




PACS gene family-related neurological diseases: limited genotypes and diverse phenotypes

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Abstract

Background The *PACS* gene family has been demonstrated to be related to intracellular vesicular trafficking. The phenotypic manifestations caused by the pathogenic variants of *PACS* include epilepsy, intellectual disability/developmental delay, and malformations, such as facial abnormalities.

Methods We identified seven new cases with pathogenic or likely pathogenic *PACS* variants using next-generation sequencing. Detailed information obtained from these patients was analyzed along with that obtained from previously reported patients.

Results With the inclusion of the newly diagnosed cases in this study, 103 cases with *PACS* gene family-related neurological diseases were reported, of which 43 were *PACS2*-related cases and the remaining were *PACS1*-related cases. Most patients had seizures, which have been reported to be effectively controlled by several types of anti-seizure medications (ASMs). The most efficacious and frequently prescribed ASMs included sodium valproate (43.3%, 13/30), oxcarbazepine/carbamazepine (26.7%, 8/30), and levetiracetam (20%, 6/30). Almost all patients had intellectual disability/developmental delay. The most common pathogenic missense variants were *PACS1* p. Arg203Trp and *PACS2* p. Glu209Lys. In addition, we report a patient carrying a likely pathogenic copy number variation (CNV) (de novo heterozygous deletion of chr14:105821380-106107443, 286 kilobase, destroyed part of the furin-binding region domain and the protein structure after it) with more severe and refractory late-onset epilepsy.

Conclusions The clinical phenotypes of the different *PACS* heterozygous missense variants were similar. The pathogenic variant sites of *PACS1* and *PACS2* were quite limited but located in different regions. A CNV destroying part of the *PACS2* gene might also be pathogenic. These findings may provide an important clue for further functional studies on the pathogenic mechanism of neurological disorders related to the *PACS* gene family.

Keywords Developmental delay · Epilepsy · Intellectual disability · *PACS1* · *PACS2*

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Introduction

The *PACS* gene family, which has been demonstrated to be involved in intracellular vesicular trafficking, organellar communication and gene expression [1–3], includes two highly homologous genes in humans, *PACS1* and *PACS2*. Variants of both genes are reportedly associated with neurological disorders. The pathogenic variants of *PACS2* are associated with developmental and epileptic encephalopathy type 66 (OMIM #618067). Since 2018 [4], 10 studies have reported a total of 38 cases of *PACS2*-related neurological disease [5–14]. Based on the available clinical records, most patients had seizures and intellectual disability (ID)/developmental delay (DD). Some patients also exhibited

structural malformations, such as facial abnormalities. The pathogenic variants of *PACSI* are associated with Schuurs-Hoeijmakers syndrome (OMIM #615009). Since 2012 [15], 59 cases of *PACSI*-related neurological diseases have been reported, whose chief presentation included facial abnormalities, ID/DD, seizures, and structural malformations [16, 17]. Thus, the clinical phenotypes resulting from the pathogenic variants of *PACSI* and *PACS2* were similar, but the pathomechanism remains to be elucidated. In this study, we reported six cases of *PACS*-related neurological disorders, including five cases of *PACS2*-related neurological disorder and one case of *PACSI*-related neurological disorder. In addition, we report that a CNV (copy number variation) (heterozygous deletion of chr14:105821380-106107443, 286 kilobase) might cause a *PACS2*-related disorder. We also collated, summarized, and analyzed all reported cases of *PACS* gene family-related neurological diseases and our newly diagnosed cases to elucidate the genotypic and phenotypic characteristics of *PACS*-related neurological disease.

Methods

Ethics statement

This study was approved by the Ethical Committee of Peking University First Hospital. The patients' parents provided written informed consent for the publication of their case details. All data were analyzed anonymously.

Participants

Pathogenic or likely pathogenic variants were identified in seven Chinese patients with epilepsy. All available data, including clinical manifestations, electroencephalography (EEG) findings, brain magnetic resonance imaging (MRI) features, and gene variant information, pertaining to these seven patients and previously reported patients with *PACS* gene family-related neurological disease were collated. Patients were excluded if they met one of the following conditions: (1) the pathogenicity of the variant was uncertain; (2) the original paper was not written in English or Chinese; and (3) the cases were found in public databases.

Variation analysis

We collected 5 mL of peripheral venous blood from the patients and their parents. Genomic DNA was extracted for trio-based whole-exome sequencing (WES). The pathogenicity of the variants was predicted using Sorting Intolerant from Tolerant, Polymorphism Phenotyping version 2,

combined annotation-dependent depletion, and Mutation Taster. Variants were evaluated against the overall population in the 1000 Genomes Project, GnomAD, and ExAC databases. Pathogenicity was classified using variant classification standards based on the 2015 American College of Medical Genetics and Genomics guidelines. Patients with a suspected causative CNV identified by WES also underwent whole genome sequencing (WGS) to identify the extent of the CNV and analyze its pathogenicity.

Results

Phenotypic summary

With the inclusion of the new cases diagnosed in the current study, 43 cases of *PACS2*-related neurological diseases and 60 cases of *PACSI*-related neurological diseases have been reported to date. Table 1 summarizes the information on all *PACS2*-related neurological disease cases, except for those with incomplete clinical data. Seto et al. summarized the information of 52 cases of *PACSI*-related neurological disease in 2020 [16], and Tenorio-Castano et al. added seven cases in 2021 [17]. Epilepsy, ID/DD, and structural abnormalities were the chief manifestations of *PACS* gene family-related neurological diseases.

Seizures

Seizures were the characteristic and most frequent first symptom of *PACS* gene family-related diseases. All 28 *PACS2*-related patients, whose detailed information was available, experienced seizures as the first symptom, whereas 60% of patients with *PACSI*-related neurological diseases had seizures [16]. Focal seizures were the principal seizure type; 59.4% (19/32) of patients with detailed records had focal seizures. Moreover, tonic seizures, generalized tonic–clonic seizures, clonic seizures, and myoclonic seizures were also observed in some patients. Most patients' seizures were easily controlled by anti-seizure medications (ASMs), and 6/22 patients with *PACS2*-related neurological diseases (with complete medication data records) had used only one ASM, and 10/22 had used two ASMs. However, detailed records of the seizures were available for only three previously reported patients with *PACSI*-related neurological diseases, and the follow-up duration was short. In 2018, Martinez-Monseny et al. reported a patient with c.607C>G (p. Arg203Trp) [18], who experienced seizure onset at 3 years of age and presented with focal seizures that were treated with oral carbamazepine. Both patients carrying c.607C>G (p. Arg203Trp) reported by Hoshino et al.

Table 1 Detailed data of 28 patients with PACS2-related neurological diseases

Case	Variant	Sex	Epilepsy	ID/DD				Physical development				Others
				Age of onset	Seizure type	Number of ASMs used	Last seizure episode	Effective ASM	Microcephaly	Facial abnormalities	Distal limb abnormalities	
1. Olson et al. [4]	p.Glu209Lys	F	6 d	Focal	1	NA	+	–	+	+	–	Myopia
2. Olson et al. [4]	p.Glu209Lys	F	4 d	GTCS	2	6 mon	+	–	+	+	–	Dextrocardia
3. Olson et al. [4]	p.Glu209Lys	M	4 d	NA	1	CBZ	+	–	+	+	–	Cryptorchidism
4. Olson et al. [4]	p.Glu209Lys	F	7 d	GTCS	4	VPA	+	–	+	+	–	Strabismus
5. Olson et al. [4]	p.Glu209Lys	M	2 d	GTCS, clonic	3	NA	+	–	+	+	–	Strabismus
6. Olson et al. [4]	p.Glu209Lys	M	2 d	SE	1	NA	+	–	+	+	–	Myopia, VSD, cryptorchidism
7. Olson et al. [4]	p.Glu209Lys	M	2 d	Focal, tonic, clonic	1	NA	+	–	+	+	–	Cryptorchidism, frequent infection, hearing difficulties
8. Olson et al. [4]	p.Glu209Lys	F	2 wk	Focal, tonic	2	NA	+	–	+	–	–	Astigmatism
9. Olson et al. [4]	p.Glu209Lys	F	2 d	Focal tonic, myoclonic, GTCS, generalized tonic	2	NA	+	–	+	–	–	Myopia, visual impairment
10. Olson et al. [4]	p.Glu209Lys	M	1 mon	Clonic, GTCS	1	NA	+	–	+	+	–	

Table 1 (continued)

Case	Variant	Sex	Epilepsy	Epilepsy				Physical development				Others		
				Age of onset	Seizure type	Number of ASMs used	Last seizure episode	Effective ASM	ID/DD	Microcephaly	Facial abnormalities	Distal limb abnormalities	Brain MRI abnormalities	Malnutrition
11. Olson et al. [4]	p.Glu209Lys	M	1 d	Focal, GTCS	2	2 mon	NA	NA	+	–	+	+	+	–
12. Olson et al. [4]	p.Glu209Lys	F	3 d	Focal tonic–clonic, SE	3	3 mon	OXC	OXC	+	–	+	+	+	–
13. Olson et al. [4]	p.Glu209Lys	F	2 wk	Focal tonic–clonic, generalized	7	3 y	NA	NA	+	–	+	–	+	–
14. Olson et al. [4]	p.Glu209Lys	F	3 d	Tonic, GTCS	4	4 y	LEV LCM	LEV LCM	+	–	+	+	+	–
15. Dentici et al. [5]	p.Glu209Lys	F	2 d	Focal→tonic	3	4 mon	CBZ	CBZ	+	–	+	–	+	–
16. Simmen et al. [3]	p.Glu209Lys	F	2 wk	Tonic	NA	2 y	CBZ CLB	CBZ CLB	+	–	+	–	+	–
17. Simmen et al. [3]	p.Glu209Lys	F	3 d	Focal or generalized tonic LGS	NA	9 y	TPM ZNS CZP LTG VPA	TPM ZNS CZP LTG VPA	+	–	+	–	–	–
18. Simmen et al. [3]	p.Glu209Lys	F	3 d	Tonic	NA	9 mon	CBZ	CBZ	–	–	+	–	±	–
19. Olson et al. [7]	p.Glu209Lys	F	7 d	Focal, tonic, clonic	2	2 y 3 mon	VPA	VPA	+	–	+	–	–	–
20. Olson et al. [7]	p.Glu209Lys	F	5 d	Focal	4	1 y	VPA LEV	VPA LEV	+	+	+	–	+	–
21. Olson et al. [7]	p.Glu209Lys	F	3 d	Tonic	2	11 mon	VPA LEV	VPA LEV	+	–	+	–	–	–
22. Wu et al. [10]	p.Glu209Lys	M	3 d	Focal	1	NA	NA	NA	+	–	–	–	–	–

Table 1 (continued)

Case	Variant	Sex	Epilepsy	Age of onset	Seizure type	Number of ASMs used	Last seizure episode	Effective ASM	ID/DD				Physical development				Others	
23.	p.Glu209Lys	M		3 d	Focal, GTCS	1	1 y 4 mon	VPA	+	-	-	+	+	-	-	-	-	Newly diagnosed
24.	p.Glu209Lys	F		3 d	Focal	2	1 y 1 mon	VPA OXC	+	-	-	-	-	+	-	-	-	Newly diagnosed
25.	p.Glu209Lys	M		< 1 mon	Focal	2	17 y	VPA LEV	Mild	-	-	-	-	-	-	-	-	Newly diagnosed
26.	p.Glu209Lys	F		2 d	Focal	2	1 y 9 mon	PB VPA	+	-	-	-	-	-	-	-	-	Newly diagnosed
27.	p.Glu209Lys	F		1 mon 7 d	Focal	2	4 mon	LEV OXC	Mild	+	+	+	+	-	-	+	+	Newly diagnosed
28. Wan et al. [2]	p.Glu211Lys	M		3 d	Tonic	3	13 mon	VGB VPA	+	-	-	-	+	+	-	-	±	-

Detailed data of other reported cases were not available from published papers. F female, M male, ASM anti-seizure medication, ASD atrial septal defect, VSD ventricular septal defect, NA not available, GTCS generalized tonic-clonic seizure, VPA sodium valproate, CBZ carbamazepine, OXC oxcarbazepine, LEV levetiracetam, LCM lacosamide, VGB vigabatrin, CLB clobazam, TPM topiramate, ZNS zonisamide, CZP clonazepam, LTG lamotrigine, PB phenobarbital, MRI magnetic resonance imaging

in 2019 had seizures [19]. Onset of tonic seizures occurred in the neonatal period in one patient, which could be completely controlled by oral phenobarbital; EEG performed during the inter-ictal period showed no abnormalities. In the other patient, seizures developed two days after birth and manifested as apnea, which was completely controlled by oral phenobarbital; the seizures recurred at 2 years and 4 months of age after ASM discontinuation and were treated with oral sodium valproate. In our study, one newly diagnosed patient with *PACSI*-related neurological disease presented with focal seizures at four days after birth and was treated with phenobarbital and levetiracetam, and no seizures were observed for up to two months. The patient experienced another seizure episode during discontinuation of phenobarbital and dose reduction of levetiracetam. Sodium valproate and levetiracetam were administered four months after birth, and complete control of the seizures was achieved again. The patient had no seizures from 6 months through 9 years of age, i.e., the present day. Several types of ASMs have been reported to be effective for the treatment of epilepsy. The most efficacious and frequently used ASMs were sodium valproate (43.3%, 13/30), oxcarbazepine/carbamazepine (26.7%, 8/30), and levetiracetam (20%, 6/30). Most patients achieved freedom from seizures. ASMs achieved complete seizure control within 6 months of age in 8 of 24 (33.3%) patients with *PACS* gene family-related neurological diseases whose complete epilepsy treatment was available (including case 25, who experienced recurrence at 17 years of age, which was controlled by levetiracetam monotherapy), and complete seizure control was achieved within 2 years of age in 18 of 24 (75%) patients. The patients' inter-ictal EEG showed slow background rhythm and abnormal wave discharge. Focal seizures of unknown origin were recorded on EEG in case 25. EEG returned to normal in case 5 one month after complete seizure control.

Intellectual disability/developmental delay

Almost all patients (88/89) with *PACS* family gene-related neurological diseases had ID/DD of varying severity. Patients with milder ID/DD (case 25) could graduate from junior high school. At the age of 20 years, this patient could communicate to the bare minimum and imitate others to prepare simple handicrafts. However, his language skills were poor. Patients with more severe ID/DD were likely to have developmental regression and lack self-care ability, language ability, or motor abilities, such as running and jumping.

Structural abnormalities

Height and weight development were normal in patients with *PACS* family gene-related neurological diseases. The

most common physical malformations were facial abnormalities, as observed in 96.6% (86/89) of patients, which were characterized by orbital hypertelorism, narrow palpebral fissures, low and flat nasal bridges, spherical nasal tips, low-set ears, thin lips, and micrognathia. Some patients had microcephaly and dysplasia of the distal limbs (fingers and toes). Abnormal neuroimaging findings were observed in most patients: 60.7% (17/28) of patients with *PACS2*-related neurological diseases exhibited abnormal findings on brain MRI, which included widening of the subarachnoid space, cerebellar dysplasia, and dysplasia of the corpus callosum. However, patients with *PACS1*-related neurological diseases often present with cerebellar vermis dysplasia on brain MRI. Several patients had other malformations. Cardiac structural abnormalities [16], such as atrial septal defect (ASD), bicuspid aortic valve, aortic constriction, aorto-pulmonary dilatation, aortic valve dysplasia, pulmonary valve dysplasia, patent ductus arteriosus, patent foramen ovale, persistent left superior vena cava, and ventricular septal defect (VSD), were observed in 42% of patients with *PACS1*-related neurological diseases; 21% had skull deformities or microcephaly; 33% had ocular abnormalities, which manifested as astigmatism, strabismus, myopia, hyperopia, and optic disk irregularity; and 7.7% had structural abnormalities of the urinary tract, which manifested as duplex kidney, extra-renal pelvis, and posterior urethral compartment. Some patients had other structural abnormalities, such as cryptorchidism, congenital diaphragmatic hernia, and pulmonary dysplasia. Abnormalities have also been reported in *PACS2*-related diseases. The patients could present with cardiovascular abnormalities, such as ASD, VSD, and dextrocardia and ocular disorders, such as myopia, strabismus, and astigmatism.

Genotype summary

Forty-three of the 103 reported patients with *PACS* gene family-related neurological diseases carried pathogenic variants of *PACS2*, while 60 carried pathogenic variants of *PACS1*. Forty-two of 43 (97.7%) patients with *PACS2*-related neurological diseases carried a missense variant, c.625G>A (p. Glu209Lys), 1/43 patient (case 28) carried a missense variant, c.607C>G (p. Glu211Lys). A total of 59/60 (98.3%) patients with *PACS1*-related neurological diseases carried the same type of missense variant c.607C>G (p. Arg203Trp), and one carried another missense variant c.608G>A (p. Arg203Gln).

The sites of pathogenic variation in the *PACS1* and *PACS2* proteins were quite localized (Fig. 1). The pathogenic variants of *PACS1* were missense variants involving Arg203. Arg203 was located in the furin-binding region (FBR), and missense variation in this amino acid may have affected the recognition or binding between the *PACS1* protein and the cargo. The pathogenic point variants of *PACS2*

were also missense variants, resulting in the substitution of the acidic amino acid glutamate for the basic amino acid lysine. The variant amino acids were located in the middle region (MR), which plays a major regulatory role. The variant amino acids were all adjacent to the auto-regulatory area of the MR, with Glu209 being the last amino acid in the auto-regulatory region.

Patient carrying chr14:105821380-106107443 heterozygous deletion

Only one patient with a de novo heterozygous deletion was identified. The clinical phenotype of this patient differed considerably from that of the other patients. The disease history was characterized by late onset. The patient had essentially normal development until 1 year of age without seizures. DD occurred at approximately 1 year of age, and focal and tonic seizures developed from the ages of 1 year and 5 months. The severity of seizures was greater, status epilepticus was a common occurrence, and the administration of sedative drugs was needed. The patient had refractory epilepsy and had used four ASMs. No seizure episodes occurred for up to one year during the administration of oxcarbazepine and levetiracetam. The patient experienced seizures one month before the last follow-up, and drug titration was underway to obtain the optimal pharmacotherapeutic regimen. The patient's facial abnormalities were similar to those of other patients with *PACS* gene family-related neurological diseases.

Our study is the first to report that a heterozygous deletion, chr14:105821380-106107443 (286 kilobase), might be associated with the pathogenesis of *PACS*-related neurological disease. Trio-WES and CNV sequencing revealed that he carried this de novo CNV, and we performed WGS, which confirmed that he carried a heterozygous deletion measuring 286 kilobases in length (chr14:105821380-106107443) (Fig. 2a). This fragment contained *PACS2*, *C14orf80*, *CRIP1*, *CRIP2*, *IGHA2*, *IGHE*, *IGHG4*, *MTA1*, *TEX22*, and *TMEM121*. This CNV destroyed part of the FBR and the protein structure after it (Fig. 2b). This heterozygous deletion has not been reported in the Database of Genomic Variants. Karyotype analysis yielded normal results. Trio-WES and WGS did not reveal any other pathogenic variants associated with this phenotype.

Discussion

In humans, the *PACS* family includes the *PACS1* and *PACS2* genes. The pathogenic variants of both genes are associated with neurological diseases with similar clinical manifestations, chief of which are epilepsy, ID/DD of varying



Fig. 1 Comparison of PACS1 and PACS2 protein structures and variant sites. *ARR* atrophin-1-related region, *FBR* furin-binding region, *MR* middle region, *CTR* C-terminal region

severity, and structural abnormalities. Epilepsy is the most common first symptom, which mainly presents as focal seizures and could be associated with a variety of seizure types and status epilepticus. Seizures are easily controlled in most patients, and seizure freedom can be achieved by ASMs in infancy. The most efficacious and frequently used ASMs were sodium valproate, oxcarbazepine/carbamazepine, and levetiracetam. In these cases, the choice of ASM was usually based on the type of seizure. Broad-spectrum ASMs, such as sodium valproate and levetiracetam, are commonly used. Several cases were characterized by focal seizures and were therefore treated with oxcarbazepine/carbamazepine. The choice of pharmacotherapy is usually not based on the causative gene because *PACS*-related seizures are usually easy to control. Some patients had started to use effective ASMs before the causative gene was identified. Almost all patients had facial abnormalities, and some even had abnormal brain MRI findings, microcephaly, and multiple visceral structural malformations. However, the phenotype of the patient with heterozygous deletion of chr14:105821380-106107443 diagnosed in our study differed from that of patients with other *PACS2*-related neurological diseases. This patient had more severe and refractory epilepsy of later onset, in addition to frequent episodes of status epilepticus. The facial abnormalities and microcephaly observed in this patient were similar to those of the other patients.

PACS1 and *PACS2* are highly homologous [1], and both are derived from the prevertebrate *PACS* gene. Both genes have been implicated in intracellular vesicular trafficking [20]. *PACS1* localizes cargo to the trans-Golgi network, whereas *PACS2* is involved in vesicular transport from the Golgi to the endoplasmic reticulum (ER) and ER to the Golgi, cell membranes, mitochondria, and other pathways. Currently, *PACS2* is also considered to be related to apoptosis, autophagy and other pathways [1–3]. *PACS1* and *PACS2* (also called *PACS-1* and *PACS-2*) both contain three regions [20], viz. the FBR, MR, and C-terminal region (CTR). The

FBR, which is the functional domain, possesses the highest degree of homology and is involved in recognition and binding between the proteins and the cargo. The MR contains nuclear localization signals and phosphorylation sites, which play a regulatory role in the function of the *PACS*, while the specific function of CTR remains unclear. The two prevalent pathogenic variants of *PACS1* were missense mutations involving Arg203. Arg203 is located in the FBR, and the missense variation in this amino acid may have affected recognition or binding between the *PACS1* protein and the cargo. Both variants affected the same CpG dinucleotide. Studies in zebrafish have suggested a dominant-negative or gain-of-function patho-mechanism in *PACS1*-related neurological disorders [15, 21]. The two pathogenic point variants in *PACS2* were one amino acid apart; both involved switching of glutamate to lysine, resulting in remarkably similar clinical phenotypes, suggesting a similar pathogenic mechanism. Both variants were located in the MR, i.e., the regulatory domain, with Glu209 being the last amino acid in the auto-regulatory region. The presence of two indels in this region suggests that the structure near this site may be highly relevant for the regulatory function of *PACS2*. Co-immunoprecipitation analysis showed that the *PACS2* p. Glu209Lys reduced the reversibility of FBR binding to client proteins, which may disturb cellular function [4]. This finding indicated that, in contrast to *PACS1*, the missense variants of *PACS2* may be loss-of-function. On the other hand, the loss-of-function constraint measure of *PACS2* was 0.279 according to the GnomAD database, which suggested that the *PACS2* gene is less resistant to loss-of-function variants.

We reported a patient with a heterozygous deletion of chr14:105821380-106107443. This fragment contained *PACS2*, *C14orf80*, *CRIP1*, *CRIP2*, *IGHA2*, *IGHG4*, *MTA1*, *TEX22*, and *TMEM121*; however, none of the other genes, except *PACS2*, has been reported to be associated with epilepsy or ID/DD, and only *PACS2*, *CRIP1*, and *MTA1* are expressed in the central nervous system. This CNV

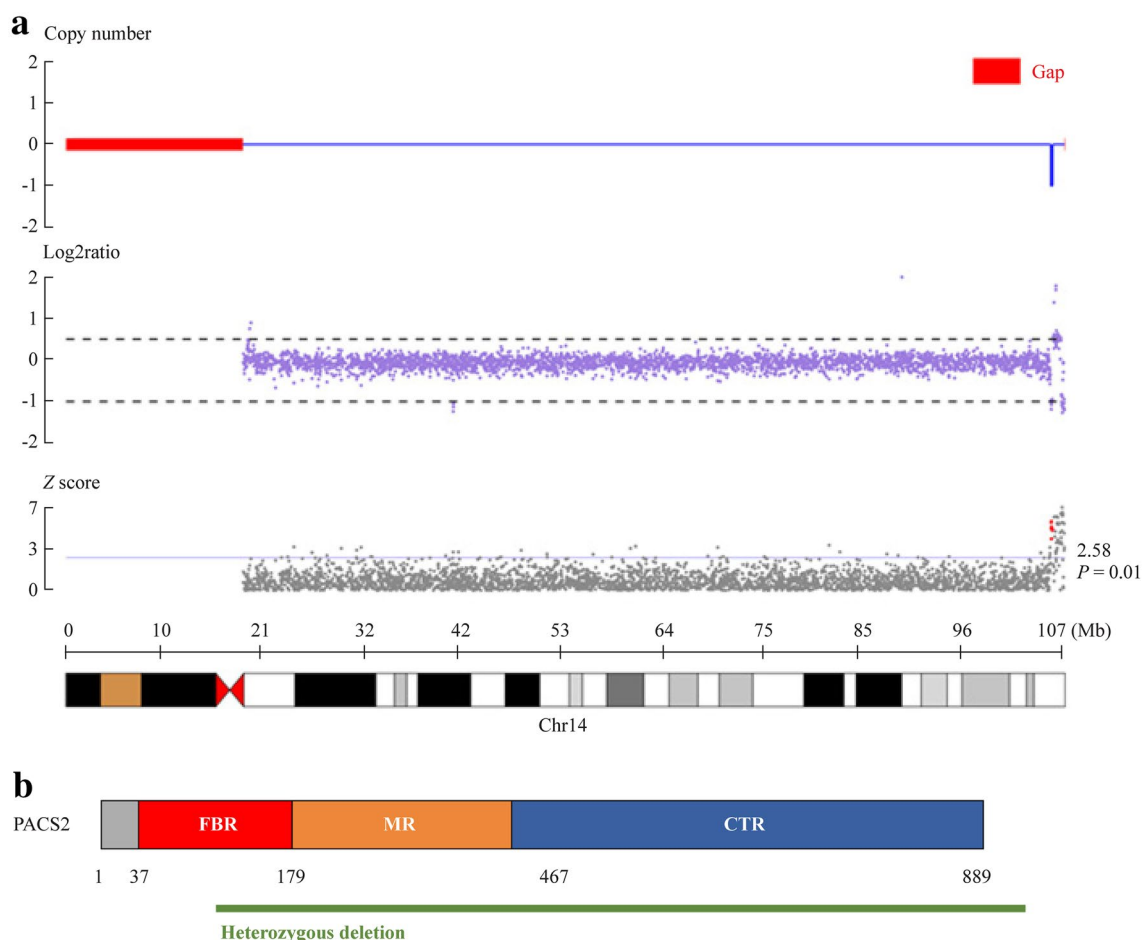


Fig. 2 The heterozygous deletion we reported. **a** WGS detected a heterozygous deletion of chr14:105821380-106107443 (286.06 kilobase); **b** this CNV caused deletion of a partial segment of PACS2,

including half of the FBR and the latter. WGS whole genome sequencing, CNV copy number variation, FBR furin-binding region, MR middle region, CTR C-terminal region

destroyed part of the FBR domain and the protein structure after it, which might significantly affect protein function, and therefore, we believe that this variant causes neurological disease mainly through its effect on PACS2. This microdeletion might be loss-of-function-like missense variants. However, more functional studies are needed. In 2012, Holder Jr et al. reported one patient carrying a 0.31-Mb microdeletion of chromosome 14q32.33 who presented with epilepsy, DD, and facial dysmorphism [22]. However, the pathogenicity could not be determined because they only performed CGH array and fragile X testing, and they did not analyze the segregation of the microdeletion in the family. The clinical phenotype of the patient with heterozygous deletion of chr14:105821380-106107443 diagnosed in our study may be different from that of patients with other PACS2-related neurological diseases. Does this suggest that PACS2 protein dose dependence in humans is related to developmental period? These findings may provide clues to elucidate the pathogenic mechanism and the precise treatment of

PACS2-related neurological diseases, although they require subsequent functional research. However, a patient with a heterozygous deletion of chr14:105821380-106107443 experienced later onset of epilepsy and severe disease. Hence, the possible pathogenic contribution of other genes in the fragment showing high expression in the nervous system, including *CRIP1* and *MTA1*, to the epileptic phenotypes or refractory mechanisms still cannot be discounted.

In conclusion, this study reported one newly diagnosed patient with *PACS1* and five patients with *PACS2*-related neurological disease in a Chinese population and reported that the heterozygous deletion of chr14:105821380-106107443 might cause *PACS2*-related disorders. We summarized the clinical phenotypes and genotypes of the cases diagnosed over the course of this study and those of previously reported cases and found that epilepsy and structural changes associated with heterozygous missense variants in all *PACS* family genes were similar. Patients carrying heterozygous deletions in the *PACS2* gene have

a more severe neurological phenotype than those with other *PACS2*-related neurological diseases. The pathogenic mutation sites in the *PACS* family genes are quite limited, but the functional domains where the common mutations of *PACS1* and *PACS2* genes are located are different, viz. the FBR (the main functional domain) and MR (the regulatory domain), respectively. However, we could not explain the mechanism underlying the variations in the severity of the clinical phenotypes and high concentrations of *PACS1* and *PACS2* pathogenic variants in patients carrying the same *PACS2* pathogenic variants. This suggests a need for further investigation with a larger sample size and further analysis into the genetic background, relationship between phenotype and environment, and biological function. Our findings have provided an important cornerstone for further functional studies and research on the pathogenic mechanisms of associated neurological diseases, in addition to useful clues for future precision treatment studies.

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Author contributions ZH and GK contributed equally to this manuscript and they were co-first authors. ZH contributed to study design, collection, evaluation and interpretation of the data, and drafting and writing of the manuscript. WS, ZYH, YZX, and WY contributed to collection, evaluation and interpretation of the data. GK and JYW contributed to study design, collection, evaluation and interpretation of the data, and revised the manuscript and approved the final version.

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Data availability Most data generated or analyzed during this study are included in this published article. Other data are available from the corresponding author on reasonable request.

Declarations

Ethical approval This study was approved by the Ethical Committee of Peking University First Hospital. The individuals' parents in this manuscript had given written informed consent to publish these cases details. All data were analyzed anonymously.

Conflict of interest Author Yu-Wu Jiang is a member of Editorial Board for *World Journal of Pediatrics*. The paper was handled by the other Editor and has undergone rigorous peer review process. Author Yu-Wu Jiang was not involved in the journal's review of, or decisions related to this manuscript. All authors declared no financial or non-financial conflict of interests related to this paper.

References

- Li C, Li L, Yang M, Zeng L, Sun L. PACS-2: a key regulator of mitochondria-associated membranes (MAMs). *Pharmacol Res*. 2020;160:105080.
- Wan L, Molloy SS, Thomas L, Liu G, Xiang Y, Rybak SL, et al. PACS-1 defines a novel gene family of cytosolic sorting proteins required for trans-Golgi network localization. *Cell*. 1998;94:205–16.
- Simmen T, Aslan JE, Blagoveshchenskaya AD, Thomas L, Wan L, Xiang Y, et al. PACS-2 controls endoplasmic reticulum-mitochondria communication and Bid-mediated apoptosis. *EMBO J*. 2005;24:717–29.
- Olson HE, Jean-Marçais N, Yang E, Heron D, Tatton-Brown K, van der Zwaag PA, et al. A recurrent *de novo* *PACS2* heterozygous missense variant causes neonatal-onset developmental epileptic encephalopathy, facial dysmorphism, and cerebellar dysgenesis. *Am J Hum Genet*. 2018;102:995–1007.
- Dentici ML, Barresi S, Niceta M, Ciolfi A, Trivisano M, Bartuli A, et al. Expanding the clinical spectrum associated with *PACS2* mutations. *Clin Genet*. 2019;95:525–31.
- Mizuno T, Miyata R, Hojo A, Tamura Y, Nakashima M, Mizuguchi T, et al. Clinical variations of epileptic syndrome associated with *PACS2* variant. *Brain Dev*. 2021;43:343–7.
- Olson HE, Jean-Marçais N, Yang E, Heron D, Tatton-Brown K, van der Zwaag PA, et al. A recurrent *de novo* *PACS2* heterozygous missense variant causes neonatal-onset developmental epileptic encephalopathy, facial dysmorphism, and cerebellar dysgenesis. *Am J Hum Genet*. 2018;103:631.
- Terrone G, Marchese F, Vari MS, Severino M, Madia F, Amadori E, et al. A further contribution to the delineation of epileptic phenotype in *PACS2*-related syndrome. *Seizure*. 2020;79:53–5.
- Zhu T, Gong X, Bei F, Ma L, Chen Y, Zhang Y, et al. Application of next-generation sequencing for genetic diagnosis in neonatal intensive care units: results of a multicenter study in China. *Front Genet*. 2020;11:565078.
- Wu MJ, Hu CH, Ma JH, Hu JS, Liu ZS, Sun D. Early infantile epileptic encephalopathy caused by *PACS2* gene variation: three cases report and literature review. *Zhonghua Er Ke Za Zhi*. 2021;59:594–9 (in Chinese).
- Wang T, Hoekzema K, Vecchio D, Wu H, Sulovari A, Coe BP, et al. Large-scale targeted sequencing identifies risk genes for neurodevelopmental disorders. *Nat Commun*. 2020;11:4932.
- Monies D, Abouelhoda M, Assoum M, Moghrabi N, Rafiullah R, Almontashiri N, et al. Lessons learned from large-scale, first-tier clinical exome sequencing in a highly consanguineous population. *Am J Hum Genet*. 2019;104:1182–201.
- Jiang T, Gao J, Jiang L, Xu L, Zhao C, Su X, et al. Application of trio-whole exome sequencing in genetic diagnosis and therapy in Chinese children with epilepsy. *Front Mol Neurosci*. 2021;14:699574.
- Bruehl AL, Nambot S, Quéré V, Vitobello A, Thevenon J, Assoum M, et al. Increased diagnostic and new genes identification outcome using research reanalysis of singleton exome sequencing. *Eur J Hum Genet*. 2019;27:1519–31.
- Schuurs-Hoeijmakers JH, Oh EC, Vissers LE, Swinkels ME, Gilissen C, Willemsen MA, et al. Recurrent *de novo* mutations in

- PACSI* cause defective cranial-neural-crest migration and define a recognizable intellectual-disability syndrome. *Am J Hum Genet.* 2012;91:1122–7.
16. Seto MT, Bertoli-Avella AM, Cheung KW, Chan KY, Yeung KS, Fung JL, et al. Prenatal and postnatal diagnosis of Schuurs-Hoeijmakers syndrome: case series and review of the literature. *Am J Med Genet A.* 2021;185:384–9.
 17. Tenorio-Castaño J, Morte B, Nevado J, Martinez-Glez V, Santos-Simarro F, García-Miñaur S, et al. Schuurs-Hoeijmakers syndrome (*PACSI* neurodevelopmental disorder): seven novel patients and a review. *Genes (Basel).* 2021;12:738.
 18. Martinez-Monseny A, Bolasell M, Arjona C, Martorell L, Yubero D, Arsmtrong J, et al. Mutation of *PACSI*: the milder end of the spectrum. *Clin Dysmorphol.* 2018;27:148–50.
 19. Hoshino Y, Enokizono T, Imagawa K, Tanaka R, Suzuki H, Fukushima H, et al. Schuurs-Hoeijmakers syndrome in two patients from Japan. *Am J Med Genet A.* 2019;179:341–3.
 20. Thomas G, Aslan JE, Thomas L, Shinde P, Shinde U, Simmen T. Caught in the act-protein adaptation and the expanding roles of the PACS proteins in tissue homeostasis and disease. *J Cell Sci.* 2017;130:1865–76.
 21. Miyake N, Ozasa S, Mabe H, Kimura S, Shiina M, Imagawa E, et al. A novel missense mutation affecting the same amino acid as the recurrent *PACSI* mutation in Schuurs-Hoeijmakers syndrome. *Clin Genet.* 2018;93:929–30.
 22. Holder JL Jr, Lotze TE, Bacino C, Cheung SW. A child with an inherited 0.31 Mb microdeletion of chromosome 14q32.33: further delineation of a critical region for the 14q32 deletion syndrome. *Am J Med Genet A.* 2012;158A:1962–6.

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