

## REVIEW

## Binocular response modulation in the lateral geniculate nucleus

Kacie Dougherty<sup>1</sup>  | Michael C. Schmid<sup>2</sup>  | Alexander Maier<sup>1</sup> 

<sup>1</sup>Department of Psychology, Center for Cognitive and Integrative Neuroscience, Vanderbilt Vision Research Center, Vanderbilt University, Nashville Tennessee 37203

<sup>2</sup>Institute of Neuroscience, Newcastle University, Newcastle upon Tyne, NE2 4HH, UK

## Correspondence

Alexander Maier, Department of Psychology, Center for Cognitive and Integrative Neuroscience, Vanderbilt Vision Research Center, Vanderbilt University, Nashville, TN 37203.  
Email: alex.maier@vanderbilt.edu

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## Abstract

The dorsal lateral geniculate nucleus of the thalamus (LGN) receives the main outputs of both eyes and relays those signals to the visual cortex. Each retina projects to separate layers of the LGN so that each LGN neuron is innervated by a single eye. In line with this anatomical separation, **visual responses of almost all of LGN neurons are driven by one eye only.** Nonetheless, many LGN neurons are sensitive to what is shown to the other eye as their visual responses differ when both eyes are stimulated compared to when the driving eye is stimulated in isolation. This, predominantly suppressive, binocular modulation of LGN responses might suggest that the LGN is the first location in the primary visual pathway where the outputs from the two eyes interact. Indeed, **the LGN features several anatomical structures that would allow for LGN neurons responding to one eye to modulate neurons that respond to the other eye.** However, it is also possible that binocular response modulation in the LGN arises indirectly as the LGN also receives input from binocular visual structures. Here we review the extant literature on the effects of binocular stimulation on LGN spiking responses, highlighting findings from cats and primates, and **evaluate the neural circuits that might mediate binocular response modulation in the LGN.**

## KEYWORDS

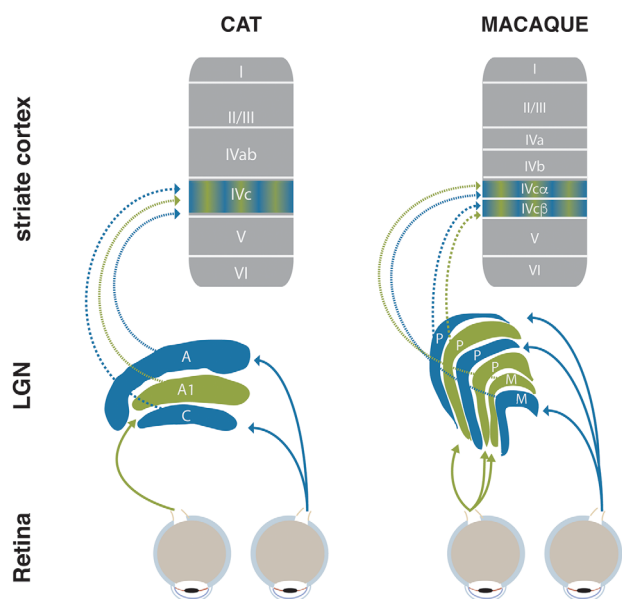
binocular combination, binocular integration, binocular vision, lateral geniculate nucleus, neurophysiology

## 1 | INTRODUCTION

Our visual perception of the world is characterized by a singular view despite the fact that each eye's perspective differs from the other. To create this unified perspective, our brains need to combine the separate outputs of the two eyes into a unified binocular signal. In order to allow for this binocular combination, outputs from one eye must meet and interact with outputs from the other eye. Knowing where the outputs from the two eyes converge is critical for our understanding of binocular vision and promises the discovery of new therapeutic targets for binocular vision disorders such as strabismus and amblyopia. The neuroanatomical framework of binocular convergence has been authoritatively summarized in seminal work by Casagrande and Boyd (1996). Here we **provide an extension of their preeminent review, adding recent developments and functional measurements that augment the structural findings.**

The primary visual pathway of mammals accommodates two structures where the outputs of the two eyes might first meet and interact (Figure 1): Retinal ganglion cells from each eye project in two isolated

streams to the dorsal lateral geniculate nucleus of the thalamus (LGN). The LGN projects to the primary visual cortex, which constitutes a bottleneck for visual input to all other cortical areas (Felleman & Van Essen, 1991; Lennie & Movshon, 2005; Markov et al., 2013; Schmid et al., 2013; Schmiedt et al., 2014). The primary visual cortex is the first structure in the primary visual pathway where almost all neurons are excited by stimulation of either eye (Hubel & Wiesel, 1962; Hubel & Wiesel, 1977; Smith et al., 1997). This feature suggests that the signals from the two eyes are combined, or merged together, in this structure (Casagrande & Kaas, 1994; Hubel & Wiesel, 1969). Unlike in primary visual cortex, almost all neurons at the previous stage of visual processing, the LGN, are excited by stimulation of one eye only. This means that only stimulation from one eye, but not the other, will result in a significant increase or decrease of in the frequency of action potentials, or spikes, in LGN neurons. Nevertheless, the LGN is a candidate structure for the two eyes' outputs to meet and interact, in that **the spike rate of a neuron that responds to the neuron's driving eye is modulated (enhanced or suppressed) when the other eye is stimulated as well.** This type of binocular modulation could serve computations that



**FIGURE 1** Simplified schematic of primary visual pathway of cat (left) and macaque monkeys (right). Retinal neurons project visual sensory information to the LGN, which is divided into several, eye-specific layers (blue and green). LGN neurons primarily project to layer 4 (arrows) as well as other sublayers of striate visual cortex. Note the differences in anatomy and nomenclature between cats and monkeys

require the two eyes' outputs to interact before they are merged together.

For example, we know that the visual system adjusts the relative strength of each eye's outputs before merging them into a single binocular signal. This fact is evidenced by the observation that our visual perception hardly changes when we close one eye, despite the fact that this action virtually halves the visual input to the visual system. In order to account for the difference in visual activation between these two viewing conditions, the brain needs to adjust the relative strength, or gain, of the signal from each eye in a way that depends on the activation of the other eye. This computational step likely takes place prior to binocular merging because the relative strength of the outputs from each eye are lost in the merged binocular signal. Indeed, several neurophysiological and psychophysical studies on this subject convergently determined that **the gain of the outputs of the eyes are adjusted while the two signals are still separate** (Baker, Meese, & Summers, 2007; Ding & Sperling, 2006; Meese, Georgeson, & Baker, 2006; Moradi & Heeger, 2009; Truchard, Ohzawa, & Freeman, 2000).

Based on the above, we can conclude that the outputs from the two eyes meet and interact somewhere in the brain before binocular combination in primary visual cortex. This neural process termed **binocular modulation** is most likely carried out by neurons that are excited by one eye only (so-called monocular neurons), but also modulate their responses depending on what is shown to the other eye. In other words, when monocular neurons are excited by a visual stimulus in their driving, "dominant eye," this visual response is either enhanced or inhibited when the other, "nondominant eye" is also stimulated. These

two types of binocular modulation have been termed binocular facilitation and binocular suppression, respectively. Based on the anatomy discussed above, binocular modulation could arise in three different ways along the primary visual pathway: (a) Binocular modulation is exclusively generated by LGN neurons. (b) Binocular modulation is exclusively generated by V1 neurons, and LGN neurons inherit binocular modulation following cortical feedback. (c) Binocular modulation is generated by both LGN and V1 neurons (Figure 2). Below, we will discuss these three alternatives, and summarize our current understanding of binocular modulation in the primary visual pathway.

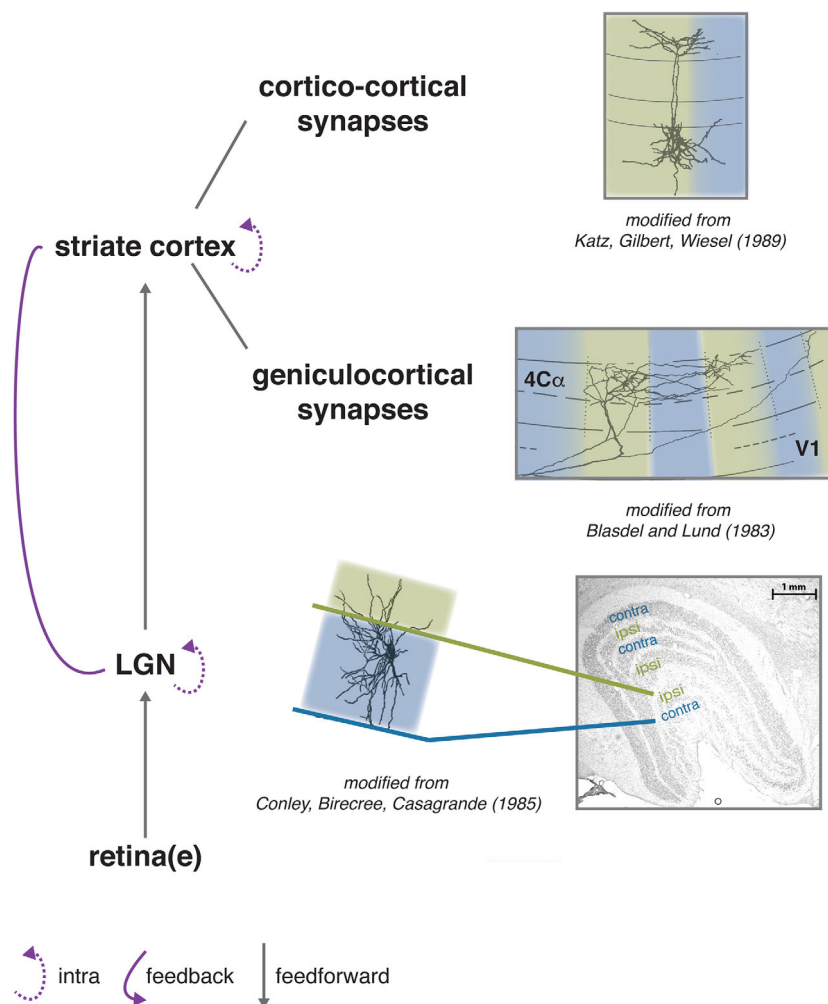
## 2 | BINOCULAR MODULATION OF LGN SPIKING RESPONSES

Over the past half century, neurophysiological studies across many eminent laboratories have probed whether LGN neurons exhibit binocular modulation. The bulk of this work was based on extracellular recordings in anesthetized cats and monkeys. These model species were chosen because their eyes are positioned on the head in a way that is similar to humans, resulting in similarly sized binocular visual fields (Heesy, 2009). Many other mammalian species feature a more lateralized position of the eyes and consequently deviate in their anatomy of binocular combination (Grieve, 2005; Howarth, Walmsley, & Brown, 2014; Jeffery, Cowey, & Kuypers, 1981; Kondo, Takada, Honda, & Mizuno, 1993; Longordo, To, Ikeda, & Stuart, 2013; Niell & Stryker, 2008; Scholl, Burge, & Priebe, 2013).

### 2.1 | Studies in cat LGN

**Only a very small number (2–11%) of cat LGN neurons increase or reduce their spiking rate through stimulation of either eye** (Bishop, Burke, & Davis, 1962; Erulkar & Fillenz, 1960; Kinston, Vadas, & Bishop, 1969) (Table 1). However, visual responses of most cat LGN neurons are significantly altered when a second visual stimulus is simultaneously presented to the nondominant eye, that is, their spike rate differs between binocular and monocular stimulation (Figure 3) (Guido, Tumosa, & Spear, 1989; Sengpiel, Blakemore, & Harad, 1995; Xue, Ramoa, Carney, & Freeman, 1987). **The retinal region of the nondominant eye that, when stimulated, elicits binocular modulation is called the nondominant eye receptive field.** More than three fourths of cat LGN cells (~82%) feature such a nondominant eye receptive field, and 88% of these neurons are suppressed by stimulation of the nondominant eye (Sanderson, Bishop, & Darian-Smith, 1971). Nondominant eye receptive fields are generally larger than dominant-eye receptive fields (Sanderson et al., 1971). And whereas dominant eye receptive fields in the LGN typically follow a center-surround organization, where stimulation of the center results in the opposite effect as stimulation of the surrounding region, **nondominant eye receptive fields feature a spatially homogeneous organization** (Sanderson et al., 1971 but see Schmielau & Singer, 1977).

Binocular modulation of cat LGN responses is greatest when the stimulus in the nondominant eye matches the spatial frequency of the



**FIGURE 2** Possible sites of binocular modulation in the primary visual pathway. 1) The outputs of the two eyes arrive in segregated eye-specific (green/blue) layers in the LGN, but some anatomical connections bridge them (top). 2) The projections of LGN neurons to primary visual cortex are also largely segregated by eye along the tangential dimension, terminating in eye-specific ocular dominance columns (green/blue). However, some of these LGN projections appear to form synapses outside their respective ocular dominance columns. 3) Projections from layer 4 neurons to other neurons within primary visual cortex are not bound to the boundaries of the ocular dominance columns. 4) Connections within the LGN or visual cortex as well as corticogeniculate feedback could provide a structural substrate for binocular modulation. Adopted from Blasdel and Lund (1983), Conley et al. (1985), Fitzpatrick, Lund, and Blasdel, (1985), and Katz, Gilbert, and Wiesel, (1989)

stimulus in the dominant eye (Sengpiel, Blakemore et al., 1995; Tong, Guido, Tumosa, Spear, & Heidenreich, 1992), though binocular modulation can be evoked across a wide range of spatial frequencies (Moore, Spear, Kim, & Xue, 1992; Sengpiel, Blakemore et al., 1995). While most studies used stimuli of relatively low spatial frequencies ( $< 0.8$  cycles per degree), binocular modulation has been observed for stimuli as high as five or more cycles per degree, suggesting that binocular modulation in cat LGN acts at high spatial acuity (Guido et al., 1989) (the peak contrast sensitivity for cats lies between 1 and 5 cycles per degree; Blake, Cool, & Crawford, 1974; Pasternak & Merigan, 1981).

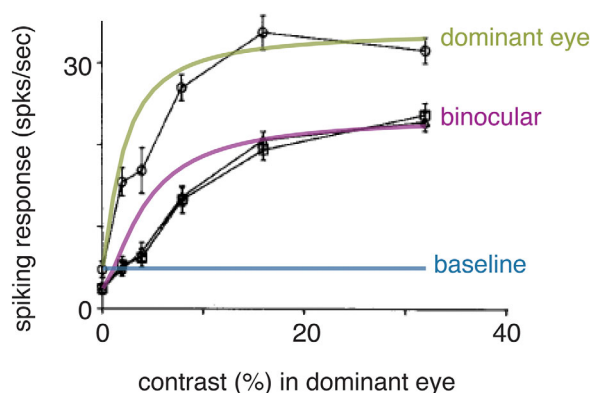
The spiking responses of most cat LGN neurons increase with increasing stimulus contrast. The resulting contrast response functions can be measured for stimuli presented to one eye alone (monocular stimulation) or for identical stimuli presented at the same position on the retina of both eyes (dioptic stimulation) (Tong et al., 1992). Assuming

binocular suppression, which dominates cat LGN, binocular modulation under these conditions could take one of three different forms:

1. Dioptic stimulation results in decreased responses relative to monocular stimulation at all contrast levels, that is, stimulating the

**TABLE 1** Summary estimates of binocular responses in cat and monkey LGN (see sections 2.1 and 2.2 for included studies).

	CAT LGN	MONKEY LGN
Exclusively monocular neurons	20%	70–90%
Interocular facilitated neurons	10%	5%
Interocular suppressed neurons	70%	10–30%
Binocular neurons	2–10%	3%



**FIGURE 3** Binocular modulation of monocular neurons. Data from a previously published example cat LGN neuron. Ordinate represents the magnitude of the neuron's spiking response to visual stimulation and abscissa plots the contrast of the visual stimulus (shown to the neuron's dominant eye). Model fits using a Naka-Rushton equation (Naka & Rushton, 1966) for binocular (purple line) and monocular (green line) responses are superimposed on the actual data (black traces). The solid blue line represents the estimated baseline firing rate of the neuron based on the activity plotted for the monocular condition at 0% contrast. Note the overall drop in response gain for the binocular stimulation condition, indicating that even though the LGN neuron can only be activated by one eye (the dominant eye), this neuron is nonetheless sensitive to stimulation of the other (nondominant) eye, resulting in an overall reduced visual response when both eyes are stimulated. No difference was found between a dioptic condition (triangles) and a dichoptic condition (squares) in which the spatial frequency in the two eyes were different. Adapted from Tong et al. (1992)

nondominant eye has the same suppressive effect for any stimulus contrast;

2. dioptic stimulation results in decreased responses relative to monocular stimulation for high contrasts only, that is, binocular suppression is limited to high contrasts;
3. dioptic stimulation results in a shift of the slope of the contrast response function relative to that for monocular stimulation, that is, the dynamic range of contrast responses is enhanced.

The majority of cat LGN cells exhibit the first type of binocular modulation (Tong et al., 1992). Therefore, binocular modulation in cat LGN appears to primarily reduce the neurons' response gain.

## 2.2 | Studies in monkey LGN

Similar to the cat, only a small minority (~3%) of neurons in monkey LGN can be driven through either eye (binocular responses have only been shown for ~30% of K neurons, which constitute ~10% of all LGN neurons) (Cheong, Tailby, Solomon, & Martin, 2013; Dacey, 1994; Zeater, Cheong, Solomon, Dreher, & Martin, 2015). As in cats, monkey LGN also contains neurons that modulate their spiking under binocular viewing (Marrocco & McClurkin, 1979; Rodieck & Dreher, 1979). And, similar to the cat, between 70% and 100% of this modulation in

monkeys takes the form of binocular suppression. One notable difference between the two species is that the fraction of LGN cells for which binocular modulation has been reported is drastically smaller in monkeys (<10–30%) compared to cats (Marrocco & McClurkin, 1979; Rodieck & Dreher, 1979) (Table 1). This difference in proportions of binocularly modulated LGN neurons might be due to differences in LGN anatomy between carnivores and primates, which we will discuss below. Another possibility is that the magnitude of binocular modulation depends on the type of visual stimulation. Specifically, investigations in cat LGN relied primarily on moving grating stimuli to isolate visual contrast responses. In contrast, almost all studies on binocular modulation in macaque monkey LGN used bars or light flashes that covered the entire visual field to evoke neural responses, which change global luminance in addition to local visual contrast (Marrocco & McClurkin, 1979; Rodieck & Dreher, 1979 but see Schroeder, Tenke, Arezzo, and Vaughan, (1990).

## 3 | PRIMATE SPECIALIZATIONS

The primate LGN is organized in parvocellular (P), magnocellular (M), and koniocellular (K) layers (Brunso-Bechtold & Casagrande, 1982; Norton, Casagrande, Irvin, Sesma, & Petry, 1988; Xu et al., 2001) (Figure 1). P cells, which make up the majority of LGN neurons, form the primary four dorsal layers of the LGN (Dreher, Fukada, & Rodieck, 1976). M cells, which constitute less than 20% of LGN neurons, are located in the two ventral-most primary layers (Hendry & Reid, 2000). K neurons, which constitute less than a tenth of LGN neurons, are almost exclusively located within the intercalated zones between the primary LGN layers (Casagrande, Yazar, Jones, & Ding, 2007; Hendry & Reid, 2000). M and P neurons can be reliably distinguished using neurophysiological criteria, including systematic differences in the transiency of their responses, selectivity to spatial and temporal frequency, color (cone) opponency, contrast response functions and recording locations within the LGN (Brunso-Bechtold & Casagrande, 1982; Norton et al., 1988; Xu et al., 2001). K neurons tend to differ in their spectral response from M and P neurons, and can be cytochemically distinguished via optogenetic targeting by their expression of CamKII, a protein kinase, which is absent in all other LGN neurons (Hendry & Yoshioka, 1994; Klein et al., 2016).

The distinction between P, M, and K neurons is particularly important for binocular vision because the physiology and anatomical connectivity of these cells differs distinctively, which might affect binocular modulation in these pathways. Several psychophysical studies have found that motion information, believed to be carried by the M pathway, is integrated across the eyes differently than other visual properties (Andrews & Blakemore, 2002; Carlson & He, 2000; Sun, Tong, Yang, Tian, & Hung, 2002). This hypothesis is corroborated by neurophysiological data (Tailby, Majaj, & Movshon, 2010), but how this finding relates to the responses of specific LGN subpopulations is unclear. Some researchers suggest that binocular modulation occurs with equal frequency in P and M layers (Marrocco & McClurkin, 1979; Schroeder et al., 1990), while others report that binocular modulation is

exclusive to the M layers (Rodieck & Dreher, 1979). Interestingly, anatomical studies in New World monkeys show that M neurons tend to be oriented orthogonally to the laminar boundaries and stay less confined to their home layer than P cells (Conley, Birecree, & Casagrande, 1985). This idiosyncratic morphology might constitute a unique mechanism to provide M cells with inputs from both eyes. No neurophysiological data to date speak to whether K neurons exhibit binocular modulation by altering their responses to the dominant eye when the nondominant is stimulated as well. Intriguingly, a small fraction (~10–30%) of K neurons respond to both eyes (Cheong et al., 2013; Zeater et al., 2015), suggesting that (some) K neurons might play a special role for binocular interactions. K neurons uniquely receive input from the superior colliculus (Stepniewska, Qi, & Kaas, 1999), which might provide them with exclusive binocular input. The superior colliculus receives inputs from both eyes as well as inputs from V1, and approximately 80% of superior colliculus neurons respond to stimuli shown to either eye in primates (Moors & Vendrik, 1979), with ~30% even responding equally strong to input from either eye (Marrocco & Li, 1977).

## 4 | SUBCORTICAL CIRCUITS SUPPORTING BINOCULAR MODULATION

### 4.1 | Potential role of intrageniculate cells

Neurons in the primary layers of the LGN receive input from one eye only (Guillery, 1970; Hayhow, 1958; Hickey & Guillery, 1974; Kaas, Guillery, & Allman, 1972; Laties & Sprague, 1966; Stone & Hansen, 1966). In order for binocular modulation to be generated locally in the LGN, the processes of any involved LGN neurons must extend across these laminar boundaries. In the cat, several types of intrageniculate cells could fulfill this criterion (Sanderson et al., 1971). First, large multipolar class I cells, located in interlaminar zones, receive binocular input from the optic tract, and also feature dendrites that extend beyond the cells' home layer (Hayhow, 1958; Laties & Sprague, 1966). Second, geniculocortical class II cells feature dendrites that extend into other layers (Guillery, 1966). Third, one type of LGN interneuron (subtype b) features axons that extend into other layers of the LGN (Tömböl, 1969). Lastly, another type of cell that is present in all major laminae has dendrites that cross into other layers and contacts other dendrites there (Famiglietti, 1970). Binocular modulation in cat LGN could arise through the activity of any of these four cell types, or a combination thereof.

Evidence of similar anatomical connections among LGN layers in primates is much sparser. However, some primate LGN neurons, particularly those close to primary laminar borders, have dendrites that extend across the border of origin into interlaminar spaces (K layers) and sometimes even into the adjacent primary layer (Campos Ortega, Glees, & Neuhoof, 1968; Saini & Garey, 1981). In addition, neurons within interlaminar zones feature dendrites that span into both neighboring laminae, suggesting potential for interactions between monocular neurons across laminar borders in primates (Guillery & Colonnier, 1970).

### 4.2 | Potential role of other subcortical structures for binocular modulation in the LGN

Both cats and primates feature a group of cell bodies superior to the LGN, known as the perigeniculate nucleus (PGN) and thalamic reticular nucleus (TRN), respectively, that receive inputs from both the LGN and early visual cortex (Ahlsén & Lindström, 1983; Ahlsén, Lindström, & Sybirska, 1978; Dubin & Cleland, 1977; Updyke, 1975). Some PGN cells respond to visual stimulation in either eye (Xue, Carney, Ramoa, & Freeman, 1988). These binocular responses in PGN are robust to ablation of areas 17, 18, and 19 (Xue et al., 1988), suggesting that PGN cells combine monocular LGN inputs locally. In turn, PGN cells inhibit LGN principal cells (Lindström, 1982). Given that LGN neurons generally reduce their responses under binocular viewing (Guido et al., 1989; Sengpiel, Blakemore et al. 1995; Xue et al., 1987), PGN/TRN neurons might play a role in subcortical binocular modulation (Funke & Eysel, 1998). Other possible subcortical routes for LGN binocular modulation involve the superior colliculus, as discussed above, as well as the pretectum or the parabigeminal nucleus (Casagrande & Boyd, 1996; Feig & Harting, 1994; Harting, Hashikawa, & Van Lieshout, 1986; Harting, Van Lieshout, Hashikawa, & Weber, 1991). Future research may discern the role of each of these pathways for binocular modulation.

## 5 | BINOCULAR MODULATION IN PRIMARY VISUAL CORTEX

An alternative to the hypothesis that the outputs from the two eyes first meet and interact in the LGN is that this meeting first occurs in primary visual cortex. LGN relay cells from M and P laminae primarily project onto granular layer 4 stellate cells in primary visual cortex (Figure 2). Layer 4 cells have been described as predominantly monocular (Hubel & Wiesel, 1968). Lesioning a geniculate layer corresponding to one eye results in patchy degeneration of cortical tissue in macaque V1 layer 4, suggesting spatial segregation of each eye's input (Hubel & Wiesel, 1972). Proline (dye) injections in one eye reveal alternating bands of ocular dominance in the granular layer, which further demonstrates that eye-specific LGN inputs to macaque V1 layer 4 are spatially distinct (Blasdel & Lund, 1983; Hubel & Wiesel, 1972). However, the spatial segregation of eye-specific inputs in V1 seems far less strict than in the LGN as some LGN afferents terminate in V1 ocular dominance bands corresponding to the other eye (Blasdel & Lund, 1983). Therefore, the signals from the two eyes might interact at the level of geniculocortical synapses in layer 4, that is, at the input stage to visual cortex. Another possibility is that binocular modulation in primary visual cortex is mediated via local interneurons. For example, inhibitory basket cells in layer 3 of cat visual cortex span ocular dominance bands (Buzás, Eysel, Adorján, & Kisvárdy, 2001). Basket cells also reside in layer 4, although it is unknown whether these neurons span ocular dominance as well (Martin, Somogyi, & Whitteridge, 1983). In any case, most neurons outside of layer 4 of primary visual cortex are driven through either eye, which suggests that the signals from the two eyes are merged when visual activation reaches these layers (Hubel & Wiesel, 1962).



## 5.1 | Single neuron studies in cat visual cortex

A large body of literature on binocular modulation in cat visual cortex has shown that binocular stimulation generally results in a reduction of activity compared to monocular stimulation, especially if the orientation of the stimuli in each eye are orthogonal (Sengpiel & Blakemore, 1994; Sengpiel & Vorobyov, 2005; Sengpiel, Baddeley, Freeman, Harrad, & Blakemore, 1998; Sengpiel, Blakemore et al., 1995; Sengpiel, Freeman, & Blakemore, 1995).

Several neurophysiological studies have probed the specific origins of binocular modulation in cat area A17 and A18. For example, Ohzawa and Freeman relied on the fact that, unlike LGN neurons (Xue et al., 1987), responses of visual cortical neurons vary with binocular disparity, i.e., a slight positional shift of the same image between the two eyes. In this study, both the relative phase (disparity) and contrast of grating stimuli shown to each eye were varied (Freeman & Ohzawa, 1990). Interestingly, the neurons' disparity tuning remained constant, even for large interocular contrast differences, such as 2.5% contrast in one eye and 50% contrast in the other eye. This result suggests that binocular modulation occurs before area 17 neurons produce action potentials since their spiking output has already been adjusted to the contrast of the stimulus shown to the other eye.

Truchard et al. (2000) developed a model for contrast encoding that incorporates monocular and binocular stages of visual processing. They also recorded spiking responses from binocular cells in area 17 in anesthetized cats while a grating of either high or low contrast was presented to one eye, and another grating of varying contrast and spatial phase was simultaneously shown to the other eye. They found that increasing the contrast of a grating in one eye results in a large reduction in monocular contrast gain, and that this reduction is largely independent of the contrast gain of the other eye (Truchard et al., 2000). In other words, their data suggest that, under binocular viewing, most contrast gain control occurs at the monocular level.

On the other hand, transfer of visual adaptation from one eye to the other (interocular transfer) has been pointed out as evidence that some interocular gain control occurs at the binocular level. Specifically, following several hundred milliseconds to several seconds of exposure to stimuli presented to one eye, responses to stimulation of the other eye are reduced for both binocular neurons (Hammond & Mouat, 1988; Maffei, Berardi, & Bisti, 1986) and monocular neurons alike (Howarth, Vorobyov, & Sengpiel, 2009). This finding suggests that monocular neurons are not only modulated by their counterparts that encode the other eye, but also by neurons that receive inputs from both eyes and therefore encode the binocular signal.

## 5.2 | Single neuron studies in macaque V1

There have been numerous studies on the effects of binocular stimulation in monkey primary visual cortex. The vast majority of them were limited to stimuli that did not match between the eyes (see Cumming & DeAngelis, 2001; Freeman, 2017; Henriksen, Tanabe, & Cumming, 2016; Leopold, Maier, Wilke, & Logothetis, 2005; Logothetis, 1998; Macknik & Martinez-Conde, 2007; Parker & Cumming, 2001; Parker,

Smith, & Krug, 2016, for review). Similar to the cat, most neurons in monkey V1 are driven through either eye (Hubel & Wiesel, 1968), and the predominant effect of binocular stimulation (outside very low contrast levels) is binocular suppression (Endo, Kaas, Jain, Smith, & Chino, 2000; Kumagami, Zhang, Smith, & Chino, 2000).

One study in monkey V1 specifically addressed the question of whether signals from the two eyes interact before reaching this area. In this experiment, a grating was presented at one contrast level to one eye, and a target of varying contrast was presented to the other eye. The authors found that when a lower contrast grating is shown to one eye, a higher contrast grating needs to be shown to the other eye to elicit a certain criterion response (Smith, Chino, Ni, & Cheng, 1997). This finding mirrors the results from the cat outlined above (Truchard et al., 2000): Given that dichoptic gratings of varying contrast can elicit the same neuronal response, binocular modulation seems to occur prior to the spiking of V1 neurons. This conclusion allows for the possibility that the site of initial binocular modulation lies either within the LGN or at the input level to V1 (such as in layer 4C), prior to spiking of most V1 neurons.

## 6 | CORTICAL FEEDBACK TO THE LGN AND ITS POTENTIAL ROLE FOR BINOCULAR MODULATION

In both cats and primates, some cells in primary visual cortex project back to the LGN (Figure 2). In cats, layer 6 neurons in areas 17, 18, and 19 project to the LGN (Gilbert & Kelly, 1975). In monkeys, ~15% of V1 layer 6 neurons project to the LGN (Fitzpatrick, Usrey, Schofield, & Einstein, 1994). To a much smaller extent, neurons in extrastriate areas, such as area V2, project to the LGN as well (Briggs, Kiley, Callaway, & Usrey, 2016). This corticogeniculate projection is retinotopically aligned with the retinogeniculate input (Ichida, Mavity-Hudson, & Casagrande, 2014; Updyke, 1975). But in contrast to retinal inputs that mostly terminate proximally on LGN neurons and activate ionotropic receptors, corticogeniculate inputs to LGN tend to terminate distally and act on metabotropic receptors, effectively serving a modulatory role (see Sherman, 2007). Given the specificity of corticogeniculate projections and their termination pattern, it is conceivable that binocular modulation in the LGN is not computed locally, but fed back from primary visual cortex. As discussed above, corticogeniculate cells also synapse on cells in the PGN/TRN, which in turn project to and inhibit cells in the LGN (Ahlsén & Lindström, 1983; Dubin & Cleland, 1977). Therefore, corticogeniculate feedback could also act through the PGN/TRN to mediate binocular modulation in the LGN.

To investigate the potential role of cortical feedback on binocular modulation in the LGN, several researchers studied whether binocular modulation is sensitive to stimulus orientation (Moore et al., 1992; Sengpiel et al., 1995; Varela & Singer, 1987). The rationale behind these studies is that neuronal orientation selectivity (orientation tuning) is of cortical origin (Scholl, Tan, Corey, & Priebe, 2013). Therefore, orientation dependence of binocular modulation in the LGN seems indicative of involvement of corticogeniculate feedback. Only one of these studies found an orientation bias of binocular inhibition of LGN

neurons (Sengpiel et al., 1995; Varela & Singer, 1987), leaving the question of feedback-mediated mechanisms unresolved. Notably, cortex could also be involved in LGN binocular modulation in an orientation-independent manner if this modulation arises by a population of cortico-geniculate neurons with varying orientation tuning or by cortical neurons without orientation tuning (Moore et al., 1992). Furthermore, a small, but measurable orientation bias has been found among retinal ganglion cells (Kuffler, 1953; Levick & Thibos, 1982) as well as geniculate neurons of both cats (Creutzfeldt & Nothdurft, 1978; Daniels, Norman, & Pettigrew, 1977; Shou, Ruan, & Zhou, 1986; Soodak, Shapley, & Kaplan, 1987; Vidyasagar & Urbas, 1982) and monkeys (Lee, Virsu, & Creutzfeldt, 1977; Smith, Chino, Ridder, Kitagawa, & Langston, 1990; Xu, Ichida, Shostak, Bonds, & Casagrande, 2002). **This orientation bias of LGN neurons survives inactivation of cortical feedback** (Thompson, Leventhal, Zhou, & Liu, 1994; Vidyasagar & Urbas, 1982). Therefore, (modest) orientation-dependent binocular modulation may not be sufficient to exclusively implicate cortical feedback for binocular modulation (Shou & Leventhal, 1989; Xu et al., 2002).

In cats, the role of the cortical feedback for binocular modulation has also been investigated during reversible or permanent inactivation of visual cortex. One of these studies found that removal of areas 17, 18, and 19 had little-to-no impact on LGN cells that exhibit binocular modulation (Sanderson et al., 1971). In line with this result, two related studies found that LGN binocular modulation remained unchanged after cooling areas 17, 18, and 19 (Pape & Eysel, 1986; Singer, 1970). However, other groups found results that stand in direct contradiction to the above. In one of these studies, areas 17 and 18 were cooled to inactivate neural activity in the deep layers that project to the LGN, and binocular modulation in the LGN was greatly diminished in the LGN under this condition (Schmielau & Singer, 1977). Another study found that removing areas 17, 18, and 19 permanently eliminated binocular modulation in the LGN (Varela & Singer, 1987). Therefore, based on the existing data, we cannot conclude how involved cortical feedback is to LGN binocular modulation.

## 6.1 | Binocular modulation and binocular rivalry

When the two eyes are simultaneously stimulated using two very different (nonfusible) stimuli at the same retinal location, perception can alternate stochastically between each eye's view (Alais & Blake, 2005; Blake & Logothetis, 2002; Leopold et al., 2005; Maier, Panagiotaropoulos, Tsuchiya, & Keliris, 2012; Tong, Meng, & Blake, 2006). For example, when one eye views a face while the other eye is looking at a house that is presented at the same retinal location, perception dynamically switches between the face and the house. In other words, although retinal stimulation remains constant, perception fluctuates under these conditions. This dynamic perceptual phenomenon termed **binocular rivalry** seems related to binocular modulation in that it involves stimulation of the two eyes as well as suppression of one eye's view at the expense of the other. However, there are several important differences between these two phenomena. First, binocular rivalry only occurs when the stimuli in the two eyes cannot be fused, whereas the kind of binocular suppression discussed here occurs even

when the stimuli are perceptually fused. Second, binocular rivalry suppresses only one eye's view, whereas the kind of binocular modulation discussed here affects the responses to both eyes. Third, binocular rivalry fluctuates over time, whereas the binocular modulation discussed here does not co-vary with these perceptual alternations. The relationship between binocular modulation and binocular rivalry remains unclear.

One benefit of binocular rivalry is that it can be utilized by experimenters to track if certain groups of visual neurons alter their activity whenever perception changes between the two simultaneously presented stimuli. Two independent studies with human volunteers found that fMRI signals in LGN strongly correlate with perceptual report (Haynes, Deichmann, & Rees, 2005; Wunderlich, Schneider, & Kastner, 2005). In contrast, neurophysiological studies in trained macaques did not show any population firing rate changes in LGN during similar stimulation conditions (Lehky & Maunsell, 1996; Wilke, Mueller, & Leopold, 2009). The involvement of LGN in binocular rivalry therefore remains unclear. An empirically supported explanation for the apparent discrepancies is that the fMRI signal is more sensitive to metabolically costly synaptic modulation than spiking output (Logothetis, Pauls, Augath, Trinath, & Oeltermann, 2001). This difference in measurement might explain why spiking activity and fMRI signals diverge when feedback modulation is too weak to elicit large changes in population spiking (Boynton, 2011; Maier et al., 2008; Schmid & Maier, 2015).

## 7 | DISCUSSION

The LGN is often conceptualized as a "monocular" structure. This classification is well-supported by anatomy and physiology that show that almost all LGN neurons receive input and are driven through only one eye (Bishop, Kozak, Levick, & Vakkur, 1962; Cleland, Dubin, & Levick, 1971; Guillery, 1970; Guillery & Kaas, 1971; Hayhow, 1958; Hickey & Guillery, 1974; Hubel & Wiesel, 1961; Kaas et al., 1972; Laties & Sprague, 1966; Stone & Hansen, 1966). **Yet, as described in this review, several LGN circuits, both local and extra-geniculate, could provide a substrate for binocular modulation of LGN neurons. Indeed, most single neuron studies in both cats and primates have found evidence for binocular modulation in the LGN.**

Convergent observations of binocular modulation in the LGN of cats and primates suggest a homologous mechanism for processing binocular inputs. Could this geniculate processing serve to adjust the gain of responses under binocular viewing? The input to the visual system is greater under binocular than monocular viewing, yet the appearance of the visual world remains largely the same under these viewing conditions. This phenomenon suggests that the brain accounts for the difference in inputs between monocular and binocular viewing, likely through a gain control mechanism. Intriguingly, in both cats and monkeys, most binocular modulation in the LGN is suppressive. Furthermore, where **contrast-dependent modulation was measured, the majority of binocular modulation suggested an adjustment in responses at all contrast levels** (Tong et al., 1992).

While both cats and monkeys show (mostly suppressive) binocular modulation in the LGN, the data also reveal several differences between these two model species. **Most strikingly, while the majority of LGN neurons in the cat modulate under binocular viewing, this binocular modulation is limited to a minority of macaque neurons.** Some of these interspecies differences in neuronal response properties might be explained by species-specific anatomical adaptations. Several researchers have drawn homologies between cat X, Y, and W cells and primate P, M, and K cells, respectively, based on similarities of their response properties (Dreher et al., 1976; Irvin, Norton, Sesma, & Casagrande, 1986; Norton & Casagrande, 1982; Schiller & Malpeli, 1978; Sherman, Wilson, Kaas, & Webb, 1976; but see Kaplan & Shapley, 1982). In cats, the superior colliculus projects to the C layers of cat LGN where W cells reside (Torrealba, Partlow, & Guillery, 1981). Similarly, in primates, the superior colliculus projects to K neurons (Stepniewska et al., 1999). Given this homology, it is possible that this tectothalamic projection serves similar roles for binocular processing for cat W and primate K neurons. However, several other observations suggest that binocular processing differs significantly between cats and primates. Chief among them is the difference in anatomical layout of the cat LGN and primate LGN. Although homologies have been drawn between X, Y, W and P, M, and K neurons, their distribution in the LGN in each species is different. X cells terminate in layers A and A1 of cat LGN, Y cells terminate in layer A and C, and W cells terminate in C layers only (Stone & Fukuda, 1974; Sur & Sherman, 1982a, b; Wilson, Rowe, & Stone, 1976). In contrast, primate P, M, and K neurons are segregated in separate layers. Second, neurons in cat LGN that cross eye-specific layers appear to be more frequent in cats than in monkeys (Guillery, 1966; Levitt, Schumer, Sherman, Spear, & Movshon, 2001; Movshon et al., 1987; Sanderson et al., 1971). Lastly, whereas cat LGN projects to three visual areas (A17, A18, and A19), primate LGN predominantly projects to the primary visual cortex (V1). Cat LGN also receives feedback from three cortical areas (A17–19), whereas primate LGN receives its cortical feedback almost exclusively from V1 (but see Briggs et al., 2016; Hendrickson, Wilson, & Ogren, 1978). Given these differences in organization, the LGN circuitry for binocular combination in cats and primates may differ significantly, which may translate directly into functional differences that have been observed across neurophysiological studies (Conway & Schiller, 1983).

Another consideration regarding the extant literature on binocular modulation is that most of the work from cat and primate LGN were performed using anesthetized preparations that relied on GABAergic action. LGN neurons are highly sensitive to altered states of arousal such as wakefulness, drowsiness, sleep, and general anesthesia (Alitto, Moore, Rathbun, & Usrey, 2011). In addition, response normalization, which is one potential mechanism by which the gain of monocular signals might be adjusted under binocular viewing, is significantly altered by such anesthetic preparations (Vaiceliunaite, Eriskien, Franzen, Katzner, & Busse, 2013). In line with this concern, direct comparison between anesthetized and awake recordings in cat LGN found that the anesthetized state produced significantly different results than the awake state (Garrahy, Salinger, MacAvoy, Schroeder, & Guido, 1982; Schroeder, Salinger, & Guido, 1988).

All findings reviewed in this paper were collected in animals with normal vision, but, as mentioned in the introduction, binocular modulation in the LGN is of great relevance for abnormal vision as well. In particular, strabismic amblyopia, a disorder of binocular vision that affects children with misaligned eyes, and causes lifelong perceptual suppression of one eye if not treated before they reach adolescence, causes “shrinkage” of LGN neurons (primarily of the affected eye) in cats, macaques and humans (Barnes et al., 2010; Cheng et al., 2008; von Noorden & Crawford, 1992; von Noorden & Middleditch, 1975; Tremain & Ikeda, 1982). However, whether and how this structural malformation affects binocular modulation among LGN neurons remains unknown as existing studies in the LGN of strabismic cats were generally limited to testing the acuity of LGN neurons related to the perceptually suppressed eye (Chino et al., 1994; Gillard-Crewther & Crewther, 1988).

An impressive amount of work on the topic of binocular modulation in the LGN has laid a strong foundation to fully elucidate the role of this structure in binocular processing. These advances will be critical to determining the origins of binocular modulation and its possible basis for interocular gain control and binocular contrast normalization (Murphy & Sillito, 1989; Pape & Eysel, 1986; Sanderson et al., 1971; Schmielau & Singer, 1977; Singer, 1970; Varela & Singer, 1987). The apparent paradox that outputs from both eyes are brought together in the LGN in retinotopic register while keeping each eye's output segregated may explain binocular modulation within this structure through mechanisms yet to be uncovered.

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## IN MEMORIAM OF VIVIEN A. CASAGRANDE

Among her many scientific breakthroughs, Vivien made fundamental contributions to the neuronal basis of binocular combination in LGN and V1 across different primate species. Much of her work laid the foundation for the research we discuss in this review. In addition to her shining legacy as a scientist, Vivien was a deeply admired colleague and an inspiring mentor. She is sorely missed.

## ORCID

Kacie Dougherty  <http://orcid.org/0000-0001-6295-0026>

Michael Schmid  <http://orcid.org/0000-0003-1424-130X>

Alexander Maier  <http://orcid.org/0000-0002-7250-502X>

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