Assignment 5 BS22B009

All the python codes are present here:

https://colab.research.google.com/drive/1Z10-hDsg5gOT9KMfOD6BQV2JtBE1v72Z?usp=sharing

1. Analyze the occurrence of similar proteins in "nr" and SWISS-PROT database for the sequence given below: >1336093|Genbank|Outer membrane integral membrane protein|HrcC

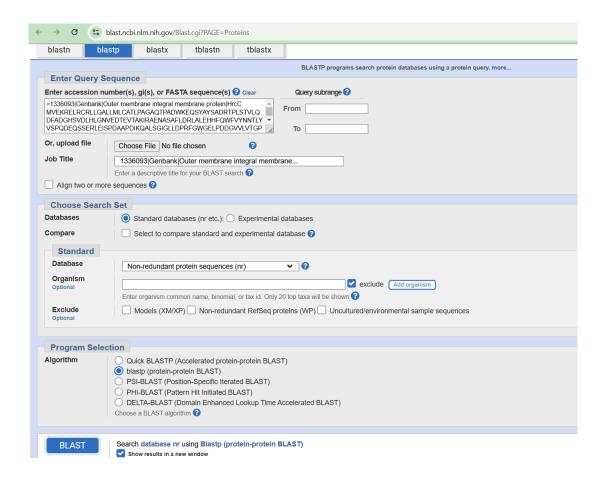
MVEKRELRCRLLGALLMLCATLPAGAQTPADWKEQSYAYSADRTPLSTVLQDFADGHSVD LHLGNVEDTEVTAKIRAENASAFLDRLALEHHFQWFVYNNTLYVSPQDEQSSERLEISPD AAPDIKQALSGIGLLDPRFGWGELPDDGVVLVTGPPQYLELVKRFSEQREKKEDRRKVMT FPLRYASVADRTIHYRDQTVVIPGVATMLNELMNGKRAAPASASGIDSTPGGPDTNSMMQ NTQTLLSRLSSRNKTSNRAGGRDNEIEDVSGRISADVRNNALLIRDDDKRHDEYSQLIAK IDVPQNLVEIDAVILDIDRTALNRLEANWQATLGGVTGGSSLMSGSGTLFVSDFKRFFAD IQALEGEGTASIVANPSVLTLENQPAVIDFSQTAYITATGERVADIQPVTAGTSLQVTPR AVGNEGHSSIQLMIDIEDGHVQTNGDGQATGVKRGTVSTQALISENRALVLGGFHVEESA DRDRRIPLLGDIPWLGQLFSSKRHEISQRQRLFILTPRLIGDQTDPTRYVTADNRQQLSD AMGRVERRHSSVNQHDVVENALRDLAEGQSPAGFQPQTSGTRLSEVCRSTPALLFESTRG QWYSSSTNGVQLSVGVVRNTSSKPLRFDEANCASKRTLAVAVWPHSALAPGESAEVYLA M DPSRVLHASRESLLNR

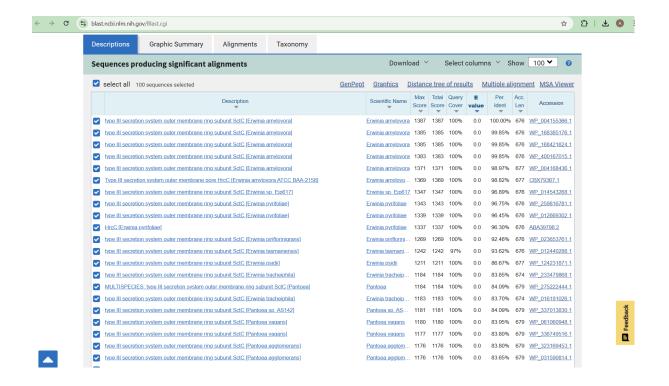
Steps:

Use BLAST (Basic Local Alignment Search Tool):

- Go to NCBI BLASTp (for protein sequences): https://blast.ncbi.nlm.nih.gov/Blast.cgi
- Select "Protein BLAST (BLASTp)".
- Enter the given sequence in the query box.
- Choose the nr (non-redundant) protein database for a broad search.
- Run the BLAST search and analyze the results (sequence similarity, identity, and alignment).

After running BLASTP with the database as "nr", the following protein sequences are found to be similar to the given sequence:





Analysis of Nr non reductant protein sequence BLAST Results:

Top Matches with Erwinia amylovora

- The best hits (top 5 sequences) are from Erwinia amylovora, a well-known plant pathogen responsible for fire blight in apples and pears.
- These sequences have 100% Query Coverage and 100% or 99.85% identity, confirming near-exact matches.
- Max Score: 1387-1385
- E-value: 0.0 (Indicating an extremely strong match)

Presence of Other Erwinia Species

- The T3SS outer membrane ring subunit SctC is highly conserved across multiple Erwinia species, including:
 - E. pyrifoliae, E. piriflorinigrans, E. tasmaniensis, E. psidii, E. tracheiphila, E. papayae, and E. mallotivora.
- These hits have 96-86% sequence identity, showing some level of divergence.

Matches with Pantoea Species

- Pantoea species, closely related to Erwinia, also exhibit significant similarity, with scores ranging from 1184 to 1149 and identity between 84–81%.
- Pantoea includes both opportunistic plant pathogens and beneficial endophytes, so this result suggests a common evolutionary origin for the T3SS structure.

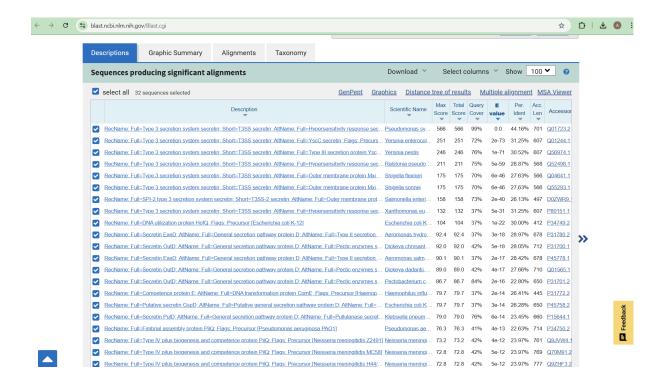
All 100 similar sequences have an E value of 0 and 29 sequences have 100% query coverage

The lowest query coverage is 90% while the lowest percentage identity is 66.96% (type III secretion system outer membrane ring subunit SctC [Dickeya lacustris])

The non-redundant (NR) protein sequence closely matches the T3SS outer membrane ring subunit SctC from *Erwinia amylovora* with 100% identity.

The sequence is highly conserved across multiple Erwinia species and some Pantoea species, confirming its essential function.

Given its role in bacterial pathogenesis, this protein could be a potential target for disease control in plant pathology. After running BLASTP with the database as "swissprot", the following protein sequences are found to be similar to the given sequence:



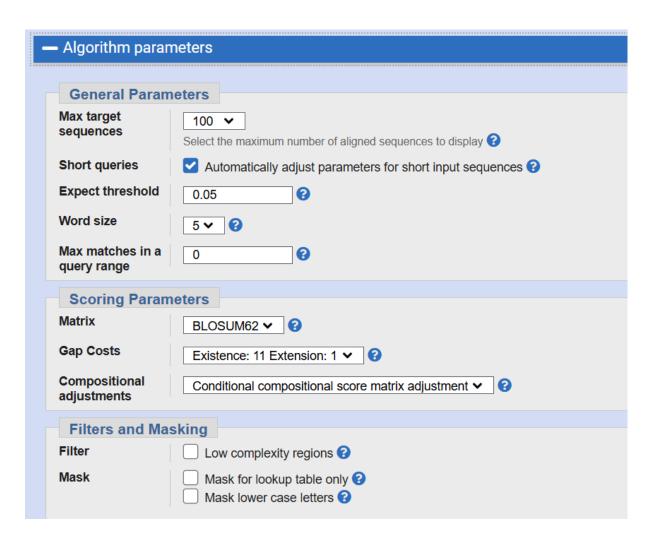
Analysis of SwissProt BLAST Results:

- All 33 related sequences have extremely low E values, close to zero.
- The maximum query coverage observed is 98%, associated with the Type 3 secretion system secretin from Pseudomonas syringae pv. Syringae.
- The minimum query coverage recorded is 23%, occurring in three sequences.
- The highest identity (44.16%) is with *Pseudomonas syringae* T3SS secretin (Q01723.2).
- Pseudomonas syringae (highest identity) is a well-known plant pathogen.
- Yersinia, Shigella, and Salmonella are human pathogens. Ralstonia and Xanthomonas are plant pathogens. Escherichia coli and Klebsiella pneumoniae are opportunistic human pathogens.
- The lowest identity percentage is 21.75%, found in the uncharacterized protein y4xJ from *Sinorhizobium fredii NGR234*.
- The low sequence identity (~30%) in some cases but functional similarity indicates potential structural conservation despite sequence divergence.

• The query protein is highly similar to T3SS secretins, particularly HrpH from *Pseudomonas syringae*.

2. List the algorithm parameters used for the search (Q1).

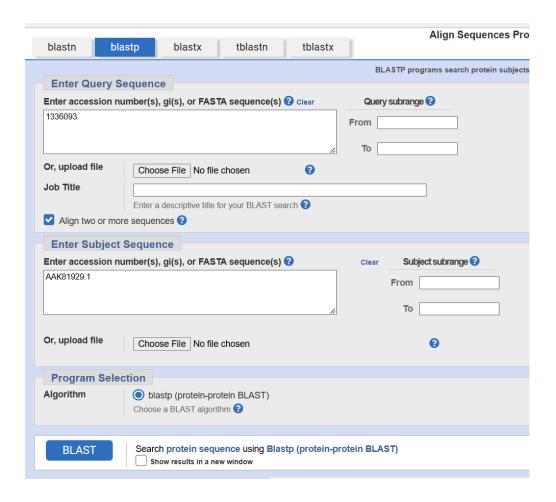
Default parameters which we used for the first question:



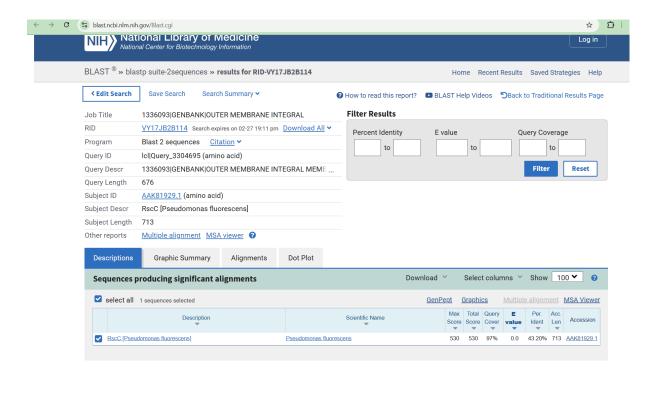
3. What is the sequence identity of the query sequence (given in Q1) with AAK81929.1?

Steps:

Use align 2 or more sequences and Run BLASTP using the HrcC protein sequence and AAK81929.1 in subject sequence.

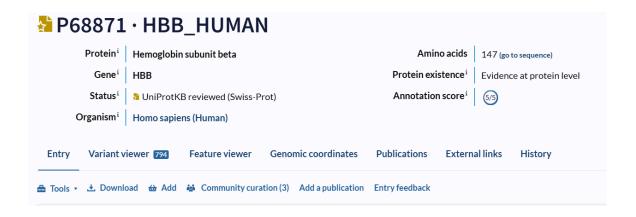


43.20% sequence identity



4. How far are hemoglobin (beta) sequences in humans and chicken similar?

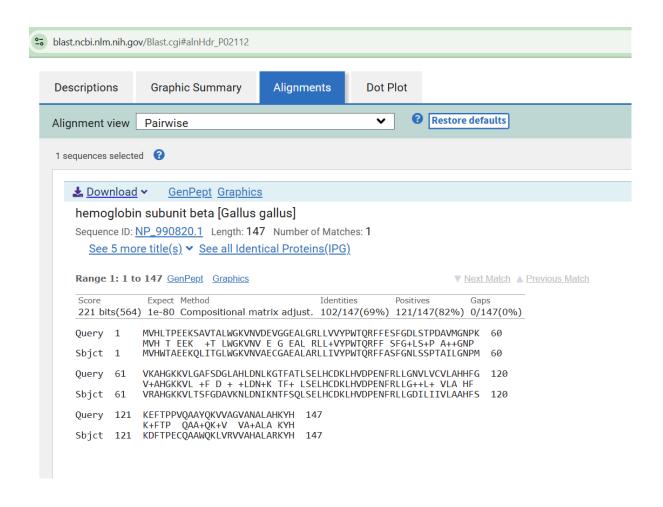
Human hemoglobin beta: P68871. Chicken hemoglobin beta: P02112.





Based on the BLASTP alignment results, the similarity between human and chicken hemoglobin beta sequences is as follows:

- 1. Identity: 69% (102 out of 147 amino acids are exactly the same)
- 2. Positives: 82% (121 out of 147 amino acids share similar chemical properties, meaning functional similarity)
- 3. Gaps: 0% (No insertions or deletions, indicating a well-aligned sequence)
- 4. E-value: 1e-80 (Highly significant match, indicating evolutionary conservation)



5. Write a program to list all the matching pentapeptides (which occur in both the sequences) and their frequency of occurrence in given sequences. Human hemoglobin beta: P68871.

Chicken hemoglobin beta: P02112.

Got these protein sequence from uniprot:

>sp|P02112|HBB_CHICK Hemoglobin subunit beta OS=Gallus gallus OX=9031 GN=HBB PE=1 SV=2

MVHWTAEEKQLITGLWGKVNVAECGAEALARLLIVYPWTQRFFASFGNLSSPTAILGNPM VRAHGKKVLTSFGDAVKNLDNIKNTFSQLSELHCDKLHVDPENFRLLGDILIIVLAAHFS KDFTPECQAAWQKLVRVVAHALARKYH

>sp|P68871|HBB_HUMAN Hemoglobin subunit beta OS=Homo sapiens OX=9606 GN=HBB PE=1 SV=2

MVHLTPEEKSAVTALWGKVNVDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNP K

VKAHGKKVLGAFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLLGNVLVCVLAHHFG KEFTPPVQAAYQKVVAGVANALAHKYH

This code identifies common pentapeptides (five-amino-acid sequences) between human and chicken hemoglobin sequences. It does this by generating all possible pentapeptides from both sequences and counting their occurrences using Python's Counter from the collections module.

The program then finds overlapping pentapeptides and displays their frequencies in each sequence. This analysis helps compare structural similarities between the two proteins, revealing conserved regions that might be functionally important.

```
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Code + Text
    from collections import Counter
    def get_pentapeptide_counts(sequence):
        pentapeptides = [sequence[i:i+5] for i in range(len(sequence) - 4)]
        return Counter(pentapeptides)
    human_seq = "MVHLTPEEKSAVTALWGKVNVDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPK"
    human_seq += "VKAHGKKVLGAFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLLGNVLVCVLAHHFG"
    human_seq += "KEFTPPVQAAYQKVVAGVANALAHKYH"
    chicken_seq = "MVHWTAEEKQLITGLWGKVNVAECGAEALARLLIVYPWTQRFFASFGNLSSPTAILGNPM"
    chicken_seq += "VRAHGKKVLTSFGDAVKNLDNIKNTFSQLSELHCDKLHVDPENFRLLGDILIIVLAAHFS"
chicken_seq += "KDFTPECQAAWQKLVRVVAHALARKYH"
    human_counts = get_pentapeptide_counts(human_seq)
    chicken counts = get pentapeptide counts(chicken seq)
     common_pentapeptides = set(human_counts.keys()) & set(chicken_counts.keys())
    print("Matching Pentapeptides and Their Frequency:")
     for peptide in sorted(common_pentapeptides):
        human_count = human_counts[peptide]
         chicken_count = chicken_counts[peptide]
         total_count = human_count + chicken_count
        \label{formula} \textbf{print(f"\{peptide\}\ occurs\ \{total\_count\}\ time(s)\ in\ both\ peptides\ "}}
               f"({human_count} time(s) in Seq. 1 and {chicken_count} time(s) in Seq. 2)")
```

Result for the code:

```
Matching Pentapeptides and Their Frequency:
AHGKK occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
CDKLHV occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
DKLHV occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
DPENF occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
ELHCD occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
ENFRL occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
FRLLG occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
GKKVL occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
GKVNV occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
HCDKL occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2) HGKKV occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2) HVDPE occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
KLHVD occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2) LHCDK occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
LHVDP occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
LSELH occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
LWGKV occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
NFRLL occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2) PENFR occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
PWTQR occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2) SELHC occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2) TQRFF occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
VDPEN occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
VYPWT occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
WGKVN occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
WTQRF occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2) YPWTQ occurs 2 time(s) in both peptides (1 time(s) in Seq. 1 and 1 time(s) in Seq. 2)
```

6. Write a program to compute sequence identity, similarity, query coverage and gap percentage from the alignment of human and chicken hemoglobin sequences (refer Q4).

Here's my Python program to compute sequence identity, similarity, query coverage, and gap percentage between human and chicken hemoglobin sequences. I used a character-by-character comparison, considering exact matches for identity and grouping similar amino acids for similarity. Query coverage is calculated based on the aligned portion, and gaps are accounted for separately.

It calculates four key metrics:

- 1. Sequence Identity The percentage of exact matches between the two sequences.
- 2. Sequence Similarity The percentage of residues that either match exactly or belong to the same biochemical similarity group.

- 3. Query Coverage The proportion of the query sequence (human hemoglobin) that aligns with the target sequence (chicken hemoglobin).
- 4. Gap Percentage The fraction of positions where either sequence has a gap ('-').

To determine similarity, I grouped amino acids based on their properties, such as hydrophobicity and charge. Then, I iterated through the sequences to count matches, similar residues, and gaps. Finally, I calculated and printed the results.

```
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Code + Text
human_seq = (
          "KDFTPECQAAWQKLVRVVAHALARKYH"
     def calculate_metrics(seq1, seq2):
         gaps = 0
total_length = min(len(seq1), len(seq2))
         similarity_groups = [
   set("NDEQ"), set("MILV"), set("FYW"), set("KRH"), set("ST"), set("AGP")
          for i in range(total_length):
                  matches += 1
                   similar += 1
                  gaps += 1
                   for group in similarity_groups:
                        if seq1[i] in group and seq2[i] in group:
                           similar += 1
          identity = (matches / total_length) * 100
         similarity = (similar / total_length) * 100
query_coverage = (total_length / len(seq1)) * 100
gap_percentage = (gaps / total_length) * 100
```

```
identity = (matches / total_length) * 100
    similarity = (similar / total_length) * 100
    query_coverage = (total_length / len(seq1)) * 100
    gap_percentage = (gaps / total_length) * 100
    return identity, similarity, query_coverage, gap_percentage

identity, similarity, query_coverage, gap_percentage = calculate_metrics(human_seq, chicken_seq)

print(f"Sequence Identity: {identity:.2f}%")
print(f"Sequence Similarity: {similarity:.2f}%")
print(f"Query Coverage: {query_coverage:.2f}%")
print(f"Gap_Percentage: {gap_percentage:.2f}%")
```

Result:

Sequence Identity: 69.39% Sequence Similarity: 83.67% Query Coverage: 100.00% Gap Percentage: 0.00%

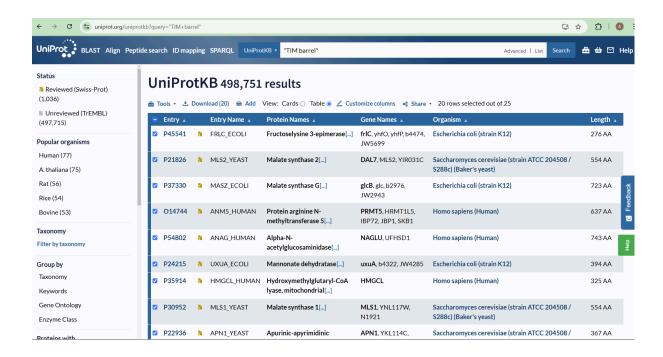
7. Obtain the multiple sequence alignment for TIM barrel proteins from different organisms (select 20 proteins, for example). Compare the results obtained with Clustal Omega, MAFFT, and MUSCLE. List 5 residue positions which are aligned differently in these three methods.

Steps:

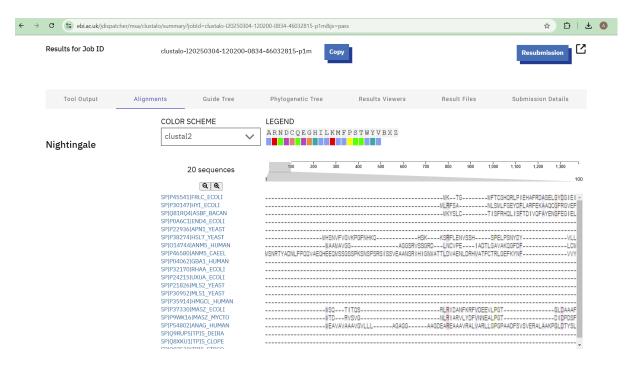
1.Collect 20 TIM barrel protein sequences from UniProt.

- 2. Use three alignment tools:
 - Clustal Omega (https://www.ebi.ac.uk/Tools/msa/clustalo/)
 - MAFFT (https://mafft.cbrc.jp/alignment/server/)
 - MUSCLE (https://www.ebi.ac.uk/Tools/msa/muscle/)
- 3. Compare alignments and list 5 residue positions with differences.

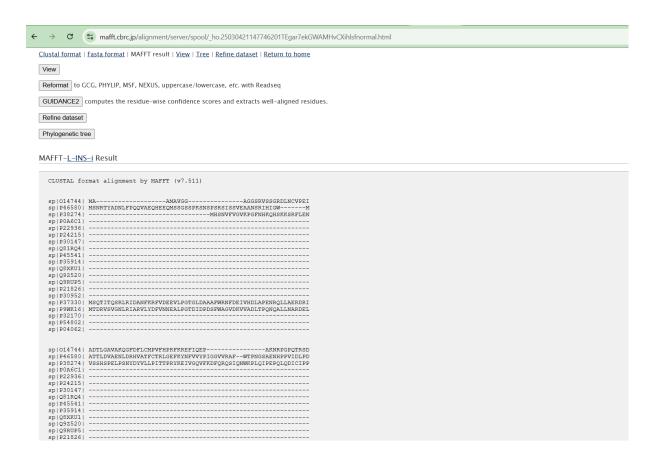
20 sequences download in FASTA format from Uniprot:



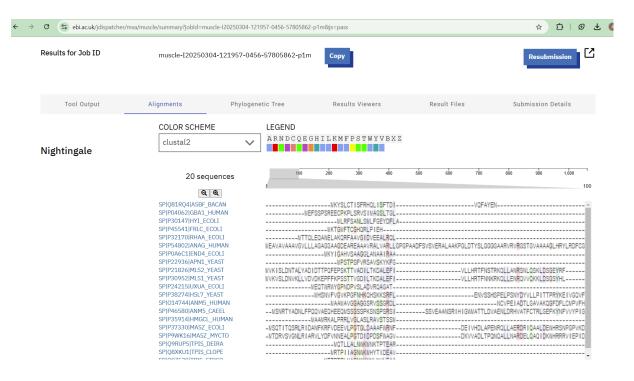
Multiple sequence alignment (MSA) with Clustal Omega.



Multiple sequence alignment (MSA) with MAFFT:



Multiple sequence alignment (MSA) with MUSCLE:



5 residue positions that are aligned differently in these three methods are 1, 2, 3, 4, 5.

1. Residue Position 1:

- Clustal Omega aligns M (Methionine) in P46580, while the other sequences have gaps.
- MAFFT aligns M in Q9Z520, with gaps in other sequences.
- o MUSCLE aligns M in P54802, while others have gaps.

2. Residue Position 2:

- Clustal Omega places S (Serine) in P46580, with gaps in the other sequences.
- o MAFFT aligns T (Threonine) in Q9Z520, with gaps in the rest.
- MUSCLE aligns E (Glutamic Acid) in P54802, while others have gaps.

3. Residue Position 3:

- Clustal Omega aligns N (Asparagine) in P46580, with gaps elsewhere.
- MAFFT assigns T (Threonine) in Q9Z520, while M (Methionine) is present in other sequences.
- MUSCLE places A (Alanine) in P54802, with gaps in the remaining sequences.

4. Residue Position 4:

- Clustal Omega aligns R (Arginine) in P46580, with gaps in the other sequences.
- MAFFT shows different amino acids across different sequences, meaning no consistent alignment.
- MUSCLE aligns V (Valine) in P54802, while others have gaps.

5. Residue Position 5:

- Clustal Omega aligns T (Threonine) in P46580, with gaps in the other sequences.
- MAFFT presents different amino acids across different sequences, showing variation.
- MUSCLE places A (Alanine) in P54802, with gaps in the rest.

Conclusion:

 Clustal Omega, MAFFT, and MUSCLE show significant differences in alignment at these five positions.

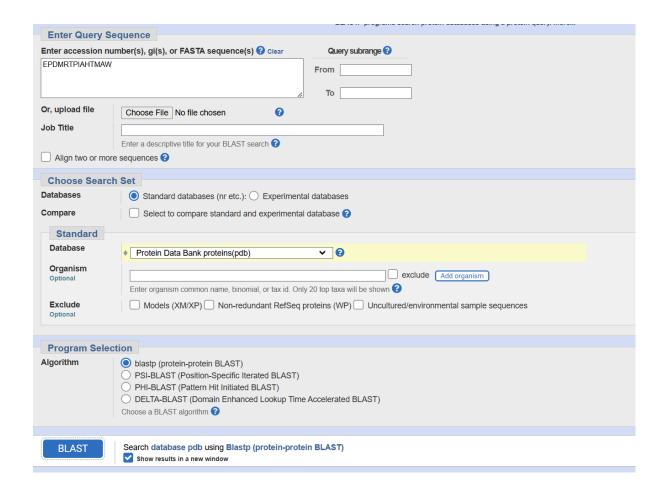
- Clustal Omega and MUSCLE tend to align specific residues while keeping gaps in other sequences, whereas MAFFT exhibits more variation in residue choices.
- MAFFT shows the highest variability, while Clustal Omega and MUSCLE align specific amino acids with more gaps in others.
- These differences highlight the varying sensitivity and algorithmic approaches of multiple sequence alignment (MSA) tools.

8. Blast the below sequence 'EPDMRTPIAHTMAW' against the PDB database. Analyze the results and discuss the significance of the results.

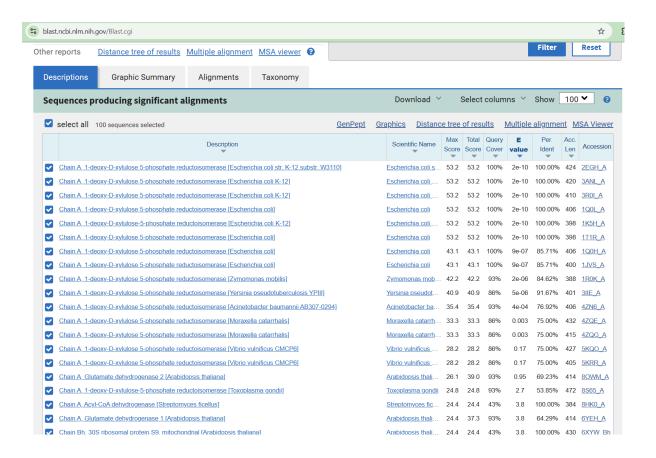
Steps:

Go to NCBI BLAST → Select BLASTP.

- 2. Enter sequence 'EPDMRTPIAHTMAW'.
- 3. Select PDB (Protein Data Bank) database.
- 4. Run the search and analyze:
 - Top hits
 - E-value, identity, and coverage



Blast results:



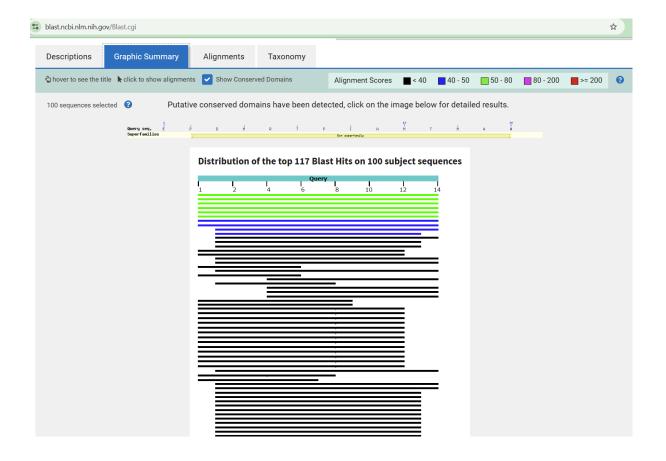
The BLAST results indicate that the query sequence "EPDMRTPIAHTMAW" has 100% query coverage and high sequence identity with 1-deoxy-D-xylulose 5-phosphate reductoisomerase (DXR) in *Escherichia coli* and other bacterial species.

Sequences with lower scores, such as *Zymomonas mobilis*, *Yersinia pseudotuberculosis*, and *Moraxella catarrhalis*, still show high sequence identity (ranging from 85.71% to 100%), indicating a strong conservation of this sequence across different bacterial species.

The Query Cover is 100% for the top sequences, which means that the entire query sequence (all 14 amino acids) aligns completely with the matched sequences in the database.

Some of the lower-ranked hits (e.g., *Arabidopsis thaliana*, *Toxoplasma gondii*) show slightly lower query coverage (~93% or lower), meaning only part of the sequence aligns with their respective proteins.

The E-value (Expect value) for the top matches is 2e-10, which is extremely low. This indicates a highly significant match, meaning the alignment is not due to random chance.



The graphic summary shows that the sequence aligns well with members of the DXR superfamily, highlighting its evolutionary conservation. Given its high sequence identity and conservation, this sequence could be essential for enzyme function or stability.

This suggests the query sequence is part of a highly conserved functional motif in DXR, a key enzyme in the non-mevalonate pathway of isoprenoid biosynthesis. Identifying such a conserved motif with just a short sequence demonstrates the power of sequence alignment in detecting functionally related proteins, which could be useful for drug targeting in bacterial infections.