

# Biological evidence supports an early and complex emergence of the Isthmus of Panama

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**The linking of North and South America by the Isthmus of Panama had major impacts on global climate, oceanic and atmospheric currents, and biodiversity, yet the timing of this critical event remains contentious. The Isthmus is traditionally understood to have fully closed by ca. 3.5 million years ago (Ma), and this date has been used as a benchmark for oceanographic, climatic, and evolutionary research, but recent evidence suggests a more complex geological formation. Here, we analyze both molecular and fossil data to evaluate the tempo of biotic exchange across the Americas in light of geological evidence. We demonstrate significant waves of dispersal of terrestrial organisms at approximately ca. 20 and 6 Ma and corresponding events separating marine organisms in the Atlantic and Pacific oceans at ca. 23 and 7 Ma. The direction of dispersal and their rates were symmetrical until the last ca. 6 Ma, when northern migration of South American lineages increased significantly. Variability among taxa in their timing of dispersal or vicariance across the Isthmus is not explained by the ecological factors tested in these analyses, including biome type, dispersal ability, and elevation preference. Migration was therefore not generally regulated by intrinsic traits but more likely reflects the presence of emergent terrain several millions of years earlier than commonly assumed. These results indicate that the dramatic biotic turnover associated with the Great American Biotic Interchange was a long and complex process that began as early as the Oligocene–Miocene transition.**

biogeography | evolution | neotropics | fossil | migration

**T**he Isthmus of Panama is the narrow strip of land that connects North and South America and divides the Atlantic and Pacific oceans. The emergence of the Isthmus initiated one of the largest episodes of biological migration between previously disconnected landmasses, the Great American Biotic Interchange (GABI) (1–5), one of the best natural experiments on invasive species. The closure of the Central American Seaway (CAS, the oceanic pathway along the tectonic boundary between South America and the Panama Block) and rise of the Isthmus have been linked to the onset of both thermohaline oceanic circulation and northern hemisphere glaciation (6–8). Despite its broad importance, the formation of the Isthmus and its impact on the rich biodiversity of the Americas remains contentious (9). Therefore, a better understanding of when the formation of the Isthmus of Panama occurred has important implications in several scientific fields across multiple disciplines.

The timing of Isthmus formation has been assessed through different proxies. Previous studies have long suggested full closure by 3.5 Ma (7, 8, 10–16). More recent geological work has suggested a longer and more complex formation, where the initial collision between South America and the Panama Block occurred between 25 and 23 Ma (17). By 20 Ma the Panama Block is suggested to have been connected to North America (18–21) and the width of the CAS to be 200 km (19, 20). Full closure of the CAS occurred by 10 Ma, ending the exchange of

deep and intermediate waters between the Caribbean and the Pacific (11, 19, 20, 22–24). However, the exchange of shallow waters between these oceans likely continued along pathways other than the CAS for many millions of years (7, 8, 10–16, 25).

Over the past two decades, hundreds of studies have assumed the Isthmus of Panama to have closed at 3.5 Ma (*SI Appendix*), causing the separation of widespread marine populations into distinct Pacific and Caribbean groups (vicariance) and the first possible dispersals between North and South America (with the exception of stochastic long-distance dispersals). Here, we address the following questions: Given the complexities and the recent evidence of a much older geological history of the Isthmus of Panama, is 3.5 Ma an adequate age for those events? Were the suggested water corridors across the Isthmus—even if shallow and narrow—indeed effective barriers against both dispersal of terrestrial organisms and conduits of marine ones until a full closure at 3.5 Ma? We address these questions using comprehensive biological data from living and fossil organisms, where the assumption is that any well-developed terrestrial corridor would lead to both more frequent biotic dispersal between North and South America as well as a division of widespread marine organisms into distinct Caribbean and Pacific lineages (26). Biological data provide a powerful tool for this purpose compared with geological evidence, which cannot inform on the subtle

## Significance

**The formation of the Isthmus of Panama, which linked North and South America, is key to understanding the biodiversity, oceanography, atmosphere, and climate in the region. Despite its importance across multiple disciplines, the timing of formation and emergence of the Isthmus and the biological patterns it created have been controversial. Here, we analyze molecular and fossil data, including terrestrial and marine organisms, to show that biotic migrations across the Isthmus of Panama began several million years earlier than commonly assumed. An earlier evolution of the Isthmus has broad implications for the mechanisms driving global climate (e.g., Pleistocene glaciations, thermohaline circulation) as well as the rich biodiversity of the Americas.**

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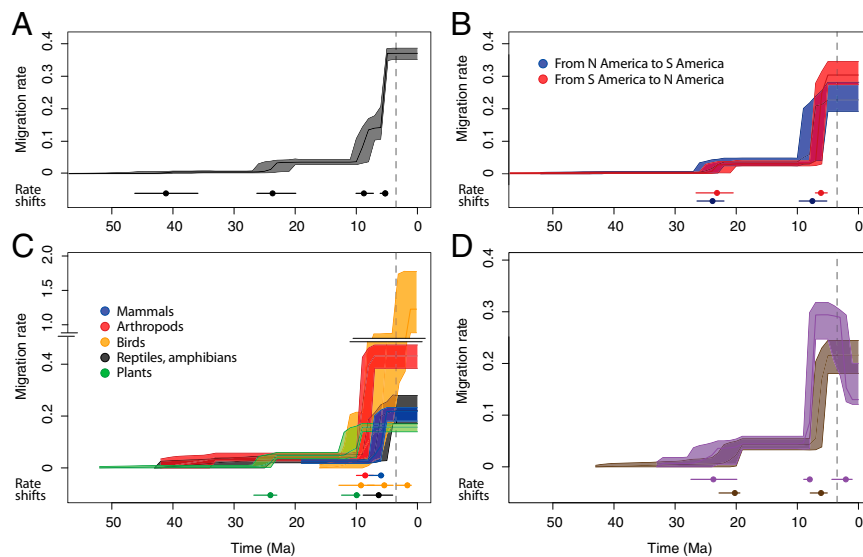
Freely available online through the PNAS open access option.

Data deposition: Raw data for phylogenies and fossil data have been deposited in Dryad, [www.datadryad.org](http://www.datadryad.org) (10.5061/dryad.6m653).

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**Fig. 1.** MRTT estimated from molecular data. The plots show rescaled migration rate for dispersal (A–C) and vicariance (D) events across the Isthmus of Panama. The rates and their temporal dynamics were estimated using maximum likelihood and 1,000 randomizations to infer 95% confidence intervals of MRTT (shaded areas) (see also *SI Appendix, Fig. S1* for a sensitivity analysis) and the time of each rate shift with their confidence intervals (circles and bars at the bottom of each panel). Dispersal between North and South America was calculated for (A) the entire dataset, (B) major taxonomic groups, and (C) direction of dispersal. (D) Comparison between vicariant events separating marine organisms in the Caribbean and the Pacific and the dispersal of terrestrial vertebrates (mammals, amphibians, nonavian reptiles, and birds) between North and South America. The number of rate shifts was selected by model testing via AICc (Table 1 and *SI Appendix, Table S10*). The dashed line indicates the generally accepted emergence of the Isthmus of Panama at 3.5 Ma.

This is shown by an improvement in the second-order AIC by several orders of magnitude (Table 1). Results from these tests demonstrate that cross-Isthmian migrations occurred in incremental pulses over a long time period, presumably correlated with the stepwise formation and emergence of the Isthmus of Panama and in stark contrast to the assumption that a majority of events occurred at, or shortly after, a 3.5 Ma closure (dashed line in Fig. 1).

We split the molecular data to compare terrestrial dispersal patterns with those of marine vicariance events. We found that terrestrial migration exhibited significant increases at ~20.23 (19.4–22.81) and 6.12 (5.1–7.89) Ma, whereas corresponding events separating marine organisms occurred at 23.73 (19.9–27.41) and 7.96 (7.75–8.96). Migration rates in marine organisms decreased by 50% after the third inferred shift at 2.06 (1.03–4.35) Ma.

We further compared our molecular results with inferences from fossil mammals, which provide the most abundant and best-studied fossil record. Although the number of fossil occurrences varies considerably through time due to differential sampling efforts and preservation rates (Fig. 2A), there is no bias toward particular time periods at the species level, except for fossils 1 million years or younger (*SI Appendix, Table S2*). The plots of the fossil data (Fig. 2B and C) show that migration events occurred with increasing frequency since ca. 10 Ma and a drastic increase in North American taxa migrating south occurred over the last 3 Ma. The common explanation of this pattern is complete closure of the Isthmus and a competitive advantage of North American mammals over South American counterparts (1). However, we consider equally possible the scenario that the major drop in global temperatures in the last 3 Ma (Fig. 2C, blue curve) acted as the triggering mechanism driving North American mammals southwards, as well as leading to the onset of northern hemisphere glaciations and rapid vegetation changes in North America. Dropping sea levels in the same period (Fig. 2C, yellow curve) likely increased land exposure along coastal plains, which should have further facilitated migration.

**Asymmetry in the Direction of Dispersal Events.** The directionality of migrations has long been an intriguing aspect of the GABI (2). In our analyses based on phylogenetic data, the asymmetric model (where migration rates and times of shift were considered as independent parameters) strongly outperformed the symmetric model [corrected  $\Delta\text{AIC}$  ( $\Delta\text{AICc}$ ) = 13.96; *SI Appendix, Table S3*]. This suggests that the migrations from South America to North America and in the opposite direction followed different dynamics. However, the estimated rates through time and the respective confidence intervals show that the rates in the two directions were largely overlapping across the long geological time and both underwent a similar rate shift around the transition between the Oligocene and Miocene (ca. 23 Ma; *SI Appendix, Table S3* and Fig. 1B). Migration rates only started to differ significantly after the most recent rate shift (around 6–7 Ma). During this time frame, lasting until the present, the overall migration rate from South America to North America is found to be around 30% higher than the rate from North America to South America.

Our analyses of the fossil mammal data recover a similar number of migrant lineages in South and North America between 11 and 3 Ma; however, these represent a higher fraction of the sampled diversity in South America (Fig. 2B and C and *SI Appendix, Table S2*). A large increase in the proportion of North American lineages in South America is observed over the last 3 Ma, resulting in about 45% of the sampled diversity of South American species (Fig. 2C). This diversity is likely affected by our definition of migrant lineages and reflects the combined effects of migration and in situ diversification.

The asymmetry detected in the fossil record contrasts with the cross-taxonomic results from molecular phylogenies, where migrations in either direction were not found to be significantly different until ca. 6 Ma, when the migration rate from South to North America exceeds that in the opposite direction (Fig. 1B and *SI Appendix, Table S3*). The causes for this discrepancy are still uncertain. The fossil record of South American mammals is mostly derived from temperate latitudes and is heavily biased toward large body-sized grazer/browser animals, whereas the extant mammal data primarily include taxa inhabiting tropical





biogeographic event, we also recorded mean crown age, the lower and upper confidence intervals of ages, and direction of dispersal, as well as the taxon's altitudinal range, dispersal capability, and biome of occurrence.

**MRTT Analyses.** Biogeographic events with associated age uncertainties were analyzed in a maximum likelihood framework to estimate rates of migration (*SI Appendix, Table S10*) and their variation through time. Although, ideally, such migration rates should be estimated on a per-capita basis (as for other macroevolutionary rates), their estimation in this context would be affected by unobserved extinct lineages and further biases would likely arise from combining data from multiple independent phylogenies. We therefore modeled migrations as random events resulting from a stochastic Poisson process, with a rate parameter that describes the expected waiting time between successive events. As a mean to standardize the migration rates across datasets, we rescaled the estimated absolute Poisson rates by the number of families considered in each analysis. Thus, the migration rates shown in the MRTT plots indicate the expected number of migration events per Ma per family. We emphasize that this standardization is not intended as a correction for temporal biases within each dataset but rather as a tool to facilitate comparisons across different analyses. To account for deviations from a constant rate (homogeneous) process, we tested different non-homogeneous Poisson processes with time-varying rates, including a model with exponentially increasing rates, which might capture potential biases owed to the exponential increase of lineages in extant-taxa phylogenies. To allow for temporal changes in the migration rates, we implemented a non-homogeneous Poisson process, in which rate shifts can occur through time. Our maximum likelihood algorithm involved (i) assessing the best-fitting number of rate shifts by a stepwise AICc procedure, (ii) the optimization of their temporal placement, and (iii) the estimation of the migration rates between shifts (*SI Appendix, 1.7*). We used simulations to assess the most appropriate AICc thresholds, thus minimizing the risk of false positives in our analyses (*SI Appendix, 1.6*). We generated and analyzed 1,000 datasets resampled from the uncertainty intervals for migration dates to generate the MRTT plots and calculate 95% confidence intervals.

In addition to jointly analyzing the full molecular dataset, we repeated the analyses on 14 subsets based on different criteria considering geographical, ecological, and taxonomic aspects. These tests allowed us to investigate the differential patterns of migrations linked to direction of migration and environment (terrestrial or marine) and to compare different taxonomic groups and their dispersal ability (Fig. 1*B* and *SI Appendix, Figs. S2–S4* and *1.8*). Additionally, we compared the AIC scores of two migration models with a model containing a single rate shift fixed at 3.5 Ma and a model containing two shifts in the Miocene, at 25 Ma (initial collision of the Panama block and South America) and 10 Ma (closure of the CAS) (*SI Appendix, 1.8* and Table 1).

**GLMM Analyses.** We used a GLMM approach on the molecular datasets of terrestrial/freshwater and marine taxa to test for intrinsic (biological, ecological) factors driving the differential timing of dispersal or vicariance. Ecological and biological variables (biome type, dispersal ability, dispersal direction, and elevation preference as derived from the literature and consultation with specialists) were treated as fixed effects, and we included a phylogenetic correction by treating taxonomic rank as a nested random effect (*SI Appendix, Tables S4–S7*). Because the molecular dates in the dataset were calibrated using a range of approaches (e.g., fossils, molecular clocks), we included calibration type as an additional variable in the model to account for any potential bias associated with how the phylogenies were time-calibrated.

**Fossil Data.** We compiled fossil occurrence data from all available publications of South American mammalian fossils and reviewed the Cenozoic American mammal fossil record in the Paleobiology Database (<https://paleobiodb.org/#/>), synthesizing data from across the entire American continent. The vetted dataset comprised 23,090 fossil records from 112 families and 3,589 species. Based on first and last appearances of each species in the fossil record, we plotted diversity trajectories through time, by counting the number of species within 1 Ma bins. To explicitly incorporate dating uncertainty of fossils, we randomized the age of each fossil 1,000 times within the temporal boundaries of the geological formation in which they were found (Fig. 2*B* and *SI Appendix, 1.4*). We classified each fossil occurrence as North or South American if the taxon or its ancestor was in either North or South America before 10 Ma, following Carillo et al. (37). We then plotted the amount of immigrant species as a proportion of the total diversity in each continent within 1 Ma time bins (Fig. 2*C*).

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1. Simpson GG (1980) *Splendid Isolation: The Curious History of South American Mammals* (Yale Univ Press, New Haven, CT).
2. Stehli FG, Webb SD (1985) *The Great American Biotic Interchange* (Plenum Press, New York).
3. Wallace AR (1876) *The Geographical Distribution of Animals. With a Study of the Relations of Living and Extinct Faunas as Elucidating the Past Changes of the Earth's Surface* (Macmillan and Co., London).
4. Webb SD (1991) Ecogeography and the Great American Interchange. *Paleobiology* 17(3):226–280.
5. Webb SD, Rancy A (1996) Late Cenozoic evolution of the neotropical mammal fauna. *Evolution and Environment in Tropical America*, eds Jackson JBS, Budd AF, Coates AG (University of Chicago Press, Chicago), pp 335–358.
6. Haug GH, et al. (2005) North Pacific seasonality and the glaciation of North America 2.7 million years ago. *Nature* 433(7028):821–825.
7. Haug GH, Tiedemann R (1998) Effect of the formation of the Isthmus of Panama on Atlantic Ocean thermohaline circulation. *Nature* 393(6686):673–676.
8. Haug GH, Tiedemann R, Zahn R, Ravelo AC (2001) Role of Panama uplift on oceanic freshwater balance. *Geology* 29(3):207–210.
9. Stone R (2013) Battle for the Americas. *Science* 341(6143):230–233.
10. Coates AG, Aubry MP, Berggren WA, Collins LS, Kunk M (2003) Early Neogene history of the Central American arc from Bocas del Toro, western Panama. *Geol Soc Am Bull* 115(3):271–287.
11. Coates AG, Collins LS, Aubry MP, Berggren WA (2004) The geology of the Darien, Panama, and the Miocene-Pliocene collision of the Panama arc with northwestern South America. *Geol Soc Am Bull* 116(11):1327–1344.
12. Coates AG, et al. (1992) Closure of the Isthmus of Panama: The near-shore marine record of Costa Rica and western Panama. *Geol Soc Am Bull* 104(7):814–828.
13. Coates AG, Obando JA (1996) The geological evolution of the Central American Isthmus. *Evolution and Environment in Tropical America*, eds Jackson JBS, Budd AF, Coates AG (University of Chicago Press, Chicago).
14. Coates AG, Stallard RF (2013) How old is the Isthmus of Panama. *Bull Mar Sci* 89(4): 801–814.
15. Duque-Caro H (1990) The Choco block in the northwestern corner of South America: Structural, tectonostratigraphic, and paleogeographic implications. *J S Am Earth Sci* 3(1):71–84.
16. Duque-Caro H (1990) Neogene stratigraphy, paleoceanography and paleobiogeography in northwest South America and the evolution of the Panama Seaway. *Palaeogeogr Palaeoclimatol Palaeoecol* 77(3–4):203–234.
17. Farris DW, et al. (2011) Fracturing of the Panamanian Isthmus during initial collision with South America. *Geology* 39(11):1007–1010.
18. MacFadden BJ (2010) Extinct peccary “Cynorca” occidentale (Tayassuidae, Tayassuinae) from the Miocene of Panama and correlations to North America. *J Paleontol* 84(2): 288–289.
19. Montes C, et al. (2012) Arc-continent collision and oroclinal formation: Closing of the Central American Seaway. *J Geophys Res* 117(B4):4105.
20. Montes C, et al. (2012) Evidence for middle Eocene and younger land emergence in central Panama: Implications for Isthmus closure. *Geol Soc Am Bull* 124(5–6):780–799.
21. Rincon A, Bloch JJ, Suarez C, MacFadden BJ, Jaramillo CA (2012) New Floridatragulines (Mammalia, Camelidae) from the Early Miocene Las Cascadas Formation, Panama. *J Vertebr Paleontol* 32(2):456–475.
22. Sepulchre P, et al. (2014) Consequences of shoaling of the Central American Seaway determined from modeling Nd isotopes. *Paleoceanography* 29(3):176–189.
23. Vargas C, Mann P (2013) Tearing and breaking off of subducted slabs as the result of collision of the Panama Arc-indenter with northwestern South America. *Bull Seismol Soc Am* 103(3):2025–2046.
24. Montes C, et al. (2015) Middle Miocene closure of the Central American Seaway. *Science* 348(6231):226–229.
25. Gutiérrez-García TA, Vázquez-Domínguez E (2013) Consensus between genes and stones in the biogeographic and evolutionary history of Central America. *Quat Res* 79(3):311–324.
26. Lessios HA (2008) The great American schism: Divergence of marine organisms after the rise of the Central American Isthmus. *Annu Rev Ecol Syst* 39(1):63–91.
27. Cody S, Richardson JE, Rull V, Ellis C, Pennington RT (2010) The Great American Biotic Interchange revisited. *Ecography* 33(2):326–332.

28. Smith BT, Klicka J (2010) The profound influence of the Late Pliocene Panamanian uplift on the exchange, diversification, and distribution of New World birds. *Ecography* 33(2):333–342.
29. Weir JT, Bermingham E, Schluter D (2009) The Great American Biotic Interchange in birds. *Proc Natl Acad Sci USA* 106(51):21737–21742.
30. Bacon CD, Mora A, Wagner WL, Jaramillo CA (2013) Testing geological models of evolution of the Isthmus of Panama in a phylogenetic framework. *Bot J Linn Soc* 171(1):287–300.
31. Smith BT, Amei A, Klicka J (2012) Evaluating the role of contracting and expanding rainforest in initiating cycles of speciation across the Isthmus of Panama. *Proc Biol Sci* 279(1742):3520–3526.
32. Akaike H (1974) A new look at the statistical model identification. *IEEE Trans Automat Contr* AC-19(6):716–723.
33. Jaramillo CA, et al. (2014) Palynological record of the last 20 million years in Panama. *Paleobotany and Biogeography: A Festschrift for Alan Graham in His 80th Year*, eds Stevens WD, Montiel OM, Raven P (Missouri Botanical Garden Press, St. Louis, MO), pp 134–253.
34. Sanmartín I, Ronquist F (2004) Southern hemisphere biogeography inferred by event-based models: Plant versus animal patterns. *Syst Biol* 53(2):216–243.
35. Müller RD, Sdrólías M, Gaina C, Steinberger B, Heine C (2008) Long-term sea-level fluctuations driven by ocean basin dynamics. *Science* 319(5868):1357–1362.
36. McLoughlin S (2001) The breakup history of Gondwana and its impact on pre-Cenozoic floristic provincialism. *Aust J Bot* 49(3):271–300.
37. Carillo JD, Forasiepi A, Jaramillo CA, Sanchez-Villagra MR (2015) Neotropical mammal diversity and the Great American Biotic Interchange: Spatial and temporal variation in South America's fossil record. *Front Genet* 5:451.
38. Hansen J, et al. (2008) Target atmospheric CO<sub>2</sub>: Where should humanity aim? *Open Atmospheric Science* 2:217–231.
39. Miller KG, et al. (2005) The Phanerozoic record of global sea-level change. *Science* 310(5752):1293–1298.

**Supporting Information:**

Materials and Methods, Results and Discussion

Figures S1-S4

Tables S1-S10

References (1-26)

**SUPPORTING INFORMATION****1. SUPPORTING MATERIALS AND METHODS****1.1 Assumptions about the Isthmus of Panama**

Hundreds of researchers have assumed a closure date for the Isthmus of Panama between 3.5 and 3.0 Ma over the last two decades. The papers most cited for this geological date are essentially from two research groups, Coates ([1-5](#)) and Haug ([6, 7](#)). Taken together, these papers alone have been cited 2092 times, according to Google Scholar (accessed March 4, 2015).

**1.2 Molecular data compilation and analyses**

We used molecular phylogenetic studies that included extant New World species published or in press until May 2013 in 29 journals (Tables S8 and S9). We identified 424 divergence time estimates in 169 phylogenies that represented unambiguous instances of dispersal across the Isthmus of Panama or vicariance events of marine taxa across the Caribbean and Pacific oceans. Our proxy for dispersal time, mean crown ages

and the confidence intervals around them, were gleaned from each phylogeny. Crown groups are defined as the least inclusive monophyletic group that contains all the extant members of a clade ([sensu 8](#)). We adopted the approach of Hoorn et al. ([9](#)) for data mining and included studies that were directly or indirectly dated with fossil calibration or known rates of nucleotide substitution and excluded studies with calibrations based on geologic events, such as the closure of the Panama Isthmus.

### **1.3 Fossil data generation**

To compile fossil mammal data, searches were conducted in Web of Knowledge (<http://wokinfo.com>) and Georef (<http://www.agiweb.org/georef/>). The key terms used in the searches included: fossil mammal(s), South America, geological epochs from the Paleocene to the Pleistocene, and the names of all South American countries. New fossil occurrences were added to the Paleobiology Database (PBDB; <http://paleodb.org/>). From this database, a total of 32,342 fossil mammal records were gathered for this study. We assigned the origin of each fossil mammal occurrence to its respective family or order using information from the fossil record and the literature.

### **1.4 Fossil data analysis**

All undetermined fossil occurrences and all records that had not been identified to species level were removed, leaving 23,090 occurrences in the analyses. The final data set (available from <http://datadryad.org>) included 3,589 species, of which 2,657 (21,104 occurrences) were found in North America and 958 (1,986 occurrences) in South America (Fig. 2A). We classified each fossil occurrence as North or South American if



the taxon or its ancestor(s) were in either North or South America before 10 Ma following Carillo et al. ([10](#)) and identified 50 South American clades in North America (494 occurrences) and 182 North American lineages in South America (439 occurrences).

All fossil occurrences were provided with minimum and maximum ages, derived from the temporal ranges of the respective geological units and reflecting the degree of uncertainty in their dating. These were treated as confidence intervals and used as uniform ranges from which the ages of the occurrences were randomly drawn ([11](#)). The ages of the fossil occurrences of each species were randomly resampled 1,000 times from the respective ranges and used to determine the first and last appearances of species, while accounting for the dating uncertainties of the fossil record. Based on the first and last appearances, we calculated the raw number of species in North and South America within time bins of 1 Myr and plotted diversity trajectories through time with 95% confidence intervals obtained from the 1,000 randomized samples (Fig. 3B, Table S2). Based on these counts the proportions of migrant species in the two continents were also quantified for each time bin (Fig. 2C, Table S2).

### **1.5 Re-estimation of divergence times**

A number of marine lineages exhibit trans-Isthmian divergences that are assumed to have taken place concomitantly with the final closure of the Isthmus of Panama ([reviewed in 12](#)). The genetic distances among these geminate species pairs have been used to calibrate molecular clocks based on a closure of the Isthmus of Panama between 3.5 and 3.0 Ma. This molecular clock has then been used to estimate the divergence times of

sister pairs that occur in the marine ecosystems on each side of the Isthmus (e.g. [13](#), [14](#), [15](#)). Because we did not include molecular dates calibrated with any estimate of Isthmus closure, we had to estimate divergence dates for marine taxa using an independent rate in order to include these taxa in our study.

We obtained genetic distances (Kimura's two-parameter distance) for trans-isthmian species pairs from Lessios (25) and applied a standard molecular clock rate of 2% mitochondrial sequence divergence per million years to convert values to absolute time. This molecular clock has substantial uncertainty surrounding it, thus divergence times for these species pairs should thus be considered as coarse approximations. Although molecular dating based on secondary rate calibrations is prone to potential caveats (e.g. [16](#), [17](#)), re-estimation allowed for a comprehensive synthesis on trans-isthmian divergences in both marine and terrestrial systems.

### **1.6 Estimating migration rates from dated migration events**

Migration events (dispersals and vicariances) extracted from dated phylogenies were used to estimate rates of migrations and their variation through time. Migrations were treated as independent events occurring through time according to a random Poisson process. In our notation, the ages of all events are measured as time before the present. For a number  $K$  of migration events, we define a vector of times of migration  $\mathbf{t} = t_1, t_2, \dots, t_K, t_{K+1}$ , where  $t_1$  and  $t_K$  represent the oldest and most recent events respectively, and  $t_{K+1} = 0$  represents time 0, i.e. the present. Assuming that the rate parameter changes over time according to a function  $\lambda(t)$ , the likelihood of  $\mathbf{t}$  based on a non-homogeneous Poisson process is:

$$P(t_1, \dots, t_K | \lambda) = \prod_{j=1}^K \exp \left( - \int_{t_j}^{t_{j+1}} \lambda(u) \, du \right) \prod_{j=1}^K \lambda(t_j) \quad , \quad (1)$$

where the first product describes the probability of the waiting times between events and the second product accounts for each migration events based on the variable rate  $\lambda$ . Note that  $t_{K+1}$  appears in the first product where it accounts for the waiting time between the last migration event and the present (i.e.  $t_K - t_{K+1}$ ), whereas it is not included amongst the migration events in the second product.

Several Poisson models were implemented, the simplest of which had one constant rate through time, i.e.  $\lambda(t) = \lambda$ , for  $t_I \leq t \leq \theta$ , that is a single free parameter estimated from the data (degrees of freedom, d.f. = 1). A model with exponentially distributed rate was generated using the following function:

$$\lambda(t) = q + l \exp(-lt) \quad , \quad (2)$$

where  $q \geq 0$  is a minimum baseline rate and  $l$  is the rate parameter of the exponential distribution. Both  $q$  and  $l$  were estimated from the data as two independent free parameters (d.f. = 2). Because the times of migration are expressed as time before present ( $t = 0$  being the present), this function models a rate that exponentially increases towards the present. To allow for more complex patterns of rate variation, models with rate shifts through time were additionally implemented. These assumed that rate changes occurred

at times of shift  $\tau$  defining time frames, while the rates were assumed to be constant within each time frame. For a set of  $T$  times of shift defining  $T+1$  time frames, a vector of rates  $\lambda_1, \dots, \lambda_{T+1}$  was defined and used to calculate the probability of Equation (1). Each rate shift introduces two additional parameters in the model, i.e. the time of shift and the migration rate following the shift. For instance, a model with one rate shift has two rate parameters  $\{\lambda_1, \lambda_2\}$  and one time of shift  $\{\tau_1\}$  (d.f. = 3).

**Implementation** - The fit of alternative models of migration were compared in a maximum likelihood framework. Because the sample size in several of the data sets was comparatively small (Table S10), we used the second-order Akaike Information Criterion (AICc; [18](#), [19](#)) in all model comparisons, and AICc thresholds determined through simulations (see below) as indicators of significant support for a more parameter-rich model against a simpler model. We used a stepwise procedure that starts with a constant rate model and subsequently adds rate shifts to determine the best number of rate shifts supported by the data (e.g., [20](#), [21](#)). At every step, the algorithm adds a rate shift in the model, estimates the parameters (migration rates and times of shift) by likelihood optimization, and computes the AICc score. If the AICc for this model is significantly lower (i.e. better) than the previous score, based on the AICc threshold, the model is retained and one additional rate shift is tested. The procedure is stopped when an additional rate shift does not yield a significant improvement of AICc score, in which case the previous (simpler) model is retained ([20](#)). For model comparison, we also ran maximum likelihood optimizations under two models with fixed times of rate shift, which we named the ‘Standard’ and the ‘Miocene’ models following Coates and Stallard

(5). The Standard model assumed a shift at 3.5 Ma and two rate parameters (before and after the shift; d.f. = 2) and the Miocene Model had shifts at 25 and 15 Ma and three rate parameters (d.f. = 3).

The migration models were implemented in a Python ([www.python.org](http://www.python.org)) script (available from the authors) to optimize the parameters using maximum likelihood. To reduce the risk of finding local optima, likelihood optimizations were repeated 500 times from different random starting parameter values. Under each model, starting times of shift were proposed as random values from a uniform distribution spanning the age of the data set and then optimized as a free parameter.

***Determining AICc significance thresholds*** - We carried out simulations in order to assess AICc thresholds indicating significant support for a parameter rich model against a simpler model. Following the procedure recently employed by Pennel et al. ([22](#)), we simulated data sets of sizes  $N = \{10, 25, 50, 75, 100, 150, 200, 250, 300, 400\}$  under a constant-rate Poisson process, i.e. drawing  $N$  times of migration from uniform distributions. The number of simulations totaled 10,000 (i.e. 1,000 of each size). The simulated migrations spanned a time frame ranging from 0 to a maximum age randomly sampled between 65 and 15 Ma. These sample sizes and time frames reflected the ranges observed in our empirical data sets. We analyzed each simulated data set and recorded the difference in AICc values ( $\Delta AICc$ ) between the (true) constant rate model and best (incorrect) model with one rate shift. We then determined AICc threshold for each data size as the 95<sup>th</sup>  $\Delta AICc$  percentile and fitted an  $x$ -shifted power function ([22](#)) to draw



AICc thresholds for any data size. Based on the best fitting function the AICc threshold for a data set of size  $N$  can be obtained from the following equation:

$$\Delta AICc_{threshold}(N) = -59.587 \times (N \times 0.573)^{-1.399} + 6.108 \quad (3)$$

Given the range of sample sizes in our empirical analyses, the AICc thresholds used here ranged between 2.87 and 6.08 log-units (Table S10).

### **1.7 Analysis of the migration events (molecular data)**

Analyses of migration rates were performed on the full data set (see above) and on different subsets based on different biogeographic, ecologic, and taxonomic criteria. The definition of subsets was also conditioned on including at least 10 migration events, in order to avoid an exceeding loss of analytical power due to sample size.

The following data sets were analyzed (sample size in parentheses):

1. All data (424): all sampled plants and animals (terrestrial and aquatic)
2. Dispersal direction of terrestrial taxa: North to South (144), South to North (154), defined as explained below
3. Terrestrial vertebrates (59) and marine taxa (86), the latter distributed either in the Caribbean Sea or in the Pacific Ocean
4. Taxonomic groups: arthropods (35), mammals (24), birds (158), non-avian reptiles (17), amphibians (18), plants (74), and freshwater fishes (14)
5. Dispersal ability (as defined below): low (61), medium (78), high (287).

Model selection was performed independently on each data set based on the mean ages of migration events (Table S8). After testing the migration models with a constant rate and an exponentially distributed rate, we used the stepwise AICc procedure described above to assess the number rate shifts (Table S10). Maximum likelihood estimations of the parameters (rates and times of rate shifts) were performed under the model selected by stepwise AICc and repeated on 1,000 replicates after randomizing the age of the migration events uniformly within the respective confidence intervals (Table S8). The rate estimates were used to calculate mean and 95% confidence interval of the migration rate within time bins of 1 Myr and to construct migration rates through time plots (MRTT, Figs. 2 and S2). 95% confidence intervals were also calculated for the times of rate shifts.

***Rate standardization*** - The migration rates estimated through Equations (1,2) represent the absolute rate of the fitted Poisson process, and represent the expected number of migration events per time unit (1 Myr) in a given data set. While ideally, such rates should be estimated on a per-lineage basis [as for other macroevolutionary rates ([e.g. 23](#))], their estimation would be likely affected by unobserved extinct lineages and further biases might arise from combining data from different phylogenies. Thus, we preferred the estimation of absolute Poisson rates, and assumed that the model with exponentially increasing rates might capture potential biases owed to the exponential increase of lineages in extant-taxa phylogenies.

As a mean to approximately standardize the migration rates across data sets, we rescaled the estimated absolute Poisson rates by the number of families considered in each analysis. Thus the migration rates showed in the MRTT plots indicate the expected number of migration events per Myr per family. We emphasize that this standardization is not intended as a correction for temporal biases within each data set (as rates are simply rescaled), but rather as tool to facilitate comparisons across different analyses. Number of families included is provided for each data set in Table S10.

An additional test was performed using the two data sets of terrestrial taxa defined by dispersal direction (see point 2 above) to assess whether significant asymmetries in migration rates could be detected between North American taxa migrating southward and South American taxa migrating northward. The likelihoods of different migration models were therefore optimized on a data set that included both subsets. We first tested three-rate models, i.e. the model selected by stepwise AICc for both subsets analyzed independently (Table S3). Secondly a symmetric model (identical rates and shift times) was compared with an asymmetric four-rate model in which rates and times of shifts were independently estimated for the two directions of migration and their AICs scores were compared. Finally to assess when were most different, we looked at the amount of overlap in the respective migration rates (and 95% confidence intervals) through different time frames.

***Sensitivity to dating errors*** - To further assess the effect of data uncertainty on the estimated dynamics of migration rates, the ages of the biogeographic events were

randomized to simulate different proportions of error. For a given relative error  $r$ , the ages of all biogeographic events were randomly redrawn from normal distributions centered in their mean value and with standard deviation equal to a fraction  $r$  of their mean. Thus, for an event  $i$  of mean age  $m_i$  new values were randomly resampled from:

$$m'_i = |\mathcal{N}(\mu = m_i, \sigma^2 = rm_i)|, \quad (4)$$

where the absolute value is used to avoid negative ages. Based on Equation (4), 100 randomized data sets with relative errors of 10, 20, and 50 % (obtained from  $r = \{0.1, 0.2, 0.5\}$ , respectively) were simulated and analyzed by the stepwise AICc algorithm described above. To assess whether our choice of simulated errors were comparable with those observed in the data we calculated the mean relative error in our molecular data. We used the available 95 % confidence intervals around the age estimates (Table S2) and assumed normal distributed errors, so that the boundaries of the confidence interval equal the mean age  $\pm 1.96 \sigma^2$ . Based on Equation 3, we calculated the relative error for all events for which a confidence interval was available (46 % of the data) and obtained an average relative error in our data equal to 21.04 % ( $r = 0.21$ ), thus falling well within the range of simulated errors. The dynamics of the migration rates through time obtained from the data sets with simulated errors were strikingly consistent with those estimated from the events' mean ages, even when the relative error was more than twice as large as that observed from the data (50 %; Fig. S1). This indicates that our findings are robust even against considerable amounts of potential error in the data (Figs. 1, S1).

## **1.8 Assessing the impact of taxon biology on dispersal time**

The taxa that migrated across the Isthmus of Panama exhibit variability in ecology and evolutionary history. Because taxon ecology and evolutionary history has been shown to impact genetic differentiation and dispersal across the landscape ([27, 24](#)), the variance in the observed dispersal times across the Isthmus of Panama may be attributable to taxon-specific factors.

To test this hypothesis, we employed a Generalized Linear Mixed (GLMM) model approach to assess the effect of elevational preference, biome type, dispersal ability, and dispersal direction on the timing of dispersal. We also included the molecular-dating calibration type used by the original studies in the model to account for potential variance in migration times associated with a particular calibration method (e.g. molecular clock vs. fossil calibration). For example, dates obtained using a molecular clock may yield more recent divergence events if the substitution rates tended to be faster than dates obtained from phylogenies calibrated with fossils. A complete description of the ecological and biological variables is listed below. We fit the data to models using the lme4 v. 0.9975-12 package in the R programming language ([25](#)).

For the models examining taxa that inhabit terrestrial and freshwater biomes we treated the variables (biome, elevational preference, dispersal ability, dispersal direction, and calibration type) as fixed effects. We were unable to identify dispersal direction for some taxa because there is inadequate information in their phylogenies. We also could not



obtain sufficiently detailed information on the elevational preferences or biome for particular taxa to accurately assign them to a category. Thus, we removed the points classified as missing or equivocal data. Because some taxa had missing data in multiple categories, the final data set analyzed was reduced from 341 to 280 points. Finally, we included a phylogenetic correction by treating taxonomy as a nested random effect to account for the non-independence of the data in each model. We started with ‘Class’, ‘Order’, and ‘Family’ as a nested random effect, but ‘Order’ was removed because the estimated variance around the parameter was 0 due to the strong correlation between ‘Order’ and ‘Class’. We obtained similar results if we removed ‘Class’ instead of ‘Order’. Irrespective of the random effect chosen, the results from our model comparisons were similar between models retaining all three taxonomic levels. The full model for taxa that inhabit the terrestrial and freshwater biomes was structured as follows:

```
glmer(log(Dispersal.Time) ~ Elevational.Preference + Dispersal.Direction + Dispersal.Ability +
Biome + Calibration +(1|Order/Family), family="gaussian")
```

For the model examining the marine taxa, we did not include habitat type or altitude because the majority of taxa inhabited shallow waters and there were inadequate sample sizes for the different levels in the ‘Biome’ category. We included all of the compiled dispersal times for marine taxa in the model (n=80). For the phylogenetic correction, we used ‘Class’, ‘Order’, and ‘Family’ as a nested random effect. The 0 estimated variance in ‘Order’ observed in the terrestrial and freshwater taxa model was not observed in the marine taxa models (Table S5). The full model for marine was structured as follows:

```
glmer(log(Dispersal.Time) ~ Dispersal.Ability + Calibration + (1|Class/Order/Family),  
family="gaussian")
```

For all models, we specified a Gaussian error distribution and we log-transformed dispersal time to reduce the residual variance. The outputs for full and reduced models are available in Tables S4 and S6.

We evaluated the importance of each fixed effect by comparing the full model versus a model where the variable was removed using the R function `anova(full.model, reduced.model, test="chisq")`. The idea being that if a variable was important in explaining the variance in dispersal times the model would be significantly worse when it was excluded (Tables S5 and S7).

We found that the biological variables could not significantly explain the variance in dispersal times across the Isthmus of Panama (Tables S5 and S7). By comparing full and reduced models in the terrestrial and freshwater taxa we found that excluding the variables elevational preference ( $\text{Pr} > \text{ChiSq} = 0.60$ ), dispersal direction ( $\text{Pr} > \text{ChiSq} = 0.30$ ), dispersal ability ( $\text{Pr} > \text{ChiSq} = 0.54$ ), or biome type ( $\text{Pr} > \text{ChiSq} = 0.20$ ) did not yield significantly different models. In the marine taxa the removal of dispersal ability ( $\text{Pr} > \text{ChiSq} = 0.83$ ) from the model did not have a significant effect. One of the confounding issues in assessing the significance of these variables was the association between biological differences and the taxonomic groups. For example, the taxa

identified as good dispersers exhibit much more recent dispersal times across the Isthmus of Panama, but the majority of the good dispersers were birds.

We found that the inclusion of calibration type improved the model, which suggests that some of the variance in dispersal times may be associated with how molecular dating was done in the original studies. In the terrestrial and freshwater taxa model calibration type was only marginally significant ( $\text{Pr} > \text{ChiSq} = 0.09$ ), whereas in the marine taxa, calibration type was significant ( $\text{Pr} > \text{ChiSq} = 6.19\text{E-}5$ ).

**Biomes** - We used World Wildlife Fund (WWF) biomes to assign taxa to broad-scale habitats that represent their climatic envelopes ([26](#)). Biome types are as follows: TSMBF – Tropical & Subtropical Moist Broadleaf Forest; TSDBF – Tropical & Subtropical Dry Broadleaf Forest; TSGSS – Tropical & Subtropical Grasslands, Savannas, and Shrublands; DXS – Deserts & Xeric Shrublands; TSCF – Tropical & Subtropical Coniferous Forests; TGSS – Temperate Grasslands, Savannas, and Shrublands; BFT – Boreal Forests/Taiga; MGS – Mangroves; and Aquatic). Marine habits include: Tropical Coral, Tropical Upwelling, Pelagic Marine (open-water dispersers), and Intertidal (for nearshore, generally sessile aquatic organisms). We condensed our variables into humid forest (TSMBF), dry forest (TSDBF), forest (multiple forest types or non-tropical forest), nonforest (TGSS, DXS TSGSS), multiple biomes (eurybiomic), or aquatic (separating freshwater and salt water).

***Elevational range*** - We assigned clades to their elevational preference: Lowland (0 – 500 m), Montane (500-2500), and Highland (>2500 m). There were only five taxa assigned to the highland category, so for the purposes of the analyses we merged Montane and Highland taxa. For clades that were distributed in multiple elevational zones, we assigned them to two broad elevation categories Lowland-Montane and Lowland-Highland.

***Dispersal direction*** - Dispersal direction was obtained from original publications. The three dispersal categories assigned were 1) North to South – represents a dispersal event where a lineage started north of the Isthmus of Panama and went south into South America, and 2) South to North – represents a dispersal event where a lineage started in South America and dispersed north of the Isthmus of Panama.

***Dispersal ability*** - We assigned taxa to three categories, which reflect coarse-scale differences in dispersal abilities. Dispersal categories were defined as Category 1 – poor; Category 2 – moderate; and Category 3 – good.

***Aquatic Organisms*** Category 1) Obligatory freshwater organisms. In general these species are restricted to freshwater habitats and are incapable of crossing a marine barrier causing them to have limited ability to disperse. Category 2) Viviparous, clonal, or nesting marine species were assumed to be relatively good dispersers but not as high as broadcast spawners. Category 3) Marine organisms whose reproductive life history traits include having pelagic eggs or larvae (i.e., early-stage young are released to be dispersed by the ocean).

*Terrestrial Animals* Category 1) Organisms that are constrained to aquatic habitats on land during any stage of life. Category 2) Organisms that spend the majority of their lives on terrestrial habitats (not aquatic), and are incapable of flight. Category 3) Organisms that are capable of flying.

*Plants* Category 1) Plants possessing heavy fruits or seeds, passive fruit dispersal, or narrow distribution. Category 2) Because of the lack of information on natural history of many tropical plant groups this category was scored as anything in between categories 1 and 2. Category 3) Plants producing spores; small or winged seeds; or dispersed by birds, usually producing berries.

***Molecular dating calibration*** - The published phylogenies used to extract dispersal events across the Isthmus of Panama were calibrated using molecular clocks, fossils, and secondary calibrations based on dating events from external phylogenies, or a “mixed” calibration, which used more than one type of calibration. Because the calibration type may influence divergence time estimates, we included a calibration type in our GLM models to account for any potential biases.

## **SUPPORTING FIGURE AND TABLE CAPTIONS**

**Figure S1.** Migration rates through time estimated on the full data set (426 biogeographic events) after introducing 10, 20, and 50 % of random error in the data. The black dashed



line indicates the rate estimate obtained from the analysis of the mean ages of the biogeographic events.

**Figure S2.** Estimated rates of dispersal across the Isthmus of Panama through time for non-avian reptiles and amphibians, birds, and mammals. The rates and their temporal dynamics were estimated using maximum likelihood and 1,000 randomizations to infer 95% confidence intervals (CI; shaded areas). Circles and bars at the bottom indicate the mean and 95% CI for times of rate shift.

**Figure S3.** Estimated rates of dispersal across the Isthmus of Panama through time for freshwater fish, arthropods, and plants. The rates and their temporal dynamics were estimated using maximum likelihood and 1,000 randomizations to infer 95% confidence intervals (CI; shaded areas). Circles and bars at the bottom indicate the mean and 95% CI for times of rate shift.

**Figure S4.** Estimated rates of dispersal across the Isthmus of Panama through time for taxa divided by low, medium, and high dispersal ability. The rates and their temporal dynamics were estimated using maximum likelihood and 1,000 randomizations to infer 95% confidence intervals (CI; shaded areas). Circles and bars at the bottom indicate the mean and 95% CI for times of rate shift.

**Table S1.** Migration rates and times of rate shifts, estimated from the analysis of phylogenetic data.

**Table S2.** Mean and 95% confidence intervals of sampled species through time based on the fossil record, and proportion of immigrant species in North and South America.

**Table S3.** Migration models estimated from analysis of phylogenetic data.

**Table S4a-f.** Generalized Linear Mixed Model output for full and reduced models examining the effects of ecological and historical variables on cross-Isthmus of Panama dispersal times in terrestrial and freshwater taxa; a) full model; b) model with elevational preference variable removed; c) model with dispersal direction variable removed; d) model with dispersal ability variable removed; e) model with biome type variable removed; and f) model with calibration type variable removed.

**Table S5.** Generalized Linear Mixed Model comparison between full and reduced models, computed to examine the effects of ecological and historical variables on dispersal times across the Isthmus of Panama for terrestrial and freshwater taxa.

**Table S6a-c.** Generalized Linear Mixed Model output for full and reduced models examining the effects of ecological and historical variables on dispersal times across the Isthmus of Panama for marine taxa: a) full model; b) model with dispersal ability variable removed; and c) model with calibration type variable removed.

**Table S7.** Generalized Linear Mixed Model comparison for full and reduced models examining the effects of ecological and historical variables on dispersal times across the Isthmus of Panama for marine taxa.

**Table S8.** Survey of approximate ages of extant taxa reported from published phylogenies. The complete data is deposited at Dryad (<http://datadryad.org>).

**Table S9.** List of journals surveyed for the present study, in which we searched for suitable dated phylogenies.

**Table S10.** Migration models estimated over the phylogenetic data for terrestrial taxa, partitioned by the direction of migration.

## REFERENCES

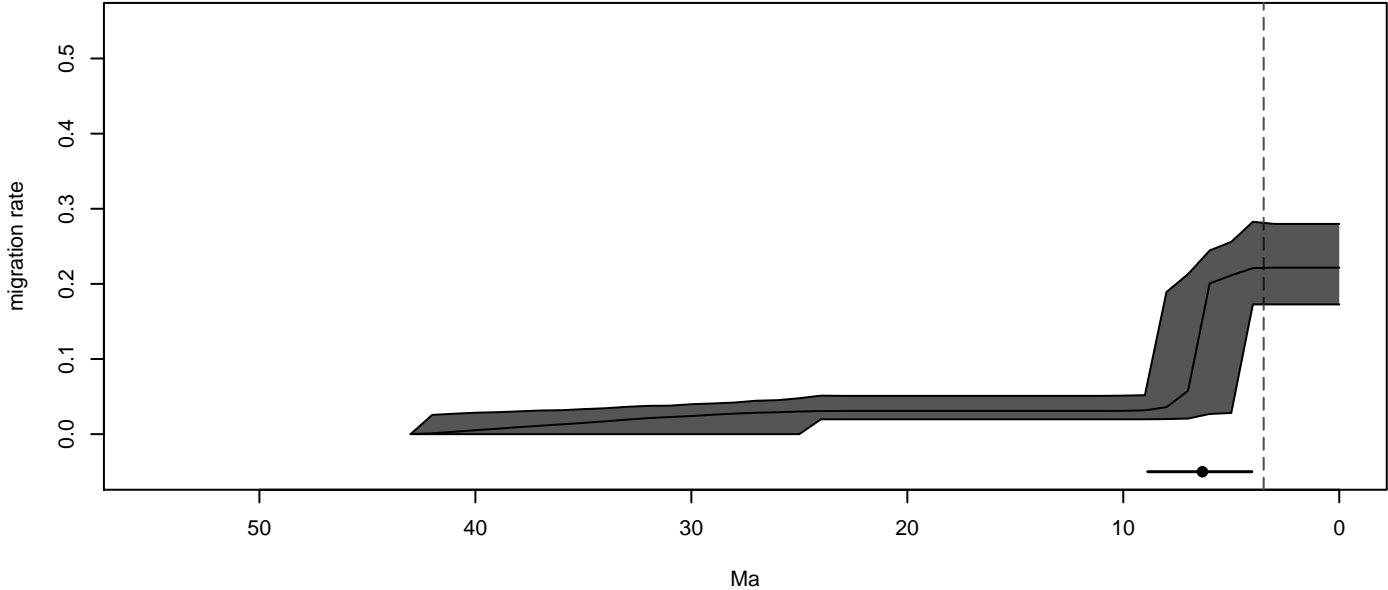
1. Coates AG, Aubry MP, Berggren WA, Collins LS, & Kunk M (2003) Early Neogene history of the Central American arc from Bocas del Toro, western Panama. *Geological Society of America Bulletin* 115(3):271-287.
2. Coates AG, Collins LS, Aubry MP, & Berggren WA (2004) The geology of the Darien, Panama, and the Miocene-Pliocene collision of the Panama arc with northwestern South America. *Geological Society of America Bulletin* 116(11):1327-1344.

3. Coates AG, *et al.* (1992) Closure of the Isthmus of Panama: The near-shore marine record of Costa Rica and western Panama. *Geological Society of America Bulletin* 104(7):814-828.
4. Coates AG & Obando JA (1996) The Geological Evolution of the Central American Isthmus. *Evolution and Environment in Tropical America*, eds Jackson JBS, Budd AF, & Coates AG (University of Chicago, Chicago).
5. Coates AG & Stallard RF (2013) How old is the Isthmus of Panama. *Bulletin of Marine Science* 89(4):801-814.
6. Haug GH & Tiedemann R (1998) Effect of the formation of the Isthmus of Panama on Atlantic Ocean thermohaline circulation. *Nature* 393(6686):673-676.
7. Haug GH, Tiedemann R, Zahn R, & Ravelo AC (2001) Role of Panama uplift on oceanic freshwater balance. *Geology* 29(3):207-210.
8. Magallon S & Sanderson MJ (2001) Absolute diversification rates in angiosperm clades. *Evolution* 55(9):1762-1780.
9. Hoorn C, *et al.* (2010) Amazonia through time: Andean uplift, climate change, landscape evolution and biodiversity. *Science* 330(6006):927-931.
10. Carillo JD, Forasiepi A, Jaramillo CA, & Sanchez-Villagra MR (2015) Neotropical mammal diversity and the Great American Biotic Interchange: spatial and temporal variation in South America's fossil record. *Frontiers in Genetics*.
11. Silvestro D, Schnitzler J, H LL, Antonelli A, & Salamin N (2014) Bayesian estimation of speciation and extinction from incomplete fossil occurrence data. *Systematic Biology* 63(3):349-367.

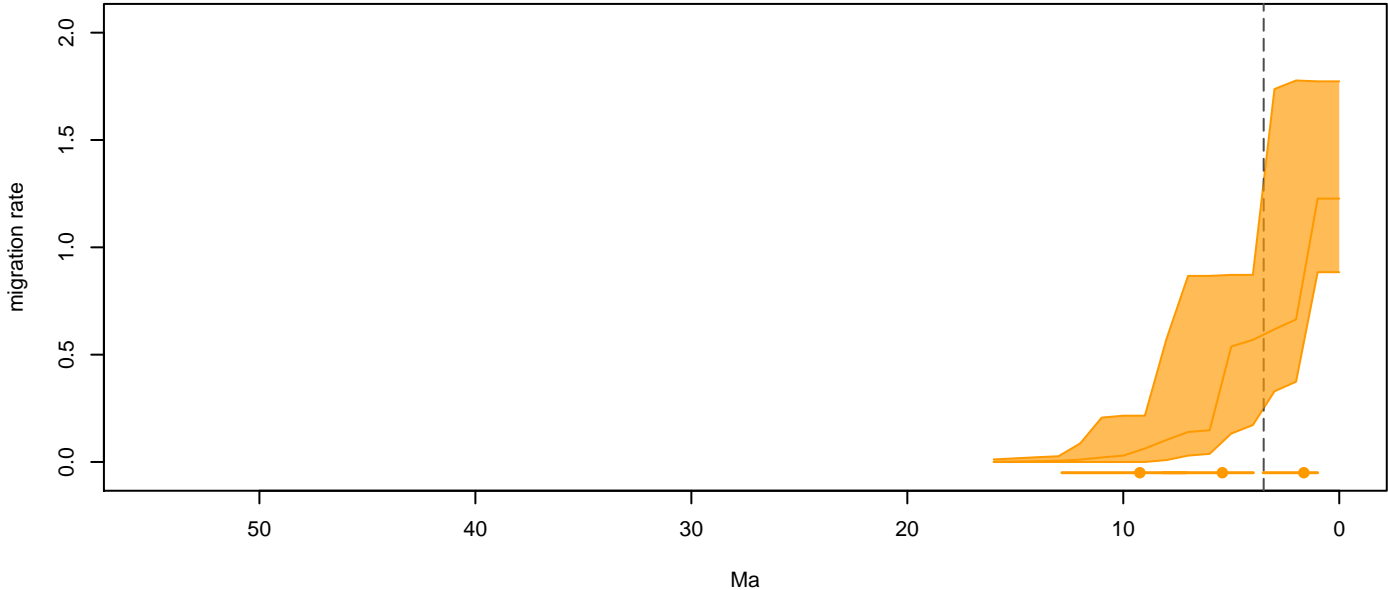
12. Lessios HA (2008) The Great American Schism: Divergence of marine organisms after the rise of the Central American Isthmus. *Annual Review of Ecology, Evolution, and Systematics* 39(1):63-91.
13. Bermingham E & Lessios HA (1993) Rate variation of protein and mitochondrial DNA evolution as revealed by sea urchins separated by the Isthmus of Panama. *Proceedings of the National Academy of Sciences, USA* 90(7):2734-2738.
14. Hickerson MJ, Stahl EA, & Lessios HA (2006) Test for simultaneous divergence using approximate Bayesian computation. *Evolution* 60(12):2435-2453.
15. Knowlton N & Weigt LA (1998) New dates and new rates for divergence across the Isthmus of Panama. *Proceedings of the Royal Society B: Biological Sciences* 265(1412):2257-2263.
16. Pulquerio MJ & Nichols RA (2007) Dates from the molecular clock: how wrong can we be? *Trends in Ecology and Evolution* 22(4):180-184.
17. Sauquet H, *et al.* (2012) Testing the impact of calibration on molecular divergence times using a fossil-rich group: the case of *Nothofagus* (Fagales). *Systematic Biology* 61(2):289-313.
18. Akaike H (1974) A new look at the statistical model identification. *IEEE Transactions on Automatic Control* AC-19(6):716-723.
19. Burnham KP & Anderson DR (2002) *Model selection and multimodel inference: a practical information-theoretic approach* (Springer, New York, NY).
20. Alfaro ME, *et al.* (2009) Nine exceptional radiations plus high turnover explain species diversity on jawed vertebrates. *Proceedings of the National Academy of Sciences, USA* 106:13410-13414.

21. Stadler T (2011) Mammalian phylogeny reveals recent diversification rate shifts. *Proceedings of the National Academy of Sciences, USA* 108(15):6187-6192.
22. Pennell MW, *et al.* (2014) geiger v2.0: an expanded suite of methods for fitting macroevolutionary models to phylogenetic trees. *Bioinformatics* 30(15):2216-2218.
23. Rabosky DL (2014) Automatic detection of key innovations, rate shifts, and diversity-dependance on phylogenetic trees. *PLoS ONE* 9(2):e89543.
24. Burney CW & Brumfield RT (2009) Ecology predicts levels of genetic differentiation in Neotropical birds. *American Naturalist* 174(3):358-368.
25. Team RDC (2008) A language and environment for statistical computing. R Foundation for Statistical Computing (Vienna, Austria).
26. Olson DM, *et al.* (2001) Terrestrial ecoregions of the world: a new map of life on Earth. *Bioscience* 51(11):933-938.

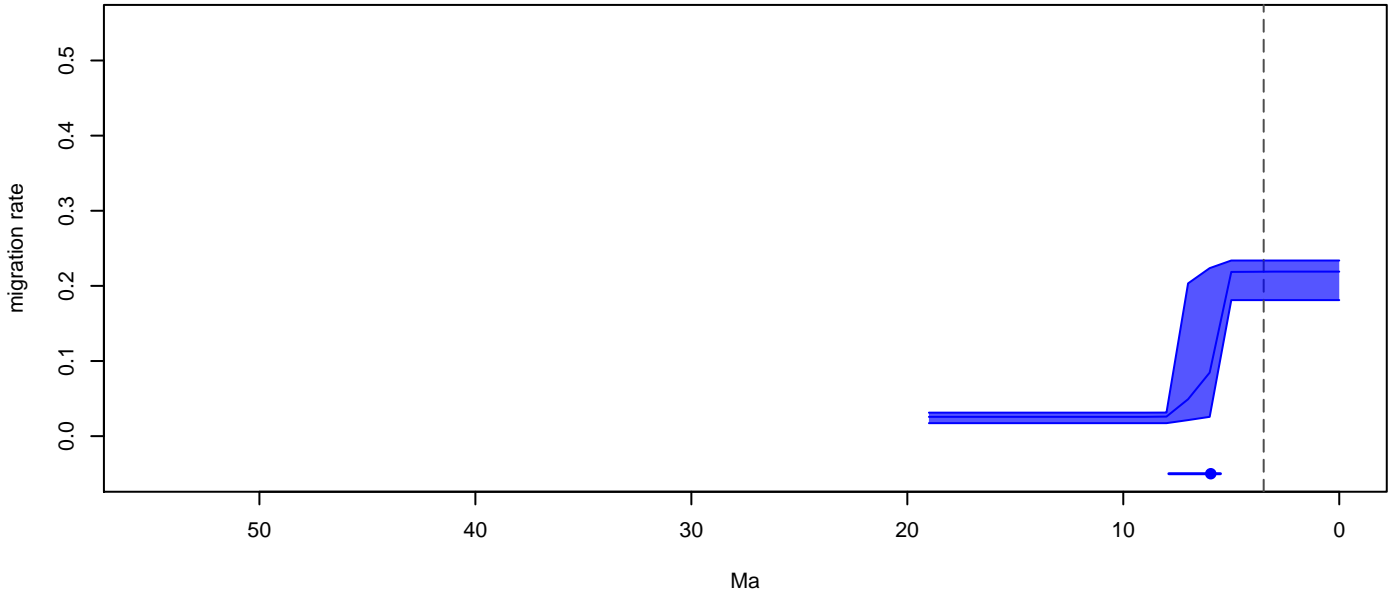
Reptiles and Amphibians



Birds

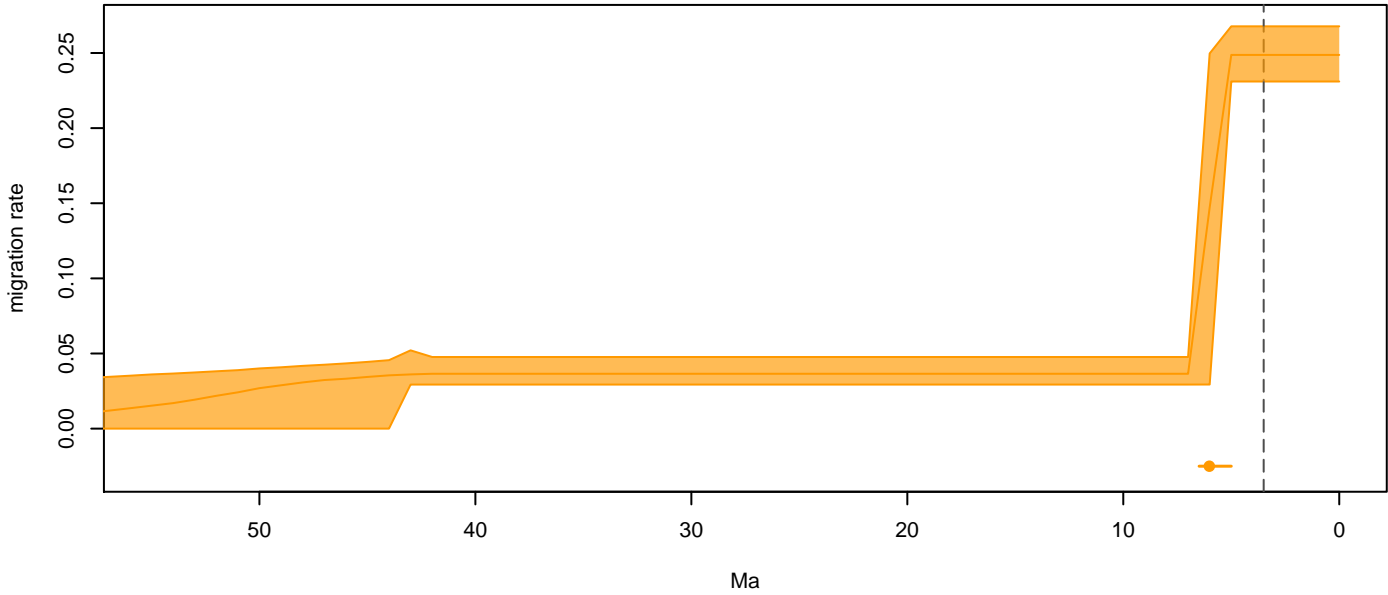


Mammals

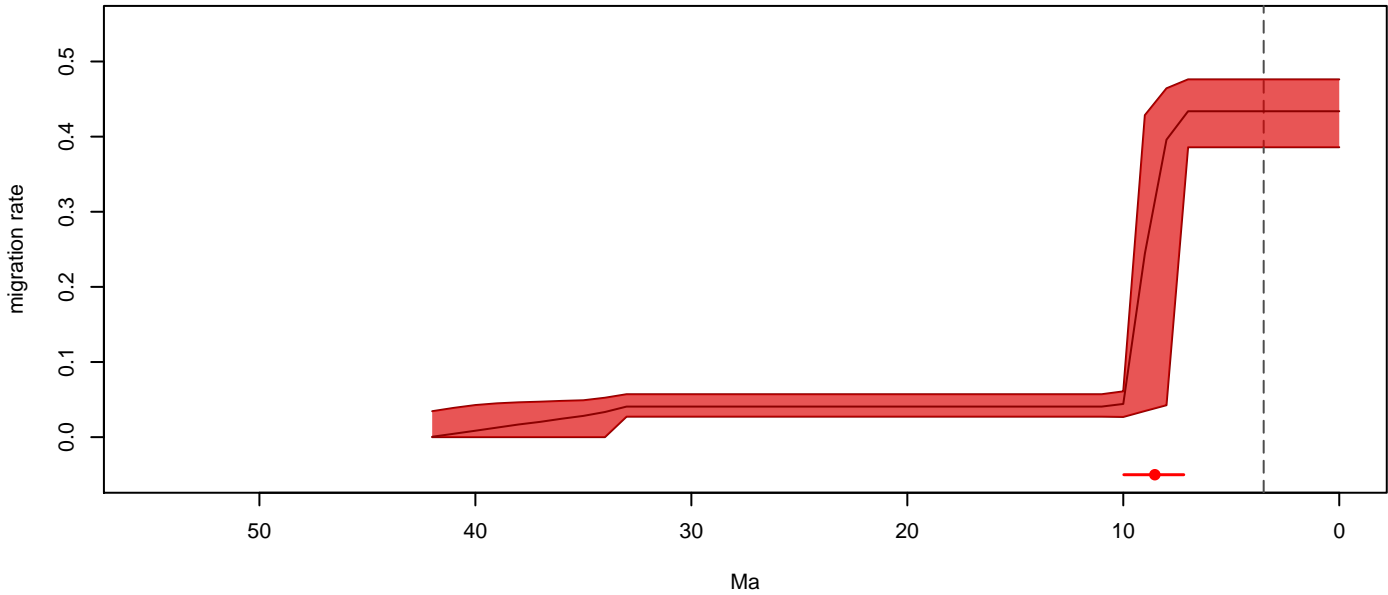




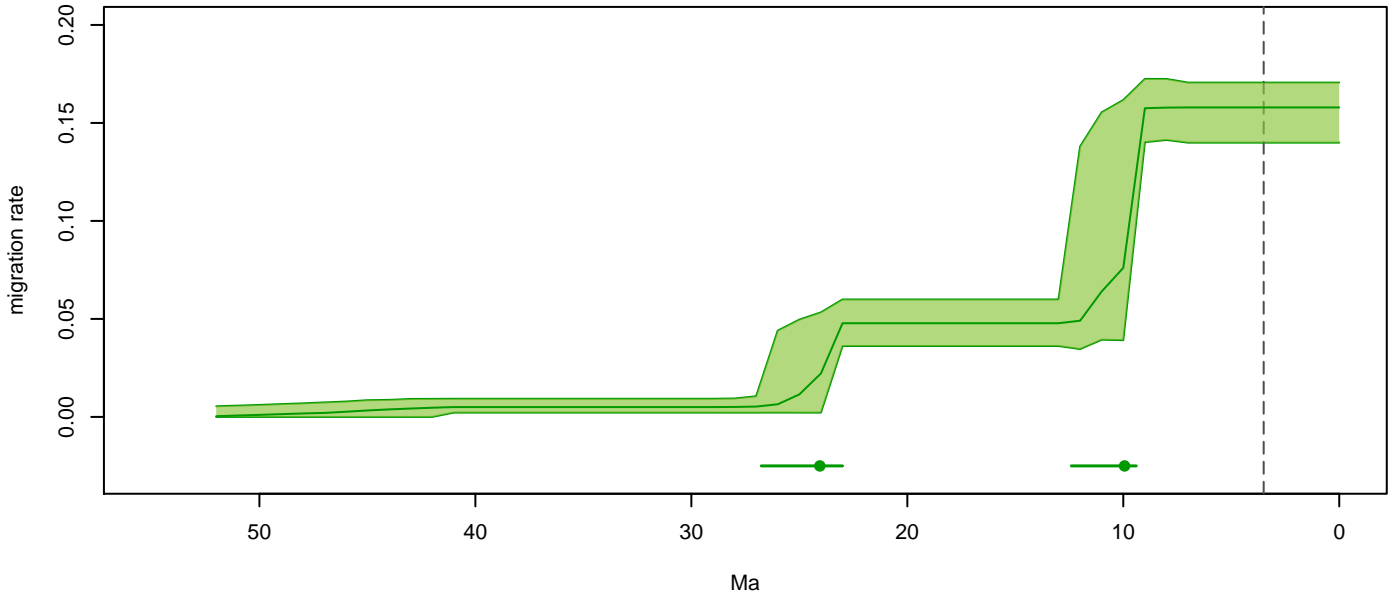
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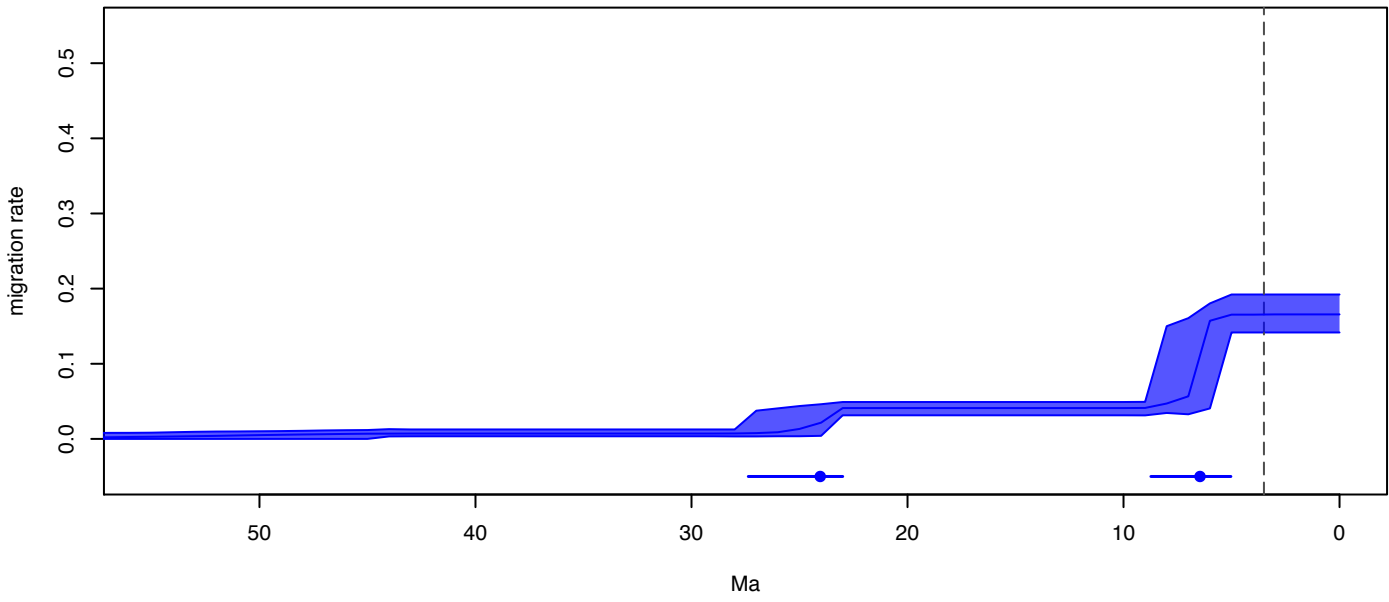
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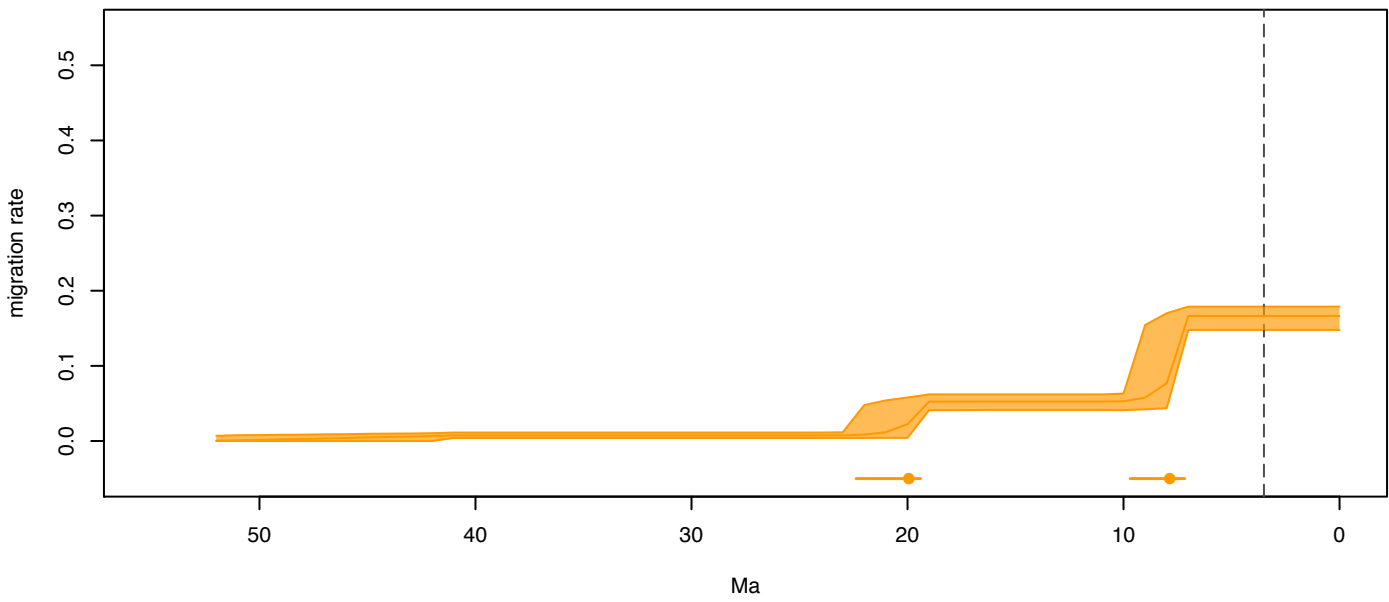
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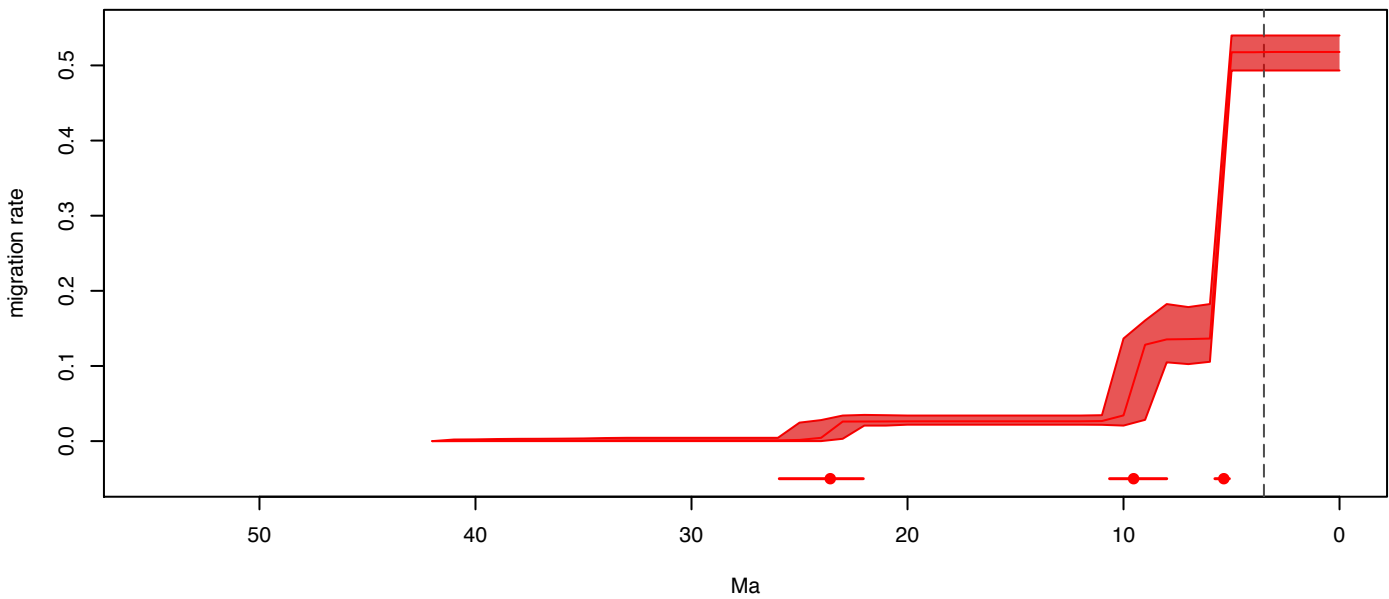
**Low dispersal ability**



**Medium dispersal ability**



**High dispersal ability**



**Table S1. Migration rates and times of rate shifts from analysis of phylogenetic data.**

Data set	Time of rate shift (Ma)				Migration rates between shifts				
All	5.263 (5.1–6.092)	8.756 (7.2–10.047)	23.696 (19.9–26.271)	41.14 (35.917–46.239)	0.371 (0.351–0.386)	0.142 (0.106–0.207)	0.036 (0.029–0.044)	0.006 (0.003–0.008)	0.001 (0–0.002)
N to S America	7.541 (5.172–9.704)	23.873 (22–26.415)			0.227 (0.192–0.281)	0.036 (0.025–0.048)	0.004 (0.002–0.007)		
S to N America	6.132 (5.1–7.048)	23.142 (20.5–26.576)			0.304 (0.274–0.345)	0.031 (0.024–0.04)	0.003 (0–0.005)		
Marine	2.061 (1.028–4.35)	7.959 (7.75–8.955)	23.733 (19.9–27.411)		0.13 (0.121–0.2)	0.292 (0.205–0.312)	0.045 (0.035–0.059)	0.006 (0–0.019)	
Terrestrial tetrapods	6.12 (5.1–7.89)	20.225 (19.4–22.813)			0.216 (0.181–0.245)	0.043 (0.033–0.054)	0.004 (0–0.01)		
Reptiles and amphibians	6.335 (4.046–8.864)				0.222 (0.173–0.28)	0.031 (0.02–0.051)			
Fresh water fish	6.016 (5–6.49)				0.249 (0.231–0.268)	0.037 (0.029–0.048)			
Arthropods	8.54 (7.2–9.975)				0.434 (0.386–0.476)	0.041 (0.027–0.057)			
Plants	9.941 (9.4–12.405)	24.051 (23–26.762)			0.158 (0.14–0.171)	0.048 (0.036–0.06)	0.005 (0.002–0.009)		
Birds	1.64 (1–3.524)	5.414 (3.977–8.177)	9.234 (7.071–12.843)		1.227 (0.884–1.773)	0.618 (0.329–1.737)	0.148 (0.038–0.867)	0.011 (0–0.086)	
Mammals	5.95 (5.5–7.89)				0.219 (0.181–0.234)	0.026 (0.017–0.031)			
Low dispersal ability	6.458 (5.03–8.733)	24.04 (23–27.37)			0.166 (0.142–0.192)	0.041 (0.031–0.049)	0.007 (0.003–0.013)		
Medium dispersal ability	7.86 (7.17–9.694)	19.943 (19.4–22.38)			0.166 (0.148–0.179)	0.053 (0.041–0.062)	0.007 (0.004–0.011)		
High dispersal ability	5.357 (5.1–5.769)	9.534 (8–10.647)	23.576 (22.047–25.937)		0.518 (0.493–0.54)	0.137 (0.106–0.182)	0.026 (0.022–0.034)	0.001 (0–0.004)	

The results of migration rate analyses are shown for the full dataset and several subsets defined by the direction of migration, taxonomic group, and dispersal ability. Maximum likelihood estimates of the times of rate shift (in Ma) and migration rates (dispersals/family/Myr) are reported based on the best fitting model selected by AICc (see text). Reported migration rates refer to the mid-point between times of shift. 95% confidence intervals are provided in parentheses for both shift times and migration rates.

**Table S2. Mean number of sampled species and 95% confidence intervals through time based on the fossil record.**

Time bin (Ma)	Species in NA	Immigrants spp. In NA	Species in SA	Immigrants spp. In SA
1-0	451.541 (438-462)	31.018 (26-34)	265.643 (256-274)	132.196 (126-139)
2-1	345.403 (326-361)	29.307 (25-32)	144.823 (134-154)	72.567 (66-79)
3-2	290.129 (273-308)	23.681 (19-27)	88.737 (76-100)	40.388 (31-48)
4-3	261.813 (245-279)	15.058 (11-19)	41.736 (35-48)	5.48 (4-8)
5-4	202.087 (183-217)	8.349 (3-12)	54.962 (48-62)	3.665 (0-6)
6-5	146.069 (132-161)	2.855 (0-5)	61.98 (53-71)	3.353 (1-5)
7-6	197.235 (179-214)	3.967 (1-6)	81.165 (68-92)	3.48 (1-5)
8-7	232.0 (215-252)	4.601 (3-7)	145.899 (132-157)	4.875 (2-7)
9-8	248.387 (229-267)	4.739 (3-7)	145.056 (133-158)	4.124 (2-6)
10-9	245.849 (225-263)	4.285 (1-6)	76.743 (67-87)	1.687 (1-3)
11-10	251.042 (231-269)	3.116 (1-4)	58.861 (48-68)	0.414 (0-1)
12-11	275.948 (260-294)	2.179 (0-3)	40.467 (31-48)	0.123 (0-1)
13-12	286.671 (269-301)	1.517 (0-3)	32.88 (26-38)	0
14-13	303.987 (286-321)	0.852 (0-2)	37.235 (30-43)	0
15-14	341.303 (326-357)	0.457 (0-1)	33.742 (28-40)	0
16-15	323.656 (305-341)	0.399 (0-1)	49.2 (42-57)	0
17-16	272.66 (255-290)	0.354 (0-1)	62.221 (52-70)	0
18-17	276.298 (255-294)	0.307 (0-1)	56.887 (47-66)	0
19-18	259.082 (238-277)	0.273 (0-1)	47.093 (37-54)	0
20-19	221.165 (203-239)	0.218 (0-1)	48.407 (41-57)	0
21-20	192.9 (173-211)	0.166 (0-1)	42.753 (34-51)	0
22-21	194.115 (175-211)	0.115 (0-1)	46.916 (36-57)	0
23-22	194.932 (177-212)	0.054 (0-0)	48.79 (38-59)	0
24-23	177.211 (162-191)	0.003 (0-0)	43.093 (34-51)	0
25-24	170.026 (155-185)	0	48.921 (40-57)	0

26-25	154.27 (141-166)	0	48.633 (40-56)	0
27-26	144.085 (130-156)	0	44.551 (37-53)	0
28-27	132.027 (119-144)	0	35.645 (27-44)	0
29-28	116.989 (105-129)	0	26.871 (17-34)	0
30-29	96.625 (84-108)	0	16.876 (11-22)	0
31-30	75.878 (65-85)	0	21.885 (15-27)	0
32-31	71.742 (64-78)	0	24.128 (18-29)	0
33-32	75.734 (69-82)	0	23.91 (18-29)	0
34-33	149.921 (143-156)	0	21.012 (15-26)	0
35-34	131.024 (120-142)	0	19.08 (14-24)	0
36-35	145.614 (132-156)	0	18.583 (13-23)	0
37-36	122.66 (109-133)	0	15.726 (10-20)	0
38-37	87.016 (76-97)	0	10.44 (7-14)	0
39-38	87.623 (78-97)	0	9.534 (6-13)	0
40-39	87.321 (77-96)	0	9.844 (6-13)	0
41-40	89.524 (78-99)	0	10.667 (6-14)	0
42-41	104.63 (93-116)	0	13.203 (9-17)	0
43-42	116.778 (105-129)	0	10.548 (7-14)	0
44-43	117.049 (102-128)	0	10.868 (6-14)	0
45-44	104.33 (90-116)	0	10.933 (6-14)	0
46-45	79.049 (66-90)	0	10.893 (6-14)	0
47-46	67.289 (58-78)	0	10.633 (6-14)	0
48-47	80.434 (70-89)	0	10.215 (6-14)	0
49-48	90.497 (81-101)	0	13.779 (8-18)	0
50-49	91.143 (79-101)	0	20.152 (14-26)	0
51-50	97.343 (84-108)	0	24.872 (17-31)	0
52-51	114.26 (101-126)	0	27.678 (20-34)	0
53-52	123.66 (109-135)	0	28.63 (22-36)	0
54-53	118.126 (106-131)	0	27.578 (20-34)	0

55-54	97.919 (85-110)	0	24.717 (19-32)	0
56-55	70.031 (58-80)	0	23.169 (17-30)	0
57-56	48.671 (44-53)	0	25.768 (20-31)	0
58-57	30.39 (24-36)	0	14.043 (9-18)	0
59-58	34.334 (28-41)	0	7.318 (3-11)	0
60-59	34.671 (27-40)	0	1.246 (0-2)	0
61-60	32.541 (26-38)	0	1.119 (0-2)	0
62-61	33.173 (27-39)	0	2.402 (0-5)	0
63-62	35.209 (32-39)	0	6.508 (3-10)	0
64-63	25.364 (18-32)	0	8.921 (6-12)	0
65-64	23.625 (19-27)	0	8.482 (5-11)	0
66-65	17.378 (12-22)	0	5.256 (2-8)	0

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The number of species in each North America (NA) and South America (SA) were counted within 1 Myr time bins and averaged after 1,000 randomizations to account for the uncertainty of the ages of the occurrences, 95% confidence intervals are provided in parentheses. The number of immigrant lineages (see definition in the text) shows a significant increase of migrant species in both continents over the past 15–10 Myrs (Fig. 2).

**Table S3. Migration models estimated over the terrestrial taxa phylogenetic data, partitioned by the direction of the migration.**

Model	Subset	d.f.	Time of rate shift (Ma)		Rescaled rates between shifts			log-likelihood	AICc
					2 Ma	15 Ma	30 Ma		
Asymmetric	NA->SA	10	7.54 (5.63–9.71)	23.87 (22.00–25.86)	<b>0.227 (0.192–0.265)</b>	0.036 (0.032–0.038)	0.004 (0.003–0.003)	345.64	-670.51
	SA->NA		6.13 (5.10–6.72)	23.14 (20.50–24.93)	<b>0.304 (0.279–0.335)</b>	0.026 (0.026–0.029)	0.002 (0.002–0.002)		
Symmetric		5	6.01 (5.1–7.24)	23.54 (22.00–25.18)	0.173 (0.153–0.190)	0.027 (0.023–0.026)	0.004 (0.003–0.003)	333.38	-656.55

Different constraints on the times of shift and rates between shifts were applied to assess the presence of significant asymmetry of the rates through time. The preferred model is the asymmetric, in which rates and their variation are estimated independently in the two directions (North to South, South to North). Times of rate shift and rescaled rates between shifts are given as mean and 95% confidence intervals (CI). The estimated rate following the most recent shift (~ 6–8 Ma) is significantly higher in direction S to N as compared with the rate estimated in the opposite direction, as suggested by the lack of overlap between the respective CI.

Table S4

**Table S4a Terrestrial and freshwater taxa - Generalized linear mixed model parameter estimates and likelihood scores using all ecological and biological variables (full model)**

Formula:  $\log(\text{Dispersal.Time}) \sim \text{elevation} + \text{Dispersal.Direction} + \text{Dispersal.Ability} + \text{Biome} + \text{Calibration} + (1 | \text{Class/Family})$

AIC	logLik	deviance	REMLdev
810.6	-386.3	754.9	772.6

**Random effects:**

Groups	Name	Variance	Std.Dev.
Family:Class	(Intercept)	0.40901	0.63954
Class	(Intercept)	0.17929	0.42342
Residual		0.66961	0.8183

Number of obs: 279, groups: Family:Class, 92; Class, 10

**Fixed effects:**

	Estimate	Std. Error	t value
(Intercept)	14.774913	0.543804	27.17
elevation-Lowland	0.198281	0.159263	1.245
elevation-Lowland.Highland	0.149247	0.258569	0.577
elevation-Lowland.Montane	0.030644	0.181332	0.169
Dispersal.Direction-south.to.n	-0.142801	0.139666	-1.022
Dispersal.Ability-moderate	0.079662	0.296096	0.269
Dispersal.Ability-poor	0.334834	0.335246	0.999
Biome-dryforest	-0.461085	0.564123	-0.817
Biome-eurybiomic	0.451573	0.496146	0.91
Biome-forest	0.026395	0.476824	0.055
Biome-humidforest	0.004105	0.473582	0.009
Biome-nonforest	0.131791	0.54127	0.243
Calibration-fossil	0.56672	0.289524	1.957
Calibration-geology	0.427819	0.468079	0.914
Calibration-mixed	0.734744	0.281032	2.614
Calibration-secondary	0.403461	0.312215	1.292

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables. The coefficients correspond to the comparison of the reference level for each categorical variable.



Table S4

**Table S4b Terrestrial and freshwater taxa - Generalized linear mixed model  
parameter estimates and likelihood scores for the model with the variable  
elevational preference removed**

Formula:  $\log(\text{Dispersal.Time}) \sim \text{Dispersal.Direction} + \text{Dispersal.Ability} + \text{Biome} + \text{Calibration} + (1 \mid \text{Class/Family})$

AIC	logLik	deviance	REMLdev
801.7	-384.9	756.7	769.7

**Random effects:**

Groups	Name	Variance	Std.Dev.
Family:Class	(Intercept)	0.39599	0.62928
Class	(Intercept)	0.16355	0.40441
Residual		0.67166	0.81955

Number of obs: 279, groups: Family:Class, 92; Class, 10

**Fixed effects:**

	Estimate	Std. Error	t value
(Intercept)	14.95854	0.51094	29.276
Dispersal.Direction-south.to.n	-0.10233	0.1353	-0.756
Dispersal.Ability-moderate	0.09552	0.29188	0.327
Dispersal.Ability-poor	0.28682	0.32874	0.872
Biome-dryforest	-0.6241	0.5399	-1.156
Biome-eurybiomic	0.31085	0.47817	0.65
Biome-forest	-0.11818	0.45846	-0.258
Biome-humidforest	-0.12928	0.45668	-0.283
Biome-nonforest	-0.04054	0.52019	-0.078
Calibration-fossil	0.62278	0.28356	2.196
Calibration-geology	0.4487	0.46506	0.965
Calibration-mixed	0.82416	0.27171	3.033
Calibration-secondary	0.43267	0.30397	1.423

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables.  
The coefficients correspond to the comparison of the reference level for each categorical variable.

Table S4

**Table S4c Terrestrial and freshwater taxa - Generalized linear mixed model  
parameter estimates and likelihood scores for the model with the  
variable dispersal direction removed**

Formula:  $\log(\text{Dispersal.Time}) \sim \text{elevation} + \text{Dispersal.Ability} + \text{Biome} + \text{Calibration} + (1 \mid \text{Class/Family})$

AIC	logLik	deviance	REMLdev
807.5	-385.8	755.9	771.5

**Random effects:**

Groups	Name	Variance	Std.Dev.
Family:Class	(Intercept)	0.43447	0.65915
Class	(Intercept)	0.17128	0.41386
Residual		0.66192	0.81358

Number of obs: 279, groups: Family:Class, 92; Class, 10

**Fixed effects:**

	Estimate	Std. Error	t value
(Intercept)	14.75308	0.5447	27.085
elevation-Lowland	0.16729	0.15539	1.077
elevation-Lowland.Highland	0.10896	0.25495	0.427
elevation-Lowland.Montane	0.01871	0.18061	0.104
Dispersal.Ability-moderate	0.08889	0.29766	0.299
Dispersal.Ability-poor	0.31293	0.33473	0.935
Biome-dryforest	-0.45371	0.56532	-0.803
Biome-eurybiomic	0.40915	0.49592	0.825
Biome-forest	-0.01219	0.47627	-0.026
Biome-humidforest	-0.0581	0.47051	-0.123
Biome-nonforest	0.10947	0.54142	0.202
Calibration-fossil	0.58433	0.29018	2.014
Calibration-geology	0.42563	0.46812	0.909
Calibration-mixed	0.74345	0.28134	2.643
Calibration-secondary	0.42094	0.3129	1.345

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables.  
The coefficients correspond to the comparison of the reference level for each categorical variable.

Table S4

**Table S4d Terrestrial and freshwater taxa - Generalized linear mixed model parameter estimates and likelihood scores for the model with the variable dispersal ability removed**

Formula:  $\log(\text{Dispersal.Time}) \sim \text{elevation} + \text{Dispersal.Direction} + \text{Biome} + \text{Calibration} + (1 \mid \text{Class/Family})$

AIC	logLik	deviance	REMLdev
806.4	-386.2	756.1	772.4

**Random effects:**

Groups	Name	Variance	Std.Dev.
Family:Class	(Intercept)	0.39985	0.63234
Class	(Intercept)	0.20901	0.45717
Residual		0.66759	0.81706

Number of obs: 279, groups: Family:Class, 92; Class, 10

**Fixed effects:**

	Estimate	Std. error	t value
(Intercept)	15.10824	0.45332	33.33
elevation-Lowland	0.17656	0.15664	1.13
elevation-Lowland.Highland	0.12935	0.25679	0.5
elevation-Lowland.Montane	0.02369	0.18021	0.13
Dispersal.Direction-south.to.n	-0.13141	0.13849	-0.95
Biome-dryforest	-0.6845	0.52201	-1.31
Biome-eurybiomic	0.24346	0.45265	0.54
Biome-forest	-0.17631	0.43577	-0.4
Biome-humidforest	-0.21213	0.42617	-0.5
Biome-nonforest	-0.08196	0.50138	-0.16
Calibration-fossil	0.56295	0.28979	1.94
Calibration-geology	0.44944	0.46611	0.96
Calibration-mixed	0.76649	0.27579	2.78
Calibration-secondary	0.39702	0.31114	1.28

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables.

Table S4

The coefficients correspond to the comparison of the reference level for each categorical variable.

**Table S4e Terrestrial and freshwater taxa - Generalized linear mixed model parameter estimates and likelihood scores for the model with the variable biome type removed**

Formula:  $\log(\text{Dispersal.Time}) \sim \text{elevation} + \text{Origin} + \text{Dispersal.Ability} + \text{Calibration} + (1 \mid \text{Class/Family})$

AIC	logLik	deviance	REMLdev
803.8	-387.9	762.1	775.8

**Random effects:**

Groups	Name	Variance	Std.Dev.
Family:Class	(Intercept)	0.4325	0.65765
Class	(Intercept)	0.16893	0.41101
Residual		0.66874	0.81777

Number of obs: 279, groups: Family:Class, 92; Class, 10

**Fixed effects:**

	Estimate	Std. error	t value
(Intercept)	14.8175	0.3123	47.44
elevation-Lowland	0.1743	0.156	1.12
elevation-Lowland.Highland	0.174	0.2575	0.68
elevation-Lowland.Montane	-0.0144	0.1756	-0.08
Dispersal.Direction-south.to.n	-0.1198	0.1358	-0.88
Dispersal.Ability-moderate	0.1346	0.2963	0.45
Dispersal.Ability-poor	0.3585	0.306	1.17
Calibration-fossil	0.5657	0.2891	1.96
Calibration-geology	0.3044	0.4601	0.66
Calibration-mixed	0.7906	0.2768	2.86
Calibration-secondary	0.4207	0.3076	1.37

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables.  
The coefficients correspond to the comparison of the reference level for each categorical variable.

Table S4

**Table S4f Terrestrial and freshwater taxa - Generalized linear mixed model parameter estimates and likelihood scores for the model with the variable calibration type removed**

Formula:  $\log(\text{Dispersal.Time}) \sim \text{elevation} + \text{Origin} + \text{Dispersal.Ability} + \text{Biome} + (1 \mid \text{Class/Family})$

AIC	logLik	deviance	REMLdev
807.2	-388.6	763	777.2

**Random effects:**

Groups	Name	Variance	Std.Dev.
Family:Class	(Intercept)	0.39996	0.63242
Class	(Intercept)	0.25192	0.50192
Residual		0.67837	0.82363

Number of obs: 279, groups: Family:Class, 92; Class, 10

**Fixed effects:**

	Estimate	Std. Error	t value
(Intercept)	14.93755	0.52779	28.302
Elevation-Lowland	0.29634	0.15432	1.92
elevation-Lowland.Highland	0.22591	0.25659	0.88
elevation-Lowland.Montane	0.04615	0.17813	0.259
Dispersal.Direction-south.to.n	-0.15094	0.13991	-1.079
Dispersal.Ability-moderate	0.16087	0.2982	0.539
Dispersal.Ability-poor	0.4534	0.33726	1.344
Biome-dryforest	-0.26803	0.57214	-0.468
Biome-eurybiomic	0.66184	0.49669	1.332
Biome-forest	0.16838	0.48349	0.348
Biome-humidforest	0.15971	0.48017	0.333
Biome-nonforest	0.27619	0.5463	0.506

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables. The coefficients correspond to the comparison of the reference level for each categorical variable.

Table S5

**Table S5 Terrestrial and freshwater taxa GLMM model comparison**

Model comparison of full model containing all variables vs a model with a variable removed

	Df	AIC	logLik	Chisq	Chi Df	Pr(>Chisq)
Elevational Preference Removed	16	788.73	-378.36			
Full Model	19	792.87	-377.43	1.856	3	0.6029
Dispersal Direction Removed	18	791.93	-377.97			
Full Model	19	792.87	-377.43	1.063	1	0.3026
Dispersal Ability Removed	17	790.12	-378.06			
Full Model	19	792.87	-377.43	1.25	2	0.5353
Biome Type Removed	14	790.15	-381.07			
Full Model	19	792.87	-377.43	7.28	5	0.2006
Calibration Type Removed	15	792.99	-381.5			
Full Model	19	792.87	-377.43	8.123	4	0.08717

Table S5

**Table S5 Terrestrial and freshwater taxa GLMM model comparison**

Model comparison of full model containing all variables vs a model with a variable removed

	Df	AIC	logLik	Chisq	Chi Df	Pr(>Chisq)
Elevational Preference Removed	16	788.73	-378.36			
Full Model	19	792.87	-377.43	1.856	3	0.6029
Dispersal Direction Removed	18	791.93	-377.97			
Full Model	19	792.87	-377.43	1.063	1	0.3026
Dispersal Ability Removed	17	790.12	-378.06			
Full Model	19	792.87	-377.43	1.25	2	0.5353
Biome Type Removed	14	790.15	-381.07			
Full Model	19	792.87	-377.43	7.28	5	0.2006
Calibration Type Removed	15	792.99	-381.5			
Full Model	19	792.87	-377.43	8.123	4	0.08717

Table S6

**Table S6a Marine taxa - Generalized linear mixed model parameter estimates and likelihood scores using the variables dispersal ability and calibration type (full model)**

 Formula:  $\log(\text{Dispersal.Time}) \sim \text{Dispersal.Ability} + \text{Calibration} + (1 \mid \text{Class/Order/Family})$ 

	AIC	logLik	deviance	REMLdev
	171.6	78.8	157.1	169.6
<b>Random effects:</b>				
Groups	Name	Variance	Std.Dev.	
Family:(Order:Class)	(Intercept)	0.12478	0.35324	
Order:Class	(Intercept)	0.36113	0.60094	
Class		0.25324	0.50323	
Residual		0.26122	0.5111	

Number of obs: 80, groups: Family:(Order:Class), 29; Order:Class, 17; Class, 7

**Fixed effects:**

	Estimate	Std. Error	t value
(Intercept)	1.1059	0.3293	3.358
Dispersal.Ability-moderate	-0.2902	0.6096	-0.476
Calibration-fossil	1.3776	0.295	4.669

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables. The coefficients correspond to the comparison of the reference level for each categorical variable.

**Table S6b Marine taxa - Generalized linear mixed model parameter estimates and likelihood scores for the model with the variable dispersal ability removed**

 Formula:  $\log(\text{Dispersal.Time}) \sim \text{Calibration} + (1 \mid \text{Class/Order/Family})$ 

	AIC	logLik	deviance	REMLdev
	170.6	-79.31	157.1	158.6
<b>Random effects:</b>				
Groups	Name	Variance	Std.Dev.	
Family:(Order:Class)	(Intercept)	0.1227	0.35029	
Order:Class	(Intercept)	0.34852	0.59036	
Class		0.19805	0.44502	
Residual		0.26186	0.51172	

Number of obs: 80, groups: Family:(Order:Class), 29; Order:Class, 17; Class, 7

**Fixed effects:**

	Estimate	Std. Error	t value
(Intercept)	1.0566	0.2915	3.625
Calibration-fossil	1.3469	0.2854	4.72

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables. The coefficients correspond to the comparison of the reference level for each categorical variable.



Table S6

**Table S6c Marine taxa - Generalized linear mixed model parameter estimates and likelihood scores for the model with the variable calibration type removed**Formula:  $\log(\text{Dispersal.Time}) \sim \text{Dispersal.Ability} + (1 \mid \text{Class/Order/Family})$ 

	AIC	logLik	deviance	REMLdev
	184.5	-86.25	173.1	172.5
<b>Random effects:</b>				
Groups	Name	Variance	Std.Dev.	
Family:(Order:Class)	(Intercept)	0.43065	0.65624	
Order:Class	(Intercept)	0.50772	0.71255	
Class		0.19064	0.43662	
Residual		0.26078	0.51067	

Number of obs: 80, groups: Family:(Order:Class), 29; Order:Class, 17; Class, 7

**Fixed effects:**

	Estimate	Std. Error	t value
(Intercept)	1.4704	0.347	4.237
Dispersal.Ability-moderate	0.4676	0.7062	0.662

Note: categorical variables with K levels are entered in the model as a sequence of K-1 variables. The coefficients correspond to the comparison of the reference level for each categorical variable.

Table S7

**Table S7 Marine taxa GLMM model comparison**

Model comparison of full model containing all variables vs a model with a variable removed

	Df	AIC	logLik	Chisq	Chi Df	Pr(>Chisq)
Dispersal Ability Removed	6	169.13	-78.563			
Full Model	7	171.08	-78.54	0.0465	1	0.8293
	Df	AIC	logLik	Chisq	Chi Df	Pr(>Chisq)
Calibration Type Removed	6	185.12	-86.561			
Full Model	7	171.08	-78.54	16.043	1	<0.0000

**Table S8. Survey of approximate ages of extant taxa reported from published phylogenies, the complete data is deposited at Dryad (<http://datadryad.org>).**

Class : Order : Family	Taxon / clade (constituent species)	Crown age (MA)	Reference
<b>Marine organisms</b>			
Actinopterygii : Mugiliformes : Mugilidae	<i>Agonostomous monticola</i>	20.7	1
Actinopterygii : Perciformes : Carangidae	<i>Trachurus</i>	7.7	2
Actinopterygii : Perciformes : Chaetodontidae	<i>Chaetodon humeralis</i> vs. <i>C. ocellatus</i>	3.4	3
Actinopterygii : Perciformes : Cichlidae	<i>Andinoacara coeruleopunctatus</i>	5	4
Actinopterygii : Perciformes : Cichlidae	<i>Caquetaia umbrifera</i> (Central America)	33.89	5
Actinopterygii : Perciformes : Cichlidae	<i>Cichlasoma facetus</i>	19.56	5
Actinopterygii : Perciformes : Cichlidae	<i>'Heros' atromaculatus</i>	3.6	6
Actinopterygii : Perciformes : Cichlidae	Central American Heroini	54.15	6
Actinopterygii : Perciformes : Cichlidae	<i>Heros' festae</i> group	41.39	6
Actinopterygii : Perciformes : Gerridae	<i>Gerres cinereus</i> vs. <i>G. cinereus</i>	2.55	7
Actinopterygii : Perciformes : Haemulidae	<i>Anisotremus interruptus</i> vs. <i>A. surinamensis</i>	0.8	7
Actinopterygii : Perciformes : Haemulidae	<i>Anisotremus taeniatus</i> vs. <i>A. virginicus</i>	2.2	7
Actinopterygii : Perciformes : Lutjanidae	<i>Lutjanus argentiventris</i> vs. <i>L. apodus</i>	1.75	7
Actinopterygii : Perciformes : Mulloidichthys	<i>Mulloidichthys dentatus</i> vs. <i>M. martinicus</i>	0.65	7
Actinopterygii : Perciformes : Poeciliidae	<i>Pseudopoecilia festae</i>	22	4
Actinopterygii : Perciformes : Poeciliidae	<i>Neoheterandria elegans</i> + <i>N. tridentiger</i>	22	4
Actinopterygii : Perciformes : Poeciliidae	<i>Poecilia reticulata</i>	3	4
Actinopterygii : Perciformes : Poeciliidae	<i>Poecilia</i> spp.	27.8	8
Actinopterygii : Perciformes : Pomacanthidae	<i>Centropyge aurantonota</i> + ( <i>C. loriculus</i> , <i>C. potteri</i> )	13.9	9
Actinopterygii : Perciformes : Pomacanthidae	<i>Pomacanthus zonipectus</i> , <i>P. parularcuatus</i>	19.9	9
Actinopterygii : Perciformes : Pomacanthidae	<i>Holacanthus passer</i> + ( <i>H. bermudensis</i> , <i>H. ciliaris</i> )	5	9
Actinopterygii : Perciformes : Pomacentridae	<i>Abudefduf concolor</i> vs. <i>A. taurus</i>	0.7	7
Actinopterygii : Perciformes : Pomacentridae	<i>Abudefduf troshelii</i> vs. <i>A. saxatilis</i>	2.25	7
Actinopterygii : Perciformes : Serranidae	<i>Rypticus bicolor</i> vs. <i>R. saponaceus</i>	1.6	7

Actinopterygii : Perciformes : Serranidae	<i>Paranthias colonus</i> vs. <i>P. furcifer</i>	2.4	7
Actinopterygii : Perciformes : Labridae	<i>Halichoeres nicholsi</i> vs. <i>H. bivittatus</i> clade	3.55	7
Actinopterygii : Perciformes : Labridae	<i>Halichoeres dispilus</i> vs. <i>H. pictus</i>	5.1	7
Actinopterygii : Perciformes : Pomacentridae	<i>Chromis atrilobata</i> vs. <i>C. multilineata</i>	4.7	7
Actinopterygii : Perciformes : Priacanthidae	<i>Heteropriacanthus cruentatus</i> vs. <i>H. cruentatus</i>	5.35	7
Actinopterygii : Siluriformes : Heptapteridae	<i>Pimelodella chagresi</i>	3.5	10
Actinopterygii : Siluriformes : Heptapteridae	<i>Pimelodella chagresi</i>	1	10
Actinopterygii : Siluriformes : Heptapteridae	<i>Rhamdia guatemalensis</i> , <i>R. laticauda</i> , <i>R. cinerascens</i>	6.05	11
Actinopterygii : Tetraodontiformes: Balistidae	<i>Melichthys niger</i> vs. <i>M. niger</i>	0.1	7
Actinopterygii : Tetraodontiformes: Balistidae	<i>Diodon hystrix</i> vs. <i>D. hystrix</i>	0.3	7
Actinopterygii : Tetraodontiformes: Monacanthidae	<i>Alutera scripta</i> vs. <i>A. scripta</i>	0.45	7
Actinopterygii: Scorpaeniformes: Scorpaenidae	<i>Scorpaena mystes</i> vs. <i>S. plumieri</i>	2.75	7
Bivalvia : Arcoida : Arcidae	<i>Arca imbricata</i> EP6-8/WA6-10	3.3	12
Bivalvia : Arcoida : Arcidae	<i>Arca imbricata</i> EP6/WA7	3.5	12
Bivalvia : Arcoida : Arcidae	<i>Arca imbricata</i> EP3/WA4-5	14	12
Bivalvia : Arcoida : Arcidae	<i>Arca imbricata</i> EP9/WA11	14	12
Bivalvia : Arcoida : Arcidae	<i>Barbatia gradata</i> , <i>B. domingensis</i>	17.4	13
Chondrichthyes : Squatiniformes : Squatinidae	<i>Squatina californica</i> , <i>S. dumeril</i>	6.11	14
Crustacea: Decapoda: Alpheidae	<i>Sesarma rhizophorae</i> vs. <i>S. reticulatum</i> clade	2.05	7
Crustacea: Decapoda: Alpheidae	<i>Alpheus antepenultimus</i> A vs. <i>A. chacei</i>	2.7	7
Crustacea: Decapoda: Alpheidae	<i>Sesarma (sulcatum, aequatoriale)</i> vs. <i>S. crassipes</i>	3.1	7
Crustacea: Decapoda: Alpheidae	<i>Alpheus rostratus</i> vs. <i>A. paracrinatus</i> spot	3.2	7
Crustacea: Decapoda: Alpheidae	<i>Alpheus colombiensis</i> vs. <i>A. estuarensis</i>	3.4	7
Crustacea: Decapoda: Alpheidae	<i>Alpheus websteri</i> vs. <i>A. websteri</i>	3.65	7
Crustacea: Decapoda: Alpheidae	<i>Alpheus cylindricus</i> vs. <i>A. cylindricus</i>	4.35	7
Crustacea: Decapoda: Alpheidae	<i>Synalpheus brevicarpus</i> vs. <i>S. brevicarpus</i>	4.35	7
Crustacea: Decapoda: Alpheidae	<i>Alpheus floridanus</i> B' vs. <i>A. floridanus</i> B	4.7	7
Crustacea: Decapoda: Alpheidae	<i>Alpheus canalis</i> -sp.B(blue) = <i>millsae</i> vs. <i>A. nuttingi</i>	4.7	7

Alpheidae				
Crustacea: Decapoda: Alpheidae	<i>Alpheus paracrinitus no spot</i> vs. <i>A. paracrinitus no spot</i>	4.75	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus panamensis</i> vs. <i>A. formosus-sp.A</i>	4.75	7	
Crustacea: Decapoda: Alpheidae	<i>Synalpheus digueti</i> vs. <i>S. minus</i>	4.75	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus bouvieri</i> vs. <i>A. bouvieri</i>	5.5	7	
Crustacea: Decapoda: Alpheidae	<i>Xiphopenaeus riveti</i> vs. <i>X. sp. 2</i>	5.55	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus malleator</i> vs. <i>A. malleator</i>	5.75	7	
Crustacea: Decapoda: Alpheidae	<i>Synalpheus fritzmuelleri</i> vs. <i>S. fritzmuelleri</i>	6.05	7	
Crustacea: Decapoda: Alpheidae	<i>Penaeus vannamei</i> vs. <i>P. duorarum clade</i>	6.05	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus umbo</i> vs. <i>A. schmitti</i>	6.6	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus saxidomus simus</i>	6.95	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus floridanus A'</i> vs. <i>A. floridanus A</i>	7.55	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus normanni</i> vs. <i>A. normanni Brazil</i>	7.75	7	
Crustacea: Decapoda: Alpheidae	<i>Emerita rathbunae</i> vs. <i>E. brasiliensis clade</i>	7.9	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus (naos, aequus)</i> vs. <i>A. christofferseni</i>	8.9	7	
Crustacea: Decapoda: Alpheidae	<i>Alpheus cristulifrons</i> vs. <i>A. cristulifrons</i>	9.45	7	
Crustacea: Decapoda: Alpheidae	<i>Austinixa felipensis</i> vs. <i>A. cristata clade</i>	9.75	7	
Crustacea: Decapoda: Alpheidae	<i>Synalpheus bannerorum</i> vs. <i>S. dominicensis</i>	11.3	7	
Echinodermata:				
Temnopleuroida:Toxopneustidae	<i>Tripneustes depressus-gratilla</i> vs. <i>T. ventricosus</i>	4.35	7	
Echinoidea: Arbacioidae:Arbaciidae	<i>Arbacia spatuligera</i> vs. <i>A. lixula</i>	6.75	7	
Echinoidea: Spatangoidae:Brissidae	<i>Meoma ventricosa grandis</i> vs. <i>M. ventricosa</i>	2.25	7	
Echinoidea:Camarodonta:Echinometridae	<i>Echinometra vanbrunti</i> vs. <i>E. lucunter</i>	5.15	7	
Echinoidea:Cidaroidae:Cidaridae	<i>Eucidaris thouarsi</i> vs. <i>E. tribuloides</i>	4.75	7	
Echinoidea:Diadematoidea : Diadematidae	<i>Diadema mexicanum</i> vs. <i>D. antillarum</i>	2.1	7	
Echinoidea:Diadematoidea: Diadematidae	<i>Astropyga pulvinata</i> vs. <i>A. magnifica</i>	4.95	7	
Echinoidea:Temnopleuroida:Toxopneustidae	<i>Lytechinus semituberculatus</i> vs. <i>L. variegatus-williamsi</i>	6.75	7	
Gastropoda : Littorinimorpha : Littorinidae	<i>Littoraria rosewateri</i> , <i>L. tessellata</i>	24.2	15	
Gastropoda : Littorinimorpha : Littorinidae	<i>Littoraria nebulosa</i> + ( <i>L. varia</i> , <i>L. zebra</i> , <i>L. irrorata</i> , <i>L. variegata</i> )	25.7	15	

Gastropoda : Littorinimorpha : Littorinidae	<i>Littoraria irrorara, L. variegata</i>	8.5	15
Gastropoda : Littorinimorpha : Littorinidae	<i>Echinolittorina angustor, E. periciliata</i>	19.2	16
Gastropoda : Littorinimorpha : Littorinidae	<i>Echinolittorina interrupta</i> + ( <i>E. aspera, E. dubiosa, E. tenuistriata</i> )	19	16
Gastropoda : Littorinimorpha : Littorinidae	<i>Echinolittorina riser</i> + ( <i>E. apicina E. paytensis</i> )	14.2	16
Gastropoda : Littorinimorpha : Littorinidae	<i>Echinolittorina conspersa, C. modesta</i> + <i>E. ziczac</i>	15.5	16
Gastropoda : Littorinimorpha : Littorinidae	<i>Echinolittorina galapagensis</i> + ( <i>E. vermeiji, E. tuberculata, E. granosa</i> )	32.5	16
Gastropoda : Neritimorpha : Neritidae	<i>Nerita scabricosta, N. peloronta / N. versicolor</i>	19.4	17
Gastropoda : Neritimorpha : Neritidae	<i>Nerita funiculata, N. tessellata / N. fulgurans / N. senegalensis</i>	23.3	17
Gastropoda : Unranked : Bullidae	<i>Bulla gouldiana, B. punctulata</i> + ( <i>B. mabiliei</i> )	16.8	18
Gastropoda : Unranked : Conidae	<i>Conus bartschii, C. brunneus</i>	14.3	19
Gastropoda : Unranked : Conidae	<i>Conus regius, C. gladiator</i> + ( <i>C. mus</i> )	6.9	19
Gastropoda : Unranked : Conidae	<i>Conus ermineus, C. purpurascens</i>	7.3	19
Gastropoda : Unranked : Muricidae	<i>Stramonita canaliculata, S. rustica, S. haemastoma</i>	13	20
Gastropoda : Unranked : Muricidae	<i>Stramonita brasiliensis, S. floridana, S. biserialis</i>	7	20
Gastropoda : Unranked : Cerithiidae	<i>Cerithium stercusmuscarum, C. lutosum</i>	3.7	21
Gastropoda : Unranked : Cerithiidae	<i>Cerithium atratum, C. uncinatum</i>	5.1	21
Gastropoda : Unranked : Potamididae	<i>Cerithidea montagnaei, c. scalariformis</i>	6.4	21
Hydrozoa : Anthomedusae : Hydridae	<i>Ophioblennius steindachneri</i> vs. <i>O. atlanticus</i>	12.4	22
Mammalia : Carnivora : Phocidae	<i>Histriophoca</i> vs. <i>Pagophilus</i>	3.4	23
<b>Terrestrial organisms</b>			
Amphibia : Anura : Brachycephalidae	<i>Pristimantis ridens</i>	17	24
Amphibia : Anura : Bufonidae	<i>Rhinella marina</i>	2.75	25
Amphibia : Anura : Dendrobatidae	<i>Phyllobates (P. lugubris, P. vittatus)</i>	3.17	26
Amphibia : Anura : Dendrobatidae	<i>Dendrobates granuliferus</i>	1.39	26
Amphibia : Anura : Dendrobatidae	<i>Dendrobates</i> I ( <i>D. arboreus, D. pumilio, D. speciosus, D. vicentei</i> )	3.01	26
Amphibia : Anura : Dendrobatidae	<i>Colostethus pratti</i>	2.94	26
Amphibia : Anura : Dendrobatidae	<i>Dendrobates</i> II ( <i>D. claudiae, D. fulguritus, D. minutus</i> )	9.31	26
Amphibia : Anura : Dendrobatidae	<i>Silverstoneia nubicola</i>	4.52	26

Amphibia : Anura : Dendrobatidae	<i>Silverstoneia flotator</i>	4.15	26
Amphibia : Anura : Dendrobatidae	<i>Colostethus panamensis</i>	3.37	26
Amphibia : Anura : Dendrobatidae	<i>Allobates talamancae</i>	3.17	26
Amphibia : Anura : Dendrobatidae	<i>Dendrobates auratus</i>	0.58	26
Amphibia : Anura : Eleutherodactylidae	<i>Craugastor (C. podiciferus, C. longirostris, C. pygmaeus, C. montanus)</i>	30.51	27
Amphibia : Anura : Eleutherodactylidae	<i>Eleutherodactylus (E. zeus, E. marnockii)</i>	19.05	27
Amphibia : Anura : Leptodactylidae	<i>Physalaemus pustulosus</i>	5.5	28
Amphibia : Anura : Microhylidae	Origin of clade containing <i>Elachistocleis</i> , <i>Gastrophryne</i> , <i>Hypopachus</i> vs	30.3	29
Amphibia : Caudata : Plethodontidae	South American <i>Bolitoglossa</i>	18	30
Amphibia : Caudata : Plethodontidae	<i>Oedipina (O. complex, O. parvipes)</i>	6	31
Arachnida : Araneae : Sicariidae	<i>Loxosceles reclusa</i> group	33	32
Arachnida : Pseudoscorpiones : Chernetidae	<i>Cordylochernes scorpioides</i>	10.88	33
Aves : Anseriformes : Anatidae	<i>Anas acuta</i> & <i>A. georgica</i>	1.33	34, 35
Aves : Anseriformes : Anatidae	<i>Anas americana</i> & <i>A. nassibilatrix</i>	0.08	34, 35
Aves : Anseriformes : Anatidae	<i>Anas flavirostris</i> & <i>A. carolinensis</i>	1.44	34, 35
Aves : Apodiformes : Trochilidae	<i>Eupherusa/Microchera/Elvira</i>	4.3	34, 36
Aves : Apodiformes : Trochilidae	<i>Bee Hummingbirds</i>	4.79	34, 36
Aves : Apodiformes : Trochilidae	<i>Mt. Gems &amp; Bee Hummingbirds</i>	12.8	34, 36
Aves : Columbiformes : Columbidae	<i>Zenaida auriculata</i> vs. <i>Z. macroura/Z. graysoni</i>	1.48	37
Aves : Columbiformes : Columbidae	<i>Zenaida asiatica</i> vs. <i>Z. meloda</i>	1.25	37
Aves : Columbiformes : Columbidae	<i>Geotrygon chiriquensis</i> vs. <i>G. goldmani</i>	0.95	37
Aves : Columbiformes : Columbidae	<i>Leptotila verreauxi</i>	0.86	37
Aves : Columbiformes : Columbidae	<i>Leptotila cassini</i>	0.11	37
Aves : Columbiformes : Columbidae	<i>Leptotila verreauxi/L. jamaicensis</i> vs. <i>L. cassini/L. plumbeiceps/L. rufaxilla/L. megalura</i>	3.44	37
Aves : Coraciiformes : Motmotidae	<i>Baryphthengus ruficapillus/marti</i> & <i>B. martii</i>	3.24	38
Aves : Cuculiformes : Cuculidae	<i>Geococcyx velox/californianus</i> & <i>Neomorphus geoffroyi</i>	7.88	34
Aves : Cuculiformes : Cuculidae	<i>Piaya cayana</i>	1.12	39
Aves : Falconiformes :	<i>Micrastur semitorquatus</i>	0.4	40

Falconidae			
Aves : Falconiformes : Falconidae	<i>Micrastur ruficollis</i>	0.75	40
Aves : Galliformes : Cracidae	<i>Crax rubra</i> & <i>Crax SA clade</i>	2.44	41, 42
Aves : Passeriformes : Cardinalidae	<i>Cardinalis sinuatus/C. cardinalis</i> & <i>C. phoeniceus</i>	4.92	38
Aves : Passeriformes : Cardinalidae	<i>Caryothraustes canadensis</i> & <i>C. poliogaster</i>	1.3	38
Aves : Passeriformes : Cardinalidae	<i>Cyanocopsa cyanoides</i>	0.76	38
Aves : Passeriformes : Cardinalidae	<i>Granatellus sallaei/venustus</i> & <i>G. pelzelni</i>	6.13	38
Aves : Passeriformes : Cardinalidae	<i>Pheucticus tibialis/chrysopheplus</i> & <i>P. aureoventris/chrysogaster</i>	1.32	38
Aves : Passeriformes : Cardinalidae	<i>Piranga flava</i>	3.34	34, 43
Aves : Passeriformes : Cardinalidae	<i>Piranga leucoptera</i>	0.73	34, 43
Aves : Passeriformes : Cardinalidae	<i>Piranga leucoptera</i> & <i>P. rubriceps</i>	3.7	34, 43
Aves : Passeriformes : Cardinalidae	<i>Rhodothraupis celaeno</i> & <i>Periporphyrus erythromelas</i>	5.2	38
Aves : Passeriformes : Cardinalidae	<i>Cyanocopsa</i>	6.06	34
Aves : Passeriformes : Cardinalidae	<i>Amaurospiza concolor</i> & <i>A. moesta/carrizalensis</i>	3.1	34
Aves : Passeriformes : Cinclidae	<i>Cinclus leucocephalus/schulzi</i> & <i>C. mexicanus</i>	4.66	34
Aves : Passeriformes : Corvidae	<i>Cyanocorax dickeyi</i> & <i>C. chrysops/cayanus</i>	2.11	34, 44
Aves : Passeriformes : Corvidae	<i>Cyanolyca pulchra</i> & <i>C. cucullata</i>	3.95	34, 44
Aves : Passeriformes : Corvidae	<i>Cyanolyca viridicyana/turcosa/armillata/pulchra/cucullata</i>	10.44	34, 44
Aves : Passeriformes : Cotingidae	<i>Querula purpurata</i>	0.64	39
Aves : Passeriformes : Emberizidae	<i>Arremonops tocuyensis</i> & <i>A. conirostris</i>	5.23	34
Aves : Passeriformes : Emberizidae	<i>Arremon aurantirostris</i>	2.76	34
Aves : Passeriformes : Emberizidae	<i>Arremonops conirostris</i>	3.53	34
Aves : Passeriformes : Emberizidae	<i>Atlapetes</i>	3.72	34
Aves : Passeriformes : Emberizidae	<i>Arremon brunneinucha</i>	1.02	34, 45
Aves : Passeriformes : Emberizidae	<i>Arremon torquatus</i> & <i>A. torquatus/atricapillus</i>	3.79	34, 45
Aves : Passeriformes : Emberizidae	<i>Chlorospingus canigularis</i>	3.5	34, 45
Aves : Passeriformes : Emberizidae	<i>Chlorospingus flavigularis</i>	2.3	34, 46
Aves : Passeriformes : Emberizidae	<i>Chlorospingus ophthalmicus</i>	3.32	34, 46
Aves : Passeriformes :	<i>Lysurus castaneiceps</i> & <i>L. crassirostris</i>	1.28	34, 47



Emberizidae			
Aves : Passeriformes : Emberizidae	<i>Arremon brunneinucha/virenticeps &amp; A. torquatus/Arremon</i>	10.5	34, 47
Aves : Passeriformes : Fringillidae	<i>Carduelis notata &amp; SA Carduelis</i>	1.55	34, 48
Aves : Passeriformes : Furnariidae	<i>Xenops minutus</i>	0.41	34
Aves : Passeriformes : Furnariidae	<i>Xiphorhynchus erythropygius</i>	0.79	34, 49
Aves : Passeriformes : Furnariidae	<i>Xiphorhynchus flavigaster &amp; X. lachrymosus</i>	2.12	34, 49
Aves : Passeriformes : Furnariidae	<i>Xiphorhynchus susurrans</i>	1.05	38
Aves : Passeriformes : Furnariidae	<i>Automolus ochrolaemus</i>	3.06	39
Aves : Passeriformes : Furnariidae	<i>Dendrocincla fuliginosa vs. D. anabatina</i>	2.6	39
Aves : Passeriformes : Furnariidae	<i>Dendrocincla fuliginosa</i>	0.16	39
Aves : Passeriformes : Furnariidae	<i>Sclerurus mexicanus</i>	7.41	39
Aves : Passeriformes : Hirundinidae	<i>Notiochelidon pileata &amp; Neochelidon tibialis</i>	2.86	34, 50
Aves : Passeriformes : Hirundinidae	<i>Petrochelidon fulva &amp; P. rufocollaris</i>	1.62	34, 50
Aves : Passeriformes : Hirundinidae	<i>Stelgidopteryx serripennis &amp; S. ruficollis</i>	3.81	34, 50
Aves : Passeriformes : Hirundinidae	<i>Tachycineta albilinea &amp; T. albiventer</i>	2.3	34, 50
Aves : Passeriformes : Icteridae	<i>Icterus graceannae &amp; I. pectoralis</i>	2.19	38
Aves : Passeriformes : Icteridae	<i>Icterus gularis &amp; I. nigrogularis</i>	0.47	38
Aves : Passeriformes : Icteridae	<i>Icterus mesomelas</i>	0.31	34, 51
Aves : Passeriformes : Icteridae	<i>Molothrus ater &amp; M. bonariensis</i>	1.13	34, 52
Aves : Passeriformes : Icteridae	<i>Psarocolius decumanus</i>	1.19	34, 53
Aves : Passeriformes : Icteridae	<i>Psarocolius montezuma &amp; P. bifasciatus</i>	0.63	34, 53
Aves : Passeriformes : Icteridae	<i>Quiscalus lugubris &amp; Q. major/mexicanus/niger</i>	1.05	38
Aves : Passeriformes : Icteridae	<i>South American clade of Black Birds</i>	6.53	34, 52
Aves : Passeriformes : Motacillidae	<i>Anthus spragueii &amp; A. lutescens</i>	4.97	34, 54
Aves : Passeriformes : Parulidae	<i>Basileuterus culicivorus</i>	2	55
Aves : Passeriformes : Parulidae	<i>Myioborus South Am. clade (M. albifacies - M. brunniceps)</i>	3.72	56
Aves : Passeriformes : Parulidae	<i>Geothlypis aequinoctialis &amp; G. poliocephala</i>	3.35	38
Aves : Passeriformes : Parulidae	<i>Geothlypis semiflava</i>	1.28	34, 57

Aves : Passeriformes : Parulidae	<i>Myioborus miniatus</i>	0.86	34, 56
Aves : Passeriformes : Parulidae	<i>Myioborus torquatus</i> & <i>M. melanocephalus/ornatus/albifrons</i>	2.54	34, 56
Aves : Passeriformes : Parulidae	<i>Phaeothlypis fulvicauda/rivularis</i>	2.45	34, 58
Aves : Passeriformes : Pipridae	<i>Lepidothrix coronata</i>	0.55	34, 59
Aves : Passeriformes : Poliophtilidae	<i>Microbates cinereiventris</i>	1.69	38
Aves : Passeriformes : Poliophtilidae	<i>Poliophtila plumbea</i> & <i>P. plumbea</i>	1.66	34
Aves : Passeriformes : Poliophtilidae	<i>Poliophtila plumbea</i> & <i>P. plumbea/albilaris</i>	4.01	38
Aves : Passeriformes : Poliophtilidae	<i>Ramphocaenus melanurus</i>	4.18	38
Aves : Passeriformes : Poliophtilidae	<i>Poliophtila dumicola/plumbea/lactea</i> & <i>P. albilaris/melanura/californica/nigriceps/plumbea/caerulea*</i>	6.13	34
Aves : Passeriformes : Thamnophilidae	<i>Thamnophilus nigriceps</i> vs. <i>T. praecox</i>	1.1	60
Aves : Passeriformes : Thamnophilidae	<i>Thamnophilus atrinucha/bridgesi</i> & <i>T. bernardi</i>	4.2	34, 60
Aves : Passeriformes : Thamnophilidae	<i>Cymbilaimus lineatus</i>	0.24	39
Aves : Passeriformes : Thamnophilidae	<i>Myrmotherula axillaris</i>	0.25	39
Aves : Passeriformes : Thraupidae	<i>Diglossa plumbea</i> vs. <i>D. baritula</i>	0.8	61
Aves : Passeriformes : Thraupidae	<i>Ramphocelus dimidiatus</i>	0.9	62
Aves : Passeriformes : Thraupidae	<i>Tachyphonus delatrii</i>	8.8	62
Aves : Passeriformes : Thraupidae	<i>Tachyphonus luctuosus</i>	5.6	62
Aves : Passeriformes : Thraupidae	<i>Lanio fulvus</i> vs. <i>L. leucothorax/L. aurantius</i>	4.6	62
Aves : Passeriformes : Thraupidae	<i>Eucometis penicillata</i>	4.2	62
Aves : Passeriformes : Thraupidae	<i>Tachyphonus rufus</i>	3.7	62
Aves : Passeriformes : Thraupidae	<i>Heterospingus rubrifrons</i> & <i>H. xanthopygius</i>	1	34, 62, 63
Aves : Passeriformes : Thraupidae	<i>Saltator atripennis</i> & <i>S. atriceps</i>	3.08	38
Aves : Passeriformes : Thraupidae	<i>Saltator grandis/coerulescens</i> & <i>S. coerulescens/striatipectus</i>	3.85	34
Aves : Passeriformes : Thraupidae	<i>Saltator grossus</i>	0.51	34
Aves : Passeriformes : Thraupidae	<i>Saltator maximus</i>	0.56	38
Aves : Passeriformes : Thraupidae	<i>Tangara fucosa</i> & <i>T. dowii</i>	3.14	34, 64
Aves : Passeriformes : Thraupidae	<i>Tangara inornata</i> & <i>T. mexicana</i>	2.42	34, 64
Aves : Passeriformes :	<i>Ramphocelus passerinii/costaricensis</i> & <i>R.</i>	2.98	34, 65

Thraupidae	<i>icteronotus</i>		
Aves : Passeriformes : Thraupidae	<i>Chlorophanes spiza</i>	0.43	39
Aves : Passeriformes : Thraupidae	<i>Tangara gyrola</i>	0.49	39
Aves : Passeriformes : Tityridae	<i>Schiffornis turdina</i>	0.62	34, 66
Aves : Passeriformes : Tityridae	<i>Tityra semifasciata</i>	0.71	39
Aves : Passeriformes : Troglodytidae	<i>Campylorhynchus chiapensis &amp; C. griseus</i>	1.11	38
Aves : Passeriformes : Troglodytidae	<i>Cantorchilus nigricapillus</i>	0.87	38
Aves : Passeriformes : Troglodytidae	<i>Henicorhina leucophrys</i>	2.67	34
Aves : Passeriformes : Troglodytidae	<i>Henicorhina leucophrys &amp; H. leucosticta</i>	5.07	34
Aves : Passeriformes : Troglodytidae	<i>Henicorhina leucosticta</i>	4.03	38
Aves : Passeriformes : Troglodytidae	<i>Pheugopedius sclateri &amp; P. felix</i>	1.15	38
Aves : Passeriformes : Troglodytidae	<i>Thryothorus thoracicus &amp; T. leucopogon</i>	2.15	34, 67
Aves : Passeriformes : Troglodytidae	<i>Troglodytes aedon</i>	2.14	38
Aves : Passeriformes : Troglodytidae	<i>Campylorhynchus (Basal NA and SA split)</i>	7.07	34
Aves : Passeriformes : Troglodytidae	<i>Troglodytes rufociliatus &amp; T. ochraceus/rufulus/solstitialis</i>	4.35	34
Aves : Passeriformes : Troglodytidae	<i>Microrcerculus marginatus</i>	4.25	39
Aves : Passeriformes : Turdidae	<i>Myadestes melanops &amp; M. coloratus</i>	0.85	34, 68
Aves : Passeriformes : Turdidae	<i>Turdus albicollis &amp; T. assimilis</i>	3.97	34, 69
Aves : Passeriformes : Turdidae	<i>Cichlopsis/Entomodestes &amp; Catharus clade</i>	6.51	34, 70
Aves : Passeriformes : Tyrannidae	<i>Mionectes oleagineus (node C)</i>	0.2	71
Aves : Passeriformes : Tyrannidae	<i>Mionectes oleagineus (SW + Central Panama clade)</i>	0.5	71
Aves : Passeriformes : Tyrannidae	<i>Mionectes oleagineus (NW Panama + SE Mexico)</i>	0.1	71
Aves : Passeriformes : Tyrannidae	<i>Elaenia flavogaster</i>	0.66	34, 72
Aves : Passeriformes : Tyrannidae	<i>Elaenia frantzii &amp; E. pallatangae</i>	1.96	34, 72
Aves : Passeriformes : Tyrannidae	<i>Myiarchus tuberculifer</i>	0.48	34, 73
Aves : Passeriformes : Tyrannidae	<i>Myiarchus tyrannulus</i>	0.52	34, 73
Aves : Passeriformes : Tyrannidae	<i>Attila spadiceus</i>	1.83	39
Aves : Passeriformes : Tyrannidae	<i>Colonia colonus</i>	0.44	39

Aves : Passeriformes : Vireonidae	<i>Cyclarhis gujanensis</i>	2.94	38
Aves : Passeriformes : Vireonidae	<i>Hylophilus decurtatus</i>	2.72	34
Aves : Passeriformes : Vireonidae	<i>Hylophilus ochraceiceps</i>	2.23	38
Aves : Passeriformes : Vireonidae	<i>Vireo leucophrys</i> & <i>V. chiriquensis</i>	1.24	34
Aves : Passeriformes : Vireonidae	<i>Vireo olivaceus</i> & <i>V. flavoviridis</i>	2.63	34
Aves : Picidae : Picinae	<i>Veniliornis</i> South Am, clade ( <i>V. affinis</i> - <i>P. mixtus</i> )	5.1	74
Aves : Picidae : Picinae	Colaptes South Am. clade	3.6	75
Aves : Piciformes : Bucconidae	<i>Notharchus tectus</i>	3.09	38
Aves : Piciformes : Ramphastidae	<i>Aulacorhynchus prasinus</i>	4.17	34, 76
Aves : Piciformes : Ramphastidae	<i>Ramphastos swainsonii</i> & <i>R. ambiguus</i>	0.38	34, 77
Aves : Piciformes : Ramphastidae	<i>Pteroglossus frantzii</i> & <i>P. sanguineus/erythropygius</i>	0.4	34, 78
Aves : Piciformes : Ramphastidae	<i>Pteroglossus frantzii/sanguineus/erythropygius/torquatus</i>	1	81
Aves : Piciformes : Semnornithidae	<i>Semnornis frantzii</i> & <i>S. ramphastinus</i>	5.05	34, 39
Aves : Psittaciformes : Psittacidae	<i>Brotogeris jugularis</i> vs. <i>Brotogeris pyrrhopterus</i>	2.01	81
Aves : Psittaciformes : Psittacidae	<i>Amazona autumnalis</i>	1.23	34, 82
Aves : Psittaciformes : Psittacidae	<i>Amazona farinosa</i>	2.71	34, 82
Aves : Psittaciformes : Psittacidae	<i>Amazona ochrocephala</i>	1	34, 82, 83
Aves : Psittaciformes : Psittacidae	<i>Forpus cyanopygius</i> & <i>F. passeririnus/xanthops/xanthopterygius/conspicillatus/coelestis</i>	3.22	38
Aves : Psittaciformes : Psittacidae	<i>Pionus senilis</i> & <i>P. chalcopterus</i>	0.31	38
Aves : Psittaciformes : Psittacidae	<i>Pyrrilia haematotis</i> & <i>P. coccinickollaris</i>	2.62	34, 84
Aves : Psittaciformes : Psittacidae	<i>Pyrrhura eisenmanni</i> & <i>Pyrrhura picta/emma</i>	0.64	34, 84
Aves : Psittaciformes : Psittacidae	<i>Pyrrhura hoffmanni</i> & <i>P. rhodocephala</i>	1.14	34, 84
Aves : Strigiformes : Strigidae	<i>Aegolius harrisii</i> & <i>A. ridgwayi/acadicus</i>	7.37	34
Aves : Trogoniformes : Trogonidae	<i>T. violaceus/T. curucui/T. surrucura</i>	3.1	85
Aves : Trogoniformes : Trogonidae	<i>Trogon viridis</i>	2.4	85
Aves : Trogoniformes : Trogonidae	<i>Trogon clathratus</i>	4.8	85
Aves : Trogoniformes : Trogonidae	<i>Trogon personatus</i>	8.9	85
Aves : Trogoniformes : Trogonidae	<i>Trogon collaris</i>	1.8	85

Aves : Trogoniformes : Trogonidae	<i>Pharomachrus antisianus/auriceps/fulgidus/pavoninus</i>	4.76	34, 85
Aves : Trogoniformes : Trogonidae	<i>Trogon rufus</i>	3.93	38
Aves: Piciformes : Capitonidae	<i>Capito squamatus &amp; C. maculicoronatus</i>	0.77	34, 39
Aves: Piciformes : Capitonidae	<i>Eubucco bourcierii</i>	0.04	34, 39
Dicot : Asterales : Boraginaceae	<i>Lithospermum</i>	3.75	86
Dicot : Asterales : Boraginaceae	<i>Heliotropium curassavicum</i>	23	87
Dicot : Asterales : Boraginaceae	<i>Heliotropium molle</i>	8	87
Dicot : Asterales : Lobelioideae	<i>Lobelia laxiflora</i>	7	88
Dicot : Brassicales : Caricaceae	South American clade	23	89
Dicot : Caryophyllales : Cactaceae	North American <i>Opuntia</i> clade	5.12	90
Dicot : Caryophyllales : Polygonaceae	<i>Ruprechtia</i> South Am. clade	1.2	91
Dicot : Chloranthales : Chloranthaceae	<i>Hedyosmum (H. costaricense - H. racemosum)</i>	19.4	92
Dicot : Cucurbitales : Cucurbitaceae	<i>Sicyos warmingii</i> to <i>Sicyos baderoa</i>	4.5	93
Dicot : Cucurbitales : Cucurbitaceae	<i>Sicyos polyacanthus</i>	3	93
Dicot : Cucurbitales : Cucurbitaceae	<i>Sicyos acarieanthus - S. quinquelobatus</i>	1.7	93
Dicot : Cucurbitales : Cucurbitaceae	<i>Sicyos malvifolius</i>	2.8	93
Dicot : Cucurbitales : Cucurbitaceae	<i>Sicyos longisetosus</i>	3.9	93
Dicot : Cucurbitales : Cucurbitaceae	<i>Sicyos andreanus</i>	12.3	93
Dicot : Cucurbitales : Cucurbitaceae	<i>Cyclanthera carthagenensis - C. australis</i>	9.4	93
Dicot : Dipsacales : Valerianaceae	<i>Valeriana</i>	23.75	94
Dicot : Fabales : Fabaceae : Faboideae	<i>Vicia montevidensis + V. stenophylla</i>	2.6	95
Dicot : Fabales : Fabaceae : Faboideae	<i>Vicia humilis + V. vicina</i>	2.65	95
Dicot : Fabales : Fabaceae : Faboideae	<i>Vicia gigantea</i>	0.75	95
Dicot : Fabales : Fabaceae : Faboideae	<i>Lathyrus pusillus</i>	2.5	95
Dicot : Fabales : Fabaceae : Faboideae	<i>Chaetocalyx</i> (South Am. clade)	5.9	96
Dicot : Fabales : Fabaceae : Faboideae	<i>Ateleia</i> ("RESI" clade)	4.24	97
Dicot : Fabales : Fabaceae : Faboideae	<i>Ateleia</i> ("GUAR" clade)	0.52	97
Dicot : Fabales : Fabaceae : Faboideae	<i>Lupinus</i> (Andean polytomy)	1.47	98

Dicot : Fabales : Fabaceae : Faboideae	<i>Phaseolus lunatus</i> Andean	1.04	99
Dicot : Fabales : Fabaceae : Faboideae	<i>Platymiscium pinnatum</i> ssp. <i>polystachyum</i>	3.9	100
Dicot : Fabales : Fabaceae : Faboideae	<i>Platymiscium yucatanum</i> - <i>P. trifoliatum</i>	5.9	100
Dicot : Gentianales : Apocynaceae	<i>Cynanchum</i>	18.6	101
Dicot : Gentianales : Rubiaceae	<i>Cinchona pubescens</i>	3	102
Dicot : Gentianales : Rubiaceae	<i>Iseria</i> ( <i>I. hypoleuca</i> , <i>I. haenkeana</i> , <i>I. laevis</i> )	14.8	102
Dicot : Lamiales : Gesneriaceae	Gloxineae widely distributed clade	6.5	103
Dicot : Lamiales : Gesneriaceae	Gloxineae Central American clade	17.5	103
Dicot : Lamiales : Gesneriaceae	Episcieae clade	11	103
Dicot : Lamiales : Lamiaceae	Clades B and C	12.65	104
Dicot : Lamiales : Lamiaceae	<i>Salvia polystachia</i> (node IV)	9.5	105
Dicot : Lamiales : Lamiaceae	Node III	11	105
Dicot : Lamiales : Lamiaceae	Node II	2.5	105
Dicot : Lamiales : Lamiaceae	Node III	6	105
Dicot : Laurales : Lauraceae	<i>Ocotea</i> complex ( <i>Endlicheria chalisea</i> - <i>Pleurothyrium insigne</i> )	18.5	106
Dicot : Magnoliales : Annonaceae	<i>Anaxagorea</i>	40	107
Dicot : Magnoliales : Annonaceae	<i>Crematosperma</i>	7.17	108
Dicot : Magnoliales : Annonaceae	<i>Guatteria</i> SAC clade ( <i>G. hyposericea</i> - <i>G. megalophylla</i> )	6.75	109
Dicot : Magnoliales : Annonaceae	<i>Klarobelia</i>	7.69	109
Dicot : Magnoliales : Annonaceae	<i>Malmea</i>	9.54	109
Dicot : Magnoliales : Annonaceae	<i>Mosannonna costaricensis</i>	17.5	110
Dicot : Malpighiales : Chrysobalanaceae	<i>Chrysobalanaceae</i>	47	111
Dicot : Malpighiales : Clusiaceae	<i>Symphonia</i> (trans-Andes clade)	5.03	112
Dicot : Mrytales : Melastomataceae	<i>Rhexia</i> / <i>Arthrostemma</i> divergence	23	113
Dicot : Piperales : Piperaceae	<i>Tildenia</i> (Central Am. clade)	20.6	114
Dicot : Ranunculales : Ranunculaceae	<i>Callianthemoides</i> - <i>Peltocalathos</i> (node 14)	8.67	115
Dicot : Rhamnales : Vitaceae	<i>Cissus gongylodes</i>	12	116
Dicot : Rhamnales : Vitaceae	<i>Cissus erosa</i> , <i>C. ulmifolia</i> , <i>C. obliqua</i>	19	116
Dicot : Rhamnales : Vitaceae	<i>Cissus amazonica</i> + <i>C. pseudoverticillata</i>	9.75	116
Dicot : Sapindales : Bursuraceae	<i>Protieae</i>	37	117
Dicot : Sapindales : Meliaceae	<i>Cedrela</i> I ( <i>C. fissilis</i> , <i>C. balansae</i> , <i>C. sp5</i> , <i>C. weberbaueri</i> , <i>C. sp2</i> )	15.7	118

Dicot : Sapindales : Meliaceae	<i>Cedrela II (C. angustifolia, C. montana)</i>	6.9	118
Dicot : Sapindales : Simaroubaceae	<i>Simarouba amara, S. versicolor</i>	9.3	119
Dicot : Sapindales : Simaroubaceae	<i>Castela erecta, C. refusa</i>	5	119
Dicot : Scrophulariales : Bignoniaceae	<i>Tanaecium crucigerum</i>	23	120
Dicot : Vitales : Vitaceae	Node 3	30.5	121
Fungi : Lecanorales : Parmeliaceae	<i>Xanthoparmelia</i> SA clade	3.44	122
Insecta : Coleoptera : Chrysomelidae	<i>Stator limbatus</i>	2.4	123
Insecta : Diptera: Culicidae	<i>Anopheles albimanus</i>	0.85	124
Insecta : Hymenoptera : Apidae	<i>Coccineobombus - Rubicundobombus</i>	7.2	125
Insecta : Hymenoptera : Apidae	<i>Funebribombus funebris</i>	7	125
Insecta : Hymenoptera : Apidae	<i>Robustobombus</i>	3.1	125
Insecta : Hymenoptera : Apidae	<i>Robustobombus volucelloides</i>	0.6	125
Insecta : Hymenoptera : Apidae	<i>Fervidobombus opifex, F. bellicosus</i>	1.5	125
Insecta : Hymenoptera : Apidae	<i>Fervidobombus atratus, F. transversalis, F. brasiliensis</i>	2	125
Insecta : Hymenoptera : Apidae	<i>Euglossa heterosticta - E. dissimula</i>	5	126
Insecta : Hymenoptera : Apidae	<i>Eulaema</i>	13	126
Insecta : Hymenoptera : Apidae	<i>Melikerria beecheii</i>	3.5	127
Insecta : Hymenoptera : Apidae	<i>Michmelia fasciata</i>	5.5	127
Insecta : Hymenoptera : Apidae	<i>Nannotrigona perilampoides</i>	4	127
Insecta : Hymenoptera : Apidae	<i>Noguierapis mirandula</i>	20.5	127
Insecta : Hymenoptera : Apidae	<i>Oxytrigona mediorufa + O. mellicolor</i>	4.5	127
Insecta : Hymenoptera : Apidae	<i>Partamona musarum + P. orizabaensis polytomy</i>	4	127
Insecta : Hymenoptera : Apidae	<i>Plebeia frontalis</i>	13	127
Insecta : Hymenoptera : Apidae	<i>Scaptotrigona meciana + S. pectoralis</i>	2	127
Insecta : Hymenoptera : Apidae	<i>Tetragona perangulata + T. zieglerei</i>	16	127
Insecta : Hymenoptera : Apidae	<i>Trigona fulviventrtris</i>	5	127
Insecta : Hymenoptera : Apidae	<i>Trigona fuscipennis</i>	3	127
Insecta : Hymenoptera : Apidae	<i>Trigona nigerrima, T. corvina</i>	8	127
Insecta : Hymenoptera :	<i>Trigonisca buyssoni</i>	9	127

Apidae			
Insecta : Hymenoptera : Apidae	<i>Battus</i>	21.66	128
Insecta : Hymenoptera : Apidae	<i>Euryades &amp; Parides</i>	27.45	128
Insecta : Lepidoptera : Nymphaelidae	Limenitis (Nearctic/Palaearctic)	9.95	127
Insecta : Lepidoptera : Nymphaelidae	clade of <i>Ithomia patilla</i> - <i>I. heraldica</i>	5.29	129
Insecta : Lepidoptera : Nymphaelidae	<i>Ithomia xenos</i>	5.48	130
Insecta : Lepidoptera : Nymphaelidae	<i>Ithomia terra</i>	4.48	130
Insecta : Lepidoptera : Nymphaelidae	<i>Napeogenes peridia</i>	5.65	130
Insecta : Lepidoptera : Nymphaelidae	<i>Napeogenes tolosa</i>	6.95	130
Insecta : Lepidoptera : Nymphaelidae	<i>Napeogenes cranto</i>	7.33	130
Insecta: Orthoptera: Acrididae	<i>Trimerotropis pallidipennis</i> NA/ <i>T. saxatilis</i> vs <i>T. ochraceipennis</i> / <i>T. pallidipennis</i> SA	0.7	129
Mammalia : Artiodactyla : Cervidae	Cervidae (South American clade)	4.15	130
Mammalia : Carnivora : Canidae	<i>South American endemic clade</i>	4.25	131
Mammalia : Carnivora : Felidae	<i>Leopardus tigrinus</i>	3.7	132
Mammalia : Carnivora : Mustelidae	<i>Lontra longicaudis</i> , <i>L. felina</i>	1.5	133
Mammalia : Carnivora : Mustelidae	<i>Pteronura brasiliensis</i> vs. <i>Clade at Node 8</i>	7.7	133
Mammalia : Carnivora : Procyonidae	<i>Nasua narica</i> vs. <i>N. nasua</i>	7.5	134
Mammalia : Carnivora : Procyonidae	<i>Procyon cancrivorus</i> vs. <i>P. lotor</i>	5.35	134
Mammalia : Carnivora: Ursidae	<i>Tremarctos ornatus</i> vs. <i>Arctodus simus</i>	5.66	135
Mammalia : Chiroptera : Phyllostomidae	<i>Desmodus rotundus</i>	1	136
Mammalia : Chiroptera : Emballonuroidea	<i>Centronycteris centralis</i>	5.5	137
Mammalia : Chiroptera : Emballonuroidea	<i>Cormura brevirostris</i>	19.4	137
Mammalia : Chiroptera : Emballonuroidea	<i>Cyttarops alecto</i>	14.6	137
Mammalia : Chiroptera : Emballonuroidea	<i>Diclidurus albus</i>	2	137
Mammalia : Chiroptera : Emballonuroidea	<i>Peropteryx kappleri</i>	1	137
Mammalia : Chiroptera : Emballonuroidea	<i>Rhynchonycteris naso</i>	19	137
Mammalia : Chiroptera : Emballonuroidea	<i>Saccopteryx bilineata</i> , <i>S. leptura</i>	5.1	137
Mammalia : Chiroptera : Vespertilionidae	<i>Myotis ruber</i> , <i>M. riparius</i> + ( <i>M. keaysi</i> )	2.8	138



Mammalia : Primates : Atelidae	<i>Alouatta pigra, A. palliata</i>	3	139
Mammalia : Primates : Atelidae	<i>Ateles geoffroyi</i> (- subsp. <i>robsutus</i> )	1.29	140
Mammalia : Primates : Cebidae	<i>Cebus</i> (Coastal and Andes Venezuelan) vs. <i>Cebus</i> (Colombian Andes and Central American)	1.46	141
Mammalia : Primates : Cebidae	<i>Saimiri oerstedii</i> vs. <i>S. s. sciureus</i>	0.77	142
Mammalia : Rodentia : Cricetidae	<i>Sigmodontinae</i>	12	143
Mammalia : Rodentia : Sciuridae	<i>Sciurus</i> South American Clade	2.8	144
Monocot : Alismatales : Hydrocharitaceae	<i>Elodea</i>	2.5	145
Monocot : Arecales : Arecaceae	<i>Astrocaryum</i> ( <i>A. murumuru</i> - <i>A. paramaca</i> )	22	146
Monocot : Arecales : Arecaceae	<i>Astrocaryum standleyanum</i>	6	147
Monocot : Arecales : Arecaceae	<i>Chamaedorea</i> ( <i>C. linearis</i> , <i>C. fragrans</i> )	9	148
Monocot : Arecales : Arecaceae	<i>Copernicia</i> crown clade	16.34	149
Monocot : Arecales : Arecaceae	<i>Geonoma</i> ( <i>G. hugonis</i> , <i>G. epetiolata</i> , <i>G. monospatha</i> , <i>G. longevaginata</i> , <i>G. scoparia</i> , <i>G. poeppigiana</i> , <i>G. undata</i> , <i>G. jussieuana</i> , <i>G. orbignyana</i> , <i>G. weberbaueri</i> )	8.8	150
Monocot : Arecales : Arecaceae	Phytelephea ( <i>Phytelephus schottii</i> , <i>P. seemanii</i> , <i>P. macrocarpa</i> , <i>P. tenuicaulis</i> , <i>P. aequatorialis</i> , <i>P. turnacana</i> , <i>Amandra decasperma</i> , <i>Aphandra natalia</i> )	17	151
Monocot : Poales : Bromeliaceae	<i>Fosterella micrantha</i>	0.6	152
Monocot : Poales : Bromeliaceae	<i>Pitcairnia wendlandii</i> + <i>P. orchidifolia</i>	4.5	153
Monocot : Poales : Bromeliaceae	<i>Tillandsia utriculata</i> - <i>T. viridiflora</i> polytomy	6	153
Monocot : Poales : Cyperaceae	<i>Oreobolus</i> South American clade	3.7	154
Monocot : Poales : Poaceae	<i>Festuca calligera</i> + ( <i>F. anicola</i> - <i>H. fragilis</i> )	3.1	155
Monocot : Poales : Poaceae	<i>Festuca purpurascens</i> + <i>F. subantarctica</i>	2	155
Pteridophytes : Polypodiales : Dryopteridaceae	SA members of clade IV	9	156
Reptilia : Squamata : Boidae	<i>Corallus annulatus</i>	10.1	157
Reptilia : Squamata : Boidae	<i>Corallus ruschenbergerii</i>	1.5	157
Reptilia : Squamata : Colubridae	<i>Leptodeira</i>	3.91	158
Reptilia : Squamata : Viperidae	<i>Atropoides</i> + <i>Cerrophidion</i>	10	159
Reptilia : Squamata : Viperidae	<i>Crotalus durissus</i>	1.54	160
Reptilia : Squamata : Viperidae	<i>Bothrops diporus</i> - <i>B. taeniatus</i>	12.5	161
Reptilia : Squamata : Viperidae	<i>Porthidium</i> (node 1)	2.5	161
Reptilia : Squamata : Viperidae	<i>Lachesis muta</i>	7.25	161

Reptilia : Squamata : Viperidae	<i>Lachesis stenophrys</i>	7.5	162
Reptilia : Testudines : Emydidae	<i>Trachemys callirostris</i>	0.98	163
Reptilia : Testudines : Emydidae	<i>Trachemys adiutrix</i> vs. <i>T. dorbigni</i>	2.28	163
Reptilia : Testudines : Geoemydidae	<i>Rhinoclemmys nasuta</i> vs <i>R. areolata</i> / <i>R. diademata</i> / <i>R. punctularia</i> / <i>R. funerea</i> / <i>R. melanosterna</i> / <i>R. annulata</i>	20.1	164
Reptilia : Testudines : Geoemydidae	<i>Rhinoclemmys diademata</i> , <i>R. punctularia</i> , <i>R. funerea</i> , <i>P. melanosterna</i> (node 6)	5.3	164
Reptilia : Testudines : Geoemydidae	<i>Rhinoclemmys annulata</i> vs. <i>R. pulcherrima</i>	15.3	164
Reptilia: Crocodylia: Alligatoridae	<i>Caiman crocodilus</i> (Central American Crown Age)	2.7	165
Reptilia:Squamata: Phyllodactylidae	<i>Phyllodactylus reissii</i> vs North American <i>Phyllodactylus</i> clade	23	166
Reptilia:Squamata: Sphaerodactylidae	<i>Gonatodes albogularis</i> vs. <i>Gonatodes vittatus</i>	13	167

## References

1. McMahan, C.D., Davis, M.P., Dominguez-Dominguez, O., Garcia-de-Leon, F.J., Doadrio, I., Piller, K.R. 2012. From the mountains to the sea: phylogeography and cryptic diversity within the mountain mullet, *Agonostomus monticola* (Teleostei: Mugilidae). *Journal of Biogeography* 40: 894-904.
2. Cárdenas L., Hernández C.E., Poulin E., Magoulas A., Kornfield I., Ojeda F.P. 2005. Origin, diversification, and historical biogeography of the genus *Trachurus* (Perciformis: Carangidae). *Molecular Phylogenetics and Evolution* 35: 496-507.
3. Bellwood D.R., Klanten S., Cowman P.F., Pratchett M.S., Konow N., van Herwerden L. 2010. Evolutionary history of the butterfly fishes (f: Chaetodonidae) and the rise of coral feeding fishes. *Journal of Evolutionary Biology* 23: 335-349.
4. Matamoros, W.A., McMahan, C.D., Chakrabarty, P., Albert, J.S., Schaefer, J.F. 2014. Derivation of the freshwater fish fauna of Central America revisited: Myer's hypothesis in the twenty-first century. *Cladistics* 31: 177-188.
5. Lopez-Fernandez, H., Arbour, J.H., Winemiller, K.O., Honeycutt, R.L. 2013. Testing for ancient adaptive radiations in Neotropical cichlid fishes. *Evolution* 67: 1321-1337.
6. Rican, O., Pialek, L., Zardoya, R., Doadrio, I., Zrzavy, J. 2012. Biogeography of the Mesoamerican Cichlidae (Teleostei: Heroini): colonization through the GAARlandia land bridge and early diversification. *Journal of Biogeography* 40: 579-593.
7. Lessios, H.A. 2008. The Great American Schism: Divergence of marine organisms after the rise of the Central American Isthmus. *Annual Review of Ecology, Evolution, and Systematics* 39: 63-91.
8. Alda, F., Reina, R.G., Doadrio, I., Bermingham, E. 2013. Phylogeny and biogeography of the *Poecilia sphenops* complex (Actinopterygii, Poeciliidae) in Central America. *Molecular Phylogenetics and Evolution* 66: 1011-1026.
9. Bellwood, D.R., van Herwerden, L., Konow, N. 2004. Evolution and biogeography of marine angelfishes (Pisces: Pomacanthidae). *Molecular Phylogenetics and Evolution* 33: 140-155.

10. Martin, A.P., Bermingham, E. 2000. Regional endemism and cryptic species revealed by molecular and morphological analysis of a widespread species of Neotropical catfish. *Proceedings of the Royal Society B* 267: 1135-1141.
11. Perdices, A., Bermingham, E., Montilla, A., Doadrio, I. 2002. Evolutionary history of the genus *Rhamdia* (Teleostei: Pimelodidae) in Central America 25: 172-189.
12. Marko P.B., Moran A.L. 2009. Out of sight, out of mind: high cryptic diversity obscures the identities and histories of geminate species in the marine bivalve *Acar*. *Journal of Biogeography* 36: 1861-1880.
13. Marko P.B. 2002. Fossil calibration of molecular clocks and the divergence times of geminate species pairs separated by the Isthmus of Panama. *Molecular Biology and Evolution* 19: 2005-2021.
14. Stelbrink B., von Rintelen T., Cliff G., Kriwet J. 2010. Molecular systematics and global phylogeography of angel sharks (genus *Squatina*). *Molecular Phylogenetics and Evolution* 54: 395-404.
15. Reid D.G., Dyal P., Williams S.T. 2010. Global diversification of mangrove fauna: a molecular phylogeny of *Littoraria* (Gastropoda: Littorinidae). *Molecular Phylogenetics and Evolution* 55: 185-201.
16. Williams S.T., Reid D.G. 2004. Speciation and diversity on tropical rocky shores: A global phylogeny of snails of the genus *Echinolittorina*. *Evolution* 58: 2227-2251.
17. Frey M.A., Vermeij G.J. 2008. Molecular phylogenies and historical biogeography of a circumtropical group of gastropods (Genus: *Nerita*): Implications for regional diversity patterns in the marine tropical. *Molecular Phylogenetics and Evolution* 48: 1067-1086.
18. Malaquias M.A.E., Reid D.G. 2009. Tethyan vicariance, relictualism and speciation: evidence from a global molecular phylogeny of the opisthobranch genus *Bulla*. *Journal of Biogeography* 36: 1760-1777.
19. Duda T.F. Jr., Kohn K.J. 2005. Species-level phylogeography and evolutionary history of the hyperdiverse marine gastropod genus *Conus*. *Molecular Phylogenetics and Evolution* 34: 257-272.
20. Claremont M., Williams S.T., Barraclough T.G., Reid D.G. 2011. The geographic scale of speciation in a marine snail with high dispersal potential. *Journal of Biogeography* 38: 1016-1032.
21. Miura O., Torchin M.E., Bermingham E. 2010. Molecular phylogenetics reveals differential divergence of coastal snails separated by the Isthmus of Panama. *Molecular Phylogenetics and Evolution* 56: 40-48.
22. Martinez, D.E., Iniguez, A.R., Percell, K.M., Willner, J.B., Signorovitch, J., Campbell, R.D. 2010. Phylogeny and biogeography of *Hydra* (Cnidaria: Hydridae) using mitochondrial and nuclear DNA sequences. *Molecular Phylogenetics and Evolution* 57: 403-410.
23. Fulton, T.L., Strobeck, C. 2010. Multiple markers and multiple individuals refine true seal phylogeny and bring molecules and morphology back in line. *Proceedings of the Royal Society B* 277: 1065-1070.
24. Wang I.J., Crawford A.J., Bermingham E. 2008. Phylogeography of the Pygmy Rain Frog (*Pristimantis ridens*) across the lowland wet forests of isthmian Central America. *Molecular Phylogenetics and Evolution* 47: 992-1004.

25. Maciel N.M., Collevatti R.G., Colli G.R., Schwartz E.F. 2010. Late Miocene diversification and phylogenetic relationships of the huge toads in the *Rhinella marina* (Linnaeus, 1758) species group (Anura: Bufonidae). *Molecular Phylogenetics and Evolution* 57: 787-797.
26. Santos J.C., Coloma L.A., Summers K., Caldwell J.P., Ree R., Cannatella D.C. 2009. Amazonian amphibian diversity is primarily derived from Late Miocene Andean lineages. *PLoS Biology*: e1000056.
27. Heinicke M.P., Duellman W.E., Hedges S.B. 2007. Major Caribbean and Central American frog faunas originated by ancient oceanic dispersal. *Proceedings of the National Academy of Science USA* 104: 10092-10097.
28. Weight L.A., Crawford A.J., Rand A.S., Ryan M.J. 2005. Biogeography of the túngara frog, *Physalaemus pustulosus*: a molecular perspective. *Molecular Ecology* 14: 3857-3876.
29. O de Sa, Streicher, J.W., Sekonyela, R., Forlani, M.C., Loader, S.P., Greenbaum, E., Richards, S., Haddad, C.F. 2012. Molecular phylogeny of microhylid frigs (Anura: Microhylidae) with emphasis on relationships among New World genera. *BMC Evolutionary Biology* 12: e241.
30. Elmer, K.R., Nonett, R.M., Wake, D.B., Lougheed, S.C. 2013. Early Miocene origin and cryptic diversification of South American salamanders. *BMC Evolutionary Biology* 13: e59.
31. Wiens J.J., Parra-Olea G., García-París M., Wake D.B. 2007. Phylogenetic history underlies elevational biodiversity patterns in tropical salamanders. *Proceedings of the Royal Society of London B* 274: 919-928.
32. Binford G.J., Callahan M.S., Bodner M.R., Rynerson M.R., Núñez P.B., Ellison C.E., Duncan R.P. 2008. Phylogenetic relationships of *Loxosceles* and *Sicarius* spiders are consistent with Western Gondwana vicariance. *Molecular Phylogenetics and Evolution* 49: 538-553.
33. Zeh J.A., Zeh D.W., Bonilla M.M. 2003. Phylogeography of the harlequin beetle-riding pseudoscorpion and the rise of the Isthmus of Panamá. *Molecular Ecology* 12: 2759-2769.
34. Smith, B.T., Klicka, J. 2010. The profound influence of the Late Pliocene Panamanian uplift on the exchange, diversification, and distribution of New World birds. *Ecography* 33: 333-342.
35. Johnson, K.J., Sorenson, M.D. 1998. Comparing molecular evolution in two mitochondrial protein coding genes (Cytochrome b and ND2) in the dabbling ducks (Tribe: Anatini). *Molecular Phylogenetics and Evolution* 10: 82-94.
36. McGuire, J.A., Witt, C.C., Altshuler, D.L., Remsen, J.V. Jr. 2007. Phylogenetic systematics and biogeography of hummingbirds: Bayesian and maximum-likelihood analysis of partitioned data and selection of an appropriate partitioning strategy. *Systematic Biology* 56: 837-856.
37. Johnson, K.P., Weckstein, J.D. 2011. The Central American land bridge as an engine of diversification in New World doves. *Journal of Biogeography* 38: 1069-1076.
38. Smith, B.T., Amei, A., Klicka, J. 2012. Evaluating the role of contracting and expanding rainforest in initiating cycles of speciation across the Isthmus of Panama. *Proceedings of the Royal Society B* 279: 3520-3526.
39. Smith, B.T., McCormack, J.E., Cuervo, A.M., Hickerson, M.J., Zleixo, A., Cadena, C.D., Perez-Eman, J., Burney, C.W., Xie, X., Harvey, M.G., Faircloth, B.C., Glenn, T.C.,

- Derryberry, E.P., Prejean, J., Fields, S., Brumfield, R.T. 2014. The drivers of tropical speciation. *Nature* 515: 406-409.
40. Fuchs, J., Chen, S., Johnson, J.A., Mindell, D.P. 2011. Pliocene diversification within the South American Forest Falcons (Falconidae: *Micrastur*). *Molecular Phylogenetics and Evolution* 60: 398-407.
  41. Pereira, S.L., Baker, A.J. 2004. Low number of mitochondrial pseudogenes in the chicken (*Gallus gallus*) nuclear genome: implications for molecular inference of population history and phylogenetics. *BMC Evolutionary Biology* 4: e17.
  42. Burns, K.J. 1998. Molecular phylogenetics of the genus *Piranga*: implications for biogeography and evolution of morphology and behavior. *The Auk* 115: 621-634.
  43. Bonaccorso, E., Peterson, A.T. 2007. A multilocus phylogeny of New World Jay genera. *Molecular Phylogenetics and Evolution* 42: 467-476.
  44. Bonaccorso, E. 2009. Historical biogeography and speciation in the Neotropical highlands: molecular phylogenetics of the Jay genus *Cyanolyca*. *Molecular Phylogenetics and Evolution* 50: 618-632.
  45. Navarro-Siguenza, A.G., Townsend Peterson, A., Nyari, A., Garcia-Deras, G.M., Garcia-Moreno, J. 2008. Phylogenography of the *Buarremon* brush-finch complex (Aves, Emberizidae) in Mesoamerica. *Molecular Phylogenetics and Evolution* 47: 21-35.
  46. Weir, J.T., Bermingham, E., Miller, M.J., Klicka, J., Gonzalez, M.A. 2008. Phylogeography of a morphologically diverse Neotropical montane species, the Common Bush Tanager (*Chlorospingus ophthalmicus*). *Molecular Phylogenetics and Evolution* 47: 650-664.
  47. Cadena, C.D. 2007. Testing the role of interspecific competition in the evolutionary origin of elevational zonation: an example with *Buarremon* brush-finches (Aves, Emberizidae) in the Neotropical mountains. *Evolution* 61: 120-1136.
  48. Arnaiz-Villena, A., Alvarez-Tejado, M., Ruz-del-Valle, V., Garcia-de-la-Torre, C., Varela, P., Recio, M.J., Ferre, S., Martinez-Laso, J. 1999. Rapid radiation of Canaries (Genus *Serinus*). *Molecular Biology and Evolution* 16: 2-11.
  49. Aleixo, A. 2002. Molecular systematics and the role of the “Varzea”-“Terre-firme” ecotone in the diversification of *Xiphorhynchus* woodcreepers (Aves: Dendrocolaptidae). *The Auk* 119: 621-640.
  50. Sheldon, F.H., Whittingham, L.A., Moyle, R.G., Slikas, B., Winkler, D.W. 2005. Phylogeny of swallows (Aves: Hirundinidae) estimated from nuclear and mitochondrial DNA sequences. *Molecular Phylogenetics and Evolution* 35: 254-270.
  51. Omland, K., Lanyon, S., Fritz, S. 1999. A molecular phylogeny of the New World Orioles (*Icterus*): the importance of dense taxon sampling. *Molecular Phylogenetics and Evolution* 12: 224-239.
  52. Lanyon, S.M., Omland, K.E. 1999. A molecular phylogeny of the blackbirds (Icteridae): five lineages revealed by cytochrome-b sequence data. *The Auk* 116: 629-639.
  53. Price, J.J., Lanyon, S.M. 2002. A robust molecular phylogeny of the oropendolas: polyphyly revealed by mitochondrial sequence data. *The Auk* 119: 335-348.
  54. Voelker, G. 1999. Dispersal, vicariance, and clocks: Historical biogeography and speciation in a Cosmopolitan passerine genus (*Anthus*: Motacillidae). *Evolution* 53: 1536-1552.

55. Gutierrez-Pinto, N., Cuervo, A.M., Miranda, J., Perez-Eman, J.L., Brumfield, R.T., Cadena, C.D. 2012. Non-monophyly and deep genetic differentiation across low-elevation barriers in a Neotropical montane bird (*Basileuterus tristriatus*; Aves: Parulidae). *Molecular Phylogenetics and Evolution* 64: 156-165.
56. Perez-Eman, J.L. 2005. Molecular phylogenetics and biogeography of the Neotropical redstarts (*Myoborus*; Aves, Parulinae). *Molecular Phylogenetics and Evolution* 37: 511-528.
57. Escalante, P., Marquez-Valdelamar, L., de la Torre, P., Laclette, J.P., Klicka, J. 2009. Evolutionary history of a prominent North American warbler clade: the *Oporornis-Geothlypis* complex. *Molecular Phylogenetics and Evolution* 53: 668-678.
58. Lovette, I.J. 2004. Molecular phylogeny and plumage signal evolution in a trans-Andean and circum-Amazonian avian species complex. *Molecular Phylogenetics and Evolution* 32: 512-523.
59. Cheviron, Z.A., Hackett, S.J., Capparella, A.P. 2005. Complex evolutionary history of a Neotropical lowland forest bird (*Lepidothrix coronate*) and its implications for historical hypotheses of the origin of Neotropical avian diversity. *Molecular Phylogenetics and Evolution* 36: 338-357.
60. Brumfield, R.T., Edwards, S.V. 2007. Evolution into and out of the Andes: A Bayesian analysis of historical diversification in *Thamnophilus* antshrikes. *Evolution* 61: 346-367.
61. Mauck III, W.M., Burns K.J. 2009. Phylogeny, biogeography, and recurrent evolution of divergent bill types in the nectar-stealing flowerpiercers (Thraupini: *Diglossa* and *Diglossopsis*). *Biological Journal of the Linnean Society* 98: 14-28.
62. Burns K.J., Racicot R.A. 2009. Molecular phylogenetics of a clade of lowland Tanagers: implications for avian participation in the Great American Interchange. *The Auk* 126: 635-648.
63. Burns, K.J. 1997. Molecular systematics of tanagers (Thraupinae): evolution and biogeography of a diverse radiation of Neotropical birds. *Molecular Phylogenetics and Evolution* 8: 334-348.
64. Weir, J.T., Schluter, D. 2007. The latitudinal gradient in recent speciation and extinction rates of birds and mammals. *Science* 315: 1574-1576.
65. Burns, K.J., Naoki, K. 2004. Molecular phylogenetics and biogeography of Neotropical tanagers in the genus *Tangara*. *Molecular Phylogenetics and Evolution* 32: 838-854.
66. Whittingham, L.A., Slikas, B., Winkler, D.W., Sheldon F.H. 2002. Phylogeny of the tree swallow genus, *Tachycineta* (Aves: Hirundinidae), by Bayesian analysis of Mitochondrial DNA sequences. *Molecular Phylogenetics and Evolution* 22: 430-441.
67. Nyari, A.S. 2007. Phylogeographic patterns, molecular and vocal differentiation, and species limits in *Schiffornis turdina* (Aves). *Molecular Phylogenetics and Evolution* 44: 154-164.
68. Mann, N.I., Dingess, K.A., Slater, P.J.B. 2005. Antiphonal four-part synchronized chorusing in a Neotropical wren. *Biology Letters* 2: 1-4.
69. Miller, M., Bermingham, E., Ricklefs, R.E. 2007. Historical biogeography of the New World Solitaires *Myadestes* (*Turdidae*). *The Auk* 124: 868-885.

70. Voelker, G., Rohwer, R., Bowie, C.K., Outlaw, D.C. 2007. Molecular systematics of a speciose, cosmopolitan songbird genus: Defining the limits of, and relationships amongs, the *Turdus* thrushes. *Molecular Phylogenetics and Evolution* 42: 422-434.
71. Klicka, J., Voelker, G., Spellman, G. 2005. A molecular phylogenetic analysis of the “true thrushes” (Aves: Turdinae). *Molecular Phylogenetics and Evolution* 34 486-500.
72. Miller, M.J., Bermingham E., Klicka J., Escalante P. Raposo do Amaral F., Weir J.T., Winker K. 2008. Out of Amazonia again and again: episodic crossing of the Andes promotes diversification in a lowland forest flycatcher. *Proceedings of the Royal Society of London B* 275: 1133-1142.
73. Rheindt F.E., Christidis L., Norman J.A. 2008. Habitat shifts in the evolutionary history of a Neotropical flycatcher lineage from forest and open landscapes. *BMC Evolutionary Biology* 8: e1193.
74. Joseph, L. Wilke, T., Bermingham, E., Alpers, D., Ricklefs, R.E. 2004. Towards a phylogenetic framework for the evolution of shakes, rattles, and rolls in *Myiarchus* tyrant-flycatchers (Aves: Passiformes: Tyrannidae). *Molecular Phylogenetics and Evolution* 31: 139-152.
75. Moore W.S., Weibel A.C., Agius A. 2006. Mitochondrial DNA phylogeny of the woodpecker genus *Veniliornis* (Picidae, Picinae) and related genera implies convergent evolution of plumage patterns. *Biological Journal of the Linnean Society* 87: 311-624.
76. Moore W.S., Overton L.C., Miglia K.J. 2011. Mitochondrial DNA based phylogeny of the woodpecker genera *Colaptes* and *Piculus*, and implications for the history of woodpecker diversification in South America. *Molecular Phylogenetics and Evolution* 58: 76-84.
77. Puebla-Olivares, F.E., Bonaccorso, F., de los Monteros, A.E., Omland, K.E., Llorente-Bousquets, J.E., Peterson A.T., Navarro-Siquenza, A.G. 2008. Speciation in the Emerald Toucanet (*Aulacorhynchus prasinus*) complex. *The Auk* 125: 39-50.
78. Weckstein, J.D. 2004. Biogeography explains cophylogenetic patterns in toucan chewing lice. *Systematic Biology* 53: 154-164.
79. Eberhard, J.R., Bermingham, E. 2005. Phylogeny and comparative biogeography of *Pionopsitta* parrots and *Pteroglossus* toucans. *Molecular Phylogenetics and Evolution* 36: 288-304.
80. Patel, S., Weckstein, J.D., Patane, J.S., Bates, J.M., Aleixo, A. 2011. Temporal and spatial diversification of *Pteroglossus aracarís* (Aves: Ramphastidae) in the neotropics: constant rate of diversification does not support an increase in radiation during the Pleistocene. *Molecular Phylogenetics and Evolution* 58: 105-115.
81. Ribas, C.C., Miyaki, C.Y., Cracraft, J. 2009. Phylogenetic relationships, diversification and biogeography in Neotropical *Brotogeris* parakeets. *Journal of Biogeography* 36: 1712-1729.
82. Russello, M.A., Amato, G. 2004. A molecular phylogeny of *Amazona*: implications for Neotropical parrot biogeography, taxonomy, and conservation. *Molecular Phylogenetics and Evolution* 30: 421-437.
83. Eberhard, J.R., Bermingham, E. 2004. Phylogeny and biogeography of the *Amazona ochrocephala* (Aves: Psittacidae) complex. *The Auk* 121: 318-332.

84. Ribas, C.C., Josphe, L., Miyaki, C.Y. 2006. Molecular systematics and patterns of diversification in *Pyrrhura* (Psittacidae), with special reference to the *Picta-Leucotis* complex. *The Auk* 123: 660-680.
85. Dacosta J.M., Klicka J. 2008. The Great American Interchange in birds: a phylogenetic perspective with the genus *Trogon*. *Molecular Ecology* 17: 1328-1343.
86. Weigend M., Gottshling M., Selvi F., Hilger H.H. 2009. Marbleseed are gromwells- Systematics and evolution of *Lithospermum* and allies (Boraginaceae tribe Lithospermeae) based on molecular and morphological data. *Molecular Phylogenetics and Evolution* 52: 755-768.
87. Luebert F., Hilger H.H., Weigend M. 2011. Diversification in the Andes: Age and origins of South American *Heliotropium* lineages (Heliotropiaceae, Boraginales). *Molecular Phylogenetics and Evolution* 61: 90-102.
88. Antonelli A. 2009. Have giant lobelias evolved several times independently? Life form shifts and historical biogeography of the cosmopolitan and highly diverse subfamily Lobelioideae (Campanulaceae). *BMC Biology* 7: e82.
89. Antunes Carvalho, F., Renner, S.S. 2012. A dated phylogeny of the papaya family (Caricaceae) reveals the crop's closest relatives and the family's biogeographic history. *Molecular Phylogenetics and Evolution* 65: 46-53.
90. Majure, L.C., Puente, R., Griffith, M.P., Judd, W.S., Soltis, P.S., Soltis, D.E. 2012. Phylogeny of *Opuntia* s.s. (Cactaceae): clade delineation, geographic origins, and reticulate evolution. *American Journal of Botany* 99: 847-864.
91. Pennington R.T., Lavin M., Prado D.E., Pendry C.A., Pell S.K., Butterworth C.A. 2004. Historical climate change and speciation: neotropical seasonally dry forest plants show patterns of both Tertiary and Quaternary diversification. *Proceedings of the Royal Society of London B* 359: 515-538.
92. Antontelli A., Sanmartín I. 2011. Mass extinction, gradual cooling, or rapid radiation? Reconstructing the spatiotemporal evolution of the ancient angiosperm genus *Hedyosmum* (Chloranthaceae) using empirical and simulated approaches. *Systematic Biology* 60: 596-615.
93. Sebastian, P. Schaefer, H., Lira, R., Telford, I.R.H., Renner, S.S. 2012. Radiation following long-distance dispersal: the contributions of time, opportunity and diaspora morphology in *Sicyos* (Cucurbitaceae). *Journal of Biogeography* 39: 1427-1438.
94. Bell C.D., Donoghue M.J. 2005. Phylogeny and biogeography of Valerianaceae (Dipsacales) with special reference to the South American valerians. *Organisms, Diversity & Evolution* 5: 147-159.
95. Schaefer, H., Hechenleitner, P., Santos-Guerra, A., Menezes de Sequeira, M., Pennington, R.T., Kenicer, G., Carine, M.A. 2012. Systematics, biogeography, and character evolution of the legume tribe Fabeae with special focus on the middle-Atlantic island lineages. *BMC Evolutionary Biology* 12: e250.
96. Pennington, R.T., Lavin, M., Prado, D.E., Pendry, C.A., Pell, S.K., Butterworth, C.A. 2004. Historical climate change and speciation: neotropical seasonally dry forest plants show patterns of both Tertiary and Quaternary diversification. *Philosophical Transactions of the Royal Society B* 359: 515-538.



97. Ireland H.E., Kite G.C., Veitch N.C., Chase M.W., Schrire B., Lavin M., Linares J., Pennington R.T. 2010. Biogeographical, ecological and morphological structure in a phylogenetic analysis of *Ateleia* (Swartzieae, Fabaceae) derived from combined molecular, morphological and chemical data. *Botanical Journal of the Linnean Society* 162: 39-53.
98. Hughes C., Eastwood R. 2006. Island radiation on a continental scale: Exceptional rates of plant diversification after uplift of the Andes. *Proceedings of the National Academy of Science USA* 103: 10334-10339.
99. Serrano-Serrano M.L., Hernández-Torres J., Castillo-Villamizar G., Debouck D.G., Chacón Sánchez M.I. 2010. Gene pools in wild Lima bean (*Phaseolus lunatus* L.) from the Americas: Evidences for an Andean origin and past migrations. *Molecular Phylogenetics and Evolution* 54: 76-87.
100. Saslis-Lagoudakis C., Chase M.W., Robinson D.N., Russell S.J., Klitgaard B.B. 2008. Phylogenetics of neotropical *Platymiscium* (Leguminosae: Dalbergieae): systematics, divergence times, and biogeography inferred from nuclear ribosomal and plastid DNA sequence data. *American Journal of Botany* 95: 1270-1286.
101. Rapini A., van den Berg C., Liede-Schumann S. 2007. Diversification of Asclepiadoideae (Apocynaceae) in the New World. *Annals of the Missouri Botanical Garden* 94: 407-422.
102. Antonelli A., Nylander J.A.A., Persson C., Sanmartín I. 2009. Tracing the impact of the Andean uplift on Neotropical plant evolution. *Proceedings of the National Academy of Science USA* 106: 9749-9754.
103. Perret, M., Chautems, A., Onofre de Araujo, A., Salamin, N. 2013. Temporal and spatial origin of Gesneriaceae in the New World inferred from plastid DNA sequences. *Botanical Journal of the Linnean Society* 171: 61-70.
104. Drew, B.T., Sytsma, K.J. 2013. The South American radiation of *Lepechinia* (Lamiaceae): phylogenetics, divergence times and evolution of dioecy. *Botanical Journal of the Linnean Society* 171: 171-190.
105. Drew, B.T., Sytsma, K.J. 2012. Phylogenetics, biogeography, and stamina evolution in the tribe Mentheae (Lamiaceae). *American Journal of Botany* 99: 933-953.
106. Chanderbali A.S., van der Werff H., Renner S.S. 2001. Phylogeny and historical biogeography of Lauraceae: Evidence from the chloroplast and nuclear genomes. *Annals of the Missouri Botanical Garden* 88: 104-134.
107. Scharaschkin T., Doyle J.A. 2005. Phylogeny and historical biogeography of *Anaxagorea* (Annonaceae) using morphology and non-coding chloroplast sequence data. *Systematic Botany* 30: 712-735.
108. Pirie M.D., Chatrou L.W., Mols J.B., Erkens R.H., Oosterhof J. 2006. 'Andean-centered' genera in the short-branch clade of Annonaceae: testing biogeographical hypotheses using phylogeny reconstruction and molecular dating. *Journal of Biogeography* 33: 31-46.
109. Erkens R.H., Chatrou L.W., Maas J.W., van der Niet T., Savolainen V. 2007. A rapid diversification of rainforest trees (*Guatteria*; Annonaceae) following dispersal from Central into South America. *Molecular Phylogenetics and Evolution* 44: 399-411.
110. Couvreur T.L.P., Pirie M.D., Chatrou L.W., Saunders M.K., Su Y.C.F., Richardson J.E., Erkens R.H. 2011. Early evolutionary history of the flowering plant family Annonaceae: steady diversification and boreotropical geodispersal. *Journal of Biogeography* 38: 664-680.

111. Bardon, L., Chamagne, J., Dexter, K.G., Sothers, C.A., Prance, G.T., Chave, J. 2013. Origin and evolution of Chrysobalanaceae. *Botanical Journal of the Linnean Society* 171: 19-37.
112. Dick C.W., Abdul-Salim K., Bermingham E. 2003. Molecular systematic analysis reveals cryptic Tertiary diversification of a widespread tropical rain forest tree. *The American Naturalist* 162: 691-703.
113. Renner S.S., Meyer K. 2001. Melastomeae come full circle: biogeographic reconstruction and molecular clock dating. *Evolution* 55: 1315-1324.
114. Symmank L. Samain M.-S., Smith J.F, Pino G., Still A., Goetghebeur P., Neinhuis C., Wanke S. 2011. The extraordinary journey of *Peperomia* subgenus *Tildenia* (Piperaceae): insights into diversification and colonization patterns from its cradle in Peru to the Trans-Mexican Volcanic Belt. *Journal of Biogeography* 38: 2337-2349.
115. Emadzade K., Hörandl E. 2011. Northern hemisphere origin, transoceanic dispersal, and diversification of Ranunculeae DC. (Ranunculaceae) in the Cenozoic. *Journal of Biogeography* 38: 517-530.
116. Liu, X.-Q., Ickert-Bond, S.M., Chen, L.-Q., Wen, J. 2013. Molecular phylogeny of *Cissus* L. of Vitaceae (the grape family) and evolution of its pantropical intercontinental disjunctions. *Molecular Phylogenetics and Evolution* 66: 43-53.
117. Weeks A., Daly D.C., Simpson B.B. 2005. The phylogenetic history and biogeography of the frankincense and myrrh family (Burseraceae) based on nuclear and chloroplast sequence data. *Molecular Phylogenetics and Evolution* 35: 85-101.
118. Muellner A.N., Pennington T.D., Koecke A.V., Renner S.S. 2010. Biogeography of *Cedrela* (Meliaceae, Sapindales) in Central and South America. *American Journal of Botany* 97: 511-518.
119. Clayton J.W., Soltis P.S., Soltis D.E. 2009. Recent long-distance dispersal overshadows ancient biogeographical patterns in a pantropical angiosperm family (Simaroubaceae, Sapindales). *Systematic Biology* 58: 395-410.
120. Lohmann, L.G., Bell, C.D., Calio, M.F., Winkworth, R.C. 2013. Pattern and timing of biogeographical history in the Neotropical tribe Bignoniaceae (Bignoniaceae). *Botanical Journal of the Linnean Society* 171: 154-170.
121. Nie, Z.L., Sun, H., Chen, Z.D., Meng, Y., Manchester, S.R., Wen, J. 2010. Molecular phylogeny and biogeographic diversification of *Parthenocissus* (Vitaceae) disjunct between Asia and North America. *American Journal of Botany* 97: 1342-1353.
122. Amo de Paz, G., Cubas, P., Crespo, A., Elix, J.A., Lumbsch, H.T. 2012. Transoceanic dispersal and subsequent diversification on separate continents shaped diversity of the *Xanthoparmelia pulla* group (Ascomycota). *PLoS One* 7: e39683.
123. Morse G.E., Farrell B.D. 2005. Interspecific phylogeography of the *Strator limbatus* species complex: The geographic context of speciation and specialization. *Molecular Phylogenetics and Evolution* 36: 201-213.
124. Loaiza, J.R., Scott, M.E., Bermingham, E., Sanjur, O.I., Wilkerson, R., Rovira, J., Gutierrez, L.A., Correa, M.M., Grijalva, M.J., Birnberg, L., Bickersmith, S., Conn, J.E. 2010. Late Pleistocene environmental changes lead to unstable demography and population divergence of *Anopheles albimanus* in the northern Neotropics. *Molecular Phylogenetics and Evolution* 57: 1341-1346.

125. Hines H. 2008. Historical biogeography, divergence times, and diversification patterns of bumble bees. *Systematic Biology* 57: 58-75.
126. Ramírez, S.R., Roubik D.W., Skov C., Pierce N.E. 2010. Phylogeny, diversification patterns and historical biogeography of euglossine orchid bees (Hymenoptera: Apidae). *Biological Journal of the Linnean Society* 100: 552-572.
127. Rasmussen C., Cameron S.A. 2010. Global stingless bee phylogeny supports ancient divergence, vicariance, and long distance dispersal. *Biological Journal of the Linnean Society* 99: 206-232.
128. Condamine, F.L., Silva-Brandao, K.L., Kergoat, G.J., Sperling, F.A.H. 2012. Biogeographic and diversification patterns of Neotropical Troidini butterflies (Papilionidae) support museum model of diversity dynamics for Amazonia. *BMC Evolutionary Biology* 12: e82.
129. Husemann, M., Guzman, N.V., Danley, P.D., Cigliano, M.M., Confalonieri, V.A. 2013. Biogeography of *Trimerotropis pallidipennis* (Acrididae: Oedipodinae): deep divergence across the Americas. *Journal of Biogeography* 40: 261-273.
130. Gilbert C., Ropiquet A., Hassanin A. 2006. Mitochondrial and nuclear phylogenies of Cervidae (Mammalia, Ruminantia): Systematics, morphology, and biogeography. 2006. *Molecular Phylogenetics and Evolution* 40: 101-117.
131. Perini F.A., Russo C.A.M., Schrago C.G. 2009. The evolution of South American endemic canids: a history of rapid diversification and morphological parallelism. *Journal of Evolutionary Biology* 23: 311-322.
132. Johnson, W.E., Pecon Slattery J., Eizirik E., Kim J.-H., Menotti Raymond M., Bonacic C., Cambre R., Crawshaw P., Nunes A., Seuánez H.N., Martins Moreira M.A., Seymour K.L., Simon F., Sawnsen W., O'Brien S.J. 1999. Disparate phylogeographic patterns of molecular genetic variation in four closely related South American small cat species. *Molecular Ecology* 8: S79-S94.
133. Koepfli K.-P., Deere K.A., Slater G.J., Begg C., Begg K., Grassman L., Lucherini M., Veron G., Wayne R.K. 2008. Multigene phylogeny of the Mustelidae: Resolving relationships, tempo and biogeographic history of a mammalian adaptive radiation. *BMC Biology* 6: e10.
134. Koepfli K.-P., Gompert M.E., Eizirik E., Ho C.-C., Linden L., Maldonado J.E., Wayne R.K. 2007. Phylogeny of the Procyonidae (Mammalia: Carnivora): Molecules, morphology and the Great American Interchange. *Molecular Phylogenetics and Evolution* 43: 1076-1095.
135. Krause, J., Unger, T., Nocon, A., Malaspinas, A.-S., Kolokotronis, S.-O., Stiller, M., Soibelzon, L., Spriggs, H., Dear, P.H., Briggs, A.W., Bray, S.C.E., O'Brian, S.J., Rabeder, G., Matheus, P., Cooper, A., Slatkin, M., Paabo, S., Hofreiter, M. 2008. Mitochondrial genomes reveal an explosive radiation of extinct and extant bears near the Miocene-Pliocene boundary. *BMC Evolutionary Biology* 8: e 220.
136. Martins, F.M., Templeton, A.R., Pavan, A.C.O., Kohlbach, B.C., Morgante, J.S. 2009. Phylogeography of the common vampire bat (*Desmodus rotundus*): marked population structure, Neotropical Pleistocene vicariance and incongruence between nuclear and mtDNA markers. *BMC Evolutionary Biology* 9: e294.

137. Lim B.K. 2008. Historical biogeography of new World emballonurid bats (tribe Diclidurini): taxon pulse diversification. *Journal of Biogeography* 35: 1385-1401.
138. Ruedi M., Mayar F. 2001. Molecular systematics of bats of the genus *Myotis* (Vespertilionidae) suggests deterministic ecomorphological convergences. *Molecular Phylogenetics and Evolution* 21: 436-448.
139. Cortés-Ortiz L., Bermingham E., Rico C., Rodríguez-Luna E., Sampaio I., Ruiz-García M. 2003. Molecular systematics and biogeography of the Neotropical monkey genus, *Alouatta*. *Molecular Phylogenetics and Evolution* 26: 64-81.
140. Collins A.C., Dubach J.M. 2000. Biogeographic and ecological forces responsible for speciation in *Ateles*. *International Journal of Primatology* 21: 421-444.
141. Lynch Alfaro, J.W., Boubli, J.P., Olson, L.E., Di Fiore, A., Wilson, B., Gutierrez-Espeleta, Chiou, K.L., Schulte, M., Neitzel, S., Ross, V., Schwochow, D., Nguyen, M.T.T., Farias, I., Janson, C.H., Alfaro, M.E. 2012. Explosive Pleistocene range expansion leads to widespread Amazonian sympatry between robust and gracile capuchin monkeys. *Journal of Biogeography* 39: 272-288.
142. Chou, K.L., Pozzi, L., Lynch Alfaro, J.W., Di Fiore, A. 2011. Pleistocene diversification of living squirrel monkeys (*Saimiri* spp.) inferred from complete mitochondrial genome sequences. *Molecular Phylogenetics and Evolution* 59: 736-745.
143. Smith M.F., Patton J.L. 1999. Phylogenetic relationships and the radiation of Sigmodontine rodents in South America: Evidence from Cytochrome *b*. *Journal of Mammalian Evolution* 6: 89-128.
144. Mercer, J.M., Roth, V.L. 2003. The effects of Cenozoic global change on squirrel phylogeny. *Science* 299: 1569-1572.
145. Chen, L.-Y., Chen, J.-M., Gituru, R.W., Wang, Q.-F. 2012. Generic phylogeny, historical biogeography and character evolution of the cosmopolitan aquatic plant family Hydrocharitaceae. *BMC Evolutionary Biology* 12: e30.
146. Eiserhardt W.L., Pinaud J.-C., Asmussen-Lange C., Hahn W.J., Bernal R., Balslev H., Borchsenius F. 2011. Phylogeny and divergence times of Bactridinae (Arecaceae, Palmae) based on plastid and nuclear chloroplast DNA sequences. *Taxon* 60: 485-498.
147. Roncal, J., Kahn, F., Millan, B., Couvreur, T.L.P., Pinaud, J.-C. 2013. Cenozoic colonization and diversification patterns of tropical American palms: evidence from *Astrocaryum* (Arecaceae). *Botanical Journal of the Linnean Society* 171: 120-139.
148. Cuenca A., Asmussen-Lange C., Borchsenius F. 2008. A dated phylogeny of the palm tribe Chamaedoreae supports Eocene dispersal between Africa, North and South America. *Molecular Phylogenetics and Evolution* 46: 760-775.
149. Bacon, C.D., Mora, A., Wagner, W., Jaramillo, C.J. 2013. Testing geological models of evolution of the Isthmus of Panama in a phylogenetic framework. *Botanical Journal of the Linnean Society* 171: 287-300.
150. Roncal J., Blach-Overgaard, Borchsenius F., Balslev H., Svenning J.-C. 2011. A dated phylogeny complements macroecological analysis to explain the diversity patterns in *Geonoma* (Arecaceae). *Biotropica* 43: 324-334.
151. Trénel P., Gustafsson M.H.G., Baker W.J., Asmussen-Lange C.B., Dransfield J., Borchsenius F. 2007. Mid-Tertiary, not Gondwanan vicariance explains distribution

- patterns in the wax palm subfamily (Ceroxyloideae: Arecaceae). *Molecular Phylogenetics and Evolution* 45: 272-288.
152. Wagner, N. Silvestro, D., Brie, D., Ibsch, P.L., Zizka, G., Weising, K., Schulte, K. 2012. Spatio-temporal evolution of *Fosterella* (Bromeliaceae) in the Central Andean biodiversity hotspot. *Journal of Biogeography* 40:869-880.
  153. Givnish T.J., Barfuss M.H.J., Van Ee B., Riina R., Schulte R., Gonsiska P.A., Jabailey R.S., Crayn D.M., Smith A.C., Winter K., Brown G.K., Evans T.M., Holst B.K., Luther H., Till W., Zizka G., Berry P.E., Sytsma K.J. 2011. Phylogeny, adaptive radiation, and historical biogeography in Bromeliaceae: Insights from an eight-locus plastid phylogeny. *American Journal of Botany* 98: 872-895.
  154. Chacón J., Madriñán S., Chase M.W., Bruhl J.J. 2006. Molecular phylogenetic of *Oreobolus* (Cyperaceae) and the origin and diversification of the American species. *Taxon* 55: 359-366.
  155. Inda L.A., Segarra-Moragues J.G., Müller J., Peterson P.M., Catalán P. 2008. Dated historical biogeography of the temperate Loliinae (Poaceae, Pooideae) grasses in the northern and southern hemispheres. *Molecular Phylogenetics and Evolution* 46: 932-957.
  156. Sessa, E.B., Zimmer, E.A., Givnish, T.J. 2012. Phylogeny, divergence times, and historical biogeography of New World *Dryopteris* (Dryopteridaceae). *American Journal of Botany* 99: 730-750.
  157. Colston, T.J., Grazziotin, F.E., Shepard, D.B., Vitt, L.J., Colii, G.R., Henderson, R.W., Hedges, S.B., Bonatto, S., Zaher, H., Noonan, B.P., Burbirnk, F.T. 2013. Molecular systematics and historical biogeography of tree boas (*Corallus* spp.). *Molecular Phylogenetics and Evolution* 66: 953-959.
  158. Daza J.M., Smith E.N., Páez V.P., Parkinson C.L. 2009. Complex evolution in the Neotropics: The origin and diversification of the widespread genus *Leptodeira* (Serpentes: Colubridae). *Molecular Phylogenetics and Evolution* 53: 653-667.
  159. Castoe T.A., Daza J.M., Smith E.N., Sasa M.M., Kuch U., Campbell J.A., Chippindale P.T., Parkinson C.L. 2009. Comparative phylogeography of pitvipers suggests a consensus of ancient Middle American highland biogeography. *Journal of Biogeography* 36: 88-103.
  160. Wüster W., Ferguson J.E., Quijada-Mascareñas J.A., Pook C.E., da Graça Salomão M., Thorpe R.S. 2005. Tracing an invasion: landbridges, refugia, and the phylogeography of the Neotropical rattlesnake (Serpentes: Viperidae: *Crotalus durissus*). *Molecular Ecology* 14: 1095-1108.
  161. Wüster W., Peppin L., Pook C.E., Walker D.E. 2008. A nesting of vipers: Phylogeny and historical biogeography of the Viperidae (Squamata: Serpentes). *Molecular Phylogenetics and Evolution* 49: 445-459.
  162. Zamudio K.R., Greene H.W. 1997. Phylogeography of the bushmaster (*Lachesis muta*: Viperidae): implications for neotropical biogeography, systematics, and conservation. *Biological Journal of the Linnean Society* 62: 421-442.
  163. Fritz U., Stuckas H., Vargas-Ramírez M., Hundsörfer A.K., Maran J., Päckert M. 2011. Molecular phylogeny of Central and South American slider turtles: implications for biogeography and systematics (Testudines: Emydidae: *Trachemys*). *Journal of Zoological Systematics and Evolutionary Research* 50: 125-136.

164. Le M., McCord W.P. 2008. Phylogenetic relationships and biogeographical history of the genus *Rhinoclemmys* Fitzinger, 1835 and the monophyly of the turtle family Geomydidae (Testudines: Testudinoidea). *Zoological Journal of the Linnean Society* 153: 751-767.
165. Venegas-Anaya, M., Crawford, A.J., Escobedo Galvan, A.H., Sanjur, O.I., Densmore III, L.D., Bermingham, E. 2008. Mitochondrial DNA phylogeography of *Caiman crocodilus* in Mesoamerica and South America. *Journal of Experimental Zoology* 309A: 614-627.
166. Gamble, T., Bauer, A.M., Colli, G.R., Greenbaum, E., Jackman, T.R., Vitt, L.J., Simons, A.M. 2011. Coming to America: multiple origins of New World geckos. *Journal of Evolutionary Biology* 24: 231-244.
167. Gamble, T., Simons, A.M., Colli, G.R., Bitt, L.J. 2008. Tertiary climate change and the diversification of the Amazonian gecko genus *Gonatodes* (Sphaerodactylidae, Squamata). *Molecular Phylogenetics and Evolution* 46: 269-277.

**Table S9. List of journals used for the present study.**

Name of scientific journal
American Journal of Botany
Biological Journal of the Linnean Society
BMC Biology
BMC Evolution Biology
Botanical Journal of the Linnean Society
Cladistics
Evolution
International Journal of Primatology
Journal of Biogeography
Journal of Evolutionary Biology
Journal of Mammalian Evolution
Journal of Zoological Systematics and Evolutionary Research
Molecular Biology and Evolution
Molecular Ecology
Molecular Phylogenetics and Evolution
Nature
Organisms Diversity and Evolution
Philosophical Transactions of the Royal Society B
Proceedings of the National Academy of Sciences, USA
Proceedings of the Royal Society B
Science
Systematic Biology
Systematic Botany
Taxon
The American Naturalist
The Auk
Zoological Journal of the Linnean Society

**Table S10. Migration models estimated from analysis of phylogenetic data.**

Data set	Sample size	Families	Model	d.f.	log likelihood	AICc	stepwise deltaAICc	delta AICc*	AICc threshold
All taxa	426	138	constant rate	1	453.175	-904.34		965.14	6.081
			2 rates	3	865.231	-1724.405	820.065	145.075	
			3 rates	5	921.034	-1831.924	107.519	37.556	
			4 rates	7	937.353	-1860.438	28.514	9.042	
			<b>5 rates</b>	<b>9</b>	<b>943.957</b>	<b>-1869.48</b>	<b>9.042</b>	<b>0</b>	
			6 rates	11	946.148	-1869.658	0.178	-0.178	
			exponential	2	458.689	-913.35		956.13	
			shift at 3.5	2	670.723	-1337.417		532.063	
NA -> SA	144	60	constant rate	1	17.232	-32.436		248.492	5.984
			2 rates	3	127.556	-248.941	216.505	31.987	
			<b>3 rates</b>	<b>5</b>	<b>145.682</b>	<b>-280.928</b>	<b>31.987</b>	<b>0</b>	
			4 rates	7	149.98	-285.136	4.208	-4.208	
			exponential	2	20.925	-37.765		243.163	
			shift at 3.5	2	78.938	-153.792		127.136	
SA -> NA	154	62	constant rate	1	7.131	-12.235		377.276	5.995
			2 rates	3	179.948	-353.737	341.502	35.774	
			<b>3 rates</b>	<b>5</b>	<b>199.958</b>	<b>-389.511</b>	<b>35.774</b>	<b>0</b>	
			4 rates	7	205.088	-395.408	5.897	-5.897	
			exponential	2	14.706	-25.333		364.178	
			shift at 3.5	2	102.847	-201.615		187.896	
Marine (Atlantic - Pacific)	86	13	constant rate	1	-2.313	6.673		94.847	5.853
			2 rates	3	38.810	-71.327	78.000	16.847	
			3 rates	5	44.363	-77.977	6.650	10.197	
			<b>4 rates</b>	<b>7</b>	<b>51.805</b>	<b>-88.174</b>	<b>10.197</b>	<b>0</b>	
			5 rates	9	53.35	-86.331	-1.843	1.843	
			exponential	2	-0.902	5.949		94.123	
			shift at 3.5	2	5.413	-6.681		81.493	
Terrestrial tetrapods	59	29	constant rate	1	-23.673	49.416		63.754	5.675
			2 rates	3	7.02	-7.604	57.02	6.734	



			<b>3 rates</b>	<b>5</b>	<b>12.735</b>	<b>-14.338</b>	<b>6.734</b>	<b>0</b>	
			4 rates	7	14.22	-12.244	-2.094	2.094	
			exponential	2	-21.867	47.948		62.286	
			shift at 3.5	2	-9.674	23.563		37.901	
Reptiles and amphibians	35	15	constant rate	1	-32.32	66.761		22.364	5.210
			<b>2 rates</b>	<b>3</b>	<b>-18.811</b>	<b>44.397</b>	<b>22.364</b>	<b>0</b>	
			3 rates	5	-15.594	43.258	1.139	-1.139	
			exponential	2	-30.999	66.372		21.975	
			shift at 3.5	2	-26.671	57.718		13.321	
Fresh water fish	14	4	constant rate	1	-31.534	65.402		3.937	2.872
			<b>2 rates</b>	<b>3</b>	<b>-26.533</b>	<b>61.465</b>	<b>3.936</b>	<b>0</b>	
			3 rates	5	-24.241	65.983	<b>-4.518</b>	4.518	
			exponential	2	-31.700	68.492		7.027	
			shift at 3.5	2	-30.904	66.898		5.433	
Arthropods	35	7	constant rate	1	-37.26	76.642		37.571	5.210
			<b>2 rates</b>	<b>3</b>	<b>-16.148</b>	<b>39.071</b>	<b>37.571</b>	<b>0</b>	
			3 rates	5	-12.573	37.215	1.856	-1.856	
			exponential	2	-35.519	75.414		36.343	
			shift at 3.5	2	-33.065	70.506		31.435	
Plants	74	31	constant rate	1	-40.41	82.876		85.756	5.793
			2 rates	3	-3.479	13.301	69.575	16.181	
			<b>3 rates</b>	<b>5</b>	<b>6.881</b>	<b>-2.88</b>	<b>16.181</b>	<b>0</b>	
			4 rates	7	9.342	-2.987	0.107	-0.107	
			exponential	2	-38.638	81.446		84.326	
			shift at 3.5	2	-31.987	68.144		71.024	
Birds	158	36	constant rate	1	236.021	-470.017		188.115	5.999
			2 rates	3	319.657	-633.158	163.141	24.974	
			3 rates	5	330.176	-649.958	16.8	8.174	
			<b>4 rates</b>	<b>7</b>	<b>336.439</b>	<b>-658.132</b>	<b>8.174</b>	<b>0</b>	
			5 rates	9	338.829	-658.443	0.311	-0.311	
			exponential	2	238.056	-472.035		186.097	

			shift at 3.5	2	293.5	-582.923		75.209	
Mammals	24	14	constant rate	1	-18.893	39.968		17.532	4.585
			<b>2 rates</b>	<b>3</b>	<b>-7.618</b>	<b>22.436</b>	<b>17.532</b>	<b>0</b>	
			3 rates	5	-6.225	25.783	-3.347	3.347	
			exponential	2	-17.809	40.19		17.754	
			shift at 3.5	2	-13.78	32.132		9.696	
Mollusc + Urchin	32	13	constant rate	1	-32.496	67.126		12.252	5.090
			2 rates	3	-25.188	57.232	9.894	2.358	
			<b>3 rates</b>	<b>5</b>	<b>-21.283</b>	<b>54.874</b>	<b>2.358</b>	<b>0</b>	
			4 rates	7	-16.522	51.71	3.164	-3.164	
			exponential	2	-32.112	68.638		13.764	
			shift at 3.5	2	-32.445	69.304		14.43	
Low dispersal ability	61	30	constant rate	1	-53.666	109.4		72.213	5.695
			2 rates	3	-23.431	53.283	56.117	16.096	
			<b>3 rates</b>	<b>5</b>	<b>-13.048</b>	<b>37.187</b>	<b>16.096</b>	<b>0</b>	
			4 rates	7	-9.672	35.457	1.73	-1.73	
			exponential	2	-51.337	106.88		69.693	
			shift at 3.5	2	-43.44	91.087		53.9	
Medium dispersal ability	78	36	constant rate	1	-38.488	79.029		92.131	5.815
			2 rates	3	0.479	5.365	73.664	18.467	
			<b>3 rates</b>	<b>5</b>	<b>11.968</b>	<b>-13.102</b>	<b>18.467</b>	<b>0</b>	
			4 rates	7	14.598	-13.595	0.493	-0.493	
			exponential	2	-36.483	77.126		90.228	
			shift at 3.5	2	-28.173	60.505		73.607	
High dispersal ability	287	77	constant rate	1	298.351	-594.688		634.561	6.061
			2 rates	3	572.303	-1138.522	543.834	90.727	
			3 rates	5	603.258	-1196.303	57.781	32.946	
			<b>4 rates</b>	<b>7</b>	<b>621.825</b>	<b>-1229.249</b>	<b>32.946</b>	<b>0</b>	
			5 rates	9	626.939	-1235.229	5.98	-5.98	
			exponential	2	303.134	-602.226		627.023	
			shift at 3.5	2	457.524	-911.005		318.244	

The results of migration rate analyses are shown for the full dataset, and several subsets defined by direction of migration, habitat, taxonomic group. Different migration models were compared by AICc values. To determine the number of rate shifts supported by the data we optimized models with increasing number of rate shifts using a stepwise AICc approach, i.e. new rate shifts were added to the model until the improvement in AICc score was lower than a threshold estimated from simulations (see text for more details). The 'stepwise deltaAICc' column shows the difference in AICc scores between the current migration model and the previous, i.e. with one fewer rate shift. Additionally, we optimized a migration model with exponentially increasing rates and a "Standard" model (assuming one rate shift fixed at time 3.5 Ma). These models were rejected by AICc in favor of models with one or more rate shifts. The overall preferred model, used for parameter estimation, is highlighted in bold and delta-AICc scores are calculated with respect to the preferred model.