## How to Compile your R Code in RMarkdown

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# Section 1: Background/Setting Up/Converting your R file to R-Markdown file

## **Background Rationale**

RMarkdown is an application that can run and save R-code into a high-quality document like PDF, HTML, Word, to name a few. First, RMarkdown runs in the R-Studio's Integrated development environment (IDE), making it convenient to access. All you need to do is to download the package using the command "install.packages("rmarkdown"). RMarkdown can run code in different coding languages, including Python or SQL. RMarkdown saves the time to write the report within R-studio,instead of exporting graphs to word documents.

The R coding language can be used for DNA Sequence Statistics, Comparative Genomics, Identifying homologous genes between two species. R Markdown is capable of compiling all these data into a presentable format for any reports. This tutorial will help you set up your RMarkdown file; There will be various R language commands to go over the *Iris dataset* by illustrating the *Iris dataset* into a pairwise plot, scatterplot, linear regression, boxplot, histogram, and heat map in order to display this dataset into an HTML and PDF outputs while using RMarkdown.

#### Set up for Rmarkdown

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- 1. Start RStudio and Open a "New Project"
- 2. Install RMarkdown
- Option 1: on the "console": code install.packages("rmarkdown")
- Option 2: on the upper tabs go to **Tools** next *Install packages* then search "rmarkdown" (default CRAN and install repositories)
- 3. Install TinyTex (to compile RMarkdown into a PDF document)
- Option 1: on the "console": code 'install.packages("tinytex") or tinytex::install\_tinytex()
- Option 2: on the upper tabs go to **Tools** next *Install packages* then search "tinytex" (default CRAN and install repositories)
- 4. Go to upper tabs **File**
- Start at "New File" next select RMarkdown then Name finally, select "HTML"

#### R file convert to RMarkdown file

- 1. Specify the code chunk with '#+'
- 2. Render the document as an RMarkdown file knitr::spin("filename.R", knit = FALSE, format = "Rmd")
- 3. This will generate an RMarkdown file called "filename.Rmd"
- 4. Knit this document, with your preferred output.

#### Text file (.txt) convert to Rmarkdown file

- 1. Save as ".Rmd" file
- 2. Now, you can use the various commands in RMarkdown

## Section 2: Formatting and Knit your outputs

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## Formatting in RMarkdown:

- 1. For Headers, the amount of "#" characters corresponds to the size of Header it is.
- 2. Links, must contain "http://" prior to the web domain in order to operate for example: http://www.uml.edu
- 3. Images ! [alt text here] (path-to-image-here) or ! [alt text here] (http://example.com/logo.png) ! [uml logo] (https://www.uml.edu/Images/uml\_vertical\_logo\_with\_black\_tagline\_tcm18-288512.png)
- 4. Also, commands can be found in the "Markdown Quick Reference" guide under the "Help" tab

## Choose your outputs

replace the current output with either of the following:

- 1. output: html\_document html file (web page)
- 2. output: pdf document pdf file(pdf)
- 3. output: ioslides presentation ioslides slideshow (html)
- 4. output: word\_document Microsoft Word (.docx)
- 5. output: beamer presentation beamer slideshow (pdf)

## Rendering/Knitting

- 1. Option 1: ''' rmarkdown::render("<insert your file path here>") '''
- 2. Option 2: ''' rmarkdown::render("<insert your file path here>","<insert output of your choice: pdf
- 3. Option 3: Knit Document button: File -> Knit Document (' '
- 4. Option 4: To knit your document: for Mac Users 'Command + Shift + K' and Windows/Linux Users 'Ctrl

## Section 3: Shortcuts and Tips

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#### **Keyboard Shortcuts**

- 1. We will create plenty of 'R Code Blocks' where R code will be evaluated and printed, a shortcut to insert a "R code Block" for Mac Users is 'Command + Option + I' and Windows/Linux Users is 'Ctrl + Alt + I'
- 2. To output your document: for Mac Users 'Command + Shift + K' and Windows/Linux Users 'Ctrl + Shift + K'
- 3. Run the current R code block: for Mac Users 'Command + shift + Enter' and Windows/Linux Users 'Ctrl + Shift + Enter'
- 4. Run all R code blocks in the .Rmd file: for Mac Users 'Command + Option + R' and and Windows/Linux Users 'Ctrl + Alt + R'

#### Altering Your R Code Blocks

- 1. eval = FALSE Show code, but do not evaluate it
- 2. fig.show = "hide" Hides plots
- 3. results = "hide" Hides printed output
- 4. include = FALSE Runs code, but suppresses all output. This is helpful for setup code.
- 5. message=TRUE Whether to display messages, FALSE does not display

This is not the definitive list of tricks, only the tutorials demostration. For more information, please refer back to R Markdown tips, tricks, and shortcuts in the references section

#### Youtube Video

https://youtu.be/8ogEaPWMwCw

# Section 4: Plotting/Modeling/Mapping with the "Iris" dataset

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### Selected Dataset

The **Iris dataset** which is one of many built-in datasets found in R. This dataset consists of 5 variables: sepal length, sepal width, petal length and petal width measured in centimeters of 50 samples each of 3 species of Iris: Iris setosa, versicolor, and virginica totaling 150 samples.

## install packages for this dataset

```
install.packages("RColorBrewer") # example of modfied R code block
install.packages("gplots") # show code only not evaluated, because these packages are already installed
```

## Load libraries for graphs

```
library(RColorBrewer) # for the colors of the graphs
library(gplots) # for heat map
```

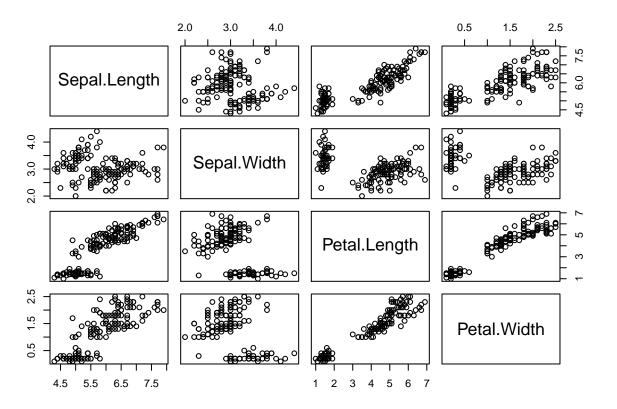
#### Load Iris Dataset

```
library(datasets)
data("iris") #"iris" is one of few built-in R datasets!
summary(iris) #output a summary data table of all 150 irises
##
    Sepal.Length
                  Sepal.Width
                                 Petal.Length
                                                Petal.Width
         :4.300 Min.
                                     :1.000
                                               Min. :0.100
## Min.
                       :2.000
                                Min.
## 1st Qu.:5.100 1st Qu.:2.800
                                1st Qu.:1.600
                                               1st Qu.:0.300
## Median :5.800 Median :3.000
                                Median :4.350
                                               Median :1.300
## Mean
        :5.843 Mean :3.057
                                Mean
                                      :3.758
                                               Mean :1.199
## 3rd Qu.:6.400 3rd Qu.:3.300
                                3rd Qu.:5.100
                                               3rd Qu.:1.800
## Max. :7.900 Max. :4.400
                                Max. :6.900
                                               Max.
                                                     :2.500
##
         Species
## setosa
            :50
## versicolor:50
## virginica:50
##
##
```

#### Plots for Iris Dataset

##

```
names(iris) # list variables
## [1] "Sepal.Length" "Sepal.Width" "Petal.Length" "Petal.Width" "Species"
pairs(iris[1:4]) # illustrate how variables compare to eachother. Excluding "Species"
```



#this function provides a list of scatterplots of each variable being "mapped" to eachother as both ind

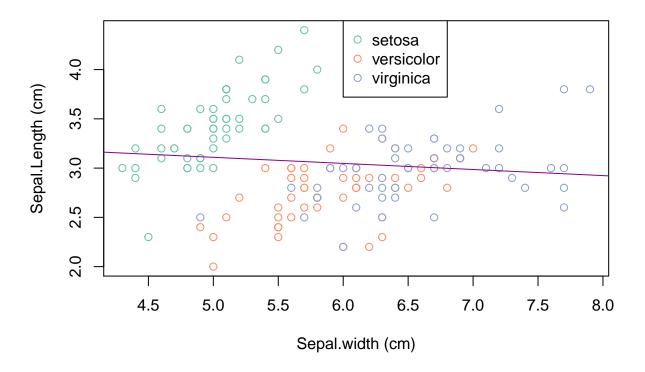
## **Exploring Rcolor Brewer**

```
library(RColorBrewer)
display.brewer.all(n=3)
brewer.pal.info # provide information of all possible palette names
```

# Regression model of Sepal\_Length vs Sepal\_Width of all three different Iris species of 150 samples

```
# Plotting the main and both axis titles
# RColorBrewer (n, name). n = number of different colors (3), name = selected from "brewer.pal.info". M
plot(Sepal.Width ~ Sepal.Length, data = iris, main = "Sepal_Length vs Sepal_Width of all three differen
# Add regression line
abline(lm(Sepal.Width ~ Sepal.Length, data = iris), col = 'darkmagenta')
# legend of of the three different species
legend(x=6.0, y=4.5, legend=levels(iris$Species), col=brewer.pal(3, "Set2"), pch=1)
```

## epal\_Length vs Sepal\_Width of all three different Iris species of 150 sa



```
#Summary table of regression data
summary(lm(Sepal.Width ~ Sepal.Length, data = iris))
```

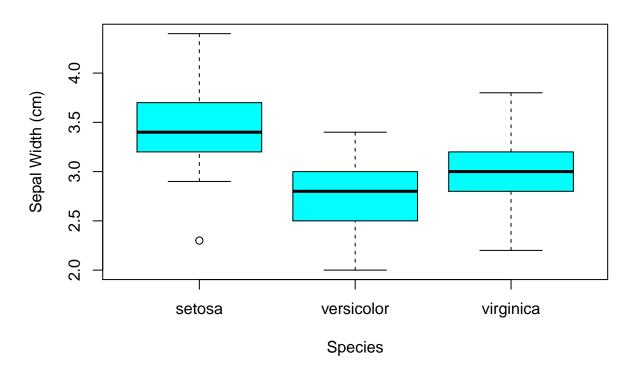
```
##
## Call:
  lm(formula = Sepal.Width ~ Sepal.Length, data = iris)
## Residuals:
##
       Min
                1Q Median
                                       Max
## -1.1095 -0.2454 -0.0167
                           0.2763
                                    1.3338
##
## Coefficients:
##
                Estimate Std. Error t value Pr(>|t|)
## (Intercept)
                 3.41895
                            0.25356
                                      13.48
                                              <2e-16 ***
## Sepal.Length -0.06188
                            0.04297
                                      -1.44
                                               0.152
                   0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
## Signif. codes:
## Residual standard error: 0.4343 on 148 degrees of freedom
## Multiple R-squared: 0.01382,
                                    Adjusted R-squared:
## F-statistic: 2.074 on 1 and 148 DF, p-value: 0.1519
```

 $\textit{\#Notice how Setosa have noticably short Sepal.width. However length its the longest compared to Versico \\ \textit{\#The regression line tells that future outcome will range around the line's value } \\$ 

## Boxplot of Sepal Width of all three different Iris species of 150 samples

boxplot(Sepal.Width ~ Species, data = iris, main = "Sepal Width of all three different Iris species of

## Sepal Width of all three different Iris species of 150 samples

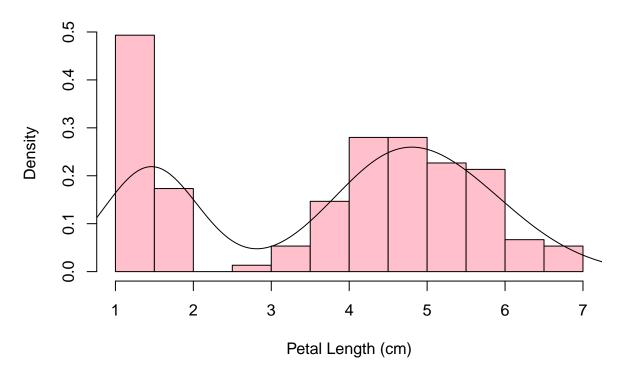


#Y-variable is mapped to X-variable
#Setosa have the miniumin sepal width as it's outlier however, it also have the greatest sepal width and

### Histogram of Petal Length of all three different Iris species of 150 samples

```
Petal_Length <- iris$Petal.Length #collect all petal length samples in all 150 Iris samples
hist(Petal_Length, main = "Petal Length of all Iris species of 150 samples", xlab = "Petal Length (cm)"
# freq = FALSE, check for population density instead of freq = TRUE which counts frequency
lines(density(iris$Petal.Length)) #create density plot line
```

## Petal Length of all Iris species of 150 samples



```
## Min. 1st Qu. Median Mean 3rd Qu. Max.
## 1.000 1.600 4.350 3.758 5.100 6.900
```

#this density plot indicates the likeliness based on frequency data where the Petal length is most like

## Heat Maps of Iris Dataset

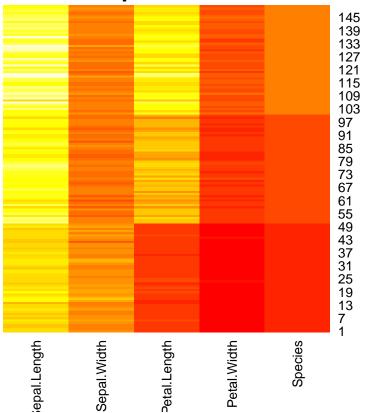
```
#heatmap() found in default R package
?heatmap # for quick reference guide

#set up class for iris dataset
class(iris)
```

## [1] "data.frame"

```
#convert iris_heat dataframe into a matrix
iris_heat <- data.matrix(iris)
heatmap(iris_heat, scale="none", cexRow=1, cexCol = 1, col= heat.colors(n=20), Rowv = NA, Colv = NA, y</pre>
```

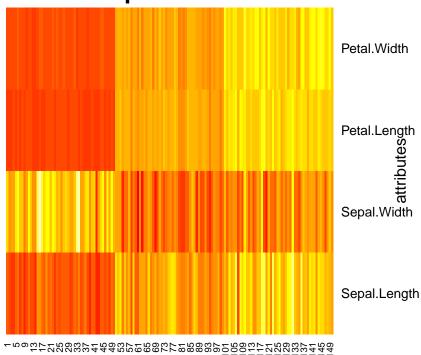




#Transpose dataset (flip x and y axis)
heatmap(t(iris[, 1:4]), scale="row", cexRow=1, col = heat.colors(n=20), Rowv = NA, Colv = NA, ylab =

sample number

# **Heatmap of Iris dataset**



```
#"scale" indicates if the values in either the row direction or column direction
# heat.colors() heat map default colors, n = number of colors. hcl.pals() to list avaliable color palet
#"Colv" for the column dendrogram and "RowV" for Row dendrogram. 'NA' does not render dendrograms in pl
#"cexRow" and "cexCol" are for scaling of the x and y axis
#red equates high and yellow equates to low value. More matching, means more similiar and related the d
# Notice samples 1-49 have lower sepal width compare to other samples, but greater value in the other a
```

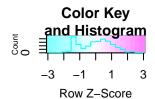
# Notice how consistent petal.width, petal.length. sepal.length is pattern wise based on the color howe

#### Iris Enhance Heat Map

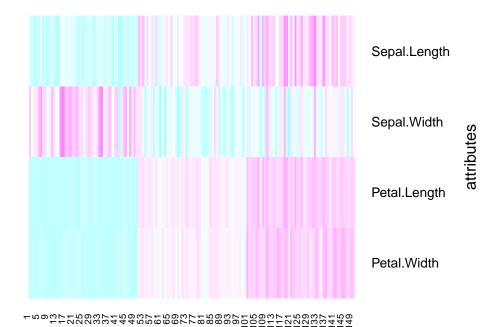
## column dendogram.

```
?heatmap.2 #reference guide
library(gplots) #enchanced heatmap
heatmap.2(t(iris[, 1:4]), trace="none", scale="row", key=TRUE, margins=c(2, 8), cexRow=1, col = cm.colo
## Warning in heatmap.2(t(iris[, 1:4]), trace = "none", scale = "row", key =
## TRUE, : Discrepancy: Rowv is FALSE, while dendrogram is 'both'. Omitting row
## dendogram.
## Warning in heatmap.2(t(iris[, 1:4]), trace = "none", scale = "row", key =
```

## TRUE, : Discrepancy: Colv is FALSE, while dendrogram is 'column'. Omitting



## **Heatmap of Iris dataset**



```
# "t(iris[,1:4])" matrix transpose that reconstruct the order of attributes and sample number
# "trace" shows you where on the range the cells are positions. options are column", "row", "both", "none"
# "key=TRUE" logical indicating whether a color-key should be shown.
# "margins = c(2,8)" gives less space to x axis labels, but more space to y axis labels
#if error of graphic error occurs debug with dev.off()
#about the data, it's very similar inference compare the to heatmap above however the presentation is a
```

## Section 5: Saving your Rmarkdown file and References

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Assuming that you have Adobe Acrobat Reader installed in your computer 1. Go to the top right corner of your PDF file 2. Go to "Downloads" 3. It'll open up in the Adobe Acrobat Reader Application/Client 4. Go to "File", Save As, rename and there you have your final PDF file of your report

Congratulations! you have learned the overall basics of how to use RMarkdown and hopefully you'll be able to incorporate these lessons of the tutorial into your assignments, projects, and overall productivity involving R and RStudio.

## References

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