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Diminished adrenal sensitivity to endogenous and exogenous adrenocorticotropic hormone in critical illness: a prospective cohort study

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Abstract

Introduction

Adrenal dysfunction may represent critical illness-related corticosteroid insufficiency (CIRCI), as evidenced by a diminished cortisol response to exogenous adrenocorticotropic hormone (ACTH), but this concept and its clinical significance remain highly controversial. We studied the adrenal response to exogenous ACTH as a function of the endogenous cortisol to ACTH ratio, a measure of adrenal sensitivity, and of clinical variables, during critical illness and recovery from the acute phase.

Methods

We prospectively included 59 consecutive septic and non-septic patients in the intensive care unit with treatment-insensitive hypotension in whom CIRCI was suspected; patients having

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received etomidate and chronic corticosteroids were excluded. An ACTH test (250 mcg) was performed, followed by a second test after \geq 7 days in acute phase survivors. Serum total and free cortisol, ACTH and clinical variables were assessed. Patients were divided according to responses (delta, Δ) of cortisol to ACTH at first and second test.

Results

Patients with low (<250 nmol/L) Δ cortisol (n = 14 to 17) had higher baseline cortisol and ACTH but lower cortisol/ACTH ratios than patients with a normal Δ cortisol (\geq 250 nmol/L) in the course of time. A low Δ cortisol in time was associated with more severe disease, culture-positive sepsis and prolonged activated prothrombin time. Results for free cortisol were similar.

Conclusions

Even though the pituitary-adrenal axis is activated following stress during critical illness, diminished adrenal sensitivity to endogenous ACTH predicts a low increase of cortisol to exogenous ACTH suggesting adrenal dysfunction, irrespective of the stage of disease. The data further suggest a role of disease severity and culture-positive sepsis.

Introduction

The stress of critical illness, and sepsis in particular, strongly activates the hypothalamicpituitary-adrenal (HPA) axis and increases circulating cortisol [1-9]. Nevertheless, the secretion and concentration of (free) cortisol may be relatively insufficient, even when not subnormal, for the severity of disease in the course of critical illness-related corticosteroid insufficiency (CIRCI), which may relate to morbidity and mortality, particularly in sepsis [2,3,6-15]. Albeit hotly debated, a relatively low increase in circulating cortisol upon exogenous adrenocorticotropic hormone (ACTH) stimulation may denote adrenal dysfunction, which is associated with severe disease and ultimate mortality and may warrant treatment by corticosteroids to increase survival from septic shock [2,6,7,11,12,14]. This may be accompanied by relatively high baseline cortisol levels but, as some studies suggest, by low cortisol/ACTH ratios indicative of diminished adrenal sensitivity even when ACTH levels do not exceed normal values [3,4,6,7,16-18]. Treatment with etomidate-inhibiting adrenal cortisol synthesis, indeed, lowers the cortisol/ACTH ratio and diminishes the cortisol response to exogenous ACTH [16,19]. The ratio is used to diagnose primary absolute adrenal insufficiency [20]. However, Boonen et al. [21] recently suggested that, during critical illness, impaired cortisol metabolism (measured in relatively few patients) contributes to elevated (free) cortisol levels and low endogenous ACTH levels via feedback inhibition, and thereby to a diminished response to exogenous ACTH in the absence of adrenal dysfunction. A diminished response to exogenous ACTH is however hard to explain on the basis of an acutely low endogenous ACTH only. Nevertheless, decreased metabolism might explain in part the dissociation between low ACTH and elevated cortisol concentrations in persistent rather than acute (as in the Boonen et al. study [21]) critical illness over days, attributed to hypersensitivity or non-ACTH stimulation of the adrenals [6,22,23]. The controversy on adrenal function in critical illness is further compounded by the allegedly poor relationship between protein-bound total and free cortisol involved in tissue effects and feedback, but increases in free cortisol may parallel increases in total cortisol irrespective of binding proteins [5,17].

Finally, the question remains whether adrenal injury contributes to dysfunction, for instance following diffuse intravascular coagulation and adrenal thrombosis or haemorrhage, as may occur during meningococcemia, other causes of sepsis and heparin treatment amongst others [2,16,24,25]. We previously showed in transversal but also in longitudinal, retrospectively analyzed observations that clinical predictors for a diminished cortisol response to exogenous ACTH are septic shock with severe disease, acidosis and organ failure on the one hand and coagulation disturbances on the other [8,15]. In any case, a relatively low exogenous ACTH response during critical illness may normalize when disease severity resolves in surviving patients, whereas a fall in response may be associated with increasing severity of disease and mortality, arguing against the ACTH test as a poorly reproducible laboratory phenomenon in the critically ill [15,26-28]. This may favor reversal and development of adrenal dysfunction in time, respectively.

The hypothesis for the present study is, first, that diminished adrenal sensitivity to endogenous ACTH attenuates responses to exogenous ACTH, irrespective of the stage of critical illness. Second, that these abnormalities characterize adrenal dysfunction and insufficiency relative to disease severity in the course of critical illness. Third, that sepsis and its sequelae is a main risk factor for adrenal dysfunction. We therefore examined these factors longitudinally in a cohort critically ill patients who were able to undergo repeated ACTH testing.

Materials and methods

Patient population and ACTH test

This prospective study was carried out in the 32-bed general intensive care unit (ICU) of a university hospital from December 2004 to March 2007. According to Dutch legislation and the institutional ethical committee (Medical Ethical Committee VUmc, chair Prof dr JA Rauwerda), there was no need for ethical approval and informed consent, since the ACTH test is routinely performed in our department for patient care, when clinically indicated, no extra blood was drawn for this study, and the results were treated anonymously. In the study period, an ACTH test was performed in 288 consecutive patients and 59 patients (20%) were included when fulfilling criteria for the current study (Figure 1). These patients were over 18 years old with a clinical suspicion of CIRCI on the basis of >6 h hypotension (<100 mm Hg systolic) requiring repeated fluid challenges and/or vasopressor/inotropic treatment and underwent a first ACTH test in the ICU (Test 1) and a second test (Test 2) prior to or after discharge form the ICU; there was a minimum interval of 7 days between tests and a minimum interval of 5 days after cessation of hydrocortisone treatment when started. Patients were excluded if they had a history of HPA disease, if they received glucocorticoid treatment within 3 months of Test 1, or if they received etomidate prior to endotracheal intubation, since etomidate is a well known inhibitor of adrenal function [2,13,14,16]. All patients underwent a short 250 mcg ACTH (tetracosactide-hexa-acetate, Synacthen, Novartis Pharma, Basel Switzerland) test. Blood samples were taken at baseline and 30 and 60 minutes after intravenous injection. Serum total cortisol was measured by competitive immunoassay (Advia Centaur, Siemens Diagnostics, Deerfield, IL). The intra- and interassay coefficients of variation are 3 and 6%, respectively, and the detection limit was 30 nmol/L (500 nmol/L = 18µg/dL). Serum free cortisol levels were measured by equilibrium dialysis of undiluted serum samples (n = 49 available) followed by radioimmunoassay (Orion Diagnostica, Espoo, Finland; <6% cross reactivity with cortisone). The intra- and interassay coefficients of variation were less than 7 and 8%, respectively. Samples for ACTH were immediately put on ice and ACTH was determined with an immunometric assay and CBG with radioimmunoassay (Immulite 2500; Siemens Medical Diagnostic Solutions, Deerfield, IL, USA; *n* ACTH 2-26 pmol/L and BioSource, Nivelles, Belgium; *n* 30-54 mg/L, respectively). We calculated the ratio of baseline cortisol/ACTH as a parameter of adrenal sensitivity [3,4,16-18]. The peak (at 30 or 60 min) minus baseline cortisol level was taken to calculate increases upon exogenous ACTH. A low response to ACTH in critical illness was defined by a subnormal total cortisol increase (<250 nmol/L) [6,7,10,12-14].

Figure 1. Flow chart of patient inclusion for this study.

Data collection

At study entry, the following parameters were recorded: time from ICU admission, age and sex, and ICD-10 diagnoses of common clinical conditions at admission. At the day of each ACTH test, the following was collected: serum total and free cortisol levels prior to and 30 and 60 minutes after 250 mcg of i.v. ACTH, serum levels of ACTH cortisol binding globulin (CBG) and albumin. Other laboratory measurements included total white blood cell count, platelet count, activated partial thromboplastin time (aPTT) and prothrombin time (PT). Interventions including type and doses of vasopressor/inotropes, intubation need for mechanical ventilation and renal replacement therapy were recorded. Sepsis was defined as the presence of systemic inflammatory response syndrome (SIRS) with a positive microbiological local (urine, trachea or other) and/or blood culture. SIRS was defined as two or more of the following criteria: a temperature of >38 C or <35.5 C, a leukocyte count >12 of <4 x10⁹/L, a heart rate >90/min, and a respiratory rate >20/min or the presence of mechanical ventilation. Suspected or microbiologically proven sources of sepsis were recorded. Disease severity was assessed by the acute physiology and chronic health evaluation score (APACHE II) and sequential organ failure assessment score (SOFA) on the days of the ACTH test, and length of ICU stay and mortality in the ICU and hospital were recorded.

Clinical management

Patients were treated according to local protocols, derived from national and international guidelines. Treatment of sepsis included antibiotics and drainage, whenever possible, and intubation, mechanical ventilation and continuous veno-venous hemofiltration (CVVH) were instituted when needed. Patients and caregivers were not blinded to ACTH test results and moderate dose corticosteroids (hydrocortisone starting 100 mg TID i.v.) was administered at the discretion of attending intensivists. For anticoagulation, for instance during CVVH, mostly heparin was used.

Statistical analysis

For categorical data the X^2 and Fisher s exact test was used (SPSS v21, IBM SPSS Inc, Chicago, Ill). Continuous variables were tested for normality with the Kolmogorov-Smirnov test and if P < 0.05 they were logarithmically transformed for further analysis with help of generalized estimating equations (GEE). This was performed to investigate the association of Δ cortisol, time and their first-order interaction to endocrine and clinical variables (in a linear model), taking repeated measurements in the same patients into account (in an exchangeable

matrix). The Mann-Whitney U test was done for other group comparisons. The Spearman correlation coefficient (r_s) was used to express relation between pooled variables. Receiver operating characteristic area under the curve (AUROC) was used to evaluate predictive values. A two-sided P < 0.05 was considered to indicate statistical significance, and exact P values <0.10 are given, unless <0.001. Data are expressed as median (range) or number (percentage).

Results

Patients

Patient characteristics are described in Table 1. In 14 patients (24%), the second test was performed after discharge from the ICU. Six patients had in both tests Δ cortisol <250 nmol/L, and 11 patients recovered and 8 deteriorated in the course of time. Thirty-four patients had a normal response in both tests. All patients received hydrocortisone after the first test awaiting results.

Table 1. Patient characteristics at admission

	n = 59
Age	68 (19-87)
Sex (m/f)	35 (59)/24(41)
Admission diagnosis:*	
Trauma and postoperative	23 (39)
Respiratory insufficiency	16 (27)
CPR	5 (9)
Sepsis	10 (17)
Renal insufficiency	4 (7)
Other	11 (19)

Median (range) or number (%) where appropriate. *Patients may have more than two. CPR: cardiopulmonary resuscitation.

ACTH and cortisol

Levels of Δ cortisol as a continuous variable, which were directly associated with Δ free cortisol (P < 0.001), were inversely associated with ACTH levels irrespective of time (P < 0.001). Similarly, baseline cortisol and cortisol/ACTH were inversely and directly associated with Δ cortisol (P = 0.042 and 0.004), respectively. Test characteristics according to Δ cortisol <, \geq 250 nmol/L on test days are described in Table 2. It shows that the response in Δ (free) cortisol to exogenous ACTH was low when baseline ACTH as well as (free) cortisol levels were relatively high but baseline (free) cortisol to ACTH ratios were low, as compared to normal responders to exogenous ACTH . CBG increased but ACTH and (free) cortisol did not decrease in time, in spite of decreasing indices of disease severity (Table 3).

Table 2. ACTH Test 1 and 2 classified according to groups of Δ cortisol <, \geq 250 nmol/L

	Test 1 Δ < 250 nmol/L n = 17	Test 1 $\Delta \ge 250 \text{ nmol/L}$ n = 42	Test 2 Δ < 250 nmol/L n = 14	Test 2 $\Delta \ge 250 \text{ nmol/L}$ n = 45	$P \Delta$ cortisol, time, interaction
ACTH, pmol/L	3.1 (1.1-18.0)	1.5 (1.1-15.0)	6.0 (1.1-14.0)	3.8 (1.0-8.9)	0.001, 0.025, 0.72
Baseline cortisol, nmol/L	470 (200-1355)	430 (75-2670)	655 (335-1890)	495 (35-2670)	0.021, 0.12, 0.30
Baseline cortisol/ACTH, nmol/pmol	148 (36-250)	238 (14-2054)	109 (48-1718)	126 (35-2054)	0.029, 0.16, 0.14
Baseline free cortisol, nmol/L	77 (63-357) (n = 9)	86 (38-256) (n = 14)	154 (114-252) (n = 6)	75 (4-151) (n = 20)	0.001, 0.91, 0.19
Baseline free cortisol/ACTH, nmol/pmol	22 (9-38)	39 (16-89)	26 (11-43)	17 (4-35)	0.042, 0.10, < 0.001
Δ cortisol, nmol/L	140 (-50-245)	388 (260-4015)	177 (-80-230)	425 (255-965)	n.a., 0.45, 0.093
Δ free cortisol, nmol/L	59 (-71-124)	151 (74-678)	40 (-34-99)	133 (32-233)	< 0.001, 0.15, 0.16
CBG, mg/L	23 (6-43)	29 (13-63)	39 (4-60)	41 (21-65)	0.060, < 0.001, 0.90
Albumin, g/L	13 (6-32)	17 (6-24)	16 (12-28)	20 (8-32)	0.25, < 0.001, 0.89
Days from admission - test	1 (0-50)	3 (0-44)	25 (9-49)	20 (9-83)	0.57*, 0.15*
Days between tests	n.a.	n.a.	18 (8-34)	16 (8-43)	0.41*
Days after stop hydrocortisone	n.a.	n.a.	7 (5-12)	7 (5-14)	0.75*

Median (range); ACTH: adrenocorticotropic hormone, CBG: cortisol binding globulin. *Mann-Whitney U test, otherwise generalized estimating equations; n.a.: not applicable.

Table 3. Clinical variables according to Δ cortisol $<, \ge 250$ nmol/L

	Test 1 Δ < 250 nmol/L	Test 1 $\Delta \ge 250 \text{ nmol/L}$	Test 2 Δ < 250 nmol/L	Test 2 $\Delta \ge 250 \text{ nmol/L}$	P Δ cortisol, time,
	n = 17	n = 42	n = 14	n = 45	interaction
Disease severity					
APACHE II	19 (14-33)	17 (5-31)	16 (10-24)	14 (5-21)	0.002, < 0.001, 0.90
SOFA	9 (2-16)	9 (2-16)	5 (0-10)	4 (0-16)	0.46, < 0.001, 0.91
MAP, mm Hg	71 (53-102)	80 (36-114)	83 (65-102)	87 (50-127)	0.20, 0.002, 0.61
Vasopressors/inotropes	13 (76)	35 (83)	4 (29)	9 (20)	0.85, < 0.001, 0.56
MV	16 (94)	40 (95)	11 (79)	28 (62)	0.81, 0.002, 0.43
Renal replacement therapy	3 (17)	9 (20)	4 (29)	5 (11)	0.19, 0.88, 0.10
Infection					
Γemperature, C	36.8 (31.0-40.1)	36.9 (31.6-38.3)	37.1 (35.4-37.6)	36.8 (35.3-38.6)	1.0, 0.44, 0.83
Leukocytes, x10 ⁹ /L	13.4 (2.4-71.9)	13.3 (2.2-36.0)	12.7 (5.4-89.7)	9.9 (2.7-32.2)	0.35, 0.12, 0.82
SIRS	16 (94)	34 (81)	10 (71)	23 (51)	0.066, 0.002, 0.79
Sepsis	11 (65)	19 (45)	4 (29)	6 (13)	0.055, 0.001, 0.89
Positive cultures	11 (65)	20 (48)	4 (29)	8 (18)	0.15, 0.002, 0.88
Coagulation					
Platelets, x10 ⁹ /L	204 (59-661)	185 (51-803)	235 (24-419)	281 (88-721)	0.084, 0.19, 0.016
PT, INR	1.53 (0.99-2.47)	1.47 (1.05-4.77)	1.17 (0.98-5.46)	1.20 (1.02-3.04)	0.81, 0.068, 0.99
aPTT, sec	60 (35-92)	48 (31-77)	55 (38-72)	51 (31-103)	0.14, 0.94, 0.037
Outcome					
Length of ICU stay, days	41 (11-130)	30 (3-104)	36 (13-92)	30 (3-130)	0.93*,0.70*
Length of hospital stay, days	60 (21-230)	51 (15-141)	58 (31-230)	54 (15-175)	0.51*,0.70*
CU mortality	5 (29)	6 (14)	2 (14)	9 (20)	0.27**,1.00**
Hospital mortality	7 (41)	14 (33)	5 (36)	16 (36)	0.57**,1.00**

Median (range) or number (%), where appropriate; APACHE: acute physiology and chronic health evaluation; SOFA: sequential organ failure assessment; MAP: mean arterial pressure; MV: mechanical ventilation; SIRS: systemic inflammatory response syndrome; PT: prothrombin time; INR: international normalized ratio; aPTT: activated partial thromboplastin time; ICU: intensive care unit. *Mann-Whitney U test, **Fisher exact test, otherwise generalized estimating equations.

Clinical correlates

The following inversely related to Δ cortisol as a continuous variable (in GEE), independent of time: APACHE II (P < 0.001), renal replacement therapy (P = 0.047), sepsis (P = 0.055), positive cultures (0.041) and the aPTT (P = 0.020). In multiple GEE, a low Δ cortisol response was associated with sepsis (P = 0.003) and a low cortisol/ACTH ratio (P = 0.008) and their first order interaction (P = 0.003). Clinical variables for Δ cortisol <, \geq 250 nmol/L on test days are described in Table 3. Low responders had elevated APACHE II score and tended to have more often SIRS and sepsis, even though decreasing in time, than normal responders. The aPTT was higher in the first test and the platelets in the second test in low vs normal responders (interaction between Δ cortisol and time). There were no differences in ICU or hospital mortality among response groups, nor in groups with persistently low or normal responses or changing from a low to normal response or vice versa.

Correlations

Baseline total and free cortisol directly correlated at $r_s=0.78$, P<0.001; total and free Δ cortisol also correlated ($r_s=0.88$, P<0.001). At baseline, ACTH directly related to total and free cortisol ($r_s=0.34$, P=0.001 and $r_s=0.52$, P=0.001, respectively). Baseline ACTH inversely related to Δ total and free cortisol ($r_s=-0.32$, P=0.002 and $r_s=-0.49$ P=0.001, respectively). Baseline (free) cortisol inversely related to Δ (free) cortisol: min $r_s=-0.26$, P=0.005 and max $r_s=-0.45$, P=0.001. Baseline total cortisol/ACTH ratio directly related to Δ (free) cortisol ($r_s=0.25$, P=0.015 to $r_s=0.53$, P<0.001; Figure 2). Total and free cortisol/ACTH ratios directly correlated at $r_s=0.82$, P<0.001.

Figure 2. The cortisol to ACTH ratio at baseline and the subsequent response in increase in circulating total (panel A) and free (panel B) cortisol upon exogenous ACTH: $r_s = 0.25$ ($r^2 = 0.06$), P = 0.015; $r_s = 0.53$ ($r^2 = 0.28$), P < 0.001, respectively, together for for Test 1 and 2 in the course of critical illness (see text; no differences between tests). This suggests that ACTH-insensitive adrenals respond less with cortisol secretion to exogenous ACTH.

Predictive values

Baseline ACTH and cortisol/ACTH predicted a low cortisol response to exogenous ACTH (AUROC 0.72, P = 0.001, and 0.66, P = 0.017, respectively).

Discussion

Our study suggests that, particularly in sepsis, diminished adrenal sensitivity to endogenous ACTH is associated with a low cortisol response to exogenous ACTH irrespective of cortisol protein binding and stage of critical illness. This may denote adrenal dysfunction and insufficiency, relative to severity of disease.

Some studies suggest that baseline total cortisol is more elevated in patients subsequently responding less to exogenous ACTH with severe illness and elevated risk for death, than in those with normal baseline levels and responses [1,3,4,7,8,10,15,27]. In studies not reporting this relationship decreased binding molecules may have attenuated total cortisol levels [3,5-7,13,14]. Our study also suggests increasing baseline and decreasing Δ cortisol with

increasing disease severity (APACHE II scores), so that adrenal function may not be well adapted to disease severity thus suggesting insufficiency. Some studies evaluated baseline endogenous ACTH or cortisol/ACTH ratios and found relatively increased values and decreased ratios, respectively, at least in the acute phase of critical illness and predicting mortality [3,4,16-18]. Our ACTH levels are in the low normal range, as reported before [6], but, nevertheless, predicted a low response to exogenous ACTH, irrespective of stage of disease. The latter, otherwise, renders a confounding effect of intercurrent hydrocortisone treatment unlikely. This is the first report, to the best of our knowledge, suggesting adrenal dysfunction on the basis of diminished adrenal sensitivity to endogenous and exogenous ACTH, as described after etomidate administration [16,19]. The time dependency of the association between a low free cortisol/ACTH ratio and a low cortisol response to exogenous ACTH can be attributed in part to the low number of observations available on free cortisol in low responders of Test 2. In any case, our results again suggest that (changes in) free cortisol parallel (changes in) total cortisol upon ACTH testing, in spite of changing binding proteins in time [17]. Hence, free and total cortisol were almost interchangeable in the current evaluation.

Particularly during the chronic or prolonged phase of critical illness (rather than in the acute phase as in the Boonen et al study [20]), low ACTH and elevated cortisol levels can be seen, suggestive of increased sensitivity to ACTH or ACTH-independent cortisol production [2,21-23]. The latter may be caused by non-ACTH pituitary stimulants or feedback suppression of ACTH following impaired cortisol metabolism and persistent hypercortisolemia [2,21-23]. Our results do not suggest reversal nor development of this so called cortisol-ACTH dissociation, since the cortisol/ACTH ratio did not change in time. Boonen et al. [21] suggested that baseline cortisol was not under control of subnormal ACTH levels and that exogenous ACTH resulted in low responses of circulating cortisol, even though only peak and not Δ s cortisol were reported. This was attributed, at least in part, to impaired cortisol metabolism with diminished feedback ACTH secretion. However, low endogenous ACTH levels in the course of panhypopituitarism or steroid treatment, for instance, variably diminish adrenal sensitivity to exogenous ACTH only after days to weeks [2,29]. In contrast, our correlation data suggest basal and particularly free cortisol levels being under persistent control of ACTH, as reported before [4,18]. Our results refute the impaired metabolism-low endogenous ACTH hypothesis either in the acute or chronic phase of critical illness to fully explain a low Δ cortisol upon exogenous ACTH, by suggesting that relatively low responses to exogenous ACTH are associated with diminished adrenal sensitivity to endogenous ACTH, irrespective of stage of disease. Nevertheless, the discrepancy between our and Boonen et al. study [19] is hard to explain, unless their ACTH values were diluted by more extensive fluid therapy or taken at different stages of disease than in our study.

Even though disease severity diminished in time in this selected cohort, the number of patients with subnormal ACTH responses only slightly decreased and tended to relate to persistent or new occurrence of SIRS and culture-positive sepsis and coagulation disturbances (prolonged aPTT), in line with other investigators suggesting sepsis as the main risk factor for adrenal dysfunction of the critically ill [3,6,7,9-14,18,26-28]. The currently prospectively studied clinical risk factors in time closely agree with those retrospectively found by us before, even in longitudinal analysis [8,15]. This favors the idea that a low Δ cortisol in time is of clinical significance, rather than reflecting poor reproducibility of the ACTH test. Adrenal injury following adrenal thrombosis or haemorrhage may underlie the association of dysfunction with coagulation disturbances (for aPTT with Δ cortisol as a continuous variable), as described in meningococcemia and the Waterhouse-Friderichsen

syndrome, since other infections may also cause adrenal thrombosis, haemorrhage and insufficiency [2,16,24,25]. Heparin treatment is a well-known risk factor too [24]. The observed endocrine results did not bear any relation with mortality in the current study (as opposed to previous ones, ref. [1,3,7,8,10,12,13,15,27]), which may be explained by patient selection of acute phase survivors allowing repeated ACTH testing within 3 weeks, on the average, by virtue of study design. Together with the exclusion of etomidate-treated patients, this selection may also explain the relatively low frequency of adrenal dysfunction of about 30% in our patients, whereas adrenal insufficiency may occur in up to 60% of those suffering from septic shock [6-8,10-12,14,15,18,27]. Finally, we do not know nor evaluated the potentially interfering phenomenon of steroid resistance in critical illness [30].

Our study has several limitations. First, we did not routinely measure cortisol metabolism nor tissue effects in our patients because this requires elaborate techniques [8]. The patients studied are a selection since we wanted to evaluate repeated ACTH tests in various stages of disease.

Conclusions

Our data in a large cohort of critically ill patients with follow-up ACTH testing suggest that even though the HPA axis is activated during critical illness and recovery from the acute phase, diminished adrenal ACTH sensitivity predicts a low increase of cortisol to exogenous ACTH suggesting adrenal dysfunction irrespective of stage of disease. The data further suggest a role of disease severity and culture-positive sepsis. The implications for treatment by corticosteroids remain unanswered by our study. Our study may nevertheless imply imaging the adrenals to look for haemorrhage or thrombosis in future studies on sepsis-induced adrenal dysfunction to help deciding on corticosteroids.

Key messages

Even though the pituitary-adrenal axis is activated following stress in the course of critical illness, diminished adrenal sensitivity for endogenous ACTH is associated with a low increase of cortisol to exogenous ACTH.

Diminished adrenal sensitivity to endogenous and exogenous ACTH suggests adrenal dysfunction.

The data further suggest a role of disease severity and culture-positive sepsis in the adrenal dysfunction and insufficiency of the critically ill.

Abbreviations

ACTH, adrenocorticotropic hormone; APACHE II, acute physiology and chronic health evaluation; aPTT, activated partial thromboplastin time; AUROC, receiver operating characteristic area under the curve; CBG, cortisol binding globulin; CIRCI, critical illness-related corticosteroid insufficiency; CVVH, continuous veno-venous hemofiltration; GEE, Generalized Estimating Equations; HPA, hypothalamic-pituitary-adrenal; ICU, intensive care unit; PT, prothrombin time; r_s, Spearman correlation coefficient; SOFA, sequential organ failure assessment; SIRS, systemic inflammatory response syndrome.

Competing interests

The authors declare that they have no competing interests.

Authors contributions

MJ participated in the design and coordination of the study, collected the data, performed statistical analyses and drafted the manuscript. NM helped to collect the data and helped to draft the manuscript. AB participated in the design of the study and helped to draft the manuscript. AG participated in the design and coordination of the study, performed statistical analyses and helped to draft the manuscript. All authors read and approved the final manuscript.

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References

- 1. Jurney TH, Cockrell Jr JL, Lindberg JS, Lamiell JM, Wade CE. Spectrum of serum cortisol response to ACTH in ICU patients. Correlation with degree of illness and mortality. Chest. 1987;92:292 5.
- 2. Arlt W, Allolio B. Adrenal insufficiency. Lancet. 2003;361:1881 93.
- 3. Manglik S, Flores E, Lubarsky L, Fernandez F, Chhibber VL, Tayek JA. Glucocorticoid insufficiency in patients who present to the hospital with severe sepsis: a prospective clinical trial. Crit Care Med. 2003;31:1668–75.
- 4. Widmer IE, Puder JJ, Konig C, Pargger H, Zerkowski HR, Girard J, et al. Cortisol response in relation to the severity of stress and illness. J Clin Endocrinol Metab. 2005;90:4579 86.
- 5. Arafah BM. Hypothalamic pituitary adrenal function during critical illness: limitations of current assessment methods. J Clin Endocrinol Metab. 2006;91:3725 45.
- 6. Annane D, Maxime V, Ibrahim F, Alvarez JC, Abe E, Boudou P. Diagnosis of adrenal insufficiency in severe sepsis and septic shock. Am J Respir Crit Care Med. 2006;174:1319 26.
- 7. De Jong MF, Beishuizen A, Spijkstra JJ, Groeneveld AB. Relative adrenal insufficiency as a predictor of disease severity, mortality, and beneficial effects of corticosteroid treatment in septic shock. Crit Care Med. 2007;35:1896 903.
- 8. De Jong MF, Beishuizen A, Spijkstra JJ, Girbes AR, van Schijndel RJ S, Twisk JW, et al. Predicting a low cortisol response to adrenocorticotrophic hormone in the critically ill: a retrospective cohort study. Crit Care. 2007;11:R61.

- 9. Lesur O, Roussy J-F, Chagnon F, Gallo-Payet N, Dumaine R, Sarret P, et al. Proven infection-related sepsis induces a differential stress response early after ICU admission. Crit Care. 2010;14:R131.
- 10. Annane D, S bille V, Troch G, Rapha l J-C, Gajdos P, Bellissant E. A 3-level prognostic classification in septic shock based on cortisol and cortisol response to corticotropin. JAMA. 2000;283:1038 45.
- 11. Marik PE, Zaloga GP. Adrenal insufficiency during septic shock. Crit Care Med. 2003;31:141 5.
- 12. Annane D, S bille V, Charpentier C, Bollaert PE, Fran ois B, Korach JM, et al. Effect of treatment with low doses of hydrocortisone and fludrocortisone on mortality in patients with septic shock. JAMA. 2002;288:862 71.
- 13. Lipiner-Friedman D, Sprung CL, Laterre PF, Weiss Y, Goodman SV, Vogeser M, et al. Adrenal function in sepsis: the retrospective Corticus cohort study. Crit Care Med. 2007;35:1012 8.
- 14. Sprung CL, Annane D, Keh D, Moreno R, Singer M, Freivogel K, et al. Hydrocortisone therapy for patients with septic shock. N Engl J Med. 2008;358:111 24.
- 15. De Jong MF, Beishuizen A, van Schijndel RJ, Girbes AR, Groeneveld AB. Risk factors and outcome of changes in adrenal response to ACTH in the course of critical illness. J Intensive Care Med. 2012;27:37 44.
- 16. Den Brinker M, Joosten KF, Liem O, de Jong FH, Hop WC, Hazelzet JA, et al. Adrenal insufficiency in meningococcal sepsis: bioavailable cortisol levels and impact of interleukin-6 levels and intubation with etomidate on adrenal function and mortality. J Clin Endocrinol Metab. 2005;90:5110 7.
- 17. Molenaar N, Johan Groeneveld AB, Dijstelbloem HM, de Jong MF, Girbes AR, Heijboer AC, et al. Assessing adrenal insufficiency of corticosteroid secretion using free versus total cortisol levels in critical illness. Intensive Care Med. 2011;37:1986 93.
- 18. Llompart-Pou JA, Raurich JM, Ayestar n I, Fern ndez-de-Castillo AG, P rez-B rcena J, Ib ez J. Response to the high-dose corticotrophin stimulation test depends on plasma adrenocotropin hormone levels in septic shock. J Crit Care. 2012;27:276 82.
- 19. Molenaar N, Bijkerk RM, Beishuizen A, Hempen CM, de Jong MF, Vermes I, et al. Steroidogenesis in the adrenal dysfunction of critical illness: impact of etomidate. Crit Care. 2012;16:R121.
- 20. Lee MKV, Vasikaran S, Doery JCG, Wijeratne N, Prentice D. Cortisol: ACTH ratio to test for primary hypoadrenalism: a pilot study. Postgrad Med J. 2013;89:617–20.
- 21. Boonen E, Vervenne H, Meersseman P, Andrew R, Mortier L, Declercq PE, et al. Reduced cortisol metabolism during critical illness. N Engl J Med. 2013;368:1477–88.

- 22. Vermes I, Beishuizen A, Hampsink RM, Haanen C. Dissociation of plasma adrenocorticotropin and cortisol levels in critically ill patients: possible role of endothelin and atrial natriuretic hormone. J Clin Endocrinol Metab. 1995;80:1238–42.
- 23. Bornstein SR, Engeland WC, Ehrhart-Bornstein M, Herman JP. Dissociation of ACTH and glucocorticoids. Trends Endocrinol Metab. 2008;19:175 80.
- 24. Dahlberg PJ, Goellner MH, Pehling GB. Adrenal insufficiency secondary to adrenal hemorrhage. Arch Intern Med. 1990;150:905 9.
- 25. Paolo Jr WF, Nosanchuk JD. Adrenal infections. Int J Infect Dis. 2006;10:343 53.
- 26. Briegel J, Schelling G, Haller M, Mraz W, Forst H, Peter K. A comparison of the adrenocortical response during septic shock and after complete recovery. Intensive Care Med. 1996;22:894 9.
- 27. Goodman S, Sprung SL, Ziegler D, Weiss YG. Cortisol changes among patients with septic shock and the relationship to ICU and hospital stay. Intensive Care Med. 2005;31:1362 9.
- 28. Guzman JA, Guzman CB. Adrenal exhaustion in septic patients with vasopressor dependency. J Crit Care. 2007;22:319 23.
- 29. Reimondo G, Bovio S, Allasino B, Terzolo M, Angeli A. Secondary hypoadrenalism. Pituitary. 2008;11:147 54.
- 30. Guerrero J, Gatica HA, Rodr guez M, Estay R, Gœcke IA. Septic serum induces glucocorticoid resistance and modifies the expression of glucocorticoid isoforms receptors: a prospective cohort study and in vitro experimental assay. Crit Care. 2013;17:R107.



