RNAlib-2.4.14

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1 Main Page	1
1.1 A Library for predicting and comparing RNA secondary structures	1
1.2 License	1
1.3 Contributors	2
2 Getting Started	3
2.1 Installation and Configuration	3
2.1.1 Installing the ViennaRNA Package	3
2.1.1.1 Quick-start	3
2.1.1.2 Installation without root privileges	3
2.1.1.3 Notes for MacOS X users	4
2.1.2 Configuring RNAlib features	4
2.1.2.1 Streaming SIMD Extension (SSE) support	4
2.1.2.2 Scripting Interfaces	4
2.1.2.3 Cluster Analysis	5
2.1.2.4 Kinfold	5
2.1.2.5 RNAforester	5
2.1.2.6 Kinwalker	5
2.1.2.7 Link Time Optimization (LTO)	5
2.1.2.8 OpenMP support	6
2.1.2.9 POSIX threads (pthread) support	6
2.1.2.10 Stochastic backtracking using Boustrophedon scheme	6
2.1.2.11 SVM Z-score filter in RNALfold	6
2.1.2.12 GNU Scientific Library	6
2.1.2.13 Disable C11/C++11 feature support	7
2.1.2.14 Enable warnings for use of deprecated symbols	7
2.1.2.15 Single precision partition function	7
2.1.2.16 Help	7
2.1.3 Linking against RNAlib	7
2.1.3.1 Compiler and Linker flags	8
2.1.3.2 The pkg-config tool	9
2.2 HelloWorld	9
2.3 HelloWorld (Perl/Python)	11
2.3.1 Perl5	11
2.3.2 Python	12
3 Concepts and Algorithms	13
3.1 RNA Structure	14
3.1.1 RNA Structures	14
3.1.2 Levels of Structure Abstraction	14
3.1.2.1 Primary Structure	14
3.1.2.2 Secondary Structure	14
3.1.2.3 Tertiary Structure	14
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3.1.2.4 Quarternary Structure	14
3.1.2.5 Pseudo-Knots	14
3.2 Distance Measures	14
3.2.1 Functions for Tree Edit Distances	15
3.2.2 Functions for String Alignment	16
3.2.3 Functions for Comparison of Base Pair Probabilities	16
3.3 Free Energy of Secondary Structures	16
3.3.1 Secondary Structure Loop Decomposition	17
3.3.1.1 Free Energy Evaluation API	18
3.3.2 Free Energy Parameters	18
3.3.2.1 Free Energy Parameters Modification API	18
3.3.3 Fine-tuning of the Energy Evaluation Model	18
3.4 Secondary Structure Folding Grammar	18
3.4.1 Secondary Structure Folding Recurrences	19
3.4.2 Additional Structural Domains	19
3.4.2.1 Structured Domains	20
3.4.2.2 Unstructured Domains	20
3.4.2.3 Domain Extension API	21
3.4.3 Constraints on the Folding Grammar	21
3.4.3.1 Hard Constraints API	21
3.4.3.2 Soft Constraints API	22
3.5 RNA Secondary Structure Landscapes	22
3.5.1 The Neighborhood of a Secondary Structure	22
3.5.2 The Secondary Structure Landscape API	22
3.6 Minimum Free Energy Algorithm(s)	22
3.6.1 Zuker's Algorithm	22
3.6.2 MFE for circular RNAs	22
3.6.3 MFE Algorithm API	22
3.7 Partition Function and Equilibrium Probabilitiy Algorithm(s)	23
3.7.1 Equilibrium Ensemble Statistics	23
3.7.2 Partition Function and Equilibrium Probability API	23
3.8 Suboptimals and (other) Representative Structures	24
3.8.1 Suboptimal Secondary Structures	24
3.8.2 Sampling Secondary Structures from the Ensemble	24
3.8.3 Structure Enumeration and Sampling API	24
3.9 RNA-RNA Interaction	24
3.9.1 rip_intro	24
3.9.2 Concatenating RNA sequences	24
3.9.3 RNA-RNA interaction as a Stepwise Process	24
3.9.4 RNA-RNA Interaction API	25
3.10 Locally Stable Secondary Structures	25
3.10.1 local_intro	25

3.10.2 local_mfe	25
3.10.3 local_pf	25
3.10.4 Locally Stable Secondary Structure API	25
3.11 Comparative Structure Prediction	25
3.11.1 Incorporate Evolutionary Information	25
3.11.2 Comparative Structure Prediction API	25
3.12 Classified DP variations	25
3.12.1 The Idea of Classified Dynamic Programming	25
3.12.2 Distance Class Partitioning	25
3.12.3 Density of States (DOS)	26
3.12.4 Classified DP API	26
3.13 RNA Sequence Design	26
3.13.1 Generate Sequences that fold into particular Secondary Structures	26
3.13.2 RNA Sequence Design API	26
3.14 Experimental Structure Probing Data	26
3.14.1 Guide the Structure Prediction using Experimental Data	26
3.14.1.1 SHAPE reactivities	26
3.14.2 Structure Probing Data API	26
3.15 Ligand Binding	26
3.15.1 Small Molecules and Proteins that bind to specific RNA Structures	26
3.15.2 ligand_binding_api	26
3.16 (Tertiary) Structure Motifs	27
3.16.1 Incorporating Higher-Order (Tertiary) Structure Motifs	27
3.16.2 RNA G-Quadruplexes	27
3.16.3 (Tertiary) Structure Motif API	27
I/O Formats	29
4.1 RNA Structure Notations	29
4.1.1 Representations of Secondary Structures	29
4.1.2 Dot-Bracket Notation (a.k.a. Dot-Parenthesis Notation)	29
4.1.3 Extended Dot-Bracket Notation	30
4.1.4 Washington University Secondary Structure (WUSS) notation	30
4.1.5 Tree Representations of Secondary Structures	31
4.1.6 Examples for Structure Parsing and Conversion	32
4.1.7 Structure Parsing and Conversion API	32
4.2 File Formats	34
4.2.1 File formats for Multiple Sequence Alignments (MSA)	34
4.2.1.1 ClustalW format	34
4.2.1.2 Stockholm 1.0 format	35
4.2.1.3 FASTA (Pearson) format	35
4.2.1.4 MAF format	36
4.2.2 File formats to manipulate the RNA folding grammar	37

10 Changelog	73
9 Contributing	71
8.3 Python Examples	. 67
8.2 Perl5 Examples	
8.1.6 Deprecated Examples	. 66
8.1.5 Other Examples	. 65
8.1.4 Application of Soft Constraints	. 64
8.1.3 Writing Callback Functions	. 64
8.1.2 First Steps with the Fold Compound	. 63
8.1.1 Hello World Examples	. 61
8.1 C Examples	. 61
8 Examples	61
7 Additional Utilities	59
6.3.5 SWIG generated Wrapper notes	. 49
6.3.4 Examples	
6.3.3 Object oriented Interface for Data Structures	
6.3.2.1 Global Variables	
6.3.2 Function Renaming	. 48
6.3.1 Introduction	. 48
6.3 Scripting Language interface(s)	. 48
6.2.2 List of available Callbacks	. 47
6.2.1 The purpose of Callback mechanisms	. 46
6.2 Callback Functions	. 46
6.1.4 Some Examples using RNAlib API v3.0	. 46
6.1.3 How to port your program to the new API	. 46
6.1.2 What are the major changes?	. 45
6.1.1 Introduction	
6.1 RNAlib API v3.0	. 45
6 API Features	45
5.3 Model Details	. 43
5.2 The 'Fold Compound'	. 43
5.1 Sequence and Structure Data	. 43
5 Basic Data Structures	43
4.3.3 Producing (colored) alignments	. 41
4.3.2 Producing (colored) dot plots for base pair probabilities	. 41
4.3.1 Producing secondary structure graphs	. 40
4.3 Plotting	. 40
4.2.2.1 Command Files	. 37

11 Deprecated List	101
12 Bug List	113
13 Module Index	115
13.1 The RNAlib API	115
14 Data Structure Index	119
14.1 Data Structures	119
15 File Index	121
15.1 File List	121
16 Module Documentation	127
16.1 Free Energy Evaluation	127
16.1.1 Detailed Description	127
16.1.2 Function Documentation	130
16.1.2.1 vrna_eval_structure()	130
16.1.2.2 vrna_eval_covar_structure()	131
16.1.2.3 vrna_eval_structure_verbose()	131
16.1.2.4 vrna_eval_structure_v()	132
16.1.2.5 vrna_eval_structure_pt()	133
16.1.2.6 vrna_eval_structure_pt_verbose()	134
16.1.2.7 vrna_eval_structure_pt_v()	134
16.1.2.8 vrna_eval_structure_simple()	135
16.1.2.9 vrna_eval_circ_structure()	136
16.1.2.10 vrna_eval_gquad_structure()	136
16.1.2.11 vrna_eval_circ_gquad_structure()	137
16.1.2.12 vrna_eval_structure_simple_verbose()	138
16.1.2.13 vrna_eval_structure_simple_v()	138
16.1.2.14 vrna_eval_circ_structure_v()	139
16.1.2.15 vrna_eval_gquad_structure_v()	140
16.1.2.16 vrna_eval_circ_gquad_structure_v()	140
16.1.2.17 vrna_eval_consensus_structure_simple()	141
16.1.2.18 vrna_eval_circ_consensus_structure()	142
16.1.2.19 vrna_eval_gquad_consensus_structure()	143
16.1.2.20 vrna_eval_circ_gquad_consensus_structure()	143
16.1.2.21 vrna_eval_consensus_structure_simple_verbose()	144
16.1.2.22 vrna_eval_consensus_structure_simple_v()	145
16.1.2.23 vrna_eval_circ_consensus_structure_v()	146
16.1.2.24 vrna_eval_gquad_consensus_structure_v()	147
16.1.2.25 vrna_eval_circ_gquad_consensus_structure_v()	147
16.1.2.26 vrna_eval_structure_pt_simple()	148
16.1.2.27 vrna_eval_structure_pt_simple_verbose()	149

16.1.2.28 vrna_eval_structure_pt_simple_v()
16.1.2.29 vrna_eval_consensus_structure_pt_simple()
16.2 Energy Evaluation for Individual Loops
16.2.1 Detailed Description
16.2.2 Function Documentation
16.2.2.1 vrna_eval_loop_pt()
16.2.2.2 vrna_eval_loop_pt_v()
16.3 Energy Evaluation for Atomic Moves
16.3.1 Detailed Description
16.3.2 Function Documentation
16.3.2.1 vrna_eval_move()
16.3.2.2 vrna_eval_move_pt()
16.4 Deprecated Interface for Free Energy Evaluation
16.4.1 Detailed Description
16.4.2 Function Documentation
16.4.2.1 energy_of_structure()
16.4.2.2 energy_of_struct_par()
16.4.2.3 energy_of_circ_structure()
16.4.2.4 energy_of_circ_struct_par()
16.4.2.5 energy_of_structure_pt()
16.4.2.6 energy_of_struct_pt_par()
16.4.2.7 energy_of_move()
16.4.2.8 energy_of_move_pt()
16.4.2.9 loop_energy()
16.4.2.10 energy_of_struct()
16.4.2.11 energy_of_struct_pt()
16.4.2.12 energy_of_circ_struct()
16.4.2.13 E_Stem()
16.4.2.14 exp_E_ExtLoop()
16.4.2.15 exp_E_Stem()
16.4.2.16 E_IntLoop()
16.4.2.17 exp_E_IntLoop()
16.5 The RNA Folding Grammar
16.5.1 Detailed Description
16.5.2 Data Structure Documentation
16.5.2.1 struct vrna_gr_aux_s
16.6 Fine-tuning of the Implemented Models
16.6.1 Detailed Description
16.6.2 Data Structure Documentation
16.6.2.1 struct vrna_md_s
16.6.3 Macro Definition Documentation
16.6.3.1 VRNA MODEL DEFAULT TEMPERATURE

16.6.3.2 VRNA_MODEL_DEFAULT_PF_SCALE	181
16.6.3.3 VRNA_MODEL_DEFAULT_BETA_SCALE	181
16.6.3.4 VRNA_MODEL_DEFAULT_DANGLES	181
16.6.3.5 VRNA_MODEL_DEFAULT_SPECIAL_HP	181
16.6.3.6 VRNA_MODEL_DEFAULT_NO_LP	182
16.6.3.7 VRNA_MODEL_DEFAULT_NO_GU	182
16.6.3.8 VRNA_MODEL_DEFAULT_NO_GU_CLOSURE	182
16.6.3.9 VRNA_MODEL_DEFAULT_CIRC	182
16.6.3.10 VRNA_MODEL_DEFAULT_GQUAD	183
16.6.3.11 VRNA_MODEL_DEFAULT_UNIQ_ML	183
16.6.3.12 VRNA_MODEL_DEFAULT_ENERGY_SET	183
16.6.3.13 VRNA_MODEL_DEFAULT_BACKTRACK	183
16.6.3.14 VRNA_MODEL_DEFAULT_BACKTRACK_TYPE	184
16.6.3.15 VRNA_MODEL_DEFAULT_COMPUTE_BPP	184
16.6.3.16 VRNA_MODEL_DEFAULT_MAX_BP_SPAN	184
16.6.3.17 VRNA_MODEL_DEFAULT_WINDOW_SIZE	184
16.6.3.18 VRNA_MODEL_DEFAULT_LOG_ML	185
16.6.3.19 VRNA_MODEL_DEFAULT_ALI_OLD_EN	185
16.6.3.20 VRNA_MODEL_DEFAULT_ALI_RIBO	185
16.6.3.21 VRNA_MODEL_DEFAULT_ALI_CV_FACT	185
16.6.3.22 VRNA_MODEL_DEFAULT_ALI_NC_FACT	186
16.6.4 Function Documentation	186
16.6.4.1 vrna_md_set_default()	186
16.6.4.2 vrna_md_update()	186
16.6.4.3 vrna_md_copy()	187
16.6.4.4 vrna_md_option_string()	187
16.6.4.5 vrna_md_defaults_reset()	187
16.6.4.6 vrna_md_defaults_temperature()	188
16.6.4.7 vrna_md_defaults_temperature_get()	188
16.6.4.8 vrna_md_defaults_betaScale()	189
16.6.4.9 vrna_md_defaults_betaScale_get()	189
16.6.4.10 vrna_md_defaults_dangles()	190
16.6.4.11 vrna_md_defaults_dangles_get()	190
16.6.4.12 vrna_md_defaults_special_hp()	190
16.6.4.13 vrna_md_defaults_special_hp_get()	191
16.6.4.14 vrna_md_defaults_noLP()	191
16.6.4.15 vrna_md_defaults_noLP_get()	191
16.6.4.16 vrna_md_defaults_noGU()	192
16.6.4.17 vrna_md_defaults_noGU_get()	192
16.6.4.18 vrna_md_defaults_noGUclosure()	193
16.6.4.19 vrna_md_defaults_noGUclosure_get()	193
16.6.4.20 vrna_md_defaults_logML()	193

16.6.4.21 vrna_md_defaults_logML_get()		194
16.6.4.22 vrna_md_defaults_circ()		194
16.6.4.23 vrna_md_defaults_circ_get()		194
16.6.4.24 vrna_md_defaults_gquad()		195
16.6.4.25 vrna_md_defaults_gquad_get()		195
16.6.4.26 vrna_md_defaults_uniq_ML()		196
16.6.4.27 vrna_md_defaults_uniq_ML_get()		196
16.6.4.28 vrna_md_defaults_energy_set()		196
16.6.4.29 vrna_md_defaults_energy_set_get()		197
16.6.4.30 vrna_md_defaults_backtrack()		197
16.6.4.31 vrna_md_defaults_backtrack_get()		197
16.6.4.32 vrna_md_defaults_backtrack_type()		198
16.6.4.33 vrna_md_defaults_backtrack_type_get()		198
16.6.4.34 vrna_md_defaults_compute_bpp()		
16.6.4.35 vrna_md_defaults_compute_bpp_get()		199
16.6.4.36 vrna_md_defaults_max_bp_span()		199
16.6.4.37 vrna_md_defaults_max_bp_span_get()	٠.	200
16.6.4.38 vrna_md_defaults_min_loop_size()	٠.	200
16.6.4.39 vrna_md_defaults_min_loop_size_get()		
16.6.4.40 vrna_md_defaults_window_size()	٠.	201
16.6.4.41 vrna_md_defaults_window_size_get()		
16.6.4.42 vrna_md_defaults_oldAliEn()		
16.6.4.43 vrna_md_defaults_oldAliEn_get()		
16.6.4.44 vrna_md_defaults_ribo()		
16.6.4.45 vrna_md_defaults_ribo_get()		
16.6.4.46 vrna_md_defaults_cv_fact()		
16.6.4.47 vrna_md_defaults_cv_fact_get()	٠.	203
16.6.4.48 vrna_md_defaults_nc_fact()		
16.6.4.49 vrna_md_defaults_nc_fact_get()		
16.6.4.50 vrna_md_defaults_sfact()		
16.6.4.51 vrna_md_defaults_sfact_get()		
16.6.4.52 set_model_details()		
16.6.5 Variable Documentation		
16.6.5.1 temperature		
16.6.5.2 pf_scale		
16.6.5.3 dangles		
16.6.5.4 tetra_loop		
16.6.5.5 noLonelyPairs		
16.6.5.6 energy_set		
16.6.5.7 do_backtrack		
16.6.5.8 backtrack_type		
16.6.5.9 nonstandards		208

16.6.5.10 max_bp_span	208
16.7 Energy Parameters	209
16.7.1 Detailed Description	209
16.7.2 Data Structure Documentation	210
16.7.2.1 struct vrna_param_s	210
16.7.2.2 struct vrna_exp_param_s	211
16.7.3 Typedef Documentation	212
16.7.3.1 paramT	212
16.7.3.2 pf_paramT	212
16.7.4 Function Documentation	212
16.7.4.1 vrna_params()	212
16.7.4.2 vrna_params_copy()	213
16.7.4.3 vrna_exp_params()	213
16.7.4.4 vrna_exp_params_comparative()	214
16.7.4.5 vrna_exp_params_copy()	214
16.7.4.6 vrna_params_subst()	215
16.7.4.7 vrna_exp_params_subst()	215
16.7.4.8 vrna_exp_params_rescale()	216
16.7.4.9 vrna_params_reset()	217
16.7.4.10 vrna_exp_params_reset()	218
16.7.4.11 get_scaled_pf_parameters()	218
16.7.4.12 get_boltzmann_factors()	219
16.7.4.13 get_boltzmann_factor_copy()	219
16.7.4.14 get_scaled_alipf_parameters()	220
16.7.4.15 get_boltzmann_factors_ali()	220
16.7.4.16 scale_parameters()	221
16.7.4.17 get_scaled_parameters()	221
16.8 Extending the Folding Grammar with Additional Domains	223
16.8.1 Detailed Description	223
16.9 Unstructured Domains	224
16.9.1 Detailed Description	224
16.9.2 Data Structure Documentation	226
16.9.2.1 struct vrna_unstructured_domain_s	226
16.9.3 Typedef Documentation	227
16.9.3.1 vrna_callback_ud_energy	227
16.9.3.2 vrna_callback_ud_exp_energy	227
16.9.3.3 vrna_callback_ud_production	228
16.9.3.4 vrna_callback_ud_exp_production	228
16.9.3.5 vrna_callback_ud_probs_add	229
16.9.3.6 vrna_callback_ud_probs_get	229
16.9.4 Function Documentation	229
16.9.4.1 vrna_ud_motifs_centroid()	229

16.9.4.2 vrna_ud_motifs_MEA()	. 230
16.9.4.3 vrna_ud_motifs_MFE()	. 230
16.9.4.4 vrna_ud_add_motif()	. 231
16.9.4.5 vrna_ud_remove()	. 232
16.9.4.6 vrna_ud_set_data()	. 232
16.9.4.7 vrna_ud_set_prod_rule_cb()	. 233
16.9.4.8 vrna_ud_set_exp_prod_rule_cb()	. 234
16.10 Structured Domains	. 236
16.10.1 Detailed Description	. 236
16.11 Constraining the RNA Folding Grammar	. 237
16.11.1 Detailed Description	. 237
16.11.2 Macro Definition Documentation	. 240
16.11.2.1 VRNA_CONSTRAINT_FILE	. 240
16.11.2.2 VRNA_CONSTRAINT_SOFT_MFE	. 240
16.11.2.3 VRNA_CONSTRAINT_SOFT_PF	. 241
16.11.2.4 VRNA_DECOMP_PAIR_HP	. 241
16.11.2.5 VRNA_DECOMP_PAIR_IL	. 242
16.11.2.6 VRNA_DECOMP_PAIR_ML	. 242
16.11.2.7 VRNA_DECOMP_ML_ML	. 243
16.11.2.8 VRNA_DECOMP_ML_STEM	. 244
16.11.2.9 VRNA_DECOMP_ML_ML	. 244
16.11.2.10 VRNA_DECOMP_ML_UP	. 245
16.11.2.11 VRNA_DECOMP_ML_ML_STEM	. 245
16.11.2.12 VRNA_DECOMP_ML_COAXIAL	. 246
16.11.2.13 VRNA_DECOMP_ML_COAXIAL_ENC	. 246
16.11.2.14 VRNA_DECOMP_EXT_EXT	. 247
16.11.2.15 VRNA_DECOMP_EXT_UP	. 247
16.11.2.16 VRNA_DECOMP_EXT_STEM	. 248
16.11.2.17 VRNA_DECOMP_EXT_EXT_EXT	. 248
16.11.2.18 VRNA_DECOMP_EXT_STEM_EXT	. 249
16.11.2.19 VRNA_DECOMP_EXT_EXT_STEM	. 249
16.11.2.20 VRNA_DECOMP_EXT_EXT_STEM1	. 250
16.11.3 Function Documentation	. 250
16.11.3.1 vrna_constraints_add()	. 250
16.11.3.2 vrna_message_constraint_options()	. 251
16.11.3.3 vrna_message_constraint_options_all()	. 253
16.12 Hard Constraints	. 254
16.12.1 Detailed Description	. 254
16.12.2 Data Structure Documentation	. 256
16.12.2.1 struct vrna_hc_s	. 256
16.12.2.2 struct vrna_hc_up_s	. 257
16.12.3 Macro Definition Documentation	257

16.12.3.1 VRNA_CONSTRAINT_DB	 25/
16.12.3.2 VRNA_CONSTRAINT_DB_ENFORCE_BP	 258
16.12.3.3 VRNA_CONSTRAINT_DB_PIPE	 258
16.12.3.4 VRNA_CONSTRAINT_DB_DOT	 258
16.12.3.5 VRNA_CONSTRAINT_DB_X	 259
16.12.3.6 VRNA_CONSTRAINT_DB_RND_BRACK	 259
16.12.3.7 VRNA_CONSTRAINT_DB_INTRAMOL	 259
16.12.3.8 VRNA_CONSTRAINT_DB_INTERMOL	 260
16.12.3.9 VRNA_CONSTRAINT_DB_GQUAD	 260
16.12.3.10 VRNA_CONSTRAINT_DB_WUSS	 260
16.12.3.11 VRNA_CONSTRAINT_DB_DEFAULT	 261
16.12.4 Typedef Documentation	 261
16.12.4.1 vrna_callback_hc_evaluate	 261
16.12.5 Function Documentation	 262
16.12.5.1 vrna_hc_init()	 262
16.12.5.2 vrna_hc_add_up()	 262
16.12.5.3 vrna_hc_add_up_batch()	 263
16.12.5.4 vrna_hc_add_bp()	 263
16.12.5.5 vrna_hc_add_bp_nonspecific()	 264
16.12.5.6 vrna_hc_free()	 264
16.12.5.7 vrna_hc_add_from_db()	 265
16.13 Soft Constraints	 266
16.13.1 Detailed Description	 266
16.13.2 Data Structure Documentation	 267
16.13.2.1 struct vrna_sc_s	 267
16.13.3 Typedef Documentation	 268
16.13.3.1 vrna_callback_sc_energy	 269
16.13.3.2 vrna_callback_sc_exp_energy	 270
16.13.3.3 vrna_callback_sc_backtrack	 271
16.13.4 Function Documentation	 271
16.13.4.1 vrna_sc_init()	 271
16.13.4.2 vrna_sc_set_bp()	 272
16.13.4.3 vrna_sc_add_bp()	 273
16.13.4.4 vrna_sc_set_up()	 273
16.13.4.5 vrna_sc_add_up()	 274
16.13.4.6 vrna_sc_remove()	 275
16.13.4.7 vrna_sc_free()	 275
16.13.4.8 vrna_sc_add_data()	 275
16.13.4.9 vrna_sc_add_f()	 276
16.13.4.10 vrna_sc_add_bt()	 276
16.13.4.11 vrna_sc_add_exp_f()	 277
16.14 The RNA Secondary Structure Landscape	 278

16.14.1 Detailed Description
16.15 Minimum Free Energy (MFE) Algorithms
16.15.1 Detailed Description
16.16 Partition Function and Equilibrium Properties
16.16.1 Detailed Description
16.16.2 Function Documentation
16.16.2.1 vrna_pf_float_precision()
16.17 Global MFE Prediction
16.17.1 Detailed Description
16.17.2 Function Documentation
16.17.2.1 vrna_mfe()
16.17.2.2 vrna_mfe_dimer()
16.17.2.3 vrna_fold()
16.17.2.4 vrna_circfold()
16.17.2.5 vrna_alifold()
16.17.2.6 vrna_circalifold()
16.17.2.7 vrna_cofold()
16.18 Local (sliding window) MFE Prediction
16.18.1 Detailed Description
16.18.2 Typedef Documentation
16.18.2.1 vrna_mfe_window_callback
16.18.3 Function Documentation
16.18.3.1 vrna_mfe_window()
16.18.3.2 vrna_mfe_window_zscore()
16.18.3.3 vrna_Lfold()
16.18.3.4 vrna_Lfoldz()
16.19 Backtracking MFE structures
16.19.1 Detailed Description
16.19.2 Function Documentation
16.19.2.1 vrna_backtrack5()
16.19.2.2 vrna_BT_hp_loop()
16.19.2.3 vrna_BT_stack()
16.19.2.4 vrna_BT_int_loop()
16.19.2.5 vrna_BT_mb_loop()
16.20 Global Partition Function and Equilibrium Probabilities
16.20.1 Detailed Description
16.20.2 Data Structure Documentation
16.20.2.1 struct vrna_dimer_pf_s
16.20.3 Function Documentation
16.20.3.1 vrna_mean_bp_distance_pr()
16.20.3.2 vrna_mean_bp_distance()
16.20.3.3 vrna_ensemble_defect()

16.20.3.4 vrna_stack_prob()	301
16.20.3.5 vrna_pf_dimer_probs()	301
16.20.3.6 vrna_pr_structure()	302
16.20.3.7 vrna_pf()	302
16.20.3.8 vrna_pf_dimer()	303
16.20.3.9 vrna_pf_fold()	304
16.20.3.10 vrna_pf_circfold()	305
16.20.3.11 vrna_pf_alifold()	305
16.20.3.12 vrna_pf_circalifold()	306
16.20.3.13 vrna_plist_from_probs()	308
16.20.3.14 vrna_positional_entropy()	308
16.20.3.15 vrna_pf_co_fold()	309
16.21 Local (sliding window) Partition Function and Equilibrium Probabilities	311
16.21.1 Detailed Description	311
16.21.2 Macro Definition Documentation	312
16.21.2.1 VRNA_PROBS_WINDOW_BPP	312
16.21.2.2 VRNA_PROBS_WINDOW_UP	313
16.21.2.3 VRNA_PROBS_WINDOW_STACKP	313
16.21.2.4 VRNA_PROBS_WINDOW_UP_SPLIT	313
16.21.2.5 VRNA_PROBS_WINDOW_PF	314
16.21.3 Typedef Documentation	314
16.21.3.1 vrna_probs_window_callback	314
16.21.4 Function Documentation	315
16.21.4.1 vrna_probs_window()	315
16.21.4.2 vrna_pfl_fold()	316
16.21.4.3 vrna_pfl_fold_cb()	317
16.21.4.4 vrna_pfl_fold_up()	317
16.21.4.5 vrna_pfl_fold_up_cb()	318
16.22 Suboptimals and Representative Structures	321
16.22.1 Detailed Description	321
16.23 Suboptimal Structures sensu Stiegler et al. 1984 / Zuker et al. 1989	322
16.23.1 Detailed Description	322
16.23.2 Function Documentation	322
16.23.2.1 vrna_subopt_zuker()	322
16.23.2.2 zukersubopt()	323
16.23.2.3 zukersubopt_par()	323
16.24 Suboptimal Structures within an Energy Band around the MFE	324
16.24.1 Detailed Description	324
16.24.2 Typedef Documentation	324
16.24.2.1 vrna_subopt_callback	324
16.24.3 Function Documentation	325
16 24 3 1 yrna_subont()	325

16.24.3.2 vrna_subopt_cb()
16.24.3.3 subopt()
16.24.3.4 subopt_circ()
16.25 Random Structure Samples from the Ensemble
16.25.1 Detailed Description
16.25.2 Macro Definition Documentation
16.25.2.1 VRNA_PBACKTRACK_DEFAULT
16.25.2.2 VRNA_PBACKTRACK_NON_REDUNDANT
16.25.3 Typedef Documentation
16.25.3.1 vrna_boltzmann_sampling_callback
16.25.3.2 vrna_pbacktrack_mem_t
16.25.4 Function Documentation
16.25.4.1 vrna_pbacktrack5()
16.25.4.2 vrna_pbacktrack5_num()
16.25.4.3 vrna_pbacktrack5_cb()
16.25.4.4 vrna_pbacktrack5_resume()
16.25.4.5 vrna_pbacktrack5_resume_cb()
16.25.4.6 vrna_pbacktrack()
16.25.4.7 vrna_pbacktrack_num()
16.25.4.8 vrna_pbacktrack_cb()
16.25.4.9 vrna_pbacktrack_resume()
16.25.4.10 vrna_pbacktrack_resume_cb()
16.25.4.11 vrna_pbacktrack_mem_free()
16.26 Compute the Structure with Maximum Expected Accuracy (MEA)
16.26.1 Detailed Description
16.26.2 Function Documentation
16.26.2.1 vrna_MEA()
16.26.2.2 vrna_MEA_from_plist()
16.26.2.3 MEA()
16.26.2.3 MEA() 34 16.27 Compute the Centroid Structure 35
v
16.27 Compute the Centroid Structure
16.27 Compute the Centroid Structure 350 16.27.1 Detailed Description 350
16.27 Compute the Centroid Structure
16.27 Compute the Centroid Structure 350 16.27.1 Detailed Description 350 16.27.2 Function Documentation 350 16.27.2.1 vrna_centroid() 350
16.27 Compute the Centroid Structure 356 16.27.1 Detailed Description 356 16.27.2 Function Documentation 356 16.27.2.1 vrna_centroid() 356 16.27.2.2 vrna_centroid_from_plist() 35
16.27 Compute the Centroid Structure 350 16.27.1 Detailed Description 350 16.27.2 Function Documentation 350 16.27.2.1 vrna_centroid() 350 16.27.2.2 vrna_centroid_from_plist() 350 16.27.2.3 vrna_centroid_from_probs() 350
16.27 Compute the Centroid Structure 35 16.27.1 Detailed Description 35 16.27.2 Function Documentation 35 16.27.2.1 vrna_centroid() 35 16.27.2.2 vrna_centroid_from_plist() 35 16.27.2.3 vrna_centroid_from_probs() 35 16.28 RNA-RNA Interaction 35
16.27 Compute the Centroid Structure 356 16.27.1 Detailed Description 356 16.27.2 Function Documentation 356 16.27.2.1 vrna_centroid() 356 16.27.2.2 vrna_centroid_from_plist() 35 16.27.2.3 vrna_centroid_from_probs() 35 16.28 RNA-RNA Interaction 35 16.28.1 Detailed Description 35
16.27 Compute the Centroid Structure 35 16.27.1 Detailed Description 35 16.27.2 Function Documentation 35 16.27.2.1 vrna_centroid() 35 16.27.2.2 vrna_centroid_from_plist() 35 16.27.2.3 vrna_centroid_from_probs() 35 16.28 RNA-RNA Interaction 35 16.28.1 Detailed Description 35 16.29 Classified Dynamic Programming Variants 35
16.27 Compute the Centroid Structure 35 16.27.1 Detailed Description 35 16.27.2 Function Documentation 35 16.27.2.1 vrna_centroid() 35 16.27.2.2 vrna_centroid_from_plist() 35 16.27.2.3 vrna_centroid_from_probs() 35 16.28 RNA-RNA Interaction 35 16.29.1 Detailed Description 35 16.29.1 Detailed Description 35 16.29.1 Detailed Description 35

16.31.1 Detailed Description
16.31.2 Data Structure Documentation
16.31.2.1 struct vrna_sol_TwoD_t
16.31.2.2 struct TwoDfold_vars
16.31.3 Typedef Documentation
16.31.3.1 vrna_sol_TwoD_t
16.31.3.2 TwoDfold_vars
16.31.4 Function Documentation
16.31.4.1 vrna_mfe_TwoD()
16.31.4.2 vrna_backtrack5_TwoD()
16.31.4.3 get_TwoDfold_variables()
16.31.4.4 destroy_TwoDfold_variables()
16.31.4.5 TwoDfoldList()
16.31.4.6 TwoDfold_backtrack_f5()
16.31.4.7 TwoDfold()
16.32 Computing Partition Functions of a Distance Based Partitioning
16.32.1 Detailed Description
16.32.2 Data Structure Documentation
16.32.2.1 struct vrna_sol_TwoD_pf_t
16.32.3 Typedef Documentation
16.32.3.1 vrna_sol_TwoD_pf_t
16.32.4 Function Documentation
16.32.4.1 vrna_pf_TwoD()
16.33 Stochastic Backtracking of Structures from Distance Based Partitioning
16.33.1 Detailed Description
16.33.2 Function Documentation
16.33.2.1 vrna_pbacktrack_TwoD()
16.33.2.2 vrna_pbacktrack5_TwoD()
16.34 Compute the Density of States
16.34.1 Detailed Description
16.34.2 Variable Documentation
16.34.2.1 density_of_states
16.35 Inverse Folding (Design)
16.35.1 Detailed Description
16.35.2 Function Documentation
16.35.2.1 inverse_fold()
16.35.2.2 inverse_pf_fold()
16.35.2.2 inverse_pf_fold() 373 16.35.3 Variable Documentation 373
16.35.3 Variable Documentation
16.35.3 Variable Documentation 373 16.35.3.1 final_cost 373

16.36.1 Detailed Description	3/5
16.36.2 Data Structure Documentation	377
16.36.2.1 struct vrna_move_s	377
16.36.3 Macro Definition Documentation	378
16.36.3.1 VRNA_MOVESET_INSERTION	378
16.36.3.2 VRNA_MOVESET_DELETION	379
16.36.3.3 VRNA_MOVESET_SHIFT	379
16.36.3.4 VRNA_MOVESET_NO_LP	379
16.36.3.5 VRNA_MOVESET_DEFAULT	379
16.36.3.6 VRNA_NEIGHBOR_CHANGE	380
16.36.3.7 VRNA_NEIGHBOR_INVALID	380
16.36.3.8 VRNA_NEIGHBOR_NEW	380
16.36.4 Typedef Documentation	380
16.36.4.1 vrna_callback_move_update	380
16.36.5 Function Documentation	381
16.36.5.1 vrna_move_init()	381
16.36.5.2 vrna_move_list_free()	381
16.36.5.3 vrna_move_apply()	382
16.36.5.4 vrna_move_is_removal()	382
16.36.5.5 vrna_move_is_insertion()	382
16.36.5.6 vrna_move_is_shift()	383
16.36.5.7 vrna_move_compare()	383
16.36.5.8 vrna_loopidx_update()	384
16.36.5.9 vrna_neighbors()	384
16.36.5.10 vrna_neighbors_successive()	385
16.36.5.11 vrna_move_neighbor_diff_cb()	386
16.36.5.12 vrna_move_neighbor_diff()	387
16.37 (Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima	388
16.37.1 Detailed Description	388
16.37.2 Data Structure Documentation	389
16.37.2.1 struct vrna_path_s	389
16.37.3 Macro Definition Documentation	390
16.37.3.1 VRNA_PATH_TYPE_DOT_BRACKET	390
16.37.3.2 VRNA_PATH_TYPE_MOVES	391
16.37.4 Function Documentation	391
16.37.4.1 vrna_path_free()	391
16.37.4.2 vrna_path_options_free()	391
16.38 Direct Refolding Paths between two Secondary Structures	393
16.38.1 Detailed Description	393
16.38.2 Function Documentation	393
16.38.2.1 vrna_path_findpath_saddle()	393
16.38.2.2 vrna_path_findpath_saddle_ub()	394

16.38.2.3 vrna_path_findpath()	95
16.38.2.4 vrna_path_findpath_ub()	96
16.38.2.5 vrna_path_options_findpath()	96
16.38.2.6 vrna_path_direct()	97
16.38.2.7 vrna_path_direct_ub()	98
16.39 Folding Paths that start at a single Secondary Structure	00
16.39.1 Detailed Description	00
16.39.2 Macro Definition Documentation	00
16.39.2.1 VRNA_PATH_STEEPEST_DESCENT	00
16.39.2.2 VRNA_PATH_RANDOM	01
16.39.2.3 VRNA_PATH_NO_TRANSITION_OUTPUT	01
16.39.2.4 VRNA_PATH_DEFAULT	01
16.39.3 Function Documentation	01
16.39.3.1 vrna_path()	02
16.39.3.2 vrna_path_gradient()	03
16.39.3.3 vrna_path_random()	03
16.40 Experimental Structure Probing Data	05
16.40.1 Detailed Description	05
16.41 SHAPE Reactivity Data	06
16.41.1 Detailed Description	06
16.41.2 Function Documentation	06
16.41.2.1 vrna_sc_add_SHAPE_deigan()	06
16.41.2.2 vrna_sc_add_SHAPE_deigan_ali()	07
16.41.2.3 vrna_sc_add_SHAPE_zarringhalam()	80
16.41.2.4 vrna_sc_SHAPE_to_pr()	09
16.42 Generate Soft Constraints from Data	10
16.42.1 Detailed Description	10
16.42.2 Macro Definition Documentation	11
16.42.2.1 VRNA_OBJECTIVE_FUNCTION_QUADRATIC	11
16.42.2.2 VRNA_OBJECTIVE_FUNCTION_ABSOLUTE	11
16.42.2.3 VRNA_MINIMIZER_CONJUGATE_FR	11
16.42.2.4 VRNA_MINIMIZER_CONJUGATE_PR	11
16.42.2.5 VRNA_MINIMIZER_VECTOR_BFGS	12
16.42.2.6 VRNA_MINIMIZER_VECTOR_BFGS2	12
16.42.2.7 VRNA_MINIMIZER_STEEPEST_DESCENT	
16.42.3 Typedef Documentation	12
16.42.3.1 progress_callback	12
16.42.4 Function Documentation	13
16.42.4.1 vrna_sc_minimize_pertubation()	13
16.43 Ligands Binding to RNA Structures	
16.43.1 Detailed Description	15
16.44 Ligands Binding to Unstructured Domains	16

16.45 Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints 417
16.45.1 Detailed Description
16.45.2 Function Documentation
16.45.2.1 vrna_sc_add_hi_motif()
16.46 Complex Structured Modules
16.46.1 Detailed Description
16.47 G-Quadruplexes
16.47.1 Detailed Description
16.47.2 Function Documentation
16.47.2.1 get_gquad_matrix()
16.47.2.2 parse_gquad()
16.47.2.3 backtrack_GQuad_IntLoop()
16.47.2.4 backtrack_GQuad_IntLoop_L()
16.48 Utilities
16.48.1 Detailed Description
16.48.2 Macro Definition Documentation
16.48.2.1 VRNA_INPUT_FASTA_HEADER
16.48.2.2 VRNA_INPUT_CONSTRAINT
16.48.3 Function Documentation
16.48.3.1 vrna_alloc()
16.48.3.2 vrna_realloc()
16.48.3.3 vrna_urn()
16.48.3.4 vrna_int_urn()
16.48.3.5 vrna_time_stamp()
16.48.3.6 get_input_line()
16.48.3.7 vrna_idx_row_wise()
16.48.3.8 vrna_idx_col_wise()
16.48.4 Variable Documentation
16.48.4.1 xsubi
16.49 Exterior Loops
16.49.1 Detailed Description
16.49.2 Typedef Documentation
16.49.2.1 vrna_mx_pf_aux_el_t
16.49.3 Function Documentation
16.49.3.1 vrna_E_ext_stem()
16.49.3.2 vrna_E_ext_loop()
16.49.3.3 vrna_exp_E_ext_stem()
16.50 Hairpin Loops
16.50.1 Detailed Description
16.50.2 Function Documentation
16.50.2.1 vrna_E_hp_loop()
16.50.2.2 yrna E ext hp loop()

16.50.2.3 vrna_eval_hp_loop()	36
16.50.2.4 E_Hairpin()	36
16.50.2.5 exp_E_Hairpin()	37
16.50.2.6 vrna_exp_E_hp_loop()	38
16.51 Internal Loops	39
16.51.1 Detailed Description	39
16.51.2 Function Documentation	39
16.51.2.1 vrna_eval_int_loop()	39
16.52 Multibranch Loops	40
16.52.1 Detailed Description	40
16.52.2 Typedef Documentation	40
16.52.2.1 vrna_mx_pf_aux_ml_t	40
16.52.3 Function Documentation	41
16.52.3.1 vrna_E_mb_loop_stack()	41
16.53 Partition Function for Two Hybridized Sequences	42
16.53.1 Detailed Description	42
16.53.2 Function Documentation	43
16.53.2.1 vrna_pf_co_fold()	43
16.53.2.2 vrna_pf_dimer_concentrations()	44
16.54 Partition Function for two Hybridized Sequences as a Stepwise Process	45
16.54.1 Detailed Description	45
16.54.2 Function Documentation	45
16.54.2.1 pf_unstru()	45
16.54.2.2 pf_interact()	46
16.55 Reading/Writing Energy Parameter Sets from/to File	48
16.55.1 Detailed Description	48
16.55.2 Macro Definition Documentation	49
16.55.2.1 VRNA_PARAMETER_FORMAT_DEFAULT	49
16.55.3 Function Documentation	49
16.55.3.1 vrna_params_load()	49
16.55.3.2 vrna_params_save()	50
16.55.3.3 vrna_params_load_from_string()	50
16.55.3.4 vrna_params_load_defaults()	51
16.55.3.5 vrna_params_load_RNA_Turner2004()	51
16.55.3.6 vrna_params_load_RNA_Turner1999()	52
16.55.3.7 vrna_params_load_RNA_Andronescu2007()	52
16.55.3.8 vrna_params_load_RNA_Langdon2018()	53
16.55.3.9 vrna_params_load_RNA_misc_special_hairpins()	53
16.55.3.10 vrna_params_load_DNA_Mathews2004()	54
16.55.3.11 vrna_params_load_DNA_Mathews1999()	54
16.55.3.12 last_parameter_file()	55
16.55.3.13 read_parameter_file()	55

16.55.3.14 write_parameter_file()	455
16.56 Converting Energy Parameter Files	456
16.56.1 Detailed Description	456
16.56.2 Macro Definition Documentation	457
16.56.2.1 VRNA_CONVERT_OUTPUT_ALL	457
16.56.2.2 VRNA_CONVERT_OUTPUT_HP	457
16.56.2.3 VRNA_CONVERT_OUTPUT_STACK	457
16.56.2.4 VRNA_CONVERT_OUTPUT_MM_HP	457
16.56.2.5 VRNA_CONVERT_OUTPUT_MM_INT	457
16.56.2.6 VRNA_CONVERT_OUTPUT_MM_INT_1N	458
16.56.2.7 VRNA_CONVERT_OUTPUT_MM_INT_23	458
16.56.2.8 VRNA_CONVERT_OUTPUT_MM_MULTI	458
16.56.2.9 VRNA_CONVERT_OUTPUT_MM_EXT	458
16.56.2.10 VRNA_CONVERT_OUTPUT_DANGLE5	458
16.56.2.11 VRNA_CONVERT_OUTPUT_DANGLE3	458
16.56.2.12 VRNA_CONVERT_OUTPUT_INT_11	459
16.56.2.13 VRNA_CONVERT_OUTPUT_INT_21	459
16.56.2.14 VRNA_CONVERT_OUTPUT_INT_22	459
16.56.2.15 VRNA_CONVERT_OUTPUT_BULGE	459
16.56.2.16 VRNA_CONVERT_OUTPUT_INT	459
16.56.2.17 VRNA_CONVERT_OUTPUT_ML	459
16.56.2.18 VRNA_CONVERT_OUTPUT_MISC	460
16.56.2.19 VRNA_CONVERT_OUTPUT_SPECIAL_HP	460
16.56.2.20 VRNA_CONVERT_OUTPUT_VANILLA	460
16.56.2.21 VRNA_CONVERT_OUTPUT_NINIO	460
16.56.2.22 VRNA_CONVERT_OUTPUT_DUMP	460
16.56.3 Function Documentation	461
16.56.3.1 convert_parameter_file()	461
16.57 Utilities to deal with Nucleotide Alphabets	462
16.57.1 Detailed Description	462
16.57.2 Data Structure Documentation	463
16.57.2.1 struct vrna_sequence_s	463
16.57.2.2 struct vrna_alignment_s	463
16.57.3 Enumeration Type Documentation	463
16.57.3.1 vrna_seq_type_e	463
16.57.4 Function Documentation	463
16.57.4.1 vrna_ptypes()	464
16.57.4.2 vrna_seq_encode()	464
16.57.4.3 vrna_seq_encode_simple()	464
16.57.4.4 vrna_nucleotide_encode()	464
16.57.4.5 vrna_nucleotide_decode()	465
16.58 (Nucleic Acid Sequence) String Utilitites	466

16.58.1 Detailed Description	66
16.58.2 Macro Definition Documentation	67
16.58.2.1 FILENAME_MAX_LENGTH	67
16.58.2.2 FILENAME_ID_LENGTH	67
16.58.3 Function Documentation	67
16.58.3.1 vrna_strdup_printf()	67
16.58.3.2 vrna_strdup_vprintf()	68
16.58.3.3 vrna_strcat_printf()	68
16.58.3.4 vrna_strcat_vprintf()	69
16.58.3.5 vrna_strsplit()	70
16.58.3.6 vrna_random_string()	71
16.58.3.7 vrna_hamming_distance()	71
16.58.3.8 vrna_hamming_distance_bound()	71
16.58.3.9 vrna_seq_toRNA()	72
16.58.3.10 vrna_seq_toupper()	72
16.58.3.11 vrna_seq_ungapped()	73
16.58.3.12 vrna_cut_point_insert()	73
16.58.3.13 vrna_cut_point_remove()	73
16.59 Secondary Structure Utilities	75
16.59.1 Detailed Description	75
16.59.2 Function Documentation	75
16.59.2.1 vrna_bp_distance()	76
16.59.2.2 vrna_refBPcnt_matrix()	77
16.59.2.3 vrna_refBPdist_matrix()	77
16.59.2.4 vrna_db_from_bp_stack()	77
16.60 Dot-Bracket Notation of Secondary Structures	79
16.60.1 Detailed Description	79
16.60.2 Macro Definition Documentation	79
16.60.2.1 VRNA_BRACKETS_ALPHA	80
16.60.2.2 VRNA_BRACKETS_RND	80
16.60.2.3 VRNA_BRACKETS_CLY	80
16.60.2.4 VRNA_BRACKETS_ANG	80
16.60.2.5 VRNA_BRACKETS_SQR	81
16.60.2.6 VRNA_BRACKETS_DEFAULT	81
16.60.2.7 VRNA_BRACKETS_ANY	81
16.60.3 Function Documentation	82
16.60.3.1 vrna_db_pack()	82
16.60.3.2 vrna_db_unpack()	82
16.60.3.3 vrna_db_flatten()	83
16.60.3.4 vrna_db_flatten_to()	83
16.60.3.5 vrna_db_from_ptable()	84
16.60.3.6 vrna db from WUSS()	84

16.60.3.7 vrna_db_from_plist()	85
16.60.3.8 vrna_db_to_element_string()	85
16.60.3.9 vrna_db_pk_remove()	86
16.61 Pair Table Representation of Secondary Structures	87
16.61.1 Detailed Description	87
16.61.2 Function Documentation	87
16.61.2.1 vrna_ptable()	87
16.61.2.2 vrna_ptable_from_string()	88
16.61.2.3 vrna_pt_pk_get()	88
16.61.2.4 vrna_ptable_copy()	89
16.61.2.5 vrna_pt_ali_get()	89
16.61.2.6 vrna_pt_snoop_get()	89
16.61.2.7 vrna_pt_pk_remove()	90
16.62 Pair List Representation of Secondary Structures	91
16.62.1 Detailed Description	91
16.62.2 Data Structure Documentation	91
16.62.2.1 struct vrna_elem_prob_s	91
16.62.3 Function Documentation	92
16.62.3.1 vrna_plist()	92
16.63 Helix List Representation of Secondary Structures	93
16.63.1 Detailed Description	93
16.63.2 Data Structure Documentation	93
16.63.2.1 struct vrna_hx_s	93
16.63.3 Function Documentation	93
16.63.3.1 vrna_hx_from_ptable()	93
16.64 Tree Representation of Secondary Structures	95
16.64.1 Detailed Description	95
16.64.2 Macro Definition Documentation	95
16.64.2.1 VRNA_STRUCTURE_TREE_HIT	95
16.64.2.2 VRNA_STRUCTURE_TREE_SHAPIRO_SHORT	96
16.64.2.3 VRNA_STRUCTURE_TREE_SHAPIRO	96
16.64.2.4 VRNA_STRUCTURE_TREE_SHAPIRO_EXT	96
16.64.2.5 VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT	96
16.64.2.6 VRNA_STRUCTURE_TREE_EXPANDED	97
16.64.3 Function Documentation	97
16.64.3.1 vrna_db_to_tree_string()	97
16.64.3.2 vrna_tree_string_unweight()	98
16.64.3.3 vrna_tree_string_to_db()	98
16.65 Multiple Sequence Alignment Utilities	01
16.65.1 Detailed Description	01
16.65.2 Data Structure Documentation	02
16.65.2.1 struct vrna pinfo s	02

16.65.3 Macro Definition Documentation)3
16.65.3.1 VRNA_MEASURE_SHANNON_ENTROPY)3
16.65.4 Function Documentation)3
16.65.4.1 vrna_aln_mpi())3
16.65.4.2 vrna_aln_pinfo())3
16.65.4.3 vrna_aln_slice())5
16.65.4.4 vrna_aln_free())5
16.65.4.5 vrna_aln_uppercase())7
16.65.4.6 vrna_aln_toRNA())7
16.65.4.7 vrna_aln_copy()	8(
16.65.4.8 vrna_aln_conservation_struct()	8(
16.65.4.9 vrna_aln_conservation_col())9
16.65.4.10 vrna_aln_consensus_sequence()	0
16.65.4.11 vrna_aln_consensus_mis()	0
16.66 Files and I/O	1
16.66.1 Detailed Description	1
16.66.2 Function Documentation	2
16.66.2.1 readribosum()	2
16.66.2.2 vrna_read_line()	2
16.66.2.3 vrna_filename_sanitize()	2
16.66.2.4 vrna_file_exists()	3
16.67 Nucleic Acid Sequences and Structures	5
16.67.1 Detailed Description	5
16.67.2 Macro Definition Documentation	5
16.67.2.1 VRNA_OPTION_MULTILINE	6
16.67.2.2 VRNA_CONSTRAINT_MULTILINE	6
16.67.3 Function Documentation	6
16.67.3.1 vrna_file_helixlist()	6
16.67.3.2 vrna_file_connect()	7
16.67.3.3 vrna_file_bpseq()	7
16.67.3.4 vrna_file_json()	8
16.67.3.5 vrna_file_fasta_read_record()	8
16.67.3.6 vrna_extract_record_rest_structure()	20
16.67.3.7 vrna_file_SHAPE_read()	20
16.67.3.8 vrna_extract_record_rest_constraint()	21
16.67.3.9 read_record()	22
16.68 Multiple Sequence Alignments	23
16.68.1 Detailed Description	23
16.68.2 Macro Definition Documentation	24
16.68.2.1 VRNA_FILE_FORMAT_MSA_CLUSTAL	24
16.68.2.2 VRNA_FILE_FORMAT_MSA_STOCKHOLM	24
16.68.2.3 VRNA_FILE_FORMAT_MSA_FASTA	24

16	6.68.2.4 VRNA_FILE_FORMAT_MSA_MAF	525
16	6.68.2.5 VRNA_FILE_FORMAT_MSA_MIS	525
16	6.68.2.6 VRNA_FILE_FORMAT_MSA_DEFAULT	525
16	6.68.2.7 VRNA_FILE_FORMAT_MSA_NOCHECK	526
16	6.68.2.8 VRNA_FILE_FORMAT_MSA_UNKNOWN	526
16	6.68.2.9 VRNA_FILE_FORMAT_MSA_APPEND	526
16	6.68.2.10 VRNA_FILE_FORMAT_MSA_QUIET	526
16	6.68.2.11 VRNA_FILE_FORMAT_MSA_SILENT	527
16.68.3 Fu	unction Documentation	527
16	6.68.3.1 vrna_file_msa_read()	527
16	6.68.3.2 vrna_file_msa_read_record()	528
16	6.68.3.3 vrna_file_msa_detect_format()	530
16	6.68.3.4 vrna_file_msa_write()	530
16.69 Command	d Files	532
16.69.1 De	etailed Description	532
16.69.2 M	acro Definition Documentation	533
16	6.69.2.1 VRNA_CMD_PARSE_HC	533
16	6.69.2.2 VRNA_CMD_PARSE_SC	533
16	6.69.2.3 VRNA_CMD_PARSE_UD	533
16	6.69.2.4 VRNA_CMD_PARSE_SD	534
16	6.69.2.5 VRNA_CMD_PARSE_DEFAULTS	534
16.69.3 Fu	unction Documentation	534
16	6.69.3.1 vrna_file_commands_read()	534
16	6.69.3.2 vrna_file_commands_apply()	535
16	6.69.3.3 vrna_commands_apply()	535
16	6.69.3.4 vrna_commands_free()	536
16.70 Plotting		537
16.70.1 De	etailed Description	537
16.70.2 Da	ata Structure Documentation	538
16	5.70.2.1 struct vrna_dotplot_auxdata_t	538
16.70.3 Fu	unction Documentation	538
16	5.70.3.1 PS_dot_plot_list()	538
16	3.70.3.2 PS_dot_plot()	539
16	5.70.3.3 vrna_file_PS_rnaplot()	539
16	5.70.3.4 vrna_file_PS_rnaplot_a()	540
16	3.70.3.5 gmlRNA()	540
16	5.70.3.6 ssv_rna_plot()	541
16	5.70.3.7 svg_rna_plot()	542
16	5.70.3.8 xrna_plot()	542
16	5.70.3.9 PS_rna_plot()	542
16	5.70.3.10 PS_rna_plot_a()	543
16	5.70.3.11 PS rna plot a gguad()	543

16.71 Layouts and Coordinates	544
16.71.1 Detailed Description	544
16.71.2 Data Structure Documentation	545
16.71.2.1 struct vrna_plot_layout_s	545
16.71.2.2 struct vrna_plot_options_puzzler_t	545
16.71.3 Macro Definition Documentation	545
16.71.3.1 VRNA_PLOT_TYPE_SIMPLE	546
16.71.3.2 VRNA_PLOT_TYPE_NAVIEW	546
16.71.3.3 VRNA_PLOT_TYPE_CIRCULAR	546
16.71.3.4 VRNA_PLOT_TYPE_TURTLE	547
16.71.3.5 VRNA_PLOT_TYPE_PUZZLER	547
16.71.4 Typedef Documentation	547
16.71.4.1 vrna_plot_layout_t	547
16.71.5 Function Documentation	547
16.71.5.1 vrna_plot_layout()	548
16.71.5.2 vrna_plot_layout_simple()	548
16.71.5.3 vrna_plot_layout_naview()	549
16.71.5.4 vrna_plot_layout_circular()	550
16.71.5.5 vrna_plot_layout_turtle()	550
16.71.5.6 vrna_plot_layout_puzzler()	551
16.71.5.7 vrna_plot_layout_free()	551
16.71.5.8 vrna_plot_coords()	552
16.71.5.9 vrna_plot_coords_pt()	553
16.71.5.10 vrna_plot_coords_simple()	554
16.71.5.11 vrna_plot_coords_simple_pt()	555
16.71.5.12 vrna_plot_coords_circular()	555
16.71.5.13 vrna_plot_coords_circular_pt()	556
16.71.5.14 vrna_plot_coords_naview()	557
16.71.5.15 vrna_plot_coords_naview_pt()	558
16.71.5.16 vrna_plot_coords_puzzler()	558
16.71.5.17 vrna_plot_coords_puzzler_pt()	559
16.71.5.18 vrna_plot_options_puzzler()	560
16.71.5.19 vrna_plot_options_puzzler_free()	560
16.71.5.20 vrna_plot_coords_turtle()	561
16.71.5.21 vrna_plot_coords_turtle_pt()	562
16.72 Annotation	563
16.72.1 Detailed Description	563
16.72.2 Function Documentation	563
16.72.2.1 vrna_annotate_covar_db()	563
16.72.2.2 vrna_annotate_covar_pairs()	563
16.73 Alignment Plots	564
16.73.1 Detailed Description	564

16.73.2 Function Documentation
16.73.2.1 vrna_file_PS_aln()
16.73.2.2 vrna_file_PS_aln_slice()
16.74 Search Algorithms
16.74.1 Detailed Description
16.74.2 Function Documentation
16.74.2.1 vrna_search_BMH_num()
16.74.2.2 vrna_search_BMH()
16.74.2.3 vrna_search_BM_BCT_num()
16.74.2.4 vrna_search_BM_BCT()
16.75 Combinatorics Algorithms
16.75.1 Detailed Description
16.75.2 Function Documentation
16.75.2.1 vrna_enumerate_necklaces()
16.75.2.2 vrna_rotational_symmetry_num()
16.75.2.3 vrna_rotational_symmetry_pos_num()
16.75.2.4 vrna_rotational_symmetry()
16.75.2.5 vrna_rotational_symmetry_pos()
16.75.2.6 vrna_rotational_symmetry_db()
16.75.2.7 vrna_rotational_symmetry_db_pos()
16.76 (Abstract) Data Structures
16.76.1 Detailed Description
16.76.2 Data Structure Documentation
16.76.2.1 struct vrna_basepair_s
16.76.2.2 struct vrna_cpair_s
16.76.2.3 struct vrna_color_s
16.76.2.4 struct vrna_data_linear_s
16.76.2.5 struct vrna_sect_s
16.76.2.6 struct vrna_bp_stack_s
16.76.2.7 struct pu_contrib
16.76.2.8 struct interact
16.76.2.9 struct pu_out
16.76.2.10 struct constrain
16.76.2.11 struct duplexT
16.76.2.12 struct node
16.76.2.13 struct snoopT
16.76.2.14 struct dupVar
16.76.3 Typedef Documentation
16.76.3.1 PAIR
16.76.3.2 plist
16.76.3.3 cpair
16.76.3.4 sect

16.76.3.5 bondT	1
16.76.4 Function Documentation	1
16.76.4.1 vrna_C11_features()	2
16.77 Messages	3
16.77.1 Detailed Description	3
16.77.2 Function Documentation	3
16.77.2.1 vrna_message_error()	3
16.77.2.2 vrna_message_verror()	4
16.77.2.3 vrna_message_warning()	4
16.77.2.4 vrna_message_vwarning()	15
16.77.2.5 vrna_message_info()	15
16.77.2.6 vrna_message_vinfo()	6
16.77.2.7 vrna_message_input_seq_simple()	6
16.77.2.8 vrna_message_input_seq()	7
16.78 Unit Conversion	9
16.78.1 Detailed Description	9
16.78.2 Enumeration Type Documentation	9
16.78.2.1 vrna_unit_energy_e	9
16.78.2.2 vrna_unit_temperature_e	0
16.78.3 Function Documentation	11
16.78.3.1 vrna_convert_energy()	11
16.78.3.2 vrna_convert_temperature()	11
16.79 The Fold Compound	13
16.79.1 Detailed Description	13
16.79.2 Data Structure Documentation	14
16.79.2.1 struct vrna_fc_s	14
16.79.3 Macro Definition Documentation	12
16.79.3.1 VRNA_STATUS_MFE_PRE	12
16.79.3.2 VRNA_STATUS_MFE_POST	13
16.79.3.3 VRNA_STATUS_PF_PRE	13
16.79.3.4 VRNA_STATUS_PF_POST	13
16.79.3.5 VRNA_OPTION_MFE	13
16.79.3.6 VRNA_OPTION_PF	14
16.79.3.7 VRNA_OPTION_EVAL_ONLY	14
16.79.4 Typedef Documentation	14
16.79.4.1 vrna_callback_free_auxdata	14
16.79.4.2 vrna_callback_recursion_status	15
16.79.5 Enumeration Type Documentation	15
16.79.5.1 vrna_fc_type_e	15
16.79.6 Function Documentation	16
16.79.6.1 vrna_fold_compound()	16
16.79.6.2 vrna_fold_compound_comparative()	7

16.79.6.3 vrna_fold_compound_free()	. 608
16.79.6.4 vrna_fold_compound_add_auxdata()	. 608
16.79.6.5 vrna_fold_compound_add_callback()	. 609
16.80 The Dynamic Programming Matrices	. 610
16.80.1 Detailed Description	. 610
16.80.2 Data Structure Documentation	. 610
16.80.2.1 struct vrna_mx_mfe_s	. 610
16.80.2.2 struct vrna_mx_pf_s	. 611
16.80.3 Enumeration Type Documentation	. 612
16.80.3.1 vrna_mx_type_e	. 612
16.80.4 Function Documentation	. 612
16.80.4.1 vrna_mx_add()	. 612
16.80.4.2 vrna_mx_mfe_free()	. 613
16.80.4.3 vrna_mx_pf_free()	. 613
16.81 Hash Tables	. 615
16.81.1 Detailed Description	. 615
16.81.2 Data Structure Documentation	. 616
16.81.2.1 struct vrna_ht_entry_db_t	. 616
16.81.3 Typedef Documentation	. 616
16.81.3.1 vrna_hash_table_t	. 617
16.81.3.2 vrna_callback_ht_compare_entries	. 617
16.81.3.3 vrna_callback_ht_hash_function	. 617
16.81.3.4 vrna_callback_ht_free_entry	. 618
16.81.4 Function Documentation	. 618
16.81.4.1 vrna_ht_init()	. 618
16.81.4.2 vrna_ht_size()	. 619
16.81.4.3 vrna_ht_collisions()	. 619
16.81.4.4 vrna_ht_get()	. 621
16.81.4.5 vrna_ht_insert()	. 621
16.81.4.6 vrna_ht_remove()	. 622
16.81.4.7 vrna_ht_clear()	. 622
16.81.4.8 vrna_ht_free()	. 623
16.81.4.9 vrna_ht_db_comp()	. 623
16.81.4.10 vrna_ht_db_hash_func()	. 624
16.81.4.11 vrna_ht_db_free_entry()	. 624
16.82 Heaps	. 626
16.82.1 Detailed Description	. 626
16.82.2 Typedef Documentation	. 627
16.82.2.1 vrna_heap_t	. 627
16.82.2.2 vrna_callback_heap_cmp	. 627
16.82.2.3 vrna_callback_heap_get_pos	. 628
16.82.2.4 vrna_callback_heap_set_pos	. 628

16.82.3 Function Documentation	628
16.82.3.1 vrna_heap_init()	628
16.82.3.2 vrna_heap_free()	629
16.82.3.3 vrna_heap_size()	630
16.82.3.4 vrna_heap_insert()	630
16.82.3.5 vrna_heap_pop()	631
16.82.3.6 vrna_heap_top()	631
16.82.3.7 vrna_heap_remove()	632
16.82.3.8 vrna_heap_update()	632
16.83 Buffers	634
16.83.1 Detailed Description	634
16.83.2 Typedef Documentation	634
16.83.2.1 vrna_callback_stream_output	635
16.83.3 Function Documentation	635
16.83.3.1 vrna_cstr()	635
16.83.3.2 vrna_cstr_free()	636
16.83.3.3 vrna_cstr_close()	636
16.83.3.4 vrna_cstr_fflush()	636
16.83.3.5 vrna_ostream_init()	637
16.83.3.6 vrna_ostream_free()	637
16.83.3.7 vrna_ostream_request()	638
16.83.3.8 vrna_ostream_provide()	638
	000
16.84 Deprecated Interface for Global MFE Prediction	
16.84 Deprecated Interface for Global MFE Prediction	640
	640 640
16.84.1 Detailed Description	640 640 641
16.84.1 Detailed Description	640 640 641 641
16.84.1 Detailed Description	640 640 641 641
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold()	640 640 641 641 642 642
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par()	640 640 641 641 642 642
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold()	640 640 641 641 642 642 643
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params()	640 640 641 641 642 643 643
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par()	640 640 641 642 642 643 643 644
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par() 16.84.2.7 export_cofold_arrays_gq()	640 640 641 642 643 643 643 644 644
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par() 16.84.2.7 export_cofold_arrays_gq() 16.84.2.8 export_cofold_arrays()	640 640 641 641 642 643 643 644 644 645
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par() 16.84.2.7 export_cofold_arrays_gq() 16.84.2.8 export_cofold_arrays() 16.84.2.9 get_monomere_mfes()	640 640 641 641 642 643 643 644 644 645 646
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par() 16.84.2.7 export_cofold_arrays_gq() 16.84.2.8 export_cofold_arrays() 16.84.2.9 get_monomere_mfes() 16.84.2.10 initialize_cofold()	640 640 641 642 643 643 643 644 644 645 646
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par() 16.84.2.7 export_cofold_arrays_gq() 16.84.2.8 export_cofold_arrays() 16.84.2.9 get_monomere_mfes() 16.84.2.10 initialize_cofold() 16.84.2.11 fold_par()	640 640 641 642 643 643 644 645 646 646 647
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par() 16.84.2.7 export_cofold_arrays_gq() 16.84.2.8 export_cofold_arrays() 16.84.2.9 get_monomere_mfes() 16.84.2.10 initialize_cofold() 16.84.2.11 fold_par() 16.84.2.12 fold()	640 640 641 642 643 643 644 644 645 646 646 647 648
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par() 16.84.2.7 export_cofold_arrays_gq() 16.84.2.8 export_cofold_arrays() 16.84.2.9 get_monomere_mfes() 16.84.2.10 initialize_cofold() 16.84.2.11 fold_par() 16.84.2.12 fold() 16.84.2.13 circfold()	640 640 641 641 642 643 643 644 645 646 646 647 648
16.84.1 Detailed Description 16.84.2 Function Documentation 16.84.2.1 alifold() 16.84.2.2 cofold() 16.84.2.3 cofold_par() 16.84.2.4 free_co_arrays() 16.84.2.5 update_cofold_params() 16.84.2.6 update_cofold_params_par() 16.84.2.7 export_cofold_arrays_gq() 16.84.2.8 export_cofold_arrays() 16.84.2.9 get_monomere_mfes() 16.84.2.10 initialize_cofold() 16.84.2.11 fold_par() 16.84.2.12 fold() 16.84.2.13 circfold() 16.84.2.14 free_arrays()	640 640 641 642 643 643 644 645 646 646 647 648 649

16.84.2.18 export_fold_arrays_par()	350
16.84.2.19 export_circfold_arrays()	350
16.84.2.20 export_circfold_arrays_par()	50
16.84.2.21 LoopEnergy()	51
16.84.2.22 HairpinE()	51
16.84.2.23 initialize_fold()	51
16.84.2.24 backtrack_fold_from_pair()	52
16.84.2.25 circalifold()	352
16.84.2.26 free_alifold_arrays()	352
16.85 Deprecated Interface for Local (Sliding Window) MFE Prediction	54
16.85.1 Detailed Description	54
16.85.2 Function Documentation	54
16.85.2.1 Lfold()	54
16.85.2.2 Lfoldz()	354
16.86 Deprecated Interface for Global Partition Function Computation	355
16.86.1 Detailed Description	555
16.86.2 Function Documentation	56
16.86.2.1 alipf_fold_par()	56
16.86.2.2 pf_fold_par()	57
16.86.2.3 pf_fold()	58
16.86.2.4 pf_circ_fold()	559
16.86.2.5 free_pf_arrays()	60
16.86.2.6 update_pf_params()	60
16.86.2.7 update_pf_params_par()	61
16.86.2.8 export_bppm()	
16.86.2.9 get_pf_arrays()	61
16.86.2.10 mean_bp_distance()	62
16.86.2.11 mean_bp_distance_pr()	62
16.86.2.12 stackProb()	63
16.86.2.13 init_pf_fold()	
16.86.2.14 co_pf_fold()	64
16.86.2.15 co_pf_fold_par()	64
16.86.2.16 compute_probabilities()	65
16.86.2.17 init_co_pf_fold()	
16.86.2.18 export_co_bppm()	
16.86.2.19 free_co_pf_arrays()	
16.86.2.20 update_co_pf_params()6	
16.86.2.21 update_co_pf_params_par()	
16.86.2.22 assign_plist_from_db()	
16.86.2.23 assign_plist_from_pr()	
16.86.2.24 alipf_fold()	
16.86.2.25 alipf_circ_fold()	69

16.86.2.26 export_ali_bppm()	369
16.86.2.27 free_alipf_arrays()	370
16.86.2.28 alipbacktrack()	370
16.86.2.29 get_alipf_arrays()	371
16.87 Deprecated Interface for Local (Sliding Window) Partition Function Computation	373
16.87.1 Detailed Description	373
16.87.2 Function Documentation	373
16.87.2.1 update_pf_paramsLP()	373
16.87.2.2 pfl_fold()	373
16.87.2.3 putoutpU_prob()	374
16.87.2.4 putoutpU_prob_bin()	375
16.88 Deprecated Interface for Stochastic Backtracking	376
16.88.1 Detailed Description	376
16.88.2 Function Documentation	376
16.88.2.1 pbacktrack()	376
16.88.2.2 pbacktrack_circ()	377
16.88.3 Variable Documentation	377
16.88.3.1 st_back	377
16.89 Deprecated Interface for Multiple Sequence Alignment Utilities	378
16.89.1 Detailed Description	378
16.89.2 Typedef Documentation	378
16.89.2.1 pair_info	378
16.89.3 Function Documentation	378
16.89.3.1 get_mpi()	378
16.89.3.2 encode_ali_sequence()	379
16.89.3.3 alloc_sequence_arrays()	379
16.89.3.4 free_sequence_arrays()	680
16.90 Deprecated Interface for Secondary Structure Utilities	381
16.90.1 Detailed Description	381
16.90.2 Function Documentation	382
16.90.2.1 b2HIT()	382
16.90.2.2 b2C()	683
16.90.2.3 b2Shapiro()	383
16.90.2.4 add_root()	
16.90.2.5 expand_Shapiro()	
16.90.2.6 expand_Full()	
16.90.2.7 unexpand_Full()	
16.90.2.8 unweight()	
16.90.2.9 unexpand_aligned_F()	
16.90.2.10 parse_structure()	
16.90.2.11 pack_structure()	
	687

	. 687
16.90.2.14 copy_pair_table()	. 688
16.90.2.15 alimake_pair_table()	. 688
16.90.2.16 make_pair_table_snoop()	. 689
16.90.2.17 bp_distance()	. 689
16.90.2.18 make_referenceBP_array()	. 689
16.90.2.19 compute_BPdifferences()	. 690
16.90.2.20 parenthesis_structure()	. 690
16.90.2.21 parenthesis_zuker()	. 691
16.90.2.22 bppm_to_structure()	. 691
16.90.2.23 bppm_symbol()	. 691
16.91 Deprecated Interface for Plotting Utilities	. 692
16.91.1 Detailed Description	. 692
16.91.2 Data Structure Documentation	. 692
16.91.2.1 struct COORDINATE	. 692
16.91.3 Function Documentation	. 692
16.91.3.1 PS_color_aln()	. 693
16.91.3.2 aliPS_color_aln()	. 693
16.91.3.3 simple_xy_coordinates()	. 693
16.91.3.4 simple_circplot_coordinates()	. 694
16.91.3.5 naview_xy_coordinates()	. 695
16.91.4 Variable Documentation	. 695
16.91.4 Variable Documentation	
	. 695
16.91.4.1 rna_plot_type	. 695 . 696
16.91.4.1 rna_plot_type	. 695 . 696 . 696
16.91.4.1 rna_plot_type	. 695 . 696 . 696 . 696
16.91.4.1 rna_plot_type	. 695 . 696 . 696 . 696 . 696
16.91.4.1 rna_plot_type	. 695 . 696 . 696 . 696 . 696
16.91.4.1 rna_plot_type . 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers	. 695 . 696 . 696 . 696 . 696 . 696
16.91.4.1 rna_plot_type	. 695 . 696 . 696 . 696 . 696 . 696
16.91.4.1 rna_plot_type . 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers	. 695 . 696 . 696 . 696 . 696 . 696
16.91.4.1 rna_plot_type . 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers . 16.92.1 Detailed Description	. 695 . 696 . 696 . 696 . 696 . 697 . 697
16.91.4.1 rna_plot_type . 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers 16.92.1 Detailed Description 16.92.2 Typedef Documentation . 16.92.2.1 path_t 16.92.3 Function Documentation . 16.92.3.1 find_saddle() . 16.92.3.2 free_path() . 16.92.3.3 get_path() . 17 Data Structure Documentation	. 695 . 696 . 696 . 696 . 696 . 697 . 697
16.91.4.1 rna_plot_type 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers 16.92.1 Detailed Description 16.92.2 Typedef Documentation 16.92.2.1 path_t 16.92.3 Function Documentation 16.92.3.1 find_saddle() 16.92.3.2 free_path() 16.92.3.3 get_path() 17 Data Structure Documentation 17.1_struct_en Struct Reference	. 695 . 696 . 696 . 696 . 696 . 697 . 697 . 699
16.91.4.1 rna_plot_type 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers 16.92.1 Detailed Description 16.92.2 Typedef Documentation 16.92.3.1 path_t 16.92.3 Function Documentation 16.92.3.1 find_saddle() 16.92.3.2 free_path() 16.92.3.3 get_path() 17.1 Data Structure Documentation 17.1 _struct_en Struct Reference 17.1.1 Detailed Description	. 695 . 696 . 696 . 696 . 696 . 697 . 697 . 699 . 699
16.91.4.1 rna_plot_type 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers 16.92.1 Detailed Description 16.92.2 Typedef Documentation 16.92.3.1 path_t 16.92.3 Function Documentation 16.92.3.1 find_saddle() 16.92.3.2 free_path() 16.92.3.3 get_path() 17.1 Data Structure Documentation 17.1_struct_en Struct Reference 17.1.1 Detailed Description 17.2 LIST Struct Reference	. 695 . 696 . 696 . 696 . 696 . 697 . 697 . 699 . 699 . 699
16.91.4.1 rna_plot_type . 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers 16.92.1 Detailed Description 16.92.2 Typedef Documentation . 16.92.2.1 path_t . 16.92.3 Function Documentation . 16.92.3.1 find_saddle() . 16.92.3.2 free_path() . 16.92.3.3 get_path() 17.1 Data Structure Documentation 17.1_struct_en Struct Reference . 17.1.1 Detailed Description 17.2 LIST Struct Reference . 17.3 LST_BUCKET Struct Reference	. 695 . 696 . 696 . 696 . 696 . 697 . 699 . 699 . 699 . 699
16.91.4.1 rna_plot_type . 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers . 16.92.1 Detailed Description . 16.92.2 Typedef Documentation . 16.92.3.1 path_t . 16.92.3 Function Documentation . 16.92.3.1 find_saddle() . 16.92.3.2 free_path() . 16.92.3.3 get_path() . 17 Data Structure Documentation . 17.1 _struct_en Struct Reference . 17.1.1 Detailed Description . 17.2 LIST Struct Reference . 17.3 LST_BUCKET Struct Reference . 17.4 Postorder_list Struct Reference .	. 695 . 696 . 696 . 696 . 696 . 697 . 699 . 699 . 699 . 700
16.91.4.1 rna_plot_type 16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers 16.92.1 Detailed Description 16.92.2 Typedef Documentation 16.92.3.1 path_t 16.92.3.1 find_saddle() 16.92.3.2 free_path() 16.92.3.3 get_path() 17 Data Structure Documentation 17.1 _struct_en Struct Reference 17.1.1 Detailed Description 17.2 LIST Struct Reference 17.3 LST_BUCKET Struct Reference 17.4 Postorder_list Struct Reference 17.4.1 Detailed Description	. 695 . 696 . 696 . 696 . 696 . 697 . 697 . 699 . 699 . 699 . 700 . 700

	17.6.1 Detailed Description	700
	17.7 TwoDpfold_vars Struct Reference	700
	17.7.1 Detailed Description	701
	17.8 vrna_dimer_conc_s Struct Reference	701
	17.8.1 Detailed Description	702
	17.9 vrna_hc_bp_storage_t Struct Reference	702
	17.9.1 Detailed Description	702
	17.10 vrna_sc_bp_storage_t Struct Reference	702
	17.10.1 Detailed Description	702
	17.11 vrna_sc_motif_s Struct Reference	703
	17.12 vrna_structured_domains_s Struct Reference	703
	17.13 vrna_subopt_sol_s Struct Reference	703
	17.13.1 Detailed Description	703
	17.14 vrna_unstructured_domain_motif_s Struct Reference	703
18	File Documentation	705
	18.1 ViennaRNA/2Dfold.h File Reference	
	18.1.1 Detailed Description	
	18.2 ViennaRNA/2Dpfold.h File Reference	
	18.2.1 Detailed Description	706
	18.2.2 Function Documentation	
	18.2.2.1 get_TwoDpfold_variables()	707
	18.2.2.2 destroy_TwoDpfold_variables()	707
	18.2.2.3 TwoDpfoldList()	
	18.2.2.4 TwoDpfold_pbacktrack()	708
	18.2.2.5 TwoDpfold_pbacktrack5()	709
	18.3 ViennaRNA/alifold.h File Reference	710
	18.3.1 Detailed Description	711
	18.3.2 Function Documentation	711
	18.3.2.1 energy_of_alistruct()	711
	18.3.2.2 update_alifold_params()	712
	18.3.3 Variable Documentation	712
	18.3.3.1 cv_fact	712
	18.3.3.2 nc_fact	712
	18.4 ViennaRNA/aln_util.h File Reference	713
	18.4.1 Detailed Description	713
	18.5 ViennaRNA/alphabet.h File Reference	713
	18.5.1 Detailed Description	713
	18.6 ViennaRNA/boltzmann_sampling.h File Reference	713
	18.6.1 Detailed Description	715
	18.7 ViennaRNA/centroid.h File Reference	715
	18.7.1 Detailed Description	715

18.7.2 Function Documentation
18.7.2.1 get_centroid_struct_pl()
18.7.2.2 get_centroid_struct_pr()
18.8 ViennaRNA/char_stream.h File Reference
18.8.1 Detailed Description
18.9 ViennaRNA/datastructures/char_stream.h File Reference
18.9.1 Detailed Description
18.10 ViennaRNA/cofold.h File Reference
18.10.1 Detailed Description
18.11 ViennaRNA/combinatorics.h File Reference
18.11.1 Detailed Description
18.12 ViennaRNA/commands.h File Reference
18.12.1 Detailed Description
18.13 ViennaRNA/concentrations.h File Reference
18.13.1 Detailed Description
18.13.2 Function Documentation
18.13.2.1 get_concentrations()
18.14 ViennaRNA/constraints.h File Reference
18.14.1 Detailed Description
18.15 ViennaRNA/constraints/hard.h File Reference
18.15.1 Detailed Description
18.15.2 Macro Definition Documentation
18.15.2.1 VRNA_CONSTRAINT_NO_HEADER
18.15.2.2 VRNA_CONSTRAINT_DB_ANG_BRACK
18.15.3 Enumeration Type Documentation
······································
18.15.3.1 vrna_hc_type_e
18.15.3.1 vrna_hc_type_e
18.15.3.1 vrna_hc_type_e
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 724
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 724 18.15.4.2 print_tty_constraint() 725
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 724 18.15.4.2 print_tty_constraint() 729 18.15.4.3 print_tty_constraint_full() 729
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 724 18.15.4.2 print_tty_constraint() 725 18.15.4.3 print_tty_constraint_full() 725 18.15.4.4 constrain_ptypes() 725
18.15.3.1 vrna_hc_type_e 72-4 18.15.4 Function Documentation 72-4 18.15.4.1 vrna_hc_add_data() 72-4 18.15.4.2 print_tty_constraint() 72-4 18.15.4.3 print_tty_constraint_full() 72-4 18.15.4.4 constrain_ptypes() 72-4 18.16 ViennaRNA/constraints/ligand.h File Reference 72-4
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 724 18.15.4.2 print_tty_constraint() 725 18.15.4.3 print_tty_constraint_full() 725 18.15.4.4 constrain_ptypes() 725 18.16 ViennaRNA/constraints/ligand.h File Reference 726 18.16.1 Detailed Description 726
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 724 18.15.4.2 print_tty_constraint() 725 18.15.4.3 print_tty_constraint_full() 725 18.15.4.4 constrain_ptypes() 725 18.16 ViennaRNA/constraints/ligand.h File Reference 726 18.16.1 Detailed Description 726 18.16.2 Typedef Documentation 726
18.15.3.1 vrna_hc_type_e 72- 18.15.4 Function Documentation 72- 18.15.4.1 vrna_hc_add_data() 72- 18.15.4.2 print_tty_constraint() 72- 18.15.4.3 print_tty_constraint_full() 72- 18.15.4.4 constrain_ptypes() 72- 18.16 ViennaRNA/constraints/ligand.h File Reference 72- 18.16.1 Detailed Description 72- 18.16.2 Typedef Documentation 72- 18.16.2.1 vrna_sc_motif_t 72-
18.15.3.1 vrna_hc_type_e 72-18.15.4 Function Documentation 72-18.15.4 Function Documentation 72-18.15.4.1 vrna_hc_add_data() 72-18.15.4.2 print_tty_constraint() 72-18.15.4.2 print_tty_constraint_full() 72-18.15.4.3 print_tty_constraint_full() 72-18.15.4.4 constrain_ptypes() 72-18.16.15.4.4 constraints/ligand.h File Reference 72-18.16.10 Detailed Description 72-18.16.2 Typedef Documentation 72-18.16.2 Typede
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 724 18.15.4.2 print_tty_constraint() 725 18.15.4.3 print_tty_constraint_full() 725 18.15.4.4 constrain_ptypes() 725 18.16 ViennaRNA/constraints/ligand.h File Reference 726 18.16.1 Detailed Description 726 18.16.2 Typedef Documentation 726 18.17 ViennaRNA/constraints/SHAPE.h File Reference 727 18.17 Updailed Description 726 18.17.1 Detailed Description 727
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 724 18.15.4.2 print_tty_constraint() 725 18.15.4.3 print_tty_constraint_full() 725 18.15.4.4 constrain_ptypes() 725 18.16 ViennaRNA/constraints/ligand.h File Reference 726 18.16.1 Detailed Description 726 18.16.2 Typedef Documentation 726 18.17 ViennaRNA/constraints/SHAPE.h File Reference 727 18.17.1 Detailed Description 726 18.17.2 Function Documentation 726
18.15.3.1 vrna_hc_type_e 724 18.15.4 Function Documentation 724 18.15.4.1 vrna_hc_add_data() 725 18.15.4.2 print_tty_constraint() 725 18.15.4.3 print_tty_constraint_full() 725 18.15.4.4 constrain_ptypes() 725 18.16 ViennaRNA/constraints/ligand.h File Reference 726 18.16.1 Detailed Description 726 18.16.2 Typedef Documentation 726 18.17 ViennaRNA/constraints/SHAPE.h File Reference 727 18.17 ViennaRNA/constraints/SHAPE.h File Reference 727 18.17.1 Detailed Description 726 18.17.2 Function Documentation 726 18.17.2.1 vrna_sc_SHAPE_parse_method() 726

18.18.2.1 vrna_sc_type_e	29
18.19 ViennaRNA/constraints_hard.h File Reference	30
18.19.1 Detailed Description	30
18.20 ViennaRNA/constraints_ligand.h File Reference	30
18.20.1 Detailed Description	30
18.21 ViennaRNA/constraints_SHAPE.h File Reference	30
18.21.1 Detailed Description	30
18.22 ViennaRNA/constraints_soft.h File Reference	30
18.22.1 Detailed Description	31
18.23 ViennaRNA/convert_epars.h File Reference	31
18.23.1 Detailed Description	31
18.24 ViennaRNA/data_structures.h File Reference	31
18.24.1 Detailed Description	31
18.25 ViennaRNA/datastructures/hash_tables.h File Reference	31
18.25.1 Detailed Description	32
18.26 ViennaRNA/datastructures/heap.h File Reference	32
18.26.1 Detailed Description	33
18.27 ViennaRNA/dist_vars.h File Reference	33
18.27.1 Detailed Description	34
18.27.2 Variable Documentation	34
18.27.2.1 edit_backtrack	34
18.27.2.2 cost_matrix	34
18.28 ViennaRNA/dp_matrices.h File Reference	34
18.28.1 Detailed Description	35
18.29 ViennaRNA/duplex.h File Reference	35
18.29.1 Detailed Description	35
18.30 ViennaRNA/edit_cost.h File Reference	35
18.30.1 Detailed Description	35
18.31 ViennaRNA/energy_const.h File Reference	36
18.31.1 Detailed Description	36
18.32 ViennaRNA/energy_par.h File Reference	36
18.32.1 Detailed Description	36
18.33 ViennaRNA/equilibrium_probs.h File Reference	36
18.33.1 Detailed Description	37
18.33.2 Function Documentation	37
18.33.2.1 vrna_pr_energy()	37
18.34 ViennaRNA/eval.h File Reference	37
18.34.1 Detailed Description	40
18.35 ViennaRNA/exterior_loops.h File Reference	40
18.35.1 Detailed Description	40
18.36 ViennaRNA/file_formats.h File Reference	40
18.36.1 Detailed Description	40

18.37 ViennaRNA/io/file_formats.h File Reference	741
18.37.1 Detailed Description	741
18.38 ViennaRNA/file_formats_msa.h File Reference	741
18.38.1 Detailed Description	742
18.39 ViennaRNA/io/file_formats_msa.h File Reference	742
18.39.1 Detailed Description	743
18.40 ViennaRNA/file_utils.h File Reference	743
18.40.1 Detailed Description	743
18.41 ViennaRNA/findpath.h File Reference	743
18.41.1 Detailed Description	743
18.42 ViennaRNA/landscape/findpath.h File Reference	743
18.42.1 Detailed Description	744
18.43 ViennaRNA/fold.h File Reference	744
18.43.1 Detailed Description	745
18.44 ViennaRNA/fold_compound.h File Reference	745
18.44.1 Detailed Description	746
18.45 ViennaRNA/fold_vars.h File Reference	746
18.45.1 Detailed Description	747
18.45.2 Variable Documentation	747
18.45.2.1 RibosumFile	747
18.45.2.2 james_rule	747
18.45.2.3 logML	747
18.45.2.4 cut_point	747
18.45.2.5 base_pair	747
18.45.2.6 pr	748
18.45.2.7 iindx	748
18.46 ViennaRNA/gquad.h File Reference	748
18.46.1 Detailed Description	748
18.47 ViennaRNA/grammar.h File Reference	748
18.47.1 Detailed Description	749
18.48 ViennaRNA/hairpin_loops.h File Reference	749
18.48.1 Detailed Description	749
18.49 ViennaRNA/interior_loops.h File Reference	749
18.49.1 Detailed Description	749
18.50 ViennaRNA/inverse.h File Reference	749
18.50.1 Detailed Description	750
18.51 ViennaRNA/landscape/move.h File Reference	750
18.51.1 Detailed Description	751
18.52 ViennaRNA/landscape/paths.h File Reference	751
18.52.1 Detailed Description	752
18.53 ViennaRNA/Lfold.h File Reference	752
18 53 1 Detailed Description	752

18.54 ViennaRNA/loop_energies.h File Reference
18.54.1 Detailed Description
18.55 ViennaRNA/loops/all.h File Reference
18.55.1 Detailed Description
18.56 ViennaRNA/loops/external.h File Reference
18.56.1 Detailed Description
18.57 ViennaRNA/loops/hairpin.h File Reference
18.57.1 Detailed Description
18.58 ViennaRNA/loops/internal.h File Reference
18.58.1 Detailed Description
18.59 ViennaRNA/loops/multibranch.h File Reference
18.59.1 Detailed Description
18.60 ViennaRNA/LPfold.h File Reference
18.60.1 Detailed Description
18.60.2 Function Documentation
18.60.2.1 init_pf_foldLP()
18.61 ViennaRNA/MEA.h File Reference
18.61.1 Detailed Description
18.62 ViennaRNA/mfe.h File Reference
18.62.1 Detailed Description
18.63 ViennaRNA/mfe_window.h File Reference
18.63.1 Detailed Description
18.64 ViennaRNA/mm.h File Reference
18.64.1 Detailed Description
18.64.2 Function Documentation
18.64.2.1 vrna_maximum_matching()
18.64.2.2 vrna_maximum_matching_simple()
18.65 ViennaRNA/model.h File Reference
18.65.1 Detailed Description
18.66 ViennaRNA/multibranch_loops.h File Reference
18.66.1 Detailed Description
18.67 ViennaRNA/naview.h File Reference
18.67.1 Detailed Description
18.68 ViennaRNA/plotting/naview.h File Reference
18.68.1 Detailed Description
18.69 ViennaRNA/neighbor.h File Reference
18.69.1 Detailed Description
18.70 ViennaRNA/landscape/neighbor.h File Reference
18.70.1 Detailed Description
18.71 ViennaRNA/params.h File Reference
18.71.1 Detailed Description
18.72 ViennaRNA/params/1.8.4_epars.h File Reference

18.72.1 Detailed Description
18.73 ViennaRNA/params/1.8.4_intloops.h File Reference
18.73.1 Detailed Description
18.74 ViennaRNA/params/basic.h File Reference
18.74.1 Detailed Description
18.75 ViennaRNA/constraints/basic.h File Reference
18.75.1 Detailed Description
18.76 ViennaRNA/utils/basic.h File Reference
18.76.1 Detailed Description
18.76.2 Function Documentation
18.76.2.1 get_line()
18.76.2.2 print_tty_input_seq()
18.76.2.3 print_tty_input_seq_str()
18.76.2.4 warn_user()
18.76.2.5 nrerror()
18.76.2.6 space()
18.76.2.7 xrealloc()
18.76.2.8 init_rand()
18.76.2.9 urn()
18.76.2.10 int_urn()
18.76.2.11 filecopy()
18.76.2.12 time_stamp()
18.77 ViennaRNA/datastructures/basic.h File Reference
18.77.1 Detailed Description
18.78 ViennaRNA/params/constants.h File Reference
18.78.1 Detailed Description
18.78.2 Macro Definition Documentation
18.78.2.1 GASCONST
18.78.2.2 K0
18.78.2.3 INF
18.78.2.4 FORBIDDEN
18.78.2.5 BONUS
18.78.2.6 NBPAIRS
18.78.2.7 TURN
18.78.2.8 MAXLOOP
18.79 ViennaRNA/params/convert.h File Reference
18.79.1 Detailed Description
18.80 ViennaRNA/params/io.h File Reference
18.80.1 Detailed Description
18.81 ViennaRNA/part_func.h File Reference
18.81.1 Detailed Description
18.81.2 Function Documentation

18.81.2.1 centroid()	34
18.81.2.2 get_centroid_struct_gquad_pr()	34
18.81.2.3 mean_bp_dist()	34
18.81.2.4 expLoopEnergy()	35
18.81.2.5 expHairpinEnergy()	35
18.82 ViennaRNA/part_func_co.h File Reference	35
18.82.1 Detailed Description	36
18.82.2 Function Documentation	36
18.82.2.1 get_plist()	36
18.83 ViennaRNA/part_func_up.h File Reference	36
18.83.1 Detailed Description	37
18.84 ViennaRNA/part_func_window.h File Reference	37
18.84.1 Detailed Description	38
18.85 ViennaRNA/perturbation_fold.h File Reference	38
18.85.1 Detailed Description	39
18.86 ViennaRNA/plot_aln.h File Reference	39
18.86.1 Detailed Description	39
18.87 ViennaRNA/plot_layouts.h File Reference	39
18.87.1 Detailed Description	39
18.88 ViennaRNA/plot_structure.h File Reference	90
18.88.1 Detailed Description	90
18.89 ViennaRNA/plot_utils.h File Reference	90
18.89.1 Detailed Description	90
18.90 ViennaRNA/plotting/alignments.h File Reference	90
18.90.1 Detailed Description)1
18.91 ViennaRNA/utils/alignments.h File Reference)1
18.91.1 Detailed Description)2
18.92 ViennaRNA/plotting/layouts.h File Reference)2
18.92.1 Detailed Description)4
18.93 ViennaRNA/plotting/probabilities.h File Reference)4
18.93.1 Detailed Description)4
18.94 ViennaRNA/plotting/RNApuzzler/RNApuzzler.h File Reference)4
18.94.1 Detailed Description) 5
18.95 ViennaRNA/plotting/RNApuzzler/RNAturtle.h File Reference) 5
18.95.1 Detailed Description) 5
18.96 ViennaRNA/plotting/structures.h File Reference) 5
18.96.1 Detailed Description	96
18.97 ViennaRNA/utils/structures.h File Reference	96
18.97.1 Detailed Description	9
18.98 ViennaRNA/profiledist.h File Reference	9
18.98.1 Function Documentation)0
18.98.1.1 profile_edit_distance())0

18.98.1.2 Make_bp_profile_bppm()
18.98.1.3 free_profile()
18.98.1.4 Make_bp_profile()
18.99 ViennaRNA/PS_dot.h File Reference
18.99.1 Detailed Description
18.100 ViennaRNA/read_epars.h File Reference
18.100.1 Detailed Description
18.101 ViennaRNA/ribo.h File Reference
18.101.1 Detailed Description
18.102 ViennaRNA/RNAstruct.h File Reference
18.102.1 Detailed Description
18.103 ViennaRNA/search/BoyerMoore.h File Reference
18.103.1 Detailed Description
18.104 ViennaRNA/sequence.h File Reference
18.104.1 Detailed Description
18.105 ViennaRNA/stream_output.h File Reference
18.105.1 Detailed Description
18.106 ViennaRNA/datastructures/stream_output.h File Reference
18.106.1 Detailed Description
18.107 ViennaRNA/string_utils.h File Reference
18.107.1 Detailed Description
18.108 ViennaRNA/stringdist.h File Reference
18.108.1 Detailed Description
18.108.2 Function Documentation
18.108.2.1 Make_swString()
18.108.2.2 string_edit_distance()
18.109 ViennaRNA/structure_utils.h File Reference
18.109.1 Detailed Description
18.110 ViennaRNA/structured_domains.h File Reference
18.110.1 Detailed Description
18.111 ViennaRNA/subopt.h File Reference
18.111.1 Detailed Description
18.111.2 Typedef Documentation
18.111.2.1 SOLUTION
18.112 ViennaRNA/svm_utils.h File Reference
18.112.1 Detailed Description
18.113 ViennaRNA/treedist.h File Reference
18.113.1 Detailed Description
18.113.2 Function Documentation
18.113.2.1 make_tree()
18.113.2.2 tree_edit_distance()
18.113.2.3 free_tree()

823

18.114 ViennaRNA/units.h File Reference	811
18.114.1 Detailed Description	811
18.115 ViennaRNA/unstructured_domains.h File Reference	811
18.115.1 Detailed Description	813
18.115.2 Function Documentation	813
18.115.2.1 vrna_ud_get_motif_size_at()	813
18.115.2.2 vrna_ud_set_prob_cb()	814
18.116 ViennaRNA/utils.h File Reference	814
18.116.1 Detailed Description	814
18.117 ViennaRNA/io/utils.h File Reference	814
18.117.1 Detailed Description	815
18.118 ViennaRNA/plotting/utils.h File Reference	815
18.118.1 Detailed Description	815
18.119 ViennaRNA/utils/strings.h File Reference	815
18.119.1 Detailed Description	816
18.119.2 Function Documentation	816
18.119.2.1 str_uppercase()	817
18.119.2.2 str_DNA2RNA()	817
18.119.2.3 random_string()	817
18.119.2.4 hamming()	817
18.119.2.5 hamming_bound()	818
18.120 ViennaRNA/walk.h File Reference	818
18.120.1 Detailed Description	818
18.121 ViennaRNA/landscape/walk.h File Reference	818
18.121.1 Detailed Description	819
Bibliography	822

Index

Chapter 1

Main Page

1.1 A Library for predicting and comparing RNA secondary structures

The core of the ViennaRNA Package ([13], [11]) is formed by a collection of routines for the prediction and comparison of RNA secondary structures. These routines can be accessed through stand-alone programs, such as RNAfold, RNAdistance etc., which should be sufficient for most users. For those who wish to develop their own programs we provide a library which can be

linked to your own code.

This document describes the library and will be primarily useful to programmers. However, it also contains details about the implementation that may be of interest to advanced users. The stand-alone programs are described in separate man pages. The latest version of the package including source code and html versions of the documentation can be found at

```
http://www.tbi.univie.ac.at/RNA
```

Date

1994-2019

Authors

Ivo Hofacker, Peter Stadler, Ronny Lorenz, and so many more

1.2 License

Disclaimer and Copyright

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Permission is granted for research, educational, and commercial use and modification so long as 1) the package and any derived works are not redistributed for any fee, other than media costs, 2) proper credit is given to the authors and the Institute for Theoretical Chemistry of the University of Vienna.

If you want to include this software in a commercial product, please contact the authors.

2 Main Page

1.3 Contributors

Over the past decades since the <code>ViennaRNA Package</code> first sprang to life as part of Ivo Hofackers PhD project, several different authors contributed more and more algorithm implementations. In 2008, Ronny Lorenz took over the extensive task to harmonize and simplify the already existing implementations for the sake of easier feature addition. This eventually lead to version 2.0 of the <code>ViennaRNA Package</code>. Since then, he (re-)implemented a large portion of the currently existing library features, such as the new, generalized constraints framework, RNA folding grammar domain extensions, and the major part of the scripting language interface. Below is a list of most people who contributed larger parts of the implementations:

- · Juraj Michalik (non-redundant Boltzmann sampling)
- Gregor Entzian (neighbor, walk)
- · Mario Koestl (worked on SWIG interface and related unit testing)
- · Dominik Luntzer (pertubation fold)
- Stefan Badelt (cofold evaluation, RNAdesign.pl, cofold findpath extensions)
- Stefan Hammer (parts of SWIG interface and corresponding unit tests)
- · Ronny Lorenz (circfold, version 2.0, generic constraints, grammar extensions, and much more)
- Hakim Tafer (RNAplex, RNAsnoop)
- Ulrike Mueckstein (RNAup)
- Stephan Bernhart (cofold, plfold, unpaired probabilities, alifold, and so many more)
- Ivo Hofacker, Peter Stadler, and Christoph Flamm (almost every implementation up to version 1.8.5)

We also want to thank the following people:

- Sebastian Bonhoeffer's implementation of partition function folding served as a precursor to our part_func.c
- Manfred Tacker hacked constrained folding into fold.c for the first time
- · Martin Fekete made the first attempts at "alignment folding"
- Andrea Tanzer and Martin Raden (Mann) for not stopping to report bugs found through comprehensive usage of our applications and RNAlib
- Thanks also to everyone else who helped testing and finding bugs, especially Christoph Flamm, Martijn Huynen, Baerbel Krakhofer, and many more

If you want to get involved in the development of the ViennaRNA Package yourself, please read the Contributing page.

Chapter 2

Getting Started

- · Installation and Configuration describes how to install and configure RNAlib for your requirements
- · HelloWorld presents some small example programs to get a first impression on how to use this library
- HelloWorld (Perl/Python) contains small examples that show how to use RNAlib even without C/C++ programming skills from within your favorite scripting language

2.1 Installation and Configuration

A documentation on how to configure the different features of RNAlib, how to install the ViennaRNA Package, and finally, how to link you own programs against RNAlib.

2.1.1 Installing the ViennaRNA Package

For best portability the ViennaRNA package uses the GNU autoconf and automake tools. The instructions below are for installing the ViennaRNA package from source. However, pre-compiled binaries for various Linux distributions, as well as for Windows users are available from Download section of the main ViennaRNA homepage.

2.1.1.1 Quick-start

Usually you'll just unpack, configure and make. To do this type:

```
tar -zxvf ViennaRNA-2.4.14.tar.gz
cd ViennaRNA-2.4.14
./configure
make
sudo make install
```

2.1.1.2 Installation without root privileges

If you do not have root privileges on your computer, you might want to install the ViennaRNA Package to a location where you actually have write access to. To do so, you can set the installation prefix of the ./configure script like so:

```
./configure --prefix=/home/username/ViennaRNA
```

This will install the entire ViennaRNA Package into a new directory ViennaRNA directly into the users username home directory.

4 Getting Started

2.1.1.3 Notes for MacOS X users

Although users will find /usr/bin/gcc and /usr/bin/g++ executables in their directory tree, these programs are not at all what they pretend to be. Instead of including the GNU programs, Apple decided to install clang/llvm in disguise. Unfortunately, the default version of clang/llvm does not support OpenMP (yet), but only complains at a late stage of the build process when this support is required. Therefore, it seems necessary to deactivate OpenMP support by passing the option –disable-openmp to the ./configure script.

Additionally, since MacOS X 10.5 the perl and python installation distributed with MacOS X always include so called universal-binaries (a.k.a. fat-binaries), i.e. binaries for multiple architecture types. In order to compile and link the programs, library, and scripting language interfaces of the ViennaRNA Package for multiple architectures, we've added a new configure switch that sets up the required changes automatically:

```
./configure --enable-universal-binary
```

Note

Note, that with link time optimization turned on, MacOS X's default compiler (llvm/clang) generates an intermediary binary format that can not easily be combined into a multi-architecture library. Therefore, the –enable-universal-binary switch turns off link time optimization!

2.1.2 Configuring RNAlib features

The ViennaRNA Package includes additional executable programs such as RNAforester, Kinfold, and Kinwalker. Furthermore, we include several features in our C-library that may be activated by default, or have to be explicitly turned on at configure-time. Below we list a selection of the available configure options that affect the features included in all executable programs, the RNAlib C-library, and the corresponding scripting language interface(s).

2.1.2.1 Streaming SIMD Extension (SSE) support

Since version 2.3.5 our sources contain code that implements a faster multibranch loop decomposition in global MFE predictions, as used e.g. in RNAfold. This implementation makes use of modern processors capability to execute particular instructions on multiple data simultaneously (SIMD - single instruction multiple data, thanks to W. B. Langdon for providing the modified code). Consequently, the time required to assess the minimum of all multibranch loop decompositions is reduced up to about one half compared to the runtime of the original implementation. This feature is enabled by default since version 2.4.11 and a dispatcher ensures that the correct implementation will be selected at runtime. If for any reason you want to disable this feature at compile-time use the following configure flag:

```
./configure --disable-simd
```

2.1.2.2 Scripting Interfaces

The ViennaRNA Package comes with scripting language interfaces for Perl 5, Python 2, and Python 3 (provided by swig), that allow one to use the implemented algorithms directly without the need of calling an executable program. The interfaces are build by default whenever the autoconf tool-chain detects the required build tools on your system. You may, however, explicitly turn off particular scripting language interface support at configure-time, for instance for Perl 5 and Python 2, before the actual installation.

Example:

```
./configure --without-perl --without-python
```

Disabling the scripting language support all-together can be accomplished using the following switch:

```
./ {\tt configure} \ -\! {\tt without-swig}
```

2.1.2.3 Cluster Analysis

The programs AnalyseSeqs and AnalyseDists offer some cluster analysis tools (split decomposition, statistical geometry, neighbor joining, Ward's method) for sequences and distance data. To also build these programs add

```
--with-cluster
```

to your configure options.

2.1.2.4 Kinfold

The Kinfold program can be used to simulate the folding dynamics of an RNA molecule, and is compiled by default. Use the

```
--without-kinfold
```

option to skip compilation and installation of Kinfold.

2.1.2.5 RNAforester

The RNAforester program is used for comparing secondary structures using tree alignment. Similar to Kinfold, use the

```
--without-forester
```

option to skip compilation and installation of RNAforester.

2.1.2.6 Kinwalker

The Kinwalker algorithm performs co-transcriptional folding of RNAs, starting at a user specified structure (default
∴ open chain) and ending at the minimum free energy structure. Compilation and installation of this program is deactivated by default. Use the

```
--with-kinwalker
```

option to enable building and installation of Kinwalker.

2.1.2.7 Link Time Optimization (LTO)

To increase the performance of our implementations, the ViennaRNA Package tries to make use of the Link Time Optimization (LTO) feature of modern C-compilers. If you are experiencing any troubles at make-time or run-time, or the configure script for some reason detects that your compiler supports this feature although it doesn't, you can deactivate it using the flag

```
./configure --disable-lto
```

Note, that GCC before version 5 is known to produce unreliable LTO code, especially in combination with SSE (see config_sse). We therefore recommend using a more recent compiler (GCC 5 or above) or to turn off one of the two features, LTO or SSE optimized code.

6 Getting Started

2.1.2.8 OpenMP support

To enable concurrent computation of our implementations and in some cases parallelization of the algorithms we make use of the OpenMP API. This interface is well understood by most modern compilers. However, in some cases it might be necessary to deactivate OpenMP support and therefore transform *RNAlib* into a C-library that is not entirely *thread-safe*. To do so, add the following configure option

```
./configure --disable-openmp
```

2.1.2.9 POSIX threads (pthread) support

To enable concurrent computation of multiple input data in RNAfold, and for our implementation of the concurrent unordered insert, ordered output flush data structure vrna_ostream_t we make use of POSIX threads. This should be supported on all modern platforms and usually does not pose any problems. Unfortunately, we use a threadpool implementation that is not compatible with Microsoft Windows yet. Thus, POSIX thread support can not be activated for Windows builds until we have fixed this problem. If you want to compile RNAfold and RNAlib without POSIX threads support for any other reasons, add the following configure option

```
./configure --disable-pthreads
```

2.1.2.10 Stochastic backtracking using Boustrophedon scheme

Stochastic backtracking for single RNA sequences, e.g. available through the RNAsubopt program, received a major speedup by implementing a Boustrophedon scheme (see this article for details). If for some reason you want to deactivate this feature, you can do that by adding the following switch to the configure script:

```
./configure --disable-boustrophedon
```

2.1.2.11 SVM Z-score filter in RNALfold

By default, RNALfold that comes with the ViennaRNA Package allows for z-score filtering of its predicted results using a support vector machine (SVM). However, the library we use to implement this feature (libsvm) is statically linked to our own RNAlib. If this introduces any problems for your own third-party programs that link against RNAlib, you can safely switch off the z-scoring implementation using

```
./configure --without-svm
```

2.1.2.12 GNU Scientific Library

The new program RNApvmin computes a pseudo-energy perturbation vector that aims to minimize the discrepancy of predicted, and observed pairing probabilities. For that purpose it implements several methods to solve the optimization problem. Many of them are provided by the GNU Scientific Library, which is why the RNApvmin program, and the RNAlib C-library are required to be linked against libgsl. If this introduces any problems in your own third-party programs that link against RNAlib, you can turn off a larger portion of available minimizers in RN← Apvmin and linking against libgsl all-together, using the switch

```
./ {\tt configure} \ -\!- {\tt without-gsl}
```

2.1.2.13 Disable C11/C++11 feature support

By default, we use C11/C++11 features in our implementations. This mainly accounts for unnamed unions/structs within *RNAlib*. The configure script automatically detects whether or not your compiler understands these features. In case you are using an older compiler, these features will be deactivated by setting a specific pre-processor directive. If for some reason you want to deactivate C11/C++11 features despite the capabilities of your compiler, use the following configure option:

```
./configure --disable-c11
```

2.1.2.14 Enable warnings for use of deprecated symbols

Since version 2.2 we are in the process of transforming the API of our *RNAlib*. Hence, several symbols are marked as *deprecated* whenever they have been replaced by the new API. By default, deprecation warnings at compile time are deactivated. If you want to get your terminal spammed by tons of deprecation warnings, enable them using:

```
./configure --enable-warn-deprecated
```

2.1.2.15 Single precision partition function

Calculation of partition functions (via RNAfold -p) uses double precision floats by default, to avoid overflow errors on longer sequences. If your machine has little memory and you don't plan to fold sequences over 1000 bases in length you can compile the package to do the computations in single precision by running

```
./configure --enable-floatpf
```

Note

Using this option is discouraged and not necessary on most modern computers.

2.1.2.16 Help

For a complete list of all ./configure options and important environment variables, type

```
./configure --help
```

For more general information on the build process see the INSTALL file.

2.1.3 Linking against RNAlib

In order to use our implemented algorithms you simply need to link your program to our *RNAlib* C-library that usually comes along with the ViennaRNA Package installation. If you've installed the ViennaRNA Package as a pre-build binary package, you probably need the corresponding development package, e.g. *viennarna-devel*, or *viennarna-dev*. The only thing that is left is to include the ViennaRNA header files into your source code, e.g.:

```
#include <ViennaRNA/mfe.h>
```

and start using our fast and efficient algorithm implementations.

See also

In the mp_example and Some Examples using RNAlib API v3.0 sections, we list a small set of example code that usually is a good starting point for your application.

8 Getting Started

2.1.3.1 Compiler and Linker flags

Of course, simply adding the ViennaRNA header files into your source code is usually not enough. You probably need to tell your compiler where to find the header files, and sometimes add additional pre-processor directives. Whenever your installation of *RNAlib* was build with default settings and the header files were installed into their default location, a simple

```
-I/usr/include
```

pre-processor/compile flag should suffice. It can even be omitted in this case, since your compiler should search this directory by default anyway. You only need to change the path from /usr/include to the correct location whenever the header files have been installed into a non-standard directory.

On the other hand, if you've compiled *RNAlib* with some non-default settings then you probably need to define some additional pre-processor macros:

• VRNA_DISABLE_C11_FEATURES . . . Disable C11/C++11 features.

Warning

Add this directive to your pre-processor/compile flags only if *RNAlib* was build with the *-disable-c11* configure option.

See also

Disable C11/C++11 feature support and vrna_C11_features()

• VRNA WARN DEPRECATED ... Enable warnings for using deprecated symbols.

Note

Adding this directive enables compiler warnings whenever you use symbols in *RNAlib* that are marked *deprecated*.

See also

Enable warnings for use of deprecated symbols and Deprecated List

• *USE_FLOAT_PF* ... Use single precision floating point operations instead of double precision in partition function computations.

Warning

Define this macro only if RNAlib was build with the -enable-floatpf configure option!

See also

Single precision partition function

Simply add the corresponding definition(s) to your pre-processor/compile flags, for instance:

```
-DVRNA_DISABLE_C11_FEATURES
```

Finally, linking against RNAlib is achieved by adding the following linker flag

```
-L/usr/lib -lRNA -fopenmp
```

Again, the path to the library, /usr/lib, may be omitted if this path is searched for libraries by default. The second flag tells the linker to include libRNA.a, and the remaining two flags activate Link Time Optimization (LTO) and OpenMP support support, respectively.

Note

Depending on your linker, the last two flags may differ.

Depending on your configure time decisions, you can drop one or both of the last flags.

In case you've compiled *RNAlib* with LTO support (See Link Time Optimization (LTO)) and you are using the same compiler for your third-party project that links against our library, you may add the

flag to enable Link Time Optimization.

2.2 HelloWorld 9

2.1.3.2 The pkg-config tool

Instead of hard-coding the required compiler and linker flags, you can also let the *pkg-config* tool automatically determine the required flags. This tool is usually packaged for any Linux distribution and should be available for MacOS X and MinGW as well. We ship a file *RNAlib2.pc* which is installed along with the static *libRNA.a* C-library and populated with all required compiler and linker flags that correspond to your configure time decisions.

The compiler flags required for properly building your code that uses RNAlib can be easily obtained via

```
pkg-config --cflags RNAlib2
```

You get the corresponding linker flags using

```
pkg-config --libs RNAlib2
```

With this widely accepted standard it is also very easy to integrate *RNAlib* in your *autotools* project, just have a look at the *PKG CHECK MODULES* macro.

2.2 HelloWorld

Below, you'll find some more or less simple C programs showing first steps into using *RNAlib*. A complete list of example C programs can be found in the C Examples section.

Simple MFE prediction for a given sequence

```
#include <stdlib.h>
#include <stdio.h>
#include <ViennaRNA/fold.h>
#include <ViennaRNA/utils/basic.h>
int
main()
{
    /* The RNA sequence */
    char *seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA";
    /* allocate memory for MFE structure (length + 1) */
    char *structure = (char *)vrna_alloc(sizeof(char) * (strlen(seq) + 1));
    /* predict Minmum Free Energy and corresponding secondary structure */
    float mfe = vrna_fold(seq, structure);
    /* print sequence, structure and MFE */
    printf("%s\n%s [ %6.2f ]\n", seq, structure, mfe);
    /* cleanup memory */
    free(structure);
    return 0;
}
```

See also

examples/helloworld_mfe.c in the source code tarball

10 Getting Started

Simple MFE prediction for a multiple sequence alignment

```
#include <stdlib.h>
#include <stdio.h>
#include <string.h>
#include <ViennaRNA/alifold.h>
#include <ViennaRNA/utils/basic.h>
#include <ViennaRNA/utils/alignments.h>
int
main()
  /\star The RNA sequence alignment \star/
  const char *sequences[] =
    "CUGCCUCACAACGUUUGUGCCUCAGUUACCCGUAGAUGUAGUGAGGGU",
    "CUGCCUCACAACAUUUGUGCCUCAGUUACUCAUAGAUGUAGUGAGGGU"
    "--CUCGACACCACU--GCCUCGGUUACCCAUCGGUGCAGUGCGGGU",
    NULL /* indicates end of alignment */
  /* compute the consensus sequence */
              *cons = consensus(sequences);
  /\star allocate memory for MFE consensus structure (length + 1) \star/
              *structure = (char *) vrna_alloc(sizeof(char) * (strlen(sequences[0]) + 1));
  /* predict Minmum Free Energy and corresponding secondary structure */
             mfe = vrna_alifold(sequences, structure);
  /\star print consensus sequence, structure and MFE \star/
  printf("%s\n%s [ %6.2f ]\n", cons, structure, mfe);
   /* cleanup memory */
  free (cons):
  free (structure);
  return 0;
```

See also

examples/helloworld_mfe_comparative.c in the source code tarball

Simple Base Pair Probability computation

```
#include <stdlib.h>
#include <stdio.h>
#include <string.h>
#include <ViennaRNA/fold.h>
#include <ViennaRNA/part_func.h>
#include <ViennaRNA/utils/basic.h>
int
main()
              *seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA";
   /\star allocate memory for pairing propensity string (length + 1) \,\star/
              *propensity = (char *) vrna_alloc(sizeof(char) * (strlen(seq) + 1));
  /* pointers for storing and navigating through base pair probabilities */
vrna_ep_t *ptr, *pair_probabilities = NULL;
               en = vrna_pf_fold(seq, propensity, &pair_probabilities);
  /* print sequence, pairing propensity string and ensemble free energy */
printf("%s\n%s [ %6.2f ]\n", seq, propensity, en);
/* print all base pairs with probability above 50% */
for (ptr = pair_probabilities; ptr->i != 0; ptr++)
     if (ptr->p > 0.5)
    printf("p(%d, %d) = %g\n", ptr->i, ptr->p;
   /* cleanup memory */
   free (pair_probabilities);
   free (propensity);
   return 0;
```

See also

 $\verb|examples/helloworld_probabilities.c| in the source code tarball|$

Deviating from the Default Model

```
#include <stdlib.h>
#include <stdio.h>
#include <string.h>
#include <ViennaRNA/model.h>
#include <ViennaRNA/fold_compound.h>
#include <ViennaRNA/utils/basic.h>
```

```
#include <ViennaRNA/utils/strings.h>
#include <ViennaRNA/mfe.h>
int
main()
  /* initialize random number generator */
  vrna_init_rand();
  /\star Generate a random sequence of 50 nucleotides \star/
         *seq = vrna_random_string(50, "ACGU");
  /\star allocate memory for MFE structure (length + 1) \,\star/
           *structure = (char *)vrna_alloc(sizeof(char) * (strlen(seq) + 1));
  /\star create a new model details structure to store the Model Settings \star/
  vrna_md_t md;
  /* ALWAYS set default model settings first! */
  vrna_md_set_default(&md);
  /\star change temperature and activate G-Quadruplex prediction \star/
  md.temperature = 25.0; /* 25 Deg Celcius */
md.gquad = 1; /* Turn-on G-Quadruples support */
  /* create a fold compound */
  vrna_fold_compound_t *fc = vrna_fold_compound(seq, &md, VRNA_OPTION_DEFAULT);
  /* predict Minmum Free Energy and corresponding secondary structure */
  printf("%s\n%s [ %6.2f ]\n", seq, structure, mfe);
  /* cleanup memory */
  free (structure);
  vrna_fold_compound_free(fc);
  return 0;
```

See also

examples/fold_compound_md.c in the source code tarball

2.3 HelloWorld (Perl/Python)

2.3.1 Perl5

Simple MFE prediction for a given sequence

```
use RNA; # The RNA sequence my $seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA"; # compute minimum free energy (MFE) and corresponding structure my ($ss, $mfe) = RNA::fold($seq); # print output printf "%s\n%s [ %6.2f ]\n", $seq, $ss, $mfe;
```

Simple MFE prediction for a multiple sequence alignment

```
use RNA;
# The RNA sequence alignment
my @sequences = (
    "CUGCCUCACAACGUUUGUGCCUCAGUUACCCGUAGAUGUAGUGAGGGU",
    "CUGCCUCACAACAUUUGUGCCUCAGUUACUCAUAGAUGUAGUGAGGGU",
    "--CUCGACACCACU--GCCUCGGUUACCCAUCGGUGCAGUGCGGGU"
);
# compute the consensus sequence
my $cons = RNA::consensus(\@sequences);
# predict Minmum Free Energy and corresponding secondary structure
my ($ss, $mfe) = RNA::alifold(\@sequences);
# print output
printf "%s\n\%s [ \%6.2f ]\n", \$cons, \$ss, \$mfe;
```

12 Getting Started

Deviating from the Default Model

```
use RNA;
# The RNA sequence
my $seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA";
# create a new model details structure
my $md = new RNA::md();
# change temperature and dangle model
$md->{temperature} = 20.0; # 20 Deg Celcius
$md->{dangles} = 1; # Dangle Model 1
# create a fold compound
my $fc = new RNA::fold_compound($seq, $md);
# predict Minmum Free Energy and corresponding secondary structure
my ($ss, $mfe) = $fc->mfe();
# print sequence, structure and MFE
printf "%s\n%s [ %6.2f ]\n", $seq, $ss, $mfe;
```

2.3.2 Python

Simple MFE prediction for a given sequence

```
import RNA
# The RNA sequence
seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA"
# compute minimum free energy (MFE) and corresponding structure
(ss, mfe) = RNA.fold(seq)
# print output
print "%s\n%s [ %6.2f ]" % (seq, ss, mfe)
```

Simple MFE prediction for a multiple sequence alignment

```
import RNA
# The RNA sequence alignment
sequences = [
    "CUGCCUCACAACGUUUGUGCCUCAGUUACCCGUAGAUGUAGUGAGGGU",
    "CUGCCUCACAACAGUUUGUGCCUCAGUUACUCAUAGAUGUAGUGAGGGU",
    "--CUCGACACCACU--GCCUCGGUUACCCAUCGGUGCAGUGCGGGU"
]
# compute the consensus sequence
cons = RNA.consensus(sequences)
# predict Minmum Free Energy and corresponding secondary structure
(ss, mfe) = RNA.alifold(sequences);
# print output
print "%s\n%s [ %6.2f ]" % (cons, ss, mfe)
```

Deviating from the Default Model

```
import RNA
# The RNA sequence
seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA"
# create a new model details structure
md = RNA.md()
# change temperature and dangle model
md.temperature = 20.0 # 20 Deg Celcius
md.dangles = 1 # Dangle Model 1
# create a fold compound
fc = RNA.fold_compound(seq, md)
# predict Minmum Free Energy and corresponding secondary structure
(ss, mfe) = fc.mfe()
# print sequence, structure and MFE
print "%s\n%s [ %6.2f ]\n" % (seq, ss, mfe)
```

Chapter 3

Concepts and Algorithms

This is an overview of the concepts and algorithms for which implementations can be found in this library.

Almost all of them rely on the physics based Nearest Neighbor Model for RNA secondary structure prediction.

- · RNA Structure gives an introduction into the different layers of abstraction for RNA structures
- · Distance Measures introduces different metrics to allow for the comparison of secondary structures
- Free Energy of Secondary Structures shows how the stability of a secondary structure can be quantified in terms of free energy
- Secondary Structure Folding Grammar explains the basic recursive decomposition scheme that is applied in secondary structure prediction
- RNA Secondary Structure Landscapes describes how transition paths between secondary structures span a landscape like graph
- Minimum Free Energy Algorithm(s) compute the most stable conformation in thermodynamic equilibrium
- Partition Function and Equilibrium Probability Algorithm(s) enable one to apply statistical mechanics to derive equilibrium probabilities of structure features
- Suboptimals and (other) Representative Structures allow for alternative description and enumeration of the structure ensemble
- RNA-RNA Interaction introduces how to model the interaction between RNA molecules
- · Locally Stable Secondary Structures offer insights into structuredness of long sequences and entire genomes
- Comparative Structure Prediction augment structure prediction with evolutionary conservation of homologous sequences
- Classified DP variations perform an *a priori* partitioning of the structure ensemble and compute various properties for the resulting classes.
- RNA Sequence Design constitutes the inverse problem of structure prediction
- Experimental Structure Probing Data can be used to guide structure prediction, for instance using SHAPE reactivity data
- Ligand Binding adds more complexity to structure prediction by modelling the interaction between small chemical compounds or proteins and the RNA
- (Tertiary) Structure Motifs extend the abstraction of secondary structure beyond canonical base pair formation

3.1 RNA Structure

- 3.1.1 RNA Structures
- 3.1.2 Levels of Structure Abstraction
- 3.1.2.1 Primary Structure
- 3.1.2.2 Secondary Structure
- 3.1.2.3 Tertiary Structure
- 3.1.2.4 Quarternary Structure
- 3.1.2.5 Pseudo-Knots

3.2 Distance Measures

A simple measure of dissimilarity between secondary structures of equal length is the base pair distance, given by the number of pairs present in only one of the two structures being compared. I.e. the number of base pairs that have to be opened or closed to transform one structure into the other. It is therefore particularly useful for comparing structures on the same sequence. It is implemented by

Compute the "base pair" distance between two secondary structures s1 and s2.

For other cases a distance measure that allows for gaps is preferable. We can define distances between structures as edit distances between trees or their string representations. In the case of string distances this is the same as "sequence alignment". Given a set of edit operations and edit costs, the edit distance is given by the minimum sum of the costs along an edit path converting one object into the other. Edit distances like these always define a metric. The edit operations used by us are insertion, deletion and replacement of nodes. String editing does not pay attention to the matching of brackets, while in tree editing matching brackets represent a single node of the tree. Tree editing is therefore usually preferable, although somewhat slower. String edit distances are always smaller or equal to tree edit distances.

The different level of detail in the structure representations defined above naturally leads to different measures of distance. For full structures we use a cost of 1 for deletion or insertion of an unpaired base and 2 for a base pair. Replacing an unpaired base for a pair incurs a cost of 1.

Two cost matrices are provided for coarse grained structures:

3.2 Distance Measures 15

The lower matrix uses the costs given in [21]. All distance functions use the following global variables:

```
int cost_matrix;
```

Specify the cost matrix to be used for distance calculations.

```
int edit_backtrack;
```

Produce an alignment of the two structures being compared by tracing the editing path giving the minimum distance.

```
char *aligned_line[4];
```

Contains the two aligned structures after a call to one of the distance functions with edit backtrack set to 1.

See also

utils.h, dist_vars.h and stringdist.h for more details

3.2.1 Functions for Tree Edit Distances

```
Tree *make_tree (char *struc)
```

Constructs a Tree (essentially the postorder list) of the structure 'struc', for use in tree edit distance().

```
float tree_edit_distance (Tree *T1, Tree *T2)
```

Calculates the edit distance of the two trees.

```
void free_tree(Tree *t)
```

Free the memory allocated for Tree t.

See also

dist_vars.h and treedist.h for prototypes and more detailed descriptions

3.2.2 Functions for String Alignment

```
swString *Make_swString (char *string)
```

Convert a structure into a format suitable for string_edit_distance().

Calculate the string edit distance of T1 and T2.

See also

dist_vars.h and stringdist.h for prototypes and more detailed descriptions

3.2.3 Functions for Comparison of Base Pair Probabilities

For comparison of base pair probability matrices, the matrices are first condensed into probability profiles which are the compared by alignment.

condense pair probability matrix into a vector containing probabilities for unpaired, upstream paired and downstream paired.

```
float profile_edit_distance ( const float *T1, const float *T2)
```

Align the 2 probability profiles T1, T2

See also

ProfileDist.h for prototypes and more details of the above functions

3.3 Free Energy of Secondary Structures

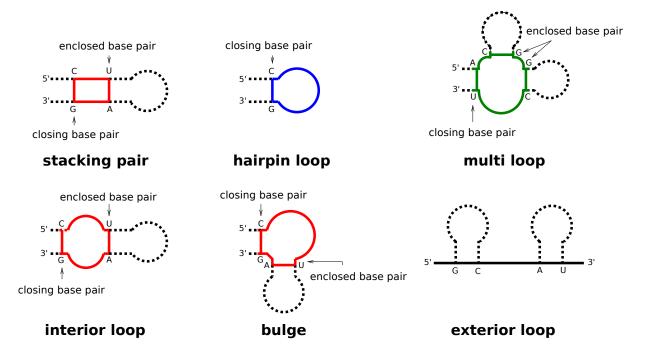
A description on how secondary structures are decomposed into individual loops to eventually evaluate their stability in terms of free energy.

3.3.1 Secondary Structure Loop Decomposition

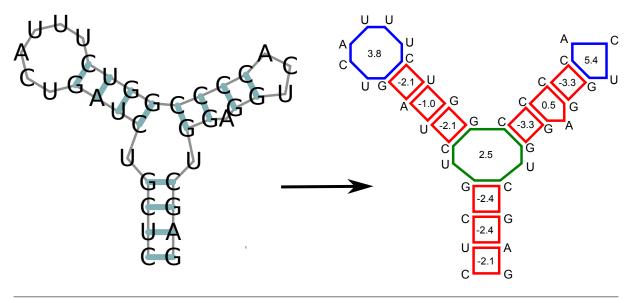
Each base pair in a secondary structure closes a loop, thereby directly enclosing unpaired nucleotides, and/or further base pairs. Our implementation distinguishes four basic types of loops:

- · hairpin loops
- · interior loops
- · multibranch loops
- · exterior loop

While the exterior loop is a special case without a closing pair, the other loops are determined by the number of base pairs involved in the loop formation, i.e. hairpin loops are 1-loops, since only a single base pair delimits the loop. interior loops are 2-loops due to their enclosing, and enclosed base pair. All loops where more than two base pairs are involved, are termed multibranch loops.



Any secondary structure can be decomposed into its loops. Each of the loops then can be scored in terms of free energy, and the free energy of an entire secondary structure is simply the sum of free energies of its loops.



3.3.1.1 Free Energy Evaluation API

While we implement some functions that decompose a secondary structure into its individual loops, the majority of methods provided in @RNAlib are dedicated to free energy evaluation. The corresponding modules are:

See also

Free Energy Evaluation, Energy Evaluation for Individual Loops

3.3.2 Free Energy Parameters

For secondary structure free energy evaluation we usually utilize the set of Nearest Neighbor Parameters also used in other software, such as *UNAfold* and *RNAstructure*. While the *RNAlib* already contains a compiled-in set of the latest *Turner 2004 Free Energy Parameters*, we defined a file format that allows to change these parameters at runtime. The ViennaRNA Package already comes with a set of parameter files containing

- Turner 1999 RNA parameters
- · Mathews 1999 DNA parameters
- · Andronescu 2007 RNA parameters
- · Mathews 2004 DNA parameters

3.3.2.1 Free Energy Parameters Modification API

See also

Energy Parameters, Reading/Writing Energy Parameter Sets from/to File

3.3.3 Fine-tuning of the Energy Evaluation Model

See also

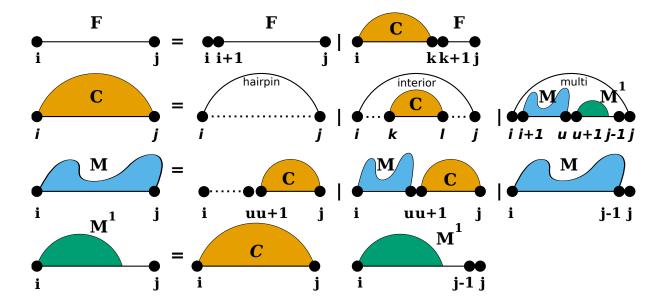
Fine-tuning of the Implemented Models

3.4 Secondary Structure Folding Grammar

A description of the basic grammar to generate secondary structures, used for almost all prediction algorithms in our library and how to modify it.

3.4.1 Secondary Structure Folding Recurrences

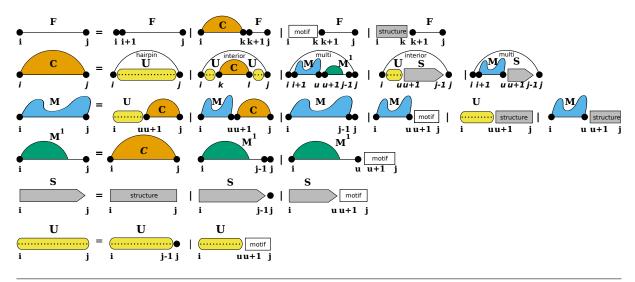
To predict secondary structures composed of the four distinguished loop types introduced before, all algorithms implemented in *RNAlib* follow a specific decomposition scheme, also known as the *RNA folding grammar*, or *Secondary Structure Folding Recurrences*.



However, compared to other RNA secondary structure prediction libraries, our implementation allows for a fine-grained control of the above recursions by constraining both, the individual derivations of the grammar as well as the evaluation of particular loop contributions. Furthermore, we provide a mechanism to extend the above grammar with additional derivation rules, so-called *Domains*.

3.4.2 Additional Structural Domains

Some applications of RNA secondary structure prediction require an extension of the *regular RNA folding grammar*. For instance one would like to include proteins and other ligands binding to unpaired loop regions while competing with conventional base pairing. Another application could be that one may want to include the formation of self-enclosed structural modules, such as *G-quadruplexes*. For such applications, we provide a pair of additional domains that extend the regular RNA folding grammar, Structured Domains and Unstructured Domains.



While unstructured domains are usually determined by a more or less precise sequence motif, e.g. the binding site for a protein, structured domains are considered self-enclosed modules with a more or less complex pairing pattern. Our extension with these two domains introduces two production rules to fill additional dynamic processing matrices S and U where we store the pre-computed contributions of structured domains (S), and unstructured domains (U).

3.4.2.1 Structured Domains

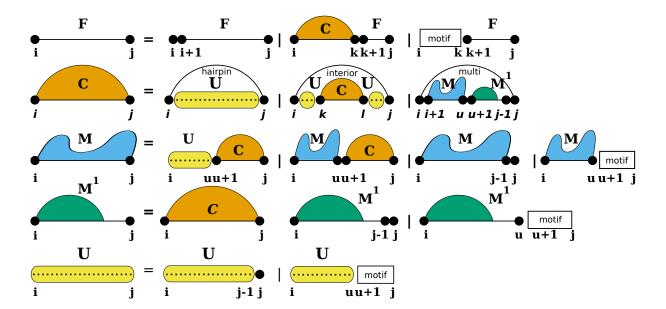
Usually, structured domains represent self-enclosed structural modules that exhibit a more or less complex base pairing pattern. This can be more or less well-defined 3D motifs, such as *G-Quadruplexes*, or loops with additional non-canonical base pair interactions, such as *kink-turns*.

Note

Currently, our implementation only provides the specialized case of *G-Quadruplexes*.

3.4.2.2 Unstructured Domains

Unstructured domains appear in the production rules of the RNA folding grammar wherever new unpaired nucleotides are attached to a growing substructure (see also [15]):



The white boxes represent the stretch of RNA bound to the ligand and represented by a more or less specific sequence motif. The motif itself is considered unable to form base pairs. The additional production rule U is used to precompute the contribution of unpaired stretches possibly bound by one or more ligands. The auxiliary DP matrix for this production rule is filled right before processing the other (regular) production rules of the RNA folding grammar.

3.4.2.3 Domain Extension API

For the sake of flexibility, each of the domains is associated with a specific data structure serving as an abstract interface to the extension. The interface uses callback functions to

- pre-compute arbitrary data, e.g. filling up additional dynamic programming matrices, and
- evaluate the contribution of a paired or unpaired structural feature of the RNA.

Implementations of these callbacks are separate for regular free energy evaluation, e.g. MFE prediction, and partition function applications. A data structure holding arbitrary data required for the callback functions can be associated to the domain as well. While *RNAlib* comes with a default implementation for structured and unstructured domains, the system is entirely user-customizable.

See also

Unstructured Domains, Structured Domains, G-Quadruplexes, Ligands Binding to Unstructured Domains

3.4.3 Constraints on the Folding Grammar

Secondary Structure constraints can be subdivided into two groups:

- · Hard Constraints
- Soft Constraints

While Hard-Constraints directly influence the production rules used in the folding recursions by allowing, disallowing, or enforcing certain decomposition steps, Soft-constraints on the other hand are used to change position specific contributions in the recursions by adding bonuses/penalties in form of pseudo free energies to certain loop configurations.

Note

Secondary structure constraints are always applied at decomposition level, i.e. in each step of the recursive structure decomposition, for instance during MFE prediction.

3.4.3.1 Hard Constraints API

Hard constraints as implemented in our library can be specified for individual loop types, i.e. the atomic derivations of the RNA folding grammar rules. Hence, the pairing behavior of both, single nucleotides and pairs of bases, can be constrained in every loop context separately. Additionally, an abstract implementation using a callback mechanism allows for full control of more complex hard constraints.

See also

Hard Constraints

3.4.3.2 Soft Constraints API

For the sake of memory efficiency, we do not implement a loop context aware version of soft constraints. The *static* soft constraints as implemented only distinguish unpaired from paired nucleotides. This is usually sufficient for most use-case scenarios. However, similar to hard constraints, an abstract soft constraints implementation using a callback mechanism exists, that allows for any soft constraint that is compatible with the RNA folding grammar. Thus, loop contexts and even individual derivation rules can be addressed separately for maximum flexibility in soft-constraints application.

See also

Soft Constraints, Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints, SHAPE Reactivity Data

3.5 RNA Secondary Structure Landscapes

A description of the implicit landscape-like network of structures that appears upon modelling the transition of one structure into another

- 3.5.1 The Neighborhood of a Secondary Structure
- 3.5.2 The Secondary Structure Landscape API

3.6 Minimum Free Energy Algorithm(s)

Computing the Minimum Free Energy (MFE), i.e. the most stable conformation in thermodynamic equilibrium

3.6.1 Zuker's Algorithm

Our library provides fast dynamic programming Minimum Free Energy (MFE) folding algorithms derived from the decomposition scheme as described by "Zuker & Stiegler (1981)" [27].

3.6.2 MFE for circular RNAs

Folding of *circular* RNA sequences is handled as a post-processing step of the forward recursions. See [12] for further details.

3.6.3 MFE Algorithm API

We provide interfaces for the prediction of

- · MFE and corresponding secondary structure for single sequences,
- · consensus MFE structures of sequence alignments, and
- · MFE structure for two hybridized RNA strands

See also

Minimum Free Energy (MFE) Algorithms, consensus_mfe_fold, mfe_cofold, Computing MFE representatives of a Distance Base

3.7 Partition Function and Equilibrium Probability Algorithm(s)

3.7.1 Equilibrium Ensemble Statistics

In contrast to methods that compute the property of a single structure in the ensemble, e.g. Minimum Free Energy Algorithm(s), the partition function algorithms always consider the entire equilibrium ensemble. For that purpose, the McCaskill algorithm [17] and its variants can be used to efficiently compute

- · the partition function, and from that
- various equilibrium probabilities, for instance base pair probabilities, probabilities of individual structure motifs, and many more.

The principal idea behind this approach is that in equilibrium, statistical mechanics and polymer theory tells us that the frequency or probability p(s) of a particular state s depends on its energy E(s) and follows a Boltzmann distribution, i.e.

$$p(s) \propto e^{-\beta E(s)}$$
 with $\beta = \frac{1}{kT}$

where $k\approx 1.987\cdot 10^{-3}\frac{kcal}{mol\ K}$ is the Boltzmann constant, and T the thermodynamic temperature. From that relation, the actual probability of state s can then be obtained using a proper scaling factor, the *canonical partition function*

$$Z = \sum_{s \in \Omega} e^{-\beta E(s)}$$

where Ω is the finite set of all states. Finally, the equilibrium probability of state s can be computed as

$$p(s) = \frac{e^{-\beta E(s)}}{Z}$$

Instead of enumerating all states exhaustively to compute Z one can apply the Secondary Structure Folding Recurrences again for an efficient computation in cubic time. An *outside* variant of the same recursions is then used to compute probabilities for base pairs, stretches of consecutive unpaired nucleotides, or structural motifs.

See also

Further details of the Partition function and Base Pair Probability algorithm can be obtained from McCaskill 1990 [17]

3.7.2 Partition Function and Equilibrium Probability API

We implement a wide variety of variants of the partition function algorithm according to McCaskill 1990 [17]. See the corresponding submodules for specific implementation details.

See also

Partition Function and Equilibrium Properties, consensus_pf_fold, Partition Function for Two Hybridized Sequences, Partition Function for two Hybridized Sequences as a Stepwise Process, local_pf_fold, Computing Partition Functions of a Distance of the Process o

3.8 Suboptimals and (other) Representative Structures

- 3.8.1 Suboptimal Secondary Structures
- 3.8.2 Sampling Secondary Structures from the Ensemble
- 3.8.3 Structure Enumeration and Sampling API

See also

Suboptimal Structures sensu Stiegler et al. 1984 / Zuker et al. 1989, Suboptimal Structures within an Energy Band around the Random Structure Samples from the Ensemble, Compute the Structure with Maximum Expected Accuracy (MEA), Compute the Centroid Structure

3.9 RNA-RNA Interaction

3.9.1 rip_intro

The function of an RNA molecule often depends on its interaction with other RNAs. The following routines therefore allows one to predict structures formed by two RNA molecules upon hybridization.

3.9.2 Concatenating RNA sequences

One approach to co-folding two RNAs consists of concatenating the two sequences and keeping track of the concatenation point in all energy evaluations. Correspondingly, many of the cofold() and co_pf_fold() routines take one sequence string as argument and use the global variable cut_point to mark the concatenation point. Note that while the RNAcofold program uses the '&' character to mark the chain break in its input, you should not use an '&' when using the library routines (set cut_point instead).

3.9.3 RNA-RNA interaction as a Stepwise Process

In a second approach to co-folding two RNAs, cofolding is seen as a stepwise process. In the first step the probability of an unpaired region is calculated and in a second step this probability of an unpaired region is multiplied with the probability of an interaction between the two RNAs. This approach is implemented for the interaction between a long target sequence and a short ligand RNA. Function pf_unstru() calculates the partition function over all unpaired regions in the input sequence. Function pf_interact(), which calculates the partition function over all possible interactions between two sequences, needs both sequence as separate strings as input.

3.9.4 RNA-RNA Interaction API

3.10 Locally Stable Secondary Structures

- 3.10.1 local_intro
- 3.10.2 local_mfe
- 3.10.3 local_pf
- 3.10.4 Locally Stable Secondary Structure API

3.11 Comparative Structure Prediction

3.11.1 Incorporate Evolutionary Information

Consensus structures can be predicted by a modified version of the fold() algorithm that takes a set of aligned sequences instead of a single sequence. The energy function consists of the mean energy averaged over the sequences, plus a covariance term that favors pairs with consistent and compensatory mutations and penalizes pairs that cannot be formed by all structures. For details see [10] and [1].

3.11.2 Comparative Structure Prediction API

3.12 Classified DP variations

3.12.1 The Idea of Classified Dynamic Programming

Usually, thermodynamic properties using the basic recursions for Minimum Free Energy Algorithm(s), Partition Function and Equilibriu and so forth, are computed over the entire structure space. However, sometimes it is desired to partition the structure space a priori and compute the above properties for each of the resulting partitions. This approach directly leads to Classified Dynamic Programming.

3.12.2 Distance Class Partitioning

The secondary structure space is divided into partitions according to the base pair distance to two given reference structures and all relevant properties are calculated for each of the resulting partitions.

See also

For further details, we refer to Lorenz et al. 2009 [14]

- 3.12.3 Density of States (DOS)
- 3.12.4 Classified DP API
- 3.13 RNA Sequence Design
- 3.13.1 Generate Sequences that fold into particular Secondary Structures
- 3.13.2 RNA Sequence Design API

See also

Inverse Folding (Design)

- 3.14 Experimental Structure Probing Data
- 3.14.1 Guide the Structure Prediction using Experimental Data
- 3.14.1.1 SHAPE reactivities
- 3.14.2 Structure Probing Data API

See also

Experimental Structure Probing Data, SHAPE Reactivity Data, pertubation

- 3.15 Ligand Binding
- 3.15.1 Small Molecules and Proteins that bind to specific RNA Structures
- 3.15.2 ligand_binding_api

In our library, we provide two different ways to incorporate ligand binding to RNA structures:

- · Ligands Binding to Unstructured Domains, and
- Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints

The first approach is implemented as an actual extension of the folding grammar. It adds auxiliary derivation rules for each case when consecutive unpaired nucleotides are evaluated. Therefore, this model is applicable to ligand binding to any loop context.

The second approach, on the other hand, uses the soft-constraints feature to change the energy evaluation of hairpin- or interior-loops. Hence, it can only be appleid when a ligand binds to a hairpin-like, or interior-loop like motif.

See also

Ligands Binding to Unstructured Domains, Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Con

- 3.16 (Tertiary) Structure Motifs
- 3.16.1 Incorporating Higher-Order (Tertiary) Structure Motifs
- 3.16.2 RNA G-Quadruplexes
- 3.16.3 (Tertiary) Structure Motif API

Chapter 4

I/O Formats

Below, you'll find a listing of different sections that introduce the most common notations of sequence and structure data, specifications of bioinformatics sequence and structure file formats, and various output file formats produced by our library.

- RNA Structure Notations describes the different notations and representations of RNA secondary structures
- · File Formats gives an overview of the file formats compatible with our library
- Plotting shows the different (PostScript) plotting functions for RNA secondary structures, feature probabilities, and multiple sequence alignments

4.1 RNA Structure Notations

4.1.1 Representations of Secondary Structures

The standard representation of a secondary structure in our library is the Dot-Bracket Notation (a.k.a. Dot-Parenthesis Notation), where matching brackets symbolize base pairs and unpaired bases are shown as dots. Based on that notation, more elaborate representations have been developed to include additional information, such as the loop context a nucleotide belongs to and to annotated pseudo-knots.

See also

Extended Dot-Bracket Notation, Washington University Secondary Structure (WUSS) notation

4.1.2 Dot-Bracket Notation (a.k.a. Dot-Parenthesis Notation)

The Dot-Bracket notation as introduced already in the early times of the ViennaRNA Package denotes base pairs by matching pairs of parenthesis () and unpaired nucleotides by dots...

Example: A simple helix of size 4 enclosing a hairpin of size 4 is annotated as

```
((((...))))
```

See also

vrna_ptable_from_string(), vrna_db_flatten(), vrna_db_flatten_to()

30 I/O Formats

4.1.3 Extended Dot-Bracket Notation

A more generalized version of the original Dot-Bracket notation may use additional pairs of brackets, such as <>, and [], and matching pairs of uppercase/lowercase letters. This allows for anotating pseudo-knots, since different pairs of brackets are not required to be nested.

Example: The follwing annotations of a simple structure with two crossing helices of size 4 are equivalent:

```
<<<[[[[...>>>]]]]
((((AAAA...))))aaaa
AAAA{{{{...aaaa}}}}
```

See also

```
vrna ptable from string(), vrna db flatten(), vrna db flatten to()
```

4.1.4 Washington University Secondary Structure (WUSS) notation

The WUSS notation, as frequently used for consensus secondary structures in Stockholm 1.0 format allows for a fine-grained annotation of base pairs and unpaired nucleotides, including pseudo-knots.

Below, you'll find a list of secondary structure elements and their corresponding WUSS annotation (See also the infernal user guide at http://eddylab.org/infernal/Userguide.pdf)

Base pairs

Nested base pairs are annotated by matching pairs of the symbols <>, (), $\{\}$, and []. Each of the matching pairs of parenthesis have their special meaning, however, when used as input in our programs, e.g. structure constraint, these details are usually ignored. Furthermore, base pairs that constitute as pseudo-knot are denoted by letters from the latin alphabet and are, if not denoted otherwise, ignored entirely in our programs.

· Hairpin loops

Unpaired nucleotides that constitute the hairpin loop are indicated by underscores, _.

```
Example:
```

Bulges and interior loops

Residues that constitute a bulge or interior loop are denoted by dashes, -.

```
Example:
```

· Multibranch loops

Unpaired nucleotides in multibranch loops are indicated by commas,.

```
Example: ((("«_____», «____»)))
```

External residues

Single stranded nucleotides in the exterior loop, i.e. not enclosed by any other pair are denoted by colons, :.

```
Example: 
«<______»>::
```

Insertions

In cases where an alignment represents the consensus with a known structure, insertions relative to the known structure are denoted by periods, .. Regions where local structural alignment was invoked, leaving regions of both target and query sequence unaligned, are indicated by tildes, \sim .

Note

These symbols only appear in alignments of a known (query) structure annotation to a target sequence of unknown structure.

· Pseudo-knots

The WUSS notation allows for annotation of pseudo-knots using pairs of upper-case/lower-case letters.

Note

Our programs and library functions usually ignore pseudo-knots entirely treating them as unpaired nucleotides, if not stated otherwise.

```
Example: «<_AAA____»>aaa
```

See also

```
vrna db from WUSS()
```

4.1.5 Tree Representations of Secondary Structures

Alternatively, one may find representations with two types of node labels, 'P' for paired and 'U' for unpaired; a dot is then replaced by '(U)', and each closed bracket is assigned an additional identifier 'P'. We call this the expanded notation. In [8] a condensed representation of the secondary structure is proposed, the so-called homeomorphically irreducible tree (HIT) representation. Here a stack is represented as a single pair of matching brackets labeled 'P' and weighted by the number of base pairs. Correspondingly, a contiguous strain of unpaired bases is shown as one pair of matching brackets labeled 'U' and weighted by its length. Generally any string consisting of matching brackets and identifiers is equivalent to a plane tree with as many different types of nodes as there are identifiers.

Bruce Shapiro proposed a coarse grained representation [20], which, does not retain the full information of the secondary structure. He represents the different structure elements by single matching brackets and labels them as

- H (hairpin loop),
- I (interior loop),
- B (bulge),
- · M (multi-loop), and
- S (stack).

We extend his alphabet by an extra letter for external elements E. Again these identifiers may be followed by a weight corresponding to the number of unpaired bases or base pairs in the structure element. All tree representations (except for the dot-bracket form) can be encapsulated into a virtual root (labeled R).

The following example illustrates the different linear tree representations used by the package:

Consider the secondary structure represented by the dot-bracket string (full tree)

```
.((..(((...)))..((..))).
```

which is the most convenient condensed notation used by our programs and library functions.

Then, the following tree representations are equivalent:

32 I/O Formats

· Expanded tree:

• HIT representation (Fontana et al. 1993 [8]):

```
((U1)((U2)((U3)P3)(U2)((U2)P2)P2)(U1)R)
```

- Coarse Grained Tree Representation (Shapiro 1988 [20]):
 - Short (with root node \mathbb{R} , without stem nodes \mathbb{S}):

```
((H)((H)M)R)
```

- Full (with root node R):

```
(((((H)S)((H)S)M)S)R)
```

– Extended (with root node \mathbb{R} , with external nodes \mathbb{E}):

```
(((((((H)S)((H)S)M)S)E)R)
```

- Weighted (with root node \mathbb{R} , with external nodes \mathbb{E}):

```
((((((H3)S3)((H2)S2)M4)S2)E2)R)
```

The Expanded tree is rather clumsy and mostly included for the sake of completeness. The different versions of Coarse Grained Tree Representations are variatios of Shapiro's linear tree notation.

For the output of aligned structures from string editing, different representations are needed, where we put the label on both sides. The above examples for tree representations would then look like:

Aligned structures additionally contain the gap character ' '.

4.1.6 Examples for Structure Parsing and Conversion

4.1.7 Structure Parsing and Conversion API

Several functions are provided for parsing structures and converting to different representations.

```
char *expand_Full(const char *structure)
```

Convert the full structure from bracket notation to the expanded notation including root.

```
char *b2HIT (const char *structure)
```

Converts the full structure from bracket notation to the HIT notation including root.

```
char *b2C (const char *structure)
```

Converts the full structure from bracket notation to the a coarse grained notation using the 'H' 'B' 'I' 'M' and 'R' identifiers.

```
char *b2Shapiro (const char *structure)
```

Converts the full structure from bracket notation to the *weighted* coarse grained notation using the 'H' 'B' 'I' 'M' 'S' 'E' and 'R' identifiers.

```
char *expand_Shapiro (const char *coarse);
```

Inserts missing 'S' identifiers in unweighted coarse grained structures as obtained from b2C().

```
char *add_root (const char *structure)
```

Adds a root to an un-rooted tree in any except bracket notation.

```
char *unexpand_Full (const char *ffull)
```

Restores the bracket notation from an expanded full or HIT tree, that is any tree using only identifiers 'U' 'P' and 'R'.

```
char *unweight (const char *wcoarse)
```

Strip weights from any weighted tree.

```
void unexpand_aligned_F (char *align[2])
```

Converts two aligned structures in expanded notation.

```
void parse_structure (const char *structure)
```

Collects a statistic of structure elements of the full structure in bracket notation.

See also

RNAstruct.h for prototypes and more detailed description

34 I/O Formats

4.2 File Formats

4.2.1 File formats for Multiple Sequence Alignments (MSA)

4.2.1.1 ClustalW format

The *ClustalW* format is a relatively simple text file containing a single multiple sequence alignment of DNA, RNA, or protein sequences. It was first used as an output format for the *clustalw* programs, but nowadays it may also be generated by various other sequence alignment tools. The specification is straight forward:

· The first line starts with the words

```
CLUSTAL W

Or

CLUSTALW
```

- · After the above header there is at least one empty line
- Finally, one or more blocks of sequence data are following, where each block is separated by at least one empty line

Each line in a blocks of sequence data consists of the sequence name followed by the sequence symbols, separated by at least one whitespace character. Usually, the length of a sequence in one block does not exceed 60 symbols. Optionally, an additional whitespace separated cumulative residue count may follow the sequence symbols. Optionally, a block may be followed by a line depicting the degree of conservation of the respective alignment columns.

Note

Sequence names and the sequences must not contain whitespace characters! Allowed gap symbols are the hyphen ("-"), and dot (".").

Warning

Please note that many programs that output this format tend to truncate the sequence names to a limited number of characters, for instance the first 15 characters. This can destroy the uniqueness of identifiers in your MSA.

Here is an example alignment in ClustalW format:

```
CLUSTAL W (1.83) multiple sequence alignment
```

AL031296.1/85969-86120 CUGCCUCACAACGUUUGUGCCUCAGUUACCCGUAGAUGUGAGGGGUAACAAUACUUAC
AANU01225121.1/438-603 CUGCCUCACAACAUUUGUGCCUCAGUUACUCAUAGAUGUAGUGAGGGGUAACAAUACUUAC
AAWR02037329.1/29294-29150 CUCUCGACACCACU--GCCUCGGUUACCCAUCGGUGCAGUGCGGGUAGUAGUACCAAU

AL031296.1/85969-86120 UCUCGUUGGUGAUAAGGAACAGCU
AANU01225121.1/438-603 UCUCGUUGGUGAUAAGGAACAGCU
AAWR02037329.1/29294-29150 GCUAAUUAGUUGUGAGGACCAACU

4.2 File Formats 35

4.2.1.2 Stockholm 1.0 format

Here is an example alignment in Stockholm 1.0 format:

```
# STOCKHOLM 1.0
#=GF AC
                    RF01293
#=GF ID ACA59
#=GF DE
                       Small nucleolar RNA ACA59
#=GF AU Wilkinson A
#=GF SE Predicted; WAR; Wilkinson A
                      Predicted; WAR; Wilkinson A
#=GF SS
#=GF GA
                      43.00
#=GF TC
                      44.90
#=GF NC
                       40.30
#=GF TP
                     Gene; snRNA; snoRNA; HACA-box;
#=GF BM cmbuild -F CM SEED
#=GF CB cmcalibrate --mpi CM
#=GF SM cmsearch --cpu 4 --verbose --nohmmonly -E 1000 -Z 549862.597050 CM SEQDB
#=GF DR snoRNABase; ACA59;
#=GF DR
                      SO; 0001263; ncRNA_gene;
#=GF DR
                       GO; 0006396; RNA processing;
#=GF DR
                    GO; 0005730; nucleolus;
#=GF RN
                      [1]
#=GF RM
                       15199136
#=GF RT
                       Human box H/ACA pseudouridylation guide RNA machinery.
#=GF RA
                      Kiss AM, Jady BE, Bertrand E, Kiss T
#=GF RL
                       Mol Cell Biol. 2004;24:5797-5807.
#=GF WK
                      Small_nucleolar_RNA
#=GF SQ
                                                        AL031296.1/85969-86120
AANU01225121.1/438-603
                                                                AAWR02037329.1/29294-29150 ---CUCGACACCACU---GCCUCGGUUACCCAUCGGUGCAGUGCGGGUAGUAGUACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGUGAGGACCAAUGCUAAUUAGUUGAGGACCAAUGCUAAUUAGUUGAGGACCAAUGCUAAUGCUAAUUAGUUGAGGACCAAUGCUAAUGCUAAUUAGUUGAGGACCAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCUAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAUGCAAU
                                                               ----(((((,<<<<<<
                                                                                                                                        _>>>>>>
                                                                                                                                                                                                     #=GC SS_cons
#=GC RF
                                                                CUGCcccaCAaCacuuquGCCUCaGUUACcCauaqquGuAGUGaGqGuqqcAaUACccaCcCucqUUqGuqqUaAGGAaCAqC
//
```

See also

Washington University Secondary Structure (WUSS) notation on legal characters for the consensus secondary structure line SS_cons and their interpretation

4.2.1.3 FASTA (Pearson) format

Note

Sequence names must not contain whitespace characters. Otherwise, the parts after the first whitespace will be dropped. The only allowed gap character is the hyphen ("-").

Here is an example alignment in FASTA format:

```
>AL031296.1/85969-86120
CUGCCUCACAACGUUUGUGCCUCAGUUACCCGUAGAUGUAGUGAGGGUAACAAUACUUAC
UCUCGUUGGUGAUAAGGAACAGCU
>AANU01225121.1/438-603
CUGCCUCACAACAUUUGUGCCUCAGUUACUCAUAGAUGUAGUGAGGGUGACAAUACUUAC
UCUCGUUGGUGAUAAGGAACAGCU
>AAWR02037329.1/29294-29150
---CUCGACACCACU---GCCUCGGUUACCCAUCGGUGCAGUGCGGGUAGUAGUACCAAU
GCUAAUUAGUUGUGAGGACCAACU
```

36 I/O Formats

4.2.1.4 MAF format

The multiple alignment format (MAF) is usually used to store multiple alignments on DNA level between entire genomes. It consists of independent blocks of aligned sequences which are annotated by their genomic location. Consequently, an MAF formatted MSA file may contain multiple records. MAF files start with a line

##maf

which is optionally extended by whitespace delimited key=value pairs. Lines starting with the character ("#") are considered comments and usually ignored.

A MAF block starts with character ("a") at the beginning of a line, optionally followed by whitespace delimited key=value pairs. The next lines start with character ("s") and contain sequence information of the form

```
s src start size strand srcSize sequence
```

where

- · src is the name of the sequence source
- start is the start of the aligned region within the source (0-based)
- · size is the length of the aligned region without gap characters
- strand is either ("+") or ("-"), depicting the location of the aligned region relative to the source
- srcSize is the size of the entire sequence source, e.g. the full chromosome
- sequence is the aligned sequence including gaps depicted by the hyphen ("-")

Here is an example alignment in MAF format (bluntly taken from the UCSC Genome browser website):

```
##maf version=1 scoring=tba.v8
# tba.v8 (((human chimp) baboon) (mouse rat))
# multiz.v7
# maf_project.v5 _tba_right.maf3 mouse _tba_C
# single_cov2.v4 single_cov2 /dev/stdin
a score=23262.0
s hq16.chr7 27578828 38 + 158545518 AAA-GGGAATGTTAACCAAATGA---ATTGTCTCTTACGGTG
s panTrol.chr6 28741140 38 + 161576975 AAA-GGGAATGTTAACCAAATGA---ATTGTCTCTTACGGTG
                116834 38 + 4622798 AAA-GGGAATGTTAACCAAATGA---GTTGTCTCTTATGGTG
s mm4.chr6
              53215344 38 + 151104725 -AATGGGAATGTTAAGCAAACGA---ATTGTCTCTCAGTGTG
s rn3.chr4
             81344243 40 + 187371129 -AA-GGGGATGCTAAGCCAATGAGTTGTTGTCTCAATGTG
a score=5062.0
s hg16.chr7
              27699739 6 + 158545518 TAAAGA
s panTrol.chr6 28862317 6 + 161576975 TAAAGA
s baboon
               241163 6 + 4622798 TAAAGA
s mm4.chr6
             53303881 6 + 151104725 TAAAGA
              81444246 6 + 187371129 taagga
s rn3.chr4
a score=6636.0
s hg16.chr7 27707221 13 + 158545518 gcagctgaaaaca
s panTrol.chr6 28869787 13 + 161576975 gcagctgaaaaca
s baboon
                249182 13 + 4622798 gcagctgaaaaca
              53310102 13 + 151104725 ACAGCTGAAAATA
s mm4.chr6
```

4.2 File Formats 37

4.2.2 File formats to manipulate the RNA folding grammar

4.2.2.1 Command Files

The RNAlib and many programs of the ViennaRNA Package can parse and apply data from so-called command files. These commands may refer to structure constraints or even extensions of the RNA folding grammar (such as Unstructured Domains). Commands are given as a line of whitespace delimited data fields. The syntax we use extends the constraint definitions used in the mfold / UNAfold software, where each line begins with a command character followed by a set of positions.

However, we introduce several new commands, and allow for an optional loop type context specifier in form of a sequence of characters, and an orientation flag that enables one to force a nucleotide to pair upstream, or downstream.

4.2.2.1.1 Constraint commands

The following set of commands is recognized:

- F ... Force
- P ... Prohibit
- A ... Allow (for non-canonical pairs)
- E ... Soft constraints for unpaired position(s), or base pair(s)

4.2.2.1.2 RNA folding grammar exensions

UD ... Add ligand binding using the Unstructured Domains feature

4.2.2.1.3 Specification of the loop type context

The optional loop type context specifier [LOOP] may be a combination of the following:

- E ... Exterior loop
- H ... Hairpin loop
- I ... Interior loop
- M ... Multibranch loop
- A . . . All loops

For structure constraints, we additionally allow one to address base pairs enclosed by a particular kind of loop, which results in the specifier [WHERE] which consists of [LOOP] plus the following character:

- i ... enclosed pair of an Interior loop
- $\ensuremath{\mathtt{m}} \dots$ enclosed pair of a Multibranch loop

If no [LOOP] or [WHERE] flags are set, all contexts are considered (equivalent to ${\tt A}$)

38 I/O Formats

4.2.2.1.4 Controlling the orientation of base pairing

For particular nucleotides that are forced to pair, the following [ORIENTATION] flags may be used:

- U ... Upstream
- D ... Downstream

If no [ORIENTATION] flag is set, both directions are considered.

4.2.2.1.5 Sequence coordinates

Sequence positions of nucleotides/base pairs are 1- based and consist of three positions i, j, and k. Alternativly, four positions may be provided as a pair of two position ranges [i:j], and [k:l] using the '-' sign as delimiter within each range, i.e. i-j, and k-l.

4.2.2.1.6 Valid constraint commands

Below are resulting general cases that are considered *valid* constraints:

1. "Forcing a range of nucleotide positions to be paired":

Syntax:

```
F i 0 k [WHERE] [ORIENTATION]
```

Description:

Enforces the set of k consecutive nucleotides starting at position i to be paired. The optional loop type specifier [WHERE] allows to force them to appear as closing/enclosed pairs of certain types of loops.

2. "Forcing a set of consecutive base pairs to form":

Syntax:

```
Fijk [WHERE]
```

Description:

Enforces the base pairs $(i, j), \dots, (i+(k-1), j-(k-1))$ to form. The optional loop type specifier [WHERE] allows to specify in which loop context the base pair must appear.

3. "Prohibiting a range of nucleotide positions to be paired":

Syntax:

```
P i O k [WHERE]
```

Description:

Prohibit a set of k consecutive nucleotides to participate in base pairing, i.e. make these positions unpaired. The optional loop type specifier [WHERE] allows to force the nucleotides to appear within the loop of specific types.

4. "Probibiting a set of consecutive base pairs to form":

Syntax:

```
Pijk [WHERE]
```

Description:

Probibit the base pairs $(i, j), \dots, (i + (k - 1), j - (k - 1))$ to form. The optional loop type specifier [WHERE] allows to specify the type of loop they are disallowed to be the closing or an enclosed pair of.

4.2 File Formats 39

5. "Prohibiting two ranges of nucleotides to pair with each other": Syntax:

```
P i-j k-l [WHERE]
```

Description:

Prohibit any nucleotide $p \in [i:j]$ to pair with any other nucleotide $q \in [k:l]$. The optional loop type specifier [WHERE] allows to specify the type of loop they are disallowed to be the closing or an enclosed pair of.

6. "Enforce a loop context for a range of nucleotide positions": Syntax:

```
C i 0 k [WHERE]
```

Description:

This command enforces nucleotides to be unpaired similar to *prohibiting* nucleotides to be paired, as described above. It too marks the corresponding nucleotides to be unpaired, however, the [WHERE] flag can be used to enforce specific loop types the nucleotides must appear in.

7. "Remove pairs that conflict with a set of consecutive base pairs": Syntax:

Cijk

Description:

Remove all base pairs that conflict with a set of consecutive base pairs $(i, j), \ldots, (i + (k - 1), j - (k - 1))$. Two base pairs (i, j) and (p, q) conflict with each other if i , or <math>p < i < q < j.

8. "Allow a set of consecutive (non-canonical) base pairs to form":

Syntax:

A i j k [WHERE]

Description:

This command enables the formation of the consecutive base pairs $(i,j),\ldots,(i+(k-1),j-(k-1))$, no matter if they are *canonical*, or *non-canonical*. In contrast to the above $\mathbb F$ and $\mathbb W$ commands, which remove conflicting base pairs, the $\mathbb A$ command does not. Therefore, it may be used to allow *non-canonical* base pair interactions. Since the RNAlib does not contain free energy contributions E_{ij} for non-canonical base pairs (i,j), they are scored as the *maximum* of similar, known contributions. In terms of a *Nussinov* like scoring function the free energy of non-canonical base pairs is therefore estimated as

$$E_{ij} = \min \left[\max_{(i,k) \in \{GC, CG, AU, UA, GU, UG\}} E_{ik}, \max_{(k,j) \in \{GC, CG, AU, UA, GU, UG\}} E_{kj} \right].$$

The optional loop type specifier [WHERE] allows to specify in which loop context the base pair may appear.

9. "Apply pseudo free energy to a range of unpaired nucleotide positions":

Syntax:

Ei0ke

Description:

Use this command to apply a pseudo free energy of e to the set of k consecutive nucleotides, starting at position i. The pseudo free energy is applied only if these nucleotides are considered unpaired in the recursions, or evaluations, and is expected to be given in kcal/mol.

10. "Apply pseudo free energy to a set of consecutive base pairs":

Syntax

Eijke

Use this command to apply a pseudo free energy of e to the set of base pairs $(i, j), \ldots, (i + (k - 1), j - (k - 1))$. Energies are expected to be given in kcal/mol.

40 I/O Formats

4.2.2.1.7 Valid domain extensions commands

1. "Add ligand binding to unpaired motif (a.k.a. unstructured domains)":

```
Syntax:
UD m e [LOOP]
```

Description:

Add ligand binding to unpaired sequence motif m (given in IUPAC format, capital letters) with binding energy e in particular loop type(s).

```
Example:
UD AAA -5.0 A
```

The above example applies a binding free energy of -5kcal/mol for a motif AAA that may be present in all loop types.

4.3 Plotting

Create Plots of Secondary Structures, Feature Motifs, and Sequence Alignments

4.3.1 Producing secondary structure graphs

Produce a secondary structure graph in PostScript and write it to 'filename'.

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename'.

Produce a secondary structure graph in Graph Meta Language (gml) and write it to a file.

Produce a secondary structure graph in SStructView format.

4.3 Plotting 41

Produce a secondary structure plot in SVG format and write it to a file.

Produce a secondary structure plot for further editing in XRNA.

```
int rna_plot_type
```

Switch for changing the secondary structure layout algorithm.

Two low-level functions provide direct access to the graph lauyouting algorithms:

Calculate nucleotide coordinates for secondary structure plot the Simple way

See also

PS dot.h and naview.h for more detailed descriptions.

4.3.2 Producing (colored) dot plots for base pair probabilities

Produce a postscript dot-plot from two pair lists.

See also

PS_dot.h for more detailed descriptions.

4.3.3 Producing (colored) alignments

Produce PostScript sequence alignment color-annotated by consensus structure.

42 I/O Formats

Chapter 5

Basic Data Structures

- Sequence and Structure Data shows the most common types for sequence or structure data
- The 'Fold Compound' is the basic, central container for our implementations of prediction-, evaluation, and other algorithms
- Model Details provides the means to store the different model parameters

5.1 Sequence and Structure Data

See also

Secondary Structure Utilities

5.2 The 'Fold Compound'

See also

The Fold Compound

5.3 Model Details

See also

Fine-tuning of the Implemented Models

44 Basic Data Structures

Chapter 6

API Features

- RNAlib API v3.0
- Callback Functions
- Scripting Language interface(s)

6.1 RNAlib API v3.0

6.1.1 Introduction

With version 2.2 we introduce the new API that will take over the old one in the future version 3.0. By then, backwards compatibility will be broken, and third party applications using RNAlib need to be ported. This switch of API became necessary, since many new features found their way into the RNAlib where a balance between threadsafety and easy-to-use library functions is hard or even impossible to establish. Furthermore, many old functions of the library are present as slightly modified copies of themself to provide a crude way to overload functions.

Therefore, we introduce the new v3.0 API very early in our development stage such that developers have enough time to migrate to the new functions and interfaces. We also started to provide encapsulation of the RNAlib functions, data structures, typedefs, and macros by prefixing them with <code>vrna_</code> and <code>VRNA_</code>, respectively. Header files should also be included using the <code>ViennaRNA/</code> namespace, e.g.

#include <ViennaRNA/fold.h>

instead of just using

as required for RNAlib 1.x and 2.x.

This eases the work for programmers of third party applications that would otherwise need to put much effort into renaming functions and data types in their own implementations if their names appear in our library. Since we still provide backward compatibility up to the last version of RNAlib 2.x, this advantage may be fully exploited only starting from v3.0 which will be released in the future. However, our plan is to provide the possibility for an early switch-off mechanism of the backward compatibility in one of our next releases of ViennaRNA Package 2.x.

6.1.2 What are the major changes?

...

46 API Features

6.1.3 How to port your program to the new API

...

6.1.4 Some Examples using RNAlib API v3.0

Examples on how to use the new v3.0 API can be found in the examples c new API section.

6.2 Callback Functions

With the new RNAlib API v3.0 we introduce so-called callback mechanisms for several functions.

6.2.1 The purpose of Callback mechanisms

Using callback mechanisms, our library enables users not only to retrieve computed data without the need for parsing complicated data structures, but also allows one to tweak our implementation to do additional tasks without the requirement of a re-implementation of basic algorithms.

Our implementation of the callback mechanisms always follows the same scheme: The user:

- · defines a function that complies with the interface we've defined, and
- · passes a pointer to said function to our implementations

In addition to the specific arguments of our callback interfaces, virtually all callbacks receive an additional *pass-through-pointer* as their last argument. This enables one to:

- · encapsulate data, and
- · provide thread-safe operations,

since this pointer is simply passed through by our library functions. It may therefore hold the address of an arbitrary, user-defined data structure.

6.2 Callback Functions 47

6.2.2 List of available Callbacks

Below, you find an enumeration of the individual callback functions that are available in RNAlib.

Global vrna_boltzmann_sampling_callback (const char *stucture, void *data)

This function will be called for each secondary structure that has been successfully backtraced from the partition function DP matrices.

Global vrna_callback_free_auxdata (void *data)

This callback is supposed to free memory occupied by an auxiliary data structure. It will be called when the vrna_fold_compound_t is erased from memory through a call to vrna_fold_compound_free() and will be passed the address of memory previously bound to the vrna_fold_compound_t via vrna_fold_compound_add_auxdata().

Global vrna callback hc evaluate (int i, int j, int k, int l, unsigned char d, void *data)

This callback enables one to over-rule default hard constraints in secondary structure decompositions.

Global vrna callback recursion status (unsigned char status, void *data)

This function will be called to notify a third-party implementation about the status of a currently ongoing recursion. The purpose of this callback mechanism is to provide users with a simple way to ensure pre- and post conditions for auxiliary mechanisms attached to our implementations.

Global vrna callback sc backtrack (int i, int j, int k, int l, unsigned char d, void *data)

This callback enables one to add auxiliary base pairs in the backtracking steps of hairpin- and interior loops.

Global vrna_callback_sc_energy (int i, int j, int k, int l, unsigned char d, void *data)

This callback enables one to add (pseudo-)energy contributions to individual decompositions of the secondary structure.

Global vrna_callback_sc_exp_energy (int i, int j, int k, int l, unsigned char d, void *data)

This callback enables one to add (pseudo-)energy contributions to individual decompositions of the secondary structure (Partition function variant, i.e. contributions must be returned as Boltzmann factors).

Global vrna_callback_ud_energy (vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, void *data)

This function will be called to determine the additional energy contribution of a specific unstructured domain, e.g. the binding free energy of some ligand.

Global vrna_callback_ud_exp_energy (vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, void *data)

This function will be called to determine the additional energy contribution of a specific unstructured domain, e.g. the binding free energy of some ligand (Partition function variant, i.e. the Boltzmann factors instead of actual free energies).

Global vrna_callback_ud_exp_production (vrna_fold_compound_t *vc, void *data)

The production rule for the unstructured domain grammar extension (Partition function variant)

Global vrna_callback_ud_probs_add (vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, FLT← _OR_DBL exp_energy, void *data)

A callback function to store equilibrium probabilities for the unstructured domain feature

Global vrna_callback_ud_probs_get (vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, int motif, void *data)

A callback function to retrieve equilibrium probabilities for the unstructured domain feature

Global vrna_callback_ud_production (vrna_fold_compound_t *vc, void *data)

The production rule for the unstructured domain grammar extension

Global vrna_mfe_window_callback (int start, int end, const char *structure, float en, void *data)

This function will be called for each hit in a sliding window MFE prediction.

Global vrna_probs_window_callback (FLT_OR_DBL *pr, int pr_size, int i, int max, unsigned int type, void *data)

This function will be called for each probability data set in the sliding window probability computation implementation of vrna_probs_window(). The argument type specifies the type of probability that is passed to this function.

48 API Features

Global vrna_subopt_callback (const char *stucture, float energy, void *data)

This function will be called for each suboptimal secondary structure that is successfully backtraced.

6.3 Scripting Language interface(s)

6.3.1 Introduction

For an easy integration into scripting languages, we provide an automatically generated interface to the RNAlib C-library, generated with SWIG.

6.3.2 Function Renaming

To provide a namespace-like separation of function symbols from our C library and third-party code, we use the prefix <code>vrna_</code> or <code>VRNA_</code> whenever possible. This, however, is not necessary for the scripting language interface, as it uses the separate namespace or package <code>RNA</code> anyway. Consequently, symbols that appear to have the <code>vrna_</code> or <code>VRNA_</code> prefix in the C-library have the corresponding prefix stripped away.

```
For instance, the C code

mfe = vrna_fold(sequence, structure);

translates to

my ($structure, $mfe) = RNA::fold($sequence)

in the Perl 5 interface, and

structure mfe = RNA fold(sequence)
```

for Python 2/3. Note, that in this example we also make use of the possibility to return multiple data at once in the scripting language, while the C library function uses additional parameters to return multiple data.

Functions that are dedicated to work on specific data structures only, e.g. the vrna_fold_compound_t, are usually not exported at all. Instead, they are attached as object methods of a corresponding class (see Object oriented Interface for Data Structures for detailed information).

6.3.2.1 Global Variables

For the Python interface(s) SWIG places global variables of the C-library into an additional namespace cvar. For instance, changing the global temperature variable thus becomes

RNA.cvar.temperature = 25

6.3.3 Object oriented Interface for Data Structures

For data structures, typedefs, and enumerations the <code>vrna_prefixes</code> are dropped as well, together with their suffixes <code>_s, _t, and _e, respectively.</code> Furthermore, data structures are usually transformed into classes and relevant functions of the C-library are attached as methods.

6.3.4 Examples

Examples on the basic usage of the scripting language interfaces can be found in the scripting_perl_examples and scripting_python_examples section.

6.3.5 SWIG generated Wrapper notes

Special notes on how functions, structures, enums, and macro definitions are actually wrapped, can be found below

Global vrna_aln_conservation_col (const char **alignment, const vrna_md_t *md_p, unsigned int options)

This function is available in an overloaded form where the last two parameters may be omitted, indicating md = *NULL*, and options = VRNA_MEASURE_SHANNON_ENTROPY, respectively.

Global vrna_aln_conservation_struct (const char **alignment, const char *structure, const vrna_md_← t *md)

This function is available in an overloaded form where the last parameter may be omitted, indicating md = NULL

Global vrna backtrack5 (vrna fold compound t *fc, unsigned int length, char *structure)

This function is attached as overloaded method **backtrack()** to objects of type *fold_compound* with default parameter length equal to the total length of the RNA.

Global vrna db flatten (char *structure, unsigned int options)

This function flattens an input structure string in-place! The second parameter is optional and defaults to VRNA BRACKETS DEFAULT.

An overloaded version of this function exists, where an additional second parameter can be passed to specify the target brackets, i.e. the type of matching pair characters all brackets will be flattened to. Therefore, in the scripting language interface this function is a replacement for vrna db flatten to().

Global vrna_db_flatten_to (char *string, const char target[3], unsigned int options)

This function is available as an overloaded version of vrna db flatten()

Global vrna_db_pk_remove (const char *structure, unsigned int options)

This function is available as an overloaded function $db_pk_remove()$ where the optional second parameter options defaults to #VRNA BRACKET ANY.

Global vrna_ensemble_defect (vrna_fold_compound_t *fc, const char *structure)

This function is attached as method **ensemble_defect()** to objects of type *fold_compound*

Global vrna_enumerate_necklaces (const unsigned int *type_counts)

This function is available as global function **enumerate_necklaces()** which accepts lists input, an produces list of lists output.

Global vrna_eval_circ_consensus_structure (const char **alignment, const char *structure)

This function is available through an overloadeded version of vrna_eval_circ_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

Global vrna_eval_circ_consensus_structure_v (const char **alignment, const char *structure, int verbosity_level, FILE *file)

This function is available through an overloaded version of vrna_eval_circ_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument. The last two arguments are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

Global vrna_eval_circ_gquad_consensus_structure (const char **alignment, const char *structure)

This function is available through an overloadeded version of vrna_eval_circ_gquad_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

Global vrna_eval_circ_gquad_consensus_structure_v (const char **alignment, const char *structure, int verbosity level, FILE *file)

This function is available through an overloaded version of vrna_eval_circ_gquad_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument. The last two arguments are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

Global vrna eval circ gquad structure (const char *string, const char *structure)

In the target scripting language, this function serves as a wrapper for vrna_eval_circ_gquad_structure_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA_VERBOSITY_QUIET and NULL, respectively.

50 API Features

Global vrna_eval_circ_gquad_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)

This function is available through an overloaded version of vrna_eval_circ_gquad_structure(). The last two arguments for this function are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

Global vrna_eval_circ_structure (const char *string, const char *structure)

In the target scripting language, this function serves as a wrapper for vrna_eval_circ_structure_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA_VERBOSITY_QUIET and NULL, respectively.

Global vrna_eval_circ_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)

This function is available through an overloaded version of vrna_eval_circ_structure(). The last two arguments for this function are optional and default to VRNA VERBOSITY QUIET and NULL, respectively.

Global vrna_eval_consensus_structure_pt_simple (const char **alignment, const short *pt)

This function is available through an overloadeded version of vrna_eval_structure_pt_simple(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

Global vrna_eval_consensus_structure_simple (const char **alignment, const char *structure)

This function is available through an overloadeded version of vrna_eval_structure_simple(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

Global vrna_eval_consensus_structure_simple_v (const char **alignment, const char *structure, int verbosity_level, FILE *file)

This function is available through an overloaded version of vrna_eval_structure_simple(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument. The last two arguments are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

Global vrna_eval_consensus_structure_simple_verbose (const char **alignment, const char *structure, FILE *file)

This function is not available. Use vrna_eval_consensus_structure_simple_v() instead!

Global vrna_eval_covar_structure (vrna_fold_compound_t *vc, const char *structure)

This function is attached as method eval covar structure() to objects of type fold compound

Global vrna_eval_gquad_consensus_structure (const char **alignment, const char *structure)

This function is available through an overloadeded version of vrna_eval_gquad_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

Global vrna_eval_gquad_consensus_structure_v (const char **alignment, const char *structure, int verbosity level, FILE *file)

This function is available through an overloaded version of vrna_eval_gquad_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument. The last two arguments are optional and default to VRNA VERBOSITY QUIET and NULL, respectively.

Global vrna_eval_gquad_structure (const char *string, const char *structure)

In the target scripting language, this function serves as a wrapper for vrna_eval_gquad_structure_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA VERBOSITY QUIET and NULL, respectively.

Global vrna_eval_gquad_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)

This function is available through an overloaded version of vrna_eval_gquad_structure(). The last two arguments for this function are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

Global vrna_eval_hp_loop (vrna_fold_compound_t *fc, int i, int j)

This function is attached as method eval_hp_loop() to objects of type fold_compound

Global vrna eval int loop (vrna fold compound t *fc, int i, int j, int k, int l)

This function is attached as method eval_int_loop() to objects of type fold_compound

Global vrna_eval_loop_pt (vrna_fold_compound_t *vc, int i, const short *pt)

This function is attached as method eval_loop_pt() to objects of type fold_compound

Global vrna_eval_move (vrna_fold_compound_t *vc, const char *structure, int m1, int m2)

This function is attached as method eval move() to objects of type fold compound

Global vrna_eval_move_pt (vrna_fold_compound_t *vc, short *pt, int m1, int m2)

This function is attached as method eval_move_pt() to objects of type fold_compound

Global vrna eval structure (vrna fold compound t *vc, const char *structure)

This function is attached as method eval_structure() to objects of type fold_compound

Global vrna eval structure pt (vrna fold compound t *vc, const short *pt)

This function is attached as method eval structure pt() to objects of type fold compound

Global vrna eval structure pt simple (const char *string, const short *pt)

In the target scripting language, this function serves as a wrapper for vrna_eval_structure_pt_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA VERBOSITY QUIET and NULL, respectively.

Global vrna_eval_structure_pt_verbose (vrna_fold_compound_t *vc, const short *pt, FILE *file)

This function is attached as method eval_structure_pt_verbose() to objects of type fold_compound

Global vrna_eval_structure_simple (const char *string, const char *structure)

In the target scripting language, this function serves as a wrapper for vrna_eval_structure_simple_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to vrna_eval_structure_simple_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to vrna_eval_structure_simple_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to vrna_eval_structure_simple_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to vrna_eval_structure_simple_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to vrna_eval_structure_simple_v() and vrna_eval_struc

Global vrna_eval_structure_simple_v (const char *string, const char *structure, int verbosity_level, FILE *file)

This function is available through an overloaded version of vrna_eval_structure_simple(). The last two arguments for this function are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

Global vrna_eval_structure_simple_verbose (const char *string, const char *structure, FILE *file)

This function is not available. Use vrna_eval_structure_simple_v() instead!

$\textbf{Global vrna_eval_structure_verbose} \ (\textbf{vrna_fold_compound_t} \ *\textbf{vc}, \ \textbf{const char} \ *\textbf{structure}, \ \textbf{FILE} \ *\textbf{file})$

This function is attached as method eval_structure_verbose() to objects of type fold_compound

Global vrna_exp_params_rescale (vrna_fold_compound_t *vc, double *mfe)

This function is attached to vrna_fc_s objects as overloaded exp_params_rescale() method.

When no parameter is passed to this method, the resulting action is the same as passing *NULL* as second parameter to <u>vrna_exp_params_rescale()</u>, i.e. default scaling of the partition function. Passing an energy in kcal/mol, e.g. as retrieved by a previous call to the *mfe()* method, instructs all subsequent calls to scale the partition function accordingly.

Global vrna exp params reset (vrna fold compound t *vc, vrna md t *md p)

This function is attached to vrna_fc_s objects as overloaded exp_params_reset() method.

When no parameter is passed to this method, the resulting action is the same as passing *NULL* as second parameter to vrna_exp_params_reset(), i.e. global default model settings are used. Passing an object of type vrna md s resets the fold compound according to the specifications stored within the vrna md s object.

Class vrna fc s

This data structure is wrapped as an object **fold_compound** with several related functions attached as methods.

A new **fold_compound** can be obtained by calling one of its constructors:

- fold_compound(seq) Initialize with a single sequence, or two concatenated sequences separated by an ampersand character '&' (for cofolding)
- fold_compound(aln) Initialize with a sequence alignment aln stored as a list of sequences (with gap characters)

The resulting object has a list of attached methods which in most cases directly correspond to functions that mainly operate on the corresponding C data structure:

52 API Features

- type() Get the type of the fold_compound (See vrna_fc_type_e)
- · length() Get the length of the sequence(s) or alignment stored within the fold compound

Global vrna_file_commands_apply (vrna_fold_compound_t *vc, const char *filename, unsigned int options)

This function is attached as method file commands apply() to objects of type fold compound

Global vrna_file_msa_detect_format (const char *filename, unsigned int options)

This function exists as an overloaded version where the options parameter may be omitted! In that case, the options parameter defaults to VRNA_FILE_FORMAT_MSA_DEFAULT.

Global vrna_file_msa_read (const char *filename, char ***names, char ***aln, char **id, char **structure, unsigned int options)

In the target scripting language, only the first and last argument, filename and options, are passed to the corresponding function. The other arguments, which serve as output in the C-library, are available as additional return values. Hence, a function call in python may look like this:

Global vrna_file_msa_read_record (FILE *fp, char ***names, char ***aln, char **id, char **structure, unsigned int options)

In the target scripting language, only the first and last argument, fp and options, are passed to the corresponding function. The other arguments, which serve as output in the C-library, are available as additional return values. Hence, a function call in python may look like this:

Global vrna_file_msa_write (const char *filename, const char **names, const char **aln, const char *id, const char *structure, const char *source, unsigned int options)

In the target scripting language, this function exists as a set of overloaded versions, where the last four parameters may be omitted. If the options parameter is missing the options default to (VRNA_FILE_FORMAT_MSA_STOCKHOLM | VRNA_FILE_FORMAT_MSA_APPEND).

Global vrna_file_PS_aln (const char *filename, const char **seqs, const char **names, const char *structure, unsigned int columns)

This function is available as overloaded function $file_PS_aln()$ with three additional parameters start, end, and offset before the columns argument. Thus, it resembles the $vrna_file_PS_aln_slice()$ function. The last four arguments may be omitted, indicating the default of start = 0, end = 0, offset = 0, and columns = 60.

Global vrna_file_PS_aln_slice (const char *filename, const char **seqs, const char **names, const char *structure, unsigned int start, unsigned int end, int offset, unsigned int columns)

This function is available as overloaded function $file_PS_aln()$ where the last four parameter may be omitted, indicating start = 0, end = 0, end = 0, end = 0, and end = 0.

Global vrna hc add from db (vrna fold compound t *vc, const char *constraint, unsigned int options)

This function is attached as method hc_add_from_db() to objects of type fold_compound

Global vrna_hc_init (vrna_fold_compound_t *vc)

This function is attached as method **hc_init()** to objects of type *fold_compound*

Global vrna_maximum_matching (vrna_fold_compound_t *fc)

This function is attached as method maximum_matching() to objects of type fold_compound (i.e. vrna_fold_compound_t).

Global vrna_maximum_matching_simple (const char *sequence)

This function is available as global function maximum_matching().

Class vrna md s

This data structure is wrapped as an object **md** with multiple related functions attached as methods.

A new set of default parameters can be obtained by calling the constructure of md:

md() – Initialize with default settings

The resulting object has a list of attached methods which directly correspond to functions that mainly operate on the corresponding C data structure:

- reset() vrna_md_set_default()
- set from globals() set model details()
- option_string() vrna_md_option_string()

Note, that default parameters can be modified by directly setting any of the following global variables. Internally, getting/setting default parameters using their global variable representative translates into calls of the following functions, therefore these wrappers for these functions do not exist in the scripting language interface(s):

Global vrna_MEA (vrna_fold_compound_t *fc, double gamma, float *mea)

This function is attached as overloaded method **MEA**(gamma = 1.) to objects of type *fold_compound*. Note, that it returns the MEA structure and MEA value as a tuple (MEA structure, MEA)

Global vrna_MEA_from_plist (vrna_ep_t *plist, const char *sequence, double gamma, vrna_md_t *md, float *mea)

This function is available as overloaded function **MEA_from_plist**(gamma = 1., md = NULL). Note, that it returns the MEA structure and MEA value as a tuple (MEA structure, MEA)

Global vrna mean bp distance (vrna fold compound t *vc)

This function is attached as method mean_bp_distance() to objects of type fold_compound

Global vrna_mfe (vrna_fold_compound_t *vc, char *structure)

This function is attached as method **mfe()** to objects of type fold_compound

Global vrna mfe dimer (vrna fold compound t *vc, char *structure)

This function is attached as method mfe dimer() to objects of type fold compound

Global vrna mfe window (vrna fold compound t *vc, FILE *file)

This function is attached as method **mfe_window()** to objects of type fold_compound

Global vrna neighbors (vrna fold compound t *vc, const short *pt, unsigned int options)

This function is attached as an overloaded method *neighbors()* to objects of type *fold_compound*. The optional parameter options defaults to VRNA MOVESET DEFAULT if it is omitted.

Global vrna_params_load (const char fname[], unsigned int options)

This function is available as overloaded function **params_load**(fname="", options=VRNA_PARAMETER_FORMAT_DEFAULT). Here, the empty filename string indicates to load default RNA parameters, i.e. this is equivalent to calling vrna_params_load_defaults().

Global vrna_params_load_defaults (void)

This function is available as overloaded function params_load().

Global vrna params load DNA Mathews1999 (void)

This function is available as function params load DNA Mathews1999().

Global vrna params load DNA Mathews2004 (void)

This function is available as function params_load_DNA_Mathews2004().

Global vrna_params_load_from_string (const char *string, const char *name, unsigned int options)

This function is available as overloaded function params_load_from_string(string, name="", options=VRNA_PARAMETER_FOR

Global vrna params load RNA Andronescu2007 (void)

This function is available as function params_load_RNA_Andronescu2007().

Global vrna params load RNA Langdon2018 (void)

This function is available as function params load RNA Langdon2018().

Global vrna_params_load_RNA_misc_special_hairpins (void)

This function is available as function params_load_RNA_misc_special_hairpins().

54 API Features

Global vrna params load RNA Turner1999 (void)

This function is available as function params_load_RNA_Turner1999().

Global vrna params load RNA Turner2004 (void)

This function is available as function params load RNA Turner2004().

Global vrna_params_reset (vrna_fold_compound_t *vc, vrna_md_t *md_p)

This function is attached to vrna_fc_s objects as overloaded params_reset() method.

When no parameter is passed to this method, the resulting action is the same as passing *NULL* as second parameter to vrna_params_reset(), i.e. global default model settings are used. Passing an object of type vrna md s resets the fold compound according to the specifications stored within the vrna md s object.

Global vrna params save (const char fname[], unsigned int options)

This function is available as overloaded function params save(fname, options=VRNA PARAMETER FORMAT DEFAULT).

Global vrna params subst (vrna fold compound t *vc, vrna param t *par)

This function is attached to vrna_fc_s objects as params_subst() method.

Global vrna path (vrna fold compound t *vc, short *pt, unsigned int steps, unsigned int options)

This function is attached as an overloaded method path() to objects of type $fold_compound$. The optional parameter options defaults to VRNA_PATH_DEFAULT if it is omitted.

Global vrna_path_direct (vrna_fold_compound_t *fc, const char *s1, const char *s2, vrna_path_options_t options)

This function is attached as an overloaded method *path_direct()* to objects of type *fold_compound*. The optional parameter options defaults to *NULL* if it is omitted.

Global vrna_path_direct_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int maxE, vrna_← path_options_t options)

This function is attached as an overloaded method $path_direct()$ to objects of type $fold_compound$. The optional parameter maxE defaults to #INT_MAX - 1 if it is omitted, while the optional parameter options defaults to NULL. In case the function did not find a path with $E_{saddle} < E_{max}$ it returns an empty list.

$\textbf{Global vrna_path_findpath} \ (\textbf{vrna_fold_compound_t} \ *fc, \ const \ char \ *s1, \ const \ char \ *s2, \ int \ width)$

This function is attached as an overloaded method *path_findpath()* to objects of type *fold_compound*. The optional parameter width defaults to 1 if it is omitted.

Global vrna path findpath saddle (vrna fold compound t *fc, const char *s1, const char *s2, int width)

This function is attached as an overloaded method $path_findpath_saddle()$ to objects of type $fold_compound$. The optional parameter width defaults to 1 if it is omitted.

Global vrna_path_findpath_saddle_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width, int maxE)

This function is attached as an overloaded method $path_findpath_saddle()$ to objects of type $fold_compound$. The optional parameter width defaults to 1 if it is omitted, while the optional parameter maxE defaults to INF. In case the function did not find a path with $E_{saddle} < E_{max}$ the function returns a NULL object, i.e. undef for Perl and None for Python.

Global vrna_path_findpath_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width, int maxE)

This function is attached as an overloaded method $path_findpath()$ to objects of type $fold_compound$. The optional parameter width defaults to 1 if it is omitted, while the optional parameter maxE defaults to INF. In case the function did not find a path with $E_{saddle} < E_{max}$ the function returns an empty list.

Global vrna_path_gradient (vrna_fold_compound_t *vc, short *pt, unsigned int options)

This function is attached as an overloaded method *path_gradient()* to objects of type *fold_compound*. The optional parameter options defaults to VRNA_PATH_DEFAULT if it is omitted.

Global vrna_path_options_findpath (int width, unsigned int type)

This function is available as overloaded function $path_options_findpath()$. The optional parameter width defaults to 10 if omitted, while the optional parameter type defaults to VRNA_PATH_TYPE_DOT_BRACKET.

Global vrna path random (vrna fold compound t *vc, short *pt, unsigned int steps, unsigned int options)

This function is attached as an overloaded method <code>path_gradient()</code> to objects of type <code>fold_compound</code>. The optional parameter <code>options</code> defaults to <code>VRNA PATH DEFAULT</code> if it is omitted.

Global vrna_pbacktrack (vrna_fold_compound_t *fc)

This function is attached as overloaded method **pbacktrack()** to objects of type *fold_compound*. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack5 (vrna_fold_compound_t *fc, unsigned int length)

This function is attached as overloaded method **pbacktrack5()** to objects of type *fold_compound*. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack5_cb (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, vrna_boltzmann_sampling_callback *cb, void *data, unsigned int options)

This function is attached as overloaded method **pbacktrack5()** to objects of type *fold_compound* where the last argument options is optional with default value options = VRNA_PBACKTRACK_DEFAULT. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack5_num (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, unsigned int options)

This function is attached as overloaded method **pbacktrack5()** to objects of type *fold_compound* where the last argument options is optional with default value options = VRNA_PBACKTRACK_DEFAULT. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack5_resume (vrna_fold_compound_t *vc, unsigned int num_samples, unsigned int length, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

This function is attached as overloaded method **pbacktrack5()** to objects of type *fold_compound*. In addition to the list of structures, this function also returns the nr_mem data structure as first element. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack5_resume_cb (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, vrna_boltzmann_sampling_callback *cb, void *data, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

This function is attached as overloaded method **pbacktrack5()** to objects of type *fold_compound*. In addition to the number of structures backtraced, this function also returns the nr_mem data structure as first element. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack_cb (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_boltzmann_compound_t *fc, unsigned int num_samples, unsigned i

This function is attached as overloaded method **pbacktrack()** to objects of type *fold_compound* where the last argument options is optional with default value options = VRNA_PBACKTRACK_DEFAULT. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack_num (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int options)

This function is attached as overloaded method **pbacktrack()** to objects of type *fold_compound* where the last argument options is optional with default value options = VRNA_PBACKTRACK_DEFAULT. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack_resume (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_← pbacktrack_mem_t *nr_mem, unsigned int options)

This function is attached as overloaded method **pbacktrack()** to objects of type *fold_compound*. In addition to the list of structures, this function also returns the nr_mem data structure as first element. See also Python Examples - Boltzmann Sampling

Global vrna_pbacktrack_resume_cb (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_← boltzmann_sampling_callback *cb, void *data, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

This function is attached as overloaded method **pbacktrack()** to objects of type *fold_compound*. In addition to the number of structures backtraced, this function also returns the nr_mem data structure as first element. See also Python Examples - Boltzmann Sampling

56 API Features

Global vrna_pf (vrna_fold_compound_t *vc, char *structure)

This function is attached as method pf() to objects of type fold compound

Global vrna_pf_dimer (vrna_fold_compound_t *vc, char *structure)

This function is attached as method **pf_dimer()** to objects of type fold_compound

Global vrna_positional_entropy (vrna_fold_compound_t *fc)

This function is attached as method positional entropy() to objects of type fold compound

Global vrna_pr_energy (vrna_fold_compound_t *vc, double e)

This function is attached as method **pr_energy()** to objects of type fold compound

Global vrna pr structure (vrna fold compound t *fc, const char *structure)

This function is attached as method pr_structure() to objects of type fold_compound

Global vrna rotational symmetry (const char *string)

This function is available as global function **rotational_symmetry()**. See vrna_rotational_symmetry_pos() for details.

Global vrna_rotational_symmetry_db (vrna_fold_compound_t *fc, const char *structure)

This function is attached as method rotational_symmetry_db() to objects of type fold_compound (i. ← e. vrna fold compound t). See vrna rotational symmetry db pos() for details.

Global vrna_rotational_symmetry_db_pos (vrna_fold_compound_t *fc, const char *structure, unsigned int **positions)

This function is attached as method **rotational_symmetry_db()** to objects of type fold_compound (i. ← e. vrna_fold_compound_t). Thus, the first argument must be omitted. In contrast to our C-implementation, this function doesn't simply return the order of rotational symmetry of the secondary structure, but returns the list position of cyclic permutation shifts that result in a rotationally symmetric structure. The length of the list then determines the order of rotational symmetry.

Global vrna_rotational_symmetry_num (const unsigned int *string, size_t string_length)

This function is available as global function **rotational_symmetry()**. See vrna_rotational_symmetry_pos() for details. Note, that in the target language the length of the list string is always known a-priori, so the parameter string_length must be omitted.

Global vrna_rotational_symmetry_pos (const char *string, unsigned int **positions)

This function is available as overloaded global function **rotational_symmetry()**. It merges the functionalities of vrna_rotational_symmetry(), vrna_rotational_symmetry_pos(), vrna_rotational_symmetry_num(), and vrna_rotational_symmetry_pos_num(). In contrast to our C-implementation, this function doesn't return the order of rotational symmetry as a single value, but returns a list of cyclic permutation shifts that result in a rotationally symmetric string. The length of the list then determines the order of rotational symmetry.

Global vrna_rotational_symmetry_pos_num (const unsigned int *string, size_t string_length, unsigned int **positions)

This function is available as global function **rotational_symmetry()**. See vrna_rotational_symmetry_pos() for details. Note, that in the target language the length of the list string is always known a-priori, so the parameter string_length must be omitted.

Global vrna_sc_add_bp (vrna_fold_compound_t *vc, int i, int j, FLT_OR_DBL energy, unsigned int options)

This function is attached as an overloaded method **sc_add_bp()** to objects of type *fold_compound*. The method either takes arguments for a single base pair (i,j) with the corresponding energy value:

Global vrna sc add bt (vrna fold compound t *vc, vrna callback sc backtrack *f)

This function is attached as method sc_add_bt() to objects of type fold_compound

Global vrna_sc_add_data (vrna_fold_compound_t *vc, void *data, vrna_callback_free_auxdata *free_data)

This function is attached as method sc_add_data() to objects of type fold_compound

Global vrna_sc_add_exp_f (vrna_fold_compound_t *vc, vrna_callback_sc_exp_energy *exp_f)

This function is attached as method sc_add_exp_f() to objects of type fold_compound

Global vrna_sc_add_f (vrna_fold_compound_t *vc, vrna_callback_sc_energy *f)

This function is attached as method **sc_add_f()** to objects of type *fold_compound*

Global vrna_sc_add_hi_motif (vrna_fold_compound_t *vc, const char *seq, const char *structure, FLT_← OR_DBL energy, unsigned int options)

This function is attached as method sc add hi motif() to objects of type fold compound

Global vrna_sc_add_SHAPE_deigan (vrna_fold_compound_t *vc, const double *reactivities, double m, double b, unsigned int options)

This function is attached as method sc_add_SHAPE_deigan() to objects of type fold_compound

Global vrna_sc_add_SHAPE_deigan_ali (vrna_fold_compound_t *vc, const char **shape_files, const int *shape file association, double m, double b, unsigned int options)

This function is attached as method sc add SHAPE deigan ali() to objects of type fold compound

Global vrna_sc_add_SHAPE_zarringhalam (vrna_fold_compound_t *vc, const double *reactivities, double b, double default_value, const char *shape conversion, unsigned int options)

This function is attached as method sc add SHAPE zarringhalam() to objects of type fold compound

Global vrna sc add up (vrna fold compound t *vc, int i, FLT OR DBL energy, unsigned int options)

This function is attached as an overloaded method $sc_add_up()$ to objects of type $fold_compound$. The method either takes arguments for a single nucleotide i with the corresponding energy value:

Global vrna_sc_init (vrna_fold_compound_t *vc)

This function is attached as method sc_init() to objects of type fold compound

Global vrna sc remove (vrna fold compound t *vc)

This function is attached as method sc_remove() to objects of type fold_compound

Global vrna_sc_set_bp (vrna_fold_compound_t *vc, const FLT_OR_DBL **constraints, unsigned int options)

This function is attached as method sc set bp() to objects of type fold compound

Global vrna_sc_set_up (vrna_fold_compound_t *vc, const FLT_OR_DBL *constraints, unsigned int options)

This function is attached as method sc_set_up() to objects of type fold_compound

Global vrna subopt (vrna fold compound t *vc, int delta, int sorted, FILE *fp)

This function is attached as method subopt() to objects of type fold_compound

Global vrna_subopt_cb (vrna_fold_compound_t *vc, int delta, vrna_subopt_callback *cb, void *data)

This function is attached as method subopt cb() to objects of type fold compound

Global vrna_subopt_zuker (vrna_fold_compound_t *vc)

This function is attached as method subopt_zuker() to objects of type fold_compound

Global vrna_ud_remove (vrna_fold_compound_t *vc)

This function is attached as method ud_remove() to objects of type fold_compound

 $\textbf{Global vrna_ud_set_data} \ (\textbf{vrna_fold_compound_t} \ *\textbf{vc}, \ \textbf{void} \ *\textbf{data}, \ \textbf{vrna_callback_free_auxdata} \ *\textbf{free_cb})$

This function is attached as method ud set data() to objects of type fold compound

Global vrna_ud_set_exp_prod_rule_cb (vrna_fold_compound_t *vc, vrna_callback_ud_exp_production *pre_cb, vrna_callback_ud_exp_energy *exp_e_cb)

This function is attached as method ud set exp prod rule cb() to objects of type fold compound

Global vrna_ud_set_prob_cb (vrna_fold_compound_t *vc, vrna_callback_ud_probs_add *setter, vrna_← callback_ud_probs_get *getter)

This function is attached as method ud_set_prob_cb() to objects of type fold_compound

Global vrna_ud_set_prod_rule_cb (vrna_fold_compound_t *vc, vrna_callback_ud_production *pre_cb, vrna_callback_ud_energy *e_cb)

This function is attached as method ud set prod rule cb() to objects of type fold compound

58 API Features

Chapter 7

Additional Utilities

60 Additional Utilities

Chapter 8

Examples

- C Examples
- · Perl5 Examples
- Python Examples

8.1 C Examples

8.1.1 Hello World Examples

helloworld mfe.c

The following is an example showing the minimal requirements to compute the Minimum Free Energy (MFE) and corresponding secondary structure of an RNA sequence

```
#include <stdlib.h>
#include <stdio.h>
#include <string.h>
#include <ViennaRNA/fold.h>
#include <ViennaRNA/utils/basic.h>
int
main()
{
    /* The RNA sequence */
    char *seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA";
    /* allocate memory for MFE structure (length + 1) */
    char *structure = (char *)vrna_alloc(sizeof(char) * (strlen(seq) + 1));
    /* predict Minmum Free Energy and corresponding secondary structure */
    float mfe = vrna_fold(seq, structure);
    /* print sequence, structure and MFE */
    printf("%s\n%s [ %6.2f ]\n", seq, structure, mfe);
    /* cleanup memory */
    free(structure);
    return 0;
}
```

See also

 $\verb|examples/helloworld_mfe.c| in the source code tarball|$

62 Examples

helloworld_mfe_comparative.c

Instead of using a single sequence as done above, this example predicts a consensus structure for a multiple sequence alignment

```
#include <stdlib.h>
#include <stdio.h>
#include <string.h>
#include <ViennaRNA/alifold.h>
#include <ViennaRNA/utils/basic.h>
#include <ViennaRNA/utils/alignments.h>
int
main()
  /* The RNA sequence alignment */
  const char *sequences[] = {
    "CUGCCUCACAACGUUUGUGCCUCAGUUACCCGUAGAUGUAGUGAGGGU", "CUGCCUCACAACAUUUGUGCCUCAGUUACUCAUAGAUGUAGUGAGGGU",
     "--CUCGACACCACU--GCCUCGGUUACCCAUCGGUGCAGUGCGGGU",
    NULL /\star indicates end of alignment \star/
  /* compute the consensus sequence */
                *cons = consensus (sequences);
  /\star allocate memory for MFE consensus structure (length + 1) \star/
               *structure = (char *)vrna_alloc(sizeof(char) * (strlen(sequences[0]) + 1));
  /\star predict Minmum Free Energy and corresponding secondary structure \star
              mfe = vrna_alifold(sequences, structure);
  /* print consensus sequence, structure and MFE */
  printf("%s\n%s [ %6.2f ]\n", cons, structure, mfe);
   /* cleanup memory */
  free (cons);
  free (structure);
  return 0;
```

See also

examples/helloworld_mfe_comparative.c in the source code tarball

helloworld_probabilities.c

This example shows how to compute the partition function and base pair probabilities with minimal implementation effort.

```
#include <stdlib.h>
#include <stdio.h>
#include <string.h>
#include <ViennaRNA/fold.h>
#include <ViennaRNA/part_func.h>
#include <ViennaRNA/utils/basic.h>
int
main()
  /\star The RNA sequence \star/
              *seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA";
  /\star allocate memory for pairing propensity string (length + 1) \star/
              *propensity = (char *)vrna_alloc(sizeof(char) * (strlen(seq) + 1));
  /* pointers for storing and navigating through base pair probabilities */
vrna_ep_t *ptr, *pair_probabilities = NULL;
float en = vrna_pf_fold(seq, propensity, &pair_probabilities);
   /\star print sequence, pairing propensity string and ensemble free energy \star/
  printf("%s\n", seq, propensity, en);
  /* print all base pairs with probability above 50%
for (ptr = pair_probabilities; ptr->i != 0; ptr++)
   if (ptr->p > 0.5)
       printf("p(%d, %d) = %g\n", ptr->i, ptr->j, ptr->p);
   /* cleanup memory */
   free (pair_probabilities);
  free (propensity);
  return 0;
```

See also

examples/helloworld_probabilities.c in the source code tarball

8.1 C Examples 63

8.1.2 First Steps with the Fold Compound

fold_compound_mfe.c

Instead of calling the simple MFE folding interface vrna_fold(), this example shows how to first create a vrna_fold_compound_t container with the RNA sequence to finally compute the MFE using this container. This is especially useful if non-default model settings are applied or the dynamic programming (DP) matrices of the MFE prediction are required for post-processing operations, or other tasks on the same sequence will be performed.

```
#include <stdio.h>
#include <ViennaRNA/fold_compound.h>
#include <ViennaRNA/utils/basic.h>
#include <ViennaRNA/utils/strings.h>
#include <ViennaRNA/mfe.h>
main()
  /* initialize random number generator */
  vrna init rand():
  /* Generate a random sequence of 50 nucleotides */
                              *seq = vrna_random_string(50, "ACGU");
  /* Create a fold compound for the sequence */
  vrna_fold_compound_t *fc = vrna_fold_compound(seq, NULL, VRNA_OPTION_DEFAULT);
/* allocate memory for MFE structure (length + 1) */
char *structure = (char *) vrna_alloc(sizeof(char) * (strlen(seq) + 1));
  /* predict Minmum Free Energy and corresponding secondary structure \star/float mfe = vrna_mfe(fc, structure);
  /* print sequence, structure and MFE */
  printf("%s\n%s [ %6.2f ]\n", seq, structure, mfe);
   /* cleanup memory */
  free (seq);
  free (structure);
  vrna_fold_compound_free(fc);
  return 0;
```

See also

examples/fold_compound_mfe.c in the source code tarball

fold_compound_md.c

In the following, we change the model settings (model details) to a temperature of 25 Degree Celcius, and activate G-Quadruplex precition.

```
#include <stdlib.h>
#include <stdio.h>
#include <string.h>
#include <ViennaRNA/model.h>
#include <ViennaRNA/fold_compound.h>
#include <ViennaRNA/utils/basic.h>
#include <ViennaRNA/utils/strings.h>
#include <ViennaRNA/mfe.h>
main()
  /* initialize random number generator */
  vrna init rand();
  /* Generate a random sequence of 50 nucleotides */
           *seq = vrna_random_string(50, "ACGU");
  /* allocate memory for MFE structure (length + 1) */
  char
           *structure = (char *) vrna_alloc(sizeof(char) * (strlen(seq) + 1));
  /* create a new model details structure to store the Model Settings */
  vrna md t md;
  /* ALWAYS set default model settings first! */
  vrna_md_set_default(&md);
  /* change temperature and activate G-Quadruplex prediction */
  md.temperature = 25.0; /* 25 Deg Celcius */
md.gquad = 1; /* Turn-on G-Quadruples support */
  /* create a fold compound */
  vrna_fold_compound(t *fc = vrna_fold_compound(seq, &md, VRNA_OPTION_DEFAULT);
  /* predict Minmum Free Energy and corresponding secondary structure */
                         mfe = vrna_mfe(fc, structure);
  /* print sequence, structure and MFE */
  printf("%s\n%s [ %6.2f ]\n", seq, structure, mfe);
  /* cleanup memory */
  free (structure);
  vrna_fold_compound_free(fc);
  return 0;
```

64 Examples

See also

examples/fold_compound_md.c in the source code tarball

8.1.3 Writing Callback Functions

callback_subopt.c

Here is a basic example how to use the callback mechanism in vrna_subopt_cb(). It simply defines a callback function (see interface definition for vrna_subopt_callback) that prints the result and increases a counter variable.

```
#include <stdlib.h>
#include <stdio.h>
#include <ViennaRNA/fold_compound.h>
#include <ViennaRNA/utils/basic.h>
#include <ViennaRNA/utils/strings.h>
#include <ViennaRNA/subopt.h>
void
subopt_callback(const char *structure,
                float
                             energy,
                void
  /\star simply print the result and increase the counter variable by 1 \star/
  if (structure)
    printf("%d.\t%s\t%6.2f\n", (*((int *)data))++, structure, energy);
int
main()
  /* initialize random number generator */
  vrna init rand();
  /* Generate a random sequence of 50 nucleotides */
                         *seq = vrna_random_string(50, "ACGU");
  /\star Create a fold compound for the sequence \star/
  vrna_fold_compound_t
                        *fc = vrna_fold_compound(seq, NULL, VRNA_OPTION_DEFAULT);
  int
                        counter = 0;
  \star call subopt to enumerate all secondary structures in an energy band of
      5 kcal/mol of the MFE and pass it the address of the callback and counter
  vrna_subopt_cb(fc, 500, &subopt_callback, (void *)&counter);
  /* cleanup memory */
  free (seq);
  vrna_fold_compound_free(fc);
  return 0;
```

See also

examples/callback_subopt.c in the source code tarball

8.1.4 Application of Soft Constraints

soft_constraints_up.c

In this example, a random RNA sequence is generated to predict its MFE under the constraint that a particular nucleotide receives an additional bonus energy if it remains unpaired.

```
#include <stdlib.h>
#include <stdio.h>
#include <ViennaRNA/fold_compound.h>
#include <ViennaRNA/utils/basic.h>
#include <ViennaRNA/utils/strings.h>
#include <ViennaRNA/constraints/soft.h>
#include <ViennaRNA/mfe.h>
int
main()
{
    /* initialize random number generator */
    vrna_init_rand();
    /* Generate a random sequence of 50 nucleotides */
```

8.1 C Examples 65

See also

examples/soft_constraints_up.c in the source code tarball

8.1.5 Other Examples

example1.c

A more extensive example including MFE, Partition Function, and Centroid structure prediction.

```
#include <stdio.h>
#include <string.h>
#include <ViennaRNA/part_func.h>
int
main(int argc,
     char *argv[])
{
    "AGACGACAAGGUUGAAUCGCACCCACAGUCUAUGAGUCGGUGACAACAUUACGAAAGGCUGUAAAAUCAAUUAUUCACCACAGGGGGCCCCCGUGUCUAG";
               *mfe_structure = vrna_alloc(sizeof(char) * (strlen(seq) + 1));
*prob_string = vrna_alloc(sizeof(char) * (strlen(seq) + 1));
  char
  /* get a vrna_fold_compound with default settings */
  vrna_fold_compound_t *vc = vrna_fold_compound(seq, NULL, VRNA_OPTION_DEFAULT);
  /* call MFE function */
                           mfe = (double)vrna_mfe(vc, mfe_structure);
  printf("%s\n^*s (%6.2f)\n^*, seq, mfe_structure, mfe);
  /* rescale parameters for Boltzmann factors */
  vrna_exp_params_rescale(vc, &mfe);
  /* call PF function */
  FLT_OR_DBL en = vrna_pf(vc, prob_string);
  /* print probability string and free energy of ensemble */
printf("%s (%6.2f)\n", prob_string, en);
   /* compute centroid structure */
  double dist;
char *cent = vrna_centroid(vc, &dist);
/* print centroid structure, its free energy and mean distance to the ensemble */
printf("%s (%6.2f d=%6.2f)\n", cent, vrna_eval_structure(vc, cent), dist);
   * free centroid structure */
  free (cent);
  /* free pseudo dot-bracket probability string */
  free (prob_string);
  /* free mfe structure */
  free (mfe_structure);
  /* free memory occupied by vrna_fold_compound */
  vrna_fold_compound_free(vc);
  return EXIT_SUCCESS;
```

See also

examples/example1.c in the source code tarball

66 Examples

8.1.6 Deprecated Examples

```
#include <stdio.h>
#include <stdlib.h>
#include <math.h>
#include <string.h>
#include "utils.h"
#include "fold_vars.h"
#include "fold.h"
#include "part_func.h'
#include "inverse.h"
#include "RNAstruct.h"
#include "treedist.h"
#include "stringdist.h"
#include "profiledist.h"
void
main()
  char
               *seq1 = "CGCAGGGAUACCCGCG", *seq2 = "GCGCCCAUAGGGACGC",
                *struct1, *struct2, *xstruc;
               e1, e2, tree_dist, string_dist, profile_dist, kT;
  float
               *T1, *T2;
  Tree
               *S1, *S2;
*pf1, *pf2;
  swString
  FLT_OR_DBL *bppm;
  /\star fold at 30C instead of the default 37C \star/
  temperature = 30.;    /* must be set *before* initializi
/* allocate memory for structure and fold */
struct1 = (char *)space(sizeof(char) * (strlen(seq1) + 1));
                               /* must be set *before* initializing */
          = fold(seq1, struct1);
  struct2 = (char *)space(sizeof(char) * (strlen(seq2) + 1));
          = fold(seq2, struct2);
  free_arrays();
                        /\star free arrays used in fold() \star/
  /\star produce tree and string representations for comparison \star/
  xstruc = expand_Full(struct1);
          = make_tree(xstruc);
         = Make_swString(xstruc);
  free (xstruc);
  xstruc = expand_Full(struct2);
T2 = make_tree(xstruc);
          = Make_swString(xstruc);
  free (xstruc);
  /* calculate tree edit distance and aligned structures with gaps */
  edit_backtrack = 1;
  tree_dist
                    = tree_edit_distance(T1, T2);
  free_tree(T1);
  free tree(T2);
  unexpand_aligned_F(aligned_line);
  printf("%s\n%s %3.2f\n", aligned_line[0], aligned_line[1], tree_dist);
  /* same thing using string edit (alignment) distance */
  string_dist = string_edit_distance(S1, S2);
  free(S1);
  free (S2):
  printf("%s mfe=%5.2f\n%s mfe=%5.2f dist=%3.2f\n",
         aligned_line[0], e1, aligned_line[1], e2, string_dist);
  /\star for longer sequences one should also set a scaling factor for
  * partition function folding, e.g: */
kT = (temperature + 273.15) * 1.98717 / 1000.; /* kT in kcal/mol */
pf_scale = exp(-e1 / kT / strlen(seq1));
  /st calculate partition function and base pair probabilities st/
  e1 = pf_fold(seq1, struct1);
  /\star get the base pair probability matrix for the previous run of pf_fold() \star/
  bppm = export_bppm();
pf1 = Make_bp_profile_bppm(bppm, strlen(seq1));
e2 = pf_fold(seq2, struct2);
  /* get the base pair probability matrix for the previous run of pf_fold() */
  bppm = export_bppm();
        = Make_bp_profile_bppm(bppm, strlen(seq2));
  free_pf_arrays();
                        /* free space allocated for pf_fold() */
  profile_dist = profile_edit_distance(pf1, pf2);
  printf("%s free energy=%5.2f\n%s free energy=%5.2f dist=%3.2f\n",
          aligned_line[0], e1, aligned_line[1], e2, profile_dist);
  free_profile(pf1);
  free_profile(pf2);
```

See also

examples/example_old.c in the source code tarball

8.2 Perl5 Examples

8.3 Python Examples 67

Hello World Examples

Using the flat interface

· MFE prediction

```
use RNA;
# The RNA sequence
my $seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA";
# compute minimum free energy (MFE) and corresponding structure
my ($ss, $mfe) = RNA::fold($seq);
# print output
printf "%s\n%s [ %6.2f ]\n", $seq, $ss, $mfe;
```

Using the object oriented interface

· MFE prediction

```
#!/usr/bin/perl
use warnings;
use strict;
use RNA;
my $seq1 = "CGCAGGGAUACCCGCG";
# create new fold_compound object
my $fc = new RNA::fold_compound($seq1);
# compute minimum free energy (mfe) and corresponding structure
my ($ss, $mfe) = $fc->mfe();
# print output
printf "%s [ %6.2f ]\n", $ss, $mfe;
```

Changing the Model Settings

Using the flat interface

· MFE prediction at different temperature and dangle model

```
use RNA;
# The RNA sequence
my $seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA";
# create a new model details structure
my $md = new RNA::md();
# change temperature and dangle model
$md->{temperature} = 20.0; # 20 Deg Celcius
$md->{dangles} = 1; # Dangle Model 1
# create a fold compound
my $fc = new RNA::fold_compound($seq, $md);
# predict Minmum Free Energy and corresponding secondary structure
my ($ss, $mfe) = $fc->mfe();
# print sequence, structure and MFE
printf "%s\n%s [ %6.2f ]\n", $seq, $ss, $mfe;
```

Using the object oriented interface

· MFE prediction at different temperature and dangle model

8.3 Python Examples

MFE Prediction (flat interface)

```
import RNA
# The RNA sequence
seq = "GAGUAGUGGAACCAGGCUAUGUUUGUGACUCGCAGACUAACA"
# compute minimum free energy (MFE) and corresponding structure
(ss, mfe) = RNA.fold(seq)
# print output
print "%s\n%s [ %6.2f ]" % (seq, ss, mfe)
```

68 Examples

MFE Prediction (object oriented interface)

```
import RNA;
sequence = "CGCAGGGAUACCCGCG"

# create new fold_compound object
fc = RNA.fold_compound(sequence)
# compute minimum free energy (mfe) and corresponding structure
(ss, mfe) = fc.mfe()
# print output
print "%s [ %6.2f ]" % (ss, mfe)
```

Suboptimal Structure Prediction

```
import RNA
sequence = "GGGGAAAACCCC"

# Set global switch for unique ML decomposition
RNA.cvar.uniq_ML = 1
subopt_data = { 'counter' : 1, 'sequence' : sequence }

# Print a subopt result as FASTA record
def print_subopt_result(structure, energy, data):
    if not structure == None:
        print ">subopt %d" % data['counter']
        print "%s" % data['sequence']
        print "%s" % data['sequence']
        print "%s" % data['counter'] + 1

# Create a 'fold_compound' for our sequence
a = RNA.fold_compound (sequence)
# Enumerate all structures 500 dacal/mol = 5 kcal/mol arround
# the MFE and print each structure using the function above
a.subopt_cb(500, print_subopt_result, subopt_data);
```

Boltzmann Sampling (a.k.a. Probabilistic Backtracing)

```
import RNA
sequence =
       "UGGGAAUAGUCUCUUCCGAGUCUCGCGGGCGACGGGCGAUCUUCGAAAGUGGAAUCCGUACUUAUACCGCCUGUGCGGACUACUAUCCUGACCACAUAGU"
A simple callback function that stores
a structure sample into a list
def store_structure(s, data):
       data.append(s)
First we prepare a fold_compound object
# create model details
md = RNA.md()
# activate unique multibranch loop decomposition
md.uniq\_ML = 1
# create fold compound object
fc = RNA.fold_compound(sequence, md)
# compute MFE
(ss, mfe) = fc.mfe()
# rescale Boltzmann factors according to MFE
fc.exp_params_rescale(mfe)
# compute partition function to fill DP matrices
fc.pf()
Now we are ready to perform Boltzmann sampling
# 1. backtrace a single sub-structure of length 10
print "%s" % fc.pbacktrack5(10)
# 2. backtrace a single sub-structure of length 50
print "%s" % fc.pbacktrack5(50)
# 3. backtrace multiple sub-structures of length 10 at once
for s in fc.pbacktrack5(20, 10):
    print "%s" % s
# 4. backtrace multiple sub-structures of length 50 at once
for s in fc.pbacktrack5(100, 50):
    print "%s" % s
# 5. backtrace a single structure (full length)
      "%s" % fc.pbacktrack()
# 6. backtrace multiple structures at once
for s in fc.pbacktrack(100):
    print "%s" % s
# 7. backtrace multiple structures non-redundantly
for s in fc.pbacktrack(100, RNA.PBACKTRACK_NON_REDUNDANT):
# 8. backtrace multiple structures non-redundantly (with resume option)
```

8.3 Python Examples 69

```
num\_samples = 500
iterations = 15
           = None # pbacktrack memory object = list()
d
s_list
for i in range(0, iterations):
   d, ss = fc.pbacktrack(num_samples, d, RNA.PBACKTRACK_NON_REDUNDANT)
s_list = s_list + list(ss)
for s in s_list:
    print "%s" % s
# 9. backtrace multiple sub-structures of length 50 in callback mode
ss = list()
   = fc.pbacktrack5(100, 50, store_structure, ss)
for s in ss:
    print "%s" % s
# 10. backtrace multiple full-length structures in callback mode
ss = list()
i = fc.pbacktrack(100, store_structure, ss)
for s in ss:
    print "%s" % s
# 11. non-redundantly backtrace multiple full-length structures in callback mode
ss = list()
   = fc.pbacktrack(100, store_structure, ss, RNA.PBACKTRACK_NON_REDUNDANT)
for s in ss:
    print "%s" % s
# 12. non-redundantly backtrace multiple full length structures
# in callback mode with resume option
ss = list()
d = None # pbacktrack memory object
for i in range(0, iterations):
   d, i = fc.pbacktrack(num_samples, store_structure, ss, d, RNA.PBACKTRACK_NON_REDUNDANT)
for s in ss:
    print "%s" % s
```

RNAfold -p -MEA equivalent

```
#!/usr/bin/python
import RNA
seq = "AUUUCCACUAGAGAAGGUCUAGAGUGUUUGUCGUUUGUCAGAAGUCCCUAUUCCAGGUACGAACACGGUGGAUAUGUUCGACGACAGGAUCGGCGCACUA"
# create fold_compound data structure (required for all subsequently applied algorithms)
fc = RNA.fold_compound(seq)
# compute MFE and MFE structure
(mfe\_struct, mfe) = fc.mfe()
# rescale Boltzmann factors for partition function computation
fc.exp params rescale (mfe)
\# compute partition function
(pp, pf) = fc.pf()
  compute centroid structure
(centroid_struct, dist) = fc.centroid()
# compute free energy of centroid structure
centroid en = fc.eval structure(centroid struct)
# compute MEA structure
(MEA_struct, MEA) = fc.MEA()
  compute free energy of MEA structure
MEA_en = fc.eval_structure(MEA_struct)
# print everything like RNAfold -p -MEA
print("%s\n%s (%6.2f)" % (seq, mfe_struct, mfe))
print("%s [%6.2f]" % (pp, pf))
print("%s [%6.2f] d=%.2f]" % (centroid_struct, centroid_en, dist))
print("%s {%6.2f MEA=%.2f}" % (MEA_struct, MEA_en, MEA))
print(" frequency of mfe structure in ensemble %g; ensemble diversity %-6.2f" %
        (fc.pr_structure(mfe_struct), fc.mean_bp_distance()))
```

Fun with Soft Constraints

```
import RNA
seq1 = "CUCGUCGCCUUAAUCCAGUGCGGGCGCUAGACAUCUAGUUAUCGCCGCAA"
# Turn-off dangles globally
RNA.cvar.dangles = 0
# Data structure that will be passed to our MaximumMatching() callback with two components:
# 1. a 'dummy' fold_compound to evaluate loop energies w/o constraints, 2. a fresh set of energy parameters
mm_data = { 'dummy': RNA.fold_compound(seq1), 'params': RNA.param() }
# Nearest Neighbor Parameter reversal functions
revert_NN = {
                                               lambda i, j, k, l, f, p: - f.eval_hp_loop(i, j) - 100, lambda i, j, k, l, f, p: - f.eval_int_loop(i, j, k, l) - 100, lambda i, j, k, l, f, p: - p.MLclosing - p.MLintern[0] - (j - i - k + l - 2) \star
      RNA.DECOMP_PAIR_HP:
      RNA.DECOMP_PAIR_IL:
      RNA.DECOMP_PAIR_ML:
           p.MLbase - 100,
                                               lambda i, j, k, l, f, p: - p.MLintern[0] - (1 - k - 1) * p.MLbase, lambda i, j, k, l, f, p: - p.MLintern[0] - (j - i - k + 1) * p.MLbase,
      RNA.DECOMP_ML_ML_STEM:
      RNA.DECOMP_ML_STEM:
      RNA.DECOMP_ML_ML:
                                               lambda i, j, k, l, f, p: - (j - i - k + 1) * p.MLbase,
```

70 Examples

```
RNA.DECOMP_ML_ML: lambda i, j, k, l, f, p: 0,
RNA.DECOMP_ML_UP: lambda i, j, k, l, f, p: - (j - i + 1) * p.MLbase,
RNA.DECOMP_EXT_STEM: lambda i, j, k, l, f, p: - f.E_ext_loop(k, l),
RNA.DECOMP_EXT_EXT: lambda i, j, k, l, f, p: 0,
RNA.DECOMP_EXT_STEM_EXT: lambda i, j, k, l, f, p: - f.E_ext_loop(i, k),
RNA.DECOMP_EXT_EXT_STEM: lambda i, j, k, l, f, p: - f.E_ext_loop(l, j),
RNA.DECOMP_EXT_EXT_STEM: lambda i, j, k, l, f, p: - f.E_ext_loop(l, j),
RNA.DECOMP_EXT_EXT_STEM1: lambda i, j, k, l, f, p: - f.E_ext_loop(l, j-1),
}

# Maximum Matching callback function (will be called by RNAlib in each decomposition step)
def MaximumMatching(i, j, k, l, data['dummy'], data['params'])
# Create a 'fold_compound' for our sequence
fc = RNA.fold_compound(seq1)
# Add maximum matching soft-constraints
fc.sc_add_f(MaximumMatching)
fc.sc_add_data(mm_data, None)
# Call MFE algorithm
(s, mm) = fc.mfe()
# print result
print "%s\n%s (MM: %d)\n" % (seq1, s, -mm)
```

Chapter 9

Contributing

If you wish to contribute to this project, please first discuss any proposed changes with the owners and main developers. You may do that either through making an issue at our official GitHub presence https://github.com/ViennaRNA/ViennaRNA, by email (rna@tbi.univie.ac.at), or any other personal communication with the core developer team.

Please note that we have a code of conduct. Please follow it in all your interactions with this project.

Reporting Bugs

- 1. Please make an issue at GitHub or notify us by emailing to rna@tbi.univie.ac.at
- 2. In your report, include as much information as possible, such that we are able to reproduce it. If possible, find a minimal example that triggers the bug.
- 3. Include the version number for the ViennaRNA Package you experience the bug with.
- 4. Include at least some minimal information regarding your operating system (Linux, Mac OS X, Windows, etc.)

Pull Request Process

- 1. Ensure that you have not checked-in any files that are automatically build!
- 2. When contributing C source code, follow our code formatting guide lines. You may use the tool uncrustify together with our config located in misc/uncrustify.cfg to accomplish that.
- 3. Only expose symbols (functions, variables, etc.) to the libraries interface that are absolutely necessary! Hide all other symbols in the corresponding object file(s) by declaring them as static.
- 4. Use the prefixes vrna_ for any symbol you add to the API of our library! Preprocessor macros in header files require the prefix in capital letters, i.e. VRNA_.
- 5. Use C-style comments at any place necessary to make sure your implementation can still be understood and followed in the future.
- 6. Add test cases for any new implementation! The test suite is located in the tests directory and is split into tests for the C-library, executable programs, and the individual scripting language interfaces.
- 7. Run make check to ensure that all other test suites still run properly with your applied changes!
- 8. When contributing via GitHub, make a personal fork of our project and create a separate branch for your changes. Then make a pull request to our user-contrib branch. Pull requests to the master branch will be rejected to keep its history clean.
- 9. Pull requests that have been successfully merged into the user-contrib branch usually find their way into the next release of the ViennaRNA Package. However, please note that the core developers may decide to include your changes in a later version.

72 Contributing

Chapter 10

Changelog

Below, you'll find a list of notable changes for each version of the ViennaRNA Package.

Version 2.4.x

Unreleased

```
Version 2.4.14 (Release date: 2019-08-13)
```

Programs

- Fix RNApvmin pertubation vector computation
- Add non-redundant sampling option to RNApvmin
- Add RNAdos program to compute density of states
- Add –P DNA convenience command line parameter to most programs to quickly load DNA parameters without any input file
- MAN: Add example section to man-page of RNAalifold

- API: Fix memory leak in vrna_path_gradient()
- API: Fix release of memory fir vrna_sequence_remove_all()
- API: Fix soft-constraints application in vrna_sc_minimize_pertubation() that prevented proper computation of the pertubation vector
- API: Add 5' and 3' neighbor nucleotide encoding arrays and name string to vrna_seq_t
- · API: Add new data structure for multiple sequence alignments
- API: Add vrna_sequence_order_update() function
- API: Add non-redundant sampling mode to vrna_sc_minimize_pertubation() through passing negative sample-sizes
- API: Add v3.0 API functions for maximum expected accuracy (MEA) computation

- · API: Include energy parameter sets into RNAlib and provide functions to load them at runtime
- API: Prepare sequence data in vrna_fold_compound_t with vrna_sequence_add()
- API: Use vrna_pbacktrack_num() instead of vrna_pbacktrack() in vrna_sc_minimize_pertubation() to speed-up sample generation
- Reduce use of global variable cut_point in RNAlib
- SWIG: Use importlib in favor of imp to determine Python 3 tag extension
- · SWIG: Update various wrapper functions
- SWIG: Add wrappers for MEA computation with $vrna_MEA$ () and $vrna_MEA_from_plist$
- SWIG: Add wrappers for vrna_pr_structure() and vrna_pr_energy()

Package

- REFMAN: Fix LaTeX code in units.h that prevented proper compilation with pdflatex
- Add an R script to create 2D landscape plots from RNA2Dfold output
- Add gengetopt to configure-time requirements to build man-pages
- Add new energy parameter file rna_misc_special_hairpins.par with additional UV-melting derived parameters for Tri- and Tetra-loops
- · Update RNA Tutorial
- · Colorize final configure script message
- REFMAN: Always use pdflatex to compile reference manual and tutorial
- EXAMPLES: Add Python script that performs computations equivalent to RNAfold -p --MEA

Version 2.4.13 (Release date: 2019-05-30)

Programs

- Fix centroid structure prediction for RNAcofold
- Fix --noLP option for RNALalifold

- API: Refactor and fix collision handling in vrna_hash_table_t
- API: Fix one access using wrong index for odd dangles in <code>loops/external.c</code>
- API: Add two missing MLbase contributions for MFE prediction in loops/multibranch.c
- · API: Refactor multiloop MFE backtracking for odd dangles
- API: Add function vrna_backtrack5 () to allow for MFE backtracking of sub-sequences starting at the 5'-end
- API: Reduce usage of global macro TURN by replacing it with min_loop_size field of vrna_md_t
- API: Add functions vrna_path_direct() and vrna_path_direct_ub() that may also return
 move lists instead of dot-bracket lists
- API: Add functions <code>vrna_pt_pk_remove()</code> and <code>vrna_db_pk_remove()</code> that remove pseudoknots from an input structure
- API: Fix invalid memory access for lonely pair mode (--nole) in comparative sliding-window MFE prediction
- SWIG: Fix access to global variable pf_smooth and pf_smooth attribute in model_details object
- SWIG: Fix Python reference counting for Py_None in interfaces/findpath.i wrapper
- · SWIG: Refactor reference counting for all Python2 and Python3 wrappers
- REFMAN: Larger updates and restructuring of reference manual

Package

- · Install example scripts and source code files, e.g. to \$prefix/share/ViennaRNA/examples
- · Properly pass GSL, PTHREADS, and MPFR flags to sub-projects
- Fix RNApuzzler header file installation
- · SWIG: Include Python 3.7 and 3.8 in list of autoconf-probed python interpreters
- SWIG: Fix wrapper building for swig >= 4.0.0

Version 2.4.12 (Release date: 2019-04-16)

Programs

- Add non-redundant stochastic backtracing option for RNAalifold
- Add --noDP option to suppress dot-plot output in RNAfold and RNAalifold
- Add RNApuzzler (4) and RNAturtle (3) secondary structure layout algorithm options to RNAfold and RNAplot
- Update help/man page of RNALfold
- Allow for multiple input files and parallel input processing in RNAheat

- API: Fix declaration of vrna_move_apply_db()
- API: Fix vrna_path() lexicographical ordering in gradient walks
- · API: Enable non-redundant stochastic backtracing for comparative structure prediction
- API: Enable stochastic backtracing for circular comparative structure prediction
- · API: Enable stochastic backtracing of subsequences (5' prefixes) for comparative structure prediction
- API: Add pf_smooth attribute to vrna_md_t data stucture to allow for disabling Boltzmann factor energy smoothing
- · API: Add functions to allow for resuming non-redundant stochastic backtracing
- · API: Add functions to retrieve multiple stochastically backtraced structures (list and callback variants)
- API: Add vrna_positional_entropy to compute vector of positional entropies
- API: Add RNApuzzler and RNAturtle secondary structure layout algorithm (Wiegreffe et al. 2018)
- API: Add v3.0 API for secondary structure layout/coordinate algorithms
- API: Add more helper/utility functions for vrna_move_t data structures
- API: Add callback-based neighborhood update function for (subsequent) vrna_move_t application
- API: Add abstract heap data structure available as <ViennaRNA/datastructures/heap.h>
- API: Refactor and speed-up gradient walk implementation available as vrna_path_gradient()
- API: Substitute vrna_file_PS_aln_sub() alignment plot function by vrna_file_PS_aln_slice() that actually slices out a sub-alignment
- API: Rename vrna_annotate_covar_struct() to vrna_annotate_covar_db() and add new function vrna_annotate_covar_db_extended() to support more bracket types

- API: Calling vrna_params_reset () now implies a call to vrna_exp_params_reset () as well
- API: Move landscape implementations into separate directory, thus headers should be included as <ViennaRNA/landscape/move.h>, <ViennaRNA/landscape/neighbor.h>, etc.
- Ensure proper rescaling of energy parameters upon temperature changes
- Refactor soft constraints implementation in stochastic backtracing
- SWIG: Wrap all non-redundant stochastic backtracing functions to scripting language interface(s)
- SWIG: Refactor stochastic backtracing interface(s)
- SWIG: Add proper constructor for objects of type vrna_ep_t
- SWIG: Sanitize alignment plot function interface(s)

Package

- · Update Ubuntu/Debian and OpenSUSE build instructions
- · Reduce intra-package dependency on non-v3.0 API

```
Version 2.4.11 (Release date: 2018-12-17)
```

Programs

- Add --commands option to RNAsubopt
- Add non-redundant Boltzmann sampling mode for RNAsubopt

- · API: Fix wrong access to base pair soft constraints in equilibrium probability computations
- API: Fix behavior of vrna_nucleotide_encode() with lowercase characters in sequence
- API: Fix behavior of encode_char() with lowercase characters in sequence
- · API: Fix forbidden GU pairs behavior in pscore computation for comparative folding
- API: Fix potential errors due to uninitialized next pointers in vrna_move_t of vrna_eval_move_

 shift_pt
- API: Add AVX 512 optimized version of MFE multibranch loop decomposition
- · API: Add functions for CPU SIMD feature detection
- API: Add dispatcher to automatically delegate exterior-/multibranch loop MFE decomposition to supported SIMD optimized implementation
- API: Add function vrna_dist_mountain() to compute mountain distance between two structures
- API: Add function vrna ensemble defect () to compute ensemble defect given a target structure
- · API: Add non-redundant Boltzmann sampling
- API: Change behavior of vrna_cstr_free () and vrna_cstr_close () to always flush output before unregistering the stream
- SWIG: Add interface for vrna_loopidx_from_ptable()

Package

- · Activate compilation for compile-time supported SIMD optimized implementations by default
- Replace -- enable-sse configure script option with -- disable-simd

```
Version 2.4.10 (Release date: 2018-09-26)
```

Programs

- Fix wrong output filename for binary opening energies in RNAplfold
- Enable G-Quadruplex support for partition function computation in RNAalifold

Library

- · Fix broken SSE4.1 support for multibranch loop MFE computation that resulted in increased run times
- Fix redundant output issue in subopt backtracking with unusually high delta energies (>=INF)
- Restore default behavior of '|' symbol in dot-bracket hard constraint strings that got lost with version 2.2.0
- · Add faster (cache-optimized) version of Nussinov Maximum Matching algorithm
- Change default linker- and loop length computations for G-Quadruplex predictions in comparative prediction modes
- Add hard constraints warning for base pairs that violate the min_loop_size of the model
- Update libsvm to version 3.23
- · API: Add functions to set auxiliary grammar extension rules
- · API: Replace upper-triangular hard constraints matrix with full matrix for cache-optimized access
- · API: Add G-Quadruplex prediction support for comparative partition function
- API: Remove VRNA_GQUAD_MISMATCH_PENALTY and VRNA_GQUAD_MISMATCH_NUM_ALI macros
- · SWIG: Fix invalid memory access in subopt () method of fold compound object when writing to file
- · SWIG: Add wrapper for Nussinov Maximum Matching algorithm

Package

Add -ftree-vectorize compile flag by default if supported

```
Version 2.4.9 (Release date: 2018-07-11)
```

Programs

- Fix interactive mode behavior for multiple sequence alignment input in RNAalifold, RNALalifold
- $\bullet \ \, \text{Allow for Stockholm formatted multiple sequence alignment input in $\tt RNAeval and RNAplot$ }$
- Allow for multiple input files in RNAeval and RNAplot
- Allow for parallel processing of input batch jobs in RNAeval and RNAplot
- Add -g option to activate G-Quadruplex support in RNAheat
- Warn on unsatisfiable hard constraints from dot-bracket string input in RNAfold, RNAcofold, and RN←
 Aalifold

Library

• Fix parameter order bug in vrna_path_findpath* functions that resulted in too large search widths

- · Fix wrong application of base pair soft constraints in partition function computations
- · Fix position ruler string in EPS alignment output files
- · Fix MFE backtracking errors that might appear under specific hard constrained base pair patterns
- Refrain from reading anything other than #=GC SS_cons to retrieve structures when parsing Stockholm
 1.0 format
- · Complete soft constraints additions to Boltzmann sampling implementation for single sequences
- Allow for disabling alignment wrapping in vrna_file_PS_aln* functions
- Do not remove G-Quadruplex annotation from WUSS formatted structure strings upon calls to vrna_db_←
 from_WUSS
- Enable G-Quadruplex related average loop energy correction terms in verbose output of vrna_eval_* functions
- Speed-up backward compatibility layer for energy evaluation functions that unnecessarily slowed down thirdparty tools using the old API
- Allow for passing dot-bracket strings with "&'strand-end identifier to simplevrna_eval_← *functions
- Remove implicitexit()` calls from global MFE backtracking implementation.

Version 2.4.8 (Release date: 2018-06-23)

Programs

- Fix compilation of RNAforester with C++17 standard
- · Fix tty input detection in RNAcofold
- · Fix bad memory access with RNAcofold -p

Library

- API: Fix incorrect unpaired probability computations in vrna_probs_window()
- API: Fix potential out-of-bounds access situations (for circular RNA folding) in eval.c
- · API: Fix comparative exterior internal loop partition function computation for circfold
- SWIG: Fix false-positive use of uninitialized value in Python3/file_py3.i

Package

- · TESTS: Add tests for special features in RNAalifold
- · TESTS: Add test case for RNAcofold -p

Version 2.4.7 (Release date: 2018-06-13)

- Allow for parallel processing across multiple input files in RNAfold
- · Allow for arbitrary number of input files in RNAalifold
- · Allow for parallel processing of input data in RNAalifold
- · Allow for arbitrary number of input files in RNAcofold
- · Allow for parallel processing of input data in RNAcofold
- Enable parallel processing in RNAfold, RNAcofold, RNAalifold for MS Windows build
- · Add centroid and MEA structure computation to RNAcofold
- Add configure time check for LTO capabilities of the linker
- Include ligand binding energies in centroid and MEA structure output of RNAfold
- · Refactor ct2db program to process multiple structures from single .ct file
- API: Enable processing of comparative fold compound with vrna pr *() functions
- API: Refactor vrna ostream t to enable NULL input in vrna ostream provide()
- API: Major refactoring in loop energy evaluations (MFE and PF)
- API: Make vrna_mx_pf_aux_el_t and vrna_mx_pf_aux_ml_s opaque pointers
- API: Make fold compound field type a const attribute
- · API: Refactor MFE post-processing for circular RNAs
- · API: Add motif name/id support for unstructured domains
- · API: Remove major part of implicit exit() calls in RNAlib
- · API: Add implementations of Boyer-Moore-Horspool search algorithm
- · API: Add implementations to determine number of rotational symmetry for strings (of objects)
- API: Make vrna_cmd_t an opaque pointer
- API: Move headers for constraints, datastructures, io, loop energy evaluation, energy parameters, plotting, search, and utilities into separate subdirectories (backward compatibility is maintained)
- · API: Add hash table data structure
- API: Fix discrepancy between comparative and single sequence -noLP predictions
- API: Add functions to replace 'old API' interface of RNAstruct.h
- API: Add functions to replace 'old API' interface of aln_util.h
- API: Add generic soft constraints support to suboptimal structure prediction sensu Wuchty et al.
- SWIG: Refactor callback execution for Python 2 / 3 interface to reduce overhead
- · SWIG: Fix configure-time check for Python 3 interface build
- SWIG: Fix Python 3 IO file stream to C FILE * conversion
- · Cosmetic changes in final configure notice
- · Major changes in source tree structure of the library
- · Add autoconf checks for maintainer tools
- Generate C strings from static PostScript files at configure time (for structure- and dot plots)
- REFMAN: Large updates in API documentation and structure of reference manual

Version 2.4.6 (Release date: 2018-04-19)

- Stabilize rounding of free energy output in RNAalifold
- API: Fix potential rounding errors for comparative free energies in eval.c and mfe.c
- API: Fix regression in exterior loop dangling end contributions for comparative base pair probabilities and Boltzmann sampling (introduced with v2.4.4)
- API: Fix regression with hard constrained base pairs for comparative structure prediction (introduced with v2.4.4)
- · TESTS: Add basic tests for RNAalifold executable
- TESTS: Ignore 'frequency of MFE structure' in RNAcofold partition function checks

Version 2.4.5 (Release date: 2018-04-17)

- · Allow for arbitrary number of input files in RNAfold
- Allow for parallel processing of input data in RNAfold (UNIX only, no Windows support yet)
- · Add SHAPE reactivity support through commandline options for RNAplfold
- · Fix unstructured domain motif detection in MFE, centroid, and MEA structures computed by RNAfold
- · Limit allowed set of commands in command file for RNAcofold to hard and soft constraints
- · API: Add functions to compute equilibrium probability of particular secondary structures
- API: Add dynamic string stream data type and associated functions
- API: Add priority-queue like data structure with unordered fill capability and ordered output callback execution
- · API: Add functions to detect unstructured domain motifs in MFE, centroid, and MEA structures
- API: Fix bug in sliding-window partition function computation with SHAPE reactivity and Deigan et al. conversion method
- API: Fix application of '<' and '>' constraint symbols in dot-bracket provided constraints (was broken since v2.4.2)
- · API: Fix MEA structure computation in the presence of unstructured domains
- · API: Stabilize order of probability entries in EPS dot-plot files
- Fix compiler warnings on wrong type of printf() in naview.c
- · Define VRNA_VERSION macro as string literal and add macros for major, minor, and patch numbers
- Stabilize parallel make of Mac OS X installer
- · Add energy parameter set from Langdon et al. 2018
- · Add autoconf checks for POSIX threads compiler/linker support
- · SWIG: Fix 'next' is a perl keyword warnings for Perl5 wrapper
- SWIG: Catch errors and throw execptions whenever scripting language provided callback functions are not applicable or fail
- SWIG: Add keyword arguments and autodoc feature for Python/Python3 wrappers

Version 2.4.4 (Release date: 2018-03-06)

- · Change verbose output for soft-constraints derived ligand binding motifs in RNAfold
- · Allow for lowercase letters in ct2db input
- · Fix bug in interior-like G-Quadruplex MFE computation for single sequences
- · Fix autoconf switch to enable deprecation warnings
- Fix bug in eval_int_loop() that prevented propagation of energy evaluation for loops with nick in strands
- · Fix several bugs for SHAPE reactivity related comparative partition function computations
- · Fix annotation of PostScript output for soft-constraint derived ligand binding motifs in RNAfold
- · Fix constraint indices for multibranch loops in unpaired probability computations of LPfold.c
- · Fix dangling end contributions in comparative partition function for exterior loops
- API: Add simplified interface for vrna_pf_dimer()
- · API: Move concentraton dependent implementation for co-folding to separate compile unit
- · API: Add new API functions for exterior loop evaluations
- · API: Add simplified interfaces for energy evaluation with G-Quadruplexes and circular RNAs
- · API: Add findpath functions that allow for specification of an upper bound for the saddle point
- · Add configure-time linker check for Python3 interface
- · Add automatic CPP suggestions for deprecated function substitutes
- · Major restucturing and constraints feature additions in loop type dependent energy evaluation functions
- · Major restructuring in MFE implementations
- · Major restructuring in PF implementations
- · Minor fixes in Boltzmann sampling implementation
- SWIG: Fix wrappers for findpath() implementation
- SWIG: Add tons of energy evaluation wrappers
- · SWIG: Fix configure-time check of Perl5 interface build capabilities
- · SWIG: Wrap functions from walk.c and neighbor.c
- · DOC: Add some missing references to manpages of executable programs
- · REFMAN: Heavy re-ordering of the RNAlib reference manual

Version 2.4.3 (Release date: 2017-11-14)

- Fix handling of dangling end contribution at sequence boundaries for sliding window base pair probability computations
- Fix handling of base pair hard constraints in sliding-window implementations
- · Fix sliding-window pair probability computations with multibranch-loop unpaired constraints
- Fix sliding-window non-specific base pair hard constraint implementation
- Fix probability computation for stochastic backtracking in RNAsubopt -stochBT_en output
- · Fix regression in comparative structure prediction for circular RNAs

- Fix LDFLAGS for scripting language interfaces in corresponding Makefiles
- · Stabilize partition function scaling by always using sfact scaling factor from model details
- Add –pf_scale commandling parameter to RNAplfold
- · Add constraint framework for single sequence circular RNA structure prediction
- · Add RNAfold test suite to check for working implementation of constraints for circular RNAs
- · Add a brief contribution guideline CONTRIBUTING.md
- · Prevent RNAplfold from creating inf/-inf output when solution set is empty with particular hard constraints
- Include RNAforester v2.0.1

Version 2.4.2 (Release date: 2017-10-13)

- · Fix G-Quadruplex energy corrections in comparative structure energy evaluations
- · Fix discrepancy in comparative exterior loop dangling end contribution of eval vs. MFE predictions
- · Fix regression in RNAup unstructuredness and interaction energy computations
- · Fix sequence length confusions when FASTA input contains carriage returns
- · Fix build problems of RNAlocmin with older compilers
- · Fix sliding-window hard constraints where single nucleotides are prohibited from pairing
- · Fix dot-bracket output string length in sliding-window MFE with G-Quadruplexes
- Fix unpaired probability computations for separate individual loop types in LPfold.c
- · Fix bad memory access in RNAsubopt with dot-bracket constraint
- Add full WUSS support for –SS_cons constraint option in RNAalifold
- · Add commandline option to RNALalifold that enables splitting of energy contributions into separate parts
- Add missing hard constraint cases to sliding-window partition function implementation
- Add CSV output option to RNAcofold
- Use the same model details for SCI computations in RNAalifold
- Abort computations in vrna_eval_structure_v() if structure has unexpected length
- Use original MSA in all output generated by RNAalifold and RNALalifold
- API: Add new functions to convert dot-bracket like structure annotations
- · API: Add various new utility functions for alignment handling and comparative structure predictions
- API: Add function vrna_strsplit() to split string into tokens
- API: Do not convert sequences of input MSA to uppercase letters in vrna_file_msa_read_record()
- API: Rename vrna_annotate_bp_covar() and vrna_annotate_pr_covar()
- API: Add new noLP neighbor generation
- SWIG: Add wrapper for functions in file_utils_msa.h
- SWIG: Add wrappers for vrna_pbacktrack() and vrna_pbacktrack5()
- SWIG: Add vrna_db_to_element_string() to scripting language interface
- · REFMAN: Fix formula to image conversion in HTML output

Version 2.4.1 (Release date: 2017-08-23)

- Fix memory leak in fold_compound methods of SWIG interface
- Fix memory leaks in double ** returning functions of SWIG Perl5 interface
- Fix memory leak in vrna_ep_t to-string() function of SWIG interface
- Regression: Fix reverting pf_scale to defaults after vrna_exp_params_rescale()
- · Regression: Fix homo-dimer partition function computation in RNAcofold
- · Add unit tests for RNAcofold executable
- · Add SHAPE reactivity support to RNAcofold
- · Add SHAPE reactivity support to RNALalifold

Version 2.4.0 (Release date: 2017-08-01)

- · Bump libsvm to version 3.22
- · Print G-Quadruplex corrections in verbose mode of RNAeval
- Change behavior of RNAfold –outfile option to something more predictable
- Unify max_bp_span usage among sliding window prediction algorithms: RNAplfold, RNALfold, and RNA
 — Lalifold now consider any base pair (i,j) with (j i + 1) <= max_bp_span
- · Add SHAPE reactivity data support to RNALfold
- Add commands-file support for RNALfold, RNAplfold (hard/soft constraints)
- · Add RNAlocmin Calculate local minima from structures via gradient walks
- Add RNA Bioinformatics tutorial (PDF version)
- · Add hard constraints to sliding-window MFE implementations (RNALfold, RNALalifold)
- Add hard constraints to sliding-window PF implementations (RNAplfold)
- · Add soft constraints to sliding-window MFE implementation for single sequences (RNALfold)
- Add soft constraints to sliding-window PF implementations (RNAplfold)
- · Add SWIG interfaces for sliding-window MFE/PF implementations
- Add proper SWIG interface for alignment and structure plotting functions
- · Add proper SWIG interface for duplexfold, duplex_subopt, and its comparative variants
- Add SWIG wrapper for vrna_exp_params_rescale()
- · Add explicit destructor for SWIG generated vrna md t objects
- Add SWIG perl5 typemap for simple nested STL vectors
- Add dummy field in vrna_structured_domains_s
- Add note about SSE optimized code in reference manual
- Add SWIG interface for findpath implementation
- Add prepare() functions for ptypes-arrays and vrna_(exp_)param_t
- Add warnings for ignored commands in function vrna_commands_apply()
- Add callback featured functions for sliding window MFE and PF implementations

- Change default behavior of adding soft constraints to a vrna_fold_compound_t (store only)
- Several fixes with respect to G-Quadruplex prediction in sliding-window MFE recursions (single sequence and comparative implementation)
- Replace comparative sliding-window MFE recursions (All hits are reported to callback and can be filtered in a post-processing step)
- API: Remove E_mb_loop_stack() and introduce new function vrna_E_mb_loop_stack() as a replacement
- API: change data type of all constraint bit-flags from char to unsigned char
- API: change data type of a2s array in comparative structure prediction from unsigned short to unsigned int
- API: Change function parameter order in vrna_probs_window() to follow the style of other callback-aware functions in RNAlib
- Move sliding-window MFE implementations to new file mfe_window.c
- · Fix building PDF Reference manual with non-standard executable paths
- · Fix redefinition of macro ON SAME STRAND() in subopt.c
- · Fix dangling end issues in sliding-window MFE implementations
- Fix regression for -canonicalBPonly switch in RNAfold/RNAcofold/RNAsubopt
- · Fix building sliding-window MFE implementation without SVM support
- Fix parsing of STOCKHOLM 1.0 MSA files that contain MSA spanning multiple blocks
- Fix Alidot link in RNAalifold manpage
- · Fix wrong pre-processor flags when enabling single-precision PF computations
- Fix unit testing perl5 interface by including builddir/tests in PERL5LIB path
- · Fix buffer overflow in hairpin loop sequence motif extraction for circular RNAs
- · Fix out-of-bounds memory access in neighbor.c
- · Restore capability to compile stand-alone findpath utility
- · Restore capability to use non-standard alphabets for structure prediction
- · Restore old-API random number functions in SWIG interface
- · Allow additional control characters in MAF MSA input that do not end a block
- · Improve reference manual
- Make functions in pair_mat.h static inline
- · Prevent users from adding out-of-range base pair soft constraints
- Inline print functions in color_output.inc
- · Start documenting callback features in reference manual
- · Re-write large portions of sliding-window PF implementation
- · Introduce soft-constraint state flag
- · Clean-up SWIG unit test framework
- Remove obsolete scripts ct2b.pl and colorrna.pl from src/Utils directory
- · Remove old RNAfold tutorial

Version 2.3.x

Version 2.3.5 (Release date: 2017-04-14)

- · Fix duplication of output filename prefix in RNAfold
- · Add V3.0 API for sliding window partition function (a.k.a. RNAPLfold)
- · Add G-Quadruplex prediction to RNALalifold
- · Add SWIG wrappers for callback-based sliding window comparative MFE prediction
- · Add SSE4.1 multiloop decomposition for single sequence MFE prediction
- · Enable RNAfold unit tests to run in paralllel
- · Enable users to turn-off base pair probability computations in RNAcofold with -a option
- · Split move set in neighbor.c

Version 2.3.4 (Release date: 2017-03-10)

- · Fix G-Quadruplex probability computation for single sequences
- · Fix double-free when using SHAPE reactivity data in RNAalifold
- · Fix out-of-bounds access in strand_number array
- Fix weighting of SHAPE reactivity data in consensus structure prediction when fewer data than sequences are present
- · Fix z-score output in RNALfold
- Substitute field name 'A0'/'B0' in data structure vrna_dimer_conc_s by 'Ac_start'/'Bc_start' to avoid clashes with termios.h (Mac OSX Python wrapper bug)
- · Minimize usage of 'unsafe' sprintf() calls
- · Enhance auto-id feature in executable programs
- · Always sanitize output file names to avoid problems due to strange FASTA headers
- · Lift restrictions of FASTA header length in RNAfold, RNAcofold, and RNAeval
- · Add ViennaRNA/config.h with pre-processor definitions of configure time choices
- · Add test-suite for RNAfold
- · Add functions to procude colored EPS structure alignments
- · Add function to write Stockholm 1.0 formatted alignments
- · Add function to sanitize file names
- Add callback based implementation for sliding-window MFE prediction (single sequences, comparative structure prediction)
- Add fast API 3.0 implementations to generate structural neighbors and perform steepest descent / random walks (Thanks to Gregor!)
- · Add parameter option to RNALalifold for colored EPS structure alignment and structure plot output
- Add parameter option to RNALalifold to write hits into Stockholm file
- · Add parameter option to RNAalifold to write Stockholm 1.0 formatted output

- · Add parameter option to RNAalifold to suppress stderr spam
- · Add auto-id feature to RNAplot, RNALfold, RNAsubopt, RNAplfold, RNAheat
- · Add SHAPE reactivity derived pseudo-energies as separate output in RNAalifold
- Add colored output to RNA2Dfold, RNALalifold, RNALfold, RNAduplex, RNAheat, RNAinverse, RNAplfold, and RNAsubopt
- · Add command line parameters to RNAsubopt to allow for specification of input/output files

Version 2.3.3 (Release date: 2017-01-24)

- · Fix multiloop contributions for comparative partition function
- · Fix building python2 extension module for OSX

Version 2.3.2 (Release date: 2017-01-18)

- Fix pair probability plist creation with G-Quadruplexes
- Allow for specification of python2/3-config at configure time
- Fix init of vrna_md_t data structure after call to set_model_details()
- · Fix bug in consensus partition function with hard constraints that force nucleotides to be paired
- · Fix compilation of functions that use ellipsis/va_list
- · Enable generic hard constraints by default
- · Fix init of partition function DP matrices for unusually short RNAs
- · Fix behavior of RNAplfold for unusually short RNAs
- · Report SCI of 0 in RNAalifold when sum of single sequence MFEs is 0
- · Avoid multiple includes of pair mat.h
- · Add configure flag to build entirely static executables

Version 2.3.1 (Release date: 2016-11-15)

- Add description for how to use unstructured domains through command files to reference manual and RN

 Afold manpage
- · Fix compilation issue for Windows platforms with MingW
- Add missing newline in non-TTY-color output of vrna_message_info()
- Fix regression in vrna_md_update() that resulted in incomplete init of reverse-basepair type array
- · Extend coverage of generic hard constraints for partition function computations
- · Fix scaling of secondary structure in EPS plot such that it always fits into bounding box
- · Several fixes and improvements for SWIG generated scripting language interface(s)

Version 2.3.0 (Release date: 2016-11-01)

- · Add grammar extension with structured and unstructured domains
- Add default implementation for unstructured domains to allow for ligand/protein binding to unpaired structure segments (MFE and PF for single sequences)
- · Introduced command files that subsume constraint definition files (currently used in RNAfold and RNAcofold)
- Replace explicit calls to asprintf() with portable equivalent functions in the library
- · Fix configure script to deal with situations where Perl module can't be build
- Fix bug in doc/Makefile.am that prevented HTML installation due to long argument list
- · Added utility functions that deal with conversion between different units
- Bugfix in SWIG wrapped generic soft constraint feature
- Add subopt() and subopt_zuker() methods to SWIG wrapped fold_compound objects
- · Bugfix multiloop decomposition in MFE for circular RNAs
- · Add separate function to compute pscore for alignments
- Renamed VRNA_VC_TYPE_* macros to VRNA_FC_TYPE_*
- Bugfix regression that prevented programs to fail on too long input sequences
- · Extend EPS dot-plot in RNAfold to include motif/binding probabilities from unstructured domains
- · Add variadic functions for error/warning/info message
- · Add ID manipulation feature to RNAeval
- Extend API for soft constraint feature for more fine-grained control
- · Add section on SWIG wrapped functions in reference manual
- · Fix bug in interior loop computations when hard constraints result in non-canonical base pairs

Version 2.2.x

Version 2.2.10 (Release date: 2016-09-06)

- · Do not 'forget' subopt results when output is not written to file handle and sorting is switched off
- Fix bad memory access in vrna_subopt() with sorted output
- Add SWIG wrappers for vrna subopt cb()
- · Correctly show if C11 features are activated in configure status
- Fix autoconf checks to allow for cross compilation again

Version 2.2.9 (Release date: 2016-09-01)

- Fix bug in partition function scaling for backward compatibility of ali_pf_fold()
- Stabilize v3.0 API when building RNAlib and third party program linking against it with compilers that use different C/C++ standards
- Add details on how to link against RNAlib to the reference manual
- Fix RNAlib2.pc
- · Fix bug for temperature setting in RNAplfold
- Use -fflat-lto-objects for static RNAlib library to allow linking without LTO
- · Fix interpretation of 'P' hard constraint for single nucleotides in constraint definition files
- Add 'A' command for hard constraints
- Fix several hard constraint corner-cases in MFE and partition function computation when nucleotides must not be unpaired
- · Fix order of hard constraints when read from input file
- · Allow for non-canonical base pairs in MFE and partition function computations if hard constraints demand it
- · Fix behavior of -without-swig configure script option
- Fix bug in hard constraints usage of exterior loop MFE prediction with odd dangles
- · Add parsers for Clustal, Stockholm, FASTA, and MAF formatted alignment files
- Enable RNAalifold to use Clustal, Stockholm, FASTA, or MAF alignments as input
- · Lift restriction of sequence number in alignments for RNAalifold
- Enable ANSI colors for TTY output in RNAfold, RNAcofold, RNAsubopt, and warnings/errors issued by RNAlib
- Add various new commandline options to manipulate sequence/alignment IDs in RNAfold, RNAcofold and RNAalifold

Version 2.2.8 (Release date: 2016-08-01)

- · Fix bad memory access in RNAalifold
- · Fix regression in RNAalifold to restore covariance contribution ratio determination for circular RNA alignments
- Changed output of RNAsubopt in energy-band enumeration mode to print MFE and energy range in kcal/mol instead of 10cal/mol
- Include latest Kinfold sources that make use of v3.0 API, therefore speeding up runtime substantially
- Re-activate warnings in RNAeval when non-canonical base pairs are encountered
- · Fix syntactic incompatibilities that potentially prevented compilation with compilers other than gcc
- · dd function to compare nucleotides encoded in IUPAC format
- · Fix regression in energy evaluation for circular RNA sequences
- · Fix regression in suboptimal structure enumeration for circular RNAs
- Allow for P i-j k-l commands in constraint definition files
- · Make free energy evaluation functions polymorphic

- · Add free energy evaluation functions that allow for specifying verbosity level
- · Secure functions in alphabet.c against NULL pointer arguments
- Fix incompatibility with swig >= 3.0.9
- Fix memory leak in swig-generated scripting language interface(s) for user-provided target language softconstraint callbacks
- Expose additional functions to swig-generated scripting language interface(s)
- · Build Python3 interface by default
- Start of more comprehensive scripting language interface documentation
- Fix linking of python2/python3 interfaces when libpython is in non-standard directory
- · Restructured viennarna.spec for RPM based distributions
- · Several syntactic changes in the implementation to minimize compiler warnings
- Fix -with-*/-without-* and -enable-*/-disable-* configure script behavior

Version 2.2.7 (Release date: 2016-06-30)

- Fix partition function scaling for long sequences in RNAfold, RNAalifold, and RNAup
- · Fix backtracking issue in RNAcofold when -noLP option is activated
- · Fix hard constraints issue for circular RNAs in generating suboptimal structures
- Rebuild reference manual only when actually required

Version 2.2.6 (Release date: 2016-06-19)

- · Plugged memory leak in RNAcofold
- · Fixed partition function rescaling bug in RNAup
- Fixed bug in RNALfold with window sizes larger than sequence length
- · Re-added SCI parameter for RNAalifold
- · Fixed backtracking issue for large G-quadruplexes in RNAalifold
- · Fixed missing FASTA id in RNAeval output
- · Added option to RNAalifold that allows to specify prefix for output files
- · Several fixes and additional functions/methods in scripting language interface(s)
- Added version information for scripting language interface(s)
- Some changes to allow for compilation with newer compilers, such as gcc 6.1

Version 2.2.5 (Release date: 2016-04-09)

- Fixed regression in RNAcofold that prohibited output of concentration computations
- Fixed behavior of RNAfold and RNAcofold when hard constraints create empty solution set (programs now abort with error message)
- · Added optional Python 3 interface
- · Added RNA::Params Perl 5 sub-package
- Update RNA::Design Perl 5 sub-package
- · Simplified usage of v3.0 API with default options
- Wrap more functions of v3.0 API in SWIG generated scripting language interfaces
- Plugged some memory leaks in SWIG generated scripting language interfaces
- Changed parameters of recursion status callback in vrna fold compound t
- · Enable definition and binding of callback functions from within SWIG target language
- · Added optional subpackage Kinwalker
- Added several configure options to ease building and packaging under MacOS X
- · Added new utility script RNAdesign.pl

Version 2.2.4 (Release date: 2016-02-19)

- · Fixed bug in RNAsubopt that occasionally produced cofolded structures twice
- · Removed debugging output in preparations of consensus structure prediction datastructures

Version 2.2.3 (Release date: 2016-02-13)

- · Added postscipt annotations for found ligand motifs in RNAfold
- · Added more documentation for the constraints features in RNAfold and RNAalifold
- Restore backward compatibility of get_alipf_arrays()

Version 2.2.2 (Release date: 2016-02-08)

· Fix regression bug that occasionally prevented backtracking with RNAcofold -noLP

Version 2.2.1 (Release date: 2016-02-06)

- · Fix regression bug that made RNAcofold -a unusable
- Fix regression bug that prohibited RNAfold to compute the MEA structure when G-Quadruplex support was switched on
- · Fix bug in Kinfold to enable loading energy parameters from file
- Fix potential use of uninitialized value in RNApdist
- · Add manpage for ct2db
- · Fix MEA computation when G-Quadruplex support is activated
- · Allow for vendor installation of the perl interface using INSTALLDIRS=vendor at configure time
- Install architecture dependent and independent files of the perl and python interface to their correct file system locations

Version 2.2.0 (Release date: 2016-01-25)

- RNAforester is now of version 2.0
- New program RNApvmin to compute pseudo-energy pertubation vector that minimizes discrepancy between observed and predicted pairing probabilities
- · SHAPE reactivity support for RNAfold, RNAsubopt, and RNAalifold
- · Ligand binding to hairpin- and interior-loop motif support in RNAfold
- New commandline option to limit maximum base pair span for RNAfold, RNAsubopt, RNAcofold, and RN←
 Aalifold
- · Bugfix in RNAheat to remove numerical instabilities
- · Bugfix in RNAplex to allow for computation of interactions without length limitation
- · Bugfix in RNAplot for simple layouts and hairpins of size 0
- (generic) hard- and soft-constraints for MFE, partition function, base pair probabilities, stochastic backtracking, and suboptimal secondary structures of single sequences, sequence alignments, and sequence dimers
- libsvm version as required for z-scoring in RNALfold is now 3.20
- · Stochastic backtracking for single sequences is faster due to usage of Boustrophedon scheme
- First polymorphic functions vrna_mfe(), vrna_pf(), and vrna_pbacktrack().
- · The FLT_OR_DBL macro is now a typedef
- New functions to convert between different secondary structure representations, such as helix lists, and R

 NAshapes abstractions
- · First object-oriented interface for new API functions in the scripting language interfaces
- new ViennaRNA-perl submodule that augments the Perl interface to RNAlib
- · Ligand binding to hairpin- and interior-loop motif support in C-library and scripting language interfaces.
- · Libraries are generated using libtool
- Linking of libraries and executables defaults to use Link Time Optimization (LTO)
- · Large changes in directory structure of the source code files

Version 2.1.x

Version 2.1.9

- · Fixed integer underflow bug in RNALfold
- · Added Sequence Conservation index (SCI) option to RNAalifold
- · Fixed bug in energy evaluation of dangling ends / terminal mismatches of exterior loops and multibranch loops
- Fixed bug in alifold partition function for circular RNAs
- · Fixed bug in alifold that scrambled backtracing with activated G-Quadruplex support
- Fixed bug in alifold backtracking for larger G-Quadruplexes

Version 2.1.8

- · Repaired incorporation of RNAinverse user provided alphabet
- · Fix missing FASTA ID in RNAeval output
- prevent race condition in parallel calls of Lfold()
- · Fixed memory bug in Lfold() that occured using long sequences and activated G-Quad support
- · Added latest version of switch.pl

Version 2.1.7

- · Fixed bug in RNALfold -z
- · Python and Perl interface are compiling again under MacOSX
- · Fixed handling of C arrays in Python interface
- · Added latest version of switch.pl
- · Make relplot.pl work with RNAcofold output

Version 2.1.6

- New commandline switches allow for elimination of non-canonical base pairs from constraint structures in RNAfold, RNAalifold and RNAsubopt
- · updated moveset functions
- · final fix for discrepancy of tri-loop evaluation between partition function and mfe
- pkg-config file now includes the OpenMP linker flag if necessary
- · New program ct2db allows for conversion of .ct files into dot-bracket notation (incl. pseudo-knot removal)

Version 2.1.5

· Fix for discrepancy between special hairpin loop evaluation in partition functions and MFE

Version 2.1.4

- Fix of G-quadruplex support in subopt()
- · Fix for discrepancy between special hairpin loop evaluation in partition functions and MFE

Version 2.1.3

- RNAfold: Bugfix for ignoring user specified energy parameter files
- · RNAcofold: Bugfix for crashing upon constrained folding without specifying a constraint structure
- RNAsubopt: Added G-quadruplex support
- RNAalifold: Added parameter option to specify base pair probability threshold in dotplot
- Fix of several G-quadruplex related bugs
- Added G-quadruplex support in subopt()

Version 2.1.2

- RNAfold: Bugfix for randomly missing probabilities in dot-plot during batch job execution
- RNAeval: Bugfix for misinterpreted G-quadruplex containing sequences where the quadruplex starts at nucleotide 1
- · RNAsubopt: Slight changes to the output of stochastic backtracking and zuker subopt
- · Fix of some memory leaks
- Bugfixes in zukersubopt(), assign_plist_from_pr()
- New threadsafe variants of putoutpU_prob*() for LPfold()
- · Provision of python2 interface support.

Version 2.1.1

 Bugfix to restore backward compatibility with ViennaRNA Package 1.8.x API (this bug also affected proper usage of the the perl interface)

Version 2.1.0

- · G-Quadruplex support in RNAfold, RNAcofold, RNALfold, RNAalifold, RNAeval and RNAplot
- · LPfold got a new option to output its computations in split-mode
- · several G-Quadruplex related functions were introduced with this release
- · several functions for moves in an RNA landscape were introduced
- new function in alipfold.c now enables access to the partition function matrices of alipf_fold()
- different numeric approach was implement for concentration dependend co-folding to avoid instabilities which occured under certain circumstances

Version 2.0.x

Version 2.0.7

- Bugfix for RNAplfold where segfault happened upon usage of -O option
- · Corrected misbehavior of RNAeval and RNAplot in tty mode

Version 2.0.6

- Bugfix for bad type casting with gcc under MacOSX (resulted in accidental "sequence too long" errors)
- · Bugfix for disappearing tri-/hexaloop contributions when read in from certain parameter files
- Bugfix for RNALfold that segfaulted on short strange sequences like AT+ repeats
- · Change of RNA2Dfold output format for stochastic backtracking

Version 2.0.5

Restored z-score computation capabilities in RNALfold

Version 2.0.4

- · Bugfix for RNAcofold partition function
- · Perl wrapper compatibility to changed RNAlib has been restored
- · Backward compatibility for partition function calls has been restored

Version 2.0.3

- Bugfix for RNAalifold partition function and base pair probabilities in v2.0.3b
- · Added Boltzmann factor scaling in RNAsubopt, RNAalifold, RNAplfold and RNAcofold
- Bugfix for alipfold() in v2.0.3b
- Restored threadsafety of folding matrix access in LPfold.c, alipfold.c, part_func.c, part_func_co.c and part
 _func_up.c
- Added several new functions regarding threadsafe function calls in terms of concurrently changing the model details
- Added pkg-config file in the distribution to allow easy checks for certain RNAlib2 versions, compiler flags and linker flags.

Version 2.0.2

- · added support for Boltzmann factor scaling in RNAfold
- · fixed fastaheader to filename bug
- · plugged some memory leaks

Version 2.0.1

- · First official release of version 2.0
- · included latest bugfixes

History

2011-03-10 Ronny Lorenz ronny@tbi.univie.ac.at

- · new naming scheme for all shipped energy parameter files
- fixed bugs that appear while compiling with gcc under MacOS X
- fixed bug in RNAup -interaction-first where the longer of the first two sequences was taken as target
- added full FASTA input support to RNAfold, RNAcofold, RNAheat, RNAplfold RNALfoldz, RNAsubopt and RNALfold

2010-11-24 Ronny Lorenz ronny@tbi.univie.ac.at

first full pre-release of version 2.0

2009-11-03 Ivo Hofacker ivo@tbi.univie.ac.at

• Fix memory corruption in PS_color_aln()

2009-09-09 Ivo Hofacker ivo@tbi.univie.ac.at

- · Fix bug in RNAplfold when -u and -L parameters are equal
- Fix double call to free_arrays() in RNAfold.c
- · Improve drawing of cofolded structures

2009-05-14 Ivo Hofacker ivo@tbi.univie.ac.at

• Fix occasional segfault in RNAalifold's print_aliout()

2009-02-24 lvo Hofacker ivo@tbi.univie.ac.at

- · Add -MEA options to RNAfold and RNAalifold
- · change energy_of_alistruct to return float not void

2009-02-24 lvo Hofacker ivo@tbi.univie.ac.at

- RNAfold will draw structures unless -noPS is used (no more "structure too long" messages)
- · Restore the "alifold.out" output from RNAalifold -p
- · RNAalifold -circ did not work due to wrong return type
- Accessibility calculation with RNAplfold would give wrong results for u<=30

2008-12-03 lvo Hofacker ivo@tbi.univie.ac.at

- Add zuker style suboptimals to RNAsubopt (-z)
- get_line() should be much faster when reading huge sequences (e.g. whole chromosomes for RNALfold)

2008-08-12 Ivo Hofacker ivo@tbi.univie.ac.at

· Add Ribosum matrices for covariance scoring in RNAalifold

2008-06-27 Ivo Hofacker ivo@tbi.univie.ac.at

- Change RNAalifold to used berni's new energy evaluation w/o gaps
- · Add stochastic backtracking in RNAalifold

2008-07-04 Ivo Hofacker ivo@tbi.univie.ac.at

• modify output of RNAup (again). Program reading RNAup output will have to updated!

2008-07-02 Ivo Hofacker ivo@tbi.univie.ac.at

• RNAplfold now computes accessibilities for all regions up to a max length simultaneously. Slightly slower when only 1 value is needed, but much faster if all of them are wanted. This entails a new output format. Programs reading accessibility output from RNAplfold need to be updated!

2008-03-31 Stephan Bernhart berni@tbi.univie.ac.at

· add cofolding to RNAsubopt

2008-01-08 Ivo Hofacker ivo@tbi.univie.ac.at

· ensure circfold works even for open chain

2007-12-13 Ulli Mueckstein ulli@tbi.univie.ac.at

 upate RNAup related files RNAup can now include the intramolecular structure of both molecules and handles constraints.

2007-12-05 Ronny Lorenz ronny@tbi.univie.ac.at

· add circfold variants in part func.c alipfold.c subopt.c

2007-09-19 Ivo Hofacker ivo@tbi.univie.ac.at

- compute the controld structure of the ensemble in RNAfold -p
- fix a missing factor 2 in mean_bp_dist(). CAUTION ensemble diversities returned by RNAfold -p are now twice as large as in earlier versions.

2007-09-04 Ivo Hofacker ivo@blini.tbi.univie.ac.at

• fix a bug in Lfold() where base number n-max-4 would never pair

2007-08-26 Ivo Hofacker ivo@tbi.univie.ac.at

- · add RNAaliduplex the alignment version of RNAduplex
- introduce a minimal distance between hits produced by duplex_subopt()

2007-07-03 Ivo Hofacker ivo@tbi.univie.ac.at

add a loop_energy() function to compute energy of a single loop

```
2007-06-23 Ivo Hofacker ivo@tbi.univie.ac.at
```

add aliLfold() and RNALalifold, alignment variant of Lfold()

2007-04-30 lvo Hofacker ivo@tbi.univie.ac.at

· add RNAup to distribution

2007-04-15 Ivo Hofacker ivo@tbi.univie.ac.at

fix segfault in colorps output (thanks to Andres Varon)

2007-03-03 Ivo Hofacker ivo@tbi.univie.ac.at

• avoid unnormalized doubles in scale[], big speedup for pf_fold() on very long sequences

2007-02-03 Ivo Hofacker ivo@tbi.univie.ac.at

• RNAalifold can now produce colored structure plots and alignment plots

2007-02-01 lvo Hofacker ivo@tbi.univie.ac.at

· Fix segfault in RNAplfold because of missing prototype

2006-12-01 Ivo Hofacker ivo@tbi.univie.ac.at

· RNAduplex would segfault when no structure base pairs are possible

2006-08-22 lvo Hofacker ivo@tbi.univie.ac.at

- · add computation stacking probabilities using RNAfold -p2
- · add -noPS option for NRAfold to supress drawing structures

2006-08-09 Stephan Bernhart berni@tbi.univie.ac.at

· RNAplfold can now compute probabilites of unpaired regions (scanning version of RNAup)

2006-06-14 Ivo Hofacker ivo@tbi.univie.ac.at

- compile library with -fpic (if available) for use as shared library in the Perl module.
- fix another bug when calling Lfold() repeatedly
- fix switch cmdline parsing in RNAalifold (-mis implied -4)

• fix bug in cofold() with dangles=0

2006-05-08 Ivo Hofacker ivo@tbi.univie.ac.at

- · fix segfault in Lfold() when calling repeatedly
- fix structure parsing in RNAstruct.c (thanks to Michael Pheasant for reporting both bugs)
- add duplexfold() and alifold() to Perl module
- · distinguish window size and max pair span in LPfold

2006-04-05 lvo Hofacker ivo@tbi.univie.ac.at

- fix performance bug in co_pf_fold()
- · use relative error for termination of Newton iteration

2006-03-02 Ivo Hofacker ivo@tbi.univie.ac.at

• add circular folding in alifold()

2006-01-18 Ivo Hofacker ivo@tbi.univie.ac.at

· cleanup berni partition cofold code, including several bug fixes

2006-01-16 Ivo Hofacker ivo@tbi.univie.ac.at

- update RNAplfold to working version
- add PS_dot_plot_turn() in PS_dot.c

2005-11-07 Ivo Hofacker ivo@tbi.univie.ac.at

· add new utilities colorna and coloraln

2005-10-11 Christoph Flamm xtof@tbi.univie.ac.at

adapt PS_rna_plot() for drawing co-folded structures

2005-07-24 Ivo Hofacker ivo@tbi.univie.ac.at

· fix a few memory problems in structure comparison routines

2005-04-30 Ivo Hofacker ivo@blini.tbi.univie.ac.at

add folding of circular RNAs

```
2005-03-11 lvo Hofacker ivo@blini.tbi.univie.ac.at

    add -mis option to RNAalifold to give "most informative sequence" as consensus

2005-02-10 Ivo Hofacker ivo@tbi.univie.ac.at

    move alifold() into the library

2004-12-22 Stephan Bernhart berni@tbi.univie.ac.at
    · add partition function version of RNAcofold
2004-12-23 Ivo Hofacker ivo@tbi.univie.ac.at
    · add RNApaln for fast structural alignments (RNApdist improvement)
2004-08-12 lvo Hofacker ivo@tbi.univie.ac.at
    · fix constrained folding in stochastic backtracking
```

2004-07-21 lvo Hofacker ivo@tbi.univie.ac.at

· add RNAduplex, to compute hybrid structures without intra-molecular pairs

2004-02-09 Ivo Hofacker ivo@tbi.univie.ac.at

- · fix bug in fold that caused segfaults when using Intel compiler
- · add computation of ensemble diversity to RNAfold

2003-09-10 lvo Hofacker ivo@tbi.univie.ac.at

· add annotation options to RNAplot

2003-08-04 Ivo Hofacker ivo@tbi.univie.ac.at

stochastic backtracking finally works. Try e.g. RNAsubopt -p 10

2003-07-18 Ivo Hofacker ivo@tbi.univie.ac.at

· add relplot.pl and rotate_ss.pl utilities for reliability annotation and rotation of rna structure plots

2003-01-29 lvo Hofacker ivo@tbi.univie.ac.at

· add RNALfold program to compute locally optimal structures with maximum pair span.

· add RNAcofold for computing hybrid structure

2002-11-07 Ivo Hofacker ivo@tbi.univie.ac.at

• change Make_bp_profile() and profile_edit_distance() to use simple (float *) arrays; makes Perl access much easier. RNApdist -B now works again

2002-10-28 Ivo Hofacker ivo@tbi.univie.ac.at

• Improved Perl module with pod documentation; allow to write things like (\$structure, \$energy) = RNA:::fold(\$seq); Compatibility warning: the ptrvalue() and related functions are gone, see the pod documentation for alternatives.

2002-10-29 Ivo Hofacker ivo@tbi.univie.ac.at

• added svg structure plots in PS_dot.c and RNAplot

2002-08-15 lvo Hofacker ivo@tbi.univie.ac.at

- · Improve reading of clustal files (alifold)
- · add a sample alifold.cgi script

2001-09-18 lvo Hofacker ivo@tbi.univie.ac.at

· moved suboptimal folding into the library, thus it's now accessible from the Perl module

2001-08-31 Ivo Hofacker ivo@tbi.univie.ac.at

· added co-folding support in energy_of_struct(), and thus RNAeval

2001-04-30 Ivo Hofacker ivo@tbi.univie.ac.at

· switch from handcrafted makefiles to automake and autoconf

2001-04-05 Ivo Hofacker ivo@tbi.univie.ac.at

• added PS_rna_plot_a to produce structure plots with annotation

2001-03-03 lvo Hofacker ivo@tbi.univie.ac.at

· add alifold; predict consensus structures from alignment

2000-09-28 Ivo Hofacker ivo@tbi.univie.ac.at

· add -d3 option to RNAfold for co-axial stacking

Chapter 11

Deprecated List

```
Global alifold (const char **strings, char *structure)
   Usage of this function is discouraged! Use vrna alifold(), or vrna mfe() instead!
Global alimake_pair_table (const char *structure)
   Use vrna_pt_ali_get() instead!
Global alipbacktrack (double *prob)
   Use vrna_pbacktrack() instead!
Global alipf_circ_fold (const char **sequences, char *structure, vrna_ep_t **pl)
   Use vrna_pf() instead
Global alipf_fold (const char **sequences, char *structure, vrna_ep_t **pl)
   Use vrna pf() instead
Global alipf_fold_par (const char **sequences, char *structure, vrna_ep_t **pl, vrna_exp_param_
   t *parameters, int calculate_bppm, int is_constrained, int is_circular)
   Use vrna_pf() instead
Global aliPS_color_aln (const char *structure, const char *filename, const char *seqs[], const char
   *names[])
   Use vrna_file_PS_aln() instead!
File aln util.h
   Use ViennaRNA/utils/alignments.h instead
Global assign plist from db (vrna ep t **pl, const char *struc, float pr)
   Use vrna_plist() instead
Global assign_plist_from_pr (vrna_ep_t **pl, FLT_OR_DBL *probs, int length, double cutoff)
   Use vrna_plist_from_probs() instead!
Global b2C (const char *structure)
   See vrna_db_to_tree_string() and VRNA_STRUCTURE_TREE_SHAPIRO_SHORT for a replacement
Global b2HIT (const char *structure)
   See vrna db to tree string() and VRNA STRUCTURE TREE HIT for a replacement
Global b2Shapiro (const char *structure)
   See vrna_db_to_tree_string() and VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT for a replacement
Global base_pair
   Do not use this variable anymore!
Global bondT
   Use vrna_bp_stack_t instead!
```

102 Deprecated List

Global bp_distance (const char *str1, const char *str2)

Use vrna_bp_distance instead

Global bppm_symbol (const float *x)

Use vrna_bpp_symbol() instead!

Global bppm to structure (char *structure, FLT OR DBL *pr, unsigned int length)

Use vrna_db_from_probs() instead!

Global centroid (int length, double *dist)

This function is deprecated and should not be used anymore as it is not threadsafe!

File char stream.h

Use ViennaRNA/datastructures/char stream.h instead

Global circalifold (const char **strings, char *structure)

Usage of this function is discouraged! Use vrna_alicircfold(), and vrna_mfe() instead!

Global circfold (const char *sequence, char *structure)

Use vrna_circfold(), or vrna_mfe() instead!

Global co_pf_fold (char *sequence, char *structure)

{Use vrna_pf_dimer() instead!}

Global co_pf_fold_par (char *sequence, char *structure, vrna_exp_param_t *parameters, int calculate_← bppm, int is constrained)

Use vrna_pf_dimer() instead!

Global cofold (const char *sequence, char *structure)

use vrna_mfe_dimer() instead

Global cofold par (const char *string, char *structure, vrna param t *parameters, int is constrained)

use vrna_mfe_dimer() instead

Global compute_BPdifferences (short *pt1, short *pt2, unsigned int turn)

Use vrna_refBPdist_matrix() instead

Global compute_probabilities (double FAB, double FEA, double FEB, vrna_ep_t *prAB, vrna_ep_t *prA, vrna_ep_t *prB, int Alength)

{ Use vrna_pf_dimer_probs() instead!}

Global constrain_ptypes (const char ∗constraint, unsigned int length, char ∗ptype, int ∗BP, int min_loop ← _size, unsigned int idx_type)

Do not use this function anymore! Structure constraints are now handled through vrna_hc_t and related functions.

File constraints.h

Use ViennaRNA/constraints/basic.h instead

File constraints_hard.h

Use ViennaRNA/constraints/hard.h instead

File constraints_ligand.h

Use ViennaRNA/constraints/ligand.h instead

File constraints_SHAPE.h

Use ViennaRNA/constraints/SHAPE.h instead

File constraints soft.h

Use ViennaRNA/constraints/soft.h instead

File convert_epars.h

Use ViennaRNA/params/convert.h instead

Global copy_pair_table (const short *pt)

Use vrna_ptable_copy() instead

Global cpair

Use vrna_cpair_t instead!

Global cv_fact

See vrna_md_t.cv_fact, and vrna_mfe() to avoid using global variables

File data structures.h

Use ViennaRNA/datastructures/basic.h instead

Global destroy TwoDfold variables (TwoDfold vars *our variables)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound ← __TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

Global destroy_TwoDpfold_variables (TwoDpfold_vars *vars)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound
_TwoD(), vrna_pf_TwoD(), and vrna_fold_compound_free() instead!

Global E Stem (int type, int si1, int sj1, int extLoop, vrna_param_t *P)

Please use one of the functions $vrna_E_ext_stem()$ and $E_MLstem()$ instead! Use the former for cases where extLoop != 0 and the latter otherwise.

File energy const.h

Use ViennaRNA/params/constants.h instead

Global energy of alistruct (const char **sequences, const char *structure, int n seq, float *energy)

Usage of this function is discouraged! Use vrna_eval_structure(), and vrna_eval_covar_structure() instead!

Global energy_of_circ_struct (const char *string, const char *structure)

This function is deprecated and should not be used in future programs Use energy_of_circ_structure() instead!

Global energy_of_circ_struct_par (const char *string, const char *structure, vrna_param_t *parameters, int verbosity_level)

Use vrna_eval_structure() or vrna_eval_structure_verbose() instead!

Global energy_of_circ_structure (const char *string, const char *structure, int verbosity_level)

Use vrna_eval_structure() or vrna_eval_structure_verbose() instead!

Global energy of move (const char *string, const char *structure, int m1, int m2)

Use vrna_eval_move() instead!

Global energy_of_move_pt (short *pt, short *s, short *s1, int m1, int m2)

Use vrna eval move pt() instead!

Global energy_of_struct (const char *string, const char *structure)

This function is deprecated and should not be used in future programs! Use energy of structure() instead!

Global energy_of_struct_par (const char *string, const char *structure, vrna_param_t *parameters, int verbosity_level)

Use vrna_eval_structure() or vrna_eval_structure_verbose() instead!

Global energy_of_struct_pt (const char *string, short *ptable, short *s, short *s1)

This function is deprecated and should not be used in future programs! Use energy of structure pt() instead!

Global energy_of_struct_pt_par (const char *string, short *ptable, short *s, short *s1, vrna_param_← t *parameters, int verbosity_level)

Use vrna_eval_structure_pt() or vrna_eval_structure_pt_verbose() instead!

Global energy_of_structure (const char *string, const char *structure, int verbosity_level)

Use vrna_eval_structure() or vrna_eval_structure_verbose() instead!

Global energy_of_structure_pt (const char *string, short *ptable, short *s, short *s1, int verbosity_level)

Use vrna_eval_structure_pt() or vrna_eval_structure_pt_verbose() instead!

File energy_par.h

Use ViennaRNA/params/default.h instead

104 Deprecated List

```
Global exp_E_ExtLoop (int type, int si1, int sj1, vrna_exp_param_t *P)
```

Use vrna_exp_E_ext_stem() instead!

Global expHairpinEnergy (int u, int type, short si1, short sj1, const char *string)

Use exp E Hairpin() from loop energies.h instead

Global expLoopEnergy (int u1, int u2, int type, int type2, short si1, short sj1, short sp1, short sq1)

Use exp_E_IntLoop() from loop_energies.h instead

Global export ali bppm (void)

Usage of this function is discouraged! The new vrna_fold_compound_t allows direct access to the folding matrices, including the pair probabilities! The pair probability array returned here reflects the one of the latest call to vrna_pf(), or any of the old API calls for consensus structure partition function folding.

```
Global export_circfold_arrays (int *Fc_p, int *FcH_p, int *FcI_p, int *FcM_p, int **fM2_p, int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p)
```

See vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

Global export_circfold_arrays_par (int *Fc_p, int *FcH_p, int *FcI_p, int *FcM_p, int **fM2_p, int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p, vrna_param_t **P_p)

See vrna mfe() and vrna fold compound t for the usage of the new API!

Global export_co_bppm (void)

This function is deprecated and will be removed soon! The base pair probability array is available through the vrna_fold_compound_t data structure, and its associated <a href="https://www.na_mx_pf_t member.

Global export_cofold_arrays (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **fc_p, int **indx_p, char **ptype_p)

folding matrices now reside within the vrna_fold_compound_t. Thus, this function will only work in conjunction with a prior call to the deprecated functions cofold() or cofold_par()

Global export_cofold_arrays_gq (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **fc_p, int **ggg_p, int **indx_p, char **ptype_p)

folding matrices now reside within the fold compound. Thus, this function will only work in conjunction with a prior call to cofold() or cofold_par()

```
Global export_fold_arrays (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p)

See vrna mfe() and vrna fold compound t for the usage of the new API!
```

```
Global export_fold_arrays_par (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype← _p, vrna_param_t **P_p)
```

See vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

File exterior loops.h

Use ViennaRNA/loops/external.h instead

File file formats.h

Use ViennaRNA/io/file formats.h instead

File file formats msa.h

Use ViennaRNA/io/file_formats_msa.h instead

File file utils.h

Use ViennaRNA/io/utils.h instead

Global filecopy (FILE *from, FILE *to)

Use vrna_file_copy() instead!

Global find_saddle (const char *seq, const char *s1, const char *s2, int width)

Use vrna path findpath saddle() instead!

File findpath.h

Use ViennaRNA/landscape/findpath.h instead

Global fold (const char *sequence, char *structure)

use vrna_fold(), or vrna_mfe() instead!

Global fold_par (const char *sequence, char *structure, vrna_param_t *parameters, int is_constrained, int is_circular)

use vrna_mfe() instead!

Global free_alifold_arrays (void)

Usage of this function is discouraged! It only affects memory being free'd that was allocated by an old API function before. Release of memory occupied by the newly introduced vrna_fold_compound_t is handled by vrna_fold_compound_free()

Global free_alipf_arrays (void)

Usage of this function is discouraged! This function only free's memory allocated by old API function calls. Memory allocated by any of the new API calls (starting with vrna_) will be not affected!

Global free arrays (void)

See vrna fold(), vrna circfold(), or vrna mfe() and vrna fold compound t for the usage of the new API!

Global free co arrays (void)

This function will only free memory allocated by a prior call of cofold() or cofold_par(). See vrna_mfe_dimer() for how to use the new API

Global free_co_pf_arrays (void)

This function will be removed for the new API soon! See vrna_pf_dimer(), vrna_fold_compound(), and vrna fold compound free() for an alternative

Global free path (vrna path t *path)

Use vrna path free() instead!

Global free_pf_arrays (void)

See vrna_fold_compound_t and its related functions for how to free memory occupied by the dynamic programming matrices

Global get_alipf_arrays (short ***S_p, short ***S5_p, short ***S3_p, unsigned short ***a2s_p, char ***← Ss_p, FLT_OR_DBL **qb_p, FLT_OR_DBL **qm_p, FLT_OR_DBL **q1k_p, FLT_OR_DBL **qln_p, short **pscore)

It is discouraged to use this function! The new vrna_fold_compound_t allows direct access to all necessary consensus structure prediction related variables!

Global get_boltzmann_factor_copy (vrna_exp_param_t *parameters)

Use vrna_exp_params_copy() instead!

Global get_boltzmann_factors (double temperature, double betaScale, vrna_md_t md, double pf_scale)

Use vrna_exp_params() instead!

Global get_boltzmann_factors_ali (unsigned int n_seq, double temperature, double betaScale, vrna_md_t md, double pf_scale)

Use vrna exp params comparative() instead!

Global get_centroid_struct_gquad_pr (int length, double *dist)

This function is deprecated and should not be used anymore as it is not threadsafe!

Global get_centroid_struct_pl (int length, double *dist, vrna_ep_t *pl)

This function was renamed to vrna_centroid_from_plist()

Global get_centroid_struct_pr (int length, double *dist, FLT_OR_DBL *pr)

This function was renamed to vrna_centroid_from_probs()

Global get_concentrations (double FEAB, double FEAA, double FEBB, double FEA, double FEB, double *startconc)

{ Use vrna_pf_dimer_concentrations() instead!}

Global get_line (FILE *fp)

Use vrna_read_line() as a substitute!

106 Deprecated List

Global get_monomere_mfes (float *e1, float *e2)

{This function is obsolete and will be removed soon!}

Global get_mpi (char *Alseq[], int n_seq, int length, int *mini)

Use vrna_aln_mpi() as a replacement

Global get_path (const char *seq, const char *s1, const char *s2, int width)

Use vrna_path_findpath() instead!

Global get plist (vrna ep t *pl, int length, double cut off)

{ This function is deprecated and will be removed soon!} use assign plist from pr() instead!

Global get_scaled_alipf_parameters (unsigned int n_seq)

Use vrna_exp_params_comparative() instead!

Global get scaled parameters (double temperature, vrna md t md)

Use vrna_params() instead!

Global get_scaled_pf_parameters (void)

Use vrna exp params() instead!

Global get_TwoDfold_variables (const char *seq, const char *structure1, const char *structure2, int circ)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound ← __TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

Global get_TwoDpfold_variables (const char *seq, const char *structure1, char *structure2, int circ)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound
_TwoD(), vrna_pf_TwoD(), and vrna_fold_compound_free() instead!

File hairpin loops.h

Use ViennaRNA/loops/hairpin.h instead

Global HairpinE (int size, int type, int si1, int sj1, const char *string)

{This function is deprecated and will be removed soon. Use E Hairpin() instead!}

Global hamming (const char *s1, const char *s2)

Use vrna_hamming_distance() instead!

Global hamming_bound (const char *s1, const char *s2, int n)

Use vrna_hamming_distance_bound() instead!

Global iindx

Do not use this variable anymore!

Global init_co_pf_fold (int length)

{ This function is deprecated and will be removed soon!}

Global init pf fold (int length)

This function is obsolete and will be removed soon!

Global init rand (void)

Use vrna_init_rand() instead!

Global initialize_cofold (int length)

{This function is obsolete and will be removed soon!}

Global initialize_fold (int length)

See vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

Global int_urn (int from, int to)

Use vrna_int_urn() instead!

File interior_loops.h

Use ViennaRNA/loops/internal.h instead

```
Global Lfold (const char *string, const char *structure, int maxdist)
   Use vrna_mfe_window() instead!
Global Lfoldz (const char *string, const char *structure, int maxdist, int zsc, double min_z)
   Use vrna_mfe_window_zscore() instead!
File loop energies.h
   Use ViennaRNA/loops/all.h instead
Global loop_energy (short *ptable, short *s, short *s1, int i)
   Use vrna_eval_loop_pt() instead!
Global LoopEnergy (int n1, int n2, int type, int type 2, int si1, int sj1, int sp1, int sq1)
   {This function is deprecated and will be removed soon. Use E_IntLoop() instead!}
Global Make bp profile (int length)
   This function is deprecated and will be removed soon! See Make_bp_profile_bppm() for a replacement
Global make pair table (const char *structure)
   Use vrna ptable() instead
Global make_pair_table_snoop (const char *structure)
   Use vrna_pt_snoop_get() instead!
Global make_referenceBP_array (short *reference_pt, unsigned int turn)
   Use vrna refBPcnt matrix() instead
Global MEA (plist *p, char *structure, double gamma)
   Use vrna_MEA() or vrna_MEA_from_plist() instead!
Global mean_bp_dist (int length)
   This function is not threadsafe and should not be used anymore. Use mean bp distance() instead!
Global mean_bp_distance (int length)
   Use vrna_mean_bp_distance() or vrna_mean_bp_distance_pr() instead!
Global mean bp distance pr (int length, FLT OR DBL *pr)
   Use vrna mean bp distance() or vrna mean bp distance pr() instead!
File multibranch_loops.h
   Use ViennaRNA/loops/multibranch.h instead
File naview.h
   Use ViennaRNA/plotting/naview.h instead
Global naview_xy_coordinates (short *pair_table, float *X, float *Y)
   Consider using vrna_plot_coords_naview_pt() instead!
Global nc fact
   See vrna_md_t.nc_fact, and vrna_mfe() to avoid using global variables
File neighbor.h
   Use ViennaRNA/landscape/neighbor.h instead
Global nrerror (const char message[])
   Use vrna_message_error() instead!
Global pack_structure (const char *struc)
   Use vrna_db_pack() as a replacement
Global PAIR
   Use vrna_basepair_t instead!
Global pair_info
```

Use vrna_pinfo_t instead!

108 **Deprecated List**

File params.h

Use ViennaRNA/params/basic.h instead

Global paramT

Use vrna_param_t instead!

Global parenthesis_structure (char *structure, vrna_bp_stack_t *bp, int length)

use vrna_parenthesis_structure() instead

Global parenthesis zuker (char *structure, vrna bp stack t *bp, int length)

use vrna_parenthesis_zuker instead

Global path_t

Use vrna_path_t instead!

Global pbacktrack_circ (char *sequence)

Use vrna pbacktrack() instead.

Global pf_circ_fold (const char *sequence, char *structure)

Use vrna pf() instead!

Global pf_fold_par (const char *sequence, char *structure, vrna_exp_param_t *parameters, int calculate ← _bppm, int is_constrained, int is_circular)

Use vrna pf() instead

Global pf_paramT

Use vrna_exp_param_t instead!

Global plist

Use vrna_ep_t or vrna_elem_prob_s instead!

File plot aln.h

Use ViennaRNA/plotting/alignments.h instead

File plot_layouts.h

Use ViennaRNA/plotting/layouts.h instead

File plot structure.h

Use ViennaRNA/plotting/structures.h instead

File plot utils.h

Use ViennaRNA/plotting/utils.h instead

Global pr

Do not use this variable anymore!

Global print_tty_constraint (unsigned int option)

Use vrna message constraints() instead!

Global print tty constraint full (void)

Use vrna_message_constraint_options_all() instead!

Global print_tty_input_seq (void)

Use vrna_message_input_seq_simple() instead!

Global print_tty_input_seq_str (const char *s)

Use vrna_message_input_seq() instead!

Global PS color aln (const char *structure, const char *filename, const char *seqs[], const char *names[])

Use vrna_file_PS_aln() instead!

File PS dot.h

Use ViennaRNA/plotting/probabilities.h instead

Global PS_dot_plot (char *string, char *file)

This function is deprecated and will be removed soon! Use PS_dot_plot_list() instead!

Global PS rna plot (char *string, char *structure, char *file) Use vrna_file_PS_rnaplot() instead! Global PS rna plot a (char *string, char *structure, char *file, char *pre, char *post) Use vrna_file_PS_rnaplot_a() instead! Global PS_rna_plot_a_gquad (char *string, char *structure, char *ssfile, char *pre, char *post) Use vrna_file_PS_rnaplot_a() instead! Global random_string (int I, const char symbols[]) Use vrna random string() instead! File read epars.h Use ViennaRNA/params/io.h instead Global read_parameter_file (const char fname[]) Use vrna params load() instead! Global read_record (char **header, char **sequence, char ***rest, unsigned int options) This function is deprecated! Use vrna file fasta read record() as a replacment. Global scale parameters (void) Use vrna_params() instead! Global sect Use vrna_sect_t instead! Global set_model_details (vrna_md_t *md) This function will vanish as soon as backward compatibility of RNAlib is dropped (expected in version 3). Use vrna md set default() instead! Global simple circulates (short *pair table, float *x, float *y) Consider switching to vrna_plot_coords_circular_pt() instead! Global simple_xy_coordinates (short *pair_table, float *X, float *Y) Consider switching to vrna_plot_coords_simple_pt() instead! **Global SOLUTION** Use vrna subopt solution tinstead! Global space (unsigned size) Use vrna_alloc() instead! Global st back set the *uniq_ML* flag in vrna_md_t before passing it to vrna_fold_compound(). Global stackProb (double cutoff) Use vrna stack prob() instead! Global str_DNA2RNA (char *sequence) Use vrna_seq_toRNA() instead! Global str_uppercase (char *sequence) Use vrna_seq_toupper() instead! File stream_output.h Use ViennaRNA/datastructures/stream_output.h instead File string utils.h Use ViennaRNA/utils/strings.h instead File structure utils.h

File svm_utils.h

Use ViennaRNA/utils/structures.h instead

Use ViennaRNA/utils/svm.h instead

110 Deprecated List

Global temperature

Use vrna_md_defaults_temperature(), and vrna_md_defaults_temperature_get() to change, and read the global default temperature settings

Global time_stamp (void)

Use vrna_time_stamp() instead!

Global TwoDfold_backtrack_f5 (unsigned int j, int k, int l, TwoDfold_vars *vars)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound
— TwoD(), vrna_mfe_TwoD(), vrna_backtrack5_TwoD(), and vrna_fold_compound_free() instead!

Global TwoDfold vars

This data structure will be removed from the library soon! Use vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

Global TwoDfoldList (TwoDfold_vars *vars, int distance1, int distance2)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound ← __TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

Global TwoDpfold_pbacktrack (TwoDpfold_vars *vars, int d1, int d2)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound ← TwoD(), vrna_pf TwoD(), vrna_pbacktrack_TwoD(), and vrna_fold_compound_free() instead!

Global TwoDpfold_pbacktrack5 (TwoDpfold_vars *vars, int d1, int d2, unsigned int length)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound ← _TwoD(), vrna_pf_TwoD(), vrna_pbacktrack5_TwoD(), and vrna_fold_compound_free() instead!

Class TwoDpfold_vars

This data structure will be removed from the library soon! Use vrna_fold_compound_t and the corresponding functions vrna_fold_compound_trop(), vrna_pf_TwoD(), and vrna_fold_compound_free() instead!

Global TwoDpfoldList (TwoDpfold vars *vars, int maxDistance1, int maxDistance2)

Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound ← _TwoD(), vrna_pf_TwoD(), and vrna_fold_compound_free() instead!

Global unpack_structure (const char *packed)

Use vrna_db_unpack() as a replacement

Global update_alifold_params (void)

Usage of this function is discouraged! The new API uses vrna_fold_compound_t to lump all folding related necessities together, including the energy parameters. Use vrna_update_fold_params() to update the energy parameters within a vrna_fold_compound_t.

Global update_co_pf_params (int length)

Use vrna_exp_params_subst() instead!

Global update co pf params par (int length, vrna exp param t *parameters)

Use vrna_exp_params_subst() instead!

Global update cofold params (void)

See vrna_params_subst() for an alternative using the new API

Global update_cofold_params_par (vrna_param_t *parameters)

See vrna_params_subst() for an alternative using the new API

Global update fold params (void)

For non-default model settings use the new API with vrna_params_subst() and vrna_mfe() instead!

Global update fold params par (vrna param t *parameters)

For non-default model settings use the new API with vrna_params_subst() and vrna_mfe() instead!

Global update_pf_params (int length)

Use vrna_exp_params_subst() instead

Global update_pf_params_par (int length, vrna_exp_param_t *parameters)

Use vrna_exp_params_subst() instead

Global urn (void)

Use vrna_urn() instead!

File utils.h

Use ViennaRNA/utils/basic.h instead

Global VRNA CONSTRAINT FILE

Use 0 instead!

Global VRNA_CONSTRAINT_MULTILINE

see vrna_extract_record_rest_structure()

Global VRNA_CONSTRAINT_NO_HEADER

This mode is not supported anymore!

Global VRNA_CONSTRAINT_SOFT_MFE

This flag has no meaning anymore, since constraints are now always stored!

Global VRNA_CONSTRAINT_SOFT_PF

Use VRNA_OPTION_PF instead!

Global vrna_exp_param_s::id

This attribute will be removed in version 3

Global vrna_extract_record_rest_constraint (char **cstruc, const char **lines, unsigned int option)

Use vrna_extract_record_rest_structure() instead!

Global vrna_fc_s::pscore_pf_compat

This attribute will vanish in the future!

Global vrna_fc_s::ptype_pf_compat

This attribute will vanish in the future! It's meant for backward compatibility only!

File walk.h

Use ViennaRNA/landscape/walk.h instead

Global warn_user (const char message[])

Use vrna_message_warning() instead!

Global write_parameter_file (const char fname[])

Use vrna_params_save() instead!

Global xrealloc (void *p, unsigned size)

Use vrna_realloc() instead!

Global zukersubopt (const char *string)

use vrna_zukersubopt() instead

Global zukersubopt_par (const char *string, vrna_param_t *parameters)

use vrna zukersubopt() instead

112 Deprecated List

Chapter 12

Bug List

Module domains_up

Although the additional production rule(s) for unstructured domains as descibed in Unstructured Domains are always treated as 'segments possibly bound to one or more ligands', the current implementation requires that at least one ligand is bound. The default implementation already takes care of the required changes, however, upon using callback functions other than the default ones, one has to take care of this fact. Please also note, that this behavior might change in one of the next releases, such that the decomposition schemes as shown above comply with the actual implementation.

Global VRNA_PROBS_WINDOW_STACKP

Currently, this flag is a placeholder doing nothing as the corresponding implementation for stack probability computation is missing.

Global vrna_subopt_zuker (vrna_fold_compound_t *vc)

Due to resizing, any pre-existing constraints will be lost!

114 Bug List

Chapter 13

Module Index

13.1 The RNAlib API

Our library is grouped into several modules, each addressing different aspects of RNA secondary structure related problems. You can find an overview of the different groups below.

Free Energy Evaluation	27
Energy Evaluation for Individual Loops	52
Exterior Loops	30
Hairpin Loops	34
Internal Loops	39
Multibranch Loops	
Energy Evaluation for Atomic Moves	
Deprecated Interface for Free Energy Evaluation	57
The RNA Folding Grammar	72
Fine-tuning of the Implemented Models	73
Energy Parameters	ე9
Reading/Writing Energy Parameter Sets from/to File	48
Converting Energy Parameter Files	56
Extending the Folding Grammar with Additional Domains	23
Unstructured Domains	24
Structured Domains	36
Constraining the RNA Folding Grammar	
Hard Constraints	54
Soft Constraints	66
The RNA Secondary Structure Landscape	78
Neighborhood Relation and Move Sets for Secondary Structures	75
(Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima	38
Direct Refolding Paths between two Secondary Structures	
Folding Paths that start at a single Secondary Structure	
Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers 69	96
Minimum Free Energy (MFE) Algorithms	79
Global MFE Prediction	32
Computing MFE representatives of a Distance Based Partitioning	56
Deprecated Interface for Global MFE Prediction	40
Local (sliding window) MFE Prediction	39
Deprecated Interface for Local (Sliding Window) MFE Prediction	54
Backtracking MFE structures	94

116 Module Index

Partition Function and Equilibrium Properties	280
Global Partition Function and Equilibrium Probabilities	. 297
Computing Partition Functions of a Distance Based Partitioning	. 365
Deprecated Interface for Global Partition Function Computation	. 655
Local (sliding window) Partition Function and Equilibrium Probabilities	. 311
Deprecated Interface for Local (Sliding Window) Partition Function Computation	673
Suboptimals and Representative Structures	321
Suboptimal Structures sensu Stiegler et al. 1984 / Zuker et al. 1989	. 322
Suboptimal Structures within an Energy Band around the MFE	
Stochastic Backtracking of Structures from Distance Based Partitioning	
Deprecated Interface for Stochastic Backtracking	
Compute the Structure with Maximum Expected Accuracy (MEA)	. 347
Compute the Centroid Structure	. 350
RNA-RNA Interaction	353
Partition Function for Two Hybridized Sequences	. 442
Partition Function for two Hybridized Sequences as a Stepwise Process	
Classified Dynamic Programming Variants	354
Distance Based Partitioning of the Secondary Structure Space	. 355
Computing MFE representatives of a Distance Based Partitioning	
Computing Partition Functions of a Distance Based Partitioning	
Stochastic Backtracking of Structures from Distance Based Partitioning	
Compute the Density of States	
Inverse Folding (Design)	
Experimental Structure Probing Data	
SHAPE Reactivity Data	
Ligands Binding to RNA Structures	
	710
Ligands Binding to Unstructured Domains	. 416 . 417
Ligands Binding to Unstructured Domains	. 416 . 417
Ligands Binding to Unstructured Domains	. 416 . 417 418
Ligands Binding to Unstructured Domains	. 416 . 417 418 . 419
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities	. 416 . 417 418 . 419 422 . 462
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilitites	. 416 . 417 418 . 419 422 . 462 . 466
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilitites Secondary Structure Utilities	. 416 . 417 418 . 419 422 . 462 . 466 . 475
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures	. 416 . 417 418 . 419 422 . 462 . 466 . 475
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures	. 416 . 417 . 418 . 419 . 422 . 462 . 466 . 475 . 479 . 487
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures	. 416 . 417 . 418 . 419 . 422 . 462 . 466 . 475 . 479 . 487 . 491
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Helix List Representation of Secondary Structures Helix List Representation of Secondary Structures	.416 .417 .418 .419 .422 .462 .466 .475 .479 .487 .491 .493
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures	. 416 . 417 . 418 . 419 . 422 . 462 . 466 . 475 . 479 . 487 . 491 . 493 . 495
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities	. 416 . 417 418 . 419 422 . 462 . 466 . 475 . 479 . 487 . 491 . 493 . 495 . 681
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilitites Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities	. 416 . 417 . 418 . 419 . 422 . 462 . 466 . 475 . 479 . 487 . 491 . 493 . 495 . 681 . 501
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Deprecated Interface for Multiple Sequence Alignment Utilities	. 416 . 417 . 418 . 419 . 462 . 466 . 475 . 479 . 487 . 491 . 493 . 495 . 681 . 501 . 678
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Deprecated Interface for Multiple Sequence Alignment Utilities Files and I/O	. 416 . 417 . 418 . 419 . 462 . 466 . 475 . 479 . 487 . 491 . 493 . 495 . 681 . 501 . 678 . 511
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Deprecated Interface for Multiple Sequence Alignment Utilities Files and I/O Nucleic Acid Sequences and Structures	. 416 . 417 . 418 . 419 . 422 . 462 . 466 . 475 . 479 . 487 . 491 . 493 . 681 . 501 . 678 . 511 . 515
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Deprecated Interface for Multiple Sequence Alignment Utilities Files and I/O	. 416 . 417 . 418 . 419 . 422 . 462 . 466 . 475 . 479 . 491 . 493 . 495 . 681 . 501 . 678 . 511 . 515 . 523
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilities Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Files and I/O Nucleic Acid Sequences and Structures Multiple Sequence Alignments	. 416 . 417 . 418 . 419 . 422 . 462 . 466 . 475 . 479 . 491 . 501 . 678 . 511 . 515 . 523 . 532
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilitites Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Files and I/O Nucleic Acid Sequences and Structures Multiple Sequence Alignments Command Files	. 416 . 417 . 418 . 419 . 462 . 466 . 475 . 479 . 487 . 491 . 681 . 501 . 678 . 511 . 515 . 523 . 532 . 537
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilitites Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Deprecated Interface for Multiple Sequence Alignment Utilities Files and I/O Nucleic Acid Sequences and Structures Multiple Sequence Alignments Command Files Plotting Layouts and Coordinates Annotation	. 416 . 417 . 418 . 419 . 422 . 462 . 466 . 475 . 479 . 491 . 501 . 678 . 511 . 515 . 523 . 532 . 537 . 544 . 563
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilitites Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Deprecated Interface for Multiple Sequence Alignment Utilities Files and I/O Nucleic Acid Sequences and Structures Multiple Sequence Alignments Command Files Plotting Layouts and Coordinates Annotation Alignment Plots	. 416 . 417 . 418 . 419 . 462 . 466 . 475 . 479 . 487 . 491 . 501 . 501 . 515 . 523 . 532 . 532 . 537 . 544 . 563 . 564
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilitites Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Deprecated Interface for Multiple Sequence Alignment Utilities Files and I/O Nucleic Acid Sequences and Structures Multiple Sequence Alignments Command Files Plotting Layouts and Coordinates Annotation Alignment Plots Deprecated Interface for Plotting Utilities	. 416 . 417 . 418 . 419 . 422 . 466 . 475 . 479 . 487 . 491 . 501 . 678 . 511 . 515 . 523 . 532 . 537 . 544 . 692
Ligands Binding to Unstructured Domains Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints Complex Structured Modules G-Quadruplexes Utilities Utilities to deal with Nucleotide Alphabets (Nucleic Acid Sequence) String Utilitites Secondary Structure Utilities Dot-Bracket Notation of Secondary Structures Pair Table Representation of Secondary Structures Pair List Representation of Secondary Structures Helix List Representation of Secondary Structures Tree Representation of Secondary Structures Deprecated Interface for Secondary Structure Utilities Multiple Sequence Alignment Utilities Deprecated Interface for Multiple Sequence Alignment Utilities Files and I/O Nucleic Acid Sequences and Structures Multiple Sequence Alignments Command Files Plotting Layouts and Coordinates Annotation Alignment Plots	. 416 . 417 . 418 . 419 . 422 . 466 . 475 . 479 . 487 . 491 . 501 . 678 . 511 . 515 . 523 . 532 . 537 . 544 . 692 . 566

13.1 The RNAlib API

(Abstract) Data Structures	 	 576
The Fold Compound	 	 593
The Dynamic Programming Matrices	 	 610
Hash Tables	 	 615
Heaps	 	 626
Buffers	 	 634
Messages	 	 583
Unit Conversion	 	 589

118 Module Index

Chapter 14

Data Structure Index

14.1 Data Structures

Here are the data structures with brief descriptions:

_struct_en	
Data structure for energy_of_move()	96
LIST	96
LST_BUCKET 69	96
Postorder_list	
Postorder data structure	00
swString	
Some other data structure	00
Tree	
Tree data structure	00
TwoDpfold_vars	
Variables compound for 2Dfold partition function folding	00
vrna_dimer_conc_s	
Data structure for concentration dependency computations	01
vrna_hc_bp_storage_t	
A base pair hard constraint	02
vrna_sc_bp_storage_t	
A base pair constraint	
vrna_sc_motif_s	03
vrna_structured_domains_s	03
vrna_subopt_sol_s	
Solution element from subopt.c	03
vrna unstructured domain motif s	03

120 Data Structure Index

Chapter 15

File Index

15.1 File List

Here is a list of all documented files with brief descriptions:

ViennaRNA/2Dfold.h	
MFE structures for base pair distance classes	705
ViennaRNA/2Dpfold.h	
Partition function implementations for base pair distance classes	706
ViennaRNA/ali_plex.h	??
ViennaRNA/alifold.h	
Functions for comparative structure prediction using RNA sequence alignments	710
ViennaRNA/aln_util.h	
Use ViennaRNA/utils/alignments.h instead	713
ViennaRNA/alphabet.h	
Functions to process, convert, and generally handle different nucleotide and/or base pair alpha-	
bets	713
ViennaRNA/boltzmann_sampling.h	
Boltzmann Sampling of secondary structures from the ensemble	713
ViennaRNA/centroid.h	
Centroid structure computation	715
ViennaRNA/char_stream.h	
Use ViennaRNA/datastructures/char_stream.h instead	716
ViennaRNA/cofold.h	
MFE implementations for RNA-RNA interaction	717
ViennaRNA/combinatorics.h	
Various implementations that deal with combinatorial aspects of objects	717
ViennaRNA/commands.h	
Parse and apply different commands that alter the behavior of secondary structure prediction	
and evaluation	718
ViennaRNA/concentrations.h	
Concentration computations for RNA-RNA interactions	719
ViennaRNA/constraints.h	
Use ViennaRNA/constraints/basic.h instead	720
ViennaRNA/constraints_hard.h	
Use ViennaRNA/constraints/hard.h instead	730
ViennaRNA/constraints_ligand.h	
Use ViennaRNA/constraints/ligand.h instead	730
ViennaRNA/constraints_SHAPE.h	
Use ViennaRNA/constraints/SHAPE h instead	730

122 File Index

ViennaRNA/constraints_soft.h	
Use ViennaRNA/constraints/soft.h instead	730
ViennaRNA/convert_epars.h Use ViennaRNA/params/convert.h instead	731
ViennaRNA/data structures.h	
Use ViennaRNA/datastructures/basic.h instead	731
ViennaRNA/dist_vars.h Global variables for Distance-Package	733
ViennaRNA/dp_matrices.h	
Functions to deal with standard dynamic programming (DP) matrices	734
ViennaRNA/duplex.h Functions for simple RNA-RNA duplex interactions	735
ViennaRNA/edit_cost.h Global variables for Edit Costs included by treedist.c and stringdist.c	735
ViennaRNA/energy const.h	
Use ViennaRNA/params/constants.h instead	736
ViennaRNA/energy_par.h	
Use ViennaRNA/params/default.h instead	736
Equilibrium Probability implementations	736
ViennaRNA/eval.h	730
Functions and variables related to energy evaluation of sequence/structure pairs	737
ViennaRNA/exterior_loops.h	
Use ViennaRNA/loops/external.h instead	740
ViennaRNA/file_formats.h Use ViennaRNA/io/file_formats.h instead	740
ViennaRNA/file_formats_msa.h	
Use ViennaRNA/io/file_formats_msa.h instead	741
ViennaRNA/file_utils.h Use ViennaRNA/io/utils.h instead	743
ViennaRNA/findpath.h	
Use ViennaRNA/landscape/findpath.h instead	743
ViennaRNA/fold.h	
MFE calculations for single RNA sequences	744
ViennaRNA/fold_compound.h	
The Basic Fold Compound API	745
ViennaRNA/fold_vars.h Here all all declarations of the global variables used throughout RNAlib	746
ViennaRNA/gquad.h	
G-quadruplexes	748
ViennaRNA/grammar.h	
Implementations for the RNA folding grammar	/48
ViennaRNA/hairpin_loops.h Use ViennaRNA/loops/hairpin.h instead	749
ViennaRNA/interior loops.h	
Use ViennaRNA/loops/internal.h instead	749
ViennaRNA/inverse.h	
Inverse folding routines	749
ViennaRNA/Lfold.h Functions for locally optimal MFE structure prediction	752
·	102
ViennaRNA/loop_energies.h Use ViennaRNA/loops/all.h instead	752
ViennaRNA/LPfold.h	
Partition function and equilibrium probability implementation for the sliding window algorithm .	756
ViennaRNA/MEA.h Computes a MEA (maximum expected accuracy) structure	757
Computes a MEA (maximum expected accuracy) structure	131

15.1 File List 123

ViennaRNA/mfe.h	
Compute Minimum Free energy (MFE) and backtrace corresponding secondary structures from RNA sequence data	757
ViennaRNA/mfe_window.h	
Compute local Minimum Free Energy (MFE) using a sliding window approach and backtrace corresponding secondary structures	758
ViennaRNA/mm.h	750
Several Maximum Matching implementations	759
ViennaRNA/model.h	750
The model details data structure and its corresponding modifiers	760
ViennaRNA/move_set.h	??
ViennaRNA/multibranch_loops.h	
Use ViennaRNA/loops/multibranch.h instead	765
ViennaRNA/naview.h	700
Use ViennaRNA/plotting/naview.h instead	761
· · · ·	765
ViennaRNA/neighbor.h	701
Use ViennaRNA/landscape/neighbor.h instead	
ViennaRNA/pair_mat.h	??
ViennaRNA/params.h	70-
Use ViennaRNA/params/basic.h instead	767
ViennaRNA/part_func.h	
Partition function implementations	782
ViennaRNA/part_func_co.h	
Partition function for two RNA sequences	785
ViennaRNA/part_func_up.h	
Implementations for accessibility and RNA-RNA interaction as a stepwise process	786
ViennaRNA/part_func_window.h	
Partition function and equilibrium probability implementation for the sliding window algorithm .	787
ViennaRNA/perturbation_fold.h	
Find a vector of perturbation energies that minimizes the discripancies between predicted and	
observed pairing probabilities and the amount of neccessary adjustments	788
observed pairing probabilities and the amount of neccessary adjustments	??
· · · · · · · · · · · · · · · · · · ·	
ViennaRNA/ PKplex.h	??
ViennaRNA/ PKplex.h	?? ??
ViennaRNA/PKplex.h	?? ??
ViennaRNA/PKplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead	?? ?? 789
ViennaRNA/PKplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h	?? ?? 789
ViennaRNA/PKplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead	789 789
ViennaRNA/PKplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h	789 789
ViennaRNA/PKplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead	789 789 789
ViennaRNA/PKplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h	789 789 789 790
ViennaRNA/Pkplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/instead ViennaRNA/ProfileAln.h	789 789 789 790 790
ViennaRNA/Pkplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/utils.h instead ViennaRNA/ProfileAln.h ViennaRNA/profiledist.h	789 789 789 790 790
ViennaRNA/Pkplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h	789 789 789 790 790 799
ViennaRNA/Pkplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h	789 789 789 790 790 799
ViennaRNA/Pkplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/utils.h instead ViennaRNA/ProfileAln.h ViennaRNA/profiledist.h ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/Ps_dot.h Use ViennaRNA/plotting/probabilities.h instead	790 790 790 790 790 801
ViennaRNA/Pkplex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/utils.h instead ViennaRNA/ProfileAln.h ViennaRNA/Profiledist.h ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/Ps_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead	790 790 790 790 790 801
ViennaRNA/Pkplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/utils.h instead ViennaRNA/ProfileAln.h ViennaRNA/Profiledist.h ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h	790 790 790 790 790 801 801
ViennaRNA/Pkplex.h ViennaRNA/plex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/utils.h instead ViennaRNA/ProfileAln.h ViennaRNA/Profiledist.h ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments	790 790 790 790 790 801 801
ViennaRNA/Pkplex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/utils.h instead ViennaRNA/ProfileAIn.h ViennaRNA/Profiledist.h ViennaRNA/Ps_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments ViennaRNA/RNAstruct.h	789 789 789 790 790 21 799 801 801
ViennaRNA/PKplex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/profileAln.h ViennaRNA/ProfileAln.h ViennaRNA/profiledist.h ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments ViennaRNA/RNAstruct.h Parsing and Coarse Graining of Structures	71 789 789 789 790 790 21 799 80° 80° 80°
ViennaRNA/Pkplex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/utils.h instead ViennaRNA/profileAln.h ViennaRNA/ProfileAln.h ViennaRNA/Ps_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments ViennaRNA/RNAstruct.h Parsing and Coarse Graining of Structures ViennaRNA/sequence.h	790 790 790 790 801 802 802
ViennaRNA/Pkplex.h ViennaRNA/plot_aln.h	790 790 790 790 790 801 802 802 804
ViennaRNA/Pkplex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/utils.h instead ViennaRNA/ProfileAln.h ViennaRNA/ProfileAln.h ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments ViennaRNA/RNAstruct.h Parsing and Coarse Graining of Structures ViennaRNA/sequence.h Functions and data structures related to sequence representations , ViennaRNA/snofold.h	790 790 790 790 790 801 802 802 804 79
ViennaRNA/PKplex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/ProfileAln.h ViennaRNA/ProfileAln.h ViennaRNA/ProfileAln.h ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments ViennaRNA/RNAstruct.h Parsing and Coarse Graining of Structures ViennaRNA/sequence.h Functions and data structures related to sequence representations, ViennaRNA/snofold.h ViennaRNA/snofold.h	789 789 789 789 790 790 91 801 802 802 804 97 97
ViennaRNA/Pkplex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/profileAln.h ViennaRNA/ProfileAln.h ViennaRNA/Ps_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/Ps_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments ViennaRNA/RNAstruct.h Parsing and Coarse Graining of Structures ViennaRNA/sequence.h Functions and data structures related to sequence representations, ViennaRNA/snofold.h ViennaRNA/snoop.h ViennaRNA/snoop.h	789 789 789 789 790 790 91 801 802 802 804 97 97
ViennaRNA/PKplex.h ViennaRNA/plot_aln.h Use ViennaRNA/plotting/alignments.h instead ViennaRNA/plot_layouts.h Use ViennaRNA/plotting/layouts.h instead ViennaRNA/plot_structure.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/plot_utils.h Use ViennaRNA/plotting/structures.h instead ViennaRNA/ProfileAln.h ViennaRNA/ProfileAln.h ViennaRNA/ProfileAln.h ViennaRNA/PS_dot.h Use ViennaRNA/plotting/probabilities.h instead ViennaRNA/read_epars.h Use ViennaRNA/params/io.h instead ViennaRNA/ribo.h Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments ViennaRNA/RNAstruct.h Parsing and Coarse Graining of Structures ViennaRNA/sequence.h Functions and data structures related to sequence representations, ViennaRNA/snofold.h ViennaRNA/snofold.h	790 790 790 790 801 802 802 804 21 21

124 File Index

ViennaRNA/string_utils.h	
Use ViennaRNA/utils/strings.h instead	805
ViennaRNA/stringdist.h	
Functions for String Alignment	806
ViennaRNA/structure_utils.h	
Use ViennaRNA/utils/structures.h instead	807
ViennaRNA/structured_domains.h	
This module provides interfaces that deal with additional structured domains in the folding gram-	
mar	807
ViennaRNA/subopt.h	
RNAsubopt and density of states declarations	807
ViennaRNA/svm_utils.h	
Use ViennaRNA/utils/svm.h instead	809
ViennaRNA/treedist.h	
Functions for Tree Edit Distances	809
ViennaRNA/ ugly_bt.h	??
ViennaRNA/units.h	
Physical Units and Functions to convert them into each other	811
ViennaRNA/unstructured domains.h	
Functions to modify unstructured domains, e.g. to incorporate ligands binding to unpaired	
stretches	811
ViennaRNA/utils.h	
Use ViennaRNA/utils/basic.h instead	814
ViennaRNA/ vrna_config.h	??
ViennaRNA/walk.h	• • •
Use ViennaRNA/landscape/walk.h instead	818
ViennaRNA/constraints/basic.h	010
Functions and data structures for constraining secondary structure predictions and evaluation .	770
ViennaRNA/constraints/hard.h	770
Functions and data structures for handling of secondary structure hard constraints	721
ViennaRNA/constraints/ligand.h	721
Functions for incorporation of ligands binding to hairpin and interior loop motifs using the soft	
constraints framework	726
ViennaRNA/constraints/SHAPE.h	720
This module provides function to incorporate SHAPE reactivity data into the folding recursions by means of soft constraints	727
ViennaRNA/constraints/soft.h	121
Functions and data structures for secondary structure soft constraints	700
	120
ViernaRNA/datastructures/basic.h	776
Various data structures and pre-processor macros	//6
ViennaRNA/datastructures/char_stream.h	710
Implementation of a dynamic, buffered character stream	/16
ViennaRNA/datastructures/hash_tables.h	704
Implementations of hash table functions	/31
ViennaRNA/datastructures/heap.h	
Implementation of an abstract heap data structure	
ViennaRNA/datastructures/lists.h	??
ViennaRNA/datastructures/stream_output.h	
An implementation of a buffered, ordered stream output data structure	805
ViennaRNA/io/file_formats.h	
Read and write different file formats for RNA sequences, structures	741
ViennaRNA/io/file_formats_msa.h	
Functions dealing with file formats for Multiple Sequence Alignments (MSA)	742
ViennaRNA/io/utils.h	
Several utilities for file handling	814
ViennaRNA/landscape/findpath.h	
A breadth-first search heuristic for optimal direct folding paths	743

15.1 File List 125

ViennaRNA/landscape/move.h	
Methods to operate with structural neighbors of RNA secondary structures	750
ViennaRNA/landscape/neighbor.h	
Methods to compute the neighbors of an RNA secondary structure	766
ViennaRNA/landscape/paths.h	
API for computing (optimal) (re-)folding paths between secondary structures	751
ViennaRNA/landscape/walk.h	
Methods to generate particular paths such as gradient or random walks through the energy	
landscape of an RNA sequence	818
ViennaRNA/loops/all.h	==-
Energy evaluation for MFE and partition function calculations	753
ViennaRNA/loops/external.h	===
Energy evaluation of exterior loops for MFE and partition function calculations	753
ViennaRNA/loops/hairpin.h	754
Energy evaluation of hairpin loops for MFE and partition function calculations	754
ViennaRNA/loops/internal.h	754
Energy evaluation of interior loops for MFE and partition function calculations	754
ViennaRNA/loops/multibranch.h	755
Energy evaluation of multibranch loops for MFE and partition function calculations	755
ViennaRNA/params/1.8.4_epars.h	767
Free energy parameters for parameter file conversion	767
ViennaRNA/params/1.8.4_intloops.h Free energy parameters for interior loop contributions needed by the parameter file conversion	
functions	768
ViennaRNA/params/basic.h	700
Functions to deal with sets of energy parameters	768
ViennaRNA/params/constants.h	700
Energy parameter constants	778
ViennaRNA/params/convert.h	770
Functions and definitions for energy parameter file format conversion	780
ViennaRNA/params/default.h	??
ViennaRNA/params/intl11.h	??
ViennaRNA/params/intl11dH.h	??
ViennaRNA/params/intl21.h	??
ViennaRNA/params/intl21dH.h	??
ViennaRNA/params/intl22.h	??
ViennaRNA/params/intl22dH.h	??
ViennaRNA/params/io.h	• •
Read and write energy parameter files	781
ViennaRNA/plotting/alignments.h	701
Various functions for plotting Sequence / Structure Alignments	790
ViennaRNA/plotting/layouts.h	
Secondary structure plot layout algorithms	792
ViennaRNA/plotting/naview.h	
Implementation of the Naview RNA secondary structure layout algorithm [5]	765
ViennaRNA/plotting/probabilities.h	
Various functions for plotting RNA secondary structures, dot-plots and other visualizations	794
ViennaRNA/plotting/structures.h	
Various functions for plotting RNA secondary structures	795
ViennaRNA/plotting/utils.h	
Various utilities to assist in plotting secondary structures and consensus structures	815
ViennaRNA/plotting/RNApuzzler/RNApuzzler.h	
Implementation of the RNApuzzler RNA secondary structure layout algorithm [23]	794
ViennaRNA/plotting/RNApuzzler/RNAturtle.h	
Implementation of the RNAturtle RNA secondary structure layout algorithm [23]	795
ViennaRNA/search/BoyerMoore.h	
Variants of the Boyer-Moore string search algorithm	803

126 File Index

ViennaRNA/utils/alignments.h	
Various utility- and helper-functions for sequence alignments and comparative structure predic-	
tion	791
ViennaRNA/utils/basic.h	
General utility- and helper-functions used throughout the ViennaRNA Package	771
ViennaRNA/utils/cpu.h	??
ViennaRNA/utils/higher_order_functions.h	??
ViennaRNA/utils/strings.h	
General utility- and helper-functions for RNA sequence and structure strings used throughout the	
ViennaRNA Package	815
ViennaRNA/utils/structures.h	
Various utility- and helper-functions for secondary structure parsing, converting, etc	796
ViennaRNA/utils/svm.h	??

Chapter 16

Module Documentation

16.1 Free Energy Evaluation

Functions and variables related to free energy evaluation of sequence/structure pairs.

16.1.1 Detailed Description

Functions and variables related to free energy evaluation of sequence/structure pairs.

Several different functions to evaluate the free energy of a particular secondary structure under a particular set of parameters and the Nearest Neighbor Energy model are available. For most of them, two different forms of representations for the secondary structure may be used:

- · The Dot-Bracket string
- · A pair table representation

Furthermore, the evaluation functions are divided into <code>basic</code> and <code>simplified</code> variants, where <code>basic</code> functions require the use of a <code>vrna_fold_compound_t</code> data structure holding the sequence string, and model configuration (settings and parameters). The <code>simplified</code> functions, on the other hand, provide often used default model settings that may be called directly with only sequence and structure data.

Finally, verbose options exist for some functions that allow one to print the (individual) free energy contributions to some FILE stream. Collaboration diagram for Free Energy Evaluation:

Modules

· Energy Evaluation for Individual Loops

Functions to evaluate the free energy of particular types of loops.

• Energy Evaluation for Atomic Moves

Functions to evaluate the free energy change of a structure after application of (a set of) atomic moves.

Deprecated Interface for Free Energy Evaluation

Deprecated Energy Evaluation functions.

128 Module Documentation

Files

· file eval.h

Functions and variables related to energy evaluation of sequence/structure pairs.

file all.h

Energy evaluation for MFE and partition function calculations.

· file external.h

Energy evaluation of exterior loops for MFE and partition function calculations.

· file hairpin.h

Energy evaluation of hairpin loops for MFE and partition function calculations.

· file internal.h

Energy evaluation of interior loops for MFE and partition function calculations.

· file multibranch.h

Energy evaluation of multibranch loops for MFE and partition function calculations.

Macros

#define VRNA VERBOSITY QUIET -1

Quiet level verbosity setting.

#define VRNA VERBOSITY DEFAULT 1

Default level verbosity setting.

Basic Energy Evaluation Interface with Dot-Bracket Structure String

float vrna_eval_structure (vrna_fold_compound_t *vc, const char *structure)

Calculate the free energy of an already folded RNA.

• float vrna_eval_covar_structure (vrna_fold_compound_t *vc, const char *structure)

Calculate the pseudo energy derived by the covariance scores of a set of aligned sequences.

• float vrna_eval_structure_verbose (vrna_fold_compound_t *vc, const char *structure, FILE *file)

Calculate the free energy of an already folded RNA and print contributions on a per-loop base.

- float vrna_eval_structure_v (vrna_fold_compound_t *vc, const char *structure, int verbosity_level, FILE *file)

 Calculate the free energy of an already folded RNA and print contributions on a per-loop base.
- float **vrna_eval_structure_cstr** (vrna_fold_compound_t *vc, const char *structure, int verbosity_level, vrna_cstr_t output_stream)

Basic Energy Evaluation Interface with Structure Pair Table

• int vrna_eval_structure_pt (vrna_fold_compound_t *vc, const short *pt)

Calculate the free energy of an already folded RNA.

• int vrna_eval_structure_pt_verbose (vrna_fold_compound_t *vc, const short *pt, FILE *file)

Calculate the free energy of an already folded RNA.

• int vrna eval structure pt v (vrna fold compound t *vc, const short *pt, int verbosity level, FILE *file)

Calculate the free energy of an already folded RNA.

Simplified Energy Evaluation with Sequence and Dot-Bracket Strings

- float vrna_eval_structure_simple (const char *string, const char *structure)
 - Calculate the free energy of an already folded RNA.
- float vrna_eval_circ_structure (const char *string, const char *structure)
 - Evaluate the free energy of a sequence/structure pair where the sequence is circular.
- float vrna eval gquad structure (const char *string, const char *structure)
 - Evaluate the free energy of a sequence/structure pair where the structure may contain G-Quadruplexes.
- float vrna_eval_circ_gquad_structure (const char *string, const char *structure)
 - Evaluate the free energy of a sequence/structure pair where the sequence is circular and the structure may contain G-Quadruplexes.
- float vrna_eval_structure_simple_verbose (const char *string, const char *structure, FILE *file)
 - Calculate the free energy of an already folded RNA and print contributions per loop.
- float vrna_eval_structure_simple_v (const char *string, const char *structure, int verbosity_level, FILE *file)

 Calculate the free energy of an already folded RNA and print contributions per loop.
- float vrna_eval_circ_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)
 - Evaluate free energy of a sequence/structure pair, assume sequence to be circular and print contributions per loop.
- float vrna_eval_gquad_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)

 Evaluate free energy of a sequence/structure pair, allow for G-Quadruplexes in the structure and print contributions per loop.
- float vrna_eval_circ_gquad_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)

Evaluate free energy of a sequence/structure pair, assume sequence to be circular, allow for G-Quadruplexes in the structure, and print contributions per loop.

Simplified Energy Evaluation with Sequence Alignments and Consensus Structure Dot-Bracket String

- float vrna_eval_consensus_structure_simple (const char **alignment, const char *structure)
 - Calculate the free energy of an already folded RNA sequence alignment.
- float vrna_eval_circ_consensus_structure (const char **alignment, const char *structure)
 - Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the sequences are circular.
- float vrna_eval_gquad_consensus_structure (const char **alignment, const char *structure)
 - Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the structure may contain G-Quadruplexes.
- float vrna_eval_circ_gquad_consensus_structure (const char **alignment, const char *structure)
 - Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the sequence is circular and the structure may contain G-Quadruplexes.
- float vrna_eval_consensus_structure_simple_verbose (const char **alignment, const char *structure, FILE *file)
 - Evaluate the free energy of a consensus structure for an RNA sequence alignment and print contributions per loop.
- float vrna_eval_consensus_structure_simple_v (const char **alignment, const char *structure, int verbosity_level, FILE *file)
 - Evaluate the free energy of a consensus structure for an RNA sequence alignment and print contributions per loop.
- - Evaluate the free energy of a consensus structure for an alignment of circular RNA sequences and print contributions per loop.
- float vrna_eval_gquad_consensus_structure_v (const char **alignment, const char *structure, int verbosity
 — level, FILE *file)
 - Evaluate the free energy of a consensus structure for an RNA sequence alignment, allow for annotated G-← Quadruplexes in the structure and print contributions per loop.
- float vrna_eval_circ_gquad_consensus_structure_v (const char **alignment, const char *structure, int verbosity level, FILE *file)
 - Evaluate the free energy of a consensus structure for an alignment of circular RNA sequences, allow for annotated G-Quadruplexes in the structure and print contributions per loop.

130 Module Documentation

Simplified Energy Evaluation with Sequence String and Structure Pair Table

• int vrna_eval_structure_pt_simple (const char *string, const short *pt)

Calculate the free energy of an already folded RNA.

• int vrna_eval_structure_pt_simple_verbose (const char *string, const short *pt, FILE *file)

Calculate the free energy of an already folded RNA.

• int vrna_eval_structure_pt_simple_v (const char *string, const short *pt, int verbosity_level, FILE *file)

Calculate the free energy of an already folded RNA.

Simplified Energy Evaluation with Sequence Alignment and Consensus Structure Pair Table

- int vrna_eval_consensus_structure_pt_simple (const char **alignment, const short *pt)

 Evaluate the Free Energy of a Consensus Secondary Structure given a Sequence Alignment.
- int vrna_eval_consensus_structure_pt_simple_verbose (const char **alignment, const short *pt, FILE *file)
- int vrna_eval_consensus_structure_pt_simple_v (const char **alignment, const short *pt, int verbosity
 — level, FILE *file)

16.1.2 Function Documentation

16.1.2.1 vrna_eval_structure()

Calculate the free energy of an already folded RNA.

This function allows for energy evaluation of a given pair of structure and sequence (alignment). Model details, energy parameters, and possibly soft constraints are used as provided via the parameter 'vc'. The vrna_fold_compound_t does not need to contain any DP matrices, but requires all most basic init values as one would get from a call like this:

```
vc = vrna_fold_compound(sequence, NULL, VRNA_OPTION_EVAL_ONLY);
```

Note

Accepts vrna fold compound t of type VRNA FC TYPE SINGLE and VRNA FC TYPE COMPARATIVE

See also

 $vrna_eval_structure_pt(), vrna_eval_structure_verbose(), vrna_eval_structure_pt_verbose(), vrna_fold_compound(), vrna_fold_compound_comparative(), vrna_eval_covar_structure()$

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
structure	Secondary structure in dot-bracket notation

Returns

The free energy of the input structure given the input sequence in kcal/mol

SWIG Wrapper Notes This function is attached as method eval structure() to objects of type fold compound

16.1.2.2 vrna_eval_covar_structure()

Calculate the pseudo energy derived by the covariance scores of a set of aligned sequences.

Consensus structure prediction is driven by covariance scores of base pairs in rows of the provided alignment. This function allows one to retrieve the total amount of this covariance pseudo energy scores. The vrna_fold_compound_t does not need to contain any DP matrices, but requires all most basic init values as one would get from a call like this:

```
vc = vrna_fold_compound_comparative(alignment, NULL, VRNA_OPTION_EVAL_ONLY);
```

Note

Accepts vrna_fold_compound_t of type VRNA_FC_TYPE_COMPARATIVE only!

See also

```
vrna_fold_compound_comparative(), vrna_eval_structure()
```

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
structure	Secondary (consensus) structure in dot-bracket notation

Returns

The covariance pseudo energy score of the input structure given the input sequence alignment in kcal/mol

SWIG Wrapper Notes This function is attached as method eval_covar_structure() to objects of type fold_← compound

16.1.2.3 vrna_eval_structure_verbose()

132 Module Documentation

```
const char * structure,
FILE * file )
#include <ViennaRNA/eval.h>
```

Calculate the free energy of an already folded RNA and print contributions on a per-loop base.

This function is a simplyfied version of vrna_eval_structure_v() that uses the default verbosity level.

See also

```
vrna_eval_structure_pt(), vrna_eval_structure_verbose(), vrna_eval_structure_pt_verbose(),
```

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
structure	Secondary structure in dot-bracket notation
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in kcal/mol

SWIG Wrapper Notes This function is attached as method eval_structure_verbose() to objects of type fold_← compound

16.1.2.4 vrna_eval_structure_v()

#include <ViennaRNA/eval.h>

Calculate the free energy of an already folded RNA and print contributions on a per-loop base.

This function allows for detailed energy evaluation of a given sequence/structure pair. In contrast to $vrna_eval_structure()$ this function prints detailed energy contributions based on individual loops to a file handle. If NULL is passed as file handle, this function defaults to print to stdout. Any positive $verbosity_level$ activates potential warning message of the energy evaluting functions, while values ≥ 1 allow for detailed control of what data is printed. A negative parameter $verbosity_level$ turns off printing all together.

Model details, energy parameters, and possibly soft constraints are used as provided via the parameter 'vc'. The fold_compound does not need to contain any DP matrices, but all the most basic init values as one would get from a call like this:

```
vc = vrna_fold_compound(sequence, NULL, VRNA_OPTION_EVAL_ONLY);
```

See also

vrna_eval_structure_pt(), vrna_eval_structure_verbose(), vrna_eval_structure_pt_verbose(),

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
structure	Secondary structure in dot-bracket notation
verbosity_level	The level of verbosity of this function
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in kcal/mol

16.1.2.5 vrna_eval_structure_pt()

Calculate the free energy of an already folded RNA.

This function allows for energy evaluation of a given sequence/structure pair where the structure is provided in pair_table format as obtained from vrna_ptable(). Model details, energy parameters, and possibly soft constraints are used as provided via the parameter 'vc'. The fold_compound does not need to contain any DP matrices, but all the most basic init values as one would get from a call like this:

```
vc = vrna_fold_compound(sequence, NULL, VRNA_OPTION_EVAL_ONLY);
```

See also

vrna_ptable(), vrna_eval_structure(), vrna_eval_structure_pt_verbose()

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details	
pt	Secondary structure as pair_table	

Returns

The free energy of the input structure given the input sequence in 10cal/mol

SWIG Wrapper Notes This function is attached as method eval_structure_pt() to objects of type fold_compound

134 Module Documentation

16.1.2.6 vrna_eval_structure_pt_verbose()

#include <ViennaRNA/eval.h>

Calculate the free energy of an already folded RNA.

This function is a simplyfied version of vrna_eval_structure_simple_v() that uses the default verbosity level.

See also

```
vrna eval structure pt v(), vrna ptable(), vrna eval structure pt(), vrna eval structure verbose()
```

Parameters

vc A vrna_fold_compound_t c		A vrna_fold_compound_t containing the energy parameters and model details
ĺ	pt	Secondary structure as pair_table
Ī	file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in 10cal/mol

SWIG Wrapper Notes This function is attached as method eval_structure_pt_verbose() to objects of type fold← compound

16.1.2.7 vrna_eval_structure_pt_v()

Calculate the free energy of an already folded RNA.

This function allows for energy evaluation of a given sequence/structure pair where the structure is provided in pair_table format as obtained from vrna_ptable(). Model details, energy parameters, and possibly soft constraints are used as provided via the parameter 'vc'. The fold_compound does not need to contain any DP matrices, but all the most basic init values as one would get from a call like this:

```
vc = vrna_fold_compound(sequence, NULL, VRNA_OPTION_EVAL_ONLY)
```

In contrast to $vrna_eval_structure_pt()$ this function prints detailed energy contributions based on individual loops to a file handle. If NULL is passed as file handle, this function defaults to print to stdout. Any positive $verbosity \leftarrow _level$ activates potential warning message of the energy evaluting functions, while values ≥ 1 allow for detailed control of what data is printed. A negative parameter $verbosity_level$ turns off printing all together.

See also

```
vrna_ptable(), vrna_eval_structure_pt(), vrna_eval_structure_verbose()
```

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
pt	Secondary structure as pair_table
verbosity_level	The level of verbosity of this function
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in 10cal/mol

16.1.2.8 vrna_eval_structure_simple()

Calculate the free energy of an already folded RNA.

This function allows for energy evaluation of a given sequence/structure pair. In contrast to vrna_eval_structure() this function assumes default model details and default energy parameters in order to evaluate the free energy of the secondary structure. Therefore, it serves as a simple interface function for energy evaluation for situations where no changes on the energy model are required.

See also

vrna_eval_structure(), vrna_eval_structure_pt(), vrna_eval_structure_verbose(), vrna_eval_structure_pt_verbose(),

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation

Returns

The free energy of the input structure given the input sequence in kcal/mol

SWIG Wrapper Notes In the target scripting language, this function serves as a wrapper for vrna_eval_structure_simple_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA_VERBOSITY_QUIET and NULL, respectively.

136 Module Documentation

16.1.2.9 vrna_eval_circ_structure()

Evaluate the free energy of a sequence/structure pair where the sequence is circular.

See also

vrna_eval_structure_simple(), vrna_eval_gquad_structure(), vrna_eval_circ_consensus_structure(), vrna_eval_circ_structure_vrna_eval_structure()

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation

Returns

The free energy of the structure given the circular input sequence in kcal/mol

SWIG Wrapper Notes In the target scripting language, this function serves as a wrapper for vrna_eval_circ_structure_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.10 vrna_eval_gquad_structure()

Evaluate the free energy of a sequence/structure pair where the structure may contain G-Quadruplexes.

G-Quadruplexes are annotated as plus signs ('+') for each G involved in the motif. Linker sequences must be denoted by dots ('.') as they are considered unpaired. Below is an example of a 2-layer G-quadruplex:

GGAAGGAAAGGAGG

++..++..++

See also

vrna_eval_structure(), vrna_eval_gquad_consensus_structure(), vrna_eval_gquad_structure() vrna_eval_structure()

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation

Returns

The free energy of the structure including contributions of G-quadruplexes in kcal/mol

SWIG Wrapper Notes In the target scripting language, this function serves as a wrapper for vrna_eval_gquad_structure_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.11 vrna_eval_circ_gquad_structure()

Evaluate the free energy of a sequence/structure pair where the sequence is circular and the structure may contain G-Quadruplexes.

G-Quadruplexes are annotated as plus signs ('+') for each G involved in the motif. Linker sequences must be denoted by dots ('.') as they are considered unpaired. Below is an example of a 2-layer G-quadruplex: GGAAGGAAAGGAGG

```
тт...тт...т
```

See also

vrna_eval_structure_simple(), vrna_eval_circ_gquad_consensus_structure(), vrna_eval_circ_gquad_structure_v(), vrna_eval_structure()

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation

Returns

The free energy of the structure including contributions of G-quadruplexes in kcal/mol

SWIG Wrapper Notes In the target scripting language, this function serves as a wrapper for vrna_eval_circ_gquad_structure_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.12 vrna_eval_structure_simple_verbose()

Calculate the free energy of an already folded RNA and print contributions per loop.

This function is a simplyfied version of vrna_eval_structure_simple_v() that uses the default verbosity level.

See also

```
vrna_eval_structure_simple_v(), vrna_eval_structure_verbose(), vrna_eval_structure_pt(), vrna_eval_structure_verbose(), vrna_eval_structure_pt verbose()
```

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in kcal/mol

SWIG Wrapper Notes This function is not available. Use vrna_eval_structure_simple_v() instead!

16.1.2.13 vrna_eval_structure_simple_v()

#include <ViennaRNA/eval.h>

Calculate the free energy of an already folded RNA and print contributions per loop.

This function allows for detailed energy evaluation of a given sequence/structure pair. In contrast to $vrna_eval_structure()$ this function prints detailed energy contributions based on individual loops to a file handle. If NULL is passed as file handle, this function defaults to print to stdout. Any positive $verbosity_level$ activates potential warning message of the energy evaluting functions, while values ≥ 1 allow for detailed control of what data is printed. A negative parameter $verbosity_level$ turns off printing all together.

In contrast to vrna_eval_structure_verbose() this function assumes default model details and default energy parameters in order to evaluate the free energy of the secondary structure. Threefore, it serves as a simple interface function for energy evaluation for situations where no changes on the energy model are required.

See also

vrna_eval_structure_verbose(), vrna_eval_structure_pt(), vrna_eval_structure_pt_verbose(),

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation
verbosity_level	The level of verbosity of this function
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in kcal/mol

SWIG Wrapper Notes This function is available through an overloaded version of vrna_eval_structure_simple().

The last two arguments for this function are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.14 vrna_eval_circ_structure_v()

#include <ViennaRNA/eval.h>

Evaluate free energy of a sequence/structure pair, assume sequence to be circular and print contributions per loop.

This function is the same as vrna_eval_structure_simple_v() but assumes the input sequence to be circularized.

See also

vrna_eval_structure_simple_v(), vrna_eval_circ_structure(), vrna_eval_structure_verbose()

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation
verbosity_level	The level of verbosity of this function
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in kcal/mol

SWIG Wrapper Notes This function is available through an overloaded version of vrna_eval_circ_structure(). The last two arguments for this function are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.15 vrna_eval_gquad_structure_v()

#include <ViennaRNA/eval.h>

Evaluate free energy of a sequence/structure pair, allow for G-Quadruplexes in the structure and print contributions per loop.

This function is the same as vrna_eval_structure_simple_v() but allows for annotated G-Quadruplexes in the dot-bracket structure input.

G-Quadruplexes are annotated as plus signs ('+') for each G involved in the motif. Linker sequences must be denoted by dots ('.') as they are considered unpaired. Below is an example of a 2-layer G-quadruplex:

```
++..++...++.++
```

See also

```
vrna_eval_structure_simple_v(), vrna_eval_gquad_structure(), vrna_eval_structure_verbose()
```

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation
verbosity_level	The level of verbosity of this function
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in kcal/mol

SWIG Wrapper Notes This function is available through an overloaded version of vrna_eval_gquad_structure().

The last two arguments for this function are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.16 vrna_eval_circ_gquad_structure_v()

#include <ViennaRNA/eval.h>

Evaluate free energy of a sequence/structure pair, assume sequence to be circular, allow for G-Quadruplexes in the structure, and print contributions per loop.

This function is the same as vrna_eval_structure_simple_v() but assumes the input sequence to be circular and allows for annotated G-Quadruplexes in the dot-bracket structure input.

G-Quadruplexes are annotated as plus signs ('+') for each G involved in the motif. Linker sequences must be denoted by dots ('.') as they are considered unpaired. Below is an example of a 2-layer G-quadruplex:

++..++...++.++

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation
verbosity_level	The level of verbosity of this function
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in kcal/mol

SWIG Wrapper Notes This function is available through an overloaded version of vrna_eval_circ_gquad_structure(). The last two arguments for this function are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.17 vrna_eval_consensus_structure_simple()

Calculate the free energy of an already folded RNA sequence alignment.

This function allows for energy evaluation for a given multiple sequence alignment and consensus structure pair. In contrast to vrna_eval_structure() this function assumes default model details and default energy parameters in order to evaluate the free energy of the secondary structure. Therefore, it serves as a simple interface function for energy evaluation for situations where no changes on the energy model are required.

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

vrna_eval_covar_structure(), vrna_eval_structure(), vrna_eval_structure_pt(), vrna_eval_structure_verbose(), vrna_eval_structure_pt_verbose()

Parameters

alignment	RNA sequence alignment in uppercase letters and hyphen ('-') to denote gaps
structure	Consensus Secondary structure in dot-bracket notation

Returns

The free energy of the consensus structure given the input alignment in kcal/mol

SWIG Wrapper Notes This function is available through an overloadeded version of vrna eval structure simple(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

16.1.2.18 vrna eval circ consensus structure()

```
int vrna_eval_circ_consensus_structure (
            const char ** alignment,
            const char * structure )
#include <ViennaRNA/eval.h>
```

Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the sequences are circular.

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

vrna_eval_covar_structure(), vrna_eval_consensus_structure_simple(), vrna_eval_gquad_consensus_structure(), vrna eval circ structure(), vrna eval circ consensus structure v(), vrna eval structure()

Parameters

alignment	RNA sequence alignment in uppercase letters
structure	Consensus secondary structure in dot-bracket notation

Returns

The free energy of the consensus structure given the circular input sequence in kcal/mol

SWIG Wrapper Notes This function is available through an overloadeded version of vrna_eval_circ_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

16.1.2.19 vrna_eval_gquad_consensus_structure()

Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the structure may contain G-Quadruplexes.

G-Quadruplexes are annotated as plus signs ('+') for each G involved in the motif. Linker sequences must be denoted by dots ('.') as they are considered unpaired. Below is an example of a 2-layer G-quadruplex:

```
++..++...++.++
```

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

vrna_eval_covar_structure(), vrna_eval_consensus_structure_simple(), vrna_eval_circ_consensus_structure(), vrna_eval_gquad_structure(), vrna_eval_gquad_consensus_structure_v(), vrna_eval_structure()

Parameters

alignment	RNA sequence alignment in uppercase letters
structure	Consensus secondary structure in dot-bracket notation

Returns

The free energy of the consensus structure including contributions of G-quadruplexes in kcal/mol

SWIG Wrapper Notes This function is available through an overloadeded version of vrna_eval_gquad_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

16.1.2.20 vrna_eval_circ_gquad_consensus_structure()

Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the sequence is circular and the structure may contain G-Quadruplexes.

```
++..++...++.++
```

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

vrna_eval_covar_structure(), vrna_eval_consensus_structure_simple(), vrna_eval_circ_consensus_structure(), vrna eval gquad structure(), vrna eval circ gquad consensus structure v(), vrna eval structure()

Parameters

alignment	RNA sequence alignment in uppercase letters
structure	Consensus secondary structure in dot-bracket notation

Returns

The free energy of the consensus structure including contributions of G-quadruplexes in kcal/mol

SWIG Wrapper Notes This function is available through an overloadeded version of vrna_eval_circ_gquad_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

16.1.2.21 vrna_eval_consensus_structure_simple_verbose()

#include <ViennaRNA/eval.h>

Evaluate the free energy of a consensus structure for an RNA sequence alignment and print contributions per loop.

This function is a simplyfied version of vrna_eval_consensus_structure_simple_v() that uses the *default* verbosity level.

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

```
vrna\_eval\_consensus\_structure\_simple\_v(), \qquad vrna\_eval\_structure\_verbose(), \qquad vrna\_eval\_structure\_pt(), \\ vrna\_eval\_structure\_pt\_verbose()
```

Parameters

alignment	RNA sequence alignment in uppercase letters. Gaps are denoted by hyphens ('-')
structure	Consensus secondary structure in dot-bracket notation
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the conensus structure given the aligned input sequences in kcal/mol

SWIG Wrapper Notes This function is not available. Use vrna eval consensus structure simple v() instead!

16.1.2.22 vrna_eval_consensus_structure_simple_v()

#include <ViennaRNA/eval.h>

Evaluate the free energy of a consensus structure for an RNA sequence alignment and print contributions per loop.

This function allows for detailed energy evaluation of a given sequence alignment/consensus structure pair. In contrast to $vrna_eval_consensus_structure_simple()$ this function prints detailed energy contributions based on individual loops to a file handle. If NULL is passed as file handle, this function defaults to print to stdout. Any positive $verbosity_level$ activates potential warning message of the energy evaluting functions, while values ≥ 1 allow for detailed control of what data is printed. A negative parameter $verbosity_level$ turns off printing all together.

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

```
vrna_eval_consensus_structure(), vrna_eval_structure()
```

Parameters

alignment	RNA sequence alignment in uppercase letters. Gaps are denoted by hyphens ('-')
structure	Consensus secondary structure in dot-bracket notation
verbosity_level	The level of verbosity of this function
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the consensus structure given the sequence alignment in kcal/mol

SWIG Wrapper Notes This function is available through an overloaded version of vrna eval structure simple(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument. The last two arguments are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.23 vrna_eval_circ_consensus_structure_v()

```
int vrna_eval_circ_consensus_structure_v (
             const char ** alignment,
             const char * structure,
             int verbosity_level,
             FILE * file )
```

#include <ViennaRNA/eval.h>

Evaluate the free energy of a consensus structure for an alignment of circular RNA sequences and print contributions per loop.

This function is identical with vrna eval consensus structure simple v() but assumed the aligned sequences to be circular.

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

vrna eval consensus structure simple v(), vrna eval circ consensus structure(), vrna eval structure()

Parameters

alignment	RNA sequence alignment in uppercase letters. Gaps are denoted by hyphens ('-')	
structure Consensus secondary structure in dot-bracket notation		
verbosity_level The level of verbosity of this function		
file	A file handle where this function should print to (may be NULL).	

Returns

The free energy of the consensus structure given the sequence alignment in kcal/mol

SWIG Wrapper Notes This function is available through an overloaded version of vrna eval circ structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument. The last two arguments are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.24 vrna_eval_gquad_consensus_structure_v()

Evaluate the free energy of a consensus structure for an RNA sequence alignment, allow for annotated G- \leftarrow Quadruplexes in the structure and print contributions per loop.

This function is identical with vrna_eval_consensus_structure_simple_v() but allows for annotated G-Quadruplexes in the consensus structure.

G-Quadruplexes are annotated as plus signs ('+') for each G involved in the motif. Linker sequences must be denoted by dots ('.') as they are considered unpaired. Below is an example of a 2-layer G-quadruplex:

```
++..++...++.++
```

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

vrna_eval_consensus_structure_simple_v(), vrna_eval_gquad_consensus_structure(), vrna_eval_structure()

Parameters

alignment RNA sequence alignment in uppercase letters. Gaps are denoted by hyphens		
structure	Consensus secondary structure in dot-bracket notation	
verbosity_level The level of verbosity of this function		
file	A file handle where this function should print to (may be NULL).	

Returns

The free energy of the consensus structure given the sequence alignment in kcal/mol

SWIG Wrapper Notes This function is available through an overloaded version of vrna_eval_gquad_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument. The last two arguments are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.25 vrna_eval_circ_gquad_consensus_structure_v()

```
const char * structure,
int verbosity_level,
FILE * file )
```

```
#include <ViennaRNA/eval.h>
```

Evaluate the free energy of a consensus structure for an alignment of circular RNA sequences, allow for annotated G-Quadruplexes in the structure and print contributions per loop.

This function is identical with vrna_eval_consensus_structure_simple_v() but assumes the sequences in the alignment to be circular and allows for annotated G-Quadruplexes in the consensus structure.

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

vrna_eval_consensus_structure_simple_v(), vrna_eval_circ_gquad_consensus_structure(), vrna_eval_structure()

Parameters

alignment	RNA sequence alignment in uppercase letters. Gaps are denoted by hyphens ('-')	
structure	structure Consensus secondary structure in dot-bracket notation	
verbosity_level The level of verbosity of this function		
file A file handle where this function should print to (may be NULL).		

Returns

The free energy of the consensus structure given the sequence alignment in kcal/mol

SWIG Wrapper Notes This function is available through an overloaded version of vrna_eval_circ_gquad_structure(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument. The last two arguments are optional and default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.26 vrna_eval_structure_pt_simple()

Calculate the free energy of an already folded RNA.

In contrast to vrna_eval_structure_pt() this function assumes default model details and default energy parameters in order to evaluate the free energy of the secondary structure. Threefore, it serves as a simple interface function for energy evaluation for situations where no changes on the energy model are required.

See also

vrna_ptable(), vrna_eval_structure_simple(), vrna_eval_structure_pt()

Parameters

string	RNA sequence in uppercase letters	
pt	Secondary structure as pair_table	

Returns

The free energy of the input structure given the input sequence in 10cal/mol

SWIG Wrapper Notes In the target scripting language, this function serves as a wrapper for vrna_eval_structure_pt_v() and, thus, allows for two additional, optional arguments, the verbosity level and a file handle which default to VRNA_VERBOSITY_QUIET and NULL, respectively.

16.1.2.27 vrna_eval_structure_pt_simple_verbose()

Calculate the free energy of an already folded RNA.

This function is a simplyfied version of vrna_eval_structure_pt_simple_v() that uses the default verbosity level.

See also

vrna_eval_structure_pt_simple_v(), vrna_ptable(), vrna_eval_structure_pt_verbose(), vrna_eval_structure_simple()

Parameters

string	RNA sequence in uppercase letters
pt	Secondary structure as pair_table
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in 10cal/mol

16.1.2.28 vrna_eval_structure_pt_simple_v()

#include <ViennaRNA/eval.h>

Calculate the free energy of an already folded RNA.

This function allows for energy evaluation of a given sequence/structure pair where the structure is provided in pair_table format as obtained from vrna_ptable(). Model details, energy parameters, and possibly soft constraints are used as provided via the parameter 'vc'. The fold_compound does not need to contain any DP matrices, but all the most basic init values as one would get from a call like this:

```
vc = vrna_fold_compound(sequence, NULL, VRNA_OPTION_EVAL_ONLY);
```

In contrast to vrna_eval_structure_pt_verbase() this function assumes default model details and default energy parameters in order to evaluate the free energy of the secondary structure. Threefore, it serves as a simple interface function for energy evaluation for situations where no changes on the energy model are required.

See also

```
vrna_ptable(), vrna_eval_structure_pt_v(), vrna_eval_structure_simple()
```

Parameters

string	RNA sequence in uppercase letters
pt	Secondary structure as pair_table
verbosity_level	The level of verbosity of this function
file	A file handle where this function should print to (may be NULL).

Returns

The free energy of the input structure given the input sequence in 10cal/mol

16.1.2.29 vrna_eval_consensus_structure_pt_simple()

Evaluate the Free Energy of a Consensus Secondary Structure given a Sequence Alignment.

Note

The free energy returned from this function already includes the covariation pseudo energies that is used fir comparative structure prediction within this library.

See also

vrna_eval_consensus_structure_simple(), vrna_eval_structure_pt(), vrna_eval_structure(), vrna_eval_covar_structure()

Parameters

alignment	RNA sequence alignment in uppercase letters. Gaps are denoted by hyphens ('-')
pt	Secondary structure in pair table format

Returns

Free energy of the consensus structure in 10cal/mol

SWIG Wrapper Notes This function is available through an overloadeded version of vrna_eval_structure_pt_simple(). Simply pass a sequence alignment as list of strings (including gaps) as first, and the consensus structure as second argument

16.2 Energy Evaluation for Individual Loops

Functions to evaluate the free energy of particular types of loops.

16.2.1 Detailed Description

Functions to evaluate the free energy of particular types of loops.

To assess the free energy contribution of a particular loop within a secondary structure, two variants are provided:

- The bare free energy E (usually in deka-calories, i.e. multiples of 10cal/mol), and
- The Boltzmann weight $q=exp(-\beta E)$ of the free energy E (with $\beta=\frac{1}{RT}$, gas constant R and temperature T)

The latter is usually required for partition function computations. Collaboration diagram for Energy Evaluation for Individual Loops:

Modules

· Exterior Loops

Functions to evaluate the free energy contributions for exterior loops.

Hairpin Loops

Functions to evaluate the free energy contributions for hairpin loops.

· Internal Loops

Functions to evaluate the free energy contributions for internal loops.

Multibranch Loops

Functions to evaluate the free energy contributions for mullibranch loops.

Files

• file all.h

Energy evaluation for MFE and partition function calculations.

· file external.h

Energy evaluation of exterior loops for MFE and partition function calculations.

· file hairpin.h

Energy evaluation of hairpin loops for MFE and partition function calculations.

· file internal.h

Energy evaluation of interior loops for MFE and partition function calculations.

file multibranch.h

Energy evaluation of multibranch loops for MFE and partition function calculations.

Functions

• int vrna_eval_loop_pt (vrna_fold_compound_t *vc, int i, const short *pt)

Calculate energy of a loop.

• int vrna_eval_loop_pt_v (vrna_fold_compound_t *vc, int i, const short *pt, int verbosity_level)

Calculate energy of a loop.

16.2.2 Function Documentation

16.2.2.1 vrna_eval_loop_pt()

Calculate energy of a loop.

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
i	position of covering base pair
pt	the pair table of the secondary structure

Returns

free energy of the loop in 10cal/mol

SWIG Wrapper Notes This function is attached as method eval_loop_pt() to objects of type fold_compound

16.2.2.2 vrna_eval_loop_pt_v()

Calculate energy of a loop.

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
i	position of covering base pair
pt	the pair table of the secondary structure
verbosity_level	The level of verbosity of this function

Returns

free energy of the loop in 10cal/mol

16.3 Energy Evaluation for Atomic Moves

Functions to evaluate the free energy change of a structure after application of (a set of) atomic moves.

16.3.1 Detailed Description

Functions to evaluate the free energy change of a structure after application of (a set of) atomic moves.

Here, atomic moves are not to be confused with moves of actual physical atoms. Instead, an atomic move is considered the smallest conformational change a secondary structure can undergo to form another, distinguishable structure. We currently support the following moves

Atomic Moves:

- · Opening (dissociation) of a single base pair
- · Closing (formation) of a single base pair
- Shifting one pairing partner of an existing pair to a different location

Collaboration diagram for Energy Evaluation for Atomic Moves:

Functions

```
• float vrna_eval_move (vrna_fold_compound_t *vc, const char *structure, int m1, int m2)

Calculate energy of a move (closing or opening of a base pair)
```

• int vrna_eval_move_pt (vrna_fold_compound_t *vc, short *pt, int m1, int m2)

Calculate energy of a move (closing or opening of a base pair)

16.3.2 Function Documentation

16.3.2.1 vrna_eval_move()

Calculate energy of a move (closing or opening of a base pair)

If the parameters m1 and m2 are negative, it is deletion (opening) of a base pair, otherwise it is insertion (opening).

See also

```
vrna_eval_move_pt()
```

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
structure	secondary structure in dot-bracket notation
m1	first coordinate of base pair
m2	second coordinate of base pair

Returns

energy change of the move in kcal/mol (INF / 100. upon any error)

SWIG Wrapper Notes This function is attached as method eval_move() to objects of type fold_compound

16.3.2.2 vrna_eval_move_pt()

#include <ViennaRNA/eval.h>

Calculate energy of a move (closing or opening of a base pair)

If the parameters m1 and m2 are negative, it is deletion (opening) of a base pair, otherwise it is insertion (opening).

See also

```
vrna_eval_move()
```

Parameters

VC	A vrna_fold_compound_t containing the energy parameters and model details
pt	the pair table of the secondary structure
m1	first coordinate of base pair
m2	second coordinate of base pair

Returns

energy change of the move in 10cal/mol

SWIG Wrapper Notes This function is attached as method eval_move_pt() to objects of type fold_compound

16.4 Deprecated Interface for Free Energy Evaluation

Deprecated Energy Evaluation functions.

16.4.1 Detailed Description

Deprecated Energy Evaluation functions.

Using the functions below is discouraged as they have been marked deprecated and will be removed from the library in the (near) future! Collaboration diagram for Deprecated Interface for Free Energy Evaluation:

Functions

float energy_of_structure (const char *string, const char *structure, int verbosity_level)

Calculate the free energy of an already folded RNA using global model detail settings.

• float energy_of_struct_par (const char *string, const char *structure, vrna_param_t *parameters, int verbosity_level)

Calculate the free energy of an already folded RNA.

• float energy_of_circ_structure (const char *string, const char *structure, int verbosity_level)

Calculate the free energy of an already folded circular RNA.

• float energy_of_circ_struct_par (const char *string, const char *structure, vrna_param_t *parameters, int verbosity_level)

Calculate the free energy of an already folded circular RNA.

• int energy_of_structure_pt (const char *string, short *ptable, short *s, short *s1, int verbosity_level)

Calculate the free energy of an already folded RNA.

• int energy_of_struct_pt_par (const char *string, short *ptable, short *s, short *s1, vrna_param_t *parameters, int verbosity_level)

Calculate the free energy of an already folded RNA.

float energy_of_move (const char *string, const char *structure, int m1, int m2)

Calculate energy of a move (closing or opening of a base pair)

int energy_of_move_pt (short *pt, short *s, short *s1, int m1, int m2)

Calculate energy of a move (closing or opening of a base pair)

int loop_energy (short *ptable, short *s, short *s1, int i)

Calculate energy of a loop.

- float energy_of_struct (const char *string, const char *structure)
- int energy of struct pt (const char *string, short *ptable, short *s, short *s1)
- float energy_of_circ_struct (const char *string, const char *structure)
- int E_Stem (int type, int si1, int sj1, int extLoop, vrna_param_t *P)

Compute the energy contribution of a stem branching off a loop-region.

- FLT_OR_DBL exp_E_ExtLoop (int type, int si1, int sj1, vrna_exp_param_t *P)
- FLT_OR_DBL exp_E_Stem (int type, int si1, int sj1, int extLoop, vrna_exp_param_t *P)
- PRIVATE int E IntLoop (int n1, int n2, int type, int type 2, int si1, int sj1, int sp1, int sq1, vrna param t*P)
- PRIVATE FLT_OR_DBL exp_E_IntLoop (int u1, int u2, int type, int type2, short si1, short sj1, short sp1, short sq1, vrna_exp_param_t *P)

Variables

· int cut_point

first pos of second seg for cofolding

int eos_debug

verbose info from energy_of_struct

16.4.2 Function Documentation

16.4.2.1 energy_of_structure()

Calculate the free energy of an already folded RNA using global model detail settings.

If verbosity level is set to a value >0, energies of structure elements are printed to stdout

Note

OpenMP: This function relies on several global model settings variables and thus is not to be considered threadsafe. See energy_of_struct_par() for a completely threadsafe implementation.

Deprecated Use vrna_eval_structure() or vrna_eval_structure_verbose() instead!

See also

```
vrna_eval_structure()
```

Parameters

string	RNA sequence
structure	secondary structure in dot-bracket notation
verbosity_level	a flag to turn verbose output on/off

Returns

the free energy of the input structure given the input sequence in kcal/mol

16.4.2.2 energy_of_struct_par()

#include <ViennaRNA/eval.h>

Calculate the free energy of an already folded RNA.

If verbosity level is set to a value >0, energies of structure elements are printed to stdout

Deprecated Use vrna eval structure() or vrna eval structure verbose() instead!

See also

```
vrna_eval_structure()
```

Parameters

string	RNA sequence in uppercase letters
structure	Secondary structure in dot-bracket notation
parameters	A data structure containing the prescaled energy contributions and the model details.
verbosity_level	A flag to turn verbose output on/off

Returns

The free energy of the input structure given the input sequence in kcal/mol

16.4.2.3 energy_of_circ_structure()

Calculate the free energy of an already folded circular RNA.

Note

OpenMP: This function relies on several global model settings variables and thus is not to be considered threadsafe. See energy_of_circ_struct_par() for a completely threadsafe implementation.

If verbosity level is set to a value >0, energies of structure elements are printed to stdout

Deprecated Use vrna eval structure() or vrna eval structure verbose() instead!

See also

```
vrna_eval_structure()
```

Parameters

string	RNA sequence
structure	Secondary structure in dot-bracket notation
verbosity_level	A flag to turn verbose output on/off

Returns

The free energy of the input structure given the input sequence in kcal/mol

16.4.2.4 energy_of_circ_struct_par()

Calculate the free energy of an already folded circular RNA.

If verbosity level is set to a value >0, energies of structure elements are printed to stdout

Deprecated Use vrna_eval_structure() or vrna_eval_structure_verbose() instead!

See also

```
vrna_eval_structure()
```

Parameters

string	RNA sequence
structure	Secondary structure in dot-bracket notation
parameters	A data structure containing the prescaled energy contributions and the model details.
verbosity_level	A flag to turn verbose output on/off

Returns

The free energy of the input structure given the input sequence in kcal/mol

16.4.2.5 energy_of_structure_pt()

```
short * ptable,
short * s,
short * s1,
int verbosity_level )
#include <ViennaRNA/eval.h>
```

Calculate the free energy of an already folded RNA.

If verbosity level is set to a value >0, energies of structure elements are printed to stdout

Note

OpenMP: This function relies on several global model settings variables and thus is not to be considered threadsafe. See energy_of_struct_pt_par() for a completely threadsafe implementation.

Deprecated Use vrna_eval_structure_pt() or vrna_eval_structure_pt_verbose() instead!

See also

```
vrna_eval_structure_pt()
```

Parameters

string	RNA sequence
ptable	the pair table of the secondary structure
s	encoded RNA sequence
s1	encoded RNA sequence
verbosity_level	a flag to turn verbose output on/off

Returns

the free energy of the input structure given the input sequence in 10kcal/mol

16.4.2.6 energy_of_struct_pt_par()

Calculate the free energy of an already folded RNA.

If verbosity level is set to a value >0, energies of structure elements are printed to stdout

Deprecated Use vrna_eval_structure_pt() or vrna_eval_structure_pt_verbose() instead!

See also

```
vrna_eval_structure_pt()
```

Parameters

string	RNA sequence in uppercase letters
ptable	The pair table of the secondary structure
s	Encoded RNA sequence
s1	Encoded RNA sequence
parameters	A data structure containing the prescaled energy contributions and the model details.
verbosity_level	A flag to turn verbose output on/off

Returns

The free energy of the input structure given the input sequence in 10kcal/mol

16.4.2.7 energy_of_move()

Calculate energy of a move (closing or opening of a base pair)

If the parameters m1 and m2 are negative, it is deletion (opening) of a base pair, otherwise it is insertion (opening).

Deprecated Use vrna_eval_move() instead!

See also

```
vrna_eval_move()
```

Parameters

string	RNA sequence
structure	secondary structure in dot-bracket notation
m1	first coordinate of base pair
m2	second coordinate of base pair

Returns

energy change of the move in kcal/mol

16.4.2.8 energy_of_move_pt()

#include <ViennaRNA/eval.h>

Calculate energy of a move (closing or opening of a base pair)

If the parameters m1 and m2 are negative, it is deletion (opening) of a base pair, otherwise it is insertion (opening).

Deprecated Use vrna_eval_move_pt() instead!

See also

```
vrna_eval_move_pt()
```

Parameters

pt	the pair table of the secondary structure
s	encoded RNA sequence
s1	encoded RNA sequence
m1	first coordinate of base pair
m2	second coordinate of base pair

Returns

energy change of the move in 10cal/mol

16.4.2.9 loop_energy()

Calculate energy of a loop.

Deprecated Use vrna_eval_loop_pt() instead!

See also

```
vrna_eval_loop_pt()
```

Parameters

ptable	the pair table of the secondary structure
s	encoded RNA sequence
s1	encoded RNA sequence
i	position of covering base pair

Returns

free energy of the loop in 10cal/mol

16.4.2.10 energy_of_struct()

#include <ViennaRNA/eval.h>

Calculate the free energy of an already folded RNA

Note

This function is not entirely threadsafe! Depending on the state of the global variable eos_debug it prints energy information to stdout or not...

Deprecated This function is deprecated and should not be used in future programs! Use energy_of_structure() instead!

See also

```
energy_of_structure, energy_of_circ_struct(), energy_of_struct_pt()
```

Parameters

string	RNA sequence
structure	secondary structure in dot-bracket notation

Returns

the free energy of the input structure given the input sequence in kcal/mol

16.4.2.11 energy_of_struct_pt()

#include <ViennaRNA/eval.h>

Calculate the free energy of an already folded RNA

Note

This function is not entirely threadsafe! Depending on the state of the global variable eos_debug it prints energy information to stdout or not...

Deprecated This function is deprecated and should not be used in future programs! Use energy_of_structure_pt() instead!

See also

```
make_pair_table(), energy_of_structure()
```

Parameters

string	RNA sequence
ptable	the pair table of the secondary structure
s	encoded RNA sequence
s1	encoded RNA sequence

Returns

the free energy of the input structure given the input sequence in 10kcal/mol

16.4.2.12 energy_of_circ_struct()

Calculate the free energy of an already folded circular RNA

Note

This function is not entirely threadsafe! Depending on the state of the global variable eos_debug it prints energy information to stdout or not...

Deprecated This function is deprecated and should not be used in future programs Use energy_of_circ_structure() instead!

See also

```
energy_of_circ_structure(), energy_of_struct(), energy_of_struct_pt()
```

Parameters

string	RNA sequence
structure	secondary structure in dot-bracket notation

Returns

the free energy of the input structure given the input sequence in kcal/mol

16.4.2.13 E_Stem()

#include <ViennaRNA/loops/external.h>

Compute the energy contribution of a stem branching off a loop-region.

This function computes the energy contribution of a stem that branches off a loop region. This can be the case in multiloops, when a stem branching off increases the degree of the loop but also *immediately interior base pairs* of an exterior loop contribute free energy. To switch the behavior of the function according to the evaluation of a multiloop-or exterior-loop-stem, you pass the flag 'extLoop'. The returned energy contribution consists of a TerminalAU penalty if the pair type is greater than 2, dangling end contributions of mismatching nucleotides adjacent to the stem if only one of the si1, sj1 parameters is greater than 0 and mismatch energies if both mismatching nucleotides are positive values. Thus, to avoid incorporating dangling end or mismatch energies just pass a negative number, e.g. -1 to the mismatch argument.

This is an illustration of how the energy contribution is assembled:

```
3' 5'
| | |
X - Y
5'-si1 sj1-3'
```

Here, (X,Y) is the base pair that closes the stem that branches off a loop region. The nucleotides si1 and sj1 are the 5'- and 3'- mismatches, respectively. If the base pair type of (X,Y) is greater than 2 (i.e. an A-U or G-U pair, the TerminalAU penalty will be included in the energy contribution returned. If si1 and sj1 are both nonnegative numbers, mismatch energies will also be included. If one of si1 or sj1 is a negative value, only 5' or 3' dangling end contributions are taken into account. To prohibit any of these mismatch contributions to be incorporated, just pass a negative number to both, si1 and sj1. In case the argument extLoop is 0, the returned energy contribution also includes the *internal-loop-penalty* of a multiloop stem with closing pair type.

See also

```
E_MLstem()
E_ExtLoop()
```

Note

This function is threadsafe

Deprecated Please use one of the functions vrna_E_ext_stem() and E_MLstem() instead! Use the former for cases where extLoop != 0 and the latter otherwise.

Parameters

type	The pair type of the first base pair un the stem
si1	The 5'-mismatching nucleotide
sj1	The 3'-mismatching nucleotide
extLoop	A flag that indicates whether the contribution reflects the one of an exterior loop or not
Р	The data structure containing scaled energy parameters

Returns

The Free energy of the branch off the loop in dcal/mol

16.4.2.14 exp_E_ExtLoop()

This is the partition function variant of E ExtLoop()

Deprecated Use vrna_exp_E_ext_stem() instead!

See also

E_ExtLoop()

Returns

The Boltzmann weighted energy contribution of the introduced exterior-loop stem

```
16.4.2.15 exp_E_Stem()
```

Compute the Boltzmann weighted energy contribution of a stem branching off a loop-region

This is the partition function variant of E_Stem()

See also

E_Stem()

Note

This function is threadsafe

Returns

The Boltzmann weighted energy contribution of the branch off the loop

16.4.2.16 E_IntLoop()

#include <ViennaRNA/loops/internal.h>

Compute the Energy of an interior-loop

This function computes the free energy ΔG of an interior-loop with the following structure:

```
3' 5'
U - V

a_n b_1

. . .
a_1 b_m

X - Y
| |
5' 3'
```

This general structure depicts an interior-loop that is closed by the base pair (X,Y). The enclosed base pair is (V,U) which leaves the unpaired bases a_1-a_n and b_1-b_n that constitute the loop. In this example, the length of the interior-loop is (n+m) where n or m may be 0 resulting in a bulge-loop or base pair stack. The mismatching nucleotides for the closing pair (X,Y) are:

5'-mismatch: a_1 3'-mismatch: b_m

and for the enclosed base pair (V,U):

5'-mismatch: b_1 3'-mismatch: a_n

Note

Base pairs are always denoted in 5'->3' direction. Thus the enclosed base pair must be 'turned arround' when evaluating the free energy of the interior-loop

See also

```
scale_parameters()
vrna_param_t
```

Note

This function is threadsafe

Parameters

n1	The size of the 'left'-loop (number of unpaired nucleotides)		
n2	The size of the 'right'-loop (number of unpaired nucleotides)		
type	The pair type of the base pair closing the interior loop		
type⊷	The pair type of the enclosed base pair		
_2			
si1	The 5'-mismatching nucleotide of the closing pair		
sj1	The 3'-mismatching nucleotide of the closing pair		
sp1	The 3'-mismatching nucleotide of the enclosed pair		
sq1	The 5'-mismatching nucleotide of the enclosed pair		
Р	The datastructure containing scaled energy parameters		

Returns

The Free energy of the Interior-loop in dcal/mol

16.4.2.17 exp_E_IntLoop()

```
PRIVATE FLT_OR_DBL exp_E_IntLoop (
    int u1,
    int u2,
    int type,
    int type2,
    short si1,
    short sj1,
    short sp1,
    short sq1,
    vrna_exp_param_t * P )
```

#include <ViennaRNA/loops/internal.h>

Compute Boltzmann weight $e^{-\Delta G/kT}$ of interior loop

multiply by scale[u1+u2+2] for scaling

See also

```
get_scaled_pf_parameters()
vrna_exp_param_t
E_IntLoop()
```

Note

This function is threadsafe

Parameters

u1	The size of the 'left'-loop (number of unpaired nucleotides)
u2	The size of the 'right'-loop (number of unpaired nucleotides)
type	The pair type of the base pair closing the interior loop
type2	The pair type of the enclosed base pair
si1	The 5'-mismatching nucleotide of the closing pair
sj1	The 3'-mismatching nucleotide of the closing pair
sp1	The 3'-mismatching nucleotide of the enclosed pair
sq1	The 5'-mismatching nucleotide of the enclosed pair
Р	The datastructure containing scaled Boltzmann weights of the energy parameters

	- 4.		
н	ет	ur	ns

The Boltzmann weight of the Interior-loop

16.5 The RNA Folding Grammar

The RNA folding grammar as implemented in RNAlib.

16.5.1 Detailed Description

The RNA folding grammar as implemented in RNAlib.

Collaboration diagram for The RNA Folding Grammar:

Modules

· Fine-tuning of the Implemented Models

Functions and data structures to fine-tune the implemented secondary structure evaluation model.

· Energy Parameters

All relevant functions to retrieve and copy pre-calculated energy parameter sets as well as reading/writing the energy parameter set from/to file(s).

· Extending the Folding Grammar with Additional Domains

This module covers simple and straight-forward extensions to the RNA folding grammar.

· Constraining the RNA Folding Grammar

This module provides general functions that allow for an easy control of constrained secondary structure prediction and evaluation.

Files

· file grammar.h

Implementations for the RNA folding grammar.

Data Structures

struct vrna_gr_aux_s

16.5.2 Data Structure Documentation

16.5.2.1 struct vrna_gr_aux_s

Data Fields

vrna_callback_gr_cond * cb_proc

A callback for pre- and post-processing of auxiliary grammar rules.

16.6 Fine-tuning of the Implemented Models

Functions and data structures to fine-tune the implemented secondary structure evaluation model.

16.6.1 Detailed Description

Functions and data structures to fine-tune the implemented secondary structure evaluation model.

Collaboration diagram for Fine-tuning of the Implemented Models:

Files

· file model.h

The model details data structure and its corresponding modifiers.

Data Structures

struct vrna_md_s

The data structure that contains the complete model details used throughout the calculations. More...

Macros

• #define VRNA MODEL DEFAULT TEMPERATURE 37.0

Default temperature for structure prediction and free energy evaluation in $^{\circ}C$

#define VRNA_MODEL_DEFAULT_PF_SCALE -1

Default scaling factor for partition function computations.

• #define VRNA MODEL DEFAULT BETA SCALE 1.

Default scaling factor for absolute thermodynamic temperature in Boltzmann factors.

• #define VRNA_MODEL_DEFAULT_DANGLES 2

Default dangling end model.

#define VRNA MODEL DEFAULT SPECIAL HP 1

Default model behavior for lookup of special tri-, tetra-, and hexa-loops.

• #define VRNA_MODEL_DEFAULT_NO_LP 0

Default model behavior for so-called 'lonely pairs'.

#define VRNA_MODEL_DEFAULT_NO_GU 0

Default model behavior for G-U base pairs.

• #define VRNA_MODEL_DEFAULT_NO_GU_CLOSURE 0

Default model behavior for G-U base pairs closing a loop.

#define VRNA_MODEL_DEFAULT_CIRC 0

Default model behavior to treat a molecule as a circular RNA (DNA)

• #define VRNA_MODEL_DEFAULT_GQUAD 0

Default model behavior regarding the treatment of G-Quadruplexes.

#define VRNA_MODEL_DEFAULT_UNIQ_ML 0

Default behavior of the model regarding unique multi-branch loop decomposition.

#define VRNA MODEL DEFAULT ENERGY SET 0

Default model behavior on which energy set to use.

• #define VRNA_MODEL_DEFAULT_BACKTRACK 1

Default model behavior with regards to backtracking of structures.

#define VRNA_MODEL_DEFAULT_BACKTRACK_TYPE 'F'

Default model behavior on what type of backtracking to perform.

#define VRNA MODEL DEFAULT COMPUTE BPP 1

Default model behavior with regards to computing base pair probabilities.

• #define VRNA_MODEL_DEFAULT_MAX_BP_SPAN -1

Default model behavior for the allowed maximum base pair span.

#define VRNA MODEL DEFAULT WINDOW SIZE -1

Default model behavior for the sliding window approach.

#define VRNA_MODEL_DEFAULT_LOG_ML 0

Default model behavior on how to evaluate the energy contribution of multi-branch loops.

• #define VRNA_MODEL_DEFAULT_ALI_OLD_EN 0

Default model behavior for consensus structure energy evaluation.

#define VRNA_MODEL_DEFAULT_ALI_RIBO 0

Default model behavior for consensus structure co-variance contribution assessment.

#define VRNA_MODEL_DEFAULT_ALI_CV_FACT 1.

Default model behavior for weighting the co-variance score in consensus structure prediction.

#define VRNA_MODEL_DEFAULT_ALI_NC_FACT 1.

Default model behavior for weighting the nucleotide conservation? in consensus structure prediction.

#define MAXALPHA 20

Maximal length of alphabet.

Typedefs

• typedef struct vrna_md_s vrna_md_t

Typename for the model details data structure vrna_md_s.

Functions

void vrna_md_set_default (vrna_md_t *md)

Apply default model details to a provided vrna md t data structure.

void vrna_md_update (vrna_md_t *md)

Update the model details data structure.

vrna_md_t * vrna_md_copy (vrna_md_t *md_to, const vrna_md_t *md_from)

Copy/Clone a vrna_md_t model.

char * vrna_md_option_string (vrna_md_t *md)

Get a corresponding commandline parameter string of the options in a vrna md t.

void vrna_md_defaults_reset (vrna_md_t *md_p)

Reset the global default model details to a specific set of parameters, or their initial values.

void vrna_md_defaults_temperature (double T)

Set default temperature for energy evaluation of loops.

double vrna_md_defaults_temperature_get (void)

Get default temperature for energy evaluation of loops.

void vrna_md_defaults_betaScale (double b)

Set default scaling factor of thermodynamic temperature in Boltzmann factors.

double vrna_md_defaults_betaScale_get (void)

Get default scaling factor of thermodynamic temperature in Boltzmann factors.

• void vrna_md_defaults_dangles (int d)

Set default dangle model for structure prediction.

int vrna_md_defaults_dangles_get (void)

Get default dangle model for structure prediction.

void vrna md defaults special hp (int flag)

Set default behavior for lookup of tabulated free energies for special hairpin loops, such as Tri-, Tetra-, or Hexa-loops.

int vrna_md_defaults_special_hp_get (void)

Get default behavior for lookup of tabulated free energies for special hairpin loops, such as Tri-, Tetra-, or Hexa-loops.

void vrna md defaults noLP (int flag)

Set default behavior for prediction of canonical secondary structures.

int vrna_md_defaults_noLP_get (void)

Get default behavior for prediction of canonical secondary structures.

void vrna md defaults noGU (int flag)

Set default behavior for treatment of G-U wobble pairs.

int vrna_md_defaults_noGU_get (void)

Get default behavior for treatment of G-U wobble pairs.

void vrna md defaults noGUclosure (int flag)

Set default behavior for G-U pairs as closing pair for loops.

int vrna_md_defaults_noGUclosure_get (void)

Get default behavior for G-U pairs as closing pair for loops.

void vrna md defaults logML (int flag)

Set default behavior recomputing free energies of multi-branch loops using a logarithmic model.

int vrna_md_defaults_logML_get (void)

Get default behavior recomputing free energies of multi-branch loops using a logarithmic model.

void vrna md defaults circ (int flag)

Set default behavior whether input sequences are circularized.

int vrna_md_defaults_circ_get (void)

Get default behavior whether input sequences are circularized.

void vrna_md_defaults_gquad (int flag)

Set default behavior for treatment of G-Quadruplexes.

int vrna_md_defaults_gquad_get (void)

Get default behavior for treatment of G-Quadruplexes.

void vrna_md_defaults_uniq_ML (int flag)

Set default behavior for creating additional matrix for unique multi-branch loop prediction.

int vrna_md_defaults_uniq_ML_get (void)

Get default behavior for creating additional matrix for unique multi-branch loop prediction.

void vrna_md_defaults_energy_set (int e)

Set default energy set.

int vrna_md_defaults_energy_set_get (void)

Get default energy set.

void vrna_md_defaults_backtrack (int flag)

Set default behavior for whether to backtrack secondary structures.

· int vrna md defaults backtrack get (void)

Get default behavior for whether to backtrack secondary structures.

void vrna_md_defaults_backtrack_type (char t)

Set default backtrack type, i.e. which DP matrix is used.

char vrna_md_defaults_backtrack_type_get (void)

Get default backtrack type, i.e. which DP matrix is used.

void vrna_md_defaults_compute_bpp (int flag)

Set the default behavior for whether to compute base pair probabilities after partition function computation.

int vrna md defaults compute bpp get (void)

Get the default behavior for whether to compute base pair probabilities after partition function computation.

• void vrna_md_defaults_max_bp_span (int span)

Set default maximal base pair span.

int vrna_md_defaults_max_bp_span_get (void)

Get default maximal base pair span.

· void vrna md defaults min loop size (int size)

Set default minimal loop size.

int vrna_md_defaults_min_loop_size_get (void)

Get default minimal loop size.

· void vrna md defaults window size (int size)

Set default window size for sliding window structure prediction approaches.

int vrna_md_defaults_window_size_get (void)

Get default window size for sliding window structure prediction approaches.

void vrna_md_defaults_oldAliEn (int flag)

Set default behavior for whether to use old energy model for comparative structure prediction.

int vrna_md_defaults_oldAliEn_get (void)

Get default behavior for whether to use old energy model for comparative structure prediction.

void vrna_md_defaults_ribo (int flag)

Set default behavior for whether to use Ribosum Scoring in comparative structure prediction.

int vrna_md_defaults_ribo_get (void)

Get default behavior for whether to use Ribosum Scoring in comparative structure prediction.

• void vrna_md_defaults_cv_fact (double factor)

Set the default co-variance scaling factor used in comparative structure prediction.

double vrna md defaults cv fact get (void)

Get the default co-variance scaling factor used in comparative structure prediction.

- void vrna md defaults nc fact (double factor)
- double vrna_md_defaults_nc_fact_get (void)
- void vrna_md_defaults_sfact (double factor)

Set the default scaling factor used to avoid under-/overflows in partition function computation.

double vrna_md_defaults_sfact_get (void)

Get the default scaling factor used to avoid under-/overflows in partition function computation.

void set_model_details (vrna_md_t *md)

Set default model details.

Variables

double temperature

Rescale energy parameters to a temperature in degC.

double pf_scale

A scaling factor used by pf_fold() to avoid overflows.

· int dangles

Switch the energy model for dangling end contributions (0, 1, 2, 3)

· int tetra_loop

Include special stabilizing energies for some tri-, tetra- and hexa-loops;.

· int noLonelyPairs

Global switch to avoid/allow helices of length 1.

int noGU

Global switch to forbid/allow GU base pairs at all.

· int no closingGU

GU allowed only inside stacks if set to 1.

· int circ

backward compatibility variable.. this does not effect anything

· int gquad

Allow G-quadruplex formation.

· int uniq ML

do ML decomposition uniquely (for subopt)

· int energy set

0 = BP; 1=any with GC; 2=any with AU-parameter

· int do_backtrack

do backtracking, i.e. compute secondary structures or base pair probabilities

char backtrack type

A backtrack array marker for inverse_fold()

• char * nonstandards

contains allowed non standard base pairs

int max_bp_span

Maximum allowed base pair span.

· int oldAliEn

use old alifold energies (with gaps)

int ribo

use ribosum matrices

• int logML

if nonzero use logarithmic ML energy in energy_of_struct

16.6.2 Data Structure Documentation

16.6.2.1 struct vrna md s

The data structure that contains the complete model details used throughout the calculations.

For convenience reasons, we provide the type name vrna_md_t to address this data structure without the use of the struct keyword

See also

vrna_md_set_default(), set_model_details(), vrna_md_update(), vrna_md_t

SWIG Wrapper Notes This data structure is wrapped as an object **md** with multiple related functions attached as methods.

A new set of default parameters can be obtained by calling the constructure of md:

• md() – Initialize with default settings

The resulting object has a list of attached methods which directly correspond to functions that mainly operate on the corresponding *C* data structure:

- reset() vrna_md_set_default()
- set_from_globals() set_model_details()
- option_string() vrna_md_option_string()

Note, that default parameters can be modified by directly setting any of the following global variables. Internally, getting/setting default parameters using their global variable representative translates into calls of the following functions, therefore these wrappers for these functions do not exist in the scripting language interface(s):

global variable	C getter	C setter
temperature	vrna_md_defaults_temperature_get()	vrna_md_defaults_temperature()
dangles	vrna_md_defaults_dangles_get()	vrna_md_defaults_dangles()
betaScale	vrna_md_defaults_betaScale_get()	vrna_md_defaults_betaScale()
tetra_loop	this is an alias of special_hp	
special_hp	vrna_md_defaults_special_hp_get()	vrna_md_defaults_special_hp()
noLonelyPairs	this is an alias of <i>noLP</i>	
noLP	vrna_md_defaults_noLP_get()	vrna_md_defaults_noLP()
noGU	vrna_md_defaults_noGU_get()	vrna_md_defaults_noGU()
no_closingGU	this is an alias of <i>noGUclosure</i>	
noGUclosure	vrna_md_defaults_noGUclosure_get()	vrna_md_defaults_noGUclosure()
logML	vrna_md_defaults_logML_get()	vrna_md_defaults_logML()
circ	vrna_md_defaults_circ_get()	vrna_md_defaults_circ()
gquad	vrna_md_defaults_gquad_get()	vrna_md_defaults_gquad()
uniq_ML	vrna_md_defaults_uniq_ML_get()	vrna_md_defaults_uniq_ML()
energy_set	vrna_md_defaults_energy_set_get()	vrna_md_defaults_energy_set()
backtrack	vrna_md_defaults_backtrack_get()	vrna_md_defaults_backtrack()
backtrack_type	vrna_md_defaults_backtrack_type_get()	vrna_md_defaults_backtrack_type()
do_backtrack	this is an alias of compute_bpp	
compute_bpp	vrna_md_defaults_compute_bpp_get()	vrna_md_defaults_compute_bpp()
max_bp_span	vrna_md_defaults_max_bp_span_get()	vrna_md_defaults_max_bp_span()
min_loop_size	vrna_md_defaults_min_loop_size_get()	vrna_md_defaults_min_loop_size()
window_size	vrna_md_defaults_window_size_get()	vrna_md_defaults_window_size()
oldAliEn	vrna_md_defaults_oldAliEn_get()	vrna_md_defaults_oldAliEn()
ribo	vrna_md_defaults_ribo_get()	vrna_md_defaults_ribo()
cv_fact	vrna_md_defaults_cv_fact_get()	vrna_md_defaults_cv_fact()
nc_fact	vrna_md_defaults_nc_fact_get()	vrna_md_defaults_nc_fact()
sfact	vrna_md_defaults_sfact_get()	vrna_md_defaults_sfact()

Data Fields

· double temperature

The temperature used to scale the thermodynamic parameters.

double betaScale

A scaling factor for the thermodynamic temperature of the Boltzmann factors.

• int pf_smooth

A flat specifying whether energies in Boltzmann factors need to be smoothed.

• int dangles

Specifies the dangle model used in any energy evaluation (0,1,2 or 3)

int special_hp

Include special hairpin contributions for tri, tetra and hexaloops.

· int noLP

Only consider canonical structures, i.e. no 'lonely' base pairs.

• int noGU

Do not allow GU pairs.

· int noGUclosure

Do not allow loops to be closed by GU pair.

• int logML

Use logarithmic scaling for multiloops.

· int circ

Assume RNA to be circular instead of linear.

· int gquad

Include G-quadruplexes in structure prediction.

· int uniq ML

Flag to ensure unique multi-branch loop decomposition during folding.

· int energy_set

Specifies the energy set that defines set of compatible base pairs.

· int backtrack

Specifies whether or not secondary structures should be backtraced.

· char backtrack_type

Specifies in which matrix to backtrack.

· int compute_bpp

Specifies whether or not backward recursions for base pair probability (bpp) computation will be performed.

char nonstandards [64]

contains allowed non standard bases

· int max_bp_span

maximum allowed base pair span

· int min_loop_size

Minimum size of hairpin loops.

· int window_size

Size of the sliding window for locally optimal structure prediction.

int oldAliEn

Use old alifold energy model.

· int ribo

Use ribosum scoring table in alifold energy model.

· double cv_fact

Co-variance scaling factor for consensus structure prediction.

· double nc_fact

Scaling factor to weight co-variance contributions of non-canonical pairs.

· double sfact

Scaling factor for partition function scaling.

• int rtype [8]

Reverse base pair type array.

• short alias [MAXALPHA+1]

alias of an integer nucleotide representation

• int pair [MAXALPHA+1][MAXALPHA+1]

Integer representation of a base pair.

16.6.2.1.1 Field Documentation

16.6.2.1.1.1 dangles

```
int vrna_md_s::dangles
```

Specifies the dangle model used in any energy evaluation (0,1,2 or 3)

If set to 0 no stabilizing energies are assigned to bases adjacent to helices in free ends and multiloops (so called dangling ends). Normally (dangles = 1) dangling end energies are assigned only to unpaired bases and a base cannot participate simultaneously in two dangling ends. In the partition function algorithm vrna_pf() these checks are neglected. To provide comparability between free energy minimization and partition function algorithms, the default setting is 2. This treatment of dangling ends gives more favorable energies to helices directly adjacent to one another, which can be beneficial since such helices often do engage in stabilizing interactions through co-axial stacking.

If set to 3 co-axial stacking is explicitly included for adjacent helices in multiloops. The option affects only mfe folding and energy evaluation (vrna_mfe() and vrna_eval_structure()), as well as suboptimal folding (vrna_subopt()) via re-evaluation of energies. Co-axial stacking with one intervening mismatch is not considered so far.

Note

Some function do not implement all dangle model but only a subset of (0,1,2,3). In particular, partition function algorithms can only handle 0 and 2. Read the documentation of the particular recurrences or energy evaluation function for information about the provided dangle model.

```
16.6.2.1.1.2 min_loop_size
```

```
int vrna_md_s::min_loop_size
```

Minimum size of hairpin loops.

Note

The default value for this field is TURN, however, it may be 0 in cofolding context.

16.6.3 Macro Definition Documentation

16.6.3.1 VRNA_MODEL_DEFAULT_TEMPERATURE

```
#define VRNA_MODEL_DEFAULT_TEMPERATURE 37.0
#include <ViennaRNA/model.h>
```

Default temperature for structure prediction and free energy evaluation in $^{\circ}C$

See also

vrna_md_t.temperature, vrna_md_defaults_reset(), vrna_md_set_default()

```
16.6.3.2 VRNA_MODEL_DEFAULT_PF_SCALE
#define VRNA_MODEL_DEFAULT_PF_SCALE -1
#include <ViennaRNA/model.h>
Default scaling factor for partition function computations.
See also
     vrna_exp_param_t.pf_scale, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.3 VRNA_MODEL_DEFAULT_BETA_SCALE
#define VRNA_MODEL_DEFAULT_BETA_SCALE 1.
#include <ViennaRNA/model.h>
Default scaling factor for absolute thermodynamic temperature in Boltzmann factors.
See also
     vrna_exp_param_t.alpha, vrna_md_t.betaScale, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.4 VRNA_MODEL_DEFAULT_DANGLES
#define VRNA_MODEL_DEFAULT_DANGLES 2
#include <ViennaRNA/model.h>
Default dangling end model.
See also
     vrna_md_t.dangles, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.5 VRNA_MODEL_DEFAULT_SPECIAL_HP
#define VRNA_MODEL_DEFAULT_SPECIAL_HP 1
#include <ViennaRNA/model.h>
Default model behavior for lookup of special tri-, tetra-, and hexa-loops.
See also
     vrna_md_t.special_hp, vrna_md_defaults_reset(), vrna_md_set_default()
```

```
16.6.3.6 VRNA_MODEL_DEFAULT_NO_LP
#define VRNA_MODEL_DEFAULT_NO_LP 0
#include <ViennaRNA/model.h>
Default model behavior for so-called 'lonely pairs'.
See also
     vrna_md_t.noLP, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.7 VRNA_MODEL_DEFAULT_NO_GU
#define VRNA_MODEL_DEFAULT_NO_GU 0
#include <ViennaRNA/model.h>
Default model behavior for G-U base pairs.
See also
     vrna_md_t.noGU, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.8 VRNA_MODEL_DEFAULT_NO_GU_CLOSURE
#define VRNA_MODEL_DEFAULT_NO_GU_CLOSURE 0
#include <ViennaRNA/model.h>
Default model behavior for G-U base pairs closing a loop.
See also
     vrna_md_t.noGUclosure, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.9 VRNA_MODEL_DEFAULT_CIRC
#define VRNA_MODEL_DEFAULT_CIRC 0
#include <ViennaRNA/model.h>
Default model behavior to treat a molecule as a circular RNA (DNA)
See also
     vrna_md_t.circ, vrna_md_defaults_reset(), vrna_md_set_default()
```

```
16.6 Fine-tuning of the Implemented Models
16.6.3.10 VRNA_MODEL_DEFAULT_GQUAD
#define VRNA_MODEL_DEFAULT_GQUAD 0
#include <ViennaRNA/model.h>
Default model behavior regarding the treatment of G-Quadruplexes.
See also
     vrna_md_t.gquad, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.11 VRNA_MODEL_DEFAULT_UNIQ_ML
#define VRNA_MODEL_DEFAULT_UNIQ_ML 0
#include <ViennaRNA/model.h>
Default behavior of the model regarding unique multi-branch loop decomposition.
See also
     vrna_md_t.uniq_ML, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.12 VRNA_MODEL_DEFAULT_ENERGY_SET
#define VRNA_MODEL_DEFAULT_ENERGY_SET 0
#include <ViennaRNA/model.h>
Default model behavior on which energy set to use.
See also
     vrna_md_t.energy_set, vrna_md_defaults_reset(), vrna_md_set_default()
```

```
16.6.3.13 VRNA_MODEL_DEFAULT_BACKTRACK
```

```
#define VRNA_MODEL_DEFAULT_BACKTRACK 1
#include <ViennaRNA/model.h>
```

Default model behavior with regards to backtracking of structures.

See also

```
vrna_md_t.backtrack, vrna_md_defaults_reset(), vrna_md_set_default()
```

```
16.6.3.14 VRNA_MODEL_DEFAULT_BACKTRACK_TYPE
#define VRNA_MODEL_DEFAULT_BACKTRACK_TYPE 'F'
#include <ViennaRNA/model.h>
Default model behavior on what type of backtracking to perform.
See also
     vrna md t.backtrack type, vrna md defaults reset(), vrna md set default()
16.6.3.15 VRNA_MODEL_DEFAULT_COMPUTE_BPP
#define VRNA_MODEL_DEFAULT_COMPUTE_BPP 1
#include <ViennaRNA/model.h>
Default model behavior with regards to computing base pair probabilities.
See also
     vrna_md_t.compute_bpp, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.16 VRNA_MODEL_DEFAULT_MAX_BP_SPAN
#define VRNA_MODEL_DEFAULT_MAX_BP_SPAN -1
#include <ViennaRNA/model.h>
Default model behavior for the allowed maximum base pair span.
See also
     vrna_md_t.max_bp_span, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.17 VRNA_MODEL_DEFAULT_WINDOW_SIZE
#define VRNA_MODEL_DEFAULT_WINDOW_SIZE -1
#include <ViennaRNA/model.h>
Default model behavior for the sliding window approach.
```

vrna_md_t.window_size, vrna_md_defaults_reset(), vrna_md_set_default()

See also

```
16.6.3.18 VRNA_MODEL_DEFAULT_LOG_ML
#define VRNA_MODEL_DEFAULT_LOG_ML 0
#include <ViennaRNA/model.h>
Default model behavior on how to evaluate the energy contribution of multi-branch loops.
See also
     vrna_md_t.logML, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.19 VRNA_MODEL_DEFAULT_ALI_OLD_EN
#define VRNA_MODEL_DEFAULT_ALI_OLD_EN 0
#include <ViennaRNA/model.h>
Default model behavior for consensus structure energy evaluation.
See also
     vrna_md_t.oldAliEn, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.20 VRNA_MODEL_DEFAULT_ALI_RIBO
#define VRNA_MODEL_DEFAULT_ALI_RIBO 0
#include <ViennaRNA/model.h>
Default model behavior for consensus structure co-variance contribution assessment.
See also
     vrna_md_t.ribo, vrna_md_defaults_reset(), vrna_md_set_default()
16.6.3.21 VRNA_MODEL_DEFAULT_ALI_CV_FACT
#define VRNA_MODEL_DEFAULT_ALI_CV_FACT 1.
```

See also

vrna_md_t.cv_fact, vrna_md_defaults_reset(), vrna_md_set_default()

Default model behavior for weighting the co-variance score in consensus structure prediction.

#include <ViennaRNA/model.h>

16.6.3.22 VRNA_MODEL_DEFAULT_ALI_NC_FACT

```
#define VRNA_MODEL_DEFAULT_ALI_NC_FACT 1.
#include <ViennaRNA/model.h>
```

Default model behavior for weighting the nucleotide conservation? in consensus structure prediction.

See also

```
vrna_md_t.nc_fact, vrna_md_defaults_reset(), vrna_md_set_default()
```

16.6.4 Function Documentation

16.6.4.1 vrna_md_set_default()

Apply default model details to a provided vrna_md_t data structure.

Use this function to initialize a vrna_md_t data structure with its default values

Parameters

md A pointer to the data structure that is about to be initialized

16.6.4.2 vrna_md_update()

Update the model details data structure.

This function should be called after changing the vrna_md_t.energy_set attribute since it re-initializes base pairing related arrays within the vrna_md_t data structure. In particular, vrna_md_t.pair, vrna_md_t.alias, and vrna_md_t.rtype are set to the values that correspond to the specified vrna_md_t.energy_set option

See also

vrna_md_t, vrna_md_t.energy_set, vrna_md_t.pair, vrna_md_t.rtype, vrna_md_t.alias, vrna_md_set_default()

16.6.4.3 vrna_md_copy()

Copy/Clone a vrna_md_t model.

Use this function to clone a given model either inplace (target container md_to given) or create a copy by cloning the source model and returning it ($md_to == NULL$).

Parameters

md_to	The model to be overwritten (if non-NULL and md_to != md_from)
md_from	The model to copy (if non-NULL)

Returns

A pointer to the copy model (or NULL if md_from == NULL)

16.6.4.4 vrna_md_option_string()

Get a corresponding commandline parameter string of the options in a vrna_md_t.

Note

This function is not threadsafe!

16.6.4.5 vrna_md_defaults_reset()

Reset the global default model details to a specific set of parameters, or their initial values.

This function resets the global default model details to their initial values, i.e. as specified by the ViennaRNA Package release, upon passing NULL as argument. Alternatively it resets them according to a set of provided parameters.

Note

The global default parameters affect all function calls of RNAlib where model details are not explicitly provided. Hence, any change of them is not considered threadsafe

Warning

This function first resets the global default settings to factory defaults, and only then applies user provided settings (if any). User settings that do not meet specifications are skipped.

See also

```
vrna_md_set_default(), vrna_md_t
```

Parameters

md⊷	A set of model details to use as global default (if NULL is passed, factory defaults are restored)
_p	

16.6.4.6 vrna md_defaults_temperature()

Set default temperature for energy evaluation of loops.

See also

vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_TEMPERATURE

Parameters

T Temperature in centigrade

16.6.4.7 vrna_md_defaults_temperature_get()

#include <ViennaRNA/model.h>

Get default temperature for energy evaluation of loops.

See also

 $vrna_md_defaults_temperature(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_T(), vrna_md_t, vrna_t, vrna$

Returns

The global default settings for temperature in centigrade

16.6.4.8 vrna_md_defaults_betaScale()

```
\begin{tabular}{ll} \beg
```

#include <ViennaRNA/model.h>

Set default scaling factor of thermodynamic temperature in Boltzmann factors.

Bolzmann factors are then computed as $exp(-E/(b \cdot kT))$.

See also

vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_BETA_SCALE

Parameters

```
b The scaling factor, default is 1.0
```

16.6.4.9 vrna_md_defaults_betaScale_get()

Get default scaling factor of thermodynamic temperature in Boltzmann factors.

See also

vrna_md_defaults_betaScale(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_BETAULT_B

Returns

The global default thermodynamic temperature scaling factor

16.6.4.10 vrna_md_defaults_dangles()

Set default dangle model for structure prediction.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_DANGLES
```

Parameters

```
d The dangle model
```

16.6.4.11 vrna_md_defaults_dangles_get()

Get default dangle model for structure prediction.

See also

```
vrna\_md\_defaults\_dangles(), vrna\_md\_defaults\_reset(), vrna\_md\_set\_default(), vrna\_md\_t, VRNA\_MODEL\_DEFAULT\_DANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGLEDANGL
```

Returns

The global default settings for the dangle model

16.6.4.12 vrna_md_defaults_special_hp()

Set default behavior for lookup of tabulated free energies for special hairpin loops, such as Tri-, Tetra-, or Hexa-loops.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_SPECIAL_HP
```

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.13 vrna_md_defaults_special_hp_get()

Get default behavior for lookup of tabulated free energies for special hairpin loops, such as Tri-, Tetra-, or Hexaloops.

See also

vrna_md_defaults_special_hp(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_SP

Returns

The global default settings for the treatment of special hairpin loops

16.6.4.14 vrna_md_defaults_noLP()

```
void vrna_md_defaults_noLP (
          int flag )
```

#include <ViennaRNA/model.h>

Set default behavior for prediction of canonical secondary structures.

See also

vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_NO_LP

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.15 vrna_md_defaults_noLP_get()

```
int vrna_md_defaults_noLP_get (
```

```
void )
#include <ViennaRNA/model.h>
```

Get default behavior for prediction of canonical secondary structures.

See also

```
vrna_md_defaults_noLP(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_NO_LP
```

Returns

The global default settings for predicting canonical secondary structures

```
16.6.4.16 vrna_md_defaults_noGU()
```

Set default behavior for treatment of G-U wobble pairs.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_NO_GU
```

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.17 vrna_md_defaults_noGU_get()

Get default behavior for treatment of G-U wobble pairs.

See also

```
vrna_md_defaults_noGU(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_NO_GU
```

Returns

The global default settings for treatment of G-U wobble pairs

16.6.4.18 vrna_md_defaults_noGUclosure()

Set default behavior for G-U pairs as closing pair for loops.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_NO_GU_CLOSURE
```

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.19 vrna_md_defaults_noGUclosure_get()

Get default behavior for G-U pairs as closing pair for loops.

See also

```
vrna_md_defaults_noGUclosure(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_I
```

Returns

The global default settings for treatment of G-U pairs closing a loop

16.6.4.20 vrna_md_defaults_logML()

Set default behavior recomputing free energies of multi-branch loops using a logarithmic model.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_LOG_ML
```

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.21 vrna_md_defaults_logML_get()

Get default behavior recomputing free energies of multi-branch loops using a logarithmic model.

See also

```
vrna_md_defaults_logML(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_LOG_N
```

Returns

The global default settings for logarithmic model in multi-branch loop free energy evaluation

16.6.4.22 vrna_md_defaults_circ()

Set default behavior whether input sequences are circularized.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_CIRC
```

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.23 vrna_md_defaults_circ_get()

```
#include <ViennaRNA/model.h>
```

Get default behavior whether input sequences are circularized.

See also

```
vrna_md_defaults_circ(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_CIRC
```

Returns

The global default settings for treating input sequences as circular

```
16.6.4.24 vrna_md_defaults_gquad()
```

Set default behavior for treatment of G-Quadruplexes.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_GQUAD
```

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.25 vrna_md_defaults_gquad_get()

Get default behavior for treatment of G-Quadruplexes.

See also

```
vrna_md_defaults_gquad(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_GQUAI
```

Returns

The global default settings for treatment of G-Quadruplexes

```
16.6.4.26 vrna_md_defaults_uniq_ML()
```

Set default behavior for creating additional matrix for unique multi-branch loop prediction.

Note

Activating this option usually results in higher memory consumption!

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_UNIQ_ML
```

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.27 vrna_md_defaults_uniq_ML_get()

Get default behavior for creating additional matrix for unique multi-branch loop prediction.

See also

```
vrna\_md\_defaults\_uniq\_ML(), vrna\_md\_defaults\_reset(), vrna\_md\_set\_default(), vrna\_md\_t, VRNA\_MODEL\_DEFAULT\_UNIQUE (), vrna\_md\_t, vrna\_t, vrn
```

Returns

The global default settings for creating additional matrices for unique multi-branch loop prediction

16.6.4.28 vrna md_defaults_energy_set()

Set default energy set.

See also

vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_ENERGY_SET

Parameters

```
e Energy set (0, 1, 2, 3)
```

16.6.4.29 vrna_md_defaults_energy_set_get()

Get default energy set.

See also

vrna_md_defaults_energy_set(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_EN

Returns

The global default settings for the energy set

16.6.4.30 vrna_md_defaults_backtrack()

Set default behavior for whether to backtrack secondary structures.

See also

vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_BACKTRACK

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.31 vrna_md_defaults_backtrack_get()

```
#include <ViennaRNA/model.h>
```

Get default behavior for whether to backtrack secondary structures.

See also

```
vrna_md_defaults_backtrack(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_BAC
```

Returns

The global default settings for backtracking structures

```
16.6.4.32 vrna_md_defaults_backtrack_type()
```

#include <ViennaRNA/model.h>

Set default backtrack type, i.e. which DP matrix is used.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_BACKTRACK_TYPE
```

Parameters

```
t The type ('F', 'C', or 'M')
```

16.6.4.33 vrna_md_defaults_backtrack_type_get()

Get default backtrack type, i.e. which DP matrix is used.

See also

```
vrna_md_defaults_backtrack_type(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_BACKTRACK_TYPE
```

Returns

The global default settings that specify which DP matrix is used for backtracking

16.6.4.34 vrna_md_defaults_compute_bpp()

Set the default behavior for whether to compute base pair probabilities after partition function computation.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_COMPUTE_BPP
```

Parameters

```
flag | On/Off switch (0 = OFF, else = ON)
```

16.6.4.35 vrna_md_defaults_compute_bpp_get()

Get the default behavior for whether to compute base pair probabilities after partition function computation.

See also

```
vrna\_md\_defaults\_compute\_bpp(), vrna\_md\_defaults\_reset(), vrna\_md\_set\_default(), vrna\_md\_t, VRNA\_MODEL\_DEFAULT\_total vrna\_md\_t, vrna\_t, vrna\_t,
```

Returns

The global default settings that specify whether base pair probabilities are computed together with partition function

16.6.4.36 vrna_md_defaults_max_bp_span()

Set default maximal base pair span.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_MAX_BP_SPAN
```

Parameters

span Maximal base pair span

16.6.4.37 vrna_md_defaults_max_bp_span_get()

Get default maximal base pair span.

See also

vrna_md_defaults_max_bp_span(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_

Returns

The global default settings for maximum base pair span

16.6.4.38 vrna_md_defaults_min_loop_size()

Set default minimal loop size.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, TURN
```

Parameters

size | Minimal size, i.e. number of unpaired nucleotides for a hairpin loop

16.6.4.39 vrna_md_defaults_min_loop_size_get()

```
#include <ViennaRNA/model.h>
```

Get default minimal loop size.

See also

```
vrna_md_defaults_min_loop_size(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, TURN
```

Returns

The global default settings for minimal size of hairpin loops

```
16.6.4.40 vrna_md_defaults_window_size()
```

```
void vrna_md_defaults_window_size (
    int size )
```

#include <ViennaRNA/model.h>

Set default window size for sliding window structure prediction approaches.

See also

 $vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_WINDOW_SIZE(), vrna_md_t, vrna_t, vrna_$

Parameters

size The size of the sliding window

16.6.4.41 vrna_md_defaults_window_size_get()

Get default window size for sliding window structure prediction approaches.

See also

vrna_md_defaults_window_size(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_V

Returns

The global default settings for the size of the sliding window

16.6.4.42 vrna_md_defaults_oldAliEn()

Set default behavior for whether to use old energy model for comparative structure prediction.

Note

This option is outdated. Activating the old energy model usually results in worse consensus structure predictions.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_ALI_OLD_EN
```

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.43 vrna_md_defaults_oldAliEn_get()

Get default behavior for whether to use old energy model for comparative structure prediction.

See also

```
vrna_md_defaults_oldAliEn(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_ALI_0
```

Returns

The global default settings for using old energy model for comparative structure prediction

16.6.4.44 vrna_md_defaults_ribo()

```
void vrna_md_defaults_ribo (
        int flag )
#include <ViennaRNA/model.h>
```

Set default behavior for whether to use Ribosum Scoring in comparative structure prediction.

See also

vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_ALI_RIBO

Parameters

```
flag On/Off switch (0 = OFF, else = ON)
```

16.6.4.45 vrna_md_defaults_ribo_get()

Get default behavior for whether to use Ribosum Scoring in comparative structure prediction.

See also

```
vrna_md_defaults_ribo(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_ALI_RIBC
```

Returns

The global default settings for using Ribosum scoring in comparative structure prediction

16.6.4.46 vrna_md_defaults_cv_fact()

Set the default co-variance scaling factor used in comparative structure prediction.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_ALI_CV_FACT
```

Parameters

```
factor The co-variance factor
```

16.6.4.47 vrna_md_defaults_cv_fact_get()

```
#include <ViennaRNA/model.h>
```

Get the default co-variance scaling factor used in comparative structure prediction.

See also

```
vrna\_md\_defaults\_cv\_fact(), vrna\_md\_defaults\_reset(), vrna\_md\_set\_default(), vrna\_md\_t, VRNA\_MODEL\_DEFAULT\_ALI\_C', vrna\_md\_t, VRNA\_MODEL\_DEFAULT\_ALI\_C', vrna\_md\_t, VRNA\_MODEL\_DEFAULT\_ALI\_C', vrna\_md\_t, vrna\_t, v
```

Returns

The global default settings for the co-variance factor

```
16.6.4.48 vrna_md_defaults_nc_fact()
```

See also

vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_ALI_NC_FACT

Parameters

factor

16.6.4.49 vrna_md_defaults_nc_fact_get()

See also

vrna_md_defaults_nc_fact(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t, VRNA_MODEL_DEFAULT_ALI_N

Returns

16.6.4.50 vrna_md_defaults_sfact()

Set the default scaling factor used to avoid under-/overflows in partition function computation.

See also

```
vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t
```

Parameters

```
factor The scaling factor (default: 1.07)
```

16.6.4.51 vrna_md_defaults_sfact_get()

Get the default scaling factor used to avoid under-/overflows in partition function computation.

See also

```
vrna_md_defaults_sfact(), vrna_md_defaults_reset(), vrna_md_set_default(), vrna_md_t
```

Returns

The global default settings of the scaling factor

16.6.4.52 set_model_details()

Set default model details.

Use this function if you wish to initialize a vrna_md_t data structure with its default values, i.e. the global model settings as provided by the deprecated global variables.

Deprecated This function will vanish as soon as backward compatibility of RNAlib is dropped (expected in version 3). Use vrna_md_set_default() instead!

Parameters

md A pointer to the data structure that is about to be initialized

16.6.5 Variable Documentation

16.6.5.1 temperature

double temperature

#include <ViennaRNA/model.h>

Rescale energy parameters to a temperature in degC.

Default is 37C. You have to call the update_..._params() functions after changing this parameter.

Deprecated Use vrna_md_defaults_temperature(), and vrna_md_defaults_temperature_get() to change, and read the global default temperature settings

See also

vrna_md_defaults_temperature(), vrna_md_defaults_temperature_get(), vrna_md_defaults_reset()

16.6.5.2 pf_scale

double pf_scale

#include <ViennaRNA/model.h>

A scaling factor used by pf_fold() to avoid overflows.

Should be set to approximately exp((-F/kT)/length), where F is an estimate for the ensemble free energy, for example the minimum free energy. You must call $update_pf_params()$ after changing this parameter. If pf_scale is -1 (the default) , an estimate will be provided automatically when computing partition functions, e.g. $pf_fold()$ The automatic estimate is usually insufficient for sequences more than a few hundred bases long.

16.6.5.3 dangles

```
int dangles
#include <ViennaRNA/model.h>
```

Switch the energy model for dangling end contributions (0, 1, 2, 3)

If set to 0 no stabilizing energies are assigned to bases adjacent to helices in free ends and multiloops (so called dangling ends). Normally (dangles = 1) dangling end energies are assigned only to unpaired bases and a base cannot participate simultaneously in two dangling ends. In the partition function algorithm pf_fold() these checks are neglected. If dangles is set to 2, all folding routines will follow this convention. This treatment of dangling ends gives more favorable energies to helices directly adjacent to one another, which can be beneficial since such helices often do engage in stabilizing interactions through co-axial stacking.

If dangles = 3 co-axial stacking is explicitly included for adjacent helices in multiloops. The option affects only mfe folding and energy evaluation (fold() and energy_of_structure()), as well as suboptimal folding (subopt()) via re-evaluation of energies. Co-axial stacking with one intervening mismatch is not considered so far.

Default is 2 in most algorithms, partition function algorithms can only handle 0 and 2

```
16.6.5.4 tetra_loop
```

```
int tetra_loop
#include <ViennaRNA/model.h>
```

Include special stabilizing energies for some tri-, tetra- and hexa-loops;.

default is 1.

16.6.5.5 noLonelyPairs

```
int noLonelyPairs
#include <ViennaRNA/model.h>
```

Global switch to avoid/allow helices of length 1.

Disallow all pairs which can only occur as lonely pairs (i.e. as helix of length 1). This avoids lonely base pairs in the predicted structures in most cases.

16.6.5.6 energy_set

```
int energy_set
#include <ViennaRNA/model.h>
```

0 = BP; 1=any with GC; 2=any with AU-parameter

If set to 1 or 2: fold sequences from an artificial alphabet ABCD..., where A pairs B, C pairs D, etc. using either GC (1) or AU parameters (2); default is 0, you probably don't want to change it.

16.6.5.7 do_backtrack

```
int do_backtrack
#include <ViennaRNA/model.h>
```

do backtracking, i.e. compute secondary structures or base pair probabilities

If 0, do not calculate pair probabilities in pf_fold(); this is about twice as fast. Default is 1.

16.6.5.8 backtrack_type

```
char backtrack_type
#include <ViennaRNA/model.h>
```

A backtrack array marker for inverse_fold()

If set to 'C': force (1,N) to be paired, 'M' fold as if the sequence were inside a multiloop. Otherwise ('F') the usual mfe structure is computed.

16.6.5.9 nonstandards

```
char* nonstandards
#include <ViennaRNA/model.h>
```

contains allowed non standard base pairs

Lists additional base pairs that will be allowed to form in addition to GC, CG, AU, UA, GU and UG. Nonstandard base pairs are given a stacking energy of 0.

16.6.5.10 max_bp_span

```
int max_bp_span
#include <ViennaRNA/model.h>
```

Maximum allowed base pair span.

A value of -1 indicates no restriction for distant base pairs.

16.7 Energy Parameters

All relevant functions to retrieve and copy pre-calculated energy parameter sets as well as reading/writing the energy parameter set from/to file(s).

16.7.1 Detailed Description

All relevant functions to retrieve and copy pre-calculated energy parameter sets as well as reading/writing the energy parameter set from/to file(s).

This module covers all relevant functions for pre-calculation of the energy parameters necessary for the folding routines provided by RNAlib. Furthermore, the energy parameter set in the RNAlib can be easily exchanged by a user-defined one. It is also possible to write the current energy parameter set into a text file. Collaboration diagram for Energy Parameters:

Modules

· Reading/Writing Energy Parameter Sets from/to File

Read and Write energy parameter sets from and to files or strings.

Files

• file basic.h

Functions to deal with sets of energy parameters.

file constants.h

Energy parameter constants.

· file convert.h

Functions and definitions for energy parameter file format conversion.

· file io.h

Read and write energy parameter files.

Data Structures

• struct vrna_param_s

The datastructure that contains temperature scaled energy parameters. More...

• struct vrna_exp_param_s

The data structure that contains temperature scaled Boltzmann weights of the energy parameters. More...

Typedefs

typedef struct vrna_param_s vrna_param_t

Typename for the free energy parameter data structure vrna_params.

typedef struct vrna exp param s vrna exp param t

Typename for the Boltzmann factor data structure vrna_exp_params.

typedef struct vrna_param_s paramT

Old typename of vrna param s.

typedef struct vrna_exp_param_s pf_paramT

Old typename of vrna_exp_param_s.

Functions

vrna_param_t * vrna_params (vrna_md_t *md)

Get a data structure containing prescaled free energy parameters.

vrna_param_t * vrna_params_copy (vrna_param_t *par)

Get a copy of the provided free energy parameters.

vrna_exp_param_t * vrna_exp_params (vrna_md_t *md)

Get a data structure containing prescaled free energy parameters already transformed to Boltzmann factors.

vrna_exp_param_t * vrna_exp_params_comparative (unsigned int n_seq, vrna_md_t *md)

Get a data structure containing prescaled free energy parameters already transformed to Boltzmann factors (alifold version)

vrna exp param t * vrna exp params copy (vrna exp param t *par)

Get a copy of the provided free energy parameters (provided as Boltzmann factors)

void vrna_params_subst (vrna_fold_compound_t *vc, vrna_param_t *par)

Update/Reset energy parameters data structure within a vrna_fold_compound_t.

void vrna_exp_params_subst (vrna_fold_compound_t *vc, vrna_exp_param_t *params)

Update the energy parameters for subsequent partition function computations.

• void vrna_exp_params_rescale (vrna_fold_compound_t *vc, double *mfe)

Rescale Boltzmann factors for partition function computations.

void vrna_params_reset (vrna_fold_compound_t *vc, vrna_md_t *md_p)

Reset free energy parameters within a vrna_fold_compound_t according to provided, or default model details.

void vrna exp params reset (vrna fold compound t *vc, vrna md t *md p)

Reset Boltzmann factors for partition function computations within a vrna_fold_compound_t according to provided, or default model details.

- vrna_exp_param_t * get_scaled_pf_parameters (void)
- vrna_exp_param_t * get_boltzmann_factors (double temperature, double betaScale, vrna_md_t md, double pf scale)

Get precomputed Boltzmann factors of the loop type dependent energy contributions with independent thermodynamic temperature.

vrna exp param t * get boltzmann factor copy (vrna exp param t *parameters)

Get a copy of already precomputed Boltzmann factors.

vrna_exp_param_t * get_scaled_alipf_parameters (unsigned int n_seq)

Get precomputed Boltzmann factors of the loop type dependent energy contributions (alifold variant)

vrna_exp_param_t * get_boltzmann_factors_ali (unsigned int n_seq, double temperature, double betaScale, vrna md t md, double pf scale)

Get precomputed Boltzmann factors of the loop type dependent energy contributions (alifold variant) with independent thermodynamic temperature.

vrna param t * scale parameters (void)

Get precomputed energy contributions for all the known loop types.

vrna_param_t * get_scaled_parameters (double temperature, vrna_md_t md)

Get precomputed energy contributions for all the known loop types.

16.7.2 Data Structure Documentation

16.7.2.1 struct vrna param s

The datastructure that contains temperature scaled energy parameters.

Collaboration diagram for vrna_param_s:

Data Fields

· double temperature

Temperature used for loop contribution scaling.

vrna_md_t model_details

Model details to be used in the recursions.

char param_file [256]

The filename the parameters were derived from, or empty string if they represent the default.

16.7.2.2 struct vrna_exp_param_s

The data structure that contains temperature scaled Boltzmann weights of the energy parameters.

Collaboration diagram for vrna_exp_param_s:

Data Fields

int id

An identifier for the data structure.

double pf_scale

Scaling factor to avoid over-/underflows.

double temperature

Temperature used for loop contribution scaling.

· double alpha

Scaling factor for the thermodynamic temperature.

· vrna_md_t model_details

Model details to be used in the recursions.

• char param_file [256]

The filename the parameters were derived from, or empty string if they represent the default.

16.7.2.2.1 Field Documentation

16.7.2.2.1.1 id

int vrna_exp_param_s::id

An identifier for the data structure.

Deprecated This attribute will be removed in version 3

```
16.7.2.2.1.2 alpha
```

```
double vrna_exp_param_s::alpha
```

Scaling factor for the thermodynamic temperature.

This allows for temperature scaling in Boltzmann factors independently from the energy contributions. The resulting Boltzmann factors are then computed by $e^{-E/(\alpha \cdot K \cdot T)}$

16.7.3 Typedef Documentation

```
16.7.3.1 paramT
```

```
typedef struct vrna_param_s paramT
#include <ViennaRNA/params/basic.h>
```

Old typename of vrna_param_s.

Deprecated Use vrna_param_t instead!

16.7.3.2 pf_paramT

```
typedef struct vrna_exp_param_s pf_paramT
#include <ViennaRNA/params/basic.h>
```

Old typename of vrna_exp_param_s.

Deprecated Use vrna_exp_param_t instead!

16.7.4 Function Documentation

16.7.4.1 vrna_params()

Get a data structure containing prescaled free energy parameters.

If a NULL pointer is passed for the model details parameter, the default model parameters are stored within the requested vrna param t structure.

See also

```
vrna_md_t, vrna_md_set_default(), vrna_exp_params()
```

Parameters

md A pointer to the model details to store inside the structure (Maybe NULL)

Returns

A pointer to the memory location where the requested parameters are stored

16.7.4.2 vrna_params_copy()

#include <ViennaRNA/params/basic.h>

Get a copy of the provided free energy parameters.

If NULL is passed as parameter, a default set of energy parameters is created and returned.

See also

```
vrna_params(), vrna_param_t
```

Parameters

par The free energy parameters that are to be copied (Maybe NULL)

Returns

A copy or a default set of the (provided) parameters

16.7.4.3 vrna_exp_params()

Get a data structure containing prescaled free energy parameters already transformed to Boltzmann factors.

This function returns a data structure that contains all necessary precomputed energy contributions for each type of loop.

In contrast to vrna_params(), the free energies within this data structure are stored as their Boltzmann factors, i.e.

```
exp(-E/kT)
```

where E is the free energy.

If a NULL pointer is passed for the model details parameter, the default model parameters are stored within the requested vrna_exp_param_t structure.

See also

vrna_md_t, vrna_md_set_default(), vrna_params(), vrna_rescale_pf_params()

Parameters

md A pointer to the model details to store inside the structure (Maybe NULL)

Returns

A pointer to the memory location where the requested parameters are stored

16.7.4.4 vrna_exp_params_comparative()

#include <ViennaRNA/params/basic.h>

Get a data structure containing prescaled free energy parameters already transformed to Boltzmann factors (alifold version)

If a NULL pointer is passed for the model details parameter, the default model parameters are stored within the requested vrna_exp_param_t structure.

See also

```
vrna_md_t, vrna_md_set_default(), vrna_exp_params(), vrna_params()
```

Parameters

n_seq	The number of sequences in the alignment
md	A pointer to the model details to store inside the structure (Maybe NULL)

Returns

A pointer to the memory location where the requested parameters are stored

16.7.4.5 vrna_exp_params_copy()

Get a copy of the provided free energy parameters (provided as Boltzmann factors)

If NULL is passed as parameter, a default set of energy parameters is created and returned.

See also

```
vrna_exp_params(), vrna_exp_param_t
```

Parameters

par The free energy parameters that are to be copied (Maybe NULL)

Returns

A copy or a default set of the (provided) parameters

16.7.4.6 vrna_params_subst()

#include <ViennaRNA/params/basic.h>

Update/Reset energy parameters data structure within a vrna_fold_compound_t.

Passing NULL as second argument leads to a reset of the energy parameters within vc to their default values. Otherwise, the energy parameters provided will be copied over into vc.

See also

```
vrna_params_reset(), vrna_param_t, vrna_md_t, vrna_params()
```

Parameters

vc	The vrna_fold_compound_t that is about to receive updated energy parameters
par	The energy parameters used to substitute those within vc (Maybe NULL)

SWIG Wrapper Notes This function is attached to vrna_fc_s objects as params_subst() method.

16.7.4.7 vrna_exp_params_subst()

#include <ViennaRNA/params/basic.h>

Update the energy parameters for subsequent partition function computations.

This function can be used to properly assign new energy parameters for partition function computations to a $vrna_fold_compound_t$. For this purpose, the data of the provided pointer params will be copied into vc and a recomputation of the partition function scaling factor is issued, if the pf_scale attribute of params is less than 1.0.

Passing NULL as second argument leads to a reset of the energy parameters within vc to their default values

See also

```
vrna_exp_params_reset(), vrna_exp_params_rescale(), vrna_exp_param_t, vrna_md_t, vrna_exp_params()
```

Parameters

VC	The fold compound data structure
params	A pointer to the new energy parameters

16.7.4.8 vrna_exp_params_rescale()

#include <ViennaRNA/params/basic.h>

Rescale Boltzmann factors for partition function computations.

This function may be used to (automatically) rescale the Boltzmann factors used in partition function computations. Since partition functions over subsequences can easily become extremely large, the RNAlib internally rescales them to avoid numerical over- and/or underflow. Therefore, a proper scaling factor s needs to be chosen that in turn is then used to normalize the corresponding partition functions $\hat{q}[i,j] = q[i,j]/s^{(j-i+1)}$.

This function provides two ways to automatically adjust the scaling factor.

- 1. Automatic guess
- 2. Automatic adjustment according to MFE

Passing NULL as second parameter activates the *automatic guess mode*. Here, the scaling factor is recomputed according to a mean free energy of 184.3*length cal for random sequences.

Note

This recomputation only takes place if the pf_scale attribute of the exp_params data structure contained in vc has a value below 1.0.

On the other hand, if the MFE for a sequence is known, it can be used to recompute a more robust scaling factor, since it represents the lowest free energy of the entire ensemble of structures, i.e. the highest Boltzmann factor. To activate this second mode of *automatic adjustment according to MFE*, a pointer to the MFE value needs to be passed as second argument. This value is then taken to compute the scaling factor as s = exp((sfact*MFE)/kT/length), where sfact is an additional scaling weight located in the vrna_md_t data structure of <code>expe_params</code> in vc.

The computed scaling factor s will be stored as pf_scale attribute of the exp_params data structure in vc.

See also

```
vrna_exp_params_subst(), vrna_md_t, vrna_exp_param_t, vrna_fold_compound_t
```

Parameters

VC	The fold compound data structure	
mfe	A pointer to the MFE (in kcal/mol) or NULL	

SWIG Wrapper Notes This function is attached to vrna_fc_s objects as overloaded **exp_params_rescale()** method.

When no parameter is passed to this method, the resulting action is the same as passing *NULL* as second parameter to <u>vrna_exp_params_rescale()</u>, i.e. default scaling of the partition function. Passing an energy in kcal/mol, e.g. as retrieved by a previous call to the *mfe()* method, instructs all subsequent calls to scale the partition function accordingly.

16.7.4.9 vrna_params_reset()

#include <ViennaRNA/params/basic.h>

Reset free energy parameters within a vrna_fold_compound_t according to provided, or default model details.

This function allows one to rescale free energy parameters for subsequent structure prediction or evaluation according to a set of model details, e.g. temperature values. To do so, the caller provides either a pointer to a set of model details to be used for rescaling, or NULL if global default setting should be used.

See also

```
vrna exp params reset(), vrna params subs()
```

Parameters

VC	The fold compound data structure	
md⊷	A pointer to the new model details (or NULL for reset to defaults)	
_p		

SWIG Wrapper Notes This function is attached to vrna_fc_s objects as overloaded params_reset() method.

When no parameter is passed to this method, the resulting action is the same as passing *NULL* as second parameter to vrna_params_reset(), i.e. global default model settings are used. Passing an object of type vrna_md_s resets the fold compound according to the specifications stored within the vrna_md_s object.

16.7.4.10 vrna_exp_params_reset()

#include <ViennaRNA/params/basic.h>

Reset Boltzmann factors for partition function computations within a vrna_fold_compound_t according to provided, or default model details.

This function allows one to rescale Boltzmann factors for subsequent partition function computations according to a set of model details, e.g. temperature values. To do so, the caller provides either a pointer to a set of model details to be used for rescaling, or NULL if global default setting should be used.

See also

```
vrna_params_reset(), vrna_exp_params_subst(), vrna_exp_params_rescale()
```

Parameters

VC	The fold compound data structure
md←	A pointer to the new model details (or NULL for reset to defaults)
_p	

SWIG Wrapper Notes This function is attached to vrna fc s objects as overloaded exp params reset() method.

When no parameter is passed to this method, the resulting action is the same as passing *NULL* as second parameter to vrna_exp_params_reset(), i.e. global default model settings are used. Passing an object of type vrna_md_s resets the fold compound according to the specifications stored within the vrna_md_s object.

16.7.4.11 get_scaled_pf_parameters()

#include <ViennaRNA/params/basic.h>

get a data structure of type vrna_exp_param_t which contains the Boltzmann weights of several energy parameters scaled according to the current temperature

Deprecated Use vrna_exp_params() instead!

Returns

The data structure containing Boltzmann weights for use in partition function calculations

16.7.4.12 get_boltzmann_factors()

#include <ViennaRNA/params/basic.h>

Get precomputed Boltzmann factors of the loop type dependent energy contributions with independent thermodynamic temperature.

This function returns a data structure that contains all necessary precalculated Boltzmann factors for each loop type contribution.

In contrast to get_scaled_pf_parameters(), this function enables setting of independent temperatures for both, the individual energy contributions as well as the thermodynamic temperature used in $exp(-\Delta G/kT)$

Deprecated Use vrna exp params() instead!

See also

```
get_scaled_pf_parameters(), get_boltzmann_factor_copy()
```

Parameters

temperature	The temperature in degrees Celcius used for (re-)scaling the energy contributions	
betaScale	A scaling value that is used as a multiplication factor for the absolute temperature of the system	
md	The model details to be used	
pf_scale	The scaling factor for the Boltzmann factors	

Returns

A set of precomputed Boltzmann factors

```
16.7.4.13 get_boltzmann_factor_copy()
```

Get a copy of already precomputed Boltzmann factors.

Deprecated Use vrna_exp_params_copy() instead!

See also

```
get_boltzmann_factors(), get_scaled_pf_parameters()
```

Parameters

parameters

Returns

A copy of the provided Boltzmann factor data set

```
16.7.4.14 get_scaled_alipf_parameters()
```

Get precomputed Boltzmann factors of the loop type dependent energy contributions (alifold variant)

Deprecated Use vrna_exp_params_comparative() instead!

16.7.4.15 get_boltzmann_factors_ali()

```
vrna_exp_param_t* get_boltzmann_factors_ali (
    unsigned int n_seq,
    double temperature,
    double betaScale,
    vrna_md_t md,
    double pf_scale )
```

#include <ViennaRNA/params/basic.h>

Get precomputed Boltzmann factors of the loop type dependent energy contributions (alifold variant) with independent thermodynamic temperature.

Deprecated Use vrna_exp_params_comparative() instead!

16.7.4.16 scale_parameters()

Get precomputed energy contributions for all the known loop types.

Note

OpenMP: This function relies on several global model settings variables and thus is not to be considered threadsafe. See get_scaled_parameters() for a completely threadsafe implementation.

Deprecated Use vrna_params() instead!

Returns

A set of precomputed energy contributions

16.7.4.17 get_scaled_parameters()

Get precomputed energy contributions for all the known loop types.

Call this function to retrieve precomputed energy contributions, i.e. scaled according to the temperature passed. Furthermore, this function assumes a data structure that contains the model details as well, such that subsequent folding recursions are able to retrieve the correct model settings

Deprecated Use vrna_params() instead!

See also

```
vrna_md_t, set_model_details()
```

Parameters

temperature	The temperature in degrees Celcius
md	The model details

Returns

precomputed energy contributions and model settings

16.8 Extending the Folding Grammar with Additional Domains

This module covers simple and straight-forward extensions to the RNA folding grammar.

16.8.1 Detailed Description

This module covers simple and straight-forward extensions to the RNA folding grammar.

Collaboration diagram for Extending the Folding Grammar with Additional Domains:

Modules

• Unstructured Domains

Add and modify unstructured domains to the RNA folding grammar.

· Structured Domains

Add and modify structured domains to the RNA folding grammar.

16.9 Unstructured Domains

Add and modify unstructured domains to the RNA folding grammar.

16.9.1 Detailed Description

Add and modify unstructured domains to the RNA folding grammar.

This module provides the tools to add and modify unstructured domains to the production rules of the RNA folding grammar. Usually this functionality is utilized for incorporating ligand binding to unpaired stretches of an RNA.

Bug Although the additional production rule(s) for unstructured domains as descibed in Unstructured Domains are always treated as 'segments possibly bound to one or more ligands', the current implementation requires that at least one ligand is bound. The default implementation already takes care of the required changes, however, upon using callback functions other than the default ones, one has to take care of this fact. Please also note, that this behavior might change in one of the next releases, such that the decomposition schemes as shown above comply with the actual implementation.

A default implementation allows one to readily use this feature by simply adding sequence motifs and corresponding binding free energies with the function vrna_ud_add_motif() (see also Ligands Binding to Unstructured Domains).

The grammar extension is realized using a callback function that

- evaluates the binding free energy of a ligand to its target sequence segment (white boxes in the figures above), or
- returns the free energy of an unpaired stretch possibly bound by a ligand, stored in the additional UDP matrix.

The callback is passed the segment positions, the loop context, and which of the two above mentioned evaluations are required. A second callback implements the pre-processing step that prepares the *U* DP matrix by evaluating all possible cases of the additional production rule. Both callbacks have a default implementation in *RNAlib*, but may be over-written by a user-implementation, making it fully user-customizable.

For equilibrium probability computations, two additional callbacks exist. One to store/add and one to retrieve the probability of unstructured domains at particular positions. Our implementation already takes care of computing the probabilities, but users of the unstructured domain feature are required to provide a mechanism to efficiently store/add the corresponding values into some external data structure. Collaboration diagram for Unstructured Domains:

Files

file unstructured_domains.h

Functions to modify unstructured domains, e.g. to incorporate ligands binding to unpaired stretches.

Data Structures

struct vrna_unstructured_domain_s

Data structure to store all functionality for ligand binding. More...

16.9 Unstructured Domains 225

Macros

#define VRNA_UNSTRUCTURED_DOMAIN_EXT_LOOP 1U

Flag to indicate ligand bound to unpiared stretch in the exterior loop.

#define VRNA_UNSTRUCTURED_DOMAIN_HP_LOOP 2U

Flag to indicate ligand bound to unpaired stretch in a hairpin loop.

#define VRNA_UNSTRUCTURED_DOMAIN_INT_LOOP 4U

Flag to indicate ligand bound to unpiared stretch in an interior loop.

#define VRNA_UNSTRUCTURED_DOMAIN_MB_LOOP 8U

Flag to indicate ligand bound to unpiared stretch in a multibranch loop.

#define VRNA UNSTRUCTURED DOMAIN MOTIF 16U

Flag to indicate ligand binding without additional unbound nucleotides (motif-only)

#define VRNA_UNSTRUCTURED_DOMAIN_ALL_LOOPS

Flag to indicate ligand bound to unpiared stretch in any loop (convenience macro)

Typedefs

typedef struct vrna unstructured domain s vrna ud t

Typename for the ligand binding extension data structure vrna_unstructured_domain_s.

 typedef int() vrna_callback_ud_energy(vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, void *data)

Callback to retrieve binding free energy of a ligand bound to an unpaired sequence segment.

 typedef FLT_OR_DBL() vrna_callback_ud_exp_energy(vrna_fold_compound_t *vc, int i, int j, unsigned int loop type, void *data)

Callback to retrieve Boltzmann factor of the binding free energy of a ligand bound to an unpaired sequence segment.

 $\bullet \ \ typedef\ void()\ vrna_callback_ud_production(vrna_fold_compound_t\ *vc,\ void\ *data) \\$

Callback for pre-processing the production rule of the ligand binding to unpaired stretches feature.

typedef void() vrna_callback_ud_exp_production(vrna_fold_compound_t *vc, void *data)

Callback for pre-processing the production rule of the ligand binding to unpaired stretches feature (partition function variant)

• typedef void() vrna_callback_ud_probs_add(vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, FLT_OR_DBL exp_energy, void *data)

Callback to store/add equilibrium probability for a ligand bound to an unpaired sequence segment.

• typedef FLT_OR_DBL() vrna_callback_ud_probs_get(vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, int motif, void *data)

Callback to retrieve equilibrium probability for a ligand bound to an unpaired sequence segment.

Functions

- vrna_ud_motif_t * vrna_ud_motifs_centroid (vrna_fold_compound_t *fc, const char *structure)
 Detect unstructured domains in centroid structure.
- vrna_ud_motif_t * vrna_ud_motifs_MEA (vrna_fold_compound_t *fc, const char *structure, vrna_ep_t *probability_list)

Detect unstructured domains in MEA structure.

vrna_ud_motif_t * vrna_ud_motifs_MFE (vrna_fold_compound_t *fc, const char *structure)

Detect unstructured domains in MFE structure.

void vrna_ud_add_motif (vrna_fold_compound_t *vc, const char *motif, double motif_en, const char *motif
 name, unsigned int loop_type)

Add an unstructured domain motif, e.g. for ligand binding.

void vrna_ud_remove (vrna_fold_compound_t *vc)

Remove ligand binding to unpaired stretches.

void vrna_ud_set_data (vrna_fold_compound_t *vc, void *data, vrna_callback_free_auxdata *free_cb)

Attach an auxiliary data structure.

• void vrna_ud_set_prod_rule_cb (vrna_fold_compound_t *vc, vrna_callback_ud_production *pre_cb, vrna_callback_ud_energy *e_cb)

Attach production rule callbacks for free energies computations.

void vrna_ud_set_exp_prod_rule_cb (vrna_fold_compound_t *vc, vrna_callback_ud_exp_production *pre
 —cb, vrna_callback_ud_exp_energy *exp_e_cb)

Attach production rule for partition function.

16.9.2 Data Structure Documentation

16.9.2.1 struct vrna_unstructured_domain_s

Data structure to store all functionality for ligand binding.

Data Fields

· int uniq_motif_count

The unique number of motifs of different lengths.

unsigned int * uniq motif size

An array storing a unique list of motif lengths.

· int motif_count

Total number of distinguished motifs.

char ** motif

Motif sequences.

char ** motif_name

Motif identifier/name.

• unsigned int * motif_size

Motif lengths.

double * motif_en

Ligand binding free energy contribution.

unsigned int * motif_type

Type of motif, i.e. loop type the ligand binds to.

vrna_callback_ud_production * prod_cb

Callback to ligand binding production rule, i.e. create/fill DP free energy matrices.

vrna_callback_ud_exp_production * exp_prod_cb

Callback to ligand binding production rule, i.e. create/fill DP partition function matrices.

vrna_callback_ud_energy * energy_cb

Callback to evaluate free energy of ligand binding to a particular unpaired stretch.

vrna_callback_ud_exp_energy * exp_energy_cb

Callback to evaluate Boltzmann factor of ligand binding to a particular unpaired stretch.

void * data

Auxiliary data structure passed to energy evaluation callbacks.

vrna callback free auxdata * free data

Callback to free auxiliary data structure.

vrna_callback_ud_probs_add * probs_add

Callback to store/add outside partition function.

vrna_callback_ud_probs_get * probs_get

Callback to retrieve outside partition function.

16.9 Unstructured Domains 227

16.9.2.1.1 Field Documentation

16.9.2.1.1.1 prod_cb

```
vrna_callback_ud_production* vrna_unstructured_domain_s::prod_cb
```

Callback to ligand binding production rule, i.e. create/fill DP free energy matrices.

This callback will be executed right before the actual secondary structure decompositions, and, therefore, any implementation must not interleave with the regular DP matrices.

16.9.3 Typedef Documentation

16.9.3.1 vrna_callback_ud_energy

```
typedef int() vrna_callback_ud_energy(vrna_fold_compound_t *vc, int i, int j, unsigned int
loop_type, void *data)
```

#include <ViennaRNA/unstructured_domains.h>

Callback to retrieve binding free energy of a ligand bound to an unpaired sequence segment.

Notes on Callback Functions This function will be called to determine the additional energy contribution of a specific unstructured domain, e.g. the binding free energy of some ligand.

Parameters

VC	The current vrna_fold_compound_t
i	The start of the unstructured domain (5' end)
j	The end of the unstructured domain (3' end)
loop_type	The loop context of the unstructured domain
data	Auxiliary data

Returns

The auxiliary energy contribution in deka-cal/mol

16.9.3.2 vrna_callback_ud_exp_energy

```
typedef FLT_OR_DBL() vrna_callback_ud_exp_energy(vrna_fold_compound_t *vc, int i, int j, unsigned
int loop_type, void *data)
```

```
#include <ViennaRNA/unstructured_domains.h>
```

Callback to retrieve Boltzmann factor of the binding free energy of a ligand bound to an unpaired sequence segment.

Notes on Callback Functions This function will be called to determine the additional energy contribution of a specific unstructured domain, e.g. the binding free energy of some ligand (Partition function variant, i.e. the Boltzmann factors instead of actual free energies).

Parameters

VC	The current vrna_fold_compound_t
i	The start of the unstructured domain (5' end)
j	The end of the unstructured domain (3' end)
loop_type	The loop context of the unstructured domain
data	Auxiliary data

Returns

The auxiliary energy contribution as Boltzmann factor

16.9.3.3 vrna_callback_ud_production

```
typedef void() vrna_callback_ud_production(vrna_fold_compound_t *vc, void *data)
#include <ViennaRNA/unstructured_domains.h>
```

Callback for pre-processing the production rule of the ligand binding to unpaired stretches feature.

Notes on Callback Functions The production rule for the unstructured domain grammar extension

16.9.3.4 vrna_callback_ud_exp_production

```
typedef void() vrna_callback_ud_exp_production(vrna_fold_compound_t *vc, void *data)
#include <ViennaRNA/unstructured_domains.h>
```

Callback for pre-processing the production rule of the ligand binding to unpaired stretches feature (partition function variant)

Notes on Callback Functions The production rule for the unstructured domain grammar extension (Partition function variant)

16.9 Unstructured Domains 229

16.9.3.5 vrna_callback_ud_probs_add

```
typedef void() vrna_callback_ud_probs_add(vrna_fold_compound_t *vc, int i, int j, unsigned int
loop_type, FLT_OR_DBL exp_energy, void *data)
```

```
#include <ViennaRNA/unstructured_domains.h>
```

Callback to store/add equilibrium probability for a ligand bound to an unpaired sequence segment.

Notes on Callback Functions A callback function to store equilibrium probabilities for the unstructured domain feature

16.9.3.6 vrna_callback_ud_probs_get

```
typedef FLT_OR_DBL() vrna_callback_ud_probs_get(vrna_fold_compound_t *vc, int i, int j, unsigned
int loop_type, int motif, void *data)
#include <ViennaRNA/unstructured_domains.h>
```

Callback to retrieve equilibrium probability for a ligand bound to an unpaired sequence segment.

Notes on Callback Functions A callback function to retrieve equilibrium probabilities for the unstructured domain feature

16.9.4 Function Documentation

16.9.4.1 vrna ud_motifs_centroid()

#include <ViennaRNA/unstructured domains.h>

Detect unstructured domains in centroid structure.

Given a centroid structure and a set of unstructured domains compute the list of unstructured domain motifs present in the centroid. Since we do not explicitly annotate unstructured domain motifs in dot-bracket strings, this function can be used to check for the presence and location of unstructured domain motifs under the assumption that the dot-bracket string is the centroid structure of the equilibrium ensemble.

See also

vrna_centroid()

Parameters

fc	The fold_compound data structure with pre-computed equilibrium probabilities and model settings
structure	The centroid structure in dot-bracket notation

Returns

A list of unstructured domain motifs (possibly NULL). The last element terminates the list with start=0, number=-1

16.9.4.2 vrna_ud_motifs_MEA()

#include <ViennaRNA/unstructured_domains.h>

Detect unstructured domains in MEA structure.

Given an MEA structure and a set of unstructured domains compute the list of unstructured domain motifs present in the MEA structure. Since we do not explicitly annotate unstructured domain motifs in dot-bracket strings, this function can be used to check for the presence and location of unstructured domain motifs under the assumption that the dot-bracket string is the MEA structure of the equilibrium ensemble.

See also

MEA()

Parameters

fc	The fold_compound data structure with pre-computed equilibrium probabilities and model settings
structure	The MEA structure in dot-bracket notation
probability_list	The list of probabilities to extract the MEA structure from

Returns

A list of unstructured domain motifs (possibly NULL). The last element terminates the list with start=0, number=-1

16.9.4.3 vrna_ud_motifs_MFE()

16.9 Unstructured Domains 231

```
#include <ViennaRNA/unstructured_domains.h>
```

Detect unstructured domains in MFE structure.

Given an MFE structure and a set of unstructured domains compute the list of unstructured domain motifs present in the MFE structure. Since we do not explicitly annotate unstructured domain motifs in dot-bracket strings, this function can be used to check for the presence and location of unstructured domain motifs under the assumption that the dot-bracket string is the MFE structure of the equilibrium ensemble.

See also

```
vrna_mfe()
```

Parameters

fc	The fold_compound data structure with model settings
structure	The MFE structure in dot-bracket notation

Returns

A list of unstructured domain motifs (possibly NULL). The last element terminates the list with start=0, number=-1

16.9.4.4 vrna_ud_add_motif()

#include <ViennaRNA/unstructured_domains.h>

Add an unstructured domain motif, e.g. for ligand binding.

This function adds a ligand binding motif and the associated binding free energy to the vrna_ud_t attribute of a vrna_fold_compound_t. The motif data will then be used in subsequent secondary structure predictions. Multiple calls to this function with different motifs append all additional data to a list of ligands, which all will be evaluated. Ligand motif data can be removed from the vrna_fold_compound_t again using the vrna_ud_remove() function. The loop type parameter allows one to limit the ligand binding to particular loop type, such as the exterior loop, hairpin loops, interior loops, or multibranch loops.

See also

VRNA_UNSTRUCTURED_DOMAIN_EXT_LOOP, VRNA_UNSTRUCTURED_DOMAIN_HP_LOOP, VRNA_UNSTRUCTURED_VRNA_UNSTRUCTURED_DOMAIN_MB_LOOP, VRNA_UNSTRUCTURED_DOMAIN_ALL_LOOPS, vrna_ud_remove()

Parameters

VC	The vrna_fold_compound_t data structure the ligand motif should be bound to
motif	The sequence motif the ligand binds to
motif_en	The binding free energy of the ligand in kcal/mol
motif_name	The name/id of the motif (may be NULL)
loop_type	The loop type the ligand binds to

16.9.4.5 vrna_ud_remove()

#include <ViennaRNA/unstructured_domains.h>

Remove ligand binding to unpaired stretches.

This function removes all ligand motifs that were bound to a vrna_fold_compound_t using the vrna_ud_add_motif() function.

Parameters

vc The vrna_fold_compound_t data structure the ligand motif data should be removed from

SWIG Wrapper Notes This function is attached as method ud_remove() to objects of type fold_compound

16.9.4.6 vrna_ud_set_data()

#include <ViennaRNA/unstructured_domains.h>

Attach an auxiliary data structure.

This function binds an arbitrary, auxiliary data structure for user-implemented ligand binding. The optional callback $free_cb$ will be passed the bound data structure whenever the $vrna_fold_compound_t$ is removed from memory to avoid memory leaks.

See also

```
vrna_ud_set_prod_rule_cb(), vrna_ud_set_exp_prod_rule_cb(), vrna_ud_remove()
```

16.9 Unstructured Domains 233

Parameters

VC	The vrna_fold_compound_t data structure the auxiliary data structure should be bound to
data	A pointer to the auxiliary data structure
free_cb	A pointer to a callback function that free's memory occupied by data

SWIG Wrapper Notes This function is attached as method ud_set_data() to objects of type fold_compound

16.9.4.7 vrna_ud_set_prod_rule_cb()

#include <ViennaRNA/unstructured_domains.h>

Attach production rule callbacks for free energies computations.

Use this function to bind a user-implemented grammar extension for unstructured domains.

The callback e_cb needs to evaluate the free energy contribution f(i,j) of the unpaired segment [i,j]. It will be executed in each of the regular secondary structure production rules. Whenever the callback is passed the VRNA_UNSTRUCTURED_DOMAIN_MOTIF flag via its loop_type parameter the contribution of any ligand that consecutively binds from position i to j (the white box) is requested. Otherwise, the callback usually performs a lookup in the precomputed B matrices. Which B matrix is addressed will be indicated by the flags VRNA_UNSTRUCTURED_DOMAIN_EXT_LOOP, VRNA_UNSTRUCTURED_DOMAIN_HP_LOOP VRNA_UNSTRUCTURED_DOMAIN_INT_LOOP, and VRNA_UNSTRUCTURED_DOMAIN_MB_LOOP. As their names already imply, they specify exterior loops (F production rule), hairpin loops and interior loops (C production rule), and multibranch loops (M and M1 production rule).

The pre_cb callback will be executed as a pre-processing step right before the regular secondary structure rules. Usually one would use this callback to fill the dynamic programming matrices ${\tt U}$ and preparations of the auxiliary data structure vrna_unstructured_domain_s.data

Parameters

VC	The vrna_fold_compound_t data structure the callback will be bound to
pre_cb	A pointer to a callback function for the \ensuremath{B} production rule
e_cb	A pointer to a callback function for free energy evaluation

SWIG Wrapper Notes This function is attached as method ud_set_prod_rule_cb() to objects of type fold_← compound

16.9.4.8 vrna_ud_set_exp_prod_rule_cb()

#include <ViennaRNA/unstructured_domains.h>

Attach production rule for partition function.

This function is the partition function companion of vrna_ud_set_prod_rule_cb().

Use it to bind callbacks to (i) fill the U production rule dynamic programming matrices and/or prepare the $vrna_unstructured_domain_s.data$, and (ii) provide a callback to retrieve partition functions for subsegments [i,j].

See also

vrna_ud_set_prod_rule_cb()

16.9 Unstructured Domains 235

Parameters

VC	The vrna_fold_compound_t data structure the callback will be bound to
pre_cb	A pointer to a callback function for the $\ensuremath{\mathbb{B}}$ production rule
exp_e_cb	A pointer to a callback function that retrieves the partition function for a segment $[i,j]$ that may be bound by one or more ligands.

SWIG Wrapper Notes This function is attached as method ud_set_exp_prod_rule_cb() to objects of type fold

_compound

16.10 Structured Domains

Add and modify structured domains to the RNA folding grammar.

16.10.1 Detailed Description

Add and modify structured domains to the RNA folding grammar.

This module provides the tools to add and modify structured domains to the production rules of the RNA folding grammar. Usually this functionality is utilized for incorporating self-enclosed structural modules that exhibit a more or less complex base pairing pattern. Collaboration diagram for Structured Domains:

Files

• file structured_domains.h

This module provides interfaces that deal with additional structured domains in the folding grammar.

16.11 Constraining the RNA Folding Grammar

This module provides general functions that allow for an easy control of constrained secondary structure prediction and evaluation.

16.11.1 Detailed Description

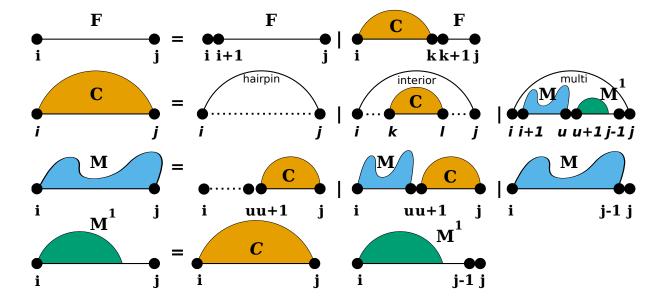
This module provides general functions that allow for an easy control of constrained secondary structure prediction and evaluation.

Secondary Structure constraints can be subdivided into two groups:

- · Hard Constraints, and
- · Soft Constraints.

While Hard-Constraints directly influence the production rules used in the folding recursions by allowing, disallowing, or enforcing certain decomposition steps, Soft-constraints on the other hand are used to change position specific contributions in the recursions by adding bonuses/penalties in form of pseudo free energies to certain loop configurations.

Secondary structure constraints are always applied at decomposition level, i.e. in each step of the recursive structure decomposition, for instance during MFE prediction. Below is a visualization of the decomposition scheme



For Hard Constraints the following option flags may be used to constrain the pairing behavior of single, or pairs of nucleotides:

- VRNA_CONSTRAINT_CONTEXT_EXT_LOOP Hard constraints flag, base pair in the exterior loop.
- VRNA_CONSTRAINT_CONTEXT_HP_LOOP Hard constraints flag, base pair encloses hairpin loop.
- VRNA_CONSTRAINT_CONTEXT_INT_LOOP Hard constraints flag, base pair encloses an interior loop.
- VRNA_CONSTRAINT_CONTEXT_INT_LOOP_ENC Hard constraints flag, base pair encloses a multi branch loop.

 VRNA_CONSTRAINT_CONTEXT_MB_LOOP - Hard constraints flag, base pair is enclosed in an interior loop.

- VRNA_CONSTRAINT_CONTEXT_MB_LOOP_ENC Hard constraints flag, base pair is enclosed in a multi branch loop.
- VRNA CONSTRAINT CONTEXT ENFORCE Hard constraint flag to indicate enforcement of constraints.
- VRNA_CONSTRAINT_CONTEXT_NO_REMOVE Hard constraint flag to indicate not to remove base pairs that conflict with a given constraint.
- VRNA_CONSTRAINT_CONTEXT_ALL_LOOPS Constraint context flag indicating any loop context.

However, for Soft Constraints we do not allow for simple loop type dependent constraining. But soft constraints are equipped with generic constraint support. This enables the user to pass arbitrary callback functions that return auxiliary energy contributions for evaluation the evaluation of any decomposition.

The callback will then always be notified about the type of decomposition that is happening, and the corresponding delimiting sequence positions. The following decomposition steps are distinguished, and should be captured by the user's implementation of the callback:

- VRNA_DECOMP_PAIR_HP Flag passed to generic softt constraints callback to indicate hairpin loop decomposition step.
- VRNA DECOMP_PAIR_IL Indicator for interior loop decomposition step.
- VRNA DECOMP PAIR ML Indicator for multibranch loop decomposition step.
- VRNA DECOMP ML ML ML Indicator for decomposition of multibranch loop part.
- VRNA_DECOMP_ML_STEM Indicator for decomposition of multibranch loop part.
- VRNA_DECOMP_ML_ML Indicator for decomposition of multibranch loop part.
- VRNA_DECOMP_ML_UP Indicator for decomposition of multibranch loop part.
- · VRNA_DECOMP_ML_ML_STEM Indicator for decomposition of multibranch loop part.
- VRNA_DECOMP_ML_COAXIAL Indicator for decomposition of multibranch loop part.
- VRNA_DECOMP_EXT_EXT Indicator for decomposition of exterior loop part.
- VRNA_DECOMP_EXT_UP Indicator for decomposition of exterior loop part.
- VRNA_DECOMP_EXT_STEM Indicator for decomposition of exterior loop part.
- VRNA_DECOMP_EXT_EXT_EXT Indicator for decomposition of exterior loop part.
- VRNA DECOMP EXT STEM EXT Indicator for decomposition of exterior loop part.
- VRNA_DECOMP_EXT_STEM_OUTSIDE Indicator for decomposition of exterior loop part.
- VRNA_DECOMP_EXT_EXT_STEM Indicator for decomposition of exterior loop part.
- VRNA_DECOMP_EXT_EXT_STEM1 Indicator for decomposition of exterior loop part.

Simplified interfaces to the soft constraints framework can be obtained by the implementations in the submodules

- SHAPE Reactivity Data and
- · ligands.

An implementation that generates soft constraints for unpaired nucleotides by minimizing the discrepancy between their predicted and expected pairing probability is available in submodule Generate Soft Constraints from Data. Collaboration diagram for Constraining the RNA Folding Grammar:

Modules

Hard Constraints

This module covers all functionality for hard constraints in secondary structure prediction.

Soft Constraints

Functions and data structures for secondary structure soft constraints.

Files

· file basic.h

Functions and data structures for constraining secondary structure predictions and evaluation.

Macros

• #define VRNA CONSTRAINT FILE 0

Flag for vrna constraints add() to indicate that constraints are present in a text file.

#define VRNA_CONSTRAINT_SOFT_MFE 0

Indicate generation of constraints for MFE folding.

• #define VRNA CONSTRAINT SOFT PF VRNA OPTION PF

Indicate generation of constraints for partition function computation.

#define VRNA_DECOMP_PAIR_HP (unsigned char)1

Flag passed to generic softt constraints callback to indicate hairpin loop decomposition step.

#define VRNA_DECOMP_PAIR_IL (unsigned char)2

Indicator for interior loop decomposition step.

#define VRNA_DECOMP_PAIR_ML (unsigned char)3

Indicator for multibranch loop decomposition step.

• #define VRNA_DECOMP_ML_ML (unsigned char)5

Indicator for decomposition of multibranch loop part.

• #define VRNA_DECOMP_ML_STEM (unsigned char)6

Indicator for decomposition of multibranch loop part.

#define VRNA_DECOMP_ML_ML (unsigned char)7

Indicator for decomposition of multibranch loop part.

#define VRNA_DECOMP_ML_UP (unsigned char)8

Indicator for decomposition of multibranch loop part.

#define VRNA_DECOMP_ML_ML_STEM (unsigned char)9

Indicator for decomposition of multibranch loop part.

• #define VRNA_DECOMP_ML_COAXIAL (unsigned char)10

Indicator for decomposition of multibranch loop part.

• #define VRNA_DECOMP_ML_COAXIAL_ENC (unsigned char)11

Indicator for decomposition of multibranch loop part.

#define VRNA_DECOMP_EXT_EXT (unsigned char)12

Indicator for decomposition of exterior loop part.

#define VRNA DECOMP EXT UP (unsigned char)13

Indicator for decomposition of exterior loop part.

#define VRNA_DECOMP_EXT_STEM (unsigned char)14

Indicator for decomposition of exterior loop part.

#define VRNA DECOMP EXT EXT EXT (unsigned char)15

Indicator for decomposition of exterior loop part.

#define VRNA_DECOMP_EXT_STEM_EXT (unsigned char)16

Indicator for decomposition of exterior loop part.

#define VRNA_DECOMP_EXT_STEM_OUTSIDE (unsigned char)17

Indicator for decomposition of exterior loop part.

• #define VRNA DECOMP EXT EXT STEM (unsigned char)18

Indicator for decomposition of exterior loop part.

#define VRNA_DECOMP_EXT_EXT_STEM1 (unsigned char)19

Indicator for decomposition of exterior loop part.

Functions

- void vrna_constraints_add (vrna_fold_compound_t *vc, const char *constraint, unsigned int options)

 Add constraints to a vrna_fold_compound_t data structure.
- void vrna message constraint options (unsigned int option)

Print a help message for pseudo dot-bracket structure constraint characters to stdout. (constraint support is specified by option parameter)

void vrna_message_constraint_options_all (void)

Print structure constraint characters to stdout (full constraint support)

16.11.2 Macro Definition Documentation

```
16.11.2.1 VRNA_CONSTRAINT_FILE
```

```
#define VRNA_CONSTRAINT_FILE 0
```

#include <ViennaRNA/constraints/basic.h>

Flag for vrna_constraints_add() to indicate that constraints are present in a text file.

See also

vrna_constraints_add()

Deprecated Use 0 instead!

16.11.2.2 VRNA_CONSTRAINT_SOFT_MFE

```
#define VRNA_CONSTRAINT_SOFT_MFE 0
```

#include <ViennaRNA/constraints/basic.h>

Indicate generation of constraints for MFE folding.

Deprecated This flag has no meaning anymore, since constraints are now always stored!

16.11.2.3 VRNA_CONSTRAINT_SOFT_PF

#define VRNA_CONSTRAINT_SOFT_PF VRNA_OPTION_PF

#include <ViennaRNA/constraints/basic.h>

Indicate generation of constraints for partition function computation.

Deprecated Use VRNA_OPTION_PF instead!

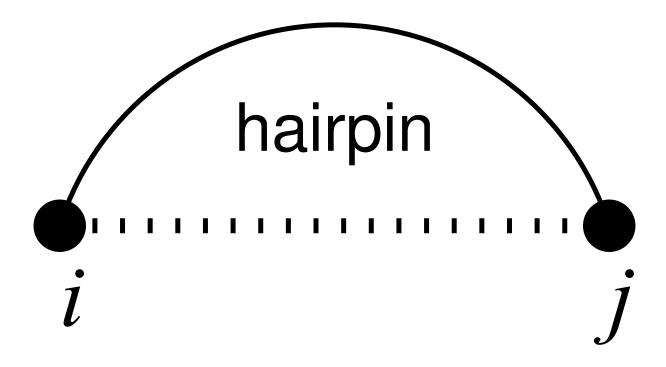
16.11.2.4 VRNA_DECOMP_PAIR_HP

#define VRNA_DECOMP_PAIR_HP (unsigned char)1

#include <ViennaRNA/constraints/basic.h>

Flag passed to generic softt constraints callback to indicate hairpin loop decomposition step.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a hairpin loop enclosed by the base pair (i, j).



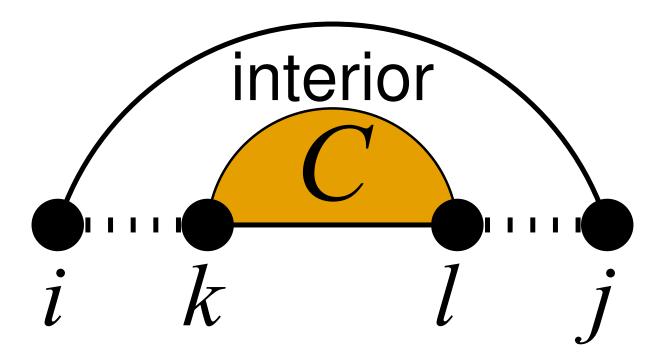
16.11.2.5 VRNA_DECOMP_PAIR_IL

#define VRNA_DECOMP_PAIR_IL (unsigned char)2

#include <ViennaRNA/constraints/basic.h>

Indicator for interior loop decomposition step.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates an interior loop enclosed by the base pair (i, j), and enclosing the base pair (k, l).



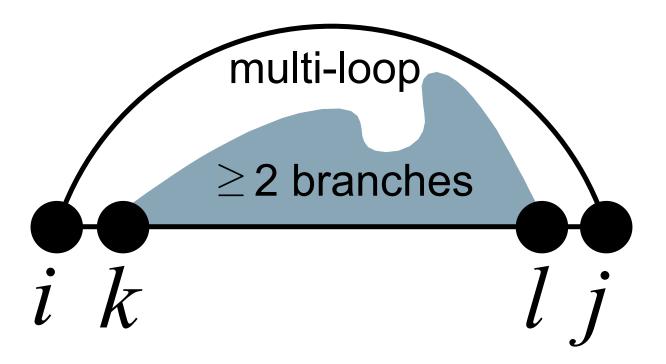
16.11.2.6 VRNA_DECOMP_PAIR_ML

#define VRNA_DECOMP_PAIR_ML (unsigned char)3

#include <ViennaRNA/constraints/basic.h>

Indicator for multibranch loop decomposition step.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a multi-branch loop enclosed by the base pair (i,j), and consisting of some enclosed multi loop content from k to l.



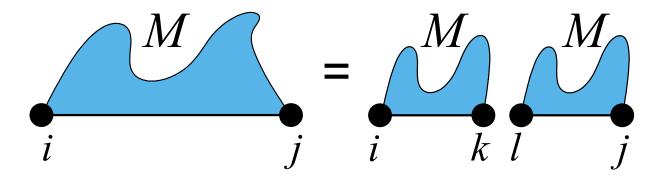
16.11.2.7 VRNA_DECOMP_ML_ML_ML

#define VRNA_DECOMP_ML_ML_ML (unsigned char)5

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of multibranch loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a multi-branch loop part in the interval [i:j], which will be decomposed into two multibranch loop parts [i:k], and [i:j].



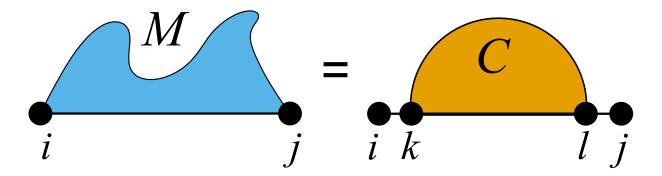
16.11.2.8 VRNA_DECOMP_ML_STEM

#define VRNA_DECOMP_ML_STEM (unsigned char) 6

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of multibranch loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a multibranch loop part in the interval [i:j], which will be considered a single stem branching off with base pair (k,l).



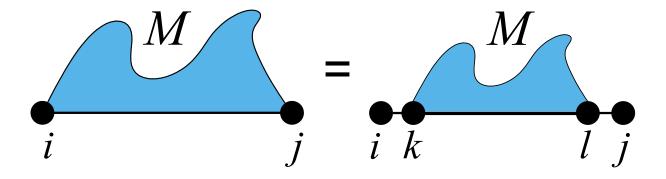
16.11.2.9 VRNA_DECOMP_ML_ML

#define VRNA_DECOMP_ML_ML (unsigned char) 7

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of multibranch loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a multi-branch loop part in the interval [i:j], which will be decomposed into a (usually) smaller multibranch loop part [k:l].



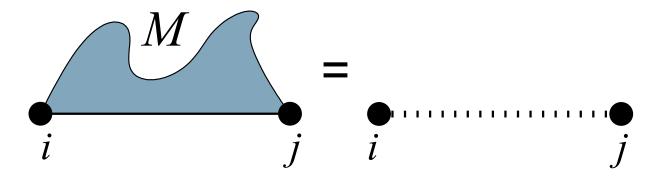
16.11.2.10 VRNA_DECOMP_ML_UP

#define VRNA_DECOMP_ML_UP (unsigned char)8

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of multibranch loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a multibranch loop part in the interval [i:j], which will be considered a multibranch loop part that only consists of unpaired nucleotides.



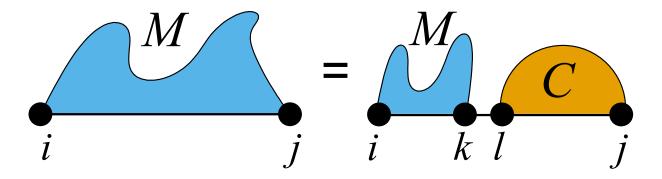
16.11.2.11 VRNA_DECOMP_ML_ML_STEM

#define VRNA_DECOMP_ML_ML_STEM (unsigned char) 9

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of multibranch loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a multibranch loop part in the interval [i:j], which will decomposed into a multibranch loop part [i:k], and a stem with enclosing base pair (l,j).



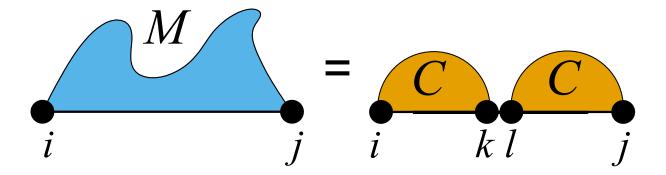
16.11.2.12 VRNA_DECOMP_ML_COAXIAL

#define VRNA_DECOMP_ML_COAXIAL (unsigned char)10

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of multibranch loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a multibranch loop part in the interval [i:j], where two stems with enclosing pairs (i,k) and (l,j) are coaxially stacking onto each other.



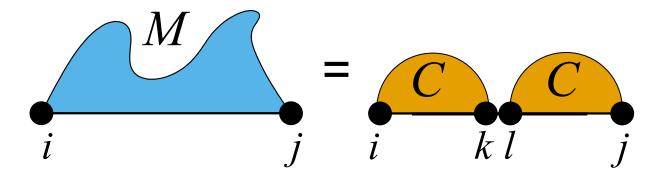
16.11.2.13 VRNA_DECOMP_ML_COAXIAL_ENC

#define VRNA_DECOMP_ML_COAXIAL_ENC (unsigned char)11

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of multibranch loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates a multi-branch loop part in the interval [i:j], where two stems with enclosing pairs (i,k) and (l,j) are coaxially stacking onto each other.



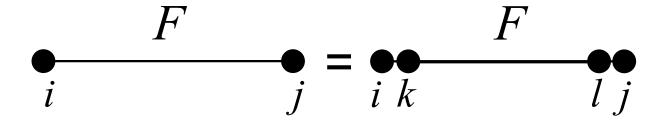
16.11.2.14 VRNA_DECOMP_EXT_EXT

#define VRNA_DECOMP_EXT_EXT (unsigned char)12

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of exterior loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates an exterior loop part in the interval [i:j], which will be decomposed into a (usually) smaller exterior loop part [k:l].



16.11.2.15 VRNA_DECOMP_EXT_UP

#define VRNA_DECOMP_EXT_UP (unsigned char)13

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of exterior loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates an exterior loop part in the interval [i:j], which will be considered as an exterior loop component consisting of only unpaired nucleotides.

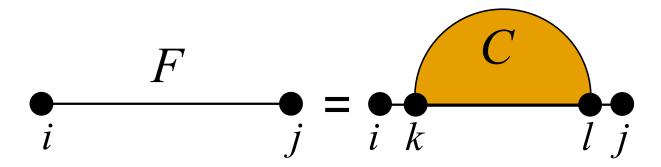
16.11.2.16 VRNA_DECOMP_EXT_STEM

#define VRNA_DECOMP_EXT_STEM (unsigned char)14

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of exterior loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates an exterior loop part in the interval [i:j], which will be considered a stem with enclosing pair (k,l).



16.11.2.17 VRNA_DECOMP_EXT_EXT_EXT

#define VRNA_DECOMP_EXT_EXT_EXT (unsigned char)15

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of exterior loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates an exterior loop part in the interval [i:j], which will be decomposed into two exterior loop parts [i:k] and [l:j].

16.11.2.18 VRNA_DECOMP_EXT_STEM_EXT

#define VRNA_DECOMP_EXT_STEM_EXT (unsigned char)16

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of exterior loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates an exterior loop part in the interval [i:j], which will be decomposed into a stem branching off with base pair (i,k), and an exterior loop part [l:j].

16.11.2.19 VRNA_DECOMP_EXT_EXT_STEM

#define VRNA_DECOMP_EXT_EXT_STEM (unsigned char)18

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of exterior loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates an exterior loop part in the interval [i:j], which will be decomposed into an exterior loop part [i:k], and a stem branching off with base pair (l,j).

$$F \longrightarrow F \longrightarrow C$$

$$i \longrightarrow f \longrightarrow k \longrightarrow j$$

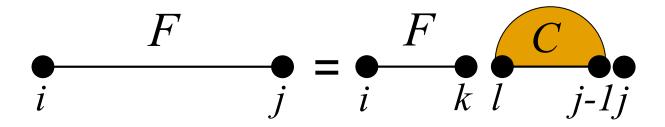
16.11.2.20 VRNA_DECOMP_EXT_EXT_STEM1

```
\verb|#define VRNA_DECOMP_EXT_EXT_STEM1 (unsigned char) 19|
```

#include <ViennaRNA/constraints/basic.h>

Indicator for decomposition of exterior loop part.

This flag notifies the soft or hard constraint callback function that the current decomposition step evaluates an exterior loop part in the interval [i:j], which will be decomposed into an exterior loop part [i:k], and a stem branching off with base pair (l,j-1).



16.11.3 Function Documentation

16.11.3.1 vrna_constraints_add()

#include <ViennaRNA/constraints/basic.h>

Add constraints to a vrna_fold_compound_t data structure.

Use this function to add/update the hard/soft constraints The function allows for passing a string 'constraint' that can either be a filename that points to a constraints definition file or it may be a pseudo dot-bracket notation indicating hard constraints. For the latter, the user has to pass the VRNA_CONSTRAINT_DB option. Also, the user has to specify, which characters are allowed to be interpreted as constraints by passing the corresponding options via the third parameter.

See also

vrna hc init(), vrna hc add up(), vrna hc add up batch(), vrna hc add bp(), vrna sc init(), vrna sc set up(), vrna sc set bp(), vrna sc add SHAPE deigan(), vrna sc add SHAPE zarringhalam(), vrna hc free(), vrna_sc_free(), VRNA_CONSTRAINT_DB, VRNA_CONSTRAINT_DB_DEFAULT, VRNA_CONSTRAINT_DB_PIPE, VRNA CONSTRAINT_DB_DOT, VRNA_CONSTRAINT_DB_X, VRNA_CONSTRAINT_DB_ANG_BRACK, VRNA CONSTRAINT DB RND BRACK, VRNA CONSTRAINT DB INTRAMOL, VRNA CONSTRAINT DB INTERMOL, VRNA CONSTRAINT DB GQUAD

The following is an example for adding hard constraints given in pseudo dot-bracket notation. Here, vc is the vrna fold compound t object, structure is a char array with the hard constraint in dot-bracket notation, and enforceConstraints is a flag indicating whether or not constraints for base pairs should be enforced instead of just doing a removal of base pair that conflict with the constraint.

```
unsigned int constraint_options = VRNA_CONSTRAINT_DB_DEFAULT; if (enforceConstraints)
 constraint_options |= VRNA_CONSTRAINT_DB_ENFORCE_BP;
if (canonicalBPonly)
 constraint_options |= VRNA_CONSTRAINT_DB_CANONICAL_BP;
vrna_constraints_add(fc, (const char *)cstruc, constraint_options);
```

In constrat to the above, constraints may also be read from file:

```
vrna_constraints_add(fc, constraints_file, VRNA_OPTION_DEFAULT);
```

See also

```
vrna_hc_add_from_db(),
                       vrna_hc_add_up(),
                                         vrna_hc_add_up_batch() vrna_hc_add_bp_unspecific(),
vrna hc add bp()
```

Parameters

VC	The fold compound
constraint	A string with either the filename of the constraint definitions or a pseudo dot-bracket notation of the hard constraint. May be NULL.
options	The option flags

16.11.3.2 vrna_message_constraint_options()

```
void vrna_message_constraint_options (
           unsigned int option )
#include <ViennaRNA/constraints/hard.h>
```

Print a help message for pseudo dot-bracket structure constraint characters to stdout. (constraint support is specified by option parameter)

```
Currently available options are:
```

```
VRNA_CONSTRAINT_DB_PIPE (paired with another base)
```

VRNA_CONSTRAINT_DB_DOT (no constraint at all)
VRNA_CONSTRAINT_DB_X (base must not pair)
VRNA_CONSTRAINT_DB_ANG_BRACK (paired downstream/upstream)

VRNA CONSTRAINT DB RND BRACK (base i pairs base j)

pass a collection of options as one value like this:

```
vrna_message_constraints(option_1 | option_2 | option_n)
```

See also

vrna_message_constraint_options_all(), vrna_constraints_add(), VRNA_CONSTRAINT_DB, VRNA_CONSTRAINT_DB_PIPE, VRNA_CONSTRAINT_DB_DOT, VRNA_CONSTRAINT_DB_X, VRNA_CONSTRAINT_DB_ANG_BRACK, VRNA_CONSTRAINT_DB_INTERMOL, VRNA_CONSTRAINT_DB_INTRAMOL

Parameters

option Option switch that tells which constraint help will be printed

16.11.3.3 vrna_message_constraint_options_all()

Print structure constraint characters to stdout (full constraint support)

See also

vrna_message_constraint_options(), vrna_constraints_add(), VRNA_CONSTRAINT_DB, VRNA_CONSTRAINT_DB_PIPE, VRNA_CONSTRAINT_DB_DOT, VRNA_CONSTRAINT_DB_X, VRNA_CONSTRAINT_DB_ANG_BRACK, VRNA_CONSTRAINT_DB_INTERMOL, VRNA_CONSTRAINT_DB_INTRAMOL

16.12 Hard Constraints

This module covers all functionality for hard constraints in secondary structure prediction.

16.12.1 Detailed Description

This module covers all functionality for hard constraints in secondary structure prediction.

Collaboration diagram for Hard Constraints:

Files

· file hard.h

Functions and data structures for handling of secondary structure hard constraints.

Data Structures

• struct vrna_hc_s

The hard constraints data structure. More...

struct vrna_hc_up_s

A single hard constraint for a single nucleotide. More...

Macros

• #define VRNA CONSTRAINT DB 16384U

Flag for vrna constraints add() to indicate that constraint is passed in pseudo dot-bracket notation.

#define VRNA CONSTRAINT DB ENFORCE BP 32768U

Switch for dot-bracket structure constraint to enforce base pairs.

#define VRNA_CONSTRAINT_DB_PIPE 65536U

Flag that is used to indicate the pipe '\' sign in pseudo dot-bracket notation of hard constraints.

#define VRNA CONSTRAINT DB DOT 131072U

dot '.' switch for structure constraints (no constraint at all)

#define VRNA_CONSTRAINT_DB_X 262144U

'x' switch for structure constraint (base must not pair)

#define VRNA_CONSTRAINT_DB_RND_BRACK 1048576U

round brackets '(',')' switch for structure constraint (base i pairs base j)

#define VRNA CONSTRAINT DB INTRAMOL 2097152U

Flag that is used to indicate the character II in pseudo dot-bracket notation of hard constraints.

#define VRNA_CONSTRAINT_DB_INTERMOL 4194304U

Flag that is used to indicate the character 'e' in pseudo dot-bracket notation of hard constraints.

#define VRNA CONSTRAINT DB GQUAD 8388608U

'+' switch for structure constraint (base is involved in a gquad)

#define VRNA_CONSTRAINT_DB_WUSS 33554432U

Flag to indicate Washington University Secondary Structure (WUSS) notation of the hard constraint string.

#define VRNA CONSTRAINT DB DEFAULT

Switch for dot-bracket structure constraint with default symbols.

#define VRNA_CONSTRAINT_CONTEXT_EXT_LOOP (unsigned char)0x01

16.12 Hard Constraints 255

Hard constraints flag, base pair in the exterior loop.

#define VRNA_CONSTRAINT_CONTEXT_HP_LOOP (unsigned char)0x02

Hard constraints flag, base pair encloses hairpin loop.

• #define VRNA_CONSTRAINT_CONTEXT_INT_LOOP (unsigned char)0x04

Hard constraints flag, base pair encloses an interior loop.

#define VRNA CONSTRAINT CONTEXT INT LOOP ENC (unsigned char)0x08

Hard constraints flag, base pair encloses a multi branch loop.

#define VRNA_CONSTRAINT_CONTEXT_MB_LOOP (unsigned char)0x10

Hard constraints flag, base pair is enclosed in an interior loop.

#define VRNA_CONSTRAINT_CONTEXT_MB_LOOP_ENC (unsigned char)0x20

Hard constraints flag, base pair is enclosed in a multi branch loop.

#define VRNA_CONSTRAINT_CONTEXT_ALL_LOOPS

Constraint context flag indicating any loop context.

Typedefs

typedef struct vrna_hc_s vrna_hc_t

Typename for the hard constraints data structure vrna hc s.

typedef struct vrna_hc_up_s vrna_hc_up_t

Typename for the single nucleotide hard constraint data structure vrna_hc_up_s.

• typedef unsigned char() vrna_callback_hc_evaluate(int i, int j, int k, int l, unsigned char d, void *data)

Callback to evaluate whether or not a particular decomposition step is contributing to the solution space.

Functions

void vrna_hc_init (vrna_fold_compound_t *vc)

Initialize/Reset hard constraints to default values.

void vrna_hc_add_up (vrna_fold_compound_t *vc, int i, unsigned char option)

Make a certain nucleotide unpaired.

• int vrna_hc_add_up_batch (vrna_fold_compound_t *vc, vrna_hc_up_t *constraints)

Apply a list of hard constraints for single nucleotides.

void vrna_hc_add_bp (vrna_fold_compound_t *vc, int i, int j, unsigned char option)

Favorize/Enforce a certain base pair (i,j)

void vrna hc add bp nonspecific (vrna fold compound t *vc, int i, int d, unsigned char option)

Enforce a nucleotide to be paired (upstream/downstream)

void vrna_hc_free (vrna_hc_t *hc)

Free the memory allocated by a vrna_hc_t data structure.

int vrna_hc_add_from_db (vrna_fold_compound_t *vc, const char *constraint, unsigned int options)

Add hard constraints from pseudo dot-bracket notation.

16.12.2 Data Structure Documentation

16.12.2.1 struct vrna hc s

The hard constraints data structure.

The content of this data structure determines the decomposition pattern used in the folding recursions. Attribute 'matrix' is used as source for the branching pattern of the decompositions during all folding recursions. Any entry in matrix[i,j] consists of the 6 LSB that allows one to distinguish the following types of base pairs:

- in the exterior loop (VRNA_CONSTRAINT_CONTEXT_EXT_LOOP)
- enclosing a hairpin (VRNA_CONSTRAINT_CONTEXT_HP_LOOP)
- enclosing an interior loop (VRNA_CONSTRAINT_CONTEXT_INT_LOOP)
- enclosed by an exterior loop (VRNA CONSTRAINT CONTEXT INT LOOP ENC)
- enclosing a multi branch loop (VRNA_CONSTRAINT_CONTEXT_MB_LOOP)
- enclosed by a multi branch loop (VRNA_CONSTRAINT_CONTEXT_MB_LOOP_ENC)

The four linear arrays 'up_xxx' provide the number of available unpaired nucleotides (including position i) 3' of each position in the sequence.

See also

vrna_hc_init(), vrna_hc_free(), VRNA_CONSTRAINT_CONTEXT_EXT_LOOP, VRNA_CONSTRAINT_CONTEXT_HP_LOOP, VRNA_CONSTRAINT_CONTEXT_INT_LOOP, VRNA_CONSTRAINT_CONTEXT_MB_LOOP, VRNA_CONTEXT_MB_LOOP, VRNA_CONTE

Collaboration diagram for vrna_hc_s:

Data Fields

int * up_ext

A linear array that holds the number of allowed unpaired nucleotides in an exterior loop.

int * up_hp

A linear array that holds the number of allowed unpaired nucleotides in a hairpin loop.

int * up int

A linear array that holds the number of allowed unpaired nucleotides in an interior loop.

• int * up_ml

A linear array that holds the number of allowed unpaired nucleotides in a multi branched loop.

vrna_callback_hc_evaluate * f

A function pointer that returns whether or not a certain decomposition may be evaluated.

void * data

A pointer to some structure where the user may store necessary data to evaluate its generic hard constraint function.

vrna_callback_free_auxdata * free_data

A pointer to a function to free memory occupied by auxiliary data.

• unsigned char * matrix

Upper triangular matrix that encodes where a base pair or unpaired nucleotide is allowed.

16.12 Hard Constraints 257

16.12.2.1.1 Field Documentation

16.12.2.1.1.1 free_data

```
vrna_callback_free_auxdata* vrna_hc_s::free_data
```

A pointer to a function to free memory occupied by auxiliary data.

The function this pointer is pointing to will be called upon destruction of the vrna_hc_s, and provided with the vrna_hc_s.data pointer that may hold auxiliary data. Hence, to avoid leaking memory, the user may use this pointer to free memory occupied by auxiliary data.

16.12.2.2 struct vrna_hc_up_s

A single hard constraint for a single nucleotide.

Data Fields

· int position

The sequence position (1-based)

unsigned char options

The hard constraint option.

16.12.3 Macro Definition Documentation

16.12.3.1 VRNA_CONSTRAINT_DB

```
#define VRNA_CONSTRAINT_DB 16384U
```

#include <ViennaRNA/constraints/hard.h>

Flag for vrna_constraints_add() to indicate that constraint is passed in pseudo dot-bracket notation.

See also

vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

```
16.12.3.2 VRNA_CONSTRAINT_DB_ENFORCE_BP
```

```
#define VRNA_CONSTRAINT_DB_ENFORCE_BP 32768U
```

```
#include <ViennaRNA/constraints/hard.h>
```

Switch for dot-bracket structure constraint to enforce base pairs.

This flag should be used to really enforce base pairs given in dot-bracket constraint rather than just weakly-enforcing them.

See also

vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

16.12.3.3 VRNA_CONSTRAINT_DB_PIPE

```
#define VRNA_CONSTRAINT_DB_PIPE 65536U
```

```
#include <ViennaRNA/constraints/hard.h>
```

Flag that is used to indicate the pipe '|' sign in pseudo dot-bracket notation of hard constraints.

Use this definition to indicate the pipe sign '|' (paired with another base)

See also

vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

16.12.3.4 VRNA_CONSTRAINT_DB_DOT

```
#define VRNA_CONSTRAINT_DB_DOT 131072U
```

#include <ViennaRNA/constraints/hard.h>

dot '.' switch for structure constraints (no constraint at all)

See also

vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

16.12 Hard Constraints 259

```
16.12.3.5 VRNA_CONSTRAINT_DB_X
#define VRNA_CONSTRAINT_DB_X 262144U
#include <ViennaRNA/constraints/hard.h>
'x' switch for structure constraint (base must not pair)
See also
     vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()
16.12.3.6 VRNA_CONSTRAINT_DB_RND_BRACK
#define VRNA_CONSTRAINT_DB_RND_BRACK 1048576U
#include <ViennaRNA/constraints/hard.h>
round brackets '(',')' switch for structure constraint (base i pairs base j)
See also
     vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()
16.12.3.7 VRNA_CONSTRAINT_DB_INTRAMOL
#define VRNA_CONSTRAINT_DB_INTRAMOL 2097152U
#include <ViennaRNA/constraints/hard.h>
Flag that is used to indicate the character 'l' in pseudo dot-bracket notation of hard constraints.
Use this definition to indicate the usage of 'I' character (intramolecular pairs only)
See also
```

vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

16.12.3.8 VRNA_CONSTRAINT_DB_INTERMOL

```
#define VRNA_CONSTRAINT_DB_INTERMOL 4194304U
```

```
#include <ViennaRNA/constraints/hard.h>
```

Flag that is used to indicate the character 'e' in pseudo dot-bracket notation of hard constraints.

Use this definition to indicate the usage of 'e' character (intermolecular pairs only)

See also

vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

16.12.3.9 VRNA CONSTRAINT DB GQUAD

```
#define VRNA_CONSTRAINT_DB_GQUAD 8388608U
```

#include <ViennaRNA/constraints/hard.h>

'+' switch for structure constraint (base is involved in a gquad)

See also

vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

Warning

This flag is for future purposes only! No implementation recognizes it yet.

16.12.3.10 VRNA_CONSTRAINT_DB_WUSS

```
#define VRNA_CONSTRAINT_DB_WUSS 33554432U
```

#include <ViennaRNA/constraints/hard.h>

Flag to indicate Washington University Secondary Structure (WUSS) notation of the hard constraint string.

This secondary structure notation for RNAs is usually used as consensus secondary structure (SS_cons) entry in Stockholm formatted files

16.12 Hard Constraints 261

16.12.3.11 VRNA_CONSTRAINT_DB_DEFAULT

```
#define VRNA_CONSTRAINT_DB_DEFAULT
```

#include <ViennaRNA/constraints/hard.h>

Value:

```
(VRNA_CONSTRAINT_DB \
| VRNA_CONSTRAINT_DB_PIPE \
| VRNA_CONSTRAINT_DB_DOT \
| VRNA_CONSTRAINT_DB_X \
| VRNA_CONSTRAINT_DB_ANG_BRACK \
| VRNA_CONSTRAINT_DB_RND_BRACK \
| VRNA_CONSTRAINT_DB_INTRAMOL \
| VRNA_CONSTRAINT_DB_INTERMOL \
| VRNA_CONSTRAINT_DB_GQUAD \
)
```

Switch for dot-bracket structure constraint with default symbols.

This flag conveniently combines all possible symbols in dot-bracket notation for hard constraints and VRNA CONSTRAINT DB

See also

vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

16.12.4 Typedef Documentation

16.12.4.1 vrna callback hc_evaluate

```
typedef unsigned char() vrna_callback_hc_evaluate(int i, int j, int k, int l, unsigned char d,
void *data)
```

#include <ViennaRNA/constraints/hard.h>

Callback to evaluate whether or not a particular decomposition step is contributing to the solution space.

This is the prototype for callback functions used by the folding recursions to evaluate generic hard constraints. The first four parameters passed indicate the delimiting nucleotide positions of the decomposition, and the parameter denotes the decomposition step. The last parameter data is the auxiliary data structure associated to the hard constraints via vrna_hc_add_data(), or NULL if no auxiliary data was added.

Notes on Callback Functions This callback enables one to over-rule default hard constraints in secondary structure decompositions.

See also

```
VRNA_DECOMP_PAIR_HP, VRNA_DECOMP_PAIR_IL, VRNA_DECOMP_PAIR_ML, VRNA_DECOMP_ML_ML_ML, VRNA_DECOMP_ML_STEM, VRNA_DECOMP_ML_ML, VRNA_DECOMP_ML_UP, VRNA_DECOMP_ML_ML_STEM, VRNA_DECOMP_ML_COAXIAL, VRNA_DECOMP_EXT_EXT, VRNA_DECOMP_EXT_UP, VRNA_DECOMP_EXT_STEM, VRNA_DECOMP_EXT_EXT_STEM, VRNA_DECOMP_EXT_EXT_STEM, VRNA_DECOMP_EXT_STEM1, vrna_hc_add_f(), vrna_hc_add_data()
```

Parameters

i	Left (5') delimiter position of substructure
j	Right (3') delimiter position of substructure
k	Left delimiter of decomposition
1	Right delimiter of decomposition
d	Decomposition step indicator
data	Auxiliary data

Returns

A non-zero value if the decomposition is valid, 0 otherwise

16.12.5 Function Documentation

Initialize/Reset hard constraints to default values.

This function resets the hard constraints to their default values, i.e. all positions may be unpaired in all contexts, and base pairs are allowed in all contexts, if they resemble canonical pairs. Previously set hard constraints will be removed before initialization.

See also

```
vrna_hc_add_bp(), vrna_hc_add_bp_nonspecific(), vrna_hc_add_up()
```

Parameters

vc	The fold compound
----	-------------------

SWIG Wrapper Notes This function is attached as method hc_init() to objects of type fold_compound

16.12 Hard Constraints 263

```
int i,
     unsigned char option )

#include <ViennaRNA/constraints/hard.h>
```

Make a certain nucleotide unpaired.

See also

vrna_hc_add_bp(), vrna_hc_add_bp_nonspecific(), vrna_hc_init(), VRNA_CONSTRAINT_CONTEXT_EXT_LOOP, VRNA_CONSTRAINT_CONTEXT_HP_LOOP, VRNA_CONSTRAINT_CONTEXT_INT_LOOP, VRNA_CONSTRAINT_CONTE VRNA_CONSTRAINT_CONTEXT_ALL_LOOPS

Parameters

vc	The vrna_fold_compound_t the hard constraints are associated with
i	The position that needs to stay unpaired (1-based)
option	The options flag indicating how/where to store the hard constraints

16.12.5.3 vrna_hc_add_up_batch()

#include <ViennaRNA/constraints/hard.h>

Apply a list of hard constraints for single nucleotides.

Parameters

VC	The vrna_fold_compound_t the hard constraints are associated with
constraints	The list off constraints to apply, last entry must have position attribute set to 0

16.12.5.4 vrna_hc_add_bp()

#include <ViennaRNA/constraints/hard.h>

Favorize/Enforce a certain base pair (i,j)

See also

vrna_hc_add_bp_nonspecific(), vrna_hc_add_up(), vrna_hc_init(), VRNA_CONSTRAINT_CONTEXT_EXT_LOOP, VRNA_CONSTRAINT_CONTEXT_HP_LOOP, VRNA_CONSTRAINT_CONTEXT_INT_LOOP, VRNA_CONSTRAINT_CONTEXT_MB_LOOP, VRNA_CONSTRAINT_CONTEXT_MB_LOOP_ENC, VRNA_CONSTRAINT_CONTEXT_BROWNERS VRNA_CONSTRAINT_CONTEXT_ALL_LOOPS

Parameters

VC	The vrna_fold_compound_t the hard constraints are associated with
i	The 5' located nucleotide position of the base pair (1-based)
j	The 3' located nucleotide position of the base pair (1-based)
option	The options flag indicating how/where to store the hard constraints

16.12.5.5 vrna_hc_add_bp_nonspecific()

#include <ViennaRNA/constraints/hard.h>

Enforce a nucleotide to be paired (upstream/downstream)

See also

```
vrna_hc_add_bp(), vrna_hc_add_up(), vrna_hc_init(), VRNA_CONSTRAINT_CONTEXT_EXT_LOOP, VRNA_CONSTRAINT_CONTEXT_HP_LOOP, VRNA_CONSTRAINT_CONTEXT_INT_LOOP, VRNA_CONSTRAINT_CONTEXT_MB_LOOP, VRNA_CONSTRAINT_CONTEXT_MB_LOOP_ENC, VRNA_CONSTRAINT_CONTEXT_ALL_LOOPS
```

Parameters

VC	The vrna_fold_compound_t the hard constraints are associated with	
i	The position that needs to stay unpaired (1-based)	
d	The direction of base pairing ($d<0$: pairs upstream, $d>0$: pairs downstream, $d==0$: no direction)	
option	The options flag indicating in which loop type context the pairs may appear	

16.12.5.6 vrna_hc_free()

16.12 Hard Constraints 265

Free the memory allocated by a vrna_hc_t data structure.

Use this function to free all memory that was allocated for a data structure of type vrna_hc_t .

See also

```
get_hard_constraints(), vrna_hc_t
```

16.12.5.7 vrna hc_add_from_db()

#include <ViennaRNA/constraints/hard.h>

Add hard constraints from pseudo dot-bracket notation.

This function allows one to apply hard constraints from a pseudo dot-bracket notation. The options parameter controls, which characters are recognized by the parser. Use the VRNA_CONSTRAINT_DB_DEFAULT convenience macro, if you want to allow all known characters

See also

VRNA_CONSTRAINT_DB_PIPE, VRNA_CONSTRAINT_DB_DOT, VRNA_CONSTRAINT_DB_X, VRNA_CONSTRAINT_DB_VRNA_CONSTRAINT_DB_RND_BRACK, VRNA_CONSTRAINT_DB_INTRAMOL, VRNA_CONSTRAINT_DB_INTERMOL, VRNA_CONSTRAINT_DB_GQUAD

Parameters

VC	The fold compound
constraint	A pseudo dot-bracket notation of the hard constraint.
options	The option flags

SWIG Wrapper Notes This function is attached as method hc_add_from_db() to objects of type fold_compound

16.13 Soft Constraints

Functions and data structures for secondary structure soft constraints.

16.13.1 Detailed Description

Functions and data structures for secondary structure soft constraints.

Soft-constraints are used to change position specific contributions in the recursions by adding bonuses/penalties in form of pseudo free energies to certain loop configurations. Collaboration diagram for Soft Constraints:

Files

· file soft.h

Functions and data structures for secondary structure soft constraints.

Data Structures

• struct vrna sc s

The soft constraints data structure. More...

Typedefs

- typedef struct vrna_sc_s vrna_sc_t
 - Typename for the soft constraints data structure vrna_sc_s.
- typedef int() vrna callback sc energy(int i, int j, int k, int l, unsigned char d, void *data)
 - Callback to retrieve pseudo energy contribution for soft constraint feature.
- typedef FLT_OR_DBL() vrna_callback_sc_exp_energy(int i, int j, int k, int l, unsigned char d, void *data)

 Callback to retrieve pseudo energy contribution as Boltzmann Factors for soft constraint feature.
- typedef vrna_basepair_t *() vrna_callback_sc_backtrack(int i, int j, int k, int l, unsigned char d, void *data)

 Callback to retrieve auxiliary base pairs for soft constraint feature.

Functions

- void vrna_sc_init (vrna_fold_compound_t *vc)
 - Initialize an empty soft constraints data structure within a vrna_fold_compound_t.
- void vrna_sc_set_bp (vrna_fold_compound_t *vc, const FLT_OR_DBL **constraints, unsigned int options)
 Set soft constraints for paired nucleotides.
- void vrna_sc_add_bp (vrna_fold_compound_t *vc, int i, int j, FLT_OR_DBL energy, unsigned int options)

 Add soft constraints for paired nucleotides.
- void vrna_sc_set_up (vrna_fold_compound_t *vc, const FLT_OR_DBL *constraints, unsigned int options) Set soft constraints for unpaired nucleotides.
- void vrna_sc_add_up (vrna_fold_compound_t *vc, int i, FLT_OR_DBL energy, unsigned int options)

 Add soft constraints for unpaired nucleotides.
- void vrna sc remove (vrna fold compound t *vc)
 - Remove soft constraints from vrna_fold_compound_t.
- void vrna_sc_free (vrna_sc_t *sc)

16.13 Soft Constraints 267

Free memory occupied by a vrna_sc_t data structure.

void vrna_sc_add_data (vrna_fold_compound_t *vc, void *data, vrna_callback_free_auxdata *free_data)

Add an auxiliary data structure for the generic soft constraints callback function.

void vrna_sc_add_f (vrna_fold_compound_t *vc, vrna_callback_sc_energy *f)

Bind a function pointer for generic soft constraint feature (MFE version)

• void vrna sc add bt (vrna fold compound t *vc, vrna callback sc backtrack *f)

Bind a backtracking function pointer for generic soft constraint feature.

void vrna_sc_add_exp_f (vrna_fold_compound_t *vc, vrna_callback_sc_exp_energy *exp_f)

Bind a function pointer for generic soft constraint feature (PF version)

16.13.2 Data Structure Documentation

```
16.13.2.1 struct vrna_sc_s
```

The soft constraints data structure.

Collaboration diagram for vrna_sc_s:

Data Fields

int ** energy up

Energy contribution for stretches of unpaired nucleotides.

FLT OR DBL ** exp energy up

Boltzmann Factors of the energy contributions for unpaired sequence stretches.

• int * up_storage

Storage container for energy contributions per unpaired nucleotide.

vrna_sc_bp_storage_t ** bp_storage

Storage container for energy contributions per base pair.

int * energy_stack

Pseudo Energy contribution per base pair involved in a stack.

• FLT_OR_DBL * exp_energy_stack

Boltzmann weighted pseudo energy contribution per nucleotide involved in a stack.

vrna_callback_sc_energy * f

A function pointer used for pseudo energy contribution in MFE calculations.

· vrna callback sc backtrack * bt

A function pointer used to obtain backtraced base pairs in loop regions that were altered by soft constrained pseudo energy contributions.

vrna_callback_sc_exp_energy * exp_f

A function pointer used for pseudo energy contribution boltzmann factors in PF calculations.

void * data

A pointer to the data object provided for for pseudo energy contribution functions of the generic soft constraints feature.

int * energy_bp

Energy contribution for base pairs.

FLT OR DBL * exp energy bp

Boltzmann Factors of the energy contribution for base pairs.

int ** energy_bp_local

Energy contribution for base pairs (sliding window approach)

FLT_OR_DBL ** exp_energy_bp_local

Boltzmann Factors of the energy contribution for base pairs (sliding window approach)

```
16.13.2.1.1 Field Documentation
```

```
16.13.2.1.1.1 f
vrna_callback_sc_energy* vrna_sc_s::f
```

A function pointer used for pseudo energy contribution in MFE calculations.

```
See also
```

```
vrna_sc_add_f()
```

```
16.13.2.1.1.2 bt
```

```
vrna_callback_sc_backtrack* vrna_sc_s::bt
```

A function pointer used to obtain backtraced base pairs in loop regions that were altered by soft constrained pseudo energy contributions.

See also

```
vrna_sc_add_bt()
```

```
16.13.2.1.1.3 exp_f
```

 ${\tt vrna_callback_sc_exp_energy* \ vrna_sc_s::exp_f}$

A function pointer used for pseudo energy contribution boltzmann factors in PF calculations.

See also

```
vrna_sc_add_exp_f()
```

16.13.3 Typedef Documentation

16.13 Soft Constraints 269

16.13.3.1 vrna_callback_sc_energy

 ${\tt typedef\ int()\ vrna_callback_sc_energy(int\ i,\ int\ j,\ int\ k,\ int\ l,\ unsigned\ char\ d,\ void\ *data)}$

#include <ViennaRNA/constraints/soft.h>

Callback to retrieve pseudo energy contribution for soft constraint feature.

This is the prototype for callback functions used by the folding recursions to evaluate generic soft constraints. The first four parameters passed indicate the delimiting nucleotide positions of the decomposition, and the parameter denotes the decomposition step. The last parameter data is the auxiliary data structure associated to the hard constraints via vrna_sc_add_data(), or NULL if no auxiliary data was added.

Notes on Callback Functions This callback enables one to add (pseudo-)energy contributions to individual decompositions of the secondary structure.

See also

VRNA_DECOMP_PAIR_HP, VRNA_DECOMP_PAIR_IL, VRNA_DECOMP_PAIR_ML, VRNA_DECOMP_ML_ML_ML, VRNA_DECOMP_ML_STEM, VRNA_DECOMP_ML_UP, VRNA_DECOMP_ML_ML_STEM, VRNA_DECOMP_ML_COAXIAL, VRNA_DECOMP_EXT_EXT, VRNA_DECOMP_EXT_UP, VRNA_DECOMP_EXT_STEM, VRNA_DECOMP_EXT_EXT, VRNA_DECOMP_EXT_STEM, VRNA_DECOMP_EXT_STEM, VRNA_DECOMP_EXT_STEM1, vrna_sc_add_f(), vrna_sc_add_exp_f(), vrna_sc_add_bt(), vrna_sc_add_data()

Parameters

i	Left (5') delimiter position of substructure
j	Right (3') delimiter position of substructure
k	Left delimiter of decomposition
1	Right delimiter of decomposition
d	Decomposition step indicator
data	Auxiliary data

Returns

Pseudo energy contribution in deka-kalories per mol

16.13.3.2 vrna_callback_sc_exp_energy

```
typedef FLT_OR_DBL() vrna_callback_sc_exp_energy(int i, int j, int k, int l, unsigned char d,
void *data)
```

#include <ViennaRNA/constraints/soft.h>

Callback to retrieve pseudo energy contribution as Boltzmann Factors for soft constraint feature.

This is the prototype for callback functions used by the partition function recursions to evaluate generic soft constraints. The first four parameters passed indicate the delimiting nucleotide positions of the decomposition, and the parameter denotes the decomposition step. The last parameter data is the auxiliary data structure associated to the hard constraints via vrna_sc_add_data(), or NULL if no auxiliary data was added.

Notes on Callback Functions This callback enables one to add (pseudo-)energy contributions to individual decompositions of the secondary structure (Partition function variant, i.e. contributions must be returned as Boltzmann factors).

See also

VRNA_DECOMP_PAIR_HP, VRNA_DECOMP_PAIR_IL, VRNA_DECOMP_PAIR_ML, VRNA_DECOMP_ML_ML_ML, VRNA_DECOMP_ML_STEM, VRNA_DECOMP_ML_UP, VRNA_DECOMP_ML_UP, VRNA_DECOMP_ML_ML_STEM, VRNA_DECOMP_ML_COAXIAL, VRNA_DECOMP_EXT_EXT, VRNA_DECOMP_EXT_UP, VRNA_DECOMP_EXT_STEM, VRNA_DECOMP_EXT_EXT, VRNA_DECOMP_EXT_STEM, VRNA_DECOMP_EXT_EXT_STEM, VRNA_DECOMP_EXT_EXT_STEM1, vrna_sc_add_exp_f(), vrna_sc_add_f(), vrna_sc_add_bt(), vrna_sc_add_data()

Parameters

k	Left delimiter of decomposition
1	Right delimiter of decomposition
d	Decomposition step indicator
data	Auxiliary data

16.13 Soft Constraints 271

Returns

Pseudo energy contribution in deka-kalories per mol

16.13.3.3 vrna_callback_sc_backtrack

```
typedef vrna_basepair_t*() vrna_callback_sc_backtrack(int i, int j, int k, int l, unsigned
char d, void *data)
```

#include <ViennaRNA/constraints/soft.h>

Callback to retrieve auxiliary base pairs for soft constraint feature.

Notes on Callback Functions This callback enables one to add auxiliary base pairs in the backtracking steps of hairpin- and interior loops.

See also

VRNA_DECOMP_PAIR_HP, VRNA_DECOMP_PAIR_IL, VRNA_DECOMP_PAIR_ML, VRNA_DECOMP_ML_ML_ML, VRNA_DECOMP_ML_STEM, VRNA_DECOMP_ML_ML, VRNA_DECOMP_ML_UP, VRNA_DECOMP_ML_ML_STEM, VRNA_DECOMP_ML_COAXIAL, VRNA_DECOMP_EXT_EXT, VRNA_DECOMP_EXT_UP, VRNA_DECOMP_EXT_STEM, VRNA_DECOMP_EXT_EXT_STEM, VRNA_DECOMP_EXT_EXT_STEM, VRNA_DECOMP_EXT_EXT_STEM1, vrna_sc_add_bt(), vrna_sc_add_exp_f(), vrna_sc_add_exp_f(), vrna_sc_add_data()

Parameters

i	Left (5') delimiter position of substructure
j	Right (3') delimiter position of substructure
k	Left delimiter of decomposition
1	Right delimiter of decomposition
d	Decomposition step indicator
data	Auxiliary data

Returns

List of additional base pairs

16.13.4 Function Documentation

16.13.4.1 vrna_sc_init()

```
#include <ViennaRNA/constraints/soft.h>
```

Initialize an empty soft constraints data structure within a vrna_fold_compound_t.

This function adds a proper soft constraints data structure to the vrna_fold_compound_t data structure. If soft constraints already exist within the fold compound, they are removed.

Note

Accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE and VRNA_FC_TYPE_COMPARATIVE

See also

```
vrna_sc_set_bp(), vrna_sc_set_up(), vrna_sc_add_SHAPE_deigan(), vrna_sc_add_SHAPE_zarringhalam(), vrna_sc_remove(), vrna_sc_add_f(), vrna_sc_add_exp_f(), vrna_sc_add_pre(), vrna_sc_add_pre(), vrna_sc_add_pre()
```

Parameters

```
vc The vrna_fold_compound_t where an empty soft constraint feature is to be added to
```

SWIG Wrapper Notes This function is attached as method sc_init() to objects of type fold_compound

```
16.13.4.2 vrna_sc_set_bp()
```

#include <ViennaRNA/constraints/soft.h>

Set soft constraints for paired nucleotides.

Note

 $\label{thm:constraints} \textbf{This function replaces any pre-exisiting soft constraints with the ones supplied in \verb|constraints||.}$

See also

```
vrna_sc_add_bp(), vrna_sc_set_up(), vrna_sc_add_up()
```

Parameters

VC	The vrna_fold_compound_t the soft constraints are associated with
constraints	A two-dimensional array of pseudo free energies in $kcal/mol$
options	The options flag indicating how/where to store the soft constraints

16.13 Soft Constraints 273

SWIG Wrapper Notes This function is attached as method sc_set_bp() to objects of type fold_compound

16.13.4.3 vrna_sc_add_bp()

#include <ViennaRNA/constraints/soft.h>

Add soft constraints for paired nucleotides.

See also

```
vrna_sc_set_bp(), vrna_sc_set_up(), vrna_sc_add_up()
```

Parameters

VC	The vrna_fold_compound_t the soft constraints are associated with
i	The 5' position of the base pair the soft constraint is added for
j	The 3' position of the base pair the soft constraint is added for
energy	The free energy (soft-constraint) in $kcal/mol$
options	The options flag indicating how/where to store the soft constraints

SWIG Wrapper Notes This function is attached as an overloaded method **sc_add_bp()** to objects of type *fold*← __*compound*. The method either takes arguments for a single base pair (i,j) with the corresponding energy value:

```
fold_compound.sc_add_bp(i, j, energy, options)
```

or an entire 2-dimensional matrix with dimensions n x n that stores free energy contributions for any base pair (i,j) with $1 \le i < j \le n$: fold_compound.sc_add_bp(matrix, options)

In both variants, the options argument is optional can may be omitted.

16.13.4.4 vrna_sc_set_up()

#include <ViennaRNA/constraints/soft.h>

Set soft constraints for unpaired nucleotides.

Note

This function replaces any pre-exisitng soft constraints with the ones supplied in constraints.

See also

```
vrna_sc_add_up(), vrna_sc_set_bp(), vrna_sc_add_bp()
```

Parameters

VC	The vrna_fold_compound_t the soft constraints are associated with
constraints	A vector of pseudo free energies in $kcal/mol$
options	The options flag indicating how/where to store the soft constraints

SWIG Wrapper Notes This function is attached as method sc_set_up() to objects of type fold_compound

16.13.4.5 vrna_sc_add_up()

#include <ViennaRNA/constraints/soft.h>

Add soft constraints for unpaired nucleotides.

See also

```
vrna_sc_set_up(), vrna_sc_add_bp(), vrna_sc_set_bp()
```

Parameters

VC	The vrna_fold_compound_t the soft constraints are associated with
i	The nucleotide position the soft constraint is added for
energy	The free energy (soft-constraint) in $kcal/mol$
options	The options flag indicating how/where to store the soft constraints

SWIG Wrapper Notes This function is attached as an overloaded method $sc_add_up()$ to objects of type $fold \leftarrow _compound$. The method either takes arguments for a single nucleotide i with the corresponding energy value:

```
fold_compound.sc_add_up(i, energy, options)
```

or an entire vector that stores free energy contributions for each nucleotide i with $1 \leq i \leq n$: fold_compound.sc_add_bp(vector, options)

In both variants, the ${\tt options}$ argument is optional can may be omitted.

16.13 Soft Constraints 275

```
16.13.4.6 vrna_sc_remove()
```

Remove soft constraints from vrna fold compound t.

Note

Accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE and VRNA_FC_TYPE_COMPARATIVE

Parameters

vc The vrna_fold_compound_t possibly containing soft constraints

SWIG Wrapper Notes This function is attached as method sc_remove() to objects of type fold_compound

```
16.13.4.7 vrna_sc_free()
```

Free memory occupied by a vrna_sc_t data structure.

Parameters

sc The data structure to free from memory

16.13.4.8 vrna_sc_add_data()

Add an auxiliary data structure for the generic soft constraints callback function.

See also

```
vrna_sc_add_f(), vrna_sc_add_exp_f(), vrna_sc_add_bt()
```

Parameters

VC	The fold compound the generic soft constraint function should be bound to
data	A pointer to the data structure that holds required data for function 'f'
free_data	A pointer to a function that free's the memory occupied by data (Maybe NULL)

SWIG Wrapper Notes This function is attached as method sc add data() to objects of type fold compound

Bind a function pointer for generic soft constraint feature (MFE version)

This function allows one to easily bind a function pointer and corresponding data structure to the soft constraint part $vrna_sc_t$ of the $vrna_fold_compound_t$. The function for evaluating the generic soft constraint feature has to return a pseudo free energy \hat{E} in dacal/mol, where 1dacal/mol = 10cal/mol.

See also

```
vrna_sc_add_data(), vrna_sc_add_bt(), vrna_sc_add_exp_f()
```

Parameters

VC	The fold compound the generic soft constraint function should be bound to
f	A pointer to the function that evaluates the generic soft constraint feature

SWIG Wrapper Notes This function is attached as method sc_add_f() to objects of type fold_compound

Bind a backtracking function pointer for generic soft constraint feature.

This function allows one to easily bind a function pointer to the soft constraint part vrna_sc_t of the vrna_fold_compound_t. The provided function should be used for backtracking purposes in loop regions that were altered via the generic soft constraint feature. It has to return an array of vrna_basepair_t data structures, were the last element in the list is indicated by a value of -1 in it's i position.

16.13 Soft Constraints 277

See also

```
vrna_sc_add_data(), vrna_sc_add_f(), vrna_sc_add_exp_f()
```

Parameters

VC	The fold compound the generic soft constraint function should be bound to
f	A pointer to the function that returns additional base pairs

SWIG Wrapper Notes This function is attached as method sc_add_bt() to objects of type fold_compound

Bind a function pointer for generic soft constraint feature (PF version)

This function allows one to easily bind a function pointer and corresponding data structure to the soft constraint part $vrna_sc_t$ of the $vrna_fold_compound_t$. The function for evaluating the generic soft constraint feature has to return a pseudo free energy \hat{E} as Boltzmann factor, i.e. $exp(-\hat{E}/kT)$. The required unit for E is cal/mol.

See also

```
vrna_sc_add_bt(), vrna_sc_add_f(), vrna_sc_add_data()
```

Parameters

VC	The fold compound the generic soft constraint function should be bound to
ехр⊷	A pointer to the function that evaluates the generic soft constraint feature
_f	

SWIG Wrapper Notes This function is attached as method sc_add_exp_f() to objects of type fold_compound

16.14 The RNA Secondary Structure Landscape

16.14.1 Detailed Description

Collaboration diagram for The RNA Secondary Structure Landscape:

Modules

Neighborhood Relation and Move Sets for Secondary Structures
 Different functions to generate structural neighbors of a secondary structure according to a particular Move Set.

• (Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima API for various RNA folding path algorithms.

16.15 Minimum Free Energy (MFE) Algorithms

Predicting the Minimum Free Energy (MFE) and a corresponding (consensus) secondary structure.

16.15.1 Detailed Description

Predicting the Minimum Free Energy (MFE) and a corresponding (consensus) secondary structure.

In a nutshell we provide two different flavors for MFE prediction:

- Global MFE Prediction to compute the MFE for the entire sequence
- · Local (sliding window) MFE Prediction to compute MFEs for each window using a sliding window approach

Each of these flavors, again, provides two implementations to either compute the MFE based on

- · single RNA (DNA) sequence(s), or
- a comparative approach using multiple sequence alignments (MSA).

For the latter, a consensus secondary structure is predicted and our implementations compute an average of free energies for each sequence in the MSA plus an additional covariance pseudo-energy term.

The implementations for Backtracking MFE structures are generally agnostic with respect to whether local or global structure prediction is in place. Collaboration diagram for Minimum Free Energy (MFE) Algorithms:

Modules

• Global MFE Prediction

Variations of the global Minimum Free Energy (MFE) prediction algorithm.

· Local (sliding window) MFE Prediction

Variations of the local (sliding window) Minimum Free Energy (MFE) prediction algorithm.

Backtracking MFE structures

Backtracking related interfaces.

Files

• file mfe.h

Compute Minimum Free energy (MFE) and backtrace corresponding secondary structures from RNA sequence data.

· file mfe_window.h

Compute local Minimum Free Energy (MFE) using a sliding window approach and backtrace corresponding secondary structures.

16.16 Partition Function and Equilibrium Properties

Compute the partition function to assess various equilibrium properties.

16.16.1 Detailed Description

Compute the partition function to assess various equilibrium properties.

Similar to our Minimum Free Energy (MFE) Algorithms, we provide two different flavors for partition function computations:

- Global Partition Function and Equilibrium Probabilities to compute the partition function for a full length sequence
- Local (sliding window) Partition Function and Equilibrium Probabilities to compute the partition function of each window using a sliding window approach

While the global partition function approach supports predictions using single sequences as well as consensus partition functions for multiple sequence alignments (MSA), we currently do not support MSA input for the local variant.

Comparative prediction computes an average of the free energy contributions plus an additional covariance pseudoenergy term, exactly as we do for the Minimum Free Energy (MFE) Algorithms implementation.

Boltzmann weights for the free energy contributions of individual loops can be found in Energy Evaluation for Individual Loops.

Our implementations also provide a stochastic backtracking procedure to draw Random Structure Samples from the Ensemble according to their equilibrium probabilty. Collaboration diagram for Partition Function and Equilibrium Properties:

Modules

· Global Partition Function and Equilibrium Probabilities

Variations of the global partition function algorithm.

· Local (sliding window) Partition Function and Equilibrium Probabilities

Scanning version using a sliding window approach to compute equilibrium probabilities.

Files

· file concentrations.h

Concentration computations for RNA-RNA interactions.

• file equilibrium_probs.h

Equilibrium Probability implementations.

· file part func.h

Partition function implementations.

• file part_func_window.h

Partition function and equilibrium probability implementation for the sliding window algorithm.

Functions

• int vrna_pf_float_precision (void)

Find out whether partition function computations are using single precision floating points.

16.16.2 Function Documentation

16.16.2.1 vrna_pf_float_precision()

Find out whether partition function computations are using single precision floating points.

See also

```
FLT_OR_DBL
```

Returns

1 if single precision is used, 0 otherwise

16.17 Global MFE Prediction

Variations of the global Minimum Free Energy (MFE) prediction algorithm.

16.17.1 Detailed Description

Variations of the global Minimum Free Energy (MFE) prediction algorithm.

We provide implementations of the global MFE prediction algorithm for

- · Single sequences,
- · Multiple sequence alignments (MSA), and
- · RNA-RNA hybrids

Collaboration diagram for Global MFE Prediction:

Modules

Computing MFE representatives of a Distance Based Partitioning

Compute the minimum free energy (MFE) and secondary structures for a partitioning of the secondary structure space according to the base pair distance to two fixed reference structures basepair distance to two fixed reference structures.

Deprecated Interface for Global MFE Prediction

Files

· file mfe.h

Compute Minimum Free energy (MFE) and backtrace corresponding secondary structures from RNA sequence data.

Basic global MFE prediction interface

float vrna_mfe (vrna_fold_compound_t *vc, char *structure)

Compute minimum free energy and an appropriate secondary structure of an RNA sequence, or RNA sequence alignment.

• float vrna_mfe_dimer (vrna_fold_compound_t *vc, char *structure)

Compute the minimum free energy of two interacting RNA molecules.

Simplified global MFE prediction using sequence(s) or multiple sequence alignment(s)

float vrna_fold (const char *sequence, char *structure)

Compute Minimum Free Energy (MFE), and a corresponding secondary structure for an RNA sequence.

float vrna_circfold (const char *sequence, char *structure)

Compute Minimum Free Energy (MFE), and a corresponding secondary structure for a circular RNA sequence.

float vrna alifold (const char **sequences, char *structure)

Compute Minimum Free Energy (MFE), and a corresponding consensus secondary structure for an RNA sequence alignment using a comparative method.

float vrna_circalifold (const char **sequences, char *structure)

Compute Minimum Free Energy (MFE), and a corresponding consensus secondary structure for a sequence alignment of circular RNAs using a comparative method.

float vrna cofold (const char *sequence, char *structure)

Compute Minimum Free Energy (MFE), and a corresponding secondary structure for two dimerized RNA sequences.

16.17.2 Function Documentation

16.17.2.1 vrna_mfe()

Compute minimum free energy and an appropriate secondary structure of an RNA sequence, or RNA sequence alignment.

Depending on the type of the provided $vrna_fold_compound_t$, this function predicts the MFE for a single sequence, or a corresponding averaged MFE for a sequence alignment. If backtracking is activated, it also constructs the corresponding secondary structure, or consensus structure. Therefore, the second parameter, structure, has to point to an allocated block of memory with a size of at least strlen(sequence) + 1 to store the backtracked MFE structure. (For consensus structures, this is the length of the alignment + 1. If NULL is passed, no backtracking will be performed.

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

See also

vrna_fold_compound_t, vrna_fold_compound(), vrna_fold(), vrna_circfold(), vrna_fold_compound_comparative(),
vrna_alifold(), vrna_circalifold()

Parameters

VC	fold compound
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to (Maybe NULL)

Returns

the minimum free energy (MFE) in kcal/mol

SWIG Wrapper Notes This function is attached as method mfe() to objects of type fold_compound

16.17.2.2 vrna_mfe_dimer()

```
#include <ViennaRNA/mfe.h>
```

Compute the minimum free energy of two interacting RNA molecules.

The code is analog to the vrna_mfe() function.

Parameters

VC	fold compound
structure	Will hold the barcket dot structure of the dimer molecule

Returns

minimum free energy of the structure

SWIG Wrapper Notes This function is attached as method mfe_dimer() to objects of type fold_compound

16.17.2.3 vrna_fold()

#include <ViennaRNA/mfe.h>

Compute Minimum Free Energy (MFE), and a corresponding secondary structure for an RNA sequence.

This simplified interface to vrna_mfe() computes the MFE and, if required, a secondary structure for an RNA sequence using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing, e.g. suboptimal backtracking, etc.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_mfe(), and the data structure vrna_mfe(), and the data structure vrna_mfe().

See also

vrna_circfold(), vrna_mfe()

Parameters

sequence	RNA sequence
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to

Returns

the minimum free energy (MFE) in kcal/mol

16.17.2.4 vrna_circfold()

#include <ViennaRNA/mfe.h>

Compute Minimum Free Energy (MFE), and a corresponding secondary structure for a circular RNA sequence.

This simplified interface to vrna_mfe() computes the MFE and, if required, a secondary structure for a circular RNA sequence using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing, e.g. suboptimal backtracking, etc.

Folding of circular RNA sequences is handled as a post-processing step of the forward recursions. See [12] for further details.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_mfe(), and the data structure vrna_mfe(), and the data structure vrna_mfe().

See also

```
vrna_fold(), vrna_mfe()
```

Parameters

sequence	RNA sequence
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to

Returns

the minimum free energy (MFE) in kcal/mol

16.17.2.5 vrna_alifold()

```
#include <ViennaRNA/mfe.h>
```

Compute Minimum Free Energy (MFE), and a corresponding consensus secondary structure for an RNA sequence alignment using a comparative method.

This simplified interface to vrna_mfe() computes the MFE and, if required, a consensus secondary structure for an RNA sequence alignment using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing, e.g. suboptimal backtracking, etc.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_mfe(), and the data structure vrna_fold_compound t instead.

See also

```
vrna_circalifold(), vrna_mfe()
```

Parameters

sequences	RNA sequence alignment
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to

Returns

the minimum free energy (MFE) in kcal/mol

16.17.2.6 vrna_circalifold()

```
float vrna_circalifold (

const char ** sequences,

char * structure )
```

#include <ViennaRNA/mfe.h>

Compute Minimum Free Energy (MFE), and a corresponding consensus secondary structure for a sequence alignment of circular RNAs using a comparative method.

This simplified interface to vrna_mfe() computes the MFE and, if required, a consensus secondary structure for an RNA sequence alignment using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing, e.g. suboptimal backtracking, etc.

Folding of circular RNA sequences is handled as a post-processing step of the forward recursions. See [12] for further details.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_mfe(), and the data structure vrna_mfe(), and the data structure vrna_fold_compound_tinstead.

See also

```
vrna alifold(), vrna mfe()
```

Parameters

sequences	Sequence alignment of circular RNAs
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to

Returns

the minimum free energy (MFE) in kcal/mol

16.17.2.7 vrna_cofold()

#include <ViennaRNA/mfe.h>

Compute Minimum Free Energy (MFE), and a corresponding secondary structure for two dimerized RNA sequences.

This simplified interface to vrna_mfe() computes the MFE and, if required, a secondary structure for two RNA sequences upon dimerization using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing, e.g. suboptimal backtracking, etc.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_mfe(), and the data structure vrna_mfe(), and the data structure vrna_fold_compound_tinstead.

See also

```
vrna_mfe_dimer(), vrna_fold_compound(), vrna_fold_compound_t, vrna_cut_point_insert()
```

Parameters

	sequence	two RNA sequences separated by the '&' character
ſ	structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to

Returns

the minimum free energy (MFE) in kcal/mol

16.18 Local (sliding window) MFE Prediction

Variations of the local (sliding window) Minimum Free Energy (MFE) prediction algorithm.

16.18.1 Detailed Description

Variations of the local (sliding window) Minimum Free Energy (MFE) prediction algorithm.

We provide implementations for the local (sliding window) MFE prediction algorithm for

- · Single sequences,
- · Multiple sequence alignments (MSA), and

Note, that our implementation scans an RNA sequence (or MSA) from the 3' to the 5' end, and reports back locally optimal (consensus) structures, the corresponding free energy, and the position of the sliding window in global coordinates.

For any particular RNA sequence (or MSA) multiple locally optimal (consensus) secondary structures may be predicted. Thus, we tried to implement an interface that allows for an effortless conversion of the corresponding hits into any target data structure. As a consequence, we provide two distinct ways to retrieve the corresponding predictions, either

- through directly writing to an open FILE stream on-the-fly, or
- through a callback function mechanism.

The latter allows one to store the results in any possible target data structure. Our implementations then pass the results through the user-implemented callback as soon as the prediction for a particular window is finished. Collaboration diagram for Local (sliding window) MFE Prediction:

Modules

• Deprecated Interface for Local (Sliding Window) MFE Prediction

Files

• file mfe_window.h

Compute local Minimum Free Energy (MFE) using a sliding window approach and backtrace corresponding secondary structures

Typedefs

• typedef void() vrna_mfe_window_callback(int start, int end, const char *structure, float en, void *data)

The default callback for sliding window MFE structure predictions.

Basic local (sliding window) MFE prediction interface

• float vrna_mfe_window (vrna_fold_compound_t *vc, FILE *file)

Local MFE prediction using a sliding window approach.

- float vrna_mfe_window_cb (vrna_fold_compound_t *vc, vrna_mfe_window_callback *cb, void *data)
- float vrna_mfe_window_zscore (vrna_fold_compound_t *vc, double min_z, FILE *file)

Local MFE prediction using a sliding window approach (with z-score cut-off)

float vrna_mfe_window_zscore_cb (vrna_fold_compound_t *vc, double min_z, vrna_mfe_window_
 zscore_callback *cb, void *data)

Simplified local MFE prediction using sequence(s) or multiple sequence alignment(s)

- float vrna_Lfold (const char *string, int window_size, FILE *file)
 Local MFE prediction using a sliding window approach (simplified interface)
- float vrna_Lfold_cb (const char *string, int window_size, vrna_mfe_window_callback *cb, void *data)
- float vrna_Lfoldz (const char *string, int window_size, double min_z, FILE *file)
 Local MFE prediction using a sliding window approach with z-score cut-off (simplified interface)
- float **vrna_Lfoldz_cb** (const char *string, int window_size, double min_z, vrna_mfe_window_zscore_← callback *cb, void *data)
- float vrna_aliLfold (const char **alignment, int maxdist, FILE *fp)
- float vrna aliLfold cb (const char **alignment, int maxdist, vrna mfe window callback *cb, void *data)

16.18.2 Typedef Documentation

16.18.2.1 vrna_mfe_window_callback

typedef void() vrna_mfe_window_callback(int start, int end, const char *structure, float en, void *data)

```
#include <ViennaRNA/mfe_window.h>
```

The default callback for sliding window MFE structure predictions.

Notes on Callback Functions This function will be called for each hit in a sliding window MFE prediction.

See also

vrna_mfe_window()

Parameters

start	provides the first position of the hit (1-based, relative to entire sequence/alignment)
end	provides the last position of the hit (1-based, relative to the entire sequence/alignment)
structure	provides the (sub)structure in dot-bracket notation
en	is the free energy of the structure hit in kcal/mol
data	is some arbitrary data pointer passed through by the function executing the callback

16.18.3 Function Documentation

16.18.3.1 vrna_mfe_window()

FILE * file)

```
#include <ViennaRNA/mfe window.h>
```

Local MFE prediction using a sliding window approach.

Computes minimum free energy structures using a sliding window approach, where base pairs may not span outside the window. In contrast to vrna_mfe(), where a maximum base pair span may be set using the vrna_md_t.max_bp_span attribute and one globally optimal structure is predicted, this function uses a sliding window to retrieve all locally optimal structures within each window. The size of the sliding window is set in the vrna_md_t.window_size attribute, prior to the retrieval of the vrna_fold_compound_t using vrna_fold_compound() with option VRNA_OPTION_WINDOW

The predicted structures are written on-the-fly, either to stdout, if a NULL pointer is passed as file parameter, or to the corresponding filehandle.

See also

 $\label{lem:compound} vrna_fold_compound(), vrna_mfe_window_zscore(), vrna_mfe(), vrna_Lfold(), vrna_Lfoldz(), VRNA_OPTION_WINDOW, vrna_md_t.max_bp_span, vrna_md_t.window_size$

Parameters

VC	The vrna_fold_compound_t with preallocated memory for the DP matrices
file	The output file handle where predictions are written to (maybe NULL)

SWIG Wrapper Notes This function is attached as method mfe_window() to objects of type fold_compound

16.18.3.2 vrna_mfe_window_zscore()

#include <ViennaRNA/mfe_window.h>

Local MFE prediction using a sliding window approach (with z-score cut-off)

Computes minimum free energy structures using a sliding window approach, where base pairs may not span outside the window. This function is the z-score version of vrna_mfe_window(), i.e. only predictions above a certain z-score cut-off value are printed. As for vrna_mfe_window(), the size of the sliding window is set in the vrna_fold_compound() with option VRNA_OPTION_WINDOW.

The predicted structures are written on-the-fly, either to stdout, if a NULL pointer is passed as file parameter, or to the corresponding filehandle.

See also

vrna_fold_compound(), vrna_mfe_window_zscore(), vrna_mfe(), vrna_Lfold(), vrna_Lfoldz(), VRNA_OPTION_WINDOW, vrna_md_t.max_bp_span, vrna_md_t.window_size

Parameters

VC	The vrna_fold_compound_t with preallocated memory for the DP matrices
min⊷	The minimal z-score for a predicted structure to appear in the output
_Z	
file	The output file handle where predictions are written to (maybe NULL)

16.18.3.3 vrna_Lfold()

Local MFE prediction using a sliding window approach (simplified interface)

This simplified interface to vrna_mfe_window() computes the MFE and locally optimal secondary structure using default options. Structures are predicted using a sliding window approach, where base pairs may not span outside the window. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_mfe_window(), and the data structure vrna_fold_compound_tinstead.

See also

```
vrna_mfe_window(), vrna_Lfoldz(), vrna_mfe_window_zscore()
```

Parameters

string	The nucleic acid sequence	
_window_size	The window size for locally optimal structures	<u> </u>
file	The output file handle where predictions are written to (if NULL, output is writter មេខាន់។) Do	xygen

16.18.3.4 vrna_Lfoldz()

#include <ViennaRNA/mfe_window.h>

Local MFE prediction using a sliding window approach with z-score cut-off (simplified interface)

This simplified interface to vrna_mfe_window_zscore() computes the MFE and locally optimal secondary structure using default options. Structures are predicted using a sliding window approach, where base pairs may not span outside the window. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing. This function is the z-score version of vrna_Lfold(), i.e. only predictions above a certain z-score cut-off value are printed.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_mfe_window(), and the data structure vrna_mfe_window(), and the data structure vrna_fold_compound_tinstead.

See also

vrna_mfe_window_zscore(), vrna_Lfold(), vrna_mfe_window()

Parameters

string	The nucleic acid sequence
window_size	The window size for locally optimal structures
min_z	The minimal z-score for a predicted structure to appear in the output
file	The output file handle where predictions are written to (if NULL, output is written to stdout)

16.19 Backtracking MFE structures

Backtracking related interfaces.

16.19.1 Detailed Description

Backtracking related interfaces.

Collaboration diagram for Backtracking MFE structures:

Functions

- float vrna_backtrack5 (vrna_fold_compound_t *fc, unsigned int length, char *structure)

 Backtrack an MFE (sub)structure.
- int vrna_BT_hp_loop (vrna_fold_compound_t *fc, int i, int j, int en, vrna_bp_stack_t *bp_stack, int *stack_
 count)

Backtrack a hairpin loop closed by (i, j).

int vrna_BT_stack (vrna_fold_compound_t *fc, int *i, int *j, int *en, vrna_bp_stack_t *bp_stack, int *stack
count)

Backtrack a stacked pair closed by (i, j).

int vrna_BT_int_loop (vrna_fold_compound_t *fc, int *i, int *j, int en, vrna_bp_stack_t *bp_stack, int *stack← count)

Backtrack an interior loop closed by (i, j).

• int vrna_BT_mb_loop (vrna_fold_compound_t *fc, int *i, int *j, int *k, int en, int *component1, int *component2)

Backtrack the decomposition of a multi branch loop closed by (i, j).

16.19.2 Function Documentation

16.19.2.1 vrna_backtrack5()

Backtrack an MFE (sub)structure.

This function allows one to backtrack the MFE structure for a (sub)sequence

Note

On error, the function returns INF / 100. and stores the empty string in structure.

Precondition

Requires pre-filled MFE dynamic programming matrices, i.e. one has to call vrna_mfe() prior to calling this function

See also

```
vrna_mfe(), vrna_pbacktrack5()
```

Parameters

fc	fold compound
length	The length of the subsequence, starting from the 5' end
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to. (Must have size of at least \$p length + 1)

Returns

The minimum free energy (MFE) for the specified length in kcal/mol and a corresponding secondary structure in dot-bracket notation (stored in structure)

SWIG Wrapper Notes This function is attached as overloaded method **backtrack()** to objects of type *fold_← compound* with default parameter length equal to the total length of the RNA.

16.19.2.2 vrna_BT_hp_loop()

```
int vrna_BT_hp_loop (
    vrna_fold_compound_t * fc,
    int i,
    int j,
    int en,
    vrna_bp_stack_t * bp_stack,
    int * stack_count )
```

#include <ViennaRNA/loops/hairpin.h>

Backtrack a hairpin loop closed by (i, j).

Note

This function is polymorphic! The provided vrna_fold_compound_t may be of type VRNA_FC_TYPE_SINGLE or VRNA_FC_TYPE_COMPARATIVE

16.19.2.3 vrna_BT_stack()

Backtrack a stacked pair closed by (i, j).

16.19.2.4 vrna_BT_int_loop()

#include <ViennaRNA/loops/internal.h>

Backtrack an interior loop closed by (i, j).

16.19.2.5 vrna_BT_mb_loop()

#include <ViennaRNA/loops/multibranch.h>

Backtrack the decomposition of a multi branch loop closed by (i, j).

Parameters

fc	The vrna_fold_compound_t filled with all relevant data for backtracking
i	5' position of base pair closing the loop (will be set to 5' position of leftmost decomposed block upon successful backtracking)
j	3' position of base pair closing the loop (will be set to 3' position of rightmost decomposed block upon successful backtracking)
k	Split position that delimits leftmost from rightmost block, [i,k] and [k+1, j], respectively. (Will be set upon successful backtracking)
en	The energy contribution of the substructure enclosed by $\left(i,j\right)$
component1	Type of leftmost block (1 = ML, 2 = C)
component2	Type of rightmost block (1 = ML, 2 = C)

Returns

1, if backtracking succeeded, 0 otherwise.

16.20 Global Partition Function and Equilibrium Probabilities

Variations of the global partition function algorithm.

16.20.1 Detailed Description

Variations of the global partition function algorithm.

We provide implementations of the global partition function algorithm for

- · Single sequences,
- · Multiple sequence alignments (MSA), and
- · RNA-RNA hybrids

Collaboration diagram for Global Partition Function and Equilibrium Probabilities:

Modules

Computing Partition Functions of a Distance Based Partitioning

Compute the partition function and stochastically sample secondary structures for a partitioning of the secondary structure space according to the base pair distance to two fixed reference structures.

Deprecated Interface for Global Partition Function Computation

Files

· file part func.h

Partition function implementations.

Data Structures

struct vrna_dimer_pf_s

Data structure returned by vrna_pf_dimer() More...

Functions

• void vrna_pf_dimer_probs (double FAB, double FA, double FB, vrna_ep_t *prAB, const vrna_ep_t *prA, const vrna_ep_t *prB, int Alength, const vrna_exp_param_t *exp_params)

Compute Boltzmann probabilities of dimerization without homodimers.

double vrna_pr_structure (vrna_fold_compound_t *fc, const char *structure)

Compute the equilibrium probability of a particular secondary structure.

vrna_ep_t * vrna_plist_from_probs (vrna_fold_compound_t *vc, double cut_off)

Create a vrna_ep_t from base pair probability matrix.

Base pair related probability computations

double vrna_mean_bp_distance_pr (int length, FLT_OR_DBL *pr)

Get the mean base pair distance in the thermodynamic ensemble from a probability matrix.

double vrna mean bp distance (vrna fold compound t *vc)

Get the mean base pair distance in the thermodynamic ensemble.

• double vrna_ensemble_defect (vrna_fold_compound_t *fc, const char *structure)

Compute the Ensemble Defect for a given target structure.

vrna_ep_t * vrna_stack_prob (vrna_fold_compound_t *vc, double cutoff)

Compute stacking probabilities.

double * vrna_positional_entropy (vrna_fold_compound_t *fc)

Compute a vector of positional entropies.

Basic global partition function interface

float vrna pf (vrna fold compound t *vc, char *structure)

Compute the partition function Q for a given RNA sequence, or sequence alignment.

vrna_dimer_pf_t vrna_pf_dimer (vrna_fold_compound_t *vc, char *structure)

Calculate partition function and base pair probabilities of nucleic acid/nucleic acid dimers.

Simplified global partition function computation using sequence(s) or multiple sequence alignment(s)

float vrna_pf_fold (const char *sequence, char *structure, vrna_ep_t **pl)

Compute Partition function Q (and base pair probabilities) for an RNA sequence using a comparative method.

• float vrna_pf_circfold (const char *sequence, char *structure, vrna_ep_t **pl)

Compute Partition function Q (and base pair probabilities) for a circular RNA sequences using a comparative method.

float vrna_pf_alifold (const char **sequences, char *structure, vrna_ep_t **pl)

Compute Partition function Q (and base pair probabilities) for an RNA sequence alignment using a comparative method

• float vrna_pf_circalifold (const char **sequences, char *structure, vrna_ep_t **pl)

Compute Partition function Q (and base pair probabilities) for an alignment of circular RNA sequences using a comparative method.

• vrna_dimer_pf_t vrna_pf_co_fold (const char *seq, char *structure, vrna_ep_t **pl)

Calculate partition function and base pair probabilities of nucleic acid/nucleic acid dimers.

16.20.2 Data Structure Documentation

16.20.2.1 struct vrna_dimer_pf_s

Data structure returned by vrna_pf_dimer()

Data Fields

double F0AB

Null model without DuplexInit.

· double FAB

all states with DuplexInit correction

· double FcAB

true hybrid states only

· double FA

monomer A

· double FB

monomer B

16.20.3 Function Documentation

16.20.3.1 vrna_mean_bp_distance_pr()

```
double vrna_mean_bp_distance_pr (
    int length,
    FLT_OR_DBL * pr )
```

#include <ViennaRNA/equilibrium_probs.h>

Get the mean base pair distance in the thermodynamic ensemble from a probability matrix.

$$\langle d \rangle = \sum_{a,b} p_a p_b d(S_a, S_b)$$

this can be computed from the pair probs p_{ij} as

$$< d > = \sum_{ij} p_{ij} (1 - p_{ij})$$

Parameters

length	The length of the sequence
pr	The matrix containing the base pair probabilities

Returns

The mean pair distance of the structure ensemble

16.20.3.2 vrna_mean_bp_distance()

#include <ViennaRNA/equilibrium_probs.h>

Get the mean base pair distance in the thermodynamic ensemble.

$$\langle d \rangle = \sum_{a,b} p_a p_b d(S_a, S_b)$$

this can be computed from the pair probs p_{ij} as

$$< d > = \sum_{ij} p_{ij} (1 - p_{ij})$$

Parameters

vc The fold compound data structure

Returns

The mean pair distance of the structure ensemble

SWIG Wrapper Notes This function is attached as method mean_bp_distance() to objects of type fold_compound

16.20.3.3 vrna_ensemble_defect()

#include <ViennaRNA/equilibrium_probs.h>

Compute the Ensemble Defect for a given target structure.

Given a target structure s, compute the average dissimilarity of a randomly drawn structure from the ensemble, i.e.:

$$ED(s) = 1 - \frac{1}{n} \sum_{ij,(i,j) \in s} p_{ij} - \frac{1}{n} \sum_{i} (1 - s_i) q_i$$

with sequence length n, the probability p_{ij} of a base pair (i,j), the probability $q_i=1-\sum_j p_{ij}$ of nucleotide i being unpaired, and the indicator variable $s_i=1$ if $\exists (i,j)\in s$, and $s_i=0$ otherwise.

Precondition

The vrna_fold_compound_t input parameter fc must contain a valid base pair probability matrix. This means that partition function and base pair probabilities must have been computed using fc before execution of this function!

See also

```
vrna_pf(), vrna_pairing_probs()
```

Parameters

fc	A fold_compound with pre-computed base pair probabilitie	
structure	A target structure in dot-bracket notation	

Returns

The ensemble defect with respect to the target structure, or -1. upon failure, e.g. pre-conditions are not met

SWIG Wrapper Notes This function is attached as method ensemble_defect() to objects of type fold_compound

16.20.3.4 vrna_stack_prob()

#include <ViennaRNA/equilibrium_probs.h>

Compute stacking probabilities.

For each possible base pair (i, j), compute the probability of a stack (i, j), (i + 1, j - 1).

Parameters

VC	The fold compound data structure with precomputed base pair probabilities
cutoff	A cutoff value that limits the output to stacks with $p > \text{cutoff}$.

Returns

A list of stacks with enclosing base pair (i,j) and probability p

16.20.3.5 vrna_pf_dimer_probs()

#include <ViennaRNA/equilibrium_probs.h>

Compute Boltzmann probabilities of dimerization without homodimers.

Given the pair probabilities and free energies (in the null model) for a dimer AB and the two constituent monomers A and B, compute the conditional pair probabilities given that a dimer AB actually forms. Null model pair probabilities are given as a list as produced by vrna plist from probs(), the dimer probabilities 'prAB' are modified in place.

Parameters

FAB	free energy of dimer AB
. ,	
FA	free energy of monomer A
FB	free energy of monomer B
prAB	pair probabilities for dimer
prA	pair probabilities monomer
Generated by Doxyg	_{en} pair probabilities monomer
Alength	Length of molecule A
exp_params	The precomputed Boltzmann factors

16.20.3.6 vrna pr structure()

#include <ViennaRNA/equilibrium_probs.h>

Compute the equilibrium probability of a particular secondary structure.

The probability p(s) of a particular secondary structure s can be computed as

$$p(s) = \frac{exp(-\beta E(s))}{Z}$$

from the structures free energy E(s) and the partition function

$$Z = \sum_{s} exp(-\beta E(s)), \text{ with } \beta = \frac{1}{RT}$$

where ${\cal R}$ is the gas constant and ${\cal T}$ the thermodynamic temperature.

Precondition

The fold compound fc must have went through a call to vrna_pf() to fill the dynamic programming matrices with the corresponding partition function.

Parameters

fc	The fold compound data structure with precomputed partition function
structure	The secondary structure to compute the probability for in dot-bracket notation

Returns

The probability of the input structure (range [0:1])

SWIG Wrapper Notes This function is attached as method pr structure() to objects of type fold compound

```
16.20.3.7 vrna_pf()
```

Compute the partition function Q for a given RNA sequence, or sequence alignment.

If structure is not a NULL pointer on input, it contains on return a string consisting of the letters "., | { } () " denoting bases that are essentially unpaired, weakly paired, strongly paired without preference, weakly upstream (downstream) paired, or strongly up- (down-)stream paired bases, respectively. If the model's compute_bpp is set to 0 base pairing probabilities will not be computed (saving CPU time), otherwise after calculations took place pr will contain the probability that bases i and j pair.

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

This function may return INF / 100. in case of contradicting constraints or numerical over-/underflow. In the latter case, a corresponding warning will be issued to stdout.

See also

vrna_fold_compound_t, vrna_fold_compound(), vrna_pf_fold(), vrna_pf_circfold(), vrna_fold_compound_comparative(), vrna_pf_alifold(), vrna_pf_circalifold(), vrna_db_from_probs(), vrna_exp_params(), vrna_aln_pinfo()

Parameters

in,out	VC	The fold compound data structure
in,out	structure	A pointer to the character array where position-wise pairing propensity will be stored.
		(Maybe NULL)

Returns

The ensemble free energy $G = -RT \cdot \log(Q)$ in kcal/mol

SWIG Wrapper Notes This function is attached as method pf() to objects of type fold_compound

16.20.3.8 vrna_pf_dimer()

Calculate partition function and base pair probabilities of nucleic acid/nucleic acid dimers.

This is the cofold partition function folding.

Note

This function may return INF / 100. for the FA, FB, FAB, FOAB members of the output data structure in case of contradicting constraints or numerical over-/underflow. In the latter case, a corresponding warning will be issued to stdout.

See also

vrna_fold_compound() for how to retrieve the necessary data structure

Parameters

VC	the fold compound data structure
structure	Will hold the structure or constraints

Returns

vrna_dimer_pf_t structure containing a set of energies needed for concentration computations.

SWIG Wrapper Notes This function is attached as method pf_dimer() to objects of type fold_compound

16.20.3.9 vrna_pf_fold()

#include <ViennaRNA/part_func.h>

Compute Partition function Q (and base pair probabilities) for an RNA sequence using a comparative method.

This simplified interface to vrna_pf() computes the partition function and, if required, base pair probabilities for an RNA sequence using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_pf(), and the data structure vrna_pf(), and the data structure vrna_pf(), and the data structure vrna_fold_compound_tinstead.

See also

```
vrna_pf_circfold(), vrna_pf(), vrna_fold_compound(), vrna_fold_compound_t
```

Parameters

sequence	RNA sequence	
structure	A pointer to the character array where position-wise pairing propensity will be stored. (Maybe NULL)	
pl	A pointer to a list of vrna_ep_t to store pairing probabilities (Maybe NULL)	

Returns

The ensemble free energy $G = -RT \cdot \log(Q)$ in kcal/mol

16.20.3.10 vrna_pf_circfold()

#include <ViennaRNA/part_func.h>

Compute Partition function ${\cal Q}$ (and base pair probabilities) for a circular RNA sequences using a comparative method.

This simplified interface to vrna_pf() computes the partition function and, if required, base pair probabilities for a circular RNA sequence using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_pf(), and the data structure vrna_pf(), and the data structure vrna_fold_compound_tinstead.

Folding of circular RNA sequences is handled as a post-processing step of the forward recursions. See [12] for further details.

See also

```
vrna_pf_fold(), vrna_pf(), vrna_fold_compound(), vrna_fold_compound_t
```

Parameters

sequence	A circular RNA sequence
structure	A pointer to the character array where position-wise pairing propensity will be stored. (Maybe NULL)
pl	A pointer to a list of vrna_ep_t to store pairing probabilities (Maybe NULL)

Returns

The ensemble free energy $G = -RT \cdot \log(Q)$ in kcal/mol

16.20.3.11 vrna_pf_alifold()

#include <ViennaRNA/part_func.h>

Compute Partition function Q (and base pair probabilities) for an RNA sequence alignment using a comparative method.

This simplified interface to vrna_pf() computes the partition function and, if required, base pair probabilities for an RNA sequence alignment using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_pf(), and the data structure vrna_pf(), and the data structure vrna_pf(), and the data structure vrna_fold_compound_tinstead.

See also

```
vrna_pf_circalifold(), vrna_pf(), vrna_fold_compound_comparative(), vrna_fold_compound_t
```

Parameters

sequences	RNA sequence alignment	
structure	A pointer to the character array where position-wise pairing propensity will be stored. (Maybe NULL)	
pl	A pointer to a list of vrna_ep_t to store pairing probabilities (Maybe NULL)	

Returns

The ensemble free energy $G = -RT \cdot \log(Q)$ in kcal/mol

16.20.3.12 vrna_pf_circalifold()

#include <ViennaRNA/part_func.h>

Compute Partition function ${\cal Q}$ (and base pair probabilities) for an alignment of circular RNA sequences using a comparative method.

This simplified interface to vrna_pf() computes the partition function and, if required, base pair probabilities for an RNA sequence alignment using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_pf(), and the data structure vrna_pf(), and the data structure vrna_fold_compound_tinstead.

Folding of circular RNA sequences is handled as a post-processing step of the forward recursions. See [12] for further details.

See also

vrna_pf_alifold(), vrna_pf(), vrna_fold_compound_comparative(), vrna_fold_compound_t

Parameters

ſ	sequences	Sequence alignment of circular RNAs
ſ	structure	A pointer to the character array where position-wise pairing propensity will be stored. (Maybe NULL)
Ī	pl	A pointer to a list of vrna_ep_t to store pairing probabilities (Maybe NULL)

Returns

The ensemble free energy $G = -RT \cdot \log(Q)$ in kcal/mol

16.20.3.13 vrna_plist_from_probs()

#include <ViennaRNA/utils/structures.h>

Create a vrna_ep_t from base pair probability matrix.

The probability matrix provided via the vrna_fold_compound_ is parsed and all pair probabilities above the given threshold are used to create an entry in the plist

The end of the plist is marked by sequence positions i as well as j equal to 0. This condition should be used to stop looping over its entries

Parameters

in	vc	The fold compound
in	cut_off	The cutoff value

Returns

A pointer to the plist that is to be created

16.20.3.14 vrna_positional_entropy()

#include <ViennaRNA/equilibrium_probs.h>

Compute a vector of positional entropies.

This function computes the positional entropies from base pair probabilities as

$$S(i) = -\sum_{i} p_{ij} \log(p_{ij}) - q_i \log(q_i)$$

with unpaired probabilities $q_i = 1 - \sum_{i} p_{ij}$.

Low entropy regions have little structural flexibility and the reliability of the predicted structure is high. High entropy implies many structural alternatives. While these alternatives may be functionally important, they make structure prediction more difficult and thus less reliable.

Precondition

This function requires pre-computed base pair probabilities! Thus, vrna pf() must be called beforehand.

Parameters

fc A fold_compound with pre-computed base pair probabilities

Returns

A 1-based vector of positional entropies S(i). (position 0 contains the sequence length)

SWIG Wrapper Notes This function is attached as method positional_entropy() to objects of type fold_compound

16.20.3.15 vrna_pf_co_fold()

#include <ViennaRNA/part_func.h>

Calculate partition function and base pair probabilities of nucleic acid/nucleic acid dimers.

This simplified interface to vrna_pf_dimer() computes the partition function and, if required, base pair probabilities for an RNA-RNA interaction using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_pf_dimer(), and the data structure vrna_fold_compound_tinstead.

See also

vrna_pf_dimer()

Parameters

seq	Two concatenated RNA sequences with a delimiting '&' in between
structure	A pointer to the character array where position-wise pairing propensity will be stored. (Maybe NULL)
pl	A pointer to a list of vrna_ep_t to store pairing probabilities (Maybe NULL)

Returns

vrna_dimer_pf_t structure containing a set of energies needed for concentration computations.

16.21 Local (sliding window) Partition Function and Equilibrium Probabilities

Scanning version using a sliding window approach to compute equilibrium probabilities.

16.21.1 Detailed Description

Scanning version using a sliding window approach to compute equilibrium probabilities.

Collaboration diagram for Local (sliding window) Partition Function and Equilibrium Probabilities:

Modules

• Deprecated Interface for Local (Sliding Window) Partition Function Computation

Files

• file part_func_window.h

Partition function and equilibrium probability implementation for the sliding window algorithm.

Macros

• #define VRNA_EXT_LOOP 1U

Exterior loop.

• #define VRNA_HP_LOOP 2U

Hairpin loop.

• #define VRNA_INT_LOOP 4U

Internal loop.

• #define VRNA MB LOOP 8U

Multibranch loop.

#define VRNA_ANY_LOOP (VRNA_EXT_LOOP | VRNA_HP_LOOP | VRNA_INT_LOOP | VRNA_MB_LOOP)
 Any loop.

#define VRNA_PROBS_WINDOW_BPP 4096U

Trigger base pairing probabilities.

#define VRNA_PROBS_WINDOW_UP 8192U

Trigger unpaired probabilities.

• #define VRNA_PROBS_WINDOW_STACKP 16384U

Trigger base pair stack probabilities.

#define VRNA_PROBS_WINDOW_UP_SPLIT 32768U

Trigger detailed unpaired probabilities split up into different loop type contexts.

• #define VRNA PROBS WINDOW PF 65536U

Trigger partition function.

Typedefs

typedef void() vrna_probs_window_callback(FLT_OR_DBL *pr, int pr_size, int i, int max, unsigned int type, void *data)

Sliding window probability computation callback.

Basic local partition function interface

• int vrna_probs_window (vrna_fold_compound_t *fc, int ulength, unsigned int options, vrna_probs_window_callback *cb, void *data)

Compute various equilibrium probabilities under a sliding window approach.

Simplified global partition function computation using sequence(s) or multiple sequence alignment(s)

- vrna_ep_t * vrna_pfl_fold (const char *sequence, int window_size, int max_bp_span, float cutoff)
 Compute base pair probabilities using a sliding-window approach.
- int vrna_pfl_fold_cb (const char *sequence, int window_size, int max_bp_span, vrna_probs_window_callback *cb, void *data)

Compute base pair probabilities using a sliding-window approach (callback version)

- double ** vrna_pfl_fold_up (const char *sequence, int ulength, int window_size, int max_bp_span)

 Compute probability of contiguous unpaired segments.
- int vrna_pfl_fold_up_cb (const char *sequence, int ulength, int window_size, int max_bp_span, vrna_probs_window_callback *cb, void *data)

Compute probability of contiguous unpaired segments.

16.21.2 Macro Definition Documentation

```
16.21.2.1 VRNA_PROBS_WINDOW_BPP
```

```
#define VRNA_PROBS_WINDOW_BPP 4096U
```

#include <ViennaRNA/part func window.h>

Trigger base pairing probabilities.

Passing this flag to vrna_probs_window() activates callback execution for base pairing probabilities. In turn, the corresponding callback receives this flag through the type argument whenever base pairing probabilities are provided.

Detailed information for the algorithm to compute unpaired probabilities can be taken from [3].

See also

vrna_probs_window()

16.21.2.2 VRNA_PROBS_WINDOW_UP

```
#define VRNA_PROBS_WINDOW_UP 8192U
#include <ViennaRNA/part_func_window.h>
```

Trigger unpaired probabilities.

Passing this flag to vrna_probs_window() activates callback execution for unpaired probabilities. In turn, the corresponding callback receives this flag through the type argument whenever unpaired probabilities are provided.

Detailed information for the algorithm to compute unpaired probabilities can be taken from [4].

See also

vrna_probs_window()

16.21.2.3 VRNA_PROBS_WINDOW_STACKP

```
#define VRNA_PROBS_WINDOW_STACKP 16384U
#include <ViennaRNA/part_func_window.h>
```

Trigger base pair stack probabilities.

Passing this flag to vrna_probs_window() activates callback execution for stacking probabilities. In turn, the corresponding callback receives this flag through the type argument whenever stack probabilities are provided.

Bug Currently, this flag is a placeholder doing nothing as the corresponding implementation for stack probability computation is missing.

See also

vrna_probs_window()

16.21.2.4 VRNA_PROBS_WINDOW_UP_SPLIT

```
#define VRNA_PROBS_WINDOW_UP_SPLIT 32768U
#include <ViennaRNA/part_func_window.h>
```

Trigger detailed unpaired probabilities split up into different loop type contexts.

Passing this flag to vrna_probs_window() activates callback execution for unpaired probabilities. In contrast to VRNA_PROBS_WINDOW_UP this flag requests unpaired probabilities to be split up into different loop type contexts. In turn, the corresponding callback receives the VRNA_PROBS_WINDOW_UP flag OR-ed together with the corresponding loop type, i.e.:

- VRNA_EXT_LOOP Exterior loop.
- VRNA_HP_LOOP Hairpin loop.
- VRNA INT LOOP Internal loop.
- VRNA_MB_LOOP Multibranch loop.
- VRNA ANY LOOP Any loop.

See also

vrna_probs_window(), VRNA_PROBS_WINDOW_UP

16.21.2.5 VRNA_PROBS_WINDOW_PF

```
#define VRNA_PROBS_WINDOW_PF 65536U
#include <ViennaRNA/part_func_window.h>
```

Trigger partition function.

Passing this flag to vrna_probs_window() activates callback execution for partition function. In turn, the corresponding callback receives this flag through it's type argument whenever partition function data is provided.

Note

Instead of actually providing the partition function Z, the callback is always provided with the corresponding enemble free energy $\Delta G = -RT \ln Z$.

See also

vrna_probs_window()

16.21.3 Typedef Documentation

16.21.3.1 vrna_probs_window_callback

```
\label{typedef} \begin{tabular}{ll} typedef void() vrna\_probs\_window\_callback(FLT\_OR\_DBL *pr, int pr\_size, int i, int max, unsigned int type, void *data) \end{tabular}
```

#include <ViennaRNA/part_func_window.h>

Sliding window probability computation callback.

Notes on Callback Functions This function will be called for each probability data set in the sliding window probability computation implementation of vrna_probs_window(). The argument type specifies the type of probability that is passed to this function.

Types:

- VRNA_PROBS_WINDOW_BPP Trigger base pairing probabilities.
- VRNA_PROBS_WINDOW_UP Trigger unpaired probabilities.
- VRNA_PROBS_WINDOW_PF Trigger partition function.

The above types usually come exclusively. However, for unpaired probabilities, the VRNA_PROBS_WINDOW_UP flag is OR-ed together with one of the loop type contexts

- VRNA_EXT_LOOP Exterior loop.
- VRNA_HP_LOOP Hairpin loop.
- VRNA_INT_LOOP Internal loop.
- VRNA_MB_LOOP Multibranch loop.
- VRNA_ANY_LOOP Any loop.

to indicate the particular type of data available through the $\ensuremath{\text{pr}}$ pointer.

See also

vrna_probs_window(), vrna_pfl_fold_up_cb()

Parameters

pr	An array of probabilities
pr_size	The length of the probability array
i	The i-position (5') of the probabilities
max	The (theoretical) maximum length of the probability array
type	The type of data that is provided
data	Auxiliary data

16.21.4 Function Documentation

16.21.4.1 vrna probs window()

#include <ViennaRNA/part_func_window.h>

Compute various equilibrium probabilities under a sliding window approach.

This function applies a sliding window scan for the sequence provided with the argument fc and reports back equilibrium probabilities through the callback function cb. The data reported to the callback depends on the options

Note

flag.

The parameter ulength only affects computation and resulting data if unpaired probability computations are requested through the options flag.

Options:

- VRNA_PROBS_WINDOW_BPP Trigger base pairing probabilities.
- VRNA_PROBS_WINDOW_UP Trigger unpaired probabilities.
- VRNA_PROBS_WINDOW_UP_SPLIT Trigger detailed unpaired probabilities split up into different loop type contexts.

Options may be OR-ed together

See also

```
vrna_pfl_fold_cb(), vrna_pfl_fold_up_cb()
```

Parameters

fc	The fold compound with sequence data, model settings and precomputed energy parameters
ulength	The maximal length of an unpaired segment (only for unpaired probability computations)
cb	The callback function which collects the pair probability data for further processing
data	Some arbitrary data structure that is passed to the callback cb
options	Option flags to control the behavior of this function

Returns

0 on failure, non-zero on success

16.21.4.2 vrna_pfl_fold()

#include <ViennaRNA/part_func_window.h>

Compute base pair probabilities using a sliding-window approach.

This is a simplified wrapper to vrna_probs_window() that given a nucleid acid sequence, a window size, a maximum base pair span, and a cutoff value computes the pair probabilities for any base pair in any window. The pair probabilities are returned as a list and the user has to take care to free() the memory occupied by the list.

Note

This function uses default model settings! For custom model settings, we refer to the function vrna_probs_window().

In case of any computation errors, this function returns \mathtt{NULL}

See also

```
vrna_probs_window(), vrna_pfl_fold_cb(), vrna_pfl_fold_up()
```

Parameters

sequence	The nucleic acid input sequence
window_size	The size of the sliding window
max_bp_span	The maximum distance along the backbone between two nucleotides that form a base pairs
cutoff	A cutoff value that omits all pairs with lower probability

Returns

A list of base pair probabilities, terminated by an entry with vrna_ep_t.i and vrna_ep_t.j set to 0

16.21.4.3 vrna_pfl_fold_cb()

Compute base pair probabilities using a sliding-window approach (callback version)

This is a simplified wrapper to vrna_probs_window() that given a nucleid acid sequence, a window size, a maximum base pair span, and a cutoff value computes the pair probabilities for any base pair in any window. It is similar to vrna_pfl_fold() but uses a callback mechanism to return the pair probabilities.

Read the details for vrna_probs_window() for details on the callback implementation!

Note

This function uses default model settings! For custom model settings, we refer to the function vrna_probs_window().

See also

```
vrna_probs_window(), vrna_pfl_fold(), vrna_pfl_fold_up_cb()
```

Parameters

sequence	The nucleic acid input sequence	
window_size	The size of the sliding window	
max_bp_span	The maximum distance along the backbone between two nucleotides that form a base pairs	
cb	The callback function which collects the pair probability data for further processing	
data	Some arbitrary data structure that is passed to the callback cb	

Returns

0 on failure, non-zero on success

16.21.4.4 vrna_pfl_fold_up()

```
int ulength,
    int window_size,
    int max_bp_span )
#include <ViennaRNA/part_func_window.h>
```

Compute probability of contiguous unpaired segments.

This is a simplified wrapper to vrna_probs_window() that given a nucleic acid sequence, a maximum length of unpaired segments (ulength), a window size, and a maximum base pair span computes the equilibrium probability of any segment not exceeding ulength. The probabilities to be unpaired are returned as a 1-based, 2-dimensional matrix with dimensions $N \times M$, where N is the length of the sequence and M is the maximum segment length. As an example, the probability of a segment of size 5 starting at position 100 is stored in the matrix entry X[100][5].

It is the users responsibility to free the memory occupied by this matrix.

Note

This function uses default model settings! For custom model settings, we refer to the function vrna_probs_window().

Parameters

sequence	The nucleic acid input sequence	
ulength	The maximal length of an unpaired segment	
window_size	The size of the sliding window	
max_bp_span	The maximum distance along the backbone between two nucleotides that form a base pairs	

Returns

The probabilities to be unpaired for any segment not exceeding ulength

16.21.4.5 vrna_pfl_fold_up_cb()

Compute probability of contiguous unpaired segments.

This is a simplified wrapper to vrna_probs_window() that given a nucleic acid sequence, a maximum length of unpaired segments (ulength), a window size, and a maximum base pair span computes the equilibrium probability of any segment not exceeding ulength. It is similar to vrna_pfl_fold_up() but uses a callback mechanism to return the unpaired probabilities.

Read the details for vrna_probs_window() for details on the callback implementation!

Note

This function uses default model settings! For custom model settings, we refer to the function vrna_probs_window().

Parameters

sequence	The nucleic acid input sequence	
ulength	The maximal length of an unpaired segment	
window_size	The size of the sliding window	
max_bp_span	The maximum distance along the backbone between two nucleotides that form a base pairs	
cb	The callback function which collects the pair probability data for further processing	
data	Some arbitrary data structure that is passed to the callback cb	

Returns

0 on failure, non-zero on success

16.22 Suboptimals and Representative Structures

Sample and enumerate suboptimal secondary structures from RNA sequence data.

16.22.1 Detailed Description

Sample and enumerate suboptimal secondary structures from RNA sequence data.

Collaboration diagram for Suboptimals and Representative Structures:

Modules

- Suboptimal Structures sensu Stiegler et al. 1984 / Zuker et al. 1989
- · Suboptimal Structures within an Energy Band around the MFE
- Random Structure Samples from the Ensemble

Functions to draw random structure samples from the ensemble according to their equilibrium probability.

- Compute the Structure with Maximum Expected Accuracy (MEA)
- · Compute the Centroid Structure

Files

· file boltzmann sampling.h

Boltzmann Sampling of secondary structures from the ensemble.

· file centroid.h

Centroid structure computation.

• file MEA.h

Computes a MEA (maximum expected accuracy) structure.

· file mm.h

Several Maximum Matching implementations.

file subopt.h

RNAsubopt and density of states declarations.

16.23 Suboptimal Structures sensu Stiegler et al. 1984 / Zuker et al. 1989

16.23.1 Detailed Description

Collaboration diagram for Suboptimal Structures sensu Stiegler et al. 1984 / Zuker et al. 1989:

Functions

vrna_subopt_solution_t * vrna_subopt_zuker (vrna_fold_compound_t *vc)

Compute Zuker type suboptimal structures.

SOLUTION * zukersubopt (const char *string)

Compute Zuker type suboptimal structures.

SOLUTION * zukersubopt_par (const char *string, vrna_param_t *parameters)

Compute Zuker type suboptimal structures.

16.23.2 Function Documentation

16.23.2.1 vrna_subopt_zuker()

Compute Zuker type suboptimal structures.

Compute Suboptimal structures according to M. Zuker [26], i.e. for every possible base pair the minimum energy structure containing the resp. base pair. Returns a list of these structures and their energies.

Note

This function internally uses the cofold implementation to compute the suboptimal structures. For that purpose, the function doubles the sequence and enlarges the DP matrices, which in fact will grow by a factor of 4 during the computation! At the end of the structure prediction, everything will be re-set to its original requriements, i.e. normal sequence, normal (empty) DP matrices.

Bug Due to resizing, any pre-existing constraints will be lost!

See also

```
vrna_subopt(), zukersubopt(), zukersubopt_par()
```

Parameters

vc fol	d compound
--------	------------

Returns

List of zuker suboptimal structures

SWIG Wrapper Notes This function is attached as method subopt_zuker() to objects of type fold_compound

16.23.2.2 zukersubopt()

Compute Zuker type suboptimal structures.

Compute Suboptimal structures according to M. Zuker, i.e. for every possible base pair the minimum energy structure containing the resp. base pair. Returns a list of these structures and their energies.

Deprecated use vrna zukersubopt() instead

Parameters

string	RNA sequence
--------	--------------

Returns

List of zuker suboptimal structures

16.23.2.3 zukersubopt_par()

Compute Zuker type suboptimal structures.

Deprecated use vrna_zukersubopt() instead

16.24 Suboptimal Structures within an Energy Band around the MFE

16.24.1 Detailed Description

Collaboration diagram for Suboptimal Structures within an Energy Band around the MFE:

Typedefs

typedef void() vrna_subopt_callback(const char *stucture, float energy, void *data)
 Callback for vrna_subopt_cb()

Functions

- vrna_subopt_solution_t * vrna_subopt (vrna_fold_compound_t *vc, int delta, int sorted, FILE *fp)
 Returns list of subopt structures or writes to fp.
- void vrna_subopt_cb (vrna_fold_compound_t *vc, int delta, vrna_subopt_callback *cb, void *data)

 Generate suboptimal structures within an energy band arround the MFE.
- SOLUTION * subopt (char *seq, char *structure, int delta, FILE *fp)

Returns list of subopt structures or writes to fp.

SOLUTION * subopt_par (char *seq, char *structure, vrna_param_t *parameters, int delta, int is_
 constrained, int is_circular, FILE *fp)

Returns list of subopt structures or writes to fp.

SOLUTION * subopt_circ (char *seq, char *sequence, int delta, FILE *fp)

Returns list of circular subopt structures or writes to fp.

Variables

· double print energy

printing threshold for use with logML

· int subopt_sorted

Sort output by energy.

16.24.2 Typedef Documentation

```
16.24.2.1 vrna_subopt_callback
```

```
typedef void() vrna_subopt_callback(const char *stucture, float energy, void *data)
#include <ViennaRNA/subopt.h>
```

Callback for vrna_subopt_cb()

Notes on Callback Functions This function will be called for each suboptimal secondary structure that is successfully backtraced.

See also

vrna_subopt_cb()

Parameters

structure	structure The suboptimal secondary structure in dot-bracket notation	
energy	The free energy of the secondary structure in kcal/mol	
data	Some arbitrary, auxiliary data address as passed to vrna_subopt_cb()	

16.24.3 Function Documentation

16.24.3.1 vrna_subopt()

#include <ViennaRNA/subopt.h>

Returns list of subopt structures or writes to fp.

This function produces **all** suboptimal secondary structures within 'delta' * 0.01 kcal/mol of the optimum, see [24]. The results are either directly written to a 'fp' (if 'fp' is not NULL), or (fp==NULL) returned in a vrna_subopt_solution_t * list terminated by an entry were the 'structure' member is NULL.

Note

This function requires all multibranch loop DP matrices for unique multibranch loop backtracing. Therefore, the supplied $vrna_fold_compound_t \lor c$ (argument 1) must be initialized with $vrna_md_t.uniq_ML = 1$, for instance like this:

```
vrna_md_t md;
vrna_md_set_default(&md);
md.uniq_ML = 1;
vrna_fold_compound_t *vc=vrna_fold_compound("GGGGGGAAAAAACCCCCCC", &md, VRNA_OPTION_DEFAULT);
```

See also

vrna_subopt_cb(), vrna_subopt_zuker()

Parameters

VC	
delta	
sorted	Sort results by energy in ascending order
fp	

Returns

SWIG Wrapper Notes This function is attached as method subopt() to objects of type fold_compound

16.24.3.2 vrna_subopt_cb()

#include <ViennaRNA/subopt.h>

Generate suboptimal structures within an energy band arround the MFE.

This is the most generic implementation of the suboptimal structure generator according to Wuchty et al. 1999 [24]. Identical to vrna_subopt(), it computes all secondary structures within an energy band delta arround the MFE. However, this function does not print the resulting structures and their corresponding free energies to a file pointer, or returns them as a list. Instead, it calls a user-provided callback function which it passes the structure in dot-bracket format, the corresponding free energy in kcal/mol, and a user-provided data structure each time a structure was backtracked successfully. This function indicates the final output, i.e. the end of the backtracking procedure by passing NULL instead of an actual dot-bracket string to the callback.

Note

This function requires all multibranch loop DP matrices for unique multibranch loop backtracing. Therefore, the supplied $vrna_fold_compound_t \lor c$ (argument 1) must be initialized with $vrna_md_t.uniq_ML = 1$, for instance like this:

```
vrna_md_t md;
vrna_md_set_default(&md);
md.uniq_ML = 1;
vrna_fold_compound_t *vc=vrna_fold_compound("GGGGGGAAAAAACCCCCC", &md, VRNA_OPTION_DEFAULT);
```

See also

vrna subopt callback, vrna subopt(), vrna subopt zuker()

Parameters

vc	fold compount with the sequence data
delta	Energy band arround the MFE in 10cal/mol, i.e. deka-calories
cb	Pointer to a callback function that handles the backtracked structure and its free energy in kcal/mol
data	Pointer to some data structure that is passed along to the callback

SWIG Wrapper Notes This function is attached as method subopt_cb() to objects of type fold_compound

16.24.3.3 subopt()

#include <ViennaRNA/subopt.h>

Returns list of subopt structures or writes to fp.

This function produces **all** suboptimal secondary structures within 'delta' * 0.01 kcal/mol of the optimum. The results are either directly written to a 'fp' (if 'fp' is not NULL), or (fp==NULL) returned in a SOLUTION * list terminated by an entry were the 'structure' pointer is NULL.

Parameters

seq	
structure	
delta	
fp	

Returns

16.24.3.4 subopt_circ()

#include <ViennaRNA/subopt.h>

Returns list of circular subopt structures or writes to fp.

This function is similar to subopt() but calculates secondary structures assuming the RNA sequence to be circular instead of linear

Parameters

seq	
sequence	
delta	
fp	

Returns

16.25 Random Structure Samples from the Ensemble

Functions to draw random structure samples from the ensemble according to their equilibrium probability.

16.25.1 Detailed Description

Functions to draw random structure samples from the ensemble according to their equilibrium probability.

Collaboration diagram for Random Structure Samples from the Ensemble:

Modules

- · Stochastic Backtracking of Structures from Distance Based Partitioning
 - Contains functions related to stochastic backtracking from a specified distance class.
- Deprecated Interface for Stochastic Backtracking

Macros

- #define VRNA_PBACKTRACK_DEFAULT 0
 - Boltzmann sampling flag indicating default backtracing mode.
- #define VRNA_PBACKTRACK_NON_REDUNDANT 1

Boltzmann sampling flag indicating non-redundant backtracing mode.

Typedefs

- typedef void() vrna_boltzmann_sampling_callback(const char *stucture, void *data)

 Callback for Boltzmann sampling.
- typedef struct vrna_pbacktrack_memory_s * vrna_pbacktrack_mem_t
 Boltzmann sampling memory data structure.

Functions

- char * vrna_pbacktrack5 (vrna_fold_compound_t *fc, unsigned int length)
 - Sample a secondary structure of a subsequence from the Boltzmann ensemble according its probability.
- char ** vrna_pbacktrack5_num (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, unsigned int options)
 - Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability
- unsigned int vrna_pbacktrack5_cb (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, vrna_boltzmann_sampling_callback *cb, void *data, unsigned int options)
 - Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.
- char ** vrna_pbacktrack5_resume (vrna_fold_compound_t *vc, unsigned int num_samples, unsigned int length, vrna_pbacktrack mem_t *nr mem, unsigned int options)
 - Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

unsigned int vrna_pbacktrack5_resume_cb (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, vrna_boltzmann_sampling_callback *cb, void *data, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

char * vrna_pbacktrack (vrna_fold_compound_t *fc)

Sample a secondary structure from the Boltzmann ensemble according its probability.

char ** vrna_pbacktrack_num (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int options)

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

 unsigned int vrna_pbacktrack_cb (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_boltzmann_sampling_callback *cb, void *data, unsigned int options)

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

char ** vrna_pbacktrack_resume (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

unsigned int vrna_pbacktrack_resume_cb (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_boltzmann_sampling_callback *cb, void *data, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

void vrna_pbacktrack_mem_free (vrna_pbacktrack_mem_t s)

Release memory occupied by a Boltzmann sampling memory data structure.

16.25.2 Macro Definition Documentation

16.25.2.1 VRNA_PBACKTRACK_DEFAULT

```
#define VRNA_PBACKTRACK_DEFAULT 0
#include <ViennaRNA/boltzmann sampling.h>
```

Boltzmann sampling flag indicating default backtracing mode.

See also

vrna_pbacktrack5_num(), vrna_pbacktrack5_cb(), vrna_pbacktrack5_resume(), vrna_pbacktrack5_resume_cb(), vrna_pbacktrack_num(), vrna_pbacktrack_cb(), vrna_pbacktrack_resume(), vrna_pbacktrack_resume_cb()

16.25.2.2 VRNA_PBACKTRACK_NON_REDUNDANT

```
#define VRNA_PBACKTRACK_NON_REDUNDANT 1
#include <ViennaRNA/boltzmann_sampling.h>
```

Boltzmann sampling flag indicating non-redundant backtracing mode.

This flag will turn the Boltzmann sampling into non-redundant backtracing mode along the lines of Michalik et al. 2017 [18]

See also

vrna_pbacktrack5_num(), vrna_pbacktrack5_cb(), vrna_pbacktrack5_resume(), vrna_pbacktrack5_resume_cb(), vrna_pbacktrack_num(), vrna_pbacktrack_cb(), vrna_pbacktrack_resume(), vrna_pbacktrack_resume_cb()

16.25.3 Typedef Documentation

16.25.3.1 vrna_boltzmann_sampling_callback

```
typedef void() vrna_boltzmann_sampling_callback(const char *stucture, void *data)
```

#include <ViennaRNA/boltzmann_sampling.h>

Callback for Boltzmann sampling.

Notes on Callback Functions This function will be called for each secondary structure that has been successfully backtraced from the partition function DP matrices.

See also

vrna_pbacktrack5_cb(), vrna_pbacktrack_cb(), vrna_pbacktrack5_resume_cb(), vrna_pbacktrack_resume_cb()

Parameters

structure	The secondary structure in dot-bracket notation
data	Some arbitrary, auxiliary data address as provided to the calling function

16.25.3.2 vrna_pbacktrack_mem_t

```
typedef struct vrna_pbacktrack_memory_s* vrna_pbacktrack_mem_t
```

#include <ViennaRNA/boltzmann_sampling.h>

Boltzmann sampling memory data structure.

This structure is required for properly resuming a previous sampling round in specialized Boltzmann sampling, such as non-redundant backtracking.

Initialize with NULL and pass its address to the corresponding functions vrna_pbacktrack5_resume(), etc.

Note

Do not forget to release memory occupied by this data structure before losing its context! Use vrna_pbacktrack_mem_free().

See also

vrna_pbacktrack5_resume(), vrna_pbacktrack_resume(), vrna_pbacktrack5_resume_cb(), vrna_pbacktrack_resume_cb(), vrna_pbacktrack_mem_free()

16.25.4 Function Documentation

16.25.4.1 vrna_pbacktrack5()

#include <ViennaRNA/boltzmann_sampling.h>

Sample a secondary structure of a subsequence from the Boltzmann ensemble according its probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a secondary structure. The parameter length specifies the length of the substructure starting from the 5' end.

The structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z = \sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Precondition

Unique multiloop decomposition has to be active upon creation of fc with vrna_fold_compound() or similar. This can be done easily by passing vrna_fold_compound() a model details parameter with vrna_md_t.uniq_ML _ 1

vrna_pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

See also

vrna_pbacktrack5_num(), vrna_pbacktrack5_cb(), vrna_pbacktrack()

Parameters

fc	The fold compound data structure	
length	The length of the subsequence to consider (starting with 5' end)	

Returns

A sampled secondary structure in dot-bracket notation (or NULL on error)

SWIG Wrapper Notes This function is attached as overloaded method **pbacktrack5()** to objects of type *fold_← compound*. See also Python Examples - Boltzmann Sampling

16.25.4.2 vrna_pbacktrack5_num()

#include <ViennaRNA/boltzmann sampling.h>

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a set of num_samples secondary structures. The parameter length specifies the length of the substructure starting from the 5' end.

Any structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z = \sum_{s} exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Using the options flag one can switch between regular (VRNA_PBACKTRACK_DEFAULT) backtracing mode, and non-redundant sampling (VRNA_PBACKTRACK_NON_REDUNDANT) along the lines of Michalik et al. 2017 [18].

Precondition

Unique multiloop decomposition has to be active upon creation of fc with $vrna_fold_compound()$ or similar. This can be done easily by passing $vrna_fold_compound()$ a model details parameter with $vrna_md_t.uniq_ML = 1$.

vrna pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

Warning

In non-redundant sampling mode (VRNA_PBACKTRACK_NON_REDUNDANT), this function may not yield the full number of requested samples. This may happen if a) the number of requested structures is larger than the total number of structures in the ensemble, b) numeric instabilities prevent the backtracking function to enumerate structures with high free energies, or c) any other error occurs.

See also

vrna_pbacktrack5(), vrna_pbacktrack5_cb(), vrna_pbacktrack_num(), VRNA_PBACKTRACK_DEFAULT, VRNA_PBACKTRACK_NON_REDUNDANT

Parameters

fc	The fold compound data structure
num_samples	The size of the sample set, i.e. number of structures
length Generated by Doxygen	The length of the subsequence to consider (starting with 5' end)
options	A bitwise OR-flag indicating the backtracing mode.

Returns

A set of secondary structure samples in dot-bracket notation terminated by NULL (or NULL on error)

SWIG Wrapper Notes This function is attached as overloaded method pbacktrack5() to objects of type fold_←
compound where the last argument options is optional with default value options =
VRNA_PBACKTRACK_DEFAULT. See also Python Examples - Boltzmann Sampling

16.25.4.3 vrna_pbacktrack5_cb()

```
unsigned int vrna_pbacktrack5_cb (
    vrna_fold_compound_t * fc,
    unsigned int num_samples,
    unsigned int length,
    vrna_boltzmann_sampling_callback * cb,
    void * data,
    unsigned int options )
```

#include <ViennaRNA/boltzmann_sampling.h>

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a set of num_samples secondary structures. The parameter length specifies the length of the substructure starting from the 5' end.

Any structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z = \sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Using the options flag one can switch between regular (VRNA_PBACKTRACK_DEFAULT) backtracing mode, and non-redundant sampling (VRNA_PBACKTRACK_NON_REDUNDANT) along the lines of Michalik et al. 2017 [18].

In contrast to vrna_pbacktrack5() and vrna_pbacktrack5_num() this function yields the structure samples through a callback mechanism.

Precondition

Unique multiloop decomposition has to be active upon creation of fc with vrna_fold_compound() or similar. This can be done easily by passing vrna_fold_compound() a model details parameter with vrna_md_t.uniq_ML = 1.

vrna_pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

Warning

In non-redundant sampling mode (VRNA_PBACKTRACK_NON_REDUNDANT), this function may not yield the full number of requested samples. This may happen if a) the number of requested structures is larger than the total number of structures in the ensemble, b) numeric instabilities prevent the backtracking function to enumerate structures with high free energies, or c) any other error occurs.

See also

vrna_pbacktrack5(), vrna_pbacktrack5_num(), vrna_pbacktrack_cb(), VRNA_PBACKTRACK_DEFAULT, VRNA_PBACKTRACK_NON_REDUNDANT

Parameters

fc	The fold compound data structure	
num_samples	The size of the sample set, i.e. number of structures	
length	The length of the subsequence to consider (starting with 5' end)	
cb	The callback that receives the sampled structure	
data	A data structure passed through to the callback cb	
options	A bitwise OR-flag indicating the backtracing mode.	

Returns

The number of structures actually backtraced

SWIG Wrapper Notes This function is attached as overloaded method pbacktrack5() to objects of type fold_←
compound where the last argument options is optional with default value options =
VRNA_PBACKTRACK_DEFAULT. See also Python Examples - Boltzmann Sampling

16.25.4.4 vrna_pbacktrack5_resume()

#include <ViennaRNA/boltzmann_sampling.h>

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a set of $num_samples$ secondary structures. The parameter length specifies the length of the substructure starting from the 5' end.

Any structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z = \sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Using the options flag one can switch between regular (VRNA_PBACKTRACK_DEFAULT) backtracing mode, and non-redundant sampling (VRNA_PBACKTRACK_NON_REDUNDANT) along the lines of Michalik et al. 2017 [18].

In contrast to vrna_pbacktrack5_cb() this function allows for resuming a previous sampling round in specialized Boltzmann sampling, such as non-redundant backtracking. For that purpose, the user passes the address of a Boltzmann sampling data structure (vrna_pbacktrack_mem_t) which will be re-used in each round of sampling, i.e. each successive call to vrna_pbacktrack5_resume_cb() or vrna_pbacktrack5_resume().

A successive sample call to this function may look like:

Precondition

Unique multiloop decomposition has to be active upon creation of fc with vrna_fold_compound() or similar. This can be done easily by passing vrna_fold_compound() a model details parameter with vrna_md_t.uniq_ML = 1.

vrna_pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

Warning

In non-redundant sampling mode (VRNA_PBACKTRACK_NON_REDUNDANT), this function may not yield the full number of requested samples. This may happen if a) the number of requested structures is larger than the total number of structures in the ensemble, b) numeric instabilities prevent the backtracking function to enumerate structures with high free energies, or c) any other error occurs.

See also

vrna_pbacktrack5_resume_cb(), vrna_pbacktrack5_cb(), vrna_pbacktrack_resume(), vrna_pbacktrack_mem_t, VRNA_PBACKTRACK_DEFAULT, VRNA_PBACKTRACK_NON_REDUNDANT, vrna_pbacktrack_mem_free

Parameters

fc	The fold compound data structure	
num_samples	The size of the sample set, i.e. number of structures	
length	The length of the subsequence to consider (starting with 5' end)	
nr_mem	The address of the Boltzmann sampling memory data structure	
options	A bitwise OR-flag indicating the backtracing mode.	

Returns

A set of secondary structure samples in dot-bracket notation terminated by NULL (or NULL on error)

SWIG Wrapper Notes This function is attached as overloaded method pbacktrack5() to objects of type fold_← compound. In addition to the list of structures, this function also returns the nr_mem data structure as first element. See also Python Examples - Boltzmann Sampling

16.25.4.5 vrna_pbacktrack5_resume_cb()

```
unsigned int vrna_pbacktrack5_resume_cb (
    vrna_fold_compound_t * fc,
    unsigned int num_samples,
    unsigned int length,
    vrna_boltzmann_sampling_callback * cb,
    void * data,
```

```
vrna_pbacktrack_mem_t * nr_mem,
unsigned int options )
```

#include <ViennaRNA/boltzmann_sampling.h>

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a set of num_samples secondary structures. The parameter length specifies the length of the substructure starting from the 5' end.

Any structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z=\sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Using the options flag one can switch between regular (VRNA_PBACKTRACK_DEFAULT) backtracing mode, and non-redundant sampling (VRNA_PBACKTRACK_NON_REDUNDANT) along the lines of Michalik et al. 2017 [18].

In contrast to vrna_pbacktrack5_resume() this function yields the structure samples through a callback mechanism.

A successive sample call to this function may look like:

```
vrna_pbacktrack_mem_t nonredundant_memory = NULL;
// sample the first 100 structures
vrna_pbacktrack5_resume_cb(fc,
                              fc->length,
                              &callback_function,
                              (void *) & callback data,
                              &nonredundant memory,
                              options);
// sample another 500 structures
vrna_pbacktrack5_resume_cb(fc,
                              fc->length,
                              &callback_function,
                              (void *) & callback data,
                              &nonredundant_memory,
                              options);
// release memory occupied by the non-redundant memory data structure
vrna_pbacktrack_mem_free(nonredundant_memory);
```

Precondition

Unique multiloop decomposition has to be active upon creation of fc with vrna_fold_compound() or similar. This can be done easily by passing vrna_fold_compound() a model details parameter with vrna_md_t.uniq_ML _ 1

vrna_pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

Warning

In non-redundant sampling mode (VRNA_PBACKTRACK_NON_REDUNDANT), this function may not yield the full number of requested samples. This may happen if a) the number of requested structures is larger than the total number of structures in the ensemble, b) numeric instabilities prevent the backtracking function to enumerate structures with high free energies, or c) any other error occurs.

See also

vrna_pbacktrack5_resume(), vrna_pbacktrack5_cb(), vrna_pbacktrack_resume_cb(), vrna_pbacktrack_mem_t, VRNA_PBACKTRACK_DEFAULT, VRNA_PBACKTRACK_NON_REDUNDANT, vrna_pbacktrack_mem_free

Parameters

fc	The fold compound data structure	
num_samples	The size of the sample set, i.e. number of structures	
length	The length of the subsequence to consider (starting with 5' end)	
cb	The callback that receives the sampled structure	
data	A data structure passed through to the callback cb	
nr_mem	The address of the Boltzmann sampling memory data structure	
options	A bitwise OR-flag indicating the backtracing mode.	

Returns

The number of structures actually backtraced

SWIG Wrapper Notes This function is attached as overloaded method pbacktrack5() to objects of type fold_← compound. In addition to the number of structures backtraced, this function also returns the nr_mem data structure as first element. See also Python Examples - Boltzmann Sampling

16.25.4.6 vrna_pbacktrack()

#include <ViennaRNA/boltzmann_sampling.h>

Sample a secondary structure from the Boltzmann ensemble according its probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a secondary structure.

The structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z=\sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Precondition

Unique multiloop decomposition has to be active upon creation of fc with vrna_fold_compound() or similar. This can be done easily by passing vrna_fold_compound() a model details parameter with vrna_md_t.uniq_ML = 1.

vrna_pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

See also

 $vrna_pbacktrack5(), vrna_pbacktrack_num, vrna_pbacktrack_cb()$

Parameters

fc The fold compound data structure

Returns

A sampled secondary structure in dot-bracket notation (or NULL on error)

SWIG Wrapper Notes This function is attached as overloaded method **pbacktrack()** to objects of type *fold_← compound*. See also Python Examples - Boltzmann Sampling

16.25.4.7 vrna_pbacktrack_num()

#include <ViennaRNA/boltzmann_sampling.h>

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a set of num_samples secondary structures.

Any structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z=\sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Using the options flag one can switch between regular (VRNA_PBACKTRACK_DEFAULT) backtracing mode, and non-redundant sampling (VRNA_PBACKTRACK_NON_REDUNDANT) along the lines of Michalik et al. 2017 [18].

Precondition

Unique multiloop decomposition has to be active upon creation of fc with vrna_fold_compound() or similar. This can be done easily by passing vrna_fold_compound() a model details parameter with vrna_md_t.uniq_ML = 1.

vrna pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

Warning

In non-redundant sampling mode (VRNA_PBACKTRACK_NON_REDUNDANT), this function may not yield the full number of requested samples. This may happen if a) the number of requested structures is larger than the total number of structures in the ensemble, b) numeric instabilities prevent the backtracking function to enumerate structures with high free energies, or c) any other error occurs.

See also

vrna_pbacktrack(), vrna_pbacktrack_cb(), vrna_pbacktrack5_num(), VRNA_PBACKTRACK_DEFAULT, VRNA_PBACKTRACK_NON_REDUNDANT

Parameters

fc	The fold compound data structure	
num_samples	The size of the sample set, i.e. number of structures	
options	A bitwise OR-flag indicating the backtracing mode.	

Returns

A set of secondary structure samples in dot-bracket notation terminated by NULL (or NULL on error)

SWIG Wrapper Notes This function is attached as overloaded method pbacktrack() to objects of type fold_←
compound where the last argument options is optional with default value options =
VRNA_PBACKTRACK_DEFAULT. See also Python Examples - Boltzmann Sampling

16.25.4.8 vrna_pbacktrack_cb()

```
unsigned int vrna_pbacktrack_cb (
    vrna_fold_compound_t * fc,
    unsigned int num_samples,
    vrna_boltzmann_sampling_callback * cb,
    void * data,
    unsigned int options )
```

#include <ViennaRNA/boltzmann_sampling.h>

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a set of num_samples secondary structures.

Any structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z=\sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Using the options flag one can switch between regular (VRNA_PBACKTRACK_DEFAULT) backtracing mode, and non-redundant sampling (VRNA_PBACKTRACK_NON_REDUNDANT) along the lines of Michalik et al. 2017 [18].

In contrast to vrna_pbacktrack() and vrna_pbacktrack_num() this function yields the structure samples through a callback mechanism.

Precondition

Unique multiloop decomposition has to be active upon creation of fc with $vrna_fold_compound()$ or similar. This can be done easily by passing $vrna_fold_compound()$ a model details parameter with $vrna_md_t.uniq_ML = 1$.

vrna_pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

Warning

In non-redundant sampling mode (VRNA_PBACKTRACK_NON_REDUNDANT), this function may not yield the full number of requested samples. This may happen if a) the number of requested structures is larger than the total number of structures in the ensemble, b) numeric instabilities prevent the backtracking function to enumerate structures with high free energies, or c) any other error occurs.

See also

vrna_pbacktrack(), vrna_pbacktrack_num(), vrna_pbacktrack5_cb(), VRNA_PBACKTRACK_DEFAULT, VRNA_PBACKTRACK_NON_REDUNDANT

Parameters

fc	The fold compound data structure	
num_samples	The size of the sample set, i.e. number of structures	
cb	The callback that receives the sampled structure	
data	A data structure passed through to the callback cb	
options	A bitwise OR-flag indicating the backtracing mode.	

Returns

The number of structures actually backtraced

SWIG Wrapper Notes This function is attached as overloaded method pbacktrack() to objects of type fold_← compound where the last argument options is optional with default value options = VRNA_PBACKTRACK_DEFAULT. See also Python Examples - Boltzmann Sampling

16.25.4.9 vrna_pbacktrack_resume()

#include <ViennaRNA/boltzmann_sampling.h>

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a set of num_samples secondary structures.

Any structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z=\sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Using the options flag one can switch between regular (VRNA_PBACKTRACK_DEFAULT) backtracing mode, and non-redundant sampling (VRNA_PBACKTRACK_NON_REDUNDANT) along the lines of Michalik et al. 2017 [18].

In contrast to vrna_pbacktrack_cb() this function allows for resuming a previous sampling round in specialized Boltzmann sampling, such as non-redundant backtracking. For that purpose, the user passes the address of a Boltzmann sampling data structure (vrna_pbacktrack_mem_t) which will be re-used in each round of sampling, i.e. each successive call to vrna_pbacktrack_resume_cb() or vrna_pbacktrack_resume().

A successive sample call to this function may look like:

Precondition

Unique multiloop decomposition has to be active upon creation of fc with vrna_fold_compound() or similar. This can be done easily by passing vrna_fold_compound() a model details parameter with vrna_md_t.uniq_ML _ 1

vrna_pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

Warning

In non-redundant sampling mode (VRNA_PBACKTRACK_NON_REDUNDANT), this function may not yield the full number of requested samples. This may happen if a) the number of requested structures is larger than the total number of structures in the ensemble, b) numeric instabilities prevent the backtracking function to enumerate structures with high free energies, or c) any other error occurs.

See also

vrna_pbacktrack_resume_cb(), vrna_pbacktrack_cb(), vrna_pbacktrack5_resume(), vrna_pbacktrack_mem_t, VRNA_PBACKTRACK_DEFAULT, VRNA_PBACKTRACK_NON_REDUNDANT, vrna_pbacktrack_mem_free

Parameters

fc	The fold compound data structure	
num_samples	The size of the sample set, i.e. number of structures	
nr_mem	The address of the Boltzmann sampling memory data structure	
options	A bitwise OR-flag indicating the backtracing mode.	

Returns

A set of secondary structure samples in dot-bracket notation terminated by NULL (or NULL on error)

SWIG Wrapper Notes This function is attached as overloaded method **pbacktrack()** to objects of type *fold_← compound*. In addition to the list of structures, this function also returns the nr_mem data structure as first element. See also Python Examples - Boltzmann Sampling

16.25.4.10 vrna_pbacktrack_resume_cb()

```
unsigned int vrna_pbacktrack_resume_cb (
    vrna_fold_compound_t * fc,
    unsigned int num_samples,
    vrna_boltzmann_sampling_callback * cb,
    void * data,
    vrna_pbacktrack_mem_t * nr_mem,
    unsigned int options )
```

#include <ViennaRNA/boltzmann_sampling.h>

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

Perform a probabilistic (stochastic) backtracing in the partition function DP arrays to obtain a set of num_samples secondary structures.

Any structure s with free energy E(s) is picked from the Boltzmann distributed ensemble according to its probability

$$p(s) = \frac{exp(-E(s)/kT)}{Z}$$

with partition function $Z=\sum_s exp(-E(s)/kT)$, Boltzmann constant k and thermodynamic temperature T.

Using the options flag one can switch between regular (VRNA_PBACKTRACK_DEFAULT) backtracing mode, and non-redundant sampling (VRNA_PBACKTRACK_NON_REDUNDANT) along the lines of Michalik et al. 2017 [18].

In contrast to vrna_pbacktrack5_resume() this function yields the structure samples through a callback mechanism.

A successive sample call to this function may look like:

```
vrna_pbacktrack_mem_t nonredundant_memory = NULL;
// sample the first 100 structures
vrna_pbacktrack5_resume_cb(fc,
                           100.
                           &callback function,
                            (void *)&callback_data,
                            &nonredundant_memory,
                           options);
// sample another 500 structures
vrna_pbacktrack5_resume_cb(fc,
                           500.
                            &callback_function,
                            (void *)&callback_data,
                           &nonredundant_memory,
                            options);
// release memory occupied by the non-redundant memory data structure
vrna_pbacktrack_mem_free(nonredundant_memory);
```

Precondition

Unique multiloop decomposition has to be active upon creation of fc with vrna_fold_compound() or similar. This can be done easily by passing vrna_fold_compound() a model details parameter with vrna_md_t.uniq_ML = 1.

vrna_pf() has to be called first to fill the partition function matrices

Note

This function is polymorphic. It accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE.

Warning

In non-redundant sampling mode (VRNA_PBACKTRACK_NON_REDUNDANT), this function may not yield the full number of requested samples. This may happen if a) the number of requested structures is larger than the total number of structures in the ensemble, b) numeric instabilities prevent the backtracking function to enumerate structures with high free energies, or c) any other error occurs.

See also

vrna_pbacktrack_resume(), vrna_pbacktrack_cb(), vrna_pbacktrack5_resume_cb(), vrna_pbacktrack_mem_t, VRNA PBACKTRACK DEFAULT, VRNA PBACKTRACK NON REDUNDANT, vrna pbacktrack mem free

Parameters

fc	The fold compound data structure
num_samples	The size of the sample set, i.e. number of structures
cb	The callback that receives the sampled structure
data	A data structure passed through to the callback cb
nr_mem	The address of the Boltzmann sampling memory data structure
options	A bitwise OR-flag indicating the backtracing mode.

Returns

The number of structures actually backtraced

SWIG Wrapper Notes This function is attached as overloaded method pbacktrack() to objects of type fold_← compound. In addition to the number of structures backtraced, this function also returns the nr_mem data structure as first element. See also Python Examples - Boltzmann Sampling

16.25.4.11 vrna_pbacktrack_mem_free()

#include <ViennaRNA/boltzmann_sampling.h>

Release memory occupied by a Boltzmann sampling memory data structure.

C	0	^	al	c	^

 $vrna_pbacktrack_mem_t, vrna_pbacktrack5_resume(), vrna_pbacktrack5_resume_cb(), vrna_pbacktrack_resume_cb(), vrna_pbacktrack_resume_cb()$

Parameters

s The non-redundancy memory data structure

16.26 Compute the Structure with Maximum Expected Accuracy (MEA)

16.26.1 Detailed Description

Collaboration diagram for Compute the Structure with Maximum Expected Accuracy (MEA):

Functions

- char * vrna_MEA (vrna_fold_compound_t *fc, double gamma, float *mea)

 Compute a MEA (maximum expected accuracy) structure.
- char * vrna_MEA_from_plist (vrna_ep_t *plist, const char *sequence, double gamma, vrna_md_t *md, float *mea)

Compute a MEA (maximum expected accuracy) structure from a list of probabilities.

float MEA (plist *p, char *structure, double gamma)

Computes a MEA (maximum expected accuracy) structure.

16.26.2 Function Documentation

16.26.2.1 vrna_MEA()

#include <ViennaRNA/MEA.h>

Compute a MEA (maximum expected accuracy) structure.

The algorithm maximizes the expected accuracy

$$A(S) = \sum_{(i,j)\in S} 2\gamma p_{ij} + \sum_{i\notin S} p_i^u$$

Higher values of γ result in more base pairs of lower probability and thus higher sensitivity. Low values of γ result in structures containing only highly likely pairs (high specificity). The code of the MEA function also demonstrates the use of sparse dynamic programming scheme to reduce the time and memory complexity of folding.

Precondition

vrna_pf() must be executed on input parameter fc

Parameters

fc	The fold compound data structure with pre-filled base pair probability matrix
gamma	The weighting factor for base pairs vs. unpaired nucleotides
mea	A pointer to a variable where the MEA value will be written to

Returns

An MEA structure (or NULL on any error)

SWIG Wrapper Notes This function is attached as overloaded method MEA(gamma = 1.) to objects of type fold

compound. Note, that it returns the MEA structure and MEA value as a tuple (MEA←
structure, MEA)

16.26.2.2 vrna_MEA_from_plist()

#include <ViennaRNA/MEA.h>

Compute a MEA (maximum expected accuracy) structure from a list of probabilities.

The algorithm maximizes the expected accuracy

$$A(S) = \sum_{(i,j)\in S} 2\gamma p_{ij} + \sum_{i\notin S} p_i^u$$

Higher values of γ result in more base pairs of lower probability and thus higher sensitivity. Low values of γ result in structures containing only highly likely pairs (high specificity). The code of the MEA function also demonstrates the use of sparse dynamic programming scheme to reduce the time and memory complexity of folding.

Note

To include G-Quadruplex support, the corresponding field in \mbox{md} must be set.

Parameters

plist	A list of base pair probabilities the MEA structure is computed from
sequence	The RNA sequence that corresponds to the list of probability values
gamma	The weighting factor for base pairs vs. unpaired nucleotides
md	A model details data structure (maybe NULL)
mea	A pointer to a variable where the MEA value will be written to

Returns

An MEA structure (or NULL on any error)

SWIG Wrapper Notes This function is available as overloaded function MEA_from_plist(gamma = 1., md = N ← ULL). Note, that it returns the MEA structure and MEA value as a tuple (MEA_structure, MEA)

16.26.2.3 MEA()

#include <ViennaRNA/MEA.h>

Computes a MEA (maximum expected accuracy) structure.

The algorithm maximizes the expected accuracy

$$A(S) = \sum_{(i,j)\in S} 2\gamma p_{ij} + \sum_{i\notin S} p_i^u$$

Higher values of γ result in more base pairs of lower probability and thus higher sensitivity. Low values of γ result in structures containing only highly likely pairs (high specificity). The code of the MEA function also demonstrates the use of sparse dynamic programming scheme to reduce the time and memory complexity of folding.

Deprecated Use vrna_MEA() or vrna_MEA_from_plist() instead!

16.27 **Compute the Centroid Structure**

16.27.1 Detailed Description

Collaboration diagram for Compute the Centroid Structure:

Functions

```
    char * vrna_centroid (vrna_fold_compound_t *vc, double *dist)
```

Get the centroid structure of the ensemble.

char * vrna_centroid_from_plist (int length, double *dist, vrna_ep_t *pl)

Get the centroid structure of the ensemble.

• char * vrna_centroid_from_probs (int length, double *dist, FLT_OR_DBL *probs)

Get the centroid structure of the ensemble.

16.27.2 Function Documentation

16.27.2.1 vrna_centroid()

```
char* vrna_centroid (
             vrna_fold_compound_t * vc,
            double * dist )
```

#include <ViennaRNA/centroid.h>

Get the centroid structure of the ensemble.

The centroid is the structure with the minimal average distance to all other structures

< $d(S)>=\sum_{(i,j)\in S}(1-p_{ij})+\sum_{(i,j)\notin S}p_{ij}$ Thus, the centroid is simply the structure containing all pairs with $p_ij>0.5$ The distance of the centroid to the ensemble is written to the memory adressed by dist.

Parameters

in	VC	The fold compound data structure
out	dist	A pointer to the distance variable where the centroid distance will be written to

Returns

The centroid structure of the ensemble in dot-bracket notation (\mathtt{NULL} on error)

16.27.2.2 vrna_centroid_from_plist()

```
char* vrna_centroid_from_plist (
            int length,
             double * dist,
             vrna_ep_t * pl )
```

#include <ViennaRNA/centroid.h>

Get the centroid structure of the ensemble.

This function is a threadsafe replacement for centroid() with a vrna_ep_t input

The centroid is the structure with the minimal average distance to all other structures

$$< d(S) > = \sum_{(i,j) \in S} (1 - p_{ij}) + \sum_{(i,j) \notin S} p_{ij}$$

< $d(S)>=\sum_{(i,j)\in S}(1-p_{ij})+\sum_{(i,j)\notin S}p_{ij}$ Thus, the centroid is simply the structure containing all pairs with $p_{ij}>0.5$ The distance of the centroid to the ensemble is written to the memory adressed by dist.

Parameters

in	length	The length of the sequence
out	dist	A pointer to the distance variable where the centroid distance will be written to
in	pl	A pair list containing base pair probability information about the ensemble

Returns

The centroid structure of the ensemble in dot-bracket notation (NULL on error)

16.27.2.3 vrna_centroid_from_probs()

```
char* vrna_centroid_from_probs (
            int length,
            double * dist,
            FLT_OR_DBL * probs )
```

#include <ViennaRNA/centroid.h>

Get the centroid structure of the ensemble.

This function is a threadsafe replacement for centroid() with a probability array input

The centroid is the structure with the minimal average distance to all other structures

< $d(S)>=\sum_{(i,j)\in S}(1-p_{ij})+\sum_{(i,j)\notin S}p_{ij}$ Thus, the centroid is simply the structure containing all pairs with $p_{ij}>0.5$ The distance of the centroid to the ensemble is written to the memory adressed by dist.

Parameters

in	length	The length of the sequence
out	dist	A pointer to the distance variable where the centroid distance will be written to
in	probs	An upper triangular matrix containing base pair probabilities (access via iindx
		vrna_idx_row_wise())

Generated by Doxygen

Returns

The centroid structure of the ensemble in dot-bracket notation (\mathtt{NULL} on error)

16.28 RNA-RNA Interaction 353

16.28 RNA-RNA Interaction

16.28.1 Detailed Description

Collaboration diagram for RNA-RNA Interaction:

Modules

• Partition Function for Two Hybridized Sequences

Partition Function Cofolding.

• Partition Function for two Hybridized Sequences as a Stepwise Process

RNA-RNA interaction as a stepwise process.

Files

· file concentrations.h

Concentration computations for RNA-RNA interactions.

• file duplex.h

Functions for simple RNA-RNA duplex interactions.

• file part_func_up.h

Implementations for accessibility and RNA-RNA interaction as a stepwise process.

16.29 Classified Dynamic Programming Variants

16.29.1 Detailed Description

Collaboration diagram for Classified Dynamic Programming Variants:

Modules

- Distance Based Partitioning of the Secondary Structure Space
- Compute the Density of States

16.30 Distance Based Partitioning of the Secondary Structure Space

16.30.1 Detailed Description

Collaboration diagram for Distance Based Partitioning of the Secondary Structure Space:

Modules

· Computing MFE representatives of a Distance Based Partitioning

Compute the minimum free energy (MFE) and secondary structures for a partitioning of the secondary structure space according to the base pair distance to two fixed reference structures basepair distance to two fixed reference structures.

· Computing Partition Functions of a Distance Based Partitioning

Compute the partition function and stochastically sample secondary structures for a partitioning of the secondary structure space according to the base pair distance to two fixed reference structures.

Stochastic Backtracking of Structures from Distance Based Partitioning

Contains functions related to stochastic backtracking from a specified distance class.

Files

· file 2Dfold.h

MFE structures for base pair distance classes.

• file 2Dpfold.h

Partition function implementations for base pair distance classes.

16.31 Computing MFE representatives of a Distance Based Partitioning

Compute the minimum free energy (MFE) and secondary structures for a partitioning of the secondary structure space according to the base pair distance to two fixed reference structures basepair distance to two fixed reference structures.

16.31.1 Detailed Description

Compute the minimum free energy (MFE) and secondary structures for a partitioning of the secondary structure space according to the base pair distance to two fixed reference structures basepair distance to two fixed reference structures.

See also

For further details, we refer to Lorenz et al. 2009 [14]

Collaboration diagram for Computing MFE representatives of a Distance Based Partitioning:

Data Structures

struct vrna_sol_TwoD_t

Solution element returned from vrna_mfe_TwoD() More...

struct TwoDfold_vars

Variables compound for 2Dfold MFE folding. More...

Typedefs

- typedef struct vrna_sol_TwoD_t vrna_sol_TwoD_t
 - Solution element returned from vrna_mfe_TwoD())
- typedef struct TwoDfold vars TwoDfold vars

Variables compound for 2Dfold MFE folding.

Functions

- vrna_sol_TwoD_t * vrna_mfe_TwoD (vrna_fold_compound_t *vc, int distance1, int distance2)
 - Compute MFE's and representative for distance partitioning.
- char * vrna_backtrack5_TwoD (vrna_fold_compound_t *vc, int k, int l, unsigned int j)

Backtrack a minimum free energy structure from a 5' section of specified length.

TwoDfold_vars * get_TwoDfold_variables (const char *seq, const char *structure1, const char *structure2, int circ)

Get a structure of type TwoDfold_vars prefilled with current global settings.

void destroy TwoDfold variables (TwoDfold vars *our variables)

Destroy a TwoDfold_vars datastructure without memory loss.

- vrna_sol_TwoD_t * TwoDfoldList (TwoDfold_vars *vars, int distance1, int distance2)
 - Compute MFE's and representative for distance partitioning.
- char * TwoDfold backtrack f5 (unsigned int j, int k, int l, TwoDfold vars *vars)

Backtrack a minimum free energy structure from a 5' section of specified length.

• vrna_sol_TwoD_t ** TwoDfold (TwoDfold_vars *our_variables, int distance1, int distance2)

16.31.2 Data Structure Documentation

16.31.2.1 struct vrna_sol_TwoD_t

Solution element returned from vrna_mfe_TwoD()

This element contains free energy and structure for the appropriate kappa (k), lambda (l) neighborhood The datastructure contains two integer attributes 'k' and 'l' as well as an attribute 'en' of type float representing the free energy in kcal/mol and an attribute 's' of type char* containg the secondary structure representative,

A value of INF in k denotes the end of a list

See also

vrna_mfe_TwoD()

Data Fields

int k

Distance to first reference.

int I

Distance to second reference.

• float en

Free energy in kcal/mol.

• char * s

MFE representative structure in dot-bracket notation.

16.31.2.2 struct TwoDfold_vars

Variables compound for 2Dfold MFE folding.

Deprecated This data structure will be removed from the library soon! Use vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

Collaboration diagram for TwoDfold_vars:

Data Fields

vrna_param_t * P

Precomputed energy parameters and model details.

· int do_backtrack

Flag whether to do backtracing of the structure(s) or not.

char * ptype

Precomputed array of pair types.

· char * sequence

The input sequence.

short * \$1

The input sequences in numeric form.

unsigned int maxD1

Maximum allowed base pair distance to first reference.

unsigned int maxD2

Maximum allowed base pair distance to second reference.

unsigned int * mm1

Maximum matching matrix, reference struct 1 disallowed.

unsigned int * mm2

Maximum matching matrix, reference struct 2 disallowed.

int * my_iindx

Index for moving in quadratic distancy dimensions.

unsigned int * referenceBPs1

Matrix containing number of basepairs of reference structure1 in interval [i,j].

unsigned int * referenceBPs2

Matrix containing number of basepairs of reference structure2 in interval [i,j].

• unsigned int * bpdist

Matrix containing base pair distance of reference structure 1 and 2 on interval [i,j].

16.31.3 Typedef Documentation

```
16.31.3.1 vrna_sol_TwoD_t

typedef struct vrna_sol_TwoD_t vrna_sol_TwoD_t

#include <ViennaRNA/2Dfold.h>
```

Solution element returned from vrna_mfe_TwoD()

This element contains free energy and structure for the appropriate kappa (k), lambda (l) neighborhood The datastructure contains two integer attributes 'k' and 'l' as well as an attribute 'en' of type float representing the free energy in kcal/mol and an attribute 's' of type char* containg the secondary structure representative,

A value of INF in k denotes the end of a list

See also

vrna_mfe_TwoD()

16.31.3.2 TwoDfold_vars

```
typedef struct TwoDfold_vars TwoDfold_vars
#include <ViennaRNA/2Dfold.h>
```

Variables compound for 2Dfold MFE folding.

Deprecated This data structure will be removed from the library soon! Use vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

16.31.4 Function Documentation

```
16.31.4.1 vrna_mfe_TwoD()
```

#include <ViennaRNA/2Dfold.h>

Compute MFE's and representative for distance partitioning.

This function computes the minimum free energies and a representative secondary structure for each distance class according to the two references specified in the datastructure 'vars'. The maximum basepair distance to each of both references may be set by the arguments 'distance1' and 'distance2', respectively. If both distance arguments are set to '-1', no restriction is assumed and the calculation is performed for each distance class possible.

The returned list contains an entry for each distance class. If a maximum basepair distance to either of the references was passed, an entry with k=l=-1 will be appended in the list, denoting the class where all structures exceeding the maximum will be thrown into. The end of the list is denoted by an attribute value of INF in the k-attribute of the list entry.

See also

```
\label{lem:compound_twoD} vrna\_fold\_compound\_free(), \quad vrna\_pf\_TwoD() \quad vrna\_backtrack5\_TwoD(), \\ vrna\_sol\_TwoD\_t, \\ vrna\_fold\_compound\_t
```

Parameters

VC	The datastructure containing all precomputed folding attributes
distance1	maximum distance to reference1 (-1 means no restriction)
distance2	maximum distance to reference2 (-1 means no restriction)

Returns

A list of minimum free energies (and corresponding structures) for each distance class

16.31.4.2 vrna_backtrack5_TwoD()

#include <ViennaRNA/2Dfold.h>

Backtrack a minimum free energy structure from a 5' section of specified length.

This function allows one to backtrack a secondary structure beginning at the 5' end, a specified length and residing in a specific distance class. If the argument 'k' gets a value of -1, the structure that is backtracked is assumed to reside in the distance class where all structures exceeding the maximum basepair distance specified in vrna_mfe_TwoD() belong to.

Note

The argument 'vars' must contain precalculated energy values in the energy matrices, i.e. a call to vrna_mfe_TwoD() preceding this function is mandatory!

See also

```
vrna_mfe_TwoD()
```

Parameters

VC	The datastructure containing all precomputed folding attributes
j	The length in nucleotides beginning from the 5' end
k	distance to reference1 (may be -1)
1	distance to reference2

16.31.4.3 get_TwoDfold_variables()

#include <ViennaRNA/2Dfold.h>

Get a structure of type TwoDfold_vars prefilled with current global settings.

This function returns a datastructure of type TwoDfold_vars. The data fields inside the TwoDfold_vars are prefilled by global settings and all memory allocations necessary to start a computation are already done for the convenience of the user

Note

Make sure that the reference structures are compatible with the sequence according to Watson-Crick- and Wobble-base pairing

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

Parameters

seq The RNA sequence	
structure1	The first reference structure in dot-bracket notation
structure2	The second reference structure in dot-bracket notation
circ	A switch to indicate the assumption to fold a circular instead of linear RNA (0=OFF, 1=ON)

Returns

A datastructure prefilled with folding options and allocated memory

16.31.4.4 destroy_TwoDfold_variables()

Destroy a TwoDfold_vars datastructure without memory loss.

This function free's all allocated memory that depends on the datastructure given.

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

Parameters

ĺ	our_variables	A pointer to the datastructure to be destroyed
	oui_variables	A pointer to the datastracture to be destroyed

16.31.4.5 TwoDfoldList()

#include <ViennaRNA/2Dfold.h>

Compute MFE's and representative for distance partitioning.

This function computes the minimum free energies and a representative secondary structure for each distance class according to the two references specified in the datastructure 'vars'. The maximum basepair distance to each of both references may be set by the arguments 'distance1' and 'distance2', respectively. If both distance arguments are set to '-1', no restriction is assumed and the calculation is performed for each distance class possible.

The returned list contains an entry for each distance class. If a maximum basepair distance to either of the references was passed, an entry with k=l=-1 will be appended in the list, denoting the class where all structures exceeding the maximum will be thrown into. The end of the list is denoted by an attribute value of INF in the k-attribute of the list entry.

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_mfe_TwoD(), and vrna_fold_compound_free() instead!

Parameters

vars	the datastructure containing all predefined folding attributes
distance1	maximum distance to reference1 (-1 means no restriction)
distance2	maximum distance to reference2 (-1 means no restriction)

16.31.4.6 TwoDfold_backtrack_f5()

```
char* TwoDfold_backtrack_f5 (
         unsigned int j,
         int k,
         int 1,
         TwoDfold_vars * vars )
```

#include <ViennaRNA/2Dfold.h>

Backtrack a minimum free energy structure from a 5' section of specified length.

This function allows one to backtrack a secondary structure beginning at the 5' end, a specified length and residing in a specific distance class. If the argument 'k' gets a value of -1, the structure that is backtracked is assumed to reside in the distance class where all structures exceeding the maximum basepair distance specified in TwoDfold() belong to.

Note

The argument 'vars' must contain precalculated energy values in the energy matrices, i.e. a call to TwoDfold() preceding this function is mandatory!

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_mfe_TwoD(), vrna_backtrack5_TwoD(), and vrna_fold_compound_free() instead!

Parameters

j	The length in nucleotides beginning from the 5' end
k	distance to reference1 (may be -1)
1	distance to reference2
vars	the datastructure containing all predefined folding attributes

16.31.4.7 TwoDfold()

#include <ViennaRNA/2Dfold.h>

16.32 Computing Partition Functions of a Distance Based Partitioning

Compute the partition function and stochastically sample secondary structures for a partitioning of the secondary structure space according to the base pair distance to two fixed reference structures.

16.32.1 Detailed Description

Compute the partition function and stochastically sample secondary structures for a partitioning of the secondary structure space according to the base pair distance to two fixed reference structures.

Collaboration diagram for Computing Partition Functions of a Distance Based Partitioning:

Data Structures

struct vrna_sol_TwoD_pf_t
 Solution element returned from vrna_pf_TwoD() More...

Typedefs

typedef struct vrna_sol_TwoD_pf_t vrna_sol_TwoD_pf_t
 Solution element returned from vrna_pf_TwoD()

Functions

• vrna_sol_TwoD_pf_t * vrna_pf_TwoD (vrna_fold_compound_t *vc, int maxDistance1, int maxDistance2)

Compute the partition function for all distance classes.

16.32.2 Data Structure Documentation

```
16.32.2.1 struct vrna_sol_TwoD_pf_t
```

Solution element returned from vrna_pf_TwoD()

This element contains the partition function for the appropriate kappa (k), lambda (l) neighborhood The datastructure contains two integer attributes 'k' and 'l' as well as an attribute 'q' of type FLT_OR_DBL

A value of INF in k denotes the end of a list

See also

```
vrna_pf_TwoD()
```

Data Fields

int k

Distance to first reference.

int I

Distance to second reference.

FLT_OR_DBL q
 partition function

16.32.3 Typedef Documentation

```
16.32.3.1 vrna_sol_TwoD_pf_t

typedef struct vrna_sol_TwoD_pf_t vrna_sol_TwoD_pf_t

#include <ViennaRNA/2Dpfold.h>
```

This element contains the partition function for the appropriate kappa (k), lambda (l) neighborhood The datastructure contains two integer attributes 'k' and 'l' as well as an attribute 'q' of type FLT_OR_DBL

A value of INF in k denotes the end of a list

Solution element returned from vrna_pf_TwoD()

See also

```
vrna_pf_TwoD()
```

16.32.4 Function Documentation

```
16.32.4.1 vrna_pf_TwoD()
```

Compute the partition function for all distance classes.

This function computes the partition functions for all distance classes according the two reference structures specified in the datastructure 'vars'. Similar to vrna_mfe_TwoD() the arguments maxDistance1 and maxDistance2 specify the maximum distance to both reference structures. A value of '-1' in either of them makes the appropriate distance restrictionless, i.e. all basepair distancies to the reference are taken into account during computation. In case there is a restriction, the returned solution contains an entry where the attribute k=l=-1 contains the partition function for all structures exceeding the restriction. A value of INF in the attribute 'k' of the returned list denotes the end of the list

See also

```
vrna fold compound TwoD(), vrna fold compound free(), vrna fold compound vrna sol TwoD pf t
```

Parameters

VC	vc The datastructure containing all necessary folding attributes and matrices	
maxDistance1	The maximum basepair distance to reference1 (may be -1)	
maxDistance2	The maximum basepair distance to reference2 (may be -1)	Generated by Doxygen

_			
R	Δtı	irn	0

A list of partition funtions for the corresponding distance classes

16.33 Stochastic Backtracking of Structures from Distance Based Partitioning

Contains functions related to stochastic backtracking from a specified distance class.

16.33.1 Detailed Description

Contains functions related to stochastic backtracking from a specified distance class.

Collaboration diagram for Stochastic Backtracking of Structures from Distance Based Partitioning:

Functions

- char * vrna_pbacktrack_TwoD (vrna_fold_compound_t *vc, int d1, int d2)
 Sample secondary structure representatives from a set of distance classes according to their Boltzmann probability.
- char * vrna_pbacktrack5_TwoD (vrna_fold_compound_t *vc, int d1, int d2, unsigned int length)
 Sample secondary structure representatives with a specified length from a set of distance classes according to their Boltzmann probability.

16.33.2 Function Documentation

16.33.2.1 vrna_pbacktrack_TwoD()

#include <ViennaRNA/2Dpfold.h>

Sample secondary structure representatives from a set of distance classes according to their Boltzmann probability.

If the argument 'd1' is set to '-1', the structure will be backtracked in the distance class where all structures exceeding the maximum basepair distance to either of the references reside.

Precondition

The argument 'vars' must contain precalculated partition function matrices, i.e. a call to vrna_pf_TwoD() preceding this function is mandatory!

See also

```
vrna_pf_TwoD()
```

Parameters

in,out	vc	The vrna_fold_compound_t datastructure containing all necessary folding attributes and matrices
in	d1	
in	d2	The distance to reference2

Returns

A sampled secondary structure in dot-bracket notation

16.33.2.2 vrna_pbacktrack5_TwoD()

#include <ViennaRNA/2Dpfold.h>

Sample secondary structure representatives with a specified length from a set of distance classes according to their Boltzmann probability.

This function does essentially the same as vrna_pbacktrack_TwoD() with the only difference that partial structures, i.e. structures beginning from the 5' end with a specified length of the sequence, are backtracked

Note

This function does not work (since it makes no sense) for circular RNA sequences!

Precondition

The argument 'vars' must contain precalculated partition function matrices, i.e. a call to vrna_pf_TwoD() preceding this function is mandatory!

See also

```
vrna_pbacktrack_TwoD(), vrna_pf_TwoD()
```

Parameters

in,out	VC	The vrna_fold_compound_t datastructure containing all necessary folding attributes and
		matrices
in	d1	The distance to reference1 (may be -1)
in	d2	The distance to reference2
in	length	The length of the structure beginning from the 5' end

Returns

A sampled secondary structure in dot-bracket notation

16.34 Compute the Density of States

16.34.1 Detailed Description

Collaboration diagram for Compute the Density of States:

Variables

• int density_of_states [MAXDOS+1]

The Density of States.

16.34.2 Variable Documentation

16.34.2.1 density_of_states

```
int density_of_states[MAXDOS+1]
#include <ViennaRNA/subopt.h>
```

The Density of States.

This array contains the density of states for an RNA sequences after a call to subopt_par(), subopt() or subopt_circ().

Precondition

Call one of the functions subopt_par(), subopt() or subopt_circ() prior accessing the contents of this array

See also

subopt_par(), subopt(), subopt_circ()

16.35 Inverse Folding (Design)

RNA sequence design.

16.35.1 Detailed Description

RNA sequence design.

Files

· file inverse.h

Inverse folding routines.

Functions

• float inverse_fold (char *start, const char *target)

Find sequences with predefined structure.

float inverse_pf_fold (char *start, const char *target)

Find sequence that maximizes probability of a predefined structure.

Variables

· char * symbolset

This global variable points to the allowed bases, initially "AUGC". It can be used to design sequences from reduced alphabets.

- · float final cost
- int give_up
- · int inv_verbose

16.35.2 Function Documentation

16.35.2.1 inverse_fold()

Find sequences with predefined structure.

This function searches for a sequence with minimum free energy structure provided in the parameter 'target', starting with sequence 'start'. It returns 0 if the search was successful, otherwise a structure distance in terms of the energy difference between the search result and the actual target 'target' is returned. The found sequence is returned in 'start'. If give_up is set to 1, the function will return as soon as it is clear that the search will be unsuccessful, this speeds up the algorithm if you are only interested in exact solutions.

Parameters

start The start sequence		The start sequence
	target	The target secondary structure in dot-bracket notation

Returns

The distance to the target in case a search was unsuccessful, 0 otherwise

16.35.2.2 inverse_pf_fold()

Find sequence that maximizes probability of a predefined structure.

This function searches for a sequence with maximum probability to fold into the provided structure 'target' using the partition function algorithm. It returns $-kT \cdot \log(p)$ where p is the frequency of 'target' in the ensemble of possible structures. This is usually much slower than inverse_fold().

Parameters

start	The start sequence
target	The target secondary structure in dot-bracket notation

Returns

The distance to the target in case a search was unsuccessful, 0 otherwise

16.35.3 Variable Documentation

16.35.3.1 final_cost

```
float final_cost

#include <ViennaRNA/inverse.h>
```

when to stop inverse_pf_fold()

```
int give_up

#include <ViennaRNA/inverse.h>

default 0: try to minimize structure distance even if no exact solution can be found

16.35.3.3 inv_verbose

int inv_verbose

#include <ViennaRNA/inverse.h>
```

print out substructure on which inverse_fold() fails

16.36 Neighborhood Relation and Move Sets for Secondary Structures

Different functions to generate structural neighbors of a secondary structure according to a particular Move Set.

16.36.1 Detailed Description

Different functions to generate structural neighbors of a secondary structure according to a particular Move Set.

This module contains methods to compute the neighbors of an RNA secondary structure. Neighbors of a given structure are all structures that differ in exactly one base pair. That means one can insert an delete base pairs in the given structure. These insertions and deletions of base pairs are usually called moves. A third move which is considered in these methods is a shift move. A shifted base pair has one stable position and one position that changes. These moves are encoded as follows:

```
• insertion: (i, j) where i,j > 0
```

- deletion: (i, j) where i,j < 0 shift: (i, j) where either i > 0, j < 0 or i < 0, j > 0

The negative position of a shift indicates the position that has changed.

```
Example:

We have given a sequence and a structure.

Sequence AAGGAAACC

Structure ..(....)

Indices 123456789

The given base pair is (3,9) and the neighbors are the insertion (4, 8), the deletion (-3,-9), the shift (3,-8)

and the shift (-4, 9).

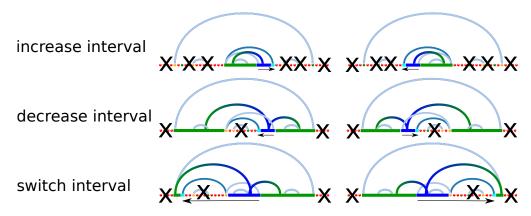
This leads to the neighbored structures:
...(...)
....(...)
....(...)
```

A simple method to construct all insertions is to iterate over the positions of a sequence twice. The first iteration has the index i in [1, sequence length], the second iteration has the index j in [i+1, sequence length]. All pairs (i,j) with compatible letters and which are non-crossing with present base pairs are valid neighbored insertion moves. Valid deletion moves are all present base pairs with negative sign. Valid shift moves are constructed by taking all paired positions as fix position of a shift move and iterating over all positions of the sequence. If the letters of a position are compatible and if it the move is non-crossing with existing base pairs, we have a valid shift move. The method of generating shift moves can be accelerated by skipping neighbored base pairs.

If we need to construct all neighbors several times for subsequent moves, we can speed up the task by using the move set of the previous structure. The previous move set has to be filtered, such that all moves that would cross the next selected move are non-crossing. Next, the selected move has to be removed. Then one has to only to generate all moves that were not possible before. One move is the inverted selected move (if it was an insertion, simply make the indices negative). The generation of all other new moves is different and depends on the selected move. It is easy for an insertion move, because we have only to include all non-crossing shift moves, that are possible with the new base pair. For that we can either iterate over the sequence or we can select all crossing shift moves in the filter procedure and convert them into shifts.

The generation of new moves given a deletion is a little bit more complex, because we can create more moves. At first we can insert the deleted pair as insertion move. Then we generate all insertions that would have crossed the deleted base pair. Finally we construct all crossing shift moves.

If the given move is a shift, we can save much time by specifying the intervals for the generation of new moves. The interval which was enclosed by the positive position of the shift move and the previous paired position is the freed interval after applying the move. This freed interval includes all positions and base pairs that we need to construct new insertions and shifts. All these new moves have one position in the freed interval and the other position in the environment of the freed interval. The environment are all position which are outside the freed interval, but within the same enclosing loop of the shift move. The environment for valid base pairs can be divided into one or more intervals, depending on the shift move. The following examples describe a few scenarios to specify the intervals of the environment.



- freed interval
- —environment for new non-crossing moves
- X intervals that would produce crossing pairs
- new shift moves from pairs in one interval to positions in the other interval
- → points to the new position of the shift move

Given the intervals of the environment and the freed interval, the new shift moves can be constructed quickly. One has to take all positions of pairs from the environment in order to create valid pairs with positions in the freed interval. The same procedure can be applied for the other direction. This is taking all paired positions within the freed interval in order to look for pairs with valid positions in the intervals of the environment. Collaboration diagram for Neighborhood Relation and Move Sets for Secondary Structures:

Files

· file move.h

Methods to operate with structural neighbors of RNA secondary structures.

file neighbor.h

Methods to compute the neighbors of an RNA secondary structure.

Data Structures

• struct vrna_move_s

An atomic representation of the transition / move from one structure to its neighbor. More...

Macros

#define VRNA MOVESET INSERTION 4

Option flag indicating insertion move.

#define VRNA_MOVESET_DELETION 8

Option flag indicating deletion move.

#define VRNA_MOVESET_SHIFT 16

Option flag indicating shift move.

#define VRNA_MOVESET_NO_LP 32

Option flag indicating moves without lonely base pairs.

• #define VRNA MOVESET DEFAULT (VRNA MOVESET INSERTION | VRNA MOVESET DELETION)

Option flag indicating default move set, i.e. insertions/deletion of a base pair.

• #define VRNA NEIGHBOR CHANGE 1

State indicator for a neighbor that has been changed.

• #define VRNA_NEIGHBOR_INVALID 2

State indicator for a neighbor that has been invalidated.

• #define VRNA_NEIGHBOR_NEW 3

State indicator for a neighbor that has become newly available.

Typedefs

- typedef struct vrna_move_s vrna_move_t
 - A single move that transforms a secondary structure into one of its neighbors.
- typedef void() vrna_callback_move_update(vrna_fold_compound_t *fc, vrna_move_t neighbor, unsigned int state, void *data)

Prototype of the neighborhood update callback.

Functions

vrna_move_t vrna_move_init (int pos_5, int pos_3)

Create an atomic move.

- void vrna_move_list_free (vrna_move_t *moves)
- void vrna_move_apply (short *pt, const vrna_move_t *m)

Apply a particular move / transition to a secondary structure, i.e. transform a structure.

int vrna_move_is_removal (const vrna_move_t *m)

Test whether a move is a base pair removal.

int vrna_move_is_insertion (const vrna_move_t *m)

Test whether a move is a base pair insertion.

int vrna_move_is_shift (const vrna_move_t *m)

Test whether a move is a base pair shift.

int vrna_move_compare (const vrna_move_t *a, const vrna_move_t *b, const short *pt)

Compare two moves.

void vrna_loopidx_update (int *loopidx, const short *pt, int length, const vrna_move_t *m)

Alters the loopIndices array that was constructed with vrna_loopidx_from_ptable().

• vrna_move_t * vrna_neighbors (vrna_fold_compound_t *vc, const short *pt, unsigned int options)

Generate neighbors of a secondary structure.

Generate neighbors of a secondary structure (the fast way)

• int vrna_move_neighbor_diff_cb (vrna_fold_compound_t *fc, short *ptable, vrna_move_t move, vrna_callback_move_update *cb, void *data, unsigned int options)

Apply a move to a secondary structure and indicate which neighbors have changed consequentially.

• vrna_move_t * vrna_move_neighbor_diff (vrna_fold_compound_t *fc, short *ptable, vrna_move_t move, vrna_move_t **invalid_moves, unsigned int options)

Apply a move to a secondary structure and indicate which neighbors have changed consequentially.

16.36.2 Data Structure Documentation

16.36.2.1 struct vrna move s

An atomic representation of the transition / move from one structure to its neighbor.

An atomic transition / move may be one of the following:

- · a base pair insertion,
- · a base pair removal, or
- a base pair shift where an existing base pair changes one of its pairing partner.

These moves are encoded by two integer values that represent the affected 5' and 3' nucleotide positions. Furthermore, we use the following convention on the signedness of these encodings:

- both values are positive for insertion moves
- · both values are negative for base pair removals
- both values have different signedness for *shift moves*, where the positive value indicates the nucleotide that stays constant, and the others absolute value is the new pairing partner

Note

A value of 0 in either field is used as list-end indicator and doesn't represent any valid move.

Collaboration diagram for vrna_move_s:

Data Fields

• int pos 5

The (absolute value of the) 5' position of a base pair, or any position of a shifted pair.

• int pos 3

The (absolute value of the) 3' position of a base pair, or any position of a shifted pair.

vrna_move_t * next

The next base pair (if an elementary move changes more than one base pair), or NULL Has to be terminated with move 0,0.

16.36.3 Macro Definition Documentation

```
16.36.3.1 VRNA_MOVESET_INSERTION
```

```
#define VRNA_MOVESET_INSERTION 4
```

#include <ViennaRNA/landscape/move.h>

Option flag indicating insertion move.

See also

vrna_neighbors(), vrna_neighbors_successive, vrna_path()

```
16.36.3.2 VRNA_MOVESET_DELETION
#define VRNA_MOVESET_DELETION 8
#include <ViennaRNA/landscape/move.h>
Option flag indicating deletion move.
See also
     vrna_neighbors(), vrna_neighbors_successive, vrna_path()
16.36.3.3 VRNA_MOVESET_SHIFT
#define VRNA_MOVESET_SHIFT 16
#include <ViennaRNA/landscape/move.h>
Option flag indicating shift move.
See also
     vrna_neighbors(), vrna_neighbors_successive, vrna_path()
16.36.3.4 VRNA_MOVESET_NO_LP
#define VRNA_MOVESET_NO_LP 32
#include <ViennaRNA/landscape/move.h>
Option flag indicating moves without lonely base pairs.
See also
     vrna_neighbors(), vrna_neighbors_successive, vrna_path()
16.36.3.5 VRNA_MOVESET_DEFAULT
#define VRNA_MOVESET_DEFAULT (VRNA_MOVESET_INSERTION | VRNA_MOVESET_DELETION)
#include <ViennaRNA/landscape/move.h>
Option flag indicating default move set, i.e. insertions/deletion of a base pair.
See also
     vrna_neighbors(), vrna_neighbors_successive, vrna_path()
```

```
16.36.3.6 VRNA_NEIGHBOR_CHANGE
#define VRNA_NEIGHBOR_CHANGE 1
#include <ViennaRNA/landscape/neighbor.h>
State indicator for a neighbor that has been changed.
See also
     vrna_move_neighbor_diff_cb()
16.36.3.7 VRNA_NEIGHBOR_INVALID
#define VRNA_NEIGHBOR_INVALID 2
#include <ViennaRNA/landscape/neighbor.h>
State indicator for a neighbor that has been invalidated.
See also
     vrna_move_neighbor_diff_cb()
16.36.3.8 VRNA_NEIGHBOR_NEW
#define VRNA_NEIGHBOR_NEW 3
#include <ViennaRNA/landscape/neighbor.h>
State indicator for a neighbor that has become newly available.
See also
     vrna_move_neighbor_diff_cb()
16.36.4 Typedef Documentation
16.36.4.1 vrna_callback_move_update
typedef void() vrna_callback_move_update(vrna_fold_compound_t *fc, vrna_move_t neighbor, unsigned
int state, void *data)
#include <ViennaRNA/landscape/neighbor.h>
Prototype of the neighborhood update callback.
See also
```

vrna_move_neighbor_diff_cb(), VRNA_NEIGHBOR_CHANGE, VRNA_NEIGHBOR_INVALID, VRNA_NEIGHBOR_NEW

Parameters

fc	The fold compound the calling function is working on	
neighbor	The move that generates the (changed or new) neighbor	
state	The state of the neighbor (move) as supplied by argument neighbor	
data	Some arbitrary data pointer as passed to vrna_move_neighbor_diff_cb()	

16.36.5 Function Documentation

```
16.36.5.1 vrna_move_init()
```

#include <ViennaRNA/landscape/move.h>

Create an atomic move.

See also

vrna_move_s

Parameters

pos	The 5' position of the move (positive for insertions, negative for removal, any value for shift moves)
_5	
pos-	The 3' position of the move (positive for insertions, negative for removal, any value for shift moves)
3	

Returns

An atomic move as specified by pos_5 and pos_3

16.36.5.2 vrna_move_list_free()

#include <ViennaRNA/landscape/move.h>

delete all moves in a zero terminated list.

16.36.5.3 vrna_move_apply()

#include <ViennaRNA/landscape/move.h>

Apply a particular move / transition to a secondary structure, i.e. transform a structure.

Parameters

in,out	pt	The pair table representation of the secondary structure
in	m	The move to apply

16.36.5.4 vrna_move_is_removal()

#include <ViennaRNA/landscape/move.h>

Test whether a move is a base pair removal.

Parameters

```
m The move to test against
```

Returns

Non-zero if the move is a base pair removal, 0 otherwise

16.36.5.5 vrna_move_is_insertion()

#include <ViennaRNA/landscape/move.h>

Test whether a move is a base pair insertion.

Parameters

m The move to test against

Returns

Non-zero if the move is a base pair insertion, 0 otherwise

16.36.5.6 vrna_move_is_shift()

#include <ViennaRNA/landscape/move.h>

Test whether a move is a base pair shift.

Parameters

m The move to test against

Returns

Non-zero if the move is a base pair shift, 0 otherwise

16.36.5.7 vrna_move_compare()

#include <ViennaRNA/landscape/move.h>

Compare two moves.

The function compares two moves a and b and returns whether move a is lexicographically smaller (-1), larger (1) or equal to move b.

If any of the moves a or b is a shift move, this comparison only makes sense in a structure context. Thus, the third argument with the current structure must be provided.

Note

This function returns 0 (equality) upon any error, e.g. missing input

Warning

Currently, shift moves are not supported!

Parameters

а	The first move of the comparison
b	The second move of the comparison
pt	The pair table of the current structure that is compatible with both moves (maybe NULL if moves are guaranteed to be no shifts)

Returns

```
-1 if a < b, 1 if a > b, 0 otherwise
```

16.36.5.8 vrna_loopidx_update()

```
void vrna_loopidx_update (
    int * loopidx,
    const short * pt,
    int length,
    const vrna_move_t * m )
```

#include <ViennaRNA/landscape/neighbor.h>

Alters the loopIndices array that was constructed with vrna_loopidx_from_ptable().

The loopIndex of the current move will be inserted. The correctness of the input will not be checked because the speed should be optimized.

Parameters

in,out	loopidx	The loop index data structure that needs an update
in	pt	A pair table on which the move will be executed
	length	The length of the structure
in	m	The move that is applied to the current structure

16.36.5.9 vrna_neighbors()

#include <ViennaRNA/landscape/neighbor.h>

Generate neighbors of a secondary structure.

This function allows one to generate all structural neighbors (according to a particular move set) of an RNA secondary structure. The neighborhood is then returned as a list of transitions / moves required to transform the current structure into the actual neighbor.

See also

vrna_neighbors_successive(), vrna_move_apply(), VRNA_MOVESET_INSERTION, VRNA_MOVESET_DELETION, VRNA_MOVESET_SHIFT, VRNA_MOVESET_DEFAULT

Parameters

in	vc	A vrna_fold_compound_t containing the energy parameters and model details
in	pt	The pair table representation of the structure
	options	Options to modify the behavior of this function, e.g. available move set

Returns

Neighbors as a list of moves / transitions (the last element in the list has both of its fields set to 0)

SWIG Wrapper Notes This function is attached as an overloaded method *neighbors()* to objects of type *fold_← compound*. The optional parameter options defaults to VRNA_MOVESET_DEFAULT if it is omitted.

16.36.5.10 vrna_neighbors_successive()

#include <ViennaRNA/landscape/neighbor.h>

Generate neighbors of a secondary structure (the fast way)

This function implements a fast way to generate all neighbors of a secondary structure that results from successive applications of individual moves. The speed-up results from updating an already known list of valid neighbors before the individual move towards the current structure took place. In essence, this function removes neighbors that are not accessible anymore and inserts neighbors emerging after a move took place.

See also

vrna_neighbors(), vrna_move_apply(), VRNA_MOVESET_INSERTION, VRNA_MOVESET_DELETION, VRNA_MOVESET_SHIFT, VRNA_MOVESET_DEFAULT

Parameters

in	VC	A vrna_fold_compound_t containing the energy parameters and model details
in	curr_move	The move that was/will be applied to prev_pt
in	prev_pt	A pair table representation of the structure before curr_move is/was applied
in	prev_neighbors	The list of neighbors of prev_pt
Congreted b	size_prev_neighbors	The size of prev_neighbors, i.e. the lists length
out	y Doxygen size_neighbors	A pointer to store the size / length of the new neighbor list
	options	Options to modify the behavior of this function, e.g. available move set

Returns

Neighbors as a list of moves / transitions (the last element in the list has both of its fields set to 0)

16.36.5.11 vrna_move_neighbor_diff_cb()

```
int vrna_move_neighbor_diff_cb (
    vrna_fold_compound_t * fc,
    short * ptable,
    vrna_move_t move,
    vrna_callback_move_update * cb,
    void * data,
    unsigned int options)
```

#include <ViennaRNA/landscape/neighbor.h>

Apply a move to a secondary structure and indicate which neighbors have changed consequentially.

This function applies a move to a secondary structure and explores the local neighborhood of the affected loop. Any changes to previously compatible neighbors that have been affected by this loop will be reported through a callback function. In particular, any of the three cases might appear:

- A previously available neighbor move has changed, usually the free energy change of the move (VRNA_NEIGHBOR_CHANGE)
- A previously available neighbor move became invalid (VRNA_NEIGHBOR_INVALID)
- A new neighbor move becomes available (VRNA_NEIGHBOR_NEW)

See also

vrna_move_neighbor_diff(), VRNA_NEIGHBOR_CHANGE, VRNA_NEIGHBOR_INVALID, VRNA_NEIGHBOR_NEW, vrna_callback_move_update

Parameters

fc	A fold compound for the RNA sequence(s) that this function operates on
ptable	The current structure as pair table
move	The move to apply
cb	The address of the callback function that is passed the neighborhood changes
data	An arbitrary data pointer that will be passed through to the callback function cb
options	Options to modify the behavior of this function, .e.g available move set

Returns

Non-zero on success, 0 otherwise

16.36.5.12 vrna_move_neighbor_diff()

```
vrna_move_t* vrna_move_neighbor_diff (
    vrna_fold_compound_t * fc,
    short * ptable,
    vrna_move_t move,
    vrna_move_t ** invalid_moves,
    unsigned int options)
```

#include <ViennaRNA/landscape/neighbor.h>

Apply a move to a secondary structure and indicate which neighbors have changed consequentially.

Similar to vrna_move_neighbor_diff_cb(), this function applies a move to a secondary structure and reports back the neighbors of the current structure become affected by this move. Instead of executing a callback for each of the affected neighbors, this function compiles two lists of neighbor moves, one that is returned and consists of all moves that are novel or may have changed in energy, and a second, invalid_moves, that consists of all the neighbor moves that become invalid, respectively.

Parameters

fc	A fold compound for the RNA sequence(s) that this function operates on
ptable	The current structure as pair table
move	The move to apply
invalid_moves	The address of a move list where the function stores those moves that become invalid
options	Options to modify the behavior of this function, .e.g available move set

Returns

A list of moves that might have changed in energy or are novel compared to the structure before application of the move

16.37 (Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima

API for various RNA folding path algorithms.

16.37.1 Detailed Description

API for various RNA folding path algorithms.

This part of our API allows for generating RNA secondary structure (re-)folding paths between two secondary structures or simply starting from a single structure. This is most important if an estimate of the refolding energy barrier between two structures is required, or a structure's corresponding local minimum needs to be determined, e.g. through a gradient-descent walk.

This part of the interface is further split into the following sections:

- · Direct Refolding Paths between two Secondary Structures, and
- · Folding Paths that start at a single Secondary Structure

Collaboration diagram for (Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima:

Modules

- · Direct Refolding Paths between two Secondary Structures
 - Heuristics to explore direct, optimal (re-)folding paths between two secondary structures.
- · Folding Paths that start at a single Secondary Structure
 - Implementation of gradient- and random walks starting from a single secondary structure.
- Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers

Files

· file findpath.h

A breadth-first search heuristic for optimal direct folding paths.

file paths.h

API for computing (optimal) (re-)folding paths between secondary structures.

• file walk.h

Methods to generate particular paths such as gradient or random walks through the energy landscape of an RNA sequence.

Data Structures

struct vrna_path_s

An element of a refolding path list. More...

Macros

• #define VRNA_PATH_TYPE_DOT_BRACKET 1U

Flag to indicate producing a (re-)folding path as list of dot-bracket structures.

#define VRNA_PATH_TYPE_MOVES 2U

Flag to indicate producing a (re-)folding path as list of transition moves.

Typedefs

typedef struct vrna_path_s vrna_path_t

Typename for the refolding path data structure vrna_path_s.

typedef struct vrna_path_options_s * vrna_path_options_t

Options data structure for (re-)folding path implementations.

Functions

void vrna_path_free (vrna_path_t *path)

Release (free) memory occupied by a (re-)folding path.

void vrna_path_options_free (vrna_path_options_t options)

Release (free) memory occupied by an options data structure for (re-)folding path implementations.

16.37.2 Data Structure Documentation

```
16.37.2.1 struct vrna_path_s
```

An element of a refolding path list.

Usually, one has to deal with an array of vrna_path_s, e.g. returned from one of the refolding-path algorithms.

Since in most cases the length of the list is not known in advance, such lists have an *end-of-list* marker, which is either:

- a value of NULL for vrna_path_s::s if vrna_path_s::type = VRNA_PATH_TYPE_DOT_BRACKET, or
- a vrna_path_s::move with zero in both fields vrna_move_t::pos_5 and vrna_move_t::pos_3 if vrna_path_s::type = VRNA PATH TYPE MOVES.

In the following we show an example for how to cover both cases of iteration:

See also

```
vrna_path_free()
```

Collaboration diagram for vrna_path_s:

Data Fields

· unsigned int type

The type of the path element.

• double en

Free energy of current structure.

char * s

Secondary structure in dot-bracket notation.

· vrna move t move

Move that transforms the previous structure into it's next neighbor along the path.

16.37.2.1.1 Field Documentation

```
16.37.2.1.1.1 type
```

```
unsigned int vrna_path_s::type
```

The type of the path element.

A value of VRNA_PATH_TYPE_DOT_BRACKET indicates that vrna_path_s::s consists of the secondary structure in dot-bracket notation, and vrna_path_s::en the corresponding free energy.

On the other hand, if the value is VRNA_PATH_TYPE_MOVES, vrna_path_s::s is NULL and vrna_path_s::move is set to the transition move that transforms a previous structure into it's neighbor along the path. In this case, the attribute vrna_path_s::en states the change in free energy with respect to the structure before application of vrna_path_s::move.

16.37.3 Macro Definition Documentation

16.37.3.1 VRNA_PATH_TYPE_DOT_BRACKET

```
#define VRNA_PATH_TYPE_DOT_BRACKET 1U
```

#include <ViennaRNA/landscape/paths.h>

Flag to indicate producing a (re-)folding path as list of dot-bracket structures.

See also

vrna_path_t, vrna_path_options_findpath(), vrna_path_direct(), vrna_path_direct_ub()

```
16.37.3.2 VRNA_PATH_TYPE_MOVES
```

```
#define VRNA_PATH_TYPE_MOVES 2U
#include <ViennaRNA/landscape/paths.h>
```

Flag to indicate producing a (re-)folding path as list of transition moves.

See also

```
vrna_path_t, vrna_path_options_findpath(), vrna_path_direct(), vrna_path_direct_ub()
```

16.37.4 Function Documentation

```
16.37.4.1 vrna_path_free()
```

#include <ViennaRNA/landscape/paths.h>

Release (free) memory occupied by a (re-)folding path.

See also

```
vrna_path_direct(), vrna_path_direct_ub(), vrna_path_findpath(), vrna_path_findpath_ub()
```

Parameters

path The refolding path to be free'd

16.37.4.2 vrna_path_options_free()

Release (free) memory occupied by an options data structure for (re-)folding path implementations.

See also

```
vrna_path_options_findpath(), vrna_path_direct(), vrna_path_direct_ub()
```

Parameters

options The options data structure to be free'd

16.38 Direct Refolding Paths between two Secondary Structures

Heuristics to explore direct, optimal (re-)folding paths between two secondary structures.

16.38.1 Detailed Description

Heuristics to explore direct, optimal (re-)folding paths between two secondary structures.

Collaboration diagram for Direct Refolding Paths between two Secondary Structures:

Functions

- int vrna_path_findpath_saddle (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width)

 Find energy of a saddle point between 2 structures (search only direct path)
- int vrna_path_findpath_saddle_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width, int maxE)

Find energy of a saddle point between 2 structures (search only direct path)

- vrna_path_t * vrna_path_findpath (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width)

 Find refolding path between 2 structures (search only direct path)
- vrna_path_t * vrna_path_findpath_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width, int maxE)

Find refolding path between 2 structures (search only direct path)

- vrna_path_options_t vrna_path_options_findpath (int width, unsigned int type)
 - Create options data structure for findpath direct (re-)folding path heuristic.
- vrna_path_t * vrna_path_direct (vrna_fold_compound_t *fc, const char *s1, const char *s2, vrna_path_options_t options)

Determine an optimal direct (re-)folding path between two secondary structures.

vrna_path_t * vrna_path_direct_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int maxE, vrna_path_options_t options)

Determine an optimal direct (re-)folding path between two secondary structures.

16.38.2 Function Documentation

16.38.2.1 vrna_path_findpath_saddle()

```
int vrna_path_findpath_saddle (
    vrna_fold_compound_t * vc,
    const char * s1,
    const char * s2,
    int width )
```

#include <ViennaRNA/landscape/findpath.h>

Find energy of a saddle point between 2 structures (search only direct path)

This function uses an inplementation of the *findpath* algorithm [7] for near-optimal direct refolding path prediction.

Model details, and energy parameters are used as provided via the parameter 'fc'. The vrna_fold_compound_t does not require memory for any DP matrices, but requires all most basic init values as one would get from a call like this:

fc = vrna_fold_compound(sequence, NULL, VRNA_OPTION_DEFAULT);

See also

vrna path findpath saddle ub(), vrna fold compound(), vrna fold compound t, vrna path findpath()

Parameters

fc	The vrna_fold_compound_t with precomputed sequence encoding and model details	
s1	The start structure in dot-bracket notation	
s2	The target structure in dot-bracket notation	
width	A number specifying how many strutures are being kept at each step during the searc	

Returns

The saddle energy in 10cal/mol

SWIG Wrapper Notes This function is attached as an overloaded method *path_findpath_saddle()* to objects of type *fold_compound*. The optional parameter width defaults to 1 if it is omitted.

16.38.2.2 vrna_path_findpath_saddle_ub()

```
int vrna_path_findpath_saddle_ub (
    vrna_fold_compound_t * vc,
    const char * s1,
    const char * s2,
    int width,
    int maxE )
```

#include <ViennaRNA/landscape/findpath.h>

Find energy of a saddle point between 2 structures (search only direct path)

This function uses an inplementation of the *findpath* algorithm [7] for near-optimal direct refolding path prediction.

Model details, and energy parameters are used as provided via the parameter 'fc'. The vrna_fold_compound_t does not require memory for any DP matrices, but requires all most basic init values as one would get from a call like this:

fc = vrna_fold_compound (sequence, NULL, VRNA_OPTION_DEFAULT);

Warning

The argument maxE (E_{max}) enables one to specify an upper bound, or maximum free energy for the saddle point between the two input structures. If no path with $E_{saddle} < E_{max}$ is found, the function simply returns maxE

See also

vrna_path_findpath_saddle(), vrna_fold_compound(), vrna_fold_compound_t, vrna_path_findpath()

Parameters

fc	The vrna_fold_compound_t with precomputed sequence encoding and model details	
s1 The start structure in dot-bracket notation		
s2	The target structure in dot-bracket notation	
width	A number specifying how many strutures are being kept at each step during the search	
maxE	An upper bound for the saddle point energy in 10cal/mol	

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Returns

The saddle energy in 10cal/mol

SWIG Wrapper Notes This function is attached as an overloaded method $path_findpath_saddle()$ to objects of type $fold_compound$. The optional parameter width defaults to 1 if it is omitted, while the optional parameter maxE defaults to INF. In case the function did not find a path with $E_{saddle} < E_{max}$ the function returns a NULL object, i.e. undef for Perl and None for Python.

16.38.2.3 vrna_path_findpath()

#include <ViennaRNA/landscape/findpath.h>

Find refolding path between 2 structures (search only direct path)

This function uses an inplementation of the *findpath* algorithm [7] for near-optimal direct refolding path prediction.

Model details, and energy parameters are used as provided via the parameter 'fc'. The vrna_fold_compound_t does not require memory for any DP matrices, but requires all most basic init values as one would get from a call like this: fc = vrna_fold_compound(sequence, NULL, VRNA_OPTION_DEFAULT);

See also

vrna_path_findpath_ub(), vrna_fold_compound(), vrna_fold_compound_t, vrna_path_findpath_saddle()

Parameters

fc	The vrna_fold_compound_t with precomputed sequence encoding and model details	
s1	The start structure in dot-bracket notation	
s2	The target structure in dot-bracket notation	
width	A number specifying how many strutures are being kept at each step during the search	

Returns

The saddle energy in 10cal/mol

SWIG Wrapper Notes This function is attached as an overloaded method path_findpath() to objects of type fold

_compound. The optional parameter width defaults to 1 if it is omitted.

16.38.2.4 vrna_path_findpath_ub()

```
vrna_path_t * vrna_path_findpath_ub (
    vrna_fold_compound_t * vc,
    const char * s1,
    const char * s2,
    int width,
    int maxE )
```

#include <ViennaRNA/landscape/findpath.h>

Find refolding path between 2 structures (search only direct path)

This function uses an inplementation of the *findpath* algorithm [7] for near-optimal direct refolding path prediction.

Model details, and energy parameters are used as provided via the parameter 'fc'. The vrna_fold_compound_t does not require memory for any DP matrices, but requires all most basic init values as one would get from a call like this:
fc = vrna_fold_compound(sequence, NULL, VRNA_OPTION_DEFAULT);

Warning

The argument maxE enables one to specify an upper bound, or maximum free energy for the saddle point between the two input structures. If no path with $E_{saddle} < E_{max}$ is found, the function simply returns *NULL*

See also

vrna_path_findpath(), vrna_fold_compound(), vrna_fold_compound_t, vrna_path_findpath_saddle()

Parameters

fc	The vrna_fold_compound_t with precomputed sequence encoding and model details	
s1	The start structure in dot-bracket notation	
s2	The target structure in dot-bracket notation	
width	A number specifying how many strutures are being kept at each step during the search	
maxE	An upper bound for the saddle point energy in 10cal/mol	

Returns

The saddle energy in 10cal/mol

SWIG Wrapper Notes This function is attached as an overloaded method $path_findpath()$ to objects of type $fold \leftarrow _compound$. The optional parameter width defaults to 1 if it is omitted, while the optional parameter maxE defaults to INF. In case the function did not find a path with $E_{saddle} < E_{max}$ the function returns an empty list.

16.38.2.5 vrna_path_options_findpath()

```
#include <ViennaRNA/landscape/paths.h>
```

Create options data structure for findpath direct (re-)folding path heuristic.

This function returns an options data structure that switches the vrna_path_direct() and vrna_path_direct_ub() API functions to use the *findpath* [7] heuristic. The parameter width specifies the width of the breadth-first search while the second parameter type allows one to set the type of the returned (re-)folding path.

Currently, the following return types are available:

- A list of dot-bracket structures and corresponding free energy (flag: VRNA_PATH_TYPE_DOT_BRACKET)
- A list of transition moves and corresponding free energy changes (flag: VRNA PATH TYPE MOVES)

See also

```
VRNA_PATH_TYPE_DOT_BRACKET, VRNA_PATH_TYPE_MOVES, vrna_path_options_free(), vrna_path_direct(), vrna_path_direct_ub()
```

Parameters

width Width of the breath-first search strategy		Width of the breath-first search strategy	
type Setting that specifies how the return (re-)folding path should be		Setting that specifies how the return (re-)folding path should be encoded]

Returns

An options data structure with settings for the findpath direct path heuristic

SWIG Wrapper Notes This function is available as overloaded function path_options_findpath(). The optional parameter width defaults to 10 if omitted, while the optional parameter type defaults to VRNA PATH TYPE DOT BRACKET.

16.38.2.6 vrna_path_direct()

```
vrna_path_t * vrna_path_direct (
    vrna_fold_compound_t * fc,
    const char * s1,
    const char * s2,
    vrna_path_options_t options )
```

#include <ViennaRNA/landscape/paths.h>

Determine an optimal direct (re-)folding path between two secondary structures.

This is the generic wrapper function to retrieve (an optimal) (re-)folding path between two secondary structures s1 and s2. The actual algorithm that is used to generate the (re-)folding path is determined by the settings specified in the options data structure. This data structure also determines the return type, which might be either:

- · a list of dot-bracket structures with corresponding free energy, or
- · a list of transition moves with corresponding free energy change

If the options parameter is passed a NULL pointer, this function defaults to the *findpath heuristic* [7] with a breadth-first search width of 10, and the returned path consists of dot-bracket structures with corresponding free energies.

See also

vrna_path_direct_ub(), vrna_path_options_findpath(), vrna_path_options_free(), vrna_path_free()

Parameters

fc	The vrna_fold_compound_t with precomputed sequence encoding and model details	
s1	The start structure in dot-bracket notation	
s2	The target structure in dot-bracket notation	
options An options data structure that specifies the path heuristic and corresponding settings (n		

Returns

An optimal (re-)folding path between the two input structures

SWIG Wrapper Notes This function is attached as an overloaded method *path_direct()* to objects of type *fold_← compound*. The optional parameter options defaults to *NULL* if it is omitted.

16.38.2.7 vrna_path_direct_ub()

#include <ViennaRNA/landscape/paths.h>

Determine an optimal direct (re-)folding path between two secondary structures.

This function is similar to vrna_path_direct(), but allows to specify an *upper-bound* for the saddle point energy. The underlying algorithms will stop determining an (optimal) (re-)folding path, if none can be found that has a saddle point below the specified upper-bound threshold maxE.

Warning

The argument maxE enables one to specify an upper bound, or maximum free energy for the saddle point between the two input structures. If no path with $E_{saddle} < E_{max}$ is found, the function simply returns *NULL*

See also

vrna_path_direct_ub(), vrna_path_options_findpath(), vrna_path_options_free(), vrna_path_free()

fc	The vrna_fold_compound_t with precomputed sequence encoding and model details		
s1	The start structure in dot-bracket notation		
s2	The target structure in dot-bracket notation		
maxE	Upper bound for the saddle point along the (re-)folding path		
options	An options data structure that specifies the path heuristic and corresponding settings (maybe NULL)		

Returns

An optimal (re-)folding path between the two input structures

SWIG Wrapper Notes This function is attached as an overloaded method $path_direct()$ to objects of type $fold_{\leftarrow}$ compound. The optional parameter <code>maxE</code> defaults to <code>#INT_MAX</code> - 1 if it is omitted, while the optional parameter <code>options</code> defaults to NULL. In case the function did not find a path with $E_{saddle} < E_{max}$ it returns an empty list.

16.39 Folding Paths that start at a single Secondary Structure

Implementation of gradient- and random walks starting from a single secondary structure.

16.39.1 Detailed Description

Implementation of gradient- and random walks starting from a single secondary structure.

Collaboration diagram for Folding Paths that start at a single Secondary Structure:

Macros

#define VRNA PATH STEEPEST DESCENT 128

Option flag to request a steepest descent / gradient path.

#define VRNA PATH RANDOM 256

Option flag to request a random walk path.

• #define VRNA PATH NO TRANSITION OUTPUT 512

Option flag to omit returning the transition path.

#define VRNA_PATH_DEFAULT (VRNA_PATH_STEEPEST_DESCENT | VRNA_MOVESET_DEFAULT)

Option flag to request defaults (steepest descent / default move set)

Functions

- vrna_move_t * vrna_path (vrna_fold_compound_t *vc, short *pt, unsigned int steps, unsigned int options)
 Compute a path, store the final structure, and return a list of transition moves from the start to the final structure.
- vrna_move_t * vrna_path_gradient (vrna_fold_compound_t *vc, short *pt, unsigned int options)
 Compute a steepest descent / gradient path, store the final structure, and return a list of transition moves from the start to the final structure.
- vrna_move_t * vrna_path_random (vrna_fold_compound_t *vc, short *pt, unsigned int steps, unsigned int options)

Generate a random walk / path of a given length, store the final structure, and return a list of transition moves from the start to the final structure.

16.39.2 Macro Definition Documentation

16.39.2.1 VRNA_PATH_STEEPEST_DESCENT

```
#define VRNA_PATH_STEEPEST_DESCENT 128
#include <ViennaRNA/landscape/walk.h>
```

Option flag to request a steepest descent / gradient path.

See also

vrna_path()

```
16.39.2.2 VRNA_PATH_RANDOM
#define VRNA_PATH_RANDOM 256
#include <ViennaRNA/landscape/walk.h>
Option flag to request a random walk path.
See also
     vrna_path()
16.39.2.3 VRNA_PATH_NO_TRANSITION_OUTPUT
#define VRNA_PATH_NO_TRANSITION_OUTPUT 512
#include <ViennaRNA/landscape/walk.h>
Option flag to omit returning the transition path.
See also
    vrna_path(), vrna_path_gradient(), vrna_path_random()
16.39.2.4 VRNA_PATH_DEFAULT
#define VRNA_PATH_DEFAULT (VRNA_PATH_STEEPEST_DESCENT | VRNA_MOVESET_DEFAULT)
#include <ViennaRNA/landscape/walk.h>
Option flag to request defaults (steepest descent / default move set)
See also
     vrna_path(), VRNA_PATH_STEEPEST_DESCENT, VRNA_MOVESET_DEFAULT
```

16.39.3 Function Documentation

16.39.3.1 vrna_path()

#include <ViennaRNA/landscape/walk.h>

Compute a path, store the final structure, and return a list of transition moves from the start to the final structure.

This function computes, given a start structure in pair table format, a transition path, updates the pair table to the final structure of the path. Finally, if not requested otherwise by using the VRNA_PATH_NO_TRANSITION_OUTPUT flag in the options field, this function returns a list of individual transitions that lead from the start to the final structure if requested.

The currently available transition paths are

- Steepest Descent / Gradient walk (flag: VRNA_PATH_STEEPEST_DESCENT)
- Random walk (flag: VRNA_PATH_RANDOM)

The type of transitions must be set through the options parameter

Note

Since the result is written to the input structure you may want to use vrna_ptable_copy() before calling this function to keep the initial structure

See also

```
vrna_path_gradient(), vrna_path_random(), vrna_ptable(), vrna_ptable_copy(), vrna_fold_compound() 
VRNA_PATH_STEEPEST_DESCENT, VRNA_PATH_RANDOM, VRNA_MOVESET_DEFAULT, VRNA_MOVESET_SHIFT, 
VRNA_PATH_NO_TRANSITION_OUTPUT
```

Parameters

in	vc	A vrna_fold_compound_t containing the energy parameters and model details
in,out	pt	The pair table containing the start structure. Used to update to the final structure after
		execution of this function
in	options	Options to modify the behavior of this function

Returns

A list of transition moves (default), or NULL (if options & VRNA_PATH_NO_TRANSITION_OUTPUT)

SWIG Wrapper Notes This function is attached as an overloaded method path() to objects of type fold_compound.

The optional parameter options defaults to VRNA PATH DEFAULT if it is omitted.

16.39.3.2 vrna_path_gradient()

```
vrna_move_t * vrna_path_gradient (
    vrna_fold_compound_t * vc,
    short * pt,
    unsigned int options)
```

#include <ViennaRNA/landscape/walk.h>

Compute a steepest descent / gradient path, store the final structure, and return a list of transition moves from the start to the final structure.

This function computes, given a start structure in pair table format, a steepest descent path, updates the pair table to the final structure of the path. Finally, if not requested otherwise by using the VRNA_PATH_NO_TRANSITION_OUTPUT flag in the options field, this function returns a list of individual transitions that lead from the start to the final structure if requested.

Note

Since the result is written to the input structure you may want to use vrna_ptable_copy() before calling this function to keep the initial structure

See also

vrna_path_random(), vrna_path(), vrna_ptable(), vrna_ptable_copy(), vrna_fold_compound() VRNA_MOVESET_DEFAULT, VRNA_MOVESET_SHIFT, VRNA_PATH_NO_TRANSITION_OUTPUT

Parameters

in	VC	A vrna_fold_compound_t containing the energy parameters and model details
in,out	pt	The pair table containing the start structure. Used to update to the final structure after
		execution of this function
in	options	Options to modify the behavior of this function

Returns

A list of transition moves (default), or NULL (if options & VRNA_PATH_NO_TRANSITION_OUTPUT)

SWIG Wrapper Notes This function is attached as an overloaded method path_gradient() to objects of type fold

_compound. The optional parameter options defaults to VRNA_PATH_DEFAULT if it is omitted.

16.39.3.3 vrna_path_random()

#include <ViennaRNA/landscape/walk.h>

Generate a random walk / path of a given length, store the final structure, and return a list of transition moves from the start to the final structure.

This function generates, given a start structure in pair table format, a random walk / path, updates the pair table to the final structure of the path. Finally, if not requested otherwise by using the VRNA_PATH_NO_TRANSITION_OUTPUT flag in the options field, this function returns a list of individual transitions that lead from the start to the final structure if requested.

Note

Since the result is written to the input structure you may want to use vrna_ptable_copy() before calling this function to keep the initial structure

See also

vrna_path_gradient(), vrna_path(), vrna_ptable(), vrna_ptable_copy(), vrna_fold_compound() VRNA_MOVESET_DEFAULT, VRNA_MOVESET_SHIFT, VRNA_PATH_NO_TRANSITION_OUTPUT

Parameters

in	vc	A vrna_fold_compound_t containing the energy parameters and model details
in,out	pt	The pair table containing the start structure. Used to update to the final structure after
		execution of this function
in	steps	The length of the path, i.e. the total number of transitions / moves
in	options	Options to modify the behavior of this function

Returns

A list of transition moves (default), or NULL (if options & VRNA_PATH_NO_TRANSITION_OUTPUT)

SWIG Wrapper Notes This function is attached as an overloaded method *path_gradient()* to objects of type *fold*__compound. The optional parameter options defaults to VRNA_PATH_DEFAULT if it is omitted.

16.40 Experimental Structure Probing Data

Include Experimental Structure Probing Data to Guide Structure Predictions.

16.40.1 Detailed Description

Include Experimental Structure Probing Data to Guide Structure Predictions.

Collaboration diagram for Experimental Structure Probing Data:

Modules

SHAPE Reactivity Data

Incorporate SHAPE reactivity structure probing data into the folding recursions by means of soft constraints.

· Generate Soft Constraints from Data

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

16.41 SHAPE Reactivity Data

Incorporate SHAPE reactivity structure probing data into the folding recursions by means of soft constraints.

16.41.1 Detailed Description

Incorporate SHAPE reactivity structure probing data into the folding recursions by means of soft constraints.

Details for our implementation to incorporate SHAPE reactivity data to guide secondary structure prediction can be found in [16] Collaboration diagram for SHAPE Reactivity Data:

Files

· file SHAPE.h

This module provides function to incorporate SHAPE reactivity data into the folding recursions by means of soft constraints.

Functions

• int vrna_sc_add_SHAPE_deigan (vrna_fold_compound_t *vc, const double *reactivities, double m, double b, unsigned int options)

Add SHAPE reactivity data as soft constraints (Deigan et al. method)

• int vrna_sc_add_SHAPE_deigan_ali (vrna_fold_compound_t *vc, const char **shape_files, const int *shape_file_association, double m, double b, unsigned int options)

Add SHAPE reactivity data from files as soft constraints for consensus structure prediction (Deigan et al. method)

• int vrna_sc_add_SHAPE_zarringhalam (vrna_fold_compound_t *vc, const double *reactivities, double b, double default_value, const char *shape_conversion, unsigned int options)

Add SHAPE reactivity data as soft constraints (Zarringhalam et al. method)

• int vrna_sc_SHAPE_to_pr (const char *shape_conversion, double *values, int length, double default_value) Convert SHAPE reactivity values to probabilities for being unpaired.

16.41.2 Function Documentation

16.41.2.1 vrna_sc_add_SHAPE_deigan()

#include <ViennaRNA/constraints/SHAPE.h>

Add SHAPE reactivity data as soft constraints (Deigan et al. method)

This approach of SHAPE directed RNA folding uses the simple linear ansatz

```
\Delta G_{\mathrm{SHAPE}}(i) = m \ln(\mathrm{SHAPE} \ \mathrm{reactivity}(i) + 1) + b
```

to convert SHAPE reactivity values to pseudo energies whenever a nucleotide i contributes to a stacked pair. A positive slope m penalizes high reactivities in paired regions, while a negative intercept b results in a confirmatory 'bonus" free energy for correctly predicted base pairs. Since the energy evaluation of a base pair stack involves two pairs, the pseudo energies are added for all four contributing nucleotides. Consequently, the energy term is applied twice for pairs inside a helix and only once for pairs adjacent to other structures. For all other loop types the energy model remains unchanged even when the experimental data highly disagrees with a certain motif.

See also

```
For further details, we refer to [6]. vrna_sc_remove(), vrna_sc_add_SHAPE_zarringhalam(), vrna_sc_minimize_pertubation()
```

Parameters

VC	The vrna_fold_compound_t the soft constraints are associated with
reactivities	A vector of normalized SHAPE reactivities
т	The slope of the conversion function
b	The intercept of the conversion function
options	The options flag indicating how/where to store the soft constraints

Returns

1 on successful extraction of the method, 0 on errors

SWIG Wrapper Notes This function is attached as method **sc_add_SHAPE_deigan()** to objects of type *fold_← compound*

16.41.2.2 vrna_sc_add_SHAPE_deigan_ali()

```
int vrna_sc_add_SHAPE_deigan_ali (
    vrna_fold_compound_t * vc,
    const char ** shape_files,
    const int * shape_file_association,
    double m,
    double b,
    unsigned int options)
```

#include <ViennaRNA/constraints/SHAPE.h>

Add SHAPE reactivity data from files as soft constraints for consensus structure prediction (Deigan et al. method)

Parameters

VC	The vrna_fold_compound_t the soft constraints are associated with
shape_files	A set of filenames that contain normalized SHAPE reactivity data
shape_file_association	An array of integers that associate the files with sequences in the alignment
m	The slope of the conversion function
b	The intercept of the conversion function
options	The options flag indicating how/where to store the soft constraints

Returns

1 on successful extraction of the method, 0 on errors

SWIG Wrapper Notes This function is attached as method sc_add_SHAPE_deigan_ali() to objects of type fold

_compound

16.41.2.3 vrna_sc_add_SHAPE_zarringhalam()

#include <ViennaRNA/constraints/SHAPE.h>

Add SHAPE reactivity data as soft constraints (Zarringhalam et al. method)

This method first converts the observed SHAPE reactivity of nucleotide i into a probability q_i that position i is unpaired by means of a non-linear map. Then pseudo-energies of the form

$$\Delta G_{\text{SHAPE}}(x,i) = \beta |x_i - q_i|$$

are computed, where $x_i=0$ if position i is unpaired and $x_i=1$ if i is paired in a given secondary structure. The parameter β serves as scaling factor. The magnitude of discrepancy between prediction and experimental observation is represented by $|x_i-q_i|$.

See also

```
For further details, we refer to [25] vrna_sc_remove(), vrna_sc_add_SHAPE_deigan(), vrna_sc_minimize_pertubation()
```

Parameters

VC	The vrna_fold_compound_t the soft constraints are associated with
reactivities	A vector of normalized SHAPE reactivities
b	The scaling factor β of the conversion function
default_value	The default value for a nucleotide where reactivity data is missing for
shape_conversion	A flag that specifies how to convert reactivities to probabilities
options	The options flag indicating how/where to store the soft constraints

Returns

1 on successful extraction of the method, 0 on errors

SWIG Wrapper Notes This function is attached as method **sc_add_SHAPE_zarringhalam()** to objects of type fold_compound

16.41.2.4 vrna_sc_SHAPE_to_pr()

Convert SHAPE reactivity values to probabilities for being unpaired.

This function parses the informations from a given file and stores the result in the preallocated string sequence and the FLT_OR_DBL array values.

See also

```
vrna_file_SHAPE_read()
```

shape_conversion	String definining the method used for the conversion process
values	Pointer to an array of SHAPE reactivities
length	Length of the array of SHAPE reactivities
default_value	Result used for position with invalid/missing reactivity values

16.42 Generate Soft Constraints from Data

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

16.42.1 Detailed Description

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

Collaboration diagram for Generate Soft Constraints from Data:

Files

file perturbation_fold.h

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

Macros

#define VRNA_OBJECTIVE_FUNCTION_QUADRATIC 0

Use the sum of squared aberrations as objective function.

#define VRNA_OBJECTIVE_FUNCTION_ABSOLUTE 1

Use the sum of absolute aberrations as objective function.

• #define VRNA_MINIMIZER_DEFAULT 0

Use a custom implementation of the gradient descent algorithm to minimize the objective function.

• #define VRNA MINIMIZER CONJUGATE FR 1

Use the GNU Scientific Library implementation of the Fletcher-Reeves conjugate gradient algorithm to minimize the objective function.

#define VRNA_MINIMIZER_CONJUGATE_PR 2

Use the GNU Scientific Library implementation of the Polak-Ribiere conjugate gradient algorithm to minimize the objective function.

#define VRNA_MINIMIZER_VECTOR_BFGS 3

Use the GNU Scientific Library implementation of the vector Broyden-Fletcher-Goldfarb-Shanno algorithm to minimize the objective function.

#define VRNA_MINIMIZER_VECTOR_BFGS2 4

Use the GNU Scientific Library implementation of the vector Broyden-Fletcher-Goldfarb-Shanno algorithm to minimize the objective function.

#define VRNA MINIMIZER STEEPEST DESCENT 5

Use the GNU Scientific Library implementation of the steepest descent algorithm to minimize the objective function.

Typedefs

• typedef void(* progress_callback) (int iteration, double score, double *epsilon)

Callback for following the progress of the minimization process.

Functions

void vrna_sc_minimize_pertubation (vrna_fold_compound_t *vc, const double *q_prob_unpaired, int objective_function, double sigma_squared, double tau_squared, int algorithm, int sample_size, double *epsilon, double initialStepSize, double minStepSize, double minImprovement, double minimizerTolerance, progress_callback callback)

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

16.42.2 Macro Definition Documentation

16.42.2.1 VRNA_OBJECTIVE_FUNCTION_QUADRATIC

```
#define VRNA_OBJECTIVE_FUNCTION_QUADRATIC 0
```

#include <ViennaRNA/perturbation_fold.h>

Use the sum of squared aberrations as objective function.

$$F(\vec{\epsilon}) = \sum_{i=1}^{n} \frac{\epsilon_i^2}{\tau^2} + \sum_{i=1}^{n} \frac{(p_i(\vec{\epsilon}) - q_i)^2}{\sigma^2} \to min$$

16.42.2.2 VRNA_OBJECTIVE_FUNCTION_ABSOLUTE

```
#define VRNA_OBJECTIVE_FUNCTION_ABSOLUTE 1
```

#include <ViennaRNA/perturbation_fold.h>

Use the sum of absolute aberrations as objective function.

$$F(\vec{\epsilon}) = \sum_{i=1}^{n} \frac{|\epsilon_i|}{\tau^2} + \sum_{i=1}^{n} \frac{|p_i(\vec{\epsilon}) - q_i|}{\sigma^2} \to min$$

16.42.2.3 VRNA_MINIMIZER_CONJUGATE_FR

```
#define VRNA_MINIMIZER_CONJUGATE_FR 1
```

#include <ViennaRNA/perturbation_fold.h>

Use the GNU Scientific Library implementation of the Fletcher-Reeves conjugate gradient algorithm to minimize the objective function.

Please note that this algorithm can only be used when the GNU Scientific Library is available on your system

16.42.2.4 VRNA MINIMIZER CONJUGATE PR

```
#define VRNA_MINIMIZER_CONJUGATE_PR 2
```

```
#include <ViennaRNA/perturbation_fold.h>
```

Use the GNU Scientific Library implementation of the Polak-Ribiere conjugate gradient algorithm to minimize the objective function.

Please note that this algorithm can only be used when the GNU Scientific Library is available on your system

16.42.2.5 VRNA_MINIMIZER_VECTOR_BFGS

```
#define VRNA_MINIMIZER_VECTOR_BFGS 3
#include <ViennaRNA/perturbation_fold.h>
```

Use the GNU Scientific Library implementation of the vector Broyden-Fletcher-Goldfarb-Shanno algorithm to minimize the objective function.

Please note that this algorithm can only be used when the GNU Scientific Library is available on your system

16.42.2.6 VRNA_MINIMIZER_VECTOR_BFGS2

```
#define VRNA_MINIMIZER_VECTOR_BFGS2 4
#include <ViennaRNA/perturbation_fold.h>
```

Use the GNU Scientific Library implementation of the vector Broyden-Fletcher-Goldfarb-Shanno algorithm to minimize the objective function.

Please note that this algorithm can only be used when the GNU Scientific Library is available on your system

16.42.2.7 VRNA_MINIMIZER_STEEPEST_DESCENT

```
#define VRNA_MINIMIZER_STEEPEST_DESCENT 5
#include <ViennaRNA/perturbation_fold.h>
```

Use the GNU Scientific Library implementation of the steepest descent algorithm to minimize the objective function.

Please note that this algorithm can only be used when the GNU Scientific Library is available on your system

16.42.3 Typedef Documentation

16.42.3.1 progress_callback

```
typedef void(* progress_callback) (int iteration, double score, double *epsilon)
#include <ViennaRNA/perturbation_fold.h>
```

Callback for following the progress of the minimization process.

iteration	The number of the current iteration
score	The score of the objective function
epsilon	The perturbation vector yielding the reported score

16.42.4 Function Documentation

16.42.4.1 vrna sc minimize pertubation()

```
void vrna_sc_minimize_pertubation (
    vrna_fold_compound_t * vc,
    const double * q_prob_unpaired,
    int objective_function,
    double sigma_squared,
    double tau_squared,
    int algorithm,
    int sample_size,
    double * epsilon,
    double initialStepSize,
    double minImprovement,
    double minimizerTolerance,
    progress_callback callback)
```

#include <ViennaRNA/perturbation_fold.h>

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

Use an iterative minimization algorithm to find a vector of perturbation energies whose incorporation as soft constraints shifts the predicted pairing probabilities closer to the experimentally observed probabilities. The algorithm aims to minimize an objective function that penalizes discripancies between predicted and observed pairing probabilities and energy model adjustments, i.e. an appropriate vector of perturbation energies satisfies

$$F(\vec{\epsilon}) = \sum_{\mu} \frac{\epsilon_{\mu}^2}{\tau^2} + \sum_{i=1}^n \frac{(p_i(\vec{\epsilon}) - q_i)^2}{\sigma^2} \to \min.$$

An initialized fold compound and an array containing the observed probability for each nucleotide to be unbound are required as input data. The parameters objective_function, sigma_squared and tau_squared are responsible for adjusting the aim of the objective function. Dependend on which type of objective function is selected, either squared or absolute aberrations are contributing to the objective function. The ratio of the parameters sigma_\circ\ squared and tau_squared can be used to adjust the algorithm to find a solution either close to the thermodynamic prediction (sigma_squared >> tau_squared) or close to the experimental data (tau_squared >> sigma_squared). The minimization can be performed by makeing use of a custom gradient descent implementation or using one of the minimizing algorithms provided by the GNU Scientific Library. All algorithms require the evaluation of the gradient of the objective function, which includes the evaluation of conditional pairing probabilites. Since an exact evaluation is expensive, the probabilities can also be estimated from sampling by setting an appropriate sample size. The found vector of perturbation energies will be stored in the array epsilon. The progress of the minimization process can be tracked by implementing and passing a callback function.

See also

For further details we refere to [22].

V	/C	Pointer to a fold compound
---	----	----------------------------

q_prob_unpaired	Pointer to an array containing the probability to be unpaired for each nucleotide
objective_function	The type of objective function to be used (VRNA_OBJECTIVE_FUNCTION_QUADRATIC / VRNA_OBJECTIVE_FUNCTION_LINEAR)
sigma_squared	A factor used for weighting the objective function. More weight on this factor will lead to a solution close to the null vector.
tau_squared	A factor used for weighting the objective function. More weight on this factor will lead to a solution close to the data provided in q_prob_unpaired.
algorithm	The minimization algorithm (VRNA_MINIMIZER_*)
sample_size	The number of sampled sequences used for estimating the pairing probabilities. A value <= 0 will lead to an exact evaluation.
epsilon	A pointer to an array used for storing the calculated vector of perturbation energies
callback	A pointer to a callback function used for reporting the current minimization progress

16.43 Ligands Binding to RNA Structures

Simple Extensions to Model Ligand Binding to RNA Structures.

16.43.1 Detailed Description

Simple Extensions to Model Ligand Binding to RNA Structures.

Collaboration diagram for Ligands Binding to RNA Structures:

Modules

- Ligands Binding to Unstructured Domains

 Add ligand binding to loop regions using the Unstructured Domains feature.
- Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints

Files

• file ligand.h

Functions for incorporation of ligands binding to hairpin and interior loop motifs using the soft constraints framework.

16.44 Ligands Binding to Unstructured Domains

Add ligand binding to loop regions using the Unstructured Domains feature.

Add ligand binding to loop regions using the Unstructured Domains feature.

Sometime, certain ligands, like single strand binding (SSB) proteins, compete with intramolecular base pairing of the RNA. In situations, where the dissociation constant of the ligand is known and the ligand binds to a consecutive stretch of single-stranded nucleotides we can use the Unstructured Domains functionality to extend the RNA folding grammar. This module provides a convenience default implementation that covers most of the application scenarios.

The function vrna_ud_add_motif() attaches a ligands sequence motif and corresponding binding free energy to the list of known ligand motifs within a vrna_fold_compound_t.domains_up attribute. The first call to this function initializes the Unstructured Domains feature with our default implementation. Subsequent calls of secondary structure prediction algorithms with the modified vrna_fold_compound_t then directly include the competition of the ligand with regules base pairing. Since we utilize the unstructured domain extension, The ligand binding model can be removed again using the vrna_ud_remove() function. Collaboration diagram for Ligands Binding to Unstructured Domains:

16.45 Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints

16.45.1 Detailed Description

Collaboration diagram for Incorporating Ligands Binding to Specific Sequence/Structure Motifs using Soft Constraints:

Functions

int vrna_sc_add_hi_motif (vrna_fold_compound_t *vc, const char *seq, const char *structure, FLT_OR_DBL energy, unsigned int options)

Add soft constraints for hairpin or interior loop binding motif.

16.45.2 Function Documentation

16.45.2.1 vrna_sc_add_hi_motif()

#include <ViennaRNA/constraints/ligand.h>

Add soft constraints for hairpin or interior loop binding motif.

Parameters

VC	The vrna_fold_compound_t the motif is applied to
seq	The sequence motif (may be interspaced by '&' character
structure	The structure motif (may be interspaced by '&' character
energy	The free energy of the motif (e.g. binding free energy)
options	Options

Returns

non-zero value if application of the motif using soft constraints was successful

SWIG Wrapper Notes This function is attached as method sc_add_hi_motif() to objects of type fold_compound

16.46 Complex Structured Modules

16.46.1 Detailed Description

Collaboration diagram for Complex Structured Modules:

Modules

• G-Quadruplexes

Various functions related to G-quadruplex computations.

Files

• file gquad.h

G-quadruplexes.

16.47 G-Quadruplexes 419

16.47 G-Quadruplexes

Various functions related to G-quadruplex computations.

16.47.1 Detailed Description

Various functions related to G-quadruplex computations.

Collaboration diagram for G-Quadruplexes:

Functions

- int * get_gquad_matrix (short *S, vrna_param_t *P)
 Get a triangular matrix prefilled with minimum free energy contributions of G-quadruplexes.
- int parse_gquad (const char *struc, int *L, int I[3])
- PRIVATE int backtrack_GQuad_IntLoop (int c, int i, int j, int type, short *S, int *ggg, int *index, int *p, int *q, vrna_param_t *P)
- PRIVATE int backtrack_GQuad_IntLoop_L (int c, int i, int j, int type, short *S, int **ggg, int maxdist, int *p, int *q, vrna_param_t *P)

16.47.2 Function Documentation

16.47.2.1 get_gquad_matrix()

#include <ViennaRNA/gquad.h>

Get a triangular matrix prefilled with minimum free energy contributions of G-quadruplexes.

At each position ij in the matrix, the minimum free energy of any G-quadruplex delimited by i and j is stored. If no G-quadruplex formation is possible, the matrix element is set to INF. Access the elements in the matrix via matrix[indx[j]+i]. To get the integer array indx see get_jindx().

See also

```
get_jindx(), encode_sequence()
```

S	The encoded sequence
Р	A pointer to the data structure containing the precomputed energy contributions

Returns

A pointer to the G-quadruplex contribution matrix

16.47.2.2 parse_gquad()

```
int parse_gquad (  \mbox{const char} \ * \ struc, \\ \mbox{int} \ * \ L, \\ \mbox{int} \ 1 [3] \ )
```

#include <ViennaRNA/gquad.h>

given a dot-bracket structure (possibly) containing gquads encoded by '+' signs, find first gquad, return end position or 0 if none found Upon return L and I[] contain the number of stacked layers, as well as the lengths of the linker regions. To parse a string with many gquads, call parse_gquad repeatedly e.g. end1 = parse_gquad(struc, &L, I); ...; end2 = parse_gquad(struc+end1, &L, I); end2+=end1; ...; end3 = parse_gquad(struc+end2, &L, I); end3+=end2; ...;

16.47.2.3 backtrack_GQuad_IntLoop()

#include <ViennaRNA/gquad.h>

backtrack an interior loop like enclosed g-quadruplex with closing pair (i,j)

С	The total contribution the loop should resemble
i	position i of enclosing pair
j	position j of enclosing pair
type	base pair type of enclosing pair (must be reverse type)
S	integer encoded sequence
999	triangular matrix containing g-quadruplex contributions
index	the index for accessing the triangular matrix
р	here the 5' position of the gquad is stored
q	here the 3' position of the gquad is stored
P	

16.47 G-Quadruplexes 421

Returns

1 on success, 0 if no gquad found

16.47.2.4 backtrack_GQuad_IntLoop_L()

#include <ViennaRNA/gquad.h>

backtrack an interior loop like enclosed g-quadruplex with closing pair (i,j) with underlying Lfold matrix

Parameters

С	The total contribution the loop should resemble
i	position i of enclosing pair
j	position j of enclosing pair
type	base pair type of enclosing pair (must be reverse type)
S	integer encoded sequence
999	triangular matrix containing g-quadruplex contributions
р	here the 5' position of the gquad is stored
q	here the 3' position of the gquad is stored
Р	the datastructure containing the precalculated contibutions

Returns

1 on success, 0 if no gquad found

16.48 Utilities

16.48.1 Detailed Description

Collaboration diagram for Utilities:

Modules

· Utilities to deal with Nucleotide Alphabets

Functions to cope with various aspects related to the nucleotide sequence alphabet.

· (Nucleic Acid Sequence) String Utilitites

Functions to parse, convert, manipulate, create, and compare (nucleic acid sequence) strings.

· Secondary Structure Utilities

Functions to create, parse, convert, manipulate, and compare secondary structure representations.

· Multiple Sequence Alignment Utilities

Functions to extract features from and to manipulate multiple sequence alignments.

· Files and I/O

Functions to parse, write, and convert various file formats and to deal with file system related issues.

Plotting

Functions for Creating Secondary Structure Plots, Dot-Plots, and More.

· Search Algorithms

Implementations of various search algorithms to detect strings of objects within other strings of objects.

Combinatorics Algorithms

Implementations to solve various combinatorial aspects for strings of objects.

(Abstract) Data Structures

All datastructures and typedefs shared among the ViennaRNA Package can be found here.

Messages

Functions to print various kind of messages.

• Unit Conversion

Functions to convert between various physical units.

Files

· file alphabet.h

Functions to process, convert, and generally handle different nucleotide and/or base pair alphabets.

· file combinatorics.h

Various implementations that deal with combinatorial aspects of objects.

· file commands.h

Parse and apply different commands that alter the behavior of secondary structure prediction and evaluation.

· file sequence.h

 $\label{prop:continuous} \textit{Functions and data structures related to sequence representations} \; ,.$

· file units.h

Physical Units and Functions to convert them into each other.

• file file_formats_msa.h

Functions dealing with file formats for Multiple Sequence Alignments (MSA)

· file utils.h

Several utilities for file handling.

file utils.h

16.48 Utilities 423

Various utilities to assist in plotting secondary structures and consensus structures.

· file alignments.h

Various utility- and helper-functions for sequence alignments and comparative structure prediction.

· file basic.h

General utility- and helper-functions used throughout the ViennaRNA Package.

· file strings.h

General utility- and helper-functions for RNA sequence and structure strings used throughout the ViennaRNA Package.

• file BoyerMoore.h

Variants of the Boyer-Moore string search algorithm.

· file char_stream.h

Implementation of a dynamic, buffered character stream.

· file stream_output.h

An implementation of a buffered, ordered stream output data structure.

Macros

#define VRNA INPUT ERROR 1U

Output flag of get_input_line(): "An ERROR has occured, maybe EOF".

• #define VRNA INPUT QUIT 2U

Output flag of get_input_line(): "the user requested quitting the program".

• #define VRNA INPUT MISC 4U

Output flag of get_input_line(): "something was read".

• #define VRNA INPUT FASTA HEADER 8U

Input/Output flag of get_input_line():

if used as input option this tells get_input_line() that the data to be read should comply with the FASTA format.

• #define VRNA_INPUT_CONSTRAINT 32U

Input flag for get_input_line():

Tell get_input_line() that we assume to read a structure constraint.

#define VRNA INPUT NO TRUNCATION 256U

Input switch for get_input_line(): "do not trunkate the line by eliminating white spaces at end of line".

• #define VRNA INPUT NO REST 512U

Input switch for vrna_file_fasta_read_record(): "do fill rest array".

#define VRNA_INPUT_NO_SPAN 1024U

Input switch for vrna_file_fasta_read_record(): "never allow data to span more than one line".

#define VRNA_INPUT_NOSKIP_BLANK_LINES 2048U

Input switch for vrna_file_fasta_read_record(): "do not skip empty lines".

• #define VRNA_INPUT_BLANK_LINE 4096U

Output flag for vrna_file_fasta_read_record(): "read an empty line".

#define VRNA_INPUT_NOSKIP_COMMENTS 128U

Input switch for get_input_line(): "do not skip comment lines".

#define VRNA INPUT COMMENT 8192U

Output flag for vrna_file_fasta_read_record(): "read a comment".

#define MIN2(A, B) ((A) < (B) ? (A) : (B))

Get the minimum of two comparable values.

#define MAX2(A, B) ((A) > (B) ? (A) : (B))

Get the maximum of two comparable values.

#define MIN3(A, B, C) (MIN2((MIN2((A), (B))), (C)))

Get the minimum of three comparable values.

#define MAX3(A, B, C) (MAX2((MAX2((A), (B))), (C)))

Get the maximum of three comparable values.

Functions

void * vrna_alloc (unsigned size)

Allocate space safely.

• void * vrna_realloc (void *p, unsigned size)

Reallocate space safely.

• void vrna_init_rand (void)

Initialize seed for random number generator.

double vrna_urn (void)

get a random number from [0..1]

• int vrna_int_urn (int from, int to)

Generates a pseudo random integer in a specified range.

char * vrna_time_stamp (void)

Get a timestamp.

- unsigned int get input line (char **string, unsigned int options)
- int * vrna_idx_row_wise (unsigned int length)

Get an index mapper array (iindx) for accessing the energy matrices, e.g. in partition function related functions.

int * vrna_idx_col_wise (unsigned int length)

Get an index mapper array (indx) for accessing the energy matrices, e.g. in MFE related functions.

Variables

• unsigned short xsubi [3]

Current 48 bit random number.

16.48.2 Macro Definition Documentation

```
16.48.2.1 VRNA_INPUT_FASTA_HEADER
```

```
#define VRNA_INPUT_FASTA_HEADER 8U
```

```
#include <ViennaRNA/utils/basic.h>
```

Input/Output flag of get_input_line():

if used as input option this tells get_input_line() that the data to be read should comply with the FASTA format.

the function will return this flag if a fasta header was read

16.48.2.2 VRNA_INPUT_CONSTRAINT

```
#define VRNA_INPUT_CONSTRAINT 32U
```

#include <ViennaRNA/utils/basic.h>

Input flag for get_input_line():

Tell get_input_line() that we assume to read a structure constraint.

16.48 Utilities 425

16.48.3 Function Documentation

```
16.48.3.1 vrna_alloc()
```

Allocate space safely.

Parameters

Returns

A pointer to the allocated memory

16.48.3.2 vrna_realloc()

Reallocate space safely.

Parameters

р	A pointer to the memory region to be reallocated
size	The size of the memory to be allocated in bytes

Returns

A pointer to the newly allocated memory

```
16.48.3.3 vrna_urn()
```

See also

```
vrna_int_urn(), vrna_init_rand()
```

Note

Usually implemented by calling erand48().

Returns

A random number in range [0..1]

16.48.3.4 vrna_int_urn()

```
int vrna_int_urn (
          int from,
          int to )
```

#include <ViennaRNA/utils/basic.h>

Generates a pseudo random integer in a specified range.

See also

```
vrna_urn(), vrna_init_rand()
```

Parameters

from	The first number in range
to	The last number in range

Returns

A pseudo random number in range [from, to]

16.48.3.5 vrna_time_stamp()

Get a timestamp.

Returns a string containing the current date in the format

```
Fri Mar 19 21:10:57 1993
```

16.48 Utilities 427

Returns

A string containing the timestamp

16.48.3.6 get_input_line()

```
#include <ViennaRNA/utils/basic.h>
```

Retrieve a line from 'stdin' savely while skipping comment characters and other features This function returns the type of input it has read if recognized. An option argument allows one to switch between different reading modes. Currently available options are:

pass a collection of options as one value like this:

```
get_input_line(string, option_1 | option_2 | option_n)
```

If the function recognizes the type of input, it will report it in the return value. It also reports if a user defined 'quit' command (-sign on 'stdin') was given. Possible return values are:

VRNA_INPUT_FASTA_HEADER, VRNA_INPUT_ERROR, VRNA_INPUT_MISC, VRNA_INPUT_QUIT

Parameters

string	A pointer to the character array that contains the line read
options	A collection of options for switching the functions behavior

Returns

A flag with information about what has been read

16.48.3.7 vrna_idx_row_wise()

Get an index mapper array (iindx) for accessing the energy matrices, e.g. in partition function related functions.

Access of a position "(i,j)" is then accomplished by using

```
(i,j) \sim iindx[i]-j
```

This function is necessary as most of the two-dimensional energy matrices are actually one-dimensional arrays throughout the ViennaRNA Package

Consult the implemented code to find out about the mapping formula;)

See also

```
vrna_idx_col_wise()
```

Parameters

```
length The length of the RNA sequence
```

Returns

The mapper array

16.48.3.8 vrna_idx_col_wise()

```
#include <ViennaRNA/utils/basic.h>
```

Get an index mapper array (indx) for accessing the energy matrices, e.g. in MFE related functions.

Access of a position "(i,j)" is then accomplished by using

```
(i,j) \sim indx[j]+i
```

This function is necessary as most of the two-dimensional energy matrices are actually one-dimensional arrays throughout the ViennaRNAPackage

Consult the implemented code to find out about the mapping formula;)

See also

```
vrna_idx_row_wise()
```

Parameters

length	The length of the RNA sequence

Returns

The mapper array

16.48.4 Variable Documentation

16.48 Utilities 429

16.48.4.1 xsubi

```
unsigned short xsubi[3]
#include <ViennaRNA/utils/basic.h>
```

Current 48 bit random number.

This variable is used by vrna_urn(). These should be set to some random number seeds before the first call to vrna_urn().

See also

vrna_urn()

16.49 Exterior Loops

Functions to evaluate the free energy contributions for exterior loops.

16.49.1 Detailed Description

Functions to evaluate the free energy contributions for exterior loops.

Collaboration diagram for Exterior Loops:

Files

· file external.h

Energy evaluation of exterior loops for MFE and partition function calculations.

Basic free energy interface

- int vrna_E_ext_stem (unsigned int type, int n5d, int n3d, vrna_param_t *p)
 - Evaluate a stem branching off the exterior loop.
- int vrna_E_ext_loop (vrna_fold_compound_t *fc, int i, int j)

Evaluate the free energy of a base pair in the exterior loop.

- int vrna_E_ext_loop_5 (vrna_fold_compound_t *fc)
- int vrna_E_ext_loop_3 (vrna_fold_compound_t *fc, int i)

Boltzmann weight (partition function) interface

- typedef struct vrna_mx_pf_aux_el_s * vrna_mx_pf_aux_el_t
 - Auxiliary helper arrays for fast exterior loop computations.
- FLT_OR_DBL vrna_exp_E_ext_stem (unsigned int type, int n5d, int n3d, vrna_exp_param_t *p)

Evaluate a stem branching off the exterior loop (Boltzmann factor version)

- struct vrna_mx_pf_aux_el_s * vrna_exp_E_ext_fast_init (vrna_fold_compound_t *fc)
- void vrna_exp_E_ext_fast_rotate (struct vrna_mx_pf_aux_el_s *aux_mx)
- void vrna_exp_E_ext_fast_free (struct vrna_mx_pf_aux_el_s *aux_mx)
- FLT_OR_DBL vrna_exp_E_ext_fast (vrna_fold_compound_t *fc, int i, int j, struct vrna_mx_pf_aux_el_s *aux_mx)
- void vrna_exp_E_ext_fast_update (vrna_fold_compound_t *fc, int j, struct vrna_mx_pf_aux_el_s *aux_←
 mx)

16.49.2 Typedef Documentation

16.49 Exterior Loops 431

```
16.49.2.1 vrna_mx_pf_aux_el_t
```

```
typedef struct vrna_mx_pf_aux_el_s* vrna_mx_pf_aux_el_t
```

#include <ViennaRNA/loops/external.h>

Auxiliary helper arrays for fast exterior loop computations.

See also

```
vrna_exp_E_ext_fast_init(), vrna_exp_E_ext_fast_rotate(), vrna_exp_E_ext_fast_free(), vrna_exp_E_ext_← fast()
```

16.49.3 Function Documentation

16.49.3.1 vrna_E_ext_stem()

```
int vrna_E_ext_stem (
          unsigned int type,
          int n5d,
          int n3d,
          vrna_param_t * p )
```

#include <ViennaRNA/loops/external.h>

Evaluate a stem branching off the exterior loop.

Given a base pair (i,j) encoded by $\it type$, compute the energy contribution including dangling-end/terminal-mismatch contributions. Instead of returning the energy contribution per-se, this function returns the corresponding Boltzmann factor. If either of the adjacent nucleotides (i-1) and (j+1) must not contribute stacking energy, the corresponding encoding must be -1.

See also

```
vrna_E_exp_stem()
```

Parameters

type	The base pair encoding
n5d	The encoded nucleotide directly adjacent at the 5' side of the base pair (may be -1)
n3d	The encoded nucleotide directly adjacent at the 3' side of the base pair (may be -1)
р	The pre-computed energy parameters

Returns

The energy contribution of the introduced exterior-loop stem

16.49.3.2 vrna_E_ext_loop()

#include <ViennaRNA/loops/external.h>

Evaluate the free energy of a base pair in the exterior loop.

Evalue the free energy of a base pair connecting two nucleotides in the exterior loop and take hard constraints into account.

Typically, this is simply dangling end contributions of the adjacent nucleotides, potentially a terminal A-U mismatch penalty, and maybe some generic soft constraint contribution for that decomposition.

Note

For dangles $== 1 \mid \mid 3$ this function also evaluates the three additional pairs (i + 1, j), (i, j - 1), and (i + 1, j - 1) and returns the minimum for all four possibilities in total.

Parameters

	fc	Fold compound to work on (defines the model and parameters)
	i	5' position of the base pair
Γ.	j	3' position of the base pair

Returns

Free energy contribution that arises when this pair is formed in the exterior loop

16.49.3.3 vrna_exp_E_ext_stem()

```
FLT_OR_DBL vrna_exp_E_ext_stem (
          unsigned int type,
          int n5d,
          int n3d,
          vrna_exp_param_t * p )
```

#include <ViennaRNA/loops/external.h>

Evaluate a stem branching off the exterior loop (Boltzmann factor version)

Given a base pair (i,j) encoded by $\it type$, compute the energy contribution including dangling-end/terminal-mismatch contributions. Instead of returning the energy contribution per-se, this function returns the corresponding Boltzmann factor. If either of the adjacent nucleotides (i-1) and (j+1) must not contribute stacking energy, the corresponding encoding must be -1.

See also

```
vrna_E_ext_stem()
```

16.49 Exterior Loops 433

Parameters

type	The base pair encoding
n5d	The encoded nucleotide directly adjacent at the 5' side of the base pair (may be -1)
n3d	The encoded nucleotide directly adjacent at the 3' side of the base pair (may be -1)
р	The pre-computed energy parameters (Boltzmann factor version)

Returns

The Boltzmann weighted energy contribution of the introduced exterior-loop stem

16.50 Hairpin Loops

Functions to evaluate the free energy contributions for hairpin loops.

16.50.1 Detailed Description

Functions to evaluate the free energy contributions for hairpin loops.

Collaboration diagram for Hairpin Loops:

Files

· file hairpin.h

Energy evaluation of hairpin loops for MFE and partition function calculations.

Basic free energy interface

int vrna_E_hp_loop (vrna_fold_compound_t *fc, int i, int j)

Evaluate the free energy of a hairpin loop and consider hard constraints if they apply.

• int vrna_E_ext_hp_loop (vrna_fold_compound_t *fc, int i, int j)

Evaluate the free energy of an exterior hairpin loop and consider possible hard constraints.

int vrna_eval_ext_hp_loop (vrna_fold_compound_t *fc, int i, int j)

Evaluate free energy of an exterior hairpin loop.

int vrna_eval_hp_loop (vrna_fold_compound_t *fc, int i, int j)

Evaluate free energy of a hairpin loop.

PRIVATE int E_Hairpin (int size, int type, int si1, int sj1, const char *string, vrna_param_t *P)

Compute the Energy of a hairpin-loop.

Boltzmann weight (partition function) interface

PRIVATE FLT_OR_DBL exp_E_Hairpin (int u, int type, short si1, short sj1, const char *string, vrna_exp_param_t *P)

Compute Boltzmann weight $e^{-\Delta G/kT}$ of a hairpin loop.

• FLT_OR_DBL vrna_exp_E_hp_loop (vrna_fold_compound_t *fc, int i, int j)

High-Level function for hairpin loop energy evaluation (partition function variant)

16.50.2 Function Documentation

16.50 Hairpin Loops 435

16.50.2.1 vrna_E_hp_loop()

#include <ViennaRNA/loops/hairpin.h>

Evaluate the free energy of a hairpin loop and consider hard constraints if they apply.

This function evaluates the free energy of a hairpin loop

In case the base pair is not allowed due to a constraint conflict, this function returns INF.

Note

This function is polymorphic! The provided vrna_fold_compound_t may be of type VRNA_FC_TYPE_SINGLE or VRNA_FC_TYPE_COMPARATIVE

Parameters

fc	The vrna_fold_compound_t that stores all relevant model settings
i	The 5' nucleotide of the base pair (3' to evaluate the pair as exterior hairpin loop)
j	The 3' nucleotide of the base pair (5' to evaluate the pair as exterior hairpin loop)

Returns

The free energy of the hairpin loop in 10cal/mol

16.50.2.2 vrna_E_ext_hp_loop()

#include <ViennaRNA/loops/hairpin.h>

Evaluate the free energy of an exterior hairpin loop and consider possible hard constraints.

Note

This function is polymorphic! The provided vrna_fold_compound_t may be of type VRNA_FC_TYPE_SINGLE or VRNA_FC_TYPE_COMPARATIVE

16.50.2.3 vrna_eval_hp_loop()

#include <ViennaRNA/loops/hairpin.h>

Evaluate free energy of a hairpin loop.

Note

This function is polymorphic! The provided vrna_fold_compound_t may be of type VRNA_FC_TYPE_SINGLE or VRNA_FC_TYPE_COMPARATIVE

Parameters

fc	The vrna_fold_compound_t for the particular energy evaluation
i	5'-position of the base pair
j	3'-position of the base pair

Returns

Free energy of the hairpin loop closed by (i, j) in deka-kal/mol

SWIG Wrapper Notes This function is attached as method eval_hp_loop() to objects of type fold_compound

16.50.2.4 E_Hairpin()

#include <ViennaRNA/loops/hairpin.h>

Compute the Energy of a hairpin-loop.

To evaluate the free energy of a hairpin-loop, several parameters have to be known. A general hairpin-loop has this structure:

```
a3 a4
a2 a5
a1 a6
X - Y
I I
5' 3'
```

16.50 Hairpin Loops 437

where X-Y marks the closing pair [e.g. a (G,C) pair]. The length of this loop is 6 as there are six unpaired nucleotides (a1-a6) enclosed by (X,Y). The 5' mismatching nucleotide is a1 while the 3' mismatch is a6. The nucleotide sequence of this loop is "a1.a2.a3.a4.a5.a6"

Note

The parameter sequence should contain the sequence of the loop in capital letters of the nucleic acid alphabet if the loop size is below 7. This is useful for unusually stable tri-, tetra- and hexa-loops which are treated differently (based on experimental data) if they are tabulated.

See also

```
scale_parameters()
vrna_param_t
```

Warning

Not (really) thread safe! A threadsafe implementation will replace this function in a future release! Energy evaluation may change due to updates in global variable "tetra_loop"

Parameters

size	The size of the loop (number of unpaired nucleotides)
type	The pair type of the base pair closing the hairpin
si1	The 5'-mismatching nucleotide
sj1	The 3'-mismatching nucleotide
string	The sequence of the loop (May be \mathtt{NULL} , otherwise mst be at least $size + 2$ long)
Р	The datastructure containing scaled energy parameters

Returns

The Free energy of the Hairpin-loop in dcal/mol

16.50.2.5 exp_E_Hairpin()

#include <ViennaRNA/loops/hairpin.h>

Compute Boltzmann weight $e^{-\Delta G/kT}$ of a hairpin loop.

multiply by scale[u+2]

See also

```
get_scaled_pf_parameters()
vrna_exp_param_t
E_Hairpin()
```

Warning

Not (really) thread safe! A threadsafe implementation will replace this function in a future release! Energy evaluation may change due to updates in global variable "tetra_loop"

Parameters

и	The size of the loop (number of unpaired nucleotides)
type	The pair type of the base pair closing the hairpin
si1	The 5'-mismatching nucleotide
sj1	The 3'-mismatching nucleotide
string	The sequence of the loop (May be \mathtt{NULL} , otherwise mst be at least $size + 2$ long)
Р	The datastructure containing scaled Boltzmann weights of the energy parameters

Returns

The Boltzmann weight of the Hairpin-loop

```
16.50.2.6 vrna_exp_E_hp_loop()
```

#include <ViennaRNA/loops/hairpin.h>

High-Level function for hairpin loop energy evaluation (partition function variant)

See also

```
vrna_E_hp_loop() for it's free energy counterpart
```

Note

This function is polymorphic! The provided vrna_fold_compound_t may be of type VRNA_FC_TYPE_SINGLE or VRNA_FC_TYPE_COMPARATIVE

16.51 Internal Loops 439

16.51 Internal Loops

Functions to evaluate the free energy contributions for internal loops.

16.51.1 Detailed Description

Functions to evaluate the free energy contributions for internal loops.

Collaboration diagram for Internal Loops:

Files

· file internal.h

Energy evaluation of interior loops for MFE and partition function calculations.

Basic free energy interface

```
• int vrna E int loop (vrna fold compound t *fc, int i, int j)
```

```
• int vrna_eval_int_loop (vrna_fold_compound_t *fc, int i, int j, int k, int l)
```

Evaluate the free energy contribution of an interior loop with delimiting base pairs (i, j) and (k, l).

- int vrna E ext int loop (vrna fold compound t *fc, int i, int j, int *ip, int *iq)
- int vrna_E_stack (vrna_fold_compound_t *fc, int i, int j)

Boltzmann weight (partition function) interface

```
• FLT OR DBL vrna exp E int loop (vrna fold compound t *fc, int i, int i)
```

```
• FLT_OR_DBL vrna_exp_E_interior_loop (vrna_fold_compound_t *fc, int i, int j, int k, int l)
```

16.51.2 Function Documentation

```
16.51.2.1 vrna_eval_int_loop()
```

#include <ViennaRNA/loops/internal.h>

Evaluate the free energy contribution of an interior loop with delimiting base pairs (i, j) and (k, l).

Note

This function is polymorphic, i.e. it accepts vrna_fold_compound_t of type VRNA_FC_TYPE_SINGLE as well as VRNA_FC_TYPE_COMPARATIVE

SWIG Wrapper Notes This function is attached as method eval int loop() to objects of type fold compound

16.52 Multibranch Loops

Functions to evaluate the free energy contributions for mullibranch loops.

16.52.1 Detailed Description

Functions to evaluate the free energy contributions for mutlibranch loops.

Collaboration diagram for Multibranch Loops:

Files

· file multibranch.h

Energy evaluation of multibranch loops for MFE and partition function calculations.

Basic free energy interface

- int vrna_E_mb_loop_stack (vrna_fold_compound_t *fc, int i, int j)
 Evaluate energy of a multi branch helices stacking onto closing pair (i,j)
- int vrna_E_mb_loop_fast (vrna_fold_compound_t *fc, int i, int j, int *dmli1, int *dmli2)
- int E ml rightmost stem (int i, int j, vrna fold compound t *fc)
- int vrna_E_ml_stems_fast (vrna_fold_compound_t *fc, int i, int j, int *fmi, int *dmli)

Boltzmann weight (partition function) interface

- typedef struct vrna_mx_pf_aux_ml_s * vrna_mx_pf_aux_ml_t
 - Auxiliary helper arrays for fast exterior loop computations.
- FLT_OR_DBL vrna_exp_E_mb_loop_fast (vrna_fold_compound_t *fc, int i, int j, vrna_mx_pf_aux_ml_t aux mx)
- vrna_mx_pf_aux_ml_t vrna_exp_E_ml_fast_init (vrna_fold_compound_t *fc)
- void vrna_exp_E_ml_fast_rotate (vrna_mx_pf_aux_ml_t aux_mx)
- void vrna_exp_E_ml_fast_free (vrna_mx_pf_aux_ml_t aux_mx)
- const FLT_OR_DBL * vrna_exp_E_ml_fast_qqm (struct vrna_mx_pf_aux_ml_s *aux_mx)
- const FLT_OR_DBL * vrna_exp_E_ml_fast_qqm1 (struct vrna_mx_pf_aux_ml_s *aux_mx)
- FLT_OR_DBL vrna_exp_E_ml_fast (vrna_fold_compound_t *fc, int i, int j, vrna_mx_pf_aux_ml_t aux_mx)

16.52.2 Typedef Documentation

```
16.52.2.1 vrna_mx_pf_aux_ml_t
typedef struct vrna_mx_pf_aux_ml_s* vrna_mx_pf_aux_ml_t
#include <ViennaRNA/loops/multibranch.h>
```

Auxiliary helper arrays for fast exterior loop computations.

See also

vrna_exp_E_ml_fast_init(), vrna_exp_E_ml_fast_rotate(), vrna_exp_E_ml_fast_free(), vrna_exp_E_ml_fast()

16.52.3 Function Documentation

```
16.52.3.1 vrna_E_mb_loop_stack()
```

#include <ViennaRNA/loops/multibranch.h>

Evaluate energy of a multi branch helices stacking onto closing pair (i,j)

Computes total free energy for coaxial stacking of (i.j) with (i+1.k) or (k+1.j-1)

16.53 Partition Function for Two Hybridized Sequences

Partition Function Cofolding.

16.53.1 Detailed Description

Partition Function Cofolding.

To simplify the implementation the partition function computation is done internally in a null model that does not include the duplex initiation energy, i.e. the entropic penalty for producing a dimer from two monomers). The resulting free energies and pair probabilities are initially relative to that null model. In a second step the free energies can be corrected to include the dimerization penalty, and the pair probabilities can be divided into the conditional pair probabilities given that a re dimer is formed or not formed. See [2] for further details.

As for folding one RNA molecule, this computes the partition function of all possible structures and the base pair probabilities. Uses the same global pf_scale variable to avoid overflows.

After computing the partition functions of all possible dimeres one can compute the probabilities of base pairs, the concentrations out of start concentrations and sofar and soaway.

Dimer formation is inherently concentration dependent. Given the free energies of the monomers A and B and dimers AB, AA, and BB one can compute the equilibrium concentrations, given input concentrations of A and B, see e.g. Dimitrov & Zuker (2004) Collaboration diagram for Partition Function for Two Hybridized Sequences:

Files

· file concentrations.h

Concentration computations for RNA-RNA interactions.

file part_func_up.h

Implementations for accessibility and RNA-RNA interaction as a stepwise process.

Typedefs

typedef struct vrna_dimer_pf_s vrna_dimer_pf_t

Typename for the data structure that stores the dimer partition functions, vrna_dimer_pf_s, as returned by vrna_pf_dimer()

· typedef struct vrna_dimer_pf_s cofoldF

Backward compatibility typedef for vrna_dimer_pf_s.

Variables

· int mirnatog

Toggles no intrabp in 2nd mol.

• double F_monomer [2]

Free energies of the two monomers.

typedef struct vrna_dimer_conc_s vrna_dimer_conc_t

Typename for the data structure that stores the dimer concentrations, vrna_dimer_conc_s, as required by vrna_pf← _dimer_concentration()

• typedef struct vrna_dimer_conc_s ConcEnt

Backward compatibility typedef for vrna_dimer_conc_s.

• vrna_dimer_conc_t * vrna_pf_dimer_concentrations (double FcAB, double FcAA, double FcBB, double FEA, double FEB, const double *startconc, const vrna_exp_param_t *exp_params)

Given two start monomer concentrations a and b, compute the concentrations in thermodynamic equilibrium of all dimers and the monomers.

Simplified global partition function computation using sequence(s) or multiple sequence alignment(s)

vrna_dimer_pf_t vrna_pf_co_fold (const char *seq, char *structure, vrna_ep_t **pl)
 Calculate partition function and base pair probabilities of nucleic acid/nucleic acid dimers.

16.53.2 Function Documentation

16.53.2.1 vrna_pf_co_fold()

Calculate partition function and base pair probabilities of nucleic acid/nucleic acid dimers.

This simplified interface to vrna_pf_dimer() computes the partition function and, if required, base pair probabilities for an RNA-RNA interaction using default options. Memory required for dynamic programming (DP) matrices will be allocated and free'd on-the-fly. Hence, after return of this function, the recursively filled matrices are not available any more for any post-processing.

Note

In case you want to use the filled DP matrices for any subsequent post-processing step, or you require other conditions than specified by the default model details, use vrna_pf_dimer(), and the data structure vrna_fold_compound_tinstead.

See also

```
vrna pf dimer()
```

Parameters

seq	Two concatenated RNA sequences with a delimiting '&' in between
structure	A pointer to the character array where position-wise pairing propensity will be stored. (Maybe NULL)
pl	A pointer to a list of vrna_ep_t to store pairing probabilities (Maybe NULL)

Returns

vrna dimer pf t structure containing a set of energies needed for concentration computations.

16.53.2.2 vrna_pf_dimer_concentrations()

#include <ViennaRNA/concentrations.h>

Given two start monomer concentrations a and b, compute the concentrations in thermodynamic equilibrium of all

This function takes an array 'startconc' of input concentrations with alternating entries for the initial concentrations of molecules A and B (terminated by two zeroes), then computes the resulting equilibrium concentrations from the free energies for the dimers. Dimer free energies should be the dimer-only free energies, i.e. the FcAB entries from the vrna_dimer_pf_t struct.

Parameters

dimers and the monomers.

FcAB	Free energy of AB dimer (FcAB entry)
FcAA	Free energy of AA dimer (FcAB entry)
FcBB	Free energy of BB dimer (FcAB entry)
FEA	Free energy of monomer A
FEB	Free energy of monomer B
startconc	List of start concentrations [a0],[b0],[a1],[b1],,[an][bn],[0],[0]
exp_params	The precomputed Boltzmann factors

Returns

vrna_dimer_conc_t array containing the equilibrium energies and start concentrations

16.54 Partition Function for two Hybridized Sequences as a Stepwise Process

RNA-RNA interaction as a stepwise process.

16.54.1 Detailed Description

RNA-RNA interaction as a stepwise process.

In this approach to cofolding the interaction between two RNA molecules is seen as a stepwise process. In a first step, the target molecule has to adopt a structure in which a binding site is accessible. In a second step, the ligand molecule will hybridize with a region accessible to an interaction. Consequently the algorithm is designed as a two step process: The first step is the calculation of the probability that a region within the target is unpaired, or equivalently, the calculation of the free energy needed to expose a region. In the second step we compute the free energy of an interaction for every possible binding site. Collaboration diagram for Partition Function for two Hybridized Sequences as a Stepwise Process:

Functions

- pu_contrib * pf_unstru (char *sequence, int max_w)
 Calculate the partition function over all unpaired regions of a maximal length.
- interact * pf_interact (const char *s1, const char *s2, pu_contrib *p_c, pu_contrib *p_c2, int max_w, char *cstruc, int incr3, int incr5)

Calculates the probability of a local interaction between two sequences.

void free_interact (interact *pin)

Frees the output of function pf_interact().

void free_pu_contrib_struct (pu_contrib *pu)

Frees the output of function pf_unstru().

16.54.2 Function Documentation

16.54.2.1 pf_unstru()

Calculate the partition function over all unpaired regions of a maximal length.

You have to call function pf_fold() providing the same sequence before calling pf_unstru(). If you want to calculate unpaired regions for a constrained structure, set variable 'structure' in function 'pf_fold()' to the constrain string. It returns a pu_contrib struct containing four arrays of dimension [i = 1 to length(sequence)][j = 0 to u-1] containing all possible contributions to the probabilities of unpaired regions of maximum length u. Each array in pu_contrib contains one of the contributions to the total probability of being unpaired: The probability of being unpaired within an exterior loop is in array pu_contrib->E, the probability of being unpaired within a hairpin loop is in array pu_contrib->H, the probability of being unpaired within an interior loop is in array pu_contrib->I and probability of being unpaired within a multi-loop is in array pu_contrib->M. The total probability of being unpaired is the sum of the four arrays of pu_contrib.

This function frees everything allocated automatically. To free the output structure call free_pu_contrib().

Parameters

sequence	
max_w	

Returns

16.54.2.2 pf_interact()

#include <ViennaRNA/part_func_up.h>

Calculates the probability of a local interaction between two sequences.

The function considers the probability that the region of interaction is unpaired within 's1' and 's2'. The longer sequence has to be given as 's1'. The shorter sequence has to be given as 's2'. Function $pf_unstru()$ has to be called for 's1' and 's2', where the probabilities of being unpaired have to be given in 'p_c' and 'p_c2', respectively. If you do not want to include the probabilities of being unpaired for 's2' set 'p_c2' to NULL. If variable 'cstruc' is not NULL, constrained folding is done: The available constrains for intermolecular interaction are: '.' (no constrain), 'x' (the base has no intermolecular interaction) and '|' (the corresponding base has to be paired intermolecularily). The parameter 'w' determines the maximal length of the interaction. The parameters 'incr5' and 'incr3' allows inclusion of unpaired residues left ('incr5') and right ('incr3') of the region of interaction in 's1'. If the 'incr' options are used, function $pf_unstru()$ has to be called with w=w+incr5+incr3 for the longer sequence 's1'.

It returns a structure of type interact which contains the probability of the best local interaction including residue in Pi and the minimum free energy in Gi, where i is the position in sequence 's1'. The member Gikjl of structure interact is the best interaction between region [k,i] k<i in longer sequence 's1' and region [j,l] j<l in 's2'. Gikjl_wo is Gikjl without the probability of beeing unpaired.

Use free interact() to free the returned structure, all other stuff is freed inside pf interact().

Parameters

16.54 F	Partition	Function 1	for two	Hybridized	Sequences	as a Ste	nwise Proc	ess

447

Returns

16.55 Reading/Writing Energy Parameter Sets from/to File

Read and Write energy parameter sets from and to files or strings.

16.55.1 Detailed Description

Read and Write energy parameter sets from and to files or strings.

Collaboration diagram for Reading/Writing Energy Parameter Sets from/to File:

Modules

· Converting Energy Parameter Files

Convert energy parameter files into the latest format.

Macros

• #define VRNA PARAMETER FORMAT DEFAULT 0

Default Energy Parameter File format.

Functions

• int vrna params load (const char fname[], unsigned int options)

Load energy parameters from a file.

int vrna_params_save (const char fname[], unsigned int options)

Save energy parameters to a file.

int vrna_params_load_from_string (const char *string, const char *name, unsigned int options)

Load energy paramters from string.

int vrna_params_load_defaults (void)

Load default RNA energy parameter set.

int vrna params load RNA Turner2004 (void)

Load Turner 2004 RNA energy parameter set.

• int vrna_params_load_RNA_Turner1999 (void)

Load Turner 1999 RNA energy parameter set.

• int vrna params load RNA Andronescu2007 (void)

Load Andronsecu 2007 RNA energy parameter set.

int vrna_params_load_RNA_Langdon2018 (void)

Load Langdon 2018 RNA energy parameter set.

int vrna_params_load_RNA_misc_special_hairpins (void)

Load Misc Special Hairpin RNA energy parameter set.

• int vrna_params_load_DNA_Mathews2004 (void)

Load Mathews 2004 DNA energy parameter set.

int vrna_params_load_DNA_Mathews1999 (void)

Load Mathews 1999 DNA energy parameter set.

const char * last_parameter_file (void)

Get the file name of the parameter file that was most recently loaded.

void read_parameter_file (const char fname[])

Read energy parameters from a file.

void write_parameter_file (const char fname[])

Write energy parameters to a file.

16.55.2 Macro Definition Documentation

16.55.2.1 VRNA_PARAMETER_FORMAT_DEFAULT

```
#define VRNA_PARAMETER_FORMAT_DEFAULT 0
#include <ViennaRNA/params/io.h>
```

Default Energy Parameter File format.

See also

vrna_params_load(), vrna_params_load_from_string(), vrna_params_save()

16.55.3 Function Documentation

16.55.3.1 vrna_params_load()

Load energy parameters from a file.

See also

Parameters

fname	The path to the file containing the energy parameters			
options	File format bit-mask (usually VRNA_PARAMETER_FORMAT_DEFAULT)			

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as overloaded function **params_load**(fname="", options=VRNA_PARAMETER_FOR Here, the empty filename string indicates to load default RNA parameters, i.e. this is equivalent to calling vrna_params_load_defaults().

16.55.3.2 vrna_params_save()

Save energy parameters to a file.

See also

```
vrna_params_load()
```

Parameters

fname	A filename (path) for the file where the current energy parameters will be written to
options	File format bit-mask (usually VRNA_PARAMETER_FORMAT_DEFAULT)

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as overloaded function params_save(fname, options=VRNA_PARAMETER_FORM

16.55.3.3 vrna_params_load_from_string()

Load energy paramters from string.

The string must follow the default energy parameter file convention! The optional name argument allows one to specify a name for the parameter set which is stored internally.

See also

```
vrna_params_load(), vrna_params_save(), vrna_params_load_defaults(), vrna_params_load_RNA_Turner2004(), vrna_params_load_RNA_Turner1999(), vrna_params_load_RNA_Andronescu2007(), vrna_params_load_RNA_Langdon2018(vrna_params_load_RNA_misc_special_hairpins(), vrna_params_load_DNA_Mathews2004(), vrna_params_DNA_Mathews2004(), vrna_params_DNA_Mathews2004(), vrna_params_DNA_Mathews2004(), vrna_params_DNA_Mathews2004(), vrna_params_DNA_Mathews2004(), vrna_params_DNA_Mathews2004(), vrna_params_DNA_Mat
```

Parameters

string	A 0-terminated string containing energy parameters		
name	A name for the parameter set in string (Maybe NULL)		
options	File format bit-mask (usually VRNA_PARAMETER_FORMAT_DEFAULT)		

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as overloaded function **params_load_from_string**(string, name="", options=VRNA_PARAMETER_FORMAT_DEFAULT).

16.55.3.4 vrna_params_load_defaults()

#include <ViennaRNA/params/io.h>

Load default RNA energy parameter set.

This is a convenience function to load the Turner 2004 RNA free energy parameters. It's the same as calling vrna_params_load_RNA_Turner2004()

See also

vrna_params_load(), vrna_params_load_from_string(), vrna_params_save(), vrna_params_load_RNA_Turner2004(), vrna_params_load_RNA_Turner1999(), vrna_params_load_RNA_Andronescu2007(), vrna_params_load_RNA_Langdon2018(vrna_params_load_RNA_misc_special_hairpins(), vrna_params_load_DNA_Mathews2004(), vrna_params_load_DNA_M

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as overloaded function params_load().

16.55.3.5 vrna_params_load_RNA_Turner2004()

Load Turner 2004 RNA energy parameter set.

See also

vrna_params_load(), vrna_params_load_from_string(), vrna_params_save(), vrna_params_load_defaults(), vrna_params_load_RNA_Turner1999(), vrna_params_load_RNA_Andronescu2007(), vrna_params_load_RNA_Langdon2018(vrna_params_load_RNA_misc_special_hairpins(), vrna_params_load_DNA_Mathews2004(), vrna_params_load_DNA_Mathews

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as function params load RNA Turner2004().

```
16.55.3.6 vrna_params_load_RNA_Turner1999()
```

#include <ViennaRNA/params/io.h>

Load Turner 1999 RNA energy parameter set.

See also

```
vrna_params_load(), vrna_params_load_from_string(), vrna_params_save(), vrna_params_load_RNA_Turner2004(), vrna_params_load_defaults(), vrna_params_load_RNA_Andronescu2007(), vrna_params_load_RNA_Langdon2018(), vrna_params_load_RNA_misc_special_hairpins(), vrna_params_load_DNA_Mathews2004(), vrna_params_DNA_Mathews2004(), vrna_pa
```

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as function params_load_RNA_Turner1999().

16.55.3.7 vrna_params_load_RNA_Andronescu2007()

Load Andronsecu 2007 RNA energy parameter set.

See also

```
vrna_params_load(), vrna_params_load_from_string(), vrna_params_save(), vrna_params_load_RNA_Turner2004(), vrna_params_load_RNA_Turner1999(), vrna_params_load_defaults(), vrna_params_load_RNA_Langdon2018(), vrna_params_load_RNA_misc_special_hairpins(), vrna_params_load_DNA_Mathews2004(), vrna_params_DNA_Mathews2004(), vrna_params_DNA_Mathews2004
```

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as function params_load_RNA_Andronescu2007().

16.55.3.8 vrna_params_load_RNA_Langdon2018()

Load Langdon 2018 RNA energy parameter set.

#include <ViennaRNA/params/io.h>

See also

vrna_params_load(), vrna_params_load_from_string(), vrna_params_save(), vrna_params_load_RNA_Turner2004(), vrna_params_load_RNA_Turner1999(), vrna_params_load_RNA_Andronescu2007(), vrna_params_load_defaults(), vrna_params_load_RNA_misc_special_hairpins(), vrna_params_load_DNA_Mathews2004(), vrna_params_load_DNA_Mathe

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as function params_load_RNA_Langdon2018().

16.55.3.9 vrna_params_load_RNA_misc_special_hairpins()

Load Misc Special Hairpin RNA energy parameter set.

See also

vrna_params_load(), vrna_params_load_from_string(), vrna_params_save(), vrna_params_load_RNA_Turner2004(), vrna_params_load_RNA_Turner1999(), vrna_params_load_RNA_Andronescu2007(), vrna_params_load_RNA_Langdon2018(vrna_params_load_defaults(), vrna_params_load_DNA_Mathews2004(), vrna_params_load_DNA_Mathews1999()

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as function params load RNA misc special hairpins().

```
16.55.3.10 vrna_params_load_DNA_Mathews2004()
```

#include <ViennaRNA/params/io.h>

Load Mathews 2004 DNA energy parameter set.

See also

vrna_params_load(), vrna_params_load_from_string(), vrna_params_save(), vrna_params_load_RNA_Turner2004(), vrna_params_load_RNA_Turner1999(), vrna_params_load_RNA_Andronescu2007(), vrna_params_load_RNA_Langdon2018(vrna_params_load_RNA_misc_special_hairpins(), vrna_params_load_defaults(), vrna_params_load_DNA_Mathews1999()

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as function params_load_DNA_Mathews2004().

```
16.55.3.11 vrna_params_load_DNA_Mathews1999()
```

Load Mathews 1999 DNA energy parameter set.

See also

vrna_params_load(), vrna_params_load_from_string(), vrna_params_save(), vrna_params_load_RNA_Turner2004(), vrna_params_load_RNA_Turner1999(), vrna_params_load_RNA_Andronescu2007(), vrna_params_load_RNA_Langdon2018(vrna_params_load_RNA_misc_special_hairpins(), vrna_params_load_DNA_Mathews2004(), vrna_params_load_defaults()

Returns

Non-zero on success, 0 on failure

SWIG Wrapper Notes This function is available as function params_load_DNA_Mathews1999().

16.55.3.12 last_parameter_file()

Get the file name of the parameter file that was most recently loaded.

Returns

The file name of the last parameter file, or NULL if parameters are still at defaults

16.55.3.13 read_parameter_file()

Read energy parameters from a file.

Deprecated Use vrna_params_load() instead!

Parameters

fname The path to the file containing the energy parameters

16.55.3.14 write_parameter_file()

Write energy parameters to a file.

Deprecated Use vrna_params_save() instead!

Parameters

fname A filename (path) for the file where the current energy parameters will be written to

16.56 Converting Energy Parameter Files

Convert energy parameter files into the latest format.

16.56.1 Detailed Description

Convert energy parameter files into the latest format.

To preserve some backward compatibility the RNAlib also provides functions to convert energy parameter files from the format used in version 1.4-1.8 into the new format used since version 2.0 Collaboration diagram for Converting Energy Parameter Files:

Files

• file 1.8.4_epars.h

Free energy parameters for parameter file conversion.

• file 1.8.4_intloops.h

Free energy parameters for interior loop contributions needed by the parameter file conversion functions.

Macros

- #define VRNA CONVERT OUTPUT ALL 1U
- #define VRNA CONVERT OUTPUT HP 2U
- #define VRNA CONVERT OUTPUT STACK 4U
- #define VRNA_CONVERT_OUTPUT_MM_HP 8U
- #define VRNA_CONVERT_OUTPUT_MM_INT 16U
- #define VRNA CONVERT OUTPUT MM INT 1N 32U
- #define VRNA CONVERT OUTPUT MM INT 23 64U
- #define VRNA_CONVERT_OUTPUT_MM_MULTI 128U
- #define VRNA_CONVERT_OUTPUT_MM_EXT 256U
- #define VRNA_CONVERT_OUTPUT_DANGLE5 512U
- #define VRNA_CONVERT_OUTPUT_DANGLE3 1024U
- #define VRNA CONVERT OUTPUT INT 11 2048U
- #define VRNA_CONVERT_OUTPUT_INT_21 4096U
- #define VRNA_CONVERT_OUTPUT_INT_22 8192U
- #define VRNA_CONVERT_OUTPUT_BULGE 16384U
- #define VRNA CONVERT OUTPUT INT 32768U
- #define VRNA_CONVERT_OUTPUT_ML 65536U
- #define VRNA_CONVERT_OUTPUT_MISC 131072U
- #define VRNA CONVERT OUTPUT SPECIAL HP 262144U
- #define VRNA_CONVERT_OUTPUT_VANILLA 524288U
- #define VRNA CONVERT OUTPUT NINIO 1048576U
- #define VRNA CONVERT OUTPUT DUMP 2097152U

Functions

• void convert_parameter_file (const char *iname, const char *oname, unsigned int options)

16.56.2 Macro Definition Documentation

```
16.56.2.1 VRNA_CONVERT_OUTPUT_ALL
#define VRNA_CONVERT_OUTPUT_ALL 1U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of a complete parameter set
16.56.2.2 VRNA_CONVERT_OUTPUT_HP
#define VRNA_CONVERT_OUTPUT_HP 2U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of hairpin contributions
16.56.2.3 VRNA_CONVERT_OUTPUT_STACK
#define VRNA_CONVERT_OUTPUT_STACK 4U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of base pair stack contributions
16.56.2.4 VRNA_CONVERT_OUTPUT_MM_HP
#define VRNA_CONVERT_OUTPUT_MM_HP 8U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of hairpin mismatch contribution
16.56.2.5 VRNA_CONVERT_OUTPUT_MM_INT
#define VRNA_CONVERT_OUTPUT_MM_INT 16U
#include <ViennaRNA/params/convert.h>
```

Flag to indicate printing of interior loop mismatch contribution

Generated by Doxygen

```
16.56.2.6 VRNA_CONVERT_OUTPUT_MM_INT_1N
#define VRNA_CONVERT_OUTPUT_MM_INT_1N 32U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of 1:n interior loop mismatch contribution
16.56.2.7 VRNA_CONVERT_OUTPUT_MM_INT_23
#define VRNA_CONVERT_OUTPUT_MM_INT_23 64U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of 2:3 interior loop mismatch contribution
16.56.2.8 VRNA_CONVERT_OUTPUT_MM_MULTI
#define VRNA_CONVERT_OUTPUT_MM_MULTI 128U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of multi loop mismatch contribution
16.56.2.9 VRNA_CONVERT_OUTPUT_MM_EXT
#define VRNA_CONVERT_OUTPUT_MM_EXT 256U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of exterior loop mismatch contribution
16.56.2.10 VRNA_CONVERT_OUTPUT_DANGLE5
#define VRNA_CONVERT_OUTPUT_DANGLE5 512U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of 5' dangle conctribution
16.56.2.11 VRNA_CONVERT_OUTPUT_DANGLE3
#define VRNA_CONVERT_OUTPUT_DANGLE3 1024U
#include <ViennaRNA/params/convert.h>
```

Flag to indicate printing of 3' dangle contribution

```
16.56.2.12 VRNA_CONVERT_OUTPUT_INT_11
#define VRNA_CONVERT_OUTPUT_INT_11 2048U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of 1:1 interior loop contribution
16.56.2.13 VRNA_CONVERT_OUTPUT_INT_21
#define VRNA_CONVERT_OUTPUT_INT_21 4096U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of 2:1 interior loop contribution
16.56.2.14 VRNA_CONVERT_OUTPUT_INT_22
#define VRNA_CONVERT_OUTPUT_INT_22 8192U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of 2:2 interior loop contribution
16.56.2.15 VRNA_CONVERT_OUTPUT_BULGE
#define VRNA_CONVERT_OUTPUT_BULGE 16384U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of bulge loop contribution
16.56.2.16 VRNA_CONVERT_OUTPUT_INT
#define VRNA_CONVERT_OUTPUT_INT 32768U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of interior loop contribution
16.56.2.17 VRNA_CONVERT_OUTPUT_ML
#define VRNA_CONVERT_OUTPUT_ML 65536U
#include <ViennaRNA/params/convert.h>
```

Generated by Doxygen

Flag to indicate printing of multi loop contribution

```
16.56.2.18 VRNA_CONVERT_OUTPUT_MISC
#define VRNA_CONVERT_OUTPUT_MISC 131072U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of misc contributions (such as terminalAU)
16.56.2.19 VRNA_CONVERT_OUTPUT_SPECIAL_HP
#define VRNA_CONVERT_OUTPUT_SPECIAL_HP 262144U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of special hairpin contributions (tri-, tetra-, hexa-loops)
16.56.2.20 VRNA_CONVERT_OUTPUT_VANILLA
#define VRNA_CONVERT_OUTPUT_VANILLA 524288U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of given parameters only
Note
     This option overrides all other output options, except VRNA_CONVERT_OUTPUT_DUMP!
16.56.2.21 VRNA_CONVERT_OUTPUT_NINIO
#define VRNA_CONVERT_OUTPUT_NINIO 1048576U
#include <ViennaRNA/params/convert.h>
Flag to indicate printing of interior loop asymmetry contribution
16.56.2.22 VRNA_CONVERT_OUTPUT_DUMP
#define VRNA_CONVERT_OUTPUT_DUMP 2097152U
#include <ViennaRNA/params/convert.h>
```

Flag to indicate dumping the energy contributions from the library instead of an input file

16.56.3 Function Documentation

16.56.3.1 convert parameter file()

Convert/dump a Vienna 1.8.4 formatted energy parameter file

The options argument allows one to control the different output modes.

Currently available options are:

VRNA_CONVERT_OUTPUT_ALL, VRNA_CONVERT_OUTPUT_HP, VRNA_CONVERT_OUTPUT_STACK
VRNA_CONVERT_OUTPUT_MM_HP, VRNA_CONVERT_OUTPUT_MM_INT, VRNA_CONVERT_OUTPUT_MM_INT_1N
VRNA_CONVERT_OUTPUT_MM_INT_23, VRNA_CONVERT_OUTPUT_MM_MULTI, VRNA_CONVERT_OUTPUT_MM_EXT
VRNA_CONVERT_OUTPUT_DANGLE5, VRNA_CONVERT_OUTPUT_DANGLE3, VRNA_CONVERT_OUTPUT_INT_11
VRNA_CONVERT_OUTPUT_INT_21, VRNA_CONVERT_OUTPUT_INT_22, VRNA_CONVERT_OUTPUT_BULGE
VRNA_CONVERT_OUTPUT_INT, VRNA_CONVERT_OUTPUT_ML, VRNA_CONVERT_OUTPUT_MISC
VRNA_CONVERT_OUTPUT_SPECIAL_HP, VRNA_CONVERT_OUTPUT_VANILLA, VRNA_CONVERT_OUTPUT_NINIO
VRNA_CONVERT_OUTPUT_DUMP

The defined options are fine for bitwise compare- and assignment-operations, e. g.: pass a collection of options as a single value like this:

```
\verb|convert_parameter_file| (ifile, ofile, option_1 \mid option_2 \mid option_n) \\
```

Parameters

iname	The input file name (If NULL input is read from stdin)
oname	The output file name (If NULL output is written to stdout)
options	The options (as described above)

16.57 Utilities to deal with Nucleotide Alphabets

Functions to cope with various aspects related to the nucleotide sequence alphabet.

16.57.1 Detailed Description

Functions to cope with various aspects related to the nucleotide sequence alphabet.

Collaboration diagram for Utilities to deal with Nucleotide Alphabets:

Files

· file alphabet.h

Functions to process, convert, and generally handle different nucleotide and/or base pair alphabets.

· file sequence.h

Functions and data structures related to sequence representations,.

Data Structures

• struct vrna_sequence_s

Data structure representing a nucleotide sequence. More...

• struct vrna_alignment_s

Typedefs

typedef struct vrna_sequence_s vrna_seq_t

Typename for nucleotide sequence representation data structure vrna_sequence_s.

Enumerations

• enum vrna_seq_type_e { VRNA_SEQ_UNKNOWN, VRNA_SEQ_RNA, VRNA_SEQ_DNA }

A enumerator used in vrna_sequence_s to distinguish different nucleotide sequences.

Functions

char * vrna_ptypes (const short *S, vrna_md_t *md)

Get an array of the numerical encoding for each possible base pair (i,j)

short * vrna_seq_encode (const char *sequence, vrna_md_t *md)

Get a numerical representation of the nucleotide sequence.

short * vrna_seq_encode_simple (const char *sequence, vrna_md_t *md)

Get a numerical representation of the nucleotide sequence (simple version)

int vrna_nucleotide_encode (char c, vrna_md_t *md)

Encode a nucleotide character to numerical value.

char vrna_nucleotide_decode (int enc, vrna_md_t *md)

Decode a numerical representation of a nucleotide back into nucleotide alphabet.

16.57.2 Data Structure Documentation

16.57.2.1 struct vrna_sequence_s

Data structure representing a nucleotide sequence.

Data Fields

vrna_seq_type_e type

The type of sequence.

• char * string

The string representation of the sequence.

• short * encoding

The integer representation of the sequence.

unsigned int length

The length of the sequence.

16.57.2.2 struct vrna_alignment_s

Collaboration diagram for vrna_alignment_s:

16.57.3 Enumeration Type Documentation

```
16.57.3.1 vrna_seq_type_e
```

enum vrna_seq_type_e

#include <ViennaRNA/sequence.h>

A enumerator used in vrna_sequence_s to distinguish different nucleotide sequences.

Enumerator

VRNA_SEQ_UNKNOWN	Nucleotide sequence represents an Unkown type.
VRNA_SEQ_RNA	Nucleotide sequence represents an RNA type.
VRNA_SEQ_DNA	Nucleotide sequence represents a DNA type.

16.57.4 Function Documentation

```
16.57.4.1 vrna_ptypes()
```

Get an array of the numerical encoding for each possible base pair (i,j)

Note

This array is always indexed in column-wise order, in contrast to previously different indexing between mfe and pf variants!

See also

```
vrna_idx_col_wise(), vrna_fold_compound_t
```

```
16.57.4.2 vrna_seq_encode()
```

Get a numerical representation of the nucleotide sequence.

```
16.57.4.3 vrna_seq_encode_simple()
```

Get a numerical representation of the nucleotide sequence (simple version)

16.57.4.4 vrna_nucleotide_encode()

Encode a nucleotide character to numerical value.

This function encodes a nucleotide character to its numerical representation as required by many functions in $R \leftarrow NA$ lib.

See also

```
vrna_nucleotide_decode(), vrna_seq_encode()
```

Parameters

С	The nucleotide character to encode	
md	The model details that determine the kind of encoding	

Returns

The encoded nucleotide

16.57.4.5 vrna_nucleotide_decode()

#include <ViennaRNA/alphabet.h>

Decode a numerical representation of a nucleotide back into nucleotide alphabet.

This function decodes a numerical representation of a nucleotide character back into nucleotide alphabet

See also

vrna_nucleotide_encode(), vrna_seq_encode()

Parameters

enc	The encoded nucleotide
md	The model details that determine the kind of decoding

Returns

The decoded nucleotide character

16.58 (Nucleic Acid Sequence) String Utilitites

Functions to parse, convert, manipulate, create, and compare (nucleic acid sequence) strings.

16.58.1 Detailed Description

Functions to parse, convert, manipulate, create, and compare (nucleic acid sequence) strings.

Collaboration diagram for (Nucleic Acid Sequence) String Utilities:

Files

· file strings.h

General utility- and helper-functions for RNA sequence and structure strings used throughout the ViennaRNA Package.

Macros

#define XSTR(s) STR(s)

Stringify a macro after expansion.

#define STR(s) #s

Stringify a macro argument.

• #define FILENAME MAX LENGTH 80

Maximum length of filenames that are generated by our programs.

• #define FILENAME ID LENGTH 42

Maximum length of id taken from fasta header for filename generation.

Functions

• char * vrna_strdup_printf (const char *format,...)

Safely create a formatted string.

char * vrna_strdup_vprintf (const char *format, va_list argp)

Safely create a formatted string.

• int vrna_strcat_printf (char **dest, const char *format,...)

Safely append a formatted string to another string.

int vrna_strcat_vprintf (char **dest, const char *format, va_list args)

Safely append a formatted string to another string.

char ** vrna_strsplit (const char *string, const char *delimiter)

Split a string into tokens using a delimiting character.

char * vrna random string (int I, const char symbols[])

Create a random string using characters from a specified symbol set.

int vrna_hamming_distance (const char *s1, const char *s2)

Calculate hamming distance between two sequences.

• int vrna_hamming_distance_bound (const char *s1, const char *s2, int n)

Calculate hamming distance between two sequences up to a specified length.

void vrna_seq_toRNA (char *sequence)

Convert an input sequence (possibly containing DNA alphabet characters) to RNA alphabet.

void vrna_seq_toupper (char *sequence)

Convert an input sequence to uppercase.

• char * vrna_seq_ungapped (const char *seq)

Remove gap characters from a nucleotide sequence.

• char * vrna_cut_point_insert (const char *string, int cp)

Add a separating '&' character into a string according to cut-point position.

char * vrna_cut_point_remove (const char *string, int *cp)

Remove a separating '&' character from a string.

16.58.2 Macro Definition Documentation

16.58.2.1 FILENAME_MAX_LENGTH

```
#define FILENAME_MAX_LENGTH 80
#include <ViennaRNA/utils/strings.h>
```

Maximum length of filenames that are generated by our programs.

This definition should be used throughout the complete ViennaRNA package wherever a static array holding filenames of output files is declared.

16.58.2.2 FILENAME ID LENGTH

```
#define FILENAME_ID_LENGTH 42
#include <ViennaRNA/utils/strings.h>
```

Maximum length of id taken from fasta header for filename generation.

this has to be smaller than FILENAME_MAX_LENGTH since in most cases, some suffix will be appended to the ID

16.58.3 Function Documentation

16.58.3.1 vrna_strdup_printf()

Safely create a formatted string.

This function is a safe implementation for creating a formatted character array, similar to *sprintf*. Internally, it uses the *asprintf* function if available to dynamically allocate a large enough character array to store the supplied content. If *asprintf* is not available, mimic it's behavior using *vsnprintf*.

Note

The returned pointer of this function should always be passed to free() to release the allocated memory

See also

```
vrna_strdup_vprintf(), vrna_strcat_printf()
```

Parameters

format	The format string (See also asprintf)
	The list of variables used to fill the format string

Returns

The formatted, null-terminated string, or NULL if something has gone wrong

16.58.3.2 vrna_strdup_vprintf()

Safely create a formatted string.

This function is the *va_list* version of vrna_strdup_printf()

#include <ViennaRNA/utils/strings.h>

Note

The returned pointer of this function should always be passed to free() to release the allocated memory

See also

```
vrna_strdup_printf(), vrna_strcat_printf(), vrna_strcat_vprintf()
```

Parameters

format	The format string (See also asprintf)
argp	The list of arguments to fill the format string

Returns

The formatted, null-terminated string, or NULL if something has gone wrong

16.58.3.3 vrna_strcat_printf()

```
#include <ViennaRNA/utils/strings.h>
```

Safely append a formatted string to another string.

This function is a safe implementation for appending a formatted character array, similar to a cobination of *strcat* and *sprintf*. The function automatically allocates enough memory to store both, the previous content stored at dest and the appended format string. If the dest pointer is NULL, the function allocate memory only for the format string. The function returns the number of characters in the resulting string or -1 in case of an error.

See also

```
vrna_strcat_vprintf(), vrna_strdup_printf(), vrna_strdup_vprintf()
```

Parameters

dest	The address of a char *pointer where the formatted string is to be appended
format	The format string (See also sprintf)
	The list of variables used to fill the format string

Returns

The number of characters in the final string, or -1 on error

16.58.3.4 vrna_strcat_vprintf()

#include <ViennaRNA/utils/strings.h>

Safely append a formatted string to another string.

This function is the *va_list* version of vrna_strcat_printf()

See also

```
vrna_strcat_printf(), vrna_strdup_printf(), vrna_strdup_vprintf()
```

Parameters

dest	The address of a char *pointer where the formatted string is to be appended
format	The format string (See also sprintf)
args	The list of argument to fill the format string

Returns

The number of characters in the final string, or -1 on error

16.58.3.5 vrna_strsplit()

#include <ViennaRNA/utils/strings.h>

Split a string into tokens using a delimiting character.

This function splits a string into an array of strings using a single character that delimits the elements within the string. The default delimiter is the ampersand '&' and will be used when NULL is passed as a second argument. The returned list is NULL terminated, i.e. the last element is NULL. If the delimiter is not found, the returned list contains exactly one element: the input string.

For instance, the following code:

```
char **tok = vrna_strsplit("GGGG&CCCC&AAAAA", NULL);
for (char **ptr = tok; *ptr; ptr++) {
  printf("%s\n", *ptr);
  free(*ptr);
}
free(tok);
```

produces this output:

```
* GGGG
* CCCC
* AAAAA
```

and properly free's the memory occupied by the returned element array.

Note

This function internally uses $strtok_r()$ and is therefore considered to be thread-safe. Also note, that it is the users responsibility to free the memory of the array and that of the individual element strings!

Parameters

string	The input string that should be split into elements	
delimiter	The delimiting character. If $\mathtt{NULL},$ the delimiter is " $\&$ "	

Returns

A NULL terminated list of the elements in the string

16.58.3.6 vrna_random_string()

Create a random string using characters from a specified symbol set.

Parameters

1	The length of the sequence
symbols	The symbol set

Returns

A random string of length 'l' containing characters from the symbolset

16.58.3.7 vrna_hamming_distance()

Calculate hamming distance between two sequences.

Parameters

5	s 1	The first sequence
5	32	The second sequence

Returns

The hamming distance between s1 and s2

16.58.3.8 vrna_hamming_distance_bound()

```
int vrna_hamming_distance_bound ( const char * s1, const char * s2, int n)
```

```
#include <ViennaRNA/utils/strings.h>
```

Calculate hamming distance between two sequences up to a specified length.

This function is similar to vrna_hamming_distance() but instead of comparing both sequences up to their actual length only the first 'n' characters are taken into account

Parameters

s1 The first sequence		The first sequence	
	s2	s2 The second sequence	
	n	The length of the subsequences to consider (starting from the 5' end)	

Returns

The hamming distance between s1 and s2

16.58.3.9 vrna_seq_toRNA()

Convert an input sequence (possibly containing DNA alphabet characters) to RNA alphabet.

This function substitudes T and t with U and u, respectively

Parameters

sequence The sequence to be converted	sequence
---------------------------------------	----------

16.58.3.10 vrna_seq_toupper()

Convert an input sequence to uppercase.

Parameters

sequence	The sequence to be converted

16.58.3.11 vrna_seq_ungapped()

Remove gap characters from a nucleotide sequence.

Parameters

sequence	The original, null-terminated nucleotide sequence
----------	---

Returns

A copy of the input sequence with all gap characters removed

16.58.3.12 vrna_cut_point_insert()

Add a separating '&' character into a string according to cut-point position.

If the cut-point position is less or equal to zero, this function just returns a copy of the provided string. Otherwise, the cut-point character is set at the corresponding position

Parameters

string	The original string
ср	The cut-point position

Returns

A copy of the provided string including the cut-point character

16.58.3.13 vrna_cut_point_remove()

#include <ViennaRNA/utils/strings.h>

Remove a separating '&' character from a string.

This function removes the cut-point indicating '&' character from a string and memorizes its position in a provided integer variable. If not '&' is found in the input, the integer variable is set to -1. The function returns a copy of the input string with the '&' being sliced out.

Parameters

string	The original string
ср	The cut-point position

Returns

A copy of the input string with the '&' being sliced out

16.59 Secondary Structure Utilities

Functions to create, parse, convert, manipulate, and compare secondary structure representations.

16.59.1 Detailed Description

Functions to create, parse, convert, manipulate, and compare secondary structure representations.

Collaboration diagram for Secondary Structure Utilities:

Modules

- · Dot-Bracket Notation of Secondary Structures
- Pair Table Representation of Secondary Structures
- Pair List Representation of Secondary Structures
- · Helix List Representation of Secondary Structures
- Tree Representation of Secondary Structures
- Deprecated Interface for Secondary Structure Utilities

Files

· file structures.h

Various utility- and helper-functions for secondary structure parsing, converting, etc.

Functions

int * vrna_loopidx_from_ptable (const short *pt)

Get a loop index representation of a structure.

int vrna_bp_distance (const char *str1, const char *str2)

Compute the "base pair" distance between two secondary structures s1 and s2.

• unsigned int * vrna_refBPcnt_matrix (const short *reference_pt, unsigned int turn)

Make a reference base pair count matrix.

unsigned int * vrna_refBPdist_matrix (const short *pt1, const short *pt2, unsigned int turn)

Make a reference base pair distance matrix.

char * vrna db from probs (const FLT OR DBL *pr, unsigned int length)

Create a dot-bracket like structure string from base pair probability matrix.

char vrna_bpp_symbol (const float *x)

Get a pseudo dot bracket notation for a given probability information.

char * vrna_db_from_bp_stack (vrna_bp_stack_t *bp, unsigned int length)

Create a dot-backet/parenthesis structure from backtracking stack.

16.59.2 Function Documentation

16.59.2.1 vrna_bp_distance()

Compute the "base pair" distance between two secondary structures s1 and s2.

The sequences should have the same length. dist = number of base pairs in one structure but not in the other same as edit distance with open-pair close-pair as move-set

Parameters

str1	First structure in dot-bracket notation
str2	Second structure in dot-bracket notation

Returns

The base pair distance between str1 and str2

16.59.2.2 vrna_refBPcnt_matrix()

#include <ViennaRNA/utils/structures.h>

Make a reference base pair count matrix.

Get an upper triangular matrix containing the number of basepairs of a reference structure for each interval [i,j] with i < j. Access it via iindx!!!

16.59.2.3 vrna_refBPdist_matrix()

#include <ViennaRNA/utils/structures.h>

Make a reference base pair distance matrix.

Get an upper triangular matrix containing the base pair distance of two reference structures for each interval [i,j] with i < j. Access it via iindx!!!

16.59.2.4 vrna_db_from_bp_stack()

#include <ViennaRNA/utils/structures.h>

Create a dot-backet/parenthesis structure from backtracking stack.

This function is capable to create dot-bracket structures from suboptimal structure prediction sensu M. Zuker

Parameters

bp	Base pair stack containing the traced base pairs
length	The length of the structure

Returns

The secondary structure in dot-bracket notation as provided in the input

16.60 Dot-Bracket Notation of Secondary Structures

16.60.1 Detailed Description

Collaboration diagram for Dot-Bracket Notation of Secondary Structures:

Macros

• #define VRNA_BRACKETS_ALPHA 4U

Bitflag to indicate secondary structure notations using uppercase/lowercase letters from the latin alphabet.

• #define VRNA BRACKETS RND 8U

Bitflag to indicate secondary structure notations using round brackets (parenthesis), ()

• #define VRNA_BRACKETS_CLY 16U

Bitflag to indicate secondary structure notations using curly brackets, {}

#define VRNA BRACKETS ANG 32U

Bitflag to indicate secondary structure notations using angular brackets, <>

• #define VRNA BRACKETS SQR 64U

Bitflag to indicate secondary structure notations using square brackets, []

#define VRNA BRACKETS DEFAULT

Default bitmask to indicate secondary structure notation using any pair of brackets.

#define VRNA BRACKETS ANY

Bitmask to indicate secondary structure notation using any pair of brackets or uppercase/lowercase alphabet letters.

Functions

char * vrna_db_pack (const char *struc)

Pack secondary secondary structure, 5:1 compression using base 3 encoding.

char * vrna_db_unpack (const char *packed)

Unpack secondary structure previously packed with vrna_db_pack()

void vrna db flatten (char *structure, unsigned int options)

Substitute pairs of brackets in a string with parenthesis.

• void vrna_db_flatten_to (char *string, const char target[3], unsigned int options)

Substitute pairs of brackets in a string with another type of pair characters.

char * vrna db from ptable (short *pt)

Convert a pair table into dot-parenthesis notation.

char * vrna_db_from_WUSS (const char *wuss)

Convert a WUSS annotation string to dot-bracket format.

char * vrna_db_from_plist (vrna_ep_t *pairs, unsigned int n)

Convert a list of base pairs into dot-bracket notation.

• char * vrna_db_to_element_string (const char *structure)

Convert a secondary structure in dot-bracket notation to a nucleotide annotation of loop contexts.

char * vrna_db_pk_remove (const char *structure, unsigned int options)

Remove pseudo-knots from an input structure.

16.60.2 Macro Definition Documentation

```
16.60.2.1 VRNA_BRACKETS_ALPHA
#define VRNA_BRACKETS_ALPHA 4U
#include <ViennaRNA/utils/structures.h>
Bitflag to indicate secondary structure notations using uppercase/lowercase letters from the latin alphabet.
See also
     vrna_ptable_from_string()
16.60.2.2 VRNA_BRACKETS_RND
#define VRNA_BRACKETS_RND 8U
#include <ViennaRNA/utils/structures.h>
Bitflag to indicate secondary structure notations using round brackets (parenthesis), ()
See also
     vrna_ptable_from_string(), vrna_db_flatten(), vrna_db_flatten_to()
16.60.2.3 VRNA_BRACKETS_CLY
#define VRNA_BRACKETS_CLY 16U
#include <ViennaRNA/utils/structures.h>
Bitflag to indicate secondary structure notations using curly brackets, { }
See also
     vrna_ptable_from_string(), vrna_db_flatten(), vrna_db_flatten_to()
16.60.2.4 VRNA_BRACKETS_ANG
#define VRNA_BRACKETS_ANG 32U
#include <ViennaRNA/utils/structures.h>
Bitflag to indicate secondary structure notations using angular brackets, <>
See also
     vrna_ptable_from_string(), vrna_db_flatten(), vrna_db_flatten_to()
```

```
16.60.2.5 VRNA_BRACKETS_SQR
```

```
#define VRNA_BRACKETS_SQR 64U
#include <ViennaRNA/utils/structures.h>
```

Bitflag to indicate secondary structure notations using square brackets, []

See also

vrna_ptable_from_string(), vrna_db_flatten(), vrna_db_flatten_to()

16.60.2.6 VRNA_BRACKETS_DEFAULT

```
#define VRNA_BRACKETS_DEFAULT
```

#include <ViennaRNA/utils/structures.h>

Value:

```
(VRNA_BRACKETS_RND | \
VRNA_BRACKETS_CLY | \
VRNA_BRACKETS_ANG | \
VRNA_BRACKETS_SQR)
```

Default bitmask to indicate secondary structure notation using any pair of brackets.

This set of matching brackets/parenthesis is always nested, i.e. pseudo-knot free, in WUSS format. However, in general different kinds of brackets are mostly used for annotating pseudo-knots. Thus special care has to be taken to remove pseudo-knots if this bitmask is used in functions that return secondary structures without pseudo-knots!

See also

vrna_ptable_from_string(), vrna_db_flatten(), vrna_db_flatten_to(), vrna_db_pk_remove() vrna_pt_pk_remove()

16.60.2.7 VRNA_BRACKETS_ANY

```
#define VRNA_BRACKETS_ANY
#include <ViennaRNA/utils/structures.h>
```

Value:

```
(VRNA_BRACKETS_RND | \
VRNA_BRACKETS_CLY | \
VRNA_BRACKETS_ANG | \
VRNA_BRACKETS_SQR | \
VRNA_BRACKETS_ALPHA)
```

Bitmask to indicate secondary structure notation using any pair of brackets or uppercase/lowercase alphabet letters.

See also

```
vrna_ptable_from_string(), vrna_db_pk_remove(), vrna_db_flatten(), vrna_db_flatten_to()
```

16.60.3 Function Documentation

Pack secondary secondary structure, 5:1 compression using base 3 encoding.

Returns a binary string encoding of the secondary structure using a 5:1 compression scheme. The string is NULL terminated and can therefore be used with standard string functions such as strcmp(). Useful for programs that need to keep many structures in memory.

See also

```
vrna_db_unpack()
```

Parameters

struc The secondary structure in dot-bracket notation

Returns

The binary encoded structure

16.60.3.2 vrna_db_unpack()

Unpack secondary structure previously packed with vrna_db_pack()

Translate a compressed binary string produced by vrna_db_pack() back into the familiar dot-bracket notation.

See also

```
vrna_db_pack()
```

Parameters

packed	The binary encoded packed secondary structure

Returns

The unpacked secondary structure in dot-bracket notation

16.60.3.3 vrna_db_flatten()

#include <ViennaRNA/utils/structures.h>

Substitute pairs of brackets in a string with parenthesis.

This function can be used to replace brackets of unusual types, such as angular brackets <> , to dot-bracket format. The options parameter is used too specify which types of brackets will be replaced by round parenthesis ().

See also

```
vrna_db_flatten_to(), VRNA_BRACKETS_RND, VRNA_BRACKETS_ANG, VRNA_BRACKETS_CLY, VRNA_BRACKETS_SQR, VRNA_BRACKETS_DEFAULT
```

Parameters

structure	The structure string where brackets are flattened in-place	
options	A bitmask to specify which types of brackets should be flattened out	

SWIG Wrapper Notes This function flattens an input structure string in-place! The second parameter is optional and defaults to VRNA_BRACKETS_DEFAULT.

An overloaded version of this function exists, where an additional second parameter can be passed to specify the target brackets, i.e. the type of matching pair characters all brackets will be flattened to. Therefore, in the scripting language interface this function is a replacement for vrna_db_flatten_to().

16.60.3.4 vrna_db_flatten_to()

Substitute pairs of brackets in a string with another type of pair characters.

This function can be used to replace brackets in a structure annotation string, such as square brackets [], to another type of pair characters, e.g. angular brackets <>.

The target array must contain a character for the 'pair open' annotation at position 0, and one for 'pair close' at position 1. Toptions parameter is used to specify which types of brackets will be replaced by the new pairs.

See also

vrna_db_flatten(), VRNA_BRACKETS_RND, VRNA_BRACKETS_ANG, VRNA_BRACKETS_CLY, VRNA_BRACKETS_SQR, VRNA_BRACKETS_DEFAULT

Parameters

string	The structure string where brackets are flattened in-place
target	The new pair characters the string will be flattened to
options	A bitmask to specify which types of brackets should be flattened out

SWIG Wrapper Notes This function is available as an overloaded version of vrna_db_flatten()

16.60.3.5 vrna_db_from_ptable()

Convert a pair table into dot-parenthesis notation.

Parameters

```
pt The pair table to be copied
```

Returns

A char pointer to the dot-bracket string

16.60.3.6 vrna_db_from_WUSS()

Convert a WUSS annotation string to dot-bracket format.

Note

This function flattens all brackets, and treats pseudo-knots annotated by matching pairs of upper/lowercase letters as unpaired nucleotides

See also

Washington University Secondary Structure (WUSS) notation

Parameters

wuss	The input string in WUSS notation
------	-----------------------------------

Returns

A dot-bracket notation of the input secondary structure

16.60.3.7 vrna_db_from_plist()

Convert a list of base pairs into dot-bracket notation.

See also

vrna_plist()

Parameters

pairs	A vrna_ep_t containing the pairs to be included in the dot-bracket string
n	The length of the structure (number of nucleotides)

Returns

The dot-bracket string containing the provided base pairs

16.60.3.8 vrna_db_to_element_string()

Convert a secondary structure in dot-bracket notation to a nucleotide annotation of loop contexts.

Parameters

structure	The secondary structure in dot-bracket notation

Returns

A string annotating each nucleotide according to it's structural context

16.60.3.9 vrna_db_pk_remove()

#include <ViennaRNA/utils/structures.h>

Remove pseudo-knots from an input structure.

This function removes pseudo-knots from an input structure by determining the minimum number of base pairs that need to be removed to make the structure pseudo-knot free.

To accomplish that, we use a dynamic programming algorithm similar to the Nussinov maxmimum matching approach.

The input structure must be in a dot-bracket string like form where crossing base pairs are denoted by the use of additional types of matching brackets, e.g. <>, $\{\}$, [], $\{\}$. Furthermore, crossing pairs may be annotated by matching uppercase/lowercase letters from the alphabet A-Z. For the latter, the uppercase letter must be the 5' and the lowercase letter the 3' nucleotide of the base pair. The actual type of brackets to be recognized by this function must be specifed through the options parameter.

Note

Brackets in the input structure string that are not covered by the options bitmask will be silently ignored!

See also

vrna_pt_pk_remove(), vrna_db_flatten(), VRNA_BRACKETS_RND, VRNA_BRACKETS_ANG, VRNA_BRACKETS_CLY, VRNA_BRACKETS_ANA_BRACKETS_ALPHA, VRNA_BRACKETS_DEFAULT, VRNA_BRACKETS_ANY

Parameters

structure	Input structure in dot-bracket format that may include pseudo-knots
options	A bitmask to specify which types of brackets should be processed

Returns

The input structure devoid of pseudo-knots in dot-bracket notation

SWIG Wrapper Notes This function is available as an overloaded function db_pk_remove () where the optional second parameter options defaults to #VRNA_BRACKET_ANY.

16.61 Pair Table Representation of Secondary Structures

16.61.1 Detailed Description

Collaboration diagram for Pair Table Representation of Secondary Structures:

Functions

• short * vrna ptable (const char *structure)

Create a pair table from a dot-bracket notation of a secondary structure.

short * vrna_ptable_from_string (const char *string, unsigned int options)

Create a pair table for a secondary structure string.

short * vrna_pt_pk_get (const char *structure)

Create a pair table of a secondary structure (pseudo-knot version)

short * vrna_ptable_copy (const short *pt)

Get an exact copy of a pair table.

short * vrna_pt_ali_get (const char *structure)

Create a pair table of a secondary structure (snoop align version)

short * vrna_pt_snoop_get (const char *structure)

Create a pair table of a secondary structure (snoop version)

short * vrna_pt_pk_remove (const short *ptable, unsigned int options)

Remove pseudo-knots from a pair table.

16.61.2 Function Documentation

16.61.2.1 vrna_ptable()

Create a pair table from a dot-bracket notation of a secondary structure.

Returns a newly allocated table, such that table[i]=j if (i.j) pair or 0 if i is unpaired, table[0] contains the length of the structure.

See also

```
vrna_ptable_from_string(), vrna_db_from_ptable()
```

Parameters

structure The secondary structure in dot-bracket notation	1
---	---

Returns

A pointer to the created pair_table

16.61.2.2 vrna_ptable_from_string()

#include <ViennaRNA/utils/structures.h>

Create a pair table for a secondary structure string.

This function takes an input string of a secondary structure annotation in Dot-Bracket Notation (a.k.a. Dot-Parenthesis Notation) or Extended Dot-Bracket Notation, and converts it into a pair table representation.

Note

This function also extracts crossing base pairs, i.e. pseudo-knots if more than a single matching bracket type is allowed through the bitmask options.

See also

vrna_ptable(), vrna_db_from_ptable(), vrna_db_flatten_to(), vrna_pt_pk_remove() VRNA_BRACKETS_RND, VRNA_BRACKETS_ANG, VRNA_BRACKETS_CLY, VRNA_BRACKETS_SQR, VRNA_BRACKETS_ALPHA, VRNA_BRACKETS_DEFAULT, VRNA_BRACKETS_ANY

Parameters

string	Secondary structure in Extended Dot-Bracket Notation
options	A bitmask to specify which brackets are recognized during conversion to pair table

Returns

A pointer to a new pair table of the provided secondary structure

16.61.2.3 vrna_pt_pk_get()

Create a pair table of a secondary structure (pseudo-knot version)

Returns a newly allocated table, such that table[i]=j if (i.j) pair or 0 if i is unpaired, table[0] contains the length of the structure.

In contrast to vrna_ptable() this function also recognizes the base pairs denoted by '[' and ']' brackets. Thus, this function behaves like

```
vrna_ptable_from_string(structure, #VRNA_BRACKET_RND | VRNA_BRACKETS_SQR)
```

See also

```
vrna_ptable_from_string()
```

Parameters

structure | The secondary structure in (extended) dot-bracket notation

Returns

A pointer to the created pair_table

16.61.2.4 vrna_ptable_copy()

```
short* vrna_ptable_copy (

const short * pt )
```

#include <ViennaRNA/utils/structures.h>

Get an exact copy of a pair table.

Parameters

```
pt The pair table to be copied
```

Returns

A pointer to the copy of 'pt'

16.61.2.5 vrna_pt_ali_get()

Create a pair table of a secondary structure (snoop align version)

16.61.2.6 vrna_pt_snoop_get()

Create a pair table of a secondary structure (snoop version)

returns a newly allocated table, such that: table[i]=j if (i.j) pair or 0 if i is unpaired, table[0] contains the length of the structure. The special pseudoknotted H/ACA-mRNA structure is taken into account.

16.61.2.7 vrna_pt_pk_remove()

Remove pseudo-knots from a pair table.

This function removes pseudo-knots from an input structure by determining the minimum number of base pairs that need to be removed to make the structure pseudo-knot free.

To accomplish that, we use a dynamic programming algorithm similar to the Nussinov maxmimum matching approach.

See also

```
vrna_db_pk_remove()
```

Parameters

ptable	Input structure that may include pseudo-knots
options	

Returns

The input structure devoid of pseudo-knots

16.62 Pair List Representation of Secondary Structures

16.62.1 Detailed Description

Collaboration diagram for Pair List Representation of Secondary Structures:

Data Structures

· struct vrna_elem_prob_s

Data structure representing a single entry of an element probability list (e.g. list of pair probabilities) More...

Macros

• #define VRNA PLIST TYPE BASEPAIR 0

A Base Pair element.

• #define VRNA PLIST TYPE GQUAD 1

A G-Quadruplex element.

#define VRNA_PLIST_TYPE_H_MOTIF 2

A Hairpin loop motif element.

• #define VRNA_PLIST_TYPE_I_MOTIF 3

An Internal loop motif element.

#define VRNA_PLIST_TYPE_UD_MOTIF 4

An Unstructured Domain motif element.

#define VRNA_PLIST_TYPE_STACK 5

A Base Pair stack element.

Typedefs

typedef struct vrna_elem_prob_s vrna_ep_t

Convenience typedef for data structure vrna_elem_prob_s.

Functions

vrna_ep_t * vrna_plist (const char *struc, float pr)

Create a vrna_ep_t from a dot-bracket string.

16.62.2 Data Structure Documentation

16.62.2.1 struct vrna_elem_prob_s

Data structure representing a single entry of an element probability list (e.g. list of pair probabilities)

See also

```
vrna_plist(), vrna_plist_from_probs(), vrna_db_from_plist(), VRNA_PLIST_TYPE_BASEPAIR, VRNA_PLIST_TYPE_GQUAD, VRNA_PLIST_TYPE_H_MOTIF, VRNA_PLIST_TYPE_I_MOTIF, VRNA_PLIST_TYPE_UD_MOTIF, VRNA_PLIST_TYPE_STACK
```

Data Fields

int i

Start position (usually 5' nucleotide that starts the element, e.g. base pair)

int j

End position (usually 3' nucleotide that ends the element, e.g. base pair)

float p

Probability of the element.

· int type

Type of the element.

16.62.3 Function Documentation

16.62.3.1 vrna_plist()

Create a vrna_ep_t from a dot-bracket string.

The dot-bracket string is parsed and for each base pair an entry in the plist is created. The probability of each pair in the list is set by a function parameter.

The end of the plist is marked by sequence positions i as well as j equal to 0. This condition should be used to stop looping over its entries

Parameters

struc	The secondary structure in dot-bracket notation
pr	The probability for each base pair used in the plist

Returns

The plist array

16.63 Helix List Representation of Secondary Structures

16.63.1 Detailed Description

Collaboration diagram for Helix List Representation of Secondary Structures:

Data Structures

struct vrna_hx_s

Data structure representing an entry of a helix list. More...

Typedefs

typedef struct vrna_hx_s vrna_hx_t
 Convenience typedef for data structure vrna_hx_s.

Functions

vrna_hx_t * vrna_hx_from_ptable (short *pt)

Convert a pair table representation of a secondary structure into a helix list.

vrna_hx_t * vrna_hx_merge (const vrna_hx_t *list, int maxdist)

Create a merged helix list from another helix list.

16.63.2 Data Structure Documentation

```
16.63.2.1 struct vrna_hx_s
```

Data structure representing an entry of a helix list.

16.63.3 Function Documentation

```
16.63.3.1 vrna_hx_from_ptable()
```

#include <ViennaRNA/utils/structures.h>

Convert a pair table representation of a secondary structure into a helix list.

Parameters

pt The secondary structure in pair table representation

Returns

The secondary structure represented as a helix list

16.64 Tree Representation of Secondary Structures

16.64.1 Detailed Description

Secondary structures can be readily represented as trees, where internal nodes represent base pairs, and leaves represent unpaired nucleotides. The dot-bracket structure string already is a tree represented by a string of parenthesis (base pairs) and dots for the leaf nodes (unpaired nucleotides).

See Tree Representations of Secondary Structures for a detailed description on tree representation of secondary structures. Collaboration diagram for Tree Representation of Secondary Structures:

Macros

- #define VRNA_STRUCTURE_TREE_HIT 1U
 - Homeomorphically Irreducible Tree (HIT) representation of a secondary structure.
- #define VRNA_STRUCTURE_TREE_SHAPIRO_SHORT 2U
 - (short) Coarse Grained representation of a secondary structure
- #define VRNA_STRUCTURE_TREE_SHAPIRO 3U
 - (full) Coarse Grained representation of a secondary structure
- #define VRNA_STRUCTURE_TREE_SHAPIRO_EXT 4U
 - (extended) Coarse Grained representation of a secondary structure
- #define VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT 5U
 - (weighted) Coarse Grained representation of a secondary structure
- #define VRNA_STRUCTURE_TREE_EXPANDED 6U
 - Expanded Tree representation of a secondary structure.

Functions

- char * vrna_db_to_tree_string (const char *structure, unsigned int type)
 - Convert a Dot-Bracket structure string into tree string representation.
- char * vrna_tree_string_unweight (const char *structure)
 - Remove weights from a linear string tree representation of a secondary structure.
- char * vrna_tree_string_to_db (const char *tree)

Convert a linear tree string representation of a secondary structure back to Dot-Bracket notation.

16.64.2 Macro Definition Documentation

16.64.2.1 VRNA_STRUCTURE_TREE_HIT

```
#define VRNA_STRUCTURE_TREE_HIT 1U
```

#include <ViennaRNA/utils/structures.h>

Homeomorphically Irreducible Tree (HIT) representation of a secondary structure.

See also

vrna_db_to_tree_string()

```
16.64.2.2 VRNA_STRUCTURE_TREE_SHAPIRO_SHORT
#define VRNA_STRUCTURE_TREE_SHAPIRO_SHORT 2U
#include <ViennaRNA/utils/structures.h>
(short) Coarse Grained representation of a secondary structure
See also
     vrna_db_to_tree_string()
16.64.2.3 VRNA_STRUCTURE_TREE_SHAPIRO
#define VRNA_STRUCTURE_TREE_SHAPIRO 3U
#include <ViennaRNA/utils/structures.h>
(full) Coarse Grained representation of a secondary structure
See also
     vrna_db_to_tree_string()
16.64.2.4 VRNA_STRUCTURE_TREE_SHAPIRO_EXT
#define VRNA_STRUCTURE_TREE_SHAPIRO_EXT 4U
#include <ViennaRNA/utils/structures.h>
(extended) Coarse Grained representation of a secondary structure
See also
    vrna_db_to_tree_string()
16.64.2.5 VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT
#define VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT 5U
#include <ViennaRNA/utils/structures.h>
(weighted) Coarse Grained representation of a secondary structure
See also
     vrna_db_to_tree_string()
```

16.64.2.6 VRNA_STRUCTURE_TREE_EXPANDED

```
#define VRNA_STRUCTURE_TREE_EXPANDED 6U
#include <ViennaRNA/utils/structures.h>
```

Expanded Tree representation of a secondary structure.

See also

```
vrna db to tree string()
```

16.64.3 Function Documentation

16.64.3.1 vrna db to tree string()

Convert a Dot-Bracket structure string into tree string representation.

This function allows one to convert a secondary structure in dot-bracket notation into one of the various tree representations for secondary structures. The resulting tree is then represented as a string of parenthesis and node symbols, similar to to the Newick format.

Currently we support conversion into the following formats, denoted by the value of parameter type:

- VRNA_STRUCTURE_TREE_HIT Homeomorphically Irreducible Tree (HIT) representation of a secondary structure. (See also Fontana et al. 1993 [8])
- VRNA_STRUCTURE_TREE_SHAPIRO_SHORT (short) Coarse Grained representation of a secondary structure (same as Shapiro 1988 [20], but with root node R and without S nodes for the stems)
- VRNA_STRUCTURE_TREE_SHAPIRO (full) Coarse Grained representation of a secondary structure (See also Shapiro 1988 [20])
- VRNA_STRUCTURE_TREE_SHAPIRO_EXT (extended) Coarse Grained representation of a secondary structure (same as Shapiro 1988 [20], but external nodes denoted as $\mathbb E$)
- VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT (weighted) Coarse Grained representation of a secondary structure (same as VRNA_STRUCTURE_TREE_SHAPIRO_EXT but with additional weights for number of unpaired nucleotides in loop, and number of pairs in stems)
- VRNA_STRUCTURE_TREE_EXPANDED Expanded Tree representation of a secondary structure.

See also

Tree Representations of Secondary Structures

Parameters

structure	The null-terminated dot-bracket structure string
type	A switch to determine the type of tree string representation

Returns

A tree representation of the input structure

16.64.3.2 vrna_tree_string_unweight()

Remove weights from a linear string tree representation of a secondary structure.

This function strips the weights of a linear string tree representation such as ${\tt HIT}$, or Coarse Grained Tree sensu Shapiro [20]

See also

```
vrna_db_to_tree_string()
```

Parameters

ſ	structure	A linear string tree representation of a secondary structure with weights]
---	-----------	---	---

Returns

A linear string tree representation of a secondary structure without weights

16.64.3.3 vrna_tree_string_to_db()

Convert a linear tree string representation of a secondary structure back to Dot-Bracket notation.

Warning

This function only accepts Expanded and HIT tree representations!

See also

vrna_db_to_tree_string(), VRNA_STRUCTURE_TREE_EXPANDED, VRNA_STRUCTURE_TREE_HIT, Tree Representations of Secondary Structures

Parameters

tree	A linear tree string representation of a secondary structure
------	--

Returns

A dot-bracket notation of the secondary structure provided in \mathtt{tree}

16.65 Multiple Sequence Alignment Utilities

Functions to extract features from and to manipulate multiple sequence alignments.

16.65.1 Detailed Description

Functions to extract features from and to manipulate multiple sequence alignments.

Collaboration diagram for Multiple Sequence Alignment Utilities:

Modules

• Deprecated Interface for Multiple Sequence Alignment Utilities

Files

· file alignments.h

Various utility- and helper-functions for sequence alignments and comparative structure prediction.

Data Structures

struct vrna_pinfo_s

A base pair info structure. More...

Macros

#define VRNA_ALN_DEFAULT 0U

Use default alignment settings.

#define VRNA_ALN_RNA 1U

Convert to RNA alphabet.

#define VRNA_ALN_DNA 2U

Convert to DNA alphabet.

#define VRNA_ALN_UPPERCASE 4U

Convert to uppercase nucleotide letters.

• #define VRNA_ALN_LOWERCASE 8U

Convert to lowercase nucleotide letters.

#define VRNA_MEASURE_SHANNON_ENTROPY 1U

Flag indicating Shannon Entropy measure.

Typedefs

typedef struct vrna_pinfo_s vrna_pinfo_t

Typename for the base pair info repesenting data structure vrna_pinfo_s.

Functions

• int vrna aln mpi (const char **alignment)

Get the mean pairwise identity in steps from ?to?(ident)

vrna_pinfo_t * vrna_aln_pinfo (vrna_fold_compound_t *vc, const char *structure, double threshold)

Retrieve an array of vrna_pinfo_t structures from precomputed pair probabilities.

char ** vrna aln slice (const char **alignment, unsigned int i, unsigned int j)

Slice out a subalignment from a larger alignment.

void vrna_aln_free (char **alignment)

Free memory occupied by a set of aligned sequences.

char ** vrna_aln_uppercase (const char **alignment)

Create a copy of an alignment with only uppercase letters in the sequences.

char ** vrna_aln_toRNA (const char **alignment)

Create a copy of an alignment where DNA alphabet is replaced by RNA alphabet.

char ** vrna_aln_copy (const char **alignment, unsigned int options)

Make a copy of a multiple sequence alignment.

- float * vrna_aln_conservation_struct (const char **alignment, const char *structure, const vrna_md_t *md)

 Compute base pair conservation of a consensus structure.
- float * vrna_aln_conservation_col (const char **alignment, const vrna_md_t *md_p, unsigned int options)

 Compute nucleotide conservation in an alignment.
- char * vrna_aln_consensus_sequence (const char **alignment, const vrna_md_t *md_p)

Compute the consensus sequence for a given multiple sequence alignment.

• char * vrna_aln_consensus_mis (const char **alignment, const vrna_md_t *md_p)

Compute the Most Informative Sequence (MIS) for a given multiple sequence alignment.

16.65.2 Data Structure Documentation

16.65.2.1 struct vrna_pinfo_s

A base pair info structure.

For each base pair (i,j) with i,j in [0, n-1] the structure lists:

- · its probability 'p'
- · an entropy-like measure for its well-definedness 'ent'
- the frequency of each type of pair in 'bp[]'
 - 'bp[0]' contains the number of non-compatible sequences
 - 'bp[1]' the number of CG pairs, etc.

Data Fields

unsigned i

nucleotide position i

unsigned j

nucleotide position j

float p

Probability.

· float ent

Pseudo entropy for $p(i,j) = S_i + S_j - p_i j * ln(p_i j)$.

short bp [8]

Frequencies of pair_types.

· char comp

1 iff pair is in mfe structure

16.65.3 Macro Definition Documentation

16.65.3.1 VRNA_MEASURE_SHANNON_ENTROPY

```
#define VRNA_MEASURE_SHANNON_ENTROPY 1U
#include <ViennaRNA/utils/alignments.h>
```

Flag indicating Shannon Entropy measure.

Shannon Entropy is defined as $H = -\sum_c p_c \cdot \log_2 p_c$

16.65.4 Function Documentation

16.65.4.1 vrna_aln_mpi()

Get the mean pairwise identity in steps from ?to?(ident)

Parameters

```
alignment Aligned sequences
```

Returns

The mean pairwise identity

16.65.4.2 vrna_aln_pinfo()

Retrieve an array of vrna_pinfo_t structures from precomputed pair probabilities.

This array of structures contains information about positionwise pair probabilies, base pair entropy and more

See also

vrna_pinfo_t, and vrna_pf()

Parameters

VC	The vrna_fold_compound_t of type VRNA_FC_TYPE_COMPARATIVE with precomputed partition
	function matrices
structure	An optional structure in dot-bracket notation (Maybe NULL)
threshold	Do not include results with pair probabilities below threshold

Returns

The vrna_pinfo_t array

16.65.4.3 vrna_aln_slice()

#include <ViennaRNA/utils/alignments.h>

Slice out a subalignment from a larger alignment.

Note

The user is responsible to free the memory occupied by the returned subalignment

See also

```
vrna_aln_free()
```

Parameters

alignment	The input alignment
i	The first column of the subalignment (1-based)
j	The last column of the subalignment (1-based)

Returns

The subalignment between column i and j

16.65.4.4 vrna_aln_free()

#include <ViennaRNA/utils/alignments.h>

Free memory occupied by a set of aligned sequences.

Parameters

16.65.4.5 vrna_aln_uppercase()

Create a copy of an alignment with only uppercase letters in the sequences.

See also

```
vrna_aln_copy
```

Parameters

alignment	The input sequence alignment (last entry must be NULL terminated)
-----------	---

Returns

A copy of the input alignment where lowercase sequence letters are replaced by uppercase letters

16.65.4.6 vrna_aln_toRNA()

Create a copy of an alignment where DNA alphabet is replaced by RNA alphabet.

See also

```
vrna_aln_copy
```

Parameters

alignment	The input sequence alignment (last entry must be <i>NULL</i> terminated)

Returns

A copy of the input alignment where DNA alphabet is replaced by RNA alphabet (T -> U)

16.65.4.7 vrna_aln_copy()

#include <ViennaRNA/utils/alignments.h>

Make a copy of a multiple sequence alignment.

This function allows one to create a copy of a multiple sequence alignment. The options parameter additionally allows for sequence manipulation, such as converting DNA to RNA alphabet, and conversion to uppercase letters.

See also

```
vrna_aln_copy(), VRNA_ALN_RNA, VRNA_ALN_UPPERCASE, VRNA_ALN_DEFAULT
```

Parameters

alignment	The input sequence alignment (last entry must be NULL terminated)
options	Option flags indicating whether the aligned sequences should be converted

Returns

A (manipulated) copy of the input alignment

16.65.4.8 vrna_aln_conservation_struct()

#include <ViennaRNA/utils/alignments.h>

Compute base pair conservation of a consensus structure.

This function computes the base pair conservation (fraction of canonical base pairs) of a consensus structure given a multiple sequence alignment. The base pair types that are considered canonical may be specified using the vrna_md_t.pair array. Passing NULL as parameter md results in default pairing rules, i.e. canonical Watson-Crick and GU Wobble pairs.

Parameters

alignment	The input sequence alignment (last entry must be NULL terminated)
structure	The consensus structure in dot-bracket notation
md	Model details that specify compatible base pairs (Maybe NULL)

Returns

A 1-based vector of base pair conservations

SWIG Wrapper Notes This function is available in an overloaded form where the last parameter may be omitted, indicating md = *NULL*

16.65.4.9 vrna_aln_conservation_col()

#include <ViennaRNA/utils/alignments.h>

Compute nucleotide conservation in an alignment.

This function computes the conservation of nucleotides in alignment columns. The simples measure is Shannon Entropy and can be selected by passing the VRNA_MEASURE_SHANNON_ENTROPY flag in the options parameter.

Note

Currently, only VRNA_MEASURE_SHANNON_ENTROPY is supported as conservation measure.

See also

VRNA_MEASURE_SHANNON_ENTROPY

Parameters

alignment	The input sequence alignment (last entry must be NULL terminated)
md	Model details that specify known nucleotides (Maybe NULL)
options	A flag indicating which measure of conservation should be applied

Returns

A 1-based vector of column conservations

SWIG Wrapper Notes This function is available in an overloaded form where the last two parameters may be omitted, indicating md = *NULL*, and options = VRNA_MEASURE_SHANNON_ENTROPY,

respectively.

16.65.4.10 vrna_aln_consensus_sequence()

#include <ViennaRNA/utils/alignments.h>

Compute the consensus sequence for a given multiple sequence alignment.

Parameters

alignment	The input sequence alignment (last entry must be NULL terminated)
md_p	Model details that specify known nucleotides (Maybe NULL)

Returns

The consensus sequence of the alignment, i.e. the most frequent nucleotide for each alignment column

16.65.4.11 vrna_aln_consensus_mis()

Compute the Most Informative Sequence (MIS) for a given multiple sequence alignment.

The most informative sequence (MIS) [9] displays for each alignment column the nucleotides with frequency greater than the background frequency, projected into IUPAC notation. Columns where gaps are over-represented are in lower case.

Parameters

alignment	The input sequence alignment (last entry must be NULL terminated)
md_p	Model details that specify known nucleotides (Maybe NULL)

Returns

The most informative sequence for the alignment

16.66 Files and I/O 511

16.66 Files and I/O

Functions to parse, write, and convert various file formats and to deal with file system related issues.

16.66.1 Detailed Description

Functions to parse, write, and convert various file formats and to deal with file system related issues.

Collaboration diagram for Files and I/O:

Modules

Nucleic Acid Sequences and Structures

Functions to read/write different file formats for nucleic acid sequences and secondary structures.

Multiple Sequence Alignments

Functions to read/write multiple sequence alignments (MSA) in various file formats.

· Command Files

Functions to parse and interpret the content of Command Files.

Files

· file commands.h

Parse and apply different commands that alter the behavior of secondary structure prediction and evaluation.

· file ribo.h

Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments.

• file file_formats.h

Read and write different file formats for RNA sequences, structures.

file file_formats_msa.h

Functions dealing with file formats for Multiple Sequence Alignments (MSA)

· file utils.h

Several utilities for file handling.

Functions

float ** get_ribosum (const char **Alseq, int n_seq, int length)

Retrieve a RiboSum Scoring Matrix for a given Alignment.

• float ** readribosum (char *name)

Read a RiboSum or other user-defined Scoring Matrix and Store into global Memory.

void vrna_file_copy (FILE *from, FILE *to)

Inefficient 'cp'.

char * vrna_read_line (FILE *fp)

Read a line of arbitrary length from a stream.

int vrna_mkdir_p (const char *path)

Recursivly create a directory tree.

char * vrna_basename (const char *path)

Extract the filename from a file path.

char * vrna_dirname (const char *path)

Extract the directory part of a file path.

• char * vrna_filename_sanitize (const char *name, const char *replacement)

Sanitize a file name.

• int vrna_file_exists (const char *filename)

Check if a file already exists in the file system.

16.66.2 Function Documentation

Read a RiboSum or other user-defined Scoring Matrix and Store into global Memory.

Read a line of arbitrary length from a stream.

Returns a pointer to the resulting string. The necessary memory is allocated and should be released using *free()* when the string is no longer needed.

Parameters

fp A file pointer to the stream where the function should read from

Returns

A pointer to the resulting string

16.66.2.3 vrna_filename_sanitize()

Sanitize a file name.

Returns a new file name where all invalid characters are substituted by a replacement character. If no replacement character is supplied, invalid characters are simply removed from the filename. File names may also never exceed a length of 255 characters. Longer file names will undergo a 'smart' truncation process, where the filenames' suffix,

16.66 Files and I/O 513

i.e. everything after the last dot '.', is attempted to be kept intact. Hence, only the filename part before the suffix is reduced in such a way that the total filename complies to the length restriction of 255 characters. If no suffix is present or the suffix itself already exceeds the maximum length, the filename is simply truncated from the back of the string.

For now we consider the following characters invalid:

- · backslash '\'
- slash '/'
- · question mark '?'
- · percent sign "
- · asterisk '*'
- · colon ':'
- pipe symbol '|'
- · double quote ""
- triangular brackets '<' and '>'

Furthermore, the (resulting) file name must not be a reserved file name, such as:

- '.'
- '..'

Note

This function allocates a new block of memory for the sanitized string. It also may return (a) NULL if the input is pointing to NULL, or (b) an empty string if the input only consists of invalid characters which are simply removed!

Parameters

name	The input file name
replacement	The replacement character, or NULL

Returns

The sanitized file name, or NULL

16.66.2.4 vrna_file_exists()

Check if a file already exists in the file system.

Parameters

filename 1	The name of (path to) the file to check for existence
------------	---

Returns

0 if it doesn't exists, 1 otherwise

16.67 Nucleic Acid Sequences and Structures

Functions to read/write different file formats for nucleic acid sequences and secondary structures.

16.67.1 Detailed Description

Functions to read/write different file formats for nucleic acid sequences and secondary structures.

Collaboration diagram for Nucleic Acid Sequences and Structures:

Files

· file file formats.h

Read and write different file formats for RNA sequences, structures.

Macros

• #define VRNA OPTION MULTILINE 32U

Tell a function that an input is assumed to span several lines.

• #define VRNA_CONSTRAINT_MULTILINE 32U

parse multiline constraint

Functions

• void vrna_file_helixlist (const char *seq, const char *db, float energy, FILE *file)

Print a secondary structure as helix list.

• void vrna_file_connect (const char *seq, const char *db, float energy, const char *identifier, FILE *file)

Print a secondary structure as connect table.

void vrna_file_bpseq (const char *seq, const char *db, FILE *file)

Print a secondary structure in bpseq format.

• void vrna_file_json (const char *seq, const char *db, double energy, const char *identifier, FILE *file)

Print a secondary structure in jsonformat.

- unsigned int vrna_file_fasta_read_record (char **header, char **sequence, char ***rest, FILE *file, unsigned int options)
- char * vrna_extract_record_rest_structure (const char **lines, unsigned int length, unsigned int option)

 Extract a dot-bracket structure string from (multiline)character array.
- int vrna_file_SHAPE_read (const char *file_name, int length, double default_value, char *sequence, double *values)

Read data from a given SHAPE reactivity input file.

• void vrna_extract_record_rest_constraint (char **cstruc, const char **lines, unsigned int option)

Extract a hard constraint encoded as pseudo dot-bracket string.

• unsigned int read_record (char **header, char **sequence, char ***rest, unsigned int options)

Get a data record from stdin.

16.67.2 Macro Definition Documentation

16.67.2.1 VRNA_OPTION_MULTILINE

```
#define VRNA_OPTION_MULTILINE 32U
#include <ViennaRNA/io/file_formats.h>
```

Tell a function that an input is assumed to span several lines.

If used as input-option a function might also be returning this state telling that it has read data from multiple lines.

See also

```
vrna_extract_record_rest_structure(), vrna_file_fasta_read_record()
```

16.67.2.2 VRNA_CONSTRAINT_MULTILINE

```
#define VRNA_CONSTRAINT_MULTILINE 32U
#include <ViennaRNA/io/file_formats.h>
```

parse multiline constraint

Deprecated see vrna_extract_record_rest_structure()

16.67.3 Function Documentation

16.67.3.1 vrna_file_helixlist()

#include <ViennaRNA/io/file_formats.h>

Print a secondary structure as helix list.

Parameters

seq	The RNA sequence
db	The structure in dot-bracket format
energy	Free energy of the structure in kcal/mol
file	The file handle used to print to (print defaults to 'stdout' if(file == NULL))

16.67.3.2 vrna_file_connect()

#include <ViennaRNA/io/file_formats.h>

Print a secondary structure as connect table.

Connect table file format looks like this:

```
* 300 ENERGY = 7.0 example
* 1 G 0 2 22 1
* 2 G 1 3 21 2
```

where the headerline is followed by 6 columns with:

- 1. Base number: index n
- 2. Base (A, C, G, T, U, X)
- 3. Index n-1 (0 if first nucleotide)
- 4. Index n+1 (0 if last nucleotide)
- 5. Number of the base to which n is paired. No pairing is indicated by 0 (zero).
- 6. Natural numbering.

Parameters

seq	The RNA sequence
db	The structure in dot-bracket format
energy	The free energy of the structure
identifier	An optional identifier for the sequence
file	The file handle used to print to (print defaults to 'stdout' if(file == NULL))

16.67.3.3 vrna_file_bpseq()

```
#include <ViennaRNA/io/file_formats.h>
```

Print a secondary structure in bpseq format.

Parameters

seq	The RNA sequence
db	The structure in dot-bracket format
file	The file handle used to print to (print defaults to 'stdout' if(file == NULL))

16.67.3.4 vrna_file_json()

#include <ViennaRNA/io/file_formats.h>

Print a secondary structure in jsonformat.

Parameters

seq	The RNA sequence
db	The structure in dot-bracket format
energy	The free energy
identifier	An identifier for the sequence
file	The file handle used to print to (print defaults to 'stdout' if(file == NULL))

16.67.3.5 vrna_file_fasta_read_record()

then vrna_file_fasta_read_record() will assume that the sequence that follows the header may span over several lines. To disable this behavior and to assign a single line to the argument 'sequence' one can pass VRNA_INPUT_NO_SPAN in the 'options' argument. If no fasta header is read in the beginning of a data block, a sequence must not span over multiple lines!

Unless the options VRNA_INPUT_NOSKIP_COMMENTS or VRNA_INPUT_NOSKIP_BLANK_LINES are passed, a sequence may be interrupted by lines starting with a comment character or empty lines.

A sequence is regarded as completely read if it was either assumed to not span over multiple lines, a secondary structure or structure constraint follows the sequence on the next line, or a new header marks the beginning of a new sequence...

All lines following the sequence (this includes comments) that do not initiate a new dataset according to the above definition are available through the line-array 'rest'. Here one can usually find the structure constraint or other information belonging to the current dataset. Filling of 'rest' may be prevented by passing VRNA_INPUT_NO_REST to the options argument.

Note

This function will exit any program with an error message if no sequence could be read!

This function is NOT threadsafe! It uses a global variable to store information about the next data block.

The main purpose of this function is to be able to easily parse blocks of data in the header of a loop where all calculations for the appropriate data is done inside the loop. The loop may be then left on certain return values,

```
e.g.:
```

```
@endverbatim
char *id, *seq, **rest;
int i;
id = seq = NULL;
rest = NULL;
while(!(vrna_file_fasta_read_record(&id, &seq, &rest, NULL, 0) & (VRNA_INPUT_ERROR | VRNA_INPUT_QUIT))){
    if(id)
        printf("%s\n", id);
        printf("%s\n", seq);
    if(rest)
        for(i=0;rest[i];i++){
            printf("%s\n", rest[i]);
            free(rest[i]);
    }
    free(cest);
    free(seq);
    free(id);
}
```

In the example above, the while loop will be terminated when vrna_file_fasta_read_record() returns either an error, EOF, or a user initiated quit request.

As long as data is read from stdin (we are passing NULL as the file pointer), the id is printed if it is available for the current block of data. The sequence will be printed in any case and if some more lines belong to the current block of data each line will be printed as well.

Note

Do not forget to free the memory occupied by header, sequence and rest!

Parameters

header	A pointer which will be set such that it points to the header of the record
sequence	A pointer which will be set such that it points to the sequence of the record
rest	A pointer which will be set such that it points to an array of lines which also belong to the record
file	A file handle to read from (if NULL, this function reads from stdin)
options	Some options which may be passed to alter the behavior of the function, use 0 for no options

Returns

A flag with information about what the function actually did read

16.67.3.6 vrna_extract_record_rest_structure()

#include <ViennaRNA/io/file_formats.h>

Extract a dot-bracket structure string from (multiline)character array.

This function extracts a dot-bracket structure string from the 'rest' array as returned by vrna_file_fasta_read_record() and returns it. All occurences of comments within the 'lines' array will be skipped as long as they do not break the structure string. If no structure could be read, this function returns NULL.

Precondition

The argument 'lines' has to be a 2-dimensional character array as obtained by vrna_file_fasta_read_record()

See also

```
vrna_file_fasta_read_record()
```

Parameters

lines	The (multiline) character array to be parsed
length	The assumed length of the dot-bracket string (passing a value $<$ 1 results in no length limit)
option	Some options which may be passed to alter the behavior of the function, use 0 for no options

Returns

The dot-bracket string read from lines or NULL

16.67.3.7 vrna_file_SHAPE_read()

```
#include <ViennaRNA/io/file_formats.h>
```

Read data from a given SHAPE reactivity input file.

This function parses the informations from a given file and stores the result in the preallocated string sequence and the double array values.

Parameters

file_name	Path to the constraints file
length	Length of the sequence (file entries exceeding this limit will cause an error)
default_value	Value for missing indices
sequence	Pointer to an array used for storing the sequence obtained from the SHAPE reactivity file
values	Pointer to an array used for storing the values obtained from the SHAPE reactivity file

16.67.3.8 vrna_extract_record_rest_constraint()

#include <ViennaRNA/io/file_formats.h>

Extract a hard constraint encoded as pseudo dot-bracket string.

Deprecated Use vrna_extract_record_rest_structure() instead!

Precondition

The argument 'lines' has to be a 2-dimensional character array as obtained by vrna_file_fasta_read_record()

See also

vrna_file_fasta_read_record(), VRNA_CONSTRAINT_DB_PIPE, VRNA_CONSTRAINT_DB_DOT, VRNA_CONSTRAINT_DB_VRNA_CONSTRAINT_DB_ANG_BRACK, VRNA_CONSTRAINT_DB_RND_BRACK

Parameters

cstruc	A pointer to a character array that is used as pseudo dot-bracket output
lines	A 2-dimensional character array with the extension lines from the FASTA input
option	The option flags that define the behavior and recognition pattern of this function

16.67.3.9 read_record()

Get a data record from stdin.

Deprecated This function is deprecated! Use vrna_file_fasta_read_record() as a replacment.

16.68 Multiple Sequence Alignments

Functions to read/write multiple sequence alignments (MSA) in various file formats.

16.68.1 Detailed Description

Functions to read/write multiple sequence alignments (MSA) in various file formats.

Collaboration diagram for Multiple Sequence Alignments:

Files

· file file formats msa.h

Functions dealing with file formats for Multiple Sequence Alignments (MSA)

Macros

• #define VRNA_FILE_FORMAT_MSA_CLUSTAL 1U

Option flag indicating ClustalW formatted files.

#define VRNA_FILE_FORMAT_MSA_STOCKHOLM 2U

Option flag indicating Stockholm 1.0 formatted files.

#define VRNA FILE FORMAT MSA FASTA 4U

Option flag indicating FASTA (Pearson) formatted files.

#define VRNA_FILE_FORMAT_MSA_MAF 8U

Option flag indicating MAF formatted files.

• #define VRNA FILE FORMAT MSA MIS 16U

Option flag indicating most informative sequence (MIS) output.

• #define VRNA_FILE_FORMAT_MSA_DEFAULT

Option flag indicating the set of default file formats.

• #define VRNA_FILE_FORMAT_MSA_NOCHECK 4096U

Option flag to disable validation of the alignment.

• #define VRNA FILE FORMAT MSA UNKNOWN 8192U

Return flag of vrna_file_msa_detect_format() to indicate unknown or malformatted alignment.

#define VRNA_FILE_FORMAT_MSA_APPEND 16384U

Option flag indicating to append data to a multiple sequence alignment file rather than overwriting it.

#define VRNA_FILE_FORMAT_MSA_QUIET 32768U

Option flag to suppress unnecessary spam messages on stderr

#define VRNA_FILE_FORMAT_MSA_SILENT 65536U

Option flag to completely silence any warnings on stderr

Functions

• int vrna_file_msa_read (const char *filename, char ***names, char ***aln, char **id, char **structure, unsigned int options)

Read a multiple sequence alignment from file.

• int vrna_file_msa_read_record (FILE *fp, char ***names, char ***aln, char **id, char **structure, unsigned int options)

Read a multiple sequence alignment from file handle.

• unsigned int vrna file msa detect format (const char *filename, unsigned int options)

Detect the format of a multiple sequence alignment file.

• int vrna_file_msa_write (const char *filename, const char **names, const char **aln, const char *id, const char *structure, const char *source, unsigned int options)

Write multiple sequence alignment file.

16.68.2 Macro Definition Documentation

```
16.68.2.1 VRNA_FILE_FORMAT_MSA_CLUSTAL
#define VRNA_FILE_FORMAT_MSA_CLUSTAL 1U
#include <ViennaRNA/io/file_formats_msa.h>
Option flag indicating ClustalW formatted files.
See also
     vrna_file_msa_read(), vrna_file_msa_read_record(), vrna_file_msa_detect_format()
16.68.2.2 VRNA_FILE_FORMAT_MSA_STOCKHOLM
#define VRNA_FILE_FORMAT_MSA_STOCKHOLM 2U
#include <ViennaRNA/io/file_formats_msa.h>
Option flag indicating Stockholm 1.0 formatted files.
See also
     vrna_file_msa_read(), vrna_file_msa_read_record(), vrna_file_msa_detect_format()
16.68.2.3 VRNA_FILE_FORMAT_MSA_FASTA
#define VRNA_FILE_FORMAT_MSA_FASTA 4U
#include <ViennaRNA/io/file_formats_msa.h>
Option flag indicating FASTA (Pearson) formatted files.
See also
     vrna_file_msa_read(), vrna_file_msa_read_record(), vrna_file_msa_detect_format()
```

```
16.68.2.4 VRNA_FILE_FORMAT_MSA_MAF
```

```
#define VRNA_FILE_FORMAT_MSA_MAF 8U
#include <ViennaRNA/io/file_formats_msa.h>
```

Option flag indicating MAF formatted files.

See also

vrna_file_msa_read(), vrna_file_msa_read_record(), vrna_file_msa_detect_format()

```
16.68.2.5 VRNA_FILE_FORMAT_MSA_MIS
```

```
#define VRNA_FILE_FORMAT_MSA_MIS 16U
#include <ViennaRNA/io/file_formats_msa.h>
```

Option flag indicating most informative sequence (MIS) output.

The default reference sequence output for an alignment is simply a consensus sequence. This flag allows to write the most informative equence (MIS) instead.

See also

```
vrna_file_msa_write()
```

```
16.68.2.6 VRNA_FILE_FORMAT_MSA_DEFAULT
```

```
#define VRNA_FILE_FORMAT_MSA_DEFAULT

#include <ViennaRNA/io/file_formats_msa.h>

Value:
( \
    VRNA_FILE_FORMAT_MSA_CLUSTAL \
    | VRNA_FILE_FORMAT_MSA_STOCKHOLM \
    | VRNA_FILE_FORMAT_MSA_FASTA \
    | VRNA_FILE_FORMAT_MSA_MAF \
```

Option flag indicating the set of default file formats.

See also

```
vrna_file_msa_read(), vrna_file_msa_read_record(), vrna_file_msa_detect_format()
```

```
16.68.2.7 VRNA_FILE_FORMAT_MSA_NOCHECK
#define VRNA_FILE_FORMAT_MSA_NOCHECK 4096U
#include <ViennaRNA/io/file_formats_msa.h>
Option flag to disable validation of the alignment.
See also
     vrna file msa read(), vrna file msa read record()
16.68.2.8 VRNA_FILE_FORMAT_MSA_UNKNOWN
#define VRNA_FILE_FORMAT_MSA_UNKNOWN 8192U
#include <ViennaRNA/io/file_formats_msa.h>
Return flag of vrna_file_msa_detect_format() to indicate unknown or malformatted alignment.
See also
     vrna_file_msa_detect_format()
16.68.2.9 VRNA_FILE_FORMAT_MSA_APPEND
#define VRNA_FILE_FORMAT_MSA_APPEND 16384U
#include <ViennaRNA/io/file_formats_msa.h>
Option flag indicating to append data to a multiple sequence alignment file rather than overwriting it.
See also
     vrna_file_msa_write()
16.68.2.10 VRNA_FILE_FORMAT_MSA_QUIET
#define VRNA_FILE_FORMAT_MSA_QUIET 32768U
#include <ViennaRNA/io/file_formats_msa.h>
Option flag to suppress unnecessary spam messages on stderr
See also
     vrna_file_msa_read(), vrna_file_msa_read_record()
```

16.68.2.11 VRNA_FILE_FORMAT_MSA_SILENT

```
#define VRNA_FILE_FORMAT_MSA_SILENT 65536U
#include <ViennaRNA/io/file_formats_msa.h>
```

Option flag to completely silence any warnings on stderr

See also

```
vrna_file_msa_read(), vrna_file_msa_read_record()
```

16.68.3 Function Documentation

16.68.3.1 vrna_file_msa_read()

Read a multiple sequence alignment from file.

This function reads the (first) multiple sequence alignment from an input file. The read alignment is split into the sequence id/name part and the actual sequence information and stored in memory as arrays of ids/names and sequences. If the alignment file format allows for additional information, such as an ID of the entire alignment or consensus structure information, this data is retrieved as well and made available. The options parameter allows to specify the set of alignment file formats that should be used to retrieve the data. If 0 is passed as option, the list of alignment file formats defaults to VRNA_FILE_FORMAT_MSA_DEFAULT.

Currently, the list of parsable multiple sequence alignment file formats consists of:

- · ClustalW format
- · Stockholm 1.0 format
- FASTA (Pearson) format
- · MAF format

Note

After successfully reading an alignment, this function performs a validation of the data that includes uniqueness of the sequence identifiers, and equal sequence lengths. This check can be deactivated by passing VRNA_FILE_FORMAT_MSA_NOCHECK in the options parameter.

It is the users responsibility to free any memory occupied by the output arguments <code>names</code>, <code>aln</code>, <code>id</code>, and <code>structure</code> after calling this function. The function automatically sets the latter two arguments to <code>NULL</code> in case no corresponding data could be retrieved from the input alignment.

See also

vrna_file_msa_read_record(), VRNA_FILE_FORMAT_MSA_CLUSTAL, VRNA_FILE_FORMAT_MSA_STOCKHOLM, VRNA_FILE_FORMAT_MSA_FASTA, VRNA_FILE_FORMAT_MSA_MAF, VRNA_FILE_FORMAT_MSA_DEFAULT, VRNA_FILE_FORMAT_MSA_NOCHECK

Parameters

filename	The name of input file that contains the alignment
names	An address to the pointer where sequence identifiers should be written to
aln	An address to the pointer where aligned sequences should be written to
id	An address to the pointer where the alignment ID should be written to (Maybe NULL)
structure	An address to the pointer where consensus structure information should be written to (Maybe NULL)
options	Options to manipulate the behavior of this function

Returns

The number of sequences in the alignment, or -1 if no alignment record could be found

SWIG Wrapper Notes In the target scripting language, only the first and last argument, filename and options, are passed to the corresponding function. The other arguments, which serve as output in the C-library, are available as additional return values. Hence, a function call in python may look like this:

```
num_seq, names, aln, id, structure = RNA.file_msa_read("msa.stk", RNA.FILE_FORMAT_MSA_STOCKHOLM)
```

After successfully reading the first record, the variable <code>num_seq</code> contains the number of sequences in the alignment (the actual return value of the C-function), while the variables <code>names</code>, <code>aln</code>, <code>id</code>, and <code>structure</code> are lists of the sequence names and aligned sequences, as well as strings holding the alignment ID and the structure as stated in the <code>SS_cons</code> line, respectively. Note, the last two return values may be empty strings in case the alignment does not provide the required data.

This function exists as an overloaded version where the options parameter may be omitted! In that case, the options parameter defaults to VRNA_FILE_FORMAT_MSA_STOCKHOLM.

16.68.3.2 vrna_file_msa_read_record()

#include <ViennaRNA/io/file_formats_msa.h>

Read a multiple sequence alignment from file handle.

Similar to vrna_file_msa_read(), this function reads a multiple sequence alignment from an input file handle. Since using a file handle, this function is not limited to the first alignment record, but allows for looping over all alignments within the input.

The read alignment is split into the sequence id/name part and the actual sequence information and stored in memory as arrays of ids/names and sequences. If the alignment file format allows for additional information, such as an ID of the entire alignment or consensus structure information, this data is retrieved as well and made available. The options parameter allows to specify the alignment file format used to retrieve the data. A single format must be specified here, see vrna_file_msa_detect_format()) for helping to determine the correct MSA file format.

Currently, the list of parsable multiple sequence alignment file formats consists of:

- · ClustalW format
- · Stockholm 1.0 format
- · FASTA (Pearson) format
- · MAF format

Note

After successfully reading an alignment, this function performs a validation of the data that includes uniqueness of the sequence identifiers, and equal sequence lengths. This check can be deactivated by passing VRNA FILE FORMAT MSA NOCHECK in the options parameter.

It is the users responsibility to free any memory occupied by the output arguments names, aln, id, and structure after calling this function. The function automatically sets the latter two arguments to NULL in case no corresponding data could be retrieved from the input alignment.

See also

vrna_file_msa_read(), vrna_file_msa_detect_format(), VRNA_FILE_FORMAT_MSA_CLUSTAL, VRNA_FILE_FORMAT_MSA_VRNA_FILE_FORMAT_MSA_DEFAULT, VRNA_FILE_FORMAT_MSA_NOCHECK

Parameters

fp	The file pointer the data will be retrieved from	
names	An address to the pointer where sequence identifiers should be written to	
aln	An address to the pointer where aligned sequences should be written to	
id	An address to the pointer where the alignment ID should be written to (Maybe NULL)	
structure	An address to the pointer where consensus structure information should be written to (Maybe NULL)	
options	Options to manipulate the behavior of this function	

Returns

The number of sequences in the alignment, or -1 if no alignment record could be found

SWIG Wrapper Notes In the target scripting language, only the first and last argument, fp and options, are passed to the corresponding function. The other arguments, which serve as output in the C-library, are available as additional return values. Hence, a function call in python may look like this:

```
f = open('msa.stk', 'r')
num_seq, names, aln, id, structure = RNA.file_msa_read_record(f, RNA.FILE_FORMAT_MSA_STOCKHOLM)
f.close()
```

After successfully reading the first record, the variable num_seq contains the number of sequences in the alignment (the actual return value of the C-function), while the variables names, aln, id, and structure are lists of the sequence names and aligned sequences, as well as strings holding the alignment ID and the structure as stated in the SS_cons line, respectively. Note, the last two return values may be empty strings in case the alignment does not provide the required data.

This function exists as an overloaded version where the options parameter may be omitted! In that case, the options parameter defaults to VRNA_FILE_FORMAT_MSA_STOCKHOLM.

16.68.3.3 vrna_file_msa_detect_format()

Detect the format of a multiple sequence alignment file.

This function attempts to determine the format of a file that supposedly contains a multiple sequence alignment (M← SA). This is useful in cases where a MSA file contains more than a single record and therefore vrna_file_msa_read() can not be applied, since it only retrieves the first. Here, one can try to guess the correct file format using this function and then loop over the file, record by record using one of the low-level record retrieval functions for the corresponding MSA file format.

Note

This function parses the entire first record within the specified file. As a result, it returns VRNA_FILE_FORMAT_MSA_UNKNOW not only if it can't detect the file's format, but also in cases where the file doesn't contain sequences!

See also

```
vrna file msa read(), vrna file stockholm read record(), vrna file clustal read record(), vrna file fasta read record()
```

Parameters

filename	The name of input file that contains the alignment
options	Options to manipulate the behavior of this function

Returns

The MSA file format, or VRNA_FILE_FORMAT_MSA_UNKNOWN

SWIG Wrapper Notes This function exists as an overloaded version where the options parameter may be omitted! In that case, the options parameter defaults to VRNA_FILE_FORMAT_MSA_DEFAULT.

16.68.3.4 vrna_file_msa_write()

Write multiple sequence alignment file.

Note

Currently, we only support Stockholm 1.0 format output

See also

VRNA_FILE_FORMAT_MSA_STOCKHOLM, VRNA_FILE_FORMAT_MSA_APPEND, VRNA_FILE_FORMAT_MSA_MIS

Parameters

filename	The output filename
names	The array of sequence names / identifies
aln	The array of aligned sequences
id	An optional ID for the alignment
structure	An optional consensus structure
source	A string describing the source of the alignment
options	Options to manipulate the behavior of this function

Returns

Non-null upon successfully writing the alignment to file

SWIG Wrapper Notes In the target scripting language, this function exists as a set of overloaded versions, where the last four parameters may be omitted. If the options parameter is missing the options default to (VRNA_FILE_FORMAT_MSA_STOCKHOLM | VRNA_FILE_FORMAT_MSA_APPEND).

16.69 Command Files

Functions to parse and interpret the content of Command Files.

16.69.1 Detailed Description

Functions to parse and interpret the content of Command Files.

Collaboration diagram for Command Files:

Files

· file commands.h

Parse and apply different commands that alter the behavior of secondary structure prediction and evaluation.

Macros

• #define VRNA CMD PARSE HC 1U

Command parse/apply flag indicating hard constraints.

#define VRNA_CMD_PARSE_SC 2U

Command parse/apply flag indicating soft constraints.

• #define VRNA CMD PARSE UD 4U

Command parse/apply flag indicating unstructured domains.

• #define VRNA_CMD_PARSE_SD 8U

Command parse/apply flag indicating structured domains.

#define VRNA_CMD_PARSE_DEFAULTS

Command parse/apply flag indicating default set of commands.

Typedefs

typedef struct vrna_command_s * vrna_cmd_t

A data structure that contains commands.

Functions

• vrna_cmd_t vrna_file_commands_read (const char *filename, unsigned int options)

Extract a list of commands from a command file.

• int vrna_file_commands_apply (vrna_fold_compound_t *vc, const char *filename, unsigned int options)

Apply a list of commands from a command file.

int vrna_commands_apply (vrna_fold_compound_t *vc, vrna_cmd_t commands, unsigned int options)

Apply a list of commands to a vrna_fold_compound_t.

void vrna_commands_free (vrna_cmd_t commands)

Free memory occupied by a list of commands.

16.69 Command Files 533

16.69.2 Macro Definition Documentation

```
16.69.2.1 VRNA_CMD_PARSE_HC
#define VRNA_CMD_PARSE_HC 1U
#include <ViennaRNA/commands.h>
Command parse/apply flag indicating hard constraints.
See also
     vrna_cmd_t, vrna_file_commands_read(), vrna_file_commands_apply(), vrna_commands_apply()
16.69.2.2 VRNA_CMD_PARSE_SC
#define VRNA_CMD_PARSE_SC 2U
#include <ViennaRNA/commands.h>
Command parse/apply flag indicating soft constraints.
See also
    vrna_cmd_t, vrna_file_commands_read(), vrna_file_commands_apply(), vrna_commands_apply()
16.69.2.3 VRNA_CMD_PARSE_UD
#define VRNA_CMD_PARSE_UD 4U
#include <ViennaRNA/commands.h>
Command parse/apply flag indicating unstructured domains.
See also
     vrna_cmd_t, vrna_file_commands_read(), vrna_file_commands_apply(), vrna_commands_apply()
```

```
16.69.2.4 VRNA_CMD_PARSE_SD
```

```
#define VRNA_CMD_PARSE_SD 8U
#include <ViennaRNA/commands.h>
```

Command parse/apply flag indicating structured domains.

See also

```
vrna_cmd_t, vrna_file_commands_read(), vrna_file_commands_apply(), vrna_commands_apply()
```

```
16.69.2.5 VRNA_CMD_PARSE_DEFAULTS
```

```
#define VRNA_CMD_PARSE_DEFAULTS
#include <ViennaRNA/commands.h>
```

Value:

Command parse/apply flag indicating default set of commands.

See also

```
vrna_cmd_t, vrna_file_commands_read(), vrna_file_commands_apply(), vrna_commands_apply()
```

16.69.3 Function Documentation

16.69.3.1 vrna_file_commands_read()

Extract a list of commands from a command file.

Read a list of commands specified in the input file and return them as list of abstract commands

See also

```
vrna_commands_apply(), vrna_file_commands_apply(), vrna_commands_free()
```

16.69 Command Files 535

Parameters

filename	The filename
options	Options to limit the type of commands read from the file

Returns

A list of abstract commands

16.69.3.2 vrna_file_commands_apply()

Apply a list of commands from a command file.

This function is a shortcut to directly parse a commands file and apply all successfully parsed commands to a vrna_fold_compound_t data structure. It is the same as:

Parameters

VC	The vrna_fold_compound_t the command list will be applied to
filename	The filename
options	Options to limit the type of commands read from the file

Returns

The number of commands successfully applied

SWIG Wrapper Notes This function is attached as method file_commands_apply() to objects of type fold_← compound

16.69.3.3 vrna_commands_apply()

Apply a list of commands to a vrna_fold_compound_t.

Parameters

VC	The vrna_fold_compound_t the command list will be applied to
commands	The commands to apply
options	Options to limit the type of commands read from the file

Returns

The number of commands successfully applied

16.69.3.4 vrna_commands_free()

#include <ViennaRNA/commands.h>

Free memory occupied by a list of commands.

Release memory occupied by a list of commands

Parameters

commands	A pointer to a list of commands

16.70 Plotting 537

16.70 Plotting

Functions for Creating Secondary Structure Plots, Dot-Plots, and More.

16.70.1 Detailed Description

Functions for Creating Secondary Structure Plots, Dot-Plots, and More.

Collaboration diagram for Plotting:

Modules

· Layouts and Coordinates

Functions to compute coordinate layouts for secondary structure plots.

Annotation

Functions to generate annotations for Secondary Structure Plots, Dot-Plots, and Others.

Alignment Plots

Functions to generate Alignment plots with annotated consensus structure.

· Deprecated Interface for Plotting Utilities

Files

· file alignments.h

Various functions for plotting Sequence / Structure Alignments.

· file layouts.h

Secondary structure plot layout algorithms.

· file naview.h

Implementation of the Naview RNA secondary structure layout algorithm [5].

· file probabilities.h

Various functions for plotting RNA secondary structures, dot-plots and other visualizations.

· file structures.h

Various functions for plotting RNA secondary structures.

• file utils.h

Various utilities to assist in plotting secondary structures and consensus structures.

· file RNApuzzler.h

Implementation of the RNApuzzler RNA secondary structure layout algorithm [23].

• file RNAturtle.h

Implementation of the RNAturtle RNA secondary structure layout algorithm [23].

Data Structures

struct vrna_dotplot_auxdata_t

Functions

• int PS_dot_plot_list (char *seq, char *filename, plist *pl, plist *mf, char *comment)

Produce a postscript dot-plot from two pair lists.

int PS_dot_plot (char *string, char *file)

Produce postscript dot-plot.

• int vrna_file_PS_rnaplot (const char *seq, const char *structure, const char *file, vrna_md_t *md_p)

Produce a secondary structure graph in PostScript and write it to 'filename'.

• int vrna_file_PS_rnaplot_a (const char *seq, const char *structure, const char *file, const char *pre, con

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename'.

• int gmlRNA (char *string, char *structure, char *ssfile, char option)

Produce a secondary structure graph in Graph Meta Language (gml) and write it to a file.

int ssv rna plot (char *string, char *structure, char *ssfile)

Produce a secondary structure graph in SStructView format.

• int svg_rna_plot (char *string, char *structure, char *ssfile)

Produce a secondary structure plot in SVG format and write it to a file.

int xrna_plot (char *string, char *structure, char *ssfile)

Produce a secondary structure plot for further editing in XRNA.

int PS rna plot (char *string, char *structure, char *file)

Produce a secondary structure graph in PostScript and write it to 'filename'.

int PS rna plot a (char *string, char *structure, char *file, char *pre, char *post)

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename'.

int PS_rna_plot_a_gquad (char *string, char *structure, char *ssfile, char *pre, char *post)

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename' (detect and draw g-quadruplexes)

16.70.2 Data Structure Documentation

```
16.70.2.1 struct vrna_dotplot_auxdata_t
```

Collaboration diagram for vrna_dotplot_auxdata_t:

16.70.3 Function Documentation

16.70.3.1 PS_dot_plot_list()

#include <ViennaRNA/plotting/probabilities.h>

Produce a postscript dot-plot from two pair lists.

This function reads two plist structures (e.g. base pair probabilities and a secondary structure) as produced by assign_plist_from_pr() and assign_plist_from_db() and produces a postscript "dot plot" that is written to 'filename'. Using base pair probabilities in the first and mfe structure in the second plist, the resulting "dot plot" represents each base pairing probability by a square of corresponding area in a upper triangle matrix. The lower part of the matrix contains the minimum free energy structure.

16.70 Plotting 539

See also

```
assign_plist_from_pr(), assign_plist_from_db()
```

Parameters

seq	The RNA sequence
filename	A filename for the postscript output
pl	The base pair probability pairlist
mf	The mfe secondary structure pairlist
comment	A comment

Returns

1 if postscript was successfully written, 0 otherwise

16.70.3.2 PS_dot_plot()

#include <ViennaRNA/plotting/probabilities.h>

Produce postscript dot-plot.

Wrapper to PS_dot_plot_list

Reads base pair probabilities produced by pf_fold() from the global array pr and the pair list base_pair produced by fold() and produces a postscript "dot plot" that is written to 'filename'. The "dot plot" represents each base pairing probability by a square of corresponding area in a upper triangle matrix. The lower part of the matrix contains the minimum free energy

Note

DO NOT USE THIS FUNCTION ANYMORE SINCE IT IS NOT THREADSAFE

Deprecated This function is deprecated and will be removed soon! Use PS_dot_plot_list() instead!

16.70.3.3 vrna_file_PS_rnaplot()

#include <ViennaRNA/plotting/structures.h>

Produce a secondary structure graph in PostScript and write it to 'filename'.

Note that this function has changed from previous versions and now expects the structure to be plotted in dot-bracket notation as an argument. It does not make use of the global base_pair array anymore.

Parameters

seq	The RNA sequence	
structure	The secondary structure in dot-bracket notation	
file	The filename of the postscript output	
md_p	Model parameters used to generate a commandline option string in the output (Maybe NULL)	

Returns

1 on success, 0 otherwise

16.70.3.4 vrna_file_PS_rnaplot_a()

#include <ViennaRNA/plotting/structures.h>

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename'.

Same as vrna_file_PS_rnaplot() but adds extra PostScript macros for various annotations (see generated PS code). The 'pre' and 'post' variables contain PostScript code that is verbatim copied in the resulting PS file just before and after the structure plot. If both arguments ('pre' and 'post') are NULL, no additional macros will be printed into the PostScript.

Parameters

seq	The RNA sequence	
structure	The secondary structure in dot-bracket notation	
file	The filename of the postscript output	
pre	PostScript code to appear before the secondary structure plot	
post	PostScript code to appear after the secondary structure plot Model parameters used to generate a commandline option string in the output (Maybe NULL)	
md_p		

Returns

1 on success, 0 otherwise

16.70.3.5 gmIRNA()

16.70 Plotting 541

```
char * structure,
char * ssfile,
char option )
```

#include <ViennaRNA/plotting/structures.h>

Produce a secondary structure graph in Graph Meta Language (gml) and write it to a file.

If 'option' is an uppercase letter the RNA sequence is used to label nodes, if 'option' equals X' or X' the resulting file will coordinates for an initial layout of the graph.

Parameters

string	The RNA sequence
structure	The secondary structure in dot-bracket notation
ssfile	The filename of the gml output
option	The option flag

Returns

1 on success, 0 otherwise

16.70.3.6 ssv_rna_plot()

#include <ViennaRNA/plotting/structures.h>

Produce a secondary structure graph in SStructView format.

Write coord file for SStructView

Parameters

string	The RNA sequence
structure	The secondary structure in dot-bracket notation
ssfile	The filename of the ssv output

Returns

1 on success, 0 otherwise

16.70.3.7 svg_rna_plot()

#include <ViennaRNA/plotting/structures.h>

Produce a secondary structure plot in SVG format and write it to a file.

Parameters

string	The RNA sequence
structure	The secondary structure in dot-bracket notation
ssfile	The filename of the svg output

Returns

1 on success, 0 otherwise

16.70.3.8 xrna_plot()

#include <ViennaRNA/plotting/structures.h>

Produce a secondary structure plot for further editing in XRNA.

Parameters

string	The RNA sequence
structure	The secondary structure in dot-bracket notation
ssfile	The filename of the xrna output

Returns

1 on success, 0 otherwise

16.70.3.9 PS_rna_plot()

16.70 Plotting 543

Produce a secondary structure graph in PostScript and write it to 'filename'.

Deprecated Use vrna_file_PS_rnaplot() instead!

```
16.70.3.10 PS_rna_plot_a()
```

#include <ViennaRNA/plotting/structures.h>

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename'.

Deprecated Use vrna_file_PS_rnaplot_a() instead!

```
16.70.3.11 PS_rna_plot_a_gquad()
```

#include <ViennaRNA/plotting/structures.h>

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename' (detect and draw g-quadruplexes)

Deprecated Use vrna_file_PS_rnaplot_a() instead!

16.71 Layouts and Coordinates

Functions to compute coordinate layouts for secondary structure plots.

16.71.1 Detailed Description

Functions to compute coordinate layouts for secondary structure plots.

Collaboration diagram for Layouts and Coordinates:

Data Structures

- struct vrna_plot_layout_s
- · struct vrna_plot_options_puzzler_t

Options data structure for RNApuzzler algorithm implementation. More...

Macros

#define VRNA PLOT TYPE SIMPLE 0

Definition of Plot type simple

#define VRNA PLOT TYPE NAVIEW 1

Definition of Plot type Naview

#define VRNA_PLOT_TYPE_CIRCULAR 2

Definition of Plot type Circular

• #define VRNA PLOT TYPE TURTLE 3

Definition of Plot type Turtle [23].

#define VRNA_PLOT_TYPE_PUZZLER 4

Definition of Plot type RNApuzzler [23].

Typedefs

typedef struct vrna_plot_layout_s vrna_plot_layout_t

RNA secondary structure figure layout.

Functions

vrna_plot_layout_t * vrna_plot_layout (const char *structure, unsigned int plot_type)

Create a layout (coordinates, etc.) for a secondary structure plot.

vrna_plot_layout_t * vrna_plot_layout_simple (const char *structure)

Create a layout (coordinates, etc.) for a simple secondary structure plot.

vrna_plot_layout_t * vrna_plot_layout_naview (const char *structure)

Create a layout (coordinates, etc.) for a secondary structure plot using the Naview Algorithm [5].

vrna_plot_layout_t * vrna_plot_layout_circular (const char *structure)

Create a layout (coordinates, etc.) for a circular secondary structure plot.

vrna_plot_layout_t * vrna_plot_layout_turtle (const char *structure)

Create a layout (coordinates, etc.) for a secondary structure plot using the Turtle Algorithm [23].

vrna_plot_layout_t * vrna_plot_layout_puzzler (const char *structure, vrna_plot_options_puzzler_t *options)

Create a layout (coordinates, etc.) for a secondary structure plot using the RNApuzzler Algorithm [23].

void vrna_plot_layout_free (vrna_plot_layout_t *layout)

Free memory occupied by a figure layout data structure.

int vrna plot coords (const char *structure, float **x, float **y, int plot type)

Compute nucleotide coordinates for secondary structure plot.

int vrna_plot_coords_pt (const short *pt, float **x, float **y, int plot_type)

Compute nucleotide coordinates for secondary structure plot.

int vrna_plot_coords_simple (const char *structure, float **x, float **y)

Compute nucleotide coordinates for secondary structure plot the Simple way

• int vrna_plot_coords_simple_pt (const short *pt, float **x, float **y)

Compute nucleotide coordinates for secondary structure plot the Simple way

• int vrna_plot_coords_circular (const char *structure, float **x, float **y)

Compute coordinates of nucleotides mapped in equal distancies onto a unit circle.

int vrna_plot_coords_circular_pt (const short *pt, float **x, float **y)

Compute nucleotide coordinates for a Circular Plot

int vrna plot coords naview (const char *structure, float **x, float **y)

Compute nucleotide coordinates for secondary structure plot using the Naview algorithm [5].

int vrna_plot_coords_naview_pt (const short *pt, float **x, float **x)

Compute nucleotide coordinates for secondary structure plot using the Naview algorithm [5].

 int vrna_plot_coords_puzzler (const char *structure, float **x, float **y, double **arc_coords, vrna_plot_options_puzzler_t *options)

Compute nucleotide coordinates for secondary structure plot using the RNApuzzler algorithm [23].

 int vrna_plot_coords_puzzler_pt (short const *const pair_table, float **x, float **y, double **arc_coords, vrna_plot_options_puzzler_t *puzzler)

Compute nucleotide coordinates for secondary structure plot using the RNApuzzler algorithm [23].

vrna_plot_options_puzzler_t * vrna_plot_options_puzzler (void)

Create an RNApuzzler options data structure.

void vrna_plot_options_puzzler_free (vrna_plot_options_puzzler_t *options)

Free memory occupied by an RNApuzzler options data structure.

int vrna_plot_coords_turtle (const char *structure, float **x, float **y, double **arc_coords)

Compute nucleotide coordinates for secondary structure plot using the RNAturtle algorithm [23].

• int vrna_plot_coords_turtle_pt (short const *const pair_table, float **x, float **y, double **arc_coords)

Compute nucleotide coordinates for secondary structure plot using the RNAturtle algorithm [23].

16.71.2 Data Structure Documentation

16.71.2.1 struct vrna_plot_layout_s

16.71.2.2 struct vrna_plot_options_puzzler_t

Options data structure for RNApuzzler algorithm implementation.

16.71.3 Macro Definition Documentation

```
16.71.3.1 VRNA_PLOT_TYPE_SIMPLE
```

```
#define VRNA_PLOT_TYPE_SIMPLE 0
#include <ViennaRNA/plotting/layouts.h>
```

Definition of Plot type simple

This is the plot type definition for several RNA structure plotting functions telling them to use **Simple** plotting algorithm

See also

```
\label{lem:condition} rna\_plot\_type, \ vrna\_file\_PS\_rnaplot\_a(), \ vrna\_file\_PS\_rnaplot(), \ svg\_rna\_plot(), \ gmlRNA(), \ ssv\_rna\_plot(), \ xrna\_plot()
```

16.71.3.2 VRNA_PLOT_TYPE_NAVIEW

```
#define VRNA_PLOT_TYPE_NAVIEW 1
#include <ViennaRNA/plotting/layouts.h>
```

Definition of Plot type Naview

This is the plot type definition for several RNA structure plotting functions telling them to use **Naview** plotting algorithm [5].

See also

```
rna_plot_type, vrna_file_PS_rnaplot_a(), vrna_file_PS_rnaplot(), svg_rna_plot(), gmlRNA(), ssv_rna_plot(),
xrna_plot()
```

16.71.3.3 VRNA_PLOT_TYPE_CIRCULAR

```
#define VRNA_PLOT_TYPE_CIRCULAR 2
#include <ViennaRNA/plotting/layouts.h>
```

Definition of Plot type Circular

This is the plot type definition for several RNA structure plotting functions telling them to produce a Circular plot

See also

```
rna_plot_type, vrna_file_PS_rnaplot_a(), vrna_file_PS_rnaplot(), svg_rna_plot(), gmlRNA(), ssv_rna_plot(),
xrna_plot()
```

```
16.71.3.4 VRNA_PLOT_TYPE_TURTLE
#define VRNA_PLOT_TYPE_TURTLE 3
#include <ViennaRNA/plotting/layouts.h>
Definition of Plot type Turtle [23].
16.71.3.5 VRNA_PLOT_TYPE_PUZZLER
#define VRNA_PLOT_TYPE_PUZZLER 4
#include <ViennaRNA/plotting/layouts.h>
Definition of Plot type RNApuzzler [23].
16.71.4 Typedef Documentation
16.71.4.1 vrna_plot_layout_t
typedef struct vrna_plot_layout_s vrna_plot_layout_t
#include <ViennaRNA/plotting/layouts.h>
RNA secondary structure figure layout.
See also
    vrna_plot_layout(), vrna_plot_layout_simple(), vrna_plot_layout_circular(), vrna_plot_layout_naview(),
    vrna_plot_layout_turtle(), vrna_plot_layout_puzzler()
```

16.71.5 Function Documentation

16.71.5.1 vrna_plot_layout()

Create a layout (coordinates, etc.) for a secondary structure plot.

This function can be used to create a secondary structure nucleotide layout that is then further processed by an actual plotting function. The layout algorithm can be specified using the plot_type parameter, and the following algorithms are currently supported:

- VRNA_PLOT_TYPE_SIMPLE
- VRNA_PLOT_TYPE_NAVIEW
- VRNA_PLOT_TYPE_CIRCULAR
- VRNA_PLOT_TYPE_TURTLE
- VRNA_PLOT_TYPE_PUZZLER

Passing an unsupported selection leads to the default algorithm VRNA_PLOT_TYPE_NAVIEW

Note

If only X-Y coordinates of the corresponding structure layout are required, consider using vrna_plot_coords() instead!

See also

```
vrna_plot_layout_free(), vrna_plot_layout_simple(), vrna_plot_layout_naview(), vrna_plot_layout_circular(), vrna_plot_layout_turtle(), vrna_plot_layout_puzzler(), vrna_plot_coords(), vrna_file_PS_rnaplot_layout()
```

Parameters

structure	The secondary structure in dot-bracket notation
plot_type	The layout algorithm to be used

Returns

The layout data structure for the provided secondary structure

16.71.5.2 vrna_plot_layout_simple()

Create a layout (coordinates, etc.) for a simple secondary structure plot.

This function basically is a wrapper to vrna_plot_layout() that passes the plot_type VRNA_PLOT_TYPE_SIMPLE.

Note

If only X-Y coordinates of the corresponding structure layout are required, consider using vrna_plot_coords_simple() instead!

See also

vrna_plot_layout_free(), vrna_plot_layout(), vrna_plot_layout_naview(), vrna_plot_layout_circular(), vrna_plot_layout_turtle(), vrna_plot_layout_puzzler(), vrna_plot_coords_simple(), vrna_file_PS_rnaplot_layout()

Parameters

structure	The secondary structure in dot-bracket notation
-----------	---

Returns

The layout data structure for the provided secondary structure

16.71.5.3 vrna_plot_layout_naview()

#include <ViennaRNA/plotting/layouts.h>

Create a layout (coordinates, etc.) for a secondary structure plot using the Naview Algorithm [5].

This function basically is a wrapper to vrna_plot_layout() that passes the plot_type VRNA_PLOT_TYPE_NAVIEW.

Note

If only X-Y coordinates of the corresponding structure layout are required, consider using vrna_plot_coords_naview() instead!

See also

vrna_plot_layout_free(), vrna_plot_layout(), vrna_plot_layout_simple(), vrna_plot_layout_circular(), vrna_plot_layout_turtle(), vrna_plot_layout_puzzler(), vrna_plot_coords_naview(), vrna_file_PS_rnaplot_layout()

Parameters

structure	The secondary structure in dot-bracket notation
-----------	---

Returns

The layout data structure for the provided secondary structure

16.71.5.4 vrna_plot_layout_circular()

Create a layout (coordinates, etc.) for a circular secondary structure plot.

This function basically is a wrapper to vrna plot layout() that passes the plot_type VRNA PLOT TYPE CIRCULAR.

Note

If only X-Y coordinates of the corresponding structure layout are required, consider using vrna_plot_coords_circular() instead!

See also

vrna_plot_layout_free(), vrna_plot_layout(), vrna_plot_layout_naview(), vrna_plot_layout_simple(), vrna_plot_layout_turtle(), vrna_plot_layout_puzzler(), vrna_plot_coords_circular(), vrna_file_PS_rnaplot_layout()

Parameters

structure	The secondary structure in dot-bracket notation
-----------	---

Returns

The layout data structure for the provided secondary structure

16.71.5.5 vrna_plot_layout_turtle()

Create a layout (coordinates, etc.) for a secondary structure plot using the Turtle Algorithm [23].

This function basically is a wrapper to vrna_plot_layout() that passes the plot_type VRNA_PLOT_TYPE_TURTLE.

Note

If only X-Y coordinates of the corresponding structure layout are required, consider using vrna_plot_coords_turtle() instead!

See also

vrna_plot_layout_free(), vrna_plot_layout(), vrna_plot_layout_simple(), vrna_plot_layout_circular(), vrna_plot_layout_naview(), vrna_plot_layout_puzzler(), vrna_plot_coords_turtle(), vrna_file_PS_rnaplot_layout()

Parameters

structure	The secondary structure in dot-bracket notation
-----------	---

Returns

The layout data structure for the provided secondary structure

16.71.5.6 vrna_plot_layout_puzzler()

Create a layout (coordinates, etc.) for a secondary structure plot using the RNApuzzler Algorithm [23].

This function basically is a wrapper to vrna_plot_layout() that passes the plot_type VRNA_PLOT_TYPE_PUZZLER.

Note

If only X-Y coordinates of the corresponding structure layout are required, consider using vrna_plot_coords_puzzler() instead!

See also

 $vrna_plot_layout_free(), vrna_plot_layout(), vrna_plot_layout_simple(), vrna_plot_layout_circular(), vrna_plot_layout_naview(), vrna_plot_layout_turtle(), vrna_plot_coords_puzzler(), vrna_file_PS_rnaplot_layout()$

Parameters

structure	The secondary structure in dot-bracket notation
-----------	---

Returns

The layout data structure for the provided secondary structure

16.71.5.7 vrna_plot_layout_free()

Free memory occupied by a figure layout data structure.

See also

vrna_plot_layout_t, vrna_plot_layout(), vrna_plot_layout_simple(), vrna_plot_layout_circular(), vrna_plot_layout_naview(), vrna_plot_layout_turtle(), vrna_plot_layout_puzzler(), vrna_file_PS_rnaplot_layout()

Parameters

```
layout The layout data structure to free
```

16.71.5.8 vrna_plot_coords()

#include <ViennaRNA/plotting/layouts.h>

Compute nucleotide coordinates for secondary structure plot.

This function takes a secondary structure and computes X-Y coordinates for each nucleotide that then can be used to create a structure plot. The parameter $plot_type$ is used to select the underlying layout algorithm. Currently, the following selections are provided:

- VRNA PLOT TYPE SIMPLE
- VRNA_PLOT_TYPE_NAVIEW
- VRNA_PLOT_TYPE_CIRCULAR
- VRNA_PLOT_TYPE_TURTLE
- VRNA_PLOT_TYPE_PUZZLER

Passing an unsupported selection leads to the default algorithm VRNA PLOT TYPE NAVIEW

Here is a simple example how to use this function, assuming variable structure contains a valid dot-bracket string:

```
float *x, *y;
if (vrna_plot_coords(structure, &x, &y)) {
  printf("all fine");
} else {
  printf("some failure occured!");
}
free(x);
free(y);
```

Note

On success, this function allocates memory for X and Y coordinates and assigns the pointers at addressess x and y to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords_pt(), vrna_plot_coords_simple(), vrna_plot_coords_naview() vrna_plot_coords_circular(), vrna_plot_coords_turtle(), vrna_plot_coords_puzzler()

Parameters

	structure	The secondary structure in dot-bracket notation
in,out	X	The address of a pointer of X coordinates (pointer will point to memory, or NULL on
		failure)
in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on
		failure)
	plot_type	The layout algorithm to be used

Returns

The length of the structure on success, 0 otherwise

16.71.5.9 vrna_plot_coords_pt()

#include <ViennaRNA/plotting/layouts.h>

Compute nucleotide coordinates for secondary structure plot.

Same as vrna_plot_coords() but takes a pair table with the structure information as input.

Note

On success, this function allocates memory for X and Y coordinates and assigns the pointers at addressess ${\bf x}$ and ${\bf y}$ to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

 $vrna_plot_coords(), vrna_plot_coords_simple_pt(), vrna_plot_coords_naview_pt() \ vrna_plot_coords_circular_pt(), vrna_plot_coords_turtle_pt(), vrna_plot_coords_puzzler_pt()$

Parameters

	pt	The pair table that holds the secondary structure
in,out	X	The address of a pointer of X coordinates (pointer will point to memory, or NULL on
		failure)
in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on
		failure)
	plot_type	The layout algorithm to be used

Returns

The length of the structure on success, 0 otherwise

16.71.5.10 vrna_plot_coords_simple()

#include <ViennaRNA/plotting/layouts.h>

Compute nucleotide coordinates for secondary structure plot the Simple way

This function basically is a wrapper to vrna_plot_coords() that passes the plot_type VRNA_PLOT_TYPE_SIMPLE.

Here is a simple example how to use this function, assuming variable structure contains a valid dot-bracket string:

```
float *x, *y;
if (vrna_plot_coords_simple(structure, &x, &y)) {
  printf("all fine");
} else {
  printf("some failure occured!");
}
free(x);
free(y);
```

Note

On success, this function allocates memory for X and Y coordinates and assigns the pointers at addressess x and y to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords(), vrna_plot_coords_simple_pt(), vrna_plot_coords_circular(), vrna_plot_coords_naview(), vrna_plot_coords_turtle(), vrna_plot_coords_puzzler()

Parameters

	structure	The secondary structure in dot-bracket notation
in,out	X	The address of a pointer of X coordinates (pointer will point to memory, or NULL on
		failure)
in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on
		failure)

Returns

The length of the structure on success, 0 otherwise

16.71.5.11 vrna_plot_coords_simple_pt()

#include <ViennaRNA/plotting/layouts.h>

Compute nucleotide coordinates for secondary structure plot the Simple way

Same as vrna_plot_coords_simple() but takes a pair table with the structure information as input.

Note

On success, this function allocates memory for X and Y coordinates and assigns the pointers at addressess x and y to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords_pt(), vrna_plot_coords_simple(), vrna_plot_coords_circular_pt(), vrna_plot_coords_naview_pt(),
vrna_plot_coords_turtle_pt(), vrna_plot_coords_puzzler_pt()

Parameters

		pt	The pair table that holds the secondary structure
Ī	in,out	х	The address of a pointer of X coordinates (pointer will point to memory, or NULL on failure)
ſ	in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on failure)

Returns

The length of the structure on success, 0 otherwise

16.71.5.12 vrna_plot_coords_circular()

#include <ViennaRNA/plotting/layouts.h>

Compute coordinates of nucleotides mapped in equal distancies onto a unit circle.

This function basically is a wrapper to vrna_plot_coords() that passes the plot_type VRNA_PLOT_TYPE_CIRCULAR.

In order to draw nice arcs using quadratic bezier curves that connect base pairs one may calculate a second tangential point P^t in addition to the actual R^2 coordinates. the simplest way to do so may be to compute a radius scaling factor rs in the interval [0,1] that weights the proportion of base pair span to the actual length of the sequence. This scaling factor can then be used to calculate the coordinates for P^t , i.e.

$$P_r^t[i] = X[i] * rs$$

and

$$P_u^t[i] = Y[i] * rs$$

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Note

On success, this function allocates memory for X and Y coordinates and assigns the pointers at addressess x and y to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords(), vrna_plot_coords_circular_pt(), vrna_plot_coords_simple(), vrna_plot_coords_naview(), vrna_plot_coords_turtle(), vrna_plot_coords_puzzler()

Parameters

	structure	The secondary structure in dot-bracket notation
in,out	X	The address of a pointer of X coordinates (pointer will point to memory, or NULL on failure)
in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on failure)

Returns

The length of the structure on success, 0 otherwise

16.71.5.13 vrna_plot_coords_circular_pt()

#include <ViennaRNA/plotting/layouts.h>

Compute nucleotide coordinates for a Circular Plot

Same as vrna_plot_coords_circular() but takes a pair table with the structure information as input.

Note

On success, this function allocates memory for X and Y coordinates and assigns the pointers at addressess x and y to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords_pt(), vrna_plot_coords_circular(), vrna_plot_coords_simple_pt(), vrna_plot_coords_naview_pt(), vrna_plot_coords_turtle_pt(), vrna_plot_coords_puzzler_pt()

Parameters

		pt	The pair table that holds the secondary structure
	in,out	X	The address of a pointer of X coordinates (pointer will point to memory, or NULL on failure)
E	in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on failure)

Returns

The length of the structure on success, 0 otherwise

16.71.5.14 vrna_plot_coords_naview()

#include <ViennaRNA/plotting/naview.h>

Compute nucleotide coordinates for secondary structure plot using the Naview algorithm [5].

This function basically is a wrapper to vrna_plot_coords() that passes the plot_type VRNA_PLOT_TYPE_NAVIEW.

Here is a simple example how to use this function, assuming variable structure contains a valid dot-bracket string:

```
float *x, *y;
if (vrna_plot_coords_naview(structure, &x, &y)) {
  printf("all fine");
} else {
  printf("some failure occured!");
}
free(x);
free(y);
```

Note

On success, this function allocates memory for X and Y coordinates and assigns the pointers at addressess x and y to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

Parameters

	structure	The secondary structure in dot-bracket notation
in,out	Х	The address of a pointer of X coordinates (pointer will point to memory, or NULL on
		failure)
in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on
		failure)

Returns

The length of the structure on success, 0 otherwise

16.71.5.15 vrna_plot_coords_naview_pt()

#include <ViennaRNA/plotting/naview.h>

Compute nucleotide coordinates for secondary structure plot using the *Naview* algorithm [5].

Same as vrna_plot_coords_naview() but takes a pair table with the structure information as input.

Note

On success, this function allocates memory for X and Y coordinates and assigns the pointers at addressess x and y to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords_pt(), vrna_plot_coords_naview(), vrna_plot_coords_circular_pt(), vrna_plot_coords_simple_pt(), vrna_plot_coords_turtle_pt(), vrna_plot_coords_puzzler_pt()

Parameters

	pt The pair table that holds the secondary structure			
Ī	in,out	х	The address of a pointer of X coordinates (pointer will point to memory, or NULL on failure	
ſ	in, out y The address of a pointer of Y coordinates (pointer will point to memory, or NULL on fail			

Returns

The length of the structure on success, 0 otherwise

16.71.5.16 vrna_plot_coords_puzzler()

#include <ViennaRNA/plotting/RNApuzzler/RNApuzzler.h>

Compute nucleotide coordinates for secondary structure plot using the RNApuzzler algorithm [23].

This function basically is a wrapper to $vrna_plot_coords()$ that passes the $plot_type\ VRNA_PLOT_TYPE_PUZZLER$.

Here is a simple example how to use this function, assuming variable structure contains a valid dot-bracket string and using the default options (options = NULL):

```
float *x, *y;
double *arcs;
if (vrna_plot_coords_puzzler(structure, &x, &y, &arcs, NULL)) {
  printf("all fine");
} else {
  printf("some failure occured!");
}
free(x);
free(y);
free(arcs);
```

Note

On success, this function allocates memory for X, Y and arc coordinates and assigns the pointers at addressess x, y and @arc_coords to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords(), vrna_plot_coords_puzzler_pt(), vrna_plot_coords_circular(), vrna_plot_coords_simple(), vrna_plot_coords_turtle(), vrna_plot_coords_naview(), vrna_plot_options_puzzler()

Parameters

	structure	The secondary structure in dot-bracket notation
in,out	Х	The address of a pointer of X coordinates (pointer will point to memory, or NULL on failure)
in,out	у	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on failure)
in,out	arc_coords	The address of a pointer that will hold arc coordinates (pointer will point to memory, or NULL on failure)
	options	The options for the RNApuzzler algorithm (or NULL)

Returns

The length of the structure on success, 0 otherwise

16.71.5.17 vrna_plot_coords_puzzler_pt()

#include <ViennaRNA/plotting/RNApuzzler/RNApuzzler.h>

Compute nucleotide coordinates for secondary structure plot using the RNApuzzler algorithm [23].

Same as vrna plot coords puzzler() but takes a pair table with the structure information as input.

Note

On success, this function allocates memory for X, Y and arc coordinates and assigns the pointers at addressess x, y and @arc_coords to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords_pt(), vrna_plot_coords_puzzler(), vrna_plot_coords_circular_pt(), vrna_plot_coords_simple_pt(), vrna_plot_coords_turtle_pt(), vrna_plot_coords_naview_pt()

Parameters

	pt	The pair table that holds the secondary structure
in,out	X	The address of a pointer of X coordinates (pointer will point to memory, or NULL on failure)
in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on failure)
in,out	arc_coords	The address of a pointer that will hold arc coordinates (pointer will point to memory, or NULL on failure)
	options	The options for the RNApuzzler algorithm (or NULL)

Returns

The length of the structure on success, 0 otherwise

16.71.5.18 vrna_plot_options_puzzler()

#include <ViennaRNA/plotting/RNApuzzler/RNApuzzler.h>

Create an RNApuzzler options data structure.

See also

vrna_plot_options_puzzler_free(), vrna_plot_coords_puzzler(), vrna_plot_coords_puzzler_pt(), vrna_plot_layout_puzzler()

Returns

An RNApuzzler options data structure with default settings

16.71.5.19 vrna_plot_options_puzzler_free()

#include <ViennaRNA/plotting/RNApuzzler/RNApuzzler.h>

Free memory occupied by an RNApuzzler options data structure.

See also

vrna_plot_options_puzzler(), vrna_plot_coords_puzzler(), vrna_plot_coords_puzzler_pt(), vrna_plot_layout_puzzler()

Parameters

options A pointer to the options data structure to free

16.71.5.20 vrna plot coords turtle()

#include <ViennaRNA/plotting/RNApuzzler/RNAturtle.h>

Compute nucleotide coordinates for secondary structure plot using the RNAturtle algorithm [23].

This function basically is a wrapper to vrna_plot_coords() that passes the plot_type VRNA_PLOT_TYPE_TURTLE.

Here is a simple example how to use this function, assuming variable structure contains a valid dot-bracket string:

```
float *x, *y;
double *arcs;
if (vrna_plot_coords_turtle(structure, &x, &y, &arcs)) {
  printf("all fine");
} else {
  printf("some failure occured!");
}
free(x);
free(y);
free(arcs);
```

Note

On success, this function allocates memory for X, Y and arc coordinates and assigns the pointers at addressess x, y and @arc_coords to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

vrna_plot_coords(), vrna_plot_coords_turtle_pt(), vrna_plot_coords_circular(), vrna_plot_coords_simple(), vrna_plot_coords_naview(), vrna_plot_coords_puzzler()

Parameters

	structure	The secondary structure in dot-bracket notation
in,out	Х	The address of a pointer of X coordinates (pointer will point to memory, or NULL on failure)
in,out	у	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on failure)
in,out	arc_coords	The address of a pointer that will hold arc coordinates (pointer will point to memory, or NULL on failure)

Returns

The length of the structure on success, 0 otherwise

16.71.5.21 vrna_plot_coords_turtle_pt()

#include <ViennaRNA/plotting/RNApuzzler/RNAturtle.h>

Compute nucleotide coordinates for secondary structure plot using the RNAturtle algorithm [23].

Same as vrna_plot_coords_turtle() but takes a pair table with the structure information as input.

Note

On success, this function allocates memory for X, Y and arc coordinates and assigns the pointers at addressess x, y and @arc_coords to the corresponding memory locations. It's the users responsibility to cleanup this memory after usage!

See also

 $vrna_plot_coords_pt(), vrna_plot_coords_turtle(), vrna_plot_coords_circular_pt(), vrna_plot_coords_simple_pt(), vrna_plot_coords_puzzler_pt(), vrna_plot_coords_naview_pt()\\$

Parameters

	pt	The pair table that holds the secondary structure
in,out	Х	The address of a pointer of X coordinates (pointer will point to memory, or NULL on failure)
in,out	У	The address of a pointer of Y coordinates (pointer will point to memory, or NULL on failure)
in,out	arc_coords	The address of a pointer that will hold arc coordinates (pointer will point to memory, or NULL on failure)

Returns

The length of the structure on success, 0 otherwise

16.72 Annotation 563

16.72 Annotation

Functions to generate annotations for Secondary Structure Plots, Dot-Plots, and Others.

16.72.1 Detailed Description

Functions to generate annotations for Secondary Structure Plots, Dot-Plots, and Others.

Collaboration diagram for Annotation:

Functions

- char ** vrna_annotate_covar_db (const char **alignment, const char *structure, vrna_md_t *md_p)

 Produce covariance annotation for an alignment given a secondary structure.
- vrna_cpair_t * vrna_annotate_covar_pairs (const char **alignment, vrna_ep_t *pl, vrna_ep_t *mfel, double threshold, vrna_md_t *md)

Produce covariance annotation for an alignment given a set of base pairs.

16.72.2 Function Documentation

16.72.2.1 vrna annotate covar db()

Produce covariance annotation for an alignment given a secondary structure.

16.72.2.2 vrna_annotate_covar_pairs()

```
vrna_cpair_t* vrna_annotate_covar_pairs (
    const char ** alignment,
    vrna_ep_t * pl,
    vrna_ep_t * mfel,
    double threshold,
    vrna_md_t * md )
```

#include <ViennaRNA/plotting/utils.h>

Produce covariance annotation for an alignment given a set of base pairs.

16.73 Alignment Plots

Functions to generate Alignment plots with annotated consensus structure.

16.73.1 Detailed Description

Functions to generate Alignment plots with annotated consensus structure.

Collaboration diagram for Alignment Plots:

Functions

• int vrna_file_PS_aln (const char *filename, const char **seqs, const char **names, const char *structure, unsigned int columns)

Create an annotated PostScript alignment plot.

• int vrna_file_PS_aln_slice (const char *filename, const char **seqs, const char **names, const char *structure, unsigned int start, unsigned int end, int offset, unsigned int columns)

Create an annotated PostScript alignment plot.

16.73.2 Function Documentation

16.73.2.1 vrna_file_PS_aln()

#include <ViennaRNA/plotting/alignments.h>

Create an annotated PostScript alignment plot.

See also

```
vrna_file_PS_aln_slice()
```

Parameters

filename	The output file name	
seqs	The aligned sequences	
names	The names of the sequences	
structure The consensus structure in dot-bracket notation		
columns	The number of columns before the alignment is wrapped as a new block (a value of 0 indicates no wrapping)	

16.73 Alignment Plots 565

SWIG Wrapper Notes This function is available as overloaded function $file_PS_aln()$ with three additional parameters start, end, and offset before the columns argument. Thus, it resembles the $vrna_file_PS_aln_slice()$ function. The last four arguments may be omitted, indicating the default of start = 0, end start = 0, and start = 0.

16.73.2.2 vrna_file_PS_aln_slice()

#include <ViennaRNA/plotting/alignments.h>

Create an annotated PostScript alignment plot.

Similar to vrna_file_PS_aln() but allows the user to print a particular slice of the alignment by specifying a start and end position. The additional offset parameter allows for adjusting the alignment position ruler value.

See also

```
vrna_file_PS_aln_slice()
```

Parameters

filename	The output file name
seqs	The aligned sequences
names	The names of the sequences
structure	The consensus structure in dot-bracket notation
start	The start of the alignment slice (a value of 0 indicates the first position of the alignment, i.e. no slicing at 5' side)
end	The end of the alignment slice (a value of 0 indicates the last position of the alignment, i.e. no slicing at 3' side)
offset	The alignment coordinate offset for the position ruler.
columns	The number of columns before the alignment is wrapped as a new block (a value of 0 indicates no wrapping)

SWIG Wrapper Notes This function is available as overloaded function file_PS_aln() where the last four parameter may be omitted, indicating start = 0, end = 0, offset = 0, and columns = 60.

16.74 Search Algorithms

Implementations of various search algorithms to detect strings of objects within other strings of objects.

16.74.1 Detailed Description

Implementations of various search algorithms to detect strings of objects within other strings of objects.

Collaboration diagram for Search Algorithms:

Files

· file BoyerMoore.h

Variants of the Boyer-Moore string search algorithm.

Functions

• const unsigned int * vrna_search_BMH_num (const unsigned int *needle, size_t needle_size, const unsigned int *haystack, size_t haystack_size, size_t start, size_t *badchars, unsigned char cyclic)

Search for a string of elements in a larger string of elements using the Boyer-Moore-Horspool algorithm.

const char * vrna_search_BMH (const char *needle, size_t needle_size, const char *haystack, size_←
t haystack size, size t start, size t *badchars, unsigned char cyclic)

Search for an ASCII pattern within a larger ASCII string using the Boyer-Moore-Horspool algorithm.

size_t * vrna_search_BM_BCT_num (const unsigned int *pattern, size_t pattern_size, unsigned int num_←
max)

Retrieve a Boyer-Moore Bad Character Table for a pattern of elements represented by natural numbers.

size_t * vrna_search_BM_BCT (const char *pattern)

Retrieve a Boyer-Moore Bad Character Table for a NULL-terminated pattern of ASCII characters.

16.74.2 Function Documentation

16.74.2.1 vrna_search_BMH_num()

#include <ViennaRNA/search/BoyerMoore.h>

Search for a string of elements in a larger string of elements using the Boyer-Moore-Horspool algorithm.

To speed-up subsequent searches with this function, the Bad Character Table should be precomputed and passed as argument badchars.

See also

```
vrna search BM BCT num(), vrna search BMH()
```

Parameters

needle	The pattern of object representations to search for	
needle_size	The size (length) of the pattern provided in needle	
haystack	The string of objects the search will be performed on	
haystack_size	The size (length) of the haystack string	
start The position within haystack where to start the search		
badchars	A pre-computed Bad Character Table obtained from vrna_search_BM_BCT_num() (If NULL, a Bad Character Table will be generated automatically)	
cyclic Allow for cyclic matches if non-zero, stop search at end of haystack otherwise		

Returns

A pointer to the first occurence of needle within haystack after position start

16.74.2.2 vrna_search_BMH()

#include <ViennaRNA/search/BoyerMoore.h>

Search for an ASCII pattern within a larger ASCII string using the Boyer-Moore-Horspool algorithm.

To speed-up subsequent searches with this function, the Bad Character Table should be precomputed and passed as argument badchars. Furthermore, both, the lengths of needle and the length of haystack should be pre-computed and must be passed along with each call.

See also

```
vrna_search_BM_BCT(), vrna_search_BMH_num()
```

Parameters

needle The NULL-terminated ASCII pattern to search for		
needle_size	The size (length) of the pattern provided in needle	
haystack	The NULL-terminated ASCII string of the search will be performed on	
haystack_size	The size (length) of the haystack string	
start The position within haystack where to start the search		
badchars	A pre-computed Bad Character Table obtained from vrna_search_BM_BCT() (If NULL, a Bad Character Table will be generated automatically)	
cyclic	Allow for cyclic matches if non-zero, stop search at end of haystack otherwise	

Returns

A pointer to the first occurence of needle within haystack after position start

16.74.2.3 vrna_search_BM_BCT_num()

#include <ViennaRNA/search/BoyerMoore.h>

Retrieve a Boyer-Moore Bad Character Table for a pattern of elements represented by natural numbers.

Note

We store the maximum number representation of an element num_max at position 0. So the actual bad character table T starts at T[1] for an element represented by number 0.

See also

```
vrna_search_BMH_num(), vrna_search_BM_BCT()
```

Parameters

pattern	The pattern of element representations used in the subsequent search
pattern_size	The size (length) of the pattern provided in pattern
num_max	The maximum number representation of an element, i.e. the size of the alphabet

Returns

A Bad Character Table for use in our Boyer-Moore search algorithm implementation(s)

16.74.2.4 vrna_search_BM_BCT()

Retrieve a Boyer-Moore Bad Character Table for a NULL-terminated pattern of ASCII characters.

Note

We store the maximum number representation of an element, i.e. 127 at position 0. So the actual bad character table \mathbb{T} starts at $\mathbb{T}[1]$ for an element represented by ASCII code 0.

See also

```
vrna_search_BMH(), vrna_search_BM_BCT_num()
```

Parameters

pattern The NULL-terminated pattern of ASCII char	racters used in the subsequent search
---	---------------------------------------

Returns

A Bad Character Table for use in our Boyer-Moore search algorithm implementation(s)

16.75 Combinatorics Algorithms

Implementations to solve various combinatorial aspects for strings of objects.

16.75.1 Detailed Description

Implementations to solve various combinatorial aspects for strings of objects.

Collaboration diagram for Combinatorics Algorithms:

Files

· file combinatorics.h

Various implementations that deal with combinatorial aspects of objects.

Functions

unsigned int ** vrna_enumerate_necklaces (const unsigned int *type_counts)
 Enumerate all necklaces with fixed content.

• unsigned int vrna_rotational_symmetry_num (const unsigned int *string, size_t string_length)

Determine the order of rotational symmetry for a string of objects represented by natural numbers.

unsigned int vrna_rotational_symmetry_pos_num (const unsigned int *string, size_t string_length, unsigned int **positions)

Determine the order of rotational symmetry for a string of objects represented by natural numbers.

unsigned int vrna_rotational_symmetry (const char *string)

Determine the order of rotational symmetry for a NULL-terminated string of ASCII characters.

• unsigned int vrna_rotational_symmetry_pos (const char *string, unsigned int **positions)

Determine the order of rotational symmetry for a NULL-terminated string of ASCII characters.

unsigned int vrna_rotational_symmetry_db (vrna_fold_compound_t *fc, const char *structure)

Determine the order of rotational symmetry for a dot-bracket structure.

unsigned int vrna_rotational_symmetry_db_pos (vrna_fold_compound_t *fc, const char *structure, unsigned int *positions)

Determine the order of rotational symmetry for a dot-bracket structure.

16.75.2 Function Documentation

16.75.2.1 vrna_enumerate_necklaces()

Enumerate all necklaces with fixed content.

This function implements A fast algorithm to generate necklaces with fixed content as published by Joe Sawada in 2003 [19].

The function receives a list of counts (the elements on the necklace) for each type of object within a necklace. The list starts at index 0 and ends with an entry that has a count of 0. The algorithm then enumerates all non-cyclic permutations of the content, returned as a list of necklaces. This list, again, is zero-terminated, i.e. the last entry of the list is a <code>NULL</code> pointer.

Parameters

S A 0-terminated list of entity counts	type_counts
--	-------------

Returns

A list of all non-cyclic permutations of the entities

SWIG Wrapper Notes This function is available as global function **enumerate_necklaces()** which accepts lists input, an produces list of lists output.

16.75.2.2 vrna_rotational_symmetry_num()

#include <ViennaRNA/combinatorics.h>

Determine the order of rotational symmetry for a string of objects represented by natural numbers.

The algorithm applies a fast search of the provided string within itself, assuming the end of the string wraps around to connect with it's start. For example, a string of the form 011011 has rotational symmetry of order 2

This is a simplified version of vrna_rotational_symmetry_pos_num() that may be useful if one is only interested in the degree of rotational symmetry but not the actual set of rotational symmetric strings.

See also

vrna_rotational_symmetry_pos_num(), vrna_rotationa_symmetry()

Parameters

string	The string of elements encoded as natural numbers
string_length	The length of the string

Returns

The order of rotational symmetry

SWIG Wrapper Notes This function is available as global function rotational_symmetry(). See vrna_rotational_symmetry_pos() for details. Note, that in the target language the length of the list string is always known a-priori, so the parameter string_length must be omitted.

16.75.2.3 vrna_rotational_symmetry_pos_num()

#include <ViennaRNA/combinatorics.h>

Determine the order of rotational symmetry for a string of objects represented by natural numbers.

The algorithm applies a fast search of the provided string within itself, assuming the end of the string wraps around to connect with it's start. For example, a string of the form 011011 has rotational symmetry of order 2

If the argument positions is not NULL, the function stores an array of string start positions for rotational shifts that map the string back onto itself. This array has length of order of rotational symmetry, i.e. the number returned by this function. The first element positions[0] always contains a shift value of 0 representing the trivial rotation.

Note

Do not forget to release the memory occupied by positions after a successful execution of this function.

See also

vrna_rotational_symmetry_num(), vrna_rotational_symmetry(), vrna_rotational_symmetry_pos()

Parameters

string	The string of elements encoded as natural numbers
string_length	The length of the string
positions	A pointer to an (undefined) list of alternative string start positions that lead to an identity mapping (may be NULL)

Returns

The order of rotational symmetry

SWIG Wrapper Notes This function is available as global function rotational_symmetry(). See vrna_rotational_symmetry_pos() for details. Note, that in the target language the length of the list string is always known a-priori, so the parameter string_length must be omitted.

16.75.2.4 vrna_rotational_symmetry()

Determine the order of rotational symmetry for a NULL-terminated string of ASCII characters.

The algorithm applies a fast search of the provided string within itself, assuming the end of the string wraps around to connect with it's start. For example, a string of the form AABAAB has rotational symmetry of order 2

This is a simplified version of vrna_rotational_symmetry_pos() that may be useful if one is only interested in the degree of rotational symmetry but not the actual set of rotational symmetric strings.

See also

vrna_rotational_symmetry_pos(), vrna_rotationa_symmetry_num()

Parameters

string	A NULL-terminated string of characters
--------	--

Returns

The order of rotational symmetry

SWIG Wrapper Notes This function is available as global function **rotational_symmetry()**. See vrna_rotational_symmetry_pos() for details.

16.75.2.5 vrna_rotational_symmetry_pos()

#include <ViennaRNA/combinatorics.h>

Determine the order of rotational symmetry for a NULL-terminated string of ASCII characters.

The algorithm applies a fast search of the provided string within itself, assuming the end of the string wraps around to connect with it's start. For example, a string of the form AABAAB has rotational symmetry of order 2

If the argument positions is not NULL, the function stores an array of string start positions for rotational shifts that map the string back onto itself. This array has length of order of rotational symmetry, i.e. the number returned by this function. The first element positions[0] always contains a shift value of 0 representing the trivial rotation.

Note

Do not forget to release the memory occupied by positions after a successful execution of this function.

See also

vrna_rotational_symmetry(), vrna_rotational_symmetry_num(), vrna_rotational_symmetry_num_pos()

Parameters

string	A NULL-terminated string of characters
positions	A pointer to an (undefined) list of alternative string start positions that lead to an identity mapping
	(may be NULL)

Returns

The order of rotational symmetry

SWIG Wrapper Notes This function is available as overloaded global function rotational_symmetry(). It merges the functionalities of vrna_rotational_symmetry(), vrna_rotational_symmetry_pos(), vrna_rotational_symmetry_num(), and vrna_rotational_symmetry_pos_num(). In contrast to our C-implementation, this function doesn't return the order of rotational symmetry as a single value, but returns a list of cyclic permutation shifts that result in a rotationally symmetric string. The length of the list then determines the order of rotational symmetry.

16.75.2.6 vrna_rotational_symmetry_db()

Determine the order of rotational symmetry for a dot-bracket structure.

Given a (permutation of multiple) RNA strand(s) and a particular secondary structure in dot-bracket notation, compute the degree of rotational symmetry. In case there is only a single linear RNA strand, the structure always has degree 1, as there are no rotational symmetries due to the direction of the nucleic acid sequence and the fixed positions of 5' and 3' ends. However, for circular RNAs, rotational symmetries might arise if the sequence consists of a concatenation of k identical subsequences.

This is a simplified version of vrna_rotational_symmetry_db_pos() that may be useful if one is only interested in the degree of rotational symmetry but not the actual set of rotational symmetric strings.

See also

vrna_rotational_symmetry_db_pos(), vrna_rotational_symmetry(), vrna_rotational_symmetry_num()

Parameters

fc	A fold_compound data structure containing the nucleic acid sequence(s), their order, and model settings
structure	The dot-bracket structure the degree of rotational symmetry is checked for

Returns

The degree of rotational symmetry of the structure (0 in case of any errors)

SWIG Wrapper Notes This function is attached as method rotational_symmetry_db() to objects of type fold
—compound (i.e. vrna_fold_compound_t). See vrna_rotational_symmetry_db_pos() for details.

16.75.2.7 vrna_rotational_symmetry_db_pos()

```
unsigned int vrna_rotational_symmetry_db_pos (
             vrna_fold_compound_t * fc,
             const char * structure,
             unsigned int ** positions )
```

#include <ViennaRNA/combinatorics.h>

Determine the order of rotational symmetry for a dot-bracket structure.

Given a (permutation of multiple) RNA strand(s) and a particular secondary structure in dot-bracket notation, compute the degree of rotational symmetry. In case there is only a single linear RNA strand, the structure always has degree 1, as there are no rotational symmetries due to the direction of the nucleic acid sequence and the fixed positions of 5' and 3' ends. However, for circular RNAs, rotational symmetries might arise if the sequence consists of a concatenation of k identical subsequences.

If the argument positions is not NULL, the function stores an array of string start positions for rotational shifts that map the string back onto itself. This array has length of order of rotational symmetry, i.e. the number returned by this function. The first element positions[0] always contains a shift value of 0 representing the trivial rotation.

Note

Do not forget to release the memory occupied by positions after a successful execution of this function.

See also

vrna_rotational_symmetry_db(), vrna_rotational_symmetry_pos(), vrna_rotational_symmetry_pos_num()

Parameters

fc	A fold_compound data structure containing the nucleic acid sequence(s), their order, and model settings
structure	The dot-bracket structure the degree of rotational symmetry is checked for
positions	A pointer to an (undefined) list of alternative string start positions that lead to an identity mapping (may be NULL)

Returns

The degree of rotational symmetry of the structure (0 in case of any errors)

SWIG Wrapper Notes This function is attached as method rotational_symmetry_db() to objects of type fold← _compound (i.e. vrna_fold_compound_t). Thus, the first argument must be omitted. In contrast to our C-implementation, this function doesn't simply return the order of rotational symmetry of the secondary structure, but returns the list position of cyclic permutation shifts that result in a rotationally symmetric structure. The length of the list then determines the order of rotational symmetry.

16.76 (Abstract) Data Structures

All datastructures and typedefs shared among the ViennaRNA Package can be found here.

16.76.1 Detailed Description

All datastructures and typedefs shared among the ViennaRNA Package can be found here.

Collaboration diagram for (Abstract) Data Structures:

Modules

· The Fold Compound

This module provides interfaces that deal with the most basic data structure used in structure predicting and energy evaluating function of the RNAlib.

The Dynamic Programming Matrices

This module provides interfaces that deal with creation and destruction of dynamic programming matrices used within the RNAlib.

· Hash Tables

Various implementations of hash table functions.

Heaps

Interface for an abstract implementation of a heap data structure.

Buffers

Functions that provide dynamically buffered stream-like data structures.

Files

• file dp_matrices.h

Functions to deal with standard dynamic programming (DP) matrices.

· file basic.h

Various data structures and pre-processor macros.

Data Structures

struct vrna_basepair_s

Base pair data structure used in subopt.c. More...

struct vrna_cpair_s

this datastructure is used as input parameter in functions of PS_dot.c More...

- struct vrna color s
- struct vrna_data_linear_s
- struct vrna_sect_s

Stack of partial structures for backtracking. More...

struct vrna_bp_stack_s

Base pair stack element. More...

struct pu_contrib

contributions to p_u More...

struct interact

interaction data structure for RNAup More...

struct pu_out

Collection of all free_energy of beeing unpaired values for output. More...

· struct constrain

constraints for cofolding More...

struct duplexT

Data structure for RNAduplex. More...

struct node

Data structure for RNAsnoop (fold energy list) More...

struct snoopT

Data structure for RNAsnoop. More...

struct dupVar

Data structure used in RNApkplex. More...

Typedefs

typedef struct vrna_basepair_s vrna_basepair_t

Typename for the base pair repesenting data structure vrna_basepair_s.

typedef struct vrna_elem_prob_s vrna_plist_t

Typename for the base pair list repesenting data structure vrna elem prob s.

typedef struct vrna_bp_stack_s vrna_bp_stack_t

Typename for the base pair stack repesenting data structure vrna_bp_stack_s.

typedef struct vrna_cpair_s vrna_cpair_t

Typename for data structure vrna_cpair_s.

typedef struct vrna_sect_s vrna_sect_t

Typename for stack of partial structures vrna_sect_s.

typedef double FLT_OR_DBL

Typename for floating point number in partition function computations.

• typedef struct vrna_basepair_s PAIR

Old typename of vrna_basepair_s.

typedef struct vrna_elem_prob_s plist

Old typename of vrna_elem_prob_s.

typedef struct vrna_cpair_s cpair

Old typename of vrna_cpair_s.

typedef struct vrna_sect_s sect

Old typename of vrna_sect_s.

typedef struct vrna_bp_stack_s bondT

Old typename of vrna_bp_stack_s.

typedef struct pu_contrib pu_contrib

contributions to p_u

· typedef struct interact interact

interaction data structure for RNAup

• typedef struct pu_out pu_out

Collection of all free_energy of beeing unpaired values for output.

• typedef struct constrain constrain

constraints for cofolding

· typedef struct node folden

Data structure for RNAsnoop (fold energy list)

typedef struct dupVar dupVar

Data structure used in RNApkplex.

Dummy symbol to check whether the library was build using C11/C++11 features.

Functions

• void vrna_C11_features (void)

16.76.2 Data Structure Documentation

longest unpaired region

```
16.76.2.1 struct vrna_basepair_s
Base pair data structure used in subopt.c.
16.76.2.2 struct vrna_cpair_s
this datastructure is used as input parameter in functions of PS_dot.c
16.76.2.3 struct vrna_color_s
16.76.2.4 struct vrna_data_linear_s
Collaboration diagram for vrna_data_linear_s:
16.76.2.5 struct vrna_sect_s
Stack of partial structures for backtracking.
16.76.2.6 struct vrna_bp_stack_s
Base pair stack element.
16.76.2.7 struct pu_contrib
contributions to p_u
Data Fields
    double ** H
          hairpin loops
    double ** I
          interior loops
    double ** M
          multi loops
    double ** E
          exterior loop

    int length

          length of the input sequence

 int w
```

```
16.76.2.8 struct interact
```

interaction data structure for RNAup

Data Fields

```
double * Pi
```

probabilities of interaction

double * Gi

free energies of interaction

· double Gikjl

full free energy for interaction between [k,i] k < i in longer seq and [j,l] j < l in shorter seq

• double Gikjl_wo

Gikjl without contributions for prob_unpaired.

int i

k<i in longer seq

int k

k<i in longer seq

int j

jjin shorter seq

int I

jjin shorter seq

int length

length of longer sequence

16.76.2.9 struct pu_out

Collection of all free_energy of beeing unpaired values for output.

Data Fields

• int len

sequence length

• int u_vals

number of different -u values

· int contribs

[-c "SHIME"]

char ** header

header line

double ** u_values

(the -u values * [-c "SHIME"]) * seq len

16.76.2.10 struct constrain

constraints for cofolding

```
16.76.2.11 struct duplexT
Data structure for RNAduplex.
16.76.2.12 struct node
Data structure for RNAsnoop (fold energy list)
Collaboration diagram for node:
16.76.2.13 struct snoopT
Data structure for RNAsnoop.
16.76.2.14 struct dupVar
Data structure used in RNApkplex.
16.76.3 Typedef Documentation
16.76.3.1 PAIR
typedef struct vrna_basepair_s PAIR
#include <ViennaRNA/datastructures/basic.h>
Old typename of vrna_basepair_s.
Deprecated Use vrna_basepair_t instead!
16.76.3.2 plist
typedef struct vrna_elem_prob_s plist
#include <ViennaRNA/datastructures/basic.h>
Old typename of <a href="mailto:vrna_elem_prob_s">vrna_elem_prob_s</a>.
```

Deprecated Use vrna_ep_t or vrna_elem_prob_s instead!

```
16.76.3.3 cpair
typedef struct vrna_cpair_s cpair
#include <ViennaRNA/datastructures/basic.h>
Old typename of vrna_cpair_s.
Deprecated Use vrna_cpair_t instead!
16.76.3.4 sect
typedef struct vrna_sect_s sect
#include <ViennaRNA/datastructures/basic.h>
Old typename of vrna_sect_s.
Deprecated Use vrna_sect_t instead!
16.76.3.5 bondT
typedef struct vrna_bp_stack_s bondT
#include <ViennaRNA/datastructures/basic.h>
Old typename of <a href="mailto:vrna_bp_stack_s">vrna_bp_stack_s</a>.
Deprecated Use vrna_bp_stack_t instead!
16.76.4 Function Documentation
```

16.76.4.1 vrna_C11_features()

#include <ViennaRNA/datastructures/basic.h>

Dummy symbol to check whether the library was build using C11/C++11 features.

By default, several data structures of our new v3.0 API use C11/C++11 features, such as unnamed unions, unnamed structs. However, these features can be deactivated at compile time to allow building the library and executables with compilers that do not support these features.

Now, the problem arises that once our static library is compiled and a third-party application is supposed to link against it, it needs to know, at compile time, how to correctly address particular data structures. This is usually implicitely taken care of through the API exposed in our header files. Unfortunately, we had some preprocessor directives in our header files that changed the API depending on the capabilities of the compiler the third-party application is build with. This in turn prohibited the use of an RNAlib compiled without C11/C++11 support in a program that compiles/links with enabled C11/C++11 support and vice-versa.

Therefore, we introduce this dummy symbol which can be used to check, whether the static library was build with C11/C++11 features.

Note

If the symbol is present, the library was build with enabled C11/C++11 features support and no action is required. However, if the symbol is missing in RNAlib >= 2.2.9, programs that link to RNAlib must define a pre-processor identifier $VRNA_DISABLE_C11_FEATURES$ before including any ViennaRNA Package header file, for instance by adding a CPPFLAG

CPPFLAGS+=-DVRNA_DISABLE_C11_FEATURES

Since

v2.2.9

16.77 Messages 583

16.77 Messages

Functions to print various kind of messages.

16.77.1 Detailed Description

Functions to print various kind of messages.

Collaboration diagram for Messages:

Functions

• void vrna_message_error (const char *format,...)

Print an error message and die.

void vrna_message_verror (const char *format, va_list args)

Print an error message and die.

• void vrna_message_warning (const char *format,...)

Print a warning message.

• void vrna_message_vwarning (const char *format, va_list args)

Print a warning message.

void vrna_message_info (FILE *fp, const char *format,...)

Print an info message.

void vrna_message_vinfo (FILE *fp, const char *format, va_list args)

Print an info message.

void vrna_message_input_seq_simple (void)

Print a line to stdout that asks for an input sequence.

• void vrna_message_input_seq (const char *s)

Print a line with a user defined string and a ruler to stdout.

16.77.2 Function Documentation

16.77.2.1 vrna_message_error()

#include <ViennaRNA/utils/basic.h>

Print an error message and die.

This function is a wrapper to *fprintf(stderr, ...)* that puts a capital **ERROR:** in front of the message and then exits the calling program.

See also

vrna_message_verror(), vrna_message_warning(), vrna_message_info()

Parameters

format	The error message to be printed
	Optional arguments for the formatted message string

16.77.2.2 vrna_message_verror()

#include <ViennaRNA/utils/basic.h>

Print an error message and die.

This function is a wrapper to *vfprintf(stderr, ...)* that puts a capital **ERROR:** in front of the message and then exits the calling program.

See also

vrna_message_error(), vrna_message_warning(), vrna_message_info()

Parameters

format	The error message to be printed
args	The argument list for the formatted message string

16.77.2.3 vrna_message_warning()

#include <ViennaRNA/utils/basic.h>

Print a warning message.

This function is a wrapper to *fprintf(stderr, ...)* that puts a capital **WARNING:** in front of the message.

See also

vrna_message_vwarning(), vrna_message_error(), vrna_message_info()

16.77 Messages 585

Parameters

format	The warning message to be printed
	Optional arguments for the formatted message string

16.77.2.4 vrna_message_vwarning()

#include <ViennaRNA/utils/basic.h>

Print a warning message.

This function is a wrapper to fprintf(stderr, ...) that puts a capital WARNING: in front of the message.

See also

vrna_message_vwarning(), vrna_message_error(), vrna_message_info()

Parameters

format	The warning message to be printed
args	The argument list for the formatted message string

16.77.2.5 vrna_message_info()

```
void vrna_message_info (
    FILE * fp,
    const char * format,
    ... )
```

#include <ViennaRNA/utils/basic.h>

Print an info message.

This function is a wrapper to fprintf(...).

See also

vrna_message_vinfo(), vrna_message_error(), vrna_message_warning()

Parameters

fp	The file pointer where the message is printed to
format	The warning message to be printed
	Optional arguments for the formatted message string

16.77.2.6 vrna_message_vinfo()

#include <ViennaRNA/utils/basic.h>

Print an info message.

This function is a wrapper to fprintf(...).

See also

vrna_message_vinfo(), vrna_message_error(), vrna_message_warning()

Parameters

fp	The file pointer where the message is printed to
format	The info message to be printed
args	The argument list for the formatted message string

16.77.2.7 vrna_message_input_seq_simple()

#include <ViennaRNA/utils/basic.h>

Print a line to stdout that asks for an input sequence.

There will also be a ruler (scale line) printed that helps orientation of the sequence positions

16.77 Messages 587

16.77.2.8 vrna_message_input_seq()

Print a line with a user defined string and a ruler to stdout.

(usually this is used to ask for user input) There will also be a ruler (scale line) printed that helps orientation of the sequence positions

Parameters

A user defined string that will be printed to stdout

16.78 Unit Conversion 589

16.78 Unit Conversion

Functions to convert between various physical units.

16.78.1 Detailed Description

Functions to convert between various physical units.

Collaboration diagram for Unit Conversion:

Files

· file units.h

Physical Units and Functions to convert them into each other.

Enumerations

```
    enum vrna_unit_energy_e {
        VRNA_UNIT_J, VRNA_UNIT_KJ, VRNA_UNIT_CAL_IT, VRNA_UNIT_DACAL_IT,
        VRNA_UNIT_KCAL_IT, VRNA_UNIT_CAL, VRNA_UNIT_DACAL, VRNA_UNIT_KCAL,
        VRNA_UNIT_G_TNT, VRNA_UNIT_KG_TNT, VRNA_UNIT_T_TNT, VRNA_UNIT_EV,
        VRNA_UNIT_WH, VRNA_UNIT_KWH }
        Energy / Work Units.
```

enum vrna_unit_temperature_e {
 VRNA_UNIT_K, VRNA_UNIT_DEG_C, VRNA_UNIT_DEG_F, VRNA_UNIT_DEG_R,
 VRNA_UNIT_DEG_N, VRNA_UNIT_DEG_DE, VRNA_UNIT_DEG_RE, VRNA_UNIT_DEG_RO }

Temperature Units.

Functions

- double vrna_convert_energy (double energy, vrna_unit_energy_e from, vrna_unit_energy_e to)

 Convert between energy / work units.
- double vrna_convert_temperature (double temp, vrna_unit_temperature_e from, vrna_unit_temperature_e to)

Convert between temperature units.

16.78.2 Enumeration Type Documentation

```
16.78.2.1 vrna_unit_energy_e
enum vrna_unit_energy_e
#include <ViennaRNA/units.h>
Energy / Work Units.
See also
vrna_convert_energy()
```

Enumerator

VRNA_UNIT_J	Joule ($1 J = 1 kg \cdot m^2 s^{-2}$)
VRNA_UNIT_KJ	Kilojoule ($1\ kJ=1,000\ J$)
VRNA_UNIT_CAL_IT	Calorie (International (Steam) Table, $1\ cal_{IT} = 4.1868\ J$)
	Decacolorie (International (Steam) Table, $1\ dacal_{IT}=10\ cal_{IT}=41.868\ J$)
VRNA_UNIT_DACAL_IT	
VRNA_UNIT_KCAL_IT	Kilocalorie (International (Steam) Table, $1\ kcal_{IT} = 4.1868\ kJ$)
VRNA_UNIT_CAL	Calorie (Thermochemical, $1 \ cal_{th} = 4.184 \ J$)
VRNA_UNIT_DACAL	Decacalorie (Thermochemical, $1 \ dacal_{th} = 10 \ cal_{th} = 41.84 \ J$)
VRNA_UNIT_KCAL	Kilocalorie (Thermochemical, 1 $kcal_{th} = 4.184 \ kJ$)
VRNA_UNIT_G_TNT	g TNT ($1~g~\mathrm{TNT} = 1,000~cal_{th} = 4,184~J$)
VRNA_UNIT_KG_TNT	kg TNT ($1~kg~\mathrm{TNT} = 1,000~kcal_{th} = 4,184~kJ$)
VRNA_UNIT_T_TNT	ton TNT ($1~t~{\rm TNT}=1,000,000~kcal_{th}=4,184~MJ$)
VRNA_UNIT_EV	Electronvolt ($1~eV = 1.602176565 \times 10^{-19}~J$)
VRNA_UNIT_WH	Watt hour ($1~W\cdot h = 1~W\cdot 3,600s = 3,600~J = 3.6~kJ$)
VRNA_UNIT_KWH	Kilowatt hour ($1~kW \cdot h = 1~kW \cdot 3,600~s = 3,600~kJ = 3.6~MJ$)

16.78.2.2 vrna_unit_temperature_e

enum vrna_unit_temperature_e

#include <ViennaRNA/units.h>

Temperature Units.

See also

vrna_convert_temperature()

Enumerator

VRNA_UNIT_K	Kelvin (K)
VRNA_UNIT_DEG_C	Degree Celcius (°C) ($[°C] = [K] - 273.15$)
VRNA_UNIT_DEG_F	Degree Fahrenheit (°F) ($[°F] = [K] imes rac{9}{5} - 459.67$)
VRNA_UNIT_DEG_R	Degree Rankine (°R) ($[^{\circ}R] = [K] imes rac{9}{5}$)
VRNA_UNIT_DEG_N	Degree Newton (°N) ($[{}^{\circ}N] = ([K] - 273.15) imes rac{33}{100}$)
VRNA_UNIT_DEG_DE	Degree Delisle (°De) ($[^{\circ}De] = (373.15 - [K]) \times \frac{3}{2}$)
VRNA_UNIT_DEG_RE	Degree Réaumur (°Ré) ($[{}^{\circ}R\acute{e}] = ([K] - 273.15) imes rac{4}{5}$)
VRNA_UNIT_DEG_RO	Degree Rømer (°Rø) ($[$ °Rø $] = ([K] - 273.15) imes rac{21}{40} + 7.5$)

16.78 Unit Conversion 591

16.78.3 Function Documentation

16.78.3.1 vrna_convert_energy()

Convert between energy / work units.

See also

```
vrna_unit_energy_e
```

Parameters

energy	Input energy value
from	Input unit
to	Output unit

Returns

Energy value in Output unit

16.78.3.2 vrna_convert_temperature()

Convert between temperature units.

See also

```
vrna_unit_temperature_e
```

Parameters

temp	Input temperature value
from	Input unit
Generated by Buttiggen unit	

Returns

Temperature value in Output unit

16.79 The Fold Compound

This module provides interfaces that deal with the most basic data structure used in structure predicting and energy evaluating function of the RNAlib.

16.79.1 Detailed Description

This module provides interfaces that deal with the most basic data structure used in structure predicting and energy evaluating function of the RNAlib.

Throughout the entire RNAlib, the vrna_fold_compound_t, is used to group information and data that is required for structure prediction and energy evaluation. Here, you'll find interface functions to create, modify, and delete vrna_fold_compound_t data structures. Collaboration diagram for The Fold Compound:

Files

· file fold compound.h

The Basic Fold Compound API.

Data Structures

• struct vrna_fc_s

The most basic data structure required by many functions throughout the RNAlib. More...

Macros

• #define VRNA STATUS MFE PRE (unsigned char)1

Status message indicating that MFE computations are about to begin.

#define VRNA_STATUS_MFE_POST (unsigned char)2

Status message indicating that MFE computations are finished.

• #define VRNA STATUS PF PRE (unsigned char)3

Status message indicating that Partition function computations are about to begin.

• #define VRNA_STATUS_PF_POST (unsigned char)4

Status message indicating that Partition function computations are finished.

#define VRNA_OPTION_DEFAULT 0U

Option flag to specify default settings/requirements.

• #define VRNA OPTION MFE 1U

Option flag to specify requirement of Minimum Free Energy (MFE) DP matrices and corresponding set of energy parameters.

• #define VRNA_OPTION_PF 2U

Option flag to specify requirement of Partition Function (PF) DP matrices and corresponding set of Boltzmann factors.

• #define VRNA_OPTION_HYBRID 4U

Option flag to specify requirement of dimer DP matrices.

#define VRNA_OPTION_EVAL_ONLY 8U

Option flag to specify that neither MFE, nor PF DP matrices are required.

#define VRNA_OPTION_WINDOW 16U

Option flag to specify requirement of DP matrices for local folding approaches.

Typedefs

- typedef struct vrna_fc_s vrna_fold_compound_t
 - Typename for the fold_compound data structure vrna_fc_s.
- typedef void() vrna_callback_free_auxdata(void *data)
 - Callback to free memory allocated for auxiliary user-provided data.
- typedef void() vrna_callback_recursion_status(unsigned char status, void *data)

Callback to perform specific user-defined actions before, or after recursive computations.

Enumerations

enum vrna fc type e { VRNA FC TYPE SINGLE, VRNA FC TYPE COMPARATIVE }

An enumerator that is used to specify the type of a vrna_fold_compound_t.

Functions

vrna_fold_compound_t * vrna_fold_compound (const char *sequence, vrna_md_t *md_p, unsigned int options)

Retrieve a vrna_fold_compound_t data structure for single sequences and hybridizing sequences.

vrna_fold_compound_t * vrna_fold_compound_comparative (const char **sequences, vrna_md_t *md_p, unsigned int options)

Retrieve a vrna_fold_compound_t data structure for sequence alignments.

void vrna_fold_compound_free (vrna_fold_compound_t *fc)

Free memory occupied by a vrna_fold_compound_t.

void vrna_fold_compound_add_auxdata (vrna_fold_compound_t *fc, void *data, vrna_callback_free_auxdata *f)

Add auxiliary data to the vrna_fold_compound_t.

void vrna_fold_compound_add_callback (vrna_fold_compound_t *fc, vrna_callback_recursion_status *f)
 Add a recursion status callback to the vrna_fold_compound_t.

16.79.2 Data Structure Documentation

16.79.2.1 struct vrna_fc_s

The most basic data structure required by many functions throughout the RNAlib.

Note

Please read the documentation of this data structure carefully! Some attributes are only available for specific types this data structure can adopt.

Warning

Reading/Writing from/to attributes that are not within the scope of the current type usually result in undefined behavior!

See also

vrna_fold_compound_t.type, vrna_fold_compound(), vrna_fold_compound_comparative(), vrna_fold_compound_free(), VRNA_FC_TYPE_SINGLE, VRNA_FC_TYPE_COMPARATIVE

SWIG Wrapper Notes This data structure is wrapped as an object **fold_compound** with several related functions attached as methods.

A new **fold_compound** can be obtained by calling one of its constructors:

- fold_compound(seq) Initialize with a single sequence, or two concatenated sequences separated by an ampersand character '&' (for cofolding)
- fold_compound(aln) Initialize with a sequence alignment aln stored as a list of sequences (with gap characters)

The resulting object has a list of attached methods which in most cases directly correspond to functions that mainly operate on the corresponding *C* data structure:

- type() Get the type of the fold_compound (See vrna_fc_type_e)
- length() Get the length of the sequence(s) or alignment stored within the fold_←
 compound

Collaboration diagram for vrna_fc_s:

Data Fields

Common data fields

• const vrna_fc_type_e type

The type of the vrna_fold_compound_t.

· unsigned int length

The length of the sequence (or sequence alignment)

int cutpoint

The position of the (cofold) cutpoint within the provided sequence. If there is no cutpoint, this field will be set to -1.

unsigned int * strand number

The strand number a particular nucleotide is associated with.

unsigned int * strand_order

The strand order, i.e. permutation of current concatenated sequence.

unsigned int * strand start

The start position of a particular strand within the current concatenated sequence.

• unsigned int * strand_end

The end (last) position of a particular strand within the current concatenated sequence.

- unsigned int strands
- vrna_seq_t * nucleotides
- vrna_msa_t * alignment
- vrna_hc_t * hc

The hard constraints data structure used for structure prediction.

vrna_mx_mfe_t * matrices

The MFE DP matrices.

vrna_mx_pf_t * exp_matrices

The PF DP matrices.

vrna param t * params

The precomputed free energy contributions for each type of loop.

vrna_exp_param_t * exp_params

The precomputed free energy contributions as Boltzmann factors.

int * iindx

DP matrix accessor.

int * jindx

DP matrix accessor.

User-defined data fields

vrna_callback_recursion_status * stat_cb

Recursion status callback (usually called just before, and after recursive computations in the library.

void * auxdata

A pointer to auxiliary, user-defined data.

vrna_callback_free_auxdata * free_auxdata

A callback to free auxiliary user data whenever the fold_compound itself is free'd.

Secondary Structure Decomposition (grammar) related data fields

• vrna sd t * domains struc

Additional structured domains.

vrna_ud_t * domains_up

Additional unstructured domains.

vrna_gr_aux_t * aux_grammar

Data fields available for single/hybrid structure prediction

Data fields for consensus structure prediction

Additional data fields for Distance Class Partitioning

These data fields are typically populated with meaningful data only if used in the context of Distance Class Partitioning

unsigned int maxD1

Maximum allowed base pair distance to first reference.

unsigned int maxD2

Maximum allowed base pair distance to second reference.

short * reference_pt1

A pairtable of the first reference structure.

short * reference_pt2

A pairtable of the second reference structure.

unsigned int * referenceBPs1

Matrix containing number of basepairs of reference structure1 in interval [i,j].

unsigned int * referenceBPs2

Matrix containing number of basepairs of reference structure2 in interval [i,j].

unsigned int * bpdist

Matrix containing base pair distance of reference structure 1 and 2 on interval [i,j].

unsigned int * mm1

Maximum matching matrix, reference struct 1 disallowed.

unsigned int * mm2

Maximum matching matrix, reference struct 2 disallowed.

Additional data fields for local folding

These data fields are typically populated with meaningful data only if used in the context of local folding

int window_size

window size for local folding sliding window approach

char ** ptype_local

Pair type array (for local folding)

16.79.2.1.1 Field Documentation

```
16.79.2.1.1.1 type
```

```
const vrna_fc_type_e vrna_fc_s::type
```

The type of the vrna_fold_compound_t.

Currently possible values are VRNA_FC_TYPE_SINGLE, and VRNA_FC_TYPE_COMPARATIVE

Warning

Do not edit this attribute, it will be automagically set by the corresponding get() methods for the vrna_fold_compound_t. The value specified in this attribute dictates the set of other attributes to use within this data structure.

16.79.2.1.1.2 stat_cb

```
vrna_callback_recursion_status* vrna_fc_s::stat_cb
```

Recursion status callback (usually called just before, and after recursive computations in the library.

See also

vrna_callback_recursion_status(), vrna_fold_compound_add_callback()

16.79.2.1.1.3 auxdata

```
void* vrna_fc_s::auxdata
```

A pointer to auxiliary, user-defined data.

See also

vrna_fold_compound_add_auxdata(), vrna_fold_compound_t.free_auxdata

16.79.2.1.1.4 free_auxdata

```
vrna_callback_free_auxdata* vrna_fc_s::free_auxdata
```

A callback to free auxiliary user data whenever the fold_compound itself is free'd.

See also

vrna_fold_compound_t.auxdata, vrna_callback_free_auxdata()

16.79.2.1.1.5 sequence

```
char* vrna_fc_s::sequence
```

The input sequence string.

Warning

Only available if

```
type==VRNA_FC_TYPE_SINGLE
```

16.79.2.1.1.6 sequence_encoding

```
short* vrna_fc_s::sequence_encoding
```

Numerical encoding of the input sequence.

See also

```
vrna_sequence_encode()
```

Warning

Only available if

```
type==VRNA_FC_TYPE_SINGLE
```

16.79.2.1.1.7 ptype

```
char* vrna_fc_s::ptype
```

Pair type array.

Contains the numerical encoding of the pair type for each pair (i,j) used in MFE, Partition function and Evaluation computations.

Note

This array is always indexed via jindx, in contrast to previously different indexing between mfe and pf variants!

Warning

Only available if

```
type==VRNA_FC_TYPE_SINGLE
```

See also

```
vrna_idx_col_wise(), vrna_ptypes()
```

```
16.79.2.1.1.8 ptype_pf_compat
```

```
char* vrna_fc_s::ptype_pf_compat
```

ptype array indexed via iindx

Deprecated This attribute will vanish in the future! It's meant for backward compatibility only!

Warning

Only available if

```
type==VRNA_FC_TYPE_SINGLE
```

16.79.2.1.1.9 sc

```
vrna_sc_t* vrna_fc_s::sc
```

The soft constraints for usage in structure prediction and evaluation.

Warning

Only available if

```
type==VRNA_FC_TYPE_SINGLE
```

16.79.2.1.1.10 sequences

```
char** vrna_fc_s::sequences
```

The aligned sequences.

Note

The end of the alignment is indicated by a NULL pointer in the second dimension

Warning

Only available if

```
type==VRNA_FC_TYPE_COMPARATIVE
```

16.79.2.1.1.11 n_seq

```
unsigned int vrna_fc_s::n_seq
```

The number of sequences in the alignment.

Warning

Only available if

```
type==VRNA_FC_TYPE_COMPARATIVE
```

16.79.2.1.1.12 cons_seq

```
char* vrna_fc_s::cons_seq
```

The consensus sequence of the aligned sequences.

Warning

Only available if

```
type==VRNA_FC_TYPE_COMPARATIVE
```

16.79.2.1.1.13 S_cons

```
short* vrna_fc_s::S_cons
```

Numerical encoding of the consensus sequence.

Warning

Only available if

```
type==VRNA_FC_TYPE_COMPARATIVE
```

16.79.2.1.1.14 S

```
short** vrna_fc_s::S
```

Numerical encoding of the sequences in the alignment.

Warning

Only available if

```
type==VRNA_FC_TYPE_COMPARATIVE
```

16.79.2.1.1.15 S5

```
short** vrna_fc_s::S5
```

S5[s][i] holds next base 5' of i in sequence s.

Warning

Only available if

type==VRNA_FC_TYPE_COMPARATIVE

16.79.2.1.1.16 S3

```
short** vrna_fc_s::S3
```

SI[s][i] holds next base 3' of i in sequence s.

Warning

Only available if

type==VRNA_FC_TYPE_COMPARATIVE

16.79.2.1.1.17 pscore

```
int* vrna_fc_s::pscore
```

Precomputed array of pair types expressed as pairing scores.

Warning

Only available if

type==VRNA_FC_TYPE_COMPARATIVE

16.79.2.1.1.18 pscore_local

```
int** vrna_fc_s::pscore_local
```

Precomputed array of pair types expressed as pairing scores.

Warning

Only available if

type==VRNA_FC_TYPE_COMPARATIVE

```
16.79.2.1.1.19 pscore_pf_compat
```

```
short* vrna_fc_s::pscore_pf_compat
```

Precomputed array of pair types expressed as pairing scores indexed via iindx.

Deprecated This attribute will vanish in the future!

Warning

Only available if

```
type==VRNA_FC_TYPE_COMPARATIVE
```

16.79.2.1.1.20 scs

```
vrna_sc_t** vrna_fc_s::scs
```

A set of soft constraints (for each sequence in the alignment)

Warning

Only available if

```
type==VRNA_FC_TYPE_COMPARATIVE
```

16.79.3 Macro Definition Documentation

16.79.3.1 VRNA_STATUS_MFE_PRE

```
#define VRNA_STATUS_MFE_PRE (unsigned char)1
#include <ViennaRNA/fold_compound.h>
```

Status message indicating that MFE computations are about to begin.

See also

vrna_fold_compound_t.stat_cb, vrna_callback_recursion_status(), vrna_mfe(), vrna_fold(), vrna_circfold(), vrna_alifold(), vrna_circalifold(), vrna_cofold()

16.79.3.2 VRNA_STATUS_MFE_POST

```
#define VRNA_STATUS_MFE_POST (unsigned char)2
#include <ViennaRNA/fold_compound.h>
```

Status message indicating that MFE computations are finished.

See also

vrna_fold_compound_t.stat_cb, vrna_callback_recursion_status(), vrna_mfe(), vrna_fold(), vrna_circfold(), vrna_alifold(), vrna_circalifold(), vrna_cofold()

16.79.3.3 VRNA_STATUS_PF_PRE

```
#define VRNA_STATUS_PF_PRE (unsigned char)3
#include <ViennaRNA/fold_compound.h>
```

Status message indicating that Partition function computations are about to begin.

See also

vrna_fold_compound_t.stat_cb, vrna_callback_recursion_status(), vrna_pf()

16.79.3.4 VRNA_STATUS_PF_POST

```
#define VRNA_STATUS_PF_POST (unsigned char)4
#include <ViennaRNA/fold_compound.h>
```

Status message indicating that Partition function computations are finished.

See also

vrna fold compound t.stat cb, vrna callback recursion status(), vrna pf()

16.79.3.5 VRNA_OPTION_MFE

```
#define VRNA_OPTION_MFE 1U
#include <ViennaRNA/fold_compound.h>
```

Option flag to specify requirement of Minimum Free Energy (MFE) DP matrices and corresponding set of energy parameters.

See also

vrna_fold_compound(), vrna_fold_compound_comparative(), VRNA_OPTION_EVAL_ONLY

16.79.3.6 VRNA_OPTION_PF

```
#define VRNA_OPTION_PF 2U
#include <ViennaRNA/fold_compound.h>
```

Option flag to specify requirement of Partition Function (PF) DP matrices and corresponding set of Boltzmann factors.

See also

vrna_fold_compound(), vrna_fold_compound_comparative(), VRNA_OPTION_EVAL_ONLY

16.79.3.7 VRNA_OPTION_EVAL_ONLY

```
#define VRNA_OPTION_EVAL_ONLY 8U
#include <ViennaRNA/fold_compound.h>
```

Option flag to specify that neither MFE, nor PF DP matrices are required.

Use this flag in conjuntion with VRNA_OPTION_MFE, and VRNA_OPTION_PF to save memory for a vrna_fold_compound_t obtained from vrna_fold_compound(), or vrna_fold_compound_comparative() in cases where only energy evaluation but no structure prediction is required.

See also

vrna_fold_compound(), vrna_fold_compound_comparative(), vrna_eval_structure()

16.79.4 Typedef Documentation

16.79.4.1 vrna_callback_free_auxdata

```
typedef void() vrna_callback_free_auxdata(void *data)
#include <ViennaRNA/fold_compound.h>
```

Callback to free memory allocated for auxiliary user-provided data.

This type of user-implemented function usually deletes auxiliary data structures. The user must take care to free all the memory occupied by the data structure passed.

Notes on Callback Functions

This callback is supposed to free memory occupied by an auxiliary data structure. It will be called when the vrna_fold_compound_t is erased from memory through a call to vrna_fold_compound_free() and will be passed the address of memory previously bound to the vrna_fold_compound_t via <a

See also

vrna_fold_compound_add_auxdata(), vrna_fold_compound_free(), vrna_fold_compound_add_callback()

Parameters

ata The data that needs to be free'd

16.79.4.2 vrna_callback_recursion_status

```
typedef void() vrna_callback_recursion_status(unsigned char status, void *data)
```

#include <ViennaRNA/fold_compound.h>

Callback to perform specific user-defined actions before, or after recursive computations.

Notes on Callback Functions This function will be called to notify a third-party implementation about the status of a currently ongoing recursion. The purpose of this callback mechanism is to provide users with a simple way to ensure pre- and post conditions for auxiliary mechanisms attached to our implementations.

See also

vrna_fold_compound_add_auxdata(), vrna_fold_compound_add_callback(), vrna_mfe(), vrna_pf(), VRNA_STATUS_MFE_PRE VRNA_STATUS_MFE_POST, VRNA_STATUS_PF_PRE, VRNA_STATUS_PF_POST

Parameters

	status	The status indicator
	data	The data structure that was assigned with vrna_fold_compound_add_auxdata()
İ	status	The status indicator

16.79.5 Enumeration Type Documentation

16.79.5.1 vrna_fc_type_e

enum vrna_fc_type_e

#include <ViennaRNA/fold_compound.h>

An enumerator that is used to specify the type of a vrna_fold_compound_t.

Enumerator

VRNA_FC_TYPE_SINGLE	Type is suitable for single, and hybridizing sequences
VRNA_FC_TYPE_COMPARATIVE	Type is suitable for sequence alignments (consensus structure prediction)

16.79.6 Function Documentation

16.79.6.1 vrna_fold_compound()

#include <ViennaRNA/fold_compound.h>

Retrieve a vrna fold compound t data structure for single sequences and hybridizing sequences.

This function provides an easy interface to obtain a prefilled vrna_fold_compound_t by passing a single sequence, or two contatenated sequences as input. For the latter, sequences need to be seperated by an '&' character like this:

```
char *sequence = "GGGG&CCCC";
```

The optional parameter md_p can be used to specify the model details for successive computations based on the content of the generated vrna_fold_compound_t. Passing NULL will instruct the function to use default model details. The third parameter options may be used to specify dynamic programming (DP) matrix requirements.

Options

- VRNA_OPTION_DEFAULT Option flag to specify default settings/requirements.
- VRNA_OPTION_MFE Option flag to specify requirement of Minimum Free Energy (MFE) DP matrices and corresponding set of energy parameters.
- VRNA_OPTION_PF Option flag to specify requirement of Partition Function (PF) DP matrices and corresponding set of Boltzmann factors.
- VRNA OPTION WINDOW Option flag to specify requirement of DP matrices for local folding approaches.

The above options may be OR-ed together.

If you just need the folding compound serving as a container for your data, you can simply pass VRNA_OPTION_DEFAULT to the option parameter. This creates a vrna_fold_compound_t without DP matrices, thus saving memory. Subsequent calls of any structure prediction function will then take care of allocating the memory required for the DP matrices. If you only intend to evaluate structures instead of actually predicting them, you may use the VRNA_OPTION_EVAL_ONLY macro. This will seriously speedup the creation of the vrna fold compound t.

Note

The sequence string must be uppercase, and should contain only RNA (resp. DNA) alphabet depending on what energy parameter set is used

See also

vrna_fold_compound_free(), vrna_fold_compound_comparative(), vrna_md_t

Parameters

sequence	A single sequence, or two concatenated sequences seperated by an '&' character	
md_p	An optional set of model details	
options	The options for DP matrices memory allocation	

Returns

A prefilled vrna fold compound t ready to be used for computations (may be NULL on error)

16.79.6.2 vrna_fold_compound_comparative()

Retrieve a vrna_fold_compound_t data structure for sequence alignments.

This function provides an easy interface to obtain a prefilled vrna_fold_compound_t by passing an alignment of sequences.

The optional parameter md_p can be used to specify the model details for successive computations based on the content of the generated vrna_fold_compound_t. Passing NULL will instruct the function to use default model details. The third parameter options may be used to specify dynamic programming (DP) matrix requirements.

Options

- VRNA OPTION DEFAULT Option flag to specify default settings/requirements.
- VRNA_OPTION_MFE Option flag to specify requirement of Minimum Free Energy (MFE) DP matrices and corresponding set of energy parameters.
- VRNA_OPTION_PF Option flag to specify requirement of Partition Function (PF) DP matrices and corresponding set of Boltzmann factors.
- VRNA_OPTION_WINDOW Option flag to specify requirement of DP matrices for local folding approaches.

The above options may be OR-ed together.

If you just need the folding compound serving as a container for your data, you can simply pass VRNA_OPTION_DEFAULT to the option parameter. This creates a vrna_fold_compound_t without DP matrices, thus saving memory. Subsequent calls of any structure prediction function will then take care of allocating the memory required for the DP matrices. If you only intend to evaluate structures instead of actually predicting them, you may use the VRNA_OPTION_EVAL_ONLY macro. This will seriously speedup the creation of the vrna_fold_compound_t.

Note

The sequence strings must be uppercase, and should contain only RNA (resp. DNA) alphabet including gap characters depending on what energy parameter set is used.

See also

vrna_fold_compound_free(), vrna_fold_compound(), vrna_md_t, VRNA_OPTION_MFE, VRNA_OPTION_PF, VRNA_OPTION_EVAL_ONLY, read_clustal()

Parameters

sequences	A sequence alignment including 'gap' characters
md_p	An optional set of model details
options	The options for DP matrices memory allocation

Returns

A prefilled vrna_fold_compound_t ready to be used for computations (may be NULL on error)

16.79.6.3 vrna_fold_compound_free()

Free memory occupied by a vrna_fold_compound_t.

See also

vrna_fold_compound(), vrna_fold_compound_comparative(), vrna_mx_mfe_free(), vrna_mx_pf_free()

Parameters

fc The vrna_fold_compound_t that is to be erased from memory

16.79.6.4 vrna_fold_compound_add_auxdata()

Add auxiliary data to the vrna_fold_compound_t.

This function allows one to bind arbitrary data to a vrna_fold_compound_t which may later on be used by one of the callback functions, e.g. vrna_callback_recursion_status(). To allow for proper cleanup of the memory occupied by this auxiliary data, the user may also provide a pointer to a cleanup function that free's the corresponding memory. This function will be called automatically when the vrna_fold_compound_t is free'd with vrna_fold_compound_free().

Note

Before attaching the arbitrary data pointer, this function will call the vrna_callback_free_auxdata() on any pre-existing data that is already attached.

See also

vrna_callback_free_auxdata()

Parameters

fc	The fold_compound the arbitrary data pointer should be associated with	
data	A pointer to an arbitrary data structure	
f	A pointer to function that free's memory occupied by the arbitrary data (May be NULL)	

16.79.6.5 vrna_fold_compound_add_callback()

Add a recursion status callback to the vrna_fold_compound_t.

Binding a recursion status callback function to a vrna_fold_compound_ t allows one to perform arbitrary operations just before, or after an actual recursive computations, e.g. MFE prediction, is performed by the RNAlib. The callback function will be provided with a pointer to its vrna_fold_compound_, and a status message. Hence, it has complete access to all variables that incluence the recursive computations.

See also

vrna_callback_recursion_status(), vrna_fold_compound_t, VRNA_STATUS_MFE_PRE, VRNA_STATUS_MFE_POST, VRNA_STATUS_PF_PRE, VRNA_STATUS_PF_POST

Parameters

fc	The fold_compound the callback function should be attached to
f	The pointer to the recursion status callback function

16.80 The Dynamic Programming Matrices

This module provides interfaces that deal with creation and destruction of dynamic programming matrices used within the RNAlib.

16.80.1 Detailed Description

This module provides interfaces that deal with creation and destruction of dynamic programming matrices used within the RNAlib.

Collaboration diagram for The Dynamic Programming Matrices:

Data Structures

· struct vrna mx mfe s

Minimum Free Energy (MFE) Dynamic Programming (DP) matrices data structure required within the vrna fold compound t. More...

· struct vrna mx pf s

Partition function (PF) Dynamic Programming (DP) matrices data structure required within the vrna_fold_compound_t.

More...

Typedefs

typedef struct vrna_mx_mfe_s vrna_mx_mfe_t

Typename for the Minimum Free Energy (MFE) DP matrices data structure vrna_mx_mfe_s.

typedef struct vrna_mx_pf_s vrna_mx_pf_t

Typename for the Partition Function (PF) DP matrices data structure vrna_mx_pf_s.

Enumerations

enum vrna mx type e { VRNA MX DEFAULT, VRNA MX WINDOW, VRNA MX 2DFOLD }

An enumerator that is used to specify the type of a polymorphic Dynamic Programming (DP) matrix data structure.

Functions

• int vrna_mx_add (vrna_fold_compound_t *vc, vrna_mx_type_e type, unsigned int options)

Add Dynamic Programming (DP) matrices (allocate memory)

void vrna_mx_mfe_free (vrna_fold_compound_t *vc)

Free memory occupied by the Minimum Free Energy (MFE) Dynamic Programming (DP) matrices.

void vrna_mx_pf_free (vrna_fold_compound_t *vc)

Free memory occupied by the Partition Function (PF) Dynamic Programming (DP) matrices.

16.80.2 Data Structure Documentation

16.80.2.1 struct vrna_mx_mfe_s

Minimum Free Energy (MFE) Dynamic Programming (DP) matrices data structure required within the vrna_fold_compound_t.

Data Fields

Common fields for MFE matrices

- vrna_mx_type_e type
- unsigned int length

Length of the sequence, therefore an indicator of the size of the DP matrices.

Default DP matrices

Note

```
These data fields are available if vrna_mx_mfe_t.type == VRNA_MX_DEFAULT
```

Local Folding DP matrices using window approach

Note

```
These data fields are available if vrna_mx_mfe_t.type == VRNA_MX_WINDOW
```

Distance Class DP matrices

Note

```
These data fields are available if vrna_mx_mfe_t.type == VRNA_MX_2DFOLD
```

```
16.80.2.2 struct vrna_mx_pf_s
```

Partition function (PF) Dynamic Programming (DP) matrices data structure required within the vrna_fold_compound_t.

Data Fields

Common fields for DP matrices

```
• vrna_mx_type_e type
```

- unsigned int length
- FLT_OR_DBL * scale
- FLT_OR_DBL * expMLbase

Default PF matrices

Note

```
These data fields are available if vrna_mx_pf_t.type == VRNA_MX_DEFAULT
```

Local Folding DP matrices using window approach

Note

```
These data fields are available if vrna_mx_mfe_t.type == VRNA_MX_WINDOW
```

Distance Class DP matrices

Note

```
These data fields are available if vrna_mx_pf_t.type == VRNA_MX_2DFOLD
```

16.80.3 Enumeration Type Documentation

```
16.80.3.1 vrna_mx_type_e
enum vrna_mx_type_e
#include <ViennaRNA/dp_matrices.h>
```

An enumerator that is used to specify the type of a polymorphic Dynamic Programming (DP) matrix data structure.

See also

```
vrna_mx_mfe_t, vrna_mx_pf_t
```

Enumerator

VRNA_MX_DEFAULT	Default DP matrices.
VRNA_MX_WINDOW	DP matrices suitable for local structure prediction using window approach.
	See also
	vrna_mfe_window(), vrna_mfe_window_zscore(), pfl_fold()
VRNA_MX_2DFOLD	DP matrices suitable for distance class partitioned structure prediction.
	See also
	vrna_mfe_TwoD(), vrna_pf_TwoD()

16.80.4 Function Documentation

Add Dynamic Programming (DP) matrices (allocate memory)

Note

Usually, there is no need to call this function, since the constructors of vrna_fold_compound_t are handling all the DP matrix memory allocation.

See also

vrna_mx_mfe_add(), vrna_mx_pf_add(), vrna_fold_compound(), vrna_fold_compound_comparative(),
vrna_fold_compound_free(), vrna_mx_pf_free(), vrna_mx_mfe_free(), vrna_mx_type_e, VRNA_OPTION_MFE,
VRNA_OPTION_PF, VRNA_OPTION_HYBRID, VRNA_OPTION_EVAL_ONLY

Parameters

VC	The vrna_fold_compound_t that holds pointers to the DP matrices
type	The type of DP matrices requested
options	Option flags that specify the kind of DP matrices, such as MFE or PF arrays, and auxiliary requirements

Returns

1 if DP matrices were properly allocated and attached, 0 otherwise

16.80.4.2 vrna_mx_mfe_free()

Free memory occupied by the Minimum Free Energy (MFE) Dynamic Programming (DP) matrices.

See also

vrna_fold_compound(), vrna_fold_compound_comparative(), vrna_fold_compound_free(), vrna_mx_pf_free()

Parameters

vc The vrna_fold_compound_t storing the MFE DP matrices that are to be erased from memory

16.80.4.3 vrna_mx_pf_free()

Free memory occupied by the Partition Function (PF) Dynamic Programming (DP) matrices.

See also

 $vrna_fold_compound(), vrna_fold_compound_comparative(), vrna_fold_compound_free(), vrna_mx_mfe_free()$

Parameters

vc The vrna_fold_compound_t storing the PF DP matrices that are to be erased from memory

16.81 Hash Tables 615

16.81 Hash Tables

Various implementations of hash table functions.

16.81.1 Detailed Description

Various implementations of hash table functions.

Hash tables are common data structures that allow for fast random access to the data that is stored within.

Here, we provide an abstract implementation of a hash table interface and a concrete implementation for pairs of secondary structure and corresponding free energy value. Collaboration diagram for Hash Tables:

Files

· file hash_tables.h

Implementations of hash table functions.

Data Structures

· struct vrna ht entry db t

Default hash table entry. More ...

Abstract interface

typedef struct vrna_hash_table_s * vrna_hash_table_t

A hash table object.

typedef int() vrna_callback_ht_compare_entries(void *x, void *y)

Callback function to compare two hash table entries.

• typedef unsigned int() vrna_callback_ht_hash_function(void *x, unsigned long hashtable_size)

Callback function to generate a hash key, i.e. hash function.

typedef int() vrna_callback_ht_free_entry(void *x)

Callback function to free a hash table entry.

• vrna_hash_table_t vrna_ht_init (unsigned int b, vrna_callback_ht_compare_entries *compare_function, vrna_callback_ht_hash_function *hash_function, vrna_callback_ht_free_entry *free_hash_entry)

Get an initialized hash table.

unsigned long vrna_ht_size (vrna_hash_table_t ht)

Get the size of the hash table.

unsigned long vrna_ht_collisions (struct vrna_hash_table_s *ht)

Get the number of collisions in the hash table.

void * vrna_ht_get (vrna_hash_table_t ht, void *x)

Get an element from the hash table.

int vrna_ht_insert (vrna_hash_table_t ht, void *x)

Insert an object into a hash table.

void vrna_ht_remove (vrna_hash_table_t ht, void *x)

Remove an object from the hash table.

void vrna_ht_clear (vrna_hash_table_t ht)

Clear the hash table.

void vrna_ht_free (vrna_hash_table_t ht)

Free all memory occupied by the hash table.

Dot-Bracket / Free Energy entries

```
int vrna_ht_db_comp (void *x, void *y)
          Default hash table entry comparison.

    unsigned int vrna_ht_db_hash_func (void *x, unsigned long hashtable_size)

          Default hash function.
    int vrna_ht_db_free_entry (void *hash_entry)
          Default function to free memory occupied by a hash entry.
16.81.2 Data Structure Documentation
16.81.2.1 struct vrna_ht_entry_db_t
```

See also

vrna_ht_init(), vrna_ht_db_comp(), vrna_ht_db_hash_func(), vrna_ht_db_free_entry()

Data Fields

• char * structure

Default hash table entry.

· float energy

16.81.2.1.1 Field Documentation

```
16.81.2.1.1.1 structure
```

```
char* vrna_ht_entry_db_t::structure
```

A secondary structure in dot-bracket notation

16.81.2.1.1.2 energy

```
float vrna_ht_entry_db_t::energy
```

The free energy of structure

16.81.3 Typedef Documentation

16.81 Hash Tables 617

```
16.81.3.1 vrna_hash_table_t
typedef struct vrna_hash_table_s* vrna_hash_table_t
#include <ViennaRNA/datastructures/hash_tables.h>
A hash table object.
See also
     vrna_ht_init(), vrna_ht_free()
16.81.3.2 vrna_callback_ht_compare_entries
typedef int() vrna_callback_ht_compare_entries(void *x, void *y)
#include <ViennaRNA/datastructures/hash_tables.h>
Callback function to compare two hash table entries.
See also
     vrna_ht_init(), vrna_ht_db_comp()
Parameters
     A hash table entry
     A hash table entry
Returns
     -1 if x is smaller, +1 if x is larger than y. 0 if x == y
16.81.3.3 vrna_callback_ht_hash_function
typedef unsigned int() vrna_callback_ht_hash_function(void *x, unsigned long hashtable_size)
#include <ViennaRNA/datastructures/hash_tables.h>
Callback function to generate a hash key, i.e. hash function.
```

vrna_ht_init(), vrna_ht_db_hash_func()

See also

Parameters

X	A hash table entry
hashtable_size	The size of the hash table

Returns

The hash table key for entry \boldsymbol{x}

```
16.81.3.4 vrna_callback_ht_free_entry
```

```
typedef int() vrna_callback_ht_free_entry(void *x)
#include <ViennaRNA/datastructures/hash_tables.h>
```

Callback function to free a hash table entry.

See also

```
vrna_ht_init(), vrna_ht_db_free_entry()
```

Parameters

```
x A hash table entry
```

Returns

0 on success

16.81.4 Function Documentation

```
16.81.4.1 vrna_ht_init()
```

```
vrna_hash_table_t vrna_ht_init (
          unsigned int b,
          vrna_callback_ht_compare_entries * compare_function,
          vrna_callback_ht_hash_function * hash_function,
          vrna_callback_ht_free_entry * free_hash_entry )
```

#include <ViennaRNA/datastructures/hash_tables.h>

Get an initialized hash table.

This function returns a ready-to-use hash table with pre-allocated memory for a particular number of entries.

16.81 Hash Tables 619

Note

If all function pointers are \mathtt{NULL} , this function initializes the hash table with *default functions*, i.e.

```
• vrna_ht_db_comp() for the compare_function,
```

- vrna_ht_db_hash_func() for the hash_function, and
- vrna_ht_db_free_entry() for the free_hash_entry

arguments.

Warning

If hash_bits is larger than 27 you have to compile it with the flag gcc -mcmodel=large.

Parameters

b	Number of bits for the hash table. This determines the size (2^b-1).
compare_function A function pointer to compare any two entr	A function pointer to compare any two entries in the hash table (may be NULL)
hash_function	A function pointer to retrieve the hash value of any entry (may be NULL)
free_hash_entry	A function pointer to free the memory occupied by any entry (may be NULL)

Returns

An initialized, empty hash table, or \mathtt{NULL} on any error

16.81.4.2 vrna_ht_size()

#include <ViennaRNA/datastructures/hash_tables.h>

Get the size of the hash table.

Parameters

```
ht The hash table
```

Returns

The size of the hash table, i.e. the maximum number of entries

16.81.4.3 vrna_ht_collisions()

```
unsigned long vrna_ht_collisions ( {\tt struct\ vrna\_hash\_table\_s\ *\ ht\ )}
```

#include <ViennaRNA/datastructures/hash_tables.h>

Get the number of collisions in the hash table.

16.81 Hash Tables 621

Parameters

ht The hash table	
-------------------	--

Returns

The number of collisions in the hash table

16.81.4.4 vrna_ht_get()

#include <ViennaRNA/datastructures/hash_tables.h>

Get an element from the hash table.

This function takes an object x and performs a look-up whether the object is stored within the hash table ht. If the object is already stored in ht, the function simply returns the entry, otherwise it returns NULL.

See also

```
vrna_ht_insert(), vrna_hash_delete(), vrna_ht_init()
```

Parameters

ht	The hash table
Х	The hash entry to look-up

Returns

The entry x if it is stored in ht, NULL otherwise

16.81.4.5 vrna_ht_insert()

#include <ViennaRNA/datastructures/hash_tables.h>

Insert an object into a hash table.

Writes the pointer to your hash entry into the table.

Warning

In case of collisions, this function simply increments the hash key until a free entry in the hash table is found.

See also

```
vrna_ht_init(), vrna_hash_delete(), vrna_ht_clear()
```

Parameters

ht	The hash table
Х	The hash entry

Returns

0 on success, 1 if the value is already in the hash table, -1 on error.

16.81.4.6 vrna_ht_remove()

Remove an object from the hash table.

Deletes the pointer to your hash entry from the table.

Note

This function doesn't free any memory occupied by the hash entry.

Parameters

ht	The hash table
X	The hash entry

16.81.4.7 vrna_ht_clear()

16.81 Hash Tables 623

Clear the hash table.

This function removes all entries from the hash table and automatically free's the memory occupied by each entry using the bound @vrna_callback_ht_free_entry() function.

See also

```
vrna_ht_free(), vrna_ht_init()
```

Parameters

```
ht The hash table
```

16.81.4.8 vrna_ht_free()

#include <ViennaRNA/datastructures/hash_tables.h>

Free all memory occupied by the hash table.

This function removes all entries from the hash table by calling the vrna_callback_ht_free_entry() function for each entry. Finally, the memory occupied by the hash table itself is free'd as well.

Parameters

```
ht The hash table
```

16.81.4.9 vrna_ht_db_comp()

```
int vrna_ht_db_comp (  \mbox{void} \ * \ x, \\ \mbox{void} \ * \ y \ ) \label{eq:vrna_ht_db_comp}
```

#include <ViennaRNA/datastructures/hash_tables.h>

Default hash table entry comparison.

This is the default comparison function for hash table entries. It assumes the both entries x and y are of type $\frac{\text{vrna_ht_entry_db_t}}{\text{and compares the structure}}$ attribute of both entries

See also

```
vrna_ht_entry_db_t, vrna_ht_init(), vrna_ht_db_hash_func(), vrna_ht_db_free_entry()
```

Parameters

X	A hash table entry of type vrna_ht_entry_db_t
У	A hash table entry of type vrna_ht_entry_db_t

Returns

-1 if x is smaller, +1 if x is larger than y. 0 if both are equal.

16.81.4.10 vrna_ht_db_hash_func()

#include <ViennaRNA/datastructures/hash_tables.h>

Default hash function.

This is the default hash function for hash table insertion/lookup. It assumes that entries are of type vrna_ht_entry_db_t and uses the Bob Jenkins 1996 mix function to create a hash key from the structure attribute of the hash entry.

See also

```
vrna_ht_entry_db_t, vrna_ht_init(), vrna_ht_db_comp(), vrna_ht_db_free_entry()
```

Parameters

X	A hash table entry to compute the key for
hashtable_size	The size of the hash table

Returns

The hash key for entry \boldsymbol{x}

16.81.4.11 vrna_ht_db_free_entry()

#include <ViennaRNA/datastructures/hash_tables.h>

Default function to free memory occupied by a hash entry.

This function assumes that hash entries are of type vrna_ht_entry_db_t and free's the memory occupied by that entry.

16.81 Hash Tables 625

See also

vrna_ht_entry_db_t, vrna_ht_init(), vrna_ht_db_comp(), vrna_ht_db_hash_func()

Parameters

hash_entry	The hash entry to remove from memory
------------	--------------------------------------

Returns

0 on success

16.82 **Heaps**

Interface for an abstract implementation of a heap data structure.

16.82.1 Detailed Description

Interface for an abstract implementation of a heap data structure.

Collaboration diagram for Heaps:

Files

· file heap.h

Implementation of an abstract heap data structure.

Typedefs

typedef struct vrna_heap_s * vrna_heap_t

An abstract heap data structure.

typedef int() vrna_callback_heap_cmp(const void *a, const void *b, void *data)

Heap compare function prototype.

typedef size_t() vrna_callback_heap_get_pos(const void *a, void *data)

Retrieve the position of a particular heap entry within the heap.

typedef void() vrna_callback_heap_set_pos(const void *a, size_t pos, void *data)

Store the position of a particular heap entry within the heap.

Functions

vrna_heap_t vrna_heap_init (size_t n, vrna_callback_heap_cmp *cmp, vrna_callback_heap_get_pos *get
 —entry_pos, vrna_callback_heap_set_pos *set_entry_pos, void *data)

Initialize a heap data structure.

void vrna_heap_free (vrna_heap_t h)

Free memory occupied by a heap data structure.

size_t vrna_heap_size (struct vrna_heap_s *h)

Get the size of a heap data structure, i.e. the number of stored elements.

void vrna_heap_insert (vrna_heap_t h, void *v)

Insert an element into the heap.

void * vrna_heap_pop (vrna_heap_t h)

Pop (remove and return) the object at the root of the heap.

• const void * vrna_heap_top (vrna_heap_t h)

Get the object at the root of the heap.

void * vrna_heap_remove (vrna_heap_t h, const void *v)

Remove an arbitrary element within the heap.

void * vrna_heap_update (vrna_heap_t h, void *v)

Update an arbitrary element within the heap.

16.82 Heaps 627

16.82.2 Typedef Documentation

16.82.2.1 vrna_heap_t

```
typedef struct vrna_heap_s* vrna_heap_t
```

#include <ViennaRNA/datastructures/heap.h>

An abstract heap data structure.

See also

vrna_heap_init(), vrna_heap_free(), vrna_heap_insert(), vrna_heap_pop(), vrna_heap_top(), vrna_heap_top(), vrna_heap_top(), vrna_heap_update()

16.82.2.2 vrna_callback_heap_cmp

```
typedef int() vrna_callback_heap_cmp(const void *a, const void *b, void *data)
#include <ViennaRNA/datastructures/heap.h>
```

Heap compare function prototype.

Use this prototype to design the compare function for the heap implementation. The arbitrary data pointer data may be used to get access to further information required to actually compare the two values a and b.

Note

The heap implementation acts as a *min-heap*, therefore, the minimum element will be present at the heap's root. In case a *max-heap* is required, simply reverse the logic of this compare function.

Parameters

а	The first object to compare
b	The second object to compare
data	An arbitrary data pointer passed through from the heap implementation

Returns

A value less than zero if a < b, a value greater than zero if a > b, and 0 otherwise

16.82.2.3 vrna_callback_heap_get_pos

```
typedef size_t() vrna_callback_heap_get_pos(const void *a, void *data)
#include <ViennaRNA/datastructures/heap.h>
```

Retrieve the position of a particular heap entry within the heap.

Parameters

а	The object to look-up within the heap
data	An arbitrary data pointer passed through from the heap implementation

Returns

The position of the element a within the heap, or 0 if it is not in the heap

16.82.2.4 vrna_callback_heap_set_pos

```
typedef void() vrna_callback_heap_set_pos(const void *a, size_t pos, void *data)
#include <ViennaRNA/datastructures/heap.h>
```

Store the position of a particular heap entry within the heap.

Parameters

а	The object whose position shall be stored
pos	The current position of a within the heap, or 0 if a was deleted
data	An arbitrary data pointer passed through from the heap implementation

16.82.3 Function Documentation

16.82.3.1 vrna_heap_init()

#include <ViennaRNA/datastructures/heap.h>

16.82 Heaps 629

Initialize a heap data structure.

This function initializes a heap data structure. The implementation is based on a *min-heap*, i.e. the minimal element is located at the root of the heap. However, by reversing the logic of the compare function, one can easily transform this into a *max-heap* implementation.

Beside the regular operations on a heap data structure, we implement removal and update of arbitrary elements within the heap. For that purpose, however, one requires a reverse-index lookup system that, (i) for a given element stores the current position in the heap, and (ii) allows for fast lookup of an elements current position within the heap. The corresponding getter- and setter- functions may be provided through the arguments get_entry_pos and set_entry_pos, respectively.

Sometimes, it is difficult to simply compare two data structures without any context. Therefore, the compare function is provided with a user-defined data pointer that can hold any context required.

Warning

If any of the arguments get_entry_pos or set_entry_pos is NULL, the operations vrna_heap_update() and vrna_heap_remove() won't work.

See also

vrna_heap_free(), vrna_heap_insert(), vrna_heap_pop(), vrna_heap_top(), vrna_heap_remove(), vrna_heap_update(), vrna_heap_t, vrna_callback_heap_cmp, vrna_callback_heap_get_pos, vrna_callback_heap_set_pos

Parameters

n	The initial size of the heap, i.e. the number of elements to store
стр	The address of a compare function that will be used to fullfill the partial order requirement
get_entry_pos	The address of a function that retrieves the position of an element within the heap (or NULL)
set_entry_pos	The address of a function that stores the position of an element within the heap (or NULL)
data	An arbitrary data pointer passed through to the compare function cmp, and the set/get functions
	get_entry_pos/set_entry_pos

Returns

An initialized heap data structure, or NULL on error

16.82.3.2 vrna_heap_free()

Free memory occupied by a heap data structure.

See also

vrna_heap_init()

Parameters

h The heap that should be free'd

16.82.3.3 vrna_heap_size()

Get the size of a heap data structure, i.e. the number of stored elements.

Parameters

h The heap data structure

Returns

The number of elements currently stored in the heap, or 0 upon any error

16.82.3.4 vrna_heap_insert()

#include <ViennaRNA/datastructures/heap.h>

Insert an element into the heap.

See also

vrna_heap_init(), vrna_heap_pop(), vrna_heap_top(), vrna_heap_free(), vrna_heap_remove(), vrna_heap_update()

Parameters

h	The heap data structure
V	A pointer to the object that is about to be inserted into the heap

16.82 Heaps 631

16.82.3.5 vrna_heap_pop()

Pop (remove and return) the object at the root of the heap.

This function removes the root from the heap and returns it to the caller.

See also

```
vrna_heap_init(), vrna_heap_top(), vrna_heap_insert(), vrna_heap_free() vrna_heap_remove(), vrna_heap_update()
```

Parameters

h The heap data structure

Returns

The object at the root of the heap, i.e. the minimal element (or NULL if (a) the heap is empty or (b) any error occurred)

16.82.3.6 vrna_heap_top()

#include <ViennaRNA/datastructures/heap.h>

Get the object at the root of the heap.

See also

vrna_heap_init(), vrna_heap_pop(), vrna_heap_insert(), vrna_heap_free() vrna_heap_remove(), vrna_heap_update()

Parameters

h The heap data structure

Returns

The object at the root of the heap, i.e. the minimal element (or NULL if (a) the heap is empty or (b) any error occurred)

16.82.3.7 vrna_heap_remove()

#include <ViennaRNA/datastructures/heap.h>

Remove an arbitrary element within the heap.

See also

```
vrna_heap_init(), vrna_callback_heap_get_pos, vrna_callback_heap_set_pos, vrna_heap_pop(), vrna_heap_free()
```

Warning

This function won't work if the heap was not properly initialized with callback functions for fast reverse-index mapping!

Parameters

h	The heap data structure
V	The object to remove from the heap

Returns

The object that was removed from the heap (or NULL if (a) it wasn't found or (b) any error occurred)

16.82.3.8 vrna_heap_update()

Update an arbitrary element within the heap.

Note

If the object that is to be updated is not currently stored in the heap, it will be inserted. In this case, the function returns NULL.

Warning

This function won't work if the heap was not properly initialized with callback functions for fast reverse-index mapping!

See also

vrna_heap_init(), vrna_callback_heap_get_pos, vrna_callback_heap_set_pos vrna_heap_pop(), vrna_heap_remove(),
vrna_heap_free()

16.82 Heaps 633

Parameters

h	The heap data structure
V	The object to update

Returns

The 'previous' object within the heap that now got replaced by v (or NULL if (a) it wasn't found or (b) any error occurred)

16.83 Buffers

Functions that provide dynamically buffered stream-like data structures.

16.83.1 Detailed Description

Functions that provide dynamically buffered stream-like data structures.

Collaboration diagram for Buffers:

Files

· file char_stream.h

Implementation of a dynamic, buffered character stream.

· file stream_output.h

An implementation of a buffered, ordered stream output data structure.

Typedefs

typedef struct vrna ordered stream s * vrna ostream t

An ordered output stream structure with unordered insert capabilities.

• typedef void() vrna_callback_stream_output(void *auxdata, unsigned int i, void *data)

Ordered stream processing callback.

Functions

• vrna cstr t vrna cstr (size t size, FILE *output)

Create a dynamic char * stream data structure.

void vrna_cstr_free (vrna_cstr_t buf)

Free the memory occupied by a dynamic char * stream data structure.

void vrna cstr close (vrna cstr t buf)

Free the memory occupied by a dynamic char * stream and close the output stream.

• void vrna_cstr_fflush (struct vrna_cstr_s *buf)

Flush the dynamic char * output stream.

• vrna_ostream_t vrna_ostream_init (vrna_callback_stream_output *output, void *auxdata)

Get an initialized ordered output stream.

void vrna_ostream_free (vrna_ostream_t dat)

Free an initialized ordered output stream.

• void vrna_ostream_request (vrna_ostream_t dat, unsigned int num)

Request index in ordered output stream.

void vrna_ostream_provide (vrna_ostream_t dat, unsigned int i, void *data)

Provide output stream data for a particular index.

16.83.2 Typedef Documentation

16.83 Buffers 635

16.83.2.1 vrna_callback_stream_output

```
typedef void() vrna_callback_stream_output(void *auxdata, unsigned int i, void *data)
#include <ViennaRNA/datastructures/stream_output.h>
```

Ordered stream processing callback.

This callback will be processed in sequential order as soon as sequential data in the output stream becomes available.

Note

The callback must also release the memory occupied by the data passed since the stream will lose any reference to it after the callback has been executed.

Parameters

auxdata	A shared pointer for all calls, as provided by the second argument to vrna_ostream_init()
i	The index number of the data passed to data
data	A block of data ready for processing

16.83.3 Function Documentation

16.83.3.1 vrna_cstr()

Create a dynamic char * stream data structure.

See also

```
vrna_cstr_free(), vrna_cstr_close(), vrna_cstr_fflush(), vrna_cstr_printf()
```

Parameters

size	The initial size of the buffer in characters
output	An optional output file stream handle that is used to write the collected data to (defaults to stdout if NULL)

16.83.3.2 vrna_cstr_free()

#include <ViennaRNA/datastructures/char_stream.h>

Free the memory occupied by a dynamic char * stream data structure.

This function first flushes any remaining character data within the stream and then free's the memory occupied by the data structure.

See also

```
vrna_cstr_close(), vrna_cstr_fflush(), vrna_cstr()
```

Parameters

buf | The dynamic char * stream data structure to free

16.83.3.3 vrna_cstr_close()

#include <ViennaRNA/datastructures/char_stream.h>

Free the memory occupied by a dynamic char * stream and close the output stream.

This function first flushes any remaining character data within the stream then closes the attached output file stream (if any), and finally free's the memory occupied by the data structure.

See also

```
vrna_cstr_free(), vrna_cstr_fflush(), vrna_cstr()
```

Parameters

buf The dynamic char * stream data structure to free

16.83.3.4 vrna_cstr_fflush()

16.83 Buffers 637

```
#include <ViennaRNA/datastructures/char_stream.h>
```

Flush the dynamic char * output stream.

This function flushes the collected char * stream, either by writing to the attached file handle, or simply by writing to stdout if no file handle has been attached upon construction using vrna_cstr().

Postcondition

The stream buffer is empty after execution of this function

See also

```
vrna_cstr(), vrna_cstr_close(), vrna_cstr_free()
```

Parameters

```
buf The dynamic char * stream data structure to flush
```

16.83.3.5 vrna_ostream_init()

#include <ViennaRNA/datastructures/stream_output.h>

Get an initialized ordered output stream.

See also

```
vrna_ostream_free(), vrna_ostream_request(), vrna_ostream_provide()
```

Parameters

output	A callback function that processes and releases data in the stream
auxdata	A pointer to auxiliary data passed as first argument to the output callback

Returns

An initialized ordered output stream

16.83.3.6 vrna_ostream_free()

```
#include <ViennaRNA/datastructures/stream_output.h>
```

Free an initialized ordered output stream.

See also

```
vrna_ostream_init()
```

Parameters

dat The output stream for which occupied memory should be free'd

16.83.3.7 vrna_ostream_request()

#include <ViennaRNA/datastructures/stream_output.h>

Request index in ordered output stream.

This function must be called prior to vrna_ostream_provide() to indicate that data associted with a certain index number is expected to be inserted into the stream in the future.

See also

```
vrna_ostream_init(), vrna_ostream_provide(), vrna_ostream_free()
```

Parameters

dat	The output stream for which the index is requested
num	The index to request data for

16.83.3.8 vrna_ostream_provide()

#include <ViennaRNA/datastructures/stream_output.h>

Provide output stream data for a particular index.

16.83 Buffers 639

Precondition

The index data is provided for must have been requested using vrna_ostream_request() beforehand.

See also

vrna_ostream_request()

Parameters

dat	The output stream for which data is provided
i	The index of the provided data
data	The data provided

16.84 Deprecated Interface for Global MFE Prediction

16.84.1 Detailed Description

Collaboration diagram for Deprecated Interface for Global MFE Prediction:

Files

· file alifold.h

Functions for comparative structure prediction using RNA sequence alignments.

· file cofold.h

MFE implementations for RNA-RNA interaction.

• file fold.h

MFE calculations for single RNA sequences.

Functions

• float cofold (const char *sequence, char *structure)

Compute the minimum free energy of two interacting RNA molecules.

• float cofold_par (const char *string, char *structure, vrna_param_t *parameters, int is_constrained)

Compute the minimum free energy of two interacting RNA molecules.

· void free_co_arrays (void)

Free memory occupied by cofold()

void update_cofold_params (void)

Recalculate parameters.

void update_cofold_params_par (vrna_param_t *parameters)

Recalculate parameters.

void export_cofold_arrays_gq (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **fc_p, int **ggg_p, int **indx_p, char **ptype_p)

Export the arrays of partition function cofold (with gquadruplex support)

void export_cofold_arrays (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **fc_p, int **indx_p, char
 **ptype_p)

Export the arrays of partition function cofold.

• void get_monomere_mfes (float *e1, float *e2)

get_monomer_free_energies

- void initialize_cofold (int length)
- float fold_par (const char *sequence, char *structure, vrna_param_t *parameters, int is_constrained, int is
 _circular)

Compute minimum free energy and an appropriate secondary structure of an RNA sequence.

float fold (const char *sequence, char *structure)

Compute minimum free energy and an appropriate secondary structure of an RNA sequence.

• float circfold (const char *sequence, char *structure)

Compute minimum free energy and an appropriate secondary structure of a circular RNA sequence.

void free_arrays (void)

Free arrays for mfe folding.

void update_fold_params (void)

Recalculate energy parameters.

void update fold params par (vrna param t *parameters)

Recalculate energy parameters.

- void export_fold_arrays (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p)
- void export_fold_arrays_par (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p, vrna_param_t **P_p)
- void export_circfold_arrays (int *Fc_p, int *FcH_p, int *FcM_p, int *FcM_p, int **fM2_p, int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p)
- void export_circfold_arrays_par (int *Fc_p, int *Fcl_p, int *Fcl_p, int *FcM_p, int **fM2_p, int **f5_p, int **e_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p, vrna_param_t **P_p)
- int LoopEnergy (int n1, int n2, int type, int type_2, int si1, int sj1, int sp1, int sq1)
- int HairpinE (int size, int type, int si1, int sj1, const char *string)
- · void initialize fold (int length)
- char * backtrack fold from pair (char *sequence, int i, int j)
- float alifold (const char **strings, char *structure)

Compute MFE and according consensus structure of an alignment of sequences.

float circalifold (const char **strings, char *structure)

Compute MFE and according structure of an alignment of sequences assuming the sequences are circular instead of linear

· void free alifold arrays (void)

Free the memory occupied by MFE alifold functions.

16.84.2 Function Documentation

16.84.2.1 alifold()

#include <ViennaRNA/alifold.h>

Compute MFE and according consensus structure of an alignment of sequences.

This function predicts the consensus structure for the aligned 'sequences' and returns the minimum free energy; the mfe structure in bracket notation is returned in 'structure'.

Sufficient space must be allocated for 'structure' before calling alifold().

Deprecated Usage of this function is discouraged! Use vrna_alifold(), or vrna_mfe() instead!

See also

```
vrna_alifold(), vrna_mfe()
```

Parameters

strings	A pointer to a NULL terminated array of character arrays	
structure	A pointer to a character array that may contain a constraining consensus structure (will be	
	overwritten by a consensus structure that exhibits the MFE)	

Returns

The free energy score in kcal/mol

16.84.2.2 cofold()

Compute the minimum free energy of two interacting RNA molecules.

The code is analog to the fold() function. If cut_point ==-1 results should be the same as with fold().

Deprecated use vrna_mfe_dimer() instead

Parameters

sequence	The two sequences concatenated
structure	Will hold the barcket dot structure of the dimer molecule

Returns

minimum free energy of the structure

16.84.2.3 cofold_par()

#include <ViennaRNA/cofold.h>

Compute the minimum free energy of two interacting RNA molecules.

Deprecated use vrna_mfe_dimer() instead

```
16.84.2.4 free_co_arrays()
```

Free memory occupied by cofold()

Deprecated This function will only free memory allocated by a prior call of cofold() or cofold_par(). See vrna_mfe_dimer() for how to use the new API

Note

folding matrices now reside in the fold compound, and should be free'd there

See also

```
vrna_fc_destroy(), vrna_mfe_dimer()
```

16.84.2.5 update_cofold_params()

Recalculate parameters.

Deprecated See vrna_params_subst() for an alternative using the new API

16.84.2.6 update_cofold_params_par()

Recalculate parameters.

Deprecated See vrna_params_subst() for an alternative using the new API

16.84.2.7 export_cofold_arrays_gq()

```
void export_cofold_arrays_gq (
    int ** f5_p,
    int ** c_p,
    int ** fML_p,
    int ** fMl_p,
    int ** fc_p,
    int ** fc_p,
    int ** ggg_p,
    int ** indx_p,
    char ** ptype_p )
#include <ViennaRNA/cofold.h>
```

Export the arrays of partition function cofold (with gquadruplex support)

Export the cofold arrays for use e.g. in the concentration Computations or suboptimal secondary structure backtracking

Deprecated folding matrices now reside within the fold compound. Thus, this function will only work in conjunction with a prior call to cofold() or cofold_par()

See also

vrna_mfe_dimer() for the new API

Parameters

f5_p	A pointer to the 'f5' array, i.e. array conatining best free energy in interval [1,j]
	1 1 2 2 2 2 2
<i>c_p</i>	A pointer to the 'c' array, i.e. array containing best free energy in interval [i,j] given that i pairs with j
fML_p	A pointer to the 'M' array, i.e. array containing best free energy in interval [i,j] for any multiloop segment with at least one stem
fM1_p	A pointer to the 'M1' array, i.e. array containing best free energy in interval [i,j] for multiloop segment with exactly one stem
fc_p	A pointer to the 'fc' array, i.e. array
ggg_p	A pointer to the 'ggg' array, i.e. array containing best free energy of a gquadruplex delimited by [i,j]
indx_p	A pointer to the indexing array used for accessing the energy matrices
ptype←	A pointer to the ptype array containing the base pair types for each possibility (i,j)
_p	

16.84.2.8 export_cofold_arrays()

Export the arrays of partition function cofold.

Export the cofold arrays for use e.g. in the concentration Computations or suboptimal secondary structure backtracking

Deprecated folding matrices now reside within the vrna_fold_compound_t. Thus, this function will only work in conjunction with a prior call to the deprecated functions cofold() or cofold_par()

See also

vrna_mfe_dimer() for the new API

Parameters

f5_p	A pointer to the 'f5' array, i.e. array conatining best free energy in interval [1,j]
<i>c_p</i>	A pointer to the 'c' array, i.e. array containing best free energy in interval [i,j] given that i pairs with j
fML_p	A pointer to the 'M' array, i.e. array containing best free energy in interval [i,j] for any multiloop segment with at least one stem
fM1_p	A pointer to the 'M1' array, i.e. array containing best free energy in interval [i,j] for multiloop segment with exactly one stem
fc_p	A pointer to the 'fc' array, i.e. array
indx_p	A pointer to the indexing array used for accessing the energy matrices
<i>ptype</i> ← _p	A pointer to the ptype array containing the base pair types for each possibility (i,j)

16.84.2.9 get_monomere_mfes()

Export monomer free energies out of cofold arrays

Deprecated {This function is obsolete and will be removed soon!}

Parameters

e1	A pointer to a variable where the energy of molecule A will be written to
e2	A pointer to a variable where the energy of molecule B will be written to

16.84.2.10 initialize_cofold()

allocate arrays for folding

Deprecated {This function is obsolete and will be removed soon!}

16.84.2.11 fold_par()

#include <ViennaRNA/fold.h>

Compute minimum free energy and an appropriate secondary structure of an RNA sequence.

The first parameter given, the RNA sequence, must be uppercase and should only contain an alphabet Σ that is understood by the RNAlib

```
(e.g. \Sigma = \{A, U, C, G\})
```

The second parameter, structure, must always point to an allocated block of memory with a size of at least strlen(sequence) + 1

If the third parameter is NULL, global model detail settings are assumed for the folding recursions. Otherwise, the provided parameters are used.

The fourth parameter indicates whether a secondary structure constraint in enhanced dot-bracket notation is passed through the structure parameter or not. If so, the characters " | x < >" are recognized to mark bases that are paired, unpaired, paired upstream, or downstream, respectively. Matching brackets " () " denote base pairs, dots "." are used for unconstrained bases.

To indicate that the RNA sequence is circular and thus has to be post-processed, set the last parameter to non-zero

After a successful call of fold_par(), a backtracked secondary structure (in dot-bracket notation) that exhibits the minimum of free energy will be written to the memory *structure* is pointing to. The function returns the minimum of free energy for any fold of the sequence given.

Note

OpenMP: Passing NULL to the 'parameters' argument involves access to several global model detail variables and thus is not to be considered threadsafe

Deprecated use vrna_mfe() instead!

See also

```
vrna_mfe(), fold(), circfold(), vrna_md_t, set_energy_model(), get_scaled_parameters()
```

Parameters

sequence	RNA sequence
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to
parameters	A data structure containing the pre-scaled energy contributions and the model details. (NULL may be passed, see OpenMP notes above)
is_constrained	Switch to indicate that a structure constraint is passed via the structure argument (0==off)
is_circular	Switch to (de-)activate post-processing steps in case RNA sequence is circular (0==off)

Returns

the minimum free energy (MFE) in kcal/mol

16.84.2.12 fold()

Compute minimum free energy and an appropriate secondary structure of an RNA sequence.

This function essentially does the same thing as fold_par(). However, it takes its model details, i.e. temperature, dangles, tetra_loop, noGU, no_closingGU, fold_constrained, noLonelyPairs from the current global settings within the library

Deprecated use vrna_fold(), or vrna_mfe() instead!

See also

fold_par(), circfold()

Parameters

sequence	RNA sequence
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to

Returns

the minimum free energy (MFE) in kcal/mol

16.84.2.13 circfold()

Compute minimum free energy and an appropriate secondary structure of a circular RNA sequence.

This function essentially does the same thing as fold_par(). However, it takes its model details, i.e. temperature, dangles, tetra_loop, noGU, no_closingGU, fold_constrained, noLonelyPairs from the current global settings within the library

Deprecated Use vrna_circfold(), or vrna_mfe() instead!

See also

```
fold_par(), circfold()
```

Parameters

sequence	RNA sequence
structure	A pointer to the character array where the secondary structure in dot-bracket notation will be written to

Returns

the minimum free energy (MFE) in kcal/mol

16.84.2.14 free_arrays()

Free arrays for mfe folding.

Deprecated See vrna_fold(), vrna_circfold(), or vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

```
16.84.2.15 update_fold_params()
```

Recalculate energy parameters.

Deprecated For non-default model settings use the new API with vrna_params_subst() and vrna_mfe() instead!

```
16.84.2.16 update_fold_params_par()
```

Recalculate energy parameters.

Deprecated For non-default model settings use the new API with vrna_params_subst() and vrna_mfe() instead!

16.84.2.17 export_fold_arrays()

Deprecated See vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

16.84.2.18 export_fold_arrays_par()

Deprecated See vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

16.84.2.19 export_circfold_arrays()

#Include <ViennaRNA/101d.n>

Deprecated See vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

16.84.2.20 export_circfold_arrays_par()

Deprecated See vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

16.84.2.21 LoopEnergy()

Deprecated {This function is deprecated and will be removed soon. Use E_IntLoop() instead!}

16.84.2.22 HairpinE()

Deprecated {This function is deprecated and will be removed soon. Use E_Hairpin() instead!}

16.84.2.23 initialize_fold()

Allocate arrays for folding

Deprecated See vrna_mfe() and vrna_fold_compound_t for the usage of the new API!

16.84.2.24 backtrack_fold_from_pair()

#include <ViennaRNA/fold.h>

16.84.2.25 circalifold()

#include <ViennaRNA/alifold.h>

Compute MFE and according structure of an alignment of sequences assuming the sequences are circular instead of linear.

Deprecated Usage of this function is discouraged! Use vrna_alicircfold(), and vrna_mfe() instead!

See also

vrna_alicircfold(), vrna_alifold(), vrna_mfe()

Parameters

strings	A pointer to a NULL terminated array of character arrays	
structure	A pointer to a character array that may contain a constraining consensus structure (will be	
	overwritten by a consensus structure that exhibits the MFE)	

Returns

The free energy score in kcal/mol

16.84.2.26 free_alifold_arrays()

Free the memory occupied by MFE alifold functions.

Deprecated Usage of this function is discouraged! It only affects memory being free'd that was allocated by an old API function before. Release of memory occupied by the newly introduced vrna_fold_compound_t is handled by vrna_fold_compound_free()

See also

vrna_fold_compound_free()

16.85 Deprecated Interface for Local (Sliding Window) MFE Prediction

16.85.1 Detailed Description

Collaboration diagram for Deprecated Interface for Local (Sliding Window) MFE Prediction:

Files

· file Lfold.h

Functions for locally optimal MFE structure prediction.

Functions

- float Lfold (const char *string, const char *structure, int maxdist)
 The local analog to fold().
- float Lfoldz (const char *string, const char *structure, int maxdist, int zsc, double min_z)

16.85.2 Function Documentation

16.85.2.1 Lfold()

The local analog to fold().

Computes the minimum free energy structure including only base pairs with a span smaller than 'maxdist'

Deprecated Use vrna_mfe_window() instead!

16.85.2.2 Lfoldz()

Deprecated Use vrna_mfe_window_zscore() instead!

16.86 Deprecated Interface for Global Partition Function Computation

16.86.1 Detailed Description

Collaboration diagram for Deprecated Interface for Global Partition Function Computation:

Files

· file part func co.h

Partition function for two RNA sequences.

Functions

• float pf_fold_par (const char *sequence, char *structure, vrna_exp_param_t *parameters, int calculate_← bppm, int is_constrained, int is_circular)

Compute the partition function Q for a given RNA sequence.

float pf fold (const char *sequence, char *structure)

Compute the partition function Q of an RNA sequence.

float pf circ fold (const char *sequence, char *structure)

Compute the partition function of a circular RNA sequence.

void free_pf_arrays (void)

Free arrays for the partition function recursions.

void update_pf_params (int length)

Recalculate energy parameters.

void update_pf_params_par (int length, vrna_exp_param_t *parameters)

Recalculate energy parameters.

FLT_OR_DBL * export_bppm (void)

Get a pointer to the base pair probability array.

int get_pf_arrays (short **S_p, short **S1_p, char **ptype_p, FLT_OR_DBL **qb_p, FLT_OR_DBL **qth_p, FLT_OR_DBL **qth_p)

Get the pointers to (almost) all relavant computation arrays used in partition function computation.

• double get_subseq_F (int i, int j)

Get the free energy of a subsequence from the q[] array.

• double mean_bp_distance (int length)

Get the mean base pair distance of the last partition function computation.

double mean_bp_distance_pr (int length, FLT_OR_DBL *pr)

Get the mean base pair distance in the thermodynamic ensemble.

vrna_ep_t * stackProb (double cutoff)

Get the probability of stacks.

void init_pf_fold (int length)

Allocate space for pf_fold()

• vrna_dimer_pf_t co_pf_fold (char *sequence, char *structure)

Calculate partition function and base pair probabilities.

vrna_dimer_pf_t co_pf_fold_par (char *sequence, char *structure, vrna_exp_param_t *parameters, int calculate_bppm, int is_constrained)

Calculate partition function and base pair probabilities.

• void compute_probabilities (double FAB, double FEA, double FEB, vrna_ep_t *prAB, vrna_ep_t *prA, vrna_ep_t *prB, int Alength)

Compute Boltzmann probabilities of dimerization without homodimers.

```
    void init_co_pf_fold (int length)
```

FLT_OR_DBL * export_co_bppm (void)

Get a pointer to the base pair probability array.

· void free co pf arrays (void)

Free the memory occupied by co_pf_fold()

void update_co_pf_params (int length)

Recalculate energy parameters.

void update_co_pf_params_par (int length, vrna_exp_param_t *parameters)

Recalculate energy parameters.

void assign_plist_from_db (vrna_ep_t **pl, const char *struc, float pr)

Create a vrna_ep_t from a dot-bracket string.

void assign_plist_from_pr (vrna_ep_t **pl, FLT_OR_DBL *probs, int length, double cutoff)

Create a vrna_ep_t from a probability matrix.

- float alipf_fold_par (const char **sequences, char *structure, vrna_ep_t **pl, vrna_exp_param_t *parameters, int calculate_bppm, int is_constrained, int is_circular)
- float alipf fold (const char **sequences, char *structure, vrna ep t **pl)

The partition function version of alifold() works in analogy to $pf_fold()$. Pair probabilities and information about sequence covariations are returned via the 'pi' variable as a list of $vrna_pinfo_t$ structs. The list is terminated by the first entry with pi.i = 0.

- float alipf_circ_fold (const char **sequences, char *structure, vrna_ep_t **pl)
- FLT_OR_DBL * export_ali_bppm (void)

Get a pointer to the base pair probability array.

void free_alipf_arrays (void)

Free the memory occupied by folding matrices allocated by alipf_fold, alipf_circ_fold, etc.

char * alipbacktrack (double *prob)

Sample a consensus secondary structure from the Boltzmann ensemble according its probability.

int get_alipf_arrays (short ***\$S_p, short ***\$S_p, short ***\$S_p, unsigned short ***a2s_p, char ***\$S ←
 _p, FLT_OR_DBL **qb_p, FLT_OR_DBL **qn_p, FLT_OR_DBL **q1k_p, FLT_OR_DBL **qln_p, short
 **pscore)

Get pointers to (almost) all relavant arrays used in alifold's partition function computation.

16.86.2 Function Documentation

16.86.2.1 alipf_fold_par()

#include <ViennaRNA/alifold.h>

Deprecated Use vrna_pf() instead

Parameters

sequences	
structure	
pl	
parameters	
calculate_bppm	
is_constrained	
is_circular	

Returns

16.86.2.2 pf_fold_par()

Compute the partition function ${\cal Q}$ for a given RNA sequence.

If structure is not a NULL pointer on input, it contains on return a string consisting of the letters " . , | { } () " denoting bases that are essentially unpaired, weakly paired, strongly paired without preference, weakly upstream (downstream) paired, or strongly up- (down-)stream paired bases, respectively. If fold_constrained is not 0, the structure string is interpreted on input as a list of constraints for the folding. The character "x" marks bases that must be unpaired, matching brackets " () " denote base pairs, all other characters are ignored. Any pairs conflicting with the constraint will be forbidden. This is usually sufficient to ensure the constraints are honored. If the parameter calculate_bppm is set to 0 base pairing probabilities will not be computed (saving CPU time), otherwise after calculations took place pr will contain the probability that bases i and j pair.

Deprecated Use vrna_pf() instead

Note

The global array pr is deprecated and the user who wants the calculated base pair probabilities for further computations is advised to use the function export_bppm()

Postcondition

After successful run the hidden folding matrices are filled with the appropriate Boltzmann factors. Depending on whether the global variable do_backtrack was set the base pair probabilities are already computed and may be accessed for further usage via the export_bppm() function. A call of free_pf_arrays() will free all memory allocated by this function. Successive calls will first free previously allocated memory before starting the computation.

See also

```
vrna_pf(), bppm_to_structure(), export_bppm(), vrna_exp_params(), free_pf_arrays()
```

Parameters

in	sequence	The RNA sequence input
in,out	structure	A pointer to a char array where a base pair probability information can be stored in a pseudo-dot-bracket notation (may be NULL, too)
in	parameters	Data structure containing the precalculated Boltzmann factors
in	calculate_bppm	Switch to Base pair probability calculations on/off (0==off)
in	is_constrained	Switch to indicate that a structure contraint is passed via the structure argument (0==off)
in	is_circular	Switch to (de-)activate postprocessing steps in case RNA sequence is circular (0==off)

Returns

The ensemble free energy $G = -RT \cdot \log(Q)$ in kcal/mol

16.86.2.3 pf_fold()

Compute the partition function Q of an RNA sequence.

If structure is not a NULL pointer on input, it contains on return a string consisting of the letters " . , | { } () " denoting bases that are essentially unpaired, weakly paired, strongly paired without preference, weakly upstream (downstream) paired, or strongly up- (down-)stream paired bases, respectively. If fold_constrained is not 0, the structure string is interpreted on input as a list of constraints for the folding. The character "x" marks bases that must be unpaired, matching brackets " () " denote base pairs, all other characters are ignored. Any pairs conflicting with the constraint will be forbidden. This is usually sufficient to ensure the constraints are honored. If do_backtrack has been set to 0 base pairing probabilities will not be computed (saving CPU time), otherwise pr will contain the probability that bases i and j pair.

Note

The global array pr is deprecated and the user who wants the calculated base pair probabilities for further computations is advised to use the function export_bppm().

OpenMP: This function is not entirely threadsafe. While the recursions are working on their own copies of data the model details for the recursions are determined from the global settings just before entering the recursions. Consider using pf_fold_par() for a really threadsafe implementation.

Precondition

This function takes its model details from the global variables provided in RNAlib

Postcondition

After successful run the hidden folding matrices are filled with the appropriate Boltzmann factors. Depending on whether the global variable do_backtrack was set the base pair probabilities are already computed and may be accessed for further usage via the export_bppm() function. A call of free_pf_arrays() will free all memory allocated by this function. Successive calls will first free previously allocated memory before starting the computation.

See also

```
pf_fold_par(), pf_circ_fold(), bppm_to_structure(), export_bppm()
```

Parameters

sequence	The RNA sequence input
structure	A pointer to a char array where a base pair probability information can be stored in a pseudo-dot-bracket notation (may be NULL, too)

Returns

The ensemble free energy $G = -RT \cdot \log(Q)$ in kcal/mol

16.86.2.4 pf_circ_fold()

Compute the partition function of a circular RNA sequence.

Note

The global array pr is deprecated and the user who wants the calculated base pair probabilities for further computations is advised to use the function export bppm().

OpenMP: This function is not entirely threadsafe. While the recursions are working on their own copies of data the model details for the recursions are determined from the global settings just before entering the recursions. Consider using pf_fold_par() for a really threadsafe implementation.

Precondition

This function takes its model details from the global variables provided in RNAlib

Postcondition

After successful run the hidden folding matrices are filled with the appropriate Boltzmann factors. Depending on whether the global variable do_backtrack was set the base pair probabilities are already computed and may be accessed for further usage via the export_bppm() function. A call of free_pf_arrays() will free all memory allocated by this function. Successive calls will first free previously allocated memory before starting the computation.

See also

vrna_pf()

Deprecated Use vrna_pf() instead!

Parameters

in	sequence	The RNA sequence input
in,out	structure	A pointer to a char array where a base pair probability information can be stored in a
		pseudo-dot-bracket notation (may be NULL, too)

Returns

The ensemble free energy $G = -RT \cdot \log(Q)$ in kcal/mol

16.86.2.5 free_pf_arrays()

Free arrays for the partition function recursions.

Call this function if you want to free all allocated memory associated with the partition function forward recursion.

Note

Successive calls of pf_fold(), pf_circ_fold() already check if they should free any memory from a previous run. **OpenMP notice:**

This function should be called before leaving a thread in order to avoid leaking memory

Deprecated See vrna_fold_compound_t and its related functions for how to free memory occupied by the dynamic programming matrices

Postcondition

All memory allocated by pf_fold_par(), pf_fold() or pf_circ_fold() will be free'd

See also

```
pf\_fold\_par(),\,pf\_fold(),\,pf\_circ\_fold()
```

16.86.2.6 update_pf_params()

Recalculate energy parameters.

Call this function to recalculate the pair matrix and energy parameters after a change in folding parameters like temperature

Deprecated Use vrna_exp_params_subst() instead

```
16.86.2.7 update_pf_params_par()
```

Recalculate energy parameters.

Deprecated Use vrna_exp_params_subst() instead

16.86.2.8 export_bppm()

Get a pointer to the base pair probability array.

Accessing the base pair probabilities for a pair (i,j) is achieved by

```
FLT_OR_DBL *pr = export_bppm();
pr_ij = pr[iindx[i]-j];
```

Precondition

Call pf fold par(), pf fold() or pf circ fold() first to fill the base pair probability array

See also

```
pf_fold(), pf_circ_fold(), vrna_idx_row_wise()
```

Returns

A pointer to the base pair probability array

16.86.2.9 get_pf_arrays()

Get the pointers to (almost) all relavant computation arrays used in partition function computation.

Precondition

In order to assign meaningful pointers, you have to call pf fold par() or pf fold() first!

See also

```
pf_fold_par(), pf_fold(), pf_circ_fold()
```

Parameters

out	S_p	A pointer to the 'S' array (integer representation of nucleotides)
out	S1_p	A pointer to the 'S1' array (2nd integer representation of nucleotides)
out	ptype⇔	A pointer to the pair type matrix
	_p	
out	qb_p	A pointer to the Q ^B matrix
out	qm_p	A pointer to the Q ^M matrix
out	q1k_p	A pointer to the 5' slice of the Q matrix ($q1k(k)=Q(1,k)$)
out	qln_p	A pointer to the 3' slice of the Q matrix ($qln(l)=Q(l,n)$)

Returns

Non Zero if everything went fine, 0 otherwise

```
16.86.2.10 mean_bp_distance()
```

Get the mean base pair distance of the last partition function computation.

Deprecated Use vrna_mean_bp_distance() or vrna_mean_bp_distance_pr() instead!

See also

```
vrna_mean_bp_distance(), vrna_mean_bp_distance_pr()
```

Parameters

length

Returns

mean base pair distance in thermodynamic ensemble

16.86.2.11 mean_bp_distance_pr()

```
#include <ViennaRNA/part_func.h>
```

Get the mean base pair distance in the thermodynamic ensemble.

This is a threadsafe implementation of mean_bp_dist()!

```
\begin{array}{l} < d> = \sum_{a,b} p_a p_b d(S_a,S_b) \\ \text{this can be computed from the pair probs } p_i j \text{ as} \\ < d> = \sum_{ij} p_{ij} (1-p_{ij}) \end{array}
```

Deprecated Use vrna_mean_bp_distance() or vrna_mean_bp_distance_pr() instead!

Parameters

length	The length of the sequence
pr	The matrix containing the base pair probabilities

Returns

The mean pair distance of the structure ensemble

16.86.2.12 stackProb()

Get the probability of stacks.

Deprecated Use vrna_stack_prob() instead!

```
16.86.2.13 init_pf_fold()
```

Allocate space for pf_fold()

Deprecated This function is obsolete and will be removed soon!

16.86.2.14 co_pf_fold()

Calculate partition function and base pair probabilities.

This is the cofold partition function folding. The second molecule starts at the cut_point nucleotide.

Note

OpenMP: Since this function relies on the global parameters do_backtrack, dangles, temperature and pf_scale it is not threadsafe according to concurrent changes in these variables! Use co_pf_fold_par() instead to circumvent this issue.

Deprecated {Use vrna_pf_dimer() instead!}

Parameters

sequence	Concatenated RNA sequences
structure	Will hold the structure or constraints

Returns

vrna dimer pf t structure containing a set of energies needed for concentration computations.

16.86.2.15 co_pf_fold_par()

Calculate partition function and base pair probabilities.

This is the cofold partition function folding. The second molecule starts at the cut_point nucleotide.

Deprecated Use vrna_pf_dimer() instead!

See also

```
get_boltzmann_factors(), co_pf_fold()
```

Parameters

sequence	Concatenated RNA sequences
structure	Pointer to the structure constraint
parameters	Data structure containing the precalculated Boltzmann factors
calculate_bppm	Switch to turn Base pair probability calculations on/off (0==off)
is_constrained	Switch to indicate that a structure contraint is passed via the structure argument (0==off)

Returns

vrna_dimer_pf_t structure containing a set of energies needed for concentration computations.

16.86.2.16 compute_probabilities()

Compute Boltzmann probabilities of dimerization without homodimers.

Given the pair probabilities and free energies (in the null model) for a dimer AB and the two constituent monomers A and B, compute the conditional pair probabilities given that a dimer AB actually forms. Null model pair probabilities are given as a list as produced by assign_plist_from_pr("), the dimer probabilities 'prAB' are modified in place.

Deprecated { Use vrna_pf_dimer_probs() instead!}

Parameters

FAB	free energy of dimer AB
FEA	free energy of monomer A
FEB	free energy of monomer B
prAB	pair probabilities for dimer
prA	pair probabilities monomer
prB	pair probabilities monomer
Alength	Length of molecule A

```
16.86.2.17 init_co_pf_fold()
```

DO NOT USE THIS FUNCTION ANYMORE

Deprecated { This function is deprecated and will be removed soon!}

16.86.2.18 export_co_bppm()

Get a pointer to the base pair probability array.

Accessing the base pair probabilities for a pair (i,j) is achieved by

```
FLT_OR_DBL *pr = export_bppm(); pr_ij = pr[iindx[i]-j];
```

Deprecated This function is deprecated and will be removed soon! The base pair probability array is available through the vrna_fold_compound_t data structure, and its associated vrna_mx_pf_t member.

See also

```
vrna_idx_row_wise()
```

Returns

A pointer to the base pair probability array

16.86.2.19 free co pf arrays()

Free the memory occupied by co_pf_fold()

Deprecated This function will be removed for the new API soon! See vrna_pf_dimer(), vrna_fold_compound(), and vrna_fold_compound_free() for an alternative

16.86.2.20 update_co_pf_params()

Recalculate energy parameters.

This function recalculates all energy parameters given the current model settings.

Deprecated Use vrna_exp_params_subst() instead!

Parameters

length Length of the current RNA sequence

16.86.2.21 update_co_pf_params_par()

Recalculate energy parameters.

This function recalculates all energy parameters given the current model settings. It's second argument can either be NULL or a data structure containing the precomputed Boltzmann factors. In the first scenario, the necessary data structure will be created automatically according to the current global model settings, i.e. this mode might not be threadsafe. However, if the provided data structure is not NULL, threadsafety for the model parameters dangles, pf_scale and temperature is regained, since their values are taken from this data structure during subsequent calculations.

Deprecated Use vrna_exp_params_subst() instead!

Parameters

length	Length of the current RNA sequence
parameters	data structure containing the precomputed Boltzmann factors

16.86.2.22 assign_plist_from_db()

Create a vrna_ep_t from a dot-bracket string.

The dot-bracket string is parsed and for each base pair an entry in the plist is created. The probability of each pair in the list is set by a function parameter.

The end of the plist is marked by sequence positions i as well as j equal to 0. This condition should be used to stop looping over its entries

Deprecated Use vrna_plist() instead

Parameters

pl	A pointer to the vrna_ep_t that is to be created
struc	The secondary structure in dot-bracket notation
pr	The probability for each base pair

16.86.2.23 assign_plist_from_pr()

#include <ViennaRNA/utils/structures.h>

Create a vrna_ep_t from a probability matrix.

The probability matrix given is parsed and all pair probabilities above the given threshold are used to create an entry in the plist

The end of the plist is marked by sequence positions i as well as j equal to 0. This condition should be used to stop looping over its entries

Note

This function is threadsafe

Deprecated Use vrna_plist_from_probs() instead!

Parameters

out	pl	A pointer to the vrna_ep_t that is to be created
in	probs	The probability matrix used for creating the plist
in	n length The length of the RNA sequence	
in	cutoff	The cutoff value

16.86.2.24 alipf_fold()

#include <ViennaRNA/alifold.h>

The partition function version of alifold() works in analogy to $pf_fold()$. Pair probabilities and information about sequence covariations are returned via the 'pi' variable as a list of $vrna_pinfo_t$ structs. The list is terminated by the first entry with pi.i = 0.

Deprecated Use vrna_pf() instead

Parameters

sequences	
structure	
pl	

Returns

16.86.2.25 alipf_circ_fold()

#include <ViennaRNA/alifold.h>

Deprecated Use vrna_pf() instead

Parameters

sequences	
structure	
pl	

Returns

16.86.2.26 export_ali_bppm()

Get a pointer to the base pair probability array.

Accessing the base pair probabilities for a pair (i,j) is achieved by

```
FLT_OR_DBL *pr = export_bppm(); pr_ij = pr[iindx[i]-j];
```

Deprecated Usage of this function is discouraged! The new vrna_fold_compound_t allows direct access to the folding matrices, including the pair probabilities! The pair probability array returned here reflects the one of the latest call to vrna_pf(), or any of the old API calls for consensus structure partition function folding.

See also

```
vrna_fold_compound_t, vrna_fold_compound_comparative(), and vrna_pf()
```

Returns

A pointer to the base pair probability array

```
16.86.2.27 free_alipf_arrays()
```

Free the memory occupied by folding matrices allocated by alipf_fold, alipf_circ_fold, etc.

Deprecated Usage of this function is discouraged! This function only free's memory allocated by old API function calls. Memory allocated by any of the new API calls (starting with vrna_) will be not affected!

See also

```
vrna_fold_compound_t, vrna_vrna_fold_compound_free()
```

16.86.2.28 alipbacktrack()

Sample a consensus secondary structure from the Boltzmann ensemble according its probability.

Deprecated Use vrna_pbacktrack() instead!

Parameters

prob	to be described (berni)
------	-------------------------

Returns

A sampled consensus secondary structure in dot-bracket notation

16.86.2.29 get_alipf_arrays()

Get pointers to (almost) all relavant arrays used in alifold's partition function computation.

Note

To obtain meaningful pointers, call alipf_fold first!

See also

```
pf_alifold(), alipf_circ_fold()
```

Deprecated It is discouraged to use this function! The new vrna_fold_compound_t allows direct access to all necessary consensus structure prediction related variables!

See also

```
vrna_fold_compound_t, vrna_fold_compound_comparative(), vrna_pf()
```

Parameters

S_p	A pointer to the 'S' array (integer representation of nucleotides)
S5_p	A pointer to the 'S5' array
S3_p	A pointer to the 'S3' array
a2s⇔	A pointer to the alignment-column to sequence position mapping array
_p	
_Ss_p	A pointer to the 'Ss' array
Generated by Pax 1966 Inter to the QB matrix	
qm_p	A pointer to the Q ^M matrix
q1k↔	A pointer to the 5' slice of the Q matrix ($q1k(k)=Q(1,k)$)
n	

Returns

Non Zero if everything went fine, 0 otherwise

16.87 Deprecated Interface for Local (Sliding Window) Partition Function Computation

16.87.1 Detailed Description

Collaboration diagram for Deprecated Interface for Local (Sliding Window) Partition Function Computation:

Files

• file LPfold.h

Partition function and equilibrium probability implementation for the sliding window algorithm.

Functions

- void update_pf_paramsLP (int length)
- vrna_ep_t * pfl_fold (char *sequence, int winSize, int pairSize, float cutoffb, double **pU, vrna_ep_t **dpp2,
 FILE *pUfp, FILE *spup)

Compute partition functions for locally stable secondary structures.

vrna_ep_t * pfl_fold_par (char *sequence, int winSize, int pairSize, float cutoffb, double **pU, vrna_ep_t
 **dpp2, FILE *pUfp, FILE *spup, vrna_exp_param_t *parameters)

Compute partition functions for locally stable secondary structures.

• void putoutpU_prob (double **pU, int length, int ulength, FILE *fp, int energies)

Writes the unpaired probabilities (pU) or opening energies into a file.

void putoutpU_prob_bin (double **pU, int length, int ulength, FILE *fp, int energies)

Writes the unpaired probabilities (pU) or opening energies into a binary file.

16.87.2 Function Documentation

16.87.2.1 update_pf_paramsLP()

Parameters

length

16.87.2.2 pfl_fold()

```
int winSize,
int pairSize,
float cutoffb,
double ** pU,
vrna_ep_t ** dpp2,
FILE * pUfp,
FILE * spup )
```

#include <ViennaRNA/LPfold.h>

Compute partition functions for locally stable secondary structures.

pfl_fold computes partition functions for every window of size 'winSize' possible in a RNA molecule, allowing only pairs with a span smaller than 'pairSize'. It returns the mean pair probabilities averaged over all windows containing the pair in 'pl'. 'winSize' should always be >= 'pairSize'. Note that in contrast to Lfold(), bases outside of the window do not influence the structure at all. Only probabilities higher than 'cutoffb' are kept.

If 'pU' is supplied (i.e is not the NULL pointer), pfl_fold() will also compute the mean probability that regions of length 'u' and smaller are unpaired. The parameter 'u' is supplied in 'pup[0][0]'. On return the 'pup' array will contain these probabilities, with the entry on 'pup[x][y]' containing the mean probability that x and the y-1 preceding bases are unpaired. The 'pU' array needs to be large enough to hold n+1 float* entries, where n is the sequence length.

If an array dpp2 is supplied, the probability of base pair (i,j) given that there already exists a base pair (i+1,j-1) is also computed and saved in this array. If pUfp is given (i.e. not NULL), pU is not saved but put out imediately. If spup is given (i.e. is not NULL), the pair probabilities in pl are not saved but put out imediately.

Parameters

sequence	RNA sequence
winSize	size of the window
pairSize	maximum size of base pair
cutoffb	cutoffb for base pairs
pU	array holding all unpaired probabilities
dpp2	array of dependent pair probabilities
pUfp	file pointer for pU
spup	file pointer for pair probabilities

Returns

list of pair probabilities

16.87.2.3 putoutpU_prob()

#include <ViennaRNA/LPfold.h>

Writes the unpaired probabilities (pU) or opening energies into a file.

Can write either the unpaired probabilities (accessibilities) pU or the opening energies -log(pU)kT into a file

Parameters

рU	pair probabilities
length	length of RNA sequence
ulength	maximum length of unpaired stretch
fp	file pointer of destination file
energies	switch to put out as opening energies

16.87.2.4 putoutpU_prob_bin()

#include <ViennaRNA/LPfold.h>

Writes the unpaired probabilities (pU) or opening energies into a binary file.

Can write either the unpaired probabilities (accessibilities) pU or the opening energies -log(pU)kT into a file

Parameters

рU	pair probabilities
length	length of RNA sequence
ulength	maximum length of unpaired stretch
fp	file pointer of destination file
energies	switch to put out as opening energies

16.88 Deprecated Interface for Stochastic Backtracking

16.88.1 Detailed Description

Collaboration diagram for Deprecated Interface for Stochastic Backtracking:

Functions

• char * pbacktrack (char *sequence)

Sample a secondary structure from the Boltzmann ensemble according its probability.

char * pbacktrack5 (char *sequence, int length)

Sample a sub-structure from the Boltzmann ensemble according its probability.

• char * pbacktrack_circ (char *sequence)

Sample a secondary structure of a circular RNA from the Boltzmann ensemble according its probability.

Variables

· int st back

Flag indicating that auxilary arrays are needed throughout the computations. This is essential for stochastic back-tracking.

16.88.2 Function Documentation

16.88.2.1 pbacktrack()

Sample a secondary structure from the Boltzmann ensemble according its probability.

Precondition

```
st_back has to be set to 1 before calling pf_fold() or pf_fold_par()
pf_fold_par() or pf_fold() have to be called first to fill the partition function matrices
```

Parameters

```
sequence The RNA sequence
```

Returns

A sampled secondary structure in dot-bracket notation

16.88.2.2 pbacktrack_circ()

Sample a secondary structure of a circular RNA from the Boltzmann ensemble according its probability.

This function does the same as pbacktrack() but assumes the RNA molecule to be circular

Precondition

```
st_back has to be set to 1 before calling pf_fold() or pf_fold_par() pf_fold_par() or pf_circ_fold() have to be called first to fill the partition function matrices
```

Deprecated Use vrna_pbacktrack() instead.

Parameters

sequence	The RNA sequence
----------	------------------

Returns

A sampled secondary structure in dot-bracket notation

16.88.3 Variable Documentation

16.88.3.1 st_back

```
int st_back
#include <ViennaRNA/part_func.h>
```

Flag indicating that auxiliary arrays are needed throughout the computations. This is essential for stochastic backtracking.

Set this variable to 1 prior to a call of pf_fold() to ensure that all matrices needed for stochastic backtracking are filled in the forward recursions

Deprecated set the *uniq_ML* flag in vrna_md_t before passing it to vrna_fold_compound().

See also

```
pbacktrack(), pbacktrack_circ
```

16.89 Deprecated Interface for Multiple Sequence Alignment Utilities

16.89.1 Detailed Description

Collaboration diagram for Deprecated Interface for Multiple Sequence Alignment Utilities:

Typedefs

• typedef struct vrna_pinfo_s pair_info

Old typename of vrna_pinfo_s.

Functions

• int get_mpi (char *Alseq[], int n_seq, int length, int *mini)

Get the mean pairwise identity in steps from ?to?(ident)

• void encode_ali_sequence (const char *sequence, short *S, short *s5, short *s3, char *ss, unsigned short *as, int circ)

Get arrays with encoded sequence of the alignment.

• void alloc_sequence_arrays (const char **sequences, short ***S, short ***S, short ***S, unsigned short ***a2s, char ***Ss, int circ)

Allocate memory for sequence array used to deal with aligned sequences.

• void free_sequence_arrays (unsigned int n_seq, short ***S, short ***S, short ***S, unsigned short ***a2s, char ***Ss)

Free the memory of the sequence arrays used to deal with aligned sequences.

16.89.2 Typedef Documentation

```
16.89.2.1 pair_info

typedef struct vrna_pinfo_s pair_info
#include <ViennaRNA/utils/alignments.h>
Old typename of vrna_pinfo_s.
```

Deprecated Use vrna_pinfo_t instead!

16.89.3 Function Documentation

Get the mean pairwise identity in steps from ?to?(ident)

Deprecated Use vrna_aln_mpi() as a replacement

Parameters

Alseq	
n_seq	The number of sequences in the alignment
length	The length of the alignment
mini	

Returns

The mean pairwise identity

16.89.3.2 encode_ali_sequence()

#include <ViennaRNA/utils/alignments.h>

Get arrays with encoded sequence of the alignment.

this function assumes that in S, S5, s3, ss and as enough space is already allocated (size must be at least sequence length+2)

Parameters

sequence	The gapped sequence from the alignment
S	pointer to an array that holds encoded sequence
s5	pointer to an array that holds the next base 5' of alignment position i
s3	pointer to an array that holds the next base 3' of alignment position i
ss	
as	
circ	assume the molecules to be circular instead of linear (circ=0)

16.89.3.3 alloc_sequence_arrays()

```
short *** S3,
unsigned short *** a2s,
char *** Ss,
int circ )
```

#include <ViennaRNA/utils/alignments.h>

Allocate memory for sequence array used to deal with aligned sequences.

Note that these arrays will also be initialized according to the sequence alignment given

See also

free_sequence_arrays()

Parameters

sequences	The aligned sequences
S	A pointer to the array of encoded sequences
S5	A pointer to the array that contains the next 5' nucleotide of a sequence position
S3	A pointer to the array that contains the next 3' nucleotide of a sequence position
a2s	A pointer to the array that contains the alignment to sequence position mapping
Ss	A pointer to the array that contains the ungapped sequence
circ	assume the molecules to be circular instead of linear (circ=0)

16.89.3.4 free_sequence_arrays()

```
void free_sequence_arrays (
    unsigned int n_seq,
    short *** S,
    short *** S5,
    short *** S3,
    unsigned short *** a2s,
    char *** Ss )
```

#include <ViennaRNA/utils/alignments.h>

Free the memory of the sequence arrays used to deal with aligned sequences.

This function frees the memory previously allocated with alloc_sequence_arrays()

See also

```
alloc_sequence_arrays()
```

Parameters

n_seq	The number of aligned sequences
S	A pointer to the array of encoded sequences
S5	A pointer to the array that contains the next 5' nucleotide of a sequence position
S3	A pointer to the array that contains the payt 2' publication of a acquence position
33	A pointer to the array that contains the next 3' nucleotide of a sequence position
a2s	A pointer to the array that contains the alignment to sequence position mapping
Ss	A pointer to the array that contains the ungapped sequence

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16.90 Deprecated Interface for Secondary Structure Utilities

16.90.1 Detailed Description

Collaboration diagram for Deprecated Interface for Secondary Structure Utilities:

Files

· file RNAstruct.h

Parsing and Coarse Graining of Structures.

Functions

char * b2HIT (const char *structure)

Converts the full structure from bracket notation to the HIT notation including root.

char * b2C (const char *structure)

Converts the full structure from bracket notation to the a coarse grained notation using the 'H' 'B' 'I' 'M' and 'R' identifiers.

char * b2Shapiro (const char *structure)

Converts the full structure from bracket notation to the weighted coarse grained notation using the 'H' 'B' 'I' 'M' 'S' 'E' and 'R' identifiers.

char * add root (const char *structure)

Adds a root to an un-rooted tree in any except bracket notation.

char * expand Shapiro (const char *coarse)

Inserts missing 'S' identifiers in unweighted coarse grained structures as obtained from b2C().

char * expand_Full (const char *structure)

Convert the full structure from bracket notation to the expanded notation including root.

char * unexpand Full (const char *ffull)

Restores the bracket notation from an expanded full or HIT tree, that is any tree using only identifiers 'U' 'P' and 'R'.

• char * unweight (const char *wcoarse)

Strip weights from any weighted tree.

void unexpand_aligned_F (char *align[2])

Converts two aligned structures in expanded notation.

void parse_structure (const char *structure)

Collects a statistic of structure elements of the full structure in bracket notation.

char * pack_structure (const char *struc)

Pack secondary secondary structure, 5:1 compression using base 3 encoding.

char * unpack structure (const char *packed)

Unpack secondary structure previously packed with pack_structure()

short * make_pair_table (const char *structure)

Create a pair table of a secondary structure.

short * copy_pair_table (const short *pt)

Get an exact copy of a pair table.

- short * alimake pair table (const char *structure)
- short * make_pair_table_snoop (const char *structure)
- int bp distance (const char *str1, const char *str2)

Compute the "base pair" distance between two secondary structures s1 and s2.

unsigned int * make_referenceBP_array (short *reference_pt, unsigned int turn)

Make a reference base pair count matrix.

• unsigned int * compute_BPdifferences (short *pt1, short *pt2, unsigned int turn)

Make a reference base pair distance matrix.

void parenthesis_structure (char *structure, vrna_bp_stack_t *bp, int length)

Create a dot-backet/parenthesis structure from backtracking stack.

void parenthesis_zuker (char *structure, vrna_bp_stack_t *bp, int length)

Create a dot-backet/parenthesis structure from backtracking stack obtained by zuker suboptimal calculation in cofold.c.

• void bppm_to_structure (char *structure, FLT_OR_DBL *pr, unsigned int length)

Create a dot-bracket like structure string from base pair probability matrix.

char bppm_symbol (const float *x)

Get a pseudo dot bracket notation for a given probability information.

Variables

• int loop size [2000]

contains a list of all loop sizes. loop_size[0] contains the number of external bases.

int helix_size [2000]

contains a list of all stack sizes.

• int loop_degree [2000]

contains the corresponding list of loop degrees.

· int loops

contains the number of loops (and therefore of stacks).

· int unpaired

contains the number of unpaired bases.

· int pairs

contains the number of base pairs in the last parsed structure.

16.90.2 Function Documentation

Converts the full structure from bracket notation to the HIT notation including root.

Deprecated See vrna_db_to_tree_string() and VRNA_STRUCTURE_TREE_HIT for a replacement

Parameters

structure

Returns

Converts the full structure from bracket notation to the a coarse grained notation using the 'H' 'B' 'I' 'M' and 'R' identifiers.

Deprecated See vrna_db_to_tree_string() and VRNA_STRUCTURE_TREE_SHAPIRO_SHORT for a replacement

Parameters

structure

Returns

16.90.2.3 b2Shapiro()

Converts the full structure from bracket notation to the *weighted* coarse grained notation using the 'H' 'B' 'I' 'M' 'S' 'E' and 'R' identifiers.

Deprecated See vrna_db_to_tree_string() and VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT for a replacement

Parameters

structure

Returns

#include <ViennaRNA/RNAstruct.h>

Adds a root to an un-rooted tree in any except bracket notation.

Parameters

structure

Returns

16.90.2.5 expand_Shapiro()

Inserts missing 'S' identifiers in unweighted coarse grained structures as obtained from b2C().

Parameters

coarse

Returns

16.90.2.6 expand_Full()

Convert the full structure from bracket notation to the expanded notation including root.

Parameters structure
Returns
16.90.2.7 unexpand_Full()
<pre>char* unexpand_Full (const char * ffull)</pre>
<pre>#include <viennarna rnastruct.h=""></viennarna></pre>
Restores the bracket notation from an expanded full or HIT tree, that is any tree using only identifiers 'U' 'P' and 'R'.
Parameters ffull
Returns
16.90.2.8 unweight()
<pre>char* unweight (const char * wcoarse)</pre>
<pre>#include <viennarna rnastruct.h=""></viennarna></pre>
Strip weights from any weighted tree.
Parameters wcoarse

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Returns

16.90.2.9 unexpand_aligned_F()

Converts two aligned structures in expanded notation.

Takes two aligned structures as produced by tree_edit_distance() function back to bracket notation with '_' as the gap character. The result overwrites the input.

Parameters

align

16.90.2.10 parse structure()

Collects a statistic of structure elements of the full structure in bracket notation.

The function writes to the following global variables: loop_size, loop_degree, helix_size, loops, pairs, unpaired

Parameters

structure

Returns

16.90.2.11 pack_structure()

Pack secondary secondary structure, 5:1 compression using base 3 encoding.

Returns a binary string encoding of the secondary structure using a 5:1 compression scheme. The string is NULL terminated and can therefore be used with standard string functions such as strcmp(). Useful for programs that need to keep many structures in memory.

Deprecated Use vrna_db_pack() as a replacement

Parameters

struc The secondary structure in dot-bracket notation

Returns

The binary encoded structure

16.90.2.12 unpack_structure()

Unpack secondary structure previously packed with pack_structure()

Translate a compressed binary string produced by pack_structure() back into the familiar dot-bracket notation.

Deprecated Use vrna_db_unpack() as a replacement

Parameters

packed	The binary encoded packed secondary structure
1	, , ,

Returns

The unpacked secondary structure in dot-bracket notation

16.90.2.13 make_pair_table()

Create a pair table of a secondary structure.

Returns a newly allocated table, such that table[i]=j if (i.j) pair or 0 if i is unpaired, table[0] contains the length of the structure.

Deprecated Use vrna_ptable() instead

Parameters

structure	The secondary structure in dot-bracket notation	1
-----------	---	---

Returns

A pointer to the created pair_table

```
16.90.2.14 copy_pair_table()
```

#include <ViennaRNA/utils/structures.h>

Get an exact copy of a pair table.

Deprecated Use vrna_ptable_copy() instead

Parameters

```
pt The pair table to be copied
```

Returns

A pointer to the copy of 'pt'

16.90.2.15 alimake_pair_table()

#include <ViennaRNA/utils/structures.h>

Pair table for snoop align

Deprecated Use vrna_pt_ali_get() instead!

16.90.2.16 make_pair_table_snoop()

returns a newly allocated table, such that: table[i]=j if (i.j) pair or 0 if i is unpaired, table[0] contains the length of the structure. The special pseudoknotted H/ACA-mRNA structure is taken into account.

Deprecated Use vrna_pt_snoop_get() instead!

16.90.2.17 bp_distance()

Compute the "base pair" distance between two secondary structures s1 and s2.

The sequences should have the same length. dist = number of base pairs in one structure but not in the other same as edit distance with open-pair close-pair as move-set

Deprecated Use vrna bp distance instead

Parameters

str1	First structure in dot-bracket notation
str2	Second structure in dot-bracket notation

Returns

The base pair distance between str1 and str2

16.90.2.18 make_referenceBP_array()

Make a reference base pair count matrix.

Get an upper triangular matrix containing the number of basepairs of a reference structure for each interval [i,j] with i < j. Access it via iindx!!!

Deprecated Use vrna_refBPcnt_matrix() instead

16.90.2.19 compute_BPdifferences()

#include <ViennaRNA/utils/structures.h>

Make a reference base pair distance matrix.

Get an upper triangular matrix containing the base pair distance of two reference structures for each interval [i,j] with i < j. Access it via iindx!!!

Deprecated Use vrna_refBPdist_matrix() instead

16.90.2.20 parenthesis_structure()

#include <ViennaRNA/utils/structures.h>

Create a dot-backet/parenthesis structure from backtracking stack.

Deprecated use vrna_parenthesis_structure() instead

Note

This function is threadsafe

16.90.2.21 parenthesis_zuker()

Create a dot-backet/parenthesis structure from backtracking stack obtained by zuker suboptimal calculation in cofold.c.

Deprecated use vrna_parenthesis_zuker instead

Note

This function is threadsafe

16.90.2.22 bppm_to_structure()

Create a dot-bracket like structure string from base pair probability matrix.

Deprecated Use vrna_db_from_probs() instead!

16.90.2.23 bppm_symbol()

Get a pseudo dot bracket notation for a given probability information.

Deprecated Use vrna_bpp_symbol() instead!

16.91 Deprecated Interface for Plotting Utilities

16.91.1 Detailed Description

Collaboration diagram for Deprecated Interface for Plotting Utilities:

Data Structures

struct COORDINATE

this is a workarround for the SWIG Perl Wrapper RNA plot function that returns an array of type COORDINATE More...

Functions

- int PS_color_aln (const char *structure, const char *filename, const char *seqs[], const char *names[])

 Produce PostScript sequence alignment color-annotated by consensus structure.
- int aliPS_color_aln (const char *structure, const char *filename, const char *seqs[], const char *names[]) PS_color_aln for duplexes.
- int simple_xy_coordinates (short *pair_table, float *X, float *Y)

Calculate nucleotide coordinates for secondary structure plot the Simple way

int simple_circplot_coordinates (short *pair_table, float *x, float *y)

Calculate nucleotide coordinates for Circular Plot

int naview_xy_coordinates (short *pair_table, float *X, float *Y)

Variables

int rna_plot_type

Switch for changing the secondary structure layout algorithm.

16.91.2 Data Structure Documentation

16.91.2.1 struct COORDINATE

this is a workarround for the SWIG Perl Wrapper RNA plot function that returns an array of type COORDINATE

16.91.3 Function Documentation

```
16.91.3.1 PS_color_aln()
```

Produce PostScript sequence alignment color-annotated by consensus structure.

Deprecated Use vrna_file_PS_aln() instead!

```
16.91.3.2 aliPS_color_aln()
```

#include <ViennaRNA/plotting/alignments.h>

PS_color_aln for duplexes.

Deprecated Use vrna_file_PS_aln() instead!

16.91.3.3 simple_xy_coordinates()

Calculate nucleotide coordinates for secondary structure plot the Simple way

See also

```
make_pair_table(), rna_plot_type, simple_circplot_coordinates(), naview_xy_coordinates(), vrna_file_PS_rnaplot_a(), vrna_file_PS_rnaplot, svg_rna_plot()
```

Deprecated Consider switching to vrna_plot_coords_simple_pt() instead!

Parameters

pair_table	The pair table of the secondary structure
X	a pointer to an array with enough allocated space to hold the x coordinates
Υ	a pointer to an array with enough allocated space to hold the y coordinates

Returns

length of sequence on success, 0 otherwise

16.91.3.4 simple_circplot_coordinates()

#include <ViennaRNA/plotting/layouts.h>

Calculate nucleotide coordinates for Circular Plot

This function calculates the coordinates of nucleotides mapped in equal distancies onto a unit circle.

Note

In order to draw nice arcs using quadratic bezier curves that connect base pairs one may calculate a second tangential point P^t in addition to the actual R^2 coordinates. the simplest way to do so may be to compute a radius scaling factor rs in the interval [0,1] that weights the proportion of base pair span to the actual length of the sequence. This scaling factor can then be used to calculate the coordinates for P^t , i.e. $P^t_x[i] = X[i] * rs$ and $P^t_y[i] = Y[i] * rs$.

See also

make_pair_table(), rna_plot_type, simple_xy_coordinates(), naview_xy_coordinates(), vrna_file_PS_rnaplot_a(), vrna_file_PS_rnaplot, svg_rna_plot()

Deprecated Consider switching to vrna_plot_coords_circular_pt() instead!

Parameters

pair_table	The pair table of the secondary structure
X	a pointer to an array with enough allocated space to hold the x coordinates
У	a pointer to an array with enough allocated space to hold the y coordinates

Returns

length of sequence on success, 0 otherwise

16.91.3.5 naview_xy_coordinates()

#include <ViennaRNA/plotting/naview.h>

Deprecated Consider using vrna_plot_coords_naview_pt() instead!

16.91.4 Variable Documentation

16.91.4.1 rna_plot_type

```
int rna_plot_type
#include <ViennaRNA/plotting/layouts.h>
```

Switch for changing the secondary structure layout algorithm.

Current possibility are 0 for a simple radial drawing or 1 for the modified radial drawing taken from the *naview* program of [5].

Note

To provide thread safety please do not rely on this global variable in future implementations but pass a plot type flag directly to the function that decides which layout algorithm it may use!

See also

VRNA PLOT TYPE SIMPLE, VRNA PLOT TYPE NAVIEW, VRNA PLOT TYPE CIRCULAR

16.92 Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers

16.92.1 Detailed Description

Collaboration diagram for Deprecated Interface for (Re-)folding Paths, Saddle Points, and Energy Barriers:

Typedefs

```
    typedef struct vrna_path_s path_t
    Old typename of vrna_path_s.
```

Functions

```
• int find_saddle (const char *seq, const char *s1, const char *s2, int width)

Find energy of a saddle point between 2 structures (search only direct path)
```

void free_path (vrna_path_t *path)

Free memory allocated by get_path() function.

vrna_path_t * get_path (const char *seq, const char *s1, const char *s2, int width)
 Find refolding path between 2 structures (search only direct path)

16.92.2 Typedef Documentation

```
16.92.2.1 path_t

typedef struct vrna_path_s path_t

#include <ViennaRNA/landscape/paths.h>
Old typename of vrna_path_s.
```

Deprecated Use vrna_path_t instead!

16.92.3 Function Documentation

Find energy of a saddle point between 2 structures (search only direct path)

Deprecated Use vrna_path_findpath_saddle() instead!

Parameters

seq	RNA sequence
s1	A pointer to the character array where the first secondary structure in dot-bracket notation will be
	written to
s2	A pointer to the character array where the second secondary structure in dot-bracket notation will be
	written to
width	integer how many strutures are being kept during the search

Returns

the saddle energy in 10cal/mol

16.92.3.2 free_path()

Free memory allocated by get_path() function.

Deprecated Use vrna_path_free() instead!

Parameters

path pointer to memory to be	e freed
------------------------------	---------

16.92.3.3 get_path()

#include <ViennaRNA/landscape/findpath.h>

Find refolding path between 2 structures (search only direct path)

Deprecated Use vrna_path_findpath() instead!

Parameters

seq	RNA sequence
s1	A pointer to the character array where the first secondary structure in dot-bracket notation will be written to
s2	A pointer to the character array where the second secondary structure in dot-bracket notation will be written to
width	integer how many strutures are being kept during the search

Returns

direct refolding path between two structures

Chapter 17

Data Structure Documentation

17.1 _struct_en Struct Reference

Data structure for energy_of_move()

17.1.1 Detailed Description

Data structure for energy_of_move()

The documentation for this struct was generated from the following file:

• ViennaRNA/move_set.h

17.2 LIST Struct Reference

Collaboration diagram for LIST:

The documentation for this struct was generated from the following file:

· ViennaRNA/datastructures/lists.h

17.3 LST_BUCKET Struct Reference

Collaboration diagram for LST_BUCKET:

The documentation for this struct was generated from the following file:

• ViennaRNA/datastructures/lists.h

17.4 Postorder_list Struct Reference

Postorder data structure.

17.4.1 Detailed Description

Postorder data structure.

The documentation for this struct was generated from the following file:

· ViennaRNA/dist vars.h

17.5 swString Struct Reference

Some other data structure.

17.5.1 Detailed Description

Some other data structure.

The documentation for this struct was generated from the following file:

ViennaRNA/dist_vars.h

17.6 Tree Struct Reference

Tree data structure.

Collaboration diagram for Tree:

17.6.1 Detailed Description

Tree data structure.

The documentation for this struct was generated from the following file:

· ViennaRNA/dist vars.h

17.7 TwoDpfold_vars Struct Reference

Variables compound for 2Dfold partition function folding.

Collaboration diagram for TwoDpfold_vars:

Data Fields

char * ptype

Precomputed array of pair types.

char * sequence

The input sequence.

short * S1

The input sequences in numeric form.

unsigned int maxD1

Maximum allowed base pair distance to first reference.

unsigned int maxD2

Maximum allowed base pair distance to second reference.

int * my_iindx

Index for moving in quadratic distancy dimensions.

int * jindx

Index for moving in the triangular matrix qm1.

unsigned int * referenceBPs1

Matrix containing number of basepairs of reference structure1 in interval [i,j].

unsigned int * referenceBPs2

Matrix containing number of basepairs of reference structure2 in interval [i,j].

unsigned int * bpdist

Matrix containing base pair distance of reference structure 1 and 2 on interval [i,j].

unsigned int * mm1

Maximum matching matrix, reference struct 1 disallowed.

unsigned int * mm2

Maximum matching matrix, reference struct 2 disallowed.

17.7.1 Detailed Description

Variables compound for 2Dfold partition function folding.

Deprecated This data structure will be removed from the library soon! Use vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_pf_TwoD(), and vrna_fold_compound_free() instead!

The documentation for this struct was generated from the following file:

ViennaRNA/2Dpfold.h

17.8 vrna_dimer_conc_s Struct Reference

Data structure for concentration dependency computations.

Data Fields

- · double Ac_start
 - start concentration A
- double Bc_start
 - start concentration B
- double ABc

End concentration AB.

17.8.1 Detailed Description

Data structure for concentration dependency computations.

The documentation for this struct was generated from the following file:

· ViennaRNA/concentrations.h

17.9 vrna_hc_bp_storage_t Struct Reference

A base pair hard constraint.

17.9.1 Detailed Description

A base pair hard constraint.

The documentation for this struct was generated from the following file:

• ViennaRNA/constraints/hard.h

17.10 vrna_sc_bp_storage_t Struct Reference

A base pair constraint.

17.10.1 Detailed Description

A base pair constraint.

The documentation for this struct was generated from the following file:

• ViennaRNA/constraints/soft.h

17.11 vrna_sc_motif_s Struct Reference

The documentation for this struct was generated from the following file:

· ViennaRNA/constraints/ligand.h

17.12 vrna structured domains s Struct Reference

The documentation for this struct was generated from the following file:

• ViennaRNA/structured_domains.h

17.13 vrna_subopt_sol_s Struct Reference

Solution element from subopt.c.

Data Fields

· float energy

Free Energy of structure in kcal/mol.

• char * structure

Structure in dot-bracket notation.

17.13.1 Detailed Description

Solution element from subopt.c.

The documentation for this struct was generated from the following file:

· ViennaRNA/subopt.h

17.14 vrna_unstructured_domain_motif_s Struct Reference

The documentation for this struct was generated from the following file:

• ViennaRNA/unstructured_domains.h

Chapter 18

File Documentation

18.1 ViennaRNA/2Dfold.h File Reference

MFE structures for base pair distance classes.

Include dependency graph for 2Dfold.h:

Data Structures

struct vrna_sol_TwoD_t

Solution element returned from vrna_mfe_TwoD() More...

struct TwoDfold_vars

Variables compound for 2Dfold MFE folding. More...

Typedefs

- typedef struct vrna_sol_TwoD_t vrna_sol_TwoD_t
 Solution element returned from vrna_mfe_TwoD()
- typedef struct TwoDfold vars TwoDfold vars

Variables compound for 2Dfold MFE folding.

Functions

vrna_sol_TwoD_t * vrna_mfe_TwoD (vrna_fold_compound_t *vc, int distance1, int distance2)

Compute MFE's and representative for distance partitioning.

char * vrna_backtrack5_TwoD (vrna_fold_compound_t *vc, int k, int l, unsigned int j)

Backtrack a minimum free energy structure from a 5' section of specified length.

TwoDfold_vars * get_TwoDfold_variables (const char *seq, const char *structure1, const char *structure2, int circ)

Get a structure of type TwoDfold_vars prefilled with current global settings.

void destroy_TwoDfold_variables (TwoDfold_vars *our_variables)

Destroy a TwoDfold vars datastructure without memory loss.

vrna_sol_TwoD_t * TwoDfoldList (TwoDfold_vars *vars, int distance1, int distance2)

Compute MFE's and representative for distance partitioning.

char * TwoDfold backtrack f5 (unsigned int j, int k, int l, TwoDfold vars *vars)

Backtrack a minimum free energy structure from a 5' section of specified length.

vrna_sol_TwoD_t ** TwoDfold (TwoDfold_vars *our_variables, int distance1, int distance2)

706 File Documentation

18.1.1 Detailed Description

MFE structures for base pair distance classes.

18.2 ViennaRNA/2Dpfold.h File Reference

Partition function implementations for base pair distance classes.

Include dependency graph for 2Dpfold.h:

Data Structures

struct vrna_sol_TwoD_pf_t

Solution element returned from vrna_pf_TwoD() More...

struct TwoDpfold_vars

Variables compound for 2Dfold partition function folding.

Typedefs

typedef struct vrna_sol_TwoD_pf_t vrna_sol_TwoD_pf_t
 Solution element returned from vrna_pf_TwoD()

Functions

- vrna_sol_TwoD_pf_t * vrna_pf_TwoD (vrna_fold_compound_t *vc, int maxDistance1, int maxDistance2)
 Compute the partition function for all distance classes.
- char * vrna_pbacktrack_TwoD (vrna_fold_compound_t *vc, int d1, int d2)

Sample secondary structure representatives from a set of distance classes according to their Boltzmann probability.

• char * vrna_pbacktrack5_TwoD (vrna_fold_compound_t *vc, int d1, int d2, unsigned int length)

Sample secondary structure representatives with a specified length from a set of distance classes according to their Boltzmann probability.

• TwoDpfold_vars * get_TwoDpfold_variables (const char *seq, const char *structure1, char *structure2, int circ)

Get a datastructure containing all necessary attributes and global folding switches.

void destroy TwoDpfold variables (TwoDpfold vars *vars)

Free all memory occupied by a TwoDpfold_vars datastructure.

vrna_sol_TwoD_pf_t * TwoDpfoldList (TwoDpfold_vars *vars, int maxDistance1, int maxDistance2)

Compute the partition function for all distance classes.

char * TwoDpfold_pbacktrack (TwoDpfold_vars *vars, int d1, int d2)

Sample secondary structure representatives from a set of distance classes according to their Boltzmann probability.

char * TwoDpfold_pbacktrack5 (TwoDpfold_vars *vars, int d1, int d2, unsigned int length)

Sample secondary structure representatives with a specified length from a set of distance classes according to their Boltzmann probability.

18.2.1 Detailed Description

Partition function implementations for base pair distance classes.

18.2.2 Function Documentation

18.2.2.1 get_TwoDpfold_variables()

Get a datastructure containing all necessary attributes and global folding switches.

This function prepares all necessary attributes and matrices etc which are needed for a call of TwoDpfold() . A snapshot of all current global model switches (dangles, temperature and so on) is done and stored in the returned datastructure. Additionally, all matrices that will hold the partition function values are prepared.

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_pf_TwoD(), and vrna_fold_compound_free() instead!

Parameters

seq	the RNA sequence in uppercase format with letters from the alphabet {AUCG}
structure1	the first reference structure in dot-bracket notation
structure2	the second reference structure in dot-bracket notation
circ	a switch indicating if the sequence is linear (0) or circular (1)

Returns

the datastructure containing all necessary partition function attributes

18.2.2.2 destroy_TwoDpfold_variables()

```
void destroy_TwoDpfold_variables ( {\tt TwoDpfold\_vars} \ * \ vars \ )
```

Free all memory occupied by a TwoDpfold_vars datastructure.

This function free's all memory occupied by a datastructure obtained from from get_TwoDpfold_variabless() or get ← _TwoDpfold_variables_from_MFE()

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound TwoD(), vrna pf TwoD(), and vrna fold compound free() instead!

See also

```
get_TwoDpfold_variables(), get_TwoDpfold_variables_from_MFE()
```

708 File Documentation

Parameters

18.2.2.3 TwoDpfoldList()

Compute the partition function for all distance classes.

This function computes the partition functions for all distance classes according the two reference structures specified in the datastructure 'vars'. Similar to TwoDfold() the arguments maxDistance1 and maxDistance2 specify the maximum distance to both reference structures. A value of '-1' in either of them makes the appropriate distance restrictionless, i.e. all basepair distancies to the reference are taken into account during computation. In case there is a restriction, the returned solution contains an entry where the attribute k=l=-1 contains the partition function for all structures exceeding the restriction. A values of INF in the attribute 'k' of the returned list denotes the end of the list

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_pf_TwoD(), and vrna_fold_compound_free() instead!

See also

```
get_TwoDpfold_variables(), destroy_TwoDpfold_variables(), vrna_sol_TwoD_pf_t
```

Parameters

vars	the datastructure containing all necessary folding attributes and matrices
maxDistance1	the maximum basepair distance to reference1 (may be -1)
maxDistance2	the maximum basepair distance to reference2 (may be -1)

Returns

a list of partition funtions for the appropriate distance classes

18.2.2.4 TwoDpfold_pbacktrack()

```
char* TwoDpfold_pbacktrack (  \begin{tabular}{ll} TwoDpfold\_vars * vars, \\ int $d1$, \\ int $d2$ ) \end{tabular}
```

Sample secondary structure representatives from a set of distance classes according to their Boltzmann probability.

If the argument 'd1' is set to '-1', the structure will be backtracked in the distance class where all structures exceeding the maximum basepair distance to either of the references reside.

Precondition

The argument 'vars' must contain precalculated partition function matrices, i.e. a call to TwoDpfold() preceding this function is mandatory!

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold compound_TwoD(), vrna_pf_TwoD(), vrna_pbacktrack_TwoD(), and vrna_fold_compound_free() instead!

See also

TwoDpfold()

Parameters

in	vars	the datastructure containing all necessary folding attributes and matrices
in	d1	the distance to reference1 (may be -1)
in	d2	the distance to reference2

Returns

A sampled secondary structure in dot-bracket notation

18.2.2.5 TwoDpfold_pbacktrack5()

Sample secondary structure representatives with a specified length from a set of distance classes according to their Boltzmann probability.

This function does essentially the same as TwoDpfold_pbacktrack() with the only difference that partial structures, i.e. structures beginning from the 5' end with a specified length of the sequence, are backtracked

Note

This function does not work (since it makes no sense) for circular RNA sequences!

710 File Documentation

Precondition

The argument 'vars' must contain precalculated partition function matrices, i.e. a call to TwoDpfold() preceding this function is mandatory!

Deprecated Use the new API that relies on vrna_fold_compound_t and the corresponding functions vrna_fold_compound_TwoD(), vrna_pf_TwoD(), vrna_pbacktrack5_TwoD(), and vrna_fold_compound_free() instead!

See also

TwoDpfold_pbacktrack(), TwoDpfold()

Parameters

in	vars	the datastructure containing all necessary folding attributes and matrices
in	d1	the distance to reference1 (may be -1)
in	d2	the distance to reference2
in	length	the length of the structure beginning from the 5' end

Returns

A sampled secondary structure in dot-bracket notation

18.3 ViennaRNA/alifold.h File Reference

Functions for comparative structure prediction using RNA sequence alignments.

Include dependency graph for alifold.h:

Functions

- float energy_of_alistruct (const char **sequences, const char *structure, int n_seq, float *energy)

 Calculate the free energy of a consensus structure given a set of aligned sequences.
- void update_alifold_params (void)

Update the energy parameters for alifold function.

- float alifold (const char **strings, char *structure)
 - Compute MFE and according consensus structure of an alignment of sequences.
- float circalifold (const char **strings, char *structure)
 - Compute MFE and according structure of an alignment of sequences assuming the sequences are circular instead of linear.
- void free_alifold_arrays (void)

Free the memory occupied by MFE alifold functions.

- float alipf_fold_par (const char **sequences, char *structure, vrna_ep_t **pl, vrna_exp_param_t *parameters, int calculate_bppm, int is_constrained, int is_circular)
- float alipf_fold (const char **sequences, char *structure, vrna_ep_t **pl)

The partition function version of alifold() works in analogy to $pf_fold()$. Pair probabilities and information about sequence covariations are returned via the 'pi' variable as a list of $vrna_pinfo_t$ structs. The list is terminated by the first entry with pi.i = 0.

- float alipf circ fold (const char **sequences, char *structure, vrna ep t **pl)
- FLT OR DBL * export all bppm (void)

Get a pointer to the base pair probability array.

void free_alipf_arrays (void)

Free the memory occupied by folding matrices allocated by alipf_fold, alipf_circ_fold, etc.

char * alipbacktrack (double *prob)

Sample a consensus secondary structure from the Boltzmann ensemble according its probability.

int get_alipf_arrays (short ***S_p, short ***S5_p, short ***S3_p, unsigned short ***a2s_p, char ***Ss←
 _p, FLT_OR_DBL **qb_p, FLT_OR_DBL **qm_p, FLT_OR_DBL **q1k_p, FLT_OR_DBL **qln_p, short
 **pscore)

Get pointers to (almost) all relavant arrays used in alifold's partition function computation.

Variables

· double cv fact

This variable controls the weight of the covariance term in the energy function of alignment folding algorithms.

· double nc fact

This variable controls the magnitude of the penalty for non-compatible sequences in the covariance term of alignment folding algorithms.

18.3.1 Detailed Description

Functions for comparative structure prediction using RNA sequence alignments.

18.3.2 Function Documentation

18.3.2.1 energy_of_alistruct()

Calculate the free energy of a consensus structure given a set of aligned sequences.

Deprecated Usage of this function is discouraged! Use vrna_eval_structure(), and vrna_eval_covar_structure() instead!

712 File Documentation

Parameters

sequences	The NULL terminated array of sequences
structure	The consensus structure
n_seq	The number of sequences in the alignment
energy	A pointer to an array of at least two floats that will hold the free energies (energy[0] will contain the free energy, energy[1] will be filled with the covariance energy term)

Returns

free energy in kcal/mol

18.3.2.2 update_alifold_params()

Update the energy parameters for alifold function.

Call this to recalculate the pair matrix and energy parameters after a change in folding parameters like temperature

Deprecated Usage of this function is discouraged! The new API uses vrna_fold_compound_t to lump all folding related necessities together, including the energy parameters. Use vrna_update_fold_params() to update the energy parameters within a vrna_fold_compound_t.

18.3.3 Variable Documentation

```
18.3.3.1 cv_fact
```

double cv_fact

This variable controls the weight of the covariance term in the energy function of alignment folding algorithms.

Deprecated See vrna_md_t.cv_fact, and vrna_mfe() to avoid using global variables

Default is 1.

18.3.3.2 nc_fact

double nc_fact

This variable controls the magnitude of the penalty for non-compatible sequences in the covariance term of alignment folding algorithms.

Deprecated See vrna_md_t.nc_fact, and vrna_mfe() to avoid using global variables

Default is 1.

18.4 ViennaRNA/aln_util.h File Reference

Use ViennaRNA/utils/alignments.h instead.

Include dependency graph for aln util.h:

18.4.1 Detailed Description

Use ViennaRNA/utils/alignments.h instead.

Deprecated Use ViennaRNA/utils/alignments.h instead

18.5 ViennaRNA/alphabet.h File Reference

Functions to process, convert, and generally handle different nucleotide and/or base pair alphabets.

Include dependency graph for alphabet.h: This graph shows which files directly or indirectly include this file:

Functions

char * vrna ptypes (const short *S, vrna md t *md)

Get an array of the numerical encoding for each possible base pair (i,j)

short * vrna_seq_encode (const char *sequence, vrna_md_t *md)

Get a numerical representation of the nucleotide sequence.

• short * vrna_seq_encode_simple (const char *sequence, vrna_md_t *md)

Get a numerical representation of the nucleotide sequence (simple version)

int vrna_nucleotide_encode (char c, vrna_md_t *md)

Encode a nucleotide character to numerical value.

char vrna_nucleotide_decode (int enc, vrna_md_t *md)

Decode a numerical representation of a nucleotide back into nucleotide alphabet.

18.5.1 Detailed Description

Functions to process, convert, and generally handle different nucleotide and/or base pair alphabets.

18.6 ViennaRNA/boltzmann_sampling.h File Reference

Boltzmann Sampling of secondary structures from the ensemble.

Include dependency graph for boltzmann_sampling.h: This graph shows which files directly or indirectly include this file:

Generated by Doxygen

Macros

• #define VRNA_PBACKTRACK_DEFAULT 0

Boltzmann sampling flag indicating default backtracing mode.

#define VRNA_PBACKTRACK_NON_REDUNDANT 1

Boltzmann sampling flag indicating non-redundant backtracing mode.

Typedefs

• typedef void() vrna_boltzmann_sampling_callback(const char *stucture, void *data)

Callback for Boltzmann sampling.

typedef struct vrna_pbacktrack_memory_s * vrna_pbacktrack_mem_t

Boltzmann sampling memory data structure.

Functions

char * vrna_pbacktrack5 (vrna_fold_compound_t *fc, unsigned int length)

Sample a secondary structure of a subsequence from the Boltzmann ensemble according its probability.

 char ** vrna_pbacktrack5_num (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, unsigned int options)

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

• unsigned int vrna_pbacktrack5_cb (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, vrna_boltzmann_sampling_callback *cb, void *data, unsigned int options)

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

• char ** vrna_pbacktrack5_resume (vrna_fold_compound_t *vc, unsigned int num_samples, unsigned int length, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

• unsigned int vrna_pbacktrack5_resume_cb (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int length, vrna_boltzmann_sampling_callback *cb, void *data, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

Obtain a set of secondary structure samples for a subsequence from the Boltzmann ensemble according their probability.

char * vrna_pbacktrack (vrna_fold_compound_t *fc)

Sample a secondary structure from the Boltzmann ensemble according its probability.

• char ** vrna_pbacktrack_num (vrna_fold_compound_t *fc, unsigned int num_samples, unsigned int options)

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

unsigned int vrna_pbacktrack_cb (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_boltzmann_sampling_callback
 *cb, void *data, unsigned int options)

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

 char ** vrna_pbacktrack_resume (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

unsigned int vrna_pbacktrack_resume_cb (vrna_fold_compound_t *fc, unsigned int num_samples, vrna_boltzmann_sampling_callback *cb, void *data, vrna_pbacktrack_mem_t *nr_mem, unsigned int options)

Obtain a set of secondary structure samples from the Boltzmann ensemble according their probability.

void vrna_pbacktrack_mem_free (vrna_pbacktrack_mem_t s)

Release memory occupied by a Boltzmann sampling memory data structure.

18.6.1 Detailed Description

Boltzmann Sampling of secondary structures from the ensemble.

A.k.a. Stochastic backtracking

18.7 ViennaRNA/centroid.h File Reference

Centroid structure computation.

Include dependency graph for centroid.h: This graph shows which files directly or indirectly include this file:

Functions

```
• char * vrna_centroid (vrna_fold_compound_t *vc, double *dist)
```

Get the centroid structure of the ensemble.

• char * vrna_centroid_from_plist (int length, double *dist, vrna_ep_t *pl)

Get the centroid structure of the ensemble.

• char * vrna_centroid_from_probs (int length, double *dist, FLT_OR_DBL *probs)

Get the centroid structure of the ensemble.

char * get_centroid_struct_pl (int length, double *dist, vrna_ep_t *pl)

Get the centroid structure of the ensemble.

char * get_centroid_struct_pr (int length, double *dist, FLT_OR_DBL *pr)

Get the centroid structure of the ensemble.

18.7.1 Detailed Description

Centroid structure computation.

18.7.2 Function Documentation

18.7.2.1 get_centroid_struct_pl()

Get the centroid structure of the ensemble.

Deprecated This function was renamed to vrna_centroid_from_plist()

18.7.2.2 get_centroid_struct_pr()

```
char* get_centroid_struct_pr (
    int length,
    double * dist,
    FLT_OR_DBL * pr )
```

Get the centroid structure of the ensemble.

Deprecated This function was renamed to vrna_centroid_from_probs()

18.8 ViennaRNA/char_stream.h File Reference

Use ViennaRNA/datastructures/char stream.h instead.

Include dependency graph for char_stream.h:

18.8.1 Detailed Description

Use ViennaRNA/datastructures/char_stream.h instead.

Deprecated Use ViennaRNA/datastructures/char_stream.h instead

18.9 ViennaRNA/datastructures/char_stream.h File Reference

Implementation of a dynamic, buffered character stream.

Include dependency graph for char_stream.h: This graph shows which files directly or indirectly include this file:

Functions

```
• vrna_cstr_t vrna_cstr (size_t size, FILE *output)
```

Create a dynamic char * stream data structure.

void vrna_cstr_free (vrna_cstr_t buf)

Free the memory occupied by a dynamic char * stream data structure.

void vrna_cstr_close (vrna_cstr_t buf)

Free the memory occupied by a dynamic char \ast stream and close the output stream.

void vrna_cstr_fflush (struct vrna_cstr_s *buf)

Flush the dynamic char * output stream.

18.9.1 Detailed Description

Implementation of a dynamic, buffered character stream.

,

18.10 ViennaRNA/cofold.h File Reference

MFE implementations for RNA-RNA interaction.

Include dependency graph for cofold.h:

Functions

• float cofold (const char *sequence, char *structure)

Compute the minimum free energy of two interacting RNA molecules.

• float cofold par (const char *string, char *structure, vrna param t *parameters, int is constrained)

Compute the minimum free energy of two interacting RNA molecules.

void free_co_arrays (void)

Free memory occupied by cofold()

void update_cofold_params (void)

Recalculate parameters.

void update_cofold_params_par (vrna_param_t *parameters)

Recalculate parameters.

void export_cofold_arrays_gq (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **fc_p, int **ggg_p, int **indx_p, char **ptype_p)

Export the arrays of partition function cofold (with gquadruplex support)

void export_cofold_arrays (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **fc_p, int **indx_p, char **ptype_p)

Export the arrays of partition function cofold.

• void get_monomere_mfes (float *e1, float *e2)

get_monomer_free_energies

void initialize_cofold (int length)

18.10.1 Detailed Description

MFE implementations for RNA-RNA interaction.

18.11 ViennaRNA/combinatorics.h File Reference

Various implementations that deal with combinatorial aspects of objects.

Include dependency graph for combinatorics.h:

Functions

unsigned int ** vrna_enumerate_necklaces (const unsigned int *type_counts)

Enumerate all necklaces with fixed content.

unsigned int vrna_rotational_symmetry_num (const unsigned int *string, size_t string_length)

Determine the order of rotational symmetry for a string of objects represented by natural numbers.

unsigned int vrna_rotational_symmetry_pos_num (const unsigned int *string, size_t string_length, unsigned int *spositions)

Determine the order of rotational symmetry for a string of objects represented by natural numbers.

unsigned int vrna_rotational_symmetry (const char *string)

Determine the order of rotational symmetry for a NULL-terminated string of ASCII characters.

unsigned int vrna_rotational_symmetry_pos (const char *string, unsigned int **positions)

Determine the order of rotational symmetry for a NULL-terminated string of ASCII characters.

• unsigned int vrna_rotational_symmetry_db (vrna_fold_compound_t *fc, const char *structure)

Determine the order of rotational symmetry for a dot-bracket structure.

unsigned int vrna_rotational_symmetry_db_pos (vrna_fold_compound_t *fc, const char *structure, unsigned int **positions)

Determine the order of rotational symmetry for a dot-bracket structure.

18.11.1 Detailed Description

Various implementations that deal with combinatorial aspects of objects.

,

18.12 ViennaRNA/commands.h File Reference

Parse and apply different commands that alter the behavior of secondary structure prediction and evaluation.

Include dependency graph for commands.h:

Macros

#define VRNA_CMD_PARSE_HC 1U

Command parse/apply flag indicating hard constraints.

#define VRNA CMD PARSE SC 2U

Command parse/apply flag indicating soft constraints.

#define VRNA_CMD_PARSE_UD 4U

Command parse/apply flag indicating unstructured domains.

#define VRNA_CMD_PARSE_SD 8U

Command parse/apply flag indicating structured domains.

• #define VRNA CMD PARSE DEFAULTS

Command parse/apply flag indicating default set of commands.

Typedefs

typedef struct vrna_command_s * vrna_cmd_t

A data structure that contains commands.

Functions

- vrna_cmd_t vrna_file_commands_read (const char *filename, unsigned int options)
 Extract a list of commands from a command file.
- int vrna_file_commands_apply (vrna_fold_compound_t *vc, const char *filename, unsigned int options)

 Apply a list of commands from a command file.
- int vrna_commands_apply (vrna_fold_compound_t *vc, vrna_cmd_t commands, unsigned int options)

 Apply a list of commands to a vrna_fold_compound_t.
- void vrna_commands_free (vrna_cmd_t commands)

Free memory occupied by a list of commands.

18.12.1 Detailed Description

Parse and apply different commands that alter the behavior of secondary structure prediction and evaluation.

, ,

18.13 ViennaRNA/concentrations.h File Reference

Concentration computations for RNA-RNA interactions.

Include dependency graph for concentrations.h: This graph shows which files directly or indirectly include this file:

Data Structures

• struct vrna dimer conc s

Data structure for concentration dependency computations.

Functions

vrna_dimer_conc_t * get_concentrations (double FEAB, double FEAA, double FEBB, double FEBB, double FEBB, double FEBB, double *startconc)

Given two start monomer concentrations a and b, compute the concentrations in thermodynamic equilibrium of all dimers and the monomers.

typedef struct vrna_dimer_conc_s vrna_dimer_conc_t

Typename for the data structure that stores the dimer concentrations, vrna_dimer_conc_s, as required by vrna_pf← dimer_concentration()

typedef struct vrna_dimer_conc_s ConcEnt

Backward compatibility typedef for vrna_dimer_conc_s.

vrna_dimer_conc_t * vrna_pf_dimer_concentrations (double FcAB, double FcAA, double FcBB, double FEA, double FEB, const double *startconc, const vrna_exp_param_t *exp_params)

Given two start monomer concentrations a and b, compute the concentrations in thermodynamic equilibrium of all dimers and the monomers.

18.13.1 Detailed Description

Concentration computations for RNA-RNA interactions.

18.13.2 Function Documentation

18.13.2.1 get_concentrations()

Given two start monomer concentrations a and b, compute the concentrations in thermodynamic equilibrium of all dimers and the monomers.

This function takes an array 'startconc' of input concentrations with alternating entries for the initial concentrations of molecules A and B (terminated by two zeroes), then computes the resulting equilibrium concentrations from the free energies for the dimers. Dimer free energies should be the dimer-only free energies, i.e. the FcAB entries from the vrna_dimer_pf_t struct.

Deprecated { Use vrna_pf_dimer_concentrations() instead!}

Parameters

FEAB	Free energy of AB dimer (FcAB entry)
FEAA	Free energy of AA dimer (FcAB entry)
FEBB	Free energy of BB dimer (FcAB entry)
FEA	Free energy of monomer A
FEB	Free energy of monomer B
startconc	List of start concentrations [a0],[b0],[a1],[b1],,[an][bn],[0],[0]

Returns

vrna_dimer_conc_t array containing the equilibrium energies and start concentrations

18.14 ViennaRNA/constraints.h File Reference

Use ViennaRNA/constraints/basic.h instead.

Include dependency graph for constraints.h:

18.14.1 Detailed Description

Use ViennaRNA/constraints/basic.h instead.

Deprecated Use ViennaRNA/constraints/basic.h instead

18.15 ViennaRNA/constraints/hard.h File Reference

Functions and data structures for handling of secondary structure hard constraints.

Include dependency graph for hard.h: This graph shows which files directly or indirectly include this file:

Data Structures

· struct vrna_hc_bp_storage_t

A base pair hard constraint.

• struct vrna hc s

The hard constraints data structure. More...

· struct vrna hc up s

A single hard constraint for a single nucleotide. More...

Macros

• #define VRNA_CONSTRAINT_NO_HEADER 0

do not print the header information line

#define VRNA_CONSTRAINT_DB 16384U

Flag for vrna_constraints_add() to indicate that constraint is passed in pseudo dot-bracket notation.

• #define VRNA CONSTRAINT DB ENFORCE BP 32768U

Switch for dot-bracket structure constraint to enforce base pairs.

#define VRNA_CONSTRAINT_DB_PIPE 65536U

Flag that is used to indicate the pipe \'sign in pseudo dot-bracket notation of hard constraints.

• #define VRNA CONSTRAINT DB DOT 131072U

dot '.' switch for structure constraints (no constraint at all)

#define VRNA_CONSTRAINT_DB_X 262144U

'x' switch for structure constraint (base must not pair)

#define VRNA_CONSTRAINT_DB_ANG_BRACK 524288U

angle brackets '<', '>' switch for structure constraint (paired downstream/upstream)

• #define VRNA CONSTRAINT DB RND BRACK 1048576U

round brackets '(',')' switch for structure constraint (base i pairs base j)

#define VRNA CONSTRAINT DB INTRAMOL 2097152U

Flag that is used to indicate the character 'l' in pseudo dot-bracket notation of hard constraints.

• #define VRNA CONSTRAINT DB INTERMOL 4194304U

Flag that is used to indicate the character 'e' in pseudo dot-bracket notation of hard constraints.

#define VRNA CONSTRAINT DB GQUAD 8388608U

'+' switch for structure constraint (base is involved in a gquad)

• #define VRNA CONSTRAINT DB WUSS 33554432U

Flag to indicate Washington University Secondary Structure (WUSS) notation of the hard constraint string.

#define VRNA_CONSTRAINT_DB_DEFAULT

Switch for dot-bracket structure constraint with default symbols.

#define VRNA_CONSTRAINT_CONTEXT_EXT_LOOP (unsigned char)0x01

Hard constraints flag, base pair in the exterior loop.

#define VRNA_CONSTRAINT_CONTEXT_HP_LOOP (unsigned char)0x02

Hard constraints flag, base pair encloses hairpin loop.

#define VRNA_CONSTRAINT_CONTEXT_INT_LOOP (unsigned char)0x04

Hard constraints flag, base pair encloses an interior loop.

#define VRNA CONSTRAINT CONTEXT INT LOOP ENC (unsigned char)0x08

Hard constraints flag, base pair encloses a multi branch loop.

• #define VRNA CONSTRAINT CONTEXT MB LOOP (unsigned char)0x10

Hard constraints flag, base pair is enclosed in an interior loop.

• #define VRNA_CONSTRAINT_CONTEXT_MB_LOOP_ENC (unsigned char)0x20

Hard constraints flag, base pair is enclosed in a multi branch loop.

#define VRNA_CONSTRAINT_CONTEXT_ENFORCE (unsigned char)0x40

Hard constraint flag to indicate enforcement of constraints.

#define VRNA CONSTRAINT CONTEXT NO REMOVE (unsigned char)0x80

Hard constraint flag to indicate not to remove base pairs that conflict with a given constraint.

#define VRNA CONSTRAINT CONTEXT NONE (unsigned char)0

Constraint context flag that forbids any loop.

#define VRNA_CONSTRAINT_CONTEXT_CLOSING_LOOPS

Constraint context flag indicating base pairs that close any loop.

#define VRNA CONSTRAINT CONTEXT ENCLOSED LOOPS

Constraint context flag indicating base pairs enclosed by any loop.

• #define VRNA CONSTRAINT CONTEXT ALL LOOPS

Constraint context flag indicating any loop context.

Typedefs

• typedef struct vrna_hc_s vrna_hc_t

Typename for the hard constraints data structure vrna_hc_s.

• typedef struct vrna hc up s vrna hc up t

Typename for the single nucleotide hard constraint data structure vrna_hc_up_s.

• typedef unsigned char() vrna_callback_hc_evaluate(int i, int j, int k, int l, unsigned char d, void *data)

Callback to evaluate whether or not a particular decomposition step is contributing to the solution space.

Enumerations

enum vrna_hc_type_e { VRNA_HC_DEFAULT, VRNA_HC_WINDOW }

The hard constraints type.

Functions

void vrna message constraint options (unsigned int option)

Print a help message for pseudo dot-bracket structure constraint characters to stdout. (constraint support is specified by option parameter)

void vrna_message_constraint_options_all (void)

Print structure constraint characters to stdout (full constraint support)

void vrna hc init (vrna fold compound t *vc)

Initialize/Reset hard constraints to default values.

• void vrna_hc_add_up (vrna_fold_compound_t *vc, int i, unsigned char option)

Make a certain nucleotide unpaired.

• int vrna hc add up batch (vrna fold compound t *vc, vrna hc up t *constraints)

Apply a list of hard constraints for single nucleotides.

void vrna_hc_add_bp (vrna_fold_compound_t *vc, int i, int j, unsigned char option)

Favorize/Enforce a certain base pair (i,j)

void vrna_hc_add_bp_nonspecific (vrna_fold_compound_t *vc, int i, int d, unsigned char option)

Enforce a nucleotide to be paired (upstream/downstream)

void vrna_hc_free (vrna_hc_t *hc)

Free the memory allocated by a vrna hc t data structure.

void vrna_hc_add_f (vrna_fold_compound_t *vc, vrna_callback_hc_evaluate *f)

Add a function pointer pointer for the generic hard constraint feature.

void vrna_hc_add_data (vrna_fold_compound_t *vc, void *data, vrna_callback_free_auxdata *f)

Add an auxiliary data structure for the generic hard constraints callback function.

• int vrna hc add from db (vrna fold compound t *vc, const char *constraint, unsigned int options)

Add hard constraints from pseudo dot-bracket notation.

· void print_tty_constraint (unsigned int option)

Print structure constraint characters to stdout. (constraint support is specified by option parameter)

void print tty constraint full (void)

Print structure constraint characters to stdout (full constraint support)

• void constrain_ptypes (const char *constraint, unsigned int length, char *ptype, int *BP, int min_loop_size, unsigned int idx_type)

Insert constraining pair types according to constraint structure string.

18.15.1 Detailed Description

Functions and data structures for handling of secondary structure hard constraints.

18.15.2 Macro Definition Documentation

18.15.2.1 VRNA_CONSTRAINT_NO_HEADER

#define VRNA_CONSTRAINT_NO_HEADER 0

do not print the header information line

Deprecated This mode is not supported anymore!

18.15.2.2 VRNA_CONSTRAINT_DB_ANG_BRACK

```
#define VRNA_CONSTRAINT_DB_ANG_BRACK 524288U
```

angle brackets '<', '>' switch for structure constraint (paired downstream/upstream)

See also

vrna_hc_add_from_db(), vrna_constraints_add(), vrna_message_constraint_options(), vrna_message_constraint_options_all()

18.15.3 Enumeration Type Documentation

```
18.15.3.1 vrna_hc_type_e
```

```
enum vrna_hc_type_e
```

The hard constraints type.

Global and local structure prediction methods use a slightly different way to handle hard constraints internally. This enum is used to distinguish both types.

Enumerator

VRNA_HC_DEFAULT	Default Hard Constraints.
VRNA_HC_WINDOW	Hard Constraints suitable for local structure prediction using window approach.
	See also
	<pre>vrna_mfe_window(), vrna_mfe_window_zscore(), pfl_fold()</pre>

18.15.4 Function Documentation

18.15.4.1 vrna_hc_add_data()

Add an auxiliary data structure for the generic hard constraints callback function.

See also

```
vrna_hc_add_f()
```

Parameters

VC	The fold compound the generic hard constraint function should be bound to
data	A pointer to the data structure that holds required data for function 'f'
f	A pointer to a function that free's the memory occupied by data (Maybe NULL)

18.15.4.2 print_tty_constraint()

```
void print_tty_constraint ( \mbox{unsigned int } option \mbox{ )} \label{eq:constraint}
```

Print structure constraint characters to stdout. (constraint support is specified by option parameter)

Deprecated Use vrna_message_constraints() instead!

Parameters

option	Option switch that tells which constraint help will be printed
--------	--

18.15.4.3 print_tty_constraint_full()

Print structure constraint characters to stdout (full constraint support)

Deprecated Use vrna_message_constraint_options_all() instead!

18.15.4.4 constrain_ptypes()

Insert constraining pair types according to constraint structure string.

Deprecated Do not use this function anymore! Structure constraints are now handled through vrna_hc_t and related functions.

Parameters

constraint	The structure constraint string
length	The actual length of the sequence (constraint may be shorter)
ptype	A pointer to the basepair type array
BP	(not used anymore)
min_loop_size	The minimal loop size (usually TURN)
idx_type	Define the access type for base pair type array (0 = indx, 1 = iindx)

18.16 ViennaRNA/constraints/ligand.h File Reference

Functions for incorporation of ligands binding to hairpin and interior loop motifs using the soft constraints framework.

Include dependency graph for ligand.h: This graph shows which files directly or indirectly include this file:

Data Structures

· struct vrna sc motif s

Typedefs

• typedef struct vrna_sc_motif_s vrna_sc_motif_t

Functions

• int vrna_sc_add_hi_motif (vrna_fold_compound_t *vc, const char *seq, const char *structure, FLT_OR_DBL energy, unsigned int options)

Add soft constraints for hairpin or interior loop binding motif.

18.16.1 Detailed Description

Functions for incorporation of ligands binding to hairpin and interior loop motifs using the soft constraints framework.

18.16.2 Typedef Documentation

```
18.16.2.1 vma_sc_motif_t

typedef struct vrna_sc_motif_s vrna_sc_motif_t

@addtogroup constraints_ligand
@brief Ligand binding to specific hairpin/interior loop like motifs using the @ref soft_constraints feature

Here is an example that adds a theophylline binding motif. Free energy
contribution is derived from @f$k_d = 0.32 \mu mol / 1 @f$, taken from
Jenison et al. 1994

@image html theo_aptamer.svg
@image latex theo_aptamer.eps
@code{.c}
```

vrna_sc_add_hi_motif(vc, "GAUACCAG&CCCUUGGCAGC", "(...((((&)...)))...)", -9.22, VRNA_OPTION_DEFAULT);

18.17 ViennaRNA/constraints/SHAPE.h File Reference

This module provides function to incorporate SHAPE reactivity data into the folding recursions by means of soft constraints.

Include dependency graph for SHAPE.h: This graph shows which files directly or indirectly include this file:

Functions

• int vrna_sc_add_SHAPE_deigan (vrna_fold_compound_t *vc, const double *reactivities, double m, double b, unsigned int options)

Add SHAPE reactivity data as soft constraints (Deigan et al. method)

• int vrna_sc_add_SHAPE_deigan_ali (vrna_fold_compound_t *vc, const char **shape_files, const int *shape_file_association, double m, double b, unsigned int options)

Add SHAPE reactivity data from files as soft constraints for consensus structure prediction (Deigan et al. method)

• int vrna_sc_add_SHAPE_zarringhalam (vrna_fold_compound_t *vc, const double *reactivities, double b, double default_value, const char *shape_conversion, unsigned int options)

Add SHAPE reactivity data as soft constraints (Zarringhalam et al. method)

• int vrna_sc_SHAPE_parse_method (const char *method_string, char *method, float *param_1, float *param_2)

Parse a character string and extract the encoded SHAPE reactivity conversion method and possibly the parameters for conversion into pseudo free energies.

• int vrna_sc_SHAPE_to_pr (const char *shape_conversion, double *values, int length, double default_value)

*Convert SHAPE reactivity values to probabilities for being unpaired.

18.17.1 Detailed Description

This module provides function to incorporate SHAPE reactivity data into the folding recursions by means of soft constraints.

18.17.2 Function Documentation

18.17.2.1 vrna_sc_SHAPE_parse_method()

Parse a character string and extract the encoded SHAPE reactivity conversion method and possibly the parameters for conversion into pseudo free energies.

Parameters

method_string	The string that contains the encoded SHAPE reactivity conversion method
method	A pointer to the memory location where the method character will be stored
param_1	A pointer to the memory location where the first parameter of the corresponding method will be stored
param_2	A pointer to the memory location where the second parameter of the corresponding method will be stored

Returns

1 on successful extraction of the method, 0 on errors

18.18 ViennaRNA/constraints/soft.h File Reference

Functions and data structures for secondary structure soft constraints.

Include dependency graph for soft.h: This graph shows which files directly or indirectly include this file:

Data Structures

- struct vrna_sc_bp_storage_t
 - A base pair constraint.
- struct vrna_sc_s

The soft constraints data structure. More...

Typedefs

- typedef struct vrna_sc_s vrna_sc_t
 - Typename for the soft constraints data structure vrna_sc_s.
- typedef int() vrna_callback_sc_energy(int i, int j, int k, int l, unsigned char d, void *data)

 Callback to retrieve pseudo energy contribution for soft constraint feature.
- typedef FLT_OR_DBL() vrna_callback_sc_exp_energy(int i, int j, int k, int l, unsigned char d, void *data)

 Callback to retrieve pseudo energy contribution as Boltzmann Factors for soft constraint feature.
- typedef vrna_basepair_t *() vrna_callback_sc_backtrack(int i, int j, int k, int l, unsigned char d, void *data)

 Callback to retrieve auxiliary base pairs for soft constraint feature.

Enumerations

enum vrna_sc_type_e { VRNA_SC_DEFAULT, VRNA_SC_WINDOW }
 The type of a soft constraint.

Functions

- void vrna_sc_init (vrna_fold_compound_t *vc)
 - Initialize an empty soft constraints data structure within a vrna fold compound t.
- void vrna_sc_set_bp (vrna_fold_compound_t *vc, const FLT_OR_DBL **constraints, unsigned int options)

 Set soft constraints for paired nucleotides.
- void vrna_sc_add_bp (vrna_fold_compound_t *vc, int i, int j, FLT_OR_DBL energy, unsigned int options)

 Add soft constraints for paired nucleotides.
- void vrna_sc_set_up (vrna_fold_compound_t *vc, const FLT_OR_DBL *constraints, unsigned int options) Set soft constraints for unpaired nucleotides.
- void vrna_sc_add_up (vrna_fold_compound_t *vc, int i, FLT_OR_DBL energy, unsigned int options)

 Add soft constraints for unpaired nucleotides.
- void vrna_sc_remove (vrna_fold_compound_t *vc)
 - Remove soft constraints from vrna_fold_compound_t.
- void vrna_sc_free (vrna_sc_t *sc)
 - Free memory occupied by a vrna_sc_t data structure.
- void vrna_sc_add_data (vrna_fold_compound_t *vc, void *data, vrna_callback_free_auxdata *free_data)

 Add an auxiliary data structure for the generic soft constraints callback function.
- void vrna_sc_add_f (vrna_fold_compound_t *vc, vrna_callback_sc_energy *f)
 - Bind a function pointer for generic soft constraint feature (MFE version)
- void vrna_sc_add_bt (vrna_fold_compound_t *vc, vrna_callback_sc_backtrack *f)
 - Bind a backtracking function pointer for generic soft constraint feature.
- void vrna_sc_add_exp_f (vrna_fold_compound_t *vc, vrna_callback_sc_exp_energy *exp_f)

Bind a function pointer for generic soft constraint feature (PF version)

18.18.1 Detailed Description

Functions and data structures for secondary structure soft constraints.

18.18.2 Enumeration Type Documentation

```
18.18.2.1 vrna_sc_type_e
```

enum vrna_sc_type_e

The type of a soft constraint.

Enumerator

VRNA_SC_DEFAULT	Default Soft Constraints.	
VRNA_SC_WINDOW	Soft Constraints suitable for local structure prediction using window approach.	
Generated by Doxygen	See also	
	vrna_mfe_window(), vrna_mfe_window_zscore(), pfl_fold()	

18.19 ViennaRNA/constraints_hard.h File Reference

Use ViennaRNA/constraints/hard.h instead.

Include dependency graph for constraints_hard.h:

18.19.1 Detailed Description

Use ViennaRNA/constraints/hard.h instead.

Deprecated Use ViennaRNA/constraints/hard.h instead

18.20 ViennaRNA/constraints_ligand.h File Reference

Use ViennaRNA/constraints/ligand.h instead.

Include dependency graph for constraints_ligand.h:

18.20.1 Detailed Description

Use ViennaRNA/constraints/ligand.h instead.

Deprecated Use ViennaRNA/constraints/ligand.h instead

18.21 ViennaRNA/constraints_SHAPE.h File Reference

Use ViennaRNA/constraints/SHAPE.h instead.

Include dependency graph for constraints_SHAPE.h:

18.21.1 Detailed Description

Use ViennaRNA/constraints/SHAPE.h instead.

Deprecated Use ViennaRNA/constraints/SHAPE.h instead

18.22 ViennaRNA/constraints_soft.h File Reference

Use ViennaRNA/constraints/soft.h instead.

Include dependency graph for constraints_soft.h:

18.22.1 Detailed Description

Use ViennaRNA/constraints/soft.h instead.

Deprecated Use ViennaRNA/constraints/soft.h instead

18.23 ViennaRNA/convert_epars.h File Reference

Use ViennaRNA/params/convert.h instead.

Include dependency graph for convert_epars.h:

18.23.1 Detailed Description

Use ViennaRNA/params/convert.h instead.

Deprecated Use ViennaRNA/params/convert.h instead

18.24 ViennaRNA/data_structures.h File Reference

Use ViennaRNA/datastructures/basic.h instead.

Include dependency graph for data_structures.h:

18.24.1 Detailed Description

Use ViennaRNA/datastructures/basic.h instead.

Deprecated Use ViennaRNA/datastructures/basic.h instead

18.25 ViennaRNA/datastructures/hash tables.h File Reference

Implementations of hash table functions.

Data Structures

struct vrna_ht_entry_db_t

Default hash table entry. More...

Functions

Dot-Bracket / Free Energy entries

```
int vrna_ht_db_comp (void *x, void *y)
```

Default hash table entry comparison.

unsigned int vrna_ht_db_hash_func (void *x, unsigned long hashtable_size)

Default hash function.

int vrna_ht_db_free_entry (void *hash_entry)

Default function to free memory occupied by a hash entry.

Abstract interface

typedef struct vrna_hash_table_s * vrna_hash_table_t

A hash table object.

typedef int() vrna_callback_ht_compare_entries(void *x, void *y)

Callback function to compare two hash table entries.

• typedef unsigned int() vrna_callback_ht_hash_function(void *x, unsigned long hashtable_size)

Callback function to generate a hash key, i.e. hash function.

typedef int() vrna_callback_ht_free_entry(void *x)

Callback function to free a hash table entry.

vrna_hash_table_t vrna_ht_init (unsigned int b, vrna_callback_ht_compare_entries *compare_function, vrna_callback_ht_hash_function *hash_function, vrna_callback_ht_free_entry *free_hash_entry)

Get an initialized hash table.

unsigned long vrna_ht_size (vrna_hash_table_t ht)

Get the size of the hash table.

unsigned long vrna_ht_collisions (struct vrna_hash_table_s *ht)

Get the number of collisions in the hash table.

void * vrna_ht_get (vrna_hash_table_t ht, void *x)

Get an element from the hash table.

int vrna_ht_insert (vrna_hash_table_t ht, void *x)

Insert an object into a hash table.

void vrna_ht_remove (vrna_hash_table_t ht, void *x)

Remove an object from the hash table.

void vrna_ht_clear (vrna_hash_table_t ht)

Clear the hash table.

void vrna_ht_free (vrna_hash_table_t ht)

Free all memory occupied by the hash table.

18.25.1 Detailed Description

Implementations of hash table functions.

18.26 ViennaRNA/datastructures/heap.h File Reference

Implementation of an abstract heap data structure.

Typedefs

typedef struct vrna_heap_s * vrna_heap_t

An abstract heap data structure.

typedef int() vrna_callback_heap_cmp(const void *a, const void *b, void *data)

Heap compare function prototype.

typedef size_t() vrna_callback_heap_get_pos(const void *a, void *data)

Retrieve the position of a particular heap entry within the heap.

typedef void() vrna_callback_heap_set_pos(const void *a, size_t pos, void *data)

Store the position of a particular heap entry within the heap.

Functions

vrna_heap_t vrna_heap_init (size_t n, vrna_callback_heap_cmp *cmp, vrna_callback_heap_get_pos *get
 —entry_pos, vrna_callback_heap_set_pos *set_entry_pos, void *data)

Initialize a heap data structure.

void vrna_heap_free (vrna_heap_t h)

Free memory occupied by a heap data structure.

size_t vrna_heap_size (struct vrna_heap_s *h)

Get the size of a heap data structure, i.e. the number of stored elements.

void vrna_heap_insert (vrna_heap_t h, void *v)

Insert an element into the heap.

void * vrna_heap_pop (vrna_heap_t h)

Pop (remove and return) the object at the root of the heap.

const void * vrna_heap_top (vrna_heap_t h)

Get the object at the root of the heap.

void * vrna_heap_remove (vrna_heap_t h, const void *v)

Remove an arbitrary element within the heap.

void * vrna_heap_update (vrna_heap_t h, void *v)

Update an arbitrary element within the heap.

18.26.1 Detailed Description

Implementation of an abstract heap data structure.

18.27 ViennaRNA/dist_vars.h File Reference

Global variables for Distance-Package.

This graph shows which files directly or indirectly include this file:

Data Structures

struct Postorder list

Postorder data structure.

struct Tree

Tree data structure.

· struct swString

Some other data structure.

Variables

· int edit backtrack

Produce an alignment of the two structures being compared by tracing the editing path giving the minimum distance.

• char * aligned_line [4]

Contains the two aligned structures after a call to one of the distance functions with edit_backtrack set to 1.

int cost_matrix

Specify the cost matrix to be used for distance calculations.

18.27.1 Detailed Description

Global variables for Distance-Package.

18.27.2 Variable Documentation

18.27.2.1 edit_backtrack

```
int edit_backtrack
```

Produce an alignment of the two structures being compared by tracing the editing path giving the minimum distance.

set to 1 if you want backtracking

18.27.2.2 cost_matrix

```
int cost_matrix
```

Specify the cost matrix to be used for distance calculations.

if 0, use the default cost matrix (upper matrix in example), otherwise use Shapiro's costs (lower matrix).

18.28 ViennaRNA/dp_matrices.h File Reference

Functions to deal with standard dynamic programming (DP) matrices.

Include dependency graph for dp_matrices.h: This graph shows which files directly or indirectly include this file:

Data Structures

• struct vrna_mx_mfe_s

Minimum Free Energy (MFE) Dynamic Programming (DP) matrices data structure required within the vrna fold compound t. More...

struct vrna_mx_pf_s

Partition function (PF) Dynamic Programming (DP) matrices data structure required within the vrna_fold_compound_t. More...

Typedefs

- typedef struct vrna_mx_mfe_s vrna_mx_mfe_t
 - Typename for the Minimum Free Energy (MFE) DP matrices data structure vrna_mx_mfe_s.
- typedef struct vrna_mx_pf_s vrna_mx_pf_t

Typename for the Partition Function (PF) DP matrices data structure vrna_mx_pf_s.

Enumerations

enum vrna mx_type_e { VRNA_MX_DEFAULT, VRNA_MX_WINDOW, VRNA_MX_2DFOLD }

An enumerator that is used to specify the type of a polymorphic Dynamic Programming (DP) matrix data structure.

Functions

- int vrna_mx_add (vrna_fold_compound_t *vc, vrna_mx_type_e type, unsigned int options)
 Add Dynamic Programming (DP) matrices (allocate memory)
- void vrna_mx_mfe_free (vrna_fold_compound_t *vc)

Free memory occupied by the Minimum Free Energy (MFE) Dynamic Programming (DP) matrices.

void vrna_mx_pf_free (vrna_fold_compound_t *vc)

Free memory occupied by the Partition Function (PF) Dynamic Programming (DP) matrices.

18.28.1 Detailed Description

Functions to deal with standard dynamic programming (DP) matrices.

18.29 ViennaRNA/duplex.h File Reference

Functions for simple RNA-RNA duplex interactions.

Include dependency graph for duplex.h:

18.29.1 Detailed Description

Functions for simple RNA-RNA duplex interactions.

18.30 ViennaRNA/edit_cost.h File Reference

global variables for Edit Costs included by treedist.c and stringdist.c

18.30.1 Detailed Description

global variables for Edit Costs included by treedist.c and stringdist.c

18.31 ViennaRNA/energy_const.h File Reference

Use ViennaRNA/params/constants.h instead.

Include dependency graph for energy_const.h:

18.31.1 Detailed Description

Use ViennaRNA/params/constants.h instead.

Deprecated Use ViennaRNA/params/constants.h instead

18.32 ViennaRNA/energy_par.h File Reference

Use ViennaRNA/params/default.h instead.

Include dependency graph for energy_par.h:

18.32.1 Detailed Description

Use ViennaRNA/params/default.h instead.

Deprecated Use ViennaRNA/params/default.h instead

18.33 ViennaRNA/equilibrium_probs.h File Reference

Equilibrium Probability implementations.

Include dependency graph for equilibrium_probs.h: This graph shows which files directly or indirectly include this file:

Functions

• void vrna_pf_dimer_probs (double FAB, double FA, double FB, vrna_ep_t *prAB, const vrna_ep_t *prA, const vrna_ep_t *prB, int Alength, const vrna_exp_param_t *exp_param_s)

Compute Boltzmann probabilities of dimerization without homodimers.

double vrna_pr_structure (vrna_fold_compound_t *fc, const char *structure)

Compute the equilibrium probability of a particular secondary structure.

• double vrna pr energy (vrna fold compound t *vc, double e)

Base pair related probability computations

- double vrna_mean_bp_distance_pr (int length, FLT_OR_DBL *pr)
 - Get the mean base pair distance in the thermodynamic ensemble from a probability matrix.
- double vrna_mean_bp_distance (vrna_fold_compound_t *vc)
 - Get the mean base pair distance in the thermodynamic ensemble.
- double vrna_ensemble_defect (vrna_fold_compound_t *fc, const char *structure)

Compute the Ensemble Defect for a given target structure.

- double * vrna_positional_entropy (vrna_fold_compound_t *fc)
 - Compute a vector of positional entropies.
- vrna_ep_t * vrna_stack_prob (vrna_fold_compound_t *vc, double cutoff)

Compute stacking probabilities.

18.33.1 Detailed Description

Equilibrium Probability implementations.

This file includes various implementations for equilibrium probability computations based on the partition function of an RNA sequence, two concatenated sequences, or a sequence alignment.

18.33.2 Function Documentation

```
18.33.2.1 vrna_pr_energy()
```

SWIG Wrapper Notes This function is attached as method pr energy() to objects of type fold compound

18.34 ViennaRNA/eval.h File Reference

Functions and variables related to energy evaluation of sequence/structure pairs.

Include dependency graph for eval.h: This graph shows which files directly or indirectly include this file:

Macros

- #define VRNA_VERBOSITY_QUIET -1
 - Quiet level verbosity setting.
- #define VRNA_VERBOSITY_DEFAULT 1

Default level verbosity setting.

Functions

- int vrna_eval_loop_pt (vrna_fold_compound_t *vc, int i, const short *pt)
 - Calculate energy of a loop.
- int vrna_eval_loop_pt_v (vrna_fold_compound_t *vc, int i, const short *pt, int verbosity_level)

Calculate energy of a loop.

- float vrna_eval_move (vrna_fold_compound_t *vc, const char *structure, int m1, int m2)
 - Calculate energy of a move (closing or opening of a base pair)
- int vrna_eval_move_pt (vrna_fold_compound_t *vc, short *pt, int m1, int m2)

Calculate energy of a move (closing or opening of a base pair)

- float energy of structure (const char *string, const char *structure, int verbosity level)
 - Calculate the free energy of an already folded RNA using global model detail settings.
- float energy_of_struct_par (const char *string, const char *structure, vrna_param_t *parameters, int verbosity_level)

Calculate the free energy of an already folded RNA.

• float energy_of_circ_structure (const char *string, const char *structure, int verbosity_level)

Calculate the free energy of an already folded circular RNA.

• float energy_of_circ_struct_par (const char *string, const char *structure, vrna_param_t *parameters, int verbosity_level)

Calculate the free energy of an already folded circular RNA.

- int energy_of_structure_pt (const char *string, short *ptable, short *s, short *s1, int verbosity_level)

 Calculate the free energy of an already folded RNA.
- int energy_of_struct_pt_par (const char *string, short *ptable, short *s, short *s1, vrna_param_t *parameters, int verbosity_level)

Calculate the free energy of an already folded RNA.

• float energy_of_move (const char *string, const char *structure, int m1, int m2)

Calculate energy of a move (closing or opening of a base pair)

int energy of move pt (short *pt, short *s, short *s1, int m1, int m2)

Calculate energy of a move (closing or opening of a base pair)

int loop energy (short *ptable, short *s, short *s1, int i)

Calculate energy of a loop.

- float energy of struct (const char *string, const char *structure)
- int energy_of_struct_pt (const char *string, short *ptable, short *s, short *s1)
- float energy_of_circ_struct (const char *string, const char *structure)

Basic Energy Evaluation Interface with Dot-Bracket Structure String

- float vrna eval structure (vrna fold compound t *vc, const char *structure)
 - Calculate the free energy of an already folded RNA.
- float vrna_eval_covar_structure (vrna_fold_compound_t *vc, const char *structure)

Calculate the pseudo energy derived by the covariance scores of a set of aligned sequences.

- float vrna_eval_structure_verbose (vrna_fold_compound_t *vc, const char *structure, FILE *file)
 - Calculate the free energy of an already folded RNA and print contributions on a per-loop base.
- float vrna_eval_structure_v (vrna_fold_compound_t *vc, const char *structure, int verbosity_level, FILE *file)

Calculate the free energy of an already folded RNA and print contributions on a per-loop base.

float vrna_eval_structure_cstr (vrna_fold_compound_t *vc, const char *structure, int verbosity_level, vrna_cstr_t output_stream)

Basic Energy Evaluation Interface with Structure Pair Table

- int vrna_eval_structure_pt (vrna_fold_compound_t *vc, const short *pt)
 - Calculate the free energy of an already folded RNA.
- int vrna_eval_structure_pt_verbose (vrna_fold_compound_t *vc, const short *pt, FILE *file)

Calculate the free energy of an already folded RNA.

• int vrna_eval_structure_pt_v (vrna_fold_compound_t *vc, const short *pt, int verbosity_level, FILE *file)

Calculate the free energy of an already folded RNA.

Simplified Energy Evaluation with Sequence and Dot-Bracket Strings

- float vrna_eval_structure_simple (const char *string, const char *structure)
 - Calculate the free energy of an already folded RNA.
- float vrna_eval_circ_structure (const char *string, const char *structure)

Evaluate the free energy of a sequence/structure pair where the sequence is circular.

- float vrna eval gquad structure (const char *string, const char *structure)
 - Evaluate the free energy of a sequence/structure pair where the structure may contain G-Quadruplexes.
- float vrna_eval_circ_gquad_structure (const char *string, const char *structure)

Evaluate the free energy of a sequence/structure pair where the sequence is circular and the structure may contain G-Quadruplexes.

- float vrna_eval_structure_simple_verbose (const char *string, const char *structure, FILE *file)

 Calculate the free energy of an already folded RNA and print contributions per loop.
- float vrna_eval_structure_simple_v (const char *string, const char *structure, int verbosity_level, FILE *file)

Calculate the free energy of an already folded RNA and print contributions per loop.

- float vrna_eval_circ_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)
 Evaluate free energy of a sequence/structure pair, assume sequence to be circular and print contributions per loop.
- float vrna_eval_gquad_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)

Evaluate free energy of a sequence/structure pair, allow for G-Quadruplexes in the structure and print contributions per loop.

float vrna_eval_circ_gquad_structure_v (const char *string, const char *structure, int verbosity_level, FILE *file)

Evaluate free energy of a sequence/structure pair, assume sequence to be circular, allow for G-Quadruplexes in the structure, and print contributions per loop.

Simplified Energy Evaluation with Sequence Alignments and Consensus Structure Dot-Bracket String

- float vrna_eval_consensus_structure_simple (const char **alignment, const char *structure)

 Calculate the free energy of an already folded RNA sequence alignment.
- float vrna_eval_circ_consensus_structure (const char **alignment, const char *structure)

Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the sequences are circular.

- float vrna_eval_gquad_consensus_structure (const char **alignment, const char *structure)
 - Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the structure may contain G-Quadruplexes.
- float vrna_eval_circ_gquad_consensus_structure (const char **alignment, const char *structure)

Evaluate the free energy of a multiple sequence alignment/consensus structure pair where the sequence is circular and the structure may contain G-Quadruplexes.

Evaluate the free energy of a consensus structure for an RNA sequence alignment and print contributions per loop.

• float vrna_eval_consensus_structure_simple_v (const char **alignment, const char *structure, int verbosity level, FILE *file)

Evaluate the free energy of a consensus structure for an RNA sequence alignment and print contributions per loop.

 float vrna_eval_circ_consensus_structure_v (const char **alignment, const char *structure, int verbosity level, FILE *file)

Evaluate the free energy of a consensus structure for an alignment of circular RNA sequences and print contributions per loop.

• float vrna_eval_gquad_consensus_structure_v (const char **alignment, const char *structure, int verbosity level, FILE *file)

Evaluate the free energy of a consensus structure for an RNA sequence alignment, allow for annotated G-← Quadruplexes in the structure and print contributions per loop.

• float vrna_eval_circ_gquad_consensus_structure_v (const char **alignment, const char *structure, int verbosity_level, FILE *file)

Evaluate the free energy of a consensus structure for an alignment of circular RNA sequences, allow for annotated G-Quadruplexes in the structure and print contributions per loop.

Simplified Energy Evaluation with Sequence String and Structure Pair Table

- int vrna_eval_structure_pt_simple (const char *string, const short *pt)
 - Calculate the free energy of an already folded RNA.
- int vrna_eval_structure_pt_simple_verbose (const char *string, const short *pt, FILE *file)
 Calculate the free energy of an already folded RNA.
- int vrna_eval_structure_pt_simple_v (const char *string, const short *pt, int verbosity_level, FILE *file)

Calculate the free energy of an already folded RNA.

Simplified Energy Evaluation with Sequence Alignment and Consensus Structure Pair Table

- int vrna_eval_consensus_structure_pt_simple (const char **alignment, const short *pt)

 Evaluate the Free Energy of a Consensus Secondary Structure given a Sequence Alignment.
- int vrna_eval_consensus_structure_pt_simple_verbose (const char **alignment, const short *pt, FILE *file)
- int vrna_eval_consensus_structure_pt_simple_v (const char **alignment, const short *pt, int verbosity_level, FILE *file)

Variables

· int cut_point

first pos of second seq for cofolding

• int eos_debug

verbose info from energy_of_struct

18.34.1 Detailed Description

Functions and variables related to energy evaluation of sequence/structure pairs.

18.35 ViennaRNA/exterior_loops.h File Reference

Use ViennaRNA/loops/external.h instead.

Include dependency graph for exterior_loops.h:

18.35.1 Detailed Description

Use ViennaRNA/loops/external.h instead.

Deprecated Use ViennaRNA/loops/external.h instead

18.36 ViennaRNA/file_formats.h File Reference

Use ViennaRNA/io/file_formats.h instead.

Include dependency graph for file_formats.h:

18.36.1 Detailed Description

Use ViennaRNA/io/file_formats.h instead.

Deprecated Use ViennaRNA/io/file_formats.h instead

18.37 ViennaRNA/io/file_formats.h File Reference

Read and write different file formats for RNA sequences, structures.

Include dependency graph for file formats.h: This graph shows which files directly or indirectly include this file:

Macros

#define VRNA OPTION MULTILINE 32U

Tell a function that an input is assumed to span several lines.

#define VRNA_CONSTRAINT_MULTILINE 32U

parse multiline constraint

Functions

 $\bullet \ \ void \ vrna_file_helixlist \ (const \ char \ *seq, \ const \ char \ *db, \ float \ energy, \ FILE \ *file)\\$

Print a secondary structure as helix list.

• void vrna_file_connect (const char *seq, const char *db, float energy, const char *identifier, FILE *file)

Print a secondary structure as connect table.

• void vrna_file_bpseq (const char *seq, const char *db, FILE *file)

Print a secondary structure in bpseq format.

• void vrna_file_json (const char *seq, const char *db, double energy, const char *identifier, FILE *file)

Print a secondary structure in jsonformat.

- unsigned int vrna_file_fasta_read_record (char **header, char **sequence, char ***rest, FILE *file, unsigned int options)
- char * vrna_extract_record_rest_structure (const char **lines, unsigned int length, unsigned int option)
 Extract a dot-bracket structure string from (multiline)character array.
- int vrna_file_SHAPE_read (const char *file_name, int length, double default_value, char *sequence, double *values)

Read data from a given SHAPE reactivity input file.

void vrna_extract_record_rest_constraint (char **cstruc, const char **lines, unsigned int option)

Extract a hard constraint encoded as pseudo dot-bracket string.

unsigned int read record (char **header, char **sequence, char ***rest, unsigned int options)

Get a data record from stdin.

18.37.1 Detailed Description

Read and write different file formats for RNA sequences, structures.

18.38 ViennaRNA/file formats msa.h File Reference

Use ViennaRNA/io/file formats msa.h instead.

Include dependency graph for file_formats_msa.h:

18.38.1 Detailed Description

Use ViennaRNA/io/file formats msa.h instead.

Deprecated Use ViennaRNA/io/file_formats_msa.h instead

18.39 ViennaRNA/io/file_formats_msa.h File Reference

Functions dealing with file formats for Multiple Sequence Alignments (MSA)

Include dependency graph for file_formats_msa.h: This graph shows which files directly or indirectly include this file:

Macros

#define VRNA FILE FORMAT MSA CLUSTAL 1U

Option flag indicating ClustalW formatted files.

#define VRNA FILE FORMAT MSA STOCKHOLM 2U

Option flag indicating Stockholm 1.0 formatted files.

#define VRNA FILE FORMAT MSA FASTA 4U

Option flag indicating FASTA (Pearson) formatted files.

• #define VRNA_FILE_FORMAT_MSA_MAF 8U

Option flag indicating MAF formatted files.

#define VRNA_FILE_FORMAT_MSA_MIS 16U

Option flag indicating most informative sequence (MIS) output.

#define VRNA_FILE_FORMAT_MSA_DEFAULT

Option flag indicating the set of default file formats.

#define VRNA FILE FORMAT MSA NOCHECK 4096U

Option flag to disable validation of the alignment.

• #define VRNA_FILE_FORMAT_MSA_UNKNOWN 8192U

Return flag of vrna_file_msa_detect_format() to indicate unknown or malformatted alignment.

• #define VRNA FILE FORMAT MSA APPEND 16384U

Option flag indicating to append data to a multiple sequence alignment file rather than overwriting it.

#define VRNA_FILE_FORMAT_MSA_QUIET 32768U

Option flag to suppress unnecessary spam messages on stderr

• #define VRNA_FILE_FORMAT_MSA_SILENT 65536U

Option flag to completely silence any warnings on stderr

Functions

• int vrna_file_msa_read (const char *filename, char ***names, char ***aln, char **id, char **structure, unsigned int options)

Read a multiple sequence alignment from file.

• int vrna_file_msa_read_record (FILE *fp, char ***names, char ***aln, char **id, char **structure, unsigned int options)

Read a multiple sequence alignment from file handle.

• unsigned int vrna_file_msa_detect_format (const char *filename, unsigned int options)

Detect the format of a multiple sequence alignment file.

• int vrna_file_msa_write (const char *filename, const char **names, const char **aln, const char *id, const char *structure, const char *source, unsigned int options)

Write multiple sequence alignment file.

18.39.1 Detailed Description

Functions dealing with file formats for Multiple Sequence Alignments (MSA)

, ,

18.40 ViennaRNA/file_utils.h File Reference

Use ViennaRNA/io/utils.h instead.

Include dependency graph for file_utils.h:

18.40.1 Detailed Description

Use ViennaRNA/io/utils.h instead.

Deprecated Use ViennaRNA/io/utils.h instead

18.41 ViennaRNA/findpath.h File Reference

Use ViennaRNA/landscape/findpath.h instead.

Include dependency graph for findpath.h:

18.41.1 Detailed Description

Use ViennaRNA/landscape/findpath.h instead.

Deprecated Use ViennaRNA/landscape/findpath.h instead

18.42 ViennaRNA/landscape/findpath.h File Reference

A breadth-first search heuristic for optimal direct folding paths.

Include dependency graph for findpath.h: This graph shows which files directly or indirectly include this file:

Functions

- int vrna_path_findpath_saddle (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width)

 Find energy of a saddle point between 2 structures (search only direct path)
- int vrna_path_findpath_saddle_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width, int maxE)

Find energy of a saddle point between 2 structures (search only direct path)

- vrna_path_t * vrna_path_findpath (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width)
 Find refolding path between 2 structures (search only direct path)
- vrna_path_t * vrna_path_findpath_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int width, int maxE)

Find refolding path between 2 structures (search only direct path)

• int find_saddle (const char *seq, const char *s1, const char *s2, int width)

Find energy of a saddle point between 2 structures (search only direct path)

void free_path (vrna_path_t *path)

Free memory allocated by get_path() function.

vrna_path_t * get_path (const char *seq, const char *s1, const char *s2, int width)

Find refolding path between 2 structures (search only direct path)

18.42.1 Detailed Description

A breadth-first search heuristic for optimal direct folding paths.

18.43 ViennaRNA/fold.h File Reference

MFE calculations for single RNA sequences.

Include dependency graph for fold.h:

Functions

float fold_par (const char *sequence, char *structure, vrna_param_t *parameters, int is_constrained, int is
 _circular)

Compute minimum free energy and an appropriate secondary structure of an RNA sequence.

float fold (const char *sequence, char *structure)

Compute minimum free energy and an appropriate secondary structure of an RNA sequence.

float circfold (const char *sequence, char *structure)

Compute minimum free energy and an appropriate secondary structure of a circular RNA sequence.

void free arrays (void)

Free arrays for mfe folding.

• void update_fold_params (void)

Recalculate energy parameters.

void update_fold_params_par (vrna_param_t *parameters)

Recalculate energy parameters.

- void export fold arrays (int **f5 p, int **c p, int **fML p, int **fM1 p, int **indx p, char **ptype p)
- void export_fold_arrays_par (int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p, vrna_param_t **P_p)
- void export_circfold_arrays (int *Fc_p, int *FcH_p, int *FcH_p, int *FcM_p, int **fM2_p, int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p)
- void export_circfold_arrays_par (int *Fc_p, int *Fcl_p, int *Fcl_p, int *FcM_p, int **fM2_p, int **f5_p, int **c_p, int **fML_p, int **fM1_p, int **indx_p, char **ptype_p, vrna_param_t **P_p)
- int LoopEnergy (int n1, int n2, int type, int type 2, int si1, int sj1, int sp1, int sq1)
- int HairpinE (int size, int type, int si1, int sj1, const char *string)
- void initialize_fold (int length)
- char * backtrack_fold_from_pair (char *sequence, int i, int j)

18.43.1 Detailed Description

MFE calculations for single RNA sequences.

18.44 ViennaRNA/fold_compound.h File Reference

The Basic Fold Compound API.

Include dependency graph for fold compound.h: This graph shows which files directly or indirectly include this file:

Data Structures

• struct vrna_fc_s

The most basic data structure required by many functions throughout the RNAlib. More...

Macros

• #define VRNA STATUS MFE PRE (unsigned char)1

Status message indicating that MFE computations are about to begin.

#define VRNA_STATUS_MFE_POST (unsigned char)2

Status message indicating that MFE computations are finished.

• #define VRNA_STATUS_PF_PRE (unsigned char)3

Status message indicating that Partition function computations are about to begin.

#define VRNA_STATUS_PF_POST (unsigned char)4

Status message indicating that Partition function computations are finished.

• #define VRNA OPTION DEFAULT 0U

Option flag to specify default settings/requirements.

#define VRNA_OPTION_MFE 1U

Option flag to specify requirement of Minimum Free Energy (MFE) DP matrices and corresponding set of energy parameters.

• #define VRNA OPTION PF 2U

Option flag to specify requirement of Partition Function (PF) DP matrices and corresponding set of Boltzmann factors.

• #define VRNA_OPTION_HYBRID 4U

Option flag to specify requirement of dimer DP matrices.

• #define VRNA_OPTION_EVAL_ONLY 8U

Option flag to specify that neither MFE, nor PF DP matrices are required.

#define VRNA_OPTION_WINDOW 16U

Option flag to specify requirement of DP matrices for local folding approaches.

Typedefs

• typedef struct vrna_fc_s vrna_fold_compound_t

Typename for the fold_compound data structure vrna_fc_s.

typedef void() vrna_callback_free_auxdata(void *data)

Callback to free memory allocated for auxiliary user-provided data.

typedef void() vrna_callback_recursion_status(unsigned char status, void *data)

Callback to perform specific user-defined actions before, or after recursive computations.

Enumerations

• enum vrna_fc_type_e { VRNA_FC_TYPE_SINGLE, VRNA_FC_TYPE_COMPARATIVE }

An enumerator that is used to specify the type of a vrna_fold_compound_t.

Functions

vrna_fold_compound_t * vrna_fold_compound (const char *sequence, vrna_md_t *md_p, unsigned int options)

Retrieve a vrna_fold_compound_t data structure for single sequences and hybridizing sequences.

vrna_fold_compound_t * vrna_fold_compound_comparative (const char **sequences, vrna_md_t *md_p, unsigned int options)

Retrieve a vrna_fold_compound_t data structure for sequence alignments.

void vrna fold compound free (vrna fold compound t *fc)

Free memory occupied by a vrna_fold_compound_t.

 void vrna_fold_compound_add_auxdata (vrna_fold_compound_t *fc, void *data, vrna_callback_free_auxdata *f)

Add auxiliary data to the vrna_fold_compound_t.

• void vrna_fold_compound_add_callback (vrna_fold_compound_t *fc, vrna_callback_recursion_status *f)

Add a recursion status callback to the vrna_fold_compound_t.

18.44.1 Detailed Description

The Basic Fold Compound API.

18.45 ViennaRNA/fold_vars.h File Reference

Here all all declarations of the global variables used throughout RNAlib.

Include dependency graph for fold vars.h: This graph shows which files directly or indirectly include this file:

Variables

int fold_constrained

Global switch to activate/deactivate folding with structure constraints.

int csv

generate comma seperated output

- char * RibosumFile
- · int james rule
- int logML
- int cut_point

Marks the position (starting from 1) of the first nucleotide of the second molecule within the concatenated sequence.

bondT * base pair

Contains a list of base pairs after a call to fold().

FLT_OR_DBL * pr

A pointer to the base pair probability matrix.

int * iindx

index array to move through pr.

18.45.1 Detailed Description

Here all all declarations of the global variables used throughout RNAlib.

18.45.2 Variable Documentation

18.45.2.1 RibosumFile

char* RibosumFile

warning this variable will vanish in the future ribosums will be compiled in instead

18.45.2.2 james_rule

int james_rule

interior loops of size 2 get energy 0.8Kcal and no mismatches, default 1

18.45.2.3 logML

int logML

use logarithmic multiloop energy function

18.45.2.4 cut_point

int cut_point

Marks the position (starting from 1) of the first nucleotide of the second molecule within the concatenated sequence.

To evaluate the energy of a duplex structure (a structure formed by two strands), concatenate the to sequences and set it to the first base of the second strand in the concatenated sequence. The default value of -1 stands for single molecule folding. The cut_point variable is also used by vrna_file_PS_rnaplot() and PS_dot_plot() to mark the chain break in postscript plots.

18.45.2.5 base_pair

bondT* base_pair

Contains a list of base pairs after a call to fold().

base_pair[0].i contains the total number of pairs.

Deprecated Do not use this variable anymore!

```
18.45.2.6 pr
```

```
FLT_OR_DBL* pr
```

A pointer to the base pair probability matrix.

Deprecated Do not use this variable anymore!

18.45.2.7 iindx

```
int* iindx
```

index array to move through pr.

The probability for base i and j to form a pair is in pr[iindx[i]-j].

Deprecated Do not use this variable anymore!

18.46 ViennaRNA/gquad.h File Reference

G-quadruplexes.

Include dependency graph for gquad.h:

Functions

- int * get_gquad_matrix (short *S, vrna_param_t *P)
 Get a triangular matrix prefilled with minimum free energy contributions of G-quadruplexes.
- int parse_gquad (const char *struc, int *L, int I[3])
- PRIVATE int backtrack_GQuad_IntLoop (int c, int i, int j, int type, short *S, int *ggg, int *index, int *p, int *q, vrna_param_t *P)
- PRIVATE int backtrack_GQuad_IntLoop_L (int c, int i, int j, int type, short *S, int **ggg, int maxdist, int *p, int *q, vrna_param_t *P)

18.46.1 Detailed Description

G-quadruplexes.

18.47 ViennaRNA/grammar.h File Reference

Implementations for the RNA folding grammar.

Include dependency graph for grammar.h: This graph shows which files directly or indirectly include this file:

Data Structures

struct vrna_gr_aux_s

18.47.1 Detailed Description

Implementations for the RNA folding grammar.

18.48 ViennaRNA/hairpin_loops.h File Reference

Use ViennaRNA/loops/hairpin.h instead.

Include dependency graph for hairpin_loops.h:

18.48.1 Detailed Description

Use ViennaRNA/loops/hairpin.h instead.

Deprecated Use ViennaRNA/loops/hairpin.h instead

18.49 ViennaRNA/interior_loops.h File Reference

Use ViennaRNA/loops/internal.h instead.

Include dependency graph for interior_loops.h:

18.49.1 Detailed Description

Use ViennaRNA/loops/internal.h instead.

Deprecated Use ViennaRNA/loops/internal.h instead

18.50 ViennaRNA/inverse.h File Reference

Inverse folding routines.

Functions

float inverse_fold (char *start, const char *target)

Find sequences with predefined structure.

float inverse_pf_fold (char *start, const char *target)

Find sequence that maximizes probability of a predefined structure.

Variables

char * symbolset

This global variable points to the allowed bases, initially "AUGC". It can be used to design sequences from reduced alphabets.

- · float final cost
- · int give up
- · int inv_verbose

18.50.1 Detailed Description

Inverse folding routines.

18.51 ViennaRNA/landscape/move.h File Reference

Methods to operate with structural neighbors of RNA secondary structures.

This graph shows which files directly or indirectly include this file:

Data Structures

struct vrna_move_s

An atomic representation of the transition / move from one structure to its neighbor. More...

Macros

#define VRNA MOVESET INSERTION 4

Option flag indicating insertion move.

#define VRNA_MOVESET_DELETION 8

Option flag indicating deletion move.

• #define VRNA_MOVESET_SHIFT 16

Option flag indicating shift move.

• #define VRNA MOVESET NO LP 32

Option flag indicating moves without lonely base pairs.

• #define VRNA_MOVESET_DEFAULT (VRNA_MOVESET_INSERTION | VRNA_MOVESET_DELETION)

Option flag indicating default move set, i.e. insertions/deletion of a base pair.

Typedefs

• typedef struct vrna_move_s vrna_move_t

A single move that transforms a secondary structure into one of its neighbors.

Functions

vrna_move_t vrna_move_init (int pos_5, int pos_3)

Create an atomic move.

- void vrna_move_list_free (vrna_move_t *moves)
- void vrna_move_apply (short *pt, const vrna_move_t *m)

Apply a particular move / transition to a secondary structure, i.e. transform a structure.

int vrna_move_is_removal (const vrna_move_t *m)

Test whether a move is a base pair removal.

int vrna_move_is_insertion (const vrna_move_t *m)

Test whether a move is a base pair insertion.

int vrna_move_is_shift (const vrna_move_t *m)

Test whether a move is a base pair shift.

int vrna_move_compare (const vrna_move_t *a, const vrna_move_t *b, const short *pt)

Compare two moves.

18.51.1 Detailed Description

Methods to operate with structural neighbors of RNA secondary structures.

18.52 ViennaRNA/landscape/paths.h File Reference

API for computing (optimal) (re-)folding paths between secondary structures.

Include dependency graph for paths.h: This graph shows which files directly or indirectly include this file:

Data Structures

struct vrna path s

An element of a refolding path list. More...

Macros

• #define VRNA_PATH_TYPE_DOT_BRACKET 1U

Flag to indicate producing a (re-)folding path as list of dot-bracket structures.

• #define VRNA_PATH_TYPE_MOVES 2U

Flag to indicate producing a (re-)folding path as list of transition moves.

Typedefs

· typedef struct vrna_path_s vrna_path_t

Typename for the refolding path data structure vrna_path_s.

typedef struct vrna_path_options_s * vrna_path_options_t

Options data structure for (re-)folding path implementations.

typedef struct vrna_path_s path_t

Old typename of vrna_path_s.

Functions

void vrna_path_free (vrna_path_t *path)

Release (free) memory occupied by a (re-)folding path.

void vrna_path_options_free (vrna_path_options_t options)

Release (free) memory occupied by an options data structure for (re-)folding path implementations.

vrna_path_options_t vrna_path_options_findpath (int width, unsigned int type)

Create options data structure for findpath direct (re-)folding path heuristic.

vrna_path_t * vrna_path_direct (vrna_fold_compound_t *fc, const char *s1, const char *s2, vrna_path_options_t options)

Determine an optimal direct (re-)folding path between two secondary structures.

vrna_path_t * vrna_path_direct_ub (vrna_fold_compound_t *fc, const char *s1, const char *s2, int maxE, vrna_path_options_t options)

Determine an optimal direct (re-)folding path between two secondary structures.

18.52.1 Detailed Description

API for computing (optimal) (re-)folding paths between secondary structures.

18.53 ViennaRNA/Lfold.h File Reference

Functions for locally optimal MFE structure prediction.

Include dependency graph for Lfold.h:

Functions

- float Lfold (const char *string, const char *structure, int maxdist)
 The local analog to fold().
- float Lfoldz (const char *string, const char *structure, int maxdist, int zsc, double min_z)

18.53.1 Detailed Description

Functions for locally optimal MFE structure prediction.

18.54 ViennaRNA/loop_energies.h File Reference

Use ViennaRNA/loops/all.h instead.

Include dependency graph for loop_energies.h:

18.54.1 Detailed Description

Use ViennaRNA/loops/all.h instead.

Deprecated Use ViennaRNA/loops/all.h instead

18.55 ViennaRNA/loops/all.h File Reference

Energy evaluation for MFE and partition function calculations.

Include dependency graph for all.h: This graph shows which files directly or indirectly include this file:

18.55.1 Detailed Description

Energy evaluation for MFE and partition function calculations.

This file contains functions for the calculation of the free energy ΔG of a hairpin- [E_Hairpin()] or interior-loop [E_IntLoop()].

The unit of the free energy returned is $10^{-2} * kcal/mol$

In case of computing the partition function, this file also supplies functions which return the Boltzmann weights $e^{-\Delta G/kT}$ for a hairpin-[exp E Hairpin()] or interior-loop [exp E IntLoop()].

18.56 ViennaRNA/loops/external.h File Reference

Energy evaluation of exterior loops for MFE and partition function calculations.

Include dependency graph for external.h: This graph shows which files directly or indirectly include this file:

Functions

```
    int E_Stem (int type, int si1, int sj1, int extLoop, vrna_param_t *P)
```

Compute the energy contribution of a stem branching off a loop-region.

- $\bullet \ \ \mathsf{FLT_OR_DBL} \ \mathsf{exp_E_ExtLoop} \ (\mathsf{int} \ \mathsf{type}, \ \mathsf{int} \ \mathsf{si1}, \ \mathsf{int} \ \mathsf{sj1}, \ \mathsf{vrna_exp_param_t} \ *\mathsf{P})$
- FLT OR DBL exp E Stem (int type, int si1, int sj1, int extLoop, vrna exp param t *P)

Basic free energy interface

```
• int vrna_E_ext_stem (unsigned int type, int n5d, int n3d, vrna_param_t *p)
```

Evaluate a stem branching off the exterior loop.

int vrna_E_ext_loop (vrna_fold_compound_t *fc, int i, int j)

Evaluate the free energy of a base pair in the exterior loop.

- int vrna_E_ext_loop_5 (vrna_fold_compound_t *fc)
- int vrna_E_ext_loop_3 (vrna_fold_compound_t *fc, int i)

Boltzmann weight (partition function) interface

```
typedef struct vrna_mx_pf_aux_el_s * vrna_mx_pf_aux_el_t
```

Auxiliary helper arrays for fast exterior loop computations.

• FLT_OR_DBL vrna_exp_E_ext_stem (unsigned int type, int n5d, int n3d, vrna_exp_param_t *p)

Evaluate a stem branching off the exterior loop (Boltzmann factor version)

- struct vrna_mx_pf_aux_el_s * vrna_exp_E_ext_fast_init (vrna_fold_compound_t *fc)
- void vrna exp E ext fast rotate (struct vrna mx pf aux el s *aux mx)
- void vrna_exp_E_ext_fast_free (struct vrna_mx_pf_aux_el_s *aux_mx)
- FLT_OR_DBL vrna_exp_E_ext_fast (vrna_fold_compound_t *fc, int i, int j, struct vrna_mx_pf_aux_el_
 s *aux mx)
- void vrna_exp_E_ext_fast_update (vrna_fold_compound_t *fc, int j, struct vrna_mx_pf_aux_el_s *aux_mx)

18.56.1 Detailed Description

Energy evaluation of exterior loops for MFE and partition function calculations.

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18.57 ViennaRNA/loops/hairpin.h File Reference

Energy evaluation of hairpin loops for MFE and partition function calculations.

Include dependency graph for hairpin.h: This graph shows which files directly or indirectly include this file:

Functions

int vrna_BT_hp_loop (vrna_fold_compound_t *fc, int i, int j, int en, vrna_bp_stack_t *bp_stack, int *stack_
 count)

Backtrack a hairpin loop closed by (i, j).

Basic free energy interface

- int vrna_E_hp_loop (vrna_fold_compound_t *fc, int i, int j)
 - Evaluate the free energy of a hairpin loop and consider hard constraints if they apply.
- int vrna_E_ext_hp_loop (vrna_fold_compound_t *fc, int i, int j)
 - Evaluate the free energy of an exterior hairpin loop and consider possible hard constraints.
- int vrna_eval_ext_hp_loop (vrna_fold_compound_t *fc, int i, int j)
 - Evaluate free energy of an exterior hairpin loop.
- int vrna_eval_hp_loop (vrna_fold_compound_t *fc, int i, int j)
 - Evaluate free energy of a hairpin loop.
- PRIVATE int E_Hairpin (int size, int type, int si1, int sj1, const char *string, vrna_param_t *P)
 Compute the Energy of a hairpin-loop.

Boltzmann weight (partition function) interface

- PRIVATE FLT_OR_DBL exp_E_Hairpin (int u, int type, short si1, short sj1, const char *string, vrna_exp_param_t *P)
- Compute Boltzmann weight $e^{-\Delta G/kT}$ of a hairpin loop.
- FLT_OR_DBL vrna_exp_E_hp_loop (vrna_fold_compound_t *fc, int i, int j)

High-Level function for hairpin loop energy evaluation (partition function variant)

18.57.1 Detailed Description

Energy evaluation of hairpin loops for MFE and partition function calculations.

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18.58 ViennaRNA/loops/internal.h File Reference

Energy evaluation of interior loops for MFE and partition function calculations.

Include dependency graph for internal.h: This graph shows which files directly or indirectly include this file:

Functions

int vrna_BT_stack (vrna_fold_compound_t *fc, int *i, int *j, int *en, vrna_bp_stack_t *bp_stack, int *stack←
 _count)

Backtrack a stacked pair closed by (i, j).

int vrna_BT_int_loop (vrna_fold_compound_t *fc, int *i, int *j, int en, vrna_bp_stack_t *bp_stack, int *stack
 _count)

Backtrack an interior loop closed by (i, j).

- PRIVATE int E_IntLoop (int n1, int n2, int type, int type_2, int si1, int sj1, int sp1, int sq1, vrna_param_t *P)
- PRIVATE FLT_OR_DBL exp_E_IntLoop (int u1, int u2, int type, int type2, short si1, short sj1, short sp1, short sq1, vrna_exp_param_t *P)

Basic free energy interface

- int vrna_E_int_loop (vrna_fold_compound_t *fc, int i, int j)
- int vrna_eval_int_loop (vrna_fold_compound_t *fc, int i, int j, int k, int l)

Evaluate the free energy contribution of an interior loop with delimiting base pairs (i, j) and (k, l).

- int vrna_E_ext_int_loop (vrna_fold_compound_t *fc, int i, int j, int *ip, int *iq)
- int vrna_E_stack (vrna_fold_compound_t *fc, int i, int j)

Boltzmann weight (partition function) interface

- FLT OR DBL vrna exp E int loop (vrna fold compound t *fc, int i, int j)
- FLT_OR_DBL vrna_exp_E_interior_loop (vrna_fold_compound_t *fc, int i, int j, int k, int l)

18.58.1 Detailed Description

Energy evaluation of interior loops for MFE and partition function calculations.

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18.59 ViennaRNA/loops/multibranch.h File Reference

Energy evaluation of multibranch loops for MFE and partition function calculations.

Include dependency graph for multibranch.h: This graph shows which files directly or indirectly include this file:

Functions

• int vrna_BT_mb_loop (vrna_fold_compound_t *fc, int *i, int *j, int *k, int en, int *component1, int *component2)

Backtrack the decomposition of a multi branch loop closed by (i, j).

Basic free energy interface

- int vrna_E_mb_loop_stack (vrna_fold_compound_t *fc, int i, int j)
 - Evaluate energy of a multi branch helices stacking onto closing pair (i,j)
- int vrna_E_mb_loop_fast (vrna_fold_compound_t *fc, int i, int j, int *dmli1, int *dmli2)
- int **E_ml_rightmost_stem** (int i, int j, vrna_fold_compound_t *fc)
- int vrna_E_ml_stems_fast (vrna_fold_compound_t *fc, int i, int j, int *fmi, int *dmli)

Boltzmann weight (partition function) interface

- typedef struct vrna_mx_pf_aux_ml_s * vrna_mx_pf_aux_ml_t
 Auxiliary helper arrays for fast exterior loop computations.
- FLT_OR_DBL vrna_exp_E_mb_loop_fast (vrna_fold_compound_t *fc, int i, int j, vrna_mx_pf_aux_ml_t aux_mx)
- vrna mx pf aux ml t vrna exp E ml fast init (vrna fold compound t *fc)
- void vrna_exp_E_ml_fast_rotate (vrna_mx_pf_aux_ml_t aux_mx)
- void vrna_exp_E_ml_fast_free (vrna_mx_pf_aux_ml_t aux_mx)
- const FLT OR DBL * vrna exp E ml fast qqm (struct vrna mx pf aux ml s *aux mx)
- const FLT_OR_DBL * vrna_exp_E_ml_fast_qqm1 (struct vrna_mx_pf_aux_ml_s *aux_mx)
- FLT_OR_DBL vrna_exp_E_ml_fast (vrna_fold_compound_t *fc, int i, int j, vrna_mx_pf_aux_ml_t aux_mx)

18.59.1 Detailed Description

Energy evaluation of multibranch loops for MFE and partition function calculations.

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18.60 ViennaRNA/LPfold.h File Reference

Partition function and equilibrium probability implementation for the sliding window algorithm.

Include dependency graph for LPfold.h:

Functions

- void update pf paramsLP (int length)
- vrna_ep_t * pfl_fold (char *sequence, int winSize, int pairSize, float cutoffb, double **pU, vrna_ep_t **dpp2,
 FILE *pUfp, FILE *spup)

Compute partition functions for locally stable secondary structures.

vrna_ep_t * pfl_fold_par (char *sequence, int winSize, int pairSize, float cutoffb, double **pU, vrna_ep_t
 **dpp2, FILE *pUfp, FILE *spup, vrna exp param t *parameters)

Compute partition functions for locally stable secondary structures.

void putoutpU_prob (double **pU, int length, int ulength, FILE *fp, int energies)

Writes the unpaired probabilities (pU) or opening energies into a file.

• void putoutpU_prob_bin (double **pU, int length, int ulength, FILE *fp, int energies)

Writes the unpaired probabilities (pU) or opening energies into a binary file.

void init_pf_foldLP (int length)

18.60.1 Detailed Description

Partition function and equilibrium probability implementation for the sliding window algorithm.

This file contains the implementation for sliding window partition function and equilibrium probabilities. It also provides the unpaired probability implementation from Bernhart et al. 2011 [4]

18.60.2 Function Documentation

18.60.2.1 init_pf_foldLP()

Dunno if this function was ever used by external programs linking to RNAlib, but it was declared PUBLIC before. Anyway, never use this function as it will be removed soon and does nothing at all

18.61 ViennaRNA/MEA.h File Reference

Computes a MEA (maximum expected accuracy) structure.

Include dependency graph for MEA.h:

Functions

- char * vrna_MEA (vrna_fold_compound_t *fc, double gamma, float *mea)

 Compute a MEA (maximum expected accuracy) structure.
- char * vrna_MEA_from_plist (vrna_ep_t *plist, const char *sequence, double gamma, vrna_md_t *md, float *mea)

Compute a MEA (maximum expected accuracy) structure from a list of probabilities.

float MEA (plist *p, char *structure, double gamma)

Computes a MEA (maximum expected accuracy) structure.

18.61.1 Detailed Description

Computes a MEA (maximum expected accuracy) structure.

18.62 ViennaRNA/mfe.h File Reference

Compute Minimum Free energy (MFE) and backtrace corresponding secondary structures from RNA sequence data.

Include dependency graph for mfe.h: This graph shows which files directly or indirectly include this file:

Functions

• float vrna_backtrack5 (vrna_fold_compound_t *fc, unsigned int length, char *structure)

Backtrack an MFE (sub)structure.

Basic global MFE prediction interface

- float vrna mfe (vrna fold compound t *vc, char *structure)
 - Compute minimum free energy and an appropriate secondary structure of an RNA sequence, or RNA sequence alignment.
- float vrna_mfe_dimer (vrna_fold_compound t *vc, char *structure)
 - Compute the minimum free energy of two interacting RNA molecules.

Simplified global MFE prediction using sequence(s) or multiple sequence alignment(s)

- float vrna_fold (const char *sequence, char *structure)
 - Compute Minimum Free Energy (MFE), and a corresponding secondary structure for an RNA sequence.
- float vrna circfold (const char *sequence, char *structure)
 - Compute Minimum Free Energy (MFE), and a corresponding secondary structure for a circular RNA sequence.
- float vrna alifold (const char **sequences, char *structure)
 - Compute Minimum Free Energy (MFE), and a corresponding consensus secondary structure for an RNA sequence alignment using a comparative method.
- float vrna_circalifold (const char **sequences, char *structure)
 - Compute Minimum Free Energy (MFE), and a corresponding consensus secondary structure for a sequence alignment of circular RNAs using a comparative method.
- float vrna_cofold (const char *sequence, char *structure)
 - Compute Minimum Free Energy (MFE), and a corresponding secondary structure for two dimerized RNA sequences.

18.62.1 Detailed Description

Compute Minimum Free energy (MFE) and backtrace corresponding secondary structures from RNA sequence data.

, This file includes (almost) all function declarations within the RNAlib that are related to MFE folding...

18.63 ViennaRNA/mfe window.h File Reference

Compute local Minimum Free Energy (MFE) using a sliding window approach and backtrace corresponding secondary structures.

Include dependency graph for mfe_window.h: This graph shows which files directly or indirectly include this file:

Typedefs

• typedef void() vrna_mfe_window_callback(int start, int end, const char *structure, float en, void *data)

The default callback for sliding window MFE structure predictions.

Functions

Basic local (sliding window) MFE prediction interface

- float vrna_mfe_window (vrna_fold_compound_t *vc, FILE *file)

 Local MFE prediction using a sliding window approach.
- float vrna mfe window cb (vrna fold compound t *vc, vrna mfe window callback *cb, void *data)
- float vrna_mfe_window_zscore (vrna_fold_compound_t *vc, double min_z, FILE *file)
 Local MFE prediction using a sliding window approach (with z-score cut-off)
- float vrna_mfe_window_zscore_cb (vrna_fold_compound_t *vc, double min_z, vrna_mfe_window_

 zscore callback *cb, void *data)

Simplified local MFE prediction using sequence(s) or multiple sequence alignment(s)

- float vrna_Lfold (const char *string, int window_size, FILE *file)
 Local MFE prediction using a sliding window approach (simplified interface)
- float vrna_Lfold_cb (const char *string, int window_size, vrna_mfe_window_callback *cb, void *data)
- float vrna_Lfoldz (const char *string, int window_size, double min_z, FILE *file)
 Local MFE prediction using a sliding window approach with z-score cut-off (simplified interface)
- float vrna_Lfoldz_cb (const char *string, int window_size, double min_z, vrna_mfe_window_zscore_
 callback *cb, void *data)
- float **vrna_aliLfold** (const char **alignment, int maxdist, FILE *fp)
- float vrna_aliLfold_cb (const char **alignment, int maxdist, vrna_mfe_window_callback *cb, void *data)

18.63.1 Detailed Description

Compute local Minimum Free Energy (MFE) using a sliding window approach and backtrace corresponding secondary structures.

, This file includes the interface to all functions related to predicting locally stable secondary structures.

18.64 ViennaRNA/mm.h File Reference

Several Maximum Matching implementations.

Include dependency graph for mm.h:

Functions

- int vrna_maximum_matching (vrna_fold_compound_t *fc)
- int vrna_maximum_matching_simple (const char *sequence)

18.64.1 Detailed Description

Several Maximum Matching implementations.

This file contains the declarations for several maximum matching implementations

18.64.2 Function Documentation

18.64.2.1 vrna_maximum_matching()

SWIG Wrapper Notes This function is attached as method maximum_matching() to objects of type $fold \leftarrow compound$ (i.e. $vrna_fold_compound_t$).

18.64.2.2 vrna_maximum_matching_simple()

SWIG Wrapper Notes This function is available as global function maximum_matching().

18.65 ViennaRNA/model.h File Reference

The model details data structure and its corresponding modifiers.

This graph shows which files directly or indirectly include this file:

Data Structures

struct vrna_md_s

The data structure that contains the complete model details used throughout the calculations. More...

Macros

#define VRNA MODEL DEFAULT TEMPERATURE 37.0

Default temperature for structure prediction and free energy evaluation in ${}^{\circ}C$

#define VRNA_MODEL_DEFAULT_PF_SCALE -1

Default scaling factor for partition function computations.

• #define VRNA MODEL DEFAULT BETA SCALE 1.

Default scaling factor for absolute thermodynamic temperature in Boltzmann factors.

#define VRNA MODEL DEFAULT DANGLES 2

Default dangling end model.

#define VRNA_MODEL_DEFAULT_SPECIAL_HP 1

Default model behavior for lookup of special tri-, tetra-, and hexa-loops.

#define VRNA MODEL DEFAULT NO LP 0

Default model behavior for so-called 'lonely pairs'.

• #define VRNA MODEL DEFAULT NO GU 0

Default model behavior for G-U base pairs.

• #define VRNA_MODEL_DEFAULT_NO_GU_CLOSURE 0

Default model behavior for G-U base pairs closing a loop.

#define VRNA_MODEL_DEFAULT_CIRC 0

Default model behavior to treat a molecule as a circular RNA (DNA)

#define VRNA MODEL DEFAULT GQUAD 0

Default model behavior regarding the treatment of G-Quadruplexes.

• #define VRNA_MODEL_DEFAULT_UNIQ_ML 0

Default behavior of the model regarding unique multi-branch loop decomposition.

• #define VRNA MODEL DEFAULT ENERGY SET 0

Default model behavior on which energy set to use.

#define VRNA MODEL DEFAULT BACKTRACK 1

Default model behavior with regards to backtracking of structures.

• #define VRNA MODEL DEFAULT BACKTRACK TYPE 'F'

Default model behavior on what type of backtracking to perform.

#define VRNA_MODEL_DEFAULT_COMPUTE_BPP 1

Default model behavior with regards to computing base pair probabilities.

#define VRNA MODEL DEFAULT MAX BP SPAN -1

Default model behavior for the allowed maximum base pair span.

#define VRNA MODEL DEFAULT WINDOW SIZE -1

Default model behavior for the sliding window approach.

#define VRNA_MODEL_DEFAULT_LOG_ML 0

Default model behavior on how to evaluate the energy contribution of multi-branch loops.

#define VRNA_MODEL_DEFAULT_ALI_OLD_EN 0

Default model behavior for consensus structure energy evaluation.

#define VRNA_MODEL_DEFAULT_ALI_RIBO 0

Default model behavior for consensus structure co-variance contribution assessment.

• #define VRNA_MODEL_DEFAULT_ALI_CV_FACT 1.

Default model behavior for weighting the co-variance score in consensus structure prediction.

#define VRNA_MODEL_DEFAULT_ALI_NC_FACT 1.

Default model behavior for weighting the nucleotide conservation? in consensus structure prediction.

• #define MAXALPHA 20

Maximal length of alphabet.

Typedefs

typedef struct vrna_md_s vrna_md_t

Typename for the model details data structure vrna_md_s.

Functions

void vrna_md_set_default (vrna_md_t *md)

Apply default model details to a provided vrna_md_t data structure.

void vrna_md_update (vrna_md_t *md)

Update the model details data structure.

vrna_md_t * vrna_md_copy (vrna_md_t *md_to, const vrna_md_t *md_from)

Copy/Clone a vrna_md_t model.

char * vrna_md_option_string (vrna_md_t *md)

Get a corresponding commandline parameter string of the options in a vrna_md_t.

void vrna_md_defaults_reset (vrna_md_t *md_p)

Reset the global default model details to a specific set of parameters, or their initial values.

void vrna md defaults temperature (double T)

Set default temperature for energy evaluation of loops.

double vrna md defaults temperature get (void)

Get default temperature for energy evaluation of loops.

• void vrna md defaults betaScale (double b)

Set default scaling factor of thermodynamic temperature in Boltzmann factors.

double vrna_md_defaults_betaScale_get (void)

Get default scaling factor of thermodynamic temperature in Boltzmann factors.

void vrna_md_defaults_dangles (int d)

Set default dangle model for structure prediction.

• int vrna_md_defaults_dangles_get (void)

Get default dangle model for structure prediction.

void vrna_md_defaults_special_hp (int flag)

Set default behavior for lookup of tabulated free energies for special hairpin loops, such as Tri-, Tetra-, or Hexa-loops.

int vrna_md_defaults_special_hp_get (void)

Get default behavior for lookup of tabulated free energies for special hairpin loops, such as Tri-, Tetra-, or Hexa-loops.

void vrna_md_defaults_noLP (int flag)

Set default behavior for prediction of canonical secondary structures.

int vrna_md_defaults_noLP_get (void)

Get default behavior for prediction of canonical secondary structures.

void vrna_md_defaults_noGU (int flag)

Set default behavior for treatment of G-U wobble pairs.

int vrna_md_defaults_noGU_get (void)

Get default behavior for treatment of G-U wobble pairs.

void vrna_md_defaults_noGUclosure (int flag)

Set default behavior for G-U pairs as closing pair for loops.

int vrna md defaults noGUclosure get (void)

Get default behavior for G-U pairs as closing pair for loops.

void vrna_md_defaults_logML (int flag)

Set default behavior recomputing free energies of multi-branch loops using a logarithmic model.

int vrna md defaults logML get (void)

Get default behavior recomputing free energies of multi-branch loops using a logarithmic model.

• void vrna_md_defaults_circ (int flag)

Set default behavior whether input sequences are circularized.

int vrna_md_defaults_circ_get (void)

Get default behavior whether input sequences are circularized.

void vrna md defaults gquad (int flag)

Set default behavior for treatment of G-Quadruplexes.

int vrna_md_defaults_gquad_get (void)

Get default behavior for treatment of G-Quadruplexes.

void vrna md defaults uniq ML (int flag)

Set default behavior for creating additional matrix for unique multi-branch loop prediction.

int vrna_md_defaults_uniq_ML_get (void)

Get default behavior for creating additional matrix for unique multi-branch loop prediction.

void vrna_md_defaults_energy_set (int e)

Set default energy set.

int vrna_md_defaults_energy_set_get (void)

Get default energy set.

void vrna md defaults backtrack (int flag)

Set default behavior for whether to backtrack secondary structures.

int vrna_md_defaults_backtrack_get (void)

Get default behavior for whether to backtrack secondary structures.

void vrna md defaults backtrack type (char t)

Set default backtrack type, i.e. which DP matrix is used.

char vrna_md_defaults_backtrack_type_get (void)

Get default backtrack type, i.e. which DP matrix is used.

void vrna_md_defaults_compute_bpp (int flag)

Set the default behavior for whether to compute base pair probabilities after partition function computation.

• int vrna_md_defaults_compute_bpp_get (void)

Get the default behavior for whether to compute base pair probabilities after partition function computation.

• void vrna_md_defaults_max_bp_span (int span)

Set default maximal base pair span.

int vrna md defaults max bp span get (void)

Get default maximal base pair span.

void vrna_md_defaults_min_loop_size (int size)

Set default minimal loop size.

int vrna_md_defaults_min_loop_size_get (void)

Get default minimal loop size.

void vrna_md_defaults_window_size (int size)

Set default window size for sliding window structure prediction approaches.

int vrna_md_defaults_window_size_get (void)

Get default window size for sliding window structure prediction approaches.

void vrna_md_defaults_oldAliEn (int flag)

Set default behavior for whether to use old energy model for comparative structure prediction.

• int vrna md defaults oldAliEn get (void)

Get default behavior for whether to use old energy model for comparative structure prediction.

void vrna_md_defaults_ribo (int flag)

Set default behavior for whether to use Ribosum Scoring in comparative structure prediction.

int vrna_md_defaults_ribo_get (void)

Get default behavior for whether to use Ribosum Scoring in comparative structure prediction.

void vrna_md_defaults_cv_fact (double factor)

Set the default co-variance scaling factor used in comparative structure prediction.

double vrna_md_defaults_cv_fact_get (void)

Get the default co-variance scaling factor used in comparative structure prediction.

- void vrna_md_defaults_nc_fact (double factor)
- double vrna_md_defaults_nc_fact_get (void)
- void vrna_md_defaults_sfact (double factor)

Set the default scaling factor used to avoid under-/overflows in partition function computation.

double vrna_md_defaults_sfact_get (void)

Get the default scaling factor used to avoid under-/overflows in partition function computation.

void set_model_details (vrna_md_t *md)

Set default model details.

Variables

· double temperature

Rescale energy parameters to a temperature in degC.

· double pf_scale

A scaling factor used by pf_fold() to avoid overflows.

· int dangles

Switch the energy model for dangling end contributions (0, 1, 2, 3)

int tetra_loop

Include special stabilizing energies for some tri-, tetra- and hexa-loops;.

· int noLonelyPairs

Global switch to avoid/allow helices of length 1.

• int noGU

Global switch to forbid/allow GU base pairs at all.

int no_closingGU

GU allowed only inside stacks if set to 1.

• int circ

backward compatibility variable.. this does not effect anything

· int gquad

Allow G-quadruplex formation.

• int uniq_ML

do ML decomposition uniquely (for subopt)

int energy_set

0 = BP; 1=any with GC; 2=any with AU-parameter

int do_backtrack

do backtracking, i.e. compute secondary structures or base pair probabilities

· char backtrack_type

A backtrack array marker for inverse_fold()

• char * nonstandards

contains allowed non standard base pairs

• int max bp span

Maximum allowed base pair span.

• int oldAliEn

use old alifold energies (with gaps)

int ribo

use ribosum matrices

• int logML

if nonzero use logarithmic ML energy in energy_of_struct

18.65.1 Detailed Description

The model details data structure and its corresponding modifiers.

18.66 ViennaRNA/multibranch_loops.h File Reference

Use ViennaRNA/loops/multibranch.h instead.

Include dependency graph for multibranch loops.h:

18.66.1 Detailed Description

Use ViennaRNA/loops/multibranch.h instead.

Deprecated Use ViennaRNA/loops/multibranch.h instead

18.67 ViennaRNA/naview.h File Reference

Use ViennaRNA/plotting/naview.h instead.

Include dependency graph for naview.h:

18.67.1 Detailed Description

Use ViennaRNA/plotting/naview.h instead.

Deprecated Use ViennaRNA/plotting/naview.h instead

18.68 ViennaRNA/plotting/naview.h File Reference

Implementation of the Naview RNA secondary structure layout algorithm [5].

This graph shows which files directly or indirectly include this file:

Functions

- int vrna_plot_coords_naview (const char *structure, float **x, float **y)
 Compute nucleotide coordinates for secondary structure plot using the Naview algorithm [5].
- int vrna_plot_coords_naview_pt (const short *pt, float **x, float **x)
 Compute nucleotide coordinates for secondary structure plot using the Naview algorithm [5].
- int naview_xy_coordinates (short *pair_table, float *X, float *Y)

18.68.1 Detailed Description

Implementation of the Naview RNA secondary structure layout algorithm [5].

18.69 ViennaRNA/neighbor.h File Reference

Use ViennaRNA/landscape/neighbor.h instead.

Include dependency graph for neighbor.h:

18.69.1 Detailed Description

Use ViennaRNA/landscape/neighbor.h instead.

Deprecated Use ViennaRNA/landscape/neighbor.h instead

18.70 ViennaRNA/landscape/neighbor.h File Reference

Methods to compute the neighbors of an RNA secondary structure.

Include dependency graph for neighbor.h: This graph shows which files directly or indirectly include this file:

Macros

• #define VRNA_NEIGHBOR_CHANGE 1

State indicator for a neighbor that has been changed.

• #define VRNA_NEIGHBOR_INVALID 2

State indicator for a neighbor that has been invalidated.

• #define VRNA NEIGHBOR NEW 3

State indicator for a neighbor that has become newly available.

Typedefs

• typedef void() vrna_callback_move_update(vrna_fold_compound_t *fc, vrna_move_t neighbor, unsigned int state, void *data)

Prototype of the neighborhood update callback.

Functions

- void vrna_loopidx_update (int *loopidx, const short *pt, int length, const vrna_move_t *m)
 - Alters the loopIndices array that was constructed with vrna_loopidx_from_ptable().
- vrna_move_t * vrna_neighbors (vrna_fold_compound_t *vc, const short *pt, unsigned int options)
 - Generate neighbors of a secondary structure.
- - Generate neighbors of a secondary structure (the fast way)
- int vrna_move_neighbor_diff_cb (vrna_fold_compound_t *fc, short *ptable, vrna_move_t move, vrna_callback_move_update *cb, void *data, unsigned int options)
 - Apply a move to a secondary structure and indicate which neighbors have changed consequentially.
- vrna_move_t * vrna_move_neighbor_diff (vrna_fold_compound_t *fc, short *ptable, vrna_move_t move, vrna_move_t **invalid_moves, unsigned int options)

Apply a move to a secondary structure and indicate which neighbors have changed consequentially.

18.70.1 Detailed Description

Methods to compute the neighbors of an RNA secondary structure.

18.71 ViennaRNA/params.h File Reference

Use ViennaRNA/params/basic.h instead.

Include dependency graph for params.h:

18.71.1 Detailed Description

Use ViennaRNA/params/basic.h instead.

Deprecated Use ViennaRNA/params/basic.h instead

18.72 ViennaRNA/params/1.8.4_epars.h File Reference

Free energy parameters for parameter file conversion.

18.72.1 Detailed Description

Free energy parameters for parameter file conversion.

This file contains the free energy parameters used in ViennaRNAPackage 1.8.4. They are summarized in:

D.H.Mathews, J. Sabina, M. ZUker, D.H. Turner "Expanded sequence dependence of thermodynamic parameters improves prediction of RNA secondary structure" JMB, 288, pp 911-940, 1999

Enthalpies taken from:

A. Walter, D Turner, J Kim, M Lyttle, P M"uller, D Mathews, M Zuker "Coaxial stckaing of helices enhances binding of oligoribonucleotides.." PNAS, 91, pp 9218-9222, 1994

D.H. Turner, N. Sugimoto, and S.M. Freier. "RNA Structure Prediction", Ann. Rev. Biophys. Biophys. Chem. 17, 167-192, 1988.

John A.Jaeger, Douglas H.Turner, and Michael Zuker. "Improved predictions of secondary structures for RNA", PNAS, 86, 7706-7710, October 1989.

L. He, R. Kierzek, J. SantaLucia, A.E. Walter, D.H. Turner "Nearest-Neughbor Parameters for GU Mismatches...." Biochemistry 1991, 30 11124-11132

A.E. Peritz, R. Kierzek, N, Sugimoto, D.H. Turner "Thermodynamic Study of Internal Loops in Oligoribonucleotides..." Biochemistry 1991, 30, 6428–6435

18.73 ViennaRNA/params/1.8.4_intloops.h File Reference

Free energy parameters for interior loop contributions needed by the parameter file conversion functions.

18.73.1 Detailed Description

Free energy parameters for interior loop contributions needed by the parameter file conversion functions.

18.74 ViennaRNA/params/basic.h File Reference

Functions to deal with sets of energy parameters.

Include dependency graph for basic.h: This graph shows which files directly or indirectly include this file:

Data Structures

struct vrna_param_s

The datastructure that contains temperature scaled energy parameters. More...

• struct vrna_exp_param_s

The data structure that contains temperature scaled Boltzmann weights of the energy parameters. More...

Typedefs

typedef struct vrna param s vrna param t

Typename for the free energy parameter data structure vrna_params.

typedef struct vrna_exp_param_s vrna_exp_param_t

Typename for the Boltzmann factor data structure vrna_exp_params.

typedef struct vrna_param_s paramT

Old typename of vrna_param_s.

typedef struct vrna_exp_param_s pf_paramT

Old typename of vrna_exp_param_s.

Functions

vrna param t * vrna params (vrna md t *md)

Get a data structure containing prescaled free energy parameters.

vrna_param_t * vrna_params_copy (vrna_param_t *par)

Get a copy of the provided free energy parameters.

vrna_exp_param_t * vrna_exp_params (vrna_md_t *md)

Get a data structure containing prescaled free energy parameters already transformed to Boltzmann factors.

vrna_exp_param_t * vrna_exp_params_comparative (unsigned int n_seq, vrna_md_t *md)

Get a data structure containing prescaled free energy parameters already transformed to Boltzmann factors (alifold version)

vrna exp_param_t * vrna_exp_params_copy (vrna_exp_param_t *par)

Get a copy of the provided free energy parameters (provided as Boltzmann factors)

void vrna_params_subst (vrna_fold_compound_t *vc, vrna_param_t *par)

Update/Reset energy parameters data structure within a vrna_fold_compound_t.

void vrna_exp_params_subst (vrna_fold_compound_t *vc, vrna_exp_param_t *params)

Update the energy parameters for subsequent partition function computations.

void vrna_exp_params_rescale (vrna_fold_compound_t *vc, double *mfe)

Rescale Boltzmann factors for partition function computations.

void vrna_params_reset (vrna_fold_compound_t *vc, vrna_md_t *md_p)

Reset free energy parameters within a vrna_fold_compound_t according to provided, or default model details.

void vrna_exp_params_reset (vrna_fold_compound_t *vc, vrna_md_t *md_p)

Reset Boltzmann factors for partition function computations within a vrna_fold_compound_t according to provided, or default model details.

- vrna exp param t * get scaled pf parameters (void)
- vrna_exp_param_t * get_boltzmann_factors (double temperature, double betaScale, vrna_md_t md, double pf_scale)

Get precomputed Boltzmann factors of the loop type dependent energy contributions with independent thermodynamic temperature.

vrna_exp_param_t * get_boltzmann_factor_copy (vrna_exp_param_t *parameters)

Get a copy of already precomputed Boltzmann factors.

vrna_exp_param_t * get_scaled_alipf_parameters (unsigned int n_seq)

Get precomputed Boltzmann factors of the loop type dependent energy contributions (alifold variant)

vrna_exp_param_t * get_boltzmann_factors_ali (unsigned int n_seq, double temperature, double betaScale, vrna md t md, double pf scale)

Get precomputed Boltzmann factors of the loop type dependent energy contributions (alifold variant) with independent thermodynamic temperature.

vrna_param_t * scale_parameters (void)

Get precomputed energy contributions for all the known loop types.

vrna_param_t * get_scaled_parameters (double temperature, vrna_md_t md)

Get precomputed energy contributions for all the known loop types.

18.74.1 Detailed Description

Functions to deal with sets of energy parameters.

18.75 ViennaRNA/constraints/basic.h File Reference

Functions and data structures for constraining secondary structure predictions and evaluation.

Include dependency graph for basic.h: This graph shows which files directly or indirectly include this file:

Macros

#define VRNA CONSTRAINT FILE 0

Flag for vrna_constraints_add() to indicate that constraints are present in a text file.

• #define VRNA CONSTRAINT SOFT MFE 0

Indicate generation of constraints for MFE folding.

#define VRNA_CONSTRAINT_SOFT_PF VRNA_OPTION_PF

Indicate generation of constraints for partition function computation.

#define VRNA_DECOMP_PAIR_HP (unsigned char)1

Flag passed to generic softt constraints callback to indicate hairpin loop decomposition step.

• #define VRNA_DECOMP_PAIR_IL (unsigned char)2

Indicator for interior loop decomposition step.

• #define VRNA DECOMP PAIR ML (unsigned char)3

Indicator for multibranch loop decomposition step.

#define VRNA DECOMP ML ML (unsigned char)5

Indicator for decomposition of multibranch loop part.

#define VRNA_DECOMP_ML_STEM (unsigned char)6

Indicator for decomposition of multibranch loop part.

#define VRNA_DECOMP_ML_ML (unsigned char)7

Indicator for decomposition of multibranch loop part.

• #define VRNA DECOMP ML UP (unsigned char)8

Indicator for decomposition of multibranch loop part.

• #define VRNA_DECOMP_ML_ML_STEM (unsigned char)9

Indicator for decomposition of multibranch loop part.

• #define VRNA_DECOMP_ML_COAXIAL (unsigned char)10

Indicator for decomposition of multibranch loop part.

#define VRNA_DECOMP_ML_COAXIAL_ENC (unsigned char)11

Indicator for decomposition of multibranch loop part.

#define VRNA_DECOMP_EXT_EXT (unsigned char)12

Indicator for decomposition of exterior loop part.

• #define VRNA_DECOMP_EXT_UP (unsigned char)13

Indicator for decomposition of exterior loop part.

#define VRNA_DECOMP_EXT_STEM (unsigned char)14

Indicator for decomposition of exterior loop part.

#define VRNA_DECOMP_EXT_EXT_EXT (unsigned char)15

Indicator for decomposition of exterior loop part.

#define VRNA_DECOMP_EXT_STEM_EXT (unsigned char)16

Indicator for decomposition of exterior loop part.

• #define VRNA_DECOMP_EXT_STEM_OUTSIDE (unsigned char)17

Indicator for decomposition of exterior loop part.

• #define VRNA_DECOMP_EXT_EXT_STEM (unsigned char)18

Indicator for decomposition of exterior loop part.

• #define VRNA_DECOMP_EXT_EXT_STEM1 (unsigned char)19

Indicator for decomposition of exterior loop part.

Functions

• void vrna_constraints_add (vrna_fold_compound_t *vc, const char *constraint, unsigned int options)

Add constraints to a vrna_fold_compound_t data structure.

18.75.1 Detailed Description

Functions and data structures for constraining secondary structure predictions and evaluation.

18.76 ViennaRNA/utils/basic.h File Reference

General utility- and helper-functions used throughout the ViennaRNA Package.

Include dependency graph for basic.h: This graph shows which files directly or indirectly include this file:

Macros

• #define VRNA INPUT ERROR 1U

Output flag of get_input_line(): "An ERROR has occured, maybe EOF".

#define VRNA_INPUT_QUIT 2U

Output flag of get_input_line(): "the user requested quitting the program".

• #define VRNA INPUT MISC 4U

Output flag of get_input_line(): "something was read".

• #define VRNA_INPUT_FASTA_HEADER 8U

Input/Output flag of get_input_line():

if used as input option this tells get input line() that the data to be read should comply with the FASTA format.

• #define VRNA_INPUT_CONSTRAINT 32U

Input flag for get_input_line():

Tell get_input_line() that we assume to read a structure constraint.

#define VRNA_INPUT_NO_TRUNCATION 256U

Input switch for get_input_line(): "do not trunkate the line by eliminating white spaces at end of line".

#define VRNA_INPUT_NO_REST 512U

Input switch for vrna_file_fasta_read_record(): "do fill rest array".

#define VRNA_INPUT_NO_SPAN 1024U

Input switch for vrna file fasta read record(): "never allow data to span more than one line".

• #define VRNA_INPUT_NOSKIP_BLANK_LINES 2048U

Input switch for vrna_file_fasta_read_record(): "do not skip empty lines".

#define VRNA INPUT BLANK LINE 4096U

Output flag for vrna_file_fasta_read_record(): "read an empty line".

#define VRNA INPUT NOSKIP COMMENTS 128U

Input switch for get_input_line(): "do not skip comment lines".

#define VRNA_INPUT_COMMENT 8192U

Output flag for vrna_file_fasta_read_record(): "read a comment".

#define MIN2(A, B) ((A) < (B) ? (A) : (B))

Get the minimum of two comparable values.

#define MAX2(A, B) ((A) > (B) ? (A) : (B))

Get the maximum of two comparable values.

#define MIN3(A, B, C) (MIN2((MIN2((A), (B))), (C)))

Get the minimum of three comparable values.

#define MAX3(A, B, C) (MAX2((MAX2((A), (B))), (C)))

Get the maximum of three comparable values.

Functions

void * vrna_alloc (unsigned size)

Allocate space safely.

void * vrna_realloc (void *p, unsigned size)

Reallocate space safely.

void vrna_init_rand (void)

Initialize seed for random number generator.

• double vrna urn (void)

get a random number from [0..1]

• int vrna_int_urn (int from, int to)

Generates a pseudo random integer in a specified range.

char * vrna_time_stamp (void)

Get a timestamp.

- unsigned int get_input_line (char **string, unsigned int options)
- int * vrna idx row wise (unsigned int length)

Get an index mapper array (iindx) for accessing the energy matrices, e.g. in partition function related functions.

int * vrna_idx_col_wise (unsigned int length)

Get an index mapper array (indx) for accessing the energy matrices, e.g. in MFE related functions.

void vrna message error (const char *format,...)

Print an error message and die.

void vrna message verror (const char *format, va list args)

Print an error message and die.

void vrna_message_warning (const char *format,...)

Print a warning message.

• void vrna_message_vwarning (const char *format, va_list args)

Print a warning message.

void vrna_message_info (FILE *fp, const char *format,...)

Print an info message.

• void vrna_message_vinfo (FILE *fp, const char *format, va_list args)

Print an info message.

void vrna_message_input_seq_simple (void)

Print a line to stdout that asks for an input sequence.

• void vrna_message_input_seq (const char *s)

Print a line with a user defined string and a ruler to stdout.

char * get_line (FILE *fp)

Read a line of arbitrary length from a stream.

void print_tty_input_seq (void)

Print a line to stdout that asks for an input sequence.

void print_tty_input_seq_str (const char *s)

Print a line with a user defined string and a ruler to stdout.

• void warn user (const char message[])

Print a warning message.

void nrerror (const char message[])

Die with an error message.

• void * space (unsigned size)

Allocate space safely.

void * xrealloc (void *p, unsigned size)

Reallocate space safely.

void init_rand (void)

Make random number seeds.

```
• double urn (void)
```

```
get a random number from [0..1]
```

• int int_urn (int from, int to)

Generates a pseudo random integer in a specified range.

• void filecopy (FILE *from, FILE *to)

Inefficient cp

char * time_stamp (void)

Get a timestamp.

Variables

• unsigned short xsubi [3]

Current 48 bit random number.

18.76.1 Detailed Description

General utility- and helper-functions used throughout the ViennaRNA Package.

18.76.2 Function Documentation

```
18.76.2.1 get_line()
```

Read a line of arbitrary length from a stream.

Returns a pointer to the resulting string. The necessary memory is allocated and should be released using *free()* when the string is no longer needed.

Deprecated Use vrna_read_line() as a substitute!

Parameters

fp A file pointer to the stream where the function should read from

Returns

A pointer to the resulting string

18.76.2.2 print_tty_input_seq()

Print a line to stdout that asks for an input sequence.

There will also be a ruler (scale line) printed that helps orientation of the sequence positions

Deprecated Use vrna message input seq simple() instead!

18.76.2.3 print_tty_input_seq_str()

```
void print_tty_input_seq_str ( {\tt const\ char\ *\ s\ )}
```

Print a line with a user defined string and a ruler to stdout.

(usually this is used to ask for user input) There will also be a ruler (scale line) printed that helps orientation of the sequence positions

Deprecated Use vrna_message_input_seq() instead!

18.76.2.4 warn_user()

Print a warning message.

Print a warning message to stderr

Deprecated Use vrna_message_warning() instead!

18.76.2.5 nrerror()

Die with an error message.

Deprecated Use vrna_message_error() instead!

Allocate space safely.

Deprecated Use vrna_alloc() instead!

Reallocate space safely.

Deprecated Use vrna_realloc() instead!

Make random number seeds.

Deprecated Use vrna_init_rand() instead!

```
18.76.2.9 urn()

double urn (

void )
```

get a random number from [0..1]

Deprecated Use vrna_urn() instead!

18.76.2.10 int_urn()

```
int int_urn (
          int from,
          int to )
```

Generates a pseudo random integer in a specified range.

Deprecated Use vrna_int_urn() instead!

18.76.2.11 filecopy()

```
void filecopy (
     FILE * from,
     FILE * to )
```

Inefficient cp

Deprecated Use vrna_file_copy() instead!

18.76.2.12 time_stamp()

```
char* time_stamp (
     void )
```

Get a timestamp.

Deprecated Use vrna_time_stamp() instead!

18.77 ViennaRNA/datastructures/basic.h File Reference

Various data structures and pre-processor macros.

Include dependency graph for basic.h: This graph shows which files directly or indirectly include this file:

Data Structures

• struct vrna_basepair_s

Base pair data structure used in subopt.c. More...

struct vrna_cpair_s

this datastructure is used as input parameter in functions of PS_dot.c More...

- struct vrna_color_s
- struct vrna_data_linear_s
- struct vrna_sect_s

Stack of partial structures for backtracking. More...

• struct vrna_bp_stack_s

Base pair stack element. More...

struct pu_contrib

contributions to p_u More...

· struct interact

interaction data structure for RNAup More...

struct pu_out

Collection of all free_energy of beeing unpaired values for output. More...

• struct constrain

constraints for cofolding More...

struct duplexT

Data structure for RNAduplex. More...

· struct node

Data structure for RNAsnoop (fold energy list) More...

struct snoopT

Data structure for RNAsnoop. More...

struct dupVar

Data structure used in RNApkplex. More...

Typedefs

typedef struct vrna_basepair_s vrna_basepair_t

Typename for the base pair repesenting data structure vrna_basepair_s.

typedef struct vrna_elem_prob_s vrna_plist_t

Typename for the base pair list repesenting data structure vrna_elem_prob_s.

typedef struct vrna_bp_stack_s vrna_bp_stack_t

Typename for the base pair stack repesenting data structure vrna_bp_stack_s.

typedef struct vrna_cpair_s vrna_cpair_t

Typename for data structure vrna_cpair_s.

• typedef struct vrna_sect_s vrna_sect_t

Typename for stack of partial structures vrna_sect_s.

typedef double FLT_OR_DBL

Typename for floating point number in partition function computations.

typedef struct vrna basepair s PAIR

Old typename of vrna_basepair_s.

typedef struct vrna_elem_prob_s plist

Old typename of vrna_elem_prob_s.

· typedef struct vrna cpair s cpair

Old typename of vrna_cpair_s.

typedef struct vrna_sect_s sect

Old typename of vrna_sect_s.

typedef struct vrna_bp_stack_s bondT

Old typename of vrna_bp_stack_s.

• typedef struct pu_contrib pu_contrib

contributions to p_u

· typedef struct interact interact

interaction data structure for RNAup

typedef struct pu_out pu_out

Collection of all free_energy of beeing unpaired values for output.

· typedef struct constrain constrain

constraints for cofolding

· typedef struct node folden

Data structure for RNAsnoop (fold energy list)

typedef struct dupVar dupVar

Data structure used in RNApkplex.

Functions

• void vrna_C11_features (void)

Dummy symbol to check whether the library was build using C11/C++11 features.

18.77.1 Detailed Description

Various data structures and pre-processor macros.

18.78 ViennaRNA/params/constants.h File Reference

Energy parameter constants.

Include dependency graph for constants.h: This graph shows which files directly or indirectly include this file:

Macros

- #define GASCONST 1.98717 /* in [cal/K] */
- #define K0 273.15
- #define INF 10000000 /* (INT MAX/10) */
- #define FORBIDDEN 9999
- #define BONUS 10000
- #define NBPAIRS 7
- #define TURN 3
- #define MAXLOOP 30

18.78.1 Detailed Description

Energy parameter constants.

18.78.2 Macro Definition Documentation

The minimum loop length

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```
18.78.2.1 GASCONST
#define GASCONST 1.98717 /* in [cal/K] */
The gas constant
18.78.2.2 K0
#define K0 273.15
0 deg Celsius in Kelvin
18.78.2.3 INF
#define INF 10000000 /* (INT_MAX/10) */
Infinity as used in minimization routines
18.78.2.4 FORBIDDEN
#define FORBIDDEN 9999
forbidden
18.78.2.5 BONUS
#define BONUS 10000
bonus contribution
18.78.2.6 NBPAIRS
#define NBPAIRS 7
The number of distinguishable base pairs
18.78.2.7 TURN
#define TURN 3
```

18.78.2.8 MAXLOOP

#define MAXLOOP 30

The maximum loop length

18.79 ViennaRNA/params/convert.h File Reference

Functions and definitions for energy parameter file format conversion.

This graph shows which files directly or indirectly include this file:

Macros

- #define VRNA_CONVERT_OUTPUT_ALL 1U
- #define VRNA_CONVERT_OUTPUT_HP 2U
- #define VRNA CONVERT OUTPUT STACK 4U
- #define VRNA_CONVERT_OUTPUT_MM_HP 8U
- #define VRNA CONVERT OUTPUT MM INT 16U
- #define VRNA_CONVERT_OUTPUT_MM_INT_1N 32U
- #define VRNA CONVERT OUTPUT MM INT 23 64U
- #define VRNA_CONVERT_OUTPUT_MM_MULTI 128U
- #define VRNA CONVERT OUTPUT MM EXT 256U
- #define VRNA CONVERT OUTPUT DANGLE5 512U
- #define VRNA_CONVERT_OUTPUT_DANGLE3 1024U
- #define VRNA CONVERT OUTPUT INT 11 2048U
- #define VRNA_CONVERT_OUTPUT_INT_21 4096U
- #define VRNA_CONVERT_OUTPUT_INT_22 8192U
- #define VRNA_CONVERT_OUTPUT_BULGE 16384U
- #define VRNA CONVERT OUTPUT INT 32768U
- #define VRNA_CONVERT_OUTPUT_ML 65536U
- #define VRNA_CONVERT_OUTPUT_MISC 131072U
- #define VRNA_CONVERT_OUTPUT_SPECIAL_HP 262144U
- #define VRNA CONVERT OUTPUT VANILLA 524288U
- #define VRNA CONVERT OUTPUT NINIO 1048576U
- #define VRNA_CONVERT_OUTPUT_DUMP 2097152U

Functions

void convert parameter file (const char *iname, const char *oname, unsigned int options)

18.79.1 Detailed Description

Functions and definitions for energy parameter file format conversion.

18.80 ViennaRNA/params/io.h File Reference

Read and write energy parameter files.

This graph shows which files directly or indirectly include this file:

Macros

• #define VRNA_PARAMETER_FORMAT_DEFAULT 0

Default Energy Parameter File format.

Functions

• int vrna_params_load (const char fname[], unsigned int options)

Load energy parameters from a file.

• int vrna_params_save (const char fname[], unsigned int options)

Save energy parameters to a file.

int vrna_params_load_from_string (const char *string, const char *name, unsigned int options)

Load energy paramters from string.

int vrna_params_load_defaults (void)

Load default RNA energy parameter set.

int vrna_params_load_RNA_Turner2004 (void)

Load Turner 2004 RNA energy parameter set.

• int vrna_params_load_RNA_Turner1999 (void)

Load Turner 1999 RNA energy parameter set.

int vrna_params_load_RNA_Andronescu2007 (void)

Load Andronsecu 2007 RNA energy parameter set.

int vrna_params_load_RNA_Langdon2018 (void)

Load Langdon 2018 RNA energy parameter set.

• int vrna_params_load_RNA_misc_special_hairpins (void)

Load Misc Special Hairpin RNA energy parameter set.

int vrna_params_load_DNA_Mathews2004 (void)

Load Mathews 2004 DNA energy parameter set.

int vrna_params_load_DNA_Mathews1999 (void)

Load Mathews 1999 DNA energy parameter set.

const char * last_parameter_file (void)

Get the file name of the parameter file that was most recently loaded.

void read_parameter_file (const char fname[])

Read energy parameters from a file.

void write_parameter_file (const char fname[])

Write energy parameters to a file.

18.80.1 Detailed Description

Read and write energy parameter files.

18.81 ViennaRNA/part_func.h File Reference

Partition function implementations.

Include dependency graph for part_func.h: This graph shows which files directly or indirectly include this file:

Data Structures

· struct vrna dimer pf s

Data structure returned by vrna_pf_dimer() More...

Typedefs

• typedef struct vrna_dimer_pf_s vrna_dimer_pf_t

Typename for the data structure that stores the dimer partition functions, vrna_dimer_pf_s, as returned by vrna_pf_dimer()

typedef struct vrna_dimer_pf_s cofoldF

Backward compatibility typedef for vrna_dimer_pf_s.

Functions

int vrna_pf_float_precision (void)

Find out whether partition function computations are using single precision floating points.

• float pf_fold_par (const char *sequence, char *structure, vrna_exp_param_t *parameters, int calculate_← bppm, int is_constrained, int is_circular)

Compute the partition function Q for a given RNA sequence.

• float pf_fold (const char *sequence, char *structure)

Compute the partition function ${\cal Q}$ of an RNA sequence.

• float pf_circ_fold (const char *sequence, char *structure)

Compute the partition function of a circular RNA sequence.

• char * pbacktrack (char *sequence)

Sample a secondary structure from the Boltzmann ensemble according its probability.

char * pbacktrack5 (char *sequence, int length)

Sample a sub-structure from the Boltzmann ensemble according its probability.

char * pbacktrack_circ (char *sequence)

Sample a secondary structure of a circular RNA from the Boltzmann ensemble according its probability.

void free_pf_arrays (void)

Free arrays for the partition function recursions.

· void update_pf_params (int length)

Recalculate energy parameters.

void update_pf_params_par (int length, vrna_exp_param_t *parameters)

Recalculate energy parameters.

FLT OR DBL * export bppm (void)

Get a pointer to the base pair probability array.

int get_pf_arrays (short **S_p, short **S1_p, char **ptype_p, FLT_OR_DBL **qb_p, FLT_OR_DBL **qtk_p, FLT_OR_DBL **qtk_p, FLT_OR_DBL **qtk_p)

Get the pointers to (almost) all relavant computation arrays used in partition function computation.

• double get subseq F (int i, int j)

Get the free energy of a subsequence from the q[] array.

double mean_bp_distance (int length)

Get the mean base pair distance of the last partition function computation.

double mean bp distance pr (int length, FLT OR DBL *pr)

Get the mean base pair distance in the thermodynamic ensemble.

vrna_ep_t * stackProb (double cutoff)

Get the probability of stacks.

void init pf fold (int length)

Allocate space for pf fold()

- char * centroid (int length, double *dist)
- char * get_centroid_struct_gquad_pr (int length, double *dist)
- double mean_bp_dist (int length)
- double expLoopEnergy (int u1, int u2, int type, int type2, short si1, short sj1, short sp1, short sq1)
- double expHairpinEnergy (int u, int type, short si1, short sj1, const char *string)

Basic global partition function interface

- float vrna_pf (vrna_fold_compound_t *vc, char *structure)
 - Compute the partition function Q for a given RNA sequence, or sequence alignment.
- vrna_dimer_pf_t vrna_pf_dimer (vrna_fold_compound_t *vc, char *structure)

Calculate partition function and base pair probabilities of nucleic acid/nucleic acid dimers.

Simplified global partition function computation using sequence(s) or multiple sequence alignment(s)

- float vrna pf fold (const char *sequence, char *structure, vrna ep t **pl)
 - Compute Partition function Q (and base pair probabilities) for an RNA sequence using a comparative method.
- float vrna_pf_circfold (const char *sequence, char *structure, vrna_ep_t **pl)
 - Compute Partition function Q (and base pair probabilities) for a circular RNA sequences using a comparative method.
- float vrna_pf_alifold (const char **sequences, char *structure, vrna_ep_t **pl)
 - Compute Partition function Q (and base pair probabilities) for an RNA sequence alignment using a comparative method
- float vrna_pf_circalifold (const char **sequences, char *structure, vrna_ep_t **pl)
 - Compute Partition function Q (and base pair probabilities) for an alignment of circular RNA sequences using a comparative method.
- vrna_dimer_pf_t vrna_pf_co_fold (const char *seq, char *structure, vrna_ep_t **pl)

Calculate partition function and base pair probabilities of nucleic acid/nucleic acid dimers.

Variables

· int st back

Flag indicating that auxilary arrays are needed throughout the computations. This is essential for stochastic backtracking.

18.81.1 Detailed Description

Partition function implementations.

, This file includes (almost) all function declarations within the **RNAlib** that are related to Partion function computations

18.81.2 Function Documentation

Deprecated This function is deprecated and should not be used anymore as it is not threadsafe!

See also

```
get_centroid_struct_pl(), get_centroid_struct_pr()
```

```
18.81.2.2 get_centroid_struct_gquad_pr()
```

Deprecated This function is deprecated and should not be used anymore as it is not threadsafe!

See also

vrna_centroid(), vrna_centroid_from_probs(), vrna_centroid_from_plist()

```
18.81.2.3 mean_bp_dist()
```

get the mean pair distance of ensemble

Deprecated This function is not threadsafe and should not be used anymore. Use mean_bp_distance() instead!

18.81.2.4 expLoopEnergy()

```
double expLoopEnergy (
    int u1,
    int u2,
    int type,
    int type2,
    short si1,
    short sj1,
    short sp1,
    short sq1 )
```

Deprecated Use exp_E_IntLoop() from loop_energies.h instead

18.81.2.5 expHairpinEnergy()

```
double expHairpinEnergy (
    int u,
    int type,
    short sil,
    short sjl,
    const char * string )
```

Deprecated Use exp_E_Hairpin() from loop_energies.h instead

18.82 ViennaRNA/part_func_co.h File Reference

Partition function for two RNA sequences.

Include dependency graph for part_func_co.h:

Functions

- vrna_dimer_pf_t co_pf_fold (char *sequence, char *structure)
 - Calculate partition function and base pair probabilities.
- vrna_dimer_pf_t co_pf_fold_par (char *sequence, char *structure, vrna_exp_param_t *parameters, int calculate_bppm, int is_constrained)

Calculate partition function and base pair probabilities.

- vrna_ep_t * get_plist (vrna_ep_t *pl, int length, double cut_off)
- void compute_probabilities (double FAB, double FEA, double FEB, vrna_ep_t *prAB, vrna_ep_t *prA, vrna_ep_t *prB, int Alength)

Compute Boltzmann probabilities of dimerization without homodimers.

- void init_co_pf_fold (int length)
- FLT_OR_DBL * export_co_bppm (void)

Get a pointer to the base pair probability array.

void free_co_pf_arrays (void)

Free the memory occupied by co_pf_fold()

void update_co_pf_params (int length)

Recalculate energy parameters.

void update_co_pf_params_par (int length, vrna_exp_param_t *parameters)

Recalculate energy parameters.

Variables

· int mirnatog

Toggles no intrabp in 2nd mol.

• double F_monomer [2]

Free energies of the two monomers.

18.82.1 Detailed Description

Partition function for two RNA sequences.

18.82.2 Function Documentation

```
18.82.2.1 get_plist()
```

DO NOT USE THIS FUNCTION ANYMORE

Deprecated { This function is deprecated and will be removed soon!} use assign_plist_from_pr() instead!

18.83 ViennaRNA/part_func_up.h File Reference

Implementations for accessibility and RNA-RNA interaction as a stepwise process.

Include dependency graph for part_func_up.h:

Functions

• pu_contrib * pf_unstru (char *sequence, int max_w)

Calculate the partition function over all unpaired regions of a maximal length.

• interact * pf_interact (const char *s1, const char *s2, pu_contrib *p_c, pu_contrib *p_c2, int max_w, char *cstruc, int incr3, int incr5)

Calculates the probability of a local interaction between two sequences.

void free_interact (interact *pin)

Frees the output of function pf_interact().

void free_pu_contrib_struct (pu_contrib *pu)

Frees the output of function pf_unstru().

18.83.1 Detailed Description

Implementations for accessibility and RNA-RNA interaction as a stepwise process.

18.84 ViennaRNA/part_func_window.h File Reference

Partition function and equilibrium probability implementation for the sliding window algorithm.

Include dependency graph for part_func_window.h: This graph shows which files directly or indirectly include this file:

Macros

#define VRNA_EXT_LOOP 1U

Exterior loop.

#define VRNA HP LOOP 2U

Hairpin loop.

• #define VRNA_INT_LOOP 4U

Internal loop.

• #define VRNA_MB_LOOP 8U

Multibranch loop.

#define VRNA_ANY_LOOP (VRNA_EXT_LOOP | VRNA_HP_LOOP | VRNA_INT_LOOP | VRNA_MB_LOOP)
 Any loop.

#define VRNA PROBS WINDOW BPP 4096U

Trigger base pairing probabilities.

#define VRNA_PROBS_WINDOW_UP 8192U

Trigger unpaired probabilities.

#define VRNA_PROBS_WINDOW_STACKP 16384U

Trigger base pair stack probabilities.

#define VRNA_PROBS_WINDOW_UP_SPLIT 32768U

Trigger detailed unpaired probabilities split up into different loop type contexts.

#define VRNA_PROBS_WINDOW_PF 65536U

Trigger partition function.

Typedefs

• typedef void() vrna_probs_window_callback(FLT_OR_DBL *pr, int pr_size, int i, int max, unsigned int type, void *data)

Sliding window probability computation callback.

Functions

Basic local partition function interface

 int vrna_probs_window (vrna_fold_compound_t *fc, int ulength, unsigned int options, vrna_probs_window_callback *cb, void *data)

Compute various equilibrium probabilities under a sliding window approach.

Simplified global partition function computation using sequence(s) or multiple sequence alignment(s)

- vrna_ep_t * vrna_pfl_fold (const char *sequence, int window_size, int max_bp_span, float cutoff)

 Compute base pair probabilities using a sliding-window approach.
- int vrna_pfl_fold_cb (const char *sequence, int window_size, int max_bp_span, vrna_probs_window_callback *cb, void *data)

Compute base pair probabilities using a sliding-window approach (callback version)

- double ** vrna_pfl_fold_up (const char *sequence, int ulength, int window_size, int max_bp_span)
 Compute probability of contiguous unpaired segments.
- int vrna_pfl_fold_up_cb (const char *sequence, int ulength, int window_size, int max_bp_span, vrna_probs_window_callback *cb, void *data)

Compute probability of contiguous unpaired segments.

18.84.1 Detailed Description

Partition function and equilibrium probability implementation for the sliding window algorithm.

, This file contains the implementation for sliding window partition function and equilibrium probabilities. It also provides the unpaired probability implementation from Bernhart et al. 2011 [4]

18.85 ViennaRNA/perturbation_fold.h File Reference

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

Include dependency graph for perturbation_fold.h:

Macros

• #define VRNA_OBJECTIVE_FUNCTION_QUADRATIC 0

Use the sum of squared aberrations as objective function.

#define VRNA_OBJECTIVE_FUNCTION_ABSOLUTE 1

Use the sum of absolute aberrations as objective function.

#define VRNA_MINIMIZER_DEFAULT 0

Use a custom implementation of the gradient descent algorithm to minimize the objective function.

#define VRNA_MINIMIZER_CONJUGATE_FR 1

Use the GNU Scientific Library implementation of the Fletcher-Reeves conjugate gradient algorithm to minimize the objective function.

#define VRNA MINIMIZER CONJUGATE PR 2

Use the GNU Scientific Library implementation of the Polak-Ribiere conjugate gradient algorithm to minimize the objective function.

• #define VRNA MINIMIZER VECTOR BFGS 3

Use the GNU Scientific Library implementation of the vector Broyden-Fletcher-Goldfarb-Shanno algorithm to minimize the objective function.

• #define VRNA_MINIMIZER_VECTOR_BFGS2 4

Use the GNU Scientific Library implementation of the vector Broyden-Fletcher-Goldfarb-Shanno algorithm to minimize the objective function.

#define VRNA MINIMIZER STEEPEST DESCENT 5

Use the GNU Scientific Library implementation of the steepest descent algorithm to minimize the objective function.

Typedefs

typedef void(* progress_callback) (int iteration, double score, double *epsilon)
 Callback for following the progress of the minimization process.

Functions

void vrna_sc_minimize_pertubation (vrna_fold_compound_t *vc, const double *q_prob_unpaired, int objective_function, double sigma_squared, double tau_squared, int algorithm, int sample_size, double *epsilon, double initialStepSize, double minStepSize, double minImprovement, double minimizerTolerance, progress callback callback)

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

18.85.1 Detailed Description

Find a vector of perturbation energies that minimizes the discripancies between predicted and observed pairing probabilities and the amount of neccessary adjustments.

18.86 ViennaRNA/plot_aln.h File Reference

Use ViennaRNA/plotting/alignments.h instead.

Include dependency graph for plot_aln.h:

18.86.1 Detailed Description

Use ViennaRNA/plotting/alignments.h instead.

Deprecated Use ViennaRNA/plotting/alignments.h instead

18.87 ViennaRNA/plot_layouts.h File Reference

Use ViennaRNA/plotting/layouts.h instead.

Include dependency graph for plot_layouts.h:

18.87.1 Detailed Description

Use ViennaRNA/plotting/layouts.h instead.

Deprecated Use ViennaRNA/plotting/layouts.h instead

18.88 ViennaRNA/plot_structure.h File Reference

Use ViennaRNA/plotting/structures.h instead.

Include dependency graph for plot_structure.h:

18.88.1 Detailed Description

Use ViennaRNA/plotting/structures.h instead.

Deprecated Use ViennaRNA/plotting/structures.h instead

18.89 ViennaRNA/plot_utils.h File Reference

Use ViennaRNA/plotting/utils.h instead.

Include dependency graph for plot_utils.h:

18.89.1 Detailed Description

Use ViennaRNA/plotting/utils.h instead.

Deprecated Use ViennaRNA/plotting/utils.h instead

18.90 ViennaRNA/plotting/alignments.h File Reference

Various functions for plotting Sequence / Structure Alignments.

This graph shows which files directly or indirectly include this file:

Functions

• int vrna_file_PS_aln (const char *filename, const char **seqs, const char **names, const char *structure, unsigned int columns)

Create an annotated PostScript alignment plot.

• int vrna_file_PS_aln_slice (const char *filename, const char **seqs, const char **names, const char *structure, unsigned int start, unsigned int end, int offset, unsigned int columns)

Create an annotated PostScript alignment plot.

- int PS_color_aln (const char *structure, const char *filename, const char *seqs[], const char *names[])

 Produce PostScript sequence alignment color-annotated by consensus structure.
- int aliPS_color_aln (const char *structure, const char *filename, const char *seqs[], const char *names[]) PS_color_aln for duplexes.

18.90.1 Detailed Description

Various functions for plotting Sequence / Structure Alignments.

18.91 ViennaRNA/utils/alignments.h File Reference

Various utility- and helper-functions for sequence alignments and comparative structure prediction.

Include dependency graph for alignments.h: This graph shows which files directly or indirectly include this file:

Data Structures

· struct vrna pinfo s

A base pair info structure. More...

Macros

• #define VRNA_ALN_DEFAULT 0U

Use default alignment settings.

• #define VRNA_ALN_RNA 1U

Convert to RNA alphabet.

#define VRNA_ALN_DNA 2U

Convert to DNA alphabet.

• #define VRNA_ALN_UPPERCASE 4U

Convert to uppercase nucleotide letters.

• #define VRNA_ALN_LOWERCASE 8U

Convert to lowercase nucleotide letters.

• #define VRNA_MEASURE_SHANNON_ENTROPY 1U

Flag indicating Shannon Entropy measure.

Typedefs

• typedef struct vrna_pinfo_s vrna_pinfo_t

Typename for the base pair info repesenting data structure vrna_pinfo_s.

• typedef struct vrna_pinfo_s pair_info

Old typename of vrna_pinfo_s.

Functions

• int vrna aln mpi (const char **alignment)

Get the mean pairwise identity in steps from ?to?(ident)

vrna_pinfo_t * vrna_aln_pinfo (vrna_fold_compound_t *vc, const char *structure, double threshold)

Retrieve an array of vrna_pinfo_t structures from precomputed pair probabilities.

char ** vrna aln slice (const char **alignment, unsigned int i, unsigned int j)

Slice out a subalignment from a larger alignment.

void vrna_aln_free (char **alignment)

Free memory occupied by a set of aligned sequences.

char ** vrna_aln_uppercase (const char **alignment)

Create a copy of an alignment with only uppercase letters in the sequences.

char ** vrna_aln_toRNA (const char **alignment)

Create a copy of an alignment where DNA alphabet is replaced by RNA alphabet.

char ** vrna_aln_copy (const char **alignment, unsigned int options)

Make a copy of a multiple sequence alignment.

- float * vrna_aln_conservation_struct (const char **alignment, const char *structure, const vrna_md_t *md)

 Compute base pair conservation of a consensus structure.
- float * vrna_aln_conservation_col (const char **alignment, const vrna_md_t *md_p, unsigned int options)

 Compute nucleotide conservation in an alignment.
- char * vrna_aln_consensus_sequence (const char **alignment, const vrna_md_t *md_p)

Compute the consensus sequence for a given multiple sequence alignment.

char * vrna_aln_consensus_mis (const char **alignment, const vrna_md_t *md_p)

Compute the Most Informative Sequence (MIS) for a given multiple sequence alignment.

int get_mpi (char *Alseq[], int n_seq, int length, int *mini)

Get the mean pairwise identity in steps from ?to?(ident)

 void encode_ali_sequence (const char *sequence, short *S, short *s5, short *s3, char *ss, unsigned short *as, int circ)

Get arrays with encoded sequence of the alignment.

 void alloc_sequence_arrays (const char **sequences, short ***S, short ***S, short ***S, unsigned short ***a2s, char ***Ss, int circ)

Allocate memory for sequence array used to deal with aligned sequences.

• void free_sequence_arrays (unsigned int n_seq, short ***S, short ***S1, unsigned short ***a2s, char ***Ss)

Free the memory of the sequence arrays used to deal with aligned sequences.

18.91.1 Detailed Description

Various utility- and helper-functions for sequence alignments and comparative structure prediction.

18.92 ViennaRNA/plotting/layouts.h File Reference

Secondary structure plot layout algorithms.

Include dependency graph for level to be This graph shows which files directly or indirectly include this file

Include dependency graph for layouts.h: This graph shows which files directly or indirectly include this file:

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Data Structures

- struct vrna_plot_layout_s
- struct COORDINATE

this is a workarround for the SWIG Perl Wrapper RNA plot function that returns an array of type COORDINATE More...

Macros

• #define VRNA PLOT TYPE SIMPLE 0

Definition of Plot type simple

• #define VRNA PLOT TYPE NAVIEW 1

Definition of Plot type Naview

• #define VRNA PLOT TYPE CIRCULAR 2

Definition of Plot type Circular

#define VRNA_PLOT_TYPE_TURTLE 3

Definition of Plot type Turtle [23].

#define VRNA PLOT TYPE PUZZLER 4

Definition of Plot type RNApuzzler [23].

Typedefs

typedef struct vrna plot layout s vrna plot layout t

RNA secondary structure figure layout.

Functions

vrna_plot_layout_t * vrna_plot_layout (const char *structure, unsigned int plot_type)

Create a layout (coordinates, etc.) for a secondary structure plot.

vrna_plot_layout_t * vrna_plot_layout_simple (const char *structure)

Create a layout (coordinates, etc.) for a simple secondary structure plot.

vrna_plot_layout_t * vrna_plot_layout_naview (const char *structure)

Create a layout (coordinates, etc.) for a secondary structure plot using the Naview Algorithm [5].

vrna plot layout t * vrna plot layout circular (const char *structure)

Create a layout (coordinates, etc.) for a circular secondary structure plot.

vrna_plot_layout_t * vrna_plot_layout_turtle (const char *structure)

Create a layout (coordinates, etc.) for a secondary structure plot using the Turtle Algorithm [23].

vrna_plot_layout_t * vrna_plot_layout_puzzler (const char *structure, vrna_plot_options_puzzler_t *options)

Create a layout (coordinates, etc.) for a secondary structure plot using the RNApuzzler Algorithm [23].

void vrna_plot_layout_free (vrna_plot_layout_t *layout)

Free memory occupied by a figure layout data structure.

int vrna_plot_coords (const char *structure, float **x, float **y, int plot_type)

Compute nucleotide coordinates for secondary structure plot.

int vrna_plot_coords_pt (const short *pt, float **x, float **y, int plot_type)

Compute nucleotide coordinates for secondary structure plot.

int vrna_plot_coords_simple (const char *structure, float **x, float **y)

Compute nucleotide coordinates for secondary structure plot the Simple way

int vrna plot coords simple pt (const short *pt, float **x, float **x)

Compute nucleotide coordinates for secondary structure plot the Simple way

int vrna_plot_coords_circular (const char *structure, float **x, float **y)

Compute coordinates of nucleotides mapped in equal distancies onto a unit circle.

int vrna_plot_coords_circular_pt (const short *pt, float **x, float **y)

Compute nucleotide coordinates for a Circular Plot

int simple_xy_coordinates (short *pair_table, float *X, float *Y)

Calculate nucleotide coordinates for secondary structure plot the Simple way

• int simple_circplot_coordinates (short *pair_table, float *x, float *y)

Calculate nucleotide coordinates for Circular Plot

Variables

• int rna_plot_type

Switch for changing the secondary structure layout algorithm.

18.92.1 Detailed Description

Secondary structure plot layout algorithms.

18.93 ViennaRNA/plotting/probabilities.h File Reference

Various functions for plotting RNA secondary structures, dot-plots and other visualizations.

Include dependency graph for probabilities.h: This graph shows which files directly or indirectly include this file:

Data Structures

· struct vrna_dotplot_auxdata_t

Functions

- int PS_dot_plot_list (char *seq, char *filename, plist *pl, plist *mf, char *comment)

 Produce a postscript dot-plot from two pair lists.
- int PS_dot_plot (char *string, char *file)

Produce postscript dot-plot.

18.93.1 Detailed Description

Various functions for plotting RNA secondary structures, dot-plots and other visualizations.

18.94 ViennaRNA/plotting/RNApuzzler/RNApuzzler.h File Reference

Implementation of the RNApuzzler RNA secondary structure layout algorithm [23].

This graph shows which files directly or indirectly include this file:

Data Structures

struct vrna_plot_options_puzzler_t

Options data structure for RNApuzzler algorithm implementation. More...

Functions

• int vrna_plot_coords_puzzler (const char *structure, float **x, float **y, double **arc_coords, vrna_plot_options_puzzler_t *options)

Compute nucleotide coordinates for secondary structure plot using the RNApuzzler algorithm [23].

 int vrna_plot_coords_puzzler_pt (short const *const pair_table, float **x, float **y, double **arc_coords, vrna_plot_options_puzzler_t *puzzler)

Compute nucleotide coordinates for secondary structure plot using the RNApuzzler algorithm [23].

vrna_plot_options_puzzler_t * vrna_plot_options_puzzler (void)

Create an RNApuzzler options data structure.

void vrna_plot_options_puzzler_free (vrna_plot_options_puzzler_t *options)

Free memory occupied by an RNApuzzler options data structure.

18.94.1 Detailed Description

Implementation of the RNApuzzler RNA secondary structure layout algorithm [23].

18.95 ViennaRNA/plotting/RNApuzzler/RNAturtle.h File Reference

Implementation of the RNAturtle RNA secondary structure layout algorithm [23].

This graph shows which files directly or indirectly include this file:

Functions

- int vrna_plot_coords_turtle (const char *structure, float **x, float **y, double **arc_coords)

 Compute nucleotide coordinates for secondary structure plot using the RNAturtle algorithm [23].
- int vrna_plot_coords_turtle_pt (short const *const pair_table, float **x, float **y, double **arc_coords)

 Compute nucleotide coordinates for secondary structure plot using the RNAturtle algorithm [23].

18.95.1 Detailed Description

Implementation of the RNAturtle RNA secondary structure layout algorithm [23].

18.96 ViennaRNA/plotting/structures.h File Reference

Various functions for plotting RNA secondary structures.

Include dependency graph for structures.h: This graph shows which files directly or indirectly include this file:

Functions

• int vrna_file_PS_rnaplot (const char *seq, const char *structure, const char *file, vrna_md_t *md_p)

Produce a secondary structure graph in PostScript and write it to 'filename'.

• int vrna_file_PS_rnaplot_a (const char *seq, const char *structure, const char *file, const char *pre, con

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename'.

int gmlRNA (char *string, char *structure, char *ssfile, char option)

Produce a secondary structure graph in Graph Meta Language (gml) and write it to a file.

int ssv_rna_plot (char *string, char *structure, char *ssfile)

Produce a secondary structure graph in SStructView format.

int svg_rna_plot (char *string, char *structure, char *ssfile)

Produce a secondary structure plot in SVG format and write it to a file.

int xrna_plot (char *string, char *structure, char *ssfile)

Produce a secondary structure plot for further editing in XRNA.

• int PS_rna_plot (char *string, char *structure, char *file)

Produce a secondary structure graph in PostScript and write it to 'filename'.

• int PS_rna_plot_a (char *string, char *structure, char *file, char *pre, char *post)

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename'.

• int PS_rna_plot_a_gquad (char *string, char *structure, char *ssfile, char *pre, char *post)

Produce a secondary structure graph in PostScript including additional annotation macros and write it to 'filename' (detect and draw g-quadruplexes)

18.96.1 Detailed Description

Various functions for plotting RNA secondary structures.

18.97 ViennaRNA/utils/structures.h File Reference

Various utility- and helper-functions for secondary structure parsing, converting, etc.

Include dependency graph for structures.h: This graph shows which files directly or indirectly include this file:

Data Structures

struct vrna_elem_prob_s

Data structure representing a single entry of an element probability list (e.g. list of pair probabilities) More...

struct vrna_hx_s

Data structure representing an entry of a helix list. More...

Macros

#define VRNA BRACKETS ALPHA 4U

Bitflag to indicate secondary structure notations using uppercase/lowercase letters from the latin alphabet.

#define VRNA_BRACKETS_RND 8U

Bitflag to indicate secondary structure notations using round brackets (parenthesis), ()

#define VRNA BRACKETS CLY 16U

Bitflag to indicate secondary structure notations using curly brackets, {}

#define VRNA BRACKETS ANG 32U

Bitflag to indicate secondary structure notations using angular brackets, <>

#define VRNA_BRACKETS_SQR 64U

Bitflag to indicate secondary structure notations using square brackets, []

#define VRNA BRACKETS DEFAULT

Default bitmask to indicate secondary structure notation using any pair of brackets.

#define VRNA BRACKETS ANY

Bitmask to indicate secondary structure notation using any pair of brackets or uppercase/lowercase alphabet letters.

#define VRNA_PLIST_TYPE_BASEPAIR 0

A Base Pair element.

#define VRNA_PLIST_TYPE_GQUAD 1

A G-Quadruplex element.

• #define VRNA PLIST TYPE H MOTIF 2

A Hairpin loop motif element.

• #define VRNA_PLIST_TYPE_I_MOTIF 3

An Internal loop motif element.

#define VRNA_PLIST_TYPE_UD_MOTIF 4

An Unstructured Domain motif element.

#define VRNA PLIST TYPE STACK 5

A Base Pair stack element.

• #define VRNA_STRUCTURE_TREE_HIT 1U

Homeomorphically Irreducible Tree (HIT) representation of a secondary structure.

#define VRNA_STRUCTURE_TREE_SHAPIRO_SHORT 2U

(short) Coarse Grained representation of a secondary structure

#define VRNA_STRUCTURE_TREE_SHAPIRO 3U

(full) Coarse Grained representation of a secondary structure

• #define VRNA_STRUCTURE_TREE_SHAPIRO_EXT 4U

(extended) Coarse Grained representation of a secondary structure #define VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT 5U

(weighted) Coarse Grained representation of a secondary structure

• #define VRNA STRUCTURE TREE EXPANDED 6U

Expanded Tree representation of a secondary structure.

Typedefs

typedef struct vrna_hx_s vrna_hx_t

Convenience typedef for data structure vrna_hx_s.

typedef struct vrna_elem_prob_s vrna_ep_t

Convenience typedef for data structure vrna_elem_prob_s.

Functions

char * vrna_db_pack (const char *struc)

Pack secondary secondary structure, 5:1 compression using base 3 encoding.

char * vrna db unpack (const char *packed)

Unpack secondary structure previously packed with vrna_db_pack()

void vrna_db_flatten (char *structure, unsigned int options)

Substitute pairs of brackets in a string with parenthesis.

void vrna db flatten to (char *string, const char target[3], unsigned int options)

Substitute pairs of brackets in a string with another type of pair characters.

char * vrna_db_from_ptable (short *pt)

Convert a pair table into dot-parenthesis notation.

char * vrna_db_from_WUSS (const char *wuss)

Convert a WUSS annotation string to dot-bracket format.

char * vrna_db_from_plist (vrna_ep_t *pairs, unsigned int n)

Convert a list of base pairs into dot-bracket notation.

char * vrna_db_to_element_string (const char *structure)

Convert a secondary structure in dot-bracket notation to a nucleotide annotation of loop contexts.

char * vrna_db_pk_remove (const char *structure, unsigned int options)

Remove pseudo-knots from an input structure.

• short * vrna_ptable (const char *structure)

Create a pair table from a dot-bracket notation of a secondary structure.

• short * vrna_ptable_from_string (const char *string, unsigned int options)

Create a pair table for a secondary structure string.

short * vrna_pt_pk_get (const char *structure)

Create a pair table of a secondary structure (pseudo-knot version)

short * vrna_ptable_copy (const short *pt)

Get an exact copy of a pair table.

short * vrna_pt_ali_get (const char *structure)

Create a pair table of a secondary structure (snoop align version)

short * vrna pt snoop get (const char *structure)

Create a pair table of a secondary structure (snoop version)

short * vrna_pt_pk_remove (const short *ptable, unsigned int options)

Remove pseudo-knots from a pair table.

vrna_ep_t * vrna_plist (const char *struc, float pr)

Create a vrna_ep_t from a dot-bracket string.

• vrna_ep_t * vrna_plist_from_probs (vrna_fold_compound_t *vc, double cut_off)

Create a vrna_ep_t from base pair probability matrix.

vrna_hx_t * vrna_hx_from_ptable (short *pt)

Convert a pair table representation of a secondary structure into a helix list.

vrna hx t * vrna hx merge (const vrna hx t *list, int maxdist)

Create a merged helix list from another helix list.

int * vrna_loopidx_from_ptable (const short *pt)

Get a loop index representation of a structure.

• int vrna bp distance (const char *str1, const char *str2)

Compute the "base pair" distance between two secondary structures s1 and s2.

unsigned int * vrna_refBPcnt_matrix (const short *reference_pt, unsigned int turn)

Make a reference base pair count matrix.

unsigned int * vrna refBPdist matrix (const short *pt1, const short *pt2, unsigned int turn)

Make a reference base pair distance matrix.

char * vrna_db_from_probs (const FLT_OR_DBL *pr, unsigned int length)

Create a dot-bracket like structure string from base pair probability matrix.

char vrna_bpp_symbol (const float *x)

Get a pseudo dot bracket notation for a given probability information.

char * vrna_db_from_bp_stack (vrna_bp_stack_t *bp, unsigned int length)

Create a dot-backet/parenthesis structure from backtracking stack.

char * vrna_db_to_tree_string (const char *structure, unsigned int type)

Convert a Dot-Bracket structure string into tree string representation.

char * vrna tree string unweight (const char *structure)

Remove weights from a linear string tree representation of a secondary structure.

char * vrna_tree_string_to_db (const char *tree)

Convert a linear tree string representation of a secondary structure back to Dot-Bracket notation.

void assign_plist_from_db (vrna_ep_t **pl, const char *struc, float pr)

Create a vrna_ep_t from a dot-bracket string.

char * pack_structure (const char *struc)

Pack secondary secondary structure, 5:1 compression using base 3 encoding.

char * unpack_structure (const char *packed)

Unpack secondary structure previously packed with pack_structure()

• short * make_pair_table (const char *structure)

Create a pair table of a secondary structure.

short * copy_pair_table (const short *pt)

Get an exact copy of a pair table.

- short * alimake pair table (const char *structure)
- short * make pair table snoop (const char *structure)
- int bp_distance (const char *str1, const char *str2)

Compute the "base pair" distance between two secondary structures s1 and s2.

• unsigned int * make referenceBP array (short *reference pt, unsigned int turn)

Make a reference base pair count matrix.

unsigned int * compute_BPdifferences (short *pt1, short *pt2, unsigned int turn)

Make a reference base pair distance matrix.

void assign plist from pr (vrna ep t **pl, FLT OR DBL *probs, int length, double cutoff)

Create a vrna_ep_t from a probability matrix.

void parenthesis_structure (char *structure, vrna_bp_stack_t *bp, int length)

Create a dot-backet/parenthesis structure from backtracking stack.

void parenthesis_zuker (char *structure, vrna_bp_stack_t *bp, int length)

Create a dot-backet/parenthesis structure from backtracking stack obtained by zuker suboptimal calculation in cofold.c.

• void bppm_to_structure (char *structure, FLT_OR_DBL *pr, unsigned int length)

Create a dot-bracket like structure string from base pair probability matrix.

char bppm_symbol (const float *x)

Get a pseudo dot bracket notation for a given probability information.

18.97.1 Detailed Description

Various utility- and helper-functions for secondary structure parsing, converting, etc.

18.98 ViennaRNA/profiledist.h File Reference

Include dependency graph for profiledist.h:

Functions

```
    float profile_edit_distance (const float *T1, const float *T2)
    Align the 2 probability profiles T1, T2
    .
```

• float * Make_bp_profile_bppm (FLT_OR_DBL *bppm, int length)

condense pair probability matrix into a vector containing probabilities for unpaired, upstream paired and downstream paired.

void print_bppm (const float *T)
 print string representation of probability profile

void free_profile (float *T)

free space allocated in Make_bp_profile

float * Make_bp_profile (int length)

18.98.1 Function Documentation

18.98.1.1 profile_edit_distance()

```
float profile_edit_distance ( {\rm const\ float\ *\ T1,} {\rm const\ float\ *\ T2\ )}
```

Align the 2 probability profiles T1, T2

.

This is like a Needleman-Wunsch alignment, we should really use affine gap-costs ala Gotoh

18.98.1.2 Make_bp_profile_bppm()

condense pair probability matrix into a vector containing probabilities for unpaired, upstream paired and downstream paired.

This resulting probability profile is used as input for profile_edit_distance

Parameters

bppm	A pointer to the base pair probability matrix	Χ
length	The length of the sequence	

Returns

The bp profile

18.98.1.3 free_profile()

```
void free_profile (
     float * T )
```

free space allocated in Make bp profile

Backward compatibility only. You can just use plain free()

18.98.1.4 Make_bp_profile()

Note

This function is NOT threadsafe

See also

Make_bp_profile_bppm()

Deprecated This function is deprecated and will be removed soon! See Make_bp_profile_bppm() for a replacement

18.99 ViennaRNA/PS_dot.h File Reference

Use ViennaRNA/plotting/probabilities.h instead.

Include dependency graph for PS_dot.h:

18.99.1 Detailed Description

Use ViennaRNA/plotting/probabilities.h instead.

Deprecated Use ViennaRNA/plotting/probabilities.h instead

18.100 ViennaRNA/read_epars.h File Reference

Use ViennaRNA/params/io.h instead.

Include dependency graph for read_epars.h:

18.100.1 Detailed Description

Use ViennaRNA/params/io.h instead.

Deprecated Use ViennaRNA/params/io.h instead

18.101 ViennaRNA/ribo.h File Reference

Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments.

This graph shows which files directly or indirectly include this file:

Functions

float ** get_ribosum (const char **Alseq, int n_seq, int length)

Retrieve a RiboSum Scoring Matrix for a given Alignment.

float ** readribosum (char *name)

Read a RiboSum or other user-defined Scoring Matrix and Store into global Memory.

18.101.1 Detailed Description

Parse RiboSum Scoring Matrices for Covariance Scoring of Alignments.

18.102 ViennaRNA/RNAstruct.h File Reference

Parsing and Coarse Graining of Structures.

Functions

char * b2HIT (const char *structure)

Converts the full structure from bracket notation to the HIT notation including root.

char * b2C (const char *structure)

Converts the full structure from bracket notation to the a coarse grained notation using the 'H' 'B' 'I' 'M' and 'R' identifiers.

char * b2Shapiro (const char *structure)

Converts the full structure from bracket notation to the weighted coarse grained notation using the 'H' 'B' 'I' 'M' 'S' 'E' and 'R' identifiers.

char * add_root (const char *structure)

Adds a root to an un-rooted tree in any except bracket notation.

char * expand_Shapiro (const char *coarse)

Inserts missing 'S' identifiers in unweighted coarse grained structures as obtained from b2C().

char * expand Full (const char *structure)

Convert the full structure from bracket notation to the expanded notation including root.

char * unexpand_Full (const char *ffull)

Restores the bracket notation from an expanded full or HIT tree, that is any tree using only identifiers 'U' 'P' and 'R'.

char * unweight (const char *wcoarse)

Strip weights from any weighted tree.

void unexpand_aligned_F (char *align[2])

Converts two aligned structures in expanded notation.

void parse structure (const char *structure)

Collects a statistic of structure elements of the full structure in bracket notation.

Variables

int loop_size [2000]

contains a list of all loop sizes. loop_size[0] contains the number of external bases.

• int helix size [2000]

contains a list of all stack sizes.

• int loop_degree [2000]

contains the corresponding list of loop degrees.

· int loops

contains the number of loops (and therefore of stacks).

· int unpaired

contains the number of unpaired bases.

int pairs

contains the number of base pairs in the last parsed structure.

18.102.1 Detailed Description

Parsing and Coarse Graining of Structures.

Example:

18.103 ViennaRNA/search/BoyerMoore.h File Reference

Variants of the Boyer-Moore string search algorithm.

Functions

• const unsigned int * vrna_search_BMH_num (const unsigned int *needle, size_t needle_size, const unsigned int *haystack, size_t haystack_size, size_t start, size_t *badchars, unsigned char cyclic)

Search for a string of elements in a larger string of elements using the Boyer-Moore-Horspool algorithm.

• const char * vrna_search_BMH (const char *needle, size_t needle_size, const char *haystack, size_← t haystack_size, size_t start, size_t *badchars, unsigned char cyclic)

Search for an ASCII pattern within a larger ASCII string using the Boyer-Moore-Horspool algorithm.

size_t * vrna_search_BM_BCT_num (const unsigned int *pattern, size_t pattern_size, unsigned int num_
 max)

Retrieve a Boyer-Moore Bad Character Table for a pattern of elements represented by natural numbers.

size_t * vrna_search_BM_BCT (const char *pattern)

Retrieve a Boyer-Moore Bad Character Table for a NULL-terminated pattern of ASCII characters.

18.103.1 Detailed Description

Variants of the Boyer-Moore string search algorithm.

,

18.104 ViennaRNA/sequence.h File Reference

Functions and data structures related to sequence representations,.

Include dependency graph for sequence.h: This graph shows which files directly or indirectly include this file:

Data Structures

• struct vrna_sequence_s

Data structure representing a nucleotide sequence. More...

struct vrna_alignment_s

Typedefs

typedef struct vrna_sequence_s vrna_seq_t
 Typename for nucleotide sequence representation data structure vrna_sequence_s.

Enumerations

enum vrna_seq_type_e { VRNA_SEQ_UNKNOWN, VRNA_SEQ_RNA, VRNA_SEQ_DNA }
 A enumerator used in vrna_sequence_s to distinguish different nucleotide sequences.

18.104.1 Detailed Description

Functions and data structures related to sequence representations,.

18.105 ViennaRNA/stream_output.h File Reference

Use ViennaRNA/datastructures/stream_output.h instead.

Include dependency graph for stream_output.h:

18.105.1 Detailed Description

Use ViennaRNA/datastructures/stream_output.h instead.

Deprecated Use ViennaRNA/datastructures/stream_output.h instead

18.106 ViennaRNA/datastructures/stream_output.h File Reference

An implementation of a buffered, ordered stream output data structure.

This graph shows which files directly or indirectly include this file:

Typedefs

- typedef struct vrna_ordered_stream_s * vrna_ostream_t
 - An ordered output stream structure with unordered insert capabilities.
- typedef void() vrna_callback_stream_output(void *auxdata, unsigned int i, void *data)

 Ordered stream processing callback.

Functions

- vrna_ostream_t vrna_ostream_init (vrna_callback_stream_output *output, void *auxdata)
 - Get an initialized ordered output stream.
- void vrna_ostream_free (vrna_ostream_t dat)
 - Free an initialized ordered output stream.
- void vrna_ostream_request (vrna_ostream_t dat, unsigned int num)
 - Request index in ordered output stream.
- void vrna_ostream_provide (vrna_ostream_t dat, unsigned int i, void *data)

Provide output stream data for a particular index.

18.106.1 Detailed Description

An implementation of a buffered, ordered stream output data structure.

18.107 ViennaRNA/string_utils.h File Reference

Use ViennaRNA/utils/strings.h instead.

Include dependency graph for string_utils.h:

18.107.1 Detailed Description

Use ViennaRNA/utils/strings.h instead.

Deprecated Use ViennaRNA/utils/strings.h instead

18.108 ViennaRNA/stringdist.h File Reference

Functions for String Alignment.

Include dependency graph for stringdist.h:

Functions

```
• swString * Make_swString (char *string)
```

Convert a structure into a format suitable for string_edit_distance().

float string_edit_distance (swString *T1, swString *T2)

Calculate the string edit distance of T1 and T2.

18.108.1 Detailed Description

Functions for String Alignment.

18.108.2 Function Documentation

18.108.2.1 Make_swString()

Convert a structure into a format suitable for string_edit_distance().

Parameters

string

Returns

18.108.2.2 string_edit_distance()

```
float string_edit_distance ( swString * T1, \\ swString * T2 )
```

Calculate the string edit distance of T1 and T2.

Parameters

T1	
T2	

Returns

18.109 ViennaRNA/structure_utils.h File Reference

Use ViennaRNA/utils/structures.h instead.

Include dependency graph for structure_utils.h:

18.109.1 Detailed Description

Use ViennaRNA/utils/structures.h instead.

Deprecated Use ViennaRNA/utils/structures.h instead

18.110 ViennaRNA/structured_domains.h File Reference

This module provides interfaces that deal with additional structured domains in the folding grammar.

This graph shows which files directly or indirectly include this file:

Data Structures

• struct vrna_structured_domains_s

18.110.1 Detailed Description

This module provides interfaces that deal with additional structured domains in the folding grammar.

18.111 ViennaRNA/subopt.h File Reference

RNAsubopt and density of states declarations.

Include dependency graph for subopt.h:

Data Structures

• struct vrna_subopt_sol_s

Solution element from subopt.c.

Macros

• #define MAXDOS 1000

Maximum density of states discretization for subopt.

Typedefs

typedef struct vrna_subopt_sol_s vrna_subopt_solution_t

Typename for the subopt solution list repesenting data structure vrna_subopt_sol_s.

typedef void() vrna_subopt_callback(const char *stucture, float energy, void *data)

Callback for vrna_subopt_cb()

typedef struct vrna_subopt_sol_s SOLUTION

Backward compatibility typedef for vrna_subopt_sol_s.

Functions

- vrna_subopt_solution_t * vrna_subopt (vrna_fold_compound_t *vc, int delta, int sorted, FILE *fp)
 Returns list of subopt structures or writes to fp.
- void vrna subopt cb (vrna fold compound t *vc, int delta, vrna subopt callback *cb, void *data)

Generate suboptimal structures within an energy band arround the MFE.

vrna_subopt_solution_t * vrna_subopt_zuker (vrna_fold_compound_t *vc)

Compute Zuker type suboptimal structures.

SOLUTION * subopt (char *seq, char *structure, int delta, FILE *fp)

Returns list of subopt structures or writes to fp.

• SOLUTION * subopt_par (char *seq, char *structure, vrna_param_t *parameters, int delta, int is_← constrained, int is_circular, FILE *fp)

Returns list of subopt structures or writes to fp.

SOLUTION * subopt_circ (char *seq, char *sequence, int delta, FILE *fp)

Returns list of circular subopt structures or writes to fp.

SOLUTION * zukersubopt (const char *string)

Compute Zuker type suboptimal structures.

SOLUTION * zukersubopt_par (const char *string, vrna_param_t *parameters)

Compute Zuker type suboptimal structures.

Variables

double print_energy

printing threshold for use with logML

int subopt_sorted

Sort output by energy.

• int density_of_states [MAXDOS+1]

The Density of States.

18.111.1 Detailed Description

RNAsubopt and density of states declarations.

18.111.2 Typedef Documentation

18.111.2.1 SOLUTION

typedef struct vrna_subopt_sol_s SOLUTION

Backward compatibility typedef for vrna_subopt_sol_s.

Deprecated Use vrna_subopt_solution_t instead!

18.112 ViennaRNA/svm_utils.h File Reference

Use ViennaRNA/utils/svm.h instead.

Include dependency graph for svm_utils.h:

18.112.1 Detailed Description

Use ViennaRNA/utils/svm.h instead.

Deprecated Use ViennaRNA/utils/svm.h instead

18.113 ViennaRNA/treedist.h File Reference

Functions for Tree Edit Distances.

Include dependency graph for treedist.h:

Functions

Tree * make_tree (char *struc)

Constructs a Tree (essentially the postorder list) of the structure 'struc', for use in tree_edit_distance().

float tree edit distance (Tree *T1, Tree *T2)

Calculates the edit distance of the two trees.

void print_tree (Tree *t)

Print a tree (mainly for debugging)

void free_tree (Tree *t)

Free the memory allocated for Tree t.

18.113.1 Detailed Description

Functions for Tree Edit Distances.

18.113.2 Function Documentation

```
18.113.2.1 make_tree()
```

Constructs a Tree (essentially the postorder list) of the structure 'struc', for use in tree_edit_distance().

Parameters

struc may be any rooted structure representation.

Returns

18.113.2.2 tree_edit_distance()

Calculates the edit distance of the two trees.

Parameters

T1	
T2	

Returns

18.113.2.3 free_tree()

Free the memory allocated for Tree t.

Parameters



18.114 ViennaRNA/units.h File Reference

Physical Units and Functions to convert them into each other.

Enumerations

```
    enum vrna_unit_energy_e {
        VRNA_UNIT_J, VRNA_UNIT_KJ, VRNA_UNIT_CAL_IT, VRNA_UNIT_DACAL_IT,
        VRNA_UNIT_KCAL_IT, VRNA_UNIT_CAL, VRNA_UNIT_DACAL, VRNA_UNIT_KCAL,
        VRNA_UNIT_G_TNT, VRNA_UNIT_KG_TNT, VRNA_UNIT_T_TNT, VRNA_UNIT_EV,
        VRNA_UNIT_WH, VRNA_UNIT_KWH }
```

Energy / Work Units.

enum vrna_unit_temperature_e {
 VRNA_UNIT_K, VRNA_UNIT_DEG_C, VRNA_UNIT_DEG_F, VRNA_UNIT_DEG_R,
 VRNA_UNIT_DEG_N, VRNA_UNIT_DEG_DE, VRNA_UNIT_DEG_RE, VRNA_UNIT_DEG_RO }

Temperature Units.

Functions

- double vrna_convert_energy (double energy, vrna_unit_energy_e from, vrna_unit_energy_e to)

 Convert between energy / work units.
- double vrna_convert_temperature (double temp, vrna_unit_temperature_e from, vrna_unit_temperature_e
 to)

Convert between temperature units.

18.114.1 Detailed Description

Physical Units and Functions to convert them into each other.

18.115 ViennaRNA/unstructured_domains.h File Reference

Functions to modify unstructured domains, e.g. to incorporate ligands binding to unpaired stretches.

Include dependency graph for unstructured_domains.h: This graph shows which files directly or indirectly include this file:

Generated by Doxygen

Data Structures

struct vrna_unstructured_domain_s

Data structure to store all functionality for ligand binding. More...

struct vrna_unstructured_domain_motif_s

Macros

• #define VRNA_UNSTRUCTURED_DOMAIN_EXT_LOOP 1U

Flag to indicate ligand bound to unpiared stretch in the exterior loop.

#define VRNA_UNSTRUCTURED_DOMAIN_HP_LOOP 2U

Flag to indicate ligand bound to unpaired stretch in a hairpin loop.

• #define VRNA UNSTRUCTURED DOMAIN INT LOOP 4U

Flag to indicate ligand bound to unpiared stretch in an interior loop.

#define VRNA_UNSTRUCTURED_DOMAIN_MB_LOOP 8U

Flag to indicate ligand bound to unpiared stretch in a multibranch loop.

#define VRNA_UNSTRUCTURED_DOMAIN_MOTIF 16U

Flag to indicate ligand binding without additional unbound nucleotides (motif-only)

• #define VRNA UNSTRUCTURED DOMAIN ALL LOOPS

Flag to indicate ligand bound to unpiared stretch in any loop (convenience macro)

Typedefs

· typedef struct vrna unstructured domain s vrna ud t

Typename for the ligand binding extension data structure vrna_unstructured_domain_s.

• typedef int() vrna_callback_ud_energy(vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, void *data)

Callback to retrieve binding free energy of a ligand bound to an unpaired sequence segment.

• typedef FLT_OR_DBL() vrna_callback_ud_exp_energy(vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, void *data)

Callback to retrieve Boltzmann factor of the binding free energy of a ligand bound to an unpaired sequence segment.

• typedef void() vrna_callback_ud_production(vrna_fold_compound_t *vc, void *data)

Callback for pre-processing the production rule of the ligand binding to unpaired stretches feature.

• typedef void() vrna_callback_ud_exp_production(vrna_fold_compound_t *vc, void *data)

Callback for pre-processing the production rule of the ligand binding to unpaired stretches feature (partition function variant)

typedef void() vrna_callback_ud_probs_add(vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type,
 FLT OR DBL exp energy, void *data)

Callback to store/add equilibrium probability for a ligand bound to an unpaired sequence segment.

• typedef FLT_OR_DBL() vrna_callback_ud_probs_get(vrna_fold_compound_t *vc, int i, int j, unsigned int loop_type, int motif, void *data)

Callback to retrieve equilibrium probability for a ligand bound to an unpaired sequence segment.

Functions

- vrna_ud_motif_t * vrna_ud_motifs_centroid (vrna_fold_compound_t *fc, const char *structure)
 Detect unstructured domains in centroid structure.
- vrna_ud_motif_t * vrna_ud_motifs_MEA (vrna_fold_compound_t *fc, const char *structure, vrna_ep_t *probability_list)

Detect unstructured domains in MEA structure.

vrna ud motif t * vrna ud motifs MFE (vrna fold compound t *fc, const char *structure)

Detect unstructured domains in MFE structure.

void vrna_ud_add_motif (vrna_fold_compound_t *vc, const char *motif, double motif_en, const char *motif
 —name, unsigned int loop_type)

Add an unstructured domain motif, e.g. for ligand binding.

int * vrna_ud_get_motif_size_at (vrna_fold_compound_t *vc, int i, unsigned int loop_type)

Get a list of unique motif sizes that start at a certain position within the sequence.

void vrna_ud_remove (vrna_fold_compound_t *vc)

Remove ligand binding to unpaired stretches.

void vrna_ud_set_data (vrna_fold_compound_t *vc, void *data, vrna_callback_free_auxdata *free_cb)

Attach an auxiliary data structure.

• void vrna_ud_set_prod_rule_cb (vrna_fold_compound_t *vc, vrna_callback_ud_production *pre_cb, vrna callback ud energy *e cb)

Attach production rule callbacks for free energies computations.

void vrna_ud_set_exp_prod_rule_cb (vrna_fold_compound_t *vc, vrna_callback_ud_exp_production *pre←
 _cb, vrna_callback_ud_exp_energy *exp_e_cb)

Attach production rule for partition function.

 void vrna_ud_set_prob_cb (vrna_fold_compound_t *vc, vrna_callback_ud_probs_add *setter, vrna_callback_ud_probs_get *getter)

18.115.1 Detailed Description

Functions to modify unstructured domains, e.g. to incorporate ligands binding to unpaired stretches.

18.115.2 Function Documentation

Get a list of unique motif sizes that start at a certain position within the sequence.

18.115.2.2 vrna_ud_set_prob_cb()

SWIG Wrapper Notes This function is attached as method ud_set_prob_cb() to objects of type fold_compound

18.116 ViennaRNA/utils.h File Reference

Use ViennaRNA/utils/basic.h instead.

Include dependency graph for utils.h:

18.116.1 Detailed Description

Use ViennaRNA/utils/basic.h instead.

Deprecated Use ViennaRNA/utils/basic.h instead

18.117 ViennaRNA/io/utils.h File Reference

Several utilities for file handling.

Include dependency graph for utils.h: This graph shows which files directly or indirectly include this file:

Functions

```
    void vrna_file_copy (FILE *from, FILE *to)
    Inefficient 'cp'.
```

char * vrna_read_line (FILE *fp)

Read a line of arbitrary length from a stream.

• int vrna_mkdir_p (const char *path)

Recursivly create a directory tree.

char * vrna basename (const char *path)

Extract the filename from a file path.

char * vrna_dirname (const char *path)

Extract the directory part of a file path.

• char * vrna_filename_sanitize (const char *name, const char *replacement)

Sanitize a file name.

• int vrna_file_exists (const char *filename)

Check if a file already exists in the file system.

18.117.1 Detailed Description

Several utilities for file handling.

,

18.118 ViennaRNA/plotting/utils.h File Reference

Various utilities to assist in plotting secondary structures and consensus structures.

Include dependency graph for utils.h: This graph shows which files directly or indirectly include this file:

Functions

- char ** vrna_annotate_covar_db (const char **alignment, const char *structure, vrna_md_t *md_p)

 Produce covariance annotation for an alignment given a secondary structure.
- vrna_cpair_t * vrna_annotate_covar_pairs (const char **alignment, vrna_ep_t *pl, vrna_ep_t *mfel, double threshold, vrna_md_t *md)

Produce covariance annotation for an alignment given a set of base pairs.

18.118.1 Detailed Description

Various utilities to assist in plotting secondary structures and consensus structures.

.

18.119 ViennaRNA/utils/strings.h File Reference

General utility- and helper-functions for RNA sequence and structure strings used throughout the ViennaRNA Package.

Include dependency graph for strings.h: This graph shows which files directly or indirectly include this file:

Macros

#define XSTR(s) STR(s)

Stringify a macro after expansion.

• #define STR(s) #s

Stringify a macro argument.

• #define FILENAME_MAX_LENGTH 80

Maximum length of filenames that are generated by our programs.

#define FILENAME_ID_LENGTH 42

Maximum length of id taken from fasta header for filename generation.

Functions

char * vrna_strdup_printf (const char *format,...)

Safely create a formatted string.

char * vrna_strdup_vprintf (const char *format, va_list argp)

Safely create a formatted string.

• int vrna_strcat_printf (char **dest, const char *format,...)

Safely append a formatted string to another string.

int vrna strcat vprintf (char **dest, const char *format, va list args)

Safely append a formatted string to another string.

char ** vrna_strsplit (const char *string, const char *delimiter)

Split a string into tokens using a delimiting character.

char * vrna random string (int I, const char symbols[])

Create a random string using characters from a specified symbol set.

int vrna_hamming_distance (const char *s1, const char *s2)

Calculate hamming distance between two sequences.

• int vrna_hamming_distance_bound (const char *s1, const char *s2, int n)

Calculate hamming distance between two sequences up to a specified length.

void vrna seg toRNA (char *sequence)

Convert an input sequence (possibly containing DNA alphabet characters) to RNA alphabet.

void vrna_seq_toupper (char *sequence)

Convert an input sequence to uppercase.

char * vrna seq ungapped (const char *seq)

Remove gap characters from a nucleotide sequence.

char * vrna cut point insert (const char *string, int cp)

Add a separating '&' character into a string according to cut-point position.

char * vrna cut point remove (const char *string, int *cp)

Remove a separating '&' character from a string.

• void str_uppercase (char *sequence)

Convert an input sequence to uppercase.

void str_DNA2RNA (char *sequence)

Convert a DNA input sequence to RNA alphabet.

• char * random_string (int I, const char symbols[])

Create a random string using characters from a specified symbol set.

int hamming (const char *s1, const char *s2)

Calculate hamming distance between two sequences.

• int hamming_bound (const char *s1, const char *s2, int n)

Calculate hamming distance between two sequences up to a specified length.

18.119.1 Detailed Description

General utility- and helper-functions for RNA sequence and structure strings used throughout the ViennaRNA Package.

18.119.2 Function Documentation

```
18.119.2.1 str_uppercase()
```

Convert an input sequence to uppercase.

Deprecated Use vrna_seq_toupper() instead!

```
18.119.2.2 str_DNA2RNA()
```

Convert a DNA input sequence to RNA alphabet.

Deprecated Use vrna_seq_toRNA() instead!

18.119.2.3 random_string()

```
char* random_string (
    int 1,
    const char symbols[] )
```

Create a random string using characters from a specified symbol set.

Deprecated Use vrna_random_string() instead!

18.119.2.4 hamming()

```
int hamming (  {\rm const~char} \ * \ s1, \\ {\rm const~char} \ * \ s2 \ )
```

Calculate hamming distance between two sequences.

Deprecated Use vrna_hamming_distance() instead!

18.119.2.5 hamming_bound()

```
int hamming_bound ( const char * s1, const char * s2, int n)
```

Calculate hamming distance between two sequences up to a specified length.

Deprecated Use vrna hamming distance bound() instead!

18.120 ViennaRNA/walk.h File Reference

Use ViennaRNA/landscape/walk.h instead.

Include dependency graph for walk.h:

18.120.1 Detailed Description

Use ViennaRNA/landscape/walk.h instead.

Deprecated Use ViennaRNA/landscape/walk.h instead

18.121 ViennaRNA/landscape/walk.h File Reference

Methods to generate particular paths such as gradient or random walks through the energy landscape of an RNA sequence.

Include dependency graph for walk.h: This graph shows which files directly or indirectly include this file:

Macros

• #define VRNA_PATH_STEEPEST_DESCENT 128

Option flag to request a steepest descent / gradient path.

• #define VRNA_PATH_RANDOM 256

Option flag to request a random walk path.

#define VRNA_PATH_NO_TRANSITION_OUTPUT 512

Option flag to omit returning the transition path.

• #define VRNA_PATH_DEFAULT (VRNA_PATH_STEEPEST_DESCENT | VRNA_MOVESET_DEFAULT)

Option flag to request defaults (steepest descent / default move set)

Functions

- vrna_move_t * vrna_path (vrna_fold_compound_t *vc, short *pt, unsigned int steps, unsigned int options)

 Compute a path, store the final structure, and return a list of transition moves from the start to the final structure.
- vrna_move_t * vrna_path_gradient (vrna_fold_compound_t *vc, short *pt, unsigned int options)
 Compute a steepest descent / gradient path, store the final structure, and return a list of transition moves from the start to the final structure.
- vrna_move_t * vrna_path_random (vrna_fold_compound_t *vc, short *pt, unsigned int steps, unsigned int options)

Generate a random walk / path of a given length, store the final structure, and return a list of transition moves from the start to the final structure.

18.121.1 Detailed Description

Methods to generate particular paths such as gradient or random walks through the energy landscape of an RNA sequence.

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822 BIBLIOGRAPHY

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Index

(Abstract) Data Structures, 576	vrna_file_PS_aln_slice, 565
bondT, 581	alimake_pair_table
cpair, 580	Deprecated Interface for Secondary Structure Utili-
PAIR, 580	ties, 688
plist, 580	alipbacktrack
sect, 581	Deprecated Interface for Global Partition Function
vrna_C11_features, 581	Computation, 670
(Nucleic Acid Sequence) String Utilitites, 466	alipf_circ_fold
FILENAME_ID_LENGTH, 467	Deprecated Interface for Global Partition Function
FILENAME_MAX_LENGTH, 467	Computation, 669
vrna_cut_point_insert, 473	alipf_fold
vrna_cut_point_remove, 473	Deprecated Interface for Global Partition Function
vrna_hamming_distance, 471	Computation, 668
vrna_hamming_distance_bound, 471	alipf_fold_par
vrna_random_string, 470	Deprecated Interface for Global Partition Function
	•
vrna_seq_toRNA, 472	Computation, 656
vrna_seq_toupper, 472	aliPS_color_aln
vrna_seq_ungapped, 473	Deprecated Interface for Plotting Utilities, 693
vrna_strcat_printf, 468	alloc_sequence_arrays
vrna_strcat_vprintf, 469	Deprecated Interface for Multiple Sequence Align-
vrna_strdup_printf, 467	ment Utilities, 679
vrna_strdup_vprintf, 468	alpha
vrna_strsplit, 470	vrna_exp_param_s, 211
(Re-)folding Paths, Saddle Points, Energy Barriers, and	Annotation, 563
Local Minima, 388	vrna_annotate_covar_db, 563
vrna_path_free, 391	vrna_annotate_covar_pairs, 563
vrna_path_options_free, 391	assign_plist_from_db
VRNA_PATH_TYPE_DOT_BRACKET, 390	Deprecated Interface for Global Partition Function
VRNA_PATH_TYPE_MOVES, 390	Computation, 667
_struct_en, 699	assign_plist_from_pr
2Dpfold.h	Deprecated Interface for Global Partition Function
destroy_TwoDpfold_variables, 707	Computation, 668
get_TwoDpfold_variables, 707	auxdata
TwoDpfold_pbacktrack, 708	vrna_fc_s, 597
TwoDpfold_pbacktrack5, 709	
TwoDpfoldList, 708	b2C
,	Deprecated Interface for Secondary Structure Utili-
add_root	ties, 683
Deprecated Interface for Secondary Structure Utili-	b2HIT
ties, 684	Deprecated Interface for Secondary Structure Utili-
alifold	ties, 682
Deprecated Interface for Global MFE Prediction,	b2Shapiro
641	Deprecated Interface for Secondary Structure Utili-
alifold.h	ties, 683
cv_fact, 712	backtrack_fold_from_pair
energy_of_alistruct, 711	Deprecated Interface for Global MFE Prediction,
nc_fact, 712	651
update_alifold_params, 712	backtrack_GQuad_IntLoop
Alignment Plots, 564	G-Quadruplexes, 420
vrna_file_PS_aln, 564	backtrack_GQuad_IntLoop_L

G-Quadruplexes, 421	Deprecated Interface for Global MFE Prediction,
backtrack_type	647
Fine-tuning of the Implemented Models, 208	Classified Dynamic Programming Variants, 354
Backtracking MFE structures, 294	co_pf_fold
vrna_backtrack5, 294	Deprecated Interface for Global Partition Function
vrna_BT_hp_loop, 295	Computation, 663
vrna_BT_int_loop, 295	co_pf_fold_par
vrna_BT_mb_loop, 296 vrna_BT_stack, 295	Deprecated Interface for Global Partition Function
base pair	Computation, 664 cofold
fold_vars.h, 747	Deprecated Interface for Global MFE Prediction,
basic.h	642
filecopy, 776	cofold_par
get_line, 773	Deprecated Interface for Global MFE Prediction,
init_rand, 775	642
int_urn, 775	Combinatorics Algorithms, 570
nrerror, 774	vrna_enumerate_necklaces, 570
print_tty_input_seq, 773	vrna_rotational_symmetry, 572
print_tty_input_seq_str, 774	vrna_rotational_symmetry_db, 574
space, 774	vrna_rotational_symmetry_db_pos, 574
time_stamp, 776	vrna_rotational_symmetry_num, 571
urn, 775	vrna_rotational_symmetry_pos, 573
warn_user, 774	vrna_rotational_symmetry_pos_num, 571
xrealloc, 775	Command Files, 532
bondT	VRNA_CMD_PARSE_DEFAULTS, 534
(Abstract) Data Structures, 581	VRNA_CMD_PARSE_HC, 533
BONUS	VRNA_CMD_PARSE_SC, 533
constants.h, 779	VRNA_CMD_PARSE_SD, 533
bp_distance Depressed Interface for Secondary Structure Utili	VRNA_CMD_PARSE_UD, 533
Deprecated Interface for Secondary Structure Utilities, 689	vrna_commands_apply, 535
bppm_symbol	vrna_commands_free, 536
Deprecated Interface for Secondary Structure Utili-	vrna_file_commands_apply, 535
ties, 691	vrna_file_commands_read, 534
bppm_to_structure	Complex Structured Modules, 418
Deprecated Interface for Secondary Structure Utili-	Compute the Centroid Structure, 350
ties, 691	vrna_centroid, 350
bt	vrna_centroid_from_plist, 350
vrna_sc_s, 268	vrna_centroid_from_probs, 351
Buffers, 634	Compute the Density of States, 371 density_of_states, 371
vrna_callback_stream_output, 634	Compute the Structure with Maximum Expected Accu-
vrna_cstr, 635	racy (MEA), 347
vrna_cstr_close, 636	MEA, 348
vrna_cstr_fflush, 636	vrna_MEA, 347
vrna_cstr_free, 635	vrna MEA from plist, 348
vrna_ostream_free, 637	compute_BPdifferences
vrna_ostream_init, 637	Deprecated Interface for Secondary Structure Utili-
vrna_ostream_provide, 638	ties, 690
vrna_ostream_request, 638	compute_probabilities
centroid	Deprecated Interface for Global Partition Function
part func.h, 784	Computation, 665
centroid.h	Computing MFE representatives of a Distance Based
get_centroid_struct_pl, 715	Partitioning, 356
get_centroid_struct_pr, 715	destroy_TwoDfold_variables, 361
circalifold	get_TwoDfold_variables, 360
Deprecated Interface for Global MFE Prediction,	TwoDfold, 364
652	TwoDfold_backtrack_f5, 362
circfold	TwoDfold_vars, 358

TwoDfoldList, 361	VRNA_CONVERT_OUTPUT_INT, 459
vrna_backtrack5_TwoD, 360	VRNA_CONVERT_OUTPUT_INT_11, 458
vrna_mfe_TwoD, 359	VRNA_CONVERT_OUTPUT_INT_21, 459
vrna_sol_TwoD_t, 358	VRNA_CONVERT_OUTPUT_INT_22, 459
Computing Partition Functions of a Distance Based Par-	VRNA_CONVERT_OUTPUT_MISC, 459
titioning, 365	VRNA_CONVERT_OUTPUT_ML, 459
vrna_pf_TwoD, 366	VRNA_CONVERT_OUTPUT_MM_EXT, 458
vrna_sol_TwoD_pf_t, 366	VRNA CONVERT OUTPUT MM HP, 457
concentrations.h	VRNA CONVERT OUTPUT MM INT, 457
get_concentrations, 720	VRNA_CONVERT_OUTPUT_MM_INT_1N, 457
cons_seq	VRNA CONVERT OUTPUT MM INT 23, 458
vrna_fc_s, 600	VRNA_CONVERT_OUTPUT_MM_MULTI, 458
constants.h	VRNA_CONVERT_OUTPUT_NINIO, 460
BONUS, 779	VRNA_CONVERT_OUTPUT_SPECIAL_HP, 460
FORBIDDEN, 779	VRNA_CONVERT_OUTPUT_STACK, 457
GASCONST, 779	VRNA_CONVERT_OUTPUT_VANILLA, 460
INF, 779	COORDINATE, 692
	copy_pair_table
K0, 779	Deprecated Interface for Secondary Structure Utili-
MAXLOOP, 779	ties, 688
NBPAIRS, 779	cost matrix
TURN, 779	dist_vars.h, 734
constrain, 579	cpair
constrain_ptypes	•
hard.h, 725	(Abstract) Data Structures, 580
Constraining the RNA Folding Grammar, 237	cut_point
VRNA_CONSTRAINT_FILE, 240	fold_vars.h, 747
VRNA_CONSTRAINT_SOFT_MFE, 240	cv_fact
VRNA_CONSTRAINT_SOFT_PF, 240	alifold.h, 712
vrna_constraints_add, 250	dangles
VRNA_DECOMP_EXT_EXT, 246	Fine-tuning of the Implemented Models, 206
VRNA_DECOMP_EXT_EXT_EXT, 248	vrna_md_s, 179
VRNA_DECOMP_EXT_EXT_STEM, 249	density of states
VRNA_DECOMP_EXT_EXT_STEM1, 249	Compute the Density of States, 371
VRNA_DECOMP_EXT_STEM, 247	Deprecated Interface for (Re-)folding Paths, Saddle
VRNA_DECOMP_EXT_STEM_EXT, 248	Points, and Energy Barriers, 696
VRNA_DECOMP_EXT_UP, 247	find_saddle, 696
VRNA_DECOMP_ML_COAXIAL, 245	free_path, 697
VRNA_DECOMP_ML_COAXIAL_ENC, 246	get_path, 697
VRNA_DECOMP_ML_ML, 244	path_t, 696
VRNA_DECOMP_ML_ML, ML, 243	Deprecated Interface for Free Energy Evaluation, 157
VRNA DECOMP ML ML STEM, 245	E_IntLoop, 168
VRNA DECOMP ML STEM, 243	E_Stem, 166
VRNA_DECOMP_ML_UP, 244	energy_of_circ_struct, 165
VRNA DECOMP PAIR HP, 241	energy_of_circ_struct_par, 160
VRNA_DECOMP_PAIR_IL, 241	
VRNA_DECOMP_PAIR_ML, 242	energy_of_circ_structure, 159
vrna_message_constraint_options, 251	energy_of_move, 162
vrna_message_constraint_options_all, 253	energy_of_move_pt, 163
convert_parameter_file	energy_of_struct, 164
Converting Energy Parameter Files, 461	energy_of_struct_par, 158
	energy_of_struct_pt, 165
Converting Energy Parameter Files, 456	energy_of_struct_pt_par, 161
convert_parameter_file, 461	energy_of_structure, 158
VRNA_CONVERT_OUTPUT_ALL, 457	energy_of_structure_pt, 160
VRNA_CONVERT_OUTPUT_BULGE, 459	exp_E_ExtLoop, 167
VRNA_CONVERT_OUTPUT_DANGLE3, 458	exp_E_IntLoop, 170
VRNA_CONVERT_OUTPUT_DANGLE5, 458	exp_E_Stem, 167
VRNA_CONVERT_OUTPUT_DUMP, 460	loop_energy, 163
VRNA_CONVERT_OUTPUT_HP, 457	Deprecated Interface for Global MFE Prediction, 640

alifold, 641	Deprecated Interface for Local (Sliding Window) MFE
backtrack_fold_from_pair, 651	Prediction, 654
circalifold, 652	Lfold, 654
circfold, 647	Lfoldz, 654
cofold, 642	Deprecated Interface for Local (Sliding Window) Partition
cofold_par, 642	Function Computation, 673
export_circfold_arrays, 650	pfl_fold, 673
export_circfold_arrays_par, 650	putoutpU_prob, 674
export_cofold_arrays, 644	putoutpU_prob_bin, 675
export_cofold_arrays_gq, 643	update_pf_paramsLP, 673
export_fold_arrays, 649	Deprecated Interface for Multiple Sequence Alignment
export_fold_arrays_par, 649	Utilities, 678
fold, 647	alloc_sequence_arrays, 679
fold_par, 646	encode_ali_sequence, 679
free_alifold_arrays, 652	free_sequence_arrays, 680
free_arrays, 648	get_mpi, 678
free_co_arrays, 642	pair_info, 678
get_monomere_mfes, 645	Deprecated Interface for Plotting Utilities, 692
HairpinE, 651	aliPS_color_aln, 693
initialize_cofold, 646	naview_xy_coordinates, 695
initialize_fold, 651	PS_color_aln, 692
LoopEnergy, 650	rna_plot_type, 695
update_cofold_params, 643	simple_circplot_coordinates, 694
update_cofold_params_par, 643	simple_xy_coordinates, 693
update_fold_params, 648	Deprecated Interface for Secondary Structure Utilities,
update_fold_params_par, 649	681
Deprecated Interface for Global Partition Function Com-	add_root, 684
putation, 655	alimake_pair_table, 688
alipbacktrack, 670	b2C, 683
alipf_circ_fold, 669	b2HIT, 682
alipf_fold, 668	b2Shapiro, 683
alipf_fold_par, 656	bp_distance, 689 bppm_symbol, 691
assign_plist_from_db, 667	bppm_to_structure, 691
assign_plist_from_pr, 668	compute_BPdifferences, 690
co_pf_fold, 663	conpute_brunerences, 690
co_pf_fold_par, 664	expand_Full, 684
compute_probabilities, 665	expand_1 dil, 004 expand_Shapiro, 684
export_ali_bppm, 669	make pair table, 687
export_bppm, 661	make_pair_table_snoop, 688
export_co_bppm, 666	make_referenceBP_array, 689
free_alipf_arrays, 670	pack structure, 686
free_co_pf_arrays, 666	parenthesis_structure, 690
free_pf_arrays, 660	parenthesis_zuker, 690
get_alipf_arrays, 671	parse_structure, 686
get_pf_arrays, 661	unexpand_aligned_F, 685
init_co_pf_fold, 665	unexpand_Full, 685
init_pf_fold, 663	unpack structure, 687
mean_bp_distance, 662	unweight, 685
mean_bp_distance_pr, 662	Deprecated Interface for Stochastic Backtracking, 676
pf_circ_fold, 659	pbacktrack, 676
pf_fold, 658	pbacktrack_circ, 677
pf_fold_par, 657	st back, 677
stackProb, 663	destroy_TwoDfold_variables
update_co_pf_params, 666	Computing MFE representatives of a Distance
update_co_pf_params_par, 667	Based Partitioning, 361
update_pf_params, 660	destroy_TwoDpfold_variables
update_pf_params_par, 660	2Dpfold.h, 707
/	'

Direct Refolding Paths between two Secondary Structures, 393 vrna_path_direct, 397	get_boltzmann_factors, 218 get_boltzmann_factors_ali, 220 get_scaled_alipf_parameters, 220
vrna_path_direct_ub, 398	get_scaled_parameters, 221
vrna path findpath, 395	get_scaled_pf_parameters, 218
vrna_path_findpath_saddle, 393	paramT, 212
vrna_path_findpath_saddle_ub, 394	pf_paramT, 212
vrna_path_findpath_ub, 395	scale_parameters, 220
vrna path options findpath, 396	
dist_vars.h	vrna_exp_params, 213 vrna_exp_params_comparative, 214
cost_matrix, 734	vrna exp params copy, 214
edit_backtrack, 734	
Distance Based Partitioning of the Secondary Structure	vrna_exp_params_rescale, 216
Space, 355	vrna_exp_params_reset, 217
do_backtrack	vrna_exp_params_subst, 215
Fine-tuning of the Implemented Models, 207	vrna_params, 212
Dot-Bracket Notation of Secondary Structures, 479	vrna_params_copy, 213
VRNA_BRACKETS_ALPHA, 479	vrna_params_reset, 217
VRNA_BRACKETS_ANG, 480	vrna_params_subst, 215
VRNA BRACKETS ANY, 481	energy_of_alistruct
VRNA BRACKETS CLY, 480	alifold.h, 711
VRNA BRACKETS DEFAULT, 481	energy_of_circ_struct
VRNA_BRACKETS_RND, 480	Deprecated Interface for Free Energy Evaluation,
VRNA_BRACKETS_SQR, 480	165
vrna_db_flatten, 483	energy_of_circ_struct_par
vrna_db_flatten_to, 483	Deprecated Interface for Free Energy Evaluation,
vrna_db_from_plist, 485	160
vrna_db_from_ptable, 484	energy_of_circ_structure
vrna_db_from_WUSS, 484	Deprecated Interface for Free Energy Evaluation, 159
vrna_db_pack, 482	
vrna_db_pk_remove, 486	energy_of_move
vrna_db_to_element_string, 485	Deprecated Interface for Free Energy Evaluation, 162
vrna_db_unpack, 482	
duplexT, 579	energy_of_move_pt Deprecated Interface for Free Energy Evaluation,
dupVar, 580	163
	energy_of_struct
E_Hairpin	Deprecated Interface for Free Energy Evaluation,
Hairpin Loops, 436	164
E_IntLoop	energy_of_struct_par
Deprecated Interface for Free Energy Evaluation,	Deprecated Interface for Free Energy Evaluation,
168	158
E_Stem	energy_of_struct_pt
Deprecated Interface for Free Energy Evaluation,	Deprecated Interface for Free Energy Evaluation,
166	165
edit_backtrack	energy_of_struct_pt_par
dist_vars.h, 734	Deprecated Interface for Free Energy Evaluation,
encode_ali_sequence Deprecated Interface for Multiple Sequence Align-	161
	energy_of_structure
ment Utilities, 679	Deprecated Interface for Free Energy Evaluation,
vrna_ht_entry_db_t, 616	158
Energy Evaluation for Atomic Moves, 155	energy_of_structure_pt
vrna_eval_move, 155	Deprecated Interface for Free Energy Evaluation,
vrna_eval_move_pt, 156	160
Energy Evaluation for Individual Loops, 152	energy_set
vrna_eval_loop_pt, 153	Fine-tuning of the Implemented Models, 207
vrna_eval_loop_pt_v, 153	equilibrium_probs.h
Energy Parameters, 209	vrna_pr_energy, 737
get_boltzmann_factor_copy, 219	exp_E_ExtLoop
3	

Deprecated Interface for Free Energy Evaluation, 167	vrna_sc_s, 268 filecopy
exp_E_Hairpin	basic.h, 776
Hairpin Loops, 437	FILENAME_ID_LENGTH
exp_E_IntLoop	(Nucleic Acid Sequence) String Utilitites, 467
Deprecated Interface for Free Energy Evaluation,	FILENAME_MAX_LENGTH
170	(Nucleic Acid Sequence) String Utilitites, 467
exp_E_Stem	Files and I/O, 511
Deprecated Interface for Free Energy Evaluation,	readribosum, 512
167	vrna_file_exists, 513
exp_f	vrna_filename_sanitize, 512
vrna_sc_s, 268	vrna_read_line, 512
expand_Full	final_cost
Deprecated Interface for Secondary Structure Utili-	Inverse Folding (Design), 373
ties, 684	find_saddle
expand_Shapiro	Deprecated Interface for (Re-)folding Paths, Saddle
Deprecated Interface for Secondary Structure Utili-	Points, and Energy Barriers, 696
ties, 684	Fine-tuning of the Implemented Models, 173
Experimental Structure Probing Data, 405	backtrack_type, 208
expHairpinEnergy	dangles, 206
part_func.h, 785	do_backtrack, 207
expLoopEnergy	energy_set, 207
part_func.h, 784	max_bp_span, 208
export_ali_bppm	noLonelyPairs, 207
Deprecated Interface for Global Partition Function	nonstandards, 208
Computation, 669	pf_scale, 206
export_bppm	set_model_details, 205
Deprecated Interface for Global Partition Function	temperature, 206
Computation, 661	tetra_loop, 207
export_circfold_arrays Deprecated Interface for Global MFE Prediction,	vrna_md_copy, 186
650	vrna_md_defaults_backtrack, 197
export_circfold_arrays_par	vrna_md_defaults_backtrack_get, 197
Deprecated Interface for Global MFE Prediction,	vrna_md_defaults_backtrack_type, 198
650	vrna_md_defaults_backtrack_type_get, 198
export_co_bppm	vrna_md_defaults_betaScale, 189
Deprecated Interface for Global Partition Function	vrna_md_defaults_betaScale_get, 189
Computation, 666	vrna_md_defaults_circ, 194
export_cofold_arrays	vrna_md_defaults_circ_get, 194
Deprecated Interface for Global MFE Prediction,	vrna_md_defaults_compute_bpp, 198
644	vrna_md_defaults_compute_bpp_get, 199
export_cofold_arrays_gq	vrna_md_defaults_cv_fact, 203
Deprecated Interface for Global MFE Prediction,	vrna_md_defaults_cv_fact_get, 203
643	vrna_md_defaults_dangles, 189
export_fold_arrays	vrna_md_defaults_dangles_get, 190
Deprecated Interface for Global MFE Prediction,	vrna_md_defaults_energy_set, 196
649	vrna_md_defaults_energy_set_get, 197
export_fold_arrays_par	vrna_md_defaults_gquad, 195
Deprecated Interface for Global MFE Prediction,	vrna_md_defaults_gquad_get, 195
649	vrna_md_defaults_logML, 193
Extending the Folding Grammar with Additional Do-	vrna_md_defaults_logML_get, 194
mains, 223	vrna_md_defaults_max_bp_span, 199
Exterior Loops, 430	vrna_md_defaults_max_bp_span_get, 200
vrna_E_ext_loop, 431	vrna_md_defaults_min_loop_size, 200
vrna_E_ext_stem, 431	vrna_md_defaults_min_loop_size_get, 200
vrna_exp_E_ext_stem, 432	vrna_md_defaults_nc_fact, 204
vrna_mx_pf_aux_el_t, 430	vrna_md_defaults_nc_fact_get, 204
f	vrna_md_defaults_noGU, 192
f	vrna_md_defaults_noGU_get, 192

	vrna_md_defaults_noGUclosure, 192 vrna_md_defaults_noGUclosure_get, 193 vrna_md_defaults_noLP, 191 vrna_md_defaults_noLP_get, 191 vrna_md_defaults_oldAliEn, 201 vrna_md_defaults_oldAliEn_get, 202 vrna_md_defaults_reset, 187 vrna_md_defaults_ribo, 202 vrna_md_defaults_ribo_get, 203 vrna_md_defaults_ribo_get, 203 vrna_md_defaults_sfact, 204 vrna_md_defaults_sfact_get, 205 vrna_md_defaults_special_hp, 190 vrna_md_defaults_special_hp_get, 191 vrna_md_defaults_temperature, 188 vrna_md_defaults_temperature_get, 188 vrna_md_defaults_uniq_ML, 195 vrna_md_defaults_uniq_ML_get, 196 vrna_md_defaults_window_size, 201 vrna_md_defaults_window_size_get, 201 vrna_md_option_string, 187 vrna_md_set_default_186	pr, 747 RibosumFile, 747 Folding Paths that start at a single Secondary Structure, 400 vrna_path, 401 VRNA_PATH_DEFAULT, 401 vrna_path_gradient, 402 VRNA_PATH_NO_TRANSITION_OUTPUT, 401 VRNA_PATH_NO_TRANSITION_OUTPUT, 401 VRNA_PATH_RANDOM, 400 vrna_path_random, 403 VRNA_PATH_STEEPEST_DESCENT, 400 FORBIDDEN constants.h, 779 Free Energy Evaluation, 127 vrna_eval_circ_consensus_structure, 142 vrna_eval_circ_consensus_structure_v, 146 vrna_eval_circ_gquad_consensus_structure_v, 147 vrna_eval_circ_gquad_structure, 137
	vrna_md_set_default, 186 vrna_md_update, 186 VRNA_MODEL_DEFAULT_ALI_CV_FACT, 185 VRNA_MODEL_DEFAULT_ALI_NC_FACT, 185 VRNA_MODEL_DEFAULT_ALI_OLD_EN, 185 VRNA_MODEL_DEFAULT_ALI_RIBO, 185 VRNA_MODEL_DEFAULT_BACKTRACK, 183 VRNA_MODEL_DEFAULT_BACKTRACK_TYPE, 183 VRNA_MODEL_DEFAULT_BETA_SCALE, 181 VRNA_MODEL_DEFAULT_CIRC, 182 VRNA_MODEL_DEFAULT_COMPUTE_BPP, 184 VRNA_MODEL_DEFAULT_DANGLES, 181 VRNA_MODEL_DEFAULT_ENERGY_SET, 183 VRNA_MODEL_DEFAULT_GQUAD, 182 VRNA_MODEL_DEFAULT_LOG_ML, 184 VRNA_MODEL_DEFAULT_NO_GU, 182 VRNA_MODEL_DEFAULT_NO_GU, 182 VRNA_MODEL_DEFAULT_NO_GU_CLOSURE, 182	vrna_eval_circ_gquad_structure_v, 140 vrna_eval_circ_structure, 135 vrna_eval_circ_structure_v, 139 vrna_eval_consensus_structure_pt_simple, 150 vrna_eval_consensus_structure_simple, 141 vrna_eval_consensus_structure_simple_v, 145 vrna_eval_consensus_structure_simple_verbose,
fold	VRNA_MODEL_DEFAULT_NO_LP, 181 VRNA_MODEL_DEFAULT_PF_SCALE, 180 VRNA_MODEL_DEFAULT_SPECIAL_HP, 181 VRNA_MODEL_DEFAULT_TEMPERATURE, 180 VRNA_MODEL_DEFAULT_UNIQ_ML, 183 VRNA_MODEL_DEFAULT_WINDOW_SIZE, 184 Deprecated Interface for Global MFE Prediction,	vrna_eval_structure_pt_verbose, 133 vrna_eval_structure_simple, 135 vrna_eval_structure_simple_v, 138 vrna_eval_structure_simple_verbose, 137 vrna_eval_structure_v, 132 vrna_eval_structure_verbose, 131 free_alifold_arrays Deprecated Interface for Global MFE Prediction,
fold_	Deprecated Interface for Global MFE Prediction, 646 _vars.h base_pair, 747 cut_point, 747 iindx, 748 james_rule, 747	free_alipf_arrays Deprecated Interface for Global Partition Function Computation, 670 free_arrays Deprecated Interface for Global MFE Prediction, 648 free_auxdata vrna_fc_s, 597
	logML, 747	free_co_arrays

Deprecated Interface for Global MFE Prediction, 642	get_input_line Utilities, 427
free_co_pf_arrays	get line
Deprecated Interface for Global Partition Function	-
Computation, 666	basic.h, 773
free_data	get_monomere_mfes Deprecated Interface for Global MFE Prediction,
vrna_hc_s, 257	645
free_path	
Deprecated Interface for (Re-)folding Paths, Saddle	get_mpi Depresented Interface for Multiple Seguence Align
Points, and Energy Barriers, 697	Deprecated Interface for Multiple Sequence Alignment Hillitias 679
free_pf_arrays	ment Utilities, 678
Deprecated Interface for Global Partition Function	get_path Personal Interface for /Pe \felding Paths, Saddle
Computation, 660	Deprecated Interface for (Re-)folding Paths, Saddle
free_profile	Points, and Energy Barriers, 697
profiledist.h, 800	get_pf_arrays
free_sequence_arrays	Deprecated Interface for Global Partition Function
Deprecated Interface for Multiple Sequence Align-	Computation, 661
ment Utilities, 680	get_plist
free tree	part_func_co.h, 786
treedist.h, 810	get_scaled_alipf_parameters
treedist.ii, oro	Energy Parameters, 220
G-Quadruplexes, 419	get_scaled_parameters
backtrack_GQuad_IntLoop, 420	Energy Parameters, 221
backtrack_GQuad_IntLoop_L, 421	get_scaled_pf_parameters
get_gquad_matrix, 419	Energy Parameters, 218
parse_gquad, 420	get_TwoDfold_variables
GASCONST	Computing MFE representatives of a Distance
constants.h, 779	Based Partitioning, 360
Generate Soft Constraints from Data, 410	get_TwoDpfold_variables
progress_callback, 412	2Dpfold.h, 707
VRNA_MINIMIZER_CONJUGATE_FR, 411	give_up
VRNA_MINIMIZER_CONJUGATE_PR, 411	Inverse Folding (Design), 373
VRNA_MINIMIZER_STEEPEST_DESCENT, 412	Global MFE Prediction, 282
VRNA MINIMIZER VECTOR BFGS, 411	vrna_alifold, 285
VRNA MINIMIZER VECTOR BFGS2, 412	vrna_circalifold, 286
VRNA_OBJECTIVE_FUNCTION_ABSOLUTE,	vrna_circfold, 285
411	vrna_cofold, 287
VRNA_OBJECTIVE_FUNCTION_QUADRATIC,	vrna_fold, 284
411	vrna_mfe, 283
vrna_sc_minimize_pertubation, 413	vrna_mfe_dimer, 283
get_alipf_arrays	Global Partition Function and Equilibrium Probabilities,
Deprecated Interface for Global Partition Function	297
Computation, 671	vrna_ensemble_defect, 300
get_boltzmann_factor_copy	vrna_mean_bp_distance, 299
Energy Parameters, 219	vrna_mean_bp_distance_pr, 299
get_boltzmann_factors	vrna_pf, 302
Energy Parameters, 218	vrna_pf_alifold, 305
get_boltzmann_factors_ali	vrna_pf_circalifold, 306
Energy Parameters, 220	vrna_pf_circfold, 304
get_centroid_struct_gquad_pr	vrna_pf_co_fold, 309
part_func.h, 784	vrna_pf_dimer, 303
get_centroid_struct_pl	vrna_pf_dimer_probs, 301
centroid.h, 715	vrna_pf_fold, 304
get_centroid_struct_pr	vrna_plist_from_probs, 308
centroid.h, 715	vrna_positional_entropy, 308
get_concentrations	vrna_pr_structure, 302
concentrations.h, 720	vrna_stack_prob, 301
get_gquad_matrix	gmlRNA
G-Quadruplexes, 419	Plotting, 540

Hairpin Loops, 434	vrna_ht_remove, 622
E_Hairpin, 436	vrna_ht_size, 619
exp_E_Hairpin, 437	Heaps, 626
vrna_E_ext_hp_loop, 435	vrna_callback_heap_cmp, 627
vrna_E_hp_loop, 434	vrna_callback_heap_get_pos, 627
vrna_eval_hp_loop, 435	vrna_callback_heap_set_pos, 628
vrna_exp_E_hp_loop, 438	vrna_heap_free, 629
HairpinE	vrna_heap_init, 628
Deprecated Interface for Global MFE Prediction,	vrna_heap_insert, 630
651	vrna_heap_pop, 630
hamming	vrna_heap_remove, 631
strings.h, 817	vrna_heap_size, 630
hamming_bound	vrna_heap_t, 627
strings.h, 817	vrna_heap_top, 631
Hard Constraints, 254	vrna_heap_update, 632
vrna_callback_hc_evaluate, 261	Helix List Representation of Secondary Structures, 493
VRNA_CONSTRAINT_DB, 257	vrna_hx_from_ptable, 493
VRNA_CONSTRAINT_DB_DEFAULT, 260	id
VRNA_CONSTRAINT_DB_DOT, 258	vrna_exp_param_s, 211
VRNA_CONSTRAINT_DB_ENFORCE_BP, 257	iindx
VRNA_CONSTRAINT_DB_GQUAD, 260	fold_vars.h, 748
VRNA_CONSTRAINT_DB_INTERMOL, 259	Incorporating Ligands Binding to Specific Se-
VRNA_CONSTRAINT_DB_INTRAMOL, 259	quence/Structure Motifs using Soft Con-
VRNA_CONSTRAINT_DB_PIPE, 258	straints, 417
VRNA_CONSTRAINT_DB_RND_BRACK, 259	vrna_sc_add_hi_motif, 417
VRNA_CONSTRAINT_DB_WUSS, 260	INF
VRNA_CONSTRAINT_DB_X, 258	constants.h, 779
vrna_hc_add_bp, 263	init_co_pf_fold
vrna_hc_add_bp_nonspecific, 264	Deprecated Interface for Global Partition Function
vrna_hc_add_from_db, 265	Computation, 665
vrna_hc_add_up, 262	init_pf_fold
vrna_hc_add_up_batch, 263	Deprecated Interface for Global Partition Function
vrna_hc_free, 264	Computation, 663
vrna_hc_init, 262	init_pf_foldLP
hard.h	LPfold.h, 757
constrain_ptypes, 725	init_rand
print_tty_constraint, 725	basic.h, 775
print_tty_constraint_full, 725	initialize_cofold
VRNA_CONSTRAINT_DB_ANG_BRACK, 723	Deprecated Interface for Global MFE Prediction,
VRNA_CONSTRAINT_NO_HEADER, 723	646
vrna_hc_add_data, 724	initialize_fold
VRNA HC DEFAULT, 724	Deprecated Interface for Global MFE Prediction,
vrna_hc_type_e, 724	651
VRNA_HC_WINDOW, 724	int_urn
Hash Tables, 615	basic.h, 775
vrna_callback_ht_compare_entries, 617	interact, 578
vrna callback ht free entry, 618	Internal Loops, 439
vrna_callback_ht_hash_function, 617	vrna_eval_int_loop, 439
vrna_hash_table_t, 616	inv_verbose
vrna_ht_clear, 622	Inverse Folding (Design), 374
vrna_ht_collisions, 619	Inverse Folding (Design), 372
vrna_ht_db_comp, 623	final_cost, 373
vrna_ht_db_free_entry, 624	give_up, 373
vrna_ht_db_hash_func, 624	inv_verbose, 374
vrna_ht_free, 623	inverse_fold, 372
vrna_ht_get, 621	inverse_loid, 372 inverse_pf_fold, 373
	_ , _
vrna_ht_init, 618	inverse_fold
vrna_ht_insert, 621	Inverse Folding (Design), 372

inverse_pf_fold Inverse Folding (Design), 373	vrna_pfl_fold, 316 vrna_pfl_fold_cb, 317 vrna_pfl_fold_up, 317
james_rule	vrna_pfl_fold_up_cb, 318
fold_vars.h, 747	vrna_probs_window, 315
1010_va15.11, 747	
K0	VRNA_PROBS_WINDOW_BPP, 312
constants.h, 779	vrna_probs_window_callback, 314
ochotamom, 770	VRNA_PROBS_WINDOW_PF, 313
last_parameter_file	VRNA_PROBS_WINDOW_STACKP, 313
Reading/Writing Energy Parameter Sets from/to	VRNA_PROBS_WINDOW_UP, 312
File, 454	VRNA_PROBS_WINDOW_UP_SPLIT, 313
Layouts and Coordinates, 544	logML
vrna_plot_coords, 552	fold_vars.h, 747
	loop_energy
vrna_plot_coords_circular, 555	Deprecated Interface for Free Energy Evaluation,
vrna_plot_coords_circular_pt, 556	163
vrna_plot_coords_naview, 557	LoopEnergy
vrna_plot_coords_naview_pt, 557	Deprecated Interface for Global MFE Prediction,
vrna_plot_coords_pt, 553	650
vrna_plot_coords_puzzler, 558	LPfold.h
vrna_plot_coords_puzzler_pt, 559	init pf foldLP, 757
vrna_plot_coords_simple, 554	LST BUCKET, 699
vrna_plot_coords_simple_pt, 554	
vrna_plot_coords_turtle, 561	Make_bp_profile
vrna_plot_coords_turtle_pt, 562	profiledist.h, 801
vrna_plot_layout, 547	Make_bp_profile_bppm
vrna_plot_layout_circular, 549	profiledist.h, 800
vrna_plot_layout_free, 551	make_pair_table
vrna_plot_layout_naview, 549	Deprecated Interface for Secondary Structure Utili-
vrna_plot_layout_puzzler, 551	ties, 687
vrna_plot_layout_simple, 548	make_pair_table_snoop
vrna_plot_layout_t, 547	Deprecated Interface for Secondary Structure Utili-
vrna_plot_layout_turtle, 550	ties, 688
vrna_plot_options_puzzler, 560	make referenceBP array
vrna_plot_options_puzzler_free, 560	Deprecated Interface for Secondary Structure Utili-
VRNA_PLOT_TYPE_CIRCULAR, 546	ties, 689
VRNA PLOT TYPE NAVIEW, 546	Make_swString
VRNA_PLOT_TYPE_PUZZLER, 547	stringdist.h, 806
VRNA_PLOT_TYPE_SIMPLE, 545	make_tree
VRNA_PLOT_TYPE_TURTLE, 546	treedist.h, 810
Lfold	
Deprecated Interface for Local (Sliding Window)	max_bp_span Fine tuning of the Implemented Models 208
MFE Prediction, 654	Fine-tuning of the Implemented Models, 208
	MAXLOOP
Lfoldz Deprecated Interface for Local (Sliding Window)	constants.h, 779 MEA
MFE Prediction, 654	Compute the Structure with Maximum Expected
ligand.h	Accuracy (MEA), 348
vrna_sc_motif_t, 726	mean_bp_dist
Ligands Binding to RNA Structures, 415	part_func.h, 784
Ligands Binding to Unstructured Domains, 416	mean_bp_distance
LIST, 699	Deprecated Interface for Global Partition Function
Local (sliding window) MFE Prediction, 289	Computation, 662
vrna_Lfold, 292	mean_bp_distance_pr
vrna_Lfoldz, 293	Deprecated Interface for Global Partition Function
vrna_mfe_window, 291	Computation, 662
vrna_mfe_window_callback, 290	Messages, 583
vrna_mfe_window_zscore, 291	vrna_message_error, 583
Local (sliding window) Partition Function and Equilibrium	vrna_message_info, 585
Probabilities, 311	vrna_message_input_seq, 586

vrna_message_input_seq_simple, 586	vrna_move_is_insertion, 382
vrna_message_verror, 584	vrna_move_is_removal, 382
vrna_message_vinfo, 586	vrna_move_is_shift, 383
vrna_message_vwarning, 585	vrna_move_list_free, 381
vrna_message_warning, 584	vrna_move_neighbor_diff, 386
min_loop_size	vrna_move_neighbor_diff_cb, 386
vrna_md_s, 180	VRNA_MOVESET_DEFAULT, 379
Minimum Free Energy (MFE) Algorithms, 279	VRNA_MOVESET_DELETION, 378
mm.h	VRNA_MOVESET_INSERTION, 378
	VRNA MOVESET NO LP, 379
vrna_maximum_matching, 760	,
vrna_maximum_matching_simple, 760	VRNA_MOVESET_SHIFT, 379
Multibranch Loops, 440	VRNA_NEIGHBOR_CHANGE, 379
vrna_E_mb_loop_stack, 441	VRNA_NEIGHBOR_INVALID, 380
vrna_mx_pf_aux_ml_t, 440	VRNA_NEIGHBOR_NEW, 380
Multiple Sequence Alignment Utilities, 501	vrna_neighbors, 384
vrna_aln_consensus_mis, 510	vrna_neighbors_successive, 385
vrna_aln_consensus_sequence, 510	node, 580
vrna_aln_conservation_col, 509	noLonelyPairs
vrna_aln_conservation_struct, 508	Fine-tuning of the Implemented Models, 207
vrna aln copy, 508	nonstandards
vrna_aln_free, 505	Fine-tuning of the Implemented Models, 208
vrna_aln_mpi, 503	nrerror
vrna_aln_pinfo, 503	basic.h, 774
vrna_aln_slice, 505	Nucleic Acid Sequences and Structures, 515
vrna_aln_toRNA, 507	read_record, 521
vrna_aln_uppercase, 507	VRNA_CONSTRAINT_MULTILINE, 516
VRNA_MEASURE_SHANNON_ENTROPY, 503	vrna_extract_record_rest_constraint, 521
Multiple Sequence Alignments, 523	vrna_extract_record_rest_structure, 520
VRNA_FILE_FORMAT_MSA_APPEND, 526	vrna_file_bpseq, 517
VRNA_FILE_FORMAT_MSA_CLUSTAL, 524	vrna_file_connect, 517
VRNA_FILE_FORMAT_MSA_DEFAULT, 525	vrna_file_fasta_read_record, 518
VRNA_FILE_FORMAT_MSA_FASTA, 524	vrna_file_helixlist, 516
VRNA_FILE_FORMAT_MSA_MAF, 524	vrna_file_json, 518
VRNA_FILE_FORMAT_MSA_MIS, 525	vrna_file_SHAPE_read, 520
VRNA_FILE_FORMAT_MSA_NOCHECK, 525	VRNA_OPTION_MULTILINE, 515
VRNA_FILE_FORMAT_MSA_QUIET, 526	
VRNA FILE FORMAT MSA SILENT, 526	pack_structure
VRNA_FILE_FORMAT_MSA_STOCKHOLM, 524	Deprecated Interface for Secondary Structure Utili-
VRNA_FILE_FORMAT_MSA_UNKNOWN, 526	ties, 686
vrna_file_msa_detect_format, 529	PAIR
vrna_file_msa_read, 527	(Abstract) Data Structures, 580
vrna_file_msa_read_record, 528	Pair List Representation of Secondary Structures, 491
vrna file msa write, 530	vrna_plist, 492
viria_iiie_iiisa_write, 550	Pair Table Representation of Secondary Structures, 487
n_seq	vrna_pt_ali_get, 489
vrna_fc_s, 599	vrna_pt_at_get, 488
naview_xy_coordinates	vrna_pt_pk_remove, 489
Deprecated Interface for Plotting Utilities, 695	vrna_pt_pk_remove, 409 vrna_pt_snoop_get, 489
-	— · — ·
NBPAIRS	vrna_ptable, 487
constants.h, 779	vrna_ptable_copy, 489
nc_fact	vrna_ptable_from_string, 488
alifold.h, 712	pair_info
Neighborhood Relation and Move Sets for Secondary	Deprecated Interface for Multiple Sequence Align-
Structures, 375	ment Utilities, 678
vrna_callback_move_update, 380	paramT
vrna_loopidx_update, 384	Energy Parameters, 212
vrna_move_apply, 381	parenthesis_structure
vrna_move_compare, 383	Deprecated Interface for Secondary Structure Utili-
vrna_move_init, 381	ties, 690

parenthesis zuker	Plotting, 537
Deprecated Interface for Secondary Structure Utili-	gmlRNA, 540
ties, 690	PS_dot_plot, 539
parse_gquad	PS_dot_plot_list, 538
G-Quadruplexes, 420	PS_rna_plot, 542
parse_structure	PS_rna_plot_a, 543
Deprecated Interface for Secondary Structure Utili-	PS_rna_plot_a_gquad, 543
ties, 686	ssv_rna_plot, 541
part_func.h	svg_rna_plot, 541
centroid, 784	vrna_file_PS_rnaplot, 539
expHairpinEnergy, 785	vrna_file_PS_rnaplot_a, 540
expLoopEnergy, 784	xrna_plot, 542
get_centroid_struct_gquad_pr, 784	Postorder_list, 700
mean_bp_dist, 784	pr
part_func_co.h	fold_vars.h, 747
get_plist, 786	print_tty_constraint
Partition Function and Equilibrium Properties, 280	hard.h, 725
vrna_pf_float_precision, 281	print_tty_constraint_full
Partition Function for Two Hybridized Sequences, 442	hard.h, 725
vrna_pf_co_fold, 443	print_tty_input_seq
vrna_pf_dimer_concentrations, 443	basic.h, 773
Partition Function for two Hybridized Sequences as a	print_tty_input_seq_str
Stepwise Process, 445	basic.h, 774
pf_interact, 446	prod_cb
pf_unstru, 445	vrna_unstructured_domain_s, 227
path_t	profile_edit_distance
Deprecated Interface for (Re-)folding Paths, Saddle	profiledist.h, 800
Points, and Energy Barriers, 696	profiledist.h
pbacktrack	free_profile, 800
Deprecated Interface for Stochastic Backtracking,	Make_bp_profile, 801
676	Make_bp_profile_bppm, 800
pbacktrack_circ	profile_edit_distance, 800
Deprecated Interface for Stochastic Backtracking,	progress_callback
677	Generate Soft Constraints from Data, 412
pf_circ_fold	PS_color_aln
Deprecated Interface for Global Partition Function	Deprecated Interface for Plotting Utilities, 692
Computation, 659	PS_dot_plot
pf_fold	Plotting, 539
Deprecated Interface for Global Partition Function	PS_dot_plot_list
Computation, 658	Plotting, 538
pf_fold_par	PS_rna_plot
Deprecated Interface for Global Partition Function	Plotting, 542
Computation, 657	PS_rna_plot_a
pf_interact	Plotting, 543
Partition Function for two Hybridized Sequences as	PS_rna_plot_a_gquad
a Stepwise Process, 446	Plotting, 543
pf_paramT	pscore
Energy Parameters, 212	vrna_fc_s, 601
pf_scale	pscore_local
Fine-tuning of the Implemented Models, 206	vrna_fc_s, 601
pf_unstru	pscore_pf_compat
Partition Function for two Hybridized Sequences as	vrna_fc_s, 601
a Stepwise Process, 445	ptype
pfl_fold	vrna_fc_s, 598
Deprecated Interface for Local (Sliding Window)	ptype_pf_compat
Partition Function Computation, 673	vrna_fc_s, 598
plist (Abetraet) Pete Structures 500	pu_contrib, 578
(Abstract) Data Structures, 580	pu_out, 579

putoutpU_prob	vrna_fc_s, 601
Deprecated Interface for Local (Sliding Window)	S5
Partition Function Computation, 674	vrna_fc_s, 600
putoutpU_prob_bin	S_cons
Deprecated Interface for Local (Sliding Window)	vrna_fc_s, 600
Partition Function Computation, 675	SC
D 0: 0 (vrna_fc_s, 599
Random Structure Samples from the Ensemble, 329	scale_parameters
vrna_boltzmann_sampling_callback, 331	Energy Parameters, 220
vrna_pbacktrack5_338	SCS
vrna_pbacktrack5, 332 vrna_pbacktrack5_cb, 334	vrna_fc_s, 602
vrna_pbacktrack5_num, 332	Search Algorithms, 566
vrna_pbacktrack5_resume, 335	vrna_search_BM_BCT, 568
vrna_pbacktrack5_resume_cb, 336	vrna_search_BM_BCT_num, 568
vrna_pbacktrack_cb, 340	vrna_search_BMH, 567
VRNA_PBACKTRACK_DEFAULT, 330	vrna_search_BMH_num, 566
vrna pbacktrack mem free, 344	Secondary Structure Utilities, 475
vrna_pbacktrack_mem_t, 331	vrna_bp_distance, 475
VRNA PBACKTRACK NON REDUNDANT, 330	vrna_db_from_bp_stack, 477
vrna_pbacktrack_num, 339	vrna_refBPcnt_matrix, 477
vrna_pbacktrack_resume, 341	vrna_refBPdist_matrix, 477
vrna_pbacktrack_resume_cb, 343	sect
random_string	(Abstract) Data Structures, 581
strings.h, 817	sequence
read_parameter_file	vrna_fc_s, 597
Reading/Writing Energy Parameter Sets from/to	sequence_encoding
File, 455	vrna_fc_s, 598
read record	sequences
Nucleic Acid Sequences and Structures, 521	vrna_fc_s, 599
Reading/Writing Energy Parameter Sets from/to File,	set_model_details
448	Fine-tuning of the Implemented Models, 205
last_parameter_file, 454	SHAPE Reactivity Data, 406
read parameter file, 455	vrna_sc_add_SHAPE_deigan, 406
VRNA_PARAMETER_FORMAT_DEFAULT, 449	vrna_sc_add_SHAPE_deigan_ali, 407
vrna_params_load, 449	vrna_sc_add_SHAPE_zarringhalam, 408
vrna_params_load_defaults, 451	vrna_sc_SHAPE_to_pr, 408
vrna_params_load_DNA_Mathews1999, 454	SHAPE.h
vrna_params_load_DNA_Mathews2004, 453	vrna_sc_SHAPE_parse_method, 728
vrna_params_load_from_string, 450	simple_circplot_coordinates
vrna_params_load_RNA_Andronescu2007, 452	Deprecated Interface for Plotting Utilities, 694
vrna_params_load_RNA_Langdon2018, 452	simple_xy_coordinates
vrna_params_load_RNA_misc_special_hairpins,	Deprecated Interface for Plotting Utilities, 693
453	snoopT, 580
vrna_params_load_RNA_Turner1999, 451	Soft Constraints, 266
vrna_params_load_RNA_Turner2004, 451	vrna_callback_sc_backtrack, 271
vrna_params_save, 449	vrna_callback_sc_energy, 268
write_parameter_file, 455	vrna_callback_sc_exp_energy, 270
readribosum	vrna_sc_add_bp, 273
Files and I/O, 512	vrna_sc_add_bt, 276
RibosumFile	vrna_sc_add_data, 275
fold_vars.h, 747	vrna_sc_add_exp_f, 277
RNA-RNA Interaction, 353	vrna_sc_add_f, 276
rna_plot_type	vrna_sc_add_up, 274
Deprecated Interface for Plotting Utilities, 695	vrna_sc_free, 275
	vrna_sc_init, 271
S	vrna_sc_remove, 274
vrna_fc_s, 600	vrna_sc_set_bp, 272
S3	vrna_sc_set_up, <mark>273</mark>

soft.h	vrna_subopt_callback, 324
VRNA_SC_DEFAULT, 729	vrna_subopt_cb, 326
vrna_sc_type_e, 729	Suboptimals and Representative Structures, 321
VRNA_SC_WINDOW, 729	svg_rna_plot
SOLUTION	Plotting, 541
subopt.h, 809	swString, 700
space	temperature
basic.h, 774	Fine-tuning of the Implemented Models, 206
ssv_rna_plot	tetra loop
Plotting, 541	Fine-tuning of the Implemented Models, 207
st_back	The Dynamic Programming Matrices, 610
Deprecated Interface for Stochastic Backtracking,	VRNA_MX_2DFOLD, 612
677 stackProb	vrna_mx_add, 612
Deprecated Interface for Global Partition Function	VRNA_MX_DEFAULT, 612
Computation, 663	vrna_mx_mfe_free, 613
stat cb	vrna_mx_pf_free, 613
vrna_fc_s, 597	vrna_mx_type_e, 612
Stochastic Backtracking of Structures from Distance	VRNA_MX_WINDOW, 612
Based Partitioning, 368	The Fold Compound, 593
vrna pbacktrack5 TwoD, 369	vrna_callback_free_auxdata, 604
vrna_pbacktrack_TwoD, 368	vrna_callback_recursion_status, 605
str_DNA2RNA	VRNA_FC_TYPE_COMPARATIVE, 605
strings.h, 817	vrna_fc_type_e, 605
str_uppercase	VRNA_FC_TYPE_SINGLE, 605
strings.h, 816	vrna_fold_compound_add_auxdata_608
string_edit_distance	vrna_fold_compound_add_auxdata, 608 vrna_fold_compound_add_callback, 609
stringdist.h, 806	vrna_fold_compound_comparative, 607
stringdist.h	vrna_fold_compound_free, 608
Make_swString, 806	VRNA_OPTION_EVAL_ONLY, 604
string_edit_distance, 806	VRNA_OPTION_MFE, 603
strings.h	VRNA OPTION PF, 603
hamming, 817	VRNA_STATUS_MFE_POST, 602
hamming_bound, 817	VRNA_STATUS_MFE_PRE, 602
random_string, 817	VRNA_STATUS_PF_POST, 603
str_DNA2RNA, 817	VRNA_STATUS_PF_PRE, 603
str_uppercase, 816	The RNA Folding Grammar, 172
structure	The RNA Secondary Structure Landscape, 278
vrna_ht_entry_db_t, 616	time_stamp
Structured Domains, 236	basic.h, 776
subopt	Tree, 700
Suboptimal Structures within an Energy Band	Tree Representation of Secondary Structures, 495
around the MFE, 326	vrna_db_to_tree_string, 497
subopt.h	VRNA_STRUCTURE_TREE_EXPANDED, 496
SOLUTION, 809	VRNA_STRUCTURE_TREE_HIT, 495
subopt_circ	VRNA_STRUCTURE_TREE_SHAPIRO, 496
Suboptimal Structures within an Energy Band around the MFE, 327	VRNA_STRUCTURE_TREE_SHAPIRO_EXT, 496
Suboptimal Structures sensu Stiegler et al. 1984 / Zuker	VRNA_STRUCTURE_TREE_SHAPIRO_SHORT,
et al. 1989, 322	495
vrna_subopt_zuker, 322	VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT
zukersubopt, 323	496 vrna_tree_string_to_db, 498
zukersubopt_par, 323	vrna_tree_string_to_db, 498
Suboptimal Structures within an Energy Band around the	tree_edit_distance
MFE, 324	treedist.h, 810
subopt, 326	treedist.h
subopt_circ, 327	free_tree, 810
vrna_subopt, 325	make_tree, 810
a_0000pt; 0=0	

trae adit diatanae 010	uppode structure
tree_edit_distance, 810 TURN	unpack_structure
	Deprecated Interface for Secondary Structure Utili-
constants.h, 779 TwoDfold	ties, 687 Unstructured Domains, 224
Computing MFE representatives of a Distance	
Based Partitioning, 364	vrna_callback_ud_energy, 227
TwoDfold_backtrack_f5	vrna_callback_ud_exp_energy, 227
Computing MFE representatives of a Distance	vrna_callback_ud_exp_production, 228
Based Partitioning, 362	vrna_callback_ud_probs_add, 228
TwoDfold_vars, 357	vrna_callback_ud_probs_get, 229
Computing MFE representatives of a Distance	vrna_callback_ud_production, 228
Based Partitioning, 358	vrna_ud_add_motif, 231
TwoDfoldList	vrna_ud_motifs_centroid, 229
Computing MFE representatives of a Distance	vrna_ud_motifs_MEA, 230
Based Partitioning, 361	vrna_ud_motifs_MFE, 230
TwoDpfold_pbacktrack	vrna_ud_remove, 232
2Dpfold.h, 708	vrna_ud_set_data, 232
TwoDpfold_pbacktrack5	vrna_ud_set_exp_prod_rule_cb, 234
2Dpfold.h, 709	vrna_ud_set_prod_rule_cb, 233
TwoDpfold_vars, 700	unstructured_domains.h
TwoDpfoldList	vrna_ud_get_motif_size_at, 813
2Dpfold.h, 708	vrna_ud_set_prob_cb, 813
type	unweight
vrna_fc_s, 597	Deprecated Interface for Secondary Structure Utili-
vrna_path_s, 390	ties, 685
	update_alifold_params
unexpand_aligned_F	alifold.h, 712
Deprecated Interface for Secondary Structure Utili-	update_co_pf_params
ties, 685	Deprecated Interface for Global Partition Function
unexpand_Full	Computation, 666
Deprecated Interface for Secondary Structure Utili-	update_co_pf_params_par
Depresaisa interiass for essertially entastars etim	
ties, 685	Deprecated Interface for Global Partition Function
	Computation, 667
ties, 685	Computation, 667 update_cofold_params
ties, 685 Unit Conversion, 589	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction,
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction,
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction,
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_F, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_F, 590 VRNA_UNIT_DEG_N, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_F, 590 VRNA_UNIT_DEG_N, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction,
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_F, 590 VRNA_UNIT_DEG_N, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_F, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_P, 590 VRNA_UNIT_DEG_N, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_N, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_G_TNT, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_G_TNT, 590 VRNA_UNIT_J, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_F, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_G_TNT, 590 VRNA_UNIT_J, 590 VRNA_UNIT_K, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_N, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_G_TNT, 590 VRNA_UNIT_J, 590 VRNA_UNIT_K, 590 VRNA_UNIT_KCAL, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660 update_pf_paramsLP
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 VRNA_UNIT_DEG_RO, 590 VRNA_UNIT_DEG_RO, 590 VRNA_UNIT_EV, 590 VRNA_UNIT_G_TNT, 590 VRNA_UNIT_K, 590 VRNA_UNIT_KCAL_IT, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_Par Deprecated Interface for Global Partition Function Computation, 660 update_pf_paramsLP Deprecated Interface for Local (Sliding Window)
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_G_TNT, 590 VRNA_UNIT_K, 590 VRNA_UNIT_KCAL_IT, 590 VRNA_UNIT_KCAL_IT, 590 VRNA_UNIT_KG_TNT, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660 update_pf_paramsLP Deprecated Interface for Local (Sliding Window) Partition Function Computation, 673
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_G_TNT, 590 VRNA_UNIT_K, 590 VRNA_UNIT_KCAL_IT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660 update_pf_paramsLP Deprecated Interface for Local (Sliding Window) Partition Function Computation, 673 urn
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_EV, 590 VRNA_UNIT_S, 590 VRNA_UNIT_K, 590 VRNA_UNIT_KCAL_IT, 590 VRNA_UNIT_KCAL_IT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KJ, 590 VRNA_UNIT_KJ, 590 VRNA_UNIT_KJ, 590 VRNA_UNIT_KWH, 590 VRNA_UNIT_KWH, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660 update_pf_paramsLP Deprecated Interface for Local (Sliding Window) Partition Function Computation, 673 urn basic.h, 775
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_EV, 590 VRNA_UNIT_K, 590 VRNA_UNIT_K, 590 VRNA_UNIT_KCAL_IT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KJ, 590 VRNA_UNIT_KWH, 590 VRNA_UNIT_KWH, 590 VRNA_UNIT_T_TNT, 590 VRNA_UNIT_T_TNT, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660 update_pf_paramsLP Deprecated Interface for Local (Sliding Window) Partition Function Computation, 673 urn basic.h, 775 Utilities, 422
ties, 685 Unit Conversion, 589 vrna_convert_energy, 591 vrna_convert_temperature, 591 VRNA_UNIT_CAL, 590 VRNA_UNIT_CAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DACAL_IT, 590 VRNA_UNIT_DEG_C, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_DE, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_R, 590 VRNA_UNIT_DEG_RE, 590 VRNA_UNIT_DEG_RO, 590 vrna_unit_energy_e, 589 VRNA_UNIT_EV, 590 VRNA_UNIT_EV, 590 VRNA_UNIT_S, 590 VRNA_UNIT_K, 590 VRNA_UNIT_KCAL_IT, 590 VRNA_UNIT_KCAL_IT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KG_TNT, 590 VRNA_UNIT_KJ, 590 VRNA_UNIT_KJ, 590 VRNA_UNIT_KJ, 590 VRNA_UNIT_KWH, 590 VRNA_UNIT_KWH, 590	Computation, 667 update_cofold_params Deprecated Interface for Global MFE Prediction, 643 update_cofold_params_par Deprecated Interface for Global MFE Prediction, 643 update_fold_params Deprecated Interface for Global MFE Prediction, 648 update_fold_params_par Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global MFE Prediction, 649 update_pf_params Deprecated Interface for Global Partition Function Computation, 660 update_pf_params_par Deprecated Interface for Global Partition Function Computation, 660 update_pf_paramsLP Deprecated Interface for Local (Sliding Window) Partition Function Computation, 673 urn basic.h, 775

vrna_idx_col_wise, 428	ViennaRNA/file_formats_msa.h, 741
vrna_idx_row_wise, 427	ViennaRNA/file_utils.h, 743
VRNA_INPUT_CONSTRAINT, 424	ViennaRNA/findpath.h, 743
VRNA_INPUT_FASTA_HEADER, 424	ViennaRNA/fold.h, 744
vrna_int_urn, 426	ViennaRNA/fold_compound.h, 745
vrna realloc, 425	ViennaRNA/fold_vars.h, 746
vrna_time_stamp, 426	ViennaRNA/gquad.h, 748
vrna urn, 425	ViennaRNA/grammar.h, 748
xsubi, 428	ViennaRNA/hairpin loops.h, 749
Utilities to deal with Nucleotide Alphabets, 462	ViennaRNA/interior_loops.h, 749
vrna_nucleotide_decode, 465	ViennaRNA/inverse.h, 749
vrna_nucleotide_encode, 464	,
vrna_ptypes, 463	ViennaRNA/io/file_formats.h, 741
VRNA_SEQ_DNA, 463	ViennaRNA/io/file_formats_msa.h, 742
vrna_seq_encode, 464	ViennaRNA/io/utils.h, 814
vrna_seq_encode_simple, 464	ViennaRNA/landscape/findpath.h, 743
	ViennaRNA/landscape/move.h, 750
VRNA_SEQ_RNA, 463	ViennaRNA/landscape/neighbor.h, 766
vrna_seq_type_e, 463	ViennaRNA/landscape/paths.h, 751
VRNA_SEQ_UNKNOWN, 463	ViennaRNA/landscape/walk.h, 818
ViennaRNA/2Dfold.h, 705	ViennaRNA/Lfold.h, 752
ViennaRNA/2Dpfold.h, 706	ViennaRNA/loop_energies.h, 752
	ViennaRNA/loops/all.h, 753
ViennaRNA/alifold.h, 710	ViennaRNA/loops/external.h, 753
ViennaRNA/aln_util.h, 713	ViennaRNA/loops/hairpin.h, 754
ViennaRNA/alphabet.h, 713	ViennaRNA/loops/internal.h, 754
ViennaRNA/boltzmann_sampling.h, 713	ViennaRNA/loops/multibranch.h, 755
ViennaRNA/centroid.h, 715	ViennaRNA/LPfold.h, 756
ViennaRNA/char_stream.h, 716	ViennaRNA/MEA.h, 757
ViennaRNA/cofold.h, 717	ViennaRNA/mfe.h, 757
ViennaRNA/combinatorics.h, 717	ViennaRNA/mfe_window.h, 758
ViennaRNA/commands.h, 718	ViennaRNA/mm.h, 759
ViennaRNA/concentrations.h, 719	
ViennaRNA/constraints.h, 720	ViennaRNA/model.h, 760
ViennaRNA/constraints/basic.h, 770	ViennaRNA/multibranch_loops.h, 765
ViennaRNA/constraints/hard.h, 721	ViennaRNA/naview.h, 765
ViennaRNA/constraints/ligand.h, 726	ViennaRNA/neighbor.h, 766
ViennaRNA/constraints/SHAPE.h, 727	ViennaRNA/params.h, 767
ViennaRNA/constraints/soft.h, 728	ViennaRNA/params/1.8.4_epars.h, 767
ViennaRNA/constraints_hard.h, 730	ViennaRNA/params/1.8.4_intloops.h, 768
ViennaRNA/constraints ligand.h, 730	ViennaRNA/params/basic.h, 768
ViennaRNA/constraints_SHAPE.h, 730	ViennaRNA/params/constants.h, 778
ViennaRNA/constraints soft.h, 730	ViennaRNA/params/convert.h, 780
ViennaRNA/convert epars.h, 731	ViennaRNA/params/io.h, 781
ViennaRNA/data structures.h, 731	ViennaRNA/part_func.h, 782
ViennaRNA/datastructures/basic.h, 776	ViennaRNA/part_func_co.h, 785
ViennaRNA/datastructures/char stream.h, 716	ViennaRNA/part func up.h, 786
ViennaRNA/datastructures/hash_tables.h, 731	ViennaRNA/part func window.h, 787
ViennaRNA/datastructures/heap.h, 732	ViennaRNA/perturbation_fold.h, 788
ViennaRNA/datastructures/stream_output.h, 805	ViennaRNA/plot_aln.h, 789
ViennaRNA/dist vars.h, 733	ViennaRNA/plot_layouts.h, 789
ViennaRNA/dp_matrices.h, 734	ViennaRNA/plot_structure.h, 790
ViennaRNA/duplex.h, 735	ViennaRNA/plot_utils.h, 790
•	ViennaRNA/plotting/alignments.h, 790
ViennaRNA/edit_cost.h, 735	
ViennaRNA/energy_const.h, 736	ViennaRNA/plotting/layouts.h, 792
ViennaRNA/energy_par.h, 736	ViennaRNA/plotting/naview.h, 765
ViennaRNA/equilibrium_probs.h, 736	ViennaRNA/plotting/probabilities.h, 794
ViennaRNA/eval.h, 737	ViennaRNA/plotting/RNApuzzler/RNApuzzler.h, 794
ViennaRNA/exterior_loops.h, 740	ViennaRNA/plotting/RNApuzzler/RNAturtle.h, 795
ViennaRNA/file_formats.h, 740	ViennaRNA/plotting/structures.h, 795

ViennaRNA/plotting/utils.h, 815	Computing MFE representatives of a Distance
ViennaRNA/profiledist.h, 799	Based Partitioning, 360
ViennaRNA/PS_dot.h, 801	vrna_basepair_s, 578
ViennaRNA/read_epars.h, 801	vrna_boltzmann_sampling_callback
ViennaRNA/ribo.h, 802	Random Structure Samples from the Ensemble,
ViennaRNA/RNAstruct.h, 802	331
ViennaRNA/search/BoyerMoore.h, 803	vrna_bp_distance
ViennaRNA/sequence.h, 804	Secondary Structure Utilities, 475
ViennaRNA/stream_output.h, 804	vrna_bp_stack_s, 578
ViennaRNA/string_utils.h, 805	VRNA_BRACKETS_ALPHA
ViennaRNA/stringdist.h, 806	Dot-Bracket Notation of Secondary Structures, 479
ViennaRNA/structure_utils.h, 807	VRNA_BRACKETS_ANG
ViennaRNA/structured_domains.h, 807	Dot-Bracket Notation of Secondary Structures, 480
ViennaRNA/subopt.h, 807	VRNA_BRACKETS_ANY
ViennaRNA/svm_utils.h, 809	Dot-Bracket Notation of Secondary Structures, 481
ViennaRNA/treedist.h, 809	VRNA_BRACKETS_CLY
ViennaRNA/units.h, 811	Dot-Bracket Notation of Secondary Structures, 480
ViennaRNA/unstructured_domains.h, 811	VRNA_BRACKETS_DEFAULT
ViennaRNA/utils.h, 814	Dot-Bracket Notation of Secondary Structures, 481
ViennaRNA/utils/alignments.h, 791	VRNA_BRACKETS_RND
ViennaRNA/utils/basic.h, 771	Dot-Bracket Notation of Secondary Structures, 480
ViennaRNA/utils/strings.h, 815	VRNA_BRACKETS_SQR
ViennaRNA/utils/structures.h, 796	Dot-Bracket Notation of Secondary Structures, 480
ViennaRNA/walk.h, 818	vrna_BT_hp_loop
vrna_alifold	Backtracking MFE structures, 295
Global MFE Prediction, 285	vrna_BT_int_loop
vrna_alignment_s, 463	Backtracking MFE structures, 295
vrna_alloc	vrna_BT_mb_loop
Utilities, 425	Backtracking MFE structures, 296
vrna_aln_consensus_mis	vrna_BT_stack
Multiple Sequence Alignment Utilities, 510	Backtracking MFE structures, 295
vrna_aln_consensus_sequence	vrna_C11_features
Multiple Sequence Alignment Utilities, 510	(Abstract) Data Structures, 581
vrna_aln_conservation_col	vrna_callback_free_auxdata
Multiple Sequence Alignment Utilities, 509	The Fold Compound, 604
vrna_aln_conservation_struct	vrna_callback_hc_evaluate
	Hard Constraints, 261
Multiple Sequence Alignment Utilities, 508 vrna aln copy	
— — · ·	vrna_callback_heap_cmp Heaps, 627
Multiple Sequence Alignment Utilities, 508	·
vrna_aln_free	vrna_callback_heap_get_pos
Multiple Sequence Alignment Utilities, 505	Heaps, 627
vrna_aln_mpi	vrna_callback_heap_set_pos
Multiple Sequence Alignment Utilities, 503	Heaps, 628
vrna_aln_pinfo	vrna_callback_ht_compare_entries
Multiple Sequence Alignment Utilities, 503	Hash Tables, 617
vrna_aln_slice	vrna_callback_ht_free_entry
Multiple Sequence Alignment Utilities, 505	Hash Tables, 618
vrna_aln_toRNA	vrna_callback_ht_hash_function
Multiple Sequence Alignment Utilities, 507	Hash Tables, 617
vrna_aln_uppercase	vrna_callback_move_update
Multiple Sequence Alignment Utilities, 507	Neighborhood Relation and Move Sets for Sec-
vrna_annotate_covar_db	ondary Structures, 380
Annotation, 563	vrna_callback_recursion_status
vrna_annotate_covar_pairs	The Fold Compound, 605
Annotation, 563	vrna_callback_sc_backtrack
vrna_backtrack5	Soft Constraints, 271
Backtracking MFE structures, 294	vrna_callback_sc_energy
vrna_backtrack5_TwoD	Soft Constraints, 268

vrna_callback_sc_exp_energy	Hard Constraints, 259
Soft Constraints, 270	VRNA_CONSTRAINT_DB_PIPE
vrna_callback_stream_output	Hard Constraints, 258
Buffers, 634	VRNA_CONSTRAINT_DB_RND_BRACK
vrna_callback_ud_energy	Hard Constraints, 259
Unstructured Domains, 227	VRNA_CONSTRAINT_DB_WUSS
vrna_callback_ud_exp_energy	Hard Constraints, 260
Unstructured Domains, 227	VRNA CONSTRAINT DB X
vrna_callback_ud_exp_production	Hard Constraints, 258
Unstructured Domains, 228	VRNA CONSTRAINT FILE
vrna_callback_ud_probs_add	Constraining the RNA Folding Grammar, 240
Unstructured Domains, 228	VRNA_CONSTRAINT_MULTILINE
vrna_callback_ud_probs_get	Nucleic Acid Sequences and Structures, 516
Unstructured Domains, 229	VRNA CONSTRAINT NO HEADER
vrna_callback_ud_production	hard.h, 723
Unstructured Domains, 228	VRNA_CONSTRAINT_SOFT_MFE
vrna_centroid	Constraining the RNA Folding Grammar, 240
Compute the Centroid Structure, 350	VRNA CONSTRAINT SOFT PF
vrna_centroid_from_plist	Constraining the RNA Folding Grammar, 240
Compute the Centroid Structure, 350	vrna_constraints_add
vrna_centroid_from_probs	Constraining the RNA Folding Grammar, 250
Compute the Centroid Structure, 351	vrna convert energy
vrna_circalifold	Unit Conversion, 591
Global MFE Prediction, 286	VRNA_CONVERT_OUTPUT_ALL
vrna_circfold	Converting Energy Parameter Files, 457
Global MFE Prediction, 285	VRNA_CONVERT_OUTPUT_BULGE
VRNA_CMD_PARSE_DEFAULTS	Converting Energy Parameter Files, 459
Command Files, 534	VRNA_CONVERT_OUTPUT_DANGLE3
VRNA_CMD_PARSE_HC	Converting Energy Parameter Files, 458
Command Files, 533	VRNA_CONVERT_OUTPUT_DANGLE5
VRNA_CMD_PARSE_SC	Converting Energy Parameter Files, 458
Command Files, 533	VRNA_CONVERT_OUTPUT_DUMP
VRNA CMD PARSE SD	Converting Energy Parameter Files, 460
Command Files, 533	VRNA_CONVERT_OUTPUT_HP
VRNA_CMD_PARSE_UD	Converting Energy Parameter Files, 457
Command Files, 533	VRNA CONVERT OUTPUT INT
vrna_cofold	Converting Energy Parameter Files, 459
Global MFE Prediction, 287	VRNA_CONVERT_OUTPUT_INT_11
vrna_color_s, 578	Converting Energy Parameter Files, 458
vrna commands apply	VRNA_CONVERT_OUTPUT_INT_21
Command Files, 535	Converting Energy Parameter Files, 459
vrna_commands_free	VRNA CONVERT OUTPUT INT 22
Command Files, 536	Converting Energy Parameter Files, 459
VRNA CONSTRAINT DB	VRNA CONVERT OUTPUT MISC
Hard Constraints, 257	Converting Energy Parameter Files, 459
VRNA_CONSTRAINT_DB_ANG_BRACK	VRNA_CONVERT_OUTPUT_ML
hard.h, 723	Converting Energy Parameter Files, 459
VRNA_CONSTRAINT_DB_DEFAULT	VRNA_CONVERT_OUTPUT_MM_EXT
Hard Constraints, 260	Converting Energy Parameter Files, 458
VRNA CONSTRAINT DB DOT	VRNA CONVERT OUTPUT MM HP
Hard Constraints, 258	Converting Energy Parameter Files, 457
VRNA_CONSTRAINT_DB_ENFORCE_BP	VRNA CONVERT OUTPUT MM INT
Hard Constraints, 257	Converting Energy Parameter Files, 457 VRNA CONVERT OUTPUT MM INT 1N
VRNA_CONSTRAINT_DB_GQUAD	
Hard Constraints, 260	Converting Energy Parameter Files, 457
VRNA_CONSTRAINT_DB_INTERMOL	VRNA_CONVERT_OUTPUT_MM_INT_23
Hard Constraints, 259	Converting Energy Parameter Files, 458
VRNA CONSTRAINT DB INTRAMOL	VRNA CONVERT OUTPUT MM MULTI

0 " 5 D . 5" 450	O
Converting Energy Parameter Files, 458	Constraining the RNA Folding Grammar, 248
VRNA_CONVERT_OUTPUT_NINIO	VRNA_DECOMP_EXT_UP
Converting Energy Parameter Files, 460	Constraining the RNA Folding Grammar, 247
VRNA_CONVERT_OUTPUT_SPECIAL_HP	VRNA_DECOMP_ML_COAXIAL
Converting Energy Parameter Files, 460	Constraining the RNA Folding Grammar, 245
VRNA_CONVERT_OUTPUT_STACK	VRNA_DECOMP_ML_COAXIAL_ENC
Converting Energy Parameter Files, 457	Constraining the RNA Folding Grammar, 246
VRNA_CONVERT_OUTPUT_VANILLA	VRNA_DECOMP_ML_ML
Converting Energy Parameter Files, 460	Constraining the RNA Folding Grammar, 244
vrna_convert_temperature	VRNA_DECOMP_ML_ML_ML
Unit Conversion, 591	Constraining the RNA Folding Grammar, 243
vrna_cpair_s, 578	VRNA_DECOMP_ML_ML_STEM
vrna_cstr	Constraining the RNA Folding Grammar, 245
Buffers, 635	VRNA_DECOMP_ML_STEM
vrna_cstr_close	Constraining the RNA Folding Grammar, 243
Buffers, 636	VRNA_DECOMP_ML_UP
vrna_cstr_fflush	Constraining the RNA Folding Grammar, 244
Buffers, 636	VRNA_DECOMP_PAIR_HP
vrna cstr free	Constraining the RNA Folding Grammar, 241
Buffers, 635	VRNA DECOMP PAIR IL
vrna_cut_point_insert	Constraining the RNA Folding Grammar, 241
(Nucleic Acid Sequence) String Utilitites, 473	VRNA_DECOMP_PAIR_ML
vrna_cut_point_remove	Constraining the RNA Folding Grammar, 242
(Nucleic Acid Sequence) String Utilitites, 473	vrna_dimer_conc_s, 701
vrna_data_linear_s, 578	vrna_dimer_pf_s, 298
vrna_db_flatten	vrna_dotplot_auxdata_t, 538
Dot-Bracket Notation of Secondary Structures, 483 vrna_db_flatten_to	vrna_E_ext_hp_loop Hairpin Loops, 435
Dot-Bracket Notation of Secondary Structures, 483	vrna_E_ext_loop
vrna_db_from_bp_stack	Exterior Loops, 431
Secondary Structure Utilities, 477	·
·	vrna_E_ext_stem
vrna_db_from_plist Dot-Bracket Notation of Secondary Structures, 485	Exterior Loops, 431
•	vrna_E_hp_loop
vrna_db_from_ptable	Hairpin Loops, 434
Dot-Bracket Notation of Secondary Structures, 484	vrna_E_mb_loop_stack
vrna_db_from_WUSS	Multibranch Loops, 441
Dot-Bracket Notation of Secondary Structures, 484	
vrna_db_pack	vrna_ensemble_defect
Dot-Bracket Notation of Secondary Structures, 482	Global Partition Function and Equilibrium Probabil-
vrna_db_pk_remove	ities, 300
Dot-Bracket Notation of Secondary Structures, 486	vrna_enumerate_necklaces
vrna_db_to_element_string	Combinatorics Algorithms, 570
Dot-Bracket Notation of Secondary Structures, 485	vrna_eval_circ_consensus_structure
vrna_db_to_tree_string	Free Energy Evaluation, 142
Tree Representation of Secondary Structures, 497	vrna_eval_circ_consensus_structure_v
vrna_db_unpack	Free Energy Evaluation, 146
Dot-Bracket Notation of Secondary Structures, 482	vrna_eval_circ_gquad_consensus_structure
VRNA_DECOMP_EXT_EXT	Free Energy Evaluation, 143
Constraining the RNA Folding Grammar, 246	vrna_eval_circ_gquad_consensus_structure_v
VRNA_DECOMP_EXT_EXT_EXT	Free Energy Evaluation, 147
Constraining the RNA Folding Grammar, 248	vrna_eval_circ_gquad_structure
VRNA_DECOMP_EXT_EXT_STEM	Free Energy Evaluation, 137
Constraining the RNA Folding Grammar, 249	vrna_eval_circ_gquad_structure_v
VRNA_DECOMP_EXT_EXT_STEM1	Free Energy Evaluation, 140
Constraining the RNA Folding Grammar, 249	vrna_eval_circ_structure
VRNA_DECOMP_EXT_STEM	Free Energy Evaluation, 135
Constraining the RNA Folding Grammar, 247	vrna_eval_circ_structure_v
VRNA_DECOMP_EXT_STEM_EXT	Free Energy Evaluation, 139

vrna_eval_consensus_structure_pt_simple Free Energy Evaluation, 150	vrna_exp_param_s, 211 alpha, 211
vrna_eval_consensus_structure_simple	id, 211
Free Energy Evaluation, 141	vrna_exp_params
vrna_eval_consensus_structure_simple_v	Energy Parameters, 213
Free Energy Evaluation, 145	vrna_exp_params_comparative
vrna_eval_consensus_structure_simple_verbose	Energy Parameters, 214
Free Energy Evaluation, 144	vrna_exp_params_copy
vrna_eval_covar_structure	Energy Parameters, 214
Free Energy Evaluation, 131	vrna_exp_params_rescale
vrna eval gquad consensus structure	Energy Parameters, 216
Free Energy Evaluation, 142	vrna_exp_params_reset
vrna_eval_gquad_consensus_structure_v	Energy Parameters, 217
Free Energy Evaluation, 146	vrna_exp_params_subst
vrna_eval_gquad_structure	Energy Parameters, 215
Free Energy Evaluation, 136	vrna_extract_record_rest_constraint
vrna_eval_gquad_structure_v	Nucleic Acid Sequences and Structures, 521
Free Energy Evaluation, 139	vrna_extract_record_rest_structure
vrna_eval_hp_loop	Nucleic Acid Sequences and Structures, 520
Hairpin Loops, 435	vrna_fc_s, 594
vrna_eval_int_loop	auxdata, 597
Internal Loops, 439	cons_seq, 600
•	free auxdata, 597
vrna_eval_loop_pt	— · · · · · · · · · · · · · · · · · · ·
Energy Evaluation for Individual Loops, 153	n_seq, 599
vrna_eval_loop_pt_v	pscore, 601
Energy Evaluation for Individual Loops, 153	pscore_local, 601
vrna_eval_move	pscore_pf_compat, 601
Energy Evaluation for Atomic Moves, 155	ptype, 598
vrna_eval_move_pt	ptype_pf_compat, 598
Energy Evaluation for Atomic Moves, 156	S, 600
vrna_eval_structure	S3, 601
Free Energy Evaluation, 130	S5, 600
vrna_eval_structure_pt	S_cons, 600
Free Energy Evaluation, 133	sc, 599
vrna_eval_structure_pt_simple	scs, 602
Free Energy Evaluation, 148	sequence, 597
vrna_eval_structure_pt_simple_v	sequence_encoding, 598
Free Energy Evaluation, 149	sequences, 599
vrna_eval_structure_pt_simple_verbose	stat_cb, 597
Free Energy Evaluation, 149	type, 597
vrna_eval_structure_pt_v	VRNA_FC_TYPE_COMPARATIVE
Free Energy Evaluation, 134	The Fold Compound, 605
vrna_eval_structure_pt_verbose	vrna_fc_type_e
Free Energy Evaluation, 133	The Fold Compound, 605
vrna_eval_structure_simple	VRNA_FC_TYPE_SINGLE
Free Energy Evaluation, 135	The Fold Compound, 605
vrna_eval_structure_simple_v	vrna_file_bpseq
Free Energy Evaluation, 138	Nucleic Acid Sequences and Structures, 517
vrna_eval_structure_simple_verbose	vrna_file_commands_apply
Free Energy Evaluation, 137	Command Files, 535
vrna_eval_structure_v	vrna_file_commands_read
Free Energy Evaluation, 132	Command Files, 534
vrna_eval_structure_verbose	vrna_file_connect
Free Energy Evaluation, 131	Nucleic Acid Sequences and Structures, 517
vrna_exp_E_ext_stem	vrna_file_exists
Exterior Loops, 432	Files and I/O, 513
vrna_exp_E_hp_loop	vrna_file_fasta_read_record
Hairpin Loops, 438	Nucleic Acid Sequences and Structures, 518

VRNA_FILE_FORMAT_MSA_APPEND	vrna_gr_aux_s, 172
Multiple Sequence Alignments, 526	vrna_hamming_distance
VRNA_FILE_FORMAT_MSA_CLUSTAL	(Nucleic Acid Sequence) String Utilitites, 471
Multiple Sequence Alignments, 524	vrna_hamming_distance_bound
VRNA_FILE_FORMAT_MSA_DEFAULT	(Nucleic Acid Sequence) String Utilitites, 471
Multiple Sequence Alignments, 525	vrna_hash_table_t
VRNA_FILE_FORMAT_MSA_FASTA	Hash Tables, 616
Multiple Sequence Alignments, 524	vrna_hc_add_bp
VRNA_FILE_FORMAT_MSA_MAF	Hard Constraints, 263
Multiple Sequence Alignments, 524	vrna_hc_add_bp_nonspecific
VRNA FILE FORMAT MSA MIS	Hard Constraints, 264
Multiple Sequence Alignments, 525	vrna_hc_add_data
VRNA_FILE_FORMAT_MSA_NOCHECK	hard.h, 724
Multiple Sequence Alignments, 525	vrna_hc_add_from_db
VRNA_FILE_FORMAT_MSA_QUIET	Hard Constraints, 265
Multiple Sequence Alignments, 526	vrna_hc_add_up
VRNA_FILE_FORMAT_MSA_SILENT	Hard Constraints, 262
Multiple Sequence Alignments, 526	vrna_hc_add_up_batch
VRNA FILE FORMAT MSA STOCKHOLM	Hard Constraints, 263
Multiple Sequence Alignments, 524	vrna_hc_bp_storage_t, 702
VRNA_FILE_FORMAT_MSA_UNKNOWN	VRNA_HC_DEFAULT
Multiple Sequence Alignments, 526	hard.h, 724
vrna_file_helixlist	
Nucleic Acid Sequences and Structures, 516	vrna_hc_free Hard Constraints, 264
·	
vrna_file_json Nucleic Acid Sequences and Structures, 518	vrna_hc_init Hard Constraints, 262
vrna_file_msa_detect_format	vrna_hc_s, 256
Multiple Sequence Alignments, 529	free_data, 257
vrna_file_msa_read	vrna_hc_type_e
Multiple Sequence Alignments, 527	hard.h, 724
vrna_file_msa_read_record	vrna_hc_up_s, 257
Multiple Sequence Alignments, 528	VRNA_HC_WINDOW
vrna_file_msa_write	hard.h, 724
Multiple Sequence Alignments, 530	vrna_heap_free
vrna_file_PS_aln Alignment Plots, 564	Heaps, 629 vrna_heap_init
vrna_file_PS_aln_slice	Heaps, 628
Alignment Plots, 565 vrna_file_PS_rnaplot	vrna_heap_insert
Plotting, 539	Heaps, 630 vrna_heap_pop
	Heaps, 630
vrna_file_PS_rnaplot_a Plotting, 540	
vrna_file_SHAPE_read	vrna_heap_remove
	Heaps, 631
Nucleic Acid Sequences and Structures, 520	vrna_heap_size
vrna_filename_sanitize	Heaps, 630
Files and I/O, 512	vrna_heap_t
vrna_fold	Heaps, 627
Global MFE Prediction, 284	vrna_heap_top
vrna_fold_compound	Heaps, 631
The Fold Compound, 606	vrna_heap_update
vrna_fold_compound_add_auxdata	Heaps, 632
The Fold Compound, 608	vrna_ht_clear
vrna_fold_compound_add_callback	Hash Tables, 622
The Fold Compound, 609	vrna_ht_collisions
vrna_fold_compound_comparative	Hash Tables, 619
The Fold Compound, 607	vrna_ht_db_comp
vrna_fold_compound_free The Fold Compound, 608	Hash Tables, 623 vrna ht db free entry
THE FUIL CUITIOUITIA. DVO	vina ili ud liee eilliv

Hash Tables 624	Fine-tuning of the Implemented Models, 194
Hash Tables, 624	· ·
vrna_ht_db_hash_func	vrna_md_defaults_circ_get
Hash Tables, 624	Fine-tuning of the Implemented Models, 194
vrna_ht_entry_db_t, 616	vrna_md_defaults_compute_bpp
energy, 616	Fine-tuning of the Implemented Models, 198
structure, 616	vrna_md_defaults_compute_bpp_get
vrna_ht_free	Fine-tuning of the Implemented Models, 199
Hash Tables, 623	vrna_md_defaults_cv_fact
vrna_ht_get	Fine-tuning of the Implemented Models, 203
Hash Tables, 621	vrna_md_defaults_cv_fact_get
vrna_ht_init	Fine-tuning of the Implemented Models, 203
Hash Tables, 618	vrna md defaults dangles
vrna_ht_insert	Fine-tuning of the Implemented Models, 189
Hash Tables, 621	vrna_md_defaults_dangles_get
vrna_ht_remove	Fine-tuning of the Implemented Models, 190
Hash Tables, 622	vrna_md_defaults_energy_set
vrna_ht_size	Fine-tuning of the Implemented Models, 196
Hash Tables, 619	vrna_md_defaults_energy_set_get
vrna_hx_from_ptable	Fine-tuning of the Implemented Models, 197
Helix List Representation of Secondary Structures,	vrna_md_defaults_gquad
493	Fine-tuning of the Implemented Models, 195
vrna_hx_s, 493	vrna_md_defaults_gquad_get
vrna_idx_col_wise	Fine-tuning of the Implemented Models, 195
Utilities, 428	vrna_md_defaults_logML
vrna_idx_row_wise	Fine-tuning of the Implemented Models, 193
Utilities, 427	vrna_md_defaults_logML_get
VRNA_INPUT_CONSTRAINT	Fine-tuning of the Implemented Models, 194
Utilities, 424	vrna_md_defaults_max_bp_span
VRNA_INPUT_FASTA_HEADER	Fine-tuning of the Implemented Models, 199
Utilities, 424	vrna_md_defaults_max_bp_span_get
vrna_int_urn	Fine-tuning of the Implemented Models, 200
Utilities, 426	vrna_md_defaults_min_loop_size
vrna_Lfold	Fine-tuning of the Implemented Models, 200
Local (sliding window) MFE Prediction, 292	vrna_md_defaults_min_loop_size_get
vrna_Lfoldz	Fine-tuning of the Implemented Models, 200
Local (sliding window) MFE Prediction, 293	vrna_md_defaults_nc_fact
vrna_loopidx_update	Fine-tuning of the Implemented Models, 204
Neighborhood Relation and Move Sets for Sec-	vrna_md_defaults_nc_fact_get
ondary Structures, 384	Fine-tuning of the Implemented Models, 204
vrna_maximum_matching	vrna_md_defaults_noGU
mm.h, 760	Fine-tuning of the Implemented Models, 192
vrna_maximum_matching_simple	vrna_md_defaults_noGU_get
mm.h, 760	Fine-tuning of the Implemented Models, 192
vrna md copy	vrna_md_defaults_noGUclosure
,	
Fine-tuning of the Implemented Models, 186	Fine-tuning of the Implemented Models, 192
vrna_md_defaults_backtrack	vrna_md_defaults_noGUclosure_get
Fine-tuning of the Implemented Models, 197	Fine-tuning of the Implemented Models, 193
vrna_md_defaults_backtrack_get	vrna_md_defaults_noLP
Fine-tuning of the Implemented Models, 197	Fine-tuning of the Implemented Models, 191
vrna_md_defaults_backtrack_type	vrna_md_defaults_noLP_get
Fine-tuning of the Implemented Models, 198	Fine-tuning of the Implemented Models, 191
vrna_md_defaults_backtrack_type_get	vrna_md_defaults_oldAliEn
Fine-tuning of the Implemented Models, 198	Fine-tuning of the Implemented Models, 201
vrna_md_defaults_betaScale	vrna_md_defaults_oldAliEn_get
Fine-tuning of the Implemented Models, 189	Fine-tuning of the Implemented Models, 202
vrna_md_defaults_betaScale_get	vrna_md_defaults_reset
Fine-tuning of the Implemented Models, 189	Fine-tuning of the Implemented Models, 187
vrna md defaults circ	vrna md defaults ribo

Fine-tuning of the Implemented Models, 202	vrna_message_verror
vrna_md_defaults_ribo_get	Messages, 584
Fine-tuning of the Implemented Models, 203	vrna_message_vinfo
vrna_md_defaults_sfact	Messages, 586
Fine-tuning of the Implemented Models, 204	vrna_message_vwarning
vrna_md_defaults_sfact_get	Messages, 585
Fine-tuning of the Implemented Models, 205	vrna_message_warning
vrna_md_defaults_special_hp	Messages, 584
Fine-tuning of the Implemented Models, 190	vrna_mfe
vrna_md_defaults_special_hp_get	Global MFE Prediction, 283
Fine-tuning of the Implemented Models, 191	vrna_mfe_dimer
vrna_md_defaults_temperature	Global MFE Prediction, 283
Fine-tuning of the Implemented Models, 188	vrna_mfe_TwoD
vrna_md_defaults_temperature_get	Computing MFE representatives of a Distance
Fine-tuning of the Implemented Models, 188	Based Partitioning, 359
vrna_md_defaults_uniq_ML	vrna_mfe_window
Fine-tuning of the Implemented Models, 195	Local (sliding window) MFE Prediction, 291
vrna_md_defaults_uniq_ML_get	vrna_mfe_window_callback
Fine-tuning of the Implemented Models, 196 vrna md defaults window size	Local (sliding window) MFE Prediction, 290 vrna mfe window zscore
Fine-tuning of the Implemented Models, 201	Local (sliding window) MFE Prediction, 291
vrna_md_defaults_window_size_get	VRNA_MINIMIZER_CONJUGATE_FR
Fine-tuning of the Implemented Models, 201	Generate Soft Constraints from Data, 411
vrna_md_option_string	VRNA_MINIMIZER_CONJUGATE_PR
Fine-tuning of the Implemented Models, 187	Generate Soft Constraints from Data, 411
vrna_md_s, 177	VRNA_MINIMIZER_STEEPEST_DESCENT
dangles, 179	Generate Soft Constraints from Data, 412
min_loop_size, 180	VRNA_MINIMIZER_VECTOR_BFGS
vrna_md_set_default	Generate Soft Constraints from Data, 411
Fine-tuning of the Implemented Models, 186	VRNA_MINIMIZER_VECTOR_BFGS2
vrna_md_update	Generate Soft Constraints from Data, 412
Fine-tuning of the Implemented Models, 186	VRNA_MODEL_DEFAULT_ALI_CV_FACT
vrna_MEA	Fine-tuning of the Implemented Models, 185
Compute the Structure with Maximum Expected	VRNA_MODEL_DEFAULT_ALI_NC_FACT
Accuracy (MEA), 347	Fine-tuning of the Implemented Models, 185
vrna_MEA_from_plist	VRNA_MODEL_DEFAULT_ALI_OLD_EN
Compute the Structure with Maximum Expected	Fine-tuning of the Implemented Models, 185
Accuracy (MEA), 348	VRNA_MODEL_DEFAULT_ALI_RIBO
vrna_mean_bp_distance	Fine-tuning of the Implemented Models, 185
Global Partition Function and Equilibrium Probabil-	VRNA_MODEL_DEFAULT_BACKTRACK
ities, 299	Fine-tuning of the Implemented Models, 183
vrna_mean_bp_distance_pr	VRNA_MODEL_DEFAULT_BACKTRACK_TYPE
Global Partition Function and Equilibrium Probabil-	Fine-tuning of the Implemented Models, 183
ities, 299	VRNA_MODEL_DEFAULT_BETA_SCALE
VRNA_MEASURE_SHANNON_ENTROPY	Fine-tuning of the Implemented Models, 181
Multiple Sequence Alignment Utilities, 503	VRNA_MODEL_DEFAULT_CIRC
vrna_message_constraint_options	Fine-tuning of the Implemented Models, 182
Constraining the RNA Folding Grammar, 251	VRNA_MODEL_DEFAULT_COMPUTE_BPP
vrna_message_constraint_options_all	Fine-tuning of the Implemented Models, 184
Constraining the RNA Folding Grammar, 253	VRNA_MODEL_DEFAULT_DANGLES
vrna_message_error	Fine-tuning of the Implemented Models, 181
Messages, 583	VRNA_MODEL_DEFAULT_ENERGY_SET
vrna_message_info	Fine-tuning of the Implemented Models, 183
Messages, 585	VRNA_MODEL_DEFAULT_GQUAD
vrna_message_input_seq	Fine-tuning of the Implemented Models, 182
Messages, 586	VRNA_MODEL_DEFAULT_LOG_ML
vrna_message_input_seq_simple	Fine-tuning of the Implemented Models, 184
Messages, 586	VRNA_MODEL_DEFAULT_MAX_BP_SPAN

Fine-tuning of the Implemented Models, 184	Neighborhood Relation and Move Sets for Sec-
VRNA_MODEL_DEFAULT_NO_GU	ondary Structures, 379
Fine-tuning of the Implemented Models, 182	VRNA_MX_2DFOLD
VRNA_MODEL_DEFAULT_NO_GU_CLOSURE	The Dynamic Programming Matrices, 612
Fine-tuning of the Implemented Models, 182	vrna_mx_add
VRNA_MODEL_DEFAULT_NO_LP	The Dynamic Programming Matrices, 612
Fine-tuning of the Implemented Models, 181	VRNA_MX_DEFAULT
VRNA_MODEL_DEFAULT_PF_SCALE	The Dynamic Programming Matrices, 612
Fine-tuning of the Implemented Models, 180	vrna_mx_mfe_free
VRNA_MODEL_DEFAULT_SPECIAL_HP	The Dynamic Programming Matrices, 613
Fine-tuning of the Implemented Models, 181	vrna_mx_mfe_s, 610
VRNA_MODEL_DEFAULT_TEMPERATURE	vrna_mx_pf_aux_el_t
Fine-tuning of the Implemented Models, 180	Exterior Loops, 430
VRNA_MODEL_DEFAULT_UNIQ_ML	vrna_mx_pf_aux_ml_t
Fine-tuning of the Implemented Models, 183	Multibranch Loops, 440
VRNA_MODEL_DEFAULT_WINDOW_SIZE	vrna_mx_pf_free
Fine-tuning of the Implemented Models, 184	The Dynamic Programming Matrices, 613
vrna_move_apply	vrna_mx_pf_s, 611
Neighborhood Relation and Move Sets for Sec-	vrna_mx_type_e The Dynamic Programming Matrices, 612
ondary Structures, 381	The Dynamic Programming Matrices, 612 VRNA_MX_WINDOW
vrna_move_compare Neighborhood Relation and Move Sets for Sec-	The Dynamic Programming Matrices, 612
ondary Structures, 383	VRNA_NEIGHBOR_CHANGE
vrna_move_init	Neighborhood Relation and Move Sets for Sec-
Neighborhood Relation and Move Sets for Sec-	ondary Structures, 379
ondary Structures, 381	VRNA_NEIGHBOR_INVALID
vrna_move_is_insertion	Neighborhood Relation and Move Sets for Sec-
Neighborhood Relation and Move Sets for Sec-	ondary Structures, 380
ondary Structures, 382	VRNA_NEIGHBOR_NEW
vrna_move_is_removal	Neighborhood Relation and Move Sets for Sec-
Neighborhood Relation and Move Sets for Sec-	ondary Structures, 380
ondary Structures, 382	vrna_neighbors
vrna_move_is_shift	Neighborhood Relation and Move Sets for Sec-
Neighborhood Relation and Move Sets for Sec-	ondary Structures, 384
ondary Structures, 383	vrna_neighbors_successive
vrna_move_list_free	Neighborhood Relation and Move Sets for Sec-
Neighborhood Relation and Move Sets for Sec-	ondary Structures, 385
ondary Structures, 381	vrna_nucleotide_decode
vrna_move_neighbor_diff	Utilities to deal with Nucleotide Alphabets, 465
Neighborhood Relation and Move Sets for Sec-	vrna_nucleotide_encode
ondary Structures, 386	Utilities to deal with Nucleotide Alphabets, 464
vrna_move_neighbor_diff_cb	VRNA_OBJECTIVE_FUNCTION_ABSOLUTE
Neighborhood Relation and Move Sets for Sec-	Generate Soft Constraints from Data, 411
ondary Structures, 386	VRNA_OBJECTIVE_FUNCTION_QUADRATIC
vrna_move_s, 377	Generate Soft Constraints from Data, 411
VRNA_MOVESET_DEFAULT	VRNA_OPTION_EVAL_ONLY
Neighborhood Relation and Move Sets for Sec-	The Fold Compound, 604
ondary Structures, 379	VRNA_OPTION_MFE
VRNA_MOVESET_DELETION	The Fold Compound, 603
Neighborhood Relation and Move Sets for Sec-	VRNA_OPTION_MULTILINE
ondary Structures, 378	Nucleic Acid Sequences and Structures, 515
VRNA_MOVESET_INSERTION	VRNA_OPTION_PF
Neighborhood Relation and Move Sets for Sec-	The Fold Compound, 603
ondary Structures, 378	vrna_ostream_free
VRNA_MOVESET_NO_LP	Buffers, 637
Neighborhood Relation and Move Sets for Sec-	vrna_ostream_init
ondary Structures, 379	Buffers, 637
VRNA_MOVESET_SHIFT	vrna_ostream_provide

Buffers, 638	Direct Refolding Paths between two Secondary
vrna_ostream_request Buffers, 638	Structures, 398 vrna path findpath
vrna_param_s, 210	Direct Refolding Paths between two Secondary
VRNA_PARAMETER_FORMAT_DEFAULT	Structures, 395
Reading/Writing Energy Parameter Sets from/to File, 449	vrna_path_findpath_saddle Direct Refolding Paths between two Secondary
vrna_params	Structures, 393
Energy Parameters, 212	vrna_path_findpath_saddle_ub
vrna_params_copy Energy Parameters, 213	Direct Refolding Paths between two Secondary Structures, 394
vrna params load	vrna_path_findpath_ub
Reading/Writing Energy Parameter Sets from/to File, 449	Direct Refolding Paths between two Secondary Structures, 395
vrna_params_load_defaults	vrna_path_free
Reading/Writing Energy Parameter Sets from/to File, 451	(Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima, 391
vrna_params_load_DNA_Mathews1999	vrna_path_gradient
Reading/Writing Energy Parameter Sets from/to File, 454	Folding Paths that start at a single Secondary Structure, 402
vrna_params_load_DNA_Mathews2004	VRNA_PATH_NO_TRANSITION_OUTPUT
Reading/Writing Energy Parameter Sets from/to File, 453	Folding Paths that start at a single Secondary Structure, 401
vrna_params_load_from_string	vrna_path_options_findpath
Reading/Writing Energy Parameter Sets from/to File, 450	Direct Refolding Paths between two Secondary Structures, 396
vrna_params_load_RNA_Andronescu2007	vrna_path_options_free
Reading/Writing Energy Parameter Sets from/to File, 452	(Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima, 391
vrna_params_load_RNA_Langdon2018	VRNA_PATH_RANDOM
Reading/Writing Energy Parameter Sets from/to File, 452	Folding Paths that start at a single Secondary Structure, 400
vrna_params_load_RNA_misc_special_hairpins	vrna_path_random
Reading/Writing Energy Parameter Sets from/to File, 453	Folding Paths that start at a single Secondary Structure, 403
vrna_params_load_RNA_Turner1999	vrna_path_s, 389
Reading/Writing Energy Parameter Sets from/to	type, 390
File, 451 vrna params load RNA Turner2004	VRNA_PATH_STEEPEST_DESCENT Folding Paths that start at a single Secondary
Reading/Writing Energy Parameter Sets from/to	Structure, 400
File, 451	VRNA_PATH_TYPE_DOT_BRACKET
vrna_params_reset Energy Parameters, 217	(Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima, 390
vrna_params_save	VRNA_PATH_TYPE_MOVES
Reading/Writing Energy Parameter Sets from/to File, 449	(Re-)folding Paths, Saddle Points, Energy Barriers, and Local Minima, 390
vrna_params_subst	vrna_pbacktrack
Energy Parameters, 215 vrna_path	Random Structure Samples from the Ensemble, 338
Folding Paths that start at a single Secondary	vrna_pbacktrack5
Structure, 401 VRNA_PATH_DEFAULT	Random Structure Samples from the Ensemble, 332
Folding Paths that start at a single Secondary	vrna_pbacktrack5_cb
Structure, 401	Random Structure Samples from the Ensemble,
vrna_path_direct	334
Direct Refolding Paths between two Secondary	
Structures, 397 vrna path direct ub	Random Structure Samples from the Ensemble,

vrna_pbacktrack5_resume	443
Random Structure Samples from the Ensemble,	vrna_pf_dimer_probs
335	Global Partition Function and Equilibrium Probabil
vrna_pbacktrack5_resume_cb	ities, 301
Random Structure Samples from the Ensemble,	vrna_pf_float_precision
336	Partition Function and Equilibrium Properties, 281
vrna_pbacktrack5_TwoD	vrna_pf_fold
Stochastic Backtracking of Structures from Dis-	Global Partition Function and Equilibrium Probabil
tance Based Partitioning, 369	ities, 304
vrna_pbacktrack_cb	vrna_pf_TwoD
Random Structure Samples from the Ensemble,	Computing Partition Functions of a Distance Based
340	Partitioning, 366
VRNA_PBACKTRACK_DEFAULT	vrna_pfl_fold
Random Structure Samples from the Ensemble,	Local (sliding window) Partition Function and Equi
330	librium Probabilities, 316
vrna_pbacktrack_mem_free	vrna_pfl_fold_cb
Random Structure Samples from the Ensemble,	Local (sliding window) Partition Function and Equi
344	librium Probabilities, 317
vrna_pbacktrack_mem_t	vrna_pfl_fold_up
Random Structure Samples from the Ensemble,	Local (sliding window) Partition Function and Equi
331	librium Probabilities, 317
VRNA_PBACKTRACK_NON_REDUNDANT	vrna_pfl_fold_up_cb
Random Structure Samples from the Ensemble,	Local (sliding window) Partition Function and Equi
330	librium Probabilities, 318
vrna_pbacktrack_num	vrna_pinfo_s, 502
Random Structure Samples from the Ensemble,	vrna_plist
339	Pair List Representation of Secondary Structures
vrna_pbacktrack_resume	492
Random Structure Samples from the Ensemble,	vrna_plist_from_probs
341	Global Partition Function and Equilibrium Probabil
vrna_pbacktrack_resume_cb	ities, 308
Random Structure Samples from the Ensemble,	vrna_plot_coords
343	Layouts and Coordinates, 552
vrna_pbacktrack_TwoD	vrna_plot_coords_circular
Stochastic Backtracking of Structures from Dis-	Layouts and Coordinates, 555
tance Based Partitioning, 368	vrna_plot_coords_circular_pt
vrna_pt	Layouts and Coordinates, 556
Global Partition Function and Equilibrium Probabil-	vrna_plot_coords_naview
ities, 302	Layouts and Coordinates, 557
vrna_pf_alifold	vrna_plot_coords_naview_pt Layouts and Coordinates, 557
Global Partition Function and Equilibrium Probabil-	vrna_plot_coords_pt
ities, 305 vrna_pf_circalifold	Layouts and Coordinates, 553
Global Partition Function and Equilibrium Probabil-	vrna_plot_coords_puzzler
ities, 306	Layouts and Coordinates, 558
vrna_pf_circfold	vrna_plot_coords_puzzler_pt
Global Partition Function and Equilibrium Probabil-	Layouts and Coordinates, 559
ities, 304	vrna_plot_coords_simple
vrna_pf_co_fold	Layouts and Coordinates, 554
Global Partition Function and Equilibrium Probabil-	vrna_plot_coords_simple_pt
ities, 309	Layouts and Coordinates, 554
Partition Function for Two Hybridized Sequences,	vrna_plot_coords_turtle
443	Layouts and Coordinates, 561
vrna_pf_dimer	vrna_plot_coords_turtle_pt
Global Partition Function and Equilibrium Probabil-	Layouts and Coordinates, 562
ities, 303	vrna_plot_layout
vrna_pf_dimer_concentrations	Layouts and Coordinates, 547
Partition Function for Two Hybridized Sequences,	vrna_plot_layout_circular
, , , , , , , , , , , , , , , , , , , ,	-, - <i>,</i> -

Layouts and Coordinates, 549	vrna_pt_ali_get
vrna_plot_layout_free	Pair Table Representation of Secondary Structures
Layouts and Coordinates, 551	489
vrna_plot_layout_naview	vrna_pt_pk_get
Layouts and Coordinates, 549	Pair Table Representation of Secondary Structures
vrna_plot_layout_puzzler	488
Layouts and Coordinates, 551	vrna_pt_pk_remove
vrna_plot_layout_s, 545	Pair Table Representation of Secondary Structures
vrna_plot_layout_simple	489
Layouts and Coordinates, 548	vrna_pt_snoop_get
vrna_plot_layout_t	Pair Table Representation of Secondary Structures
Layouts and Coordinates, 547	489
vrna_plot_layout_turtle	vrna_ptable
Layouts and Coordinates, 550	Pair Table Representation of Secondary Structures
vrna_plot_options_puzzler	487
Layouts and Coordinates, 560	vrna_ptable_copy
vrna_plot_options_puzzler_free	Pair Table Representation of Secondary Structures
Layouts and Coordinates, 560	489
vrna_plot_options_puzzler_t, 545	vrna_ptable_from_string
VRNA_PLOT_TYPE_CIRCULAR	Pair Table Representation of Secondary Structures
Layouts and Coordinates, 546	488
VRNA_PLOT_TYPE_NAVIEW	vrna_ptypes
Layouts and Coordinates, 546	Utilities to deal with Nucleotide Alphabets, 463
VRNA_PLOT_TYPE_PUZZLER	vrna_random_string
Layouts and Coordinates, 547	(Nucleic Acid Sequence) String Utilitites, 470
VRNA_PLOT_TYPE_SIMPLE	vrna_read_line
Layouts and Coordinates, 545	Files and I/O, 512
VRNA_PLOT_TYPE_TURTLE	vrna_realloc
Layouts and Coordinates, 546	Utilities, 425
vrna_positional_entropy	vrna_refBPcnt_matrix
Global Partition Function and Equilibrium Probabil-	Secondary Structure Utilities, 477
ities, 308	vrna_refBPdist_matrix
vrna_pr_energy	Secondary Structure Utilities, 477
equilibrium_probs.h, 737	vrna_rotational_symmetry
vrna_pr_structure	Combinatorics Algorithms, 572
Global Partition Function and Equilibrium Probabil-	vrna_rotational_symmetry_db
·	Combinatorics Algorithms, 574
ities, 302	vrna_rotational_symmetry_db_pos
vrna_probs_window	Combinatorics Algorithms, 574
Local (sliding window) Partition Function and Equi-	
librium Probabilities, 315	vrna_rotational_symmetry_num
VRNA_PROBS_WINDOW_BPP	Combinatorics Algorithms, 571
Local (sliding window) Partition Function and Equi-	vrna_rotational_symmetry_pos
librium Probabilities, 312	Combinatorics Algorithms, 573
vrna_probs_window_callback	vrna_rotational_symmetry_pos_num
Local (sliding window) Partition Function and Equi-	Combinatorics Algorithms, 571
librium Probabilities, 314	vrna_sc_add_bp
VRNA_PROBS_WINDOW_PF	Soft Constraints, 273
Local (sliding window) Partition Function and Equi-	vrna_sc_add_bt
librium Probabilities, 313	Soft Constraints, 276
VRNA_PROBS_WINDOW_STACKP	vrna_sc_add_data
Local (sliding window) Partition Function and Equi-	Soft Constraints, 275
librium Probabilities, 313	vrna_sc_add_exp_f
VRNA_PROBS_WINDOW_UP	Soft Constraints, 277
Local (sliding window) Partition Function and Equi-	vrna_sc_add_f
librium Probabilities, 312	Soft Constraints, 276
VRNA_PROBS_WINDOW_UP_SPLIT	vrna_sc_add_hi_motif
Local (sliding window) Partition Function and Equi-	Incorporating Ligands Binding to Specific Se
librium Probabilities, 313	auence/Structure Motifs using Soft Con-

straints, 417	vrna seg toupper
vrna_sc_add_SHAPE_deigan	(Nucleic Acid Sequence) String Utilitites, 472
SHAPE Reactivity Data, 406	vrna seq type e
vrna_sc_add_SHAPE_deigan_ali	Utilities to deal with Nucleotide Alphabets, 463
SHAPE Reactivity Data, 407	vrna_seq_ungapped
vrna sc add SHAPE zarringhalam	(Nucleic Acid Sequence) String Utilitites, 473
SHAPE Reactivity Data, 408	VRNA_SEQ_UNKNOWN
vrna_sc_add_up	Utilities to deal with Nucleotide Alphabets, 463
Soft Constraints, 274	vrna_sequence_s, 463
vrna_sc_bp_storage_t, 702	vrna_sol_TwoD_pf_t, 365
VRNA_SC_DEFAULT	Computing Partition Functions of a Distance Based
soft.h, 729	Partitioning, 366
vrna_sc_free	vrna_sol_TwoD_t, 357
Soft Constraints, 275	Computing MFE representatives of a Distance
vrna_sc_init	Based Partitioning, 358
Soft Constraints, 271	vrna_stack_prob
vrna_sc_minimize_pertubation	Global Partition Function and Equilibrium Probabil-
Generate Soft Constraints from Data, 413	ities, 301
vrna_sc_motif_s, 703	VRNA_STATUS_MFE_POST
vrna_sc_motif_t	The Fold Compound, 602
ligand.h, 726	VRNA_STATUS_MFE_PRE
vrna_sc_remove	The Fold Compound, 602
Soft Constraints, 274	VRNA_STATUS_PF_POST
vrna_sc_s, 267	The Fold Compound, 603
bt, 268	VRNA_STATUS_PF_PRE
exp_f, 268	The Fold Compound, 603
f, 268	vrna_strcat_printf
vrna_sc_set_bp	(Nucleic Acid Sequence) String Utilitites, 468
Soft Constraints, 272	vrna_strcat_vprintf
vrna_sc_set_up	(Nucleic Acid Sequence) String Utilitites, 469
Soft Constraints, 273	vrna_strdup_printf
vrna_sc_SHAPE_parse_method	(Nucleic Acid Sequence) String Utilitites, 467
SHAPE.h, 728	vrna_strdup_vprintf
vrna_sc_SHAPE_to_pr	(Nucleic Acid Sequence) String Utilitites, 468
SHAPE Reactivity Data, 408	vrna_strsplit
vrna_sc_type_e	(Nucleic Acid Sequence) String Utilitites, 470
soft.h, 729	VRNA_STRUCTURE_TREE_EXPANDED
VRNA_SC_WINDOW	Tree Representation of Secondary Structures, 496
soft.h, 729	VRNA_STRUCTURE_TREE_HIT
vrna_search_BM_BCT	Tree Representation of Secondary Structures, 495
Search Algorithms, 568	VRNA_STRUCTURE_TREE_SHAPIRO
vrna_search_BM_BCT_num	Tree Representation of Secondary Structures, 496
Search Algorithms, 568	VRNA_STRUCTURE_TREE_SHAPIRO_EXT
vrna_search_BMH	Tree Representation of Secondary Structures, 496
Search Algorithms, 567	VRNA_STRUCTURE_TREE_SHAPIRO_SHORT
vrna_search_BMH_num	Tree Representation of Secondary Structures, 495
Search Algorithms, 566	VRNA_STRUCTURE_TREE_SHAPIRO_WEIGHT
vrna_sect_s, 578	Tree Representation of Secondary Structures, 496
VRNA_SEQ_DNA	vrna_structured_domains_s, 703
Utilities to deal with Nucleotide Alphabets, 463	vrna_subopt
vrna_seq_encode	Suboptimal Structures within an Energy Band
Utilities to deal with Nucleotide Alphabets, 464	around the MFE, 325
vrna_seq_encode_simple	vrna_subopt_callback
Utilities to deal with Nucleotide Alphabets, 464	Suboptimal Structures within an Energy Band
VRNA_SEQ_RNA	around the MFE, 324
Utilities to deal with Nucleotide Alphabets, 463	vrna_subopt_cb
vrna_seq_toRNA (Nuclain Acid Sequence) String Litilities, 472	Suboptimal Structures within an Energy Band
(Nucleic Acid Sequence) String Utilitites, 472	around the MFE, 326

vrna_subopt_sol_s, 703	VRNA_UNIT_J
vrna_subopt_zuker	Unit Conversion, 590
Suboptimal Structures sensu Stiegler et al. 1984 /	VRNA_UNIT_K
Zuker et al. 1989, 322	Unit Conversion, 590
vrna_time_stamp	VRNA_UNIT_KCAL
Utilities, 426	Unit Conversion, 590
vrna_tree_string_to_db	VRNA_UNIT_KCAL_IT
Tree Representation of Secondary Structures, 498	Unit Conversion, 590
vrna_tree_string_unweight	VRNA_UNIT_KG_TNT
Tree Representation of Secondary Structures, 498	Unit Conversion, 590
vrna_ud_add_motif	VRNA_UNIT_KJ
Unstructured Domains, 231	Unit Conversion, 590
vrna_ud_get_motif_size_at	VRNA_UNIT_KWH
unstructured_domains.h, 813	Unit Conversion, 590
vrna_ud_motifs_centroid	VRNA_UNIT_T_TNT
Unstructured Domains, 229	Unit Conversion, 590
vrna_ud_motifs_MEA	vrna_unit_temperature_e
Unstructured Domains, 230	Unit Conversion, 590
vrna ud motifs MFE	VRNA_UNIT_WH
Unstructured Domains, 230	Unit Conversion, 590
vrna ud remove	vrna_unstructured_domain_motif_s, 703
Unstructured Domains, 232	vrna_unstructured_domain_s, 226
vrna_ud_set_data	prod_cb, 227
Unstructured Domains, 232	vrna_urn
vrna_ud_set_exp_prod_rule_cb	Utilities, 425
Unstructured Domains, 234	
vrna_ud_set_prob_cb	warn_user
unstructured_domains.h, 813	basic.h, 774
vrna_ud_set_prod_rule_cb	write_parameter_file
Unstructured Domains, 233	Reading/Writing Energy Parameter Sets from/to
VRNA_UNIT_CAL	File, 455
Unit Conversion, 590	xrealloc
VRNA_UNIT_CAL_IT	basic.h, 775
Unit Conversion, 590	xrna plot
VRNA_UNIT_DACAL	Plotting, 542
Unit Conversion, 590	xsubi
VRNA_UNIT_DACAL_IT	Utilities, 428
Unit Conversion, 590	Cuitios, 120
VRNA_UNIT_DEG_C	zukersubopt
Unit Conversion, 590	Suboptimal Structures sensu Stiegler et al. 1984
VRNA_UNIT_DEG_DE	Zuker et al. 1989, 323
Unit Conversion, 590	zukersubopt_par
VRNA_UNIT_DEG_F	Suboptimal Structures sensu Stiegler et al. 1984
Unit Conversion, 590	Zuker et al. 1989, 323
VRNA UNIT DEG N	
Unit Conversion, 590	
VRNA_UNIT_DEG_R	
Unit Conversion, 590	
VRNA_UNIT_DEG_RE	
Unit Conversion, 590	
VRNA_UNIT_DEG_RO	
Unit Conversion, 590	
vrna_unit_energy_e	
Unit Conversion, 589	
VRNA_UNIT_EV	
Unit Conversion, 590	
VRNA_UNIT_G_TNT	
Unit Conversion, 590	