



**Figure 3.21.** The effectiveness factor decreases with increases in enzyme loading or with increases in particle diameter. Point A represents the value of the effectiveness factor for a particle radius of 10  $\mu\text{m}$  with an enzyme loading of  $100 \text{ mg}/\text{cm}^3$ , an enzyme activity of  $100 \mu\text{mol}/\text{min}$  per mg enzyme, a substrate diffusivity of  $5 \times 10^{-6} \text{ cm}^2/\text{s}$ , and a bulk substrate concentration tenfold higher than  $K_m$ .

When designing immobilized enzyme systems using a particular support, the main variables are  $V_m$  and  $R$ , since the substrate concentration,  $K_m$ , and  $D_e$  are fixed. The particle size ( $R$ ) should be as small as possible within the constraints of particle integrity, resistance to compression, and the nature of the particle recovery systems. The maximum reaction rate is determined by enzyme activity and concentration in the support. High enzyme content will result in high enzyme activity per unit of reaction volume but low effectiveness factor. On the other hand, low enzyme content will result in lower enzyme activity per unit volume but a high effectiveness factor. For maximum conversion rates, particle size should be small ( $D_p \leq 10 \mu\text{m}$ ) and enzyme loading should be optimized. As depicted in the example in Fig. 3.21,  $D_p \leq 10 \mu\text{m}$  and enzyme loadings of less than  $10 \text{ mg}/\text{cm}^3$  are required for high values of the effectiveness factor ( $\eta \geq 0.8$ ).

### Example 3.5

D. Thornton and co-workers studied the hydrolysis of sucrose at  $\text{pH} = 4.5$  and  $25^\circ\text{C}$  using crude invertase obtained from baker's yeast in free and immobilized form. The following initial velocity data were obtained with 408 units of crude enzyme (1 unit = quantity of enzyme hydrolyzing 1  $\mu\text{mol}$  of sucrose/min when incubated with 0.29 M sucrose in a buffer at pH 4.5 and  $25^\circ\text{C}$ ).