ECES T580 Lab 1

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Part 1.1.1 Download sequence with accession number and load into MATLAB

Retrieve sequence, save to file

```
clc;close all; clear;
```

Check if file already exists, if it doesn't. Download file from genbank

```
if exist('nm_000520.txt') == 2
    % Bring file into matlab
    s = genbankread('nm_000520.txt');
else
    % Grab file and then bring it into matlab
    getgenbank('nm_000520', 'toFile', 'nm_000520.txt');
    s = genbankread('nm_000520.txt');
end
```

Extract sequence

```
seq = getgenbank('nm_000520', 'SequenceOnly', true);
```

Part 1.2.1 Getting basic statistics

Format long sequence output

```
seqdisp(s.Sequence)

ans =
   46×71 char array
```

```
' 1 TCACATCACA ACGACTTGTG GTTTTAATCC TCCGTTTTTC TGCTTCTGAA GTTACTTCAG'
' 61 CCTGGCAAGT CCTTTACCTC CCCGTAGGCC TGGCGAGCTG CATCACAACA TTCAAGATTC'
' 121 ACCCTAGAGC CATCTGGGAA ACTTTCTTCT CCAGGTCGCC CTGCGTCCTC GCCTCCCCAC'
' 181 CCCGTTCTTC TCGAGTCGGG TGAGCTGTCT AGTTCCATCA CGGCCGGCAC GGCCGCAGGGG'
' 241 GTGGCCGGT ATTTACTGCT CTACTGGGCC CGTGAACAGT CTGGCGAGCC GAGCAGTTGC'
' 301 CGACGCCCGG CACAATCCGC TGCACGTAGC AGGAGCCTCA GGTCCAGGCC GGAAGTGAAA'
' 361 GGGCAGGGTG TGGGTCCTCC TGGGGTCGCA GGCCCAGAGC CGCCTCTGGT CACGTGATTC'
```

```
' 421 GCCGATAAGT CACGGGGGCG CCGCTCACCT GACCAGGGTC TCACGTGGCC AGCCCCCTCC'
' 481 GAGAGGGAG ACCAGCGGGC CATGACAAGC TCCAGGCTTT GGTTTTCGCT GCTGCTGGCG'
      GCAGCGTTCG CAGGACGGCC GACGGCCCTC TGGCCCTGGC CTCAGAACTT CCAAACCTCC'
      GACCAGCGCT ACGTCCTTTA CCCGAACAAC TTTCAATTCC AGTACGATGT CAGCTCGGCC'
 661
      GCGCAGCCCG GCTGCTCAGT CCTCGACGAG GCCTTCCAGC GCTATCGTGA CCTGCTTTTC'
721
      GGTTCCGGGT CTTGGCCCCG TCCTTACCTC ACAGGGAAAC GGCATACACT GGAGAAGAAT'
      GTGTTGGTTG TCTCTGTAGT CACACCTGGA TGTAACCAGC TTCCTACTTT GGAGTCAGTG'
' 841 GAGAATTATA CCCTGACCAT AAATGATGAC CAGTGTTTAC TCCTCTCTGA GACTGTCTGG'
901
      GGAGCTCTCC GAGGTCTGGA GACTTTTAGC CAGCTTGTTT GGAAATCTGC TGAGGGCACA'
      TTCTTTATCA ACAAGACTGA GATTGAGGAC TTTCCCCGCT TTCCTCACCG GGGCTTGCTG'
1021
      TTGGATACAT CTCGCCATTA CCTGCCACTC TCTAGCATCC TGGACACTCT GGATGTCATG'
'1081 GCGTACAATA AATTGAACGT GTTCCACTGG CATCTGGTAG ATGATCCTTC CTTCCCATAT'
'1141 GAGAGCTTCA CTTTTCCAGA GCTCATGAGA AAGGGGTCCT ACAACCCTGT CACCCACATC'
'1201 TACACAGCAC AGGATGTGAA GGAGGTCATT GAATACGCAC GGCTCCGGGG TATCCGTGTG'
'1261 CTTGCAGAGT TTGACACTCC TGGCCACACT TTGTCCTGGG GACCAGGTAT CCCTGGATTA'
     CTGACTCCTT GCTACTCTGG GTCTGAGCCC TCTGGCACCT TTGGACCAGT GAATCCCAGT'
'1381 CTCAATAATA CCTATGAGTT CATGAGCACA TTCTTCTTAG AAGTCAGCTC TGTCTTCCCA'
'1441 GATTTTTATC TTCATCTTGG AGGAGATGAG GTTGATTTCA CCTGCTGGAA GTCCAACCCA'
'1501 GAGATCCAGG ACTTTATGAG GAAGAAAGGC TTCGGTGAGG ACTTCAAGCA GCTGGAGTCC'
1621
     CAGGAGGTGT TTGATAATAA AGTAAAGATT CAGCCAGACA CAATCATACA GGTGTGGCGA'
'1681 GAGGATATTC CAGTGAACTA TATGAAGGAG CTGGAACTGG TCACCAAGGC CGGCTTCCGG'
'1741 GCCCTTCTCT CTGCCCCCTG GTACCTGAAC CGTATATCCT ATGGCCCTGA CTGGAAGGAT'
'1801 TTCTACATAG TGGAACCCCT GGCATTTGAA GGTACCCCTG AGCAGAAGGC TCTGGTGATT'
'1861 GGTGGAGAGG CTTGTATGTG GGGAGAATAT GTGGACAACA CAAACCTGGT CCCCAGGCTC'
'1921 TGGCCCAGAG CAGGGGCTGT TGCCGAAAGG CTGTGGAGCA ACAAGTTGAC ATCTGACCTG'
'1981 ACATTTGCCT ATGAACGTTT GTCACACTTC CGCTGTGAAT TGCTGAGGCG AGGTGTCCAG'
'2041 GCCCAACCCC TCAATGTAGG CTTCTGTGAG CAGGAGTTTG AACAGACCTG AGCCCCAGGC'
'2101 ACCGAGGAGG GTGCTGGCTG TAGGTGAATG GTAGTGGAGC CAGGCTTCCA CTGCATCCTG'
'2161 GCCAGGGGAC GGAGCCCCTT GCCTTCGTGC CCCTTGCCTG CGTGCCCCTG TGCTTGGAGA'
'2221 GAAAGGGGCC GGTGCTGGCG CTCGCATTCA ATAAAGAGTA ATGTGGCATT TTTCTATAAT'
'2281 AAACATGGAT TACCTGTGTT TAAAAAAAAA AGTGTGAATG GCGTTAGGGT AAGGGCACAG'
'2341
      CCAGGCTGGA GTCAGTGTCT GCCCCTGAGG TCTTTTAAGT TGAGGGCTGG GAATGAAACC'
'2401 TATAGCCTTT GTGCTGTTCT GCCTTGCCTG TGAGCTATGT CACTCCCCTC CCACTCCTGA'
'2461 CCATATTCCA GACACCTGCC CTAATCCTCA GCCTGCTCAC TTCACTTCTG CATTATATCT'
'2521 CCAAGGCGTT GGTATATGGA AAAAGATGTA GGGGCTTGGA GGTGTTCTGG ACAGTGGGGA'
'2581 GGGCTCCAGA CCCAACCTGG TCACAGAAGA GCCTCTCCCC CATGCATACT CATCCACCTC'
'2641 CCTCCCCTAG AGCTATTCTC CTTTGGGTTT CTTGCTGCTT CAATTTTATA CAACCATTAT'
     TTAAATATTA TTAAACACAT ATTGTTCTCT AGGAAAAAAA AAAAAAAAA A
'2701
```

Count nucleotides in sequence

```
[seq_counts] = basecount(s.Sequence)

seq_counts =
  struct with fields:

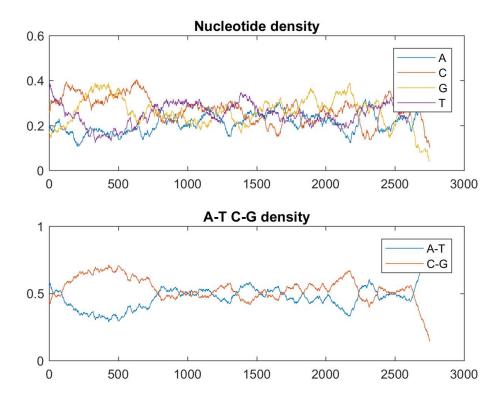
A: 593
  C: 750
  G: 716
```

Plot density of nucleotides along sequence

T: 692

```
seq_density_n20 = ntdensity(s.Sequence)
```

```
seq_density_n20 =
  struct with fields:
    A: [1×2751 double]
    C: [1×2751 double]
    G: [1×2751 double]
    T: [1×2751 double]
```



Count dimers in the nucleotide sequence

```
figure(2)
[Dimers, Percent] = dimercount(s.Sequence, 'chart', 'pie')
```

```
Dimers =

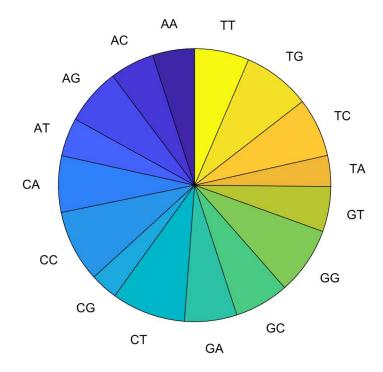
struct with fields:

AA: 137
AC: 145
AG: 185
AT: 125
CA: 184
CC: 235
CG: 90
CT: 241
GA: 171
GC: 178
```

GG: 220 GT: 147 TA: 101 TC: 192 TG: 221 TT: 178

Percent =

0.0498	0.0527	0.0673	0.0455
0.0669	0.0855	0.0327	0.0876
0.0622	0.0647	0.0800	0.0535
0.0367	0.0698	0.0804	0.0647



Count 3-mer in nucleotide sequence

```
trimer = nmercount(s.Sequence, 3)
```

```
trimer =
  64×2 cell array
    'ctg'
              [95]
    'cct'
              [81]
    'tgg'
              [77]
    'gag'
              [68]
    'ggc'
              [68]
    'ccc'
              [66]
    'tcc'
              [66]
    'agg'
              [65]
    'cag'
              [65]
```

```
'tct'
         [63]
'gct'
         [62]
'ctc'
         [61]
'ctt'
         [61]
'gcc'
         [59]
'ttc'
         [58]
'gga'
         [55]
'tga'
         [55]
'cca'
         [54]
'ggg'
         [53]
'gtg'
         [51]
'ttt'
         [51]
'cac'
        [50]
'tgt'
         [48]
'aaa'
         [46]
'aca'
         [46]
'tca'
         [46]
'agc'
         [45]
'acc'
         [44]
'ggt'
         [44]
'gtc'
         [43]
'aga'
         [42]
'gaa'
        [42]
'ttg'
        [42]
'tgc'
        [41]
'cat'
         [39]
'gca'
         [38]
'gac'
         [37]
'act'
         [35]
'tat'
         [35]
'aag'
         [34]
'att'
         [34]
'ccg'
         [34]
'agt'
         [33]
'atg'
        [33]
'gtt'
        [32]
'caa'
       [30]
'aac'
         [29]
'ata'
         [29]
'atc'
        [29]
'tac'
         [29]
'aat'
         [27]
'tta'
         [27]
'cgg'
         [25]
'cgc'
         [24]
         [24]
'cta'
'gat'
        [24]
'cgt'
         [22]
'gta'
         [21]
'acg'
         [20]
'cga'
         [19]
'gcg'
         [19]
'taa'
         [19]
'tag'
         [18]
```

'tcg'

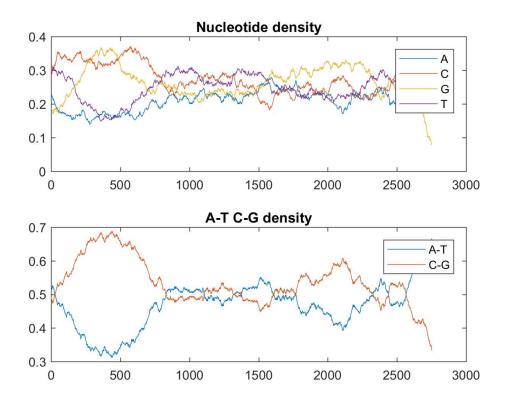
[17]

Try different window sizes for ntdensity function Default is length(seq)/20 Lets try doing length(seq)/10, using the round() function since the window length must be an integer

```
figure(3)
seq_density_n10 = ntdensity(s.Sequence, 'WINDOW', round(length(s.Sequence)/10))
```

```
seq_density_n10 =
  struct with fields:
    A: [1×2751 double]
    C: [1×2751 double]
    G: [1×2751 double]
```

T: [1×2751 double]

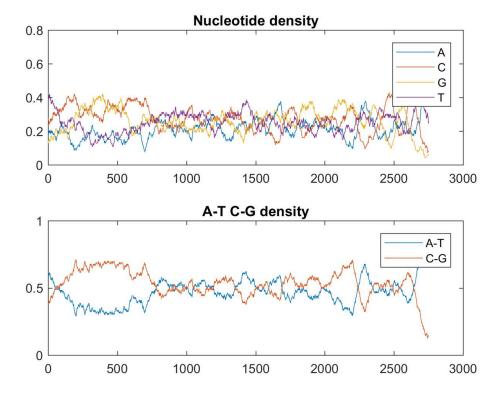


And we can try length(seq)/30

```
figure(4)
seq_density_n30 = ntdensity(s.Sequence, 'WINDOW', round(length(s.Sequence)/30))
```

```
seq_density_n30 =
struct with fields:

A: [1×2751 double]
C: [1×2751 double]
G: [1×2751 double]
T: [1×2751 double]
```



The window sizes for each of these three values are as follows: length(seq)/10 = 275 length(seq)/20 = 137 length(seq)/30 = 91 The advance of having the larger window size is that the image is more more clear to see (A bit zoomed in). However larger window sizes require you to pan to look at specfic points of interest. Smaller window sizes allow for one to see large amounts of data within one space

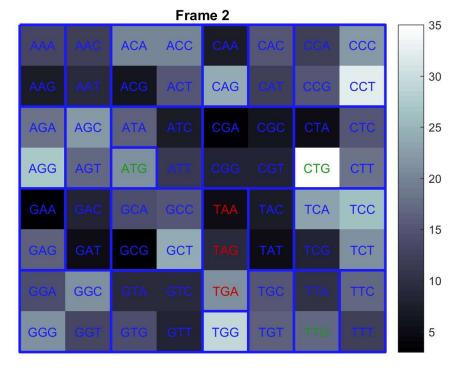
Part 1.3.1: Condon Count and ORF

Return codon counts for each of the six reading frames Plot the results in a heat map

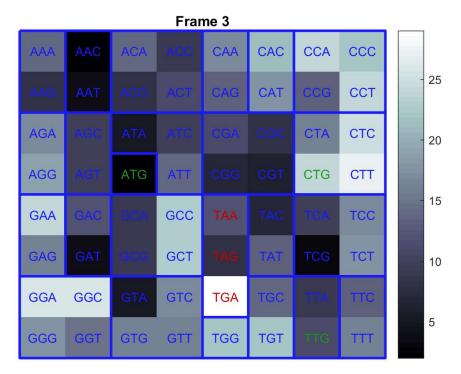
```
figure(5)
rlcodons_1 = codoncount(s.Sequence, 'frame', 1, 'figure', true);
title('Frame 1')
figure(6)
rlcodons_2 = codoncount(s.Sequence, 'frame', 2, 'figure', true);
title('Frame 2')
figure(7)
rlcodons_3 = codoncount(s.Sequence, 'frame', 3, 'figure', true);
title('Frame 3')
```

Frame 1 35 CCA CCC 30 AAG CAG CCT CTC 25 AGG CTG 20 GCC TCC 15 GAG TCT 10 GGC TTC 5 GTG TGG TTT

Genetic Code: Standard



Genetic Code: Standard



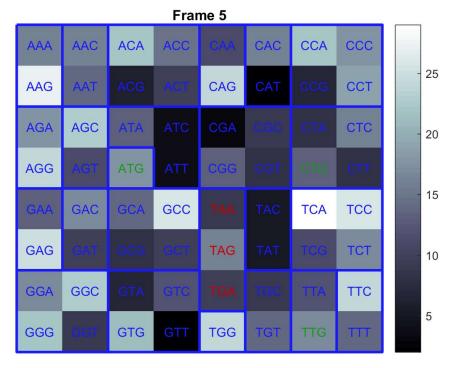
Genetic Code: Standard

To get the other 3 frames we take the reverse complement

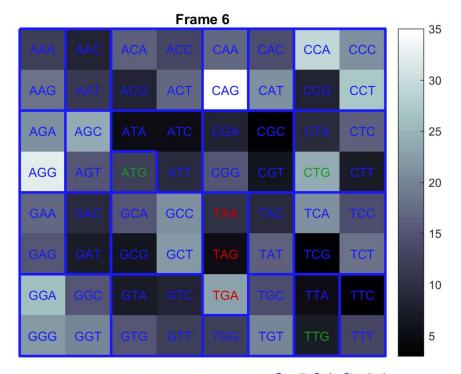
```
figure(8)
rlcodons_4 = codoncount(s.Sequence, 'Reverse',true,'frame', 1, 'figure', true);
title('Frame 4')
figure(9)
rlcodons_5 = codoncount(s.Sequence, 'Reverse',true,'frame', 2, 'figure', true);
title('Frame 5')
figure(10)
rlcodons_6 = codoncount(s.Sequence, 'Reverse',true,'frame', 3, 'figure', true);
title('Frame 6')
```

Frame 4 35 AAA CCA CAC 30 CCT CAG CTC **AGA** 25 AGG CTG CTT 20 GCC GAA 15 GAG 10 **GGA** GGC 5 GGG TGG TTT

Genetic Code: Standard



Genetic Code: Standard

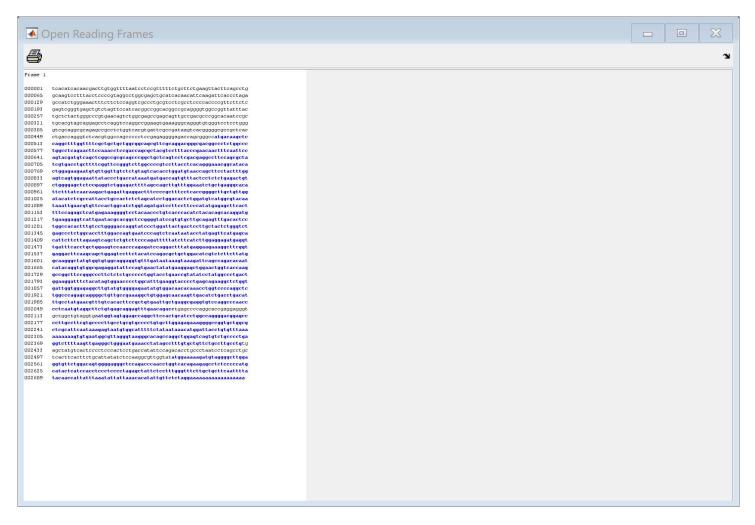


Genetic Code: Standard

Part 1.3.1: Find ORFs based on length

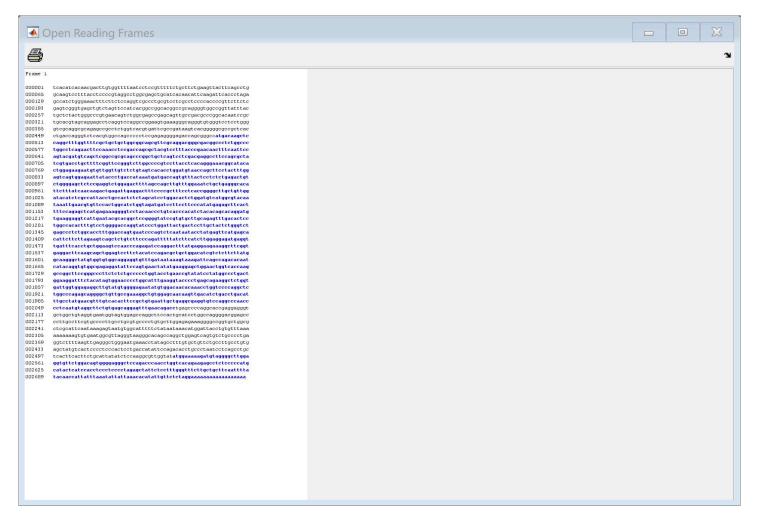
Find ORFS of length > 50 in Frame 1 of the given sequence

```
orf_min_50 = seqshoworfs(s.Sequence, 'Frames', 1, 'MinimumLength', 50);
```



Find ORFS of length > 500 in Frame 1 of the given sequence

```
orf_min_500 = seqshoworfs(s.Sequence, 'Frames', 1, 'MinimumLength', 500);
```



Part 1.3.2 Finding optimum threshold for ORFs

we can find the optimum length of ORF

Get total number of stop codons within each frame

```
total1 = rlcodons_1.TGA + rlcodons_1.TAG + rlcodons_1.TAA;
total2 = rlcodons_2.TGA + rlcodons_2.TAG + rlcodons_2.TAA;
total3 = rlcodons_3.TGA + rlcodons_3.TAG + rlcodons_3.TAA;
total4 = rlcodons_4.TGA + rlcodons_4.TAG + rlcodons_4.TAA;
total5 = rlcodons_5.TGA + rlcodons_5.TAG + rlcodons_5.TAA;
total6 = rlcodons_6.TGA + rlcodons_6.TAG + rlcodons_6.TAA;
total5 = total1 + total2 + total3 + total4 + total5 + total6
```

```
totalStop =
```

Get total number of codons in the sequence

```
c1 = struct2cel1(rlcodons_1);
c2 = struct2cel1(rlcodons_2);
c3 = struct2cel1(rlcodons_3);
c4 = struct2cel1(rlcodons_4);
c5 = struct2cel1(rlcodons_5);
```

```
c6 = struct2cell(rlcodons_6);
TotalCodons1 = CalculateTotal(c1, length(c1));
TotalCodons2 = CalculateTotal(c2, length(c2));
TotalCodons3 = CalculateTotal(c3, length(c3));
TotalCodons4 = CalculateTotal(c4, length(c4));
TotalCodons5 = CalculateTotal(c5, length(c5));
TotalCodons6 = CalculateTotal(c6, length(c6));
TotalCodons = TotalCodons1 + TotalCodons2 + TotalCodons3 + TotalCodons4 + TotalCodons5 + TotalCodons6
```

Find optimal length

```
k =

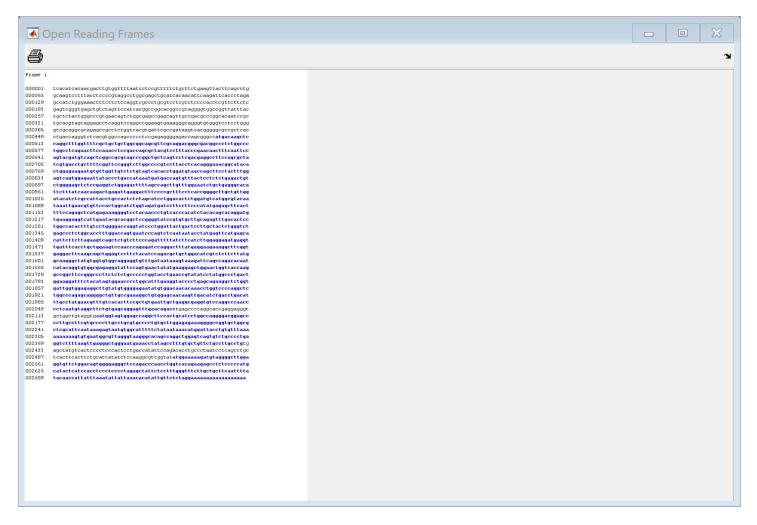
85.6391

k =

87.6391
```

Find ORFs with the given estimated numberorf_min_50 = seqshoworfs(s.Sequence, 'Frames', 1, 'MinimumLength', 50);

```
orf_estimated = seqshoworfs(s.Sequence, 'Frames', 1, 'MinimumLength', k);
```



General Functions

Quick function to calculate total within frame cell array

```
function total = CalculateTotal(array, length)
total = 0;
for n=1:length
   total = total + array{n};
end
end
```

```
TotalCodons = 5498
```

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