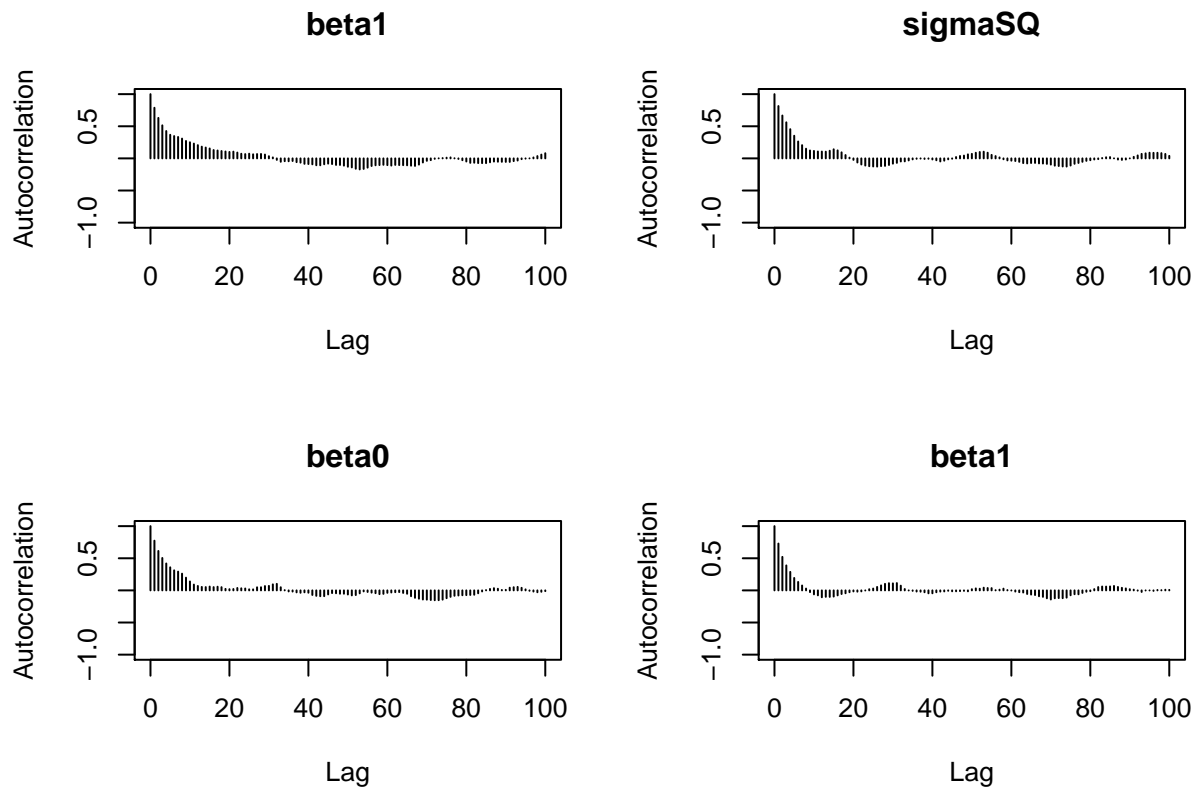
Iterations
Trace of beta1



Iterations

```
coda::autocorr.plot(mc, lag.max = 100, auto.layout = TRUE)
```





```
gelman.diag(mc, autoburnin = FALSE)
```

```
## Potential scale reduction factors:
##
##      Point est. Upper C.I.
## sigmaSQ      1.00      1.01
## beta0        1.01      1.01
## beta1        1.00      1.00
##
## Multivariate psrf
##
## 1
```

```
geweke.diag(mc)
```

```
## [[1]]
##
## Fraction in 1st window = 0.1
## Fraction in 2nd window = 0.5
##
## sigmaSQ  beta0  beta1
## -0.40871  0.06296  0.40829
##
##
## [[2]]
##
## Fraction in 1st window = 0.1
## Fraction in 2nd window = 0.5
##
## sigmaSQ  beta0  beta1
```

```

## -0.2999  0.2575 -0.3738
##
##
## [[3]]
##
## Fraction in 1st window = 0.1
## Fraction in 2nd window = 0.5
##
## sigmaSQ    beta0    beta1
## 0.2173    0.8936    0.2712
##
##
## [[4]]
##
## Fraction in 1st window = 0.1
## Fraction in 2nd window = 0.5
##
## sigmaSQ    beta0    beta1
## 0.9673    0.7504    0.4157

# extracting the MCMC samples from the soda_fit object
samples = extract(r_fit_A)
ncycles = length(samples[[1]])
T <- length(df$price)

# each row of yrep is a sample from the pp distribution
yrep = matrix(0, ncol = T, nrow = ncycles)
for (i in seq_len(T)) {
  mui = samples$beta0 + samples$beta1 * df$lcarat[i]
  yrep[, i] = rnorm(ncycles, mean = mui, sd = sqrt(samples$sigmaSQ))
}

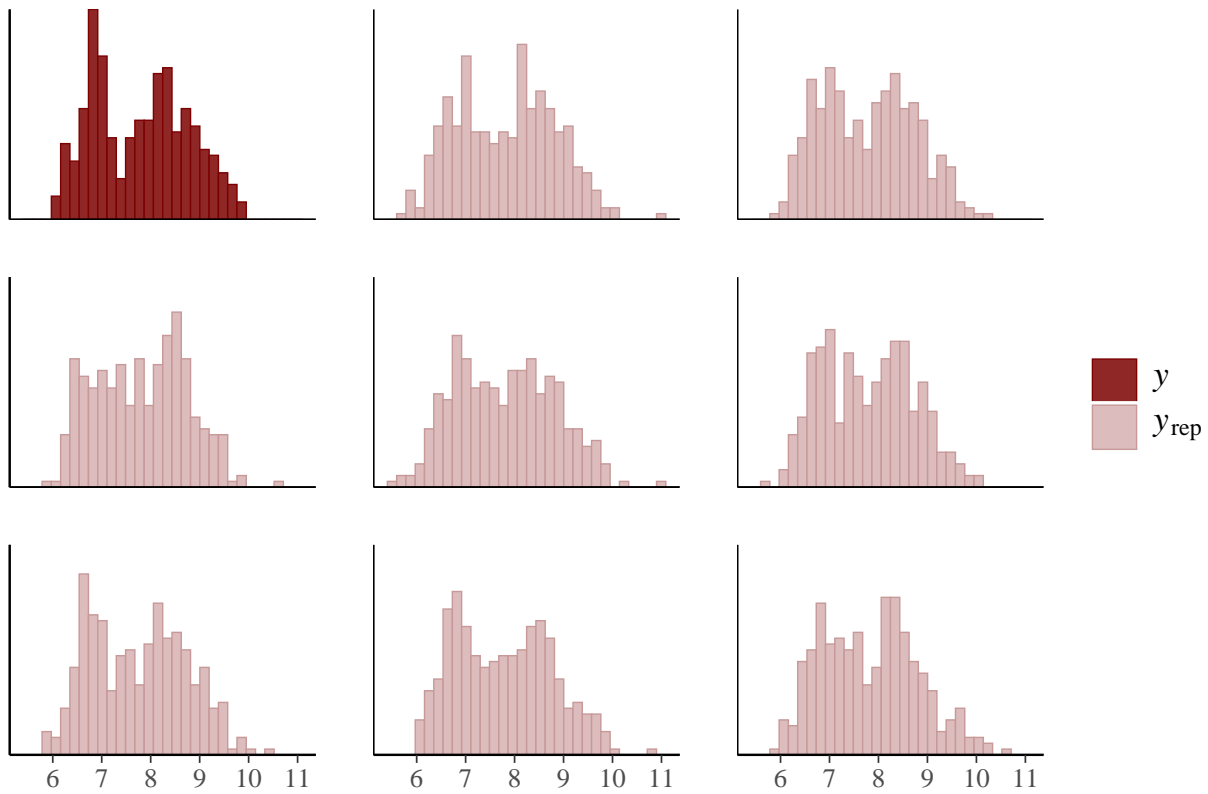
# approximate posterior predictive density for an observation
# with the same covariates as observation 1
#plot(density(yrep[,1]))
color_scheme_set("red")

# posterior predictive check
y = df$lprice
ppc_hist(y, yrep[sample(1:ncycles, 8),]) + ggtitle("Model A: histogram comparing the response to 8 repl.

## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.

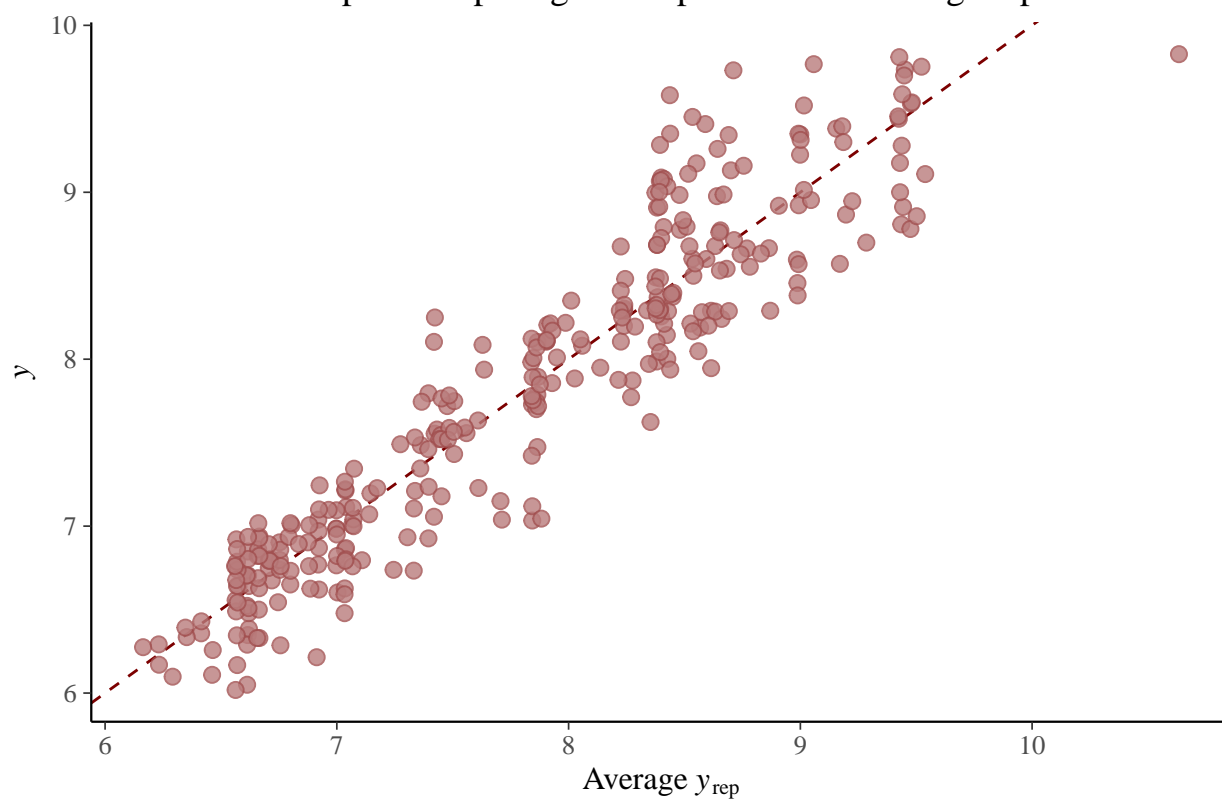
```


Model A: histogram comparing the response to 8 replicated data sets



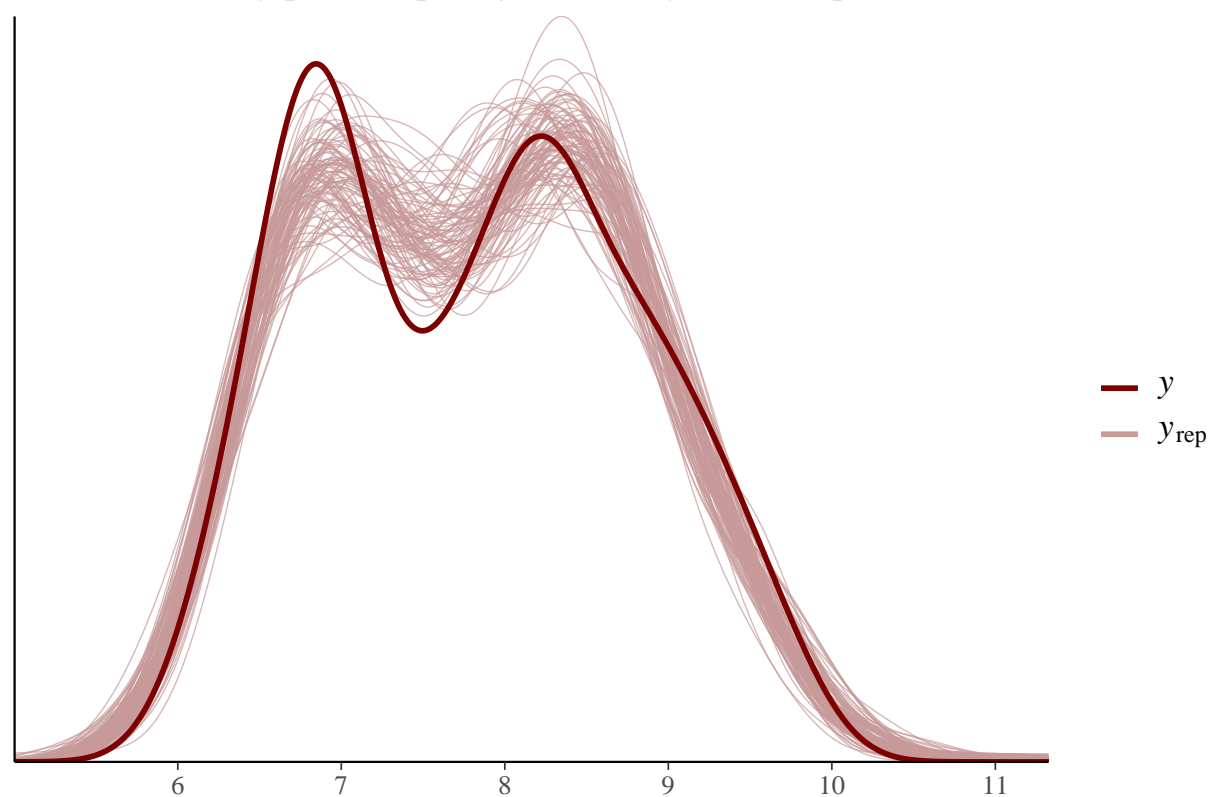
```
# scatterplot of y vs average yrep
ppc_scatter_avg(y, yrep) + ggtitle("Model A: scatterplot comparing the response to the average replicat")
```

Model A: scatterplot comparing the response to the average replicated data se



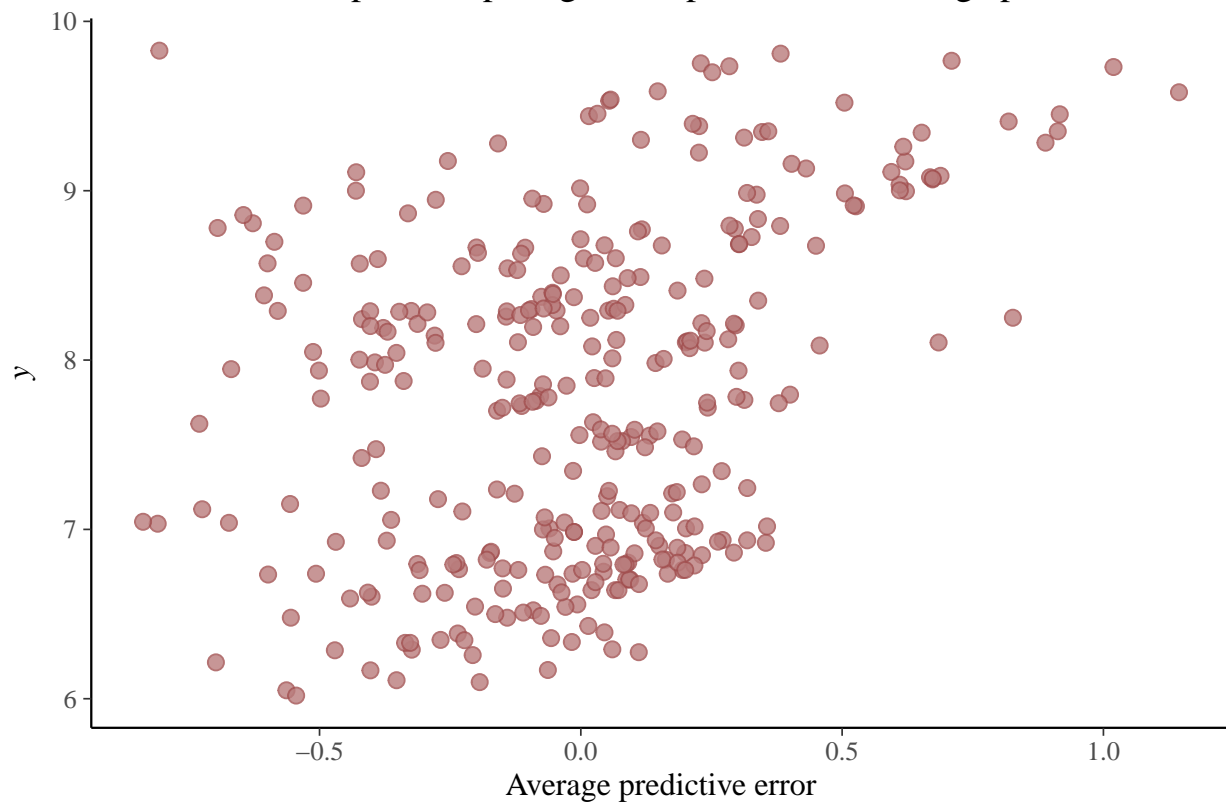
```
ppc_dens_overlay(y, yrep = yrep[sample(1:ncycles, 100),]) + ggtitle("Model A: density plot comparing the response to the average replicated data")
```

Model A: density plot comparing the density of the response to the densities of 10



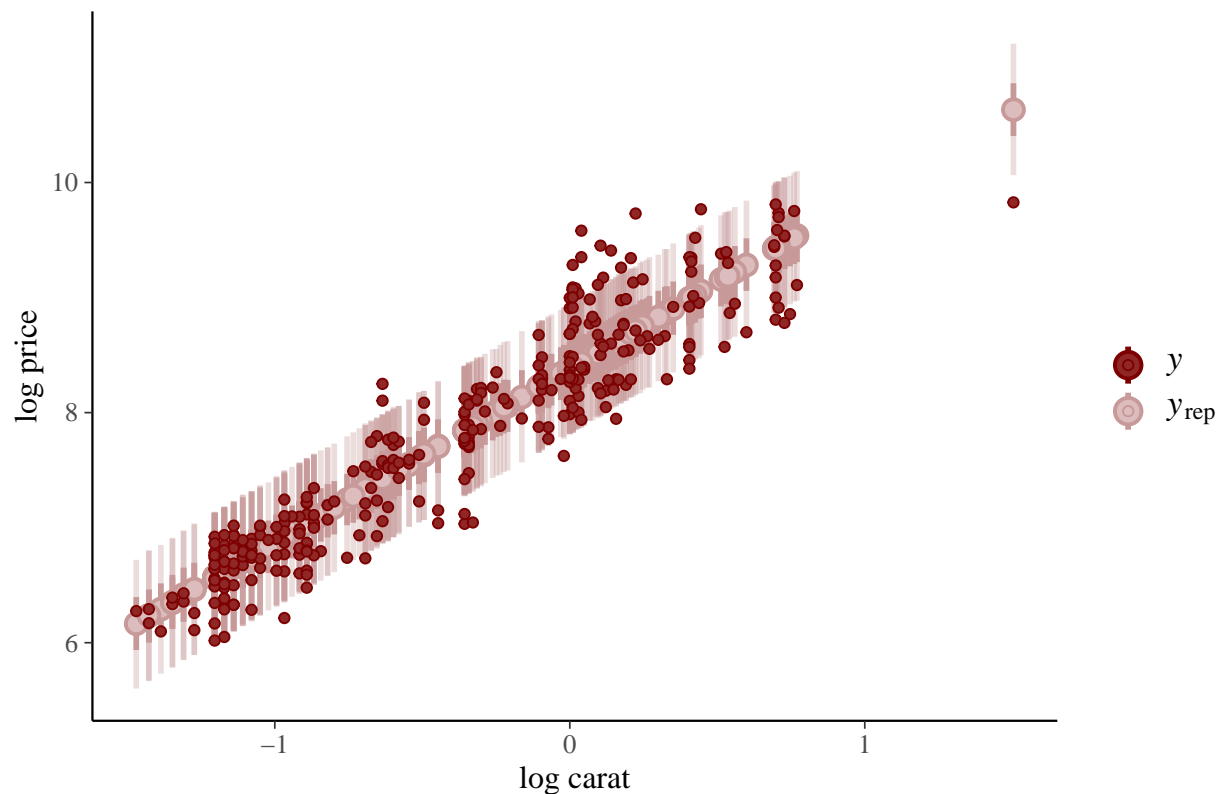
```
ppc_error_scatter_avg(y, yrep = yrep) + ggtitle("Model A: scatter plot comparing the response to the av
```

Model A: scatter plot comparing the response to the average predictive error



```
ppc_intervals(y = y, yrep = yrep, x = df$lcarat) + ggtitle("Model A: Posterior predictive intervals to
```

Model A: Posterior predictive intervals to the observed data values



```
# extracting the MCMC samples from the soda_fit object
samples = extract(r_fit_B)
ncycles = length(samples[[1]])
T <- length(df$price)

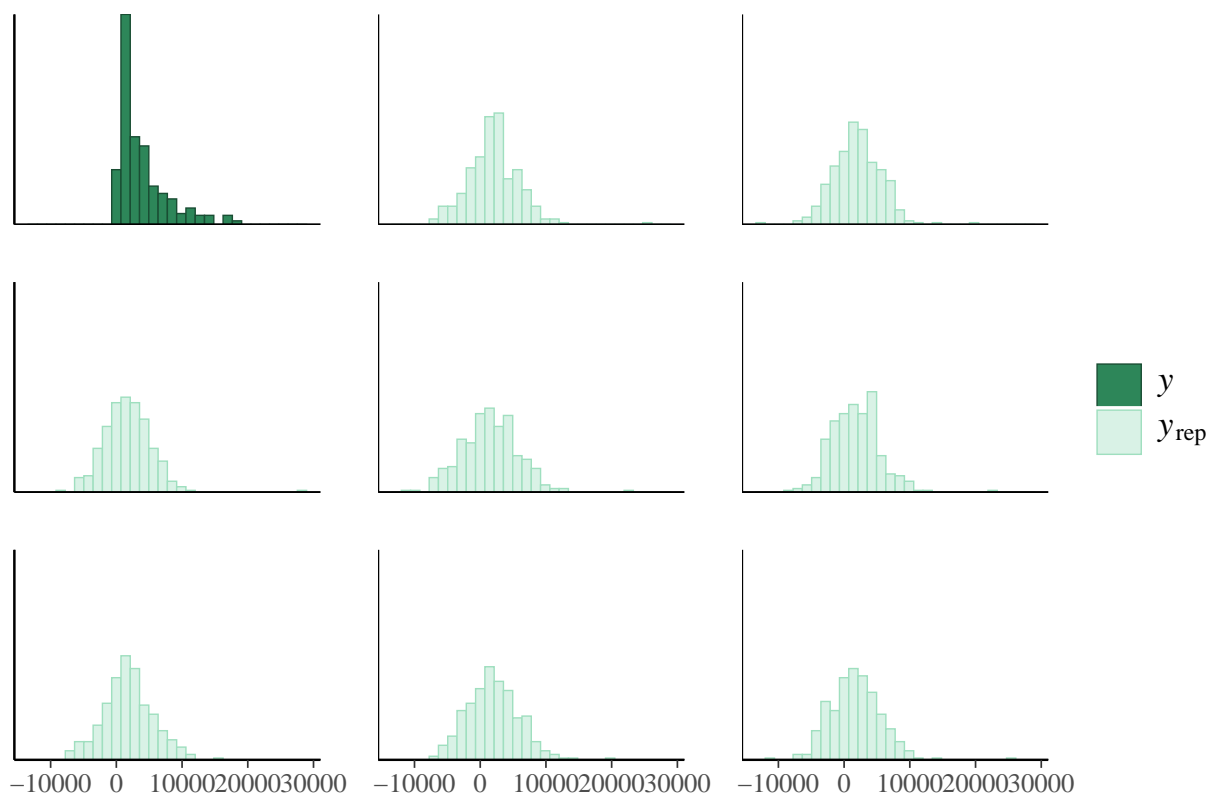
# each row of yrep is a sample from the pp distribution
yrep = matrix(0, ncol = T, nrow = ncycles)
for (i in seq_len(T)) {
  mui = samples$beta0 + samples$beta1 * df$carat[i] + samples$beta2 * df$carat[i] * df$carat[i]
  yrep[, i] = rnorm(ncycles, mean = mui, sd = sqrt(samples$sigmaSQ))
}

# approximate posterior predictive density for an observation
# with the same covariates as observation 1
#plot(density(yrep[,1]))
color_scheme_set("green")

# posterior predictive check
y = df$price
ppc_hist(y, yrep[sample(1:ncycles, 8),]) + ggtitle("Model B: histogram comparing the response to 8 repl.

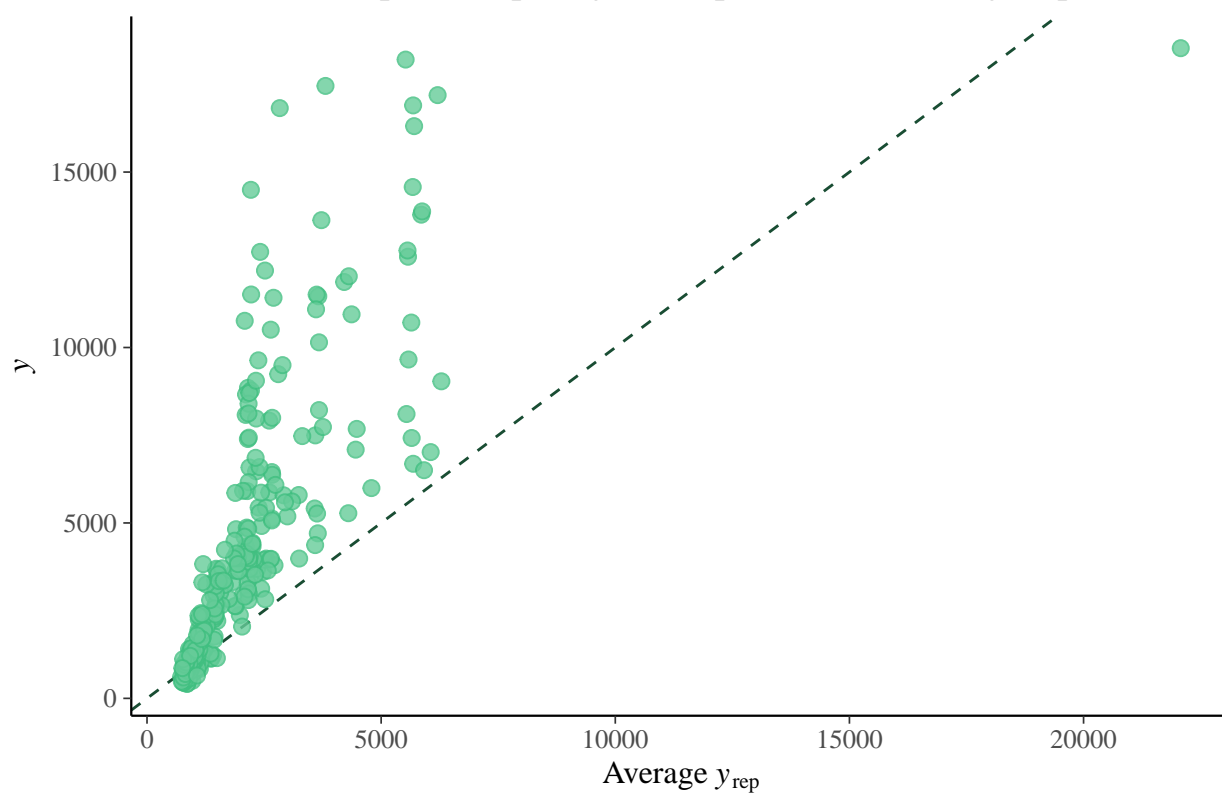
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

Model B: histogram comparing the response to 8 replicated data sets



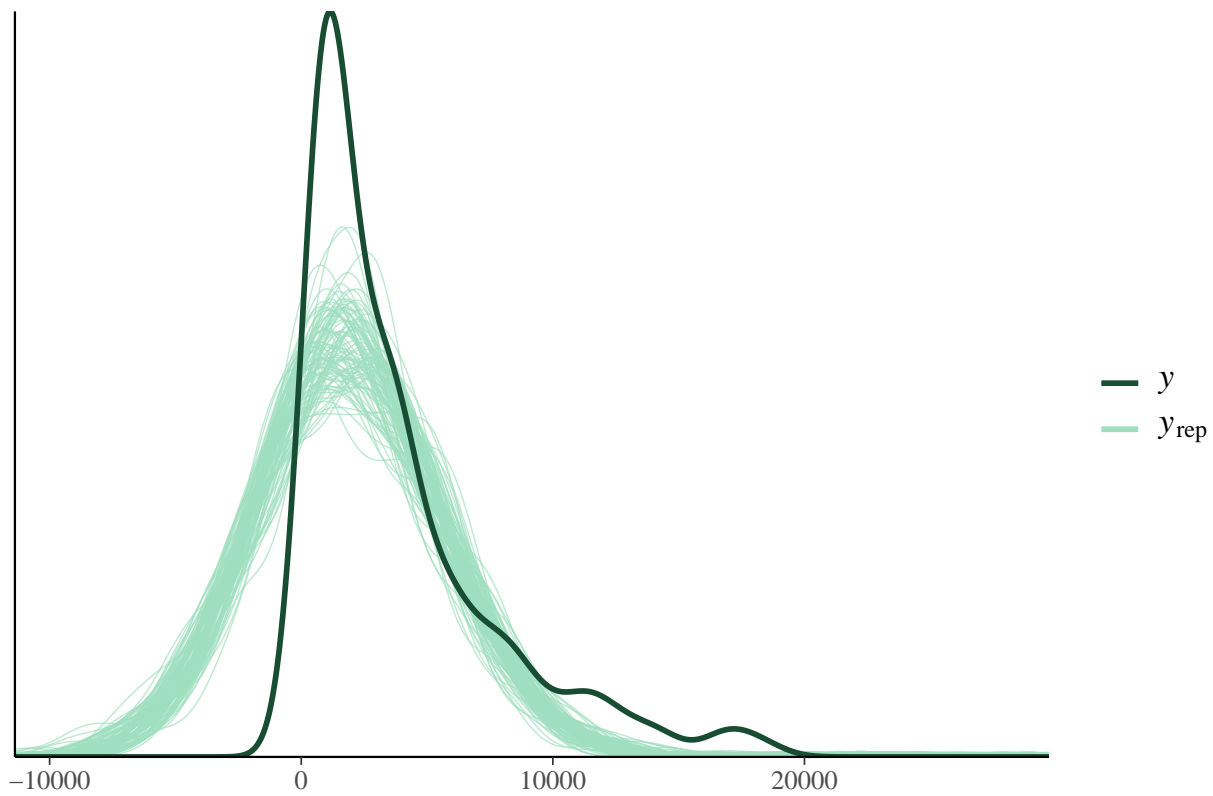
```
# scatterplot of y vs average yrep
ppc_scatter_avg(y, yrep) + ggtitle("Model B: scatterplot comparing the response to the average replicat")
```

Model B: scatterplot comparing the response to the average replicated data



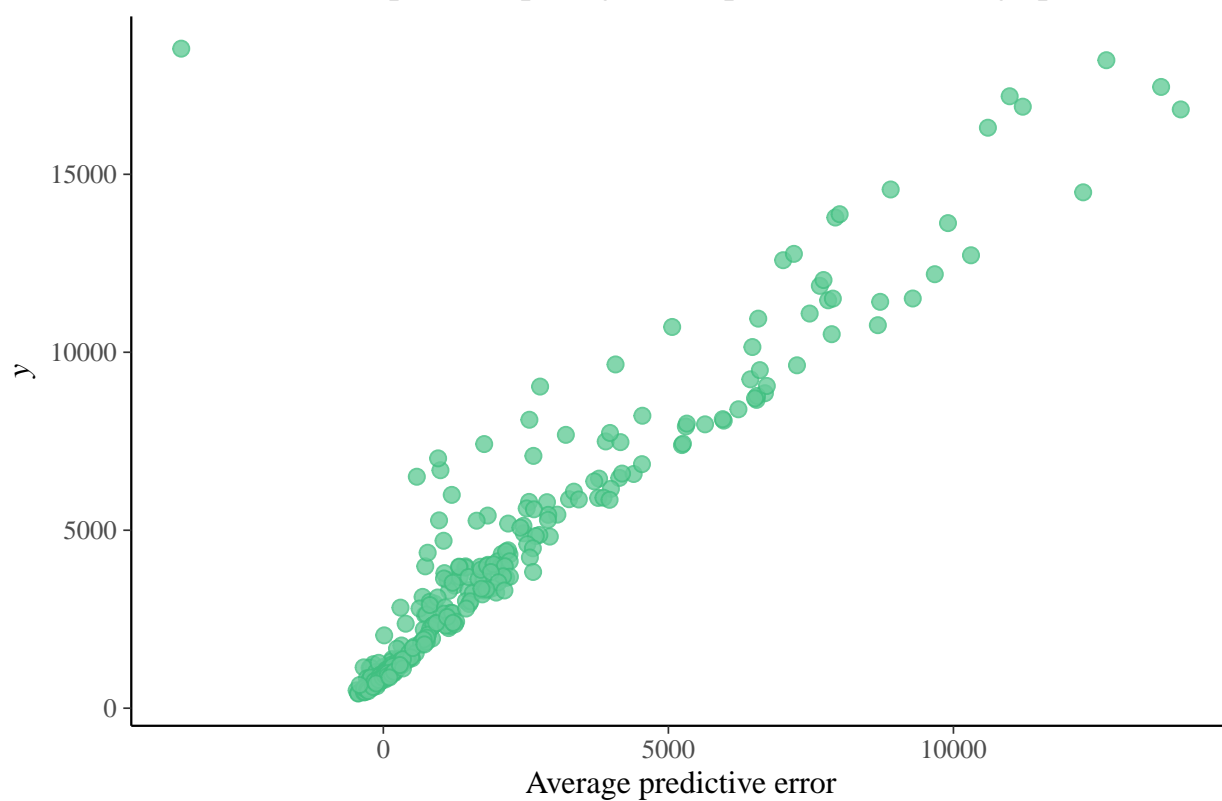
```
ppc_dens_overlay(y, yrep = yrep[sample(1:ncycles, 100),]) + ggtitle("Model B: density plot comparing the response to the average replicated data")
```

Model B: density plot comparing the density of the response to the densities of 10



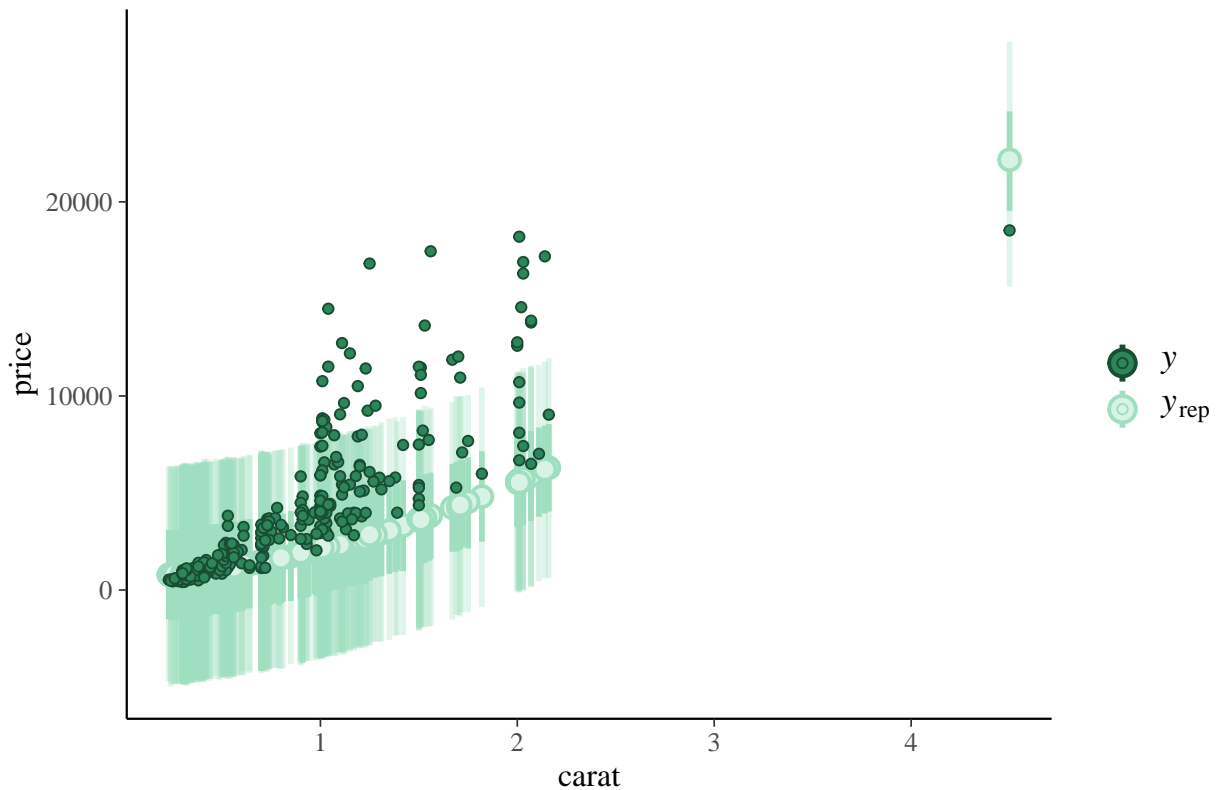
```
ppc_error_scatter_avg(y, yrep = yrep) + ggtitle("Model B: scatter plot comparing the response to the av
```


Model B: scatter plot comparing the response to the average predictive error



```
ppc_intervals(y = y, yrep = yrep, x = df$carat) + ggtitle("Model B: Posterior predictive intervals to t
```

Model B: Posterior predictive intervals to the observed data values



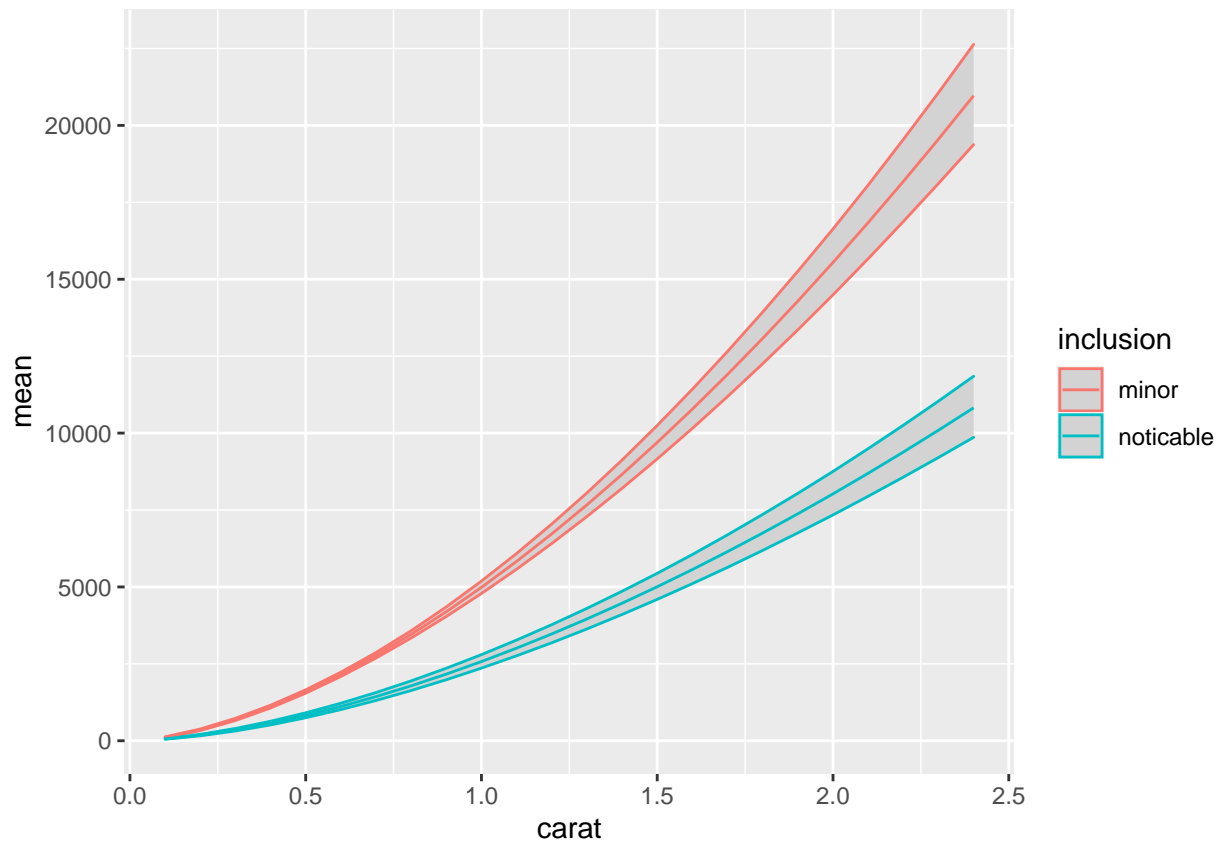
```
sim_cond_mu_1 = rstan::extract(r_fit_D)$cond_mu_1
df_sim_cond_mu_1 = data.frame(
  carat = seq(0.1, 2.4, by=0.1),
  mean = apply(sim_cond_mu_1, 2, mean),
  lo0025 = apply(sim_cond_mu_1, 2, quantile, 0.025),
  hi0975 = apply(sim_cond_mu_1, 2, quantile, 0.975),
  inclusion = "noticable"
)

sim_cond_mu_0 = rstan::extract(r_fit_D)$cond_mu_0
df_sim_cond_mu_0 = data.frame(
  carat = seq(0.1, 2.4, by=0.1),
  mean = apply(sim_cond_mu_0, 2, mean),
  lo0025 = apply(sim_cond_mu_0, 2, quantile, 0.025),
  hi0975 = apply(sim_cond_mu_0, 2, quantile, 0.975),
  inclusion = "minor"
)

df_sim = rbind(df_sim_cond_mu_0, df_sim_cond_mu_1)
rm(df_sim_cond_mu_0, df_sim_cond_mu_1)

ggplot(df_sim,
  aes(x = carat,
    y = mean, colour = inclusion)) +
  geom_ribbon(aes(ymin = lo0025,
    ymax = hi0975),
    fill = "lightgrey") +
```

```
geom_line()
```



```
# samples = extract(r_fit_D)
# ncycles = length(samples[[1]])
# T <- length(df$price)
# #each row of yrep is a sample from the pp distribution
# yrep = matrix(0, ncol = T, nrow = ncycles)
# for (i in seq_len(T)) {
#   mu_i = samples$beta0 + samples$beta1 * df$carat[i] +
#     samples$beta2 * as.integer(df$inclusions[i]=="noticeable")
#   yrep[, i] = rlnorm(ncycles, mean = mu_i, sd = sqrt(samples$sigmaSQ))
# }
# df$notic = as.integer(df$inclusions == "noticeable")
# color_scheme_set("blue")
# ppc_intervals_grouped(y = df$price, yrep = yrep, x = df$carat, prob = 0.5, prob_outer = .95, group =
# #
# #
# find_ep = function(yrep, min = 0.1, max = 2.4, step=0.1) {
#   ep = array(0, dim=c((max-min+step)/step, 5), dimnames = list(c(), c("carat_min", "carat_max", "2.5%",
#   n=0
#   for (i in seq(min+step, max, by=step)) {
#     n=n+1
#     subset = ((df$carat>(i-step)) & (df$carat<=i))
#     yrep_subset = yrep[,subset]
#     ep[n,1] = i-step
#     ep[n,2] = i
#     q = quantile(yrep_subset, probs = c(0.025, 0.975))
```

```

#   ep[n,3] = q[[1]]
#   ep[n,4] = mean(yrep_subset)
#   ep[n,5] = q[[2]]
# }
# return(ep)
# }
# find_ep(yrep)
# temp = ppc_intervals_data(y = df$price, yrep = yrep, x = df$lcarat, prob = 0.5, prob_outer = .95, gro

cat("STAN Model E WAIC", waic_loo(r_fit_E)[1])

## Warning:
## 46 (92.0%) p_waic estimates greater than 0.4. We recommend trying loo instead.

## Warning: Accessing waic using '$' is deprecated and will be removed in a
## future release. Please extract the waic estimate from the 'estimates' component
## instead.

## Warning: Some Pareto k diagnostic values are too high. See help('pareto-k-diagnostic') for details.

## Warning: Accessing looic using '$' is deprecated and will be removed in a
## future release. Please extract the looic estimate from the 'estimates' component
## instead.

## STAN Model E WAIC 68766.75

cat("STAN Model E LOOIC", waic_loo(r_fit_E)[2])

## Warning:
## 46 (92.0%) p_waic estimates greater than 0.4. We recommend trying loo instead.

## Warning: Accessing waic using '$' is deprecated and will be removed in a
## future release. Please extract the waic estimate from the 'estimates' component
## instead.

## Warning: Some Pareto k diagnostic values are too high. See help('pareto-k-diagnostic') for details.

## Warning: Accessing looic using '$' is deprecated and will be removed in a
## future release. Please extract the looic estimate from the 'estimates' component
## instead.

## STAN Model E LOOIC 61810.17

setwd('/Users/AM/Documents/_CU Masters/2020 fall Bayesian_7393/code/Bayesian_Statistics_Class_Code/Exam
r_fit_E = readRDS(file = "Model_E_2020-12-07.rda")

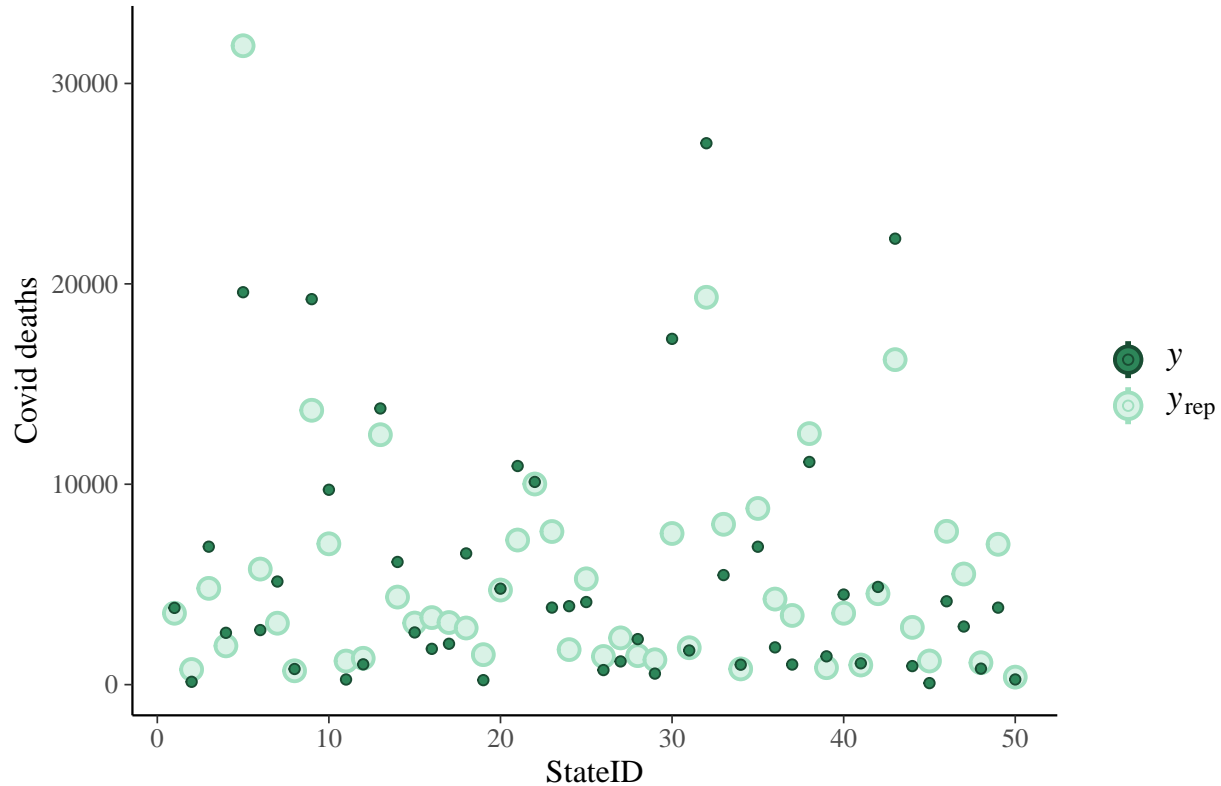
samples = extract(r_fit_E)
ncycles = length(samples[[1]])
T <- length(df_covid$bs)

# each row of yrep is a sample from the pp distribution
yrep = matrix(0, ncol = T, nrow = ncycles)
for (i in seq_len(T)) {
  log_l = log(df_covid$population[i]) +
    samples$beta0 +
    samples$beta1 * df_covid$income[i] +
    samples$beta2 * df_covid$bs[i]
  yrep[, i] = rpois(ncycles, lambda = exp(log_l))
}

```

```
ppc_intervals(y = df_covid$deaths, yrep = yrep, prob = 0.5, prob_outer = .95) +
  ggtitle("Model E: Posterior predictive intervals to the observed data values") +
  xlab("StateID") + ylab("Covid deaths")
```

Model E: Posterior predictive intervals to the observed data values



```
dif = ppc_intervals_data(y = df_covid$deaths, yrep = yrep, prob = 0.5, prob_outer = .95)
dif$ratio = abs(dif$y_obs - dif$m) / dif$y_obs
cat("the worst prediction seems to be for StateID", which.max(dif$ratio),
    df_covid$state_name[which.max(dif$ratio)])
```

the worst prediction seems to be for StateID 45 Vermont

```
dif$abs = abs(dif$y_obs - dif$m)
cat("the worst prediction seems to be for StateID", which.max(dif$abs),
    df_covid$state_name[which.max(dif$abs)])
```

the worst prediction seems to be for StateID 5 California

```
-0.000054 * 30697
```

```
## [1] -1.657638
```

```
(24191 - 39682) * -0.000054
```

```
## [1] 0.836514
```

```
(17.50 - 39.00) * 0.060348
```

```
## [1] -1.297482
```

```
stan_mod_F = "
data {
```

```

int T;
int deaths[T]; // COVID-19 deaths
vector[T] log_pop; // log of U.S. states population,
vector[T] income; // median income (USD)
vector[T] bs; // percentage of the population with bachelor's degrees.
}

parameters {
  real beta0;
  real beta1;
  real beta2;
  real phi;
}
transformed parameters {
  real eta[T];
  for (i in 1:T) {
    eta[i] = log_pop[i] + beta0 + beta1 * income[i] + beta2 * bs[i];
  }
}

model {
  beta0 ~ normal(0, 5);
  beta1 ~ normal(0, 5);
  beta2 ~ normal(0, 5);
  phi ~ gamma(0.01, 0.01);

  for (i in 1:T) {
    deaths[i] ~ neg_binomial_2_log(eta[i], phi);
  }
}
generated quantities {
  vector[T] log_lik;

  for (i in 1:T) {
    log_lik[i] = neg_binomial_2_log_lpmf(deaths[i] | eta[i], phi);
  }
}

"

T <- length(df_covid$bs)
set.seed(95)

model_name = "Model_F_"
stan_dat_F = list(T = T, deaths = df_covid$deaths, log_pop = log(df_covid$population),
                  income = df_covid$income, bs = df_covid$bs)
r_fit_F = stan(model_code = stan_mod_F, data = stan_dat_F,
               iter = 5000, chains = 2,
               control = list(max_treedepth = 21))

```

Trying to compile a simple C file

```

## Running /Library/Frameworks/R.framework/Resources/bin/R CMD SHLIB foo.c
## clang -mmacosx-version-min=10.13 -I"/Library/Frameworks/R.framework/Resources/include" -DNDEBUG -I
## In file included from <built-in>:1:
## In file included from /Library/Frameworks/R.framework/Versions/4.0/Resources/library/StanHeaders/inc
## In file included from /Library/Frameworks/R.framework/Versions/4.0/Resources/library/RcppEigen/inclu
## In file included from /Library/Frameworks/R.framework/Versions/4.0/Resources/library/RcppEigen/inclu
## /Library/Frameworks/R.framework/Versions/4.0/Resources/library/RcppEigen/include/Eigen/src/Core/util
## namespace Eigen {
## ~
## /Library/Frameworks/R.framework/Versions/4.0/Resources/library/RcppEigen/include/Eigen/src/Core/util
## namespace Eigen {
## ~
## ;
## In file included from <built-in>:1:
## In file included from /Library/Frameworks/R.framework/Versions/4.0/Resources/library/StanHeaders/inc
## In file included from /Library/Frameworks/R.framework/Versions/4.0/Resources/library/RcppEigen/inclu
## /Library/Frameworks/R.framework/Versions/4.0/Resources/library/RcppEigen/include/Eigen/Core:96:10: f
## #include <complex>
## ~~~~~
## 3 errors generated.
## make: *** [foo.o] Error 1
##
## SAMPLING FOR MODEL '95322de15106226da123bef5b1199d71' NOW (CHAIN 1).
## Chain 1:
## Chain 1: Gradient evaluation took 2.7e-05 seconds
## Chain 1: 1000 transitions using 10 leapfrog steps per transition would take 0.27 seconds.
## Chain 1: Adjust your expectations accordingly!
## Chain 1:
## Chain 1:
## Chain 1: Iteration: 1 / 5000 [ 0%] (Warmup)
## Chain 1: Iteration: 500 / 5000 [ 10%] (Warmup)
## Chain 1: Iteration: 1000 / 5000 [ 20%] (Warmup)
## Chain 1: Iteration: 1500 / 5000 [ 30%] (Warmup)
## Chain 1: Iteration: 2000 / 5000 [ 40%] (Warmup)
## Chain 1: Iteration: 2500 / 5000 [ 50%] (Warmup)
## Chain 1: Iteration: 2501 / 5000 [ 50%] (Sampling)
## Chain 1: Iteration: 3000 / 5000 [ 60%] (Sampling)
## Chain 1: Iteration: 3500 / 5000 [ 70%] (Sampling)
## Chain 1: Iteration: 4000 / 5000 [ 80%] (Sampling)
## Chain 1: Iteration: 4500 / 5000 [ 90%] (Sampling)
## Chain 1: Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 1:
## Chain 1: Elapsed Time: 139.651 seconds (Warm-up)
## Chain 1: 39.5936 seconds (Sampling)
## Chain 1: 179.244 seconds (Total)
## Chain 1:
##
## SAMPLING FOR MODEL '95322de15106226da123bef5b1199d71' NOW (CHAIN 2).
## Chain 2: Rejecting initial value:
## Chain 2: Error evaluating the log probability at the initial value.
## Chain 2: Exception: neg_binomial_2_log_lpmf: Precision parameter is -1.43325, but must be > 0! (in
##
## Chain 2:
## Chain 2: Gradient evaluation took 1.4e-05 seconds

```

```
## Chain 2: 1000 transitions using 10 leapfrog steps per transition would take 0.14 seconds.
## Chain 2: Adjust your expectations accordingly!
## Chain 2:
## Chain 2:
## Chain 2: Iteration: 1 / 5000 [ 0%] (Warmup)
## Chain 2: Iteration: 500 / 5000 [ 10%] (Warmup)
## Chain 2: Iteration: 1000 / 5000 [ 20%] (Warmup)
## Chain 2: Iteration: 1500 / 5000 [ 30%] (Warmup)
## Chain 2: Iteration: 2000 / 5000 [ 40%] (Warmup)
## Chain 2: Iteration: 2500 / 5000 [ 50%] (Warmup)
## Chain 2: Iteration: 2501 / 5000 [ 50%] (Sampling)
## Chain 2: Iteration: 3000 / 5000 [ 60%] (Sampling)
## Chain 2: Iteration: 3500 / 5000 [ 70%] (Sampling)
## Chain 2: Iteration: 4000 / 5000 [ 80%] (Sampling)
## Chain 2: Iteration: 4500 / 5000 [ 90%] (Sampling)
## Chain 2: Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 2:
## Chain 2: Elapsed Time: 120.099 seconds (Warm-up)
## Chain 2: 38.6467 seconds (Sampling)
## Chain 2: 158.746 seconds (Total)
## Chain 2:
```

```
summary(r_fit_F, pars = c("beta0", "beta1", "beta2", "phi"), prob = c(0.025, 0.975))$summary
```

```
##              mean      se_mean      sd      2.5%      97.5%
## beta0 -4.352149141 1.716854e-02 7.685934e-01 -5.789551446 -2.757540e+00
## beta1 -0.000044994 1.199203e-06 5.047454e-05 -0.000144384  5.430147e-05
## beta2  0.039237130 8.639763e-04 3.806285e-02 -0.035612120  1.139286e-01
## phi    2.529451696 1.211503e-02 4.869649e-01  1.685299747  3.591891e+00
##              n_eff      Rhat
## beta0 2004.133 1.002128
## beta1 1771.574 1.001698
## beta2 1940.881 1.000783
## phi   1615.646 1.001139
```

```
#sso <- launch_shinystan(r_fit_F)
```

```
file_name = paste(model_name, as.character(Sys.Date()), ".rda", sep="")
```

```
setwd('/Users/AM/Documents/_CU Masters/2020 fall Bayesian_7393/code/Bayesian_Statistics_Class_Code/Exam
```

```
cat("STAN Model F WAIC", waic_loo(r_fit_F)[1])
```

```
## Warning:
## 2 (4.0%) p_waic estimates greater than 0.4. We recommend trying loo instead.

## Warning: Accessing waic using '$' is deprecated and will be removed in a
## future release. Please extract the waic estimate from the 'estimates' component
## instead.

## Warning: Accessing looic using '$' is deprecated and will be removed in a
## future release. Please extract the looic estimate from the 'estimates' component
## instead.
```

```
## STAN Model F WAIC 895.8925
```

```
cat("STAN Model F LOOIC", waic_loo(r_fit_F)[2])
```

```
## Warning:
## 2 (4.0%) p_waic estimates greater than 0.4. We recommend trying loo instead.
```



```
## Warning: Accessing waic using '$' is deprecated and will be removed in a
## future release. Please extract the waic estimate from the 'estimates' component
## instead.

## Warning: Accessing looic using '$' is deprecated and will be removed in a
## future release. Please extract the looic estimate from the 'estimates' component
## instead.

## STAN Model F LOOIC 896.0408
```