

## Solutions for 8.6 Exercises

### 1. Aspirin and stroke

```
(a) > stroke <- data.frame(treat=c("aspirin", "control"), y=c(63,43), n=c(78, 77))
> stroke$rate <- stroke$y/stroke$n
> stroke
      treat y  n      rate
1 aspirin 63 78 0.8076923
2 control 43 77 0.5584416
> aspirin.1 <- glm(rate ~ factor(treat)
+                  , family = binomial
+                  , weight = n, data=stroke)
> summary(aspirin.1)

Call:
glm(formula = rate ~ factor(treat), family = binomial, data = stroke,
     weights = n)
```

Deviance Residuals:

```
[1]  0  0
```

Coefficients:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	1.4351	0.2873	4.995	5.88e-07 ***
factor(treat)control	-1.2002	0.3677	-3.264	0.00110 **

(Dispersion parameter for binomial family taken to be 1)

```
Null deviance: 1.1354e+01 on 1 degrees of freedom
Residual deviance: 1.5321e-14 on 0 degrees of freedom
AIC: 13.132
```

Number of Fisher Scoring iterations: 3

There is a significant difference (at the 0.05 significance level) between the rate of stroke for the aspirin and control groups ( $P = 0.0011$ ).

```
(b) > exp(summary(aspirin.1)$coef[2,1])
[1] 0.3011204
> exp(summary(aspirin.1)$coef[2,1]+1.96*c(-1,1)*summary(aspirin.1)$coef[2,2])
[1] 0.1464681 0.6190666
```

The estimated odds of a stroke in the aspirin group are lower by a factor of 0.30 compared with the control group (95% CI: [0.146, 0.619]). Or, equivalently, the odds of stroke are higher in the control group by a factor of 3.32, compared with the aspirin group (95% CI: [1.62, 6.83]). This interval does not include 1 and so is consistent with the results of Lab 3, Question 6, where the confidence interval for the difference in the proportions did not include 0. Both indicate a significant difference between the aspirin and control groups.

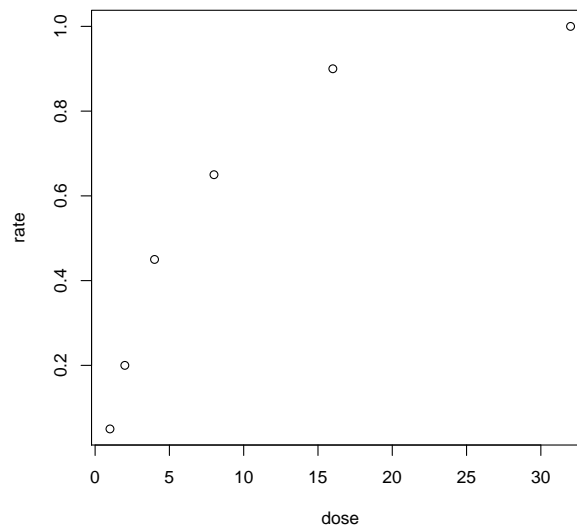
## 2. Control of budworm

(a) Enter the data

```
> moths <- data.frame(dose = c(1, 2, 4, 8, 16, 32),  
+ dead = c(1, 4, 9, 13, 18, 20), total = rep(20, 6))  
> moths$rate <- moths$dead / 20
```

Firstly, examine a plot of the mortality rate of the moths as a function of dose.

```
> plot(rate ~ dose, data = moths)
```



Fit a logistic regression model and look at the summary:

```
> moths.1 <- glm( rate ~ dose  
+                 , family = binomial  
+                 , weight = total  
+                 , data = moths )
```

```
> summary(moths.1)
```

Call:

```
glm(formula = rate ~ dose, family = binomial, data = moths, weights = total)
```

Deviance Residuals:

1	2	3	4	5	6
-1.5729	-0.0954	1.1798	0.3631	-0.7789	0.1426

Coefficients:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-1.92771	0.40195	-4.796	1.62e-06 ***
dose	0.29723	0.06254	4.752	2.01e-06 ***

---

Signif. codes: 0 \*\*\* 0.001 \*\* 0.01 \* 0.05 . 0.1 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 71.138 on 5 degrees of freedom  
Residual deviance: 4.634 on 4 degrees of freedom  
AIC: 22.981

Number of Fisher Scoring iterations: 5

The very small  $P$ -value for the Wald test indicates that the estimate of the coefficient for dose is significantly different to 0.

(b) `> exp(0.29723)`

```
[1] 1.346125
```

For every unit increase in dose, the odds of a moth dying are estimated to increase by a factor of 1.35, 35%.

(c) The coefficients are:

```
> moths.1$coef
```

```
(Intercept)      dose  
-1.9277147    0.2972343
```

We solve  $0 = -1.92771 + 0.29723 \times \text{dose}$

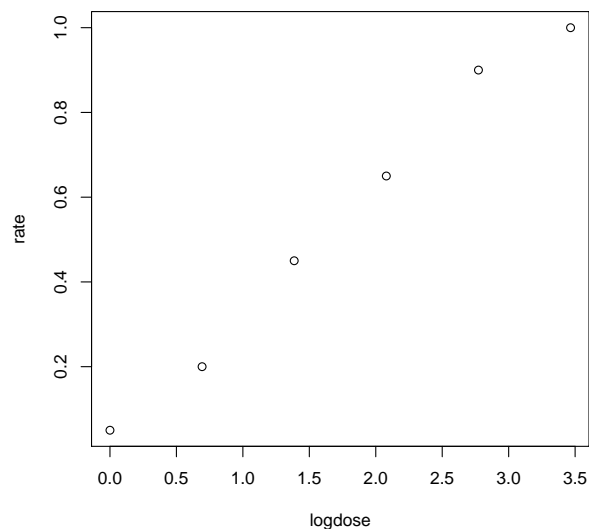
The LD50 is:

```
> 1.92771/0.29723
```

```
[1] 6.485584
```

(d) `> moths$logdose<-log(moths$dose)`

```
> plot(rate ~ logdose, data = moths)
```



```
> moths.2 <- glm( rate ~ logdose
+               , family = binomial
+               , weight = rep( 20, 6)
+               , data = moths )
> summary(moths.2)

Call:
glm(formula = rate ~ logdose, family = binomial, data = moths,
    weights = rep(20, 6))

Deviance Residuals:
    1      2      3      4      5      6 
-0.12505  0.30463  0.22204 -0.71011 -0.02679  1.10375 

Coefficients:
              Estimate Std. Error z value Pr(>|z|)
(Intercept)  -2.8186     0.5480  -5.143 2.70e-07 ***
logdose       1.8163     0.3059   5.937 2.91e-09 ***
---
Signif. codes:  0 *** 0.001 ** 0.01 * 0.05 . 0.1 1

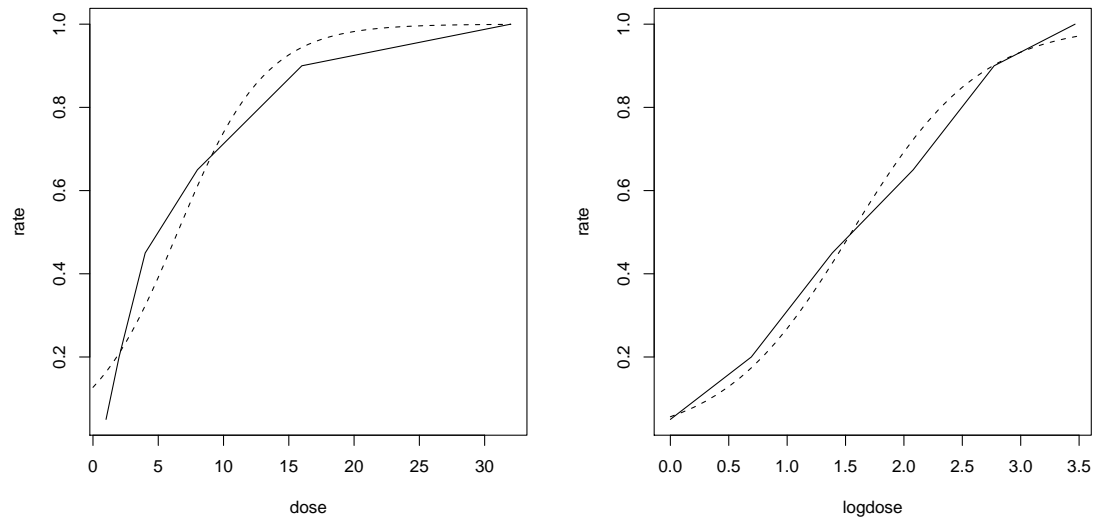
(Dispersion parameter for binomial family taken to be 1)

Null deviance: 71.1376  on 5  degrees of freedom
Residual deviance:  1.8810  on 4  degrees of freedom
AIC: 20.228

Number of Fisher Scoring iterations: 4

The P-value is 3 orders of magnitude smaller, and additionally, the residual deviance of 1.88 is smaller than the residual deviance resulting from fitting dose (4.63), so fitting log(dose) provides a better fit. The following plot confirms the better fit:

> par(mfrow = c(1, 2))
> plot(rate ~ dose, type = "l", data = moths)
> dse <- seq(0, 32, 0.5)
> lines(dse, predict(moths.1, newdata = data.frame(dose = dse),
+   type = c("response")), lty = 2)
> plot(rate ~ logdose, type = "l", data = moths)
> dse <- seq(0, 3.5, 0.1)
> lines(dse, predict(moths.2, newdata = data.frame(logdose = dse),
+   type = c("response")), lty = 2)
```



### 3. Powdery mildew on broccoli

```
(a) > broccoli <- data.frame(treat=c(rep("control",5),
+ rep("fungicide1",5), rep("fungicide2",5)),
+ alive=c(3,2,2,1,1,10,8,15,14,8,17,14,13,10,16),
+ emerged=c(17,15,16,11,15,16,11,18,16,12,18,15,16,13,16))
> broccoli$rate <- broccoli$alive/broccoli$emerged
> broccoli
```

	treat	alive	emerged	rate
1	control	3	17	0.17647059
2	control	2	15	0.13333333
3	control	2	16	0.12500000
4	control	1	11	0.09090909
5	control	1	15	0.06666667
6	fungicide1	10	16	0.62500000
7	fungicide1	8	11	0.72727273
8	fungicide1	15	18	0.83333333
9	fungicide1	14	16	0.87500000
10	fungicide1	8	12	0.66666667
11	fungicide2	17	18	0.94444444
12	fungicide2	14	15	0.93333333
13	fungicide2	13	16	0.81250000
14	fungicide2	10	13	0.76923077
15	fungicide2	16	16	1.00000000

```
> tapply(broccoli$rate,broccoli$treat,mean)
      control fungicide1 fungicide2 
0.1184759  0.7454545  0.8919017
```

```
(b) > broccoli.1 <- glm(rate ~ treat,
+                        family = binomial,
+                        weight = emerged,
+                        data = broccoli )
> summary(broccoli.1)

Call:
glm(formula = rate ~ treat, family = binomial, data = broccoli,
     weights = emerged)

Deviance Residuals:
    Min       1Q   Median       3Q      Max
-1.33513  -0.68944   0.04118   0.68408   1.86087

Coefficients:
              Estimate Std. Error z value Pr(>|z|)
(Intercept)    -1.9772     0.3557  -5.559 2.71e-08 ***
treatfungicide1  3.0941     0.4475   6.915 4.69e-12 ***
treatfungicide2  4.1462     0.5155   8.043 8.79e-16 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 127.743  on 14  degrees of freedom
Residual deviance: 12.010  on 12  degrees of freedom
AIC: 53.03
```

Number of Fisher Scoring iterations: 4

The very small  $P$ -values show a highly significant difference between the control and each of the two fungicides, consistent with the means from part (a).

```
(c) > exp(summary(broccoli.1)$coef[2, 1])
[1] 22.0679

> exp(summary(broccoli.1)$coef[2, 1] + 1.96 * c(-1, 1)
+      * summary(broccoli.1)$coef[2, 2])
[1] 9.180438 53.046738
```

#### 4. Aboriginal deaths in custody

```
(a) > deaths <- read.csv("../data/deaths.csv")
> deaths <- data.frame(Year = rep(c(1990,1995),2),
+ Indigenous = rep(c("Yes","No"),each=2),
+ Prisoners = c(2041,2907,12264,14501),
+ Deaths = c(6,17,27,42),
+ Population = c(168317,190438,13141817,13995940))
> deaths$rate <- deaths$Deaths/deaths$Prisoners
> deaths
```

	Year	Indigenous	Prisoners	Deaths	Population	rate
1	1990	Yes	2041	6	168317	0.002939735
2	1995	Yes	2907	17	190438	0.005847953
3	1990	No	12264	27	13141817	0.002201566
4	1995	No	14501	42	13995940	0.002896352

```
> deaths.1<- glm( rate ~ Year
+                 , family = binomial
+                 , weight = Prisoners
+                 , data = deaths )
> summary(deaths.1)
```

Call:

```
glm(formula = rate ~ Year, family = binomial, data = deaths,
    weights = Prisoners)
```

Deviance Residuals:

	1	2	3	4
	0.5715	2.0662	-0.2450	-1.0476

Coefficients:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-159.61579	86.77194	-1.839	0.0658 .
Year	0.07716	0.04353	1.772	0.0763 .

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 8.9920 on 3 degrees of freedom  
Residual deviance: 5.7532 on 2 degrees of freedom  
AIC: 28.797

Number of Fisher Scoring iterations: 4

Although the mortality rate increases from 1990 to 1995 in both groups of prisoners, the increase is not significant at the 0.05 level the  $P$ -value from the test.

```
(b) > deaths.2 <- glm( rate ~ Indigenous
+                 , family = binomial
```

```
+           , weight = Prisoners
+           , data = deaths )
> summary(deaths.2)

Call:
glm(formula = rate ~ Indigenous, family = binomial, data = deaths,
    weights = Prisoners)
```

Deviance Residuals:

1	2	3	4
-1.2175	0.9140	-0.8434	0.7413

Coefficients:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-5.9582	0.1205	-49.428	<2e-16 ***
IndigenousYes	0.5916	0.2413	2.452	0.0142 *

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 8.9920 on 3 degrees of freedom  
Residual deviance: 3.5784 on 2 degrees of freedom  
AIC: 26.622

Number of Fisher Scoring iterations: 4

From the  $P$ -value there was a significantly greater mortality rate for indigenous prisoners at the 0.05 significance level.

- (c) The appropriate quantity is an odds ratio. For indigenous vs non-indigenous prisoners:

```
> exp(summary(deaths.2)$coef[2, 1])
[1] 1.806836
> exp(summary(deaths.2)$coef[2, 1] + 1.96 * c(-1, 1) * summary(deaths.2)$coef[2,
+ 2])
[1] 1.126017 2.899295
```

Interpretation: for indigenous prisoners, the odds of death in custody are 1.81 times greater than for non-indigenous prisoners (95% CI:[1.13, 2.90]).

- (d) 

```
> p1 <- 6/2041
> p2 <- 27/12264
> (odds.ratio <- (p1/(1 - p1))/(p2/(1 - p2)))
[1] 1.336282
> (relative.risk <- p1/p2)
[1] 1.335293
```

These are very similar, which is the case when  $p_1$  and  $p_2$  are very small, as they are here.