

# Evaluation of Dimension-Reduction Methods (Huang et al. 2022)

Huang *et al.* propose five complementary strategies to evaluate DR methods on transcriptomic data <sup>1</sup> <sup>2</sup>. Each strategy has a clear purpose, specific implementation steps, and uses one or more quantitative metrics or illustrative datasets. Below we summarize each strategy, how it was carried out, and what was learned.

## Local structure preservation

Local structure preservation checks whether nearby points or same-class points remain neighbors after embedding <sup>3</sup>. The authors implement this in two ways:

- **Supervised (labeled).** Using labeled data  $(x_i, y_i)$ , they project the high-dimensional points  $x_i$  into 2D with each DR method. Then they train a classifier (typically SVM with an RBF kernel or k-NN with  $k = 5$  <sup>4</sup>) on a subset of the low-d embedding and test it on held-out points. High classification accuracy means points of the same label stayed close in 2D (homophily) <sup>5</sup>. For example, on the MNIST digit dataset, t-SNE, UMAP, TriMap and PaCMAP all gave high SVM/k-NN accuracy (closer to the original cluster separability), whereas ForceAtlas2 performed poorly <sup>6</sup>. Across many single-cell datasets, nearly all DR methods (except ForceAtlas2) achieved high accuracy: t-SNE, UMAP, PaCMAP and related methods typically scored best <sup>7</sup>.
- **Unsupervised (no labels).** When ground-truth labels are not available, they measure how well local neighborhoods are preserved. Concretely, for each point  $i$ , they find its  $k = 5$  nearest neighbors in high-dimensional space  $N(i)$  and its  $k = 5$  neighbors in the 2D embedding  $N'(i)$ . They compute the fraction  $|N(i) \cap N'(i)|/k$  for each  $i$  and average over all points <sup>8</sup>. A higher average means more of each point's original neighbors remain neighbors. Using this metric, t-SNE and its accelerated version (art-SNE) preserved the largest fraction of neighbors, while PCA (a purely linear method) preserved the fewest <sup>9</sup>.

**Key findings:** Methods designed for local clustering (t-SNE, UMAP, PaCMAP) excel at keeping class-neighbors together, yielding high classification accuracy and neighbor preservation <sup>7</sup> <sup>9</sup>. In contrast, ForceAtlas2 and PHATE underperform on these local metrics.

## Global structure preservation

Global structure preservation checks whether overall relationships and cluster arrangements are maintained. The authors use both qualitative and quantitative evaluations for this:

- **Qualitative examples.** They test on datasets with known large-scale structure (e.g. a synthetic “mammoth” point cloud, and simulated Gaussian-cluster data with a linear or hierarchical arrangement). By projecting these into 2D, they visually inspect whether connected parts stay together or false separations appear <sup>10</sup>. For instance, on the 3D “Mammoth” dataset, t-SNE and

UMAP fragmented the shape into disconnected clusters, whereas methods like TriMap and ForceAtlas2 preserved the overall form <sup>10</sup>. Similarly, on a Gaussian-linear test, methods like TriMap, PaCMAP and ForceAtlas2 showed a smooth color gradient reflecting the original high-dim structure, while t-SNE/UMAP did not <sup>11</sup>. On a harder hierarchical Gaussian dataset, ForceAtlas2 (and PaCMAP) did best at keeping micro-clusters of the same meso-cluster nearby <sup>12</sup>.

*Figure:* Qualitative examples of global-structure evaluation. In Huang *et al.*, a 3D “mammoth” shape (a) is projected into 2D by different DR methods (b); TriMap and ForceAtlas2 preserve the mammoth’s connected outline, whereas t-SNE and UMAP break it into disjoint pieces <sup>10</sup>. Similar tests on synthetic Gaussian data (c,d) highlight which methods retain large-scale cluster arrangements <sup>11</sup>.

- **Quantitative metrics.** The paper defines several metrics that compare distances or cluster relationships in high- and low-dimensional spaces:
- **Random Triplet Accuracy:** Sample many random triplets of points and check if the ordering of their pairwise distances is the same in high-D and low-D (i.e. if the closest pair remains closest, etc). The metric is the fraction of triplets with preserved order <sup>13</sup>. (They use a sample of triplets for tractability.) A higher triplet accuracy means global distance relations are well-preserved. On this metric, TriMap scored highest (consistent with its optimizing a triplet loss), while PCA, PaCMAP and ForceAtlas2 also performed well. In contrast, t-SNE, UMAP and PHATE scored relatively lower <sup>14</sup>.
- **Distance Spearman Correlation:** Compute all (or many) pairwise distances in the original data and in the embedding, then compute Spearman’s rank correlation between these two distance vectors <sup>15</sup>. A high correlation means the global distance ranking is maintained. Again, TriMap, PCA, PaCMAP and ForceAtlas2 scored well on most datasets, while t-SNE, UMAP (and art-SNE) tended to score worse <sup>15</sup>.
- **Cluster-centric metrics:** They also measure how well inter-cluster relationships are preserved by comparing cluster centroids. One metric is **k-Nearest Class Preservation:** for each class centroid, see how many of its  $k$  nearest-neighbor centroids in high-D remain its nearest neighbors in 2D <sup>16</sup>. (They set  $k = \lfloor (C + 2)/4 \rfloor$ , where  $C$  is the number of classes, to adapt to dataset size.) Another is **Centroid Distance Correlation:** compute the Spearman correlation between the set of all inter-centroid distances in high-D and in low-D <sup>17</sup>. These metrics again favored methods like TriMap and ForceAtlas2. On k-nearest class preservation, PCA and ForceAtlas2 were most consistent across datasets <sup>18</sup>, and on centroid-distance correlation TriMap and ForceAtlas2 scored highest <sup>17</sup>.

**Key findings:** Methods optimized for global geometry (TriMap, ForceAtlas2, PaCMAP) outperform local-only methods in these global metrics <sup>14</sup> <sup>17</sup>. By contrast, t-SNE and UMAP – which focus on local neighborhoods – often produce high-quality local clusters but badly distort the large-scale layout (creating “false” clusters) <sup>10</sup> <sup>15</sup>.

## Sensitivity to parameter choices

This evaluation checks whether small changes in DR tuning parameters lead to large changes in the embedding (which would indicate instability) <sup>2</sup>. The steps are:

1. **Select datasets with known structure** (e.g. Kazer *et al.* [immune cells] and Stuart *et al.* [hematopoietic progenitors]) <sup>19</sup>.

2. **Vary key parameters** for each algorithm. For example, for t-SNE they varied *perplexity*, for UMAP varied *n\_neighbors*, and for TriMap varied *n\_inliers*. Each parameter controls the “spread” or locality of the embedding forces <sup>19</sup>.
3. **Generate multiple embeddings**. For each choice of parameter (within a reasonable range), run the DR algorithm from the same preprocessed input (they used PCA to 70 PCs initially).
4. **Compare cluster relationships**. Examine how distances between known cell-type clusters change. In their figures, changing t-SNE’s perplexity or UMAP’s *n\_neighbors* caused the distance between certain cell clusters (e.g. two types of dendritic cells, or two progenitor types) to vary dramatically <sup>19</sup>. In some cases a single cell type even split into two clusters under one setting but not another (black circles in Fig.5a) <sup>19</sup>.
5. **Interpret sensitivity**. Large shifts in cluster arrangement indicate that the DR method is sensitive to its parameters. The authors note that t-SNE and UMAP showed strong sensitivity (cluster distances moved a lot) <sup>19</sup> <sup>20</sup>. By contrast, TriMap and PaCMAP embeddings were more stable across parameter changes in these examples (their colored clusters stayed in roughly the same relative positions) <sup>20</sup>.

*Figure:* Example of parameter-sensitivity tests (from Huang *et al.*). Changing t-SNE’s perplexity or UMAP’s *n\_neighbors* dramatically alters the distance between certain cell clusters (circled), demonstrating instability <sup>19</sup>. In the lower panel, UMAP’s output on two progenitor cell types shifts markedly as *n\_neighbors* changes, whereas TriMap’s output is more consistent <sup>20</sup>.

**Key point:** The authors do not compute a single metric here, but emphasize visual inspection. Methods whose embeddings change dramatically with parameter tweaks are deemed “sensitive”. Their results showed that popular methods like t-SNE and UMAP are often highly sensitive, whereas TriMap/PaCMAP tend to be more robust to parameter variation <sup>19</sup> <sup>20</sup>.

## Sensitivity to preprocessing choices

This test examines how the embedding changes when the input preprocessing is varied. Transcriptomic data usually undergo log-normalization and PCA before DR, but alternative schemes exist. The study’s steps were:

1. **Vary PCA dimensionality**. After log-normalizing the data, they applied PCA to reduce to different numbers of principal components (e.g. 30, 50, 70, 100) before running the DR methods <sup>21</sup>. They tested this on two scRNA-seq datasets (Kazer and Stuart). For each method and PC setting, they generated the 2D embedding.
2. **Compare cluster arrangement**. They focused on known related cell types (e.g. two dendritic cell subtypes) and measured the distance between those clusters in the embedding. As shown in Fig.6, t-SNE and UMAP produced very different cluster distances when using 50 vs 70 PCs, whereas TriMap and PaCMAP embeddings were nearly unchanged <sup>21</sup>. In one case, UMAP with 50 PCs placed two related progenitor clusters far apart, whereas with 70 PCs they were close (splitting one cluster into two) <sup>21</sup>.
3. **Vary preprocessing pipeline**. Beyond PCA, they tested three pipelines on one dataset: (a) **raw data + PCA**, (b) **log-normalization + PCA**, and (c) **GLM-PCA** (a specialized PCA for count data) <sup>22</sup>. Again, they applied each DR method to each preprocessed input.
4. **Check stability of results**. In Fig.7, t-SNE and UMAP embeddings changed dramatically between the three pipelines – the distance between two dendritic-cell clusters (mDC vs pDC) varied wildly and

many outliers appeared under GLM-PCA <sup>22</sup> <sup>23</sup> . PaCMAP's embeddings, however, showed only minor differences across preps.

*Figure:* Example of preprocessing-sensitivity tests. On the Stuart *et al.* dataset, three preprocessing pipelines were tried (raw+PCA, log-normalized+PCA, GLM-PCA). The black-circled DC clusters (mDC vs pDC) move relative to each other in t-SNE/UMAP embeddings as the pipeline changes, whereas PaCMAP's result is more consistent <sup>22</sup> .

**Key point:** An ideal DR method would be robust to such choices. Huang *et al.* found that t-SNE and UMAP were *not* robust: their embeddings (cluster distances, outlier count, etc.) changed a lot with different preprocessing <sup>22</sup> . By contrast, TriMap and especially PaCMAP were relatively insensitive to the number of PCs and to using GLM-PCA <sup>21</sup> <sup>22</sup> .

## Computational efficiency and scalability

This strategy measures running time of each method on datasets of increasing size <sup>24</sup> . The procedure was:

1. **Select datasets of various sizes.** They used multiple scRNA-seq datasets (from a few thousand to >1 million cells) discussed throughout the paper <sup>24</sup> .
2. **Time each DR algorithm.** For each method, run it with default settings to embed each dataset from  $N \times d$  to  $N \times 2$ , recording the elapsed time. (For methods like ForceAtlas2 that require a graph, they included graph construction time for fairness <sup>24</sup> <sup>25</sup> .)
3. **Compare run times.** In Fig.8, PaCMAP consistently took the least time on almost all datasets, especially the largest ones <sup>26</sup> . UMAP, TriMap and t-SNE also scaled reasonably well up to  $\sim 10^5$ – $10^6$  cells (t-SNE using a fast implementation) <sup>26</sup> . In contrast, art-SNE and PHATE were much slower: art-SNE ran out of memory on the  $\sim 10^6$ -cell datasets, and PHATE failed to finish within 24 hours on those large sets <sup>26</sup> <sup>25</sup> .

*Figure:* Running time vs. sample size for various DR algorithms (log-log scale). PaCMAP (purple) is fastest across all sizes, UMAP (green) and TriMap (red) also scale well. art-SNE (brown) and PHATE (pink) become impractical on the largest datasets <sup>26</sup> .

**Key findings:** PaCMAP is the fastest for large datasets, enabling rapid exploration of very large single-cell datasets <sup>26</sup> . UMAP, TriMap and optimized t-SNE are also reasonably fast. ForceAtlas2, art-SNE and PHATE are significantly slower and may even fail on multi-million-cell data under typical resource limits <sup>26</sup> <sup>25</sup> . This has practical importance: a faster method allows users to rerun DR many times (e.g. for parameter tuning) and handle modern large-scale transcriptomic datasets.

**Summary:** Huang *et al.*'s five-fold evaluation framework systematically probes different aspects of DR quality. Local-preservation tests (supervised kNN/SVM accuracy and neighbor overlap) identify how well nearby points stay together <sup>6</sup> <sup>9</sup> . Global-preservation tests (triplet/distance correlations and cluster-centroid metrics) quantify how well overall geometry is maintained <sup>13</sup> <sup>17</sup> . Sensitivity analyses reveal which algorithms yield stable embeddings across parameter or preprocessing changes <sup>19</sup> <sup>22</sup> . And runtime comparisons highlight practical efficiency and scalability <sup>24</sup> . Together, these evaluations show that methods like TriMap, ForceAtlas2 and PaCMAP are strong on global structure and robustness, while t-SNE/UMAP excel at local clustering but often distort global layouts and require careful parameter tuning <sup>10</sup> <sup>19</sup> .

**Sources:** All information above is drawn from the sections "Evaluation 1–5" of Huang *et al.* (2022), *Communications Biology*, which details the purpose, methodology, metrics, and findings for each evaluation strategy <sup>3</sup> <sup>24</sup> .

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<sup>1</sup> <sup>2</sup> <sup>3</sup> <sup>4</sup> <sup>5</sup> <sup>6</sup> <sup>7</sup> <sup>8</sup> <sup>9</sup> <sup>10</sup> <sup>11</sup> <sup>12</sup> <sup>13</sup> <sup>14</sup> <sup>15</sup> <sup>16</sup> <sup>17</sup> <sup>18</sup> <sup>19</sup> <sup>20</sup> <sup>21</sup> <sup>22</sup> <sup>23</sup> <sup>24</sup> <sup>25</sup> <sup>26</sup> Towards a comprehensive evaluation of dimension reduction methods for transcriptomic data visualization | *Communications Biology*

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