Package 'dStruct'

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Description More about what it does (maybe more than one line) Use four spaces when indenting paragraphs within the Description.					
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2 dCombs

calcDis

Calculates d score.

Description

Calculates d score.

Usage

calcDis(x)

Arguments

Х

A numeric vector or matrix.

Value

If input is a numeric vector, a number is returned. For a matrix, a numeric vector is returned.

 dCombs

Assesses within-group or between-group variation.

Description

Assesses within-group or between-group variation.

Usage

```
dCombs(rdf, combs)
```

Arguments

rdf Data.frame of reactivities for each sample.

combs Data.frame with each column containing groupings of samples.

Value

Nucleotide-wise d score.

dStruct 3

dStruct	Performs de novo discovery of differentially reactive regions.	

Description

Performs de novo discovery of differentially reactive regions.

Usage

```
dStruct(rdf, reps_A, reps_B, batches = F, min_length = 11,
  check_signal_strength = T, check_nucs = T, check_quality = T,
  quality = "auto", evidence = 0, signal_strength = 0.1,
  within_combs = NULL, between_combs = NULL, ind_regions = T, gap = 1,
  get_FDR = T, proximity_assisted = F, proximity = 10,
  proximity_defined_length = 30)
```

Arguments

E	guments	
	rdf	Dataframe of reactivities for each sample.
	reps_A	Number of replicates of group A.
	reps_B	Number of replicates of group B.
	batches	Logical suggesting if replicates of group A and B were performed in batches and are labelled accordingly. If TRUE, a heterogeneous/homogeneous subset may not have multiple samples from the same batch.
	min_length	Minimum length of constructed regions.
check_signal_strength		
		Logical, if TRUE, construction of regions must be based on nucleotides that have a minimum absolute value of reactivity.
	check_nucs	Logical, if TRUE, constructed regions must have a minimum number of nucleotides participating in Wilcoxon signed rank test.
	check_quality	Logical, if TRUE, check constructed regions for quality.
	quality	Worst allowed quality for a region to be tested.
	evidence	Minimum evidence of increase in variation from within-group comparisons to between-group comparisons for a region to be tested.
	signal_strength	1
		Threshold for minimum signal strength.
	within_combs	Data.frame with each column containing groupings of replicates of groups A or B, which will be used to assess within-group variation.
	between_combs	Dataframe with each column containing groupings of replicates of groups A and

B, which will be used to assess between-group variation.

ind_regions Logical, if TRUE, test each region found in the transcript separately.

gap Integer. Join regions if they are separated by these many nucleotides.

get_FDR Logical, if FALSE, FDR is not reported.

 $proximity_assisted$

Logical, if TRUE, proximally located regions are tested together.

4 dStruct.guided

proximity Maximum distance between constructed regions for them to be considered proximal.

proximity_defined_length

If performing a "proximity-assisted" test, minimum end-to-end length of a region to be tested.

Value

Constructs regions, reports p-values and FDR for them.

dStruct.guided Performs guided discovery of differentially reactive regions.

Description

Performs guided discovery of differentially reactive regions.

Usage

```
dStruct.guided(rdf, reps_A, reps_B, batches = F, within_combs = NULL,
  between_combs = NULL, check_quality = TRUE, quality = "auto",
  evidence = 0)
```

Arguments

rdf	Dataframe of reactivities for each sample. Each column must be labelled as A1, A2,, B1, B2,
reps_A	Number of replicates of group A.
reps_B	Number of replicates of group B.
batches	Logical suggesting if replicates of group A and B were performed in batches and are labelled accordingly. If TRUE, a heterogeneous/homogeneous subset may not have multiple samples from the same batch.
within_combs	Data.frame with each column containing groupings of replicates of groups A or B, which will be used to assess within-group variation.
between_combs	Dataframe with each column containing groupings of replicates of groups A and B, which will be used to assess between-group variation.
check_quality	Logical, if TRUE, check regions for quality.
quality	Worst allowed quality for a region to be tested.
evidence	Minimum evidence of increase in variation from within-group comparisons to

Value

p-value for the tested region, estimated using one-sided Wilcoxon signed rank test.

between-group comparisons for a region to be tested.

5 dStructome

dStructome	Performs de novo discovery of differentially reactive regions for transcriptome-wide data.
	transcriptome-wae aata.

Description

Performs de novo discovery of differentially reactive regions for transcriptome-wide data.

Usage

```
dStructome(rl, reps_A, reps_B, batches = F, min_length = 11,
 check_signal_strength = T, check_nucs = T, check_quality = T,
 quality = "auto", evidence = 0, signal_strength = 0.1,
 within_combs = NULL, between_combs = NULL, ind_regions = T, gap = 1,
 processes = "auto", method = "denovo", proximity_assisted = F,
 proximity = 10, proximity_defined_length = 30)
```

Arg

 $ind_regions$

processes ${\tt method}$

gap

guments			
guments			
rl	List of dataframes of reactivities for each sample.		
reps_A	Number of replicates of group A.		
reps_B	Number of replicates of group B.		
batches	Logical suggesting if replicates of group A and B were performed in batches and are labelled accordingly. If TRUE, a heterogeneous/homogeneous subset may not have multiple samples from the same batch.		
min_length	Minimum length of constructed regions.		
check_signal_strength			
	Logical, if TRUE, construction of regions must be based on nucleotides that have a minimum absolute value of reactivity.		
check_nucs	Logical, if TRUE, constructed regions must have a minimum number of nucleotides participating in Wilcoxon signed rank test.		
check_quality	Logical, if TRUE, check constructed regions for quality.		
quality	Worst allowed quality for a region to be tested.		
evidence	Minimum evidence of increase in variation from within-group comparisons to between-group comparisons for a region to be tested.		
signal_strengt	h		
	Threshold for minimum signal strength.		
within_combs	Data.frame with each column containing groupings of replicates of groups A or B, which will be used to assess within-group variation.		
between_combs	Dataframe with each column containing groupings of replicates of groups A and B, which will be used to assess between-group variation.		

Logical, if TRUE, test each region found in the transcript separately.

Integer. Join regions if they are separated by these many nucleotides.

Character specifying either guided or de novo discovery approach.

Number of parallel processes to use.

6 getCombs

proximity_assisted

Logical, if TRUE, proximally located regions are tested together.

proximity Maximum distance between constructed regions for them to be considered prox-

imal.

 $\verb"proximity_defined_length"$

If performing a "proximity-assisted" test, minimum end-to-end length of a re-

gion to be tested.

Value

Constructs regions, reports p-values and FDR for them.

getCombs	Identifies subgroupings of replicates for assessing within-group and
	between-group variation.

Description

Identifies subgroupings of replicates for assessing within-group and between-group variation.

Usage

```
getCombs(reps_A, reps_B, batches = F, between_combs = NULL,
   within_combs = NULL)
```

Arguments

reps_A	Number of replicates of group A.
reps_B	Number of replicates of group B.
batches	Logical suggesting if replicates of group A and B were performed in batches and are labelled accordingly. If TRUE, a heterogeneous/homogeneous subset may not have multiple samples from the same batch.
between_combs	Dataframe with each column containing groupings of replicates of groups A and B, which will be used to assess between-group variation.
within_combs	Data.frame with each column containing groupings of replicates of groups A or B, which will be used to assess within-group variation.

Value

List of two dataframes, containing groupings for within-group and between-group variation.

getContigRegions 7

getContigRegions	Identifies contiguous regions from a list of nucleotide indices.

Description

Identifies contiguous regions from a list of nucleotide indices.

Usage

```
getContigRegions(x, gap = 0)
```

Arguments

x A vector of integers.

gap Allowed gap to merge regions.

Value

Dataframe storing start and stop sites of continguous regions.

getRegions Constructs potential differentially reactive regions.

Description

Constructs potential differentially reactive regions.

Usage

```
getRegions(d_within, d_spec, rdf, min_length = 11,
  check_signal_strength = T, check_nucs = T, check_quality = T,
  quality = 0.5, evidence = 0, signal_strength = 0.1)
```

Arguments

d_within Nucleotide-wise d score for within-group variation.d_spec Nucleotide-wise d score for between-group variation.

rdf Dataframe of reactivities for each sample.
min_length Minimum length of constructed regions.

check_signal_strength

Logical, if TRUE, construction of regions must be based on nucleotides that

have a minimum absolute value of reactivity.

check_nucs Logical, if TRUE, constructed regions must have a minimum number of nu-

cleotides participating in Wilcoxon signed rank test.

check_quality Logical, if TRUE, check constructed regions for quality.

quality Worst allowed quality for a region to be tested.

evidence Minimum evidence of increase in variation from within-group comparisons to

between-group comparisons for a region to be tested.

signal_strength

Threshold for minimum signal strength.

8 plot_dStructurome

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Returns normalizer for reactivity vector.

Description

Assesses normalization factor for raw reactivities using 2-8 % method.

Usage

```
normalizer(raw.estimates)
```

Arguments

```
raw.estimates A vector of raw reactivities.
```

Value

The normalization factor.

plot_dStructurome

Plots differentially reactive regions.

Description

Plots differentially reactive regions.

Usage

```
plot_dStructurome(rl, diff_regions, outfile, fdr = 0.05, ylim = c(-0.05, 3))
```

Arguments

rl List of dataframes of reactivities for each sample.

diff_regions Dataframe of regions with significance of differentially reactivity.

outfile The name for pdf file which will be saved.

fdr FDR threshold for plotted regions.

ylim Y-axis limits for plots.

Value

Saves a PDF for all differentially reactive regions. Returns NULL.

two.eight.normalize 9

two.eight.normalize Normalizes reactivity vector.

Description

Normalizes raw reactivities using 2-8 % method.

Usage

```
two.eight.normalize(raw.estimates)
```

Arguments

raw.estimates A vector of raw reactivities.

Value

A vector of normalized reactivities.

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