CellMix FAQ and HowTos

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Abstract

This vignette contains hints and pointers on how to perform common tasks with the *CellMix* package. In particular, it will incorporate answers to user queries that would come up over time.

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1 Marker lists

The *CellMix* package ships with a set of marker lists gathered from a variety of public databases. This section shows how to perform some common task with these gene lists.

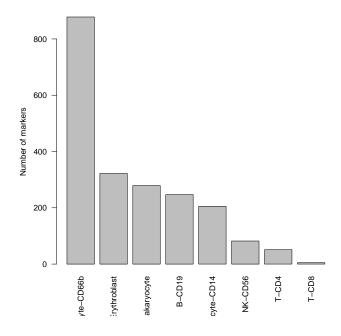
1.1 How to list all available marker gene lists?

1.2 How to load a registered marker gene list?

```
# load HaemAtlas markers
m <- cellMarkers("HaemAtlas")
# or
m <- MarkerList("HaemAtlas")</pre>
```

1.3 How to get a summary view of a marker gene list?

```
# load
m <- MarkerList("HaemAtlas")</pre>
# show summary
summary(m)
## <object of class MarkerList>
## Types: 8 ['B-CD19', 'Erythroblast', ..., 'T-CD8']
## Mode: character
## Markers: 2069
## IDtype: .Illumina ['ILMN_1793637', 'ILMN_1663575', ..., 'ILMN_1815673']
## Source: illuminaHumanv2.db
## Breakdown:
##
              B-CD19
                           {\tt Erythroblast~Granulocyte-CD66b}
                                                                Megakaryocyte
##
                                    322
                 247
                                                       878
                                                                          279
                                                     T-CD4
                                                                        T-CD8
##
       Monocyte-CD14
                                NK-CD56
##
                 205
                                      82
                                                        51
                                                                            5
# plot number of markers for each cell type
barplot(m)
```



1.4 How to subset marker gene lists?

```
# subset the cell types
summary(m[1:3])
## <object of class MarkerList>
## Types: 3 ['B-CD19', 'Erythroblast', 'Granulocyte-CD66b']
## Mode: character
## Markers: 1447
## IDtype: .Illumina ['ILMN_1793637', 'ILMN_1663575', ..., 'ILMN_1857413']
## Source: illuminaHumanv2.db
## Breakdown:
##
             B-CD19
                        Erythroblast Granulocyte-CD66b
##
                247
                                  322
                                                    878
# Take only first n markers of each cell type
summary(m[, 1:3])
## <object of class MarkerList>
## Types: 8 ['B-CD19', 'Erythroblast', ..., 'T-CD8']
## Mode: character
## Markers: 24
## IDtype: .Illumina ['ILMN_1793637', 'ILMN_1663575', ..., 'ILMN_1726589']
## Source: illuminaHumanv2.db
## Breakdown:
##
             B-CD19
                         Erythroblast Granulocyte-CD66b
                                                            Megakaryocyte
##
                             3 3
                                                                        3
##
                              NK-CD56
                                                 T-CD4
                                                                    T-CD8
      Monocyte-CD14
# subset markers that are present in some dataset => this converts/maps
# IDs if necessary
x <- ExpressionMix("GSE11058")
subset(m, x, verbose = TRUE)
## # Converting 2069 markers from Annotation (illuminaHumanv2.db) to Annotation (hgu133plus2.db)
## # Processing 2069 markers from Annotation (illuminaHumanv2.db) to Annotation (hgu133plus2.db)
## Matching character marker list against 54675 strings ['1007_s_at', '1053_at', ...,
'AFFX-TrpnX-M_at'] \dots
## OK [1659/1659 matche(s)]
## <object of class: MarkerList>
## Types: B-CD19, Erythroblast, ..., T-CD8 (total: 8)
## Mode: character
## setName: HaemAtlas
## geneIds: 206513_at, 207655_s_at, ..., 221126_at (total: 1659)
## geneIdType: Annotation (hgu133plus2.db)
## collectionType: Null
## geneValues: NA
## details: use 'details(object)'
```

1.5 How to convert MarkerList objects into plain lists?

MarkerList objects can be converted to plain list objects using the methods geneIds, or geneValues if one wants to keep numeric scores associated with each marker:

```
# load marker list that contains scores
ml <- cellMarkers("TIGER")</pre>
# plain list dropping values
1 <- geneIds(ml)</pre>
str(head(1))
## List of 6
                 : chr [1:199] "Hs.405866" "Hs.281295" "Hs.534503" "Hs.549507" ...
## $ bladder
   $ blood
                 : chr [1:413] "Hs.552036" "Hs.557039" "Hs.23118" "Hs.448401" ...
                 : chr [1:114] "Hs.518726" "Hs.2936" "Hs.1584" "Hs.98785" ...
   $ bone
## $ bone_marrow: chr [1:269] "Hs.474119" "Hs.458263" "Hs.289232" "Hs.200929"
## $ brain : chr [1:342] "Hs.7124" "Hs.12440" "Hs.13284" "Hs.20945" ...
                 : chr [1:216] "Hs.343864" "Hs.74082" "Hs.256632" "Hs.528920" ...
# plain list keeping values
1 <- geneValues(ml)</pre>
str(head(1))
## List of 6
                 : Named num [1:199] 99.9 78.5 52.3 47.1 45.8 ...
## $ bladder
    ..- attr(*, "names")= chr [1:199] "Hs.405866" "Hs.281295" "Hs.534503" "Hs.549507" ...
                 : Named num [1:413] 68.3 68.3 66.4 64.7 63 ...
##
   $ blood
    ..- attr(*, "names")= chr [1:413] "Hs.552036" "Hs.557039" "Hs.23118" "Hs.448401" ...
##
##
   $ bone
                : Named num [1:114] 50.7 40.6 40 28.9 27.4 ...
    ..- attr(*, "names")= chr [1:114] "Hs.518726" "Hs.2936" "Hs.1584" "Hs.98785" ...
## $ bone_marrow: Named num [1:269] 88.6 71.3 58.3 54.4 50.2 ...
    ..- attr(*, "names")= chr [1:269] "Hs.474119" "Hs.458263" "Hs.289232" "Hs.200929" ...
##
                 : Named num [1:342] 7.27 7.27 7.27 7.27 ...
## $ brain
    ..- attr(*, "names")= chr [1:342] "Hs.7124" "Hs.12440" "Hs.13284" "Hs.20945" ...
## $ cervix
                 : Named num [1:216] 66.1 44.5 41.3 34.4 33.1 ...
   ..- attr(*, "names")= chr [1:216] "Hs.343864" "Hs.74082" "Hs.256632" "Hs.528920" ...
```

1.6 How to create MarkerList objects

MarkerList objects can be manually created from a variety of format/object types, using the factory generic MarkerList():

2 Datasets

The *CellMix* package ships a with a set of pre-processing pipelines for some public datasets on GEO, that can be used as benchmark data for gene expression deconvolution methods.

2.1 How to list all available datasets?

2.2 How to load a dataset?

Datasets are loaded using the function ExpressionMix. This requires an internet connection. The first call will create a data directory in the user home directory, where all files related to datasets will be stored (GSE matrix files, GPL files, cache files, etc..)

```
# load GSE29832 from Gong et al. (2011)
mix <- ExpressionMix("GSE29832")</pre>
mix
## ExpressionMix (storageMode: lockedEnvironment)
## assayData: 54675 features, 15 samples
   element names: exprs
## protocolData: none
## phenoData
     sampleNames: GSM739214 GSM739215 ... GSM739228 (15 total)
##
##
     varLabels: title geo_accession ... Biorep (36 total)
     varMetadata: labelDescription
## featureData
    featureNames: 1007_s_at 1053_at ... AFFX-TrpnX-M_at (54675
##
##
      total)
    fvarLabels: ID GB_ACC ... Gene Ontology Molecular Function (16
##
##
      total)
##
     fvarMetadata: Column Description labelDescription
## experimentData: use 'experimentData(object)'
## Annotation: hgu133plus2.db
## Composition: 'Blood', 'Breast' (2 total)
```

2.3 How to retrieve data from an ExpressionMix object?

ExpressionMix objects are containers for multiple types of data, which can be retrieved with deticated methods. The idea is to hold both gene expression and cell composition data in a single object, facilitating common dataset operations (e.g. subsetting features or samples).

```
# dimensions of an ExpressionMix object
dim(mix)

## Features Samples Components
## 54675 15 2
```

Expression data: it is stored as an ExpressionSet object and is accessible via eset or exprs, if only the expression matrix is needed:

```
class(eset(mix))

## [1] "ExpressionSet"

## attr(,"package")

## [1] "Biobase"

class(exprs(mix))

## [1] "matrix"

dim(exprs(mix))

## [1] 54675 15
```

Cell proportions: if available, they are stored in the mixuture coefficient matrix of the embedded NMF model and are accessible with the method coef:

```
dim(coef(mix))
## [1] 2 15
```

Cell-specific signatures if available, they are stored in the basis matrix of the embedded NMF model and are accessible with the method basis:

```
dim(basis(mix))
## [1] 54675 2
```

3 Deconvolution methods

3.1 How to list all available methods?