

# Individual Project

Angelo LaCommare-Soto

2025-05-11

## Project Title:

The effects of seasonal fish assemblages on eelgrass (*Zostera marina*) bed health within the impacted San Francisco Bay Estuary

## Introduction:

Global seagrass habitat has declined by 19% since 1880 (Dunic et al., 2021). In San Francisco Bay (SFB), the native seagrass commonly known as eelgrass (*Zostera marina*) faces harmful eutrophication-induced microalgal blooms, along with intense mechanical damage, biological invasion, and pollution (Kelly et al., 2019; Cohen & Carlton, 1998; Fonseca et al., 2017). The high ecological value of eelgrass beds combined with their dwindling coverage has prompted scientists to investigate how rapidly changing environmental conditions will affect the suite of organisms that benefit from and contribute to the health of eelgrass.

Local studies have demonstrated that fishes can improve eelgrass performance (Best & Stachowicz, 2012; Carr & Boyer, 2014) via a trophic cascade in which a fish predator directly consumes or alters the grazing behavior of an epifaunal invertebrate that consumes eelgrass leaves, such as *Ampithoe valida* (Harper et al., 2022; Lewis & Boyer, 2014; Reynolds et al., 2012; Ayala, 2021). Current studies are investigating the effects of climate change-induced carbon dioxide increases in baywater predicted by 2100 on the herbivorous feeding preferences of common epifauna like *A. valida*. And while the community composition of epifauna in eelgrass beds in SFB has recently been examined (Ayala, 2021), informing researchers about present day conditions, the same cannot be said about the community composition of fish, as the last monitoring of fish in littoral areas occurred in 1987 (Baxter et al., 1999).

As climate change continues to alter baywater conditions, community fish assemblages and trophic interactions must be evaluated to establish a baseline of the top-down effects of fish on the eelgrass habitat. Therefore, my study will pair environmental DNA (eDNA) and beach seining to assess the composition of fish communities within eelgrass beds. The community survey phase will also be intertwined with a feeding experiment phase that will elucidate trophic interactions between fishes and epifauna throughout changing seasonal conditions. My study will give coastal managers insight into the role that water conditions, food availability, and nursery fish communities play in determining the performance of crucial eelgrass habitat now and under conditions brought about by climate change.

## Guiding Question:

What is the composition of the fish communities in eelgrass beds over a seasonal cycle and how are they involved in the food web of the eelgrass system in SFB?

## Aims:

1. Ascertain how fish assemblages and epifaunal invertebrate communities compare in eelgrass beds over a seasonal cycle and across SFB.

2. Determine the relative seasonal consumption rates of epifauna among some of the most common fishes in eelgrass beds across SFB.

## Statistical Analysis

For my statistical analysis assignment, I will focus on my second aim.

H0: There is no difference in consumption rates of epifauna among seasons and fish individuals per species.

HA: In general, there will be higher consumption rates of epifauna during summer and fall when higher water temperatures increase metabolic rates and larger prey abundances create an environment for reduced resource partitioning, which allow fishes to have more generalized feeding preferences. The species and number of individuals per tank will demonstrate variable epifaunal consumption rates depending on the species-specific prey capture strategies.

I determined relatively early on that a generalized linear model would be the best type of model to analyze my data given the bounded counts that my experiments entail. Using a GLM under Poisson assumptions, I created a model for each epifaunal consumption variable under several predictor variables, such as fish\_mass\_tot, tank, season, and of course treatment. I noticed that AIC scores dropped as I included the interaction of more variables. Ultimately, after re-reading my hypothesis for this project, I decided to settle on a model that describes the interaction between season and treatment on the consumption of epifauna without much focus on the magnitude of the AIC score since these predictors fell more in line with my scientific question. While analyzing the model output, I noticed that solo *P. ornata* individuals consumed, on average, many more gammarid amphipods during summer relative to other seasons for the same treatment and the overall control. In order to showcase this finding, I envisioned a boxplot demonstrating consumption counts for gammarid amphipods subset by season, with the summer *P. ornata* box and whisker plot not overlapping with the control treatment and the other seasons of *P. ornata* treatments. Given that this worked for gammarid amphipods and to maintain consistency, I figured I would also apply the same figure outline for both caprellid amphipod and *Pentidotea resicata* loss.

Below the references is some of the work I have completed to simulate the data values that I can expect during my experiments. More information on my experiments and simulated dataset is available in the dataset card available here: <https://github.com/AngeloLCS/EpifaunaExperimentBIOL710/blob/070cc3c0aeba8b0e9b3f4680e01527374d516e12/RCode/Input/DatasetCard.md>

```
library(tidyverse)
```

```
## Warning: package 'ggplot2' was built under R version 4.4.3
```

```
## Warning: package 'purrr' was built under R version 4.4.3
```

```
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
```

```
## v dplyr      1.1.4      v readr      2.1.5
```

```
## v forcats    1.0.0      v stringr    1.5.1
```

```
## v ggplot2    3.5.2      v tibble     3.2.1
```

```
## v lubridate  1.9.4      v tidyr      1.3.1
```

```
## v purrr      1.0.4
```

```
## -- Conflicts ----- tidyverse_conflicts() --
```

```
## x dplyr::filter() masks stats::filter()
```

```
## x dplyr::lag()     masks stats::lag()
```

```
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors
```

```
getwd()
```

```
## [1] "C:/Users/Angelo L/Documents/GitHub/EpifaunaExperimentBIOL710/RCode/Input"
```

```

# Importing Data
epifauna <- read.csv('epifauna.csv', stringsAsFactors = TRUE )
str(epifauna)

## 'data.frame':    243 obs. of  11 variables:
## $ season      : Factor w/ 3 levels "fall","spring",...: 2 2 2 2 2 2 2 2 2 ...
## $ day         : int  1 1 1 1 1 1 1 1 1 1 ...
## $ tank        : Factor w/ 27 levels "\xd1","A","B",...: 4 15 22 20 11 8 10 23 3 27 ...
## $ treatment   : Factor w/ 11 levels "2_C. aggregata",...: 7 7 7 6 6 6 1 1 1 9 ...
## $ gamm_loss   : int  5 4 4 27 6 1 18 7 11 11 ...
## $ capr_loss   : int  9 0 10 27 26 32 46 33 17 11 ...
## $ pent_loss   : int  3 0 0 1 2 3 1 2 2 5 ...
## $ fish1_mass  : num  NA NA NA 9.39 8.51 8.37 4.67 3.87 3.54 9.17 ...
## $ fish2_mass  : num  NA NA NA NA NA NA 4.42 4 3.86 NA ...
## $ fish_mass_tot: num  NA NA NA 9.39 8.51 8.37 9.09 7.87 7.4 9.17 ...
## $ tank_vol    : num  0.04 0.04 0.04 0.04 0.04 0.04 0.04 0.04 0.04 0.04 ...

# Exploring distribution of data of interest (fish biomass and epifaunal consumption)
gamm_f <- ggplot(epifauna, aes(x=gamm_loss))+
  geom_histogram()
#gamm_f

capr_f <- ggplot(epifauna, aes(x=capr_loss))+
  geom_histogram()
#capr_f

pent_f <- ggplot(epifauna, aes(x=pent_loss))+
  geom_histogram()
#pent_f

fishbm_f <- ggplot(epifauna, aes(x=fish_mass_tot))+
  geom_histogram()
#fishbm_f

# Releveling variables for simpler model output interpretation
epifauna$season <- fct_relevel(epifauna$season, 'spring', 'summer')
epifauna$treatment <- relevel(epifauna$treatment, 'Control')

#Statistical tests gammarid amphipods
#GLM with Poisson family due to count data and right skewness of the data
gamm_glm <- glm(gamm_loss ~ treatment*season, family = "poisson", data = epifauna)
summary(gamm_glm)

##
## Call:
## glm(formula = gamm_loss ~ treatment * season, family = "poisson",
##      data = epifauna)
##
## Coefficients: (6 not defined because of singularities)
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)    1.09861    0.19245   5.709 1.14e-08
## treatment2_C. aggregata 1.81615    0.20751   8.752 < 2e-16

```

## treatment2_H. guttulata	2.04170	0.20456	9.981	< 2e-16
## treatment2_L. armatus	1.92452	0.20602	9.342	< 2e-16
## treatment2_P. californicus	2.19722	0.19920	11.030	< 2e-16
## treatment2_P. ornata	2.15090	0.20334	10.578	< 2e-16
## treatmentC. aggregata	1.77307	0.20815	8.518	< 2e-16
## treatmentH. guttulata	2.20133	0.20282	10.854	< 2e-16
## treatmentL. armatus	1.99243	0.20515	9.712	< 2e-16
## treatmentP. californicus	1.75539	0.20466	8.577	< 2e-16
## treatmentP. ornata	1.69460	0.20938	8.093	5.80e-16
## seasonsummer	-0.07696	0.27756	-0.277	0.7816
## seasonfall	0.03637	0.26972	0.135	0.8927
## treatment2_C. aggregata:seasonsummer	0.26329	0.29675	0.887	0.3749
## treatment2_H. guttulata:seasonsummer	0.26488	0.29297	0.904	0.3659
## treatment2_L. armatus:seasonsummer	0.16487	0.29562	0.558	0.5770
## treatment2_P. californicus:seasonsummer	NA	NA	NA	NA
## treatment2_P. ornata:seasonsummer	0.06395	0.29277	0.218	0.8271
## treatmentC. aggregata:seasonsummer	0.23380	0.29783	0.785	0.4324
## treatmentH. guttulata:seasonsummer	-0.01753	0.29264	-0.060	0.9522
## treatmentL. armatus:seasonsummer	-0.06964	0.29654	-0.235	0.8143
## treatmentP. californicus:seasonsummer	NA	NA	NA	NA
## treatmentP. ornata:seasonsummer	0.69233	0.29583	2.340	0.0193
## treatment2_C. aggregata:seasonfall	0.01070	0.29073	0.037	0.9706
## treatment2_H. guttulata:seasonfall	NA	NA	NA	NA
## treatment2_L. armatus:seasonfall	0.12777	0.28765	0.444	0.6569
## treatment2_P. californicus:seasonfall	NA	NA	NA	NA
## treatment2_P. ornata:seasonfall	0.09662	0.28432	0.340	0.7340
## treatmentC. aggregata:seasonfall	0.10966	0.29065	0.377	0.7059
## treatmentH. guttulata:seasonfall	NA	NA	NA	NA
## treatmentL. armatus:seasonfall	0.19683	0.28601	0.688	0.4913
## treatmentP. californicus:seasonfall	NA	NA	NA	NA
## treatmentP. ornata:seasonfall	0.18269	0.29158	0.627	0.5310
##				
## (Intercept)	***			
## treatment2_C. aggregata	***			
## treatment2_H. guttulata	***			
## treatment2_L. armatus	***			
## treatment2_P. californicus	***			
## treatment2_P. ornata	***			
## treatmentC. aggregata	***			
## treatmentH. guttulata	***			
## treatmentL. armatus	***			
## treatmentP. californicus	***			
## treatmentP. ornata	***			
## seasonsummer				
## seasonfall				
## treatment2_C. aggregata:seasonsummer				
## treatment2_H. guttulata:seasonsummer				
## treatment2_L. armatus:seasonsummer				
## treatment2_P. californicus:seasonsummer				
## treatment2_P. ornata:seasonsummer				
## treatmentC. aggregata:seasonsummer				
## treatmentH. guttulata:seasonsummer				
## treatmentL. armatus:seasonsummer				
## treatmentP. californicus:seasonsummer				

```
## treatmentP. ornata:seasonsummer          *
## treatment2_C. aggregata:seasonfall
## treatment2_H. guttulata:seasonfall
## treatment2_L. armatus:seasonfall
## treatment2_P. californicus:seasonfall
## treatment2_P. ornata:seasonfall
## treatmentC. aggregata:seasonfall
## treatmentH. guttulata:seasonfall
## treatmentL. armatus:seasonfall
## treatmentP. californicus:seasonfall
## treatmentP. ornata:seasonfall
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for poisson family taken to be 1)
##
## Null deviance: 2992.2 on 242 degrees of freedom
## Residual deviance: 2147.0 on 216 degrees of freedom
## AIC: 3281.4
##
## Number of Fisher Scoring iterations: 5
```

*#Statistical tests caprellid amphipods*

```
capr_glm <- glm(capr_loss ~ treatment*season, family = "poisson", data = epifauna)
summary(capr_glm)
```

```
##
## Call:
## glm(formula = capr_loss ~ treatment * season, family = "poisson",
##      data = epifauna)
##
## Coefficients: (6 not defined because of singularities)
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)    1.79176    0.13608  13.167 < 2e-16
## treatment2_C. aggregata    1.52444    0.15017  10.152 < 2e-16
## treatment2_H. guttulata    1.43155    0.15147   9.451 < 2e-16
## treatment2_L. armatus      1.37231    0.15236   9.007 < 2e-16
## treatment2_P. californicus  1.65694    0.17939   9.236 < 2e-16
## treatment2_P. ornata       1.12300    0.15666   7.168 7.59e-13
## treatmentC. aggregata      1.69119    0.14809  11.420 < 2e-16
## treatmentH. guttulata      1.33403    0.15296   8.721 < 2e-16
## treatmentL. armatus        1.61313    0.14903  10.825 < 2e-16
## treatmentP. californicus    1.82280    0.17718  10.288 < 2e-16
## treatmentP. ornata         1.57554    0.14950  10.539 < 2e-16
## seasonsummer      -0.37807    0.21341  -1.772 0.076475
## seasonfall        -0.37807    0.21341  -1.772 0.076475
## treatment2_C. aggregata:seasonsummer  0.04079    0.23501   0.174 0.862213
## treatment2_H. guttulata:seasonsummer  0.32817    0.23371   1.404 0.160272
## treatment2_L. armatus:seasonsummer    0.55804    0.23273   2.398 0.016493
## treatment2_P. californicus:seasonsummer    NA         NA         NA         NA
## treatment2_P. ornata:seasonsummer    0.59395    0.23754   2.500 0.012403
## treatmentC. aggregata:seasonsummer   -0.19012    0.23448  -0.811 0.417482
## treatmentH. guttulata:seasonsummer    0.51464    0.23384   2.201 0.027745
## treatmentL. armatus:seasonsummer     0.06421    0.23299   0.276 0.782851
```

```

## treatmentP. californicus:seasonsummer      NA      NA      NA      NA
## treatmentP. ornata:seasonsummer             0.10181  0.23330  0.436 0.662540
## treatment2_C. aggregata:seasonfall          0.52063  0.23038  2.260 0.023828
## treatment2_H. guttulata:seasonfall          NA      NA      NA      NA
## treatment2_L. armatus:seasonfall            0.35431  0.23462  1.510 0.131010
## treatment2_P. californicus:seasonfall        NA      NA      NA      NA
## treatment2_P. ornata:seasonfall             0.88285  0.23496  3.757 0.000172
## treatmentC. aggregata:seasonfall            0.04023  0.23183  0.174 0.862238
## treatmentH. guttulata:seasonfall            NA      NA      NA      NA
## treatmentL. armatus:seasonfall              0.32112  0.23052  1.393 0.163615
## treatmentP. californicus:seasonfall          NA      NA      NA      NA
## treatmentP. ornata:seasonfall              -0.21664  0.23732 -0.913 0.361316
##
## (Intercept)                                ***
## treatment2_C. aggregata                    ***
## treatment2_H. guttulata                    ***
## treatment2_L. armatus                      ***
## treatment2_P. californicus                 ***
## treatment2_P. ornata                      ***
## treatmentC. aggregata                     ***
## treatmentH. guttulata                     ***
## treatmentL. armatus                       ***
## treatmentP. californicus                  ***
## treatmentP. ornata                       ***
## seasonsummer                             .
## seasonfall                               .
## treatment2_C. aggregata:seasonsummer
## treatment2_H. guttulata:seasonsummer
## treatment2_L. armatus:seasonsummer        *
## treatment2_P. californicus:seasonsummer
## treatment2_P. ornata:seasonsummer         *
## treatmentC. aggregata:seasonsummer
## treatmentH. guttulata:seasonsummer        *
## treatmentL. armatus:seasonsummer
## treatmentP. californicus:seasonsummer
## treatmentP. ornata:seasonsummer
## treatment2_C. aggregata:seasonfall        *
## treatment2_H. guttulata:seasonfall
## treatment2_L. armatus:seasonfall
## treatment2_P. californicus:seasonfall
## treatment2_P. ornata:seasonfall          ***
## treatmentC. aggregata:seasonfall
## treatmentH. guttulata:seasonfall
## treatmentL. armatus:seasonfall
## treatmentP. californicus:seasonfall
## treatmentP. ornata:seasonfall
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for poisson family taken to be 1)
##
## Null deviance: 2675.9 on 242 degrees of freedom
## Residual deviance: 1901.0 on 216 degrees of freedom
## AIC: 3066.5

```

```
##
## Number of Fisher Scoring iterations: 5

#Statistical tests pentidotea isopods
pent_glm <- glm(pent_loss ~ treatment*season, family = "poisson", data = epifauna)
summary(pent_glm)

##
## Call:
## glm(formula = pent_loss ~ treatment * season, family = "poisson",
##      data = epifauna)
##
## Coefficients: (6 not defined because of singularities)
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)    0.28768    0.28867   0.997  0.31898
## treatment2_C. aggregata    0.45953    0.36873   1.246  0.21267
## treatment2_H. guttulata    0.60614    0.35887   1.689  0.09122
## treatment2_L. armatus      1.15268    0.33113   3.481  0.00050
## treatment2_P. californicus  0.96141    0.32609   2.948  0.00320
## treatment2_P. ornata       1.27629    0.32648   3.909 9.26e-05
## treatmentC. aggregata      0.60614    0.35887   1.689  0.09122
## treatmentH. guttulata      0.60614    0.35887   1.689  0.09122
## treatmentL. armatus        1.36524    0.32343   4.221 2.43e-05
## treatmentP. californicus    1.01857    0.32357   3.148  0.00164
## treatmentP. ornata         0.65059    0.35611   1.827  0.06771
## seasonsummer              0.51083    0.36515   1.399  0.16183
## seasonfall                0.08004    0.40032   0.200  0.84152
## treatment2_C. aggregata:seasonsummer 0.18232    0.46074   0.396  0.69231
## treatment2_H. guttulata:seasonsummer 0.26933    0.44679   0.603  0.54663
## treatment2_L. armatus:seasonsummer -0.27721    0.42483  -0.653  0.51407
## treatment2_P. californicus:seasonsummer NA          NA          NA          NA
## treatment2_P. ornata:seasonsummer -0.58315    0.42613  -1.368  0.17117
## treatmentC. aggregata:seasonsummer 0.06169    0.45214   0.136  0.89147
## treatmentH. guttulata:seasonsummer 0.31015    0.44586   0.696  0.48665
## treatmentL. armatus:seasonsummer -0.48977    0.41886  -1.169  0.24228
## treatmentP. californicus:seasonsummer NA          NA          NA          NA
## treatmentP. ornata:seasonsummer 0.20383    0.44507   0.458  0.64698
## treatment2_C. aggregata:seasonfall 0.98285    0.48070   2.045  0.04089
## treatment2_H. guttulata:seasonfall NA          NA          NA          NA
## treatment2_L. armatus:seasonfall 0.28970    0.45250   0.640  0.52202
## treatment2_P. californicus:seasonfall NA          NA          NA          NA
## treatment2_P. ornata:seasonfall -0.23032    0.45884  -0.502  0.61569
## treatmentC. aggregata:seasonfall 0.72074    0.47552   1.516  0.12960
## treatmentH. guttulata:seasonfall NA          NA          NA          NA
## treatmentL. armatus:seasonfall -0.01817    0.44892  -0.040  0.96772
## treatmentP. californicus:seasonfall NA          NA          NA          NA
## treatmentP. ornata:seasonfall 0.42205    0.47963   0.880  0.37889
##
## (Intercept)
## treatment2_C. aggregata
## treatment2_H. guttulata .
## treatment2_L. armatus ***
## treatment2_P. californicus **
## treatment2_P. ornata ***
```

```

## treatmentC. aggregata .
## treatmentH. guttulata .
## treatmentL. armatus ***
## treatmentP. californicus **
## treatmentP. ornata .
## seasonsummer
## seasonfall
## treatment2_C. aggregata:seasonsummer
## treatment2_H. guttulata:seasonsummer
## treatment2_L. armatus:seasonsummer
## treatment2_P. californicus:seasonsummer
## treatment2_P. ornata:seasonsummer
## treatmentC. aggregata:seasonsummer
## treatmentH. guttulata:seasonsummer
## treatmentL. armatus:seasonsummer
## treatmentP. californicus:seasonsummer
## treatmentP. ornata:seasonsummer
## treatment2_C. aggregata:seasonfall *
## treatment2_H. guttulata:seasonfall
## treatment2_L. armatus:seasonfall
## treatment2_P. californicus:seasonfall
## treatment2_P. ornata:seasonfall
## treatmentC. aggregata:seasonfall
## treatmentH. guttulata:seasonfall
## treatmentL. armatus:seasonfall
## treatmentP. californicus:seasonfall
## treatmentP. ornata:seasonfall
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for poisson family taken to be 1)
##
##      Null deviance: 594.11  on 242  degrees of freedom
## Residual deviance: 460.27  on 216  degrees of freedom
## AIC: 1207.7
##
## Number of Fisher Scoring iterations: 5

```

```

#Property plots
# plot(gamm_glm)
# plot(capr_glm)
# plot(pent_glm)

```

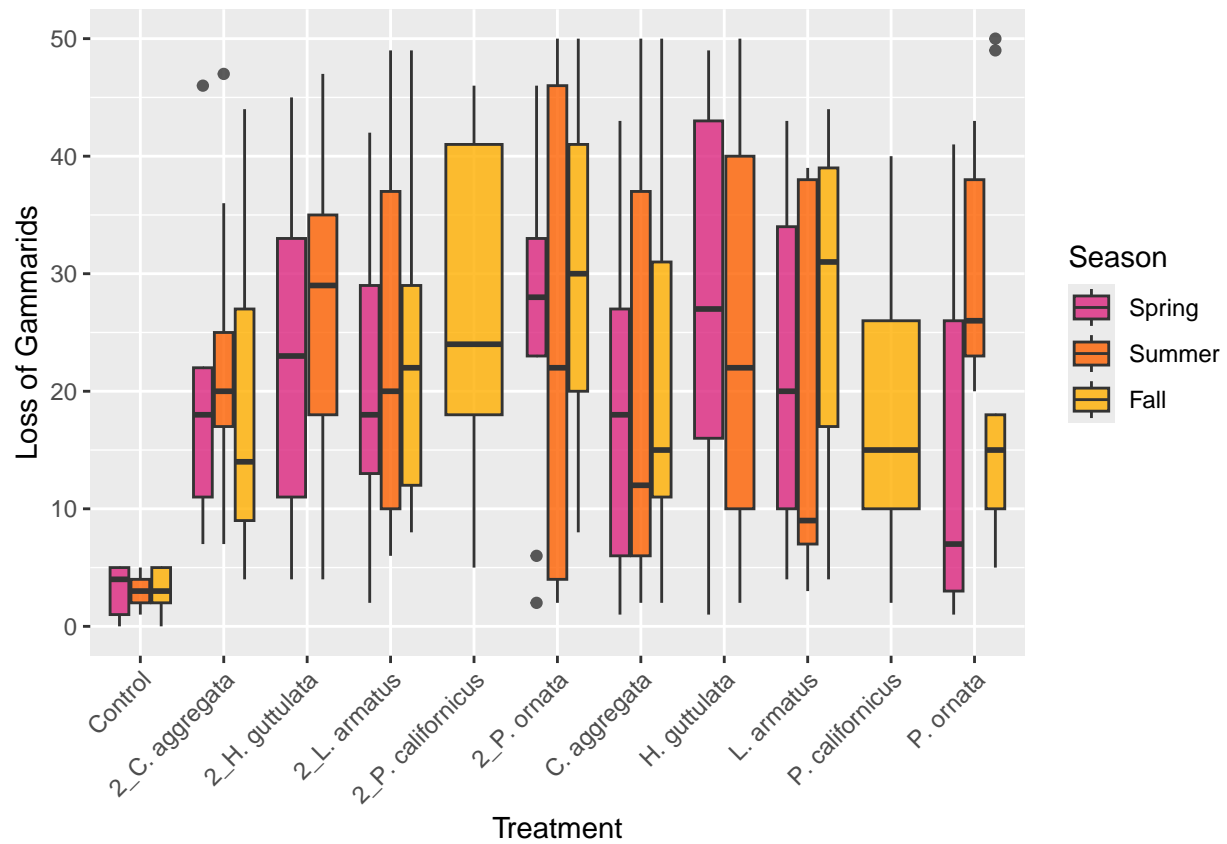
```

#Figures
gamm_f2 <- ggplot(epifauna, aes(x = treatment, y = gamm_loss, fill=season)) +
  geom_boxplot(alpha = 0.8) +
  scale_fill_manual(
    values=c('#DC267F', '#FE6100', '#FFB000'),
    label =c('Spring', 'Summer', 'Fall')
  )+
  labs(
    x = "Treatment",
    y = "Loss of Gammarids",
    fill = 'Season') +

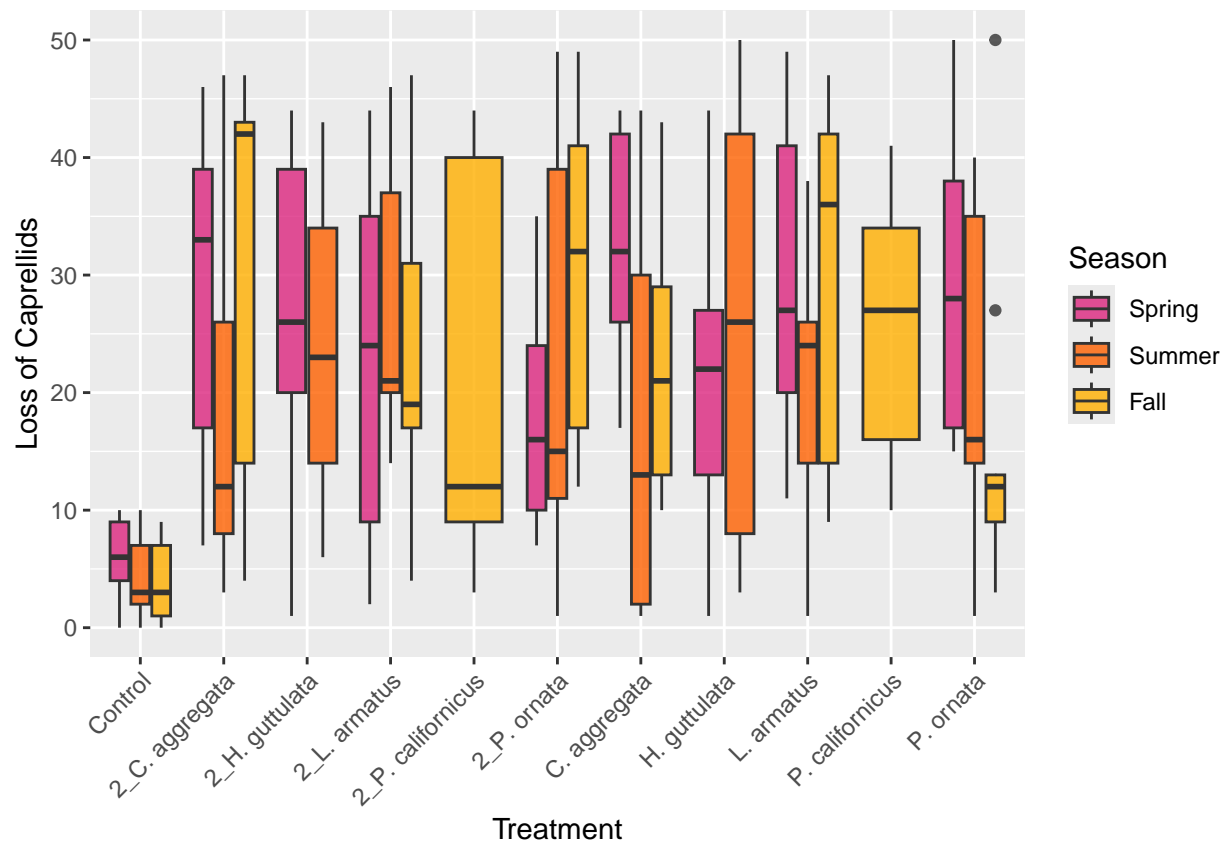
```



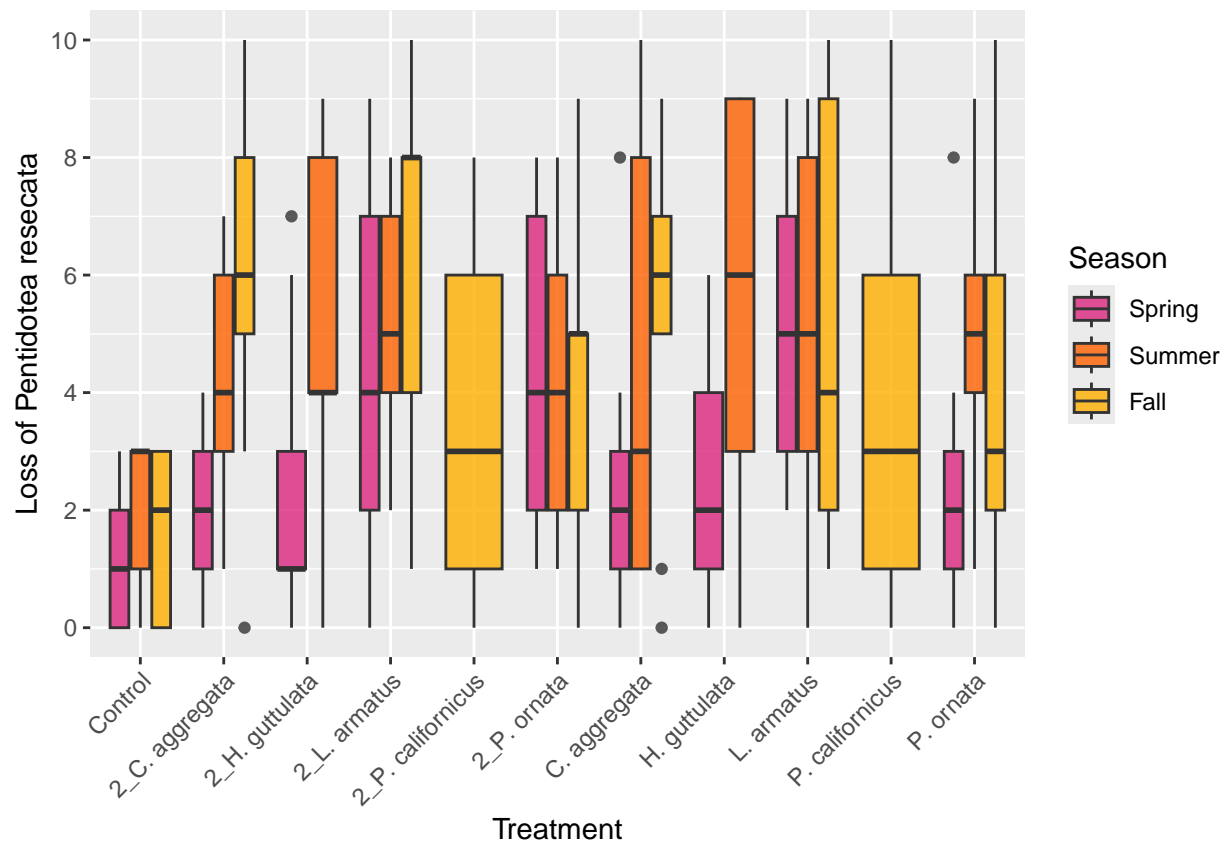
```
theme(axis.text.x = element_text(angle = 45, vjust = 1, hjust = 1))
gamm_f2
```



```
capr_f2 <- ggplot(epifauna, aes(x = treatment, y = capr_loss, fill=season)) +
  geom_boxplot(alpha = 0.8) +
  scale_fill_manual(
    values=c('#DC267F', '#FE6100', '#FFB000'),
    label =c('Spring', 'Summer', 'Fall')
  )+
  labs(
    x = "Treatment",
    y = "Loss of Caprellids",
    fill = 'Season') +
  theme(axis.text.x = element_text(angle = 45, vjust = 1, hjust = 1))
capr_f2
```



```
pent_f2 <- ggplot(epifauna, aes(x = treatment, y = pent_loss, fill=season)) +
  geom_boxplot(alpha = 0.8) +
  scale_fill_manual(
    values=c('#DC267F', '#FE6100', '#FFB000'),
    label =c('Spring', 'Summer', 'Fall')
  )+
  labs(
    x = "Treatment",
    y = "Loss of Pentidotea resecata",
    fill = 'Season') +
  theme(axis.text.x = element_text(angle = 45, vjust = 1, hjust = 1))+
  scale_y_continuous(breaks=c(0, 2, 4, 6, 8, 10))
pent_f2
```



## References:

- Ayala, G. S. 2021. Eelgrass (*Zostera marina*) invertebrate community recovery following an extreme low salinity event. Master's thesis. San Francisco State University, San Francisco, CA.
- Baxter, R., Hieb, K., DeLeon, S., Fleming, K., & Orsi, J. (1999). Report on the 1980-1995 Fish, Shrimp, and Crab Sampling in the San Francisco Estuary, California. <https://nrm.dfg.ca.gov/FileHandler.ashx?DocumentID=227169> [download link]
- Best, R. J. & Stachowicz, J.J. 2012. "Trophic Cascades in Seagrass Meadows Depend on Mesograzer Variation in Feeding Rates, Predation Susceptibility, and Abundance." *Marine Ecology Progress Series* 456 (June): 29–42. <https://doi.org/10.3354/meps09678>.
- Carr, L. A. & Boyer, K. E. 2014. "Variation at Multiple Trophic Levels Mediates a Novel Seagrass-Grazer Interaction." *Marine Ecology Progress Series* 508 (August): 117–28. <https://doi.org/10.3354/meps10855>.
- Cohen, A. N. & Carlton, J. T. 1998. "Accelerating Invasion Rate in a Highly Invaded Estuary." *Science* 279 (5350): 555–58. <https://doi.org/10.1126/science.279.5350.555>.
- Dunic, J. C., Brown, C. J., Connolly, R. M., Turschwell, M. P., & Côté, I. M. 2021. "Long Term Declines and Recovery of Meadow Area Across the World's Seagrass Bioregions." *Global Change Biology* 27 (17): 4096–4109. <https://doi.org/10.1111/gcb.15684>.
- Fonseca, M., Piniak, G. A., & Cosentino-Manning, N. 2017. "Susceptibility of Seagrass to Oil Spills: A Case Study with Eelgrass, *Zostera Marina* in San Francisco Bay, USA." *Marine Pollution Bulletin* 115 (1-2): 29–38. <https://doi.org/10.1016/j.marpolbul.2016.11.029>.
- Harper, K. E., Scheinberg, L. A., Boyer, K. E., & Sotka, E. E. 2022. "Global Distribution of Cryptic Native, Introduced and Hybrid Lineages in the Widespread Estuarine Amphipod *Ampithoe valida*." *Conservation*

*Genetics* 23 (4): 791–806. <https://doi.org/10.1007/s10592-022-01452-8>.

Kelly, J. J., Orr, D., & Takekawa, J. Y. 2019. “Quantification of Damage to Eelgrass (*Zostera marina*) Beds and Evidence-Based Management Strategies for Boats Anchoring in San Francisco Bay.” *Environmental Management* 64 (1): 20–26. <https://doi.org/10.1007/s00267-019-01169-4>.

Lewis, J. T. & Boyer, K. E. 2014. “Grazer Functional Roles, Induced Defenses, and Indirect Interactions: Implications for Eelgrass Restoration in San Francisco Bay.” *Diversity* 6 (4): 751–70. <https://doi.org/10.3390/d6040751>.

Reynolds, L. K., Carr, L. A., & Boyer, K. E. 2012. “A Non-Native Amphipod Consumes Eelgrass Inflorescences in San Francisco Bay.” *Marine Ecology Progress Series* 451 (April): 107–18. <https://doi.org/10.3354/meps09569>.

### Data Simulation Methods:

```
#vector <- rnorm(100, mean = 1.5, sd = 0.7)
#vector

# ctrl_gamm <- sample(0:5, 27, replace = TRUE)
# ctrl_capr <- sample(0:10, 27, replace = TRUE)
# ctrl_pent <- sample(0:3, 27, replace = TRUE)
#
# ctrl_gamm <- as.matrix(ctrl_gamm)
# ctrl_capr <- as.matrix(ctrl_capr)
# ctrl_pent <- as.matrix(ctrl_pent)

# tank <- c("A", "B", "C", "D", "E", "F", "G", "H", "I", "J", "K", "L", "M", "N", "Ñ", "O", "P", "Q", "R", "S", "T", "U", "V", "W", "X", "Y", "Z")
#
# tank_rand1 <- sample(tank, 27)
# tank_rand1
# tank_rand2 <- sample(tank, 27)
# tank_rand3 <- sample(tank, 27)
# tank_rand4 <- sample(tank, 27)
# tank_rand5 <- sample(tank, 27)
# tank_rand6 <- sample(tank, 27)
# tank_rand7 <- sample(tank, 27)
# tank_rand8 <- sample(tank, 27)
# tank_rand9 <- sample(tank, 27)
#
# tank_rand <- rbind(tank_rand1, tank_rand2, tank_rand3, tank_rand4, tank_rand5, tank_rand6, tank_rand7, tank_rand8, tank_rand9)
# tank_rand
# tank_rand <- as.matrix(c(apply(tank_rand, 1, c)))
#for (i in tank) {
#  print(sample(tank, 27))
#}
```

```
# gamm <- sample(1:50, 243, replace = TRUE)
# capr <- sample(1:50, 243, replace = TRUE)
# pent <- sample(0:10, 243, replace = TRUE)
#
# gamm <- as.matrix(gamm)
# capr <- as.matrix(capr)
# pent <- as.matrix(pent)
```

```

# sp_cym <- runif(n=230, min=3.5, max=10)
# sp_lept <- runif(n=27, min=3.7, max=11.4)
# sp_hyps <- runif(n=27, min=3.3, max=9.3)
# sp_pho <- runif(n=27, min=1.4, max=5.9)
#
# sp_cym <- as.matrix(sp_cym)
# sp_lept <- as.matrix(sp_lept)
# sp_hyps <- as.matrix(sp_hyps)
# sp_pho <- as.matrix(sp_pho)

```

```

# rec.list <- function(len){
#   if(length(len) == 1){
#     vector("list", len)
#   } else {
#     lapply(1:len[1], function(...) rec.list(len[-1]))
#   }
# }

```

```

# set.seed(5094)
#
# season <- c('spring', 'summer', 'fall')
# tank <- c("A", "B", "C", "D", "E", "F", "G", "H", "I", "J", "K", "L", "M", "N", "Ñ", "O", "P", "Q", "R")
# treatment <- c("control", "control", "control", "C_aggregata", "C_aggregata", "C_aggregata", "2C_aggregata")
#
# Data_full <- rec.list(c(length(season), 3, length(tank)))
#
# for (x in 1:length(season)) {
#   for (u in 1:3){#day
#     for (t in 1:length(tank)){
#
#       pent_loss <- sample(1:10, 1, replace=TRUE)
#       gamm_loss <- sample(1:50, 1, replace=TRUE)
#       capr_loss <- sample(1:50, 1, replace=TRUE)
#       treat <- sample(treatment, 1, replace=FALSE)
#       # fish1_mass<-
#       #   if (treat == 'control'){
#       #     NA
#       #   } else if (season == 'spring' & treat == 'C_aggregata' ){
#       #     runif(n=1, min=3.5, max=10)
#       #   } else {
#       #     10
#       #   }
#
#       Data_full[[x]][[u]][[t]] <- data_frame(
#         'pent_loss'=pent_loss,
#         'gamm_loss'=gamm_loss,
#         'capr_loss'= capr_loss,
#         'treatment' = treat)#,
#         # 'fish1_mass' = fish1_mass)
#     } #tank
#     names(Data_full[[x]][[u]]) <- tank
#   } #day
# } #season

```

```
# names(Data_full) <- season
#
#
# Test <- unlist(Data_full)
#
# Test2 <- data.frame(Reduce(rbind, Data_full))
#
# Test3 <- ldply(Data_full, data_frame())
```