

Workflows for Visualization

Introduction

Visual analogies often play a key role in research, helping us articulate hypotheses and understand results of hypothesis tests. Yet, while hypothesis testing is a core component of science education, visualization, and, more generally, the acts of perception and imagination that necessarily precede hypothesis testing, do not receive as much attention in our curricula, despite their critical importance.

For one thing, the basics of making good figures, which are explicitly known and practiced by most successful scientists, are rarely formally taught. Indeed, I believe the association between knowing how to make a good figure and being a successful scientist is causal.

More generally, as biologists we are often working with complex systems. Visualizing complex systems requires us to bring into focus specific emergent properties of a system, by mapping to a simpler context, and by leaving most things out. In this sense, visualizing complex systems is similar to building mathematical models. Indeed visualizing complex systems requires some mathematics, and often catalyzes further quantitative analyses.

Here's a cute argument: a huge component of our brains is evolved for spatial reasoning - for instance, to help us find food and not become food for another creature. Think of this like the GPC (graphics processing card) of your brain-computer. Your internal GPC is so much more powerful than your abstract processing abilities (the CPU of your brain-computer). We can do better science by working better with our GPCs.

Example of a basic visualization workflow

Here we chronicle the journey from a simple dataset to an acceptable figure, in base R.

Setup

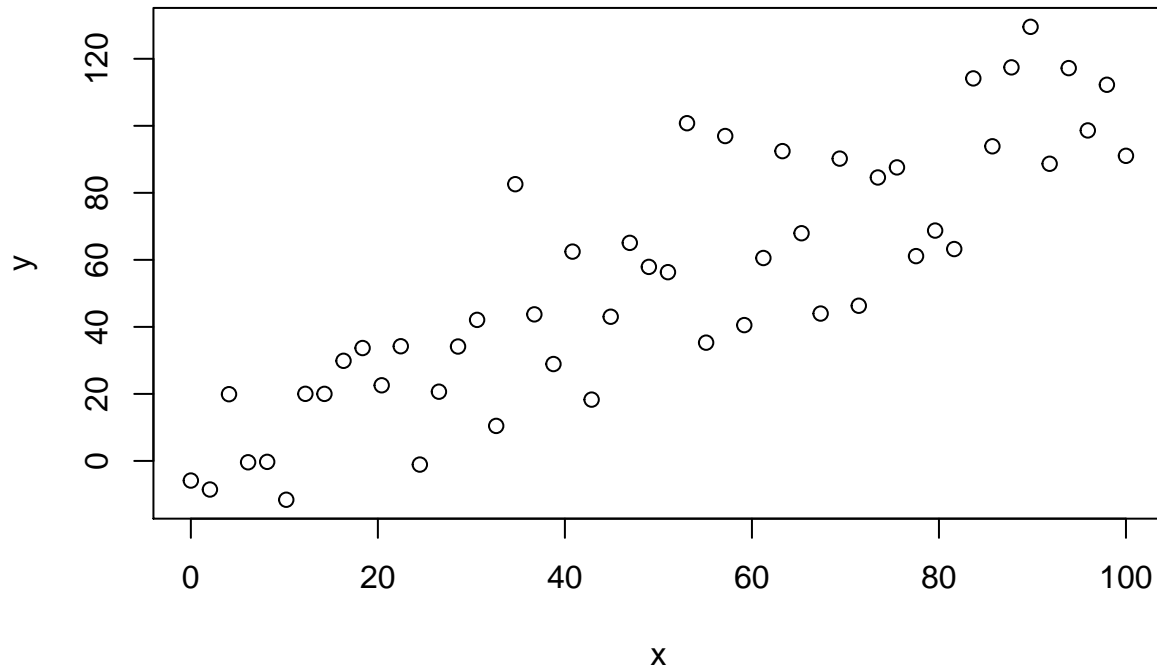
We will start by generating some data, as an additive combination of a linear "signal" and some white noise

```
n <- 50
a <- 1
b <- 2
sig <- 20
noise <- rnorm(n, 0, sig)

x <- seq(0, 100, length.out = n)
signal <- a * x + b
y <- signal + noise
```

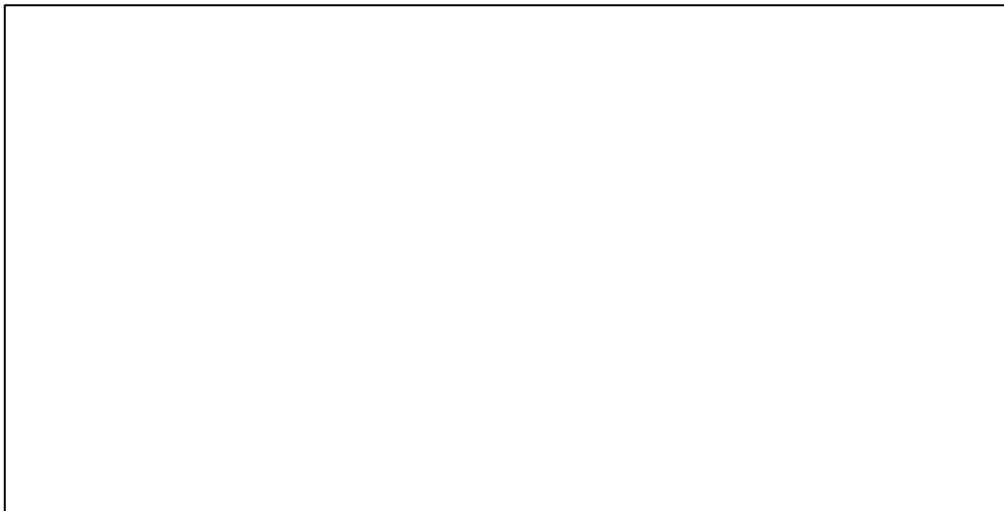
At this point it is worth spending some time in the help files for `plot()` and `par()` if you have not already. This is just to get a sense of the potential to control things, and how the basic control process works. Most people need to continually refer to these help files as they work on specific applications.

```
plot(x, y)
```



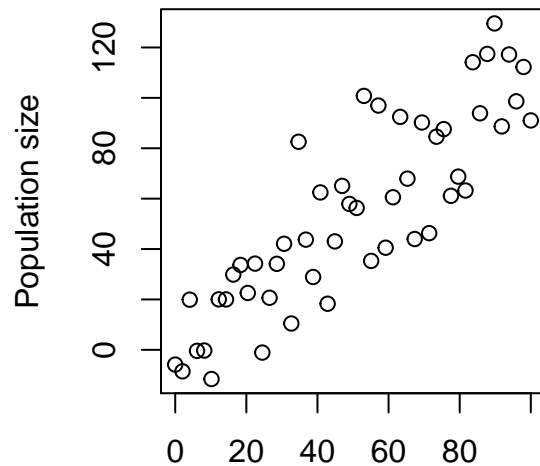
Starting a plot from the VERY beginning - (i.e. all default elements turned off) - if you want to control everything manually.

```
plot(x, y,  
     type = "n", # do not even plot the data  
     #bty = "n", # box type is "n" means don't draw a box around the plot  
     xaxt = "n", # similarly, don't make an x axis...  
     yaxt = "n", # "... or a y axis"  
     xlab = "", # and axis labels are blank  
     ylab = ""  
)
```



Before tweaking optional things, there are some minimum standards to take care of. Every figure should have informative axis labels. And these need to be large enough to be read when the figure appears in a published article. To this end, it can be helpful to control the size of a figure right from the start. If you have not already, spend some time in the help files for `plot()` and `par()`.

```
par(fin = c(4,4))           # figure dimensions (width, height)
par(mai = c(1,1,1,1))      # plot margins (bottom,left,top,right)
plot(x, y,
     xlab = "Years since Mastodon reintroduction",
     ylab = "Population size")
```



Years since Mastodon reintroduction

By labelling the axes I have just realized there are y values that make no sense given what y is: population size. This is a simplistic example of how disciplined practices in making figures (e.g. always having informative axis labels from the start) can improve the whole scientific process: now we have an opportunity to improve our data stream by figuring out what to do with our negative y values. For our purposes here, let's just remove them:

```
y[y < 0] <- NA
```

Now let's start work on minimizing the amount of effort a reader's brain has to do to "get" the figure. There are some initial things that apply to every figure that we can do right away. Then in the next section we can look at making specific messages come forward.

Little things can go a long way. Custom tick marks are usually needed to make professional-grade figures. To do that, we need just enough tick labels to orient the reader to the "space" of the plot and help them quickly measure distances; any tickmarks beyond that just become visual noise the reader has to work to filter out. We start by turning off the default axes. Then we add our own, with axis limits hardcoded.

From a reproducibility standpoint, hard-coding is a liability. The alternative is to write code that determines what the best limits should be on the fly, but we will not be distracted by that now.

```
par(fin = c(4,4))
par(mai = c(1,1,1,1))
plot(x, y,
     xlab = "Years since Mastodon reintroduction",
     ylab = "Population size",
     xaxt = "n",
     yaxt = "n",
     xlim = c(0, 100),      # hardcoded axis limits...
```

```

ylim = c(0, 120)
)
axis(1, c(0, 50, 100))    # ... and tick locations
axis(2, c(0, 60, 130))

```



Tune for impact

Now that we have something that meets minimum formatting requirements we can ask: what are we trying to say with this figure and how can we make that pop? Let us say our main message here is that the Mastodon population is increasing in a predictable way. That would be the first line of a figure legend: “i.e. Figure 1: Following reintroduction in 2023, the Mastodon population has increased in a predictable way.” This is also the first sentence you would say in a presentation of this slide. The task at hand is to make the visual say that loud and clear. We need a regression line.

```
fit <- lm(y ~ x)
```

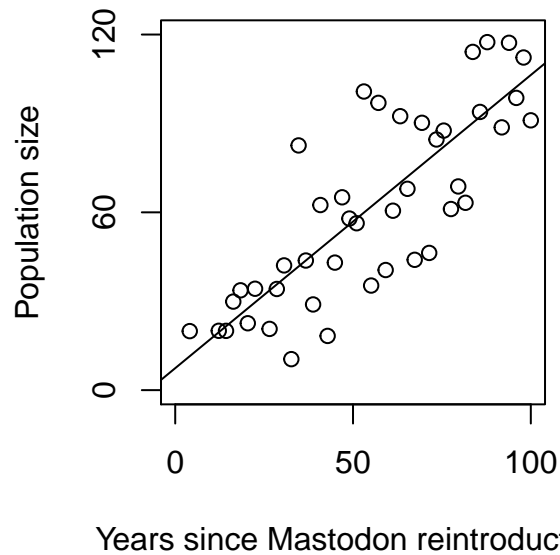
We could add this to the figure a number of ways, for example, using the function `abline()` and supplying the fitted model as input.

```

par(fin = c(4,4))
par(mai = c(1,1,1,1))
plot(x, y,
     xlab = "Years since Mastodon reintroduction",
     ylab = "Population size",
     xaxt = "n",
     yaxt = "n",
     xlim = c(0, 100),
     ylim = c(0, 120)
)
axis(1, c(0, 50, 100))
axis(2, c(0, 60, 120))

abline(fit) # add linear regression line to an existing plot

```



We get a lot more control of how the predictions are displayed if we generate an additional dataset that is the prediction, then plot that. For example, using `abline()` causes the fitted line to extend beyond the data, which is not only statistically bad (we are typically interpolating, not extrapolating when fitting a linear regression model), but also doesn't make biological or physical sense (we can't go to negative time since the mastodon reintroduction).

First generate the predictions:

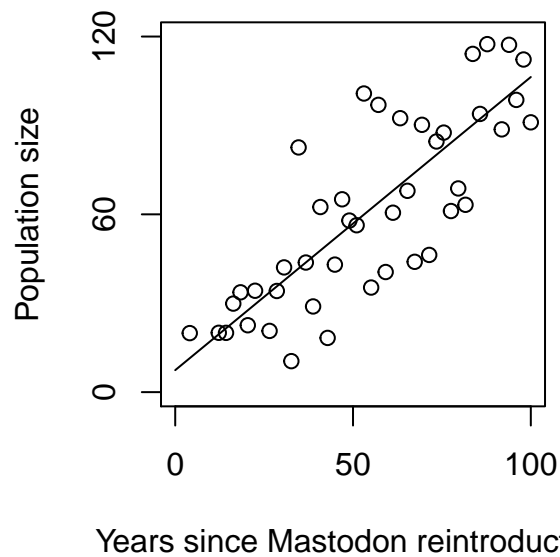
```
npred <- 10
xpred <- seq(min(x), # points at which we want to predict
             max(x),
             length.out = npred)

ypred <- predict(fit, # predicting corresponding y values and se
                 newdata = data.frame(x = xpred),
                 se.fit = TRUE)
```

Then plot:

```
par(fin = c(4,4))
par(mai = c(1,1,1,1))
plot(x, y,
     xlab = "Years since Mastodon reintroduction",
     ylab = "Population size",
     xaxt = "n",
     yaxt = "n",
     xlim = c(0, 100),
     ylim = c(0, 120)
)
axis(1, c(0, 50, 100))
axis(2, c(0, 60, 120))

lines(xpred, ypred$fit)
```



This also allows us to make the plot more informative and visually appealing:

```
# Original code, modified to not plot the points at first
par(fin = c(4,4))
par(mai = c(1,1,1,1))
plot(x, y,
      type = "n",
      xlab = "Years since Mastodon reintroduction",
      ylab = "Population size",
      xaxt = "n",
      yaxt = "n",
      xlim = c(0, 100),
      ylim = c(0, 120)
)
axis(1, c(0, 50, 100))
axis(2, c(0, 60, 120))

# Draw a polygon for the confidence envelope
xpoly <- c(xpred, rev(xpred))
ypoly <- c(ypred$fit + 5 * ypred$se.fit,
           rev(ypred$fit - 5 * ypred$se.fit))

polygon(x = xpoly, y = ypoly,
        border = NA,
        col = grey(0.8))

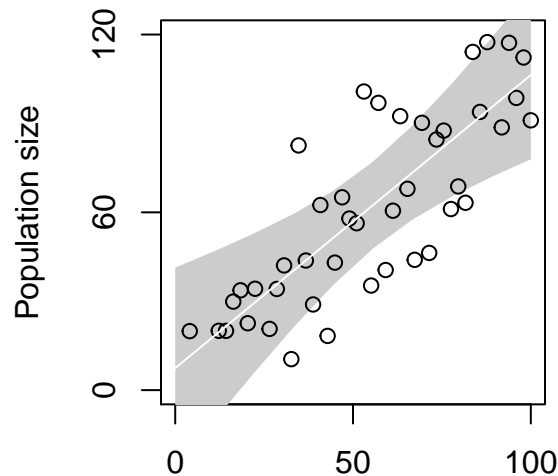
# Add in the points and line
points(x,y)
lines(xpred, ypred$fit, col = 'white')
```

#first plot with points missing

#add confidence envelope as bottom layer

#then data

#then trendline



Years since Mastodon reintroduction

That takes care of our primary message: “Mastodon population is increasing in a predictably-linear way.” What about secondary messages? Suppose we are interested in what caused the population to grow especially quickly or slowly in some years. We can use symbol color and shape to highlight points that are outside the confidence envelope.

To do that we need to have the model prediction and standard error for each observation point, rather than at 10 evenly spaced points as we had before.

```
yhat <- predict(fit, newdata = data.frame(x = x), se.fit = T)
```

Now identify points that are above (boom years) and below (bust years) the confidence envelope.

```
is_boom_year <- y > yhat$fit + 5*yhat$se.fit
is_bust_year <- y < yhat$fit - 5*yhat$se.fit
```

Use symbology to reinforce the message. To do that we first specify size, shape and color as vectors, mapping from data. Then we call the plot function. It’s generally wiser to do this outside rather inside the plot function call because the latter is harder to read and debug.

```
# Default vectors for symbology
cex <- rep(1, n)      # size
col <- rep("black", n) # color
pch <- rep(21, n)      # shape. See ?points

# Change to map non-default symbology to data
cex[is_boom_year || is_bust_year] <- 2
col[is_boom_year] <- "green"
col[is_bust_year] <- "red"
pch[is_boom_year] <- 2
pch[is_bust_year] <- 6
```

The most important thing is that it pops: do not rely on subtle differences in color or shape to get across key results. One way to achieve that is to simultaneously alter multiple features, such as changing shape, color and size together (but be careful not to overdo this and create confusion when working with multiple legends, and be sure to remain consistent across figures).

```
# Vectors for symbology
cex <- rep(0.8, n)      # size
```

```

col <- rep("black", n) # color
bg <- rep("white", n)  # background color
pch <- rep(21, n)      # shape. See ?points

# Change symbology to highlight features in data
cex[is_boom_year || is_bust_year] <- 1.3
bg[is_boom_year] <- "seagreen"
bg[is_bust_year] <- "orange"
col[is_boom_year] <- "darkgreen"
col[is_bust_year] <- "red"
pch[is_boom_year] <- 24
pch[is_bust_year] <- 25

# Make plot first without points
par(fin = c(4,4))
par(mai = c(1,1,1,1))
plot(x, y,
     type = "n",
     xlab = "Years since Mastodon reintroduction",
     ylab = "Population size",
     xaxt = "n",
     yaxt = "n",
     xlim = c(0, 100),
     ylim = c(0, 120)
    )
axis(1, c(0, 50, 100))
axis(2, c(0, 60, 120))

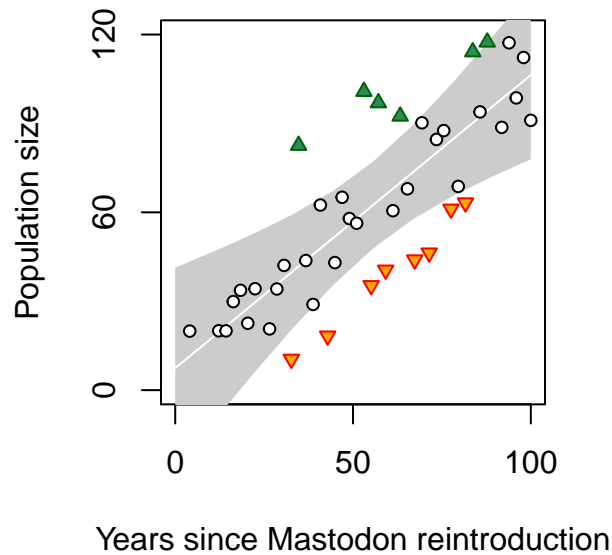
# Draw a polygon for the confidence envelope as bottom layer
xpoly <- c(xpred, rev(xpred))
ypoly <- c(ypred$fit + 5 * ypred$se.fit,
          rev(ypred$fit - 5 * ypred$se.fit))

polygon(x = xpoly, y = ypoly,
        border = NA,
        col = grey(0.8))

# Add trendline
lines(xpred, ypred$fit, col = 'white')

# Add in the points, with their symbologies pre-specified above (no calculations here)
points(x,y,
       pch = pch,
       col = col,
       bg = bg,
       cex = cex)

```

Now this figure says:

1. “Mastodon population is increasing in a predictably linear way.”
2. “We are going to focus on the years where growth was particularly fast or slow relative to those predictions.”

Other graphs in your paper should use the same color scheme and symbology to denote analyses pertaining to the boom and bust years.

Final touches

There are still a few things that might make this figure just a little more visually appealing:

1. The length of the axis tickmarks.
2. The size of the axis labels relative to the axis titles.
3. The space between the axis tickmarks and the axis labels
4. The space between the axis titles and the box.
5. A solid box (that doesn't have the grey of the confidence overlapping it).

```
# Vectors for symbology
cex <- rep(0.8, n)      # size
col <- rep("black", n)   # color
bg <- rep("white", n)    # background color
pch <- rep(21, n)        # shape. See ?points

# Change symbology to highlight features in data
cex[is_boom_year || is_bust_year] <- 1.3
bg[is_boom_year] <- "seagreen"
bg[is_bust_year] <- "orange"
col[is_boom_year] <- "darkgreen"
col[is_bust_year] <- "red"
pch[is_boom_year] <- 24
pch[is_bust_year] <- 25

# Make plot first without points
```

```

par(fin = c(4,4),
    mai = c(1,1,1,1),
    tcl = -0.3,      # length of tickmarks
    mgp = c(1.2, 0.3, 0),
    cex.axis = 0.8) # distance of axis title, axis label, and axis line to box
plot(x, y,
     type = "n",
     xlab = "Years since Mastodon reintroduction",
     ylab = "Population size",
     xaxt = "n",
     yaxt = "n",
     xlim = c(0, 100),
     ylim = c(0, 120)
    )
axis(1, c(0, 50, 100))
axis(2, c(0, 60, 120))

# Draw a polygon for the confidence envelope as bottom layer
xpoly <- c(xpred, rev(xpred))
ypoly <- c(ypred$fit + 5 * ypred$se.fit,
           rev(ypred$fit - 5 * ypred$se.fit))

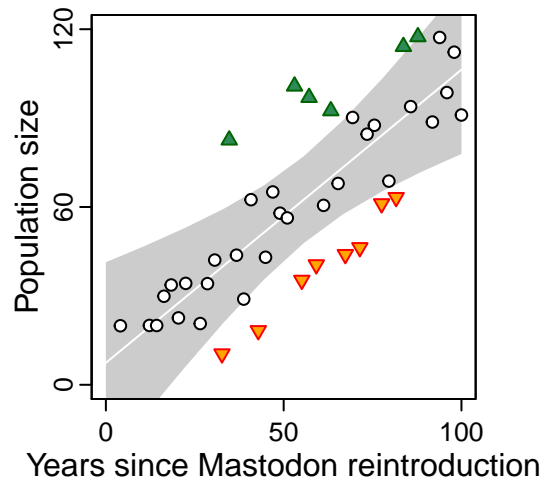
polygon(x = xpoly, y = ypoly,
        border = NA,
        col = grey(0.8))

# Add trendline
lines(xpred, ypred$fit, col = 'white')

# Add in the points, with their symbologies pre-specified above (no calculations here)
points(x,y,
       pch = pch,
       col = col,
       bg = bg,
       cex = cex)

box(lwd = 1) # place a box over the plot

```



Remark

A corollary of this approach is that figures should have a small number of clear, concrete messages. For any figure you show your audience, you should be able to explicitly populate a (short) list like the one above (“This figure says:”). Having done that, you can evaluate how efficiently the figure delivers its message. A figure does not have to show every aspect of the data, or even most aspects. Every display element should be critical to the core message. In other words, if you can leave it out with compromising the core message, you should do so. If you are worried you are misleading readers by leaving something out, include a note in the figure legend about what was omitted, and consider including a more complete (less efficient) version of the figure in the supplemental materials.