## A Practical Beginner's Guide to Proteomics

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## **Abstract**

## Introduction

[paragraph about what proteomics means today]

[history of proteomics? how we got here]

[paragraph about what proteomics can do] A wide range of questions are addressable with proteomics experiments, which translates to a wide range of variations of proteomics workflows. Sometimes identifying what proteins are present is desired, and sometimes the quantities of as many proteins as possible are desired.

### **Protein Extraction**

Discussion of methods for protein extraction and solubilizaition.

- 1. Choice of Lysis buffer
- Urea can cause chemical modifications
- 2. chemicals to avoid
- 3. removal of contaminations, Protein Precipitation
- 4. protein alkylation
- choices of reduction and alkylation reagents, TCEP/DTT/2BME, Chloroacetamide/iodoacetamide, nethyl maleimide

## **Proteolysis**

- 1. discussion of protein sequence coverage is determined by the choice of proteolysis
- 2. why trypsin is the most common choice (charge and length character)
- 3. theoretical studies of proteolysis and enzyme [1]
- 4. Challenges associated with alternative enzyme choices (non-specific and semi-specific enzymes)
- 5. Alternative enzyme choices (one paragraph each?) LysC
- 6. GluC
- 7. AspN
- 8. Alpha-lytic protease [2] and how it enables mapping human SUMO sites [3].
- 9. others?

## **Peptide and Protein Labeling**

Discussion of methods to isotopically label peptides or proteins that enable quantification

- 1. SILAC/SILAM
- 2. iTRAQ
- 3. TMT
- 4. dimethyl labeling

## **Peptide or Protein Enrichment**

### Protein enrichment (e.g. for protein protein interactions)

- colP
- APEX
- bioID
- bioplex

### Peptide enrichment

- antibody enrichments of modifications, e.g. lysine acetylation [4].
- TiO2 and Fe enrichment of phosphorylation
- Glycosylation
- SISCAPA

## **Methods for Peptide Purification**

- 1. Reverse phase including tips and cartridges
- 2. stage tips
- 3. in stage tip (iST)
- 4. SP2, SP3
- 5. s traps

## **Experiment Design**

This section should discuss trade offs and balancing them to design an experiment. 1. constraints: Each experiment will have different constraints, which may include the number of samples needed for analysis, or desire to quantify a specific subset of proteins within a sample. 2. sample size 3. statistics 4. costs

#### References

# 1. <i>In Silico</i> Proteome Cleavage Reveals Iterative Digestion Strategy for High Sequence Coverage

Jesse G Meyer

ISRN Computational Biology (2014-04-22) https://doi.org/gb6s2r

DOI: https://doi.org/10.1155/2014/960902

#### 2. Expanding Proteome Coverage with Orthogonal-specificity α-Lytic Proteases

Jesse G Meyer, Sangtae Kim, David A Maltby, Majid Ghassemian, Nuno Bandeira, Elizabeth A Komives

Molecular & Cellular Proteomics (2014-03) https://doi.org/f5vgcg

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## 3. Site-specific identification and quantitation of endogenous SUMO modifications under native conditions.

Ryan J Lumpkin, Hongbo Gu, Yiying Zhu, Marilyn Leonard, Alla S Ahmad, Karl R Clauser, Jesse G Meyer, Eric J Bennett, Elizabeth A Komives

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## 4. Simultaneous Quantification of the Acetylome and Succinylome by 'One-Pot' Affinity Enrichment

Nathan Basisty, Jesse G Meyer, Lei Wei, Bradford W Gibson, Birgit Schilling *PROTEOMICS* (2018-08-19) <a href="https://doi.org/gn4cmb">https://doi.org/gn4cmb</a>

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