

Processing Stool Sample Whole Genome Sequencing Data

Ankan Choudhury

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Introduction

This code includes the bash code for performing Whole-genome Sequence analysis on the Stool samples using Trimmomatic, Bowtie, Kraken, Bracken, Kaiju, Metaphlan, and Humann. The paired sequences (“raw”) were first trimmed using Trimmomatic v0.39, which trimmed the sequences and separated out into paired (“trimmed”) and unpaired, sequences who do not have a matching read pairing with it. The trimmed sequences were then filtered for host reads (*Homo sapiens*) using Bowtie2 resultant sequences (“trimmed-filtered”) were used for taxonomic classification. Taxonomic classification was done at all three levels of sequence refinement were done using Kraken2, Kaiju, and Metaphlan. Kraken2 classification used standard Kraken database and was normalized by Bracken using standard Bracken database. Kaiju classification was done using the Kaiju RefSeq database. Metaphlan 4.1.1 used its own ChocoPhlan database (updated October 2024). The trimmed-filtered sequences were also assembled into contigs using Megahit, and the assembled sequences (“trimmed-filtered-contigs”) were also classified using Kraken2-Bracken algorithm. The raw, trimmed, trimmed-filtered sequences were also functionally annotated using Humann 3.9. The resultant taxonomic classification data and functional annotation data were transformed and compiled into phyloseq objects in R.

Prerequisites

1. Install qiime2-metagenome-2024.10
2. Install Trimmomatic 0.39
3. Install Metaphlan 4.4.1
4. Install Humann 3.9
5. Install R 4.3.1 or higher

1. Getting the sequences

All the code chunks given below should be made into a .sh files and executed by batch submission in kodiak server as they usually take long time to run.

```
ZIP_FILE="061824KGmetagenome.zip" # The zip file containing the sequences
EXTRACT_DIR="extracted_files"
DEST_DIR="demultiplexed/raw-paired"
mkdir -p "$EXTRACT_DIR" "$DEST_DIR"

unzip -o "$ZIP_FILE" -d "$EXTRACT_DIR"

find "$EXTRACT_DIR" -type f -name "*.fastq.gz" -exec mv {} "$DEST_DIR" \;
```

For subsequent operations, please use a custom temporary folder when using kodiak server as if the temporary files generated during any operation exceeds the given space, it will shut the operation down. Add this piece of code to your bash script every time

```
TEMP_DIR="tmp"
mkdir -p "$TEMP_DIR"
export TMPDIR="$TEMP_DIR"
export TMP="$TEMP_DIR"
export TEMP="$TEMP_DIR"
```

2. Trimming the sequences using Trimmomatic v0.39

Install Trimmomatic v0.39 prior to this. Include the path where trimmomatic-0.39.jar executable file is stored.

```
cd demultiplexed

mkdir -p trimmed-paired # to store trimmed and pair matched sequences
mkdir -p trimmed-unpaired # to store unmatched sequences

for f in $(find raw-paired -maxdepth 1 -name "*R1_001.fastq.gz" -printf "%f\n"
→ | sed 's/1_001.fastq.gz//' | sort -u)
do
java -jar /path/to/Trimmomatic-0.39/trimmomatic-0.39.jar PE -phred33
→ ${f}1_001.fastq.gz ${f}2_001.fastq.gz trimmed-paired/${f}1_001.fastq.gz
→ trimmed-unpaired/${f}1_001.fastq.gz trimmed-paired/${f}2_001.fastq.gz
→ trimmed-unpaired/${f}2_001.fastq.gz LEADING:3 TRAILING:3
→ SLIDINGWINDOW:4:15 MINLEN:35;
echo Processed: ${f}1_001.fastq.gz & ${f}2_001.fastq.gz
done
```

Storing the trimmed sequences as a qiime2 artifact (.qza)

```
source activate qiime2-metagenome-2024.10 #activating qiime2 metagenome
→ distribution

qiime tools import \
  --type 'SampleData[PairedEndSequencesWithQuality]' \
  --input-path demultiplexed/trimmed-paired \
  --input-format CasavaOneEightSingleLanePerSampleDirFmt \
  --output-path demux-paired/trimmed.qza

qiime demux summarize \
  --i-data demux-paired/trimmed.qza \
  --o-visualization demux-paired/trimmed.qzv #summary visualization of the
→ sequences
```

3. Filtering the sequences with Bowtie2

Bowtie2 requires a Bowtie database built from genomic data provided by user. Currently, the database used here was built from Human Genome Data gathered from NCBI. If you are using Bowtie2 standalone installation then you

can download it from the Bowtie indices for H. sapiens, GRCh38 no-alt analysis set from here. Here Bowtie2 was used from the qiime2 metagenome distribution platform, so the database has to be in a qimme2 artifact (.qza) format. For that, we can download the FASTA version of H. sapiens genome GRCh38 in NCBI from here, and build it into Bowtie2 database format using the following bash script.

```
cd /path/to/where/it/is/stored/GRCh38_FASTA

source activate qiime2-metagenome-2024.10

qiime tools import \
    ↪ .qza format #Importing the genome stored in .fna to
    --input-path GRCh38.fna \
    --output-path GRCh38.qza \
    --type 'FeatureData[Sequence]'

qiime quality-control bowtie2-build \
    --i-sequences GRCh38.qza \
    --o-database Hsapiens_BowTie2Index.qza #Building Bowtie2 Index from it
```

After BowTie2 Indices are built, we can use that to filter human reads from the sequences using the following bash script.

```
source activate qiime2-metagenome-2024.10

qiime quality-control filter-reads \
    --i-demultiplexed-sequences demux-paired/trimmed.qza \
    ↪ trimmed sequences #Input
    --i-database /path/to/where/it/is/stored/Hsapiens_BowTie2Index.qza \
    ↪ #Bowtie2 Index built from Human genome
    --o-filtered-sequences demux-paired/trimmed-filtered.qza #Output
    ↪ trimmed & filtered sequences
```

4. Assembling the sequences into contigs using Megahit

The assemble contigs can be used for taxonomic classification using Kraken2 and functional annotation using eggNOG.

```
mkdir -p contigs

source activate qiime2-metagenome-2024.10

qiime assembly assemble-megahit \
    --i-seqs demux-paired/trimmed-filtered.qza \
    ↪ filtered sequences #Input trimmed and
    --p-presets "meta-sensitive" \
    ↪ species in the metagenome #Preset which looks for more rarer
    --p-num-cpu-threads 24 \
    ↪ #number of cpus to be used
    --p-num-partitions 1 \
    --o-contigs contigs/trimmed-filtered-contigs.qza \
    --verbose
```

5. Taxonomic classification using Kraken2

We can use Kraken2 to classify both sequences and contigs using the same basic script. Kraken2 is a taxonomic classification system using exact k-mer matches to achieve high accuracy and fast classification speeds. This classifier matches each k-mer within a query sequence to the lowest common ancestor (LCA) of all genomes containing the given k-mer. You can install Kraken2 as a stand-alone, but here we have used kraken2 through the qiime2 metagenome platform's moshpit distribution package. Before that we will have to build databses for Kraken2 and Bracken using this script. We will be storing the database in the qiime cache. A cache will store the "data" of a dataset in a separate data folder with a random generated name, which is stored in a separate "key" file named after the dataset in the keys folder. In a qiime artifact (.qza), they are all bundled up together into one thing that has to zipped and unzipped everytime you use it.

```
source activate qiime2-metagenome-2024.10

qiime tools cache-create --cache cache          #creating the cache

qiime moshpit build-kraken-db \
  --p-collection standard \                   #Using the standard Kraken2 and Bracken
  ↪ database
  --o-kraken2-database cache:kraken_standard \
  --o-bracken-database cache:bracken_standard \
  --verbose
```

After the databases are built we can go ahead and classify the data using Kraken2.

```
source activate qiime2-metagenome-2024.10

qiime moshpit classify-kraken2 \
  --i-seqs contigs/trimmed-filtered.qza \      #Input sequences/contigs
  --i-kraken2-db cache:kraken_standard \
  --p-threads 40 \                             #Number of threads to use in computation
  --p-confidence 0.1 \                         #Confidence level (0 to 1) of the matches that
  ↪ will be made, higher the value more stringent the algorithm
  --p-minimum-base-quality 0 \                 #Minimum base quality at which Kraken2
  ↪ classifies a read
  --o-hits cache:kraken_hits-trimmed-filtered \ #Kraken2 hits stored
  ↪ in cache format
  --o-reports cache:kraken_reports-trimmed-filtered \ #Kraken2 reports
  ↪ stored in cache format
  --use-cache cache \
  --verbose \
  --p-memory-mapping False
```

Kraken outputs contain a feature table (.biom), taxonomy file (.txt) and kraken reports (.txt). Kraken reports contain taxon information on each line: Percentage of fragments covered by the clade root, number of fragments covered by clade root, Number of fragments assigned directly to this taxon, a rank code: indicating (U)nclassified, (R)oot, (D)omain, (K)ingdom, (P)hylum, (C)lass, (O)rder, (F)amily, (G)enus, or (S)pecies, NCBI taxonomic ID number, and taxonomic annotation.

```
## 26.51 1453057 1453057 U 0 unclassified
## 73.49 4028832 338 R 1 root
## 73.49 4028400 1671 R1 131567 cellular organisms
## 73.44 4025812 61371 D 2 Bacteria
```

##	55.30	3031565	15	D1	1783270	FCB group
##	55.30	3031550	47	D2	68336	Bacteroidota/Chlorobiota group
##	55.30	3031503	10014	P	976	Bacteroidota
##	55.12	3021465	124	C	200643	Bacteroidia
##	55.11	3021340	251536	O	171549	Bacteroidales
##	49.56	2716871	99066	F	815	Bacteroidaceae
##	29.01	1590028	202688	G	909656	Phocaeicola
##	25.00	1370421	1370421	S	821	Phocaeicola vulgatus
##	0.28	15136	14805	S	357276	Phocaeicola dorei
##	0.01	331	331	S1	997877	Phocaeicola dorei CL03T12C01
##	0.03	1782	1782	S	387090	Phocaeicola coprophilus
##	0.00	1	0	S	376805	Phocaeicola salanitronis
##	0.00	1	1	S1	667015	Phocaeicola salanitronis DSM 18170
##	18.75	1027777	505940	G	816	Bacteroides
##	5.42	297365	294611	S	820	Bacteroides uniformis
##	0.05	2754	2754	S1	997890	Bacteroides uniformis CL03T12C01
##	3.27	179117	179117	S	28116	Bacteroides ovatus
##	0.36	19970	19970	S	818	Bacteroides thetaiotaomicron
##	0.10	5697	652	G1	2646097	unclassified Bacteroides
##	0.03	1600	1600	S	2528203	Bacteroides sp. A1C1
##	0.02	1100	1100	S	2785531	Bacteroides sp. HF-162

Renormalization with Bracken

Bracken (Bayesian Reestimation of Abundance with Kraken) is a highly accurate statistical method that computes the abundance of species in DNA sequences from a metagenomics sample. Bracken is a companion program to Kraken 2. While Kraken classifies reads to multiple levels in the taxonomic tree, Bracken allows estimation of abundance at a single level using those classifications (e.g. Bracken can estimate abundance of species within a sample). Bracken uses a Bracken database, the length of your reads and the kraken reports to give you a feature frequency table that renormalizes the data by dropping the unclassified reads and according to the Bayesian probability of the kraken hits being correct.

```
source activate qiime2-metagenome-2024.10

mkdir kraken-outputs

name="trimmed-filtered"

qiime moshpit estimate-bracken \
  --i-bracken-db cache:bracken_standard \
  --p-read-len 150 \
  --i-kraken-reports cache:kraken_reports-$name \
  --o-reports kraken-outputs/bracken-reports-$name.qza \
  --o-taxonomy kraken-outputs/taxonomy-bracken-$name.qza \
  --o-table kraken-outputs/table-bracken-$name.qza
```

The Bracken reports are very similar to Kraken2 reports except for having no unclassified reads

##	100.00	4028748	0	R	1	root
##	100.00	4028645	0	R1	131567	cellular organisms
##	99.95	4026906	0	D	2	Bacteria
##	76.64	3087472	0	D1	1783270	FCB group
##	76.64	3087472	0	D2	68336	Bacteroidota/Chlorobiota group

##	76.64	3087472	0	P	976	Bacteroidota
##	76.64	3087439	0	C	200643	Bacteroidia
##	76.64	3087438	0	O	171549	Bacteroidales
##	75.25	3031475	0	F	815	Bacteroidaceae
##	45.53	1834358	0	G	909656	Phocaeicola
##	44.98	1812114	1812114	S	821	Phocaeicola vulgatus
##	0.50	20141	20141	S	357276	Phocaeicola dorei
##	0.05	2101	2101	S	387090	Phocaeicola coprophilus
##	0.00	1	1	S	376805	Phocaeicola salanitronis
##	29.71	1197116	0	G	816	Bacteroides
##	13.24	533218	533218	S	820	Bacteroides uniformis
##	12.94	521396	521396	S	28116	Bacteroides ovatus
##	0.74	29918	0	G1	2646097	unclassified Bacteroides
##	0.25	10241	10241	S	2528203	Bacteroides sp. AlC1
##	0.24	9713	9713	S	2785531	Bacteroides sp. HF-162

The taxonomy file contains the NCBI taxa ID number and the string of taxonomic names leading to the species or the final taxonomic ranks for that ID.

##	Feature ID	Taxon
##	821	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Phocaeicola
##	357276	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Phocaeicola
##	387090	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Phocaeicola
##	376805	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Phocaeicola
##	820	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	28116	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	2528203	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	2785531	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	2763022	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	2847299	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	556259	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	2755405	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	3117552	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	2709390	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	818	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	817	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	371601	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	2792859	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides
##	2715212	d__Bacteria;k__Bacteria;p__Bacteroidota;c__Bacteroidia;o__Bacteroidales;f__Bacteroidaceae;g__Bacteroides

The feature table (.biom) is a BIOM format matrix table with Feature IDs as the row number and the number of reads for that feature in each sample as column data.

##	1-MDcol002-5-2	11-MDcol002-label-12-on-bottle
##	1002546	0
##	1002689	0
##	1005039	0
##	1005665	0
##	1006	0
##	1006155	0
##	14-MDCOL003-Stool-1B-Baseline-BSB-2-4-1-4	15-MDcol004-3-1-pretreat
##	1002546	0
##	1002689	0

##	1005039			0		0
##	1005665			0		0
##	1006			0		0
##	1006155			0		0
##		17-MDCOL006-3-1A	19-MDCOL006-3-2A	21-MDCOL006-2-pre-chemo-A		
##	1002546	0	0		0	
##	1002689	0	0		0	
##	1005039	0	0		0	
##	1005665	0	0		0	
##	1006	0	0		0	
##	1006155	0	0		0	
##		23-MDcol033-1A	25-MDcol036-1A	27-MDcol025-1B	28-7A-1-1	29-7A
##	1002546	0	0	0	0	0
##	1002689	0	0	0	0	0
##	1005039	0	0	0	0	0
##	1005665	0	0	0	0	0
##	1006	0	0	0	0	0
##	1006155	0	0	0	0	0
##		3-MDcol002-5-1	30-8B	31-8B	33-6-2	36-40-1B
##	1002546	0	0	0	0	0
##	1002689	0	0	0	1	0
##	1005039	0	0	1	0	0
##	1005665	0	0	0	0	0
##	1006	0	0	0	0	59
##	1006155	0	0	0	6	0
##		40-MDCol002-7-43	42-51-1B	45-MDCOL005-1B	46-MDCOL015-1a	48-MDCOL039-1a
##	1002546	0	0	0	3	0
##	1002689	0	0	0	0	0
##	1005039	0	0	0	0	0
##	1005665	0	0	1	0	0
##	1006	0	0	0	0	0
##	1006155	0	0	0	0	0
##		5-MDCOL055-Stool-1A-Baseline-BSB-2-4-1-2	50-MDCOL027-1a	52-MDCOL021-1b		
##	1002546		0	0		0
##	1002689		0	0		0
##	1005039		0	0		0
##	1005665		0	0		0
##	1006		0	0		0
##	1006155		0	0		0
##		54-MDCOL028-1b	60-MDcol006-4-1A	63-MDcol003-3-2		
##	1002546	1	0	0		
##	1002689	0	0	0		
##	1005039	0	0	0		
##	1005665	1	0	0		
##	1006	0	0	0		
##	1006155	0	0	0		
##		65-MDCOL003-Post-Chemo-Collection-Cycle-1-A	66-MDCOL004-1A			
##	1002546		0	0		
##	1002689		0	0		
##	1005039		0	0		
##	1005665		0	0		
##	1006		0	0		
##	1006155		0	0		
##		68-MDCOL003-3-1-Before-Chemo-2-3A				

```

## 1002546 0
## 1002689 0
## 1005039 0
## 1005665 0
## 1006 0
## 1006155 0
## 7-MDCOL058-Stool-1A-Baseline-BSB-2-4-1-3 70-MDCOL004-3-2-Post-Chemo-A
## 1002546 0 0
## 1002689 0 0
## 1005039 0 0
## 1005665 0 0
## 1006 0 0
## 1006155 0 0
## 72-MDCOL003-4-1-4A-Before-Chemo-6 75-MDCOL053-1-1-1-Unknown
## 1002546 0 0
## 1002689 0 0
## 1005039 0 0
## 1005665 0 0
## 1006 0 0
## 1006155 0 0
## 9-MDCOL062-Stool-1A-Baseline-BSB-2-4-1-3
## 1002546 0
## 1002689 0
## 1005039 0
## 1005665 0
## 1006 0
## 1006155 0

```

Creating phylogenetic trees from the bracken data

We can generate phylogenetic trees from the bracken data using a package called gracken, which can be installed by `pip install gracken` or from this link (<https://github.com/jonasoh/gracken>). But gracken uses scientific names as phylogenetic tree tip labels whereas here the feature table contains the NCBI taxa IDs, so you can replace the executable python file for gracken in the miniconda path “miniconda/lib/python3.9/site-packages/gracken/” with this file gracken.py, which contains an updated code. Once you have it installed, this code will create the phylogenetic tree for each dataset (“raw”, “trimmed”, “filtered”...etc) and an OTU table , but we can ignore that.

```

mkdir -p bracken-outputs

source activate qiime2-metagenome-2024.10

for f in $(find kraken-outputs -maxdepth 2 -name "table-bracken*")
do
  #Converting the bracken feature tables from a .qza to folder format
  filename=$(basename "$f" .qza)
  qiime tools export \
    --input-path $f \
    --output-path bracken-outputs/$filename
  echo $filename
done

for f in $(find kraken-outputs -maxdepth 2 -name "taxonomy-bracken*")
do
  #Converting the bracken taxonomy from a .qza to folder format

```



```

do
filename=$(basename "$f" .qza)
qiime tools export \
  --input-path $f \
  --output-path bracken-outputs/$filename
echo $filename
done

for f in $(find kraken-outputs -maxdepth 2 -name "bracken-reports*")
→ #Converting the bracken reports from a .qza to folder format
do
filename=$(basename "$f" .qza)
qiime tools export \
  --input-path $f \
  --output-path bracken-outputs/$filename
echo $filename
done

find bracken-outputs -type d -name "bracken-reports-*" | while read dir; do
→ #Create a phylogenetic tree for each dataset using the bracken reports
  gracken --input_dir $dir --out_prefix $dir/output --mode bracken
done

```

6. Taxonomic Classification using Kaiju

Kaiju is another taxonomic classification tool that assigns each sequencing read to a taxon in the NCBI taxonomy by comparing it to a reference protein database containing microbial and viral protein sequences. By using protein-level classification, Kaiju achieves a higher sensitivity compared with methods based on nucleotide comparison. You can use any of the several reference protein databases, such as complete genomes from NCBI RefSeq or the microbial subset of the NCBI BLAST non-redundant protein database nr, optionally also including fungi and microbial eukaryotes. We have used the NCBI RefSeq and have run the kaiju classification through the qiime2 metagenome platform's moshpit distribution package. The databases can be built in the following manner and stored as a cache artifact.

```

source activate qiime2-metagenome-2024.10
qiime moshpit fetch-kaiju-db \
  --p-database-type 'refseq' \
  --o-database cache:KaijuDB_refseq \
  --verbose

```

After the database has been built we can classify the datasets by Kaiju to any taxonomic level of choice. Here we have classified it to the species level, hence any reads that could not be assigned to any species level with enough confidence but can be assigned to other taxonomic ranks will be put in the “cannot be assigned to a (non-viral) species” category, which is different than the “unclassified” category, which contains reads that cannot be assigned to any taxonomic category at all. To classify using Kaiju, use the following code.

```

source activate qiime2-metagenome-2024.10
qiime moshpit classify-kaiju \
  --i-seqs /data/choudhurya/COLON_MD/contigs/filtered-contigs.qza \
  --i-db cache:KaijuDB_refseq \
  --p-z 40 \
  --p-a 'greedy' \
  --p-c 0 \

```

```
--o-abundances cache:kaiju_abun-trimmed-filtered \           #Stores the
↳ abundance table
--o-taxonomy cache:kaiju_taxonomy-trimmed-filtered \         #Stores the
↳ taxonomy file
--use-cache cache \
--verbose
```

The output from Kaiju is a BIOM formatted feature table (.biom) and a taxonomy file.

Feature Table from Kaiju:

```
##          1-MDcol002-5-2 11-MDcol002-label-12-on-bottle
## 0          107510          128875
## 100886          1062          1096
## 102148          0          0
## 10239          297          343
## 1042156          753          1060
## 105841          836          1093
##          14-MDCOL003-Stool-1B-Baseline-BSB-2-4-1-4 15-MDcol004-3-1-pretreat
## 0          92791          160866
## 100886          568          856
## 102148          0          733
## 10239          2204          84395
## 1042156          2081          2218
## 105841          1655          937
##          17-MDCOL006-3-1A 19-MDCOL006-3-2A 21-MDCOL006-2-pre-chemo-A
## 0          66562          69952          166779
## 100886          0          0          1156
## 102148          0          0          0
## 10239          203          382          520
## 1042156          0          0          0
## 105841          455          861          3894
##          23-MDcol033-1A 25-MDcol036-1A 27-MDcol025-1B 28-7A-1-1 29-7A
## 0          74759          103900          117685          118836 78654
## 100886          0          872          843          710 477
## 102148          0          0          0          0 0
## 10239          34575          514          17332          790 737
## 1042156          621          1146          1993          902 487
## 105841          3625          6632          1159          908 504
##          3-MDcol002-5-1 30-8B 31-8B 33-6-2 36-40-1B 38-S-Swanson
## 0          154682 42309 129571 103313 171345 234863
## 100886          828 294 982 635 1406 2094
## 102148          0 0 546 395 0 0
## 10239          612 448 569 260 513 2846
## 1042156          1068 396 1662 1230 1234 3697
## 105841          1192 387 1981 1360 1506 2760
##          40-MDcol002-7-43 42-51-1B 45-MDCOL005-1B 46-MDCOL015-1a 48-MDCOL039-1a
## 0          189502 115185 118140 134839 135318
## 100886          1769 600 0 572 867
## 102148          0 0 0 851 0
## 10239          388 816 8264 317 4951
## 1042156          1597 827 1787 1930 2703
## 105841          1913 1098 2959 6296 1430
```

##	5-MDCOL055-Stool-1A-Baseline-BSB-2-4-1-2	50-MDCOL027-1a	52-MDCOL021-1b
## 0	60205	164777	56860
## 100886	449	7126	464
## 102148	0	794	1648
## 10239	215	5659	58741
## 1042156	462	2630	1060
## 105841	438	1403	21535
##	54-MDCOL028-1b	60-MDcol006-4-1A	63-MDcol003-3-2
## 0	85864	101022	37472
## 100886	0	0	0
## 102148	809	0	0
## 10239	1693	242	13305
## 1042156	0	1153	0
## 105841	1179	5277	0
##	65-MDCOL003-Post-Chemo-Collection-Cycle-1-A	66-MDCOL004-1A	
## 0	75672	10878	
## 100886	807	51	
## 102148	0	82	
## 10239	1695	3027	
## 1042156	2534	104	
## 105841	1480	76	
##	68-MDCOL003-3-1-Before-Chemo-2-3A		
## 0	63818		
## 100886	425		
## 102148	0		
## 10239	3053		
## 1042156	1489		
## 105841	1098		
##	7-MDCOL058-Stool-1A-Baseline-BSB-2-4-1-3	70-MDCOL004-3-2-Post-Chemo-A	
## 0	47845	2676	
## 100886	329	0	
## 102148	0	0	
## 10239	152	725	
## 1042156	396	24	
## 105841	385	21	
##	72-MDCOL003-4-1-4A-Before-Chemo-6	75-MDCOL053-1-1-1-Unknown	
## 0	48410	88527	
## 100886	377	846	
## 102148	0	0	
## 10239	406	1170	
## 1042156	737	1557	
## 105841	864	746	
##	9-MDCOL062-Stool-1A-Baseline-BSB-2-4-1-3		
## 0	110766		
## 100886	998		
## 102148	0		
## 10239	768		
## 1042156	2019		
## 105841	1065		

Taxonomy from Kaiju:

##	Feature.ID
## 1	0

```
## 2      100886
## 3      102148
## 4      10239
## 5      1042156
## 6      105841
##
## 1
## 2 d__Bacteria;p__Bacillota;c__Erysipelotrichia;o__Erysipelotrichales;f__Coprobacillaceae
## 3      d__Bacteria;p__Bacillota;c__Erysipelotrichia;o__Erysipelotrichales;f__Erysipelotr
## 4
## 5      d__Bacteria;p__Bacillota;c__Clostridia;o__Eubacteriales;f__Clostr
## 6      d__Bacteria;p__Bacillota;c__Clostridia;o__Lachnospirales;f__Lachn
```

7. Taxonomic Classification using Metaphlan

MetaPhlAn 4 relies on unique clade-specific marker genes identified from ~1M microbial genomes for taxonomic assignment of metgenome sequence reads. Metaphlan 4.1.1 was used as a standalone installation as a miniconda environment, whose installation information can be gathered from here (<https://github.com/biobakery/MetaPhlAn/wiki/MetaPhlAn-4.1>). Metaphlan automatically downloads its latest database “ChocoPhlan” when you use it for the first time, but you can also do it manually with this `$ metaphlan --install --index <latest database index> --bowtie2db <database folder>`. The latest database indices can be found here (http://cmprod1.cibio.unitn.it/biobakery4/metaphlan_databases/). Once everything is installed, the following code was used to perform classification using Metaphlan for our datasets.

```
name="trimmed-filtered"          #dataset name

source activate qiime2-metagenome-2024.10

qiime tools export \              #Fetching the dataset from .qza format to a folder
  format
  --input-path demux-paired/$name \
  --output-path demultiplexed/$name

conda activate mpa                #Activating the miniconda environment, here named as
  "mpa"

mkdir -p MetaphlanAnalysis/BowTieOutput/$name MetaphlanAnalysis/Output/$name
  # folders for storing Metaphlan output and the intermediated Bowtie
  alignment output

folder="demultiplexed/$name"      #folder storing our sequences

# Loop over sample names
for f in $(find ${folder} -maxdepth 1 -name "*R1_001.fastq.gz" | sed 's|.|/|'|
  # | sed 's/R1_001.fastq.gz//' | sort -u)
do
  # Define output filename without _S[number]_L[number]_R
  clean_f=$(echo ${f} | sed -E 's/_S[0-9]+_L[0-9]+_/_//')

  # Run MetaPhlAn
  metaphlan ${folder}/${f}R1_001.fastq.gz,${folder}/${f}R2_001.fastq.gz
    --bowtie2db <database folder> --bowtie2out
    MetaphlanAnalysis/BowTieOutput/$name/${clean_f}.bowtie2.bz2
    --input_type fastq --nproc 28 -o
    MetaphlanAnalysis/Output/$name/${clean_f}.output.txt
```

```
echo ${clean_f} is done
done
```

A typical Metaphlan output contains taxonomic relative abundances

```
## [1] #mpa_vOct22_CHOCOPhAnSGB_202403
## [2] #/data/choudhurya/miniconda/envs/mpa/bin/metaphlan demultiplexed/trimmed-filtered-p
## [3] #10709265 reads processed
## [4] #SampleID    Metaphlan_Analysis
## [5] #clade_name  NCBI_tax_id relative_abundance  additional_species
## [6] k__Bacteria 2    100.0
## [7] k__Bacteria|p__Firmicutes    2|1239    51.68535
## [8] k__Bacteria|p__Bacteroidetes    2|976    46.48729
## [9] k__Bacteria|p__Actinobacteria    2|201174    1.22089
## [10] k__Bacteria|p__Proteobacteria    2|1224    0.60648
## [11] k__Bacteria|p__Bacteroidetes|c__Bacteroidia    2|976|200643    46.42399
## [12] k__Bacteria|p__Firmicutes|c__Clostridia    2|1239|186801    29.73363
## [13] k__Bacteria|p__Firmicutes|c__CFGB3002    2|1239|    15.59072
## [14] k__Bacteria|p__Firmicutes|c__CFGB3012    2|1239|    2.14771
## [15] k__Bacteria|p__Firmicutes|c__Negativicutes    2|1239|909932    1.73986
## [16] k__Bacteria|p__Firmicutes|c__CFGB3005    2|1239|    1.38625
## [17] k__Bacteria|p__Actinobacteria|c__Coriobacteriia    2|201174|84998    1.22089
## [18] k__Bacteria|p__Proteobacteria|c__Betaproteobacteria    2|1224|28216    0.46794
## [19] k__Bacteria|p__Firmicutes|c__CFGB3054    2|1239|    0.25919
## [20] k__Bacteria|p__Firmicutes|c__Firmicutes_unclassified    2|1239|    0.24734
```

The file has a five line header (lines beginning with #). We've added [i]s (line numbers) to the header text above to help call out their roles more clearly.

1. Indicates the version of the marker gene database that MetaPhlAn used in this run. There are currently ~1.1M unique clade-specific marker genes identified from ~100k reference genomes (~99,500 bacterial and archaeal and ~500 eukaryotic) included in the database.
2. Provides a copy of the command that was run to produce this profile. This includes the path to the MetaPhlAn executable, the input file that was analyzed, and any custom parameters that were configured.
3. Indicates the number of sample reads processed.
4. Lists your sample name.
5. Provides column headers from the profile data that follows. More specifically, [5] includes four headers to be familiar with:

clade_name: The taxonomic lineage of the taxon reported on this line, ranging from Kingdom (e.g. Bacteria/Archaea) to species genome bin (SGB). Taxon names are prefixed to help indicate their rank: Kingdom: k__, Phylum: p, **Class:** c, Order: o, **Family:** f, Genus: g, **Species:** s, and SGB: t__.

NCBI_tax_id: The NCBI-equivalent taxon IDs of the named taxa from clade_name.

relative_abundance: The taxon's relative abundance in %. Since typical shotgun-sequencing-based taxonomic profile is relative (i.e. it does not provide absolute cell counts), clades are hierarchically summed. Each taxonomic level will sum to 100%. That is, the sum of all kingdom-level clades is 100%, the sum of all phylum-level clades is 100%, and so forth.

additional_species: Additional species names for cases where the metagenome profile contains clades that represent multiple species. The species listed in column 1 is the representative species in such cases.

8. Functional annotation using Humann 3.9

HUMAnN is a pipeline for efficiently and accurately profiling the presence/absence and abundance of microbial pathways in a community from metagenomic or metatranscriptomic sequencing data (typically millions of short DNA/RNA reads). This process, referred to as functional profiling, aims to describe the metabolic potential of a microbial community and its members. You can install Humann 3.9 as a stand-alone through here (<https://github.com/biobakery/humann>). Once installed, it automatically downloads the ChocoPhlan database (if not already installed) to do taxonomic analysis using Metaphlan on its first run, but you will have to download the translated search database for annotation. You can do it using this code `humann_databases --download uniref uniref90_diamond $INSTALL_LOCATION`, which will download the full UniRef90 database used in our analysis. After that, run this code to annotate the sequences using HUMAnN. (P.S: Unlike MetaPhlan, HUMAnN cannot parse paired read sequences, so we will have to first concatenate both reads from each sample into a single .fastq file and then send it off for functional annotation)

```
name="trimmed-filtered"           #Dataset name

mkdir -p HumannAnalysis/HumannOutput/$name      #where the output is to be
→ stored

folder="demultiplexed/$name"           #Dataset folder

cd $folder
forward=$(echo *R1*.fastq.gz)
reverse=$(echo *R2*.fastq.gz)

mkdir -p concatenated

for f in $forward
do
    for r in $reverse
    do
        clean_f=$(echo ${f} | sed -E
        → 's/_S[0-9]+_L[0-9]+_R[0-9]_[0-9]+.fastq.gz//')
        clean_r=$(echo ${r} | sed -E
        → 's/_S[0-9]+_L[0-9]+_R[0-9]_[0-9]+.fastq.gz//')

        if [ $clean_f = $clean_r ]; then
            cat $f $r > concatenated/$clean_f.fastq.gz      #concatenating
            → the forward and reverse reads of the same sample
            humann --input concatenated/$clean_f.fastq.gz --threads 16
            → --output HumannAnalysis/HumannOutput/$name
            fi
        done
    done
done
rm -rf concatenated              #deleting the folder storing the concatenated reads
```

HUMAnN output consists of three files: Gene families file (`$SAMPLENAME_genefamilies.tsv`), Path Coverage file (`$SAMPLENAME_pathcoverage.tsv`), and Path Abundance file (`$SAMPLENAME_pathabundance.tsv`).

The **gene families file** contains the the abundance of each gene family in the community. Gene families are groups of evolutionarily-related protein-coding sequences that often perform similar functions. Gene family abundance at the community level is stratified to show the contributions from known and unknown species. Individual species' abundance contributions sum to the community total abundance. Gene family abundance is reported in RPK (reads per kilobase) units to normalize for gene length; RPK units reflect relative gene (or transcript) copy number in the community.

```
##                                     Gene-Family
## 1                                     UNMAPPED
## 2                                     UniRef90_A0A174NA57
## 3 UniRef90_A0A174NA57|g__Bacteroides.s__Bacteroides_uniformis
## 4                                     UniRef90_A0A395VRV3
## 5     UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_ovatus
## 6     UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_vulgatus
## 1-MDcol002-5-2_Abundance-RPKs
## 1                                     2364890.000
## 2                                     55247.258
## 3                                     55247.258
## 4                                     49394.382
## 5                                     25032.358
## 6                                     8720.539
```

The **path coverage file** provides information on the completeness of metabolic pathways in a given sample. It contains two main columns: Pathway, which lists metabolic pathway identifiers, and Coverage, which represents the proportion of the pathway detected in the sample as a value between 0 and 1. A coverage value of 0.00 indicates that the pathway is absent, 0.50 suggests that half of its reactions are present, and 1.00 means the pathway is fully reconstructed. These pathway identifiers are derived from the MetaCyc database.

```
##                                     Pathway
## 1                                     UNMAPPED
## 2                                     UNINTEGRATED
## 3 UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## 4     UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus
## 5     UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus
## 6     UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
## 1-MDcol002-5-2_Coverage
## 1                                     1
## 2                                     1
## 3                                     1
## 4                                     1
## 5                                     1
## 6                                     1
```

The **path abundance file** details the abundance of each pathway in the community as a function of the abundances of the pathway's component reactions, with each reaction's abundance computed as the sum over abundances of genes catalyzing the reaction. Pathway abundance is proportional to the number of complete "copies" of the pathway in the community but unlike gene abundance, a pathway's community-level abundance is not necessarily the sum of its stratified abundance values. It consists of two main columns: Pathway, which contains metabolic pathway identifiers from MetaCyc, and Abundance, which represents the cumulative abundance of reactions contributing to the pathway

```
##                                     Pathway 1-MDcol002-5-2_Abundance
## 1                                     UNMAPPED 778748.2
## 2                                     UNINTEGRATED 2481983.1
## 3 UNINTEGRATED|g__Bacteroides.s__Bacteroides_vulgatus 917463.5
## 4 UNINTEGRATED|g__Bacteroides.s__Bacteroides_uniformis 464620.0
## 5     UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus 354114.6
## 6                                     UNINTEGRATED|unclassified 147854.1
```

After gathering all the data, we will move to performing further data transformation in R in steps 9 onwards.

Required packages for the following R code

```
library(biomformat)
library(tidyverse)
library(data.table)
library(fs)
library(phyloseq)
library(writexl)
library(phylostratr)
library(ape)
library(taxize)
```

9. Transforming Kaiju Data

The following code is to read the Kaiju feature table (.biom) and Kaiju taxonomy, merge them together to create an abundance table that has both the taxonomy and the reads in each sample for each taxa, provided all the biom files and taxonomy files are kept in a folder named “Kaiju Taxonomy Abundance”.

```
Kaijubioms<-dir("Kaiju Taxonomy Abundance",pattern="*biom")      #Listing all
→ biom files from the kaiju folder
Kaijutaxa<-dir("Kaiju Taxonomy Abundance",pattern="kaiju_taxonomy*") #Listing
→ all taxonomy files from the kaiju folder
for (biom in Kaijubioms){
  for(taxa in Kaijutaxa){
    if(substring(biom,21,nchar(biom)-5)==substring(taxa,16,nchar(taxa)-4))
      name=substring(biom,21,nchar(biom)-5)
    OTUtable<-as.data.frame(as.matrix(biom_data(read_biom(paste("Kaiju
→ Taxonomy Abundance",biom,sep="/"))))%>% #Reading biom files
      rownames_to_column("Feature.ID")
    taxatable<-read.table(paste("Kaiju Taxonomy
→ Abundance",taxa,sep="/"),sep='\t',header = TRUE)      #Reading the
→ taxonomy file
    Abundance<-inner_join(OTUtable,taxatable)%>%           #merging them together
→ and separating the taxonomy string into separate columns for each
→ taxonomic ranks
      select(-"Feature.ID")%>%
      relocate(Taxon)%>%
      mutate(Taxon=str_replace_all(Taxon,".__",""))%>%

    → separate(Taxon,c("Domain","Phylum","Class","Order","Family","Genus","Species"),se
    mutate(Phylum=if_else(is.na(Phylum),Domain,Phylum),
      Class=if_else(is.na(Class),Phylum,Class),
      Order=if_else(is.na(Order),Class,Order),
      Family=if_else(is.na(Family),Order,Family),
      Genus=if_else(is.na(Genus),Family,Genus),
      Species=if_else(is.na(Species),Genus,Species))
    write_xlsx(Abundance,paste("Kaiju Taxonomy
→ Abundance/kaiju_abun-",name,".xlsx",sep=""))      #Writing the table as
→ an excel file
  }
}
```


Sample Kaiju feature table

##	Feature.ID	1-MDcol002-5-2	11-MDCol002-label-12-on-bottle					
## 1	0	107567			128946			
## 2	100886	1063			1096			
## 3	102148	0			0			
## 4	10239	297			344			
## 5	1042156	754			1061			
## 6	105841	837			1093			
##	14-MDCOL003-Stool-1B-Baseline-BSB-2-4-1-4	15-MDcol004-3-1-pretreat						
## 1		92867			164303			
## 2		568			856			
## 3		0			734			
## 4		2207			84670			
## 5		2083			2220			
## 6		1655			938			
##	17-MDCOL006-3-1A	19-MDCOL006-3-2A	21-MDCOL006-2-pre-chemo-A	23-MDcol033-1A				
## 1	66642	70074		166933		74818		
## 2	0	0		1157		0		
## 3	0	0		0		0		
## 4	204	383		520		34634		
## 5	0	0		0		622		
## 6	455	861		3895		3628		
##	25-MDcol036-1A	27-MDcol025-1B	28-7A-1-1	29-7A	3-MDcol002-5-1	30-8B	31-8B	
## 1	104008	117792	118937	78699	154777	42339	129679	
## 2	872	844	710	478	828	294	983	
## 3	0	0	0	0	0	0	546	
## 4	515	17373	791	737	612	448	572	
## 5	1147	1995	903	487	1068	396	1662	
## 6	6639	1165	908	504	1193	387	1983	
##	33-6-2	36-40-1B	38-S-Swanson	40-MDCol002-7-43	42-51-1B	45-MDCOL005-1B		
## 1	103400	171450	235039	189626	115278	118226		
## 2	635	1406	2095	1769	600	0		
## 3	395	0	0	0	0	0		
## 4	261	513	2850	388	818	8271		
## 5	1231	1234	3699	1598	828	1787		
## 6	1360	1506	2764	1913	1098	2961		
##	46-MDCOL015-1a	48-MDCOL039-1a	5-MDCOL055-Stool-1A-Baseline-BSB-2-4-1-2					
## 1	134994	135414				60250		
## 2	572	867				449		
## 3	853	0				0		
## 4	319	4953				215		
## 5	1931	2704				462		
## 6	6302	1430				438		
##	50-MDCOL027-1a	52-MDCOL021-1b	54-MDCOL028-1b	60-MDcol006-4-1A	63-MDcol003-3-2			
## 1	164912	56900	86452	101341	37508			
## 2	7133	466	0	0	0			
## 3	794	1650	810	0	0			
## 4	5663	58791	1695	331	13311			
## 5	2632	1061	0	1153	0			
## 6	1403	21548	1187	5279	0			
##	65-MDCOL003-Post-Chemo-Collection-Cycle-1-A	66-MDCOL004-1A						
## 1		77198		33754				
## 2		807		0				

## 3	0	0
## 4	1801	5560
## 5	2536	0
## 6	1480	0
## 68-MDCOL003-3-1-Before-Chemo-2-3A	7-MDCOL058-Stool-1A-Baseline-BSB-2-4-1-3	
## 1	63548	47689
## 2	425	331
## 3	0	0
## 4	3058	152
## 5	1490	397
## 6	1099	385
## 70-MDCOL004-3-2-Post-Chemo-A	72-MDCOL003-4-1-4A-Before-Chemo-6	
## 1	13449	48576
## 2	0	377
## 3	0	0
## 4	3811	416
## 5	0	737
## 6	0	865
## 75-MDCOL053-1-1-1-Unknown	9-MDCOL062-Stool-1A-Baseline-BSB-2-4-1-3	
## 1	88590	110828
## 2	847	999
## 3	0	0
## 4	1170	768
## 5	1560	2020
## 6	747	1068

Sample Kaiju taxonomy file

##	Feature.ID
## 1	0
## 2	100886
## 3	102148
## 4	10239
## 5	1042156
## 6	105841
##	
## 1	d__belong to a (non-
## 2	d__Bacteria;p__Bacillota;c__Erysipelotrichia;o__Erysipelotrichales;f__Coproba
## 3	d__Bacteria;p__Bacillota;c__Erysipelotrichia;o__Erysipelotrichales;f__Erysipelotr
## 4	
## 5	d__Bacteria;p__Bacillota;c__Clostridia;o__Eubacteriales;f__Clostr
## 6	d__Bacteria;p__Bacillota;c__Clostridia;o__Lachnospirales;f__Lachn

Sample Kaiju combined abundance table

##	Domain
## 1	belong to a (non-viral) species with less than 0.01% of all reads
## 2	Bacteria
## 3	Bacteria
## 4	Viruses
## 5	Bacteria
## 6	Bacteria
##	Phylum

```

## 1 belong to a (non-viral) species with less than 0.01% of all reads
## 2 Bacillota
## 3 Bacillota
## 4 Viruses
## 5 Bacillota
## 6 Bacillota
## Class
## 1 belong to a (non-viral) species with less than 0.01% of all reads
## 2 Erysipelotrichia
## 3 Erysipelotrichia
## 4 Viruses
## 5 Clostridia
## 6 Clostridia
## Order
## 1 belong to a (non-viral) species with less than 0.01% of all reads
## 2 Erysipelotrichales
## 3 Erysipelotrichales
## 4 Viruses
## 5 Eubacteriales
## 6 Lachnospirales
## Family
## 1 belong to a (non-viral) species with less than 0.01% of all reads
## 2 Coprobacillaceae
## 3 Erysipelotrichaceae
## 4 Viruses
## 5 Clostridiaceae
## 6 Lachnospiraceae
## Genus
## 1 belong to a (non-viral) species with less than 0.01% of all reads
## 2 Catenibacterium
## 3 Solobacterium
## 4 Viruses
## 5 Clostridium
## 6 Anaerostipes
## Species
## 1 belong to a (non-viral) species with less than 0.01% of all reads
## 2 Catenibacterium mitsuokai
## 3 Solobacterium moorei
## 4 Viruses
## 5 Clostridium sp. SY8519
## 6 Anaerostipes caccae
## 1-MDcol002-5-2 11-MDcol002-label-12-on-bottle
## 1 107567 128946
## 2 1063 1096
## 3 0 0
## 4 297 344
## 5 754 1061
## 6 837 1093
## 14-MDCOL003-Stool-1B-Baseline-BSB-2-4-1-4 15-MDcol004-3-1-pretreat
## 1 92867 164303
## 2 568 856
## 3 0 734
## 4 2207 84670
## 5 2083 2220

```

## 6			1655		938	
##	17-MDCOL006-3-1A	19-MDCOL006-3-2A	21-MDCOL006-2-pre-chemo-A	23-MDcol033-1A		
## 1	66642	70074		166933	74818	
## 2	0	0		1157	0	
## 3	0	0		0	0	
## 4	204	383		520	34634	
## 5	0	0		0	622	
## 6	455	861		3895	3628	
##	25-MDcol036-1A	27-MDcol025-1B	28-7A-1-1	29-7A	3-MDcol002-5-1	30-8B
## 1	104008	117792	118937	78699	154777	42339
## 2	872	844	710	478	828	294
## 3	0	0	0	0	0	0
## 4	515	17373	791	737	612	448
## 5	1147	1995	903	487	1068	396
## 6	6639	1165	908	504	1193	387
##	33-6-2	36-40-1B	38-S-Swanson	40-MDCol002-7-43	42-51-1B	45-MDCOL005-1B
## 1	103400	171450	235039	189626	115278	118226
## 2	635	1406	2095	1769	600	0
## 3	395	0	0	0	0	0
## 4	261	513	2850	388	818	8271
## 5	1231	1234	3699	1598	828	1787
## 6	1360	1506	2764	1913	1098	2961
##	46-MDCOL015-1a	48-MDCOL039-1a	5-MDCOL055-Stool-1A-Baseline-BSB-2-4-1-2			
## 1	134994	135414			60250	
## 2	572	867			449	
## 3	853	0			0	
## 4	319	4953			215	
## 5	1931	2704			462	
## 6	6302	1430			438	
##	50-MDCOL027-1a	52-MDCOL021-1b	54-MDCOL028-1b	60-MDcol006-4-1A	63-MDcol003-3-2	
## 1	164912	56900	86452	101341	37508	
## 2	7133	466	0	0	0	
## 3	794	1650	810	0	0	
## 4	5663	58791	1695	331	13311	
## 5	2632	1061	0	1153	0	
## 6	1403	21548	1187	5279	0	
##	65-MDCOL003-Post-Chemo-Collection-Cycle-1-A	66-MDCOL004-1A				
## 1		77198	33754			
## 2		807	0			
## 3		0	0			
## 4		1801	5560			
## 5		2536	0			
## 6		1480	0			
##	68-MDCOL003-3-1-Before-Chemo-2-3A	7-MDCOL058-Stool-1A-Baseline-BSB-2-4-1-3				
## 1		63548			47689	
## 2		425			331	
## 3		0			0	
## 4		3058			152	
## 5		1490			397	
## 6		1099			385	
##	70-MDCOL004-3-2-Post-Chemo-A	72-MDCOL003-4-1-4A-Before-Chemo-6				
## 1		13449			48576	
## 2		0			377	
## 3		0			0	

## 4	3811	416
## 5	0	737
## 6	0	865
## 75-MDCOL053-1-1-1-Unknown	9-MDCOL062-Stool-1A-Baseline-BSB-2-4-1-3	
## 1	88590	110828
## 2	847	999
## 3	0	0
## 4	1170	768
## 5	1560	2020
## 6	747	1068

Creating Phyloseq Objects for the Kaiju data

The following code packages the feature table and the taxonomy file into a single phyloseq object for the datasets (“raw”, “trimmed-filtered” etc). A Phyloseq object usually contains,

- a **sample metadata**, which contains the sample IDs and metadata pertaining to it.
- a **feature table**, which is a matrix where the rows are the taxa/feature and columns are the sample IDs with read count for each taxa/feature.
- a **taxonomy file**, which is a matrix where the rows are the taxa/feature and columns are the taxonomy path for each taxa/feature.
- a **phylogenetic tree**, a phylogenetic tree with the taxa/feature as the tip labels.

The phyloseq object can be made with one or more of these data file.

```
Kaijubiom<-dir("Kaiju Taxonomy Abundance",pattern="*biom")      #Listing all
↳ biom files from the kaiju folder
Kaijutaxa<-dir("Kaiju Taxonomy Abundance",pattern="kaiju_taxonomy*")  #Listing
↳ all taxonomy files from the kaiju folder
dir.create("PhyloseqOutputs/KaijuPhyloseq")
for (biom in Kaijubiom){
  for(taxa in Kaijutaxa){
    if(substring(biom,21,nchar(biom)-5)==substring(taxa,16,nchar(taxa)-4))
      name=substring(biom,21,nchar(biom)-5)
    OTUtable<-as.matrix(biom_data(read_biom(paste("Kaiju Taxonomy
↳ Abundance",biom,sep="/")))) #Reading biom files
    taxatable<-read.table(paste("Kaiju Taxonomy
↳ Abundance",taxa,sep="/"),sep='\t',header = TRUE)%>%      #Reading the
↳ taxonomy file
      mutate(Taxon=str_replace_all(Taxon,".____",""))%>%

    ↳ separate(Taxon,c("Domain","Phylum","Class","Order","Family","Genus","Species"),se
    mutate(Phylum=if_else(is.na(Phylum),Domain,Phylum),
      Class=if_else(is.na(Class),Phylum,Class),
      Order=if_else(is.na(Order),Class,Order),
      Family=if_else(is.na(Family),Order,Family),
      Genus=if_else(is.na(Genus),Family,Genus),
      Species=if_else(is.na(Species),Genus,Species))%>%
    column_to_rownames("Feature.ID")%>%
    as.matrix()
    physeq <- phyloseq(otu_table(OTUtable, taxa_are_rows = TRUE),
↳ tax_table(taxatable)) #Compiling the phyloseq object

    ↳ saveRDS(physeq,paste("PhyloseqOutputs/KaijuPhyloseq/phyloseq_kaiju-",name,".rds",se
    ↳ #Saving it as an .RDS file
```

```
}
}
```

Summary of the sample phyloseq object

```
##      Length      Class      Mode
##           1 phyloseq         S4

## phyloseq-class experiment-level object
## otu_table()      OTU Table:         [ 621 taxa and 37 samples ]
## tax_table()      Taxonomy Table:    [ 621 taxa by 7 taxonomic ranks ]
```

10. Transforming Kraken2-Bracken Data

The following code is to read the Bracken feature table (.biom) and Bracken taxonomy, merge them together to create an abundance table that has both the taxonomy and the reads in each sample for each taxa.

```
brackentables<-dir(path="BrackenOutputs",pattern="table-bracken*")    #Listing
  ↳ all the folders containing the bracken feature table
brackentaxonomy<-dir(path="BrackenOutputs",pattern="taxonomy-bracken*")
  ↳ #Listing all the folders containing the bracken taxonomy

dir.create("Kraken-Bracken Taxonomy Abundance")
for(each in brackentables){
  name<-substring(each,15)
  for(each2 in brackentaxonomy){
    name2<-substring(each2,18)
    if(name==name2)

    ↳ {table<-as.data.frame(as.matrix(biom_data(read_biom(paste("BrackenOutputs",each,"feature
    ↳ #Reading biom file
      rownames_to_column("Feature.ID")

    ↳ taxonomy<-read.table(paste("BrackenOutputs",each2,"taxonomy.tsv",sep='/'),sep="\t",head
    ↳ = TRUE)%>% #Reading taxonomy file
      mutate(Feature.ID=as.character(Feature.ID))
    taxatable<-inner_join(table,taxonomy)%>% #Joining into an abundance table
      select(-"Feature.ID")%>%
      relocate(Taxon)%>%
      mutate(Taxon=str_replace_all(Taxon,".____",""))%>%

      ↳ separate(Taxon,c("Domain","Kingdom","Phylum","Class","Order","Family","Genus","Sp
    write_xlsx(taxatable,paste("Kraken-Bracken Taxonomy
    ↳ Abundance/Kraken-Bracken_abun-",name,".xlsx",sep="")) #Writing the
    ↳ abundance table as excel files
  }
}
```

Sample Bracken feature table, taxonomy and combined abundance table

##	Feature.ID	1-MDcol002-5-2	11-MDcol002-label-12-on-bottle				
## 1	100886	5	8				
## 2	102148	0	0				
## 3	102684	0	0				
## 4	1042156	0	0				
## 5	104609	0	0				
## 6	1051631	0	0				
##	14-MDCOL003-Stool-1B-Baseline-BSB-2-4-1-4	15-MDcol004-3-1-pretreat					
## 1		0	1				
## 2		0	2				
## 3		0	0				
## 4		0	3				
## 5		0	0				
## 6		1	0				
##	17-MDCOL006-3-1A	19-MDCOL006-3-2A	21-MDCOL006-2-pre-chemo-A	23-MDcol033-1A			
## 1	9	5	12	5			
## 2	0	0	0	0			
## 3	0	0	0	0			
## 4	0	0	1	1			
## 5	0	0	0	0			
## 6	0	0	0	0			
##	25-MDcol036-1A	27-MDcol025-1B	28-7A-1-1	29-7A	3-MDcol002-5-1	30-8B	31-8B
## 1	1	2	11	7	3	4	19
## 2	0	0	0	0	0	0	0
## 3	0	0	0	0	0	0	0
## 4	0	3	0	0	0	0	1
## 5	0	0	0	0	0	0	0
## 6	0	0	0	0	0	0	0
##	33-6-2	36-40-1B	38-S-Swanson	40-MDcol002-7-43	42-51-1B	45-MDCOL005-1B	
## 1	7	8	10	2	2	0	
## 2	0	0	0	0	0	0	
## 3	0	0	0	0	0	0	
## 4	0	0	0	3	0	0	
## 5	0	0	0	0	0	0	
## 6	0	0	0	1	0	2	
##	46-MDCOL015-1a	48-MDCOL039-1a	5-MDCOL055-Stool-1A-Baseline-BSB-2-4-1-2				
## 1	4	8				4	
## 2	0	0				0	
## 3	0	1				0	
## 4	3	8				0	
## 5	0	0				0	
## 6	0	0				0	
##	50-MDCOL027-1a	52-MDCOL021-1b	54-MDCOL028-1b	60-MDcol006-4-1A	63-MDcol003-3-2		
## 1	55	2	0	4	0		
## 2	1	40	0	0	0		
## 3	0	0	0	0	0		
## 4	5	10	0	0	0		
## 5	0	0	0	0	0		
## 6	0	0	0	0	0		
##	65-MDCOL003-Post-Chemo-Collection-Cycle-1-A	66-MDCOL004-1A					
## 1		2	0				
## 2		0	1				
## 3		0	0				
## 4		7	0				

## 5		1	0
## 6		5	0
##	68-MDCOL003-3-1-Before-Chemo-2-3A	7-MDCOL058-Stool-1A-Baseline-BSB-2-4-1-3	
## 1		1	4
## 2		0	1
## 3		0	0
## 4		16	0
## 5		1	0
## 6		0	0
##	70-MDCOL004-3-2-Post-Chemo-A	72-MDCOL003-4-1-4A-Before-Chemo-6	
## 1		0	0
## 2		0	0
## 3		0	0
## 4		0	19
## 5		0	0
## 6		0	0
##	75-MDCOL053-1-1-1-Unknown	9-MDCOL062-Stool-1A-Baseline-BSB-2-4-1-3	
## 1		24	18
## 2		0	0
## 3		0	0
## 4		11	4
## 5		0	0
## 6		0	0

Feature.ID

1 40520
2 418240
3 2479767
4 89014
5 33035
6 1532

##

1 d__Bacteria;k__Bacteria;p__Bacillota;c__Clostridia;o__Lachnospirales;f__Lachnospiraceae
2 d__Bacteria;k__Bacteria;p__Bacillota;c__Clostridia;o__Lachnospirales;f__Lachnospiraceae
3 d__Bacteria;k__Bacteria;p__Bacillota;c__Clostridia;o__Lachnospirales;f__Lachnospiraceae
4 d__Bacteria;k__Bacteria;p__Bacillota;c__Clostridia;o__Lachnospirales;f__Lachnospiraceae
5 d__Bacteria;k__Bacteria;p__Bacillota;c__Clostridia;o__Lachnospirales;f__Lachnospiraceae
6 d__Bacteria;k__Bacteria;p__Bacillota;c__Clostridia;o__Lachnospirales;f__Lachnospiraceae

##	Domain	Kingdom	Phylum	Class
## 1	Bacteria	Bacteria	Bacillota	Erysipelotrichia
## 2	Bacteria	Bacteria	Bacillota	Erysipelotrichia
## 3	Bacteria	Bacteria	Bacillota	Bacilli
## 4	Bacteria	Bacteria	Bacillota	Clostridia
## 5	Bacteria	Bacteria	Pseudomonadota	Gammaproteobacteria
## 6	Viruses	Heunggongvirae	Uroviricota	Caudoviricetes
##		Order		Family
## 1		Erysipelotrichales		Coprobacillaceae
## 2		Erysipelotrichales		Erysipelotrichaceae
## 3		Lactobacillales		Streptococcaceae
## 4		Eubacteriales		Clostridiaceae
## 5		Vibrionales		Vibrionaceae
## 6	containing	Aliceevansviridae	Aliceevansviridae	
##			Genus	Species

## 1	Catenibacterium	Catenibacterium mitsuokai					
## 2	Solobacterium	Solobacterium moorei					
## 3	Streptococcus	Streptococcus infantarius					
## 4	Clostridium	Clostridium sp. SY8519					
## 5	Vibrio	Vibrio penaeicida					
## 6	containing Streptococcus phage YMC-2011	Streptococcus phage YMC-2011					
##	1-MDcol002-5-2	11-MDcol002-label-12-on-bottle					
## 1	5	8					
## 2	0	0					
## 3	0	0					
## 4	0	0					
## 5	0	0					
## 6	0	0					
##	14-MDCOL003-Stool-1B-Baseline-BSB-2-4-1-4	15-MDcol004-3-1-pretreat					
## 1	0	1					
## 2	0	2					
## 3	0	0					
## 4	0	3					
## 5	0	0					
## 6	1	0					
##	17-MDCOL006-3-1A	19-MDCOL006-3-2A	21-MDCOL006-2-pre-chemo-A	23-MDcol033-1A			
## 1	9	5	12	5			
## 2	0	0	0	0			
## 3	0	0	0	0			
## 4	0	0	1	1			
## 5	0	0	0	0			
## 6	0	0	0	0			
##	25-MDcol036-1A	27-MDcol025-1B	28-7A-1-1	29-7A	3-MDcol002-5-1	30-8B	31-8B
## 1	1	2	11	7	3	4	19
## 2	0	0	0	0	0	0	0
## 3	0	0	0	0	0	0	0
## 4	0	3	0	0	0	0	1
## 5	0	0	0	0	0	0	0
## 6	0	0	0	0	0	0	0
##	33-6-2	36-40-1B	38-S-Swanson	40-MDcol002-7-43	42-51-1B	45-MDCOL005-1B	
## 1	7	8	10	2	2	0	
## 2	0	0	0	0	0	0	
## 3	0	0	0	0	0	0	
## 4	0	0	0	3	0	0	
## 5	0	0	0	0	0	0	
## 6	0	0	0	1	0	2	
##	46-MDCOL015-1a	48-MDCOL039-1a	5-MDCOL055-Stool-1A-Baseline-BSB-2-4-1-2				
## 1	4	8	4				
## 2	0	0	0				
## 3	0	1	0				
## 4	3	8	0				
## 5	0	0	0				
## 6	0	0	0				
##	50-MDCOL027-1a	52-MDCOL021-1b	54-MDCOL028-1b	60-MDcol006-4-1A	63-MDcol003-3-2		
## 1	55	2	0	4	0		
## 2	1	40	0	0	0		
## 3	0	0	0	0	0		
## 4	5	10	0	0	0		
## 5	0	0	0	0	0		

```
## 6          0          0          0          0          0
## 65-MDCOL003-Post-Chemo-Collection-Cycle-1-A 66-MDCOL004-1A
## 1          2          0
## 2          0          1
## 3          0          0
## 4          7          0
## 5          1          0
## 6          5          0
## 68-MDCOL003-3-1-Before-Chemo-2-3A 7-MDCOL058-Stool-1A-Baseline-BSB-2-4-1-3
## 1          1          4
## 2          0          1
## 3          0          0
## 4          16         0
## 5          1          0
## 6          0          0
## 70-MDCOL004-3-2-Post-Chemo-A 72-MDCOL003-4-1-4A-Before-Chemo-6
## 1          0          0
## 2          0          0
## 3          0          0
## 4          0          19
## 5          0          0
## 6          0          0
## 75-MDCOL053-1-1-1-Unknown 9-MDCOL062-Stool-1A-Baseline-BSB-2-4-1-3
## 1          24         18
## 2          0          0
## 3          0          0
## 4          11         4
## 5          0          0
## 6          0          0
```

Creating Phyloseq Objects for the Kraken2-Bracken data

The following code packages the feature table, taxonomy file, and the phylogenetic tree (obtained from kraken, see Step 5) into a single phyloseq object for the datasets (“raw”, “trimmed-filtered” etc).

```
brackentables<-dir(path="BrackenOutputs",pattern="table-bracken*") #Listing
  ↳ all the folders containing the bracken feature table
brackentaxonomy<-dir(path="BrackenOutputs",pattern="taxonomy-bracken*")
  ↳ #Listing all the folders containing the bracken taxonomy
brackentree<-dir(path="BrackenOutputs",pattern="bracken-reports*") #Listing
  ↳ all the folders containing the bracken reports and tree

dir.create("PhyloseqOutputs/BrackenPhyloseq")
counter=0
for(each in brackentables){
  name<-substring(each,15)

  ↳ table<-as.matrix(biom_data(read_biom(paste("BrackenOutputs",each,"feature-table.biom",sep="")))
  ↳ #Reading biom file

  ↳ taxonomy<-read.table(paste("BrackenOutputs/taxonomy-bracken-",name,"/taxonomy.tsv",sep=""))
  ↳ = TRUE)%>% #Reading taxonomy file
  mutate(Taxon=str_replace_all(Taxon,".__",""))%>%
```

```

    ↪ separate(Taxon, c("Domain", "Phylum", "Class", "Order", "Family", "Genus", "Species"), sep="|")
    mutate(Phylum=if_else(is.na(Phylum), Domain, Phylum),
           Class=if_else(is.na(Class), Phylum, Class),
           Order=if_else(is.na(Order), Class, Order),
           Family=if_else(is.na(Family), Order, Family),
           Genus=if_else(is.na(Genus), Family, Genus),
           Species=if_else(is.na(Species), Genus, Species)) %>%
    column_to_rownames("Feature.ID") %>%
    as.matrix()

    ↪ tree<-read_tree(paste("BrackenOutputs/bracken-reports-", name, "/output.tree", sep=""))
    ↪ #Reading the phylogenetic tree for that dataset
    counter=counter+1
    physeqBracken<-phyloseq(otu_table(table, taxa_are_rows =
    ↪ TRUE), tax_table(taxonomy)) #creating the phyloseq object
    physeqBracken<-merge_phyloseq(physeqBracken, tree) #Adding the phylogenetic
    ↪ tree to the phyloseq object

    ↪ saveRDS(physeqBracken, paste("PhyloseqOutputs/BrackenPhyloseq/phyloseq_bracken-", name,
    ↪ #Saving it as an .RDS file

}

```

Summary of the phyloseq object generated from the Kraken2-Bracken data

```

##      Length      Class      Mode
##           1 phyloseq         S4

## phyloseq-class experiment-level object
## otu_table()      OTU Table:         [ 812 taxa and 37 samples ]
## tax_table()      Taxonomy Table:    [ 812 taxa by 7 taxonomic ranks ]
## phy_tree()       Phylogenetic Tree: [ 812 tips and 234 internal nodes ]

```

10. Transforming Metaphlan Data and creating phyloesq objects from it

Since, MetaPhlan data does not have the regular feature table and taxonomy format as Kraken-2 or Kaiju, we will have to transform the data to make it fit that format. Also, we will use this tree generated from all the species level features in ChocoPhlan found [here](#).

```

MetaphlanOutputs<-dir("MetaphlanOutput") #Folder where the MetaPhlan
    ↪ outputs are stored
dir.create("PhyloseqOutputs/MetaphlanPhyloseq")
MetaphlanTree<-read_tree("20201013_mpav30_speciesMod.nwk") #The Tree made
    ↪ from ChocoPhlan database
new_names <- MetaphlanTree$tip.label
new_names<-new_names%>%
    str_replace_all("s__", "")
MetaphlanTree$tip.label<-new_names
for(folder in MetaphlanOutputs){ #Compiling all the outputs from different
    ↪ sample into one feature abundance table and one taxonomy file
    Alltaxonomy<-NULL

```

```

AllOTU<-NULL
for(file in list.files(paste("MetaphlanOutput", folder, sep="/"))){
  content<-readLines(paste("MetaphlanOutput", folder, file, sep="/"))[-c(1:4)]
  → #Parsing the MetaPhlan files
  id<-substring(basename(file), 1, nchar(basename(file))-11) #Stores the
  → Sample ID
  totalreads<-readLines(paste("MetaphlanOutput", folder, file, sep="/"))[3]
  → #Parsing the total reads for that sample
  totalreads<-as.numeric(gsub("^#(\\d+).*", "\\1", totalreads))
  table<-read.table(text=content, header=FALSE, sep="\t") %>% #Transforming
  → the Metaphlan file into feature table with taxonomy
    select(-4) %>%
    mutate(V1=str_replace_all(V1, ".__", ""),
           V1 = str_replace_all(V1, "\\|", ";"),
           reads=round(V3/100*totalreads)) %>%

    → separate(V1, c("Kingdom", "Phylum", "Class", "Order", "Family", "Genus", "Species"), sep=";",
    unique() %>%
    filter(!is.na(Species))
  taxonomy<-table[, c(1:7)] #Storing the taxonomy
  OTU<-table[, c(7, 10)] %>% #Storing the feature table
    unique() %>%
    mutate(Sample=id)
  Alltaxonomy<-rbind(Alltaxonomy, taxonomy) #Adding taxonomy extracted
  → from that sample with the rest in that dataset
  AllOTU<-rbind(AllOTU, OTU) #Adding feature table extracted from that
  → sample with the rest in that dataset
}

taxa_table<-Alltaxonomy %>% #Prepping taxatable for phyloseq
  arrange("Species") %>%
  unique()
rownames(taxa_table)<-taxa_table$Species
otu_table<-AllOTU %>% #Prepping OTU/feature table for phyloseq
  group_by(Sample, Species) %>%
  summarize(reads=sum(reads)) %>%
  pivot_wider(id_cols = "Species", values_from = "reads", names_from =
  → "Sample", values_fill = 0) %>%
  column_to_rownames("Species")
physeqMetaphlan<-phyloseq(otu_table(as.matrix(otu_table)), taxa_are_rows =
  → TRUE), tax_table(as.matrix(taxa_table)) #Creating the phyloseq object
physeqMetaphlan<-merge_phyloseq(physeqMetaphlan, phy_tree(MetaphlanTree))
  → #Adding the tree to the phyloseq object

  → saveRDS(physeqMetaphlan, paste("PhyloseqOutputs/MetaphlanPhyloseq/phyloseq_Metaphlan-",
  → #Saving the phyloseq object as an .RDS file
}

```

Sample Metaphlan output for a sample from our datasets

```

## [1] #mpa_vOct22_CHOCOPhlanSGB_202403
## [2] #/data/choudhurya/miniconda/envs/mpa/bin/metaphlan demultiplexed/trimmed-filtered-p
## [3] #10709265 reads processed
## [4] #SampleID Metaphlan_Analysis

```

```
## [5] #clade_name NCBI_tax_id relative_abundance additional_species
## [6] k__Bacteria 2 100.0
## [7] k__Bacteria|p__Firmicutes 2|1239 51.68535
## [8] k__Bacteria|p__Bacteroidetes 2|976 46.48729
## [9] k__Bacteria|p__Actinobacteria 2|201174 1.22089
## [10] k__Bacteria|p__Proteobacteria 2|1224 0.60648
## [11] k__Bacteria|p__Bacteroidetes|c__Bacteroidia 2|976|200643 46.42399
## [12] k__Bacteria|p__Firmicutes|c__Clostridia 2|1239|186801 29.73363
## [13] k__Bacteria|p__Firmicutes|c__CFGB3002 2|1239| 15.59072
## [14] k__Bacteria|p__Firmicutes|c__CFGB3012 2|1239| 2.14771
## [15] k__Bacteria|p__Firmicutes|c__Negativicutes 2|1239|909932 1.73986
## [16] k__Bacteria|p__Firmicutes|c__CFGB3005 2|1239| 1.38625
## [17] k__Bacteria|p__Actinobacteria|c__Coriobacteriia 2|201174|84998 1.22089
## [18] k__Bacteria|p__Proteobacteria|c__Betaproteobacteria 2|1224|28216 0.46794
## [19] k__Bacteria|p__Firmicutes|c__CFGB3054 2|1239| 0.25919
## [20] k__Bacteria|p__Firmicutes|c__Firmicutes_unclassified 2|1239| 0.24734
```

Sample taxonomy and feature table extracted from the MetaPhlan outputs

##	Kingdom	Phylum	Class	Order
## Phocaeicola_vulgatus	Bacteria	Bacteroidetes	Bacteroidia	Bacteroidales
## GGB9464_SGB14857	Bacteria	Firmicutes	CFGB3002	OFGB3002
## Bacteroides_uniformis	Bacteria	Bacteroidetes	Bacteroidia	Bacteroidales
## Blautia_wexlerae	Bacteria	Firmicutes	Clostridia	Eubacteriales
## Bacteroides_ovatus	Bacteria	Bacteroidetes	Bacteroidia	Bacteroidales
## Vescimonas_coprocola	Bacteria	Firmicutes	Clostridia	Eubacteriales
##	Family	Genus	Species	
## Phocaeicola_vulgatus	Bacteroidaceae	Phocaeicola	Phocaeicola_vulgatus	
## GGB9464_SGB14857	FGB3002	GGB9464	GGB9464_SGB14857	
## Bacteroides_uniformis	Bacteroidaceae	Bacteroides	Bacteroides_uniformis	
## Blautia_wexlerae	Lachnospiraceae	Blautia	Blautia_wexlerae	
## Bacteroides_ovatus	Bacteroidaceae	Bacteroides	Bacteroides_ovatus	
## Vescimonas_coprocola	Oscillospiraceae	Vescimonas	Vescimonas_coprocola	

##	1-MDcol002-5-2	11-MDcol002-label-12-on-bottle
## Adlercreutzia_equolifaciens	74804	146604
## Agathobaculum_butyriciproducens	814	0
## Alistipes_communis	24059	0
## Amedibacillus_dolichus	11373	12777
## Anaerobutyricum_soehngenii	68025	85720
## Anaerostipes_hadrus	31240	14025
##	14-MDCOL003-Stool-1B-Baseline-BSB-2-4-1-4	
## Adlercreutzia_equolifaciens		0
## Agathobaculum_butyriciproducens		0
## Alistipes_communis		0
## Amedibacillus_dolichus		0
## Anaerobutyricum_soehngenii		0
## Anaerostipes_hadrus		0
##	15-MDcol004-3-1-pretreat	17-MDCOL006-3-1A
## Adlercreutzia_equolifaciens	50268	288766
## Agathobaculum_butyriciproducens	3402	0
## Alistipes_communis	35327	0
## Amedibacillus_dolichus	0	0

Summary of the phyloseq object generated from the MetaPhlan data

11. Creating Phyloseq Objects with the HUMAnN data

```
HumannOutput <- dir("HumannOutput") #Folder containing all the HUMAnN outputs
dir.create("PhyloseqOutputs/HumannPhyloseq")
for (folder in HumannOutput) {
  AllCoverage <- data.frame()
  AllAbundance <- data.frame()
  counter = 1
  for (file in list.files(paste("HumannOutput", folder,
    sep = "/"), pattern = "*_pathabundance.tsv")) {
    id <- substring(basename(file), 1, nchar(basename(file)) -
      18) #Parsing the Sample ID
    abundance <- read.delim(paste("HumannOutput",
      folder, file, sep = "/"), sep = "\t", header = TRUE,
      comment.char = "") #Reading the pathway abundance file for a
      ↪ sample
    df <- colnames(abundance) %>%
      str_replace_all("X\\.\\.\\.\\.", "") %>%
      str_replace_all("X", "") %>%
      str_replace_all("\\.", "_") %>%
      str_replace_all("_Abundance", "")
    colnames(abundance) <- df
```

```

coverage <- read.delim(paste("HumannOutput/",
  folder, "/", id, "_pathcoverage.tsv", sep = ""),
  sep = "\t", header = TRUE, comment.char = "") #Reading the
  ↳ pathway coverage file for a sample
df <- colnames(coverage) %>%
  str_replace_all("X\\\\.\\.\\.\"", "") %>%
  str_replace_all("X", "") %>%
  str_replace_all("\\\\.\"", "-") %>%
  str_replace_all("_Coverage", "")
colnames(coverage) <- df

# Combining the pathway abundances and
# pathway coverage data with those from
# the previous samples
if (counter == 1) {
  AllAbundance <- abundance
} else {
  AllAbundance <- left_join(AllAbundance,
    abundance)
}
if (counter == 1) {
  AllCoverage <- coverage
} else {
  AllCoverage <- left_join(AllCoverage, coverage)
}
counter = counter + 1
}

AllAbundance <- AllAbundance %>%
  column_to_rownames("Pathway") %>%
  as.matrix()
AllCoverage <- AllCoverage %>%
  column_to_rownames("Pathway") %>%
  as.matrix()
phyloseqcoverage <- phyloseq(otu_table(AllCoverage,
  taxa_are_rows = TRUE)) #Creating phyloseq object for the pathway
  ↳ abundance data
physeqabundance <- phyloseq(otu_table(AllAbundance,
  taxa_are_rows = TRUE)) #Creating phyloseq object for the pathway
  ↳ coverage data
saveRDS(physeqabundance,
  ↳ paste("PhyloseqOutputs/HumannPhyloseq/phyloseq_humann-",
    folder, "_pathwayabundance.rds", sep = ""))
saveRDS(physeqcoverage,
  ↳ paste("PhyloseqOutputs/HumannPhyloseq/phyloseq_humann-",
    folder, "_pathwaycoverage.rds", sep = ""))
}

for (folder in HumannOutput) {
  AllRPK <- data.frame()
  counter = 1
  for (file in list.files(paste("HumannOutput", folder,
    sep = "/"), pattern = "*_genefamilies.tsv")) {
    id <- substring(basename(file), 1, nchar(basename(file)) -

```


## UNMAPPED	778748.2	
## UNINTEGRATED	2481983.1	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	917463.5	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	464620.0	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	354114.6	
## UNINTEGRATED unclassified	147854.1	
##	11_MDCol002_label_12_on_bottle	
## UNMAPPED	812515.6	
## UNINTEGRATED	2338562.7	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	740989.9	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	205282.2	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	248232.9	
## UNINTEGRATED unclassified	229298.6	
##	14_MDCOL003_Stool_1B_Baseline_BSB_2	
## UNMAPPED	72	
## UNINTEGRATED	31	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus		
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis		
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		
## UNINTEGRATED unclassified		30
##	15_MDcol004_3_1_pretreat	
## UNMAPPED	853278.4	
## UNINTEGRATED	2607555.3	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	187785.6	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	193568.8	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	27733.6	
## UNINTEGRATED unclassified	406532.7	
##	17_MDCOL006_3_1A	
## UNMAPPED	640092.1	
## UNINTEGRATED	2365706.0	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	255956.6	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	486901.1	
## UNINTEGRATED unclassified	176732.7	
##	19_MDCOL006_3_2A	
## UNMAPPED	728047.75	
## UNINTEGRATED	2412350.76	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	84115.85	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	78634.97	
## UNINTEGRATED unclassified	178803.49	
##	21_MDCOL006_2_pre_chemo_A	
## UNMAPPED	1170228.6	
## UNINTEGRATED	4808488.1	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	240857.1	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	828564.4	
## UNINTEGRATED unclassified	532576.9	
##	23_MDcol033_1A	
## UNMAPPED	691496.1	
## UNINTEGRATED	2425535.6	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	159044.4	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	178257.6	

## UNINTEGRATED unclassified	141054.7	
##	25_MDcol036_1A	
## UNMAPPED	1068527.81	
## UNINTEGRATED	4715990.30	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	665352.47	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	85896.49	
## UNINTEGRATED unclassified	245014.75	
##	27_MDcol025_1B	28_7A_1_1
## UNMAPPED	945989.87	902644.2
## UNINTEGRATED	3843549.24	2664917.9
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	313605.33	665346.6
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	271299.80	716802.5
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	59771.61	354245.5
## UNINTEGRATED unclassified	191873.16	216273.1
##	29_7A	3_MDcol002_5_1
## UNMAPPED	630687.3	1054449.8
## UNINTEGRATED	2027313.2	2771711.1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	617776.9	502390.7
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	642820.4	658787.2
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	274152.5	399946.3
## UNINTEGRATED unclassified	150555.6	308745.2
##	30_8B	31_8B
## UNMAPPED	364662.41	860531.46
## UNINTEGRATED	1176143.90	1898317.92
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	265896.80	62848.75
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	357642.20	259350.24
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	154126.96	113944.27
## UNINTEGRATED unclassified	65569.98	186109.70
##	33_6_2	36_40_1B
## UNMAPPED	683345.04	1168856.3
## UNINTEGRATED	1265336.05	3293921.9
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	50452.27	896384.5
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	136902.02	369106.3
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	64743.90	644273.4
## UNINTEGRATED unclassified	150448.35	296277.0
##	38_S_Swanson	
## UNMAPPED	1317047.4	
## UNINTEGRATED	2714109.3	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	125167.7	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	NA	
## UNINTEGRATED unclassified	524893.7	
##	40_MDCol002_7_43	42_51_1B
## UNMAPPED	1234938.85	795759.2
## UNINTEGRATED	3188195.55	2269103.2
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	1000793.98	961751.5
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	NA	331637.7
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	77151.05	257447.8
## UNINTEGRATED unclassified	336153.66	229043.3
##	45_MDCOL005_1B	
## UNMAPPED	1246331.44	
## UNINTEGRATED	6680423.14	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	597264.54	

## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	68584.03	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	NA	
## UNINTEGRATED unclassified	243317.20	
##	46_MDCOL015_1a	
## UNMAPPED	911520.5	
## UNINTEGRATED	2033420.0	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	NA	
## UNINTEGRATED unclassified	300806.1	
##	48_MDCOL039_1a	
## UNMAPPED	994944.84	
## UNINTEGRATED	3228306.39	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	305187.68	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	405561.98	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	56772.45	
## UNINTEGRATED unclassified	243334.71	
##	5_MDCOL055_Stool_1A_Baseline_BSB_2	
## UNMAPPED	4334	
## UNINTEGRATED	14783	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	3570	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	2953	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	4328	
## UNINTEGRATED unclassified	810	
##	50_MDCOL027_1a	
## UNMAPPED	904651.65	
## UNINTEGRATED	2385396.13	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	40831.38	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	26875.49	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	12807.89	
## UNINTEGRATED unclassified	324132.36	
##	52_MDCOL021_1b	
## UNMAPPED	608032.696	
## UNINTEGRATED	2086633.441	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	6378.159	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	6021.116	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	14722.860	
## UNINTEGRATED unclassified	173575.249	
##	54_MDCOL028_1b	
## UNMAPPED	940612.29	
## UNINTEGRATED	5641354.19	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	NA	
## UNINTEGRATED unclassified	96868.25	
##	60_MDcol006_4_1A	
## UNMAPPED	709619.7	
## UNINTEGRATED	2663982.1	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	1151220.5	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	371673.6	
## UNINTEGRATED unclassified	208778.9	
##	63_MDcol003_3_2	
## UNMAPPED	552487.6	

## UNINTEGRATED	3076573.7	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	NA	
## UNINTEGRATED unclassified	199483.9	
##	65_MDCOL003_Post_Chemo_Collection_	
## UNMAPPED		
## UNINTEGRATED		
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus		
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis		
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		
## UNINTEGRATED unclassified		
##	66_MDCOL004_1A	
## UNMAPPED	102994.388	
## UNINTEGRATED	211972.628	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	24242.876	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	18036.559	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	6011.369	
## UNINTEGRATED unclassified	31441.128	
##	68_MDCOL003_3_1_Before_Chemo_2_3A	
## UNMAPPED	575372.3	
## UNINTEGRATED	2622809.6	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	NA	
## UNINTEGRATED unclassified	221928.5	
##	7_MDCOL058_Stool_1A_Baseline_BSB_2	
## UNMAPPED	328	
## UNINTEGRATED	689	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	146	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	47	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	1102	
## UNINTEGRATED unclassified	912	
##	70_MDCOL004_3_2_Post_Chemo_A	
## UNMAPPED	34068.790	
## UNINTEGRATED	39832.621	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	6643.549	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	4221.045	
## UNINTEGRATED unclassified	8543.140	
##	72_MDCOL003_4_1_4A_Before_Chemo_6	
## UNMAPPED	461003.24	
## UNINTEGRATED	1808251.90	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	186720.43	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	26477.84	
## UNINTEGRATED unclassified	104787.82	
##	75_MDCOL053_1_1_1_Unknown	
## UNMAPPED	595072.92	
## UNINTEGRATED	1816052.92	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_vulgatus	49070.33	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_uniformis	11643.02	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	26855.97	
## UNINTEGRATED unclassified	223668.29	

```

##
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_vulgatus
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_uniformis
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
## UNINTEGRATED|unclassified

```

Sample pathway coverage file

```

##
## 1
## 2
## 3 UNINTEGRATED|g__Agathobaculum.s__Agathobaculum_butyriciproducens
## 4 UNINTEGRATED|g__Akkermansia.s__Akkermansia_muciniphila
## 5 UNINTEGRATED|g__Alistipes.s__Alistipes_finegoldii
## 6 UNINTEGRATED|g__Alistipes.s__Alistipes_onderdonkii
## 9-MDCOL062-Stool-1A-Baseline-BSB-2-4-1-3
## 1 1
## 2 1
## 3 1
## 4 1
## 5 1
## 6 1

```

Combined pathway coverage of all samples in a dataset

```

##
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
##
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
##
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
##
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus

```

## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1
##	17-MDCOL006-3-1A	
## UNMAPPED		1
## UNINTEGRATED		1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1
##	19-MDCOL006-3-2A	
## UNMAPPED		1
## UNINTEGRATED		1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1
##	21-MDCOL006-2-pre-chemo-A	
## UNMAPPED		1
## UNINTEGRATED		1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1
##	23-MDcol033-1A	
## UNMAPPED		1
## UNINTEGRATED		1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		NA
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		NA
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1
##	25-MDcol036-1A	
## UNMAPPED		1
## UNINTEGRATED		1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		NA
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		NA
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1
##	27-MDcol025-1B	
## UNMAPPED		1
## UNINTEGRATED		1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1
##	28-7A-1-1 29-7A	
## UNMAPPED		1 1
## UNINTEGRATED		1 1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		1 1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		1 1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		1 1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1 1
##	3-MDcol002-5-1	
## UNMAPPED		1
## UNINTEGRATED		1

## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens			1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus			1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus			1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus			1
##	30-8B	31-8B	33-6-2
## UNMAPPED	1	1	1
## UNINTEGRATED	1	1	1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	1	1	1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	1	1	1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	1	1	1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	1	1	1
##	36-40-1B		
## UNMAPPED		1	
## UNINTEGRATED		1	
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		1	
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		1	
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		1	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1	
##	38-S-Swanson		
## UNMAPPED		1	
## UNINTEGRATED		1	
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		1	
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		1	
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		1	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		NA	
##	40-MDCol002-7-43		
## UNMAPPED			1
## UNINTEGRATED			1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens			1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus			1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus			1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus			1
##	42-51-1B		
## UNMAPPED		1	
## UNINTEGRATED		1	
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens		NA	
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus		NA	
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus		NA	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus		1	
##	45-MDCOL005-1B		
## UNMAPPED			1
## UNINTEGRATED			1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens			1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus			NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus			1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus			NA
##	46-MDCOL015-1a		
## UNMAPPED			1
## UNINTEGRATED			1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens			NA
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus			1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus			NA
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus			NA
##	48-MDCOL039-1a		

## UNMAPPED	1
## UNINTEGRATED	1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	1
##	5-MDCOL055-Stool-1A-Baseli
## UNMAPPED	
## UNINTEGRATED	
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	
##	50-MDCOL027-1a
## UNMAPPED	1
## UNINTEGRATED	1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	1
##	52-MDCOL021-1b
## UNMAPPED	1
## UNINTEGRATED	1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	NA
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	NA
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	1
##	54-MDCOL028-1b
## UNMAPPED	1
## UNINTEGRATED	1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	NA
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	NA
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	NA
##	60-MDcol006-4-1A
## UNMAPPED	1
## UNINTEGRATED	1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	1
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	1
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	1
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	1
##	63-MDcol003-3-2
## UNMAPPED	1
## UNINTEGRATED	1
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	NA
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	NA
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	NA
## UNINTEGRATED g__Bacteroides.s__Bacteroides_ovatus	NA
##	65-MDCOL003-Post-Chemo-Col
## UNMAPPED	
## UNINTEGRATED	
## UNINTEGRATED g__Adlercreutzia.s__Adlercreutzia_equolifaciens	
## UNINTEGRATED g__Anaerostipes.s__Anaerostipes_hadrus	
## UNINTEGRATED g__Asaccharobacter.s__Asaccharobacter_celatus	


```

## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
## 66-MDCOL004-1A
## UNMAPPED 1
## UNINTEGRATED 1
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens NA
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus NA
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus NA
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus 1
## 68-MDCOL003-3-1-Before-Cher
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
## 7-MDCOL058-Stool-1A-Baselin
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
## 70-MDCOL004-3-2-Post-Chemo-
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens M
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus M
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus M
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
## 72-MDCOL003-4-1-4A-Before-C
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus
## 75-MDCOL053-1-1-1-Unknown
## UNMAPPED 1
## UNINTEGRATED 1
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens NA
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus 1
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus NA
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus 1
## 9-MDCOL062-Stool-1A-Baselin
## UNMAPPED
## UNINTEGRATED
## UNINTEGRATED|g__Adlercreutzia.s__Adlercreutzia_equolifaciens
## UNINTEGRATED|g__Anaerostipes.s__Anaerostipes_hadrus
## UNINTEGRATED|g__Asaccharobacter.s__Asaccharobacter_celatus
## UNINTEGRATED|g__Bacteroides.s__Bacteroides_ovatus

```

Sample gene family abundance file

```
## Gene-Family
```

```

## 1                                UNMAPPED
## 2                                UniRef90_A5ZYV3
## 3                                UniRef90_A5ZYV3|g__Blautia.s__Blautia_obeum
## 4                                UniRef90_U3JBQ2
## 5 UniRef90_U3JBQ2|g__Eggerthella.s__Eggerthella_lenta
## 6                                UniRef90_A0A174PA28
## 9-MDCOL062-Stool-1A-Baseline-BSB-2-4-1-3
## 1                                1987609.00
## 2                                19139.59
## 3                                19139.59
## 4                                15087.70
## 5                                15087.70
## 6                                13552.05

```

Combined gene family abundances of all samples in a dataset

```

##                                1-MDcol002-5-2
## UNMAPPED                                2364890.000
## UniRef90_A0A174NA57                                55247.258
## UniRef90_A0A174NA57|g__Bacteroides.s__Bacteroides_uniformis 55247.258
## UniRef90_A0A395VRV3                                49394.382
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_ovatus 25032.358
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_vulgatus 8720.539
##                                11-MDCol002-label-12-on-bott
## UNMAPPED                                2374486.0
## UniRef90_A0A174NA57                                43582.0
## UniRef90_A0A174NA57|g__Bacteroides.s__Bacteroides_uniformis 43582.0
## UniRef90_A0A395VRV3                                31559.4
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_ovatus 7825.7
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_vulgatus 5414.0
##                                14-MDCOL003-Stool-1B-Baselin
## UNMAPPED
## UniRef90_A0A174NA57
## UniRef90_A0A174NA57|g__Bacteroides.s__Bacteroides_uniformis
## UniRef90_A0A395VRV3
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_ovatus
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_vulgatus
##                                15-MDcol004-3-1-pretreat
## UNMAPPED                                2462301.0000
## UniRef90_A0A174NA57                                12391.8543
## UniRef90_A0A174NA57|g__Bacteroides.s__Bacteroides_uniformis 12391.8543
## UniRef90_A0A395VRV3                                6052.5065
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_ovatus 2456.2799
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_vulgatus 823.7582
##                                17-MDCOL006-3-1A
## UNMAPPED                                1570111.000
## UniRef90_A0A174NA57                                NA
## UniRef90_A0A174NA57|g__Bacteroides.s__Bacteroides_uniformis NA
## UniRef90_A0A395VRV3                                5697.658
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_ovatus NA
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_vulgatus NA
##                                19-MDCOL006-3-2A
## UNMAPPED                                1791726.000
## UniRef90_A0A174NA57                                NA

```

## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	629.729
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	NA
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	NA
##	21-MDCOL006-2-pre-chemo-A
## UNMAPPED	2.798869e+06
## UniRef90_A0A174NA57	NA
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	1.730327e+04
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	7.697973e+00
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	NA
##	23-MDcol033-1A
## UNMAPPED	1.816065e+06
## UniRef90_A0A174NA57	1.139728e+01
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	1.754432e+04
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	8.162051e+03
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	NA
##	25-MDcol036-1A
## UNMAPPED	2.876898e+06
## UniRef90_A0A174NA57	3.411955e+02
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	1.683431e+04
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	1.683046e+04
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	4.744231e-01
##	27-MDcol025-1B
## UNMAPPED	2.535351e+06
## UniRef90_A0A174NA57	9.803922e+00
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	6.165756e+03
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	6.164478e+03
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	6.428122e-01
##	28-7A-1-1
## UNMAPPED	2736361.000
## UniRef90_A0A174NA57	74789.702
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	74789.702
## UniRef90_A0A395VRV3	34024.889
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	17735.630
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	5423.871
##	29-7A
## UNMAPPED	1980954.000
## UniRef90_A0A174NA57	46810.359
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	46810.359
## UniRef90_A0A395VRV3	29657.473
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	15926.570
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	4748.206
##	3-MDcol002-5-1
## UNMAPPED	3126697.000
## UniRef90_A0A174NA57	60087.977
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	60087.977
## UniRef90_A0A395VRV3	38175.660
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	9597.700
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	6431.994
##	30-8B

## UNMAPPED	1105105.000
## UniRef90_A0A174NA57	25387.917
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	25387.917
## UniRef90_A0A395VRV3	21139.826
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	10124.156
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	2733.621
##	31-8B
## UNMAPPED	2514157.000
## UniRef90_A0A174NA57	21054.698
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	21054.698
## UniRef90_A0A395VRV3	14503.126
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	6558.171
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	2147.991
##	33-6-2
## UNMAPPED	1998210.000
## UniRef90_A0A174NA57	14323.268
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	14323.268
## UniRef90_A0A395VRV3	6942.182
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	3888.373
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	1008.496
##	36-40-1B
## UNMAPPED	3434720.00
## UniRef90_A0A174NA57	78682.87
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	78682.87
## UniRef90_A0A395VRV3	56618.00
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	13359.20
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	10429.88
##	38-S-Swanson
## UNMAPPED	3.951244e+06
## UniRef90_A0A174NA57	4.437334e+01
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	3.114029e+03
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	NA
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	1.426303e+03
##	40-MDCol002-7-43
## UNMAPPED	3609851.0000
## UniRef90_A0A174NA57	478.7466
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	41862.4500
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	22803.9210
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	9008.1536
##	42-51-1B
## UNMAPPED	2458377.000
## UniRef90_A0A174NA57	39347.805
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	39347.805
## UniRef90_A0A395VRV3	45337.774
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	24467.937
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	6390.123
##	45-MDCOL005-1B
## UNMAPPED	2.619226e+06
## UniRef90_A0A174NA57	NA
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	1.742780e+04
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	NA

## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	1.037072e+01
##	46-MDCOL015-1a
## UNMAPPED	2508626
## UniRef90_A0A174NA57	NA
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	NA
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	NA
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	NA
##	48-MDCOL039-1a
## UNMAPPED	2.862189e+06
## UniRef90_A0A174NA57	1.139754e+00
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	2.077782e+03
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	1.541726e+03
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	1.107623e+02
##	5-MDCOL055-Stool-1A-Baseline
## UNMAPPED	
## UniRef90_A0A174NA57	
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	
## UniRef90_A0A395VRV3	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	
##	50-MDCOL027-1a
## UNMAPPED	2.569471e+06
## UniRef90_A0A174NA57	NA
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	8.392341e+02
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	8.490451e+01
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	1.747656e+02
##	52-MDCOL021-1b
## UNMAPPED	1332322.0000
## UniRef90_A0A174NA57	NA
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	811.9280
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	NA
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	259.6574
##	54-MDCOL028-1b
## UNMAPPED	1.875517e+06
## UniRef90_A0A174NA57	NA
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	1.006431e+01
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	NA
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	NA
##	60-MDcol006-4-1A
## UNMAPPED	2136531.000
## UniRef90_A0A174NA57	NA
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA
## UniRef90_A0A395VRV3	7466.442
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	NA
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	NA
##	63-MDcol003-3-2
## UNMAPPED	1172962.000
## UniRef90_A0A174NA57	NA
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	NA

## UniRef90_A0A395VRV3	1801.322
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	NA
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	NA
##	65-MDCOL003-Post-Chemo-Coll
## UNMAPPED	
## UniRef90_A0A174NA57	
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	
## UniRef90_A0A395VRV3	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	
##	66-MDCOL004-1A
## UNMAPPED	274343.0000
## UniRef90_A0A174NA57	2498.5665
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	2498.5665
## UniRef90_A0A395VRV3	1859.1191
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	909.3233
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	236.6618
##	68-MDCOL003-3-1-Before-Chemo
## UNMAPPED	12
## UniRef90_A0A174NA57	
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	
## UniRef90_A0A395VRV3	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	
##	7-MDCOL058-Stool-1A-Baseline
## UNMAPPED	
## UniRef90_A0A174NA57	
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	
## UniRef90_A0A395VRV3	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	
##	70-MDCOL004-3-2-Post-Chemo-A
## UNMAPPED	97034.0000
## UniRef90_A0A174NA57	1436.5401
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	1436.5401
## UniRef90_A0A395VRV3	225.2466
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	102.7223
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	49.8463
##	72-MDCOL003-4-1-4A-Before-Cl
## UNMAPPED	11706
## UniRef90_A0A174NA57	18
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	18
## UniRef90_A0A395VRV3	40
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	
##	75-MDCOL053-1-1-1-Unknown
## UNMAPPED	1578382.0000
## UniRef90_A0A174NA57	1906.2450
## UniRef90_A0A174NA57 g__Bacteroides.s__Bacteroides_uniformis	1906.2450
## UniRef90_A0A395VRV3	1064.6096
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_ovatus	171.7574
## UniRef90_A0A395VRV3 g__Bacteroides.s__Bacteroides_vulgatus	255.1828
##	9-MDCOL062-Stool-1A-Baseline
## UNMAPPED	

```
## UniRef90_A0A174NA57
## UniRef90_A0A174NA57|g__Bacteroides.s__Bacteroides_uniformis
## UniRef90_A0A395VRV3
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_ovatus
## UniRef90_A0A395VRV3|g__Bacteroides.s__Bacteroides_vulgatus
```