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# PREDICTIVE CODING OF GENOMIC INFORMATION DATA

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## 1 Preface

It should be understood at the outset that although an illustrative implementation of one or more embodiments are provided below, the disclosed systems and/or methods may be implemented using any number of techniques, whether currently known or in existence. The disclosure should in no way be limited to the illustrative implementations, drawings, and techniques illustrated below, including the exemplary designs and implementations illustrated and described herein, but may be modified within the scope of the appended claims along with their full scope of equivalents.

## 2 Introduction

Due to novel high-throughput next-generation sequencing (NGS) technologies, the sequencing of huge amounts of genetic information has become affordable. On account of this flood of data, IT costs may become a major obstacle compared to sequencing costs. High-performance compression of genomic data is required to reduce the storage size and transmission costs.

Raw sequencing data (mainly the base sequences – also known as reads - and the quality scores) is stored in so-called FASTQ files, whereas mapped sequencing data is stored in so-called SAM files. SAM files are plain-text human-readable files containing the reads and quality scores as well as mapping information with respect to some reference genome. Mapped sequencing data represented in SAM files contains more redundancy, as typically multiple reads are mapped to the same location on the reference genome. The average amount of reads mapping to the same location is referred to as *coverage*.

It has been shown by Tembe et al. [7] and Deorowicz and Grabowski [8], that splitting the data into separate streams for sequence reads, quality scores etc. (and compressing them independently) yields significant gains over general-purpose (dictionary-based [2]) programs like *gzip* or *bzip2*. All recent contributions to the topic of FASTQ and/or SAM file compression employ this scheme of separately compressing the sequence reads and quality scores (as well as the other field present in FASTQ or SAM files).

Based on statistical analysis performed on a reference data set (issued by the appointed MPEG ad-hoc group on genome compression), new proposals for sequence and quality score compression in SAM files have been developed.

### 2.1 Outline

We propose a sequence compressor which assumes aligned (position-sorted) data. Our sequence compressor combines the mapping positions, the CIGAR strings, and the actual nucleotide sequences to implicitly assemble local parts of the donor genome to compress the sequence reads. The compressor does not need a reference genome and is able to perform compression “on-the-fly” using solely a *sliding window* (permanently updated short-time memory of arbitrary size) as context for the local assembly.

Regarding the compression of quality scores, we propose predictive compression schemes (which may be based on e.g. Markov models) with a variable or fixed context to boost prediction performance. Due to the large alphabet of quality scores (and thus a huge memory consumption of any prediction model), we furthermore propose employing FIR filter-based compression for quality scores. The proposed FIR filter for quality scores can also be applied on any other data present, e.g. locations, cigar strings, sequence reads, or auxiliary data.

## 3 Block-based Compression of Nucleotide Sequences

### 3.1 Prior Work

Current implementations (e.g. *Quip* [6] and *sam\_comp* [3]) use an order-2 arithmetic coder (AC) to compress the nucleotide sequences. In order to exploit the redundancy present in the data, an arithmetic coder needs *a priori* knowledge. This practically excludes random access to the compressed data as the prediction performance increases with the amount of data the prediction model can be trained on.

The authors of *DeeZ* [5] made a new approach, called “local assembly”. *DeeZ* nevertheless performs the local assembly with respect to an external reference (given in FASTA format) and without subsequent AC.

The authors of *Quip* [6] implemented an assembly-based compression scheme for nucleotide sequences, too. They use by default the first 2.5 million reads to assemble so-called contigs, which are then used in place of a reference sequence to encode aligned reads. This algorithm does not require a reference – this means that the resulting compressed files are entirely self-contained – but has the drawback that the best match of a specific read to the previously assembled reference has to be found in the fixed non-adaptive contig repository.

### 3.2 Proposed Compression Mechanism

We propose a compression scheme using a short-time memory of arbitrary size which may be implemented as a sliding window to perform a so-called “local implicit assembly”. In contrast to *DeeZ*, our compressor does not require a reference genome and is able to work on smaller block granularities as it uses a short-time memory as reference. In contrast to *Quip*, we do not need any reads to assemble so-called contigs in advance and we are able to adapt to changes in data statistics. We perform the compression using only the currently available nucleotide sequences in the short-time memory. This short-time memory can be re-initialized at any time, enabling random access to any part of the compressed data without the need of decoding the preceding compressed data.

Our approach is based on correlations between consecutive reads. As all reads in a SAM file are aligned to their mapping position, all reads mapped to the same position should be similar, except for gene mutations (SNP’s) or sequencing errors, respectively mapping errors. These aberrations are coded in the CIGAR string. An example is given in Figure 1. Consecutive reads are plotted line by line. All reads mapped to the same position are aligned. As SAM files are stored sorted by mapping position, a read mapping to a different location than the read before leads to a step in the plot. If plotted over a large block, this leads to the staircase property visible in the plotted figure.

Our approach proposes to encode sequence reads block-wise exploiting the joint coding of the mapping positions, the CIGAR strings and the sequence reads itself, whereas the block size might be fixed or variable and of arbitrary size (which includes the compression of the input file as a single block). The first sequence read in a block is encoded without prediction, as well as the corresponding mapping position and the CIGAR string. However, it might also be coded against some arbitrary reference. Some subsequent read  $seq_i, i > 1$ , is then aligned to a previous read  $seq_j, j < i$ , using its CIGAR string and its mapping position.

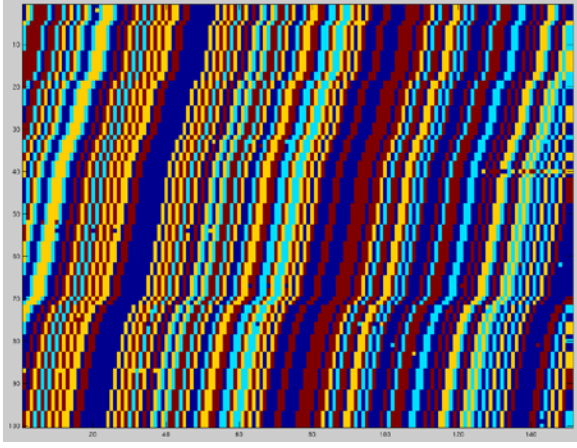


Figure 1: Consecutive nucleotide sequences from a SAM file. The different colors represent the nucleotides.

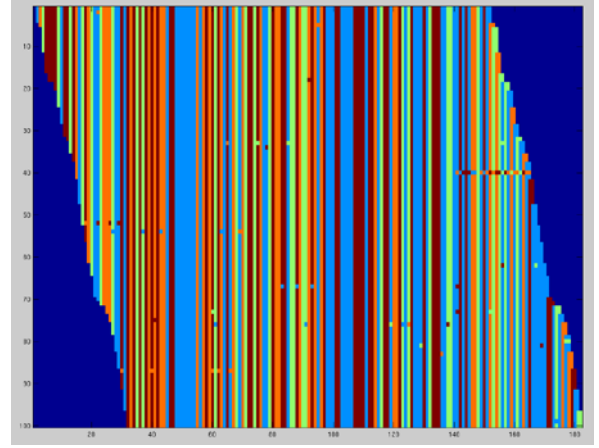


Figure 2: The nucleotide sequences after the `expand` function, stored as a sliding window in the circular buffer. The noisy 'dots' represent genome mutations or sequencing errors described in the CIGAR string.

Thereafter, we compute the difference  $d_i$  (containing the differences of the mapping position and the nucleotide sequence from the aligned read  $seq_i$  to read  $seq_j$  and pass it to an entropy coder, e.g. an order-0 arithmetic coder. At the decoder, read  $seq_i$  can be reconstructed from read  $seq_j$  by applying  $d_i$  to reconstruct the nucleotide sequence, the CIGAR string, and the mapping position.

A possible encoder structure is shown in Figure 3.

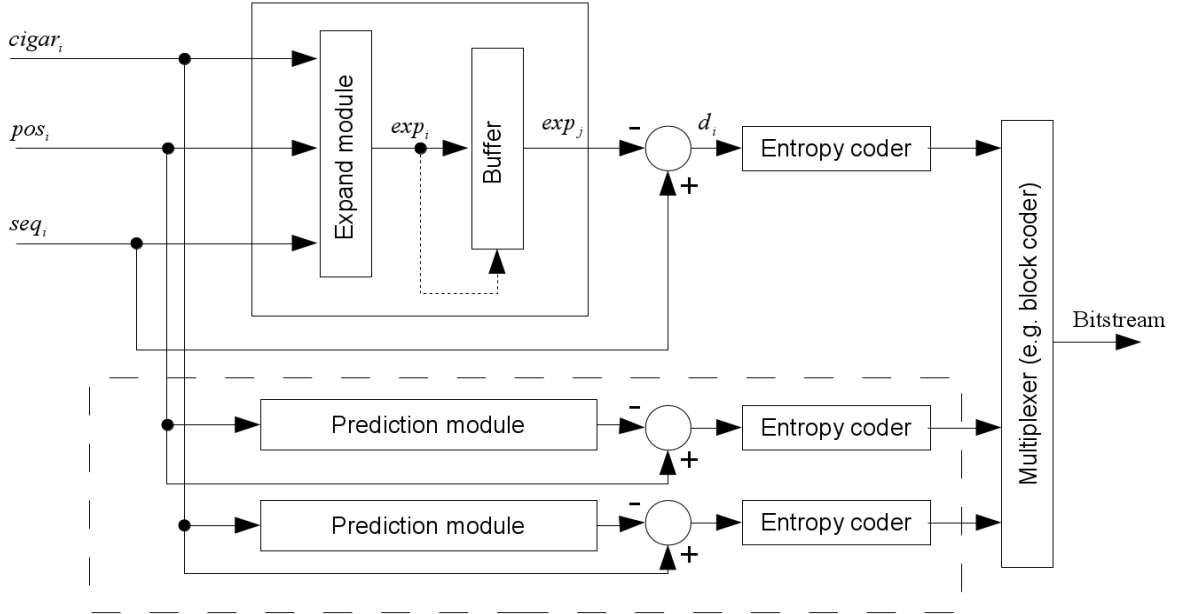


Figure 3: Possible encoder structure of the proposed sequence compression algorithm

The alignment of read  $seq_i$  to read  $seq_j$  is done with a function we call `expand`. The output of `expand` is called  $exp_i$ . The CIGAR strings and mapping positions of both reads are used to unwind the sequence such that they would both directly align (please refer to the SAM specification [1], Section 1.4ff for a detailed explanation of the CIGAR string syntax). In general, unwinding the sequence consists of a series of insertion, deletion, and modification operations as described by the CIGAR string. A possible output of the `expand` function is visualized in Figure 2. Consequently, `expand` condenses the position, the CIGAR string, and the sequence into a joint representative code word. As an

alternative, the last step could be omitted and these SAM fields could be processed separately. A short example of the working principle of `expand` is given in Figure 4.

POS	CIGAR	SEQ	
7	8M2I4M1D3M	TTAGATAAAGGATACTG	< read 1
9	3S6M1P1I4M	AAAAGATAAGGATA	< read 2
6789...	< POS		
8M	2I4M	1D3M	
TTAGATAAAGGATA	CTG		< read 1 expanded
AAAAGATAAGGATA			< read 2 expanded
3S 6M	1P1I4M		

Figure 4: Working principle of the function `expand`

The previous read  $seq_j$  is by default read  $seq_{i-1}$ . This context read  $seq_j = seq_{i-1}$  might be highly erroneous or not aligned to the reference. We address this issue by keeping track of  $N$  previous reads (sliding window) to be able to select the best matching expanded read  $exp_j$  to encode read  $seq_i$ . Sequence reads which are not aligned to the reference might be directly passed to the entropy coder or retained and encoded separately later, e.g. at the end of each block. The selected matching read might be signaled or estimated at the decoder. A possible implementation of the algorithm in pseudo code is shown in Figure 5.

```

Input: block_size, threshold, pos[L], cigar[L], seq[L], L (no. of SAM records)
Output: binary stream (from entropy coder)
new circbuf(N); // circular buffer of size N (sliding window)

l = 0;           // line count
b = block_size; // block line count

while (l < L)
| if (b == block_size) then // start new block
| | clear_circbuf();
| | reset_entropy_coder();
| | entropy_coder(pos[l], cigar[l], seq[l]);
| | circbuf.push(pos[l], cigar[l], seq[l]);
| | b = 1;
| else
| | if (cigar(l) != '*') then // sequence is mapped
| | | min_entropy = +inf;
| | | do
| | | | seq_exp = expand(pos[l], cigar[l], seq[l]);
| | | | (pos_ref, cigar_ref, seq_ref, ref_index) = circbuf.get_next();
| | | | ref_exp = expand(pos_ref, cigar_ref, seq_ref);
| | | | d = compute_differences(seq_exp, ref_exp);
| | | | if (entropy(d) < min_entropy) then
| | | | | min_entropy = entropy(d);
| | | | | ref_index_best = ref_index;
| | | | | d_best = d;
| | | | endif
| | | while (entropy(d) > threshold) && circbuf.has_next();
| | | entropy_coder(ref_index_best, d_best);
| | | else
| | | | entropy_coder(pos[l], cigar[l], seq[l]);
| | | endif
| | | circbuf.push(pos(l), cigar(l), seq(l));
| | endif
| | b++;
| | l++;
endwhile

```

Figure 5: Algorithm outline

## 4 Predictive Coding of Quality Scores

### 4.1 Prior Work

The quality scores yield the largest first-order entropy among all field present in a typical SAM file, mainly due to their large alphabet. The authors of *Fqzcomp* [3] state amongst other things the following dependencies for a sequence of quality scores:

- Any score  $q_i$  has a strong correlation to the direct previous quality scores  $q_{i-1}, q_{i-2}, \dots$ , decreasing the further back we go [4].
- Sequences as a whole tend to be good or bad.

Current SAM compression implementations exploit this observations by employing a Markov model with the memory ranging over the direct predecessors in the quality score stream.

### 4.2 Modeling the Source with a Markov Chain

Common SAM file compression approaches [4, 5] noted, that quality scores usually contain a trailing sequence of low quality values. We made this observations, too, and propose first to trim the trailing low quality values.

Current SAM file compressors model the quality score source as a Markov chain, predicting the current symbol with a maximum likelihood predictor using the  $N$  direct predecessors.

The maximum likelihood predictor selects in each state  $q_1, \dots, q_N$  the symbol  $a_i$  with the greatest conditional probability  $P$  as prediction value  $q_{N+1}^{(p)}$ :

$$P\left(q_{N+1}^{(p)} = a_i | q_1, \dots, q_N\right) \geq P(q_{N+1} = a_k | q_1, \dots, q_N) \forall k$$

#### 4.2.1 Drawbacks

The maximum likelihood predictor consumes a huge amount of memory since we have to keep track of  $k^N * k$  relative symbol frequencies (having an alphabet of size  $k$ ). This model also needs a large amount of data to train in order to tightly fit the model to the data.

### 4.3 Proposed Compression Schemes

As an alternative to building the prediction memory during the decoding process, we propose the possibility to store the prediction table (generated during compression) e.g. in the file header to speed up decompression as an alternative or in combination. The prediction tables for the individual blocks may then be encoded, e.g. delta-encoded with subsequent AC. This enables random access to the compressed data while retaining the compression performance of the prediction model (e.g. Markov model) by providing the fully trained prediction memory, or parts of it, (built from the full input data at the encoder) to the decoder, which can then be directly used from the start of the decoding process.

The following subsections describe alternatives to predictive quality score compression which might be combined to yield the best compression performance. As an additional alternative, we propose to directly compress the quality scores without prediction employing solely an entropy coder, e.g. an AC of arbitrary order.

#### 4.3.1 Extended Line Context Approach

We made the observation, that in some datasets subsequent quality score lines are “similar” in terms of a small Levensthein distance between pairs of quality score lines.

We respond to this using a so-called *line context*, i.e. a structure holding some number of previous lines from the current block (this is also known as “sliding window”). The prediction context (i.e. a

Markov model) is divided into one part containing the memory locations on the current line (“intra-line memory”) and one part containing the memory locations on some specific line from the line context (“inter-line memory”) as depicted in Figure 6.

line  $\hat{i}$  DECEEEECDDDDDDDDDDDDDDDDDDDDDDDEDCDDDDDDDD  
 $\vdots$   
line  $i$  DDDDDDDDCDDDDDDDBDDDDDDDDDEEEEDDD

Figure 6: Visualization of the intra-line (dark blue) and inter-line (light blue) context used to predict the current symbol (green).

Choosing the right line from the line context to boost prediction performance is crucial. We propose to compute an identification vector for each quality score line (or parts of it) in the line context. As an example, this could be a three-dimensional vector, containing e.g. the length, the arithmetic mean, and the variance of a quality score line. In the process of encoding line  $i$ , we first select the one specific line  $\hat{i}$  from the line context with the least identification vector distance, whereas any known vector distance measurement might be applied, e.g. the three-dimensional Euclidean distance. The vector components might be individually weighted. Consecutively, the intra-line memory is placed on line  $i$  and the inter-line memory is placed on the selected line  $\hat{i}$  from the line context.

### 4.3.2 Dictionary-based Substring Matching Approach

To encode the quality score  $q_{i,j}$  in column  $j$  and line  $i$ , we propose to select the substring  $s$  of  $N$  preceding symbols  $s = q_{i,j-N-1}, \dots, q_{i,j-1}$ . We then propose to search for a similar substring  $\hat{s} = q_{\lambda,\gamma}, \dots, q_{\lambda,\gamma+N} \equiv s$  in the line context and to use  $q_{\lambda,\gamma+N+1}$  as the prediction value  $q_{i,j}^{(p)}$  for  $q_{i,j}$ .

Alternatively, we propose to use a prediction value  $q_{i,j}^{(p)}$  calculated as a mean or median or as an arbitrary combination from  $M$  substring matches  $\hat{s}_m, 1 \leq m \leq M$ , which might be weighted by any reasonable weighting factors, or any other reasonable measure, whereas any suitable norm might be applied. This principle is briefly pointed out in Figure 7.

DECEE**E****E**CDD**D**DDDDDDDDDDDDDDDCDDDDDDDD  
:  
DDDDDDDDCDDDDDBD**E****E**CDD**D**DEEEEDDD

Figure 7: The green symbol is predicted from the blue symbol, if a matching substring containing  $M=5$  symbols (red) is found in the line context.

This approach for quality score compression can also be applied on any other data present, e.g. locations, cigar strings, sequence reads, or auxiliary data.

### 4.3.3 Markov Context-Extending Substring Matching Approach

As an alternative, we propose to extend the context of any of the above mentioned predictors with the prediction value  $^s q_{i,j}^{(p)} = q_{\lambda, \gamma+N+1}$  found by substring matching within the line context.



Regarding the prediction model from Section 4.2, or any other prediction model, we therefore propose to extend the state/context  $q_1, \dots, q_n$  with  ${}^s q_{i,j}^{(p)}$  to form an extended state  $q_1, \dots, q_n, {}^s q_{i,j}^{(p)}$ . Regarding the prediction model from Section 4.3.1, we propose to extend the context (consisting of the inter- and intra-line memories) with  ${}^s q_{i,j}^{(p)}$ .

#### 4.4 Linear Prediction with FIR Filters

Due to the large alphabet of the quality scores (typically approximately 30 symbols, although more are theoretically possible), Markov models are limited to some low order  $N$  (typically  $N \leq 3$ ).

As an alternative, we propose to use an FIR filter to predict the current symbol. While it is on the one hand costly to compute the FIR filter coefficients, on the other hand, filter coefficients might be shared between multiple quality score lines.

The basic layout of a predictive encoder can be viewed in Figure 8.

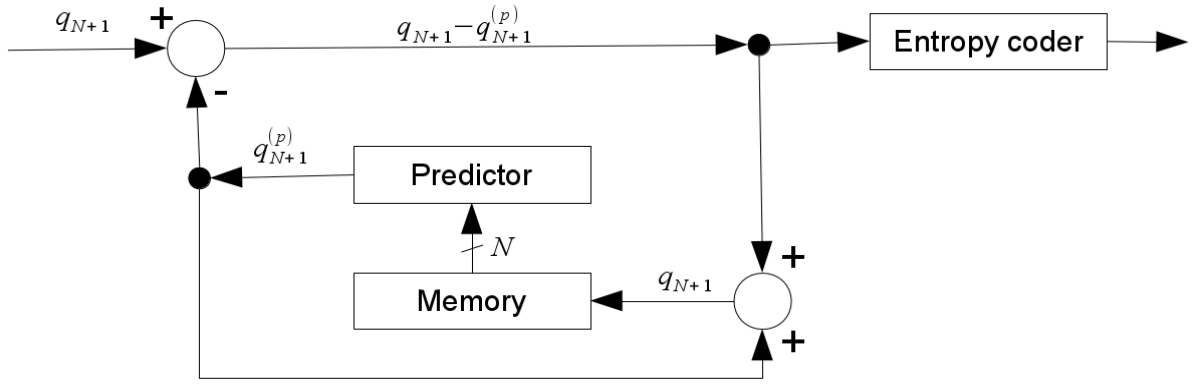


Figure 8: Linear predictive encoder schematic

The linear predictor computes the current prediction value  $q_{N+1}^{(p)}$  by evaluating the linear series of previous symbols  $q_n$  weighted with predictor coefficients  $a_n$ .

$$q_{N+1}^{(p)} = \sum_{n=1}^N a_n q_n$$

One possible approach to compute the predictor coefficients is to minimize the mean squared error which yields the following condition to compute the filter coefficients.

$$\frac{\partial}{\partial a_n} \left( E \left[ \left( q_{N+1} - q_{N+1}^{(p)} \right)^2 \right] \right) = 0$$

Thus, we obtain  $l$  sets of predictor coefficients for a quality score line of length  $l$ . Using the line context from Section 4.3.1, it could be beneficial to share previously computed predictor coefficients among multiple lines.

As an alternative to the minimum-mean-square-error filtering it should be possible to employ any other known method to compute the predictor coefficients, e.g. with minimum-variance prediction.

This approach for quality score compression can also be applied on any other data present, e.g. locations, cigar strings, sequence reads, or auxiliary data.

## 4.5 Entropy Coding of Prediction Errors

The histogram of the prediction errors approximately shows a two-sided geometric distribution. We therefore propose to use subsequent Rice coding [9] as entropy coding step to obtain the binary representation of the data. This reduces the encoder complexity, as recent compressors such as *Quip* [6] employ arithmetic coding.

## 5 Remark

While several embodiments have been provided in the present disclosure, it should be understood that the disclosed systems and methods might be embodied in many other specific forms without departing from the spirit or scope of the present disclosure. The present examples are to be considered as illustrative and not restrictive, and the intention is not to be limited to the details given herein. For example, the various elements or components may be combined or integrated in another system or certain features may be omitted, or not implemented.

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## **1 Introduction**

Due to novel high-throughput next-generation sequencing (NGS) technologies, the sequencing of huge amounts of genetic information has become affordable. On account of this flood of data, IT costs may become a major obstacle compared to sequencing costs. High-performance compression of genomic data is required to reduce the storage size and transmission costs.

Raw sequencing data (mainly the base sequences – also known as reads - and the quality scores) is stored in so-called FASTQ files, whereas mapped sequence data is stored in so-called SAM files. SAM files are plain-text human-readable files containing the reads and quality scores as well as mapping information with respect to some reference genome. Mapped sequencing data represented in SAM files contains more redundancy, as typically multiple reads are mapped to the same location on the reference genome. The average amount of reads mapping to the same location is referred to as *coverage*.

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Based on statistical analysis performed on the reference data set issued by this ad-hoc group during the 111<sup>th</sup> MPEG meeting in Geneva, new proposals for sequence and quality score compression in SAM files have been developed.

## 1.1 Outline

We propose a sequence compressor which assumes aligned and position-sorted data. Our sequence compressor combines the mapping positions, the CIGAR strings and the actual nucleotide sequences to implicitly assemble local parts of the donor genome and compress the sequence reads. The compressor does not need a reference genome and is able to perform compression “on-the-fly” using solely a *sliding window* as context for the local assembly.

Regarding the compression of quality scores, we propose a couple of compression schemes based on Markov models with a variable context to boost prediction performance. Due to the large alphabet of quality scores (and thus a huge memory consumption of any Markov model), we furthermore propose employing FIR filter-based compression for quality scores.

## 2 Software Framework

We developed a compression framework called *tsc* (C99-compliant) to evaluate compression mechanisms for the different fields present in a SAM file. *Tsc* splits the SAM file into separate streams for each field [1]. Dedicated compression algorithms can then be employed in a block-by-block manner on each field or on a combination of fields. New compression modules for e.g. quality score compression can easily be mounted.

We propose to use the *tsc* framework as a starting point for a standardization effort. Presumably, a combination of compression algorithms from various sources will yield the best overall compression ratio for SAM files as well as FASTQ files (the *tsc* framework can easily be extended to accept FASTQ files, too).

### 3 Block-based Compression of Nucleotide Sequences

#### 3.1 Prior Work

Current implementations (e.g. *Quip* [6] and *sam\_comp* [3]) use an order-2 arithmetic coder (AC) to compress the nucleotide sequences. In order to exploit the redundancy present in the data, an AC needs *a priori* knowledge. This practically excludes random access to the compressed data as the prediction performance increases with the amount of data the Markov model can be trained on.

The authors of *DeeZ* [5] made a new approach, called “local assembly” (implicit assembly of the donor genome using the mapping information present in SAM files). *DeeZ* nevertheless performs the local assembly with respect to an external reference (given in the FASTA format).

The authors of *Quip* [6] implemented an assembly-based compression scheme for nucleotide sequences, too. They use by default the first 2.5 million reads to assemble so-called contigs which are then used in place of a reference sequence to encode aligned reads. This algorithm does not require an external reference – this means that the resulting compressed files are entirely self-contained – but has the drawback that the best match of a specific read to the previously assembled reference has to be found.

#### 3.2 Proposed Compression Mechanism

We propose a compression scheme using a sliding window to perform a “local block-based implicit assembly”. In contrast to *DeeZ*, our compressor does not require a reference genome and is able to work on smaller block granularities as it uses a “sliding window” as context (of course smaller block sizes have a negative impact on compression ratio). In contrast to *Quip*, we do not need any reads to assemble so-called contigs in advance. We perform the compression using only the currently available nucleotide sequences in the sliding window. Our compression scheme therefore is less complex, but should nevertheless yield comparable compression ratios.

Our approach is based on correlations between consecutive reads. As all reads in a SAM file are aligned to their mapping position, all reads mapped to the same position should be similar, except for gene mutations (SNP's) or sequencing errors respectively mapping errors. These aberrations are coded in the CIGAR string.

Figure 1 and figure 2 illustrate the structure of mapped sequence reads. Consecutive reads are plotted line by line. All reads mapped to the same position are aligned. As SAM files are stored sorted by mapping position, a read mapping to a different location than the read before leads to a step in the plot. If plotted over a large block, this leads to the staircase property visible in the plotted figure.

Our approach proposes to encode sequence reads block-wise exploiting the joint coding of the mapping positions, the CIGAR strings and the sequence reads itself, whereas the block size might be fixed or variable and of arbitrary size. The first sequence read in a block is encoded without prediction, as well as the corresponding mapping position and the CIGAR string. However, it might also be coded against some reference. Some subsequent read  $i, i > 1$ , is then aligned to a previous read  $j, j < i$ , using its CIGAR string and its mapping position. Thereafter, we compute the difference  $d$  (containing the differences of the mapping position and the

nucleotide sequence from the aligned read  $i$  to read  $j$ ) and pass it to an entropy coder, e.g. an order-0 arithmetic coder. At the decoder, read  $i$  can be reconstructed from read  $j$  by applying  $d$  to reconstruct the nucleotide sequence, the CIGAR string, and the mapping position.

The alignment of read  $i$  to read  $j$  is done with a function called `expand`. The CIGAR strings and mapping positions of both reads are used to unwind the sequence such that they would both directly align to some external reference (please refer to the SAM file specification [1] for a detailed explanation of the CIGAR string syntax). Consequently, `expand` condenses the position, the CIGAR string, and the sequence into a joint representative code word. As an alternative, these SAM fields could be compressed using separate entropy coders.

The previous read  $j$  is by default read  $i - 1$ . This context read  $j = i - 1$  might be highly erroneous or not aligned to the reference. We address this issue by keeping track of  $N$  previous reads (sliding window) to be able to select the best matching read to encode read  $i$ .

Sequence reads which are not aligned to the reference might be directly passed to the entropy coder or retained and encoded separately later, e.g. at the end of each block.

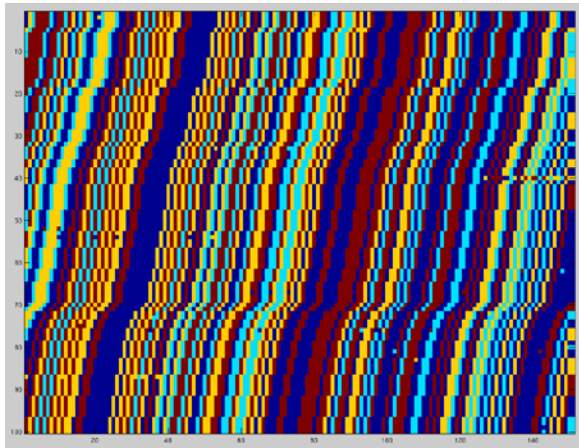


Figure 1: Consecutive nucleotide sequences from a SAM file. The different colors represent the nucleotides.

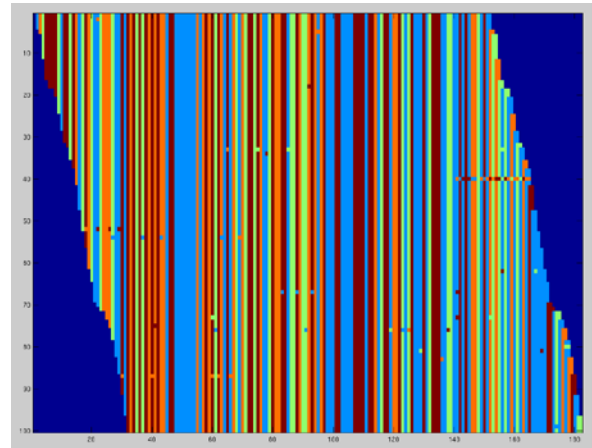


Figure 2: The nucleotide sequences after the `expand` function, stored as a sliding window in the circular buffer. The noisy 'dots' represent genome mutations or sequencing errors described in the CIGAR string.

**Note:** Per block, the initial read and the trailing substrings of subsequent reads can be regarded as locally assembled genome.

### 3.3 Results

First results have been obtained using data from the genomic information database issued by this ad-hoc group during the MPEG meeting 111 in Geneva.

The proposed sequence compression algorithm has been integrated into the *tsc* framework from section 2. We used a block size of 100,000 (sequence reads per block) and a sliding window holding 100 reads. Subsequent entropy coding is performed using an order-0 AC. The results are compared against *Quip* and *DeeZ*. *Quip* uses a high-order Markov chain. The nucleotide at a given position is predicted using the preceding twelve positions. *DeeZ* uses its own unique compression algorithm (implicit assembly of the donor genome).

Our algorithm (which does not require a reference genome) generally seems to be on par with *DeeZ*. Employing our algorithm on the MiSeq\_Ecoli\_DH10B and RNAseq datasets yields even better compression ratios than *DeeZ*. These results seem to be on account of the subsequent AC. The compression of the ERR317482 dataset yields a slightly poorer compression ratio than *DeeZ*. This might be on account of the relatively small sliding window we use in contrast to the full local assembly performed by *DeeZ*.

Dataset	Compressed SEQ size		
	tsc	Quip	DeeZ
MiSeq_Ecoli_DH10B	<b>2.22%</b>	33.11%	8.93%
ERR317482	<b>17.07%</b>	34.62%	12.21%
RNAseq	<b>7.58%</b>	31.94%	9.98%

## 4 Predictive Coding of Quality Scores

### 4.1 Prior Work

The quality scores yield the largest first-order entropy among all field present in a typical SAM file, mainly due to their large alphabet.

The authors of *Fqzcomp* [3] state the following dependencies for a sequence of quality scores:

- Any score  $q_i$  has a strong correlation to the direct previous quality score  $q_{i-1}, q_{i-2}, \dots$ , decreasing the further back we go [4].
- A known issue with the Illumina base-caller causes many sequences to end with quality score 2 (“#”).
- There is a correlation between position and quality values. Quality values are typically reducing along the length of the sequence.
- Sequences as a whole tend to be good or bad.

The authors of *Fastqz* [3] additionally state that it was observed that the quality values tend to start with a common maximum value.

Current SAM compression implementations exploit this observations by employing a Markov model with the memory ranging over the direct predecessors in the quality score stream.

### 4.2 Observations

In terms of performing some statistical analysis regarding the quality scores, we converted the quality score lines into a single stream of quality scores.

A snapshot of some typical quality score lines and their histogram is shown in figures 3 and 4.

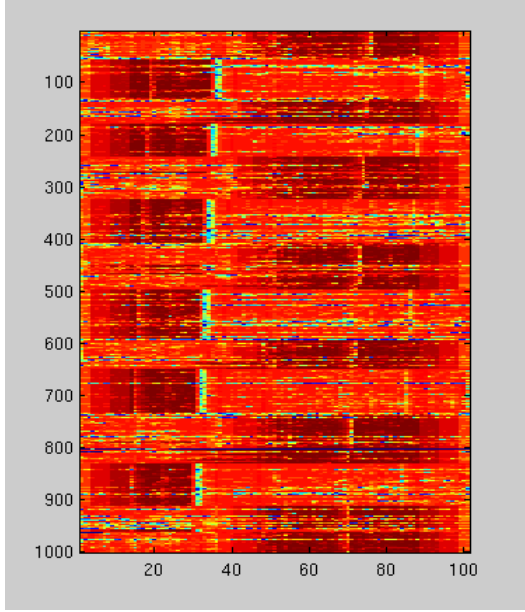


Figure 3: Snapshot of 1,000 quality score line from a SAM file. The data has been produced by an Illumina machine (constant read lengths).

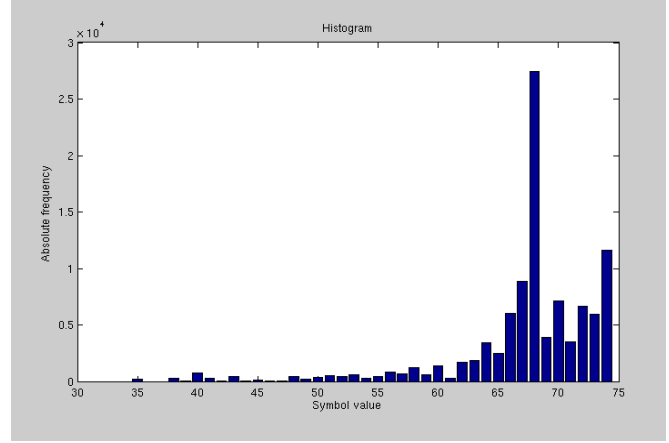


Figure 4: Typical quality score histogram

The autocorrelation function of such a stream is shown in figure 5. The auto-correlation does not resemble a delta impulse and thus the power spectral density is not constant. This means, that the source has some kind of memory. Thus, it is highly suitable to model the source with a Markov chain.

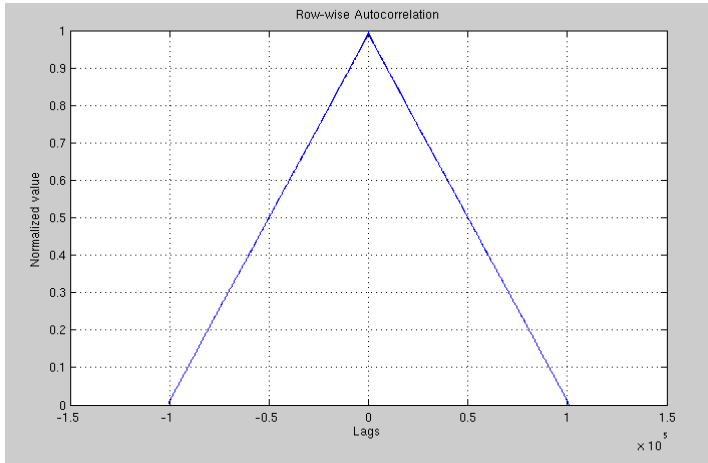


Figure 5: Autocorrelation function of a quality score stream

### 4.3 Modeling the Source with a Markov Chain

Current SAM file compressors model the quality score source as a Markov chain, predicting the current symbol with a maximum likelihood predictor using the  $N$  direct predecessors.

The maximum likelihood predictor selects in each state  $q_1, \dots, q_N$  the symbol  $a_i$  with the greatest conditional probability  $P$  as prediction value  $q_{N+1,p}$ :

$$P(q_{N+1,p} = a_i | q_1, \dots, q_N) \geq P(q_{N+1} = a_k | q_1, \dots, q_N) \forall k$$



### 4.3.1 Drawbacks and Recommendations

The maximum likelihood predictor consumes a huge amount of memory since we have to keep track of  $k^N * k$  relative symbol frequencies (having an alphabet of size  $k$ ). This model also needs a large amount of data to train in order to tightly fit the model to the data. We propose to store the prediction table (generated during compression) in the file header to speed up decompression. Prediction tables for the individual blocks could then be delta-encoded. This would enable random access to the compressed data while retaining the compression performance of the Markov model.

## 4.4 Proposed Compression Schemes

### 4.4.1 Dictionary-based Substring Matching Approach

The authors of *Fqzcomp* [3] state, that quality score lines as a whole tend to be “bad” or “good”. Additionally, we made the observation, that in some datasets subsequent quality score lines are “similar” in terms of a small Levensthein distance between pairs of quality score lines.

We respond to this using a so-called line context, i.e. a structure holding some number of previous lines from the current block (this can be regarded as *sliding window*).

To encode the quality score  $q_{i,j}$  in column  $j$  and line  $i$ , we propose to select the substring  $s$  of  $N$  preceding symbols  $s = q_{i,j-N-1}, \dots, q_{i,j-1}$ . We then propose to search for a similar substring  $\hat{s} = q_{\lambda,\gamma}, \dots, q_{\lambda,\gamma+N} \equiv s$  in the line context and to use  $q_{\lambda,\gamma+N+1}$  as the prediction value  $q_{i,j}^{(p)}$  for  $q_{i,j}$ .

Furthermore, we propose to use a prediction value  $q_{i,j}^{(p)}$  calculated as a e.g. mean or median from  $M$  substring matches  $\hat{s}_m$ .

## 4.5 Entropy Coding of Prediction Errors

The histogram of the prediction errors of the Markov or FIR predictor approximately shows a two-sided geometric distribution. We therefore propose to use subsequent Rice coding [9] as an alternative to arithmetic coding to obtain the binary representation of the data. This reduces the encoder complexity, as recent compressors such as *Quip* [6] employ arithmetic coding.

## 4.6 Results

First results have been obtained using data from the genomic information database issued by this ad-hoc group during the MPEG meeting 111 in Geneva.

Similar to the proposed sequence compressor in section 3.2, the quality score compressors have been integrated into the *tsc* framework. Again, we used a block size of 100,000 reads per block and a sliding window holding 100 reads. The results are compared against *Quip* and *DeeZ* (using a modified order-2 AC respectively an order-2 AC).

“tsc O0” refers to a simple order-0 AC. “tsc O1” refers to the dictionary-based substring matching approach from section 4.4.1 with a subsequent order-0 AC.

Surprisingly, the order-0 AC shows the best compression performance. Every attempt to predict quality score values (either by substring matching (“tsc O1”) or by employing higher-order AC's), yields poorer overall compression ratios.

Dataset	Compressed SEQ size			
	tsc O0	tsc O1	Quip	DeeZ
MiSeq_Ecoli_DH10B	<b>51.98%</b>	56.97%	57.63%	77.58%
ERR317482	<b>50.65%</b>	65.00%	53.59%	71.50%
RNAseq	<b>46.72%</b>	54.81%	53.92%	72.80%

## 5 Recommendations

Due to recent efforts made in the domain of lossy quality score compression, we recommend to further investigate lossy, predictive quality score compression with an emphasize on rate-distortion performance.

## 6 References

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