

# class11.Rmd

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```
db <- read.csv("Data Export Summary.csv", row.names = 1)
head(db)
```

##	X.ray	NMR	EM	Multiple.methods	Neutron	Other	Total
## Protein (only)	142303	11804	5999	177	70	32	160385
## Protein/Oligosaccharide	8414	31	979	5	0	0	9429
## Protein/NA	7491	274	1986	3	0	0	9754
## Nucleic acid (only)	2368	1372	60	8	2	1	3811
## Other	149	31	3	0	0	0	183
## Oligosaccharide (only)	11	6	0	1	0	4	22

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

87.6% structures in PDB solved by X-ray and 4.92% by EM.

```
method.sums <- colSums(db)
method.sums
```

##	X.ray	NMR	EM	Multiple.methods
##	160736	13518	9027	194
##	Neutron	Other	Total	
##	72	37	183584	

```
XRay <- sum(db$X.ray)
EM <- sum(db$EM)
total <- sum(db$Total)
```

```
(XRay/total) *100
```

```
## [1] 87.55447
```

```
(EM/total) *100
```

```
## [1] 4.917095
```

Q2: What proportion of structures in the PDB are protein?

87.4% are protein only structures in the PDB.

```
type.sums <- rowSums(db)
type.sums
```

```
##           Protein (only) Protein/Oligosaccharide           Protein/NA
##           320770           18858           19508
##   Nucleic acid (only)           Other Oligosaccharide (only)
##           7622           366           44
```

```
total1 <- sum(type.sums)
(type.sums["Protein (only)"]/total1)*100
```

```
## Protein (only)
##           87.36328
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

There are currently 1828 HIV-1 protease structures in the current PDB.

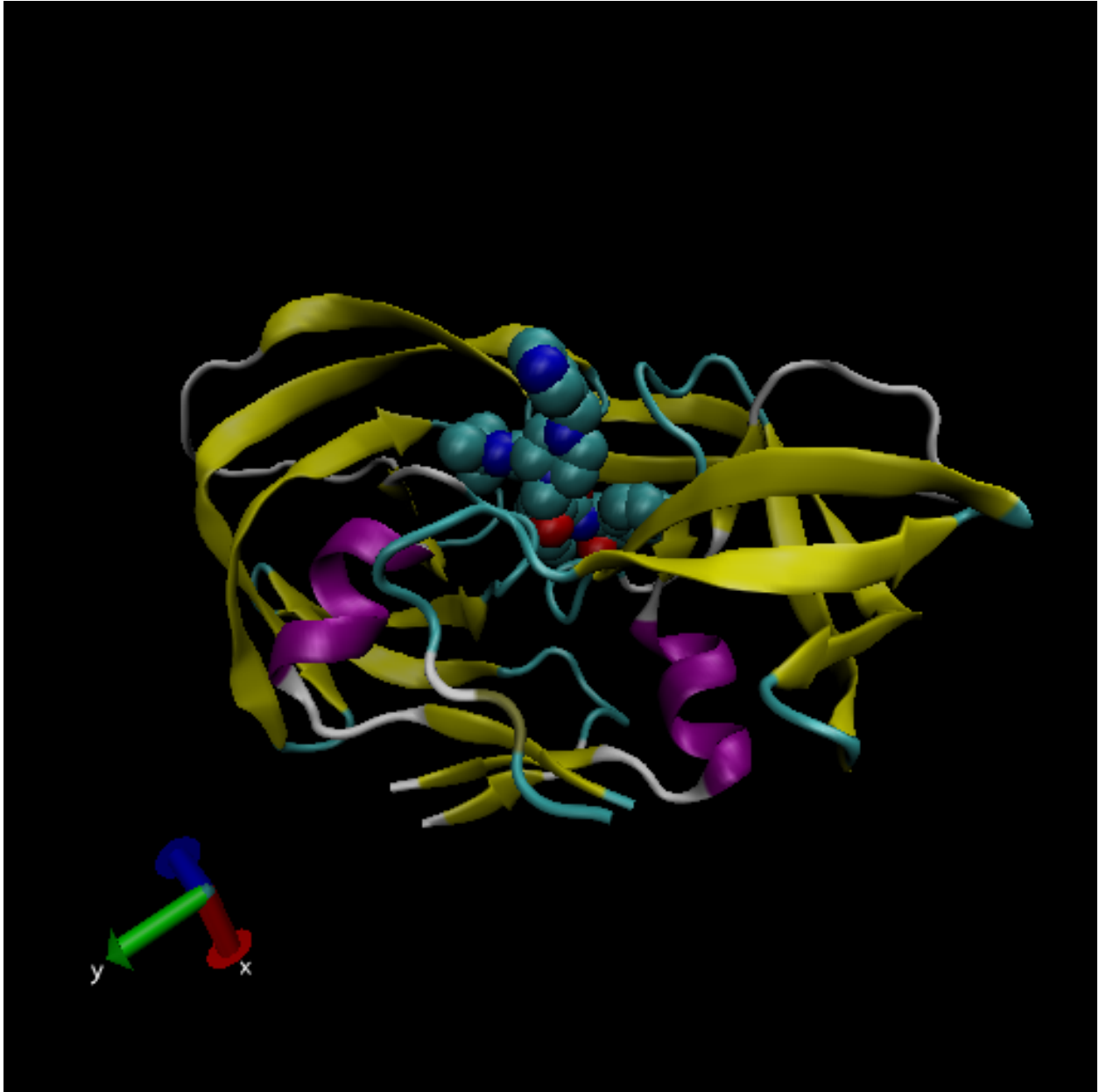
Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

The hydrogen atoms per water molecule (2) are not shown because they are very small atoms compared to the oxygen (the one actually shown) of the water molecules and the other atoms of the structure.

Q5: There is a conserved water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have (see note below)?

residue number MK1902:O4

## VMD structure visualization image



Just write text but you have some **formatting** options.

I need to load library

```
library(bio3d)
```

```
pdb <- read.pdb("1hsg")
```

```
## Note: Accessing on-line PDB file
```

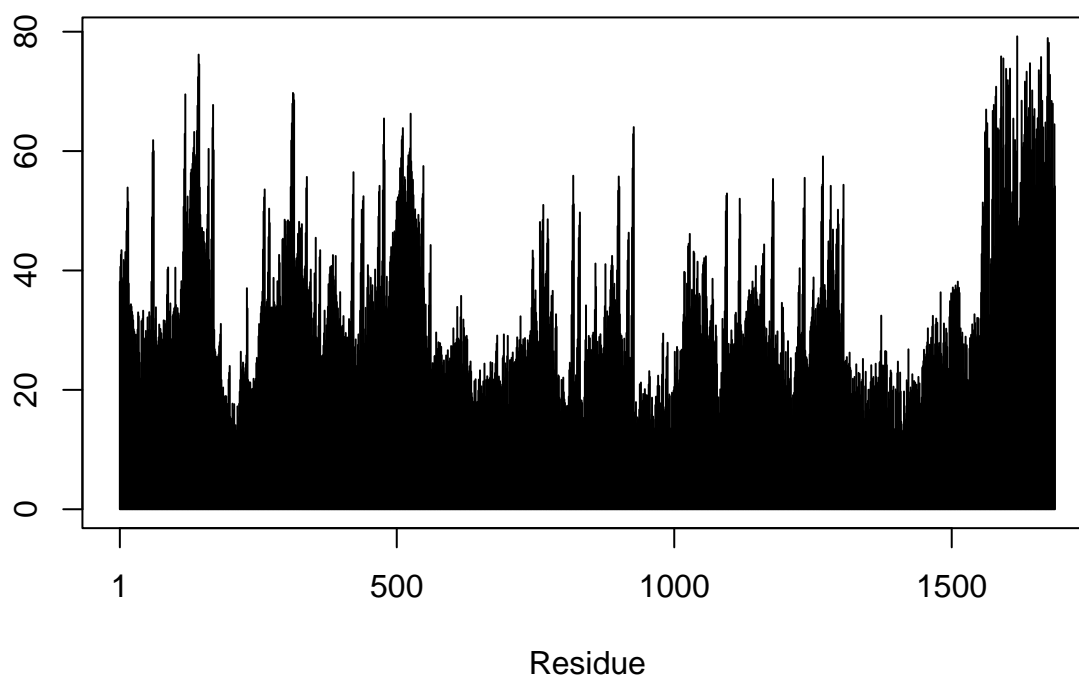
```
pdb
```

```
##
## Call: read.pdb(file = "1hsg")
##
## Total Models#: 1
## Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
##
## Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
## Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
##
## Non-protein/nucleic Atoms#: 172 (residues: 128)
## Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
##
## Protein sequence:
## PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
## QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
## ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
## VNIIGRNLLTQIGCTLNF
##
## + attr: atom, xyz, seqres, helix, sheet,
## calpha, remark, call
```

Plot B Factor

```
plot.bio3d(pdb$atom$b, sse = pdb)
```

```
## Warning in plotb3(...): Length of input 'sse' does not equal the length of input
## 'x'; Ignoring 'sse'
```



The ATOM records

```
head(pdb$atom)
```

```
##      type eleno elety  alt resid chain resno insert      x      y      z o      b
## 1 ATOM      1      N <NA>  PRO      A      1  <NA> 29.361 39.686 5.862 1 38.10
## 2 ATOM      2      CA <NA>  PRO      A      1  <NA> 30.307 38.663 5.319 1 40.62
## 3 ATOM      3      C  <NA>  PRO      A      1  <NA> 29.760 38.071 4.022 1 42.64
## 4 ATOM      4      O <NA>  PRO      A      1  <NA> 28.600 38.302 3.676 1 43.40
## 5 ATOM      5      CB <NA>  PRO      A      1  <NA> 30.508 37.541 6.342 1 37.87
## 6 ATOM      6      CG <NA>  PRO      A      1  <NA> 29.296 37.591 7.162 1 38.40
##      segid elesy charge
## 1 <NA>      N  <NA>
## 2 <NA>      C  <NA>
## 3 <NA>      C  <NA>
## 4 <NA>      O  <NA>
## 5 <NA>      C  <NA>
## 6 <NA>      C  <NA>
```