

Class09

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Section 1

```
db<- read.csv("Data Export Summary.csv")
db
```

	Molecular.Type	X.ray	EM	NMR	Multiple.methods	Neutron	Other
1	Protein (only)	154,766	10,155	12,187	191	72	32
2	Protein/Oligosaccharide	9,083	1,802	32	7	1	0
3	Protein/NA	8,110	3,176	283	6	0	0
4	Nucleic acid (only)	2,664	94	1,450	12	2	1
5	Other	163	9	32	0	0	0
6	Oligosaccharide (only)	11	0	6	1	0	4
	Total						
1		177,403					
2		10,925					
3		11,575					
4		4,223					
5		204					
6		22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
Xray.tot <-sum(as.numeric(gsub(",", "", db$X.ray)))
EM.tot <- sum(as.numeric(gsub(",", "", db$EM)))
NMR.tot <- sum(as.numeric(gsub(",", "", db$NMR)))
```

Hmm.. I am doing the same thing over and over. Time to write a function.

```
#I will work with 'x' as input
sum_comma <- function(x) {
  # Substitute the comma and convert ot numeric
  sum(as.numeric(gsub(",", "", x)))
}
```

For Xray:

```
round(sum_comma(db$X.ray) / sum_comma((db$Total)), 3)
```

```
[1] 0.855
```

85.5% are Xrays

For EM:

```
round(sum_comma(db$EM) / sum_comma((db$Total)), 3)
```

```
[1] 0.075
```

7.5% are EMs

Q2: What proportion of structures in the PDB are protein?

```
round(sum_comma(db$Total[1])/ sum_comma(db$Total),3)
```

```
[1] 0.868
```

86.8% of structures in PBD are protein

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

SKIPPED! (It appears that there are 2064 HIV-1 protease structures in the current PBD.)

Section 2

Using Molstar

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

The structure is too low a resolution to see H atoms. You need a sub 1 Angstrom resolution to see Hydrogen.

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

HOH308

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.



Figure 1: HIV-PR structure from MERK with a bound drug

Section 3

Working with Structures in R

We can read THE bio3d package to read and perform bioinformatics calculations on PDB structures.

```
library(bio3d)

pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
  Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

```
Protein sequence:
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

Q7: How many amino acid residues are there in this pdb object?

There are 198 amino acid residues.

Q8: Name one of the two non-protein residues?

Two of the non-protein residues are HOH(127) and MK1(1)

Q9: How many protein chains are in this structure?

There are 2 protein chains in this structure.

```
attributes(pdb)
```

```
$names
```

```
[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"
```

```
$class
```

```
[1] "pdb" "sse"
```

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

Predicting Functional Motions

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
```

```
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
```

```
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 244 (residues: 244)
```

```
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

```
Protein sequence:
```

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV  
DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQM  
TAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

```
+ attr: atom, xyz, seqres, helix, sheet,  
      calpha, remark, call
```

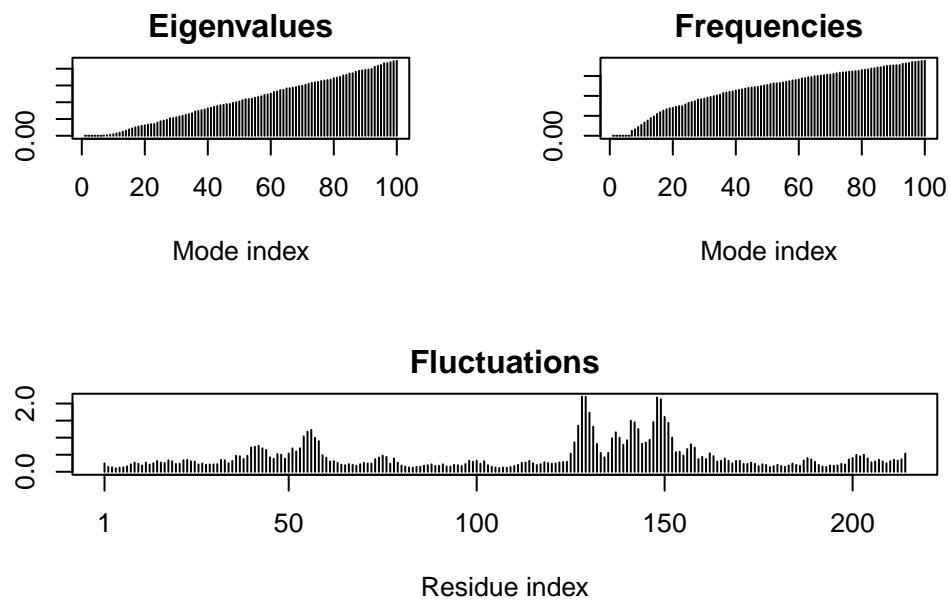
Perform a prediction of flexibility with a technique called the NMA (Normal MOde Analysis)

```
# Perform flexibility prediction  
m <- nma(adk)
```

```
Building Hessian... Done in 0.045 seconds.
```

```
Diagonalizing Hessian... Done in 0.406 seconds.
```

```
plot(m)
```



Write out a “movie”(a.k.a trajectory) of the motion for viewing in Molstar

```
mktrj(m, file="adk_m7.pdb")
```