

# HeatMap

Asad

2023-05-15

```
library(ComplexHeatmap)
```

```
## Loading required package: grid

## =====
## ComplexHeatmap version 2.10.0
## Bioconductor page: http://bioconductor.org/packages/ComplexHeatmap/
## Github page: https://github.com/jokergoo/ComplexHeatmap
## Documentation: http://jokergoo.github.io/ComplexHeatmap-reference
##
## If you use it in published research, please cite:
## Gu, Z. Complex heatmaps reveal patterns and correlations in multidimensional
## genomic data. Bioinformatics 2016.
##
## The new InteractiveComplexHeatmap package can directly export static
## complex heatmaps into an interactive Shiny app with zero effort. Have a try!
##
## This message can be suppressed by:
## suppressPackageStartupMessages(library(ComplexHeatmap))
## =====
```

```
library(dplyr)
```

```
##
## Attaching package: 'dplyr'

## The following objects are masked from 'package:stats':
##
## filter, lag

## The following objects are masked from 'package:base':
##
## intersect, setdiff, setequal, union
```

```
library(tidyverse)
```

```
## -- Attaching packages ----- tidyverse 1.3.1 --
```

```
## v ggplot2 3.4.2      v purrr  1.0.1
## v tibble  3.2.1      v stringr 1.5.0
## v tidyr   1.3.0      v forcats 1.0.0
## v readr   2.1.2

## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()
```

```
library(ggplot2)
library(AnnotationDbi)
```

```
## Loading required package: stats4
```

```
## Loading required package: BiocGenerics
```

```
##
## Attaching package: 'BiocGenerics'
```

```
## The following objects are masked from 'package:dplyr':
##
##   combine, intersect, setdiff, union
```

```
## The following objects are masked from 'package:stats':
##
##   IQR, mad, sd, var, xtabs
```

```
## The following objects are masked from 'package:base':
##
##   anyDuplicated, append, as.data.frame, basename, cbind, colnames,
##   dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep,
##   grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget,
##   order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank,
##   rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply,
##   union, unique, unsplit, which.max, which.min
```

```
## Loading required package: Biobase
```

```
## Welcome to Bioconductor
##
##   Vignettes contain introductory material; view with
##   'browseVignettes()'. To cite Bioconductor, see
##   'citation("Biobase")', and for packages 'citation("pkgname")'.
```

```
## Loading required package: IRanges
```

```
## Loading required package: S4Vectors
```

```
##
## Attaching package: 'S4Vectors'
```

```

## The following object is masked from 'package:tidyr':
##
##     expand

## The following objects are masked from 'package:dplyr':
##
##     first, rename

## The following object is masked from 'package:utils':
##
##     findMatches

## The following objects are masked from 'package:base':
##
##     expand.grid, I, unname

##
## Attaching package: 'IRanges'

## The following object is masked from 'package:purrr':
##
##     reduce

## The following objects are masked from 'package:dplyr':
##
##     collapse, desc, slice

## The following object is masked from 'package:grDevices':
##
##     windows

##
## Attaching package: 'AnnotationDbi'

## The following object is masked from 'package:dplyr':
##
##     select

library(org.Hs.eg.db)

##

library(circlize)

## =====
## circlize version 0.4.15
## CRAN page: https://cran.r-project.org/package=circlize
## Github page: https://github.com/jokergoo/circlize
## Documentation: https://jokergoo.github.io/circlize\_book/book/
##

```

```
## If you use it in published research, please cite:
## Gu, Z. circlize implements and enhances circular visualization
## in R. Bioinformatics 2014.
##
## This message can be suppressed by:
## suppressPackageStartupMessages(library(circlize))
## =====
```

```
library(magick)
```

```
## Linking to ImageMagick 6.9.12.3
## Enabled features: cairo, freetype, fftw, ghostscript, heic, lcms, pango, raw, rsvg, webp
## Disabled features: fontconfig, x11
```

```
library(RColorBrewer)
setwd('E:/R-Programming-Practices/Data Visualization/Heatmap')
```

## Heatmap from gene expression data (like output from DESeq2)

#Read count data

```
norm.counts<- read.csv('Normalized read counts.csv', row.names = 1)
z.data<- t(apply(norm.counts, 1, scale)) #Convert to z scores
colnames(z.data)<-rep(c('Healthy', 'COVID-19'), c(9,36)) #Change column names
z.data<- data.frame(z.data)
```

## Read DEGs matrix and stratify

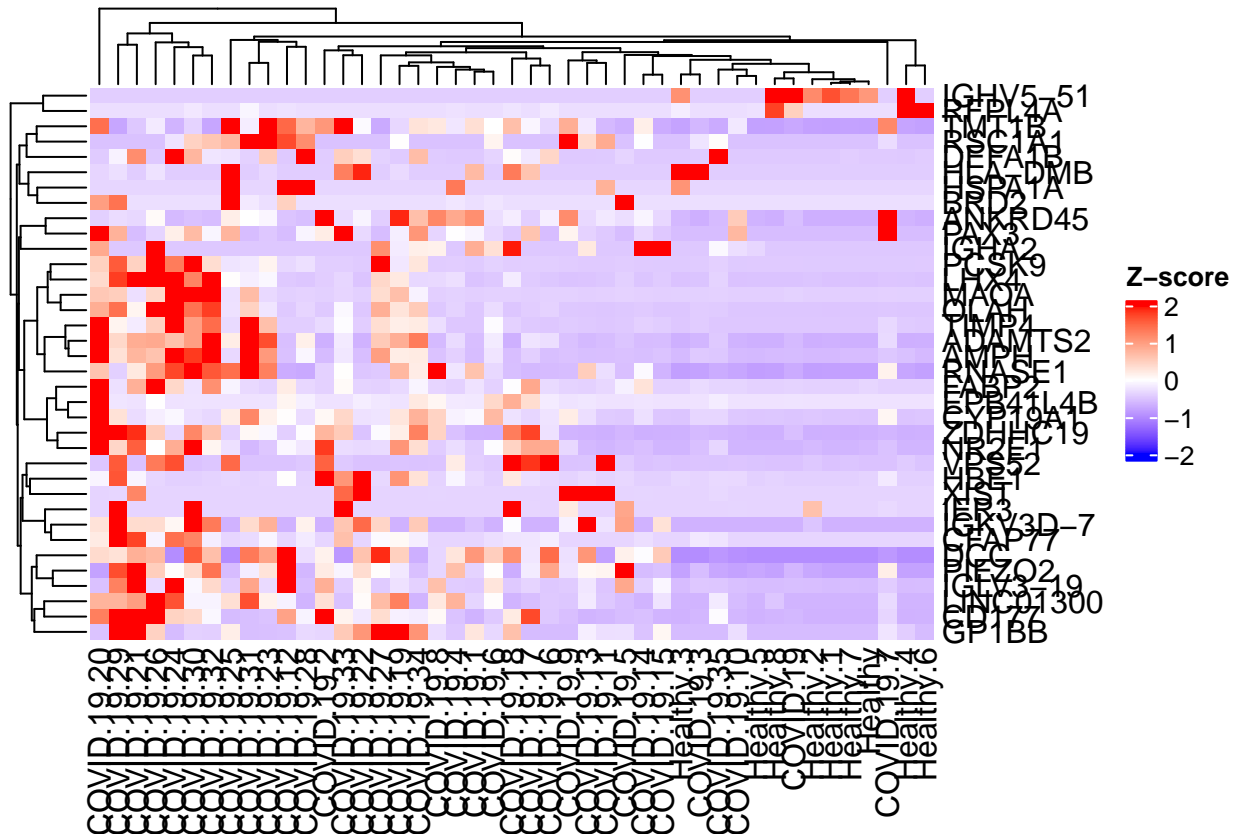
```
sig.genes<- read.csv('Significant_DEGs.csv', row.names = 1)
sig.genes<- sig.genes[order(sig.genes$padj),]
sig.genes_final<- filter(sig.genes, sig.genes$padj<0.001 & abs(sig.genes$log2FoldChange)>=5)
data.final<- z.data[row.names(sig.genes_final),]
data.final<- na.omit(data.final)
```

## Plot heatmap

```
Heatmap(data.final, cluster_rows = T, cluster_columns = T,
        column_labels = colnames(data.final), row_labels =
        sig.genes[rownames(data.final),]$symbol, name = 'Z-score')
```

```
## Warning: The input is a data frame, convert it to a matrix.
```





Heatmap from random data

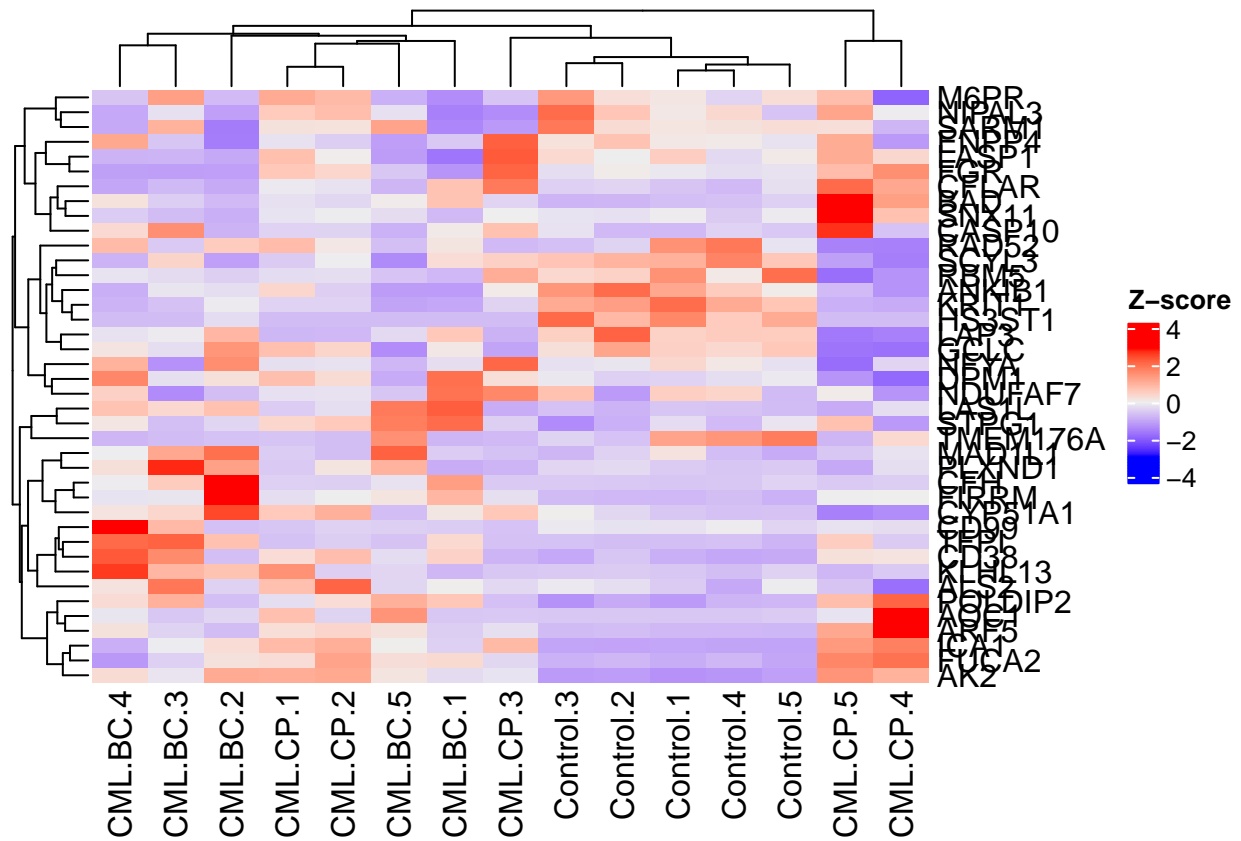
```
data2<- read.csv('Voom_CPM.csv', row.names = 1)
symbol<- mapIds(org.Hs.eg.db, keys = rownames(data2),
               keytype = 'ENSEMBL', column = 'SYMBOL')
```

## 'select()' returned 1:many mapping between keys and columns

```
symbol<- as.data.frame(symbol)

data2.z<- t(apply(data2,1,scale))
colnames(data2.z)<- colnames(data2)
data2.z<- data2.z[1:40,]

Heatmap(data2.z, row_labels = symbol[rownames(data2.z),], name = 'Z-score')
```



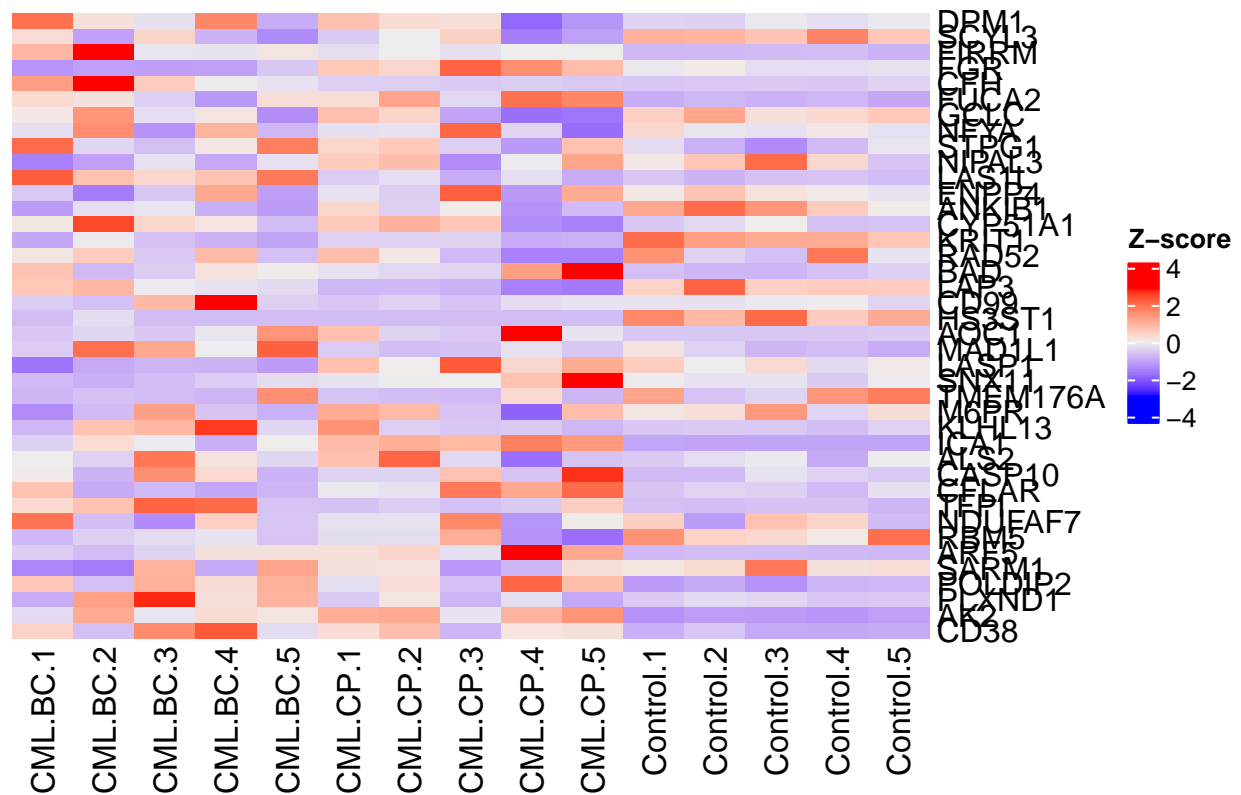
Split heatmap based on clustering

```
Heatmap(data2.z, row_labels = symbol[rownames(data2.z),], name = 'Z-score',
        column_km = 2)
```





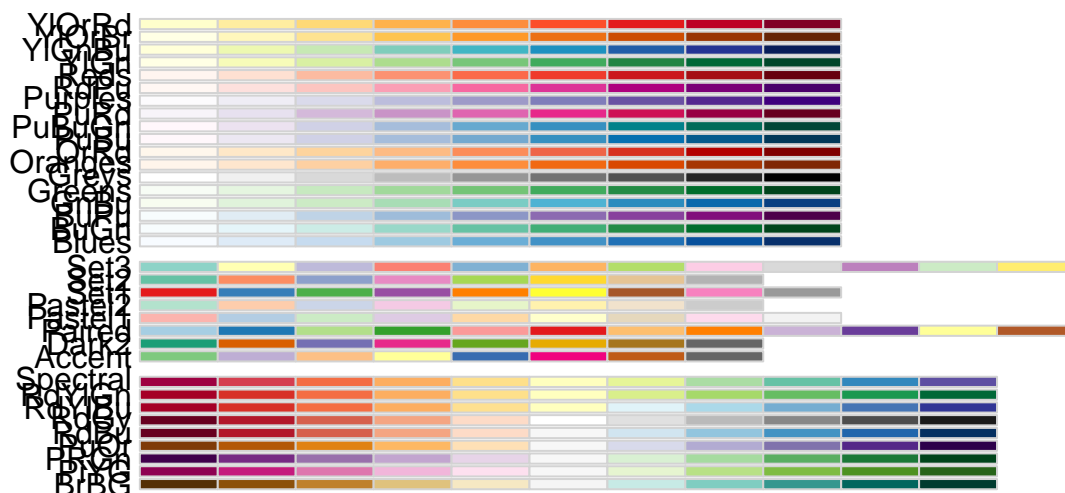
## DEGs in CML Patients vs Healthy Individuals



## Adding annotation bar

```
#Prepare data
Char1<- c(rep('Healthy', 5), 'New', 'Relapse', 'New', 'Relapse','New', 'Relapse',
           'New', 'Relapse','New', 'Relapse')
Char2<- colnames(data2)

#Adjust color
display.brewer.all()
```



```
brewer.pal(n=7, "Dark2")
```

```
## [1] "#1B9E77" "#D95F02" "#7570B3" "#E7298A" "#66A61E" "#E6AB02" "#A6761D"
```

```
ann<- data.frame(Char2, Char1)
colnames(ann)<- c('Study', 'State')
rownames(ann)<- ann[,1]
detach(package:org.Hs.eg.db, unload = TRUE)
detach(package:AnnotationDbi, unload = TRUE)
ann2<- ann %>% select('State')
col<- list('State'=c('Healthy'='#1B9E77', 'New'='#D95F02', 'Relapse'='#7570B3'))

#Adjust column annotation
colAnn <- HeatmapAnnotation(df = ann2,name = 'Disease State',
                           which = 'col',
                           col = col,
                           annotation_width = unit(c(1, 4), 'cm'),
                           gap = unit(1, 'mm'))

#Heatmap
Heatmap(data2.z, top_annotation = colAnn, name='Z-score', row_labels =
        symbol[rownames(data2.z),],
        column_names_gp = grid::gpar(fontsize = 8, fontface='bold'),
        row_names_gp = grid::gpar(fontsize = 8, fontface='bold'),
        column title = 'DEGs in CML Patients vs Healthy Individuals')
```

Heatmap showing Z-score expression data for 20 genes across 16 samples. The samples are clustered into three groups: CML (BC and CP subtypes), Controls, and CML (CP subtype). The genes are clustered into three groups: Healthy (green), New (orange), and Relapse (purple). The color scale ranges from -4 (blue) to 4 (red).

**State**

- Healthy
- New
- Relapse

**Z-score**

4  
2  
0  
-2  
-4

**Gene List:**

- M6PR
- MBP13
- ADAM1
- ADAM10
- ADAM11
- ADAM12
- ADAM13
- ADAM14
- ADAM15
- ADAM16
- ADAM17
- ADAM18
- ADAM19
- ADAM20
- ADAM21
- ADAM22
- ADAM23
- ADAM24
- ADAM25
- ADAM26
- ADAM27
- ADAM28
- ADAM29
- ADAM30
- ADAM31
- ADAM32
- ADAM33
- ADAM34
- ADAM35
- ADAM36
- ADAM37
- ADAM38
- ADAM39
- ADAM40
- ADAM41
- ADAM42
- ADAM43
- ADAM44
- ADAM45
- ADAM46
- ADAM47
- ADAM48
- ADAM49
- ADAM50
- ADAM51
- ADAM52
- ADAM53
- ADAM54
- ADAM55
- ADAM56
- ADAM57
- ADAM58
- ADAM59
- ADAM60
- ADAM61
- ADAM62
- ADAM63
- ADAM64
- ADAM65
- ADAM66
- ADAM67
- ADAM68
- ADAM69
- ADAM70
- ADAM71
- ADAM72
- ADAM73
- ADAM74
- ADAM75
- ADAM76
- ADAM77
- ADAM78
- ADAM79
- ADAM80
- ADAM81
- ADAM82
- ADAM83
- ADAM84
- ADAM85
- ADAM86
- ADAM87
- ADAM88
- ADAM89
- ADAM90
- ADAM91
- ADAM92
- ADAM93
- ADAM94
- ADAM95
- ADAM96
- ADAM97
- ADAM98
- ADAM99
- ADAM100
- ADAM101
- ADAM102
- ADAM103
- ADAM104
- ADAM105
- ADAM106
- ADAM107
- ADAM108
- ADAM109
- ADAM110
- ADAM111
- ADAM112
- ADAM113
- ADAM114
- ADAM115
- ADAM116
- ADAM117
- ADAM118
- ADAM119
- ADAM120
- ADAM121
- ADAM122
- ADAM123
- ADAM124
- ADAM125
- ADAM126
- ADAM127
- ADAM128
- ADAM129
- ADAM130
- ADAM131
- ADAM132
- ADAM133
- ADAM134
- ADAM135
- ADAM136
- ADAM137
- ADAM138
- ADAM139
- ADAM140
- ADAM141
- ADAM142
- ADAM143
- ADAM144
- ADAM145
- ADAM146
- ADAM147
- ADAM148
- ADAM149
- ADAM150
- ADAM151
- ADAM152
- ADAM153
- ADAM154
- ADAM155
- ADAM156
- ADAM157
- ADAM158
- ADAM159
- ADAM160
- ADAM161
- ADAM162
- ADAM163
- ADAM164
- ADAM165
- ADAM166
- ADAM167
- ADAM168
- ADAM169
- ADAM170
- ADAM171
- ADAM172
- ADAM173
- ADAM174
- ADAM175
- ADAM176
- ADAM177
- ADAM178
- ADAM179
- ADAM180
- ADAM181
- ADAM182
- ADAM183
- ADAM184
- ADAM185
- ADAM186
- ADAM187
- ADAM188
- ADAM189
- ADAM190
- ADAM191
- ADAM192
- ADAM193
- ADAM194
- ADAM195
- ADAM196
- ADAM197
- ADAM198
- ADAM199
- ADAM200

```
data3<- read.csv('Expression Atlas.csv')

plot<- ggplot(data3, aes(x=Genes, y= Disease, fill=log2FC))+geom_tile()+
  scale_fill_gradient2(high = 'red', mid='white', low = 'blue') +
  theme(axis.text.x = element_text(angle = 90, size = 8, face =
'bold', hjust = 1), axis.text.y = element_text(size = 8, face =
'bold'), panel.background = element_rect(colour = 'black'),
  panel.grid.major = element_line(color = 'white'))

plot
```

