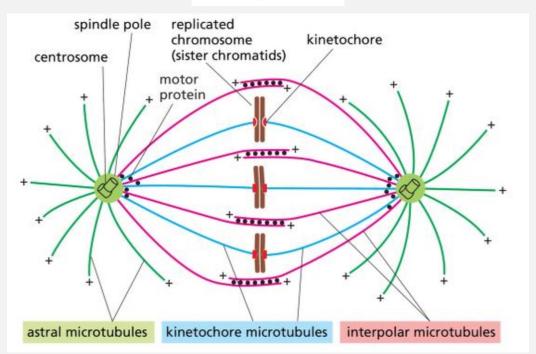
# Characterization of Motor Functions and Nuclear Localization of Truncated CtCIN8

By: Daniel Kim Jiarong Liang

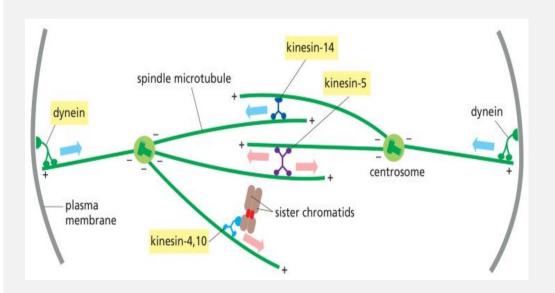
# Kinesin-5 Motor Proteins are critical for mitotic spindle assembly & elongation during Mitotic Anaphase

### Anaphase



Panel 17-1, Bruce Albert, et. al., Molecular Biology of the Cell, 5/d, Garland Science, 2008, Page 1073

### Homotetrameric Cin8 facilitates the antiparallel movement of Microtubules



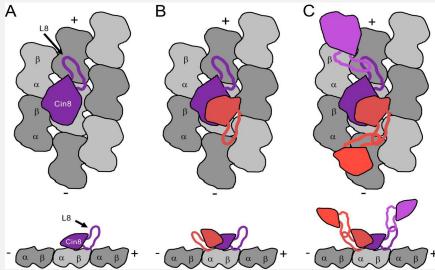
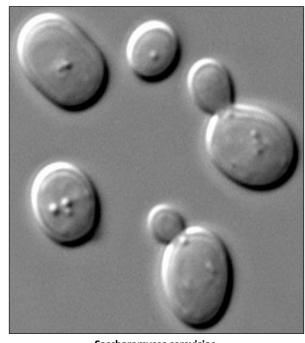
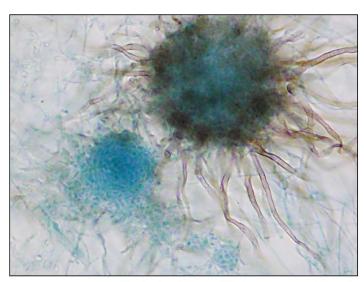


Figure 17-30, Bruce Albert, et. al., Molecular Biology of the Cell, 5/d, Garland Science, 2008, Page 1077 Figure 8. K. Bell, et. al. *Jol. Bio. Chem.* Vol 292 (35), 2007

# Structural and Functional Conservation of Kinesin-5 CIN8 in Saccharomyces Cerevisiae & Chaetomium Thermophilum



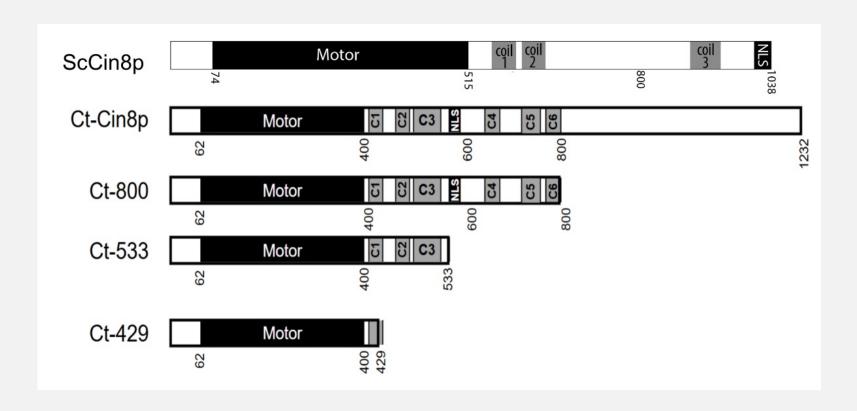
Saccharomyces cerevisiae Budding yeast



Chaetonium thermophilum Filamentous fungus

Prof. Isabelle Leblanc, Fa2024 MCB C110L, Lecture 1, 2024.

# Nuclear Localization Signal located in different domains of CtCin8 & ScCin8



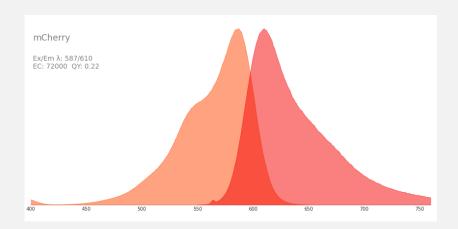
Prof. Isabelle Leblanc, Fa2024 MCB C110L, Lecture 1 & 2, 2024.

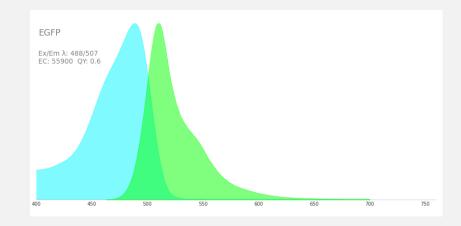
Visualizing Protein Localization

# DDY904 Strain Contains mCherry (RFP) - Tagged Tub Gene

#### DDY904:

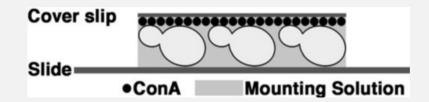
MATα ura3-52 leu2-3,112 his3Δ200 lys2-802 mCherry (RFP)-tagged tubulin





RFP - mCherry, FBBase.org, 2004. GFP - EGFP, FBBase.org, 1996

### Visualizing Subcellular Localization



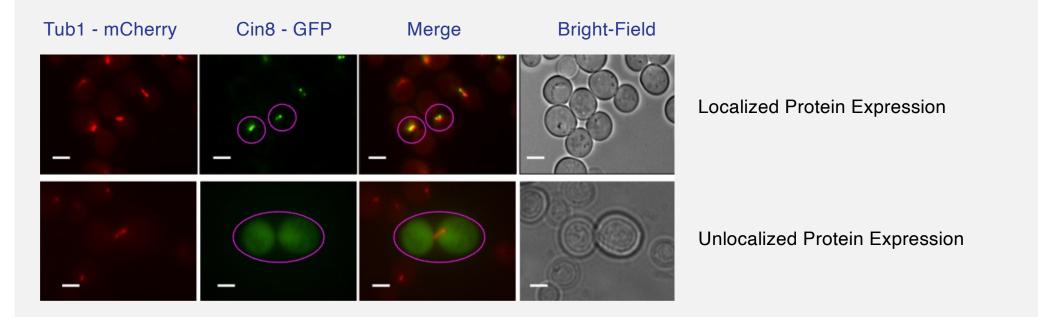
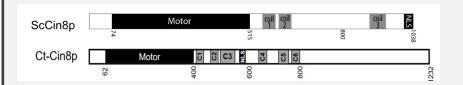
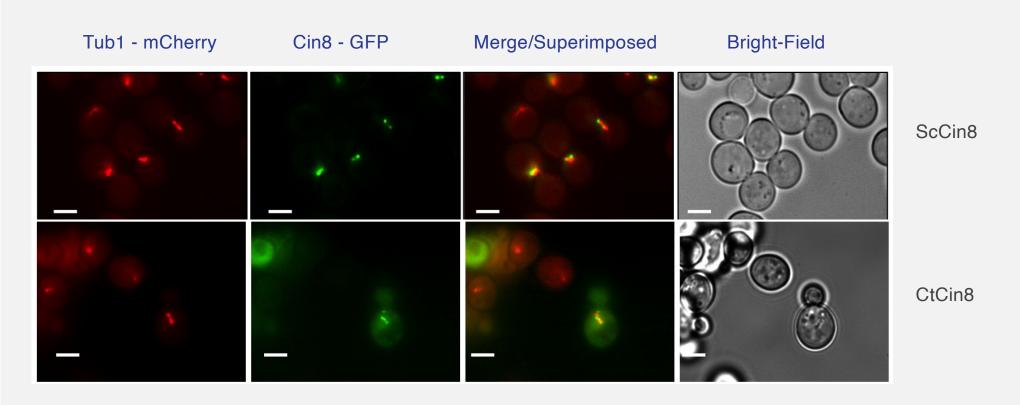


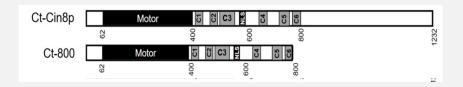
Figure 1, Cytoskeleton, C.Sing, et. al., Research Gate, Jan. 2022, Page 88

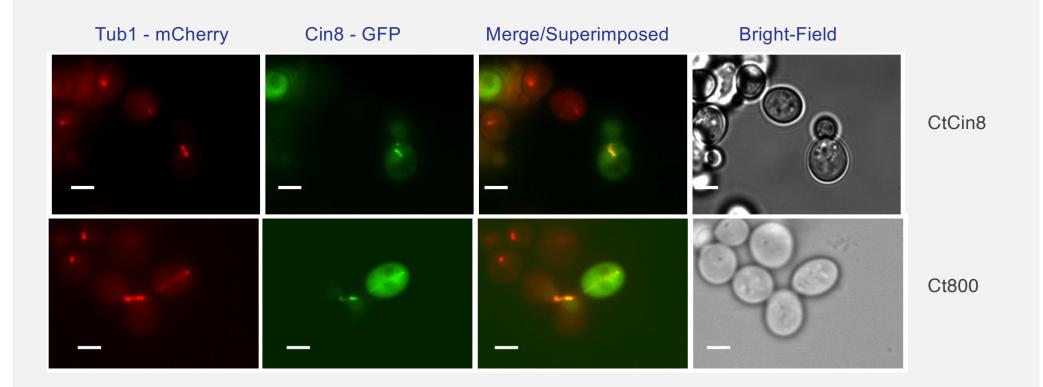
Full-Length CtCin8 exhibits high degree of localization.



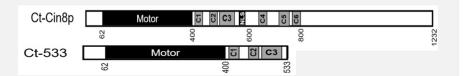


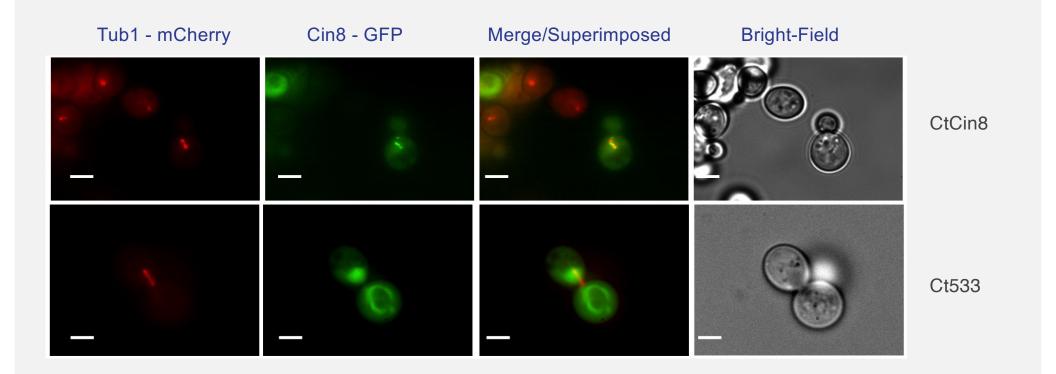
Ct800 demonstrates a slightly lower degree of localization than Full-Length Cin8.





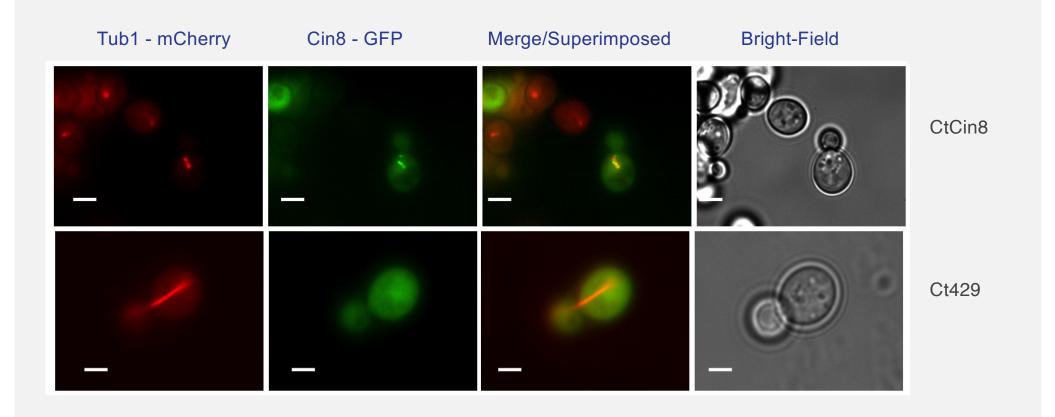
Ct533 exhibits much lower levels of Cin8-GFP localization than full-length CtCin8.





Ct429 displays virtually no Cin8-GFP localization





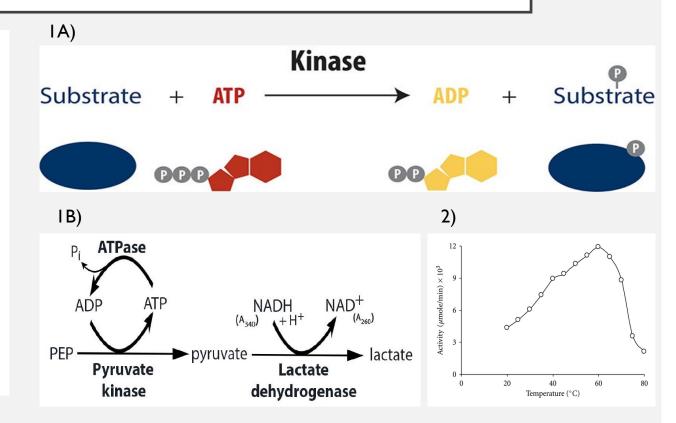
Malachite Assay to measure enzyme Kinetics

# Measuring Kinetic Properties of Cin8

• 1)

- A) ATPase Activity
- B) NADH Coupled Assay

2) Temperature Dependence Assay



BellBrookLabs, Activity Assays, Dec. 2018

R. Yadav., et. al., L. aegytiaca, ResearchGate.com, Jan. 2011

K. Sozanski, et. al., Motor Domain Diffusion, ResearchGate.com, Nov. 2015

# Column Chromatography

#### Purification Wash Buffer (W1, W3, & W4):

50 mM Hepes pH: 7.5

150 mM KCI

5 mM MgCl<sub>2</sub>

20 mM Imidazole

10 % Glycerol

1 mM DTT

0.1 mM ADP pH 7.0

#### **Purification Elution Buffer:**

50 mM Hepes pH: 7.5

150 mM KCI

5 mM MgCl<sub>2</sub>

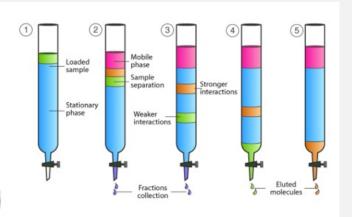
250 mM Imidazole

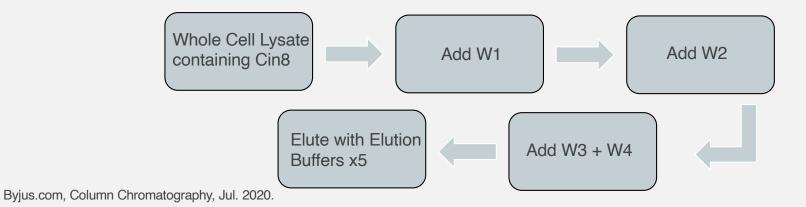
10 % Glycerol

1 mM DTT

0.1 mM ADP

Wash Buffer (W2): except 1 mM ATP replacing ADP (same as Wash buffer above)





# Malachite Green as a Phosphate Sensitive Dye

Upon addition of Malachite Green, the presence of Phosphate makes initially yellow dye turn green.

12 
$$H_2MoO_4$$
 + Malachite green = Yellow ( $\lambda_{max}$  = 446nm)  
 $H_2PO_4(MoO_3)_{12}$  + Malachite green = Green ( $\lambda_{max}$  = 650nm)

Prof. Isabelle Leblanc, Fa2024 MCB C110L, Lecture 1 & 2, 2024.

# Quenching Continuous Assay EA & SA

#### 34% Sodium Citrate

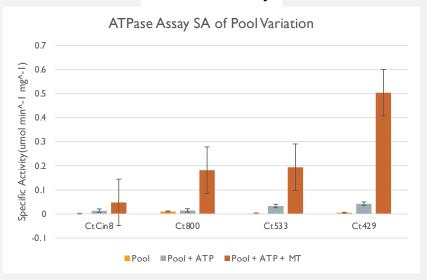
Prevents further color development of the continuous Malachite Green Assay

Enzyme Activity (**EA**) =  $\mathbf{U} = \mathbf{A}/\mathbf{I}\boldsymbol{\varepsilon} * \mathbf{V} / \mathbf{time}$ ( $\boldsymbol{\varepsilon}$ \_Malachite Green = 99.4 M^-1 \* C^-1)

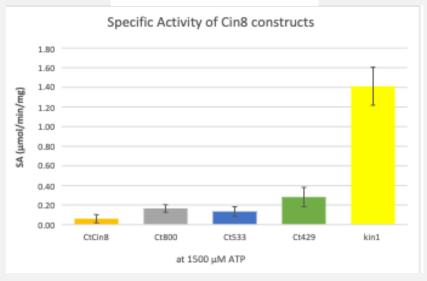
Specific Activity (SA) = EA/mass

# ATPase/NADH Assay

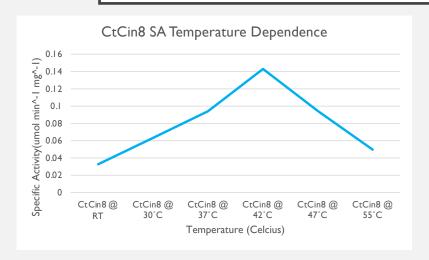
### ATPase Assay

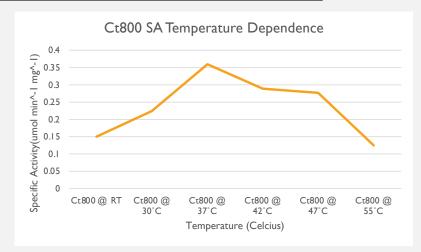


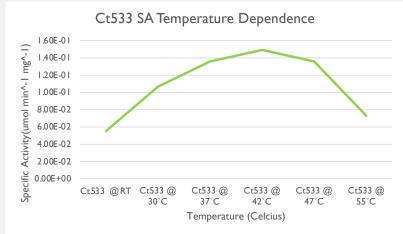
#### NADH Coupled Assay

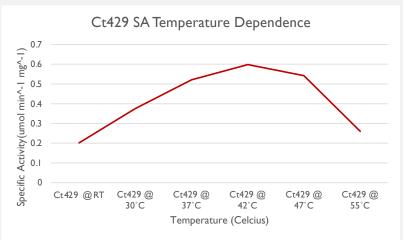


# Temperature Binding Assay









Rescuing Growth in Auxin and Cincreasin Conditions

# Y119 contained Aid-6Flag Tag and Kip1 \Delta

#### Y119

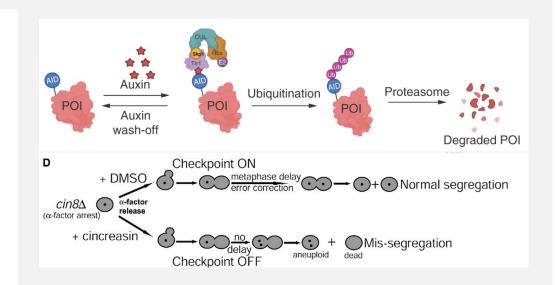
MATα CIN8-AID-6FLAG::pHyg TIR1::LEU2 kip1Δ::HIS3 ura3-52 leu2-3, 112, lys1-801 his3Δ200

#### Auxin

 Recognizes & Binds to AID sequence to induce degradation via ubiquitination

#### Cincreasin

Removes the cell cycle checkpoint for spindle assembly



K. Phanindhar, et. al., Biotechniques, Vol 74. (4), 2023 R. Dorer, et. al., Current Biology, Vol. 15, Jun. 2005, Pg 1073

# Phenotype Analysis of Cin8 Predictions

+: Growth

-: No Growth

?: Uncertain Growth

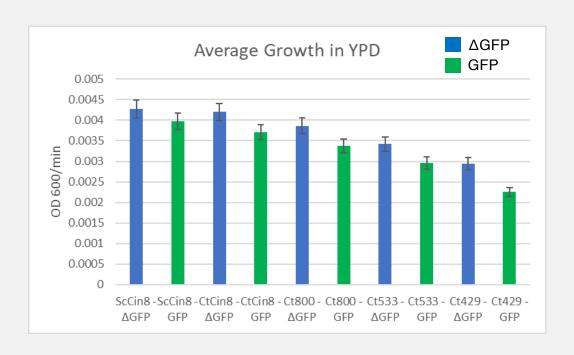
Plasmid	YPD	Auxin	Cincreasin	Auxin + Cinc.
ScCin8	+	+	+	+
CtCin8	+	?	?	?
Ct800	+	?	?	?
Ct533	+	?	?	?
Ct429	+	-	?	-

### Plate Reader's Results



- GFP/ΔGFP Cin8 Incubated in YPD
- Experiments done in Triplicate
- Reading Taken per 15 minutes:
- Average Growth per 2hr of the fastest growth period

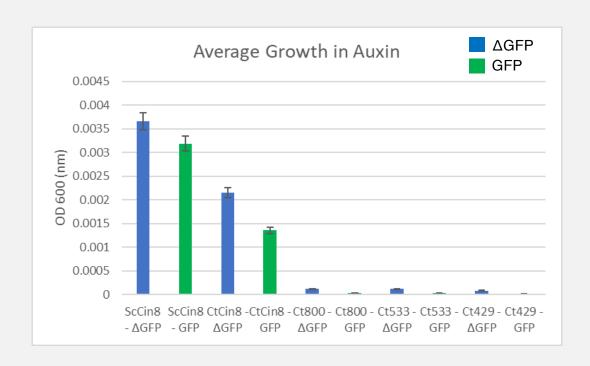
### Average Growth in YPD exhibits Size Dependent Growth



In YPD setting, Cin8 Constructs demonstrate size dependent growth

↑ Genome Size = ↑ Growth

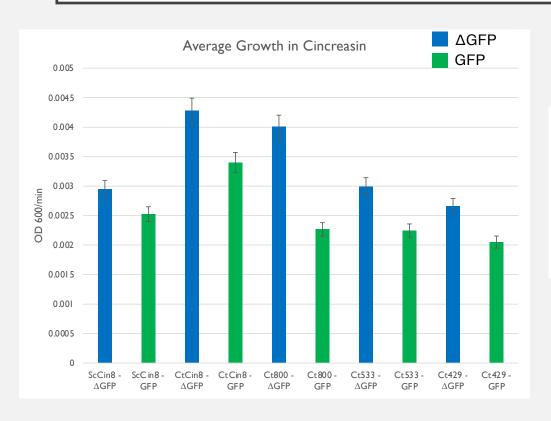
# Auxin influences growth of full length ScCin8 & CtCin8 less than YPD



No significant impact to full length constructs of ScCin8 and CtCin8

Truncated constructs exhibit less growth in YPD

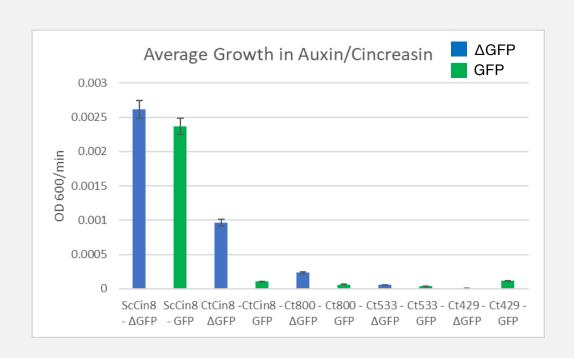
# Cincreasin influences growth of full length ScCin8 & CtCin8 less than Auxin



No significant impact to full length constructs of ScCin8 and CtCin8

Truncated constructs exhibit less growth in YPD but more than Auxin

# Average Growth in Aux/Cin exhibits Similar Growth Pattern as Auxin-only conditions



No significant impact to full length constructs of ScCin8 and CtCin8

Truncated constructs similar growth to Auxin only conditions

# **Future Direction**

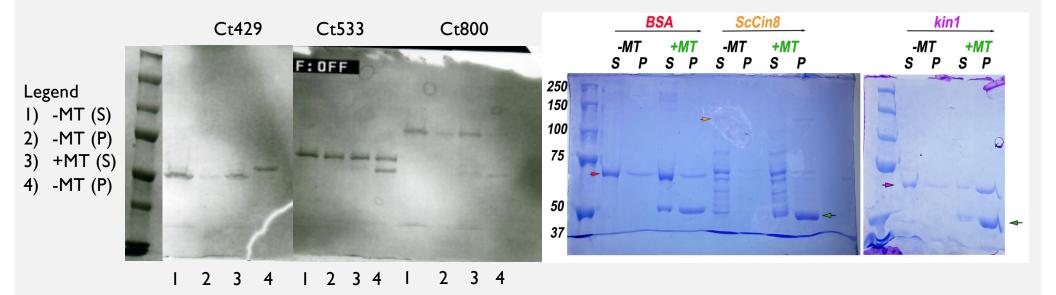
- 3D Crystallization of Truncated CtCin8
  - How do the missing coiled domains affect CtCin8 folding?

Are there any conserved regions to ScCin8?

Q/A

# **APPENDIX**

# MT Binding Assay



Note: No strong representation of Full Length CtCin8