Free radical scavenging activity screening of medicinal plants from Tripura, Northeast India

Rajendra Kshirsagar^{1*} and Shakti Upadhyay²

¹Drug Discovery & Development, Reliance Pharmaceuticals Pvt Ltd, Dhirubhai Ambani Life Sciences Center R-282, TTC Industrial Area of MIDC, Thane, Belapur Road, Rabale, Navi Mumbai - 400 701, Maharashtra, India ²35/1002, Seawood Estate, Palmbeach Marg, Nerul, Navi Mumbai - 400 706 *Correspondent author, E-mail: rajendra_kshirsagar@relbio.com; Fax No.: +9122-27068499 **Received 17 March 2008; Accepted 5 January 2009

Abstract

Antioxidative effects of 123 extracts (Direct methanolic and sequential per ether, dichloromethane, ethyl acetate, methanol) prepared from 32 plants species (59 plant samples) collected from Tripura, Northeast India have been studied. Their ability of scavenging free radicals was measured by DPPH reduction spectrophotometric assay. Sixteen extracts showed strong antioxidant capabilities, which were, subjected for their dose dependent activity at different concentrations to calculate ${\rm IC}_{50}$ values.

Keywords: Antioxidant, DPPH, Free radical scavenging activity, Medicinal plants, Northeast India, Tripura.

IPC code; **Int. cl.**⁸ — A61K 36/00, A61P 17/18

Introduction

Antioxidants, which scavenge active oxygen species (free radicals), are found in a variety of foodstuffs and are commonly referred to as scavengers. Many antioxidants are plant based and play an important role in protecting plants that are exposed to strong sunlight and live under severe oxygen stress. Antioxidants also play an important role in human health because the biologic defense mechanisms cannot operate under severe oxygen stress. According to recent research, activated oxygen is thought to be a major factor in ageing, hardening of the arteries, diabetes, cancer and tissue injury of skin. Indeed approximately 90% of age-related diseases are linked to activated oxygen. When human skin is exposed to ultraviolet rays active oxygen (free radical) is generated, which is

scavenged by excess melanin. Pigmentation from excess melanin can cause the appearance of spots and freckles on the skin¹. Plants are rich source of antioxidants, which is evinced through many reports on medicinal plants with antioxidant potential²⁻⁵. Almost all the sources for such phytochemical studies are from published or unpublished ethnobotanical knowledge⁶⁻⁹.

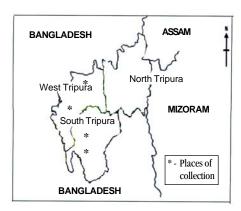
Study area

Tripura is endowed with vast natural resources. The State has a total area of 10,169 sq. km out of which 6,168 sq. km is occupied by forest (exclusive of tea gardens). Geographically, it lies in a strategic zone as it falls in between the Indo-Malayan and Indo-Chinese biological realms. Thus, Tripura stands at the gateway to floral and faunal confluence. Tripura

also lies in bio-geographical zone of nine big North East hills. Tripura occupies 0.32 per cent area of India and accounts for 12.78 per cent of the plant resources species found in the country. Tripura has been listed as one of the 26 endemic centres in India. Scientific studies have revealed that it possesses 1545 plant species with 28 varieties, 379 tree species, 320 shrubs, 581 herbs, 165 climbers, 34 ferns, 45 epiphytes, four parasites and 16 climbing shrubs out of which seven are endangered, seven are endemic and 18 rare species. The State also has 24 species of orchids and 266 species of medicinal plants. Scientific study has also shown that the Maximum Plant Diversity Index lies at 5.23, one of the highest in India. Rare species like *Angiopteris erecta* Desv., a tree fern and Gnetum montanum Markgraf, a climbing gymnosperm occupy a key position. At present, most of the wild species have been confined to the sanctuaries due to habitat destruction. Tripura has 603.65 sq. km of forests within the four sanctuaries—Sipahijala (18.4 sq. km), *Gumti* (389.54 sq. km), Trishna (194.71 sq. km) and Roa (0.86 sq. km). The forest density of Tripura in terms of percentage is higher than the national figure. It is 17.35 per cent, the national figure being 11.73 per cent. The

Vol 8(2) March-April 2009______

Research Paper



Map of Tripura showing places of collection

percentage wise figure of open forests lies at 35.4 per cent, which is greater than the national figure of 7.61 per cent. Overall, 52.79 per cent of the total land is covered by forest whereas the national figure stands at 19.47 per cent. The forests of Tripura have been classified as

evergreen and deciduous. The evergreen type has been diminishing¹⁰.

Materials and Methods Plant material

Botanical species or allied species of reported medicinal plants were collected from West and South Tripura districts of Tripura State (see Map), Northeast India during May 2005. While collecting samples the herbarium voucher specimens were also collected for future reference. Mounted herbarium specimens with proper labels and accession numbers were authenticated by using Flora of Tripura, Manipur, Mizoram¹¹⁻¹³ and deposited at Reliance herbarium of Pharmaceuticals Pvt. Ltd.



Aquillaria malaccensis



Grewia sapida



Pterospermum semisagittatum



Litsea glutinosa







Syzygium cerasoides

Dhirubhai Ambani Life Sciences Center, Navi Mumbai. Collected samples were chopped in small pieces, shade dried and pulverized to coarse powder (10 mm sieve size)¹⁴. The plant species with family, local names, accession number, plant part/s studied and percentage antioxidant activity in direct methanol extract is detailed in Table 1.

Table 1: Antioxidant (AO) activity for direct methanol extracts of plant samples

S. No.	Name of the species/Family/ Acc. No.	Local name	Plant part/s	AO activity (%)
1.	Alocasia fornicata (Roxb.)	Baibing	Rhizome	41.06
	Schott/ Araceae/ RLS-12&13		Aerial part	21.63
2.	Alpinia malaccensis Rosc. Zingiberaceae/ RLS-5	Aiphal	Aerial part	5.69
3.	Alpinia officinarum Hance	Aichal	Rhizome	94.02
	Zingiberaceae/ RLS-3&4		Aerial part	74.97
4.	Aquilaria malaccensis Lam.	Agar	Stem	43.30
	Thymelaeaceae/ RLS-44&45		Aerial part	92.03
5.	Artocarpus chaplasha Roxb.	Harikothong	Stem	47.70
	Moraceae/ RLS-30		Leaf	76.94
6.	Callicarpa arborea Roxb.	Hnahkiah	Stem	53.65
	Verbenaceae/ RLS-22		Leaf	47.20
7.	Cassia nodosa BuchHam. ex Roxb.	Tisibi	Stem	78.96
	Caesalpiniaceae/ RLS-31		Twig	71.44
8.	Cassia renigera Wall. ex Benth.	Radhachura	Stem	36.11
	Caesalpiniaceae/ RLS-52		Twig	17.65
9.	Clerodendrum indicum (Linn.) Kuntze	Kuthap	Aerial part	47.07
	Verbenaceae/ RLS-8&9		Root	1.0
10.	Dalbergia volubilis Roxb.	Dadbari	Stem	86.44
	Fabaceae/RLS-36&37		Twig	88.93
11.	Diplazium polypodioides Blume	Chakawkei	Aerial part	5.87
	Athyriaceae/ RLS-24	-chii		

118 Natural Product Radiance

S. No.	Name of the species/Family/ Acc. No.	Local name	Plant part/s	AO activity (
12.	Dipterocarpus turbinatus Gaertn. f.	Garjan	Stem	75.86
	Dipterocarpaceae/ RLS-1&2		Leaf	10.76
		Fruit	Fruit	97.26
13.	Garuga pinnata Roxb.	Kanjikara	Stem	87.56
	Burseraceae/ RLS-21	ľ	Leaf	65.01
14.	Grewia nervosa (Lour.) Panigrahi	Yongkomla	Stem	67.09
	Tiliaceae/ RLS-9&10		Twig	89.51
5.	Grewia sapida Roxb. ex DC.	Yongkomla	Stem	95.46
	Tiliaceae/ RLS-38&39		Twig	97.01
6.	Hydnocarpus kurzii (King) Warb.	Khavitur	Stem	45.96
	Flacourtiaceae/ RLS-50&51		Twig	53.27
			Fruit	47.95
7.	Litsea glutinosa (Lour.) C.B. Robinson	Kalimendi	Stem	90.57
	Lauraceae/ RLS-32&33		Twig	41.53
.8.	Mallotus tetracoccus (Roxb.) Kurz	Thingkhei	Stem	23.02
	Euphorbiaceae/ RLS-16&17		Leaf	31.42
9.	Melastoma malabathricum Linn.	Builukham	Aerial part	36.58
.,.	Melastomataceae/ RLS-40		, recruit pure	30.30
80.	Mitragyna rotundifolia (Roxb.)	Viteaval	Stem	91.51
	O. Kuntze Rubiaceae/ RLS-15	Viicavai	Twig	93.58
1.	Murraya koenigii (Linn.) Spreng.	Karipatta	Aerial part	41.75
	Rutaceae/ RLS-79	Raripana	neriai part	11.//
2.	Persicaria hydropiper (Linn.) Opiz	Leipung	Whole plant	46.68
14.	syn. Polygonum hydropiper Linn.	Leipung	whole plant	40.06
	Polygonaceae/ RLS-19			
3.	Phlogacanthus thyrsiflorus Nees	Nongmang	Stem	33.70
ال.	Acanthaceae/ RLS-48&49	-kha	Twig	14.43
4.	Psidium guineense Sw.	Sibiki	Aerial part	67.34
4.	Myrtaceae/ RLS-06	Sibiki	Aeriai part	07.34
5.	Pterospermum semisagittatum Buch	Bandarhola	Stem	96.99
ار.	Ham. ex Roxb./Sterculiaceae/ RLS-46&47	Банаатнога	Twig	50.25
6.	Saraca asoca (Roxb.) de Wilde	Malhawih	Stem	95.52
0.	Caesalpiniaceae/ RLS-26&27	mamawin	Leaf	79.50
7.	Schima wallichii (DC.) Korth.	Kanak	Stem	96.46
	Theaceae/ RLS-29	Runuk	Leaf	96.72
8.	Spilanthes paniculata Wall. ex DC.	Athlo	Whole plant	23.37
_0.	Asteraceae/ RLS-25	1111110	"Hote plant	23.37
9.	Sterculia villosa Roxb.	Udal	Stem	42.70
⊿ ∫.	Sterculiaceae/ RLS-28	Kanak	Leaf	24.62
0.	Syzygium cerasoides (Roxb.) Raiz.	Botijam	Stem	93.60
, , ,	Myrtaceae/ RLS-34&35		Leaf	94.65
1.	Vitex peduncularis Wall. ex Schauer.	Awal	Stem	24.47
	Verbenaceae/ RLS-13&14		Twig	24.96
32.	Wendlandia wallichii Wight & Arn.	Lengpat	Stem	96.99
	Rubiaceae/ RLS-42&43		Twig	95.71
	Standard-Curcumin			99.48
	Standard-Catechin			99.47
	Standard-Trolox			100

Chemicals

Pet ether, dichloro methane, ethyl acetate, methanol were purchased from Thomas baker, DPPH, catechin, curcumin, trolox from Sigma Aldrich Inc. and DMSO from s. d. fine-chem Ltd.

Extraction and evaluation of extracts

Powdered samples were subjected to cold extraction by using methanol. The samples were charged in percolators and solvent is added to it in 1:6 proportions at room temperature with intermittent agitation and filtered. The process repeated for thrice and accumulated solution from methanol was concentrated to dryness under reduced pressure and controlled temperature (42-45°C) using rotary evaporator. Then methanol extracts were evaluated in DPPH assay for antioxidative property as has been evinced in Table 1.

Antioxidant screening

One mg of sample was dissolved in 100 ml of DMSO and methanol was added to it to make concentration of 1 mg/ml. 50 µl of sample solution was taken in a micro titer plate and 200 µl of DPPH solution (10 mM concentration of 2, 2-diphenyl-1-picryl-hydrazyl prepared in methanol AR grade) was added to it. The plate was incubated in dark for half an hour. Methanol blank (250µl) and DPPH blank

Research Paper

(200 μl DPPH + 50 μl methanol) was used for calculating the percentage antioxidant activity. Absorbance measured at 540 on Eliza plate reader. Antioxidant activity in percentage was calculated by the formula: 1-(Absorbance of sample/Absorbance of DPPH) × 100 (Ref. 15). Dose dependent study was carried out by same protocol but with following concentrations; 1, 0.5, 0.25, 0.12 and 0.062 mg/ml.

Results

The antioxidant activity of various plants species in direct methanol extract is given in Table 1.

Only those crude samples, which had shown more than 90% antioxidant activity in direct methanol extract, were sequentially extracted. Solvents used for sequential extraction were, petroleum ether (60-80), dichloro methane, ethyl acetate and methanol. The process of extraction followed for all the extracts preparation is same as that of direct methanolic extraction. The name of the species, its respective plant part, type of extract studied and percentage antioxidant activity has been enumerated in Table 2.

The extracts that showed potential activity (>90%) were further studied for their IC_{50} values, which are enumerated in Table 3.

Discussion

In the present study, out of 123 extracts prepared from 59 plant samples, 59 methanolic extracts and 54 successively prepared extracts were tested for antioxidant activity using standard in vitro model. Interestingly, successively prepared two ethyl acetate and 16 methanol extracts showed strong antioxidant nature in DPPH in vitro assay. Out of 59 methanolic extract samples, which were screened in DPPH assay, 16 samples showed more than 90% antioxidant activity. Those 16 crude samples were then freshly subjected for successive extraction by using pet ether (60-80), dichloro methane, ethyl acetate

Table 2: Antioxidant activity (AO) profile of potential sequential extracts

S. No.	Name of the species with plant part	Percentage AO in PE	Percentage AO in DCM	Percentage AO in EA	Percentage AO in Me
1.	Alpinia officinarum-Rhizome	15.03	21.15	72.67	95.36
2.	Aquilaria malaccensis - Aerial Part	2.96	7.66	10.37	92.62
3.	Grewia sapida - Twig	45.94	54.26	56.24	82.38
4.	Grewia sapida - Stem	34.65	9.50	79.41	95.05
5.	Grewia nervosa - Twig	4.16	31.68	33.27	98.40
6.	Litsea glutinosa - Stem	28.31	56.97	92.89	91.39
7.	Mitragyna rotundifolia - Twig	2.55	7.52	39.05	97.34
8.	Mitragyna rotundifolia - Stem	6.39	17.65	50.25	93.03
9.	Pterospermum semisagittatum - Stem	32.67	85.15	72.56	94.65
10.	Saraca asoca - Stem	23.37	28.32	44.55	93.07
11.	Schima wallichii - Leaf	23.56	34.85	78.81	96.63
12.	Schima wallichii - Stem	32.29	51.49	96.37	97.84
13.	Syzygium cerasoides - Leaf	1.37	12.44	48.30	95.39
14.	Syzygium cerasoides - Stem	6.45	7.97	11.58	93.13
15.	Wendlandia wallichii - Twig	20.20	14.65	28.32	95.07
16.	Wendlandia wallichii - Stem	17.96	23.56	81.58	93.47
	Standard - Curcumin				99.48
	Standard - Catechin				99.47
	Standard - Trolox				100

PE-pet ether extract, DCM-dichloro methane extract, EA-ethyl acetate extract, Me-methanol extract

Table 3: Promising extracts with their IC_{50} values

S. No.	Name of the species with plant part	Type of extract	IC ₅₀ value in mg
1.	Alpinia officinarum - Rhizome	Methanol	0.42
2.	Aquilaria malaccensis - Aerial Part	Methanol	0.05
3.	Grewia nervosa - Twig	Methanol	0.044
4.	Grewia sapida - Twig	Methanol	0.04
5.	Grewia sapida - Stem	Methanol	0.045
6.	Litsea glutinosa - Stem	Methanol	0.10
7.	Mitragyna rotundifolia - Twig	Methanol	0.034
8.	Mitragyna rotundifolia - Stem	Methanol	0.11
9.	Pterospermum semisagittatum - Stem	Methanol	0.044
10.	Saraca asoka - Stem	Methanol	0.06
11.	Schima wallichii - Stem	Ethyl acetate	0.062
12.	Schima wallichii - Leaf	Methanol	0.039
13.	Syzygium cerasoides - Stem	Methanol	0.13
14.	Syzygium cerasoides - Leaf	Methanol	0.041
15.	Wendlandia wallichii - Twig	Methanol	0.05
16.	Wendlandia wallichii - Stem	Methanol	0.12
	Standard - Curcumin		0.048
	Standard – Catechin		0.033
	Standard - Trolox		0.031

and methanol. Out of all successively prepared extracts only 12 % ethyl extracts showed almost same antioxidant activity as that of alcoholic extracts. Therefore, it can be said that the efficiency of each species differs depending on the particular assay methodology, reflecting complexity of the mechanisms involved in total antioxidant activity. The remarkable example of such diversity is the extracts of Litsea glutinosa (stem) and Schima wallichii (stem). Potent antioxidant extracts and compounds are known to increase the levels of catalase and SOD and decrease the level of TBARs in blood and tissues when compared with CCl₄ treatment¹⁶.

There is antioxidant serum in the market by the name 'Juice Beauty' which is a polyherbal preparation and one

of the phytoconstituents is *Litsea cubeba* Pers.¹⁷. Leaves of *L. glutinosa* are used in nausea, vomiting and possess antidiarrhoeal activity. Decoction of bark may be applied to the sores and scabies and also aches and pains. Juice of crushed leaves may be applied to relieve the sore eyes^{18, 19}. This corroborates the antioxidant credentials of the *L. glutinosa* reported in this paper.

The butanol fraction of *Schima wallichii* leaves have been shown to have antimutagenic and antioxidant activities ²⁰. In the present study, we have found that leaf has antioxidant activity in methanol fraction and the stem has both in ethyl acetate and methanol fractions.

Water extract of *Syzygium gratum* (Wt.) S.N. Mitra (leaf)²¹ and ethanolic extract of *S. cuminii* (Linn.)

Skeels (Stem bark)²² are reported to have antioxidant and anti-inflammatory properties. In the present study we found *S. cerasoides* stem is having antioxidant property in methanolic fraction.

Antioxidative compounds were isolated from the methanol extract of fresh rhizome of *Alpinia officinarum*²³. In the present study it is noticed that dry rhizomes of same species from Tripura also have potent antioxidant property. Likewise the ethyl acetate extract of heartwood of *Aquilaria agallocha* also possess antioxidant activity²⁴ and the activity in methanol extract of aerial parts is also noticed by us.

Conclusion

All the selected extracts, subjected to dose dependent studies to calculate IC₅₀ values are from successive methanolic extraction. Therefore, it could be concluded that only direct methanol extract is not sufficient for investigations in antioxidant property, but every sample has to be successively extracted with different solvents with increasing polarities as is evident in case of two plants, viz. *Litsea glutinosa* (stem) and *Schima wallichii* (stem) in which successive ethyl acetate extracts have shown equal antioxidant activity as that of successive methanol.

Mitragyna rotundifolia (twig), Schima wallichii (leaf), Syzygium cerasoides (stem) have good activity in dose dependent study i.e. 50% reduction of DPPH is achieved at the dose of 0.03 mg/ml that is comparable to the standards (which are pure molecules), viz. Catechin and Trolox. Moreover, their activities are better than that of Curcumin for which 50% reduction is a chieved by

Research Paper

0.04 mg/ml. *Grewia nervosa* (twig), *G. sapida* (twig, stem), *Pterospermum semisagittatum* (stem) and *Syzygium cerasoides* (leaf) have same activity profile as that of Curcumin. Therefore, further isolation of antioxidant constituents and *in vivo* studies are warranted for above samples. Further phytochemical work on isolation of bioactive molecules is going on.

Acknowledgements

The authors gratefully acknowledge the encouragement and support of Reliance Life Sciences Pvt. Ltd, in carrying out the research work. They are also thankful to Medicinal Plant Board and Forest Department, Tripura for their logistic support during plant collection.

References

- Tominaga Hitoshi, Kobayashi Yuka, Goto Takashi, Kasemura Kazuo and Nomura Masato, DPPH Radical-scavenging effect of several phenylpropanoid compounds and their glycoside derivatives, Yakugaku Zasshi, 2005, 125(4), 371-375.
- Atawodi SE, Antioxidant potential of African medicinal plants, Afr J Biotechnol, 2005, 4 (2), 128-133.
- 3. Katalinic V, Milos M, Kulisic T and Jukic M, Screening of 70 medicinal plant extracts for antioxidant capacity and total phenols, *Food Chem*, 2006, **94**, 550-557.
- Badami Shrishailappa, Moorkoth Sudheer, Rammanoharsingh Rai Sujay, Kannan Elango and Bhojraj Suresh, Antioxidant activity of Caesalpinia sappan heartwood, Biol Pharm Bull, 2003, 26(11), 1534-1537.
- Shyur, Lie-Fen, Tsung Jieh-Hen, Chen Je-Hsin, Chiu Chih-Yang and Lo Chiu-Ping, Antioxidant properties of extracts from medicinal plants popularly used in Taiwan, *Int J Appl Sci Eng*, 2005, 3(3), 195-202.

- Majumdar Koushik and Datta BK, A study on ethnomedicinal usage of plants among the folklore herbalists and Tripuri medical practitioners: Part-II, *Nat Prod Rad*, 2007, 6, 66-73.
- Majumdar Koushik and Datta BK, Medicinal Plants Prescribe by Different Tribal and Nontribal Medicine Men of Tripura State, *Indian* J Trad Knowledge, 2007, 5(4), 559-562.
- Kshirsagar RD and Singh NP, Ethnobotany of Mysore and Coorg, Karnataka State, Bishen Singh Mahendra Pal Singh, Dehra Dun, 2007.
- Kshirsagar Rajendra and Saklani Arvind, Ethnomedicinal plants for diabetes, jaundice and rheumatism in Karnataka and their comparison with northeast India, *In*: Advances in Ethnobotany, by AP Das and AK Pandey (Eds), Bishen Singh Mahendra Pal Singh, Dehra Dun, 2007, pp. 95-116.
- Mazumder Pradip, Bio-Diversity of Tripura, http://www.tripurainfo.com/opinion/feature/ pradipmazumdar3.shtml.
- 11. Deb DB, Flora of Tripura, Vol. 1 & 2, Todays and Tomorrow's Printers & Publishers, New Delhi, 1981-1983.
- Singh NP, Singh KP and Singh DK, Flora of Mizoram, Vol. 1, Botanical Survey of India, Kolkata, 2002.
- 13. Singh NP, Chauhan AS and Mondal MS, Flora of Manipur, Vol. 1, Botanical Survey of India, Kolkata, 2000.
- 14. Mukherjee Pulok K, Quality Control of Herbal Drugs An approach to evaluation of botanicals, Business Horizons Pharmaceutical Publishers, New Delhi, 2002.
- 15. Molyneux Philip, The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin J Sci Technol*, 2004, **26**(2), 211-219.
- Badami Shrishailappa, Rai Sujay P and Suresh
 Antioxidant activity of Aporosa lindleyana root, J Ethnopharmacol,
 2005, 101, 180-184.

- 17. Juice Beauty Antioxidant serum 2 oz AM/ PM (http://www.skin-one.com/juice-beauty-antioxidant-serum.html).
- 18. John Farnham, Aboriginal Traditional medicines, http://www.geocities.com/BookofWisdomandMagick5/Litsea.html.
- 19. Sahoo J, Sahu PK and Tripathi DK, Antidiarrhoeal activity of *Litsea glutinosa* leaves, *Adv Pharmacol Toxicol*, 2006, 7(2), 53-55.
- Didi Jauhari Purwadiwarsa, Anas Subarnas, Cucu Hadiansyah and Supriyatna, The investigation on anti-mutagenic and antioxidant activity of Schima wallichii (DC.) Korth leaves, Cermin Dunia Kedokt, 2000, 127, 18-21.
- 21. Kukongviriyapan U, Luangaram S and Leekhaosoong K, Antioxidant and vascular protective activities of *Cratoxylum formosum*, *Syzygium gratum* and *Limnophila aromatica*, *Biol Pharm Bull*, 2007, **30**(4), 661-666.
- 22. Muruganandan S, Srinivasan K, Chandra S, Tandan SK, Lal J and Raviprakash V, Antiinflammatory activity of *Syzygium cumini* bark, *Fitoterapia*, 2001, 72, 369-375.
- Tram Ngoc Ly, Makoto Shimoyamada, Koji Kato and Ryo Yamauchi, Antioxidative compounds isolated from the rhizomes of smaller galanga (*Alpinia officinarum* Hance), *BioFactors*, 2004, 21(1-4), 305-308.
- 24. Miniyar PB, Chitre TS, Deuskar HJ, Karve SS and Jain KS, Antioxidant activity of ethyl acetate extract of *Aquilaria agallocha* on nitrite-induced methaemoglobin formation, *Int J Green Pharm*, 2008, **2**(2), 116-117.

Natural Product Radiance