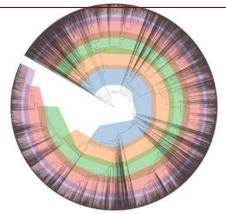
#### **ENVIRONMENTAL METAGENOMICS**

Physalia course, online, 11-15 November 2024

#### Long read assembly

Nikolay Oskolkov, Lund University, NBIS SciLifeLab Samuel Aroney, Queensland University of Technology





NB: original course material courtesy: Dr. Antti Karkman, University of Helsinki Dr. Igor Pessi, Finnish Environment Institute (SYKE) As. Prof. Luis Pedro Coelho

### A bit about me

Bachelor at University of Queenslar

DPhil at Oxford University, UK

Internship with Zooniverse – Software development

- Post-doc at Queensland University of Technology, Australia
  - $\circ \quad \text{ In the Woodcroft group} \to \to$
  - sandpiper.qut.edu.au

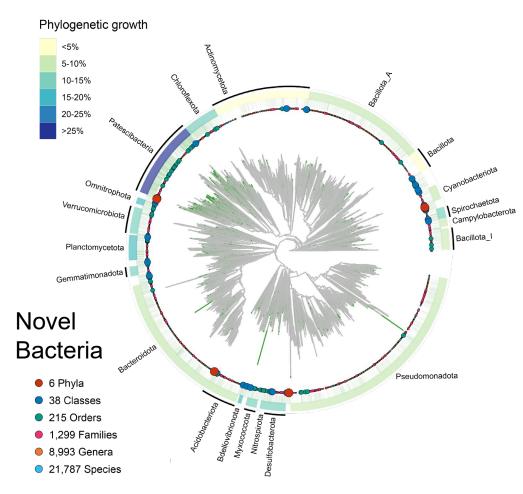




### A bit about my work

- Method development
- Global-scale genome recovery
- Permafrost thaw



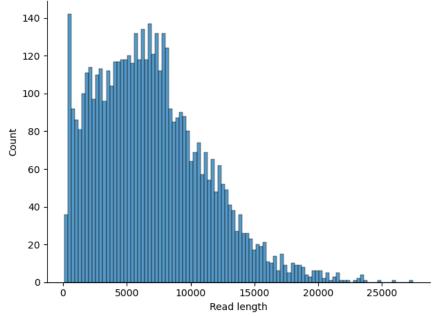


# Long reads are longer than short reads

When short reads first appeared, they were 35bps!

Nowadays, short reads are 150-300bp and long reads are longer





# We can get better assemblies with long reads!

Some individual reads are longer than contigs from short-read assembly

It can cover repeats and other hard to assemble regions

Home > BMC Genomics > Article

Metagenomic assemblies
tend to break around
antibiotic resistance genes

Research | Open access | Published: 14 October 2024
Volume 25, article number 959, (2024) Cite this article

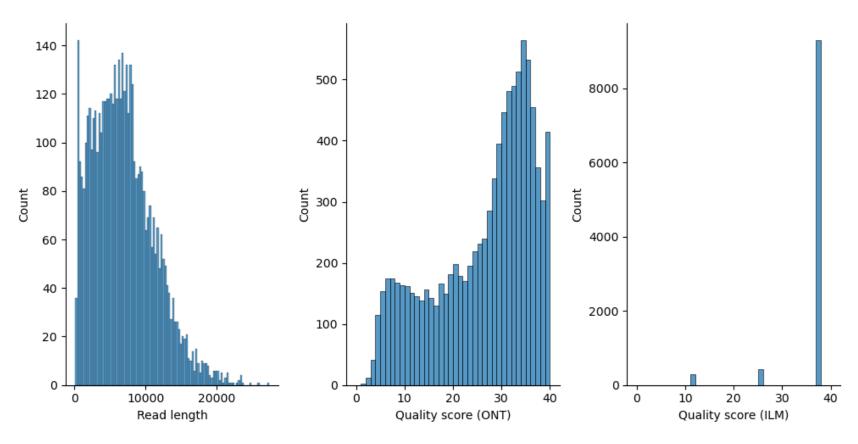
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RefSeq Non-RefSeq counts Log10 ONT MAGs MAGs

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# Why do we even use short reads then?



### Best of both worlds: do both!

In fact, the data I showed before was from hybrid assembly!

#### How do you handle hybrid assembly?

- 1. Polish the long-reads with Illumina
- 2. Use a hybrid assembler (that takes ONT + ILM)
- 3. Polish the output of a long-read assembler with ILM
- 4. Use a short read assembler, then scaffold with the long reads
- 5. ...?





### ONT data is in FASTQ format (like Illumina)

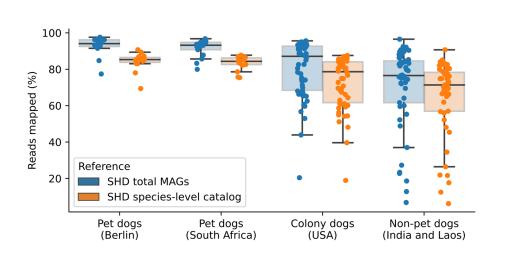
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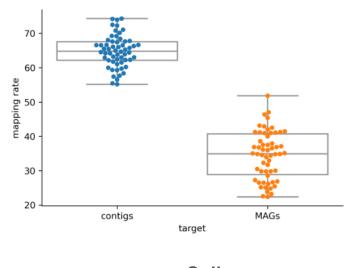
# How do results look like with long-reads?

		#contig	Average length	N50
Sample A	mNGS	277,884	1,865	8,778
	PacBio-HiFi	2,575	99,842	269,406
	Nanopore	3,414	65,685	199,639
Sample B	mNGS	146,857	2,402	28,310
	PacBio-HiFi	1,447	157,228	1,081,788
	Nanopore	1,795	123,512	658,841
Sample C	mNGS	170,813	1,956	18,485
	PacBio-HiFi	822	171,215	1,270,126
	Nanopore	1,370	126,533	891,411



### Results depend on the habitat!





**Soil** (40 Gbp + 40 Gbp) x 52 samples

**Dog gut** (20 Gbp + 20 Gbp) x 50 samples



In dog guts, 20 Gbp was enough to get excellent representation of the community, but in soil even double that was not enough! — could also be Euks!

From Anna Cuscó (dog) & Yiqian Duan (soil)

# A few other notes on long-reads

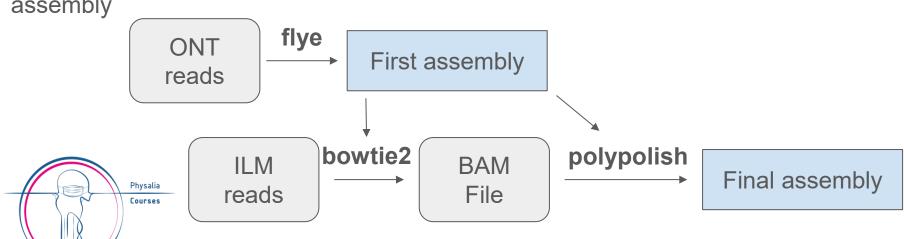
- Still a very rapidly evolving field (both in the wetlab & in the drylab)
- Differences between ONT versions matter a lot
- We might be able to get even more information in the future
  - Methylation
  - Non-canonical nucleotides
- Long-reads are the future



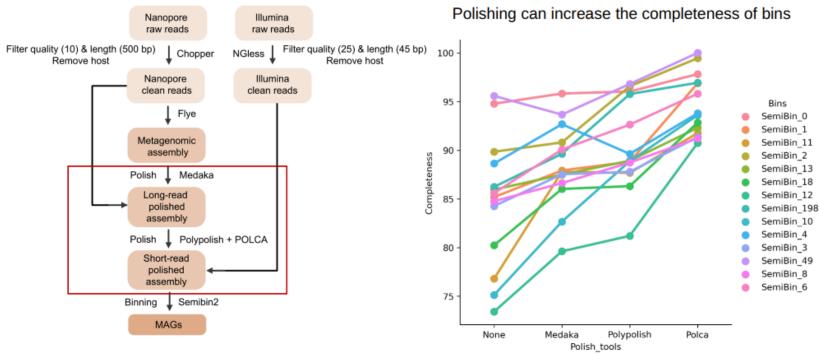
#### **Exercises overview**

- 1. Assemble with **flye** (long-read only)
- 2. Polish with **polypolish** (using the short reads)

In order to polish, we need to align the short reads to the contigs from the **flye** assembly



#### Real workflow





From <u>Yiqian Duan</u>