

An Introduction to Machine Learning

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Contents

Chapter 1

About the course

1.1 Overview

Machine learning gives computers the ability to learn without being explicitly programmed. It encompasses a broad range of approaches to data analysis with applicability across the biological sciences. Lectures will introduce commonly used algorithms and provide insight into their theoretical underpinnings. In the practicals students will apply these algorithms to real biological data-sets using the R language and environment.

During this course you will learn about:

- Some of the core mathematical concepts underpinning machine learning algorithms: matrices and linear algebra; Bayes' theorem.
- Classification (supervised learning): partitioning data into training and test sets; feature selection; logistic regression; support vector machines; artificial neural networks; decision trees; nearest neighbours, cross-validation.
- Exploratory data analysis (unsupervised learning): dimensionality reduction, anomaly detection, clustering.

After this course you should be able to:

- Understand the concepts of machine learning.
- Understand the strengths and limitations of the various machine learning algorithms presented in this course.
- Select appropriate machine learning methods for your data.
- Perform machine learning in R.

1.2 Registration

Bioinformatics Training: An Introduction to Machine Learning

1.3 Prerequisites

- Some familiarity with R would be helpful.
- For an introduction to R see An Introduction to Solving Biological Problems with R course.

1.4 Github

bioinformatics-training/intro-machine-learning

1.5 License

GPL-3

1.6 Contact

If you have any **comments**, **questions** or **suggestions** about the material, please contact the authors: Sudhakaran Prabakaran, Matt Wayland and Chris Penfold.

1.7 Colophon

This book was produced using the **bookdown** package (?), which was built on top of R Markdown and **knitr** (?).

Chapter 2

Introduction

You can label chapter and section titles using `{#label}` after them, e.g., we can reference Chapter 2. If you do not manually label them, there will be automatic labels anyway, e.g., Chapter ??.

Figures and tables with captions will be placed in `figure` and `table` environments, respectively.

```
par(mar = c(4, 4, .1, .1))  
plot(pressure, type = 'b', pch = 19)
```

Reference a figure by its code chunk label with the `fig:` prefix, e.g., see Figure 2.1. Similarly, you can reference tables generated from `knitr::kable()`, e.g., see Table 2.1.

```
knitr::kable(  
  head(iris, 20), caption = 'Here is a nice table!',  
  booktabs = TRUE  
)
```



Figure 2.1: Here is a nice figure!

Table 2.1: Here is a nice table!

Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
5.1	3.5	1.4	0.2	setosa
4.9	3.0	1.4	0.2	setosa
4.7	3.2	1.3	0.2	setosa
4.6	3.1	1.5	0.2	setosa
5.0	3.6	1.4	0.2	setosa
5.4	3.9	1.7	0.4	setosa
4.6	3.4	1.4	0.3	setosa
5.0	3.4	1.5	0.2	setosa
4.4	2.9	1.4	0.2	setosa
4.9	3.1	1.5	0.1	setosa
5.4	3.7	1.5	0.2	setosa
4.8	3.4	1.6	0.2	setosa
4.8	3.0	1.4	0.1	setosa
4.3	3.0	1.1	0.1	setosa
5.8	4.0	1.2	0.2	setosa
5.7	4.4	1.5	0.4	setosa
5.4	3.9	1.3	0.4	setosa
5.1	3.5	1.4	0.3	setosa
5.7	3.8	1.7	0.3	setosa
5.1	3.8	1.5	0.3	setosa

Chapter 3

Linear models and matrix algebra

3.1 Exercises

Solutions to exercises can be found in appendix ??

Chapter 4

Linear and non linear logistic regression

4.1 Exercises

Solutions to exercises can be found in appendix ??.

Chapter 5

Nearest neighbours

5.1 Example one

5.2 Example two

5.3 Exercises

Solutions to exercises can be found in appendix ??.

Chapter 6

Decision trees and random forests

6.1 Exercises

Solutions to exercises can be found in appendix ??.

Chapter 7

Support vector machines

7.1 Exercises

Solutions to exercises can be found in appendix ??

Chapter 8

Artificial neural networks

8.1 Exercises

Solutions to exercises can be found in appendix ??.

Chapter 9

Dimensionality reduction

9.1 Linear Dimensionality Reduction

9.1.1 Principle Component Analysis

9.1.2 Horeshoe effect

9.2 Nonlinear Dimensionality Reduction

9.2.1 t-SNE

9.2.2 Gaussian Process Latent Variable Models

9.2.3 GPLVMs with informative priors

9.3 Exercises

Solutions to exercises can be found in appendix ??.

Chapter 10

Clustering

10.1 Introduction

Hierarchic (produce dendrogram) vs partitioning methods

10.2 Distance metrics

Minkowski distance:

$$distance(x, y, p) = \left(\sum_{i=1}^n abs(x_i - y_i)^p \right)^{1/p} \quad (10.1)$$

Graphical explanation of euclidean, manhattan and max (Chebyshev?)

10.2.1 Image segmentation

10.3 Hierarchic methods

10.3.1 Linkage algorithms

Make one section panel of three dendrograms one table

Single linkage - nearest neighbours linkage Complete linkage - furthest neighbours linkage Average linkage - UPGMA (Unweighted Pair Group Method with Arithmetic Mean)

Table 10.1: Example distance matrix

	A	B	C	D
B	2			
C	6	5		
D	10	10	5	
E	9	8	3	4

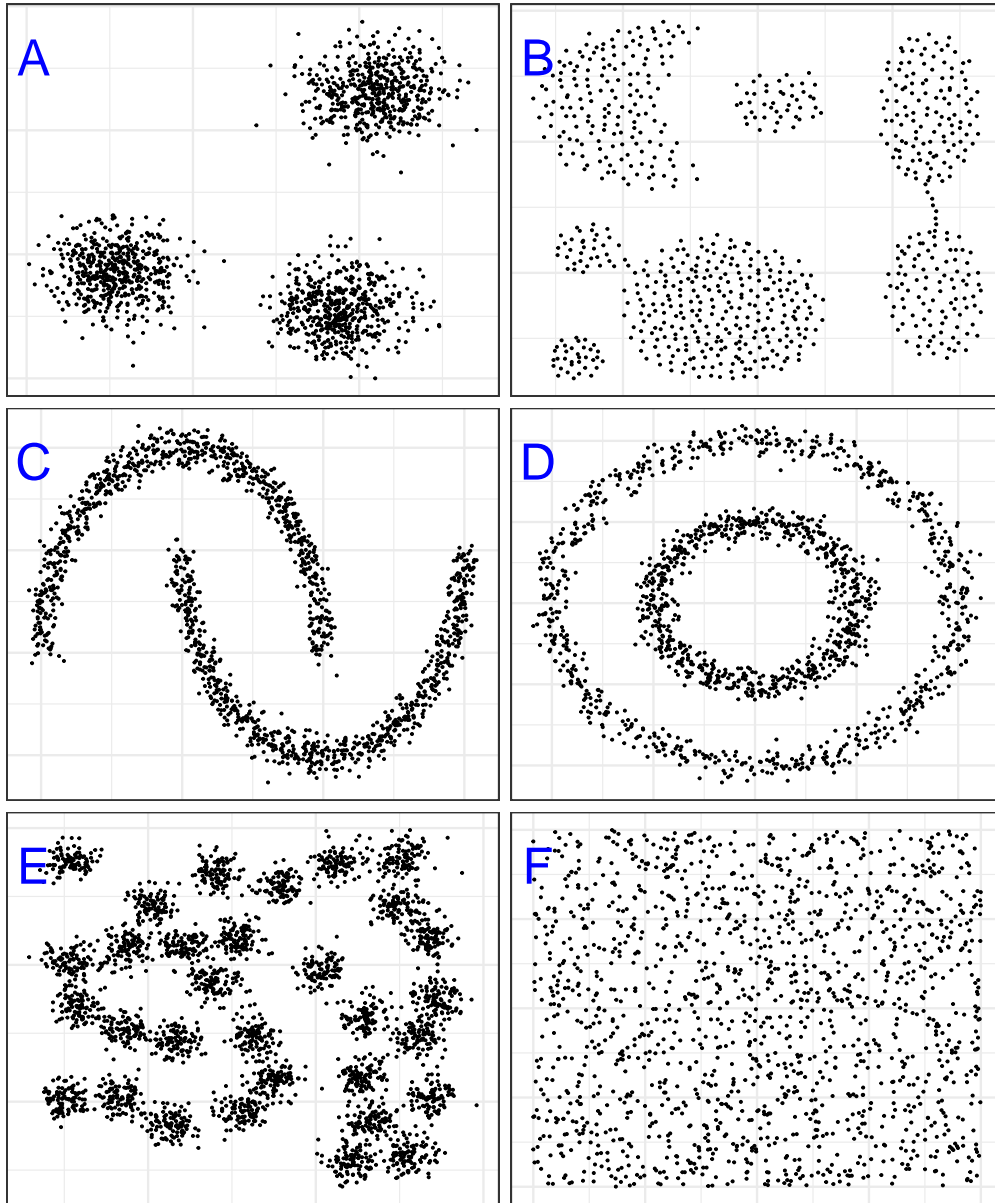
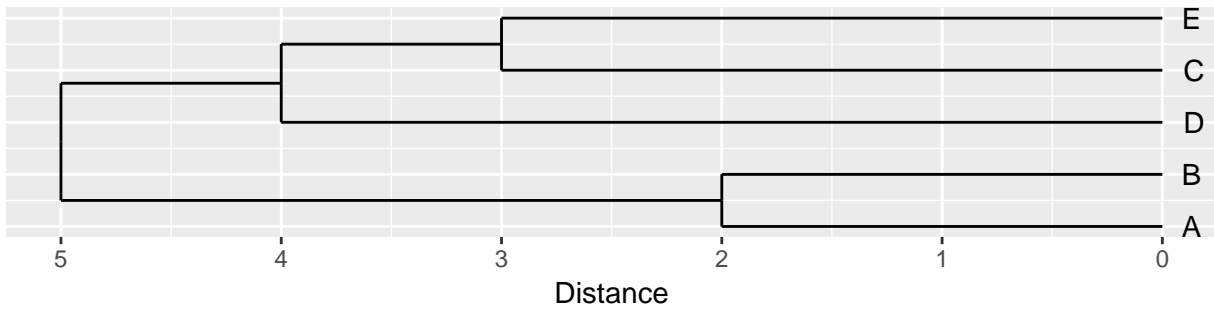


Figure 10.1: Example clusters. **A**, *blobs*; **B**, *aggregation* [Gionis2007]; **C**, *noisy moons*; **D**, *noisy circles*; **E**, *D31* [Veenman2002]; **F**, *no structure*.

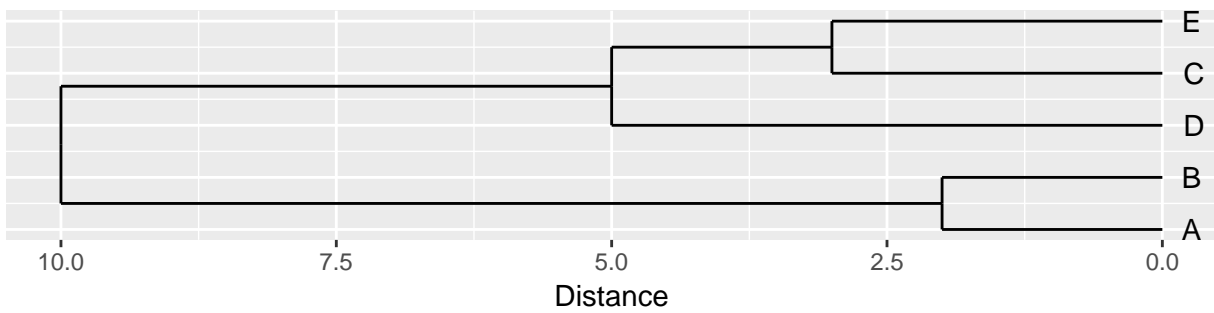
Table 10.2: Merge distances for objects in the example distance matrix using three different linkage methods.

Groups	Single	Complete	Average
A,B,C,D,E	0	0	0
(A,B),C,D,E	2	2	2
(A,B),(C,E),D	3	3	3
(A,B)(C,D,E)	4	5	4.5
(A,B,C,D,E)	5	10	8

Single linkage



Complete linkage



Average linkage

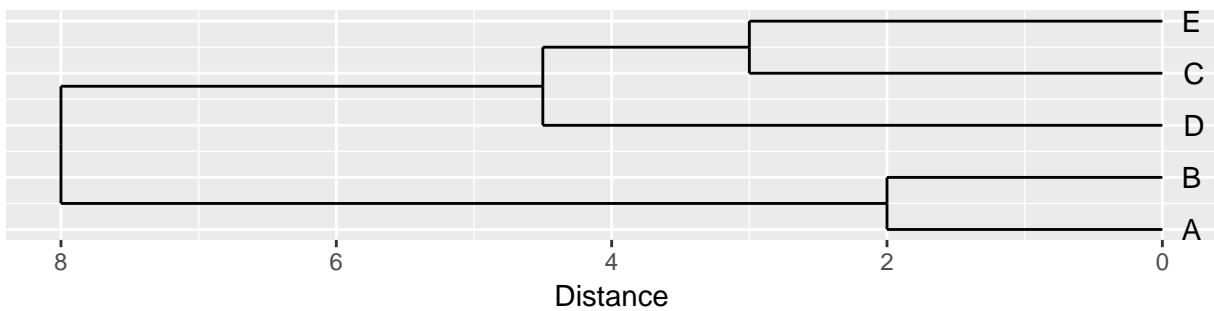


Figure 10.2: Dendrograms for the example distance matrix using three different linkage methods.

10.3.2 Example: clustering toy data sets

```

library(RColorBrewer)
library(dendextend)

##
## -----
## Welcome to dendextend version 1.5.2
## Type citation('dendextend') for how to cite the package.
##
## Type browseVignettes(package = 'dendextend') for the package vignette.
## The github page is: https://github.com/talgalili/dendextend/
##
## Suggestions and bug-reports can be submitted at: https://github.com/talgalili/dendextend/issues
## Or contact: <tal.galili@gmail.com>
##
## To suppress this message use: suppressPackageStartupMessages(library(dendextend))
## -----

##
## Attaching package: 'dendextend'

## The following object is masked from 'package:ggdendro':
##
##     theme_dendro

## The following object is masked from 'package:stats':
##
##     cutree

library(ggplot2)
library(GGally)

cluster_colours <- brewer.pal(8,"Dark2")

blobs <- read.csv("data/example_clusters/blobs.csv", header=F)

aggregation <- read.table("data/example_clusters/aggregation.txt")
noisy_moons <- read.csv("data/example_clusters/noisy_moons.csv", header=F)
noisy_circles <- read.csv("data/example_clusters/noisy_circles.csv", header=F)
no_structure <- read.csv("data/example_clusters/no_structure.csv", header=F)

hclust_plots <- function(data_set, n){
  d <- dist(data_set[,1:2])
  dend <- as.dendrogram(hclust(d, method="average"))
  clusters <- cutree(dend,n,order_clusters_as_data=F)
  dend <- color_branches(dend, clusters=clusters, col=cluster_colours[1:n])
  clusters <- clusters[order(as.numeric(names(clusters)))]
  labels(dend) <- rep("", length(data_set[,1]))
  ggd <- as.ggdend(dend)
  ggd$nodes <- ggd$nodes[!(1:length(ggd$nodes[,1])),]
  plotPair <- list(ggplot(ggd),
    ggplot(data_set, aes(V1,V2)) + geom_point(col=cluster_colours[clusters], size=0.2))
  return(plotPair)
}

```

```

plotList <- c(
  hclust_plots(aggregation, 7),
  hclust_plots(noisy_moons, 2),
  hclust_plots(noisy_circles, 2),
  hclust_plots(no_structure, 3)
)

pm <- ggmatrix(
  plotList, nrow=4, ncol=2, showXAxisPlotLabels = F, showYAxisPlotLabels = F, xAxisLabels=c("dendrogram")
) + theme_bw()

pm

```

10.3.3 Example: gene expression profiling of human tissues

Load required libraries

```

library(RColorBrewer)
library(dendextend)

```

Load data

```
load("data/tissues_gene_expression/tissuesGeneExpression.rda")
```

Inspect data

```
table(tissue)
```

```
## tissue
## cerebellum      colon endometrium hippocampus      kidney      liver
##           38          34          15          31          39          26
## placenta
##           6
```

```
dim(e)
```

```
## [1] 22215  189
```

Compute distance between each sample

```
d <- dist(t(e))
```

perform hierarchical clustering

```

hc <- hclust(d, method="average")
plot(hc, labels=tissue, cex=0.5, hang=-1, xlab="", sub="")

```

use dendextend library to plot dendrogram with colour labels

```

tissue_type <- unique(tissue)
dend <- as.dendrogram(hc)
dend_colours <- brewer.pal(length(unique(tissue)), "Dark2")
names(dend_colours) <- tissue_type
labels(dend) <- tissue[order.dendrogram(dend)]
labels_colors(dend) <- dend_colours[tissue][order.dendrogram(dend)]
labels_cex(dend) = 0.5
plot(dend, horiz=T)

```

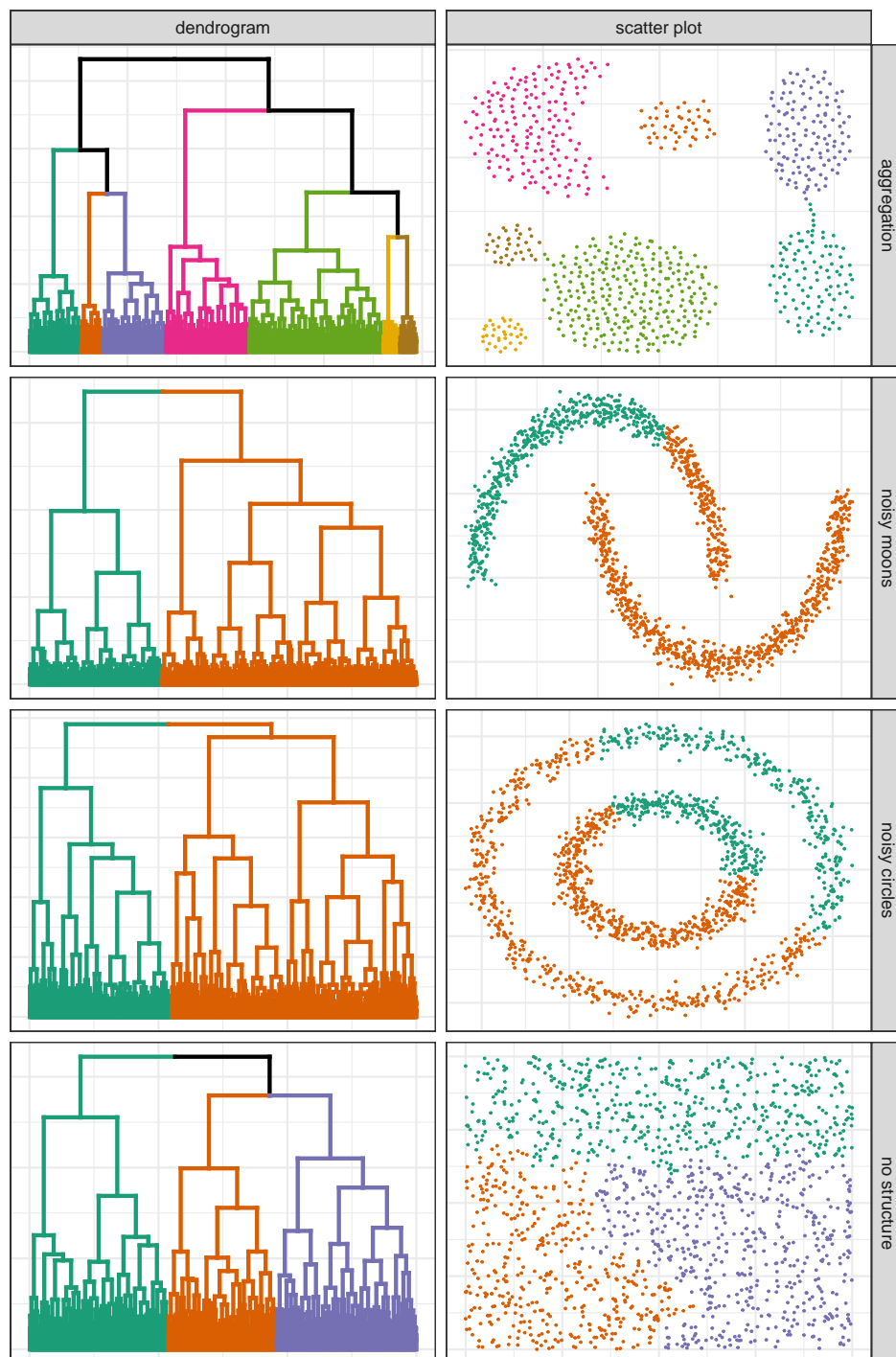


Figure 10.3: Hierarchical clustering of toy data-sets.

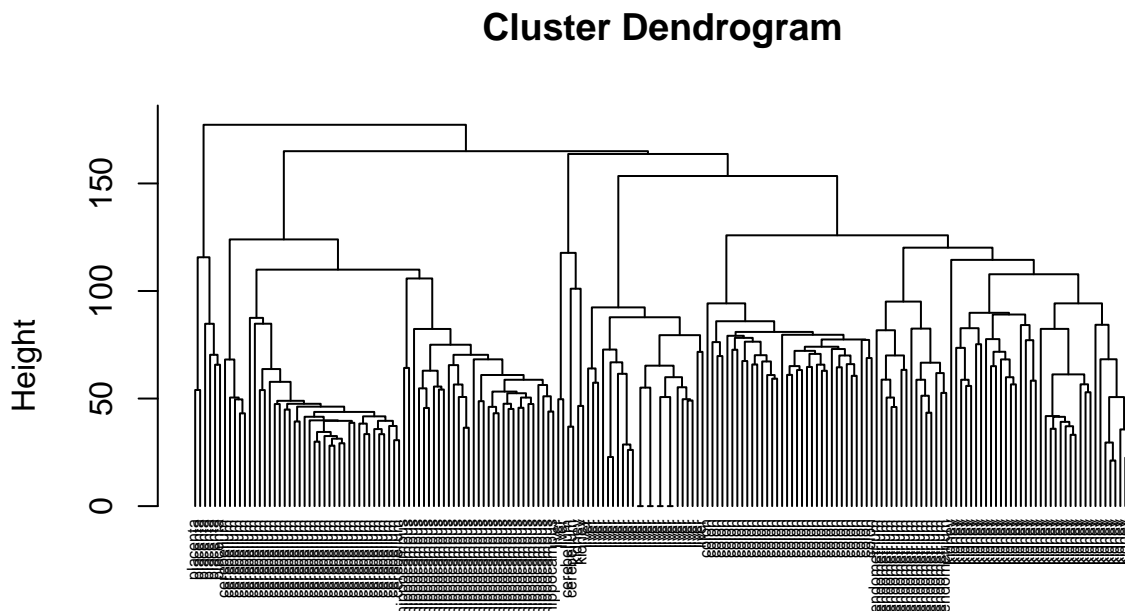


Figure 10.4: Clustering of tissue samples based on gene expression profiles.

Define clusters by cutting tree at a specific height

```
plot(dend, horiz=T)
abline(v=125, lwd=2, lty=2, col="blue")
```

```
hclusters <- cutree(dend, h=125)
table(tissue, cluster=hclusters)
```

```
##           cluster
## tissue      1  2  3  4  5  6
## cerebellum  0 36  0  0  2  0
## colon       0  0 34  0  0  0
## endometrium 15  0  0  0  0  0
## hippocampus  0 31  0  0  0  0
## kidney      37  0  0  0  2  0
## liver        0  0  0 24  2  0
## placenta     0  0  0  0  0  6
```

Select a specific number of clusters.

```
plot(dend, horiz=T)
abline(v = heights_per_k.dendrogram(dend)["8"], lwd = 2, lty = 2, col = "blue")
```

```
hclusters <- cutree(dend, k=8)
table(tissue, cluster=hclusters)
```

```
##           cluster
## tissue      1  2  3  4  5  6  7  8
## cerebellum  0 31  0  0  2  0  5  0
## colon       0  0 34  0  0  0  0  0
## endometrium  0  0  0  0  0 15  0  0
## hippocampus  0 31  0  0  0  0  0  0
## kidney      37  0  0  0  2  0  0  0
## liver        0  0  0 24  2  0  0  0
```

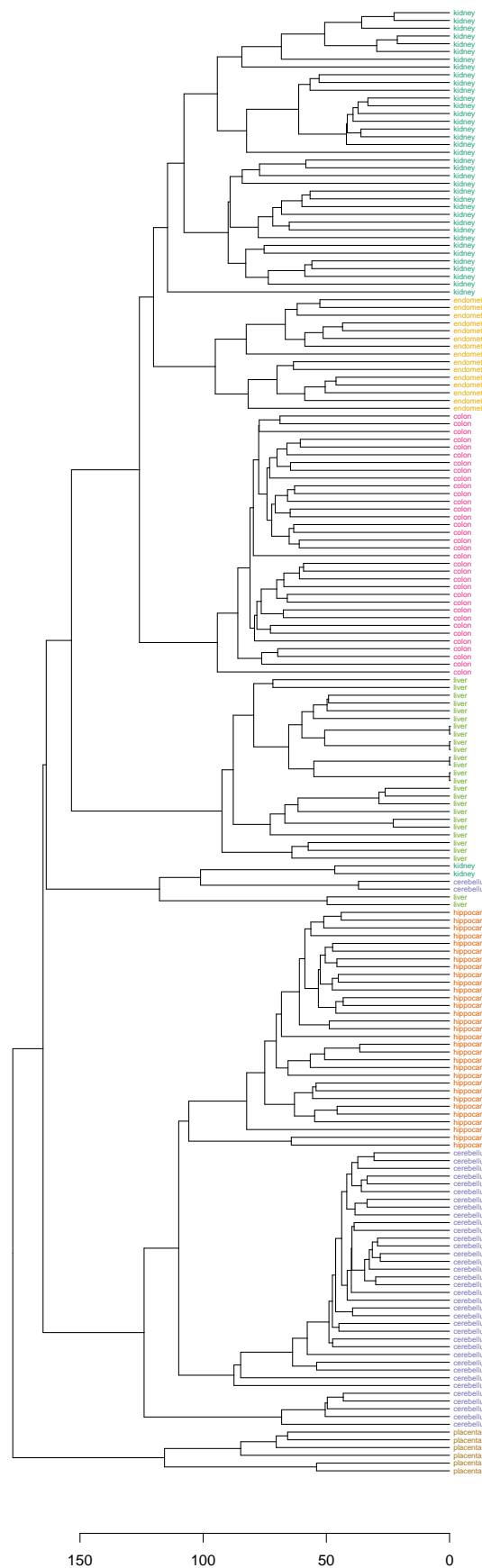


Figure 10.5: Clustering of tissue samples based on gene expression profiles with labels coloured by tissue type.

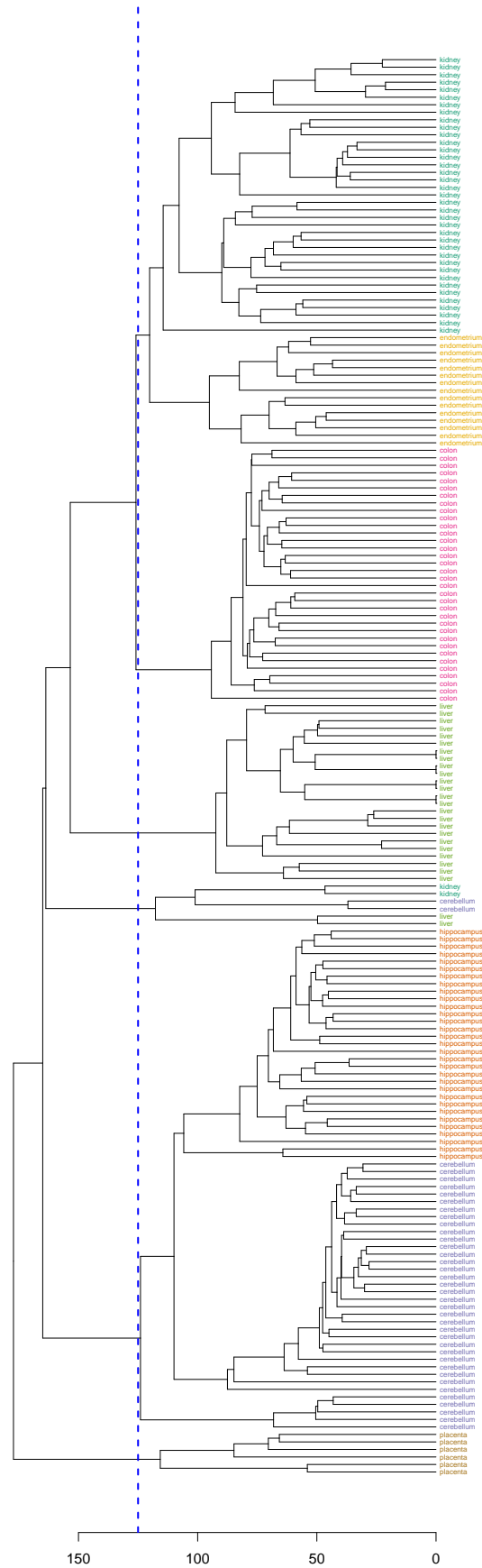


Figure 10.6: Clusters found by cutting tree at a height of 125

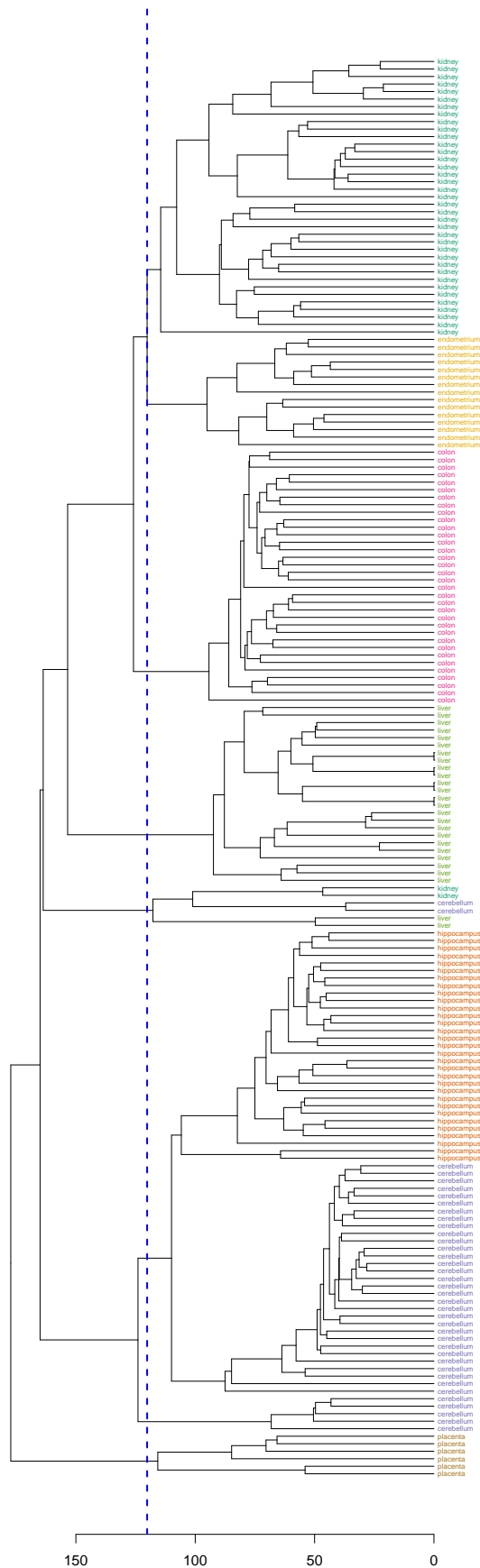


Figure 10.7: Selection of eight clusters from the dendrogram


```
## placenta      0  0  0  0  0  0  0  0  6
```

10.4 Partitioning methods

10.4.1 K-means

10.4.1.1 Algorithm

Pseudocode

to illustrate range of different types of data that can be clustered - image segmentation

The default setting of the **kmeans** function is to perform a maximum of 10 iterations and if the algorithm fails to converge a warning is issued. The maximum number of iterations is set with the argument **iter.max**.

10.4.1.2 Choosing initial cluster centres

```
library(RColorBrewer)
point_shapes <- c(15,17,19)
point_colours <- brewer.pal(3,"Dark2")
point_size = 1.5
center_point_size = 8

blobs <- as.data.frame(read.csv("data/example_clusters/blobs.csv", header=F))

good_centres <- as.data.frame(matrix(c(2,8,7,3,12,7), ncol=2, byrow=T))
bad_centres <- as.data.frame(matrix(c(13,13,8,12,2,2), ncol=2, byrow=T))

good_result <- kmeans(blobs[,1:2], centers=good_centres)
bad_result <- kmeans(blobs[,1:2], centers=bad_centres)

plotList <- list(
  ggplot(blobs, aes(V1,V2)) + geom_point(col=point_colours[good_result$cluster], shape=point_shapes[good_result$cluster], size=point_size),
  ggplot(blobs, aes(V1,V2)) + geom_point(col=point_colours[bad_result$cluster], shape=point_shapes[bad_result$cluster], size=point_size)
)

pm <- ggmatrix(
  plotList, nrow=1, ncol=2, showXAxisPlotLabels = T, showYAxisPlotLabels = T, xAxisLabels=c("A", "B")
) + theme_bw()

pm
```

Convergence to a local minimum can be avoided by starting the algorithm multiple times, with different random centres. The **nstart** argument to the **k-means** function can be used to specify the number of random sets and optimal solution will be selected automatically.

10.4.1.3 Choosing k

```
cluster_colours <- brewer.pal(9,"Set1")
k <- 1:9
res <- lapply(k, function(i){kmeans(blobs[,1:2], i, nstart=50)})
```

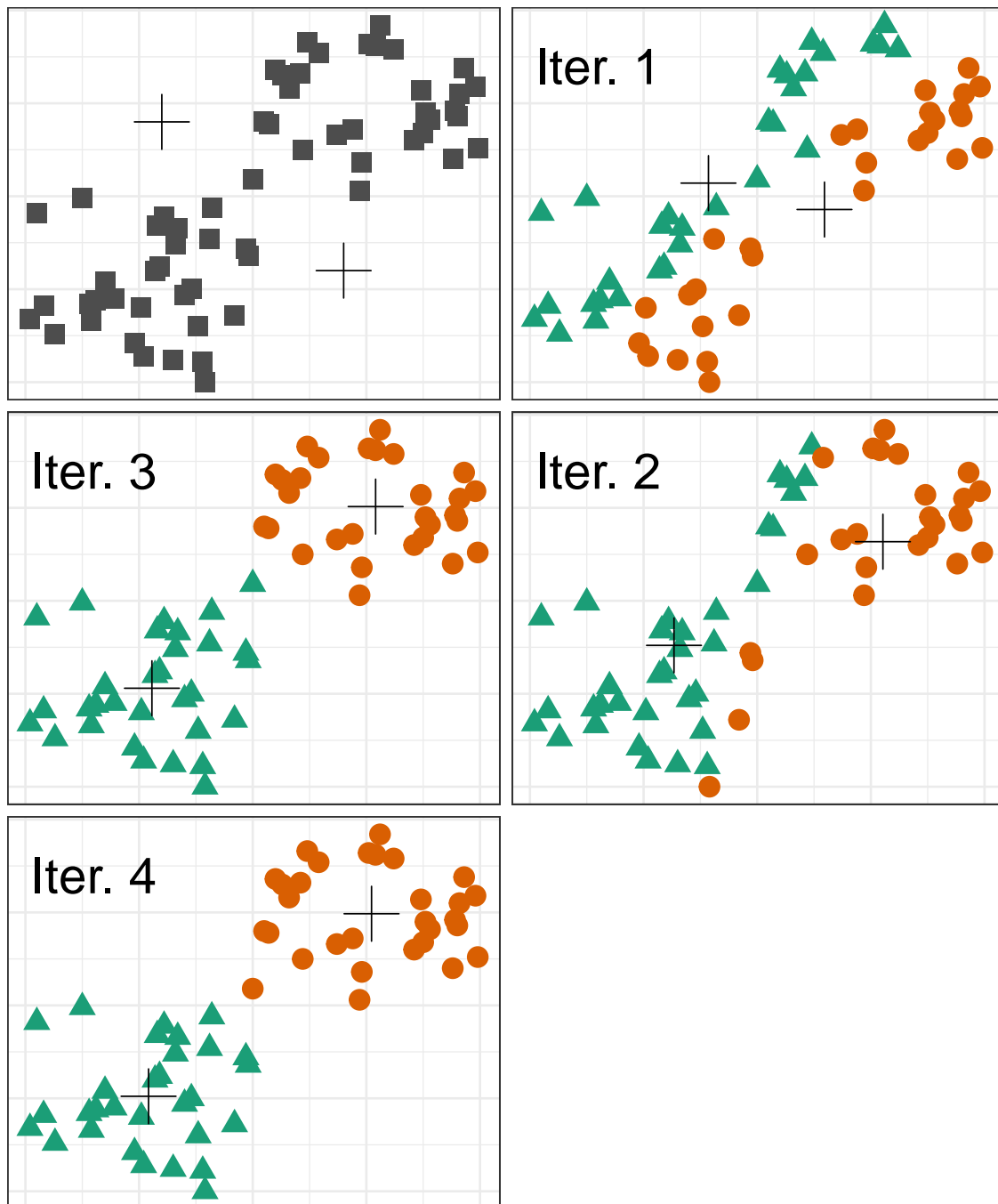


Figure 10.8: Iterations of the k-means algorithm

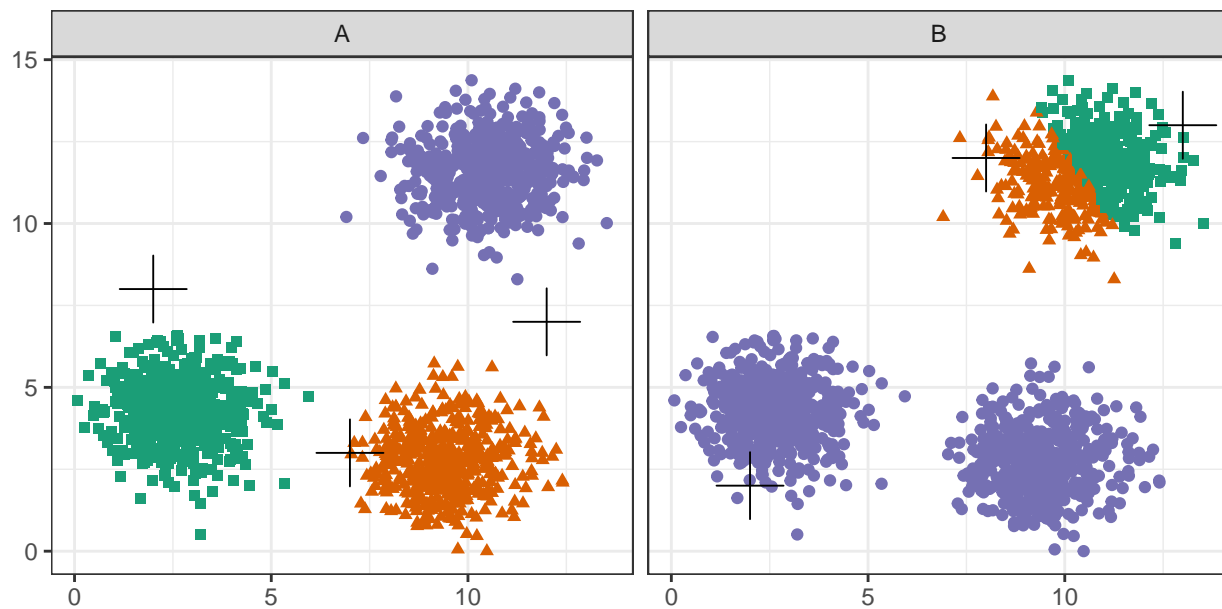


Figure 10.9: Initial centres determine clusters. The starting centres are shown as crosses. **A**, real clusters found; **B**, convergence to a local minimum.

```
plotList <- lapply(k, function(i){
  ggplot(blobs, aes(V1, V2)) +
    geom_point(col=cluster_colours[res[[i]]$cluster], size=1) +
    geom_point(data=as.data.frame(res[[i]]$centers), aes(V1,V2), shape=3, col="black", size=5) +
    annotate("text", x=2, y=13, label=paste("k=", i, sep=""), size=8, col="black") +
    theme_bw()
})

pm <- ggmatrix(
  plotList, nrow=3, ncol=3, showXAxisPlotLabels = T, showYAxisPlotLabels = T
) + theme_bw()

pm

tot_withinss <- sapply(k, function(i){res[[i]]$tot.withinss})
qplot(k, tot_withinss, geom=c("point", "line"), ylab="Total within-cluster sum of squares") + theme_bw()
```

N.B. we have set `nstart=50` so that the algorithm is started 50 times wi

10.4.2 DBSCAN

Density-based spatial clustering of applications with noise

10.4.2.1 Algorithm

Abstract DBSCAN algorithm in pseudocode (?):

```
1 Compute neighbours of each point and identify core points    // Identify core points
```

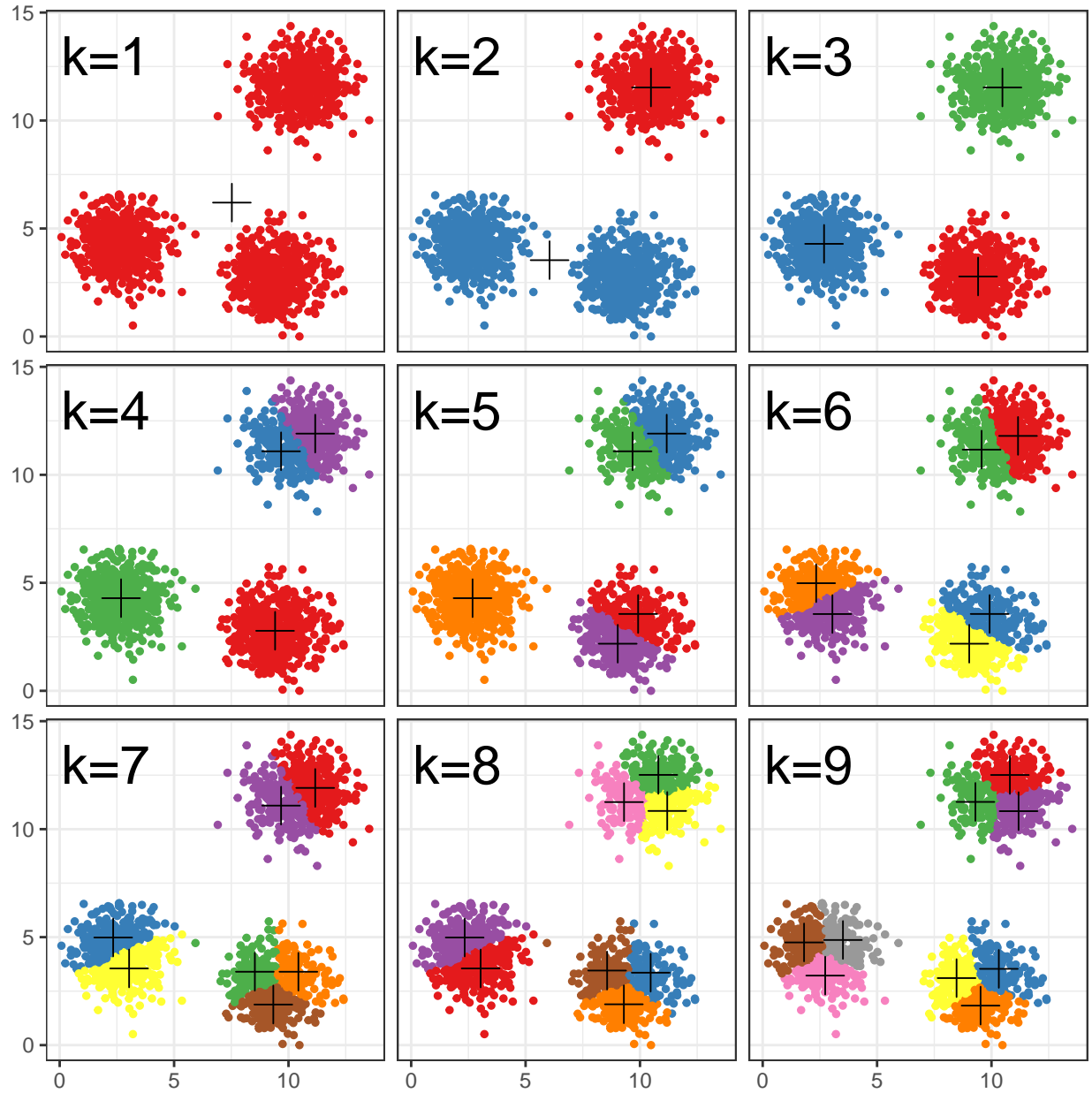


Figure 10.10: K-means clustering of the blobs data set using a range of values of k from 1-9. Cluster centres indicated with a cross.