

BAMSurgeon: Methods for spike-in mutations on BAM files

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1 Introduction

1.1 Software Dependencies

BAMSurgeon requires the following software packages be available:

1. samtools/wgsim/tabix (<http://samtools.sourceforge.net/>)
2. pysam (<http://code.google.com/p/pysam/> or `pip install pysam`)
3. bwa (<http://bio-bwa.sourceforge.net/>)
4. velvet (<http://www.ebi.ac.uk/~zerbino/velvet/>)
5. exonerate (<https://github.com/nathanweeks/exonerate>)
6. picard (<http://http://picard.sourceforge.net/>)

For install instructions please see the included README.md or have a look at the Dockerfile.

1.2 Known bugs and limitations

BAMSurgeon is under rapid development driven by suggesting and bug reports from the mutation calling community. The following is an incomplete TODO list of things needing attention:

1. Adding SVs above 30kb is very slow
2. Simulated SVs cannot be specified exactly
3. Heterozygous mutations in SV regions tend to increase or decrease in allele fraction due to one or the other haplotype being assembled and used as the basis for read coverage simulation. This can lead to false-positive mutations in SV regions, so we suggest masking these regions or simulating SVs and SNVs/INDELs on separate BAMs.

2 Adding SNVs to existing BAM alignments (addsnv)

2.1 Usage

```
usage: addsnv.py [-h] -v VARFILENAME -f BAMFILENAME -r REFFASTA -o OUTBAMFILE
                  [-s SNVFRAC] [-m MUTFRAC] [-n NUMSNVS] [-c CNVFILE]
                  [-d COVERDIFF] [-z HAPLOSIZE] [-p PROCS]
```

```

[--picardjar PICARDJAR] [--mindepth MINDEPTH]
[--maxdepth MAXDEPTH] [--minmutreads MINMUTREADS]
[--avoidreads AVOIDREADS] [--nomut] [--ignoresnps]
[--ignoreref] [--force] [--single] [--maxopen MAXOPEN]
[--requirepaired] [--tagreads] [--skipmerge] [--ignorepileup]
[--aligner ALIGNER] [--alignopts ALIGNOPTS] [--tmpdir TMPDIR]
[--seed SEED]

```

adds SNVs to reads, outputs modified reads as .bam along with mates

optional arguments:

```

-h, --help          show this help message and exit
-v VARFILENAME, --varfile VARFILENAME
                    Target regions to try and add a SNV, as BED
-f BAMFILENAME, --bamfile BAMFILENAME
                    sam/bam file from which to obtain reads
-r REFFASTA, --reference REFFASTA
                    reference genome, fasta indexed with bwa index -a
                    stdsw_and_ samtools faidx
-o OUTBAMFILE, --outbam OUTBAMFILE
                    .bam file name for output
-s SNVFRAC, --snvfrac SNVFRAC
                    maximum allowable linked SNP MAF (for avoiding
                    haplotypes) (default = 1)
-m MUTFRAC, --mutfrac MUTFRAC
                    allelic fraction at which to make SNVs (default = 0.5)
-n NUMSNVS, --numsnvs NUMSNVS
                    maximum number of mutations to try (default: entire
                    input)
-c CNVFILE, --cnvfile CNVFILE
                    tabix-indexed list of genome-wide absolute copy number
                    values (e.g. 2 alleles = no change)
-d COVERDIFF, --coverdiff COVERDIFF
                    allow difference in input and output coverage
                    (default=0.9)
-z HAPLOSIZE, --haplosize HAPLOSIZE
                    haplotype size (default = 0)
-p PROCS, --procs PROCS
                    split into multiple processes (default=1)
--picardjar PICARDJAR
                    path to picard.jar, required for most aligners
--mindepth MINDEPTH
                    minimum read depth to make mutation (default = 10)
--maxdepth MAXDEPTH
                    maximum read depth to make mutation (default = 2000)
--minmutreads MINMUTREADS
                    minimum number of mutated reads to output per site
--avoidreads AVOIDREADS
                    file of read names to avoid (mutations will be skipped
                    if overlap)

```

```

--ignoresnps      make mutations even if there are non-reference alleles
                  sharing the relevant reads
--force          force mutation to happen regardless of nearby SNP or
                  low coverage
--single         input BAM is single-ended (default is paired-end)
--maxopen MAXOPEN maximum number of open files during merge (default
                  1000)
--requirepaired  skip mutations if unpaired reads are present
--tagreads       add BS tag to altered reads
--skipmerge      final output is tmp file to be merged
--ignorepileup   do not check pileup depth in mutation regions
--aligner ALIGNER supported aligners:
                  backtrack,mem,novoalign,gsnap,bowtie2
--alignopts ALIGNOPTS
                  aligner-specific options as comma delimited list of
                  option1:value1,option2:value2,...
--tmpdir TMPDIR  temporary directory (default=addsnv.tmp)
--seed SEED      seed random number generation

```

2.2 Description

Single nucleotide changes are introduced to an existing BAM alignment using `addsnv.py` as outlined in Figure 1. Input consists of a list of locations where SNVs will be made, a target BAM alignment, and a reference genome indexed using `bwa` (preferably the same genome used to align the reads in the BAM file).

2.3 Input

Mutation input (-v/-varfile) Input to `addsnv.py` (-v/-varfile) is a three column format plus an extra optional column for the desired variant allele fraction (VAF). Input coordinates are 1-based, meaning they will match between input to -v/-varfile, VCF, and `samtools mpileup`. The default VAF is specified via -m/-mutfrac. For example:

```

22 34166720 34166720 0.25
22 33908770 33908770 0.15
22 33714964 33714964 0.5
22 33769483 33769483 0.75
22 33958087 33958087 0.01
22 34264774 34264774 0.25
22 33702084 33702084 0.2

```

Specific bases can also be specified using a fifth input column:

```

22 34166720 34166720 0.25 A
22 33908770 33908770 0.15 T
22 33714964 33714964 0.5 G
22 33769483 33769483 0.75 C

```

The above input will cause addsnv to attempt to add SNVs at the specified locations, using the VAF specified in the fourth column if available, otherwise defaulting to 0.5 or whatever is specified by `-m/-mutfrac` (e.g. the last three lines). If a position is specified that spans more than one base, the position of the SNV input will be chosen at random from that interval.

BAM Alignment input (`-f/-bamfile`) BAMs ideally should adhere to SAM specification i.e. they should validate via picard's `ValidateSamFile`. BAMs should be sorted in coordinate order and indexed. BAMs should consist either of entirely paired-end reads or entirely single-end (fragment) reads using the `-single` option, mixing paired and single reads yields unpredictable results. Currently, bwa backtrack (`bwa aln`) and bwa mem are supported, although it should be straightforward enough to add further aligners through modification of the `remap_paired` function. In practice we have been able to spike mutations into RNA-seq MapSplice BAMs, for instance.

Reference genome (`-r/-reference`) The reference genome should be the **same as that used to create the target BAM file**, specifically the chromosome names and lengths in the reference FASTA must be the same as in the BAM header. The reference must be indexed for bwa (`bwa index`) and indexed with samtools (`samtools faidx`).

Avoiding existing SNPs with `-s/-snpfrac` (optional, default=1.0) In an attempt to preserve haplotype structure and avoid confusion by spiking mutations on top of existing heterozygous alleles, the `-s/-snpfrac` cutoff may be useful. For a selected mutation site S and specified cutoff f starting from the leftmost base L of the leftmost read containing S and ending at the rightmost base R of the rightmost read containing S , the minor allele frequency M for every intervening base is calculated for every base in the interval $[L, R]$. If $M < f$ for all bases in $[L, R]$ the mutation at S is allowed, and skipped otherwise. Note, do not confuse this with `-m/-mutfrac`, which is the target variant allele frequency for mutations unless overridden in the mutation input file. By default this function is turned off (threshold minor allele frequency set to 1.0). Setting this too low can yield few successful mutations as sequence errors will inhibit successful spike-in, we recommend a setting of 0.1 for Illumina GA/HiSeq reads.

CNV file (`-c/-cnvfile`) (optional) It is possible to simulate adding mutations to a single haplotype for copy-number variant regions. For example, if a region has absolute copy number 3 (instead of 2), a mutation into one haplotype should have an apparent VAF of roughly 0.33 instead of 0.5. This optional file should be in BED-3 format plus a fourth column representing the absolute copy number for the region defined by the chromosome/contig, start, and end fields. The genome is assumed to be diploid thus an absolute copy number of 2 will not adjust the variant allele fractions. Absolute copy numbers below 2 will adjust VAFs higher and absolute copy numbers above 2 will adjust VAFs correspondingly lower.

Additional options

- `-d/-coverdiff` : threshold on output coverage divided by input coverage for mutation sites. Mutations can cause alignments to change, if this is undesired set this high, if this is part of the fun, set this low.
- `-z/-haplosize` : specifies size of haplotype block to consider when adding more than 1 proximal mutation. For instance, if two SNVs are spiked in 10bp apart and `-haplosize` is 10 or greater,

the two SNVs will be on the same haplotype (i.e. share the same reads for reads covering both positions)

- `-p/-procs` : BAMSurgeon programs currently spawn a new process for each spikein, to speed things along the processes can be run concurrently with the number of simultaneous jobs set by this parameter.
- `-picardjar` : path to `picard.jar` provided by picard tools, this is required for the some aligner options.
- `-mindepth` : minimum depth at sites for mutations, sites with fewer reads in pileup column will be dropped. Default is 10 reads.
- `-maxdepth` : sites with greater than this number of covering reads will be dropped. Default is 2000 reads. (May need to increase for targeted sequencing data)
- `-minmutreads` : minimum number of mutated reads per site. If this level of coverage cannot be obtained, mutations will be dropped.
- `-avoidreads` : `addsnv` and `addindel` can be passed a list of read names to avoid when making mutations. Any sites selected that overlap these reads will be dropped. This feature exists as a means to ensure mutations across multiple runs do not overlap.
- `-force` : force the addition of a mutation even if it fails thresholds set by `-mindepth` and `-c/-coverdiff`. Not recommended for most use cases.
- `-single` : covered above, assume reads are single-end (i.e. fragment reads).
- `-maxopen` : set the maximum number of open files (check your system limit with `ulimit -a`)
- `-requirepaired` : drop mutations where a paired end in the region cannot be located. However, it is probably better to correct the problem in the BAM file than to resort to using this option.
- `-skipmerge` : instead of replacing the mutated and realigned reads back into the original BAM, which is normally the final step, leave the `addsnv`.
- `-aligner` : choose one of the supported aligners, default is `backtrack` (for `bwa backtrack`). Additional aligners can be added through modification of `bs/aligners.py`, and I take requests. Aligner specific options are handled through `-alignopts`
- `-alignopts` : aligner-specific options are specified in a comma-delimited list as `option1:value1,option2:value2,et`. Required options are checked by the the function `checkoptions` in `bs/aligners.py`. For example, if `-aligner novoalign` is specified, `-alignopts novoref:path to novoalign reference;` should also be specified.
- `-tagreads` : adds a 'BS' tag to the reads altered by `bamsurgeon`, useful in debugging
- `-seed` : sets a random seed for reproducibility
- options not listed here are either new and the manual has not been updated as yet, or they are there for debugging and will be removed in the future.

3 Adding INDELs to existing BAM alignments (addindel)

3.1 Usage

```
usage: addindel.py [-h] -v VARFILENAME -f BAMFILENAME -r REFFASTA -o
                  OUTBAMFILE [-s SNVFRAC] [-m MUTFRAC] [-n NUMSNVS]
                  [-c CNVFILE] [-d COVERDIFF] [-p PROCS]
                  [--picardjar PICARDJAR] [--mindepth MINDEPTH]
                  [--maxdepth MAXDEPTH] [--minmutreads MINMUTREADS]
                  [--avoidreads AVOIDREADS] [--nomut] [--det] [--force]
                  [--single] [--maxopen MAXOPEN] [--requirerepaired]
                  [--aligner ALIGNER] [--alignopts ALIGNOPTS] [--tagreads]
                  [--skipmerge] [--ignorepileup] [--tmpdir TMPDIR]
                  [--seed SEED]
```

adds INDELs to reads, outputs modified reads as .bam along with mates

optional arguments:

```
-h, --help            show this help message and exit
-v VARFILENAME, --varfile VARFILENAME
                        Target regions to try and add a SNV, as BED
-f BAMFILENAME, --bamfile BAMFILENAME
                        sam/bam file from which to obtain reads
-r REFFASTA, --reference REFFASTA
                        reference genome, fasta indexed with bwa index -a
                        stdsw_and_ samtools faidx
-o OUTBAMFILE, --outbam OUTBAMFILE
                        .bam file name for output
-s SNVFRAC, --snvfrac SNVFRAC
                        maximum allowable linked SNP MAF (for avoiding
                        haplotypes) (default = 1)
-m MUTFRAC, --mutfrac MUTFRAC
                        allelic fraction at which to make SNVs (default = 0.5)
-n NUMSNVS, --numsnvs NUMSNVS
                        maximum number of mutations to try (default: entire
                        input)
-c CNVFILE, --cnvfile CNVFILE
                        tabix-indexed list of genome-wide absolute copy number
                        values (e.g. 2 alleles = no change)
-d COVERDIFF, --coverdiff COVERDIFF
                        allow difference in input and output coverage
                        (default=0.1)
-p PROCS, --procs PROCS
                        split into multiple processes (default=1)
--picardjar PICARDJAR
                        path to picard.jar
--mindepth MINDEPTH    minimum read depth to make mutation (default = 10)
--maxdepth MAXDEPTH    maximum read depth to make mutation (default = 2000)
```

```

--minmutreads MINMUTREADS          minimum number of mutated reads to output per site
--avoidreads AVOIDREADS            file of read names to avoid (mutations will be skipped
                                   if overlap)
--det                               deterministic base changes: make transitions only
--force                             force mutation to happen regardless of nearby SNP or
                                   low coverage
--single                           input BAM is single-ended (default is paired-end)
--maxopen MAXOPEN                  maximum number of open files during merge (default
                                   1000)
--requirepaired                    skip mutations if unpaired reads are present
--tagreads                          add BS tag to altered reads
--aligner ALIGNER                  supported aligners:
                                   backtrack,mem,novoalign,gsnap,bowtie2
--alignopts ALIGNOPTS              aligner-specific options as comma delimited list of
                                   option1:value1,option2:value2,...
--skipmerge                         final output is tmp file to be merged
--ignorepileup                     do not check pileup depth in mutation regions
--tmpdir TMPDIR                    temporary directory (default=addindel.tmp)
--seed SEED                         seed random number generation

```

3.2 Description

Short insertions and deletions (commonly referred to as INDELs) can be added to BAM files using `addindel.py`. The options are essentially the same as for `addsnv.py`, but the syntax of the input file is slightly different:

```

22 33694303 33694304 0.50 INS ATGGC
22 33815273 33815274 0.25 INS CG
22 33920490 33920491 0.33 INS ATTTGCTTAGCTGAGGGCTTAGGCTTAGCATGCGTA
22 33944542 33944543 0.50 DEL
22 34006643 34006653 0.50 DEL
22 33958087 33958090 0.40 DEL

```

The first four columns are the same as for `addsnv.py` input but the fourth column (VAF) is mandatory. The fifth column must be either 'INS' or 'DEL' (for insertion or deletion). For insertions there must be a sixth column specifying the sequence to be inserted at the position indicated in the second column (the position in the third column does not matter for insertions). In the above example, the first three columns direct `addindel.py` to (1) add an insertion of "ATGGC" at chromosome 22 position 33694303 with 50% VAF, (2) add an insertion of "CG" at position 338152273 with 25% VAF, and (3) add an insertion of "ATTTGC..." with 33% VAF.

For deletions, the sequence from the start to the end (columns two and three) is deleted. Lines 4,5, and 6 above indicate deletions of 1 bp, 10bp, and 3 bp, respectively.

4 Adding SVs to existing BAM alignments (addsv.py)

4.1 Usage

```
usage: addsv.py [-h] -v VARFILENAME -f BAMFILENAME -r REFFASTA -o OUTBAMFILE
               [-l MAXLIBSIZE] [-k KMERSIZE] [-s SVFRAC]
               [--minctglen MINCTGLEN] [-n MAXMUTS] [-c CNVFILE]
               [--ismean ISMEAN] [--issd ISSD] [-p PROCS] [--inslib INSLIB]
               [--delay DELAY] [--noref] [--aligner ALIGNER]
               [--alignopts ALIGNOPTS] [--tagreads] [--skipmerge]
               [--keepsecondary] [--debug] [--tmpdir TMPDIR] [--seed SEED]
```

adds SVs to reads, outputs modified reads as .bam along with mates

optional arguments:

```
-h, --help                show this help message and exit
-v VARFILENAME, --varfile VARFILENAME
                           whitespace-delimited target regions to try and add a
                           SV: chrom,start,stop,action,seqfile (if
                           insertion),TSDlength (if insertion)
-f BAMFILENAME, --bamfile BAMFILENAME
                           sam/bam file from which to obtain reads
-r REFFASTA, --reference REFFASTA
                           reference genome, fasta indexed with bwa index -a
                           stdsw_and_ samtools faidx
-o OUTBAMFILE, --outbam OUTBAMFILE
                           .bam file name for output
-l MAXLIBSIZE, --maxlibsize MAXLIBSIZE
                           maximum fragment length of seq. library
-k KMERSIZE, --kmer KMERSIZE
                           kmer size for assembly (default = 31)
-s SVFRAC, --svfrac SVFRAC
                           allele fraction of variant (default = 1.0)
--maxctglen MAXCTGLEN
                           maximum contig length for assembly - can increase if
                           velvet is compiled with LONGSEQUENCES
-n MAXMUTS                maximum number of mutations to make
-c CNVFILE, --cnvfile CNVFILE
                           tabix-indexed list of genome-wide absolute copy number
                           values (e.g. 2 alleles = no change)
--ismean ISMEAN           mean insert size (default = estimate from region)
--issd ISSD               insert size standard deviation (default = estimate
                           from region)
-p PROCS, --procs PROCS
                           split into multiple processes (default=1)
--inslib INSLIB           FASTA file containing library of possible insertions,
                           use INS RND instead of INS filename to pick one
--delay DELAY             time delay between jobs (try to avoid thrashing disks)
```



```

--aligner ALIGNER      supported aligners: backtrack,mem,novoalign
--alignopts ALIGNOPTS  aligner-specific options as comma delimited list of
                        option1:value1,option2:value2,...
--skipmerge            do not merge spike-in reads back into original BAM
--keepsecondary        keep secondary reads in final BAM
--debug               output read tracking info to debug file, retain all
                        intermediates
--tmpdir TMPDIR        temporary directory (default=addsv.tmp)
--seed SEED            seed random number generation

```

4.2 Description

Larger structural variants (insertions, deletions, duplications, inversions, and compound rearrangements) are added to existing BAM alignments using `addsv.py` as described in Figure 3. Input consists of a list of regions where SVs will be made along with a specification of each variant, and as with `addsnv` and `addindel`, a target BAM alignment, and a reference genome indexed using `bwa` are required.

4.3 Input

The input mutation list consists of four columns: chromosome, start of region, end of region, and a controlled-vocabulary description of the mutation. A mutation will not be made if the largest contig obtained from local assembly of the specified region is less than 3 times the maximum expected library size (specified by `-l/--maxlibsize`). The mutation description starts with either `INS`, `DEL`, `DUP`, `TRN`, or `INV` for insertion, deletion, duplication, transduction, and inversion, respectively and is followed by mutation-specific options. An example follows.

```

22 33871043 33884754 DEL 0.9
22 33971043 33984754 INS myseq.fa
22 34071043 34084754 INV
22 34171043 34184754 INS ACTATCGTGGGCTTATGGCTTAGCATTACGGTTATTTTGCCTGATCGCGGGGGCTACATCATTTCATG
22 34271043 34284754 DUP 3
22 34371043 34384754 INS RND

```

Transductions are also possible using the following syntax:

```

22 33607508 33607508 TRN 22 34314981 34314981

```

This functionality is currently under development. This example will join the 6kbp region centered on 22:33607508 and the 6kbp region centered on 22:34314981. Future enhancements to transductions will include the ability to specify the orientation of both breakends in the junction and more granular control of the junction location.

For insertions, `INS` should be followed by either a FASTA file containing the sequence to be inserted, by the nucleotide sequence itself, or by `RND` if a library of potential insertion sequences is passed in FASTA format via the `-inslib` option. For example, `INS ATG` would insert the sequence ‘‘ATG’’ in the middle of the largest contig obtained from the specified region, while `INS LINE1.fa` would insert the sequence in the FASTA-formatted file `LINE1.fa` into the largest contig. For

deletions, DEL should be followed by the fraction of the largest contig to be deleted, where 1.0 indicates a deletion of the largest assembled region minus one library length on each end. e.g. DEL 0.5. Inversions (INV) have no further options - the region inside the largest contig will be inverted. Duplications (DUP) have one optional parameter, an integer specifying the number of times the sequence of the largest contig should be duplicated, e.g. DUP 2 specifies the region is duplicated twice. In general, we recommend that short insertions and deletions (those shorter than one read length) be created with addindel.py instead of addsv.py as addindel.py does not require reconstruction of the region through sequence assembly and is thus less error-prone.

Compound variants are also possible by chaining a number of mutations together in a comma-delimited list, e.g. DUP 1, DEL 0.5, INS AAATCC, INV would duplicate the region inside the largest contig once, delete half the width of the region, insert the sequence AAATCC, and invert the region (not currently supported with transductions). Good luck!

4.4 Output

Where mutations add sequence (e.g. duplications), new reads are created that will be added to the original BAM, and where mutations remove sequence (e.g. deletions) those reads are marked as excluded (excluded read names are recorded in the file specified by `-x/--excluded`). Excluded reads are not carried over from the original BAM to the mutated BAM, creating the copy number effect associated with deletion.

Output consists of a BAM file containing the spike-in mutations and a directory of log files describing which bases were changed in which reads (one log per mutation) and the local assemblies for each region where a mutation was made. Currently, the directory containing output logs for SV spike-ins can be transformed into VCF format using the script `makevcf_sv.py` in the `etc/` subdirectory. The spike-in process may be done concurrently through the use of the `-p/--procs` option. Here are some extended descriptions for additional options not mentioned yet:

- `-s/--snvfrac` : equivalent to `-m/--mutfrac` for `addindel.py` and `addsnv.py` (will fix)
- `-k/--kmer` : kmer size to be fed to velvet, which is used for assembly
- `-n` : only attempt this many mutations
- `-ismean/--issd` : the mean and standard deviation of the library size should be estimated from the original bam and specified through these arguments. This can be accomplished a number of ways, I suggest `CollectInsertSizeMetrics` from the `picard` tools.
- `-inslib` : a FASTA file can be specified and insertions can be tagged with RND instead of a sequence or file. If this is done, a sequence will be chosen at random from the FASTA file specified by `-inslib` and inserted.
- `-skipmerge` : same meaning as for `addsnv/addindel`

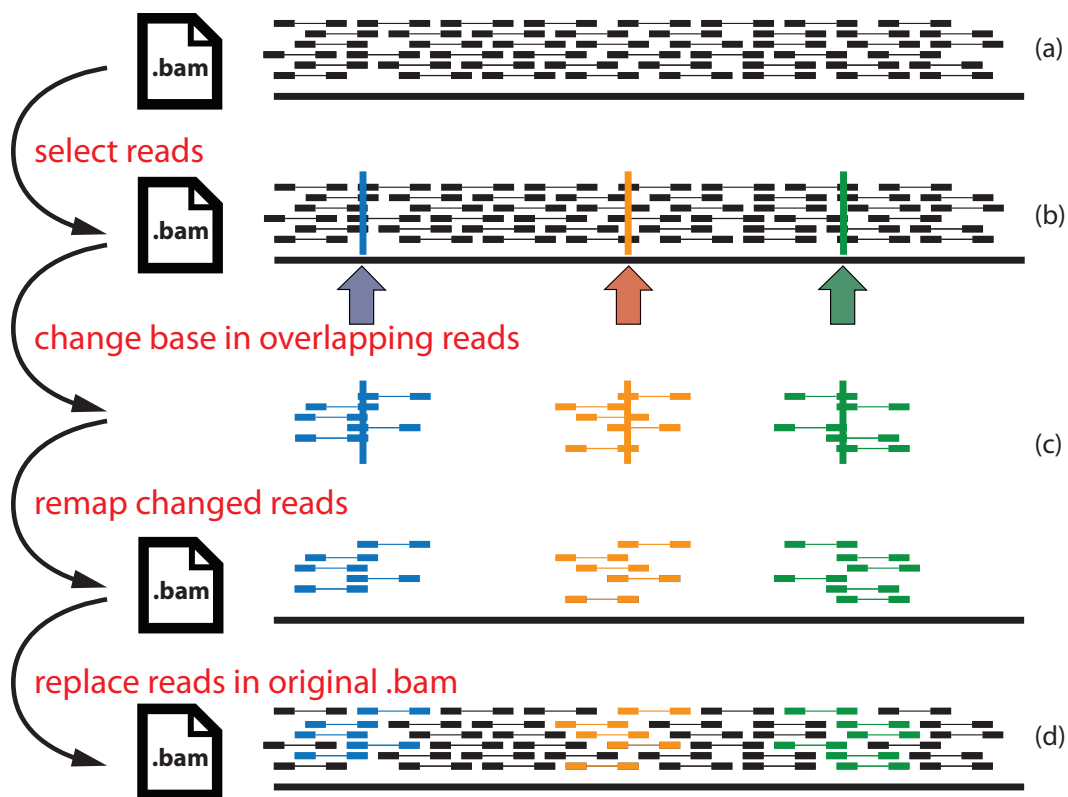


Figure 1: Method for adding SNVs to existing BAM alignments. Starting with the original BAM alignment (a) and a list of positions (b), reads overlapping the chosen positions are selected the target base change is made (c). Altered reads are re-mapped the the reference genome using bwa, and the modified, realigned reads replace the unmodified versions in the original BAM (d).

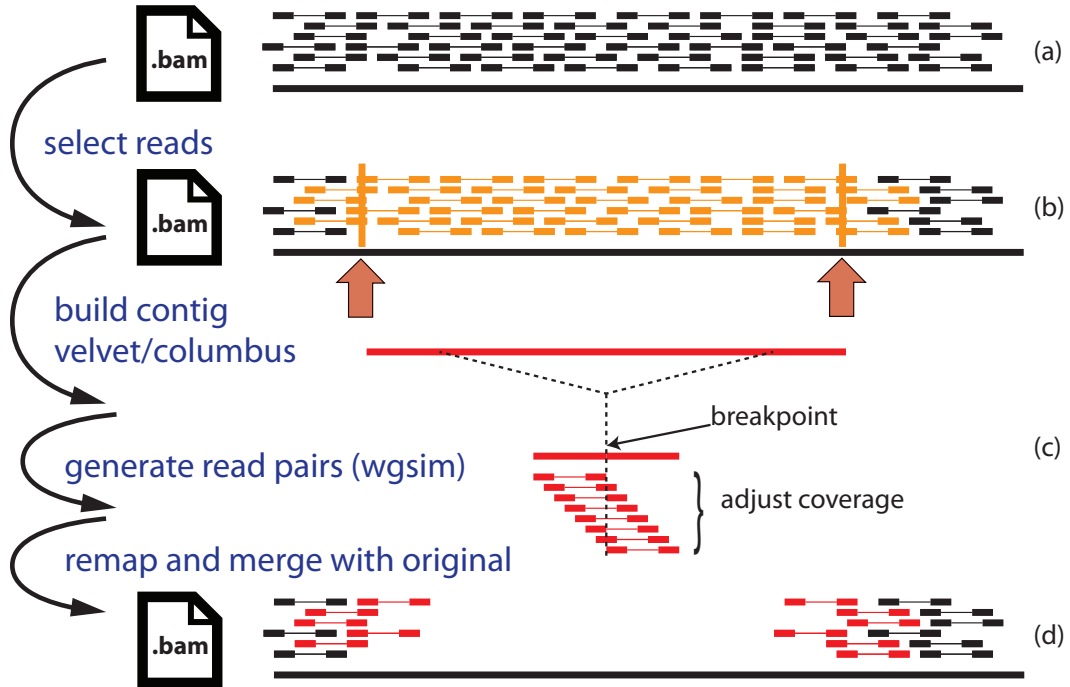


Figure 2: Method for adding multi-nucleotide variants (indels and SVs) to existing BAMs: deletion example. Starting with the original BAM alignment (a) regions, e.g. (b), are assembled using Velvet/Columbus (<http://www.ebi.ac.uk/zerbino/velvet/>) and the desired mutation(s) are created in the largest contig. Read coverage is simulated over the contig using wgsim (<https://github.com/lh3/wgsim>, also included with samtools) with parameters `-e 0 -r 0 -R 0` to suppress additional mutations, other parameters are set based on the input BAM and desired coverage. Simulated coverage is scaled based on whether the mutation is adds sequence (duplication) or removes sequence (deletions), and replaced into the original BAM while excluded reads are removed from the original BAM (d).

5 Testing

The `test/` directory contains scripts useful for testing bamsurgeon's various modes, the primary ones are as follows:

5.1 Testing addsnv.py

The script `test/test_snv.sh` may be run with no parameters to display a usage statement:

```
usage: ./test_snv.sh <number of SNPs> <number of threads> <reference indexed with bwa index>
```

Or for testing with BWA MEM:

```
./test_snv_bwamem.sh <number of SNPs> <number of threads> <reference indexed with bwa index> <genome>
```

The number of SNPs can be up to 100 and are selected from the file `test_data/random_snvs.txt`, number of threads must be specified, use 1 to run on a single core, and the final argument must be a path to a human genome (GRCh37) reference indexed with `bwa index -a bwtsv`. The reference **should not** use the 'chr' prefix on the chromosome names.

5.2 Testing addindel.py

The script `test/test_sv.sh` may be run with no parameters to display a usage statement:

```
usage: ./test_indel.sh <number of threads> <reference indexed with bwa index>
```

5.3 Testing addsv.py

The script `test/test_sv.sh` may be run with no parameters to display a usage statement:

```
usage: ./test_sv.sh <number of threads> <reference indexed with bwa index>
```

6 Other Considerations

6.1 System Requirements

For each process (specified by `-p/--procs`) `add*.py` will require 4GB RAM. So for 8 concurrent mutation processes, you should have 32GB RAM. System requirements may decrease in the future as we find ways to improve efficiency. For more recent versions of `bwa`, it is possible to load the reference index into shared memory with `'bwa shm'` which seems to improve performance.

6.2 Alignments

Reads should be re-aligned using the same aligner and parameters as were used to create the original BAM file. By default, `bamsurgeon` assumes alignments were performed using `bwa aln` with parameters `-q 5 -l 32 -k 3 -o 1`. The specific parameters are not currently specifiable on the command line but may be altered through modification of the functions `remap_paired` (for paired reads) or `remap_single`.

7 Acknowledgements

We would like to thank everyone who has submitted a pull request or bug report through github, the synapse forums for the DREAM Somatic Mutation Calling challenge, or by other means.