

ScienceDirect



Neuromodulation of hippocampal long-term synaptic plasticity

Jon Palacios-Filardo and Jack R Mellor



Multiple neuromodulators including acetylcholine, noradrenaline, dopamine and serotonin are released in response to uncertainty to focus attention on events where the predicted outcome does not match observed reality. In these situations, internal representations need to be updated, a process that requires long-term synaptic plasticity. Through a variety of common and divergent mechanisms, it is recently shown that all these neuromodulators facilitate the induction and/or expression of long-term synaptic plasticity within the hippocampus. Under physiological conditions, this may be critical for suprathreshold induction of plasticity endowing neuromodulators with a gating function and providing a mechanism by which neuromodulators enable the targeted updating of memory with relevant information to improve the accuracy of future predictions.

Address

Centre for Synaptic Plasticity, School of Physiology, Pharmacology and Neuroscience, University of Bristol, Bristol BS8 1TD, UK

Corresponding author: Mellor, Jack R (Jack.Mellor@Bristol.ac.uk)

Current Opinion in Neurobiology 2019, 54:37-43

This review comes from a themed issue on **Neurobiology of learning** and plasticity

Edited by Scott Waddell and Jesper Sjöström

For a complete overview see the Issue and the Editorial

Available online 10th September 2018

https://doi.org/10.1016/j.conb.2018.08.009

0959-4388/© 2018 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

Introduction

In competitive environments, it is important to make rapid accurate decisions in response to changing situations. This requires neuronal networks to accurately predict outcomes based on prior knowledge. The hippocampus plays a key role in the formation, updating and retrieval of memories which form the basis of outcome prediction. Memories are formed in the hippocampus by creating ensembles of strongly coupled neurons that encode a specific event or episode by the process of long-term synaptic plasticity which can be either potentiation (LTP) or depression (LTD). However, nowhere near all experienced events are turned into memories and therefore a filter operates to ensure only the most salient events are memorised. But what defines salience in this

context? It is proposed that salience is signalled when a predicted outcome does not match the observed outcome leading to a state of uncertainty which may be reduced by updating the relevant memories [1,2]. Under this theoretical framework, uncertainty triggers the release of neuromodulators that act as the neurobiological filter controlling which memories are formed and updated [3]. Interestingly, the increase in uncertainty and release of neuromodulators often results from, and therefore occurs after, the salient event. This means that the mechanisms controlling synaptic plasticity need to incorporate an extended temporal window of interaction between a salient event and release of neuromodulator [4–6].

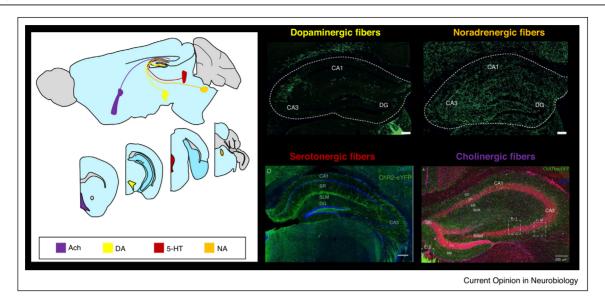
This review focuses on the mechanisms by which the neuromodulators acetylcholine, dopamine, noradrenaline and serotonin modulate long-term synaptic plasticity in the hippocampus, highlighting common and divergent themes. For more in-depth reviews on the action of each individual neuromodulator on synaptic plasticity please see [7–10]. We propose that all the neuromodulators facilitate long-term synaptic plasticity often via common mechanisms but with differences potentially reflecting distinct aspects of uncertainty encoded by each neuromodulator.

Acetylcholine

Acetylcholine release in the hippocampus is prominently associated with learning and memory with acetylcholine depletion or cholinergic receptor blockade resulting in memory deficits. Cholinergic fibers from the medial septum/diagonal band of Broca release acetylcholine into the hippocampus (Figure 1) in response to arousal and primary reinforcement cues [11,12] which can also be described as expected and unexpected uncertainty [1].

Acetylcholine activates nicotinic ionotropic (principally α4β2, α3β4 and α7 in the hippocampus) and muscarinic (M1–4) G-protein coupled receptors (GPCRs) in different cell types and subcellular locations generating a myriad of potential signalling pathways. Increasingly, these pathways are seen to act in a coordinated manner to facilitate the induction and expression of LTP and LTD (Figure 2). Most of these pathways act to enhance NMDA receptor (NMDAR) activity but through several separate mechanisms. Firstly, muscarinic M1 receptors enhance postsynaptic excitability and NMDAR activity by inhibiting potassium channels including voltage activated Kv7 [13], Kv4.2 [14] and calcium activated SK

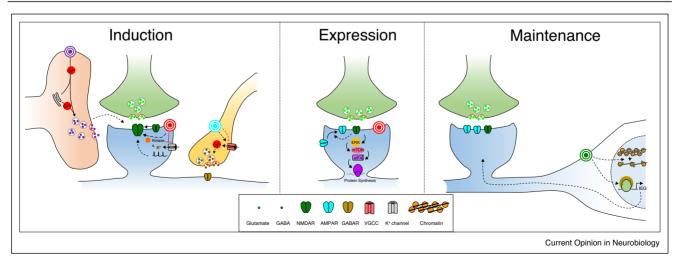
Figure 1



Neuromodulator projections to the hippocampus. (Left) Location of nuclei containing the cell bodies that give rise to neuromodulator projections for Acetylcholine (Ach), Dopamine (DA), Serotonin (5-HT) and Noradrenaline (NA). Sagittal (top) and Coronal (bottom) sections. Data from Allen Brain Atlas for mouse brain. (Right) Images of neuromodulatory projection fibers within the hippocampus labelled with eYFP targeted by viral injection into VTA and LC of TH-cre mice (top left and right respectively), YFP tagged channel rhodopsin in ePet1-cre:Ai32 mice (bottom left) and GFP in ChAT-tauGFP mice (bottom right). Images taken from Refs. [38**,65**,77]. Scale bars 200 µm.

channels [15,16] which facilitates the induction of LTP [13,16–18]. Secondly, acetylcholine also enhances NMDARs directly via a muscarinic receptor initiated signalling cascade where increased IP3 levels activate a series of protein kinases, including CAMKII, PKC and SRC, to facilitate induction of LTP [19] or through a direct interaction between nicotinic receptors and NMDARs [20]. However, it is reported that M1 receptors can also depress NMDAR activity through IP3 induced calcium release from internal stores resulting in dynamin dependent endocytosis of NMDAR [21]. Thirdly, a role for astrocytes has recently been uncovered where activation of astrocytic α7 nicotinic receptors causes the release of D-serine to enhance NMDAR activity and facilitate

Figure 2



Mechanisms for neuromodulator facilitation of long-term synaptic plasticity. Major mechanisms for the regulation of long-term synaptic plasticity include: facilitation of induction by stimulation of astrocytes (purple), increased postsynaptic excitability and NMDAR activity (red) and disinhibition (blue); facilitation of expression by regulation of AMPAR function at the postsynaptic membrane (red); facilitation of maintenance by regulation of gene transcription (green).

LTP induction [22**]. Fourthly, acetylcholine alters inhibitory interneuron activity and synaptic transmission with the potential to fundamentally reconfigure network excitability and favour plasticity at one set of excitatory synapses over another because subtypes of interneurons are able to precisely control specific neuronal dendritic regions. Due to the complexity of interneuron subtypes and functions and their responsiveness to acetylcholine, predicting the exact outcome of acetylcholine release is difficult but broadly an emerging view indicates that acetylcholine increases interneuron firing rates either directly [23] or via activation of astrocytes [24] and increases spontaneous GABA release [25]. However, evoked release of GABA is supressed by activation of presynaptic M2 receptors [26] or by postsynaptic activation of M1 receptors causing retrograde release of endocannabinoids [27]. Although the precise outcome of cholinergic modulation of inhibition is unclear a common feature is its transient nature, supporting a primary role in the induction of synaptic plasticity which is common to most of the mechanisms that facilitate NMDARs. There are a few exceptions to this rule where reports suggest that the expression of synaptic plasticity can also be facilitated by acetylcholine [19]. Furthermore, the facilitation of LTD by acetylcholine is likely via effects on the expression pathway and not through facilitation of NMDAR [21,28,29**].

Overall, the emerging picture indicates that acetylcholine facilitates LTP and LTD in the hippocampus and this facilitation becomes critical when LTP is induced by stimulation protocols that closely mirror endogenous activity patterns [17,28,29**,30] suggesting that in many situations acetylcholine is required for LTP induction in vivo.

Noradrenaline

Noradrenaline release from fibers originating in the Locus Coeruleus (LC) (Figure 1) is strongly associated with novelty or salience [31], otherwise described as unexpected uncertainty [1], and, like acetylcholine, evidence indicates a role in the formation of memories since reduction in noradrenaline or blockade of noradrenergic receptors causes memory impairments [32].

Noradrenaline activates α 1-adrenergic, α 2-adrenergic and β-adrenergic GPCRs with the β receptors, signalling via Gs and cAMP, exerting the major impact on LTP and LTD. Noradrenaline facilitates LTP in CA1 [33–35] but there is also strong evidence for a role enhancing LTP and LTD within dentate gyrus—CA3 circuits [36,37] which links to evidence of increased density of noradrenergic fibers in these regions [38°°]. There are several proposed mechanisms for the facilitation of LTP and LTD (Figure 2). Similar to acetylcholine, noradrenaline enhances NMDAR activity by inhibiting potassium channels including Kv1.1, Kv4.2 and SK channels [33,39,40] and by direct phosphorylation of NMDARs by PKA [41]. Compared to acetylcholine, there is a paucity of data on the actions of noradrenaline on inhibitory interneuron populations, so this area remains to be fully explored. Noradrenaline, via PKA, also phosphorylates AMPA receptors (AMPARs) promoting their insertion into the postsynaptic membrane [42,43]. PKA activation can also recruit protein synthesis [44] representing a delayed noradrenergic mechanism that can contribute to an expanded time window for modulation of LTP (Figure 2).

Despite strong in vitro and in vivo evidence supporting a role for noradrenaline in facilitating LTP and memory formation in the hippocampus, recent data suggests that dopamine rather than noradrenaline released by LC neurons is the important neuromodulator [38°,45,46]. This controversial finding fundamentally questions the role played by noradrenaline in hippocampal LTP and memory and will require further corroboration.

Dopamine

Dopamine release has classically been associated with reward and prediction errors as well as novelty and salience that potentially lead to heightened uncertainty levels [2]. These ideas are principally based on dopamine release patterns in striatum from projections from the Ventral Tegmental Area (VTA) [47] and that selective stimulation of VTA dopaminergic projections induces synaptic plasticity and hippocampal-dependent behavioural changes [48,49°°]. However, projections from the VTA are relatively sparse in the hippocampus with the suggestion that the major source of dopamine in the hippocampus actually arises from LC projections (Figure 1) [38°,45,46].

Dopamine exerts its action through 5 GPCRs clustered into two families: D1-like (D1 and D5) and D2-like (D2, D3 and D4) resulting in increased and decreased cAMP levels respectively. There is evidence for widespread expression of these receptors throughout the hippocampus but with some region-specific differences [50] and dimerization occurs between subtypes and with other neuromodulatory receptors including noradrenergic and cholinergic receptors making receptor pharmacology difficult to interpret [51]. Since the D1-like and D2-like receptors mechanistically act in opposition to each other, dopamine can both facilitate and depress synaptic plasticity but in general the D1 receptors appear to have most influence on plasticity processes decreasing overall cellular excitability but causing disinhibition [52] and modulation of NMDAR signalling [53] leading to facilitation of LTP and LTD (Figure 2) [54–58]. However, stimulation of endogenous dopamine release from VTA projections can produce different effects to exogenous dopamine agonists [49**] questioning the physiological relevance of pharmacological receptor activation.

Dopamine can also facilitate LTP and LTD when the receptors are activated after a plasticity inducing event (often with a considerable delay) [54] via engagement of gene transcription and protein translation (Figure 2). The proposed mechanism suggests that translated proteins are then translocated to synapses primed for plasticity by coincident stimulation [59]. Recent evidence also suggests a role for dopamine in the long-term regulation of inhibition in CA3 which is important for memory consolidation [60] and combines a delayed action for dopamine with changes to the inhibitory network. Such mechanisms are attractive because they align the timescales of behavioural learning with the cellular mechanisms of synaptic plasticity where reward, novelty and salience are often determined after the event.

Serotonin

Serotonin has classically been associated with mood but recent evidence suggests a prominent role in adaptation and plasticity where serotonin release occurs in response to prediction errors thereby promoting learning in conditions of uncertainty [61]. Serotonin is released by fibers projecting diffusely from the median raphe nucleus (MRN) into all parts of the hippocampus (Figure 1).

Serotonin (5-HT) receptors are composed of seven families (5-HT₁ to 5-HT₇) which are coupled to GPCRs except for the ionotropic 5-HT₃ receptors and all 5-HT receptors are expressed in the hippocampus. Serotonin has been shown to increase cellular excitability by several mechanisms (Figure 2), such as inhibition of K⁺ channels [62], and serotonin also enhances NMDAR activation [63] and facilitates LTP expression in CA1 [64,65**,66]. Stimulation of interneurons and astrocytes by activation of 5-HT receptors has been reported [67,68] but it is not yet clear whether this can also facilitate LTP. Conversely, 5-HT is mainly inhibitory in CA3 and prevents LTD or LTP at mossy fiber-CA3 synapses [69] highlighting the cell specific nature of serotonergic signalling.

Similarity and divergence across neuromodulatory systems

The neuromodulators acetylcholine, dopamine, nor-adrenaline and serotonin have historically been viewed as distinct systems signalling different behavioural states but increasingly this compartmentalised perspective is being broken down as overlaps in function and behavioural response are found. Observations of neuromodulator co-release [38**] and co-activation of receptor populations fundamentally questions this discrete model for neuromodulator function. Moreover, an alternative overarching theoretical framework suggests that each neuromodulatory system encodes aspects of uncertainty signalling the need to update memory representations to make better future predictions and reduce uncertainty [1,2]. A central prediction of this theoretical framework is that the release of these neuromodulators facilitates long-term

synaptic plasticity in the hippocampus. In this review we outline evidence to support this prediction where each neuromodulator triggers mechanisms that facilitate LTP and/or LTD. Thus, there is considerable support for a common function for the separate neuromodulatory systems and indeed for some common mechanisms (e.g. enhancing NMDAR activity) but there are also evident differences.

Three potential mechanistic similarities for NMDAR enhancement are highlighted (Figure 2). The first is the pivotal role played by SK channels which control spine NMDAR activity and can be inhibited by acetylcholine, noradrenaline and metabotropic glutamate receptors [15,16,40,70] and may also be inhibited by other neuromodulators although this has yet to be tested. The inhibition of SK channels leads to the gating of LTP induction. The second is reconfiguration of interneuron activity and inhibitory transmission thereby differentially altering excitability and activation of NMDARs along dendrites and across cell types. The complexity of the interneuron network makes unravelling the impact of neuromodulators difficult, but data is emerging for several of the neuromodulators considered here [23,52,67] and it will be interesting to see how the reconfiguration of hippocampal interneuron networks modulates LTP and LTD. The third is the role played by astrocytes potentially coupling circadian rhythms of neuromodulator release to induction of synaptic plasticity. This has been demonstrated for acetylcholine [22°] and there is good evidence for excitation of astrocytes by noradrenergic, dopaminergic and 5-HT receptors [68,71,72]. Another common feature of the neuromodulators is the facilitation of both LTP and LTD (although often at different synapses) which may reflect the importance of both forms of plasticity for memory formation and consolidation.

Three divergent mechanisms are also highlighted (Figure 2). Firstly, the common mechanism of reconfiguring interneuron networks and inhibitory transmission may vary considerably in the details. For example, acetylcholine excites somatostatin and parvalbumin expressing oriens-lacunosum-moleculare interneurons decreasing distal dendritic excitability in CA1 pyramidal neurons [23] whereas serotonin excites cholecystokinin expressing interneurons at the stratum radiatum/lacunosum moleculare border causing widespread inhibition [67]. Secondly, although not described in this review, the precise hippocampal regions and synapses for each neuromodulatory mechanism vary considerably. Interestingly, this may mirror the differential innervation of hippocampal regions by neuromodulator projections both in terms of subregions (e.g. dentate gyrus, CA3, CA1) and the longitudinal axis of the hippocampus dorsal to ventral where these regions perform distinct tasks and are differentially innervated by neuromodulators (Figure 1). As an example, the expression of SK channels in CA1 pyramidal cells (and

therefore the regulation of NMDARs and threshold for LTP induction) is higher in the ventral than dorsal hippocampus [73**] providing a mechanism for differential control of LTP by neuromodulators along the dorsalventral axis. Finally, as well as spatial differentiation, there is evidence that the different neuromodulator systems may vary in their ability to facilitate plasticity on a delayed timescale. Dopamine [54,59,60] and to a lesser extent noradrenaline [42-44] can act on delayed timescales providing a retroactive mechanism for facilitating synaptic plasticity which has also been demonstrated at synapses in the neocortex and striatum [74–76]. Conversely, acetylcholine and serotonin act principally on shorter timescales to facilitate induction of plasticity. This implies that acetylcholine and serotonin play a greater role signalling expected uncertainty and create a prolonged state of arousal during which plasticity is facilitated. Conversely, dopamine and noradrenaline signal unexpected uncertainty and create a transient arousal that signals retroactively to facilitate plasticity in response to recent events [1,2]. These considerations raise the intriguing question of how separate neuromodulator systems interact to control plasticity [29**].

Acknowledgements

The authors would like to acknowledge helpful discussions with D Bannerman, R Moran, M Walton and members of the Mellor group. Supported by the Wellcome Trust and BBSRC.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as

- of outstanding interest
- Yu AJ, Dayan P: Uncertainty, neuromodulation, and attention. Neuron 2005, 46:681-692
- Parr T, Friston KJ: Uncertainty, epistemics and active inference. J R Soc Interf 2017, 14.
- Hasselmo ME: The role of acetylcholine in learning and memory. Curr Opin Neurobiol 2006, 16:710-715
- Fremaux N, Gerstner W: Neuromodulated spike-timingdependent plasticity, and theory of three-factor learning rules. Front Neural Circuits 2015, 9:85.
- Sajikumar S, Frey JU: Late-associativity, synaptic tagging, and the role of dopamine during LTP and LTD. Neurobiol Learn Mem 2004, 82:12-25
- Lisman J, Grace AA, Duzel E: A neoHebbian framework for episodic memory; role of dopamine-dependent late LTP. Trends Neurosci 2011, 34:536-547.
- Teles-Grilo Ruivo LM, Mellor JR: Cholinergic modulation of hippocampal network function. Front Synaptic Neurosci 2013,
- O'Dell TJ, Connor SA, Guglietta R, Nguyen PV: beta-Adrenergic receptor signaling and modulation of long-term potentiation in the mammalian hippocampus. Learn Mem 2015, 22:461-471.
- Hagena H, Manahan-Vaughan D: The serotonergic 5-HT4 receptor: a unique modulator of hippocampal synaptic information processing and cognition. Neurobiol Learn Mem 2017, **138**:145-153.

- 10. Edelmann E, Lessmann V: Dopaminergic innervation and modulation of hippocampal networks. Cell Tissue Res 2018, 373
- 11. Teles-Grilo Ruivo LM, Baker KL, Conway MW, Kinsley PJ, Gilmour G, Phillips KG, Isaac JT, Lowry JP, Mellor JR: Coordinated acetylcholine release in prefrontal cortex and hippocampus is associated with arousal and reward on distinct timescales. Cell Rep 2017, 18:905-917.
- 12. Hangya B, Ranade SP, Lorenc M, Kepecs A: Central cholinergic neurons are rapidly recruited by reinforcement feedback. Cell 2015, 162:1155-1168.
- 13. Petrovic MM, Nowacki J, Olivo V, Tsaneva-Atanasova K Randall AD. Mellor JR: Inhibition of post-synaptic Kv7/KCNQ/M channels facilitates long-term potentiation in the hippocampus. PLoS One 2012. 7:e30402.
- 14. Losonczy A, Makara JK, Magee JC: Compartmentalized dendritic plasticity and input feature storage in neurons. Nature 2008, 452:436-440.
- Giessel AJ, Sabatini BL: M1 muscarinic receptors boost synaptic potentials and calcium influx in dendritic spines by inhibiting postsynaptic SK channels. Neuron 2010, 68:936-947.
- 16. Buchanan KA, Petrovic MM, Chamberlain SE, Marrion NV, Mellor JR: Facilitation of long-term potentiation by muscarinic M(1) receptors is mediated by inhibition of SK channels. Neuron 2010, 68:948-963.
- 17. Isaac JT, Buchanan KA, Muller RU, Mellor JR: Hippocampal place cell firing patterns can induce long-term synaptic plasticity in vitro. J Neurosci 2009, 29:6840-6850.
- 18. Dennis SH, Pasqui F, Colvin EM, Sanger H, Mogg AJ, Felder CC, Broad LM, Fitzjohn SM, Isaac JT, Mellor JR: Activation of muscarinic M1 acetylcholine receptors induces long-term potentiation in the hippocampus. Cereb Cortex 2016, 26:414-
- 19. Fernandez de Sevilla D, Buno W: The muscarinic long-term enhancement of NMDA and AMPA receptor-mediated transmission at Schaffer collateral synapses develop through different intracellular mechanisms. J Neurosci 2010, 30:11032-
- 20. Li S. Nai Q. Lipina TV. Roder JC. Liu F: alpha7nAchR/NMDAR coupling affects NMDAR function and object recognition. Mol Brain 2013, 6:58.
- 21. Jo J, Son GH, Winters BL, Kim MJ, Whitcomb DJ, Dickinson BA, Lee YB, Futai K, Amici M, Sheng M et al.: Muscarinic receptors induce LTD of NMDAR EPSCs via a mechanism involving hippocalcin, AP2 and PSD-95. Nat Neurosci 2010, 13:1216-1224.
- 22. Papouin T, Dunphy JM, Tolman M, Dineley KT, Haydon PG: Septal cholinergic neuromodulation tunes the astrocyte-dependent gating of hippocampal NMDA receptors to wakefulness. Neuron 2017, 94:840-854 e847.

Using transgenic manipulation of astrocytes, this paper reveals a novel mechanism for neuromodulation of synaptic plasticity where cholinergic transmission tunes the gating of NMDARs through astrocyte-dependent release of D-serine.

- Haam J, Zhou J, Cui G, Yakel JL: Septal cholinergic neurons gate hippocampal output to entorhinal cortex via oriens lacunosum moleculare interneurons. Proc Natl Acad Sci U S A 2018. 115:F1886-F1895.
- 24. Pabst M, Braganza O, Dannenberg H, Hu W, Pothmann L, Rosen J, Mody I, van Loo K, Deisseroth K, Becker AJ et al.: Astrocyte intermediaries of septal cholinergic modulation in the hippocampus. Neuron 2016, 90:853-865.
- Tang AH, Karson MA, Nagode DA, McIntosh JM, Uebele VN, Renger JJ, Klugmann M, Milner TA, Alger BE: Nerve terminal nicotinic acetylcholine receptors initiate quantal GABA release from perisomatic interneurons by activating axonal Ttype (Cav3) Ca(2)(+) channels and Ca(2)(+) release from stores. *J Neurosci* 2011, **31**:13546-13561.
- 26. Szabo GG, Holderith N, Gulyas Al, Freund TF, Hajos N: Distinct synaptic properties of perisomatic inhibitory cell types and their different modulation by cholinergic receptor activation in

- the CA3 region of the mouse hippocampus. Eur J Neurosci 2010. 31:2234-2246.
- Dominguez S, Fernandez de Sevilla D, Buno W: Postsynaptic activity reverses the sign of the acetylcholine-induced longterm plasticity of GABAA inhibition. Proc Natl Acad Sci U S A 2014, 111:E2741-2750.
- Gu Z, Yakel JL: Timing-dependent septal cholinergic induction of dynamic hippocampal synaptic plasticity. Neuron 2011, 71:155-165
- Brzosko Z, Zannone S, Schultz W, Clopath C, Paulsen O:
 Sequential neuromodulation of Hebbian plasticity offers mechanism for effective reward-based navigation. Elife 2017,

Combining slice electrophysiology with computational modelling, this paper shows how delayed combinations of synaptic plasticity induction with activation of cholinergic and dopaminergic receptors can modulate the expression of long-term synaptic plasticity.

- Sugisaki E, Fukushima Y, Tsukada M, Aihara T: Cholinergic modulation on spike timing-dependent plasticity in hippocampal CA1 network. Neuroscience 2011, 192:91-101.
- 31. Sara SJ: The locus coeruleus and noradrenergic modulation of cognition. *Nat Rev Neurosci* 2009, **10**:211-223.
- 32. Murchison CF, Zhang XY, Zhang WP, Ouyang M, Lee A, Thomas SA: A distinct role for norepinephrine in memory retrieval. *Cell* 2004, **117**:131-143.
- Liu Y, Cui L, Schwarz MK, Dong Y, Schluter OM: Adrenergic gate release for spike timing-dependent synaptic potentiation. Neuron 2017, 93:394-408.
- Qian H, Matt L, Zhang M, Nguyen M, Patriarchi T, Koval OM, Anderson ME, He K, Lee HK, Hell JW: beta2-Adrenergic receptor supports prolonged theta tetanus-induced LTP. J Neurophysiol 2012, 107:2703-2712.
- Lin YW, Min MY, Chiu TH, Yang HW: Enhancement of associative long-term potentiation by activation of betaadrenergic receptors at CA1 synapses in rat hippocampal slices. J Neurosci 2003, 23:4173-4181.
- Hansen N, Manahan-Vaughan D: Hippocampal long-term potentiation that is elicited by perforant path stimulation or that occurs in conjunction with spatial learning is tightly controlled by beta-adrenoreceptors and the locus coeruleus. Hippocampus 2015, 25:1285-1298.
- Hansen N, Manahan-Vaughan D: Locus coeruleus stimulation facilitates long-term depression in the dentate gyrus that requires activation of beta-adrenergic receptors. Cereb Cortex 2015, 25:1889-1896.
- 38. Takeuchi T, Duszkiewicz AJ, Sonneborn A, Spooner PA,
- Yamasaki M, Watanabe M, Smith CC, Fernandez G, Deisseroth K, Greene RW et al.: Locus coeruleus and dopaminergic consolidation of everyday memory. Nature 2016, 537:357-362.

Using optogenetic control of LC neurons and pharmacology, this paper argues that dopamine and not noradrenaline release from LC neurons controls LTP facilitation and memory formation in the hippocampus proposing a critical role for neuromodulator co-release. Similar results are shown in Refs. [40,41].

- Yuan LL, Adams JP, Swank M, Sweatt JD, Johnston D: Protein kinase modulation of dendritic K+ channels in hippocampus involves a mitogen-activated protein kinase pathway. J Neurosci 2002, 22:4860-4868.
- Faber ES, Delaney AJ, Power JM, Sedlak PL, Crane JW, Sah P: Modulation of SK channel trafficking by beta adrenoceptors enhances excitatory synaptic transmission and plasticity in the amygdala. J Neurosci 2008, 28:10803-10813.
- Murphy JA, Stein IS, Lau CG, Peixoto RT, Aman TK, Kaneko N, Aromolaran K, Saulnier JL, Popescu GK, Sabatini BL et al.: Phosphorylation of Ser1166 on GluN2B by PKA is critical to synaptic NMDA receptor function and Ca2+ signaling in spines. J Neurosci 2014, 34:869-879.
- 42. Hu H, Real E, Takamiya K, Kang MG, Ledoux J, Huganir RL, Malinow R: Emotion enhances learning via norepinephrine

- regulation of AMPA-receptor trafficking. Cell 2007, 131:160-173.
- Joiner ML, Lise MF, Yuen EY, Kam AY, Zhang M, Hall DD, Malik ZA, Qian H, Chen Y, Ulrich JD et al.: Assembly of a beta2adrenergic receptor — GluR1 signalling complex for localized cAMP signalling. EMBO J 2010, 29:482-495.
- Tenorio G, Connor SA, Guevremont D, Abraham WC, Williams J, O'Dell TJ, Nguyen PV: 'Silent' priming of translation-dependent LTP by ss-adrenergic receptors involves phosphorylation and recruitment of AMPA receptors. Learn Mem 2010, 17:627-638.
- Wagatsuma A, Okuyama T, Sun C, Smith LM, Abe K, Tonegawa S: Locus coeruleus input to hippocampal CA3 drives single-trial learning of a novel context. Proc Natl Acad Sci U S A 2018, 115: E310-E316.
- Kempadoo KA, Mosharov EV, Choi SJ, Sulzer D, Kandel ER: Dopamine release from the locus coeruleus to the dorsal hippocampus promotes spatial learning and memory. Proc Natl Acad Sci U S A 2016, 113:14835-14840.
- Howe MW, Dombeck DA: Rapid signalling in distinct dopaminergic axons during locomotion and reward. Nature 2016, 535:505-510.
- McNamara CG, Tejero-Cantero A, Trouche S, Campo-Urriza N, Dupret D: Dopaminergic neurons promote hippocampal reactivation and spatial memory persistence. Nat Neurosci 2014. 17:1658-1660.
- 49. Rosen ZB, Cheung S, Siegelbaum SA: Midbrain dopamine
 neurons bidirectionally regulate CA3-CA1 synaptic drive. Nat Neurosci 2015, 18:1763-1771.

In this paper, exogenous pharmacological activation of dopamine receptors is shown to have a different effect to optogenetically stimulated endogenous dopamine release on synaptic transmission in the hippocampus.

- Wei X, Ma T, Cheng Y, Huang CCY, Wang X, Lu J, Wang J: Dopamine D1 or D2 receptor-expressing neurons in the central nervous system. Addict Biol 2018, 23:569-584.
- Perreault ML, Hasbi A, O'Dowd BF, George SR: Heteromeric dopamine receptor signaling complexes: emerging neurobiology and disease relevance. Neuropsychopharmacology 2014, 39:156-168.
- Hammad H, Wagner JJ: Dopamine-mediated disinhibition in the CA1 region of rat hippocampus via D3 receptor activation. J Pharmacol Exp Ther 2006, 316:113-120.
- Varela JA, Hirsch SJ, Chapman D, Leverich LS, Greene RW: D1/D5 modulation of synaptic NMDA receptor currents. J Neurosci 2009, 29:3109-3119.
- Brzosko Z, Schultz W, Paulsen O: Retroactive modulation of spike timing-dependent plasticity by dopamine. Elife 2015, 4.
- Yang K, Dani JA: Dopamine D1 and D5 receptors modulate spike timing-dependent plasticity at medial perforant path to dentate granule cell synapses. J Neurosci 2014, 34:15888-15907
- Edelmann E, Lessmann V: Dopamine modulates spike timingdependent plasticity and action potential properties in CA1 pyramidal neurons of acute rat hippocampal slices. Front Synaptic Neurosci 2011, 3:6.
- Zhang JC, Lau PM, Bi GQ: Gain in sensitivity and loss in temporal contrast of STDP by dopaminergic modulation at hippocampal synapses. Proc Natl Acad Sci U S A 2009, 106:13028-13033.
- Lemon N, Manahan-Vaughan D: Dopamine D1/D5 receptors gate the acquisition of novel information through hippocampal long-term potentiation and long-term depression. J Neurosci 2006, 26:7723-7729.
- Moncada D: Evidence of VTA and LC control of protein synthesis required for the behavioral tagging process. Neurobiol Learn Mem 2017, 138:226-237.
- Karunakaran S, Chowdhury A, Donato F, Quairiaux C, Michel CM, Caroni P: PV plasticity sustained through D1/5 dopamine

- signaling required for long-term memory consolidation. Nat Neurosci 2016, 19:454-464
- 61. Matias S, Lottem E, Duque GP, Mainen ZF: Activity patterns of serotonin neurons underlying cognitive flexibility. Flife 2017, 6.
- 62. Celada P, Puig MV, Artigas F: Serotonin modulation of cortical neurons and networks. Front Integr Neurosci 2013, 7:25
- 63. Vasefi MS, Yang K, Li J, Kruk JS, Heikkila JJ, Jackson MF MacDonald JF, Beazely MA: Acute 5-HT7 receptor activation increases NMDA-evoked currents and differentially alters NMDA receptor subunit phosphorylation and trafficking in hippocampal neurons. Mol Brain 2013, 6:24.
- 64. Cai X, Kallarackal AJ, Kvarta MD, Goluskin S, Gaylor K, Bailey AM, Lee HK, Huganir RL, Thompson SM: Local potentiation of excitatory synapses by serotonin and its alteration in rodent models of depression. Nat Neurosci 2013. 16:464-472.
- 65. Teixeira CM, Rosen ZB, Suri D, Sun Q, Hersh M, Sargin D,
 Dincheva I, Morgan AA, Spivack S, Krok AC et al.: Hippocampal 5-HT input regulates memory formation and schaffer collateral excitation. Neuron 2018, 98:992-1004 e1004

This paper shows that optogenetic manipulation of serotonin input to hippocampus controls synaptic transmission in CA1 and memory formation.

- Kemp A, Manahan-Vaughan D: The 5-hydroxytryptamine4 receptor exhibits frequency-dependent properties in synaptic plasticity and behavioural metaplasticity in the hippocampal CA1 region in vivo. Cereb Cortex 2005, 15:1037-1043.
- Varga V, Losonczy A, Zemelman BV, Borhegyi Z, Nyiri G Domonkos A, Hangya B, Holderith N, Magee JC, Freund TF: Fast synaptic subcortical control of hippocampal circuits. Science 2009, 326:449-453.
- 68. Zhang S, Li B, Lovatt D, Xu J, Song D, Goldman SA, Nedergaard M, Hertz L, Peng L: 5-HT2B receptors are expressed on astrocytes from brain and in culture and are a chronic target for all five conventional 'serotonin-specific reuptake inhibitors'. Neuron Glia Biol 2010, 6:113-125.
- 69. Twarkowski H, Hagena H, Manahan-Vaughan D: The 5hydroxytryptamine4 receptor enables differentiation of informational content and encoding in the hippocampus. Hippocampus 2016, 26:875-891.

- 70. Tigaret CM, Olivo V, Sadowski JH, Ashby MC, Mellor JR: Coordinated activation of distinct Ca(2+) sources and metabotropic glutamate receptors encodes Hebbian synaptic plasticity. Nat Commun 2016, 7:10289.
- 71. Jennings A, Tyurikova O, Bard L, Zheng K, Semyanov A Henneberger C, Rusakov DA: Dopamine elevates and lowers astroglial Ca(2+) through distinct pathways depending on local synaptic circuitry. Glia 2017, 65:447-459.
- Carriba P, Pardo L, Parra-Damas A, Lichtenstein MP, Saura CA, Pujol A, Masgrau R, Galea E: ATP and noradrenaline activate CREB in astrocytes via noncanonical Ca(2+) and cyclic AMP independent pathways. Glia 2012, 60:1330-1344.
- 73. Babiec WE, Jami SA, Guglietta R, Chen PB, O'Dell TJ: Differential regulation of NMDA receptor-mediated transmission by SK channels underlies dorsal-ventral differences in dynamics of Schaffer collateral synaptic function. J Neurosci 2017, 37:1950-

In this paper, SK channels are shown to be pivotal for the induction of NMDAR-dependent plasticity, but the degree of control varies with SK channel expression along the dorsal-ventral axis.

- Yagishita S, Hayashi-Takagi A, Ellis-Davies GC, Urakubo H, Ishii S, Kasai H: A critical time window for dopamine actions on the structural plasticity of dendritic spines. Science 2014, **345**:1616-1620.
- 75. Shindou T, Shindou M, Watanabe S, Wickens J: A silent eligibility trace enables dopamine-dependent synaptic plasticity for reinforcement learning in the mouse striatum. Eur J Neurosci 2018 http://dx.doi.org/10.1111/ejn.13921. [Epub ahead of print].
- 76. He K, Huertas M, Hong SZ, Tie X, Hell JW, Shouval H, Kirkwood A: Distinct eligibility traces for LTP and LTD in cortical synapses. Neuron 2015. 88:528-538.
- 77. Yi F, Catudio-Garrett E, Gabriel R, Wilhelm M, Erdelyi F, Szabo G, Deisseroth K, Lawrence J: Hippocampal "cholinergic interneurons" visualized with the choline acetyltransferase promoter: anatomical distribution, intrinsic membrane properties, neurochemical characteristics, and capacity for cholinergic modulation. Front Synaptic Neurosci 2015, 7:4