

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/345435958>

Assessing tiger corridor functionality with landscape genetics and modelling across Terai–Arc Landscape, India

Preprint · October 2020

DOI: 10.1101/2020.10.24.353789

CITATIONS

4

READS

735

6 authors, including:



Suvankar Biswas

WWF India

21 PUBLICATIONS 98 CITATIONS

SEE PROFILE



Supriya Bhatt

Wildlife Institute of India

15 PUBLICATIONS 91 CITATIONS

SEE PROFILE



Bivash Pandav

Wildlife Institute of India

116 PUBLICATIONS 1,705 CITATIONS

SEE PROFILE

**Assessing tiger corridor functionality with landscape genetics and modelling across
Terai-Arc Landscape, India**

Suvankar Biswas[#], Supriya Bhatt[#], Debanjan Sarkar[#], Gautam Talukdar[#], Bivash Pandav^{#*}
and Samrat Mondol^{#*}

[#] Wildlife Institute of India, Dehradun, India

*Corresponding authors:

Samrat Mondol, Ph.D., Wildlife Institute of India, Dehradun, India. Email-
samrat@wii.gov.in

Bivash Pandav, Ph.D., Wildlife Institute of India, Dehradun, India. Email-
bivash.pandav@wii.gov.in

Abstract

India led the global tiger conservation initiatives since last decade and has doubled its wild tiger population to 2967 (2603-3346). The survival of these growing populations residing inside the continuously shrinking habitats is a major concern, which can only be tackled through focused landscape-scale conservation planning across five major extant Indian tiger landscapes. The Terai-Arc landscape (TAL) is one of the ‘global priority’ tiger conservation landscapes holding 22% of the country’s wild tigers. We used intensive field-sampling, genetic analyses and GIS modelling to investigate tiger population structure, source-sink dynamics and functionality of the existing corridors across TAL. Genetic analyses with 219 tigers revealed three low, but significantly differentiated tiger subpopulations. Overall, we identified Seven source and 10 sink areas in TAL through genetic migrant and gene flow analyses. GIS modelling identified total 19 (10 high, three medium and six low conductance) corridors in this landscape, with 10 being critical to maintain landscape connectivity. We suggest urgent management attention towards 2707 sq. km. non-protected habitat, mitigation measures associated with ongoing linear infrastructure developments and transboundary coordination with Nepal to ensure habitat and genetic connectivity and long-term sustainability of tigers in this globally important landscape.

Keywords: Tiger dispersal, Source-sink dynamics, *Panthera tigris tigris*, Landscape connectivity, Anthropogenic impacts, Priority conservation areas

Introduction

The tiger (*Panthera tigris*) exemplifies one of the major conservation efforts globally. Once distributed across ~30 present-day nations ranging from Armenia to Indonesia and the Russian Far East to India covering a variety of habitats¹, their distribution and numbers have drastically reduced to less than 4000 wild individuals at the beginning of the twentieth century largely due to habitat loss and human persecution^{2,3}. Despite country-specific conservation efforts, the wild tiger population size was found to be >3200 individuals in 2010⁴, leading to a commitment from the heads of 13 tiger-range countries to double their tiger number by 2022⁵. As majority of the remaining tigers persisted as small, and often isolated populations^{2,6} conservation strategies mostly focused on landscape-based approaches to attain recovery goals where habitat improvement, enhanced protection measures, prey augmentation were targeted^{2,3,7,8}. Several priority landscapes were identified as ‘Tiger Conservation Landscapes’ (TCLs)², where identification of key source populations and consolidation and improvement of surrounding habitats were emphasized^{9–11}. Given that ~80% of global wild tiger population living in most wide variety of habitats were found in India at that time¹², the success of the tiger recovery plan and the future survival of the species was mostly dependent on India’s conservation actions. In 2019, India announced the news of doubling its tiger number since 2006 (population estimate of 1411 (1165-1675) in 2006 to 2967 (2603-3346) in 2018) within its existing habitats¹³. However, these increasing tiger populations now face enormous challenges from increasing human density, rapid urbanisation, expanding agriculture, aggressive infrastructure development and economic growth^{14,15}, and their future persistence will depend on the balance between the developmental demands and conservation requirements.

Managing wide-ranging, territorial species like tiger at landscape level requires in-depth understanding of key source populations⁴, identification of potential tiger habitats

surrounding these source populations and ensuring habitat connectivity within the landscape^{9,10}. At local level, measures of population dynamics including changes in recruitment and mortality, differential rates of immigration and emigration across sex and age classes, population turnover rates etc. are essential, whereas at landscape scale most emphasis should be given on habitat connectivity to enhance gene flow and reduce the risks from inbreeding^{16–18}. Both of these are of critical importance in the Indian scenario as most of the available protected land (~5% as per the Wildlife Protection Act 1972) is extremely fragmented¹⁹ and maintaining connectivity by identifying and managing the critical corridors will play the key role in future tiger conservation^{4,20}. However, generating such detailed information is challenging, time consuming and resource intensive at any scale. In the Indian subcontinent, long-term ecological studies have already identified priority landscapes for tiger conservation (for example, Western Ghats, central India, north-eastern India, Sundarbans and the alluvial Terai flood plains in the Himalayan foothills) that support high potential tiger densities and relatively larger population sizes¹³. Earlier studies have helped us to understand source-sink population dynamics at landscape-scale in Western Ghats^{8,11,21} and central Indian landscapes^{17,22–26} along with Terai habitats of Nepal^{27,28}, but such information is missing from the Indian part of the Terai-Arc landscape (TAL), where population connectivity and their relationship with currently available habitats is poorly understood.

In this paper, we used a combination of intensive field surveys, non-invasive genetic tools and GIS modeling to assess the tiger population connectivity across the TAL. More specifically, we investigated (1) tiger presence and population structure across this landscape covering protected as well as non-protected habitats; (2) population connectivity in terms of tiger source-sink population dynamics in TAL; and (3) identify the critical corridors that helps maintaining the metapopulation dynamics. We addressed these questions by using 219 genetically identified unique tigers across TAL and details from already identified corridors in this landscape²⁹.

Methods

Research permissions and ethical considerations

All required permissions for field survey and faecal sampling were provided by the Forest Departments of Uttarakhand (Permit no: 90/5-6), Uttar Pradesh (Permit no: 1127/23-2-12(G) and 1891/23-2-12) and Bihar (Permit no: Wildlife-589). Due to the non-invasive nature (faecal sample based) of the work, no ethical clearance was required in this study.

Study area

This study was conducted in the Indian part of the TAL. The TAL is the only prime tiger habitat found along the foothills of Himalayas^{20,29}. Covering approximately 28000 km² of forested habitat in the northern India and southern part of Nepal, this region is one of the most important global tiger conservation landscapes^{20,29}. The Indian TAL is a linear 900 km long and 50 km wide landscape with 15000 km² tiger habitat encompassing the north Indian states of Haryana (westernmost), Himachal Pradesh, Uttarakhand, Uttar Pradesh and Bihar (easternmost) (Fig. 1a). Situated at the Himalayan foothills, the habitat supports tropical moist deciduous forests dominated by Sal (*Shorea robusta*), tall Terai swamp grasslands and permanently moist reed swamps³⁰. This landscape is identified as a ‘Global priority’ tiger conservation landscape (TCL)² and retains about 22% of India's wild tiger population¹³. As recent as 2003, tigers were reported across this entire landscape²⁹ but currently found only in the states of Uttarakhand, Uttar Pradesh and Bihar¹³.

This transboundary tiger habitat in TAL comprises a network of Protected Areas (PAs) and multiple use Forest Divisions (FDs) that maintain habitat connectivity through forest corridors^{20,29}. The first comprehensive landscape-scale study carried out on tiger distribution in Indian TAL by Johnsingh *et al.* (2004)²⁹ highlighted the issue of habitat fragmentation. The study also identified nine tiger habitat blocks (THBs) and 13 structural corridors that potentially facilitate tiger dispersals (Fig. 1a). Currently this entire landscape has 13 PAs

(including six Tiger Reserves (TRs), 12 FDs and three Social Forestry Divisions (SFDs)). According to the latest tiger population estimation report¹³ this landscape hosts 646 (567-726) individual tigers, with a ~33% population increase since the last estimation (n=485 (427-543)³¹). All relevant details for the THBs are provided in Supplementary Table S1.

Field sampling

In this study, we aimed to use both direct and indirect approaches (intensive field sampling, genetic analyses and GIS tools) to assess tiger presence, source-sink dynamics and functional connectivity across the entire tiger habitats in TAL. To achieve this goal at such a large landscape scale, it is important to conduct intensive sampling throughout the target study area. We conducted extensive field surveys covering all the THBs (including both PAs as well as FDs) of Uttarakhand, Uttar Pradesh and Bihar between December 2014 to May 2018 (Supplementary Table S1). During field surveys, a team of 5-6 experienced trackers walked this entire region searching for large carnivore faecal samples along all the potential trails and other habitats in this region. Sampling was mostly conducted between November-May i.e. winter and summer seasons every year. The total effort included ~9500 km of foot survey for faecal sampling.

In the field, we categorized large feline carnivore faeces (tiger and leopard) through morphological features such as size, shape, diameter as well as associated field signs (track and scrape marks etc.). We used a dry sampling approach described in Biswas *et al.* (2019)³² for the entire study. Samples were collected with GPS coordinates and kept in dry, dark boxes in the field for a maximum period of one month before they were shipped to the laboratory, where they were stored in -20 °C freezers until further processing. During the sampling period, we collected a total of 1608 relatively fresh large carnivore faecal samples from the study area. The locations of the samples are provided in Supplementary Fig. S1.

DNA extraction, species, individual and sex identification

We extracted DNA from all the field-collected faeces using protocol described in Biswas *et al.* (2019)³². In brief, we swabbed the top layer of each sample twice with PBS-soaked sterile cotton swabs. Each swab was subsequently lysed overnight at 56 °C with 330 µl ATL buffer and 30 µl of proteinase K (20 mg/ml). Following digestion, swabs were removed and the remaining extraction was followed by the Qiagen DNeasy Tissue Kit (Qiagen Inc., Hilden, Germany) standard protocol. DNA was eluted twice in 100 µl of TE buffer (pH 7.8). For every set of samples (n=22) two extraction negatives were included to monitor contamination.

For species identification, we combined two earlier described tiger-specific molecular markers^{33,34}. The PCR reactions were carried out in 10 µl reaction volumes containing 4 µl of Qiagen master mix (Qiagen Inc., Hilden, Germany), 3 µl of BSA, 1 µl of primer mix and 2 µl of template DNA. The PCR conditions included an initial denaturation (95 °C for 15 min); 50 cycles of denaturation (95 °C for 30 sec); annealing (50 °C for tiger and 57 °C for leopard for 30 sec); extension (72 °C for 30 sec); followed by a final extension (72 °C for 15 min). PCR products were visualized in 2% agarose gels and species-specific band patterns were used for unambiguous species identification. Only tiger samples were subsequently used for individual level analyses.

For individual identification, we used an already standardized panel of 13 microsatellite markers described in Mondol *et al.* (2009)³⁵, Mondol *et al.* (2012)³⁶. These markers were amplified as 10 µl multiplex reactions containing 4 µl of Qiagen multiplex master mix (Qiagen Inc., Hilden, Germany), 3 µl of BSA, 1 µl of primer mix (2 µM concentration) and 3 µl of template DNA. The PCR conditions included an initial denaturation (95 °C for 15 min); 45 cycles of denaturation (94 °C for 30 sec); annealing (Ta for 30 sec); extension (72 °C for 30 sec); followed by a final extension (72 °C for 15 min) in an ABI thermocycler. Post-amplification, 1 µl of PCR product was mixed with 9 µl mix of Hi-Di formamide and 500

LIZ (Applied Biosystems, Foster City, CA, USA) and genotyped in ABI 3500XL DNA fragment analyzer. For genotype validation, we performed a modified multiple tube approach³⁷. Each locus was genotyped and alleles were scored four independent times and a quality index³⁵ was calculated. Only loci with 0.75 or higher quality were retained for further analyses. For allele calling, we expanded the earlier tiger microsatellite allele bins for the north Indian population in program GENEMARKER 2.6.7. (Softgenetics Inc., State College, PA, USA) and scored the alleles manually.

We performed molecular sexing for only individually identified tigers using two sex chromosome markers Amelogenin³⁸ and SRY³⁹. PCR reactions were carried out in 10 µl volumes containing 4 µl of Qiagen master mix (QIAGEN Inc.), 3 µl of BSA, 1 µl of primer mix (3 µM concentration) and 2 µl of template DNA. The PCR conditions included an initial denaturation (95 °C for 15 min); 50 cycles of denaturation (95 °C for 30 sec); annealing (Ta for 30 sec); extension (72 °C for 30 sec), followed by a final extension (72 °C for 15 min) in an ABI thermocycler. PCR products were run in 3% agarose gel and sex identification was done visually through sex-specific banding patterns. For validation, we repeated the entire process twice and only samples provided consensus results were considered further.

Data analyses

Microsatellite summary statistics

We calculated average amplification success as the percent positive PCR for each locus, as described by Broquet and Petit (2004)⁴⁰. We quantified allelic dropout and false allele (FA) rates manually as the number of allelic dropouts or FAs over the total number of amplifications, respectively⁴⁰, as well as using MICROCHECKER v 2.2.3.⁴¹.

Post data quality assessment, we selected only those samples with good quality data for at least nine loci (out of 13) for further analyses. We identified unique tigers by removing samples with identical genotypes using the 'Identity analysis' module of program CERVUS

202 ⁴² and removed the ‘genetic recaptures’ from the dataset. We used the program GIMLET ⁴³ to
203 calculate the probability of identity for siblings (PID_(sibs)) and unbiased (PID_(unbiased)) for all
204 the individuals. Any allele with >10% frequency across all amplified loci were rechecked and
205 confirmed. We calculated all summary statistics for genetic diversity as well as Hardy
206 Weinberg equilibrium and linkage disequilibrium using program ARLEQUIN 3.1 ⁴⁴.

207 ***Habitat connectivity in TAL***

208 Assessing tiger connectivity directly (through radio-telemetry or camera trapping) at a
209 landscape level is challenging due to their secretive behaviour, and logistical constraints
210 resulting in infrequent information ⁴⁵. The TAL tiger habitat has 13 identified corridors ²⁹ and
211 we aimed to evaluate their current functionality. We took three independent indirect
212 approaches to understand habitat connectivity in TAL. First, we generated individual level
213 tiger genetic information across the entire tiger landscape and used this data to assess any
214 spatial population structure (indicating loss of connectivity). Subsequently, we conducted a
215 number of specific analyses to evaluate source-sink dynamics (using directional gene flow
216 and migrant analyses as proxies of habitat connectivity) involving all the THBs within this
217 landscape to corroborate the population structure results. Finally, we modelled the
218 connectivity between PAs of TAL following Circuit Theory and identified critical corridors
219 to maintain the functionality of this landscape.

220 For spatial clustering, we performed a Bayesian analysis using program TESS2.3.1 ⁴⁶ with an
221 admixture model with 60000 sweeps and 10000 burn in. We evaluated the most likely
222 number of cluster (K), testing values between 2 to 10 ⁴⁷, where clusters were identified
223 through average deviance information criterion (DIC) value for each K. Further, we
224 calculated genetic differentiation (F_{st} and G_{st}) among tiger genetic subpopulations identified
225 through spatial clustering analyses in GenALEx version 6.5 ⁴⁸.

226 To assess tiger source-sink dynamics, we performed two different analyses focusing on
227 ‘detection of genetic migrants’ and ‘assessment of rate and direction of gene flow’ within
228 each tiger subpopulation. To detect tiger dispersals, we used two different approaches that
229 use allele frequencies to detect migrant individuals. First, we used prior population
230 information in the USEPOPINFO option implemented in STRUCTURE 2.3.2 ⁴⁹ to detect
231 first-generation migrants in our sampled populations. We detected the number of clusters (K
232 between 1-10) based on 10 independent runs with 500,000 iterations and a burn-in of 50,000
233 assuming an admixture model. We considered the membership coefficient (q) above 0.9 as a
234 realistic cut-off value to assign an individual to a population ²⁴. We assigned different
235 migration rates (MIGPRIOR 0.01, 0.05, 0.1) as a sensitivity test. We ran the analysis with
236 two separate dataset: a) individuals grouped as populations according to their sampling
237 locations and b) individuals grouped as genetic clusters from our initial run. These different
238 runs helped us to evaluate the consistency of the results across different genetic groups
239 created.

240 Further, we used the ‘Migrant detection’ function in program GENECLASS 2.0 ⁵⁰ to identify
241 the first-generation migrant tigers. Here, we used a Bayesian approach as described by
242 Rannala and Mountain (1997) ⁵¹ along with the resampling method of Paetkau *et al.* (2004) ⁵²
243 for likelihood computation ($L_{\text{home}}/L_{\text{max}}$). The run parameters included 10000 simulated
244 individuals with a threshold alpha value of 0.01 (Type 1 error) ⁵². This method allows
245 detection of migrants even when the overall differentiation between populations is low. Apart
246 from first generation migrant detection, we did individual assignments in GENECLASS
247 using Bayesian criterion of Rannala and Mountain (1997) ⁵¹ in combination without
248 resampling with rest of the parameters same as described above. Subsequently, we measured
249 the magnitude and direction of recent gene flow using the Bayesian MCMC based approach
250 implemented in BAYESASS v.3.0.4 ⁵³, with run parameters of 1000000 iterations and

100000 burn-in. This approach is generally used to detect recent gene flow (5-7 generations) between populations⁵⁴.

We developed habitat permeability layer to understand potential tiger connectivity in TAL, using the maximum entropy modelling approach in the program MaxEnt v 3.4.1⁵⁵. Habitat permeability layer and focal nodes (regions between which connectivity is to be modelled) are the primary input data for connectivity analyses, indicating the difficulty experienced by an individual in moving across a landscape⁵⁶. We used tiger presence points and environmental variables in MaxEnt v 3.4.1⁵⁵ to develop a habitat permeability layer, where each pixel is a proxy of the likelihood that individuals will move through that cell (i.e. conductance). After removing the spatial cluster between point locations, we used a total 465 presence points along with five environmental variables based on tiger's ecology and habitat requirements, viz. forest cover, distance to protected areas, distance from water, road, and settlements (Supplementary Table S2). For running the model, we split the presence points as training (70%) and testing (30%) during cross-validate run with 10 replicates and rest of the settings were kept as default. We used the average of all outputs (n=10 runs) as the habitat permeability layer for Circuitscape. Subsequently, we used export to Circuitscape tool (http://www.jennessent.com/arcgis/Circuitscape_Exp.htm) in ArcGIS 10.7 to export the focal nodes and habitat permeability layer to ASCII raster layers of equal extents, cell sizes, and spatial references. We used a total of 21 focal nodes (Supplementary Table S3) and habitat permeability layer in Circuitscape v.4.0⁵⁶ to measure the corridor conductance across TAL.

We provided weightage to each of the focal nodes based on tiger source-sink populations as identified in this study. We set the node weightages in four categories that ranged from 0 (nodes with no tiger presence), 0.25 (nodes that act as sink populations), 0.5-0.75 (nodes those are moderate-level source populations) and 1 (nodes those are primary source populations) (Supplementary Table S3) during analyses. Subsequently, based on conductance

value we classified the output corridors into three categories: low functioning (0-0.30), medium functioning (0.31-0.70) and high functioning (0.71-1.0).

To identify the corridor bottlenecks, we used ‘Pinchpoint mapper’ in Linkage mapper toolbox V2.0 in ArcGIS 10.7. We converted the habitat permeability layer into a resistance surface using SDM toolbox ⁵⁷ in ArcGIS 10.7 (www.esri.com) for running the analysis. Finally, we modelled the least-cost pathways (LCPs) for tiger dispersal using Linkage mapper toolbox V2.0 in ArcGIS 10.7.

Results

Tiger data from TAL

Out of 1608 field-collected large carnivore faecal samples, we extracted DNA from 1524 samples (94.78%). Remaining 84 samples (5.22%) showed fungal growth across the top-layer and were not processed. Post tiger-specific assays we identified 743 tiger faeces (48.75%) (Supplementary Fig. S2) for downstream individual identification. Using 13 microsatellite panel ^{35,36} we subsequently identified 219 unique tiger individuals across TAL (Fig. 1b), covering approximately 35% of the estimated tiger population from this landscape (n=646 (567-726) ¹³). Locus-wise mean success rate across all samples ranged between 53-97% (Table 1). We found no large-scale signatures of allele drop out or null alleles. The mean allelic dropout and false allele rates as 0.01 and 0.03, respectively (Table 1), indicating an overall low genotyping error rate across this marker panel. All 13 loci were polymorphic for TAL tigers with mean number of alleles, observed heterozygosity and allelic size range were found to be of 14.76 ± 2.83 , 0.35 ± 0.17 and 45.38 ± 10.96 , respectively (Table 1). The cumulative $PID_{(sibs)}$ and $PID_{(unbiased)}$ values (2.24×10^{-6} and 5.99×10^{-17} , respectively) suggest a statistically strong result for unambiguous tiger individual identification (Table 1). We identified a total of 35 genetic recaptures across TAL. The details of area-wise genetic

recaptures are as follows: Rajaji TR- 10, Corbett TR- 11, Pilibhit TR- 2, Lansdowne FD- 5, Hardwar FD- 1, Ramnagar FD- 5 and Terai East FD- 1.

Of all the identified unique individuals (n=219) ~71% were from Uttarakhand (n=156, 35% of the state's population of 442 (393-491)), whereas ~20% were from Uttar Pradesh (n=44, 25% of the state's population of 173 (148-198)) and ~9% from Bihar (n=19, 61% of the state's population of 31 (26-37))¹³. We confirmed tiger presence in 12 of the 13 earlier described THBs, representing 10 PAs, nine FDs and two SFDs (Supplementary Fig. S2, Supplementary Table S1). THB VII and part of THB IV did not show any tiger evidence (Supplementary Fig. S2). We achieved an 88% success rate in molecular sexing among the unique tigers (n=193, 89 males and 104 females, respectively). Interestingly, 54 of these males were found inside PAs and 35 individuals were from non-PAs (FDs and SFDs), whereas 59 females were from PAs and 45 from outside PAs, indicating that both sexes were using non-protected habitats across TAL.

Population structure and genetic differentiation of tigers across TAL

Our Bayesian clustering analyses with 219 individual tiger genetic data revealed four genetic lineages (K=4) across TAL. When examined closely, we found that Uttarakhand state holds two of these four genetic lineages as geographically mixed populations, whereas the remaining two lineages are roughly separated in states of Uttar Pradesh and Bihar, respectively, making three genetic subpopulations across TAL (Fig. 1b). Following the description of 'Tiger Habitat Blocks (THBs)' mentioned by Johnsingh *et al.* (2004)²⁹, we report these three genetic subpopulations as 'Tiger Genetic Blocks (TGBs)' while reporting the results.

Overall, these TGBs roughly correspond to the tiger population in the states of Uttarakhand (TGB I), Uttar Pradesh (covering mostly TGB II and small parts of TGB I and III) and Bihar (TGB III), respectively (Fig. 1b). The samples collected from THB I, II and III (n=171)

formed the first genetic subpopulation (TGB I) (Figs. 1a, 1b). The TGB I is spread between the Rajaji TR (western side) to Nandhaur Wildlife Sanctuary (WLS) (eastern side) in Uttarakhand, covered THB I, II and III and retains the highest tiger population in TAL¹³ (Figs. 1a, 1b). This block represents six PAs, eight FDs, one SFD and nine earlier described corridors (Figs. 1a, 1b, Supplementary Table S1)²⁹. The TGB II includes the samples collected from THB IV, V and VI (n=26) and spread between Pilibhit SFD (western side) to Katarniaghat WLS (eastern side) in Uttar Pradesh (Figs. 1a, 1b). This block retains two PAs, one FD, one SFD and three corridors (Figs. 1a, 1b, Supplementary Table S1)²⁹. Finally, the samples collected from THB VIII and THB IX (n=22) together formed the TGB III and included only two PAs, Sohagibarwa WLS (western side) in eastern Uttar Pradesh and Valmiki TR (eastern side) in Bihar (Figs. 1a, 1b, Supplementary Table S1).

Our analyses revealed that the TGBs are genetically differentiated (Fst and Gst) at low, but significant levels. The Fst value ranged between 0.079-0.107, while the Gst value ranged between 0.067-0.087 (Table 2). Comparative summary statistic analyses among these TGBs showed that TGB I has the highest mean number of alleles (10.15 ± 3.06) when compared to TGB II (7.00 ± 2.11) and TGB III (9.08 ± 2.24), respectively. However, TGB III showed higher observed heterozygosity and the allelic size range values (0.43 ± 0.20 and 38.00 ± 14.29 , respectively) than TGB I (0.34 ± 0.20 and 24.46 ± 07.37 , respectively) and TGB II (0.29 ± 0.12 and 24.77 ± 09.30 , respectively) (Table 3).

Gene flow and tiger source-sink populations within TGBs

We analysed the genetic data to understand the tiger source-sink population dynamics (which also confirmed connectivity among populations) within each of the three already identified TGBs in TAL. Within the TGB I, ‘genetic migrant detection’ and gene flow analysis identified two major habitat complexes with genetic connectivity: The Rajaji-Lansdowne-Haridwar region and the Corbett-Ramnagar-Terai FDs-Haldwani region. In the Rajaji-

350 Lansdowne-Haridwar habitat complex we identified five first generation migrant tigers
351 (Supplementary Table S4): two migrants each from Lansdowne FD to Rajaji TR and
352 Haridwar FD and one from Rajaji TR to Lansdowne FD (Supplementary Table S4). Further
353 Bayesian analyses show higher rates of gene flow between Lansdowne FD to Rajaji TR and
354 Rajaji TR to Haridwar FD, whereas Najibabad SFD shows signatures of immigration from
355 both Rajaji TR and Lansdowne FD (Fig. 2a). Combining this information, we interpret that
356 both Rajaji TR and Lansdowne FD act as source tiger populations and Haridwar FD and
357 Najibabad SFD are sink populations in this habitat complex, respectively (Fig. 2a).

358 Similarly, in the Corbett-Ramnagar-Terai FDs-Haldwani habitat complex we identified 12
359 first-generation migrant tigers (Supplementary Table S4), showing extensive genetic
360 connectivity among the sampled areas within the habitat complex. We found three migrants
361 from Ramnagar FD to Corbett TR and two migrants to Haldwani FD (Supplementary Table
362 S4). We detected one migrant tiger from Terai West FD to Ramnagar FD (Supplementary
363 Table S4). We found two migrants from Terai Central FD to Terai West FD, one migrant to
364 Ramnagar FD and one to Champawat FD (Supplementary Table S4). We also found one
365 migrant individual from Haldwani FD to Terai East FD (Supplementary Table S4). In
366 addition, we detected one migrant individual from Najibabad SFD to Amangarh TR
367 (Supplementary Table S4). Bayesian analyses show higher gene flow from Corbett TR to
368 adjoining Najibabad SFD and Amangarh TR (Fig. 2a). Corbett TR also shows a high rate of
369 bidirectional gene flow with Ramnagar FD and Terai West FD. Similarly, Terai West FD is
370 highly interconnected with Amangarh TR, Terai Central FD and Ramnagar FD (Fig. 2a). The
371 other areas such as Haldwani FD, Ramnagar FD, Terai Central FD and Terai East FD have
372 lower rates of gene flow among them (Fig. 2a). Combined together, this information suggests
373 that Corbett TR is the main source population for Ramnagar FD, Amangarh TR, Terai West
374 FD and Najibabad SFD, whereas Ramnagar FD is the source tiger population for Haldwani,

Champawat and Terai East FDs (Fig. 2a). Within the two major tiger habitat complex in TGB I (Rajaji-Lansdowne-Haridwar and Corbett-Ramnagar-Terai FDs-Haldwani) connectivity is maintained through the Najibabad SFD, which acts as a sink population for both Corbett TR and Lansdowne FD (Fig. 2a, Supplementary Table S4).

Within TGB II, the tiger habitats are comparatively more fragmented than TGB I and we identified 10 first-generation migrant tigers (Supplementary Table S4). We detected two migrants from Pilibhit TR to Dudhwa National Park (NP) (part of Dudhwa TR) and one migrant to Kishanpur WLS (part of Dudhwa TR) (Supplementary Table S4). We also found two migrants from Dudhwa NP to Pilibhit TR, one migrant to Kishanpur WLS (part of Dudhwa TR) and two migrants to South Kheri FD (buffer zone of Dudhwa TR) (Supplementary Table S4). Further, we detected two migrants from Kishanpur WLS to Pilibhit SFD (Supplementary Table S4). We measured higher gene flow from Pilibhit TR to Dudhwa NP, Kishanpur WLS, South Kheri FD and Pilibhit SFD (Fig. 2b). We also estimated higher rate of bidirectional gene flow among Dudhwa NP, Kishanpur WLS and Katarniaghat WLS, along with unidirectional gene flow to adjoining South Kheri FD (Fig. 2b). This information suggests Pilibhit TR and Dudhwa TR are the major source tiger populations, and South Kheri FD and Pilibhit SFD are the sink populations in the TGB II.

Finally, within TGB III we detected two first-generation migrants from west Sohagibarwa WLS to Valmiki TR (Supplementary Table S4). This result was surprising as within this habitat block Valmiki TR has the highest tiger population¹³. However, the Bayesian analysis results indicate higher gene flow from Valmiki TR and east Sohagibarwa WLS to west Sohagibarwa WLS (Fig. 2c). Combined together, we interpret that Valmiki TR-east Sohagibarwa WLS is the source population for west Sohagibarwa WLS in TGB III.

Corridor connectivity across TAL

Our habitat permeability analyses indicated certain habitat variables such as distance to forest cover and protected areas are the main governing factors of tiger dispersal (Supplementary Table S5). Out of the 13 earlier-described corridors ²⁹, twelve showed conductance for tiger dispersal (Fig. 3a, Supplementary Table S6). Towards the western end of TAL Yamuna river corridor did not show any signatures of habitat conductance (Fig. 3a, Supplementary Table S6). The remaining corridors showed high to low conductances (Fig. 3a, Supplementary Table S6). The details of the least-cost pathways (LCP) are provided in Supplementary Table S6.

In addition, we identified seven new corridors in TAL. The first corridor (high conductance) connects Rajaji TR (eastern part) with Najibabad SFD through Lansdowne FD (Fig. 3a, Supplementary Table S6). The second corridor (high conductance) connects Jhilmil Jheel CR with Najibabad SFD through Haridwar FD (Fig. 3a, Supplementary Table S6). The third corridor (high conductance) connects Corbett TR with Najibabad SFD (Fig. 3a, Supplementary Table S6), making these areas very well connected with each other within TGB I. In TGB II, we identified two new corridors (high conductance), where the first one connected the northern and southern part of Pilibhit TR, and the second one connected Pilibhit TR to Kishanpur WLS through North Kheri FD, respectively (Fig. 3a, Supplementary Table S6). Finally, two new corridors were identified in TGB III, the first one (low conductance) connecting western and eastern part of Sohagibarwa WLS and the easternmost corridor (high conductance) connecting two parts of Valmiki TR, respectively (Fig. 3a, Supplementary Table S6). The corridor bottleneck analysis identified 10 critical tiger dispersal corridors in TAL (Fig. 3a, Supplementary Table S6).

Discussion

We used a multidisciplinary approach to evaluate all tiger corridor functionalities at a landscape level across the TAL. To the best of our knowledge, this is the first study using

genetic analyses and corridor modelling to understand tiger source-sink dynamics and connectivity at a landscape level in TAL. We conducted extensive faecal sampling across all tiger habitats and dispersal corridors and confirmed presence of tiger in most of the earlier described THBs in TAL. These areas include already known TRs and FDs¹³ along with new areas in this landscape (Supplementary Fig. S2, Supplementary Table S1). Our field surveys corroborated with early reports of tiger absence beyond western part of Rajaji TR, Suhelwa WLS, and remaining areas between Dudhwa TR and Sohagibarwa WLS (Fig. 1b, Supplementary Fig. S2)^{13,29}, possibly due to high anthropogenic pressure and habitat loss. Further, we confirmed tiger presence from Jhilmil Jheel Conservation Reserve (CR) (Uttarakhand), Pilibhit SFD and Najibabad SFD (Uttar Pradesh) where they were not reported earlier (Supplementary Fig. S2)¹³. This expansion in tiger habitat occupancy is possibly due to increasing tiger dispersals to newer areas driven by recent increase in their population in this landscape¹³. Tigers are territorial and high resource-demanding (in terms of both food and space) animals and such dispersal events are thus normal⁵⁸. In the Indian subcontinent such dispersal events have been reported from other landscapes also (Northwest India⁵⁹, Central India^{17,24,25}, Nepal-TAL^{27,28}) and ~35% of the country's tiger population is reported to live outside the existing TRs¹³. Interestingly, we found that both male and female tigers beyond protected areas including relatively marginal habitats of FDs and SFDs indicating both-sex dispersal events in TAL, as found in other habitats (Central India²⁵, Nepal-TAL²⁸).

Our genetic analyses identified three genetic subpopulations (named as Tiger Genetic Blocks or TGBs) in TAL. This was an unexpected result considering we found tiger presence in majority of the protected and non-protected areas, except some parts of the central and the eastern TAL (Fig. 1b). The easternmost subpopulation TGB III covering Sohagibarwa WLS and Valmiki TR is physically separated from the central and western TAL without any forest

connectivity²⁹. Tigers from Valmiki TR have also been reported earlier as genetically unique⁶⁰. Even in the Nepal part of TAL, three genetic subpopulations have been reported those are connected with TGB II and TGB III²⁸, indicating genetic discontinuity across transboundary sections of TAL in this region. However, the pattern of genetic discontinuity between TGB I and TGB II within the seemingly continuous habitat of central and western TAL was surprising. Similar patterns of tiger population structure has been earlier reported from other tiger landscapes in the subcontinent (for example, Central Indian landscape^{61,25}, Western TAL⁶², Nepal-TAL²⁸). Even in the same TAL landscape Bhatt *et al.* (2020)⁶³ has reported two leopard genetic subpopulations, possibly driven by landscape features. When compared with this study, it makes ecological sense that TAL leopards showed less population structure than tigers (three TGBs) as they are more habitat generalist and even found in human-dominated areas⁶⁴. Unlike the other major tiger landscapes in India, both of these sympatric species face one common problem related to this linear landscape where one-dimensional space (resulting in restricted movement opportunities) combined with very high human density makes the entire landscape extremely vulnerable to fragmentation at a relatively small temporal scale²⁹. We strongly feel that the genetic separation between TGB I and II is driven by human-disturbance mediated loss of corridor functionality leading to population differentiation at small, but significant levels (Table 2).

Within each of these TGBs the genetic data helped us to understand the source-sink dynamics and fine-scale population connectivity patterns between the protected and non-protected areas. Our analyses with tiger data from TGB I identified both migrant individuals (n=17) and signatures of high gene flow among majority of the TRs and other surrounding areas (Fig. 2a, Supplementary Table S4), indicating functional corridors within this region. We feel that the complex, undulating forested Shivalik and Bhabhar habitat found in this region has helped in tiger movements as hypothesized by Johnsingh *et al.* (2004)²⁹. Similar pattern was

also identified in Western Ghats and central Indian landscapes where tigers were reported to use rough terrain for dispersal and avoided human habitations^{21,65}. In addition, this area also retains the largest number of tigers, mostly within the PAs¹³ and recent increase in tiger population has probably resulted in migration of the surplus individuals to newer areas. Our source-sink dynamics analyses results also corroborate this pattern, where we found that majority of the sink populations in TGB I are in the flat, Terai region in the southern part (Fig. 2a). The Terai habitat also supports very high human density and known to support poor tiger populations in recent past²⁹. Similarly, Terai habitat dominated TGB II (where majority of the tiger populations are found within PAs) showed good connectivity (Fig. 2b, Supplementary Table S4). Our analyses indicated that mostly the PAs are source population and FD/SFDs are the sink populations (Fig. 2b), showing a contrasting pattern from TGB I where even FDs were source populations (like Lansdowne and Ramnagar FDs) (Fig. 2a). In TGB III, the largest tiger population Valmiki TR was the source whereas the western Sohagibarwa WLS was the only available sink habitat (Fig. 2c).

The Circuitscape analyses provide strong support and possible explanations to the genetic connectivity patterns we observed in this landscape. Overall, we identified a total of 10 high, three medium and six low conductance tiger corridors across TAL (Fig. 3a, Supplementary Table S6), all of them are outside PAs (Fig. 3a). First, the genetic discontinuity between TGB I and II is possibly due to loss of functionality of the Kilpura-Khatima-Surai corridor (marked as corridor J in Fig. 1a)²⁹. This corridor structurally connects the Champawat-Haldwani-Terai East FD complex of TGB I with Pilibhit-Dudhwa TR complex of TGB II but currently dysfunctional (Fig. 1a). Earlier Jonsingh *et al.* (2004)²⁹ reported no tiger use in this corridor, which has been further confirmed by Jhala *et al.* (2020)¹³. We believe that this loss of connectivity between TGB I and II is possibly resulting from developmental activities (e.g. highway, railway tracks, canal etc.) and habitat loss (from human settlements and

urbanization etc.) along the corridor (Fig. 3a). Some recent evidences suggest tiger dispersal events in this corridor ⁶⁶ indicating hope for this corridor but strong management interventions are required to maintain its functionality and ensure one panmictic population between Rajaji to Dudhwa TRs (Fig. 3a). Next, within the TGB I the southern part of Gola river corridor (marked as corridor I in Fig. 1a) connecting Ramnagar-Terai Central FDs complex with Champawat-Haldwani-Terai West FDs complex was identified as a weak corridor (Fig. 3a). While the northern part of this corridor showed medium conductance, the southern part is heavily affected by anthropogenic activities like human settlements, urbanization, industrialization and large-scale boulder mining ^{29,67} leading to low genetic exchange across it (Fig. 2a). Further, we identified that the Chilla-Motichur corridor (marked as corridor C in Fig. 1a) that connects eastern and western parts of Rajaji TR as a weak connecting link (Fig. 3a) ^{13,29}. Despite showing high conductance in Circuitscape analysis there is no functional connectivity through this corridor, possibly due to intense anthropogenic activities ^{29,68,69}. Further, one of the most important output of this study is the identification of 10 critical bottlenecks distributed across the high, medium and low conductivity corridors (Fig. 3a). Due to the linear geography of the TAL where only one-directional movements are feasible, these corridor bottlenecks demand immediate conservation attentions. Except these identified regions we could not find any other alternative paths (high Himalayan area in north and human-dominated Terai flats in south) and thus maintaining connectivity through these identified corridors are the only way these tiger populations can survive in near future ⁶⁷.

Finally, another important aspect of the results from this study is that despite having a large tiger number and relatively connected populations the TAL tigers have relatively low genetic variation compared to other tiger landscapes within India and Nepal-TAL. Different measures of genetic diversity in TAL showed lower values when compared with central India (N_A-

12.43 ± 2.99 ⁶¹, 11.71 ²⁴, 9.1 ± 2.2 ²⁵; H_O - 0.65 ± 0.09 ⁶¹, 0.54 ²⁴, 0.70 ± 0.06 ²⁵ and ASR -
 28.8 ± 10.5 ²⁵) and Western Ghats (N_A - $3-12$ ³⁴ and H_O - $0.33-0.68$ ³⁴) as well as with earlier
studies in TAL (N_A - 6.69 and H_O - 0.50 ⁶²) and Nepal-TAL (N_A - $3.0 \pm 0.33-4.0 \pm 0.32$ and
 H_O - $0.46 \pm 0.08-0.58 \pm 0.06$ ²⁸). While such patterns can be possible due to different panel of
markers, the same markers have shown higher genetic variation in central India²⁴ and
Western Ghats^{34,70}. Interestingly, similar pattern of lower genetic variation (compared to
other regions) has also been reported in leopards from TAL, which was attributed to severe
population decline in recent years⁶³. TAL has a long history of extensive trophy and bounty
hunting of tigers and leopards since Mughal and British era⁷¹, possibly leading to a small
founder population. Though the current population has increased in recent years¹³ the small
founder population is probably the reason behind lower genetic variations. However, TGB I
showed higher genetic variations ($N_{A(TGBI)}$ - 10.15 ± 3.06 , $H_{O(TGBI)}$ - 0.34 ± 0.2 and $ASR_{(TGBI)}$ -
 24.46 ± 7.37) than TGB II (Table 3), possibly because it hosts the largest tiger population
(~500 individuals) in this landscape¹³. Surprisingly, the small tiger population in TGB III
showed relatively higher genetic variation than in TGB II (Table 3). This is possibly due to
gene flow from larger tiger populations of Chitwan NP and Parsa Wildlife Reserve of Nepal
²⁷, which helped to retain high genetic diversity in the TGB III. The genetic variation of TGB
II can possibly be improved by ensuring tiger connectivity through Kilpura-Khatima-Surai
corridor from TGB I along with Shuklaphanta NP and Bardia NP of Nepal²⁷.

Conservation implications

Considering the linear shape of tiger habitats in TAL, fragmentation has always been a major
concern for conservation^{29,72}. Maintaining the integrity of this landscape with very high
human density and associated linear developments will remain the most important challenge
for long-term persistence of tiger in this globally high priority tiger conservation landscape²⁹.
Our results on tiger population structure, connectivity and source-sink dynamics thus have

critical management implications specially in the backdrop of recent increase in tiger numbers in TAL. One of the encouraging point is the revealing of only three TGBs compared to the eight distinct THBs described by Johnsingh *et al.* (2004)²⁹, indicating considerable amount genetic exchange within these THBs. However, within the TGBs we identified 10 critical corridors (Fig. 3a, Supplementary Table S6) that require urgent attention failing which integrity of TAL will be severely compromised. Further analyses show that a total of 2707 sq. km. habitat (Fig. 3b, 3c, 3d, Table 4) needs appropriate conservation planning to ensure panmictic tiger populations in TAL. Appropriate mitigation measures associated with ongoing linear infrastructure developments, roads in particular, are a must in this highly sensitive conservation landscape⁶⁹. In addition, TGB II and III requires transboundary cooperation between India and Nepal to ensure maintenance of genetic variation and source-sink dynamics in this population.

We also anticipate another potentially important upcoming tiger conservation challenge in the form of increasing tiger-human interactions. Our results show that significant proportion of tigers in TAL are found outside PAs (Supplementary Fig. S2) (possibly driven by recent increase in tiger population), co-existing with significant amount of human density and associated livestock¹³. It is likely that the incidences of tiger-human conflict will increase around the sink habitats and corridors in coming years and active management of such conflict situations will be critical for these tigers living in marginalized habitats. Further, adequate attention towards habitat and prey management in the FDs and SFDs where large number of tigers are found (at least in TGB I and II) will play important role in maintaining connectivity of the entire landscape.

India played a leading role in tiger conservation and has achieved a rare global success in population recovery of a large-bodied, apex carnivore¹³. However, the future of these tiger populations depend on appropriate management of the ever-shrinking habitats and

maintaining the existing populations as metapopulations. Our results from TAL showed the functionality of the existing corridors and point out critical areas where immediate conservation attention is needed. We believe that a focused approach to address such concerns will improve the long-term sustainability of the tiger populations in TAL.

Acknowledgement

We acknowledge the Forest Departments of Uttarakhand, Uttar Pradesh and Bihar for providing necessary permits to carry out the research. Our thanks to the Forest Department officials and frontline staffs for their support and assistance during field surveys. We acknowledge help from Annu, Bura, Abbhi, Ranjhu, Ammi, Inam, Imam, Shrutarshi, Shiv, Tista, Shrushti, Prajak, Harshvardhan, Lakhshminarayan, Ankit, Rakesh, Sultan and Nimisha for their help during field surveys. We appreciate technical help from Tista, Zenab, Aamir and Madhanraj (laboratory work) and other lab members. We thank the Director, Dean and Nodal Officer of Wildlife Forensics and Conservation Genetics Cell of Wildlife Institute of India for their support.

This research was funded by Wildlife Conservation Trust-Panthera Global Cat Alliance Grants and Department of Science and Technology, Government of India (grant no EMR/2014/000982). Samrat Mondol was supported by the Department of Science and Technology INSPIRE Faculty Award (No.IFA12-LSBM-47). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Author contribution

S Mondol and B Pandav conceived the study idea, generated funds and supervised the study. S Biswas and S Bhatt performed the sampling and data generation. Data was analysed by S Biswas (molecular and GIS data), D Sarkar (GIS), G Talukdar (GIS) and S Mondol

(molecular data). S Mondol, S Biswas and B Pandav wrote the initial manuscript and all authors approved the final draft.

Competing interest

The authors declare no competing interests.

Reference

1. Hemmer, H. *The phylogeny of the tiger (Panthera tigris)*. (Tigers of the World, 1987).
2. Sanderson, E. W. *et al. Setting Priorities for Tiger Conservation: 2005-2015. Tigers of the World* (2006).
3. Dinerstein, E. *et al.* The Fate of Wild Tigers. *Bioscience* **57**, 508–514 (2007).
4. Walston, J. *et al.* Bringing the tiger back from the brink-the six percent solution. *PLoS Biol.* **8**, e1000485 (2010).
5. Initiative, G. T. Global tiger recovery program 2010–2022. (2011).
6. Ranganathan, J., Chan, K. M. A., Karanth, K. U. & Smith, J. L. D. Where can tigers persist in the future? A landscape-scale, density-based population model for the Indian subcontinent. *Biol. Conserv.* **141**, 67–77 (2008).
7. Karanth, K. U. & Sunquist, M. E. Prey Selection by Tiger, Leopard and Dhole in Tropical Forests. *J. Anim. Ecol.* **64**, 439 (1995).
8. Wikramanayake, E. *et al.* A landscape-based conservation strategy to double the wild tiger population. *Conserv. Lett.* **4**, 219–227 (2011).
9. Joshi, A. R. *et al.* Tracking changes and preventing loss in critical tiger habitat. *Sci. Adv.* **2**, e1501675 (2016).
10. Gubbi, S., Mukherjee, K., Swaminath, M. H. & Poornesha, H. C. Providing more protected space for tigers *Panthera tigris*: A landscape conservation approach in the Western Ghats, southern India. *Oryx* **50**, 336–343 (2016).
11. Gubbi, S. *et al.* From intent to action: A case study for the expansion of tiger

- conservation from southern India. *Glob. Ecol. Conserv.* **9**, 11–20 (2017).
12. Goodrich, J. *et al.* *Panthera tigris*. The IUCN red list of threatened species 2015: e.T15955A50659951. (2015).
 13. Jhala, Y. V., Qureshi, Q. & Nayak, A. K. *Status of tigers, copredators and prey in India, 2018*. (2020).
 14. Mondal, P. & Nagendra, H. Trends of forest dynamics in tiger landscapes across Asia. *Environ. Manage.* **48**, 781–794 (2011).
 15. Gubbi, S., Poornesha, H. C. & Madhusudan, M. D. Impact of vehicular traffic on the use of highway edges by large mammals in a South Indian wildlife reserve. *Curr. Sci.* **102**, 1047–1051 (2012).
 16. Brook, B. W., Tonkyn, D. W., O’Grady, J. J. & Frankham, R. Contribution of inbreeding to extinction risk in threatened species. *Conserv. Ecol.* **6**, (2002).
 17. Sharma, S. *et al.* Forest corridors maintain historical gene flow in a tiger metapopulation in the highlands of central India. *Proc. R. Soc. B Biol. Sci.* **280**, 14 (2013).
 18. Norén, K. *et al.* Evolution, ecology and conservation—revisiting three decades of Arctic fox population genetic research. *Polar Res.* **36**, (2017).
 19. India, W. I. of. Wildlife Institute of India National Wildlife Database (2020) Dehradun, India. (2020).
 20. Wikramanayake, E. *et al.* Designing a conservation landscape for tigers in human-dominated environments. *Conserv. Biol.* **18**, 839–844 (2004).
 21. Reddy, P. A., Puyravaud, J. P., Cushman, S. A. & Segu, H. Spatial variation in the response of tiger gene flow to landscape features and limiting factors. *Anim. Conserv.* **22**, 472–480 (2019).
 22. Wikramanayake, E. *et al.* A landscape-based conservation strategy to double the wild

- 648 tiger population. *Conserv. Lett.* **4**, 219–227 (2011).
- 649 23. Rathore, C. S., Dubey, Y., Shrivastava, A., Pathak, P. & Patil, V. Opportunities of
650 habitat connectivity for tiger (*Panthera tigris*) between Kanha and Pench national parks
651 in Madhya Pradesh, India. *PLoS One* **7**, (2012).
- 652 24. Joshi, A., Vaidyanathan, S., Mondol, S., Edgaonkar, A. & Ramakrishnan, U.
653 Connectivity of Tiger (*Panthera tigris*) Populations in the Human-Influenced Forest
654 Mosaic of Central India. *PLoS One* **8**, e77980 (2013).
- 655 25. Yumnam, B. *et al.* Prioritizing Tiger Conservation through Landscape Genetics and
656 Habitat Linkages. *PLoS One* **9**, e111207 (2014).
- 657 26. Thatte, P., Joshi, A., Vaidyanathan, S., Landguth, E. & Ramakrishnan, U. Maintaining
658 tiger connectivity and minimizing extinction into the next century: Insights from
659 landscape genetics and spatially-explicit simulations. *Biol. Conserv.* **218**, 181–191
660 (2018).
- 661 27. Thapa, K. *et al.* Tigers in the Terai: Strong evidence for meta-population dynamics
662 contributing to tiger recovery and conservation in the Terai Arc Landscape. *PLoS One*
663 **12**, e0177548 (2017).
- 664 28. Thapa, K. *et al.* Assessment of genetic diversity, population structure, and gene flow of
665 tigers (*Panthera tigris tigris*) across Nepal's Terai Arc Landscape. *PLoS One* **13**, 1–25
666 (2018).
- 667 29. Johnsingh, A. J. T. *et al.* *Conservation Status of Tiger and Associated Species in the*
668 *Terai Arc Landscape , India.* (2004).
- 669 30. Champion, H. G. & Seth, S. K. *A revised survey of the forest types of India.* (1968).
- 670 31. Jhala, Y. V., Qureshi, Q. & Gopal, R. *The status of tigers in india, 2014.* (2015).
- 671 32. Biswas, S. *et al.* A Practive Faeces Collection Protocol for Multidisciplinary Research
672 in Wildlife Science. *Curr. Sci.* **116**, 1878–1885 (2019).

- 673 33. Mukherjee, N., Mondol, S., Andheria, A. & Ramakrishnan, U. Rapid multiplex PCR
674 based species identification of wild tigers using non-invasive samples. *Conserv. Genet.*
675 **8**, 1465–1470 (2007).
- 676 34. Mondol, S. *et al.* Identifying species, sex and individual tigers and leopards in the
677 Malenad-Mysore Tiger Landscape, Western Ghats, India. *Conserv. Genet. Resour.* **7**,
678 353–361 (2015).
- 679 35. Mondol, S. *et al.* Evaluation of non-invasive genetic sampling methods for estimating
680 tiger population size. *Biol. Conserv.* **142**, 2350–2360 (2009).
- 681 36. Mondol, S., Thatte, P., Yadav, P. & Ramakrishnan, U. A set of miniSTRs for
682 population genetic analyses of tigers (*Panthera tigris*) with cross-species amplification
683 for seven other Felidae. *Conserv. Genet. Resour.* **4**, 63–66 (2012).
- 684 37. Miquel, C. *et al.* Quality indexes to assess the reliability of genotypes in studies using
685 noninvasive sampling and multiple-tube approach. *Mol. Ecol. Notes* **6**, 985–988
686 (2006).
- 687 38. Pilgrim, K. L., Mckelvey, K. S., Riddle, A. E. & Schwartz, M. K. Felid sex
688 identification based on noninvasive genetic samples. *Mol. Ecol. Notes* **5**, 60–61 (2005).
- 689 39. DeCandia, A., Gaughran, S., Caragiulo, A. & Amato, G. A novel molecular method
690 for noninvasive sex identification of order Carnivora. *Conserv. Genet. Resour.* 1–3
691 (2016). doi:10.1007/s12686-016-0525-z
- 692 40. Broquet, T. & Petit, E. Quantifying genotyping errors in noninvasive population
693 genetics. *Mol. Ecol.* **13**, 3601–3608 (2004).
- 694 41. Van Oosterhout, C., Hutchinson, W. F., Wills, D. P. M. & Shipley, P. MICRO-
695 CHECKER: software for identifying and correcting genotyping errors in microsatellite
696 data. *Mol. Ecol. Notes* **4**, 535–538 (2004).
- 697 42. Kalinowski, S. T., Taper, M. L. & Marshall, T. C. Revising how the computer program

- cervus accommodates genotyping error increases success in paternity assignment. *Mol. Ecol.* **16**, 1099–1106 (2007).
43. Valiere, N. Gimlet: a computer program for analysing genetic individual identification data. *Mol. Ecol. Notes* **2**, 377–379 (2002).
44. Excoffier, L., Laval, G. & Schneider, S. *Arlequin ver 3.1: An Integrated Software Package for Population Genetics Data Analysis. Manual* **9**, (2005).
45. Janečka, J. E. *et al.* Evidence for male-biased dispersal in bobcats *Lynx rufus* using relatedness analysis. *Wildlife Biol.* **13**, 38–47 (2007).
46. Chen, C., Durand, E., Forbes, F. & Francois, O. Bayesian clustering algorithms ascertaining spatial population structure: a new computer program and a comparison study. *Mol. Ecol. Notes* **7**, 747–756 (2007).
47. Vergara, M. *et al.* Inferring Population Genetic Structure in Widely and Continuously Distributed Carnivores: The Stone Marten (*Martes foina*) as a Case Study. *PLoS One* **10**, e0134257 (2015).
48. Peakall, R. & Smouse, P. E. GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research--an update. *Bioinformatics* **28**, 2537–2539 (2012).
49. Pritchard, J. K., Stephens, M. & Donnelly Peter. Inference of population structure using multilocus genotype data. *Genetics* **155**, 945–959 (2000).
50. Piry, S. *et al.* GENECLASS2: A software for genetic assignment and first-generation migrant detection. *J. Hered.* **95**, 536–539 (2004).
51. Rannala, B. & Mountain, J. L. Detecting immigration by using multilocus genotypes. *Proc. Natl. Acad. Sci. U. S. A.* **94**, 9197–9201 (1997).
52. Paetkau, D., Robert, S., Michael, B. & Arnaud, E. Genetic assignment methods for the direct, real-time estimation of migration rate: a simulation-based exploration of

- 723 accuracy and power. *Mol. Ecol.* **13**, 55–65 (2004).
- 724 53. Wilson, G. A. & Rannala, B. Bayesian inference of recent migration rates using
725 multilocus genotypes. *Genetics* **163**, 1177–1191 (2003).
- 726 54. Smith, J. L. D. & Mcdougal, C. The Contribution of Variance in Lifetime
727 Reproduction to Effective Population Size in Tigers. *Conserv. Biol.* **5**, 484–490 (1991).
- 728 55. Philips, S. J. and & Dudík, M. Modeling of species distributions with Maxent: new
729 extensions and a comprehensive evaluation. *Ecography (Cop.)*. **31**, 161–175 (2008).
- 730 56. McRae, B. H., Shah, V. B. & Edelman, A. *Circuitscape: Modeling Landscape*
731 *Connectivity to Promote Conservation and Human Health.* (2016).
732 doi:10.13140/RG.2.1.4265.1126
- 733 57. Brown, J. L. SDMtoolbox: a python-based GIS toolkit for landscape genetic,
734 biogeographic and species distribution model analyses. *Methods Ecol. Evol.* **5**, 694–
735 700 (2014).
- 736 58. Smith, J. L. D. The Role of Dispersal in Structuring the Chitwan Tiger Population.
737 *Behaviour* **124**, 165–195 (1993).
- 738 59. Reddy, P. A. *et al.* Genetic evidence of tiger population structure and migration within
739 an isolated and fragmented landscape in northwest India. *PLoS One* **7**, (2012).
- 740 60. Kolipakam, V., Singh, S., Pant, B., Qureshi, Q. & Jhala, Y. V. Genetic structure of
741 tigers (*Panthera tigris tigris*) in India and its implications for conservation. *Glob. Ecol.*
742 *Conserv.* **20**, e00710 (2019).
- 743 61. Sharma, S. *et al.* Spatial genetic analysis reveals high connectivity of tiger (*Panthera*
744 *tigris*) populations in the Satpura-Maikal landscape of Central India. *Ecol. Evol.* **3**,
745 48–60 (2013).
- 746 62. Singh, S. K. *et al.* Fine-scale population genetic structure of the Bengal tiger (*Panthera*
747 *tigris tigris*) in a human-dominated western Terai Arc Landscape, India. *PLoS One* **12**,

- 748 e0174371 (2017).
- 749 63. Bhatt, S., Biswas, S., Karanth, K., Pandav, B. & Mondol, S. Genetic analyses reveal
750 population structure and recent decline in leopards (*Panthera pardus fusca*) across the
751 Indian subcontinent. *PeerJ* **2020**, 1–27 (2020).
- 752 64. Athreya, V., Odden, M., Linnell, J. D. C. & Karanth, K. U. Translocation as a Tool for
753 Mitigating Conflict with Leopards in Human-Dominated Landscapes of India.
754 *Conserv. Biol.* **25**, 133–141 (2010).
- 755 65. Reddy, P. A., Cushman, S. A., Srivastava, A., Sarkar, M. S. & Shivaji, S. Tiger
756 abundance and gene flow in Central India are driven by disparate combinations of
757 topography and land cover. *Divers. Distrib.* **23**, 863–874 (2017).
- 758 66. Anwar, M. & Borah, J. Functional status of a wildlife corridor with reference to tiger
759 in Terai Arc Landscape of India. *Trop. Ecol.* **60**, 525–531 (2020).
- 760 67. Qureshi, Q. *et al.* *Connecting tiger populations for long-term conservation. National*
761 *Tiger Conservation Authority* (2014).
- 762 68. Harihar, A. & Pandav, B. Influence of connectivity, wild prey and disturbance on
763 occupancy of tigers in the human-dominated western Terai Arc landscape. *PLoS One*
764 **7**, 1–10 (2012).
- 765 69. Harihar, A., Ghosh-Harihar, M. & Macmillan, D. C. Losing time for the tiger *Panthera*
766 *tigris*: Delayed action puts a globally threatened species at risk of local extinction.
767 *Oryx* **52**, 78–88 (2018).
- 768 70. Mondol, S. Phylogeography and Population Genetics of the Bengal Tiger (*Panthera*
769 *tigris tigris*) in the Indian Subcontinent. (2011).
- 770 71. Rangarajan, M. & Shahabuddin, G. Displacement and relocation from protected areas:
771 Towards a biological and historical synthesis. *Conserv. Soc.* **4**, 359–378 (2006).
- 772 72. Kanagaraj, R., Wiegand, T., Kramer-Schadt, S., Anwar, M. & Goyal, S. P. Assessing

habitat suitability for tiger in the fragmented Terai Arc Landscape of India and Nepal.
Ecography (Cop.). **34**, 970–981 (2011).

Figure legends

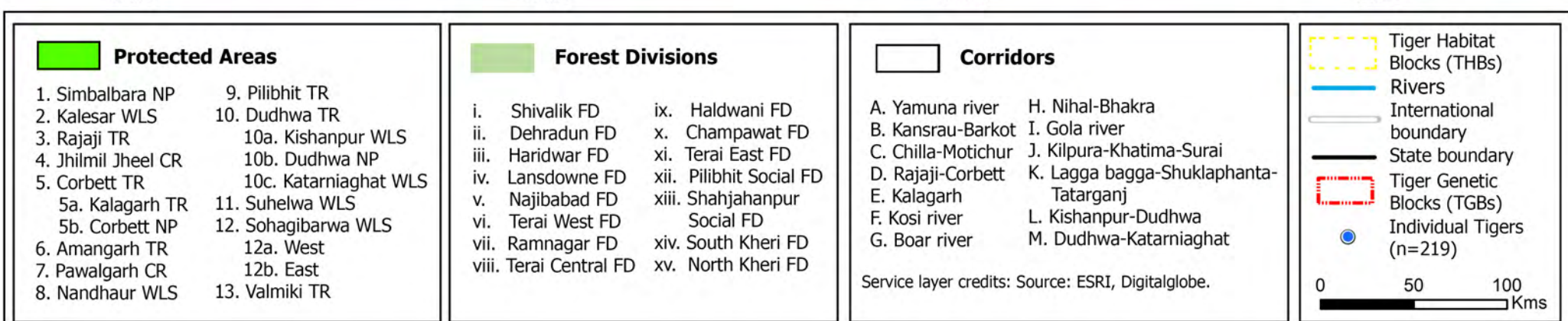
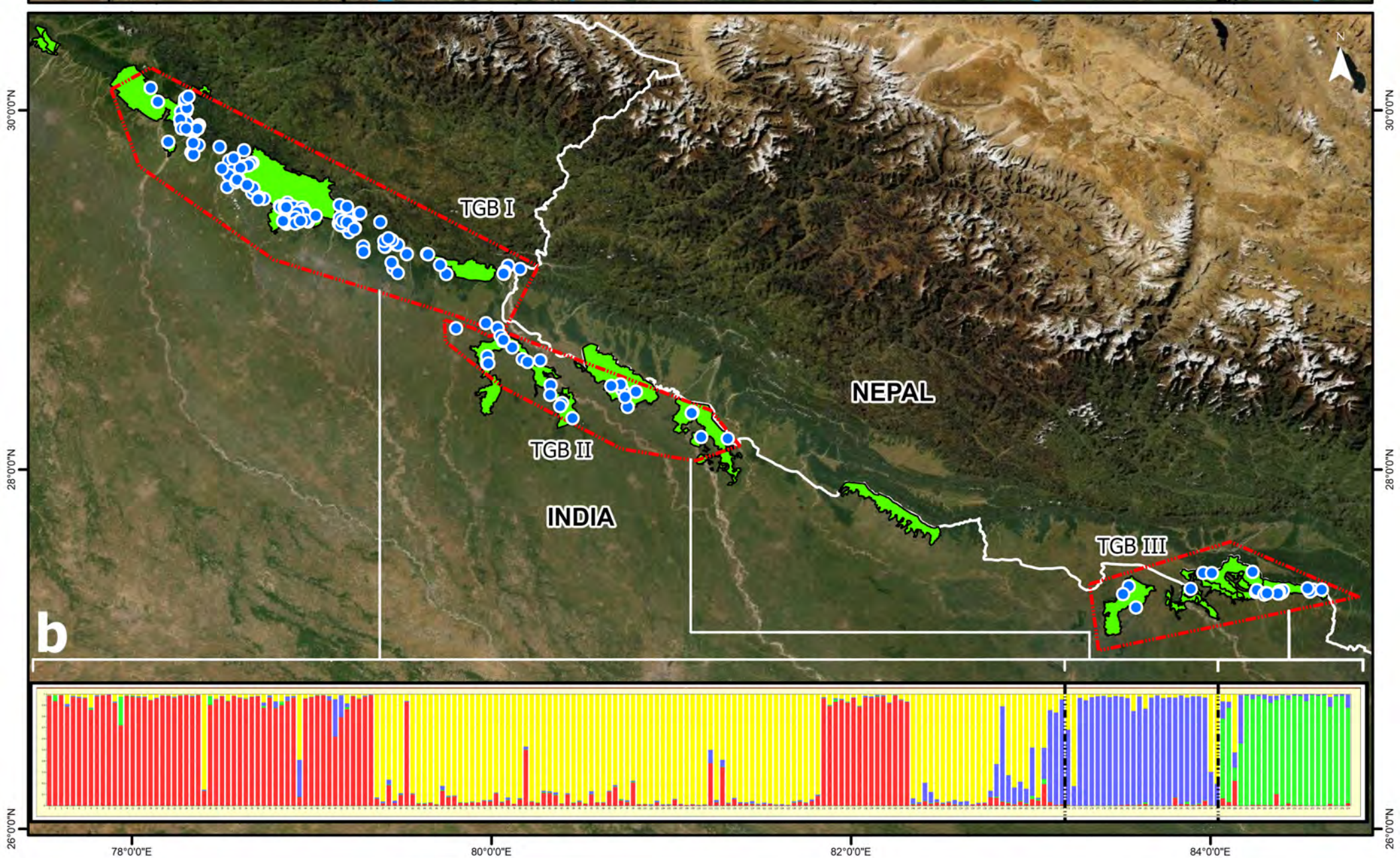
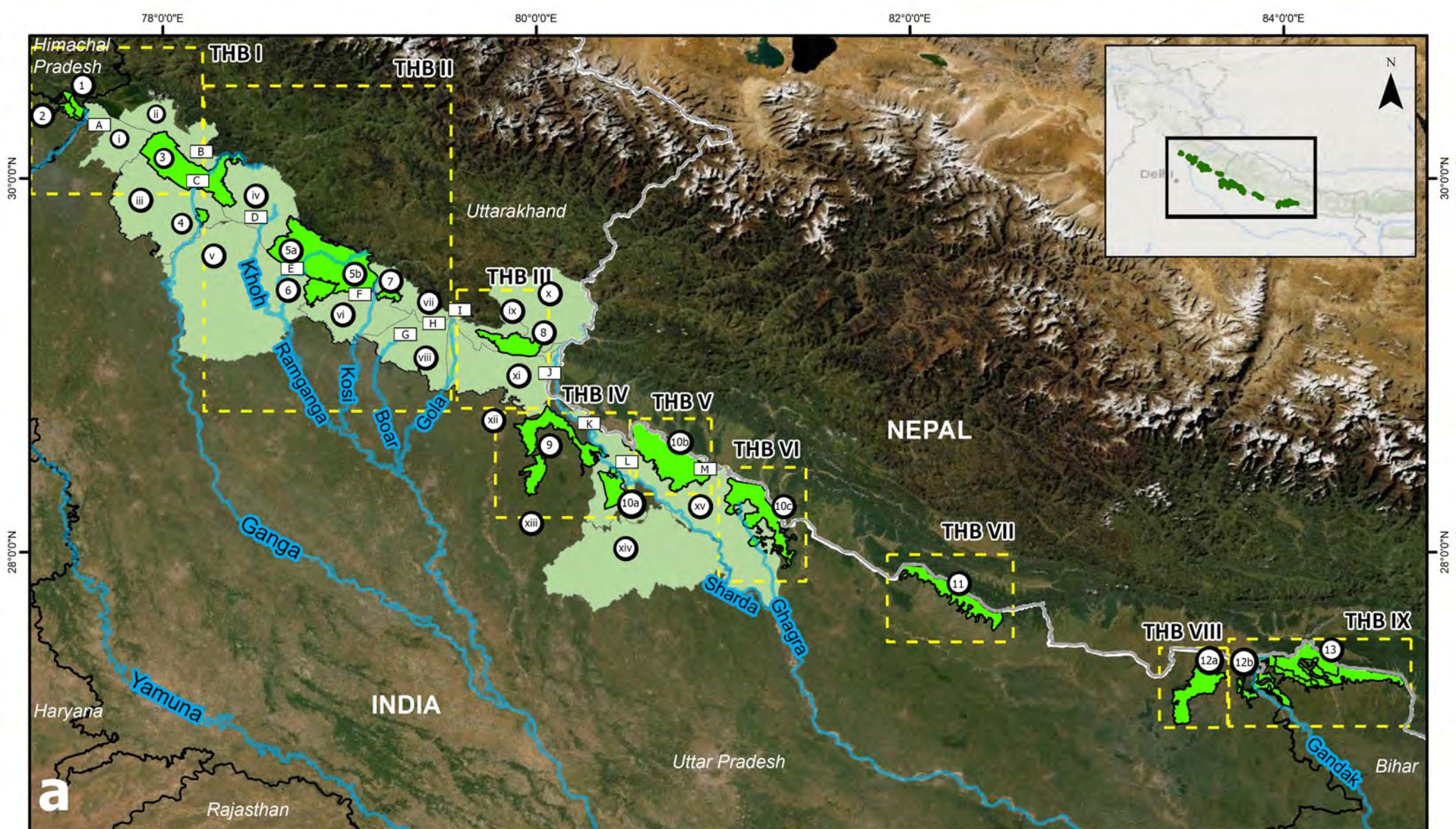
Figure 1: The tiger habitats within the Indian part of Terai-Arc landscape (TAL), encompassing both protected (National parks, Tiger Reserves, Conservation Reserves and Wildlife Sanctuaries) as well as non-protected areas (Forest Divisions and Social Forestry Divisions). The top plot (a) shows the entire landscape with marked ‘Tiger Habitat Blocks’ (THBs) and identified corridors (Johnsingh et al., 2004)²⁹. The bottom plot (b) presents the ‘Tiger Genetic Blocks’ (TGBs) along with the genetic structure results from program TESS⁴⁵. These TGBs roughly correspond to the western, central and eastern parts of the landscape.

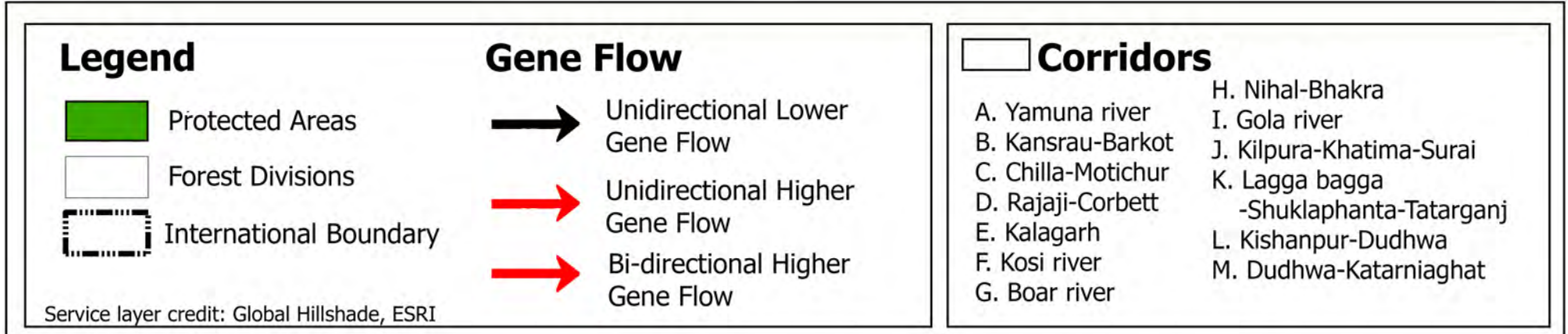
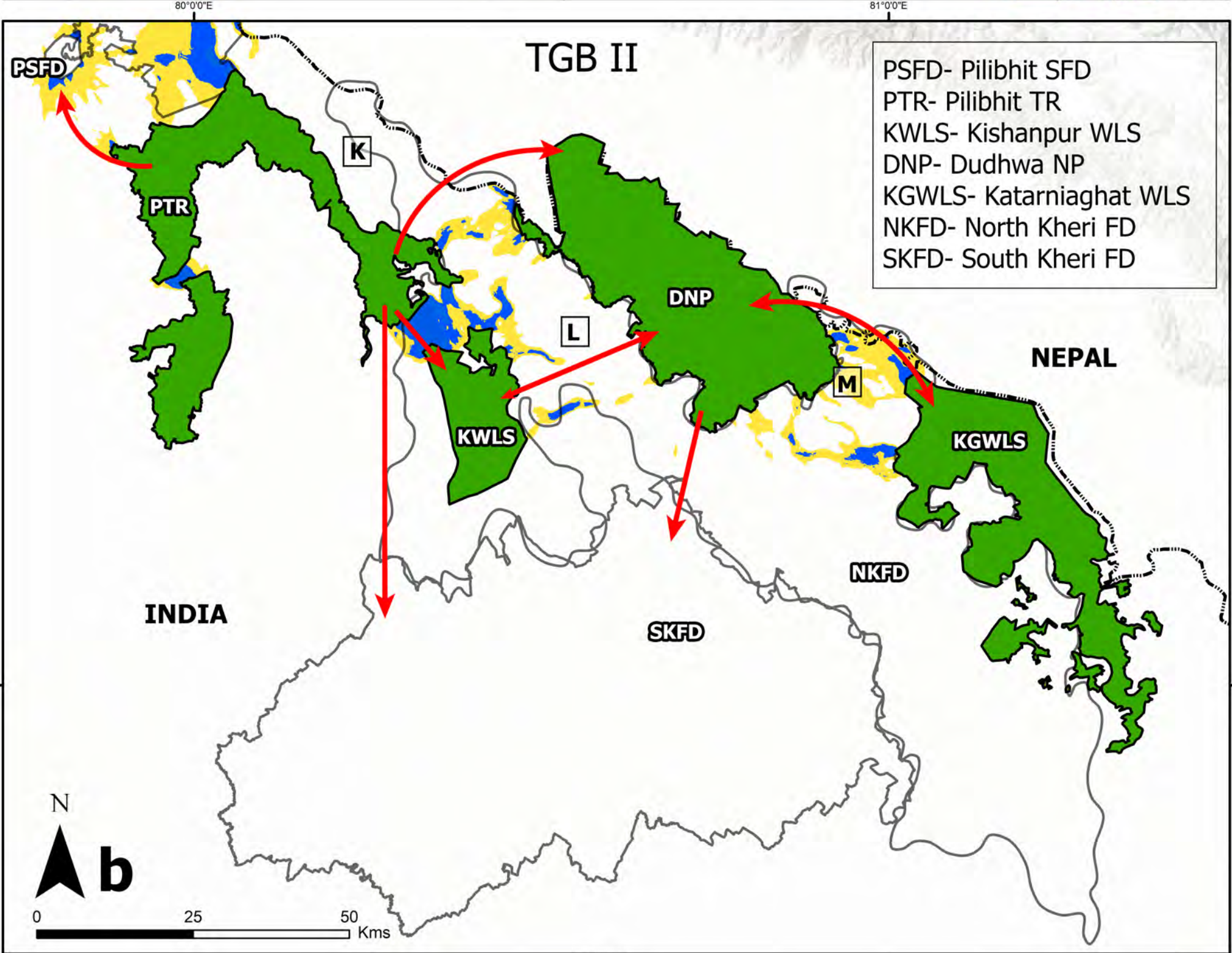
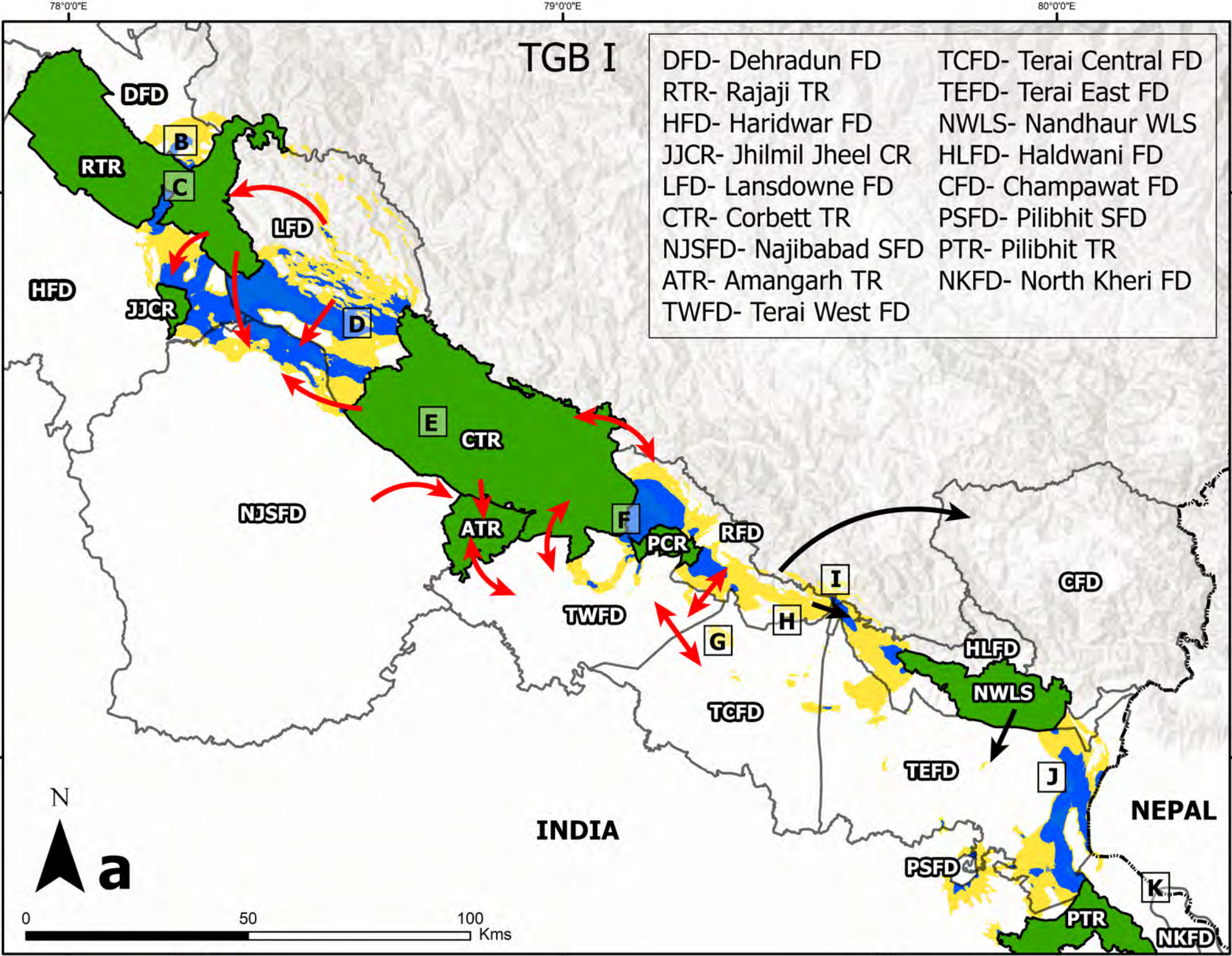
Figure 2: Assessment of tiger source-sink dynamics within each TGB in the Indian part of TAL. The direction and magnitude of gene flow has been presented by different color allows among the protected and non-protected areas. The top plot (a), middle plot (b) and the bottom plot (c) show the gene flow patterns in TGB I, II and III, respectively.

Figure 3: Results of the CIRCUITSCAPE analyses to identify the corridor conductances across TAL. Both ‘Least Cost Pathways (LCPs)’ as well as the critical corridors (pinchpoints) are shown here in (a). The critical corridor areas to maintain contiguous landscape and require urgent management attention are highlighted in (b), (c) and (d). Refer Table 4 for details of these critical areas.

Supplementary fig. 1: The locations of large carnivore faeces across TAL.

Supplementary fig. 2: The locations of genetically identified tiger faecal samples across TAL.





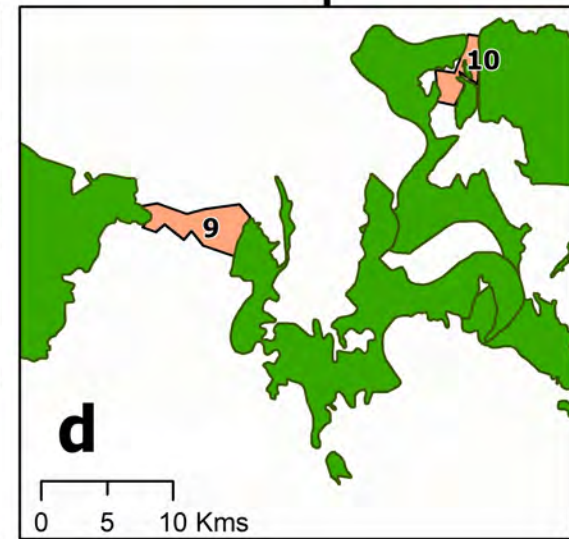
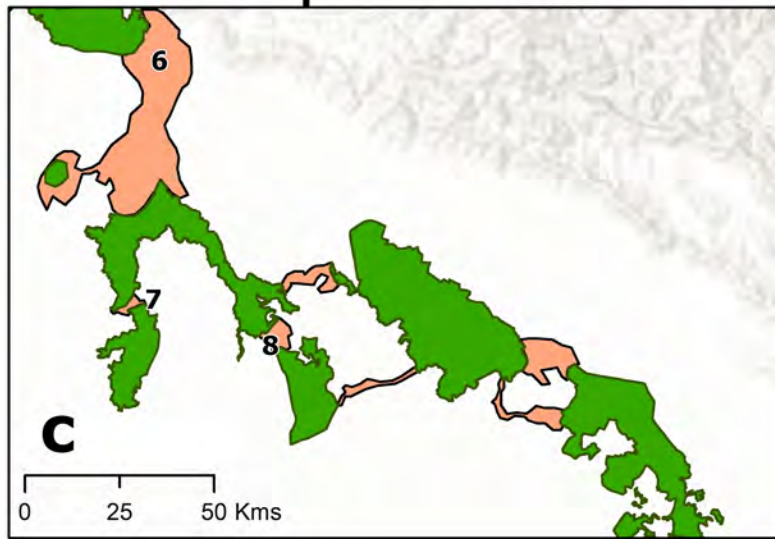
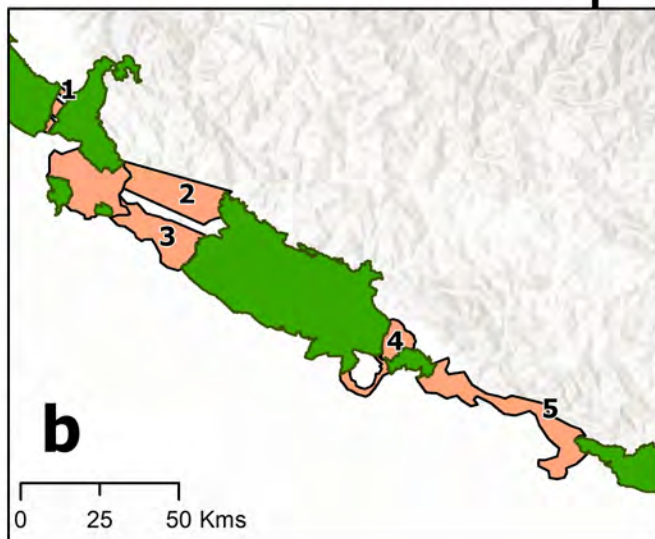
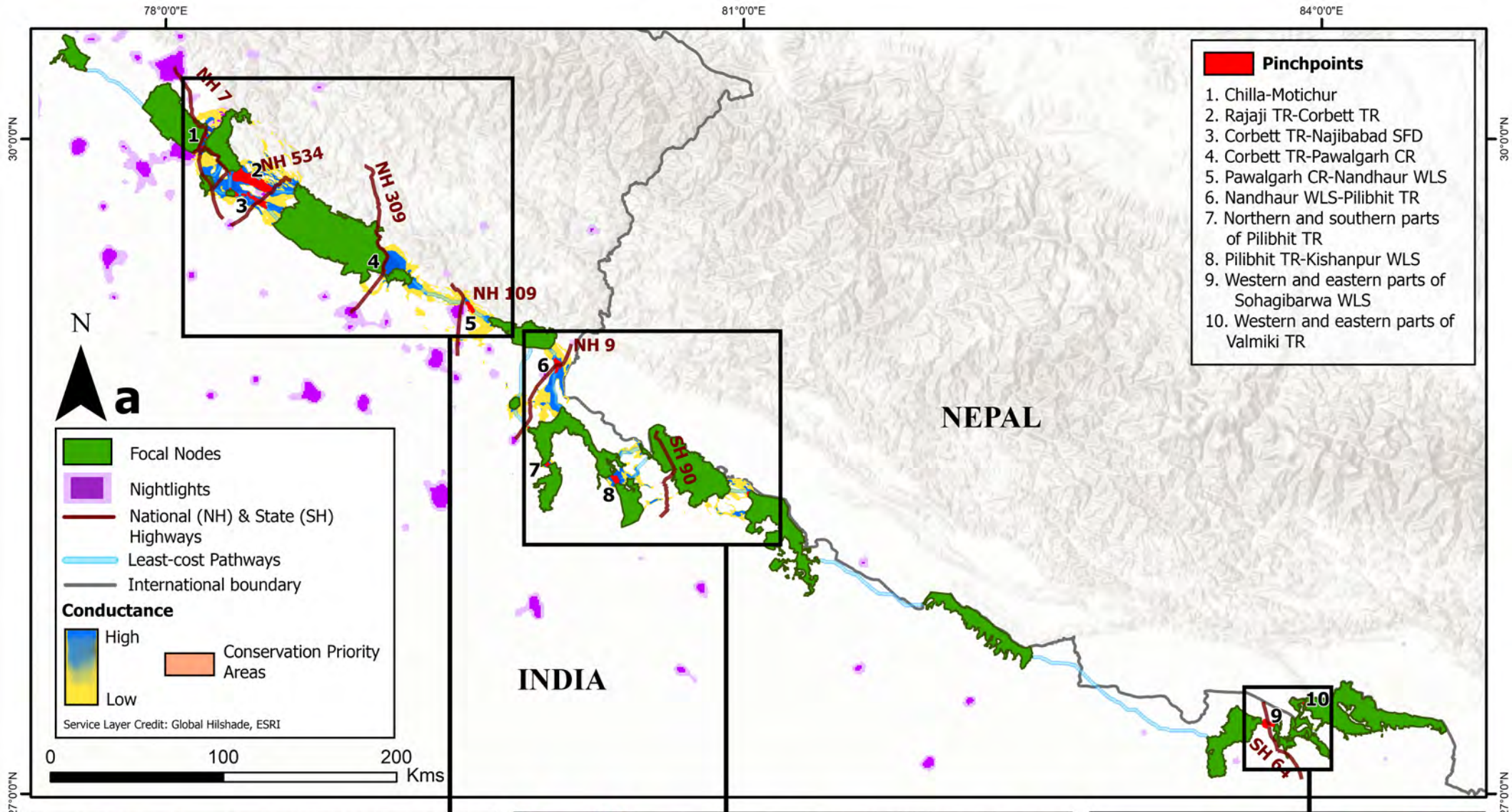


Table 1: Genetic diversity and genotyping error details for the individual tigers (n=219) from TAL.

Locus	Repeat length	N _A	ASR	H _E	H _O	Cumulative PID _(unbiased)	Cumulative PID _(sibs)	ADO	FA	Success rate (%)	Reference
FCA090	2	19	38	0.88	0.31	2.266e-02	3.142e-01	0.011	0.030	77	35
msFCA506	2	12	38	0.85	0.26	7.991e-04	1.045e-01	0.003	0.050	59	36
FCA672	2	15	42	0.87	0.58	2.036e-05	3.330e-02	0.008	0.037	94	35
FCA304	2	14	42	0.73	0.17	2.305e-06	1.373e-02	0.003	0.024	92	35
FCA628	2	20	54	0.83	0.41	1.007e-07	4.721e-03	0.012	0.026	53	35
FCA232	2	16	32	0.70	0.27	1.340e-08	2.042e-03	0.005	0.028	97	35
FCA230	2	13	52	0.82	0.12	7.273e-10	7.226e-04	0.010	0.026	94	35
FCA279	2	16	36	0.87	0.51	1.761e-11	2.298e-04	0.000	0.031	91	35
FCA441	4	14	60	0.77	0.54	1.301e-12	8.834e-05	0.014	0.030	75	35
FCA069	2	16	40	0.79	0.30	7.176e-14	3.248e-05	0.014	0.014	79	35
msFCA453	4	12	72	0.71	0.62	8.955e-15	1.374e-05	0.010	0.010	94	36
msF115	4	09	48	0.60	0.08	1.869e-15	6.860e-06	0.019	0.019	56	36
msHDZ170	2	16	36	0.86	0.33	5.999e-17	2.245e-06	0.045	0.046	83	36
Mean		14.76 (2.83)	45.38 (10.96)	0.79 (0.08)	0.35 (0.16)			0.012	0.028	80.3	

N_A- Number of alleles, ASR- Allelic size range, H_E- Expected heterozygosity, H_O- Observed heterozygosity, PID- Probability of identity, ADO- Allelic drop out, FA- False allele

Table 2: Genetic differentiation (pairwise F_{st} and G_{st}) for three TGBs across TAL. The upper diagonal presents pairwise G_{st} values and lower diagonal presents the pairwise F_{st} values

	TGB I	TGB II	TGB III
TGB I	0	0.078*	0.067*
TGB II	0.089*	0	0.087*
TGB III	0.079*	0.107*	0

* p-value= 0.001

Table 3: Comparison of different genetic diversity indices among three identified TGBs in TAL.

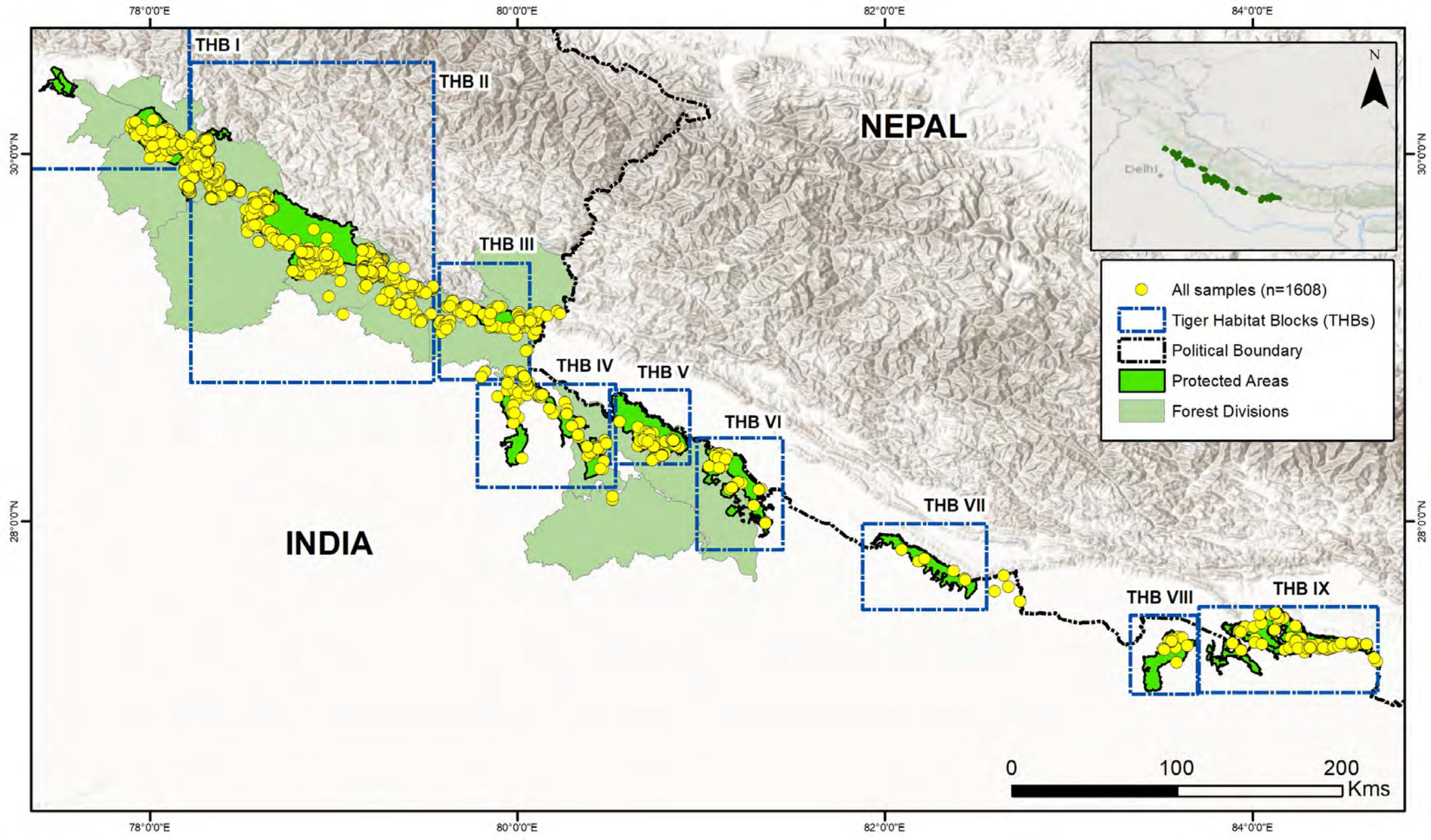
	TGB I (n=173)				TGB II (n=24)				TGB III (n=22)			
Locus	N _A	ASR	H _E	H _O	N _A	ASR	H _E	H _O	N _A	ASR	H _E	H _O
FCA090	12	28	0.85	0.31	08	18	0.70	0.29	09	28	0.78	0.37
msFCA506	11	24	0.83	0.21	08	16	0.72	0.42	08	34	0.84	0.38
FCA672	13	28	0.87	0.62	10	24	0.88	0.27	12	42	0.90	0.63
FCA304	11	28	0.73	0.14	05	20	0.62	0.30	05	10	0.73	0.32
FCA628	13	30	0.79	0.39	09	36	0.89	0.43	09	54	0.79	0.59
FCA232	11	22	0.68	0.20	07	20	0.53	0.43	11	32	0.81	0.71
FCA230	09	26	0.80	0.10	06	18	0.72	0.04	10	52	0.76	0.36
FCA279	13	24	0.86	0.53	09	22	0.59	0.20	11	32	0.77	0.73
FCA441	11	40	0.75	0.57	06	36	0.74	0.31	07	60	0.83	0.53
FCA069	03	26	0.71	0.33	10	28	0.89	0.30	09	32	0.84	0.15
msFCA453	04	08	0.65	0.70	05	20	0.69	0.38	10	60	0.82	0.33
msF115	10	16	0.44	0.03	05	48	0.74	0.35	05	24	0.67	0.06
msHDZ170	05	18	0.85	0.35	03	16	0.24	0.08	12	34	0.90	0.36
Mean	10.15 (3.06)	24.46 (7.37)	0.75 (0.11)	0.34 (0.20)	07.00 (2.11)	24.77 (9.30)	0.69 (0.17)	0.29 (0.12)	9.08 (2.24)	38.00 (14.29)	0.80 (0.06)	0.43 (0.20)

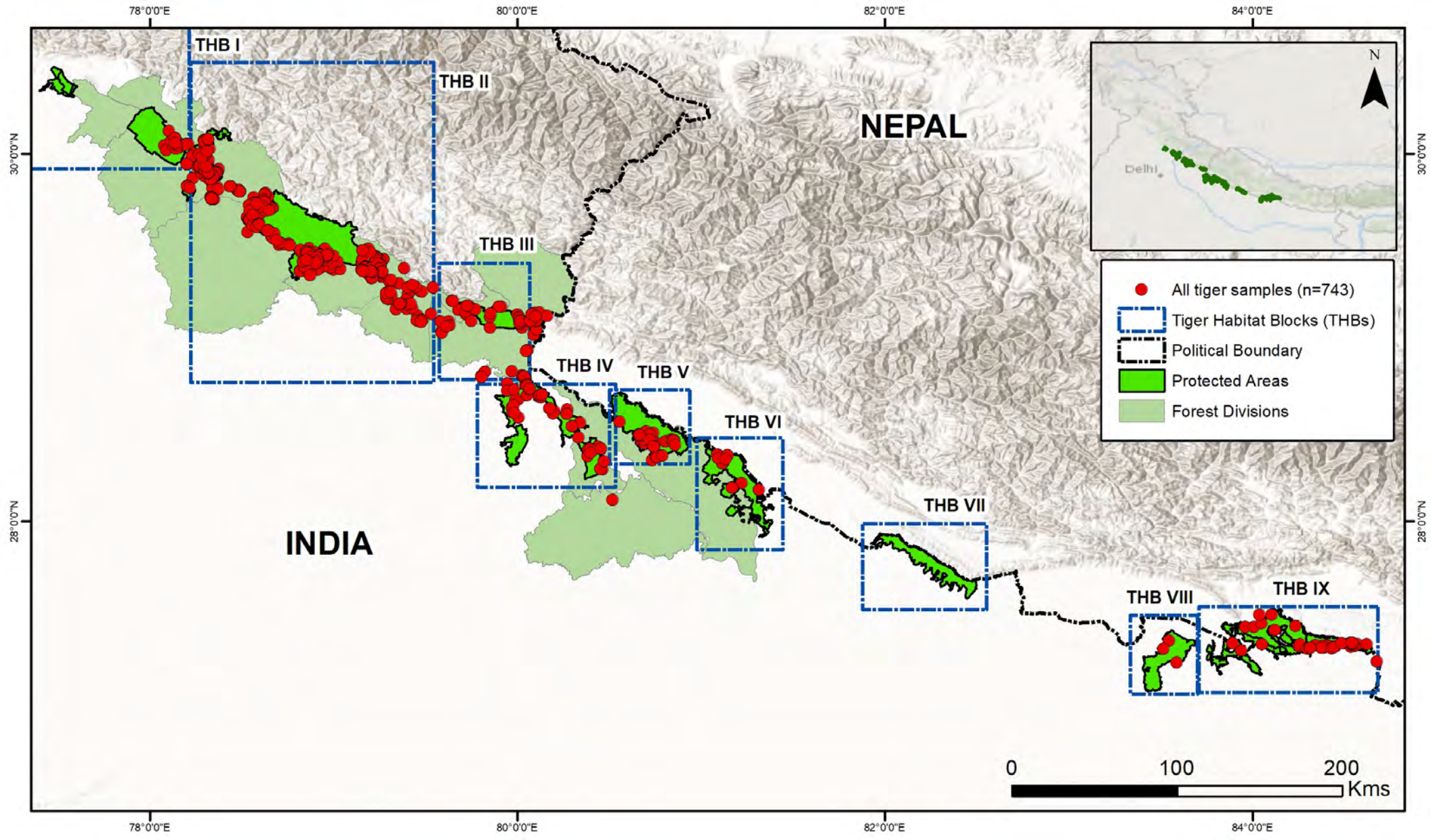
N_A- Number of alleles, ASR- Allelic size range, H_E- Expected heterozygosity, H_O- Observed heterozygosity

Table 4: Details of the critical corridor regions identified in this study.

SL	Geographic location	Area (sq.km.)	State
1	Connecting western and eastern parts of Rajaji TR (northern block)	9.73	Uttarakhand
2	Connecting western and eastern parts of Rajaji TR (central block)	14.12	Uttarakhand
3	Connecting western and eastern parts of Rajaji TR (southern block)	9.74	Uttarakhand
4	Connecting eastern Rajaji TR, Jhilmil Jheel CR and Najibabad SFD through Haridwar and Lansdowne FDs	345.07	Uttarakhand and Uttar Pradesh
5	Connecting Corbett TR and Najibabad SFD through Lansdowne FD and Najibabad SFD	267.16	Uttarakhand and Uttar Pradesh
6	Connecting Rajaji and Corbett TRs through Lansdowne FD	328.32	Uttarakhand
7	Connecting Corbett TR and Pawalgarh CR through Terai West FD	53.54	Uttarakhand
8	Connecting Corbett TR and Pawalgarh CR through Ramnagar FD	84.66	Uttarakhand
9	Connecting Pawalgarh CR and Nandhaur WLS through Ramnagar, Haldwani and Terai East FDs	405.37	Uttarakhand
10	Connecting Nandhaur WLS and Pilibhit TR through Haldwani and Terai East FDs and Pilibhit SFD	828.77	Uttarakhand and Uttar Pradesh
11	Connecting norther and southern parts of Pilibhit TR	17.4	Uttar Pradesh
12	Connecting Pilibhit TR and Dudhwa NP through North Kheri FD	56.76	Uttar Pradesh
13	Connecting Pilibhit TR and Kishanpur WLS through North Kheri FD	44.88	Uttar Pradesh
14	Connecting Kishanpur WLS and Dudhwa NP through North Kheri FD	34.83	Uttar Pradesh
15	Connecting Dudhwa NP and Katarniaghat WLS through North Kheri FD (northern part)	115.42	Uttar Pradesh
16	Connecting Dudhwa NP and Katarniaghat WLS through North Kheri FD (southern part)	64.46	Uttar Pradesh
17	Connecting western and eastern parts of Sohagibarwa WLS	18.41	Uttar Pradesh
18	Connecting western and eastern parts of Valmiki TR	8.9	Bihar
Total		2707.54	

TR- Tiger Reserve, CR- Conservation Reserve, SFD- Social Forestry Division, FD- Forest Division, WLS- Wildlife Sanctuary, NP- National Park





Supplementary Table S1: Information of tiger populations, including habitat block details, state, protection status, corridors and identified genetic subpopulations (TGBs) across TAL.

SL No	THB	State	PA, FD, SFD and Corridors	Tiger presence (Unique tigers) (Reference)	TGB
1	THB I	HP	PA: Simbalbara NP	No (a)	
			Corridor: Yamuna river	No (b)	
		HR	PA: Kalesar WLS	No (a)	
		UP	FD: Shivalik FD	No (b)	
		UK	PA: Rajaji TR (western part)	Yes (2) (a)	TGB I
			FD: Dehradun FD	No (b)	
			Corridor: Kansrao-Barkot, Chilla-Motichur	No (b), No (b)	TGB I
2	THB II	UP	PA: Amangarh TR	Yes (20) (a)	TGB I
			SFD: Najibabad SFD	Yes (5) (e)	
		UK	PA: Rajaji TR (eastern part), Jhimil Jheel CR, Corbett TR, Pawalgarh CR	Yes (37) (a), Yes (2) (e), Yes (231) (a), Yes (c)	
			FD: Haridwar FD, Lansdowne FD, Terai West FD, Ramnagar FD, Terai Central FD	No (b), Yes (34) (a), Yes (39) (a), Yes (37) (a), Yes (5) (a)	
			Corridors: Rajaji-Corbett, Kalagarh, Kosi river, Boar river, Nihal-Bhakra, Gola river	Yes (b), Yes (b), Yes (b), Yes (b), Yes (b), No (b)	
3	THB III	UK	PA: Nandhaur WLS	Yes (c)	TGB I
			FD: Haldwani FD, Champawat FD, Terai East FD	Yes (22) (a), Yes (9) (a), Yes (23) (a),	
			Corridor: Kilpura-Khatima-Surai	Yes (d)	
		UP	SFD: Pilibhit SFD	Yes (2) (e)	
4	THB IV	UK	FD: Terai East (eastern part)	Yes (c, e)	TGB II
		UP	PA: Pilibhit TR, Kishanpur WLS (part of Dudhwa TR)	Yes (57) (a), Yes (33) (a)	
			FD: North Kheri FD, South Kheri FD	No (a, e), Yes (2) (a)	
			SFD: Shahjahanpur SFD	No (b),	
			Corridor: Lagga Bagga-Shuklaphanta-Tatarganj, Kishanpur-Dudhwa	Yes (b), Yes (b),	
5	THB V	UP	PA: Dudhwa NP (part of Dudhwa TR)	Yes (20) (a)	TGB II
			Corridor: Dudhwa-Katerniaghat	No (b)	
6	THB VI	UP	PA: Katerniaghat WLS (part of Dudhwa TR)	Yes (29) (a)	
7	THB VII	UP	PA: Suhelwa WLS	No (a)	
8	THB VIII	UP	PA: Sohagibarwa WLS (western part)	Yes (a)	TGB III
9	THB XI	UP	PA: Sohagibarwa WLS (eastern part)	Yes (a)	
		BR	PA: Valmiki TR	Yes (33) (a)	

THB- Tiger habitat block, PA- Protected area, FD- Forest division, SFD- Social forestry division TGB - Tiger genetic block, NP- National park, WLS- Wildlife sanctuary, TR- Tiger reserve, CR- Conservation reserve, HP- Himachal Pradesh, HR- Haryana, UK- Uttarakhand, UP- Uttar Pradesh, BR- Bihar.

a= Jhala et al. 2020, b= Johnsingh et al. 2004, c= Jhala et al. 2015, d= Anwar and Borah 2020, e= This study

Supplementary Table S2: Details of habitat variables used to model tiger habitat permeability in TAL.

Source file	Covariates used	Source	Unit
Forest cover	Distance from open/dense forest	Forest survey of India	Meter
Protected area (PA)	Distance from PA	World database of protected areas	Meter
Road network	Distance from road	Diva-GIS	Meter
Settlement	Distance from settlement	Columbia-Village data	Meter
River network	Distance from river	Diva-GIS	Meter

Supplementary Table S3: Details of nodes and their weightages used in Circuitscape analysis.

SL	Name of the node	Protection status of nodes	Weightage (0-1)
1	Kalesar-Simbalbara complex	Protected area	0
2	Rajaji TR- western part	Protected area	0.50
3	Rajaji TR- eastern part	Protected area	1
4	Jhilmil Jheel CR	Protected area	0.25
5	Najibabad SFD	Non-protected area	0.25
6	Corbett TR	Protected area	1
7	Pawalgarh CR	Protected area	1
8	Nandhaur WLS	Protected area	0.75
9	Pilibhit SFD	Non-protected area	0.50
10	Pilibhit TR	Protected area	0.75
11	Pilibhit TR- southern part	Protected area	0.25
12	Kishanpur WLS	Protected area	0.75
13	Dudhwa NP	Protected area	0.75
14	Katarniaghat Wildlife Sanctuary	Protected area	0.25
15	Suhelwa Wildlife Sanctuary	Protected area	0
16	Sohagibarwa WLS- western part	Protected area	0.25
17	Sohagibarwa WLS- eastern part	Protected area	0.75
18	Sohagibarwa WLS- southern part	Protected area	0
19	Valmiki TR- western part	Protected area	0.75
20	Valmiki TR- central part	Protected area	0.75
21	Valmiki TR- eastern part	Protected area	0.75

TR= Tiger Reserve, CR= Conservation Reserve, SFD= Social Forestry Division, WLS= Wildlife Sanctuary, NP= National Park

Supplementary Table S4: Detail of the first-generation migrant tigers identified through GENECLASS and STRUCTURE in this study.

SL	Sampled location	Migrant from	GENECLASS likelihood computation (L home/L max)	<i>p</i> Resident	STRUCTURE USEPOPINFO (MIGPRIOR 0.05)
TGB I					
1	Rajaji TR	Lansdowne FD	3.645	0.003	NA
2	Rajaji TR	Lansdowne FD	3.695	0.004	NA
3	Lansdowne FD	Rajaji TR	8.089	0.000	NA
4	Haridwar FD	Lansdowne FD	NA	NA	0.936
5	Haridwar FD	Lansdowne FD	NA	NA	0.937
6	Terai West FD	Terai Central FD	7.816	0.001	NA
7	Terai West FD	Terai Central FD	5.653	0.006	NA
8	Ramnagar FD	Terai West FD	6.329	0.006	0.945
9	Ramnagar FD	Terai Central FD	6.307	0.004	NA
10	Haldwani FD	Ramnagar FD	2.347	0.006	0.960
11	Champawat FD	Terai Central FD	4.993	0.000	NA
12	Terai East FD	Haldwani FD	2.374	0.000	NA
13	Corbett TR	Ramnagar FD	NA	NA	0.828
14	Corbett TR	Ramnagar FD	NA	NA	0.820
15	Corbett TR	Ramnagar FD	NA	NA	0.922
16	Amangarh TR	Najibabad SFD	NA	NA	0.995
17	Haldwani FD	Ramnagar FD	NA	NA	0.994
TGB II					
18	Pilibhit TR	Dudhwa NP	4.293	0.008	NA
19	Pilibhit TR	Dudhwa NP	4.259	0.010	NA
20	Kishanpur WLS	Dudhwa NP	2.647	0.000	NA
21	Katarniaghat WLS	Pilibhit TR	3.773	0.000	0.981
22	Pilibhit SFD	Kishanpur WLS	NA	NA	0.985
23	Pilibhit SFD	Kishanpur WLS	NA	NA	0.986
24	Dudhwa NP	Pilibhit TR	NA	NA	0.942
25	Dudhwa NP	Pilibhit TR	NA	NA	0.985
26	South Kheri FD	Dudhwa NP	NA	NA	0.984
27	South Kheri FD	Dudhwa NP	NA	NA	0.989

TGB III					
28	Valmiki TR	Sohagibarwa WLS	5.204	0.001	NA
29	Valmiki TR	Sohagibarwa WLS	2.799	0.007	NA

TGB= Tiger Genetic Block, TR= Tiger Reserve, NP= National Park, WLS= Wildlife Sanctuary, FD= Forest Division, SFD= Social Forestry Division

Supplementary Table S5: Contribution of habitat variables in tiger dispersal across TAL.

Environmental variables	Contribution (%)
Distance from forest	66.8
Distance from protected areas	23.8
Distance from settlements	4.6
Distance from river	3.3
Distance from road	1.5

Supplementary Table S6: Details of least-cost pathways, corridor conductance and critical corridors of tiger dispersal identified across the TAL

SL	Corridors identified between	Length of the Least-cost pathways (km)	Conductance of corridors	Critical corridors
1	Simbalbara-Kaleser complex and Rajaji TR (western part)	41.5	NA	NA
2	Kansrau (western part of Rajaji TR) and Barkot (Dehradun FD)	No	Low	No
3	Chilla (eastern part) and Motichur (western part) ranges of Rajaji TR	3.9	High	Yes
4	Rajaji TR (eastern part) and Jhilmil Jheel CR	9.5	High	No
5	Rajaji TR (eastern part) and Najibabad SFD	9.9	High	No
6	Jhilmil Jheel CR and Najibabad SFD	9.5	High	No
7	Rajaji TR (eastern part) and Corbett TR	36.6	High	Yes
8	Corbett TR and Najibabad SFD	31.2	High	Yes
9	Corbett TR and Pawalgarh CR	1.9	High	Yes
10	Terai West and Terai Central FDs	NA	Low	No
11	Terai Central FD and Ramnagar FDs	NA	Low	No
12	Pawalgarh CR and Nandhaur WLS	53.8	Low	Yes
13	Nandhaur WLS and Pilibhit SFD	32.2	NA	NA
14	Nandhaur WLS and Pilibhit TR	38.6	Medium	Yes
15	Pilibhit SFD and Pilibhit TR	14	NA	NA
16	Northern and southern parts of Pilibhit TR	1.4	High	Yes
17	Pilibhit TR and Kishanpur WLS	5.6	High	Yes
18	Pilibhit TR and Dudhwa NP	14.8	Low	No
19	Kishanpur WLS and Dudhwa NP	16.2	Medium	No
20	Dudhwa NP and Katarniaghat WLS	11.5	Medium	No
21	Katarniaghat WLS and Suhelwa WLS	71.2	NA	NA
22	Suhelwa WLS and west Sohagibarwa WLS	120.3	NA	NA
23	Western and east Sohagibarwa WLS	7.4	Low	Yes
24	Western and eastern parts of Valmiki TR	0.5	High	Yes

TR- Tiger Reserve, CR- Conservation Reserve, SFD- Social Forestry Division, FD- Forest Division, WLS- Wildlife Sanctuary, NP- National Park