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# Modeling and numerical simulation of action potential patterns in human atrial tissues

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## Abstract

Electrophysiology of the heart is the subject of a vast interdisciplinary literature, from medical sciences through bio-engineering, physiology, chemistry and physics. The difficulty in having access to direct measures on real patients entailed the coupling of such studies with numerical simulations. Several works have been done on this topic, focusing mainly on the behavior of the ventricles. In this paper we focus on atrial simulation: we present a reaction-diffusion model coupled with the simple Fitz-Hugh-Nagumo (FHN) model and the more complex Courtemanche-Ramirez-Nattel (CRN) model, which has been derived explicitly for human atrial cells. Numerical experiments are performed with both the bidomain and the monodomain models to simulate the evolution of a complete heartbeat.

## 1 Introduction

The basic property of neural cells to produce signals is called Action Potential (AP). It consists of a sudden variation in the transmembrane potential, called upstroke, followed by a recovering of the resting condition. It shows different shapes and amplitudes according to the different kind of excitable media to which the cells belong to, and in the large muscle cells makes it possible the simultaneous contraction of the whole cell. An action potential propagates keeping the same shape and amplitude all along an entire neural or muscular fiber. Cardiac cells are characterized by a transmembrane potential that is negative at rest, owing to the fact that the concentration of potassium ions  $[K^+]_i$  inside the cardiac cell is remarkably higher than the outside concentration  $[K^+]_e$ , and show two kinds of action potentials: the quick and the slow response.

The quick response is typical in the myocardium fibers (both atrial and ventricular) and in the Purkinje fibers, which are fibers specialized in the conduction. The quick response cells are characterized by a negative transmembrane potential at rest (around -90mV), and by a rapid depolarization (positive overshoot), where the potential difference changes sign and the internal potential overtakes the external one of around 20mV: such phase is called Phase 0. Immediately after that (Phase 1) a short period of partial repolarization takes place, followed by a plateau (Phase 2) which lasts for around 0.2 seconds. The potential gets progressively more negative (Phase 3) until it reaches again the resting value. The repolarization procedure is far slower than the depolarization one, and the interval between the end of the repolarization and the next action potential is called Phase 4.

The slow response is the one taking place in the Sinoatrial Nodus (SA), the natural pacemaker of the heart, and in the Atrioventricular Nodus (AV), the tissue meant to transfer the pulse from atria to ventricles. The slow response cells are characterized by a resting potential less negative (around -50mV), and by a smaller slope and amplitude in the overshoot of the action potential, the absence of the Phase 1, and by a relative refractory period that continues during Phase 4.

The Action Potential propagates across the heart in a heterogeneous way. The pulse moves from the Sinoatrial Nodus (SA), and propagates through the ordinary myocardic fibers of the right atrium, while the Bachmann's bundle drives the pulse towards the left atrium. Some action potentials propagate downwards and reach the Atrioventricular Nodus (AV), which is, under normal conditions, the only gate for the pulse

to propagate from atria to ventricles. Moreover, the conduction is slower in atria than in ventricles ( $1 \text{ ms}^{-1}$  versus  $4 \text{ ms}^{-1}$ ).

The electrical activity of the heart as a whole is thus characterized by a complex multiscale structure, ranging from the microscopic activity of ion channels in the cellular membrane to the macroscopic properties of the anisotropic propagation of the excitation and recovery fronts in the whole heart. The most complete model of such a complex setting is the anisotropic Bidomain model (see [9, 23]), that consists of a system of two degenerate parabolic reaction-diffusion equations describing the intra and extracellular potentials in the cardiac muscle, coupled with a system of ordinary differential equations describing the ionic currents flowing through the cellular membrane, that are associated to the nonlinear reaction term. This model is computationally very expensive because of the involvement of different space and time scales, and a simplified tissue model is the anisotropic Monodomain system, consisting of a parabolic reaction-diffusion equation describing the propagation of the transmembrane potential coupled with an ionic model, which has been widely used in literature (see for instance [21, 17]).

If, on the one hand, a wide literature is available for ventricular models (see for instance [4, 5, 18] and references therein), on the other hand less has been done on atria, although Atrial Fibrillation (AF) is the most commonly sustained arrhythmia, for which clinical treatment remains the most problematic. Knowledge of the human atrial Action Potential and of its ionic currents is thus of critical importance to understand the electrical properties of atrial tissues in both normal and pathological conditions.

In this paper we describe the Fitz-Hugh-Nagumo (FHN) cell model in the Rogers-McCulloch variant, which is well suited to capture the excitation wavefront, but behaves very poorly in the description of the plateau phase, and the Courtemanche-Ramirez-Nattel (CRN) model, that is especially designed for human atrial cells. These models are then coupled with both monodomain and bidomain simulations on a two dimensional slab with anisotropic conduction.

The rest of the paper is organised as follows. In Section 2 we give a brief review of the mathematical models, the anisotropic bidomain and monodomain ones. In Section 3 we describe the ionic currents and the modified FHN and the CRN membrane models. In Section 4 we give the variational formulation of the problem and its finite element approximation. Finally, in Section 5, some numerical simulations on a two dimensional cartesian slab are presented, varying both the cardiac tissue model (monodomain and bidomain) and the ionic model (FHN and CRN).

## 2 Description of the model

The conductivity of the cardiac cells depends upon their orientation, and in the most general case the conductivity tensor is anisotropic. The structure of the cardiac cells can be modeled, following Le Grice *et al.* ([13]) as a sequence of muscular layers going from endocardium to epicardium (see also [26]). In any point  $\mathbf{x}$  it is thus possible to identify an orthonormal triplet of directions,  $\mathbf{a}_l(\mathbf{x})$ ,  $\mathbf{a}_t(\mathbf{x})$ ,  $\mathbf{a}_n(\mathbf{x})$ , with  $\mathbf{a}_l(\mathbf{x})$  parallel to the fibers direction,  $\mathbf{a}_t(\mathbf{x})$  e  $\mathbf{a}_n(\mathbf{x})$  tangent and orthogonal respectively to the radial lamination, both transversal with respect to the fiber axis.

### 2.1 The bidomain model

The bidomain model consists in representing the cardiac tissue as the superposition of two media which are continuous and anisotropic, the intra-cellular and the extra-cellular one, coexisting at each point  $\mathbf{x}$  and separated by a cell membrane. Such model has been derived, by an homogenization technique, starting from a periodic assembling of elongated cells surrounded by extracellular space and connected by end-to-end or side-to-side junctions (for the mathematical details we refer to [12, 6]).

Denoting by  $\sigma_l^{i,e}$ ,  $\sigma_t^{i,e}$ , and  $\sigma_n^{i,e}$  the conductivity coefficients in the  $\mathbf{a}_l$ ,  $\mathbf{a}_t$  and  $\mathbf{a}_n$  directions, the conductivity tensor is given by

$$D_{i,e}(\mathbf{x}) = \sigma_l^{i,e} \mathbf{a}_l(\mathbf{x}) \mathbf{a}_l^T(\mathbf{x}) + \sigma_t^{i,e} \mathbf{a}_t(\mathbf{x}) \mathbf{a}_t^T(\mathbf{x}) + \sigma_n^{i,e} \mathbf{a}_n(\mathbf{x}) \mathbf{a}_n^T(\mathbf{x}),$$

for the intra- and extra-cellular medium respectively.

The intra-cellular and extra-cellular electric potentials, which we denote by  $u_i$  and  $u_e$ , are governed in the bidomain model by a degenerate reaction diffusion system of parabolic type, coupled with an ODE system describing the ionic gating variables  $w$  and the ionic concentration  $c$ . In the following we denote

$I_m = c_m \partial_t v + I_{ion}(v, w, c)$  the membrane current per unit volume, where  $c_m = \chi C_m$ ,  $I_{ion} = \chi i_{ion}$ ,  $\chi$  being the membrane area per tissue volume,  $C_m$  the surface capacity and  $i_{ion}$  the membrane ionic current per unit area.

Let then  $I_e^{app}$  be an applied extra-cellular current per unit volume which satisfies the compatibility condition  $\int_{\Omega} I_e^{app} = 0$  and let  $\mathbf{j}_{i,e} = -D_{i,e} \nabla u_{i,e}$  be the intra- and extra-cellular current density: the conservation of the total current entails  $\text{div } \mathbf{J}_i = -I_m$  and  $\text{div } \mathbf{J}_e = I_m - I_e^{app}$ .

The bidomain model in the variables  $(u_i(\mathbf{x}, t), u_e(\mathbf{x}, t))$  and  $v(\mathbf{x}, t) = u_i(\mathbf{x}, t) - u_e(\mathbf{x}, t)$ , for an insulated domain  $\Omega \subset \mathbb{R}^3$ , reads then

$$\begin{cases} c_m \partial_t v - \text{div} (D_i \nabla u_i) + I_{ion}(v, w, c) = 0 & \text{in } \Omega \times (0, T) \\ -c_m \partial_t v - \text{div} (D_e \nabla u_e) - I_{ion}(v, w, c) = -I_e^{app} & \text{in } \Omega \times (0, T) \\ \partial_t w - R(v, w) = 0 & \text{in } \Omega \times (0, T) \\ \partial_t c - S(v, w, c) = 0 & \text{in } \Omega \times (0, T) \\ \mathbf{n}^T D_{i,e} \nabla u_{i,e} = 0 & \text{in } \partial\Omega \times (0, T) \\ v(\mathbf{x}, 0) = v_0(\mathbf{x}) \quad w(\mathbf{x}, 0) = w_0(\mathbf{x}) \quad c(\mathbf{x}, 0) = c_0(\mathbf{x}) & \text{in } \Omega. \end{cases} \quad (2.1)$$

The above system uniquely determines  $v$ , whereas the potentials  $u_i$  and  $u_e$  are determined modulus an additive constant depending on time and linked to the reference potential. Such potential is chosen as the mean extracellular potential in the cardiac volume, by imposing a zero mean condition  $\int_{\Omega} u_e = 0$ .

System (2.1) can be rewritten in terms of extracellular and transmembrane potentials. From the second equation in (2.1) we get

$$c_m \partial_t v = -\text{div} (D_e \nabla u_e) - I_{ion}(v, w, c) - I_e^{app},$$

and inserting it into the first one provides

$$-\text{div} [D_i \nabla u_i + D_e \nabla u_e] = -I_e^{app}.$$

So far, adding and subtracting  $\text{div} (D_i \nabla u_e)$ , we obtain the formulation in terms of  $v$  and  $u_e$ :

$$\begin{cases} -c_m \partial_t v - \text{div} (D_e \nabla u_e) - I_{ion}(v, w, c) = -I_e^{app} & \text{in } \Omega \times (0, T) \\ -\text{div} [(D_i + D_e) \nabla u_e] = \text{div} (D_i \nabla v) - I_e^{app} & \text{in } \Omega \times (0, T) \end{cases} \quad (2.2)$$

## 2.2 The simplified monodomain model

If we assume the anisotropy ratio to be the same in the two media, intra- and extra-cellular, namely we assume  $D_i = \lambda D_e$  with  $\lambda$  constant, the bidomain model reduces to the monodomain one. If we let

$$D = \frac{\lambda}{1 + \lambda} D_i, \quad I^{app} = \frac{\lambda}{1 + \lambda} I_i^{app} + \frac{1}{1 + \lambda} I_e^{app},$$

the simplified monodomain model consists of a single reaction-diffusion equation of parabolic type for the transmembrane potential  $v$ .

$$\begin{cases} c_m \partial_t v - \text{div} (D(\mathbf{x}) \nabla v) + I_{ion}(v, w, c) = I^{app} & \text{in } \Omega \times (0, T) \\ \partial_t w - R(v, w) = 0 & \text{in } \Omega \times (0, T) \\ \partial_t c - S(v, w, c) = 0 & \text{in } \Omega \times (0, T) \\ \mathbf{n}^T D \nabla v = 0 & \text{in } \partial\Omega \times (0, T) \\ v(\mathbf{x}, 0) = v_0(\mathbf{x}), \quad w(\mathbf{x}, 0) = w_0(\mathbf{x}), \quad c(\mathbf{x}, 0) = c_0(\mathbf{x}) & \text{in } \Omega. \end{cases} \quad (2.3)$$

## 3 Ionic currents and membrane models

The ionic currents appearing in both the monodomain and the bidomain model rely on the choice of the membrane model for the cell conductivity. The earliest model appeared in the work on nerve action potential by Hodgkin and Huxley ([10]), which earned them the Nobel prize in Medicine in 1963. Models of this type

have then been developed for the cardiac action potential, where the variation in time of the membrane potential  $v$  (under the assumption of an equipotential cell) is given by

$$\frac{dv}{dt} = -\frac{I_{ion} - I_{st}}{C_m}, \quad (3.1)$$

where  $I_{ion}$  and  $I_{st}$  are the total ionic current and stimulus current across the membrane, respectively, and  $C_m$  is the total membrane capacitance. In (3.1) the ionic current through the channels in the membrane depends on the transmembrane potential  $v$  and on  $M$  gating and concentration variables  $w \in \mathbb{R}^M$ , and reads

$$I_{ion}(v, w) = \sum_{k=1}^N G_k(v) \prod_{j=1}^M w_j^{p_{jk}} (v - v_k(w)),$$

$G_k(v)$  being the membrane conductance,  $v_k$  being the reversal potential for the  $k$ -th current and  $p_{jk}$  being integers, and where the dynamics of the gating and concentration variables is described by a system of ODE's

$$\frac{\partial w}{\partial t} = R(v, w), \quad w(\mathbf{x}, 0) = w_0(\mathbf{x}).$$

In such models, if  $w_j$  is a gating variable, the right hand side  $R_j(v, w)$  has a special structure and the corresponding ODE is given by

$$\frac{\partial w_j}{\partial t} = R_j(v, w) = R_j(v, w_j) = \alpha_j(v)(1 - w_j) - \beta_j(v)w_j, \quad (3.2)$$

with  $\alpha_j(v), \beta_j(v) > 0$ ,  $0 < w_j < 1$ .

Many refinements of the original Hodgkin-Huxley model, obtained by fitting improved experimental data with more complex models, have been developed for the modeling of ventricular cells: among them, we recall the model by Beeler and Reuter (1977, with 4 ionic currents and 7 gating and concentrations variables), and the phase-I Luo-Rudy (1991, with  $N = 6$  and  $M = 7$ ). In this direction, the most recent published model of mammalian ventricular cells is the phase-II Luo-Rudy (1994, [15]), which is based on measurements from guinea pig. Simpler models of reduced complexity have also been proposed, where only 1 or 2 gating variables are considered.

If, on the one hand, several models are available to describe the behaviour of ventricular cells, less has been done focusing on atrial cells. Atria differ from ventricles under several aspects. First of all, the thickness of the wall in atria is far less significant than in ventricles, while the speed of conduction is much larger in ventricles. A different ionic current and membrane model should therefore be used when dealing with atria. In this direction, models of atrial cells based on animal data only have been published (see [8, 20, 27]): the most recent of such model is the one proposed by Lindblad *et al.* (1996, [14]), which is based on measurements from rabbit atrial cells. Although these models have provided valuable insights into the mechanisms underlying the action potential generation in animals, the significant interspecies differences with respect to human being and the amount of available human data led scientists to develop mathematical models of the action potential based on ionic current data measured directly in human atrial cells. The most recently published models in this direction are the ones proposed by Nygren *et al.* in [16] and by Courtemanche *et al.* in [7]. These two models handle the atrial cell as a capacitor connected in parallel with variable resistances and batteries representing the ionic channels and driving forces.

### 3.1 The FHN cell model

The most used and simplest ionic model is the Fitz-Hugh-Nagumo (FHN), consisting of 1 ionic current and 1 gating variable. Assuming the potential  $v$  to be zero at rest, the ionic current uses only one recovery variable:

$$I_{ion}(v, w) = g(v) + \beta w,$$

where  $\beta > 0$ ,  $g(v)$  is a cubic-like function and  $w$  satisfies

$$\frac{\partial w}{\partial t} = \eta v - \gamma w,$$

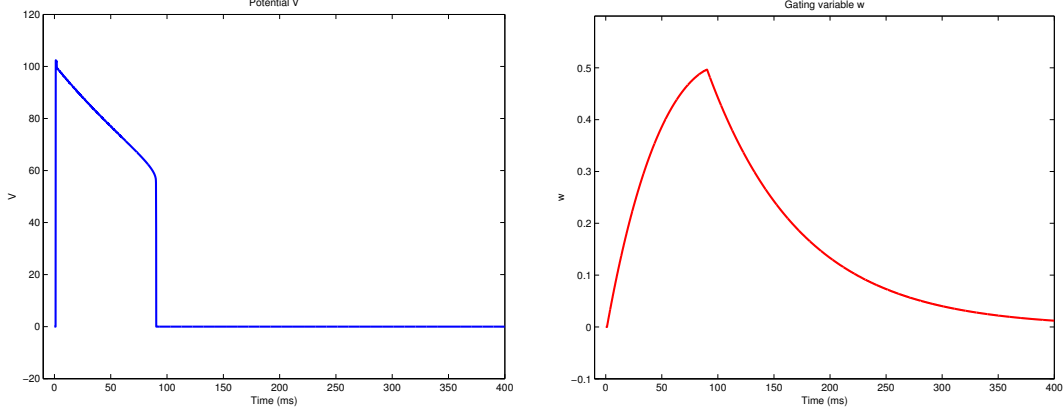


Figure 1: Time evolution of the potential  $v$  and the gating variable  $w$  in the Rogers-McCulloch variant of the FHN model

with  $\eta, \gamma > 0$ .

An improvement of this model is given by the following variant by Rogers and McCulloch ([21]):

$$I_{ion}(v, w) = Gv \left( 1 - \frac{v}{v_{th}} \right) \left( 1 - \frac{v}{v_p} \right) + \eta_1 vw,$$

$$\frac{\partial w}{\partial t} = \eta_2 \left( \frac{v}{v_p} - \eta_3 w \right),$$

where  $G, \eta_1, \eta_2, \eta_3$  are positive coefficients,  $v_{th}$  is a threshold potential, and  $v_p$  is the peak potential. In Figure 1 we report the time evolution of the potential  $v$  and of the gating variable  $w$  for the Rogers-McCulloch variant of the FHN model.

If, on the one hand, such models are well suited to describe the positive overshoot in the quick depolarization phase, on the other hand they provide only a coarse approximation in the plateau and repolarization phases of the action potential, and behave too poorly when accuracy in the description of the action potential is needed.

### 3.2 The CRN atrial cell model

One of the most accurate models for atrial cells is the CRN (Courtemanche, Ramirez and Nattel, [7]) one, in which the total ionic current is given by the sum

$$I_{ion} = I_{Na} + I_{K1} + I_{to} + I_{Kur} + I_{Kr} + I_{Ks} + I_{Ca,L} + I_{p,Ca} + I_{NaK} + I_{NaCa} + I_{b,Na} + I_{b,Ca}. \quad (3.3)$$

In the above expression, the total ionic current take into account the fast depolarizing  $Na^+$  current, the rectifier  $K^+$  currents, the L-type  $Ca^{2+}$  current, the background currents, the intracellular concentrations  $[Ca^{2+}]_i$ ,  $[Na^+]_i$ , and  $[K^+]_i$ , the action of the NaK and NaCa pumps, and the handling of the intracellular calcium concentration by the sarcoplasmic reticulum system (SR). In the model, no extracellular cleft space is included, the membrane capacitance is  $c_m = 100pF$ , the length and diameter of the cells are set to 100 and 16  $\mu m$ , respectively, and the cell compartment volumes are the same ones used in the phase-II Luo-Rudy model (LR2, [15]). We give hereafter a brief description of the ionic currents. In that order, we denote with  $E_X$  the equilibrium potential for ion  $X$ , and with  $g_X$  its maximal conductance. From Nerst equation,  $E_X$  is given by

$$E_X = \frac{RT}{zF} \log \frac{[X]_e}{[X]_i},$$

where  $R$  is the gas constant,  $T$  is the absolute temperature,  $F$  is the Faraday constant,  $z = 1$  for  $Na^+$  and  $K^+$ ,  $z = 2$  for  $Ca^{2+}$ , and  $[X]_e$  and  $[X]_i$  denote the external and internal concentration of ion  $X$ .

### Fast Na<sup>+</sup> current

It is a large, rapid inward current which is the major responsible of the depolarization phase at the beginning of the action potential. It is given by

$$I_{Na} = g_{Na} m^3 h j (v - E_{Na}),$$

where  $m$ ,  $h$ , and  $j$  are gating variables, whose coefficients in equation (3.2) are given by

$$\alpha_m = \begin{cases} .32 \frac{v + 47.13}{1 - \exp[-.1(v + 47.13)]} \\ 3.2, & \text{if } v = -47.13 \end{cases} \quad \beta_m = .08 \exp\left(-\frac{v}{11}\right)$$

$$\alpha_h = \begin{cases} .135 \exp\left(-\frac{v + 80}{6.8}\right) \\ 0, & \text{if } v \geq -40 \end{cases} \quad \beta_h = \begin{cases} 3.56 \exp(.079v) + 3.1 \times 10^5 \exp(.35v) \\ 7.69 \left[1 + \exp\left(-\frac{v + 10.66}{11.1}\right)\right]^{-1}, & \text{if } v \geq -40 \end{cases}$$

$$\alpha_j = \begin{cases} [-127, 140 \exp(.2444v) - 3.474 \times 10^{-5} \exp(-.04391v)] \frac{v + 37.78}{1 + \exp[.311(v + 79.23)]} \\ 0, & \text{if } v \geq -40 \end{cases}$$

$$\beta_j = \begin{cases} .1212 \frac{\exp(-.01052v)}{1 + \exp[-.1378(v + 40.14)]} \\ .3 \frac{\exp[-2.535 \times 10^{-7}v]}{1 + \exp[-.1(v + 32)]} & \text{if } v \geq -40 \end{cases}$$

### Inward rectifier K<sup>+</sup> current

This current plays a major role in the late repolarization phase of the AP and in determining resting membrane potential and resistance. It is given by

$$I_{K1} = \frac{g_{K1}(v - E_K)}{1 + \exp(.07(v + 80))} \quad g_{K1} = .09$$

### Ultrarapid rectifier K<sup>+</sup> current:

$$I_{Kur} = g_{Kur} u_a^3 u_i (v - E_K), \quad g_{Kur} = .005 + \frac{.05}{1 + \exp\left(\frac{v - 15}{-13}\right)},$$

where  $u_a$  and  $u_i$  are gating variables, which satisfy equation (3.2) with coefficients

$$\alpha_{ua} = .65 \left[ \exp\left(-\frac{v + 10}{8.5}\right) + \exp\left(-\frac{v - 30}{59.0}\right) \right]^{-1} \quad \beta_{ua} = .65 \left[ 2.5 + \exp\left(\frac{v + 82}{17}\right) \right]^{-1}$$

$$\alpha_{ui} = \left[ 21 + \exp\left(-\frac{v - 185}{28}\right) \right]^{-1} \quad \beta_{ui} = \exp\left(\frac{v - 158}{16}\right)$$

### Transient outward $K^+$ current

$$I_{to} = g_{to} o_a^3 o_i (v - E_K), \quad g_{to} = .1652,$$

where  $o_a$  and  $o_i$  are gating variables, which satisfy equation (3.2) with coefficients

$$\begin{aligned} \alpha_{oa} &= .65 \left[ \exp \left( -\frac{v+10}{8.5} \right) + \exp \left( -\frac{v-30}{59.} \right) \right]^{-1} & \beta_{oa} &= .65 \left[ 2.5 + \exp \left( \frac{v+82}{17.} \right) \right]^{-1} \\ \alpha_{oi} &= \left[ 18.53 + \exp \left( \frac{v+113.7}{10.95} \right) \right]^{-1} & \beta_{oi} &= \left[ 18.53 + \exp \left( -\frac{v+1.26}{7.44} \right) \right]^{-1} \end{aligned}$$

### Rapid and slow delayed rectifier $K^+$ currents

$$\begin{aligned} I_{Kr} &= \frac{g_{Kr} x_r (v - E_K)}{1 + \exp \left( \frac{v+15}{22.4} \right)} & g_{Kr} &= .0294 \\ I_{Ks} &= g_{Ks} x_s^2 (v - E_K) & g_{Ks} &= .129 \end{aligned}$$

where  $x_r$  and  $x_s$  are gating variables, which satisfy equation (3.2) with coefficients

$$\begin{aligned} \alpha_{xr} &= .0003 \frac{v+14.1}{1 - \exp \left( \frac{v+14.1}{5} \right)} & \beta_{xr} &= 7.3898 \times 10^{-5} \frac{v-3.3328}{\exp \left( \frac{v-3.3328}{5.1237} \right)} \\ \alpha_{xs} &= 4 \times 10^{-5} \frac{v-19.9}{1 - \exp \left( -\frac{v-19.9}{17} \right)} & \beta_{xs} &= 3.5 \times 10^{-5} \frac{v-19.9}{\exp \left( \frac{v-19.9}{9} - 1 \right)} \end{aligned}$$

### L-type $Ca^{2+}$ current

$$I_{Ca,L} = g_{Ca,L} d f f_{Ca} (v - 65.0) \quad g_{Ca,L} = .1238$$

where  $d$ ,  $f$  and  $f_{Ca}$  are gating variables, which satisfy equation

$$\frac{dy}{dt} = -\frac{y^\infty - y}{\tau_y} \quad (3.4)$$

where  $y$  is any of  $d$ ,  $f$ ,  $f_{Ca}$ , with coefficients

$$\begin{aligned} \tau_d &= \frac{1 - \exp \left( -\frac{v+10}{6.24} \right)}{.035 (v+10) \left[ 1 + \exp \left( -\frac{v+10}{6.24} \right) \right]} & d^\infty &= \left[ 1 + \exp \left( -\frac{v+10}{8} \right) \right]^{-1} \\ \tau_f &= 9 \left[ .0197 \exp \left[ -.0337^2 (v+10)^2 \right] + .02 \right]^{-1} & f^\infty &= \left[ 1 + \exp \left( \frac{v+28}{6.9} \right) \right]^{-1} \\ \tau_{f_{Ca}} &= 2 & f_{Ca}^\infty &= \left( 1 + \frac{[Ca^{2+}]_i}{.00035} \right)^{-1} \end{aligned}$$

### $Ca^{2+}$ pump current

$$I_{p,Ca} = I_{p,Ca(max)} \frac{[Ca^{2+}]_i}{.0005 + [Ca^{2+}]_i} \quad I_{p,Ca(max)} = .275$$

### $Na^+$ - $K^+$ pump current

$$I_{NaK} = I_{NaK(max)} f_{NaK} \frac{1}{1 + \left( \frac{K_{m,Na(i)}}{[Na^+]_i} \right)^{3/2}} \frac{[K^+]_o}{[K^+]_o + K_{m,K(o)}} \quad I_{NaK(max)} = .6,$$

where  $K_{m,Na(i)}$  and  $K_{m,K(o)}$  are the  $[Na^+]_i$  and  $[K^+]_o$  half saturation constants, respectively, while

$$f_{NaK} = \left[ 1 + .1245 \exp \left( -.1 \frac{Fv}{RT} \right) + .0052 \left[ \exp \left( \frac{[Na^+]_o}{67.3} \right) - 1 \right] \exp \left( -\frac{Fv}{RT} \right) \right]^{-1}$$



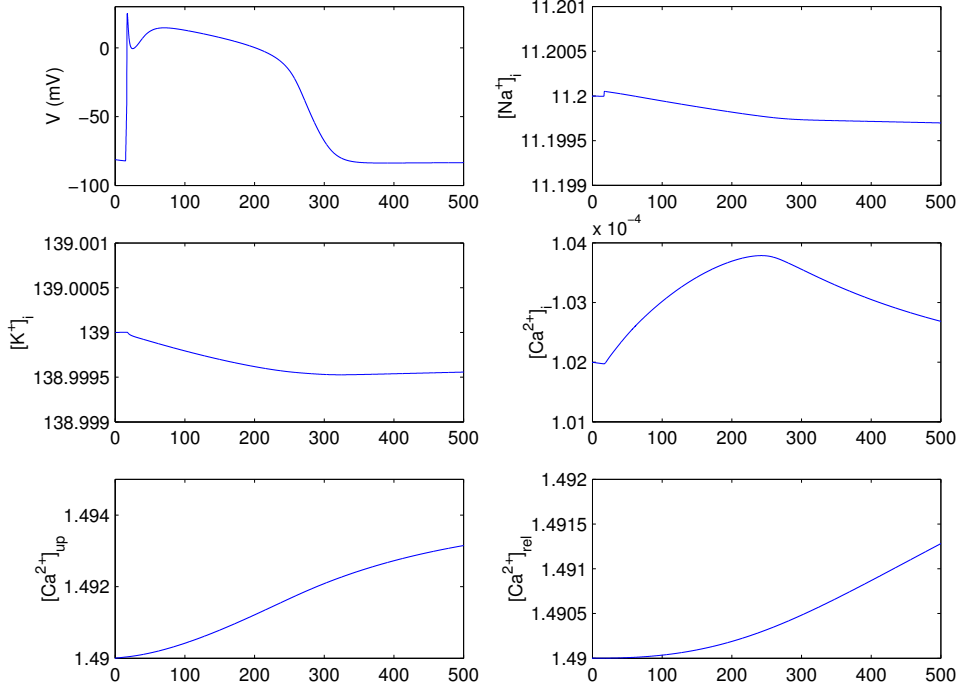


Figure 2: CRN model: potential and concentration variables.

### Na<sup>+</sup>-Ca<sup>2+</sup> exchanger

$$I_{\text{NaCa}} = I_{\text{NaCa(max)}} \frac{\exp\left[\gamma \frac{vF}{RT}\right] [\text{Na}^+]_i^3 [\text{Ca}^{2+}]_o - \exp\left[(\gamma - 1) \frac{vF}{RT}\right] [\text{Na}^+]_o^3 [\text{Ca}^{2+}]_i}{(K_{\text{m,Na}}^3 + [\text{Na}^+]_o^3)(K_{\text{m,Ca}} + [\text{Ca}^{2+}]_o)(1 + k_{\text{sat}} \exp\left[(\gamma - 1) \frac{vF}{RT}\right])}$$

where  $I_{\text{NaCa(max)}} = 1600.0$ ,  $K_{\text{m,Na}}$  and  $K_{\text{m,Ca}}$  are the  $[\text{Na}^+]_i$  and  $[\text{Ca}^{2+}]_o$  half saturation constants, respectively,  $k_{\text{sat}} = .1$  is a saturation factor, and  $\gamma = .35$  is a voltage dependence parameter.

### Background Ca<sup>2+</sup> and Na<sup>+</sup> currents

$$I_{\text{b,Ca}} = g_{\text{b,Ca}}(v - E_{\text{Ca}})$$

$$I_{\text{b,Na}} = g_{\text{b,Na}}(v - E_{\text{Na}})$$

where  $g_{\text{b,Ca}}$  and  $g_{\text{b,Na}}$  are the maximal conductances for Ca<sup>2+</sup> and Na<sup>+</sup>, respectively.

### Dynamics of the concentration variables and the calcium buffers

The concentration variables considered in the CRN model are the internal concentrations  $[\text{Na}^+]_i$ ,  $[\text{K}^+]_i$  and  $[\text{Ca}^{2+}]_i$ , and the uptake and release calcium concentration by the sarcoplasmic reticulum,  $[\text{Ca}^{2+}]_{\text{up}}$ ,  $[\text{Ca}^{2+}]_{\text{rel}}$ . Their dynamics is governed by the following equations

$$\frac{d[\text{Na}^+]_i}{dt} = \frac{-3I_{\text{NaK}} - 3I_{\text{NaCa}} - I_{\text{b,Na}} - I_{\text{Na}}}{FV_i}$$

$$\frac{d[\text{K}^+]_i}{dt} = \frac{2I_{\text{NaK}} - I_{\text{K1}} - I_{\text{to}} - I_{\text{Kur}} - I_{\text{Kr}} - I_{\text{Ks}}}{FV_i}$$

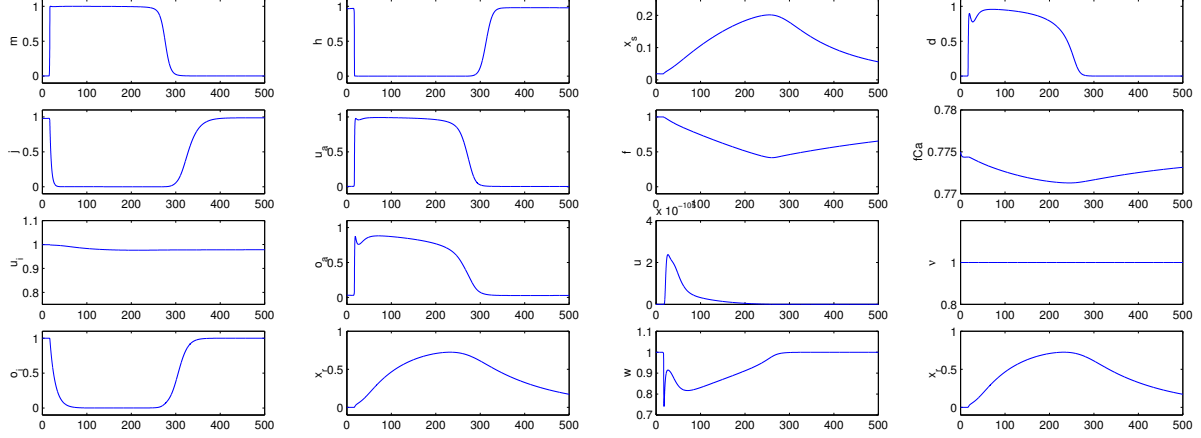


Figure 3: CRN model: gating variables.

$$\frac{d[Ca^{2+}]_i}{dt} = \left[ \frac{2I_{NaCa} - I_{p,Ca} - I_{Ca,L} - I_{b,Ca}}{2FV_i} + \frac{V_{up}(I_{up,leak} - I_{up}) + I_{rel}V_{rel}}{V_i} \right] \times \left[ 1 + \frac{[Trpn]_{max}K_{m,Trpn}}{([Ca^{2+}]_i + K_{m,Trpn})^2} + \frac{[Cmdn]_{max}K_{m,Cmdn}}{([Ca^{2+}]_i + K_{m,Cmdn})^2} \right]^{-1} \quad (3.5)$$

$$\frac{d[Ca^{2+}]_{up}}{dt} = I_{up} - I_{up,leak} - I_{tr} \frac{V_{rel}}{V_{up}} \quad (3.6)$$

$$\frac{d[Ca^{2+}]_{rel}}{dt} = (I_{tr} - I_{rel}) \left[ 1 + \frac{[Csqn]_{max}K_{m,Csqn}}{([Ca^{2+}]_{rel} + K_{m,Csqn})^2} \right]^{-1}, \quad (3.7)$$

where  $V_i$  is the intracellular volume,  $V_{up}$  and  $V_{rel}$  are the volumes of the uptake and release compartments of the sarcoplasmic reticulum (SR),  $[Trpn]_{max}$  and  $[Cmdn]_{max}$  are the total concentrations of troponin and calmodulin in myoplasm,  $[Csqn]_{max}$  is the total concentration of calsequestrin in the release compartment of SR, while  $K_{m,Trpn}$ ,  $K_{m,Cmdn}$  and  $K_{m,Csqn}$  are their half saturation constants, respectively.

The  $Ca^{2+}$  Leak current by the JSR in equations (3.5) and (3.6) above is given by

$$I_{up,leak} = \frac{[Ca^{2+}]_{up}}{[Ca^{2+}]_{up,max}} I_{up,max},$$

where  $[Ca^{2+}]_{up,max}$  and  $I_{up,max}$  are the maximal  $[Ca^{2+}]$  concentration in NSR and the maximal  $[Ca^{2+}]$  uptake rate for  $I_{up}$ , respectively.

The  $Ca^{2+}$  Uptake current by the JSR in equations (3.5) and (3.6) above is given by

$$I_{up} = \frac{I_{up,max}}{1 + (K_{up}/[Ca^{2+}]_i)},$$

where  $K_{up}$  is the  $[Ca^{2+}]_i$  half saturation constant.

The  $Ca^{2+}$  Release current from the JSR in equations (3.5) and (3.7) above is given by

$$I_{rel} = k_{rel}u^2vw([Ca^{2+}]_{rel} - [Ca^{2+}]_i),$$

where  $u$ ,  $v$  and  $w$  are gating variables whose dynamics is given by equation (3.4), with coefficients

$$\tau_u = 8.0 \quad u^\infty = \left[ 1 + \exp \left( -\frac{F_n - 3.4175 \times 10^{-13}}{13.67 \times 10^{-16}} \right) \right]^{-1}$$

$$\tau_v = 1.91 + 2.09 \left[ 1 + \exp \left( -\frac{F_n - 3.4175 \times 10^{-13}}{13.67 \times 10^{-16}} \right) \right]^{-1} \quad v^\infty = 1 - \left[ 1 + \exp \left( -\frac{F_n - 6.835 \times 10^{-14}}{13.67 \times 10^{-16}} \right) \right]^{-1}$$

$$\tau_w = 6.0 \frac{1 - \exp\left(-\frac{v-7.9}{5}\right)}{\left[1 + .3 \exp\left(-\frac{v-7.9}{5}\right)\right] (v - 7.9)} \quad w^\infty = 1 - \left[1 + \exp\left(-\frac{v-40}{17}\right)\right]^{-1},$$

where we have set

$$F_n = 10^{-12} V_{\text{rel}} I_{\text{rel}} - \frac{5 \times 10^{-13}}{F} \left( \frac{1}{2} I_{\text{Ca,L}} - \frac{1}{5} I_{\text{NaCa}} \right).$$

Finally, the *Transfer current from NSR to JSR* in equations (3.6) and (3.7) above is given by

$$I_{\text{tr}} = \frac{[\text{Ca}^{2+}]_{\text{up}} - [\text{Ca}^{2+}]_{\text{rel}}}{\tau_{\text{tr}}},$$

where  $\tau_{\text{tr}} = 180$ .

In Figure 2 and 3 we report the time evolution of the potential and of the gating and concentration variables.

## 4 Variational formulation and discretization of the model

In this section we outline a variational formulation of the monodomain and the bidomain models.

Let  $N$  be the number of gating variables, let  $M$  be the number of concentration variables of the ionic model, and let  $H^1(\Omega)$  be the usual Sobolev space over  $\mathbb{R}$ . The variational formulation of the monodomain model reads as follows.

Given  $v_0, w_0, c_0 \in L^2(\Omega)$ ,  $I^{app} \in L^2(\Omega \times (0, T))$ , find  $v \in W^{1,1}(0, T; H)$   $w \in L^2(0, T; [L^2(\Omega)]^N)$ , and  $c \in L^2(0, T; [L^2(\Omega)]^M)$  such that,  $\forall t \in (0, T)$

$$\begin{cases} c_m \frac{\partial}{\partial t}(v(t), \phi) + a(v(t), \phi) + (I_{ion}(v(t), w(t), c(t)), \phi) = (I^{app}, \phi) & \forall \phi \in H \\ \frac{\partial}{\partial t}(w(t), \psi) = (R(v(t), w(t)), \psi) & \forall \psi \in [L^2(\Omega)]^N \\ \frac{\partial}{\partial t}(c(t), \xi) = (S(v(t), w(t), c(t)), \xi) & \forall \xi \in [L^2(\Omega)]^M, \end{cases} \quad (4.1)$$

with suitable initial conditions on  $v, w, c$ , as given in (2.3). Here  $(\cdot, \cdot)$  and  $a(\cdot, \cdot)$  denote the inner product in  $L^2(\Omega)$

$$(\eta, \chi) = \int_{\Omega} \eta \chi \, dx \quad \forall \eta, \chi \in L^2(\Omega),$$

and the elliptic bilinear form

$$a(\lambda, \mu) = \int_{\Omega} (\nabla \lambda)^T D(\mathbf{x}) \nabla \mu \, dx \quad \forall \lambda, \mu \in H^1(\Omega),$$

respectively.

In a similar way, the variational formulation of the bidomain model can be written as follows.

Given  $v_0, w_0, c_0 \in L^2(\Omega)$ ,  $I^{app} \in L^2(\Omega \times (0, T))$ , find  $u_{i,e} \in W^{1,1}(0, T; H)$ ,  $w \in L^2(0, T; [L^2(\Omega)]^N)$ , and  $c \in L^2(0, T; [L^2(\Omega)]^M)$  such that  $\frac{\partial v}{\partial t} \in L^2(0, T; L^2(\Omega))$ ,  $\frac{\partial w}{\partial t} \in L^2(0, T; [L^2(\Omega)]^N)$ ,  $\frac{\partial c}{\partial t} \in L^2(0, T; [L^2(\Omega)]^M)$ , and  $\forall t \in (0, T)$

$$\begin{cases} c_m \frac{\partial}{\partial t}(v(t), \phi_i) + a_i(u_i(t), \phi_i) + (I_{ion}(v(t), w(t), c(t)), \phi_i) = 0 & \forall \phi_i \in H \\ -c_m \frac{\partial}{\partial t}(v(t), \phi_e) + a_e(u_e(t), \phi_e) - (I_{ion}(v(t), w(t), c(t)), \phi_e) = -(I_e^{app}, \phi) & \forall \phi_e \in H \\ \frac{\partial}{\partial t}(w(t), \psi) = (R(v(t), w(t)), \psi) & \forall \psi \in [L^2(\Omega)]^N \\ \frac{\partial}{\partial t}(c(t), \xi) = (S(v(t), w(t), c(t)), \xi) & \forall \xi \in [L^2(\Omega)]^M \\ v(\mathbf{x}, t) = u_i(\mathbf{x}, t) - u_e(\mathbf{x}, t), \end{cases} \quad (4.2)$$

with suitable initial conditions on  $v, w, c$ , as given in (2.1). Here  $(\cdot, \cdot)$  denotes again the inner product in  $L^2(\Omega)$ , while  $a_{i,e}(\cdot, \cdot)$  denote the elliptic bilinear forms

$$a_{i,e}(\lambda, \mu) = \int_{\Omega} (\nabla \lambda)^T D_{i,e}(\mathbf{x}) \nabla \mu \, dx \quad \forall \lambda, \mu \in H^1(\Omega),$$

respectively.

If, on the one hand, several theoretical results on reaction-diffusion equations can be applied to the monodomain model, on the other hand less is known on degenerate reaction-diffusion systems such as the bidomain model. We refer the reader to [6] for existence, uniqueness and regularity results, both at the continuous and the semi-discrete level, and to [24] for a convergence analysis of finite elements approximations. Both papers deals with the Fitz-Hugh-Nagumo (FHN) model of the gating system.

More results are known on the related eikonal approximation describing the propagation of excitation front (see for instance [2, 3, 11]), and a mathematical analysis of the bidomain model using  $\Gamma$ -convergence theory can be found in [1].

#### 4.1 Semi-discrete formulation

Let  $\mathcal{T}_h$  be a triangulation of  $\Omega \subset \mathbb{R}^d$  ( $d = 2, 3$ ), namely  $\Omega = \bigcup_{j=1}^N K_j$ , where each  $K_j = T_{K_j}(E) \in \mathcal{T}_h$ ,  $E$  is the reference element, a simplex (namely the triangle with vertices  $(0, 0)$ ,  $(1, 0)$ , and  $(0, 1)$  when  $d = 2$  or the tetrahedron with vertices  $(0, 0, 0)$ ,  $(1, 0, 0)$ ,  $(0, 1, 0)$ , and  $(0, 0, 1)$  when  $d = 3$ ) or the unit cube  $[0, 1]^d$  ( $d = 2, 3$ ), and where  $T_{K_j}$  is an invertible affine map. We define  $h$  as the maximum diameter of the elements of the triangulation. The associated finite element spaces  $X_h$  and  $Y_h$  (see e.g. [19] for an introduction to finite element methods) are defined as

$$X_h = \left\{ \varphi_h \in C^0(\Omega) \mid \varphi_h|_{K_j} \circ T_{K_j} \in \mathbb{P}_1(E) \right\}, \quad Y_h = \left\{ \varphi_h \in C^0(\Omega) \mid \varphi_h|_{K_j} \circ T_{K_j} \in \mathbb{Q}_1(E) \right\},$$

where  $\mathbb{P}_1(E)$  is the space of polynomials of degree at most one on  $E$ , whereas  $\mathbb{Q}_1(E)$  is the space of polynomials of degree at most one with respect to each variable on  $E$ .

A semi-discrete problem in space is then obtained by applying a Galerkin procedure, using as finite dimensional space  $V_h = X_h$  or  $V_h = Y_h$ , and choosing a basis for  $V_h$ . Let then  $\{\varphi_i\}$  be the finite element basis, and let  $M = (m_{kl})$ ,  $A = (a_{kl})$  and  $A_{i,e} = (a_{kl}^{i,e})$  be the symmetric mass and stiffness matrices defined by

$$m_{kl} = \sum_{j=1}^N \int_{K_j} \varphi_k \varphi_l \, dx, \\ a_{kl} = \sum_{j=1}^N \int_{K_j} (\nabla \varphi_k)^T D(\mathbf{x}) \nabla \varphi_l \, dx \quad a_{kl}^{i,e} = \sum_{j=1}^N \int_{K_j} (\nabla \varphi_k)^T D_{i,e}(\mathbf{x}) \nabla \varphi_l \, dx.$$

Numerical evaluation of such integrals is obtained by means of a 3rd order Gaussian rule. In the monodomain formulation, the finite elements approximation of the transmembrane potential  $\mathbf{v}_h$  is the solution of

$$c_m M \frac{\partial \mathbf{v}_h}{\partial t} + A \mathbf{v}_h + M I_{ion}^h(\mathbf{v}_h, \mathbf{w}_h, \mathbf{c}_h) = M I_h^{app}. \quad (4.3)$$

On the other hand, the bidomain formulation for the finite element problem can be written in compact form as

$$\chi c_m \mathcal{M} \frac{\partial}{\partial t} \begin{pmatrix} \mathbf{u}_{i,h} \\ \mathbf{u}_{e,h} \end{pmatrix} + \mathcal{A} \begin{pmatrix} \mathbf{u}_{i,h} \\ \mathbf{u}_{e,h} \end{pmatrix} + \chi \begin{pmatrix} M I_{ion}^h(\mathbf{v}_h, \mathbf{w}_h, \mathbf{c}_h) \\ -M I_{ion}^h(\mathbf{v}_h, \mathbf{w}_h, \mathbf{c}_h) \end{pmatrix} = \begin{pmatrix} 0 \\ M I_{e,h}^{app} \end{pmatrix} \quad (4.4)$$

where the matrices  $\mathcal{M}$  and  $\mathcal{A}$  are given by

$$\mathcal{M} = \begin{pmatrix} M & -M \\ -M & M \end{pmatrix} \quad \mathcal{A} = \begin{pmatrix} A_i & 0 \\ 0 & A_e \end{pmatrix}. \quad (4.5)$$

Both equations (4.3) and (4.4) are coupled with the semidiscrete formulation of the dynamics of the gating and concentration variables

$$\frac{\partial \mathbf{w}_h}{\partial t} = R(\mathbf{v}_h, \mathbf{w}_h), \quad \frac{\partial \mathbf{c}_h}{\partial t} = S(\mathbf{v}_h, \mathbf{w}_h, \mathbf{c}_h).$$

The bidomain formulation can be written alternatively in terms of  $\mathbf{v}_h$  and  $\mathbf{u}_{e,h}$  (or  $\mathbf{v}_h$  and  $\mathbf{u}_{i,h}$ ) by adding the two equations and substituting  $\mathbf{u}_{i,h} = \mathbf{v}_h + \mathbf{u}_{e,h}$  in the first equation, obtaining

$$\begin{cases} c_m M \frac{\partial \mathbf{v}_h}{\partial t} + A_i \mathbf{v}_h + A_i \mathbf{u}_{e,h} + M I_{ion}^h(\mathbf{v}_h, \mathbf{w}_h, \mathbf{c}_h) = M I_{i,h}^{app} \\ A_i \mathbf{v}_h + (A_e + A_i) \mathbf{u}_{e,h} = M(I_{i,h}^{app} - I_{e,h}^{app}). \end{cases} \quad (4.6)$$

Equation (4.6) above is a Differential-Algebraic Equation (DAE), that separates the differential variable  $\mathbf{v}_h$  from the algebraic one  $\mathbf{u}_{e,h}$ . Such approach has been firstly used in [2] and [22], and sequently by many other authors.

## 4.2 Fully discrete approximation

In order to have a fully discrete approximation of the problem, we integrate in time systems (4.3) and (4.4) by means of a semi-implicit Euler scheme: the linear diffusion term is discretized implicitly, while the nonlinear reaction term (the ionic current  $I_{ion}$ ) is treated explicitly. The mass matrix  $M$  is lumped to diagonal form by standard techniques. The ordinary differential system for the gating variables is integrated exactly after linearization around the potential at the previous time step.

The  $j$ -th gating variable at time step  $n+1$  is given by

$$\mathbf{w}_j^{n+1} = \mathbf{w}_{j\infty}(\mathbf{v}_h^n) + (\mathbf{w}_j^n - \mathbf{w}_{j\infty}(\mathbf{v}_h^n)) \exp\left(-\frac{\Delta t}{\tau \mathbf{w}_j(\mathbf{v}_h^n)}\right),$$

where

$$\mathbf{w}_{j\infty} = \alpha_j(\mathbf{v}^n) \tau \mathbf{w}_j(\mathbf{v}^n) \quad \tau \mathbf{w}_j(\mathbf{v}^n) = \frac{1}{\alpha_j(\mathbf{v}^n) + \beta_j(\mathbf{v}^n)}$$

The system for the concentration variables is then integrated by a backward Euler scheme

$$\frac{\mathbf{c}_h^{n+1} - \mathbf{c}_h^n}{\Delta t} = S(\mathbf{v}_h^n, \mathbf{w}_h^{n+1}, \mathbf{c}_h^n).$$

This allows us to decouple the ODE system by solving with respect to the gating and concentration variables first, given the potential at the previous time step  $\mathbf{v}_h^n$ , and then solving, in the monodomain case, for  $\mathbf{v}_h^{n+1}$

$$\chi c_m M \frac{\mathbf{v}_h^{n+1} - \mathbf{v}_h^n}{\Delta t} + A \mathbf{v}_h^{n+1} + \chi M I_{ion}^h(\mathbf{v}_h^n, \mathbf{w}_h^{n+1}, \mathbf{c}_h^{n+1}) = M I_h^{app}$$

and, in the bidomain case, for  $\mathbf{u}_{i,h}^{n+1}$  and  $\mathbf{u}_{e,h}^{n+1}$

$$\begin{cases} \chi c_m M \frac{\mathbf{v}_h^{n+1} - \mathbf{v}_h^n}{\Delta t} + A_i \mathbf{u}_{i,h}^{n+1} + \chi M I_{ion}^h(\mathbf{v}_h^n, \mathbf{w}_h^{n+1}, \mathbf{c}_h^{n+1}) = 0 \\ -\chi c_m M \frac{\mathbf{v}_h^{n+1} - \mathbf{v}_h^n}{\Delta t} + A_e \mathbf{u}_{e,h}^{n+1} - \chi M I_{ion}^h(\mathbf{v}_h^n, \mathbf{w}_h^{n+1}, \mathbf{c}_h^{n+1}) = -M I_{e,h}^{app}, \end{cases}$$

where  $\mathbf{v}_h^{n+1} = \mathbf{u}_{i,h}^{n+1} - \mathbf{u}_{e,h}^{n+1}$ . With this choice (notice that one could solve for the potential first and update successively the gating and concentration variables), the semi-implicit method in the monodomain case requires to solve the linear system

$$\chi \left( \frac{c_m}{\Delta t} M + A \right) \mathbf{v}_h^{n+1} = \frac{c_m}{\Delta t} M \mathbf{v}_h^n - \chi M I_{ion}^h(\mathbf{v}_h^n, \mathbf{w}_h^{n+1}, \mathbf{c}_h^{n+1}) + M I_h^{app},$$

while, in the bidoman case, the associated linear system is

$$\left( \frac{\chi c_m}{\Delta t} \mathcal{M} + \mathcal{A} \right) \begin{pmatrix} \mathbf{u}_{i,h}^{n+1} \\ \mathbf{u}_{e,h}^{n+1} \end{pmatrix} = \frac{\chi c_m}{\Delta t} \mathcal{M} \begin{pmatrix} \mathbf{u}_{i,h}^n \\ \mathbf{u}_{e,h}^n \end{pmatrix} - \chi \begin{pmatrix} M I_{ion}^h(\mathbf{v}_h^n, \mathbf{w}_h^{n+1}, \mathbf{c}_h^{n+1}) \\ -M I_{ion}^h(\mathbf{v}_h^n, \mathbf{w}_h^{n+1}, \mathbf{c}_h^{n+1}) \end{pmatrix} + \begin{pmatrix} 0 \\ -M I_{e,h}^{app} \end{pmatrix},$$

where the matrices  $\mathcal{M}$  and  $\mathcal{A}$  are the ones defined in (4.5). The transmembrane potential  $\mathbf{v}_h^{n+1}$  is uniquely determined, as in the continuous model, while  $\mathbf{u}_{i,h}^{n+1}$  and  $\mathbf{u}_{e,h}^{n+1}$  are determined up to the same additive time-dependent constant with respect to a reference potential. Such constant can be determined by imposing the

condition  $\mathbf{1}^T M \mathbf{u}_{e,h}^{n+1} = 0$ .

Notice that the semi-implicit scheme above leads to a linear system with symmetric positive definite matrix in the monodomain case. On the other hand, in the bidomain case, the resulting linear system involves a symmetric positive semidefinite matrix, with a one dimensional kernel spanned by  $(\mathbf{1}, \mathbf{1})^T$ . These systems are solved by a preconditioned conjugate gradient algorithm (PCG), using as initial guess the solution at the previous time step.

General	$\chi = 10^3 \text{ cm}^{-1}$	$C_m = 10^{-3} \text{ mF/cm}^2$
Monodomain	$\sigma_l = 1.2 \cdot 10^{-3} \Omega^{-1} \text{ cm}^{-1}$	$\sigma_t = 2.5562 \cdot 10^{-4} \Omega^{-1} \text{ cm}^{-1}$
Bidomain	$\sigma_l^e = 2 \cdot 10^{-3} \Omega^{-1} \text{ cm}^{-1}$	$\sigma_t^e = 1.3514 \cdot 10^{-4} \Omega^{-1} \text{ cm}^{-1}$
	$\sigma_l^i = 3 \cdot 10^{-3} \Omega^{-1} \text{ cm}^{-1}$	$\sigma_t^i = 3.1525 \cdot 10^{-4} \Omega^{-1} \text{ cm}^{-1}$
FHN model	$G = 1.5 \Omega^{-1} \text{ cm}^{-2}$	$\eta_1 = 4.4 \Omega^{-1} \text{ cm}^{-1}$
	$v_{th} = 13 \text{ mV}$	$\eta_2 = 0.012$
	$v_p = 100 \text{ mV}$	$\eta_3 = 1$
CRN model	As in the original paper [7]	

Table 1: Parameter calibration for the numerical simulations

## 5 Numerical simulations

In this section we describe some numerical simulations. The computational domain is  $\Omega = [0, 1] \times [0, 1]$ , and the problem is discretized by  $\mathbb{P}_1$  elements on an unstructured grid. We consider a two dimensional slab since the thickness of the atrial wall is far less significant than that of the ventricular one, and a two dimensional approximation is thus reasonable. We run monodomain and bidomain simulations, with both the modified FHN and the CRN cell model. In Table 1 we report the parameter calibration for the tests. The numerical simulations are run on MATLAB<sup>®</sup> 6.5.

In the first test we considered a vertex stimulation of the slab. We show the spread of excitation and the repolarization, in Figure 4 for the monodomain-FHN model, and, in Figure 5 for the monodomain-CRN model. For the latter model, the spike-and-dome profile of the Action Potential is more evident, as well as the longer time needed to recover the resting value of the potential, owing to the presence of the plateau phase, that is not captured by the simple FHN model.

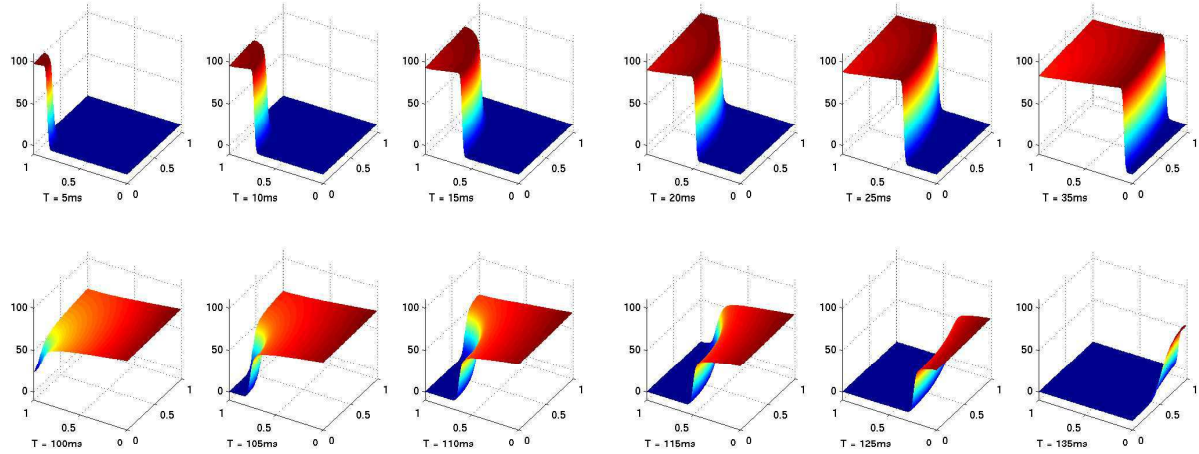


Figure 4: Monodomain FHN on a 2d slab: excitation and repolarization

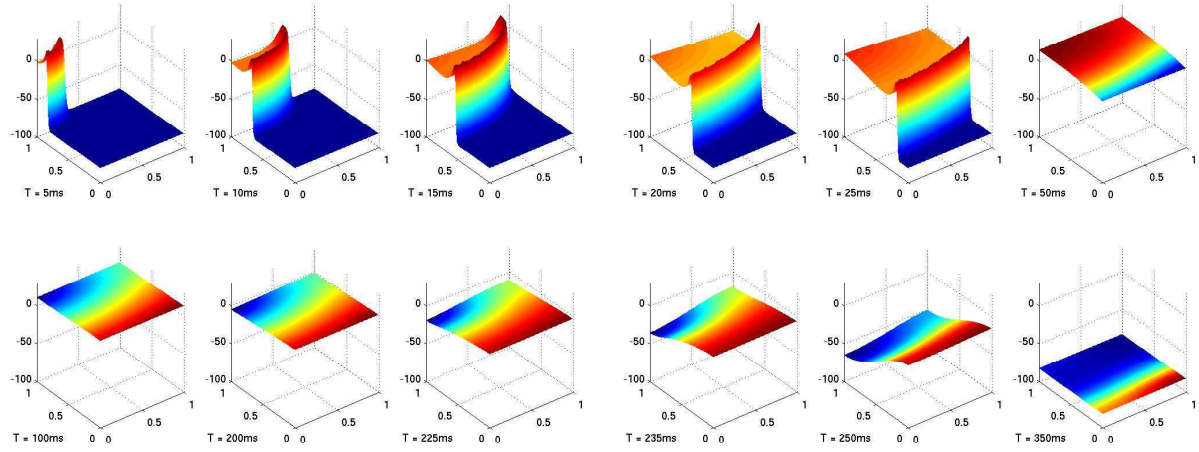


Figure 5: Monodomain CRN on a 2d slab: excitation and repolarization

In the second test we considered a central stimulation of the slab. In Figure 6 we show the transmembrane potential  $v$  as well as the intracellular and extracellular potentials  $u_i$  and  $u_e$ , for the bidomain FHN model, at point  $(.2333, .5667)$  of the slab. In Figure 7 we show the spread of excitation and the action potential at point  $(.8, .7)$  for the bidomain CRN model.

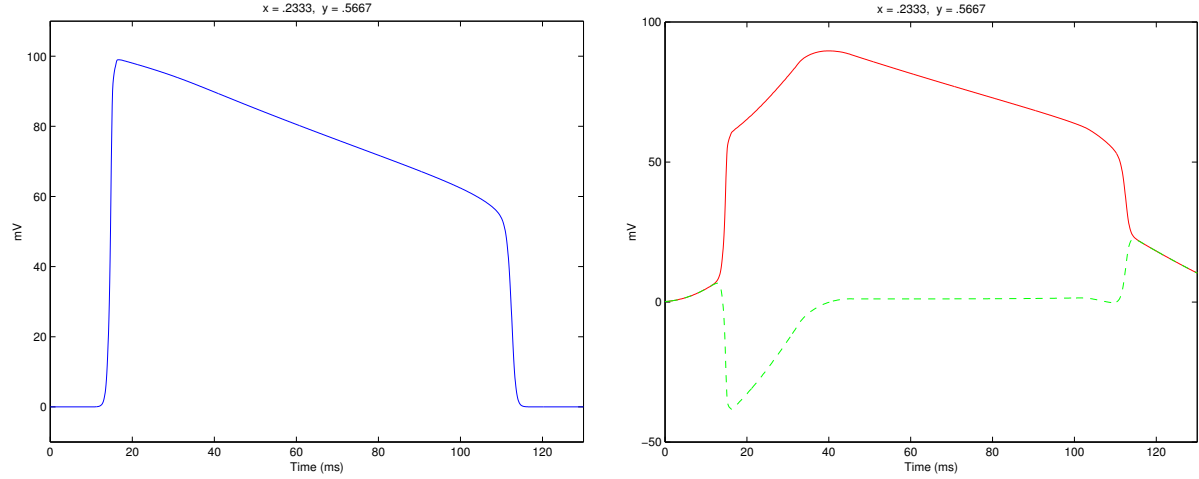


Figure 6: Bidomain FHN on a 2d slab: transmembrane potential  $v$  (left), intracellular  $u_i$  (solid line) and extracellular  $u_e$  (dotted line) potential at point  $(.2333, .5667)$

In the third test, in order to simulate the presence of arteries or veins, we considered a two dimensional slab with a circular hole embedded. Since the vessel walls (both arterious and venous) are not excitable, we impose a non-conducting condition ( $\mathbf{n} \cdot \nabla v = 0$ ) on the border of the hole. We show the spread of excitation and the repolarization for the monodomain FHN model in Figure 8, and for the monodomain CRN in Figure 9. In Figure 10, a closer look allows to better appreciate the distortion of the wavefront in the neighbourhood of the hole for both the FHN and CRN models.

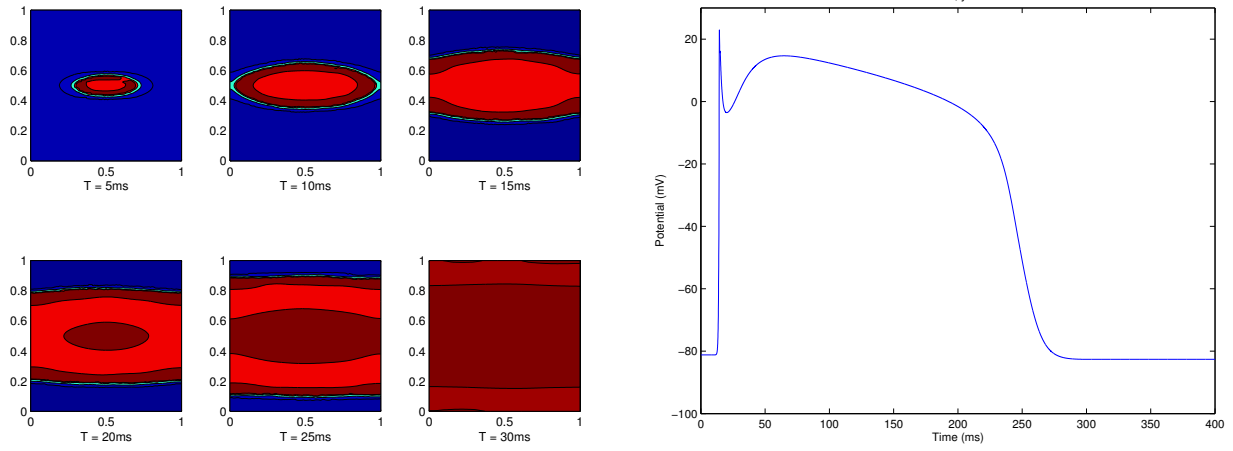


Figure 7: Bidomain CRN on a 2d slab: excitation spread (left) and transmembrane potential (right)

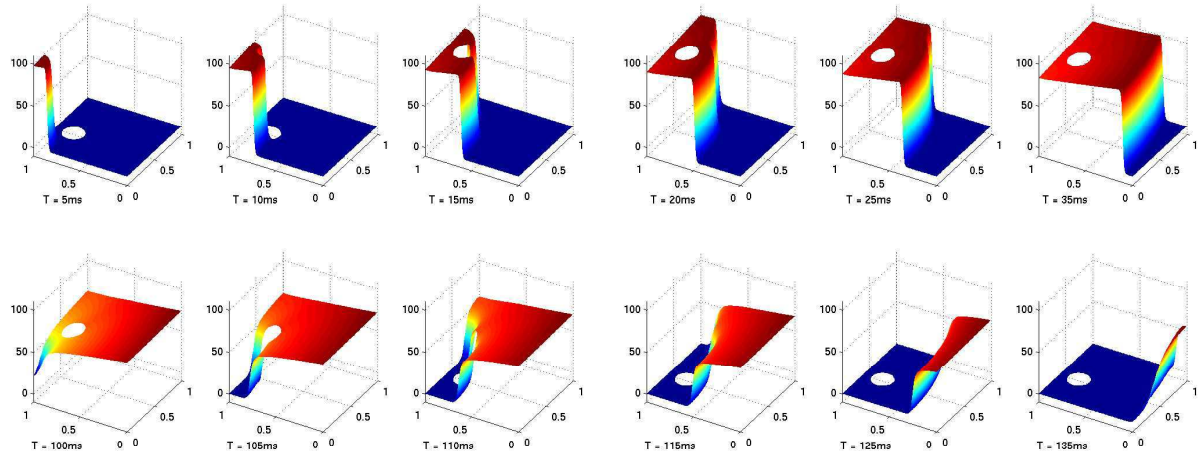


Figure 8: Monodomain FHN on a 2d slab with an hole: excitation and repolarization



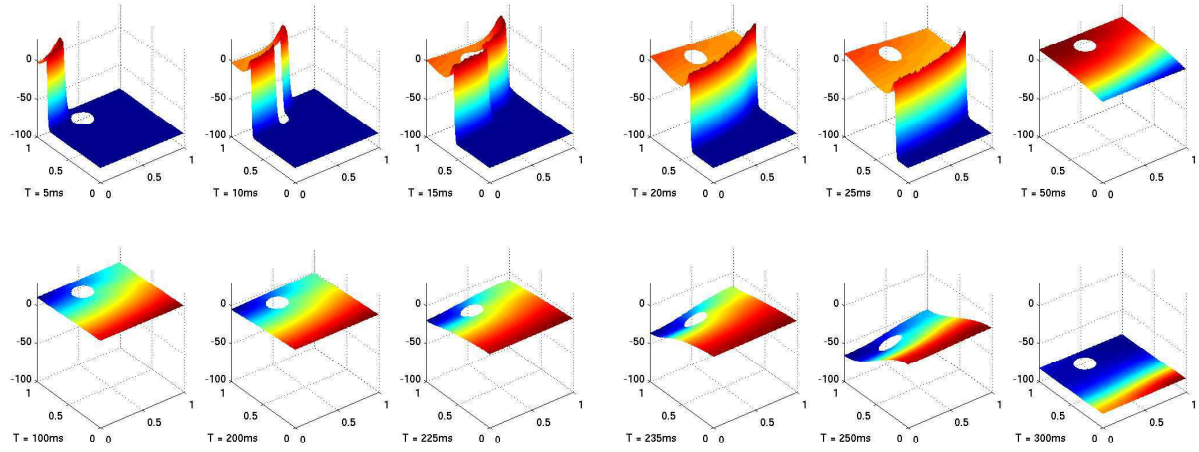


Figure 9: Monodomain CRN on a 2d slab with an hole: excitation and repolarization

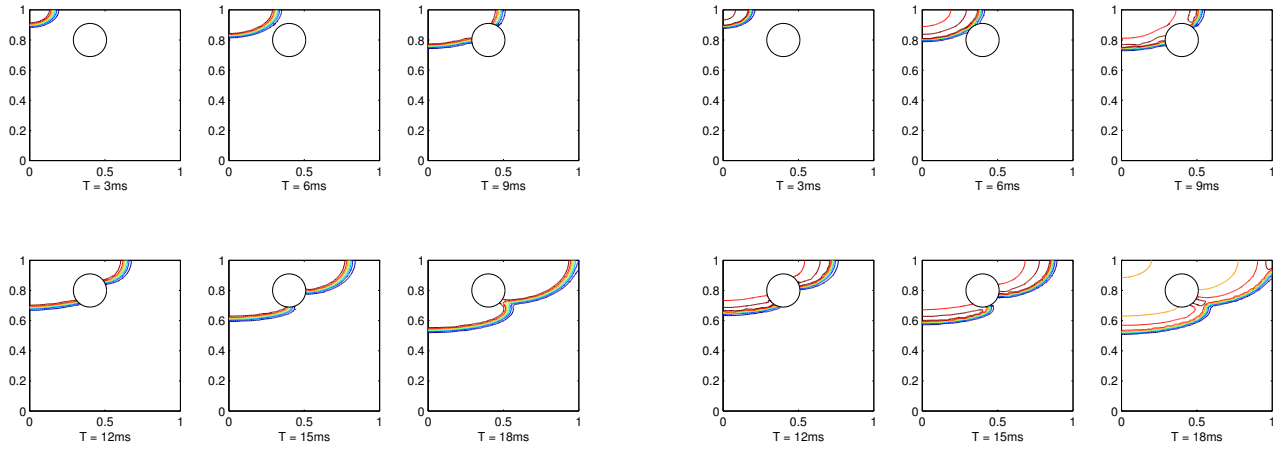


Figure 10: Propagation wavefront around an hole: monodomain FHN (left) and CRN (right) models

Finally, to analyze the cost of the two models, we compare their computational complexity in the first two test cases. The slab is discretized by an unstructured triangular grid consisting of 7744 nodes and 15166 elements. We report in Table 2 the CPU time needed to simulate complete a heartbeat (around 400msec) on a Dell Poweredge 4600 server. The higher level of accuracy in the description of the Action Potential provided by the CRN model requires a smaller step in the time integration. Moreover, at each time step with the CRN ionic model, one has to update 15 gating and 5 concentration variables on each point of the grid, whereas with the FHN model only one gating variable has to be updated at each point of the grid. No adaptivity is introduced in the integration scheme for this comparison, and the fixed time steps are  $\Delta t = 0.1\text{msec}$  for the FHN models, and  $\Delta t = 0.05\text{msec}$  for the CRN models.

FHN		CRN	
Monodomain	Bidomain	Monodomain	Bidomain
3.1e+03	5.8e+03	5.6e+04	7.2e+04

Table 2: CPU time (in sec) for monodomain and bidomain models to simulate a complete heartbeat

## 6 Conclusions

We presented here an approach to simulate the propagation of the excitation fronts in the atrial cells, based on nonlinear models of reaction-diffusion type, considering both the monodomain and the bidomain approach. The ionic currents are expressed by the simple modified FHN model (in the Rogers-McCulloch variant), and by the more sophisticated CRN model, especially designed for human atrial cells. Numerical simulations on a two dimensional slab are given to show the behaviour of the excitation spread and the repolarization phase. Future work will focus on more general domains, on the simulation of spiral waves, and on the coupling of these models with models of pacemaker cells as the ones provided by O. Doessel and his collaborators (see for instance [25]).

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