# Package 'Giotto'

December 7, 2019

```
Title Spatial single-cell transcriptomics pipeline.
Version 0.1.3
Description Pipeline to process, analyze and visualize (spatial) single-cell expression data.
License MIT + file LICENSE
Encoding UTF-8
LazyData true
RoxygenNote 7.0.1
Depends data.table (>= 1.12.2),
      ggplot2 (>= 3.1.1),
      base (>= 3.5.1),
      utils (>= 3.5.1),
      R (>= 3.5.1)
Imports Rtsne (>= 0.15),
      uwot (>= 0.0.0.9010),
      multinet (>= 3.0.2),
      FactoMineR (>= 1.34),
      factoextra (>= 1.0.5),
      cowplot (>= 0.9.4),
      grDevices,
      RColorBrewer (>= 1.1-2),
      jackstraw (>= 1.3),
      dbscan (>= 1.1-3),
      ggalluvial (>= 0.9.1),
      scales (>= 1.0.0),
      ComplexHeatmap (>= 1.20.0),
      qvalue (>= 2.14.1),
      lfa (>= 1.12.0),
      igraph (>= 1.2.4.1),
      plotly,
      reticulate,
      magrittr,
      limma,
      ggdendro,
      smfishHmrf,
      matrixStats (>= 0.55.0),
      IRanges,
      devtools,
```

reshape2

2 R topics documented:

```
Suggests knitr,
rmarkdown,
MAST,
scran (>= 1.10.1),
png,
tiff,
biomaRt
```

# biocViews

VignetteBuilder knitr

Remotes lambdamoses/smfishhmrf-r

# ${\sf R}$ topics documented:

addCellMetadata
addCellStatistics
addGeneMetadata
addGeneStatistics
addHMRF
addNetworkLayout
addStatistics
adjustGiottoMatrix
aes_string2
allCellCellcommunicationsScores
all_plots_save_function
annotateGiotto
annotateSpatialNetwork
annotate_spatlocs_with_spatgrid_2D
annotate_spatlocs_with_spatgrid_3D
average_gene_gene_expression_in_groups
binGetSpatialGenes
calculateHVG
calculateMetaTable
calculateMetaTableCells
calculate_spatial_genes_python
cellProximityBarplot
cellProximityEnrichment
cellProximityHeatmap
cellProximityNetwork
cellProximitySpatPlot
cellProximitySpatPlot2D
cellProximitySpatPlot3D
cellProximityVisPlot
cellProximityVisPlot_2D_ggplot
cellProximityVisPlot_2D_plotly
cellProximityVisPlot_3D_plotly
changeGiottoInstructions
clusterCells
combineMetadata
convertEnsemblToGeneSymbol
createGiottoInstructions 46

createGiottoObject	47
createHeatmap_DT	49
createNearestNetwork	50
createSpatialEnrich	51
createSpatialGrid	52
createSpatialGrid_2D	53
createSpatialGrid_3D	54
createSpatialNetwork	55
create_average_detection_DT	56
create_average_DT	56
create_cell_type_random_cell_IDs	57
create_cluster_matrix	58
create_dimObject	58
decide_cluster_order	59
detectSpatialPatterns	60
dimCellPlot	61
dimCellPlot2D	63
dimGenePlot	66
dimGenePlot2D	68
dimGenePlot3D	70
dimPlot	72
dimPlot2D	75
dimPlot2D_single	78
dimPlot3D	80
direction_test_CPG	82
doHclust	
doHMRF	84
doKmeans	85
doLeidenCluster	87
doLeidenSubCluster	
doLouvainCluster	
doLouvainCluster_community	90
doLouvainCluster_multinet	92
doLouvainSubCluster	93
doLouvainSubCluster_community	94
doLouvainSubCluster_multinet	96
doRandomWalkCluster	97
doSNNCluster	98
dt_to_matrix	99
exportGiottoViewer	100
exprOnlyCellCellcommunicationScores	101
extended_gini_fun	
extractNearestNetwork	102
fDataDT	103
filterCombinations	103
filterCPGscores	
filterDistributions	
filterGiotto	
findGiniMarkers	
findGiniMarkers_one_vs_all	
findMarkers	
findMarkers one vs all	

findMastMarkers
findMastMarkers one vs all
findScranMarkers
findScranMarkers_one_vs_all
find_grid_2D
find grid 3D
find_grid_x
find_grid_y
find grid z
fish_function
fish_function2
FSV_show
GenePattern_show
general_save_function
get10Xmatrix
getCellProximityGeneScores
getClusterSimilarity
getDendrogramSplits
getDistinctColors
getGeneToGeneScores
get_cell_to_cell_sorted_name_conversion
get_interaction_gene_enrichment
get_specific_interaction_gene_enrichment
ggplot_save_function
giotto-class
hyperGeometricEnrich
iterCluster
iterLeidenCluster
iterLouvainCluster
iterLouvainCluster_community
iterLouvainCluster_multinet
kmeans binarize
loadHMRF
makeSignMatrixPAGE
makeSignMatrixRank
make_simulated_network
mergeClusters
mygini_fun
nnDT to kNN
node clusters
normalizeGiotto
OR function2
PAGEEnrich
pDataDT
plotCPGscores
plotGTGscores
plotHeatmap
plotly_axis_scale_2D
plotly_axis_scale_3D
plotly_grid
plotly_network
plotMetaDataCellsHeatmap

plotMetaDataHeatmap	
plotPCA	
plotPCA_2D	
plotPCA_3D	
plotTSNE	
plotTSNE_2D	
plotTSNE_3D	. 166
plotUMAP	. 168
plotUMAP_2D	
plotUMAP_3D	
plot_network_layer_ggplot	
plot_point_layer_ggplot	
plot_spat_point_layer_ggplot	. 176
print.giotto	. 178
rankEnrich	. 178
rank_binarize	. 179
readGiottoInstructions	. 179
removeCellAnnotation	. 180
removeGeneAnnotation	. 180
replaceGiottoInstructions	
runPCA	
runtSNE	
runUMAP	
selectPatternGenes	
select_expression_values	
show,giotto-method	
showClusterDendrogram	
showClusterHeatmap	
showCPGscores	
showGeneExpressionProximityScore	
showGiottoInstructions	
showGTGscores	
showIntExpressionProximityScore	
showPattern	
showPattern2D	
showPattern3D	
showPatternGenes	
showProcessingSteps	
showTopGeneToGene	
signPCA	
spatCellPlot	
spatCellPlot2D	
spatDimCellPlot	
spatDimCellPlot2D	
spatDimGenePlot	
spatDimGenePlot2D	
spatDimGenePlot3D	
spatDimPlot	
spatDimPlot2D	
spatDimPlot3D	
spatGenePlot	
spatGenePlot2D	
~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	. <u>-</u>

spatGenePlot3D	
Spatial_AEH	237
Spatial_DE	238
spatPlot	239
spatPlot2D	242
spatPlot2D_single	245
spatPlot3D	248
specificCellCellcommunicationScores	
split_dendrogram_in_two	
stitchFieldCoordinates	252
subClusterCells	253
subsetGiotto	
subsetGiottoLocs	
viewHMRFresults	
viewHMRFresults2D	
viewHMRFresults3D	
violinPlot	
visDimGenePlot	
visDimGenePlot_2D_ggplot	262
visDimGenePlot_3D_plotly	
visDimPlot	
visDimPlot_2D_ggplot	
visDimPlot_2D_plotly	
visDimPlot_3D_plotly	
visForceLayoutPlot	
visGenePlot	
visGenePlot_2D_ggplot	
visGenePlot_3D_plotly	
visPlot	
visPlot_2D_ggplot	
visPlot_2D_plotly	
visPlot_3D_plotly	
visSpatDimGenePlot	
visSpatDimGenePlot_2D	
visSpatDimGenePlot_3D	
visSpatDimPlot	
visSpatDimPlot_2D	
visSpatDimPlot_3D	
writeHMRFresults	
write_giotto_viewer_annotation	
write_giotto_viewer_dim_reduction	
write_giotto_viewer_numeric_annotation	303

Index 304

addCellMetadata 7

addCellMetadata addCellMetadata

## **Description**

adds cell metadata to the giotto object

#### Usage

```
addCellMetadata(
  gobject,
  new_metadata,
  by_column = FALSE,
  column_cell_ID = NULL
)
```

# Arguments

gobject giotto object

new\_metadata new cell metadata to use (data.table, data.frame, ...)

by\_column merge metadata based on cell\_ID column in pDataDT (default = FALSE)

column\_cell\_ID column name of new metadata to use if by\_column = TRUE

# **Details**

You can add additional cell metadata in two manners: 1. Provide a data.table or data.frame with cell annotations in the same order as the cell\_ID column in pDataDT(gobject) 2. Provide a data.table or data.frame with cell annotations and specificy which column contains the cell IDs, these cell IDs need to match with the cell\_ID column in pDataDT(gobject)

## Value

giotto object

# **Examples**

addCellMetadata(gobject)

addCellStatistics addCellStatistics

# Description

adds cells statistics to the giotto object

8 addGeneMetadata

#### Usage

```
addCellStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

# Arguments

## **Details**

This function will add the following statistics to cell metadata:

- nr\_genes: Denotes in how many genes are detected per cell
- perc\_genes: Denotes what percentage of genes is detected per cell
- total\_expr: Shows the total sum of gene expression per cell

## Value

```
giotto object if return_gobject = TRUE
```

## **Examples**

```
addCellStatistics(gobject)
```

addGeneMetadata

addGeneMetadata

#### **Description**

adds gene metadata to the giotto object

## Usage

```
addGeneMetadata(gobject, new_metadata, by_column = F, column_gene_ID = NULL)
```

```
gobject giotto object

new_metadata new metadata to use

by_column merge metadata based on gene_ID column in fDataDT

column_cell_ID column name of new metadata to use if by_column = TRUE
```

addGeneStatistics 9

#### **Details**

You can add additional gene metadata in two manners: 1. Provide a data.table or data.frame with gene annotations in the same order as the gene\_ID column in fDataDT(gobject) 2. Provide a data.table or data.frame with gene annotations and specificy which column contains the gene IDs, these gene IDs need to match with the gene\_ID column in fDataDT(gobject)

#### Value

giotto object

# **Examples**

addGeneMetadata(gobject)

addGeneStatistics

addGeneStatistics

#### **Description**

adds gene statistics to the giotto object

#### Usage

```
addGeneStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

# **Arguments**

#### **Details**

This function will add the following statistics to gene metadata:

- nr\_cells: Denotes in how many cells the gene is detected
- per\_cells: Denotes in what percentage of cells the gene is detected
- total\_expr: Shows the total sum of gene expression in all cells
- mean\_expr: Average gene expression in all cells
- mean\_expr\_det: Average gene expression in cells with detectable levels of the gene

10 addHMRF

#### Value

```
giotto object if return_gobject = TRUE
```

# **Examples**

addGeneStatistics(gobject)

addHMRF

addHMRF

# Description

Add selected results from doHMRF to the giotto object

# Usage

```
addHMRF(gobject, HMRFoutput, k = NULL, betas_to_add = NULL, hmrf_name = NULL)
```

# Arguments

gobject giotto object

 $\label{eq:hmrf} HMRF \ output \ from \ do HMRF()$ 

k number of domains

name specify a custom name

# **Details**

Description ...

#### Value

giotto object

# **Examples**

addHMRF(gobject)

addNetworkLayout 11

addNetworkLayout

addNetworkLayout

# Description

Add a network layout for a selected nearest neighbor network

# Usage

```
addNetworkLayout(
  gobject,
  nn_network_to_use = NULL,
  network_name = NULL,
  layout_type = c("drl"),
  options_list = NULL,
  layout_name = "layout",
  return_gobject = TRUE
)
```

# **Arguments**

# Details

Description of layouts and options.

#### Value

giotto object with updated layout for selected NN network

# **Examples**

```
addNetworkLayout(gobject)
```

12 adjustGiottoMatrix

addStatistics

addStatistics

# **Description**

adds genes and cells statistics to the giotto object

## Usage

```
addStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

# **Arguments**

## **Details**

See addGeneStatistics and addCellStatistics

# Value

```
giotto object if return_gobject = TRUE, else a list with results
```

# **Examples**

```
addStatistics(gobject)
```

adjustGiottoMatrix adjustGiottoMatrix

# Description

normalize and/or scale expresion values of Giotto object

aes\_string2

## Usage

```
adjustGiottoMatrix(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  batch_columns = NULL,
  covariate_columns = NULL,
  return_gobject = TRUE,
  update_slot = c("custom")
)
```

## **Arguments**

```
gobject giotto object

expression_values

expression values to use

batch_columns metadata columns that represent different batch (max = 2)

covariate_columns

metadata columns that represent covariates to regress out

return_gobject boolean: return giotto object (default = TRUE)

update_slot expression slot that will be updated (default = custom)
```

#### **Details**

This function implements the limma::removeBatchEffect function to remove known batch effects and to adjust expression values according to provided covariates.

#### Value

giotto object

# **Examples**

```
adjustGiottoMatrix(gobject)
```

```
aes_string2 aes_string2
```

# Description

makes sure aes\_string can also be used with names that start with numeric values

```
aes_string2(...)
```

```
all \textit{CellCellcommunications} Scores \\ all \textit{CellCellcommunications} Scores
```

## **Description**

All Cell-Cell communication scores based on spatial expression of interacting cells

# Usage

```
allCellCellcommunicationsScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "cell_types",
  random_iter = 100,
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  min_observations = 2,
  verbose = c("a little", "a lot", "none")
)
```

#### **Arguments**

```
giotto object to use
gobject
spatial_network_name
                  spatial network to use for identifying interacting cells
cluster_column cluster column with cell type information
random\_iter
                  number of iterations
gene_set_1
                  first specific gene set from gene pairs
                  second specific gene set from gene pairs
gene_set_2
log2FC_addendum
                  addendum to add when calculating log2FC
min_observations
                  minimum number of interactions needed to be considered
verbose
                  verbose
```

## **Details**

Details will follow.

#### Value

Cell-Cell communication scores for gene pairs based on spatial interaction

## **Examples**

```
allCellCellcommunicationsScores(gobject)
```

```
all_plots_save_function 
 all_plots_save_function
```

# Description

Function to automatically save plots to directory of interest

# Usage

```
all_plots_save_function(
  gobject,
  plot_object,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  default_save_name = "giotto_plot",
  save_format = NULL,
  show_saved_plot = F,
  ncol = 1,
  nrow = 1,
  scale = 1,
  base_width = NULL,
  base_height = NULL,
  base_aspect_ratio = NULL,
  units = NULL,
  dpi = NULL,
  limitsize = TRUE,
)
```

```
gobject
                  giotto object
                  object to plot
plot_object
save_dir
                  directory to save to
save_folder
                  folder in save_dir to save to
save_name
                  name of plot
save_format
                  format (e.g. png, tiff, pdf, ...)
show_saved_plot
                  load & display the saved plot
ncol
                  number of columns
                  number of rows
nrow
scale
                  scale
base_width
                  width
base_height
                  height
{\tt base\_aspect\_ratio}
                  aspect ratio
```

16 annotateGiotto

```
units units
dpi Plot resolution
```

limitsize When TRUE (the default), ggsave will not save images larger than 50x50 inches,

to prevent the common error of specifying dimensions in pixels.

... additional parameters to ggplot\_save\_function or general\_save\_function

#### See Also

```
Giotto::general_save_function
```

## **Examples**

```
all_plots_save_function(gobject)
```

annotateGiotto

annotateGiotto

# **Description**

Converts cluster results into provided annotation.

# Usage

```
annotateGiotto(
  gobject,
  annotation_vector = NULL,
  cluster_column = NULL,
  name = "cell_types"
)
```

#### **Arguments**

## Details

You need to specifify which (cluster) column you want to annotate and you need to provide an annotation vector like this:

- 1. identify the cell type of each cluster
- 2. create a vector of these cell types, e.g. cell\_types = c('T-cell', 'B-cell', 'Stromal')
- 3. provide original cluster names to previous vector, e.g. names(cell\_types) = c(2, 1, 3)

# Value

giotto object

## **Examples**

```
annotateGiotto(gobject)
```

 $annotate {\tt Spatial Network}$ 

annotateSpatialNetwork

# Description

Annotate spatial network with cell metadata information.

# Usage

```
annotateSpatialNetwork(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column
)
```

# **Arguments**

# Value

annotated network in data.table format

# **Examples**

```
annotateSpatialNetwork(gobject)
```

```
annotate\_spatlocs\_with\_spatgrid\_2D \\ annotate\_spatlocs\_with\_spatgrid\_2D
```

# Description

annotate spatial locations with 2D spatial grid information

# Usage

```
annotate_spatlocs_with_spatgrid_2D(spatloc, spatgrid)
```

```
spatloc spatial_locs slot from giotto object spatgrid selected spatial_grid slot from giotto object
```

#### Value

annotated spatial location data.table

# **Examples**

```
annotate_spatlocs_with_spatgrid_2D()
```

```
annotate\_spatlocs\_with\_spatgrid\_3D \\ annotate\_spatlocs\_with\_spatgrid\_3D
```

# Description

annotate spatial locations with 3D spatial grid information

# Usage

```
annotate_spatlocs_with_spatgrid_3D(spatloc, spatgrid)
```

# **Arguments**

```
spatloc spatial_locs slot from giotto object spatgrid selected spatial_grid slot from giotto object
```

## Value

annotated spatial location data.table

## **Examples**

```
annotate_spatlocs_with_spatgrid_3D()
```

```
average_gene_gene_expression_in_groups

average_gene_gene_expression_in_groups
```

# **Description**

calculate average expression per cluster

```
average_gene_gene_expression_in_groups(
  gobject,
  cluster_column = "cell_types",
  gene_set_1,
  gene_set_2
)
```

binGetSpatialGenes 19

## **Arguments**

```
gobject giotto object to use
cluster_column cluster column with cell type information
gene_set_1 first specific gene set from gene pairs
gene_set_2 second specific gene set from gene pairs
```

#### **Details**

Details will follow.

#### Value

data.table with average expression scores for each cluster

#### **Examples**

```
average_gene_gene_expression_in_groups(gobject)
```

binGetSpatialGenes

binGetSpatialGenes

#### **Description**

Rapid computation of genes that are spatially clustered

#### Usage

```
binGetSpatialGenes(
  gobject,
  bin_method = c("kmeans", "rank"),
  expression_values = c("normalized", "scaled", "custom"),
  subset_genes = NULL,
  spatial_network_name = "spatial_network",
  nstart = 3,
  iter_max = 10,
  percentage_rank = 10,
  do_fisher_test = F,
  community_expectation = 5,
  verbose = F
)
```

20 calculateHVG

nstart kmeans: nstart parameter iter\_max kmeans: iter.max parameter

do\_fisher\_test perform fisher test

community\_expectation

cell degree expectation in spatial communities

verbose be verbose

rank\_percentage

percentage of top cells for binarization

## **Details**

We provide two ways to identify spatial genes based on gene expression binarization. Both methods are identicial except for how binarization is performed.

- 1. binarize: Each gene is binarized (0 or 1) in each cell with **kmeans** (k = 2) or based on **rank** percentile
- 2. network: Alll cells are connected through a k-nearest neighbor network
- 3. contingency table: A contingency table is calculated based on all pairwise cell-cell interactions (0-0, 0-1, 1-0 or 1-1)
- 4. For each gene an odds-ratio (OR) and fisher.test (optional) is calculated

Additionally 2 other statistics are provided:

- Number of cells with high expression (binary = 1)
- total and ratio of highly connected cells: Cells with a connectivity higher than community\_expectation

By selecting a subset of likely spatial genes (e.g. highly variable genes) the function will be much faster.

## Value

data.table with results (see details)

# **Examples**

binGetSpatialGenes(gobject)

calculateHVG calculateHVG

# **Description**

compute highly variable genes

calculateHVG 21

#### Usage

```
calculateHVG(
      gobject,
      expression_values = c("normalized", "scaled", "custom"),
      method = c("cov_groups", "cov_loess"),
      reverse_log_scale = FALSE,
      logbase = 2,
      expression_threshold = 0,
      nr_expression_groups = 20,
      zscore_threshold = 1.5,
      HVGname = "hvg",
      difference_in_cov = 0.1,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "HVGplot",
      return_gobject = TRUE
    )
Arguments
    gobject
                     giotto object
    expression_values
                     expression values to use
                     method to calculate highly variable genes
    method
    reverse_log_scale
                     reverse log-scale of expression values (default = FALSE)
    logbase
                     if reverse_log_scale is TRUE, which log base was used?
    expression_threshold
                     expression threshold to consider a gene detected
    nr_expression_groups
                     number of expression groups for cov_groups
    zscore_threshold
                     zscore to select hvg for cov_groups
    HVGname
                     name for highly variable genes in cell metadata
    difference_in_cov
                     minimum difference in coefficient of variance required
    show_plot
                     show plot
                     return ggplot object
    return_plot
                     directly save the plot [boolean]
    save_plot
                     list of saving parameters from all_plots_save_function()
    save_param
    default_save_name
                     default save name for saving, don't change, change save_name in save_param
```

return\_gobject boolean: return giotto object (default = TRUE)

22 calculateMetaTable

#### **Details**

Currently we provide 2 ways to calculate highly variable genes: 1. high coeff of variance (COV) within groups:

First genes are binned (*nr\_expression\_groups*) into average expression groups and the COV for each gene is converted into a z-score within each bin. Genes with a z-score higher than the threshold (*zscore\_threshold*) are considered highly variable.

# 2. high COV based on loess regression prediction:

A predicted COV is calculated for each gene using loess regression (COV~log(mean expression)) Genes that show a higher than predicted COV (*difference\_in\_cov*) are considered highly variable.

#### Value

giotto object highly variable genes appended to gene metadata (fDataDT)

#### **Examples**

```
calculateHVG(gobject)
```

calculateMetaTable

calculateMetaTable

#### **Description**

calculates the average gene expression for one or more (combined) annotation columns.

## Usage

```
calculateMetaTable(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metadata_cols = NULL,
  selected_genes = NULL
)
```

#### **Arguments**

```
gobject giotto object
expression_values
expression values to use
metadata_cols annotation columns found in pDataDT(gobject)
selected_genes subset of genes to use
```

# Value

data.table with average expression values for each gene per (combined) annotation

## **Examples**

```
calculateMetaTable(gobject)
```

calculateMetaTableCells 23

```
calculateMetaTableCells
```

calculateMetaTableCells

## **Description**

calculates the average metadata values for one or more (combined) annotation columns.

#### Usage

```
calculateMetaTableCells(
  gobject,
  value_cols = NULL,
  metadata_cols = NULL,
  spat_enr_names = NULL
)
```

## **Arguments**

```
gobject giotto object
value_cols metadata or enrichment value columns to use
metadata_cols annotation columns found in pDataDT(gobject)
spat_enr_names which spatial enrichment results to include
```

#### Value

data.table with average metadata values per (combined) annotation

#### **Examples**

```
calculateMetaTableCells(gobject)
```

```
calculate\_spatial\_genes\_python \\ calculate\_spatial\_genes\_python
```

## **Description**

Calculate spatial genes using distance matrix.

```
calculate_spatial_genes_python(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metric = "euclidean",
  subset_genes = NULL,
  rbp_p = 0.95,
  examine_top = 0.3,
  python_path = NULL
)
```

## **Arguments**

```
gobject giotto object
expression_values
expression values to use

metric distance metric to use
subset_genes only run on this subset of genes
rbp_p fractional binarization threshold
examine_top top fraction to evaluate with silhouette
python_path specify specific path to python if required
```

#### **Details**

Description of how we compute spatial pattern genes.

#### Value

data.table with spatial scores

## **Examples**

```
calculate_spatial_genes_python(gobject)
```

```
cellProximityBarplot cellProximityBarplot
```

# Description

Create barplot from cell-cell proximity scores

```
cellProximityBarplot(
  gobject,
  CPscore,
  min_orig_ints = 5,
  min_sim_ints = 5,
  p_val = 0.05,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityBarplot"
)
```

#### **Arguments**

gobject giotto object CPscore, output from cellProximityEnrichment() **CPscore** filter on minimum original cell-cell interactions min\_orig\_ints min\_sim\_ints filter on minimum simulated cell-cell interactions p\_val p-value show\_plot show plot return\_plot return ggplot object save\_plot directly save the plot [boolean] save\_param list of saving parameters from all\_plots\_save\_function() default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

## **Details**

This function creates a barplot that shows the spatial proximity enrichment or depletion of cell type pairs.

#### Value

ggplot barplot

# **Examples**

```
cellProximityBarplot(CPscore)
```

```
cellProximityEnrichment
```

cellProximityEnrichment

# **Description**

Compute cell-cell interaction enrichment (observed vs expected)

## Usage

```
cellProximityEnrichment(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column,
  number_of_simulations = 100
)
```

#### **Details**

Spatial proximity enrichment or depletion between pairs of cell types is calculated by calculating the observed over the expected frequency of cell-cell proximity interactions. The expected frequency is the average frequency calculated from a number of spatial network simulations. Each individual simulation is obtained by random permutations of the cell type labels of each node (cell) in the spatial network.

#### Value

List of cell Proximity scores (CPscores) in data.table format. The first data.table (raw\_sim\_table) shows the raw observations of both the original and simulated networks. The second data.table (enrichm\_res) shows the enrichment results.

## **Examples**

```
cellProximityEnrichment(gobject)
```

```
cellProximityHeatmap cellProximityHeatmap
```

#### **Description**

Create heatmap from cell-cell proximity scores

# Usage

```
cellProximityHeatmap(
  gobject,
  CPscore,
  scale = T,
  order_cell_types = T,
  color_breaks = NULL,
  color_names = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityHeatmap")
```

```
gobject giotto object

CPscore CPscore, output from cellProximityEnrichment()

scale scale cell-cell proximity interaction scores

order_cell_types

order cell types based on enrichment correlation

color_breaks numerical vector of length 3 to represent min, mean and maximum

color_names character color vector of length 3
```

cellProximityNetwork 27

```
show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function()

default_save_name

default save name for saving, don't change, change save_name in save_param
```

#### **Details**

This function creates a heatmap that shows the spatial proximity enrichment or depletion of cell type pairs.

#### Value

ggplot heatmap

# **Examples**

```
cellProximityHeatmap(CPscore)
```

```
cellProximityNetwork cellProximityNetwork
```

#### **Description**

Create network from cell-cell proximity scores

```
cellProximityNetwork(
  gobject,
  CPscore,
  remove_self_edges = FALSE,
  self_loop_strength = 0.1,
  color_depletion = "lightgreen",
  color_enrichment = "red",
  rescale_edge_weights = TRUE,
  edge_weight_range_depletion = c(0.1, 1),
  edge_weight_range_enrichment = c(1, 5),
  layout = "Fruchterman",
  only_show_enrichment_edges = F,
  edge_width_range = c(0.1, 2),
  node_size = 4,
  node_text_size = 6,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityNetwork"
)
```

#### **Arguments**

color for depleted cen-cen interactions

color\_enrichment

color for enriched cell-cell interactions

rescale\_edge\_weights

rescale edge weights (boolean)

edge\_weight\_range\_depletion

numerical vector of length 2 to rescale depleted edge weights

edge\_weight\_range\_enrichment

numerical vector of length 2 to rescale enriched edge weights

layout algorithm to use to draw nodes and edges

only\_show\_enrichment\_edges

show only the enriched pairwise scores

edge\_width\_range

range of edge width

node\_size size of nodes

node\_text\_size size of node labels

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

# **Details**

This function creates a network that shows the spatial proximity enrichment or depletion of cell type pairs.

#### Value

igraph plot

# **Examples**

cellProximityNetwork(CPscore)

cellProximitySpatPlot 29

cellProximitySpatPlot cellProximitySpatPlot

## **Description**

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

# Usage

```
cellProximitySpatPlot(gobject, ...)
```

```
gobject
                  giotto object
interaction_name
                  cell-cell interaction name
cluster_column cluster column with cell clusters
sdimx
                  x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimy
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
point_size_select
                  size of selected points
point_select_border_col
                  border color of selected points
point_select_border_stroke
                  stroke size of selected points
point_size_other
                  size of other points
```

#### **Details**

Description of parameters.

#### Value

ggplot

#### See Also

cellProximitySpatPlot2D and cellProximitySpatPlot3D for 3D

## **Examples**

```
cellProximitySpatPlot(gobject)
```

```
cellProximitySpatPlot2D
```

cellProximitySpatPlot2D

# Description

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

```
cellProximitySpatPlot2D(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  cell_color = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
```

```
spatial_network_name = "spatial_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 1,
  show_legend = T,
 point_size_select = 2,
  point_select_border_col = "black",
  point_select_border_stroke = 0.05,
 point_size_other = 1,
 point_alpha_other = 0.3,
  point_other_border_col = "lightgrey",
 point_other_border_stroke = 0.01,
  show_plot = NA,
 return_plot = NA,
  save_plot = NA,
  save_param = list(),
 default_save_name = "cellProximitySpatPlot2D"
)
```

```
giotto object
gobject
interaction_name
                  cell-cell interaction name
cluster_column cluster column with cell clusters
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
                  color for cells (see details)
cell_color
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
point_size_select
                  size of selected points
```

```
point_select_border_col
                  border color of selected points
point_select_border_stroke
                  stroke size of selected points
point_size_other
                  size of other points
point_other_border_col
                  border color of other points
point_other_border_stroke
                  stroke size of other points
show_plot
                  show plots
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

#### **Details**

Description of parameters.

#### Value

ggplot

# **Examples**

```
cellProximitySpatPlot2D(gobject)
```

```
cell Proximity Spat Plot 3D \\ cell Proximity Spat Plot 2D
```

## **Description**

Visualize 3D cell-cell interactions according to spatial coordinates in plotly mode

```
cellProximitySpatPlot3D(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = T,
```

```
show_network = T,
      show_other_network = F,
     network_color = NULL,
      spatial_network_name = "spatial_network",
      show_grid = F,
     grid_color = NULL,
      spatial_grid_name = "spatial_grid",
      show_legend = T,
      point_size_select = 4,
     point_size_other = 2,
     point_alpha_other = 0.5,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
     x_ticks = NULL,
     y_ticks = NULL,
      z_ticks = NULL,
      show_plot = NA,
     return_plot = NA,
      save_plot = NA,
     save_param = list(),
     default_save_name = "cellProximitySpatPlot3D",
   )
Arguments
   gobject
                    giotto object
   interaction_name
                    cell-cell interaction name
   cluster_column cluster column with cell clusters
   sdimx
```

```
x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimy
                  z-axis dimension name (default = 'sdimz')
sdimz
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
                  color of spatial grid
grid_color
spatial_grid_name
                  name of spatial grid to use
show_legend
                  show legend
```

cellProximityVisPlot

## **Details**

Description of parameters.

#### Value

plotly

## **Examples**

```
cellProximitySpatPlot3D(gobject)
```

```
cellProximityVisPlot cellProximityVisPlot
```

# **Description**

Visualize cell-cell interactions according to spatial coordinates

```
cellProximityVisPlot(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
```

cellProximityVisPlot 35

```
coord_fix_ratio = 1,
      show_legend = T,
      point_size_select = 2,
      point_select_border_col = "black",
      point_select_border_stroke = 0.05,
      point_size_other = 1,
      point_alpha_other = 0.3,
      point_other_border_col = "lightgrey",
      point_other_border_stroke = 0.01,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      plot_method = c("ggplot", "plotly"),
    )
Arguments
   gobject
                     giotto object
    interaction_name
                     cell-cell interaction name
    cluster_column cluster column with cell clusters
                     x-axis dimension name (default = 'sdimx')
    sdimx
                     y-axis dimension name (default = 'sdimy')
    sdimy
    sdimz
                     z-axis dimension name (default = 'sdimz')
    cell_color
                     color for cells (see details)
    cell_color_code
                     named vector with colors
    color_as_factor
                     convert color column to factor
                     show underlying spatial network
    show_network
    network_color
                     color of spatial network
    spatial_network_name
                     name of spatial network to use
    show_grid
                     show spatial grid
    grid_color
                     color of spatial grid
    spatial_grid_name
                     name of spatial grid to use
    coord_fix_ratio
                     fix ratio between x and y-axis
    {\sf show\_legend}
                     show legend
   point_size_select
                     size of selected points
   {\tt point\_select\_border\_col}
```

border color of selected points

## **Details**

Description of parameters.

#### Value

ggplot or plotly

#### **Examples**

cellProximityVisPlot(gobject)

```
cell Proximity VisPlot\_2D\_ggplot \\ cell Proximity VisPlot\_2D\_ggplot
```

## **Description**

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

```
cellProximityVisPlot_2D_ggplot(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 1,
  show_legend = T,
  point_size_select = 2,
```

```
point_select_border_col = "black",
 point_select_border_stroke = 0.05,
 point_size_other = 1,
 point_alpha_other = 0.3,
 point_other_border_col = "lightgrey",
 point_other_border_stroke = 0.01,
)
```

```
gobject
                  giotto object
interaction_name
                  cell-cell interaction name
cluster_column cluster column with cell clusters
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
point_size_select
                  size of selected points
point_select_border_col
                  border color of selected points
point_select_border_stroke
                  stroke size of selected points
point_size_other
                  size of other points
point_other_border_col
                  border color of other points
\verb"point_other_border_stroke"
                  stroke size of other points
```

### **Details**

Description of parameters.

#### Value

ggplot

## **Examples**

```
cellProximityVisPlot_2D_ggplot(gobject)
```

```
cell Proximity VisPlot\_2D\_plotly \\ cell Proximity VisPlot\_2D\_plotly
```

## Description

Visualize 2D cell-cell interactions according to spatial coordinates in plotly mode

```
cellProximityVisPlot_2D_plotly(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  point_size_select = 2,
  point_size_other = 1,
  point_alpha_other = 0.3,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
 y_ticks = NULL,
)
```

## **Arguments**

gobject giotto object

interaction\_name

cell-cell interaction name

cluster\_column cluster column with cell clusters

sdimx x-axis dimension name (default = 'sdimx')

sdimy y-axis dimension name (default = 'sdimy')

cell\_color color for cells (see details)

cell\_color\_code

named vector with colors

color\_as\_factor

convert color column to factor

show\_other\_cells

decide if show cells not in network

show\_network show underlying spatial network

network\_color color of spatial network

spatial\_network\_name

name of spatial network to use

show\_grid show spatial grid

grid\_color color of spatial grid

spatial\_grid\_name

name of spatial grid to use

show\_legend show legend

point\_size\_select

size of selected points

coord\_fix\_ratio

fix ratio between x and y-axis

## **Details**

Description of parameters.

### Value

plotly

## **Examples**

cellProximityVisPlot\_2D\_plotly(gobject)

```
cell Proximity VisPlot\_3D\_plotly \\ cell Proximity VisPlot\_3D\_plotly
```

## **Description**

Visualize 3D cell-cell interactions according to spatial coordinates in plotly mode

## Usage

```
cellProximityVisPlot_3D_plotly(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  point_size_select = 2,
  point_size_other = 1,
  point_alpha_other = 0.5,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
)
```

```
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
                  show legend
show_legend
point_size_select
                  size of selected points
coord_fix_ratio
                  fix ratio between x and y-axis
```

## **Details**

Description of parameters.

## Value

plotly

## **Examples**

```
cellProximityVisPlot_3D_plotly(gobject)
```

 ${\tt change} {\tt GiottoInstructions}$ 

change Giot to Instructions

## Description

Function to change one or more instructions from giotto object

```
changeGiottoInstructions(
  gobject,
  params = NULL,
  new_values = NULL,
  return_gobject = TRUE
)
```

42 clusterCells

### **Arguments**

```
gobject giotto object
params parameter(s) to change
new_values new value(s) for parameter(s)
return_gobject (boolean) return giotto object
```

#### Value

named vector with giotto instructions

## **Examples**

changeGiottoInstructions()

clusterCells

clusterCells

### **Description**

cluster cells using a NN-network and community detection algorithms

```
clusterCells(
  gobject,
 cluster_method = c("leiden", "louvain_community", "louvain_multinet", "randomwalk",
    "sNNclust", "kmeans", "hierarchical"),
  name = "cluster_name",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  pyth_leid_resolution = 1,
  pyth_leid_weight_col = "weight",
 pyth_leid_part_type = c("RBConfigurationVertexPartition", "ModularityVertexPartition"),
  pyth_leid_init_memb = NULL,
  pyth_leid_iterations = 1000,
  pyth_louv_resolution = 1,
  pyth_louv_weight_col = NULL,
  python_louv_random = F,
  python_path = NULL,
  louvain_gamma = 1,
  louvain_omega = 1,
  walk_steps = 4,
  walk_clusters = 10,
  walk_weights = NA,
  sNNclust_k = 20,
  sNNclust_eps = 4,
  sNNclust_minPts = 16,
  borderPoints = TRUE,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
```

clusterCells 43

```
dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
     dim_reduction_name = "pca",
     dimensions_to_use = 1:10,
     distance_method = c("original", "pearson", "spearman", "euclidean", "maximum",
        "manhattan", "canberra", "binary", "minkowski"),
     km_centers = 10,
     km_iter_max = 100,
     km_nstart = 1000,
      km_algorithm = "Hartigan-Wong",
     hc_agglomeration_method = c("ward.D2", "ward.D", "single", "complete", "average",
        "mcquitty", "median", "centroid"),
     hc_k = 10,
     hc_h = NULL
      return_gobject = TRUE,
      set_seed = T,
      seed_number = 1234,
   )
Arguments
                    giotto object
   gobject
   cluster_method community cluster method to use
                    name for new clustering result
   name
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
                    name of NN network to use
   network_name
   pyth_leid_resolution
                    resolution for leiden
   pyth_leid_weight_col
                    column to use for weights
   pyth_leid_part_type
                    partition type to use
   pyth_leid_init_memb
                    initial membership
   pyth_leid_iterations
                    number of iterations
   pyth_louv_resolution
                    resolution for louvain
   pyth_louv_weight_col
                    python louvain param: weight column
   python_louv_random
                    python louvain param: random
   python_path
                    specify specific path to python if required
   louvain_gamma
                    louvain param: gamma or resolution
   louvain_omega
                    louvain param: omega
   walk_steps
                    randomwalk: number of steps
   walk_clusters
                    randomwalk: number of clusters
   walk_weights
                    randomwalk: weight column
```

44 clusterCells

sNNclust\_k SNNclust: k neighbors to use

sNNclust\_eps SNNclust: epsilon

sNNclust\_minPts

SNNclust: min points

borderPoints SNNclust: border points

expression\_values

expression values to use

 $genes\_to\_use = NULL,$ 

dim\_reduction\_to\_use

dimension reduction to use

 $\operatorname{dim\_reduction\_name}$ 

name of reduction 'pca',

dimensions\_to\_use

dimensions to use

distance\_method

distance method

 ${\tt km\_centers} \qquad \quad {\tt kmeans} \; {\tt centers} \\$ 

km\_iter\_max kmeans iterations

km\_nstart kmeans random starting points

km\_algorithm kmeans algorithm

hc\_agglomeration\_method

hierarchical clustering method

hc\_k hierachical number of clusters

hc\_h hierarchical tree cutoff

return\_gobject boolean: return giotto object (default = TRUE)

set\_seed set seed

seed\_number number for seed

... additional parameters

### **Details**

Description of different clustering methods.

### Value

giotto object appended with new cluster

## **Examples**

clusterCells(gobject)

combineMetadata 45

combineMetadata

combineMetadata

## **Description**

This function combines the cell metadata with spatial locations and enrichment results from createSpatialEnrich

## Usage

```
combineMetadata(gobject, spat_enr_names = NULL)
```

## **Arguments**

```
gobject Giotto object
spat_enr_names names of spatial enrichment results to include
```

### Value

Extended cell metadata in data.table format.

## **Examples**

```
combineMetadata(gobject)
```

```
convertEnsemblToGeneSymbol
```

convert Ensembl To Gene Symbol

## **Description**

This function convert ensembl gene IDs from a matrix to official gene symbols

### Usage

```
convertEnsemblToGeneSymbol(matrix, species = c("mouse", "human"))
```

## **Arguments**

matrix an expression matrix with ensembl gene IDs as rownames species species to use for gene symbol conversion

## **Details**

This function requires that the biomaRt library is installed

### Value

expression matrix with gene symbols as rownames

46 createGiottoInstructions

### **Examples**

```
convertEnsemblToGeneSymbol(matrix)
```

```
createGiottoInstructions
```

createGiottoInstructions

## Description

Function to set global instructions for giotto functions

## Usage

```
createGiottoInstructions(
  python_path = NULL,
  show_plot = NULL,
  return_plot = NULL,
  save_plot = NULL,
  save_dir = NULL,
  plot_format = NULL,
  dpi = NULL,
  units = NULL,
  height = NULL,
  width = NULL
```

## **Arguments**

```
python_path
                  path to python binary to use
show_plot
                  print plot to console, default = TRUE
return_plot
                  return plot as object, default = TRUE
                  automatically save plot, dafault = FALSE
save_plot
save_dir
                  path to directory where to save plots
                  resolution for raster images
dpi
height
                  height of plots
width
                  width of plots
```

## Value

named vector with giotto instructions

```
createGiottoInstructions()
```

createGiottoObject 47

## **Description**

Function to create a giotto object

### Usage

```
createGiottoObject(
  raw_exprs,
  spatial_locs = NULL,
  norm_expr = NULL,
  norm_scaled_expr = NULL,
  custom_expr = NULL,
  cell_metadata = NULL,
  gene_metadata = NULL,
  spatial_network = NULL,
  spatial_network_name = NULL,
  spatial_grid = NULL,
  spatial_grid_name = NULL,
  spatial_enrichment = NULL,
  spatial_enrichment_name = NULL,
  dimension_reduction = NULL,
  nn_network = NULL,
  offset_file = NULL,
  instructions = NULL
)
```

```
matrix with raw expression counts [required]
raw_exprs
spatial_locs
                  data.table or data.frame with coordinates for cell centroids
norm_expr
                  normalized expression values
norm_scaled_expr
                  scaled expression values
                  custom expression values
custom_expr
                  cell annotation metadata
cell_metadata
                  gene annotation metadata
gene_metadata
spatial_network
                  list of spatial network(s)
spatial_network_name
                  list of spatial network name(s)
spatial_grid
                  list of spatial grid(s)
spatial_grid_name
                  list of spatial grid name(s)
spatial_enrichment
                  list of spatial enrichment score(s) for each spatial region
```

48 createGiottoObject

```
spatial_enrichment_name
```

list of spatial enrichment name(s)

dimension\_reduction

list of dimension reduction(s)

nn\_network list of nearest neighbor network(s)

offset\_file file used to stitch fields together (optional)

instructions list of instructions or output result from createGiottoInstructions

#### **Details**

[Requirements] To create a giotto object you need to provide at least a matrix with genes as row names and cells as column names. To include spatial information about cells (or regions) you need to provide a data.table or data.frame with coordinates for all spatial dimensions. This can be 2D (x and y) or 3D (x, y, x). The row order for the cell coordinates should be the same as the column order for the provided expression data.

[Instructions] Additionally an instruction file, generated manually or with createGiottoInstructions can be provided to instructions, if not a default instruction file will be created for the Giotto object.

[Multiple fields] In case a dataset consists of multiple fields, like seqFISH+ for example, an offset file can be provided to stitch the different fields together. stitchFieldCoordinates can be used to generate such an offset file.

[**Processed data**] Processed count data, such as normalized data, can be provided using one of the different expression slots (norm\_expr, norm\_scaled\_expr, custom\_expr).

[Metadata] Cell and gene metadata can be provided using the cell and gene metadata slots. This data can also be added afterwards using the addGeneMetadata or addCellMetadata functions.

[Other information] Additional information can be provided through the appropriate slots:

- · spatial networks
- · spatial girds
- · spatial enrichments
- · dimensions reductions
- · nearest neighbours networks

### Value

giotto object

```
createGiottoObject(raw_exprs, spatial_locs)
```

createHeatmap\_DT 49

 ${\tt createHeatmap\_DT}$ 

createHeatmap\_DT

## **Description**

creates order for clusters

### Usage

```
createHeatmap_DT(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cluster_cor_method = "pearson",
  cluster_hclust_method = "ward.D",
  gene_order = c("custom", "correlation"),
  gene_custom_order = NULL,
  gene_cor_method = "pearson",
  gene_hclust_method = "complete"
)
```

## **Arguments**

```
gobject
                 giotto object
expression_values
                 expression values to use
genes
                 genes to use
cluster_column name of column to use for clusters
cluster_order
                 method to determine cluster order
cluster_custom_order
                 custom order for clusters
cluster_cor_method
                 method for cluster correlation
cluster_hclust_method
                 method for hierarchical clustering of clusters
                 method to determine gene order
gene_order
gene_custom_order
                 custom order for genes
gene_cor_method
                 method for gene correlation
gene_hclust_method
                 method for hierarchical clustering of genes
```

## **Details**

Creates input data.tables for plotHeatmap function.

50 createNearestNetwork

### Value

list

#### **Examples**

```
createHeatmap_DT(gobject)
```

createNearestNetwork createNearestNetwork

## **Description**

create a nearest neighbour network

# Usage

```
createNearestNetwork(
  gobject,
  type = c("sNN", "kNN"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  genes_to_use = NULL,
  expression_values = c("normalized", "scaled", "custom"),
  name = "sNN.pca",
  return_gobject = TRUE,
  k = 30,
  minimum_shared = 5,
  top_shared = 3,
  verbose = T,
  ...
)
```

```
gobject
                 giotto object
                 sNN or kNN
type
dim_reduction_to_use
                 dimension reduction method to use
dim_reduction_name
                 name of dimension reduction set to use
dimensions_to_use
                 number of dimensions to use as input
                 if dim_reduction_to_use = NULL, which genes to use
genes_to_use
expression_values
                 expression values to use
                 arbitrary name for NN network
name
return_gobject boolean: return giotto object (default = TRUE)
                 number of k neighbors to use
```

createSpatialEnrich 51

```
minimum_shared minimum shared neighbors
top_shared keep at ...
verbose be verbose
... additional parameters
```

### **Details**

Description of nearest neighbor network creation and filter steps.

#### Value

giotto object with updated NN network

## **Examples**

```
createNearestNetwork(gobject)
```

 $create Spatial Enrich \qquad \textit{create Spatial Enrich}$ 

## **Description**

Function to calculate gene signature enrichment scores per spatial position using a hypergeometric test.

### Usage

```
createSpatialEnrich(
  gobject,
  enrich_method = c("PAGE", "rank", "hypergeometric"),
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore"),
  name = "PAGE",
  return_gobject = TRUE
)
```

```
gobject Giotto object

enrich_method method for gene signature enrichment calculation

sign_matrix Matrix of signature genes for each cell type / process

expression_values

expression values to use

reverse_log_scale

reverse expression values from log scale

logbase log base to use if reverse_log_scale = TRUE
```

52 createSpatialGrid

### **Details**

For details see the individual functions:

```
PAGE: PAGEEnrichPAGE: rankEnrichPAGE: hyperGeometricEnrich
```

### Value

Giotto object or enrichment results if return\_gobject = FALSE

## **Examples**

```
createSpatialEnrich(gobject)
```

 $create {\tt Spatial Grid} \qquad \textit{create Spatial Grid}$ 

### **Description**

Create a spatial grid.

## Usage

```
createSpatialGrid(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  sdimz_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

createSpatialGrid\_2D 53

### **Details**

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

#### Value

giotto object with updated spatial grid slot

### **Examples**

```
createSpatialGrid(gobject)
```

```
createSpatialGrid\_2D createSpatialGrid\_2D
```

## **Description**

create a spatial grid for 2D spatial data.

## Usage

```
createSpatialGrid_2D(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

# Arguments

### **Details**

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

### Value

giotto object with updated spatial grid slot

```
createSpatialGrid_2D(gobject)
```

```
createSpatialGrid\_3D \quad \textit{createSpatialGrid\_3D}
```

## **Description**

Create a spatial grid for 3D spatial data.

## Usage

```
createSpatialGrid_3D(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  sdimz_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

## **Arguments**

```
gobject giotto object

sdimx_stepsize stepsize along the x-axis

sdimy_stepsize stepsize along the y-axis

sdimz_stepsize stepsize along the z-axis

minimum_padding

minimum padding on the edges

name name for spatial grid (default = 'spatial_grid')

return_gobject boolean: return giotto object (default = TRUE)
```

## **Details**

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

### Value

giotto object with updated spatial grid slot

```
createSpatialGrid_3D(gobject)
```

createSpatialNetwork 55

```
createSpatialNetwork createSpatialNetwork
```

### **Description**

Create a spatial network based on cell centroid physical distances.

## Usage

```
createSpatialNetwork(
  gobject,
  k = 4,
  dimensions = "all",
  maximum_distance = NULL,
  minimum_k = 0,
  name = "spatial_network",
  verbose = F,
  return_gobject = TRUE
)
```

### **Arguments**

gobject giotto object

k number of nearest neighbors based on physical distance

dimensions which spatial dimensions to use (default = all)

maximum\_distance

distance cuttof for nearest neighbors to consider

minimum\_k minimum nearest neighbours if maximum\_distance != NULL

name name for spatial network (default = 'spatial\_network')

verbose verbose

return\_gobject boolean: return giotto object (default = TRUE)

#### **Details**

Creates a spatial network connecting single-cells based on their physical distance to each other. Number of neighbors can be determined by k, maximum distance from each cell with or without setting a minimum k for each cell.

**dimensions:** default = 'all' which takes all possible dimensions. Alternatively you can provide a character vector that specififies the spatial dimensions to use, e.g. c("sdimx', "sdimy") or a numerical vector, e.g. 2:3

**maximum\_distance:** to create a network based on maximum distance only, you also need to set k to a very high value, e.g. k = 100

#### Value

giotto object with updated spatial network slot

```
createSpatialNetwork(gobject)
```

56 create\_average\_DT

```
create\_average\_detection\_DT \\ create\_average\_detection\_DT
```

## **Description**

calculates average gene detection for a cell metadata factor (e.g. cluster)

## Usage

```
create_average_detection_DT(
  gobject,
  meta_data_name,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0
)
```

### **Arguments**

## Value

data.table with average gene epression values for each factor

## Description

calculates average gene expression for a cell metadata factor (e.g. cluster)

## Usage

```
create_average_DT(
  gobject,
  meta_data_name,
  expression_values = c("normalized", "scaled", "custom")
)
```

```
gobject giotto object
meta_data_name name of metadata column to use
expression_values
 which expression values to use
```

### Value

data.table with average gene epression values for each factor

## Description

creates randomized cell ids within a selection of cell types

## Usage

```
create_cell_type_random_cell_IDs(
  gobject,
  cluster_column = "cell_types",
  needed_cell_types
)
```

## **Arguments**

```
gobject giotto object to use

cluster_column cluster column with cell type information

needed_cell_types

vector of cell type names for which a random id will be found
```

## **Details**

Details will follow.

## Value

list of randomly sampled cell ids with same cell type composition

```
create_cell_type_random_cell_IDs(gobject)
```

58 create\_dimObject

```
create_cluster_matrix create_cluster_matrix
```

## **Description**

creates aggregated matrix for a given clustering

## Usage

```
create_cluster_matrix(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  gene_subset = NULL
)
```

## **Examples**

```
create_cluster_matrix(gobject)
```

create\_dimObject

create\_dimObject

## **Description**

Creates an object that stores a dimension reduction output

## Usage

```
create_dimObject(
  name = "test",
  reduction_method = NULL,
  coordinates = NULL,
  misc = NULL
)
```

## **Arguments**

```
name arbitrary name for object
reduction_method
method used to reduce dimensions
coordinates accepts the coordinates after dimension reduction
misc any additional information will be added to this slot
```

### Value

number of distinct colors

decide\_cluster\_order 59

```
decide_cluster_order
```

## **Description**

creates order for clusters

## Usage

```
decide_cluster_order(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cor_method = "pearson",
  hclust_method = "ward.D"
)
```

## **Arguments**

```
gobject giotto object
expression_values
expression values to use
genes genes to use
cluster_column name of column to use for clusters
cluster_order method to determine cluster order
cluster_custom_order
custom order for clusters

cor_method method for correlation
hclust_method method for hierarchical clustering
```

## **Details**

Calculates order for clusters.

### Value

custom

```
decide_cluster_order(gobject)
```

60 detectSpatialPatterns

detectSpatialPatterns detectSpatialPatterns

## Description

Identify spatial patterns through PCA on average expression in a spatial grid.

# Usage

```
detectSpatialPatterns(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  spatial_grid_name = "spatial_grid",
  min_cells_per_grid = 4,
  scale_unit = F,
  ncp = 100,
  show_plot = T,
  PC_zscore = 1.5
)
```

## **Arguments**

### **Details**

Steps to identify spatial patterns:

- 1. average gene expression for cells within a grid, see createSpatialGrid
- 2. perform PCA on the average grid expression profiles
- 3. convert variance of principlal components (PCs) to z-scores and select PCs based on a z-score threshold

## Value

spatial pattern object 'spatPatObj'

```
detectSpatialPatterns(gobject)
```

dimCellPlot 61

dimCellPlot

dim Cell Plot

### **Description**

Visualize cells according to dimension reduction coordinates

```
dimCellPlot(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  cell_annotation_values,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show\_center\_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  legend_text = 8,
  axis_text = 8,
  axis_title = 8,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
```

62 dimCellPlot

```
default_save_name = "dimCellPlot"
Arguments
    gobject
                     giotto object
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    spat_enr_names names of spatial enrichment results to include
    cell_annotation_values
                     numeric cell annotation columns
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
                     size of center points
    label_size
                     size of labels
    label_fontface font of labels
                     column to use for alpha of the edges
    edge_alpha
                     size of point (cell)
    point_size
```

color of border around points

point\_border\_col

dimCellPlot2D 63

```
point_border_stroke
                  stroke size of border around points
                  show legend
show_legend
legend_text
                  size of legend text
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default\_save\_name
                  default save name for saving, don't change, change save_name in save_param
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
title
                  title for plot, defaults to cell_color parameter
```

### **Details**

Description of parameters. For 3D plots see dimCellPlot2D

### Value

ggplot

## **Examples**

```
dimCellPlot(gobject)
```

dimCellPlot2D

dimCellPlot2D

## **Description**

Visualize cells according to dimension reduction coordinates

```
dimCellPlot2D(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  spat_enr_names = NULL,
  cell_annotation_values,
  show_NN_network = F,
```

dimCellPlot2D

```
nn_network_to_use = "sNN",
     network_name = "sNN.pca",
     cell_color_gradient = c("blue", "white", "red"),
     gradient_midpoint = NULL,
     gradient_limits = NULL,
     select_cell_groups = NULL,
     select_cells = NULL,
      show_other_cells = T,
     other_cell_color = "lightgrey",
     other_point_size = 0.5,
     show_cluster_center = F,
     show_center_label = T,
     center_point_size = 4,
     center_point_border_col = "black",
     center_point_border_stroke = 0.1,
     label_size = 4,
     label_fontface = "bold",
     edge_alpha = NULL,
     point_size = 1,
     point_border_col = "black",
     point_border_stroke = 0.1,
     show_legend = T,
     legend_text = 8,
     axis_text = 8,
     axis_title = 8,
     cow_n_col = 2,
     cow_rel_h = 1,
     cow_rel_w = 1,
     cow_align = "h",
     show_plot = NA,
     return_plot = NA,
     save_plot = NA,
     save_param = list(),
     default_save_name = "dimCellPlot2D"
   )
Arguments
                   giotto object
   gobject
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
                    dimension to use on x-axis
   dim1_to_use
   dim2_to_use
                    dimension to use on y-axis
   spat_enr_names names of spatial enrichment results to include
   cell_annotation_values
                    numeric cell annotation columns
   show_NN_network
                    show underlying NN network
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
```

network\_name name of NN network to use, if show NN network = TRUE cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels edge\_alpha column to use for alpha of the edges size of point (cell) point\_size point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_legend show legend legend\_text size of legend text axis\_text size of axis text size of axis title axis\_title show\_plot show plot return\_plot return ggplot object save\_plot directly save the plot [boolean] list of saving parameters from all\_plots\_save\_function() save\_param default\_save\_name default save name for saving, don't change, change save\_name in save\_param cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors title for plot, defaults to cell\_color parameter title

66 dimGenePlot

### **Details**

Description of parameters. For 3D plots see dimPlot3D

### Value

ggplot

### **Examples**

```
dimCellPlot2D(gobject)
```

dimGenePlot

dimGenePlot

### **Description**

Visualize cells and gene expression according to dimension reduction coordinates

```
dimGenePlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = "red",
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dimGenePlot"
```

dimGenePlot 67

#### **Arguments**

gobject giotto object expression\_values gene expression values to use genes genes to show dim\_reduction\_to\_use dimension reduction to use dim\_reduction\_name dimension reduction name dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name column to use for alpha of the edges edge\_alpha scale\_alpha\_with\_expression scale expression with ggplot alpha parameter point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points size of point (cell) midpoint cow\_n\_col cowplot param: how many columns cowplot param: relative height cow\_rel\_h cow\_rel\_w cowplot param: relative width cow\_align cowplot param: how to align show\_legend show legend show\_plot show plots return\_plot return ggplot object save\_plot directly save the plot [boolean] save\_param list of saving parameters from all\_plots\_save\_function() default\_save\_name default save name for saving, don't change, change save\_name in save\_param parameters for cowplot::save\_plot()

### **Details**

Description of parameters.

### Value

ggplot

68 dimGenePlot2D

### See Also

```
dimGenePlot3D
```

### **Examples**

```
dimGenePlot(gobject)
```

dimGenePlot2D

dimGenePlot2D

## **Description**

Visualize cells and gene expression according to dimension reduction coordinates

```
dimGenePlot2D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = "red",
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dimGenePlot2D"
)
```

dimGenePlot2D 69

#### **Arguments**

gobject giotto object expression\_values gene expression values to use genes genes to show dim\_reduction\_to\_use dimension reduction to use dim\_reduction\_name dimension reduction name dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name column to use for alpha of the edges edge\_alpha scale\_alpha\_with\_expression scale expression with ggplot alpha parameter point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points size of point (cell) midpoint cow\_n\_col cowplot param: how many columns cowplot param: relative height cow\_rel\_h cow\_rel\_w cowplot param: relative width cow\_align cowplot param: how to align show\_legend show legend show\_plot show plots return\_plot return ggplot object save\_plot directly save the plot [boolean] save\_param list of saving parameters from all\_plots\_save\_function() default\_save\_name default save name for saving, don't change, change save\_name in save\_param parameters for cowplot::save\_plot()

### **Details**

Description of parameters.

### Value

ggplot

70 dimGenePlot3D

### See Also

```
dimGenePlot3D
```

### **Examples**

```
dimGenePlot2D(gobject)
```

dimGenePlot3D

dimGenePlot3D

## Description

Visualize cells and gene expression according to dimension reduction coordinates

```
dimGenePlot3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = 3,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  cluster_column = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  edge_alpha = NULL,
  point_size = 2,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dimGenePlot3D"
)
```

dimGenePlot3D 71

## **Arguments**

gobject giotto object

expression\_values

gene expression values to use

genes genes to show

dim\_reduction\_to\_use

dimension reduction to use

dim\_reduction\_name

dimension reduction name

dim1\_to\_use dimension to use on x-axis
dim2\_to\_use dimension to use on y-axis
dim3\_to\_use dimension to use on z-axis

show\_NN\_network

show underlying NN network

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

edge\_alpha column to use for alpha of the edges

point\_size size of point (cell)

show\_legend show\_plot show plots

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

... parameters for cowplot::save\_plot()

### **Details**

Description of parameters.

## Value

ggplot

## **Examples**

dimGenePlot3D(gobject)

72 dimPlot

dimPlot

dimPlot

### **Description**

Visualize cells according to dimension reduction coordinates

```
dimPlot(
  gobject,
  group_by = NULL,
  group_by_subset = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  legend_text = 8,
  axis_text = 8,
  axis_title = 8,
  title = NULL,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
```

dimPlot 73

```
cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dimPlot"
Arguments
    gobject
                     giotto object
    group_by_subset
                     subset the group_by factor column
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
                     dimension to use on y-axis
    dim2_to_use
    spat_enr_names names of spatial enrichment results to include
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
```

74 dimPlot

center\_point\_size

size of center points

label\_size size of labels label\_fontface font of labels

edge\_alpha column to use for alpha of the edges

point\_size size of point (cell)

point\_border\_col

color of border around points

point\_border\_stroke

stroke size of border around points

show\_legend show legend

legend\_text size of legend text
axis\_text size of axis text
axis\_title size of axis title

title title for plot, defaults to cell\_color parameter

cow\_n\_col cowplot param: how many columns

cow\_rel\_h cowplot param: relative height
cow\_rel\_w cowplot param: relative width
cow\_align cowplot param: how to align

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

groub\_by create multiple plots based on cell annotation column

#### **Details**

Description of parameters, see dimPlot2D. For 3D plots see dimPlot3D

#### Value

ggplot

## **Examples**

dimPlot(gobject)

dimPlot2D 75

dimPlot2D

dimPlot2D

#### **Description**

Visualize cells according to dimension reduction coordinates

```
dimPlot2D(
  gobject,
  group_by = NULL,
  group_by_subset = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  title = NULL,
  show_legend = T,
  legend_text = 8,
  axis_text = 8,
  axis_title = 8,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
```

76 dimPlot2D

```
cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dimPlot2D"
Arguments
    gobject
                     giotto object
    group_by_subset
                     subset the group_by factor column
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
                     dimension to use on y-axis
    dim2_to_use
    spat_enr_names names of spatial enrichment results to include
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
```

dimPlot2D 77

center\_point\_size

size of center points

label\_size size of labels label\_fontface font of labels

edge\_alpha column to use for alpha of the edges

point\_size size of point (cell)

point\_border\_col

color of border around points

point\_border\_stroke

stroke size of border around points

title title for plot, defaults to cell\_color parameter

show\_legend show legend

legend\_text size of legend text
axis\_text size of axis text
axis\_title size of axis title

cow\_n\_col cowplot param: how many columns

cow\_rel\_h cowplot param: relative height
cow\_rel\_w cowplot param: relative width

cow\_align cowplot param: how to align

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

groub\_by create multiple plots based on cell annotation column

#### **Details**

Description of parameters. For 3D plots see dimPlot3D

#### Value

ggplot

## **Examples**

dimPlot2D(gobject)

78 dimPlot2D\_single

dimPlot2D\_single

dimPlot2D\_single

## **Description**

Visualize cells according to dimension reduction coordinates

```
dimPlot2D_single(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  title = NULL,
  show_legend = T,
  legend_text = 8,
  axis_text = 8,
  axis_title = 8,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dimPlot2D_single"
```

dimPlot2D\_single 79

)

```
gobject
                  giotto object
dim_reduction_to_use
                  dimension reduction to use
dim_reduction_name
                  dimension reduction name
dim1_to_use
                  dimension to use on x-axis
                  dimension to use on y-axis
dim2_to_use
spat_enr_names names of spatial enrichment results to include
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
network_name
                  name of NN network to use, if show_NN_network = TRUE
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
```

80 dimPlot3D

```
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
title
                  title for plot, defaults to cell_color parameter
show_legend
                  show legend
                  size of legend text
legend_text
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

#### **Details**

Description of parameters. For 3D plots see dimPlot3D

#### Value

ggplot

## **Examples**

```
dimPlot2D_single(gobject)
```

dimPlot3D

dimPlot3D

## **Description**

Visualize cells according to dimension reduction coordinates

```
dimPlot3D(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = 3,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 2,
```

dimPlot3D 81

```
show_NN_network = F,
      nn_network_to_use = "sNN",
      network_name = "sNN.pca",
      color_as_factor = T,
      cell_color = NULL,
      cell_color_code = NULL,
      show_cluster_center = F,
      show_center_label = T,
      center_point_size = 4,
      label_size = 4,
      edge_alpha = NULL,
      point_size = 3,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dim3D"
Arguments
                     giotto object
   gobject
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
   dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
    show_other_cells
                     display not selected cells
   other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
    color_as_factor
                     convert color column to factor
                     color for cells (see details)
    cell_color
    cell_color_code
                     named vector with colors
    show_cluster_center
                     plot center of selected clusters
```

82 direction\_test\_CPG

```
show_center_label
```

plot label of selected clusters

center\_point\_size

size of center points

label\_size size of labels

edge\_alpha column to use for alpha of the edges

point\_size size of point (cell)

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

show\_legend show legend

#### **Details**

Description of parameters.

#### Value

plotly

# **Examples**

dimPlot3D(gobject)

direction\_test\_CPG

direction\_test\_CPG

## **Description**

shows direction of change

# Usage

```
direction_test(x, min_fdr = 0.05)
```

## **Examples**

```
direction_test_CPG()
```

doHclust 83

doHclust doHclust

#### **Description**

cluster cells using hierarchical clustering algorithm

```
doHclust(
      gobject,
      expression_values = c("normalized", "scaled", "custom"),
      genes_to_use = NULL,
      dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
      dim_reduction_name = "pca",
      dimensions_to_use = 1:10,
      distance_method = c("pearson", "spearman", "original", "euclidean", "maximum",
      "manhattan", "canberra", "binary", "minkowski"),
agglomeration_method = c("ward.D2", "ward.D", "single", "complete", "average",
         "mcquitty", "median", "centroid"),
      k = 10,
      h = NULL
      name = "hclust",
      return_gobject = TRUE,
      set_seed = T,
      seed_number = 1234
    )
Arguments
    gobject
                     giotto object
    expression_values
                     expression values to use
                     subset of genes to use
    genes_to_use
    dim_reduction_to_use
```

```
dimension reduction to use
dim_reduction_name
                  dimensions reduction name
dimensions_to_use
                  dimensions to use
distance_method
                  distance method
{\it agglomeration\_method}
                  agglomeration method for hclust
                  number of final clusters
k
h
                  cut hierarchical tree at height = h
                  name for hierarchical clustering
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                  set seed
                  number for seed
seed_number
                  additional parameters
```

84 doHMRF

#### **Details**

Description on how to use Kmeans clustering method.

#### Value

giotto object appended with new cluster

## **Examples**

```
doHclust(gobject)
```

doHMRF

doHMRF

## **Description**

Run HMRF

# Usage

```
doHMRF(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  spatial_network_name = "spatial_network",
  spatial_genes = NULL,
  spatial_dimensions = c("sdimx", "sdimy", "sdimz"),
  dim_reduction_to_use = NULL,
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "test",
  k = 10,
  betas = c(0, 2, 50),
  tolerance = 1e-10,
  zscore = c("none", "rowcol", "colrow"),
  numinit = 100,
  python_path = NULL,
  output_folder = NULL,
  overwrite_output = TRUE
)
```

doKmeans 85

```
dim_reduction_to_use
                 use another dimension reduction set as input
dim_reduction_name
                 name of dimension reduction set to use
dimensions_to_use
                 number of dimensions to use as input
                 name of HMRF run
name
                 number of HMRF domains
k
betas
                 betas to test for
tolerance
                 tolerance
zscore
                 zscore
                 number of initializations
numinit
                 python path to use
python_path
                 output folder to save results
output_folder
overwrite_output
                 overwrite output folder
```

## **Details**

Description of HMRF parameters ...

#### Value

Creates a directory with results that can be viewed with viewHMRFresults

#### **Examples**

```
doHMRF(gobject)
```

doKmeans doKmeans

## **Description**

cluster cells using kmeans algorithm

86 doKmeans

```
nstart = 1000,
algorithm = "Hartigan-Wong",
name = "kmeans",
return_gobject = TRUE,
set_seed = T,
seed_number = 1234
)
```

## **Arguments**

gobject giotto object

expression\_values

expression values to use

genes\_to\_use subset of genes to use

dim\_reduction\_to\_use

dimension reduction to use

dim\_reduction\_name

dimensions reduction name

dimensions\_to\_use

dimensions to use

 ${\tt distance\_method}$ 

distance method

centers number of final clusters

iter\_max kmeans maximum iterations

nstart kmeans nstart

algorithm kmeans algorithm

name name for kmeans clustering

return\_gobject | boolean: return giotto object (default = TRUE)

set\_seed set seed

seed\_number number for seed

... additional parameters

# **Details**

Description on how to use Kmeans clustering method.

# Value

giotto object appended with new cluster

# **Examples**

```
doKmeans(gobject)
```

doLeidenCluster 87

doLeidenCluster doLeidenCluster

# Description

cluster cells using a NN-network and the Leiden community detection algorithm

# Usage

```
doLeidenCluster(
  gobject,
  name = "leiden_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = "weight",
  partition_type = c("RBConfigurationVertexPartition", "ModularityVertexPartition"),
  init_membership = NULL,
  n_iterations = 1000,
  return_gobject = TRUE,
  set_seed = T,
  seed_number = 1234,
  ...
)
```

# **Arguments**

```
giotto object
gobject
                 name for cluster
name
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
python_path
                 specify specific path to python if required
resolution
                 resolution
weight_col
                 weight column
partition_type partition type to use
init_membership
                 initial membership of cells
n_iterations
                 number of interations
return_gobject boolean: return giotto object (default = TRUE)
set\_seed
                 set seed
seed_number
                 number for seed
                 additional parameters
```

## **Details**

Description of Leiden clustering method.

88 doLeidenSubCluster

#### Value

giotto object appended with new cluster

#### **Examples**

```
doLeidenCluster(gobject)
```

doLeidenSubCluster

doLeidenSubCluster

#### **Description**

subcluster cells using a NN-network and the Leiden algorithm

# Usage

```
doLeidenSubCluster(
  gobject,
  name = "sub_pleiden_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 0.5,
  n_{iterations} = 500,
  python_path = NULL,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T,
)
```

```
gobject giotto object

name name for new clustering result

cluster_column cluster column to subcluster

selected_clusters

only do subclustering on these clusters

hvg_param parameters for calculateHVG

hvg_min_perc_cells

threshold for detection in min percentage of cells
```

doLouvainCluster 89

```
hvg_mean_expr_det
                 threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                 forces all genes to be HVG and to be used as input for PCA
min_nr_of_hvg minimum number of HVG, or all genes will be used as input for PCA
                 parameters for runPCA
pca_param
                 parameters for parameters for createNearestNetwork
nn_param
                 number of k for createNearestNetwork
k_neighbors
resolution
                 resolution of Leiden clustering
                 number of iterations
n_iterations
                 specify specific path to python if required
python_path
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
return_gobject boolean: return giotto object (default = TRUE)
verbose
                 verbose
                 additional parameters
```

#### **Details**

Description of Leiden clustering method.

#### Value

giotto object appended with new cluster

## **Examples**

```
doLeidenSubCluster(gobject)
```

doLouvainCluster doLouvainCluster

## **Description**

cluster cells using a NN-network and the Louvain algorithm.

```
doLouvainCluster(
  gobject,
  version = c("community", "multinet"),
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = NULL,
```

```
gamma = 1,
omega = 1,
louv_random = F,
return_gobject = TRUE,
set_seed = F,
seed_number = 1234,
...
)
```

## **Arguments**

giotto object gobject implemented version of Louvain clustering to use version name for cluster name nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use network\_name specify specific path to python if required python\_path resolution resolution gamma gamma omega omega return\_gobject boolean: return giotto object (default = TRUE) set seed set\_seed number for seed seed\_number additional parameters

#### **Details**

Louvain clustering using the community or multinet implementation of the louvain clustering algorithm.

#### Value

giotto object appended with new cluster

## **Examples**

```
doLouvainCluster(gobject)
```

```
\label{lower} do Louvain {\it Cluster\_community} \\ do Louvain {\it Cluster\_community} \\
```

# **Description**

cluster cells using a NN-network and the Louvain algorithm from the community module in Python

#### Usage

```
doLouvainCluster_community(
   gobject,
   name = "louvain_clus",
   nn_network_to_use = "sNN",
   network_name = "sNN.pca",
   python_path = NULL,
   resolution = 1,
   weight_col = NULL,
   louv_random = F,
   return_gobject = TRUE,
   set_seed = F,
   seed_number = 1234,
   ...
)
```

## **Arguments**

```
giotto object
gobject
                 name for cluster
name
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
                 specify specific path to python if required
python_path
resolution
                 resolution
                 weight column
weight_col
louv_random
                 random
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                 set seed
seed_number
                 number for seed
                 additional parameters
```

#### **Details**

Description of Leiden clustering method.

# Value

giotto object appended with new cluster

## **Examples**

```
doLouvainCluster_community(gobject)
```

```
\label{lower} do Louvain Cluster\_multinet \\ do Louvain Cluster\_multinet
```

#### **Description**

cluster cells using a NN-network and the Louvain algorithm from the multinet package in R.

# Usage

```
doLouvainCluster_multinet(
  gobject,
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  weight_col = NULL,
  gamma = 1,
  omega = 1,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

# Arguments

```
gobject
                 giotto object
                 name for cluster
name
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use
                 gamma
gamma
                 omega
omega
return_gobject boolean: return giotto object (default = TRUE)
                 set seed
set\_seed
seed_number
                 number for seed
                 additional parameters
python_path
                 specify specific path to python if required
```

# **Details**

See louvain algorithm from the multinet package in R.

#### Value

giotto object appended with new cluster

# **Examples**

```
doLouvainCluster_multinet(gobject)
```

doLouvainSubCluster 93

 ${\tt doLouvainSubCluster} \qquad {\tt doLouvainSubCluster}$ 

#### **Description**

subcluster cells using a NN-network and the Louvain algorithm

## Usage

```
doLouvainSubCluster(
  gobject,
  name = "sub_louvain_clus",
  version = c("community", "multinet"),
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 0.5,
  gamma = 1,
  omega = 1,
  python_path = NULL,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T,
)
```

```
giotto object
gobject
                  name for new clustering result
name
                  version of Louvain algorithm to use
version
cluster_column cluster column to subcluster
selected_clusters
                  only do subclustering on these clusters
                  parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                  threshold for detection in min percentage of cells
hvg_mean_expr_det
                  threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                  forces all genes to be HVG and to be used as input for PCA
```

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for createNearestNetwork

k\_neighbors number of k for createNearestNetwork resolution resolution for community algorithm

gamma gamma omega omega

python\_path specify specific path to python if required

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

network\_name name of NN network to use

return\_gobject boolean: return giotto object (default = TRUE)

verbose verbose

... additional parameters

#### **Details**

Description of Louvain clustering method.

#### Value

giotto object appended with new cluster

## **Examples**

```
doLouvainSubCluster(gobject)
```

```
\label{lower} do Louvain SubCluster\_community \\ do Louvain SubCluster\_community
```

## **Description**

subcluster cells using a NN-network and the Louvain community detection algorithm

```
pca_param = list(expression_values = "normalized", scale_unit = T),
nn_param = list(dimensions_to_use = 1:20),
k_neighbors = 10,
resolution = 0.5,
python_path = NULL,
nn_network_to_use = "sNN",
network_name = "sNN.pca",
return_gobject = TRUE,
verbose = T,
...
)
```

#### **Arguments**

giotto object gobject name name for new clustering result cluster\_column cluster column to subcluster selected\_clusters only do subclustering on these clusters parameters for calculateHVG hvg\_param hvg\_min\_perc\_cells threshold for detection in min percentage of cells hvg\_mean\_expr\_det threshold for mean expression level in cells with detection use\_all\_genes\_as\_hvg forces all genes to be HVG and to be used as input for PCA min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA parameters for runPCA pca\_param parameters for parameters for createNearestNetwork nn\_param k\_neighbors number of k for createNearestNetwork resolution resolution python\_path specify specific path to python if required nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use network\_name return\_gobject boolean: return giotto object (default = TRUE)

#### **Details**

verbose

Description of Leiden clustering method.

verbose

additional parameters

#### Value

giotto object appended with new cluster

# **Examples**

```
doLouvainSubCluster_community(gobject)
```

```
\label{lower_multinet} do Louvain SubCluster\_multinet \\ do Louvain SubCluster\_multinet
```

# Description

subcluster cells using a NN-network and the Louvain multinet detection algorithm

#### Usage

```
doLouvainSubCluster_multinet(
  gobject,
  name = "sub_louvain_mult_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  gamma = 1,
  omega = 1,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T,
)
```

```
giotto object
gobject
                  name for new clustering result
name
cluster_column cluster column to subcluster
selected clusters
                  only do subclustering on these clusters
                  parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                  threshold for detection in min percentage of cells
hvg_mean_expr_det
                  threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                  forces all genes to be HVG and to be used as input for PCA
                  minimum number of HVG, or all genes will be used as input for PCA
min_nr_of_hvg
                  parameters for runPCA
pca_param
```

doRandomWalkCluster 97

```
parameters for parameters for createNearestNetwork
nn_param
k_neighbors
                 number of k for createNearestNetwork
gamma
                 gamma
omega
                 omega
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
return_gobject boolean: return giotto object (default = TRUE)
                 verbose
verbose
                 additional parameters
```

specify specific path to python if required

#### **Details**

python\_path

Description of Louvain clustering method.

#### Value

giotto object appended with new cluster

## **Examples**

```
doLouvainSubCluster_multinet(gobject)
```

 $do Random Walk Cluster \qquad do Random Walk Cluster$ 

## **Description**

Cluster cells using a random walk approach.

```
doRandomWalkCluster(
  gobject,
  name = "random_walk_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  walk_steps = 4,
  walk_clusters = 10,
  walk_weights = NA,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

98 doSNNCluster

## **Arguments**

```
gobject
                 giotto object
name
                 name for cluster
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use
walk_steps
                 number of walking steps
                 number of final clusters
walk_clusters
walk_weights
                 cluster column defining the walk weights
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                 set seed
seed_number
                 number for seed
                 additional parameters
```

#### **Details**

See random walk algorithm from the igraph package in R.

#### Value

giotto object appended with new cluster

## **Examples**

doRandomWalkCluster(gobject)

doSNNCluster

doSNNCluster

## **Description**

Cluster cells using a SNN cluster approach.

```
doSNNCluster(
  gobject,
  name = "sNN_clus",
  nn_network_to_use = "kNN",
  network_name = "kNN.pca",
  k = 20,
  eps = 4,
  minPts = 16,
  borderPoints = TRUE,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

dt\_to\_matrix 99

# **Arguments**

gobject giotto object name name for cluster

nn\_network\_to\_use

type of NN network to use (only works on kNN)

 $network\_name \qquad name \ of \ kNN \ network \ to \ use$ 

k Neighborhood size for nearest neighbor sparsification to create the shared NN

graph.

eps Two objects are only reachable from each other if they share at least eps nearest

neighbors.

minPts minimum number of points that share at least eps nearest neighbors for a point

to be considered a core points.

borderPoints should borderPoints be assigned to clusters like in DBSCAN?

return\_gobject boolean: return giotto object (default = TRUE)

set\_seed set seed

seed\_number number for seed
... additional parameters

#### **Details**

See sNNclust algorithm from dbscan package

#### Value

giotto object appended with new cluster

## **Examples**

doSNNCluster(gobject)

#### **Description**

converts data.table to matrix

## Usage

```
dt_to_matrix(x)
```

#### **Examples**

 $dt_{to_matrix(x)}$ 

100 exportGiottoViewer

exportGiottoViewer

exportGiottoViewer

## **Description**

compute highly variable genes

#### Usage

```
exportGiottoViewer(
  gobject,
  output_directory = NULL,
  spat_enr_names = NULL,
  factor_annotations,
  numeric_annotations,
  dim_reductions,
  dim_reduction_names,
  expression_values = c("scaled", "normalized", "custom"),
  dim_red_rounding = NULL,
  dim_red_rescale = c(-20, 20),
  expression_rounding = 2,
  overwrite_dir = T,
  verbose = T
)
```

```
gobject
                  giotto object
output_directory
                  directory where to save the files
spat_enr_names spatial enrichment results to include for annotations
factor_annotations
                  giotto cell annotations to view as factor
numeric_annotations
                  giotto cell annotations to view as numeric
dim_reductions high level dimension reductions to view
dim_reduction_names
                  specific dimension reduction names
expression_values
                  expression values to use in Viewer
dim_red_rounding
                  numerical indicating how to round the coordinates
dim_red_rescale
                  numericals to rescale the coordinates
expression_rounding
                  numerical indicating how to round the expression data
                  overwrite files in the directory if it already existed
overwrite_dir
verbose
                  be verbose
```

#### **Details**

Giotto Viewer expects the results from Giotto Analyzer in a specific format, which is provided by this function. To include enrichment results from createSpatialEnrich include the provided spatial enrichment name (default PAGE or rank) and add the gene signature names (.e.g cell types) to the numeric annotations parameter.

#### Value

writes the necessary output to use in Giotto Viewer

# **Examples**

```
exportGiottoViewer(gobject)
```

```
expr Only Cell Cell communication Scores \\ expr Only Cell Cell communication Scores
```

#### **Description**

Cell-Cell communication scores based on expression only

## Usage

```
exprOnlyCellCellcommunicationScores(
  gobject,
  cluster_column = "cell_types",
  random_iter = 100,
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  verbose = T
)
```

# Arguments

```
gobject giotto object to use

cluster_column cluster column with cell type information

random_iter number of iterations

gene_set_1 first specific gene set from gene pairs

gene_set_2 second specific gene set from gene pairs

log2FC_addendum

addendum to add when calculating log2FC

verbose verbose
```

## Details

Details will follow.

102 extractNearestNetwork

#### Value

Cell-Cell communication scores for gene pairs based on expression only

#### **Examples**

```
exprOnlyCellCellcommunicationScores(gobject)
```

```
extended_gini_fun
     extended_gini_fun
```

# Description

calculate gini coefficient on a minimum length vector

# Usage

```
extended_gini_fun(x, weights = rep(1, length = length(x)), minimum_length = 16)
```

#### Value

gini coefficient

```
extractNearestNetwork extractNearestNetwork
```

# **Description**

Extracts a NN-network from a Giotto object as an igraph object

## Usage

```
extractNearestNetwork(
  gobject,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca"
)
```

# Arguments

```
\begin{array}{ccc} \mbox{gobject} & \mbox{giotto object} \\ \mbox{nn_network\_to\_use} & \mbox{kNN or sNN} \\ \mbox{network\_name} & \mbox{name of NN network to be used} \end{array}
```

## Value

igraph object

# **Examples**

```
extractNearestNetwork(gobject)
```

fDataDT

fDataDT

fDataDT

# Description

show gene metadata

# Usage

```
fDataDT(gobject)
```

# **Arguments**

gobject

giotto object

## Value

data.table with gene metadata

# **Examples**

```
pDataDT(gobject)
```

filterCombinations

filterCombinations

# Description

Shows how many genes and cells are lost with combinations of thresholds.

```
filterCombinations(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_thresholds = c(1, 2),
  gene_det_in_min_cells = c(5, 50),
  min_det_genes_per_cell = c(200, 400),
  scale_x_axis = "identity",
  x_axis_offset = 0,
  scale_y_axis = "identity",
  y_axis_offset = 0,
  show_plot = TRUE
)
```

104 filterCPGscores

#### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
expression_thresholds
                  all thresholds to consider a gene expressed
gene_det_in_min_cells
                  minimum number of cells that should express a gene to consider that gene fur-
min_det_genes_per_cell
                  minimum number of expressed genes per cell to consider that cell further
                  ggplot transformation for x-axis (e.g. log2)
scale_x_axis
x_axis_offset x-axis offset to be used together with the scaling transformation
scale_y_axis
                  ggplot transformation for y-axis (e.g. log2)
y_axis_offset y-axis offset to be used together with the scaling transformation
show_plot
                  show plot
```

#### Details

Creates a scatterplot that visualizes the number of genes and cells that are lost with a specific combination of a gene and cell threshold given an arbitrary cutoff to call a gene expressed. This function can be used to make an informed decision at the filtering step with filterGiotto.

#### Value

list of data.table and ggplot object

## **Examples**

```
filterCombinations(gobject)
```

filter CPG scores

filterCPGscores

#### **Description**

visualize Cell Proximity Gene enrichment scores

```
filterCPGscores(
   CPGscore,
   min_cells = 5,
   min_fdr = 0.05,
   min_spat_diff = 0.2,
   min_log2_fc = 0.5,
   keep_int_duplicates = TRUE,
   direction = c("both", "up", "down")
```

filterDistributions 105

#### **Arguments**

#### **Details**

This function filters the output from getCellProximityGeneScores based on false-discovery rate, minimum absolute difference, minimum log fold-change and direction of change.

## Value

Gene to gene scores in data.table format

## **Examples**

```
filterCPGscores(CPGscore)
```

```
filterDistributions filterDistributions
```

# **Description**

show gene or cell distribution after filtering on expression threshold

```
filterDistributions(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_threshold = 1,
  detection = c("genes", "cells"),
  plot_type = c("histogram", "violin"),
  nr_bins = 30,
  fill_color = "lightblue",
  scale_axis = "identity",
  axis_offset = 0,
  show_plot = TRUE
)
```

106 filterGiotto

# **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
expression\_threshold
                  threshold to consider a gene expressed
detection
                  consider genes or cells
                  type of plot
plot_type
nr_bins
                  number of bins for histogram plot
fill_color
                  fill color for plots
scale_axis
                  ggplot transformation for axis (e.g. log2)
                  offset to be used together with the scaling transformation
axis_offset
show_plot
                  show plot
```

#### Value

ggplot object

## **Examples**

filterDistributions(gobject)

filterGiotto filterGiotto

## **Description**

filter Giotto object based on expression threshold

# Usage

```
filterGiotto(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_threshold = 1,
  gene_det_in_min_cells = 100,
  min_det_genes_per_cell = 100,
  verbose = F
)
```

```
gobject giotto object
expression_values
expression values to use
expression_threshold
threshold to consider a gene expressed
```

findGiniMarkers 107

#### **Details**

The function filterCombinations can be used to explore the effect of different parameter values.

#### Value

giotto object

# **Examples**

filterGiotto(gobject)

findGiniMarkers

findGiniMarkers

#### **Description**

Identify marker genes for selected clusters based on gini detection and expression scores.

# Usage

```
findGiniMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank_score = 1
)
```

```
gobject giotto object
expression_values
gene expression values to use
cluster_column clusters to use
subset_clusters
selection of clusters to compare
group_1 group 1 cluster IDs from cluster_column for pairwise comparison
group_2 group 2 cluster IDs from cluster_column for pairwise comparison
```

```
min_expr_gini_score
filter on minimum gini coefficient for expression
min_det_gini_score
filter minimum gini coefficient for detection
detection_threshold
detection threshold for gene expression
rank_score
rank scores to include
```

#### **Details**

Description of parameters.

#### Value

data.table with marker genes

## **Examples**

```
findGiniMarkers(gobject)
```

## **Description**

Identify marker genes for all clusters based on gini detection and expression scores.

# Usage

```
findGiniMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  min_genes = 10,
  verbose = TRUE
)
```

```
gobject giotto object
expression_values
gene expression values to use
cluster_column clusters to use
subset_clusters
selection of clusters to compare
```

findMarkers 109

```
min_expr_gini_score
filter on minimum gini coefficient on expression

min_det_gini_score
filter on minimum gini coefficient on detection

detection_threshold
detection threshold for gene expression

min_genes minimum genes to keep per cluster, overrides pval and logFC

verbose be verbose
```

#### **Details**

Description of parameters.

#### Value

data.table with marker genes

### **Examples**

```
findGiniMarkers_one_vs_all(gobject)
```

findMarkers findMarkers

# Description

Identify marker genes for selected clusters.

```
findMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  method = c("scran", "gini", "mast"),
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank\_score = 1,
  group_1_name = NULL,
  group_2_name = NULL,
  adjust_columns = NULL,
)
```

#### **Arguments**

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
method
                  method to use to detect differentially expressed genes
subset_clusters
                  selection of clusters to compare
                  group 1 cluster IDs from cluster_column for pairwise comparison
group_1
                  group 2 cluster IDs from cluster_column for pairwise comparison
group_2
min_expr_gini_score
                  gini: filter on minimum gini coefficient for expression
min_det_gini_score
                  gini: filter minimum gini coefficient for detection
{\tt detection\_threshold}
                  gini: detection threshold for gene expression
rank_score
                  gini: rank scores to include
group_1_name
                  mast: custom name for group_1 clusters
                  mast: custom name for group_2 clusters
group_2_name
adjust_columns mast: column in pDataDT to adjust for (e.g. detection rate)
                  additional parameters for the findMarkers function in scran or zlm function in
                  MAST
```

#### **Details**

Wrapper for findScranMarkers, findGiniMarkers and FindMastMarkers.

#### Value

data.table with marker genes

# **Examples**

```
findMarkers(gobject)
```

```
findMarkers_one_vs_all
```

findMarkers\_one\_vs\_all

# Description

Identify marker genes for all clusters.

#### Usage

```
findMarkers_one_vs_all(
 gobject,
  expression_values = c("normalized", "scaled", "custom"),
 cluster_column,
  subset_clusters = NULL,
 method = c("scran", "gini", "mast"),
 pval = 0.01,
 logFC = 0.5,
 min\_genes = 10,
 min_expr_gini_score = 0.5,
 min_det_gini_score = 0.5,
 detection_threshold = 0,
 rank_score = 1,
 adjust_columns = NULL,
 verbose = TRUE,
)
```

### **Arguments**

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster column clusters to use
subset_clusters
                  selection of clusters to compare
method
                  method to use to detect differentially expressed genes
                  scran & mast: filter on minimal p-value
pval
logFC
                  scan & mast: filter on logFC
                  minimum genes to keep per cluster, overrides pval and logFC
min_genes
min_expr_gini_score
                  gini: filter on minimum gini coefficient for expression
min\_det\_gini\_score
                  gini: filter minimum gini coefficient for detection
detection\_threshold
                  gini: detection threshold for gene expression
                  gini: rank scores to include
rank_score
adjust_columns mast: column in pDataDT to adjust for (e.g. detection rate)
verbose
                  be verbose
                  additional parameters for the findMarkers function in scran or zlm function in
                  MAST
```

#### **Details**

Wrapper for findScranMarkers\_one\_vs\_all, findGiniMarkers\_one\_vs\_all and FindMastMarkers\_one\_vs\_all.

#### Value

data.table with marker genes

112 findMastMarkers

#### **Examples**

```
findMarkers_one_vs_all(gobject)
```

findMastMarkers

findMastMarkers

#### **Description**

Identify marker genes for selected clusters based on the MAST package.

### Usage

```
findMastMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  group_1 = NULL,
  group_1_name = NULL,
  group_2 = NULL,
  group_2_name = NULL,
  adjust_columns = NULL,
  ...
)
```

#### **Arguments**

```
gobject giotto object
expression_values
gene expression values to use

cluster_column clusters to use
group_1 group 1 cluster IDs from cluster_column for pairwise comparison
group_1_name custom name for group_1 clusters
group_2 group 2 cluster IDs from cluster_column for pairwise comparison
group_2_name custom name for group_2 clusters
adjust_columns column in pDataDT to adjust for (e.g. detection rate)
... additional parameters for the zlm function in MAST
```

### **Details**

This is a minimal convenience wrapper around the MAST functions to detect differentially expressed genes.

#### Value

data.table with marker genes

```
findMastMarkers(gobject)
```

```
\label{lem:cone_vs_all} find \textit{MastMarkers\_one\_vs\_all} \\ find \textit{MastMarkers\_one\_vs\_all}
```

### Description

Identify marker genes for all clusters based on the MAST package.

# Usage

```
findMastMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  adjust_columns = NULL,
  pval = 0.001,
  logFC = 1,
  min_genes = 10,
  verbose = TRUE,
  ...
)
```

#### **Arguments**

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
subset_clusters
                  selection of clusters to compare
adjust_columns column in pDataDT to adjust for (e.g. detection rate)
pval
                  filter on minimal p-value
logFC
                  filter on logFC
                  minimum genes to keep per cluster, overrides pval and logFC
min_genes
verbose
                  be verbose
                  additional parameters for the zlm function in MAST
. . .
```

### Details

This is a minimal convenience wrapper around the MAST functions to detect differentially expressed genes.

#### Value

data.table with marker genes

```
findMastMarkers_one_vs_all(gobject)
```

114 findScranMarkers

findScranMarkers

findScranMarkers

### **Description**

Identify marker genes for selected clusters based on scran's implementation of findMarkers.

### Usage

```
findScranMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  ...
)
```

# Arguments

```
gobject giotto object
expression_values
gene expression values to use

cluster_column clusters to use
subset_clusters
selection of clusters to compare
group_1 group 1 cluster IDs from cluster_column for pairwise comparison
group_2 group 2 cluster IDs from cluster_column for pairwise comparison
additional parameters for the findMarkers function in scran
```

#### **Details**

This is a minimal convenience wrapper around the findMarkers function from the scran package.

#### Value

data.table with marker genes

```
findScranMarkers(gobject)
```

```
find Scran Markers\_one\_vs\_all \\ find Scran Markers\_one\_vs\_all
```

# Description

Identify marker genes for all clusters in a one vs all manner based on scran's implementation of findMarkers.

#### Usage

```
findScranMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  pval = 0.01,
  logFC = 0.5,
  min_genes = 10,
  verbose = TRUE,
  ...
)
```

### **Arguments**

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
subset_clusters
                  subset of clusters to use
pval
                  filter on minimal p-value
logFC
                  filter on logFC
                  minimum genes to keep per cluster, overrides pval and logFC
min_genes
verbose
                  be verbose
                  additional parameters for the findMarkers function in scran
. . .
```

### Details

This is a minimal convenience wrapper around the findMarkers function from the scran package.

### Value

data.table with marker genes

```
findScranMarkers_one_vs_all(gobject)
```

find\_grid\_y

find\_grid\_2D

 $find\_grid\_2D$ 

# Description

find grid location in 2D

# Usage

```
find_grid_2D(grid_DT, x_loc, y_loc)
```

 $find\_grid\_3D$ 

find\_grid\_3D

# Description

find grid location in 3D

# Usage

```
find_grid_3D(grid_DT, x_loc, y_loc, z_loc)
```

find\_grid\_x

find\_grid\_x

# Description

find grid location on x-axis

# Usage

```
find_grid_x(grid_DT, x_loc)
```

find\_grid\_y

 $find\_grid\_y$ 

# Description

find grid location on y-axis

```
find_grid_y(grid_DT, y_loc)
```

find\_grid\_z

find\_grid\_z

 $find\_grid\_z$ 

# Description

find grid location on z-axis

# Usage

```
find_grid_z(grid_DT, z_loc)
```

fish\_function

fish\_function

# Description

perform fisher exact test

# Usage

```
fish_function(x_to, x_from)
```

fish\_function2

 $fish\_function2$ 

# Description

perform fisher exact test

```
fish_function2(A, B, C, D)
```

118 FSV\_show

FSV\_show

FSV\_show

# Description

Visualize spatial varible genes caculated by spatial\_DE

# Usage

```
FSV_show(
  results,
  ms_results = NULL,
  size = c(4, 2, 1),
  color = c("blue", "green", "red"),
  sig_alpha = 0.5,
  unsig_alpha = 0.5
)
```

# **Arguments**

results results caculated by spatial\_DE

ms\_results ms\_results caculated by spatial\_DE

size indicate different levels of qval

color indicate different SV features

sig\_alpha transparency of significant genes

unsig\_alpha transparency of unsignificant genes

# **Details**

Description of parameters.

# Value

nothing

```
FSV_show(results)
```

GenePattern\_show 119

GenePattern\_show

GenePattern\_show

# Description

Visualize genes distribution patterns calculated by spatial\_AEH

### Usage

```
GenePattern_show(
  gobject = NULL,
  AEH_results = NULL,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_alpha = 1,
  low_color = "blue",
  mid_color = "white",
  high_color = "red",
  midpoint = 0
)
```

# **Arguments**

gobject giotto object results from spatial\_AEH AEH\_results sdimx x axis of spatial locus sdimy y axis of spatial locus point\_size size of points to indicate cells transparency of points to indicate cells point\_alpha low\_color color to indicate low score level color to indicate middle score level mid\_color high\_color color to indicate high score level point to set mid\_color midpoint

# Details

Description of parameters.

### Value

nothing

```
GenePattern_show(gobject,AEH_results)
```

120 general\_save\_function

```
general_save_function general_save_function
```

# Description

Function to automatically save plots to directory of interest

### Usage

```
general_save_function(
  gobject,
  plot_object,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  default_save_name = "giotto_plot",
  save_format = c("png", "tiff", "pdf", "svg"),
  show_saved_plot = F,
  base_width = NULL,
  base_height = NULL,
  base_aspect_ratio = NULL,
  units = NULL,
  dpi = NULL,
  ...
)
```

# Arguments

```
gobject
                  giotto object
                  non-ggplot object to plot
plot_object
save_dir
                  directory to save to
save_folder
                  folder in save_dir to save to
                  name of plot
save_name
save_format
                  format (e.g. png, tiff, pdf, ...)
show_saved_plot
                  load & display the saved plot
base_width
                  width
base_height
                  height
base_aspect_ratio
                  aspect ratio
units
                  units
dpi
                  Plot resolution
```

```
general_save_function(gobject)
```

get10Xmatrix 121

get10Xmatrix

get10Xmatrix

### **Description**

This function creates an expression matrix from a 10X structured folder

### Usage

```
get10Xmatrix(path_to_data)
```

### **Arguments**

```
path_to_data path to the 10X folder
```

#### **Details**

A typical 10X folder is named raw\_feature\_bc\_matrix or raw\_feature\_bc\_matrix. It has 3 files:

- barcodes
- features.tsv.gz
- matrix.mtx.gz

#### Value

expression matrix from 10X

### **Examples**

```
get10Xmatrix(10Xmatrix)
```

```
{\it get Cell Proximity Gene Scores} \\ {\it get Cell Proximity Gene Scores}
```

### **Description**

Compute cell-cell interaction enrichment (observed vs expected)

```
getCellProximityGeneScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "louvain_clus.1",
  selected_genes = NULL,
  expression_values = c("normalized", "scaled", "custom"),
  do_diff_test = TRUE,
  diff_test = c("t.test", "wilcox"),
  minimum_unique_cells = NA,
```

```
fold_change_addendum = 0.1,
  in_two_directions = TRUE,
  exclude_selected_cells_from_test = F,
  verbose = T
)
```

### **Arguments**

```
gobject
                  giotto object
spatial_network_name
                  name of spatial network to use
cluster_column name of column to use for clusters
selected_genes selection of genes to perform calculations for
expression_values
                  expression values to use
do_diff_test
                  perform differential test
diff_test
                  which differential expression test
minimum_unique_cells
                  minimum number of cells needed to proceed
fold_change_addendum
                  constant to add when calculating log2 fold-change
in_two_directions
                  shows enrichment in both directions: cell1-cell2, cell2-cell1
exclude_selected_cells_from_test
                  exclude certain cells from test
verbose
                  verbose
```

#### **Details**

Function to calculate if genes are differentially expressed in cell types when they interact (according to physical proximity) with other cell types. The results data.table contains the following columns:

- genes: All or selected list of tested genes
- cell\_expr\_1: average gene expression in cell type 1 from unified\_int cell-cell interaction
- cell\_expr\_2: average gene expression in cell type 2 from unified\_int cell-cell interaction
- comb\_expr: combined average gene expression in cell type 1 and 2 from unified\_int cell-cell interaction
- all\_cell\_expr\_1: average gene expression for all cells from cell type 1
- all\_cell\_expr\_2: average gene expression for all cells from cell type 2
- all\_comb\_expr: combined average gene expression for all cells from cell type 1 and 2
- pval\_1: p-value from test between interacting cells and all cells from cell type 1
- pval\_2: p-value from test between interacting cells and all cells from cell type 2
- cell\_type\_1: first cell type of cell-cell interaction
- cell\_type\_2: second cell type of cell-cell interaction
- interaction: the cell-cell interaction, based on physical proximity
- nr\_1: number of cell type 1 in the unified cell-cell interaction
- nr\_2: number of cell type 2 in the unified cell-cell interaction

getClusterSimilarity 123

- all\_nr\_1: number of all cell type 1 in the whole dataset
- all\_nr\_2: number of all cell type 2 in the whole dataset
- diff\_spat: difference between comb\_expr and all\_comb\_expr
- diff\_spat\_1: difference between cell\_expr\_1 and all\_cell\_expr\_1
- diff\_spat\_2: difference between cell\_expr\_1 and all\_cell\_expr\_1
- log2fc\_spat\_1: fold-change of diff\_spat\_1
- log2fc\_spat\_2: fold-change of diff\_spat\_2
- log2fc\_spat: fold-change of diff\_spat
- type\_int: type of interaction
- unified\_int: interaction with alphabetically sorted cell type 1 and cell type 2
- unif\_int\_rank: 1 or 2
- fdr\_1: fdr from test between interacting cells and all cells from cell type 1
- fdr\_2: fdr from test between interacting cells and all cells from cell type 2

#### Value

Cell Proximity Gene scores (CPGscores) in data.table format

#### **Examples**

```
getCellProximityGeneScores(gobject)
```

```
getClusterSimilarity
```

### **Description**

Creates data.table with pairwise correlation scores between each cluster.

### Usage

```
getClusterSimilarity(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman")
)
```

### **Arguments**

```
gobject giotto object
expression_values
expression values to use
cluster_column name of column to use for clusters
cor correlation score to calculate distance
```

124 getDendrogramSplits

#### **Details**

Creates data.table with pairwise correlation scores between each cluster and the group size (# of cells) for each cluster. This information can be used together with mergeClusters to combine very similar or small clusters into bigger clusters.

### Value

data.table

### **Examples**

```
getClusterSimilarity(gobject)
```

```
getDendrogramSplits getDendrogramSplits
```

# Description

Split dendrogram at each node and keep the leave (label) information..

### Usage

```
getDendrogramSplits(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  h = NULL,
  h_color = "red",
  show_dend = TRUE,
  verbose = TRUE
)
```

### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
distance
                  distance method to use for hierarchical clustering
h
                  height of horizontal lines to plot
h_color
                  color of horizontal lines
                  show dendrogram
show_dend
verbose
                  be verbose
```

getDistinctColors 125

#### **Details**

Creates a data.table with three columns and each row represents a node in the dendrogram. For each node the height of the node is given together with the two subdendrograms. This information can be used to determine in a hierarchical manner differentially expressed marker genes at each node.

### Value

data.table object

### **Examples**

```
getDendrogramSplits(gobject)
```

getDistinctColors

getDistinctColors

### **Description**

Returns a number of distint colors based on the RGB scale

#### Usage

```
getDistinctColors(n)
```

### **Arguments**

n

number of colors wanted

#### Value

number of distinct colors

```
{\tt getGeneToGeneScores}
```

getGeneToGeneScores

### **Description**

Compute gene-gene enrichment scores.

```
getGeneToGeneScores(
   CPGscore,
   selected_genes = NULL,
   specific_genes_1 = NULL,
   specific_genes_2 = NULL,
   min_cells = 5,
   min_fdr = 0.05,
   min_spat_diff = 0.2,
   min_log2_fc = 0.5,
```

```
direction = c("both", "up", "down"),
fold_change_addendum = 0.1,
  verbose = TRUE
)
```

### **Arguments**

```
CPGscore
                  CPGscore, output from getCellProximityGeneScores()
selected_genes select subset of genes
specific_genes_1
                  specific source genes (see details)
specific_genes_2
                  specific target genes (see details)
                  min number of cells threshold
min_cells
min_spat_diff
                  spatial difference threshold
                  log2 fold-change threshold
min_log2_fc
direction
                  up or downregulation or both
fold_change_addendum
                  constant to add when calculating log2 fold-change
verbose
                  verbose
min_pval
                  p-value threshold
```

#### **Details**

Give more details ...

#### Value

Gene to gene scores in data.table format

# Examples

```
getGeneToGeneScores(CPGscore)
```

```
{\it get\_cell\_to\_cell\_sorted\_name\_conversion} \\ {\it get\_cell\_to\_cell\_sorted\_name\_conversion}
```

# Description

creates unified cell-cell interaction names

# Usage

```
get_cell_to_cell_sorted_name_conversion(all_cell_types)
```

```
get_cell_to_cell_sorted_name_conversion()
```

```
{\it get\_interaction\_gene\_enrichment} \\ {\it get\_interaction\_gene\_enrichment}
```

### **Description**

Computes gene enrichment between all interactions

# Usage

```
get_interaction_gene_enrichment(
  spatial_network,
  unified_int_col = "unified_int",
  source_col = "source_clus",
  source_IDs = "from",
  neighb_col = "neighb_clus",
  neighb_IDs = "to",
  expression_matrix,
  cell_annotation,
  annotation_ID = "uniq_ID",
  cell_type_col,
  do_diff_test = T,
  diff_test = c("t.test", "wilcox"),
  minimum_unique_cells = NA,
  exclude_selected_cells_from_test = T,
  verbose = T
)
```

### **Examples**

```
get_interaction_gene_enrichment()
```

```
{\tt get\_specific\_interaction\_gene\_enrichment} \\ {\tt get\_specific\_interaction\_gene\_enrichment}
```

### **Description**

Computes gene enrichment between specified interaction

```
get_specific_interaction_gene_enrichment(
  sub_spatial_network,
  source_col = "source_clus",
  source_IDs = "from",
  neighb_col = "neighb_clus",
  neighb_IDs = "to",
  expression_matrix,
```

128 ggplot\_save\_function

```
interaction_name = "to_specify",
  cell_annotation,
  annotation_ID = "uniq_ID",
  cell_type_col,
  do_diff_test = T,
  diff_test = c("t.test", "wilcox"),
  minimum_unique_cells = NA,
  exclude_selected_cells_from_test = T
)
```

### **Examples**

```
get_specific_interaction_gene_enrichment()
```

```
ggplot_save_function ggplot_save_function
```

### **Description**

Function to automatically save plots to directory of interest

# Usage

```
ggplot_save_function(
  gobject,
  plot_object,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  default_save_name = "giotto_plot",
  save_format = NULL,
  show_saved_plot = F,
  ncol = 1,
  nrow = 1,
  scale = 1,
  base_width = NULL,
  base_height = NULL,
  base_aspect_ratio = NULL,
  units = NULL,
  dpi = NULL,
  limitsize = TRUE,
)
```

### **Arguments**

```
gobject giotto object
plot_object ggplot object to plot
save_dir directory to save to
save_folder folder in save_dir to save to
```

giotto-class 129

save\_name name of plot

save\_format (e.g. png, tiff, pdf, ...)

show\_saved\_plot

load & display the saved plot

ncol number of columns nrow number of rows

scale scale
base\_width width
base\_height height
base\_aspect\_ratio

aspect ratio

units units

dpi Plot resolution

limitsize When TRUE (the default), ggsave will not save images larger than 50x50 inches,

to prevent the common error of specifying dimensions in pixels.

#### See Also

```
cowplot::save_plot
```

### **Examples**

ggplot\_save\_function(gobject)

giotto-class S4 giotto Class

### **Description**

Framework of giotto object to store and work with spatial expression data

#### **Slots**

```
raw_exprs raw expression counts

norm_expr normalized expression counts

norm_scaled_expr normalized and scaled expression counts

custom_expr custom normalized counts

spatial_locs spatial location coordinates for cells

cell_metadata metadata for cells

gene_metadata metadata for genes

cell_ID unique cell IDs

gene_ID unique gene IDs

spatial_network spatial network in data.table/data.frame format

spatial_grid spatial grid in data.table/data.frame format

dimension_reduction slot to save dimension reduction coordinates
```

nn\_network nearest neighbor network in igraph format parameters slot to save parameters that have been used instructions slot for global function instructions offset\_file offset file used to stitch together image fields OS\_platform Operating System to run Giotto analysis on

hyperGeometricEnrich hyperGeometricEnrich

### **Description**

Function to calculate gene signature enrichment scores per spatial position using a hypergeometric test.

# Usage

```
hyperGeometricEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

#### Arguments

# Details

The enrichment score is calculated based on the p-value from the hypergeometric test, -log10(p-value).

#### Value

data.table with enrichment results

```
hyperGeometricEnrich(gobject)
```

iterCluster 131

iterCluster

iterCluster

### **Description**

cluster cells iteratively

#### Usage

```
iterCluster(
 gobject,
 cluster_method = c("leiden", "louvain_community", "louvain_multinet"),
 nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 20,
 resolution = 1,
  gamma = 1,
 omega = 1,
 python_path = NULL,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 name = "iter_clus",
 return_gobject = TRUE,
)
```

### **Arguments**

```
giotto object
gobject
cluster_method clustering algorithm to use
nr_rounds
                 number of iterative rounds
                 parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                 threshold for detection in min percentage of cells
hvg_mean_expr_det
                 threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                 forces all genes to be HVG and to be used as input for PCA
min_nr_of_hvg minimum number of HVG, or all genes will be used as input for PCA
                 parameters for runPCA
pca_param
                 parameters for parameters for runPCA
nn_param
```

iterLeidenCluster

```
k_neighbors
                 k for nn-network
resolution
                 resolution
gamma
                 gamma
omega
                 omega
                 python path to use for Leiden clustering
python_path
nn_network_to_use
                 NN network to use
                 NN network name
network_name
                 name of clustering
name
return_gobject boolean: return giotto object (default = TRUE)
                 additional parameters
```

#### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

### **Examples**

```
iterCluster(gobject)
```

iterLeidenCluster iterLeidenCluster

# **Description**

cluster cells iteratively

```
iterLeidenCluster(
 gobject,
 name = "iter_clus",
 nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
   = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 20,
 resolution = 1,
 n_{iterations} = 1000,
 python_path = NULL,
```

iterLeidenCluster 133

```
nn_network_to_use = "sNN",
network_name = "sNN.pca",
return_gobject = TRUE,
...
)
```

#### **Arguments**

gobject giotto object

name of clustering

nr\_rounds number of iterative rounds
hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

hvg\_mean\_expr\_det

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for runPCA

 $k\_neighbors$  k for nn-network

resolution resolution for Leiden clustering

n\_iterations number of iterations for Leiden clustering

python\_path python path to use for Leiden clustering

nn\_network\_to\_use

NN network to use

network\_name NN network name

return\_gobject boolean: return giotto object (default = TRUE)

... additional parameters

### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

```
iterLeidenCluster(gobject)
```

134 iterLouvainCluster

iterLouvainCluster

*iterLouvainCluster* 

### **Description**

cluster cells iteratively

#### Usage

```
iterLouvainCluster(
 gobject,
 version = c("community", "multinet"),
 nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
   = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 20,
 resolution = 1,
  gamma = 1,
 omega = 1,
 python_path = NULL,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 name = "iter_clus",
 return_gobject = TRUE,
)
```

### **Arguments**

```
gobject
                  giotto object
version
                  louvain clustering algorithm to use
nr_rounds
                  number of iterative rounds
                  parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                  threshold for detection in min percentage of cells
hvg_mean_expr_det
                  threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                  forces all genes to be HVG and to be used as input for PCA
min_nr_of_hvg minimum number of HVG, or all genes will be used as input for PCA
                  parameters for runPCA
pca_param
                  parameters for parameters for runPCA
nn_param
```

```
k_neighbors
                 k for nn-network
resolution
                 resolution
                 gamma
gamma
omega
                 omega
                 python path to use for Leiden clustering
python_path
nn_network_to_use
                 NN network to use
network_name
                 NN network name
                 name of clustering
name
return_gobject boolean: return giotto object (default = TRUE)
                 additional parameters
```

#### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

### **Examples**

```
iterLouvainCluster(gobject)
```

```
iter Louvain Cluster\_community\\iter Louvain Cluster\_community
```

# Description

cluster cells iteratively

```
network_name = "sNN.pca",
name = "iter_clus",
return_gobject = TRUE,
...
)
```

### **Arguments**

gobject giotto object

nr\_rounds number of iterative rounds
hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

 $hvg\_mean\_expr\_det$ 

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for runPCA

 $k\_neighbors$  k for nn-network

resolution resolution for Leiden clustering

python\_path python path to use for Leiden clustering

nn\_network\_to\_use

NN network to use

network\_name NN network name name of clustering

return\_gobject boolean: return giotto object (default = TRUE)

... additional parameters

#### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

### **Examples**

iterLouvainCluster\_community(gobject)

```
iter Louvain Cluster\_multinet
```

iterLouvainCluster\_multinet

### **Description**

cluster cells iteratively

#### Usage

```
iterLouvainCluster_multinet(
 gobject,
 nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
   = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 20,
  gamma = 1,
  omega = 1,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 name = "iter_clus",
 return_gobject = TRUE,
)
```

# Arguments

```
gobject
                 giotto object
nr_rounds
                 number of iterative rounds
hvg_param
                 parameters for calculateHVG
hvg_min_perc_cells
                 threshold for detection in min percentage of cells
hvg_mean_expr_det
                 threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                 forces all genes to be HVG and to be used as input for PCA
                 minimum number of HVG, or all genes will be used as input for PCA
min_nr_of_hvg
                 parameters for runPCA
pca_param
                 parameters for parameters for runPCA
nn_param
                 k for nn-network
k_neighbors
                 gamma
gamma
omega
                 omega
```

138 loadHMRF

```
nn_network_to_use
```

NN network to use

 $\begin{array}{ll} \mbox{network\_name} & NN \mbox{ network name} \\ \mbox{name} & \mbox{name of clustering} \end{array}$ 

return\_gobject boolean: return giotto object (default = TRUE)

... additional parameters

python\_path python path to use for Leiden clustering

### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

# **Examples**

iterLouvainCluster\_multinet(gobject)

kmeans\_binarize

kmeans\_binarize

# Description

create binarized scores using kmeans

# Usage

```
kmeans_binarize(x, nstart = 3, iter.max = 10)
```

loadHMRF

loadHMRF

# Description

load previous HMRF

```
loadHMRF(
  name_used = "test",
  output_folder_used,
  k_used = 10,
  betas_used,
  python_path_used
```

makeSignMatrixPAGE 139

### **Arguments**

 $name\_used \hspace{1.5cm} name\ of\ HMRF\ that\ was\ run$ 

output\_folder\_used

output folder that was used

k\_used number of HMRF domains that was tested

betas\_used betas that were tested

python\_path\_used

python path that was used

# **Details**

Description of HMRF parameters ...

#### Value

reloads a previous ran HMRF from doHRMF

# **Examples**

loadHMRF(gobject)

makeSignMatrixPAGE mak

makeSignMatrixPAGE

# Description

Function to convert list of signature genes (e.g. for cell types or processes) into a binary matrix format that can be used with the PAGE enrichment option.

# Usage

```
makeSignMatrixPAGE(sign_names, sign_list)
```

### **Arguments**

sign\_names vector with names for each provided gene signature

sign\_list list of genes (signature)

#### Value

matrix

#### See Also

**PAGEEnrich** 

### **Examples**

 ${\tt makeSignMatrixPAGE()}$ 

make Sign Matrix Rank

makeSignMatrixRank

# Description

Function to convert a single-cell count matrix and a corresponding single-cell cluster vector into a rank matrix that can be used with the Rank enrichment option.

### Usage

```
makeSignMatrixRank(sc_matrix, sc_cluster_ids, gobject = NULL)
```

### **Arguments**

```
sign_names vector with names for each provided gene signature sign_list list of genes (signature)
```

### Value

matrix

### See Also

rankEnrich

### **Examples**

```
makeSignMatrixRank()
```

```
make_simulated_network
```

make\_simulated\_network

# Description

Simulate random network.

# Usage

```
make_simulated_network(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column,
  number_of_simulations = 100
)
```

```
make_simulated_network(gobject)
```

mergeClusters 141

mergeClusters

mergeClusters

#### **Description**

Merge selected clusters based on pairwise correlation scores and size of cluster.

### Usage

```
mergeClusters(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  new_cluster_name = "merged_cluster",
  min_cor_score = 0.8,
  max_group_size = 20,
  force_min_group_size = 10,
  return_gobject = TRUE,
  verbose = TRUE
)
```

#### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
new_cluster_name
                  new name for merged clusters
min_cor_score min correlation score to merge pairwise clusters
max_group_size max cluster size that can be merged
force_min_group_size
                  size of clusters that will be merged with their most similar neighbor(s)
return_gobject return giotto object
verbose
                  be verbose
```

# **Details**

Merge selected clusters based on pairwise correlation scores and size of cluster. To avoid large clusters to merge the max\_group\_size can be lowered. Small clusters can be forcibly merged with their most similar pairwise cluster by adjusting the force\_min\_group\_size parameter. Clusters smaller than this value will be merged independent on the provided min\_cor\_score value.

A giotto object is returned by default, if FALSE then the merging vector will be returned.

#### Value

Giotto object

nnDT\_to\_kNN

# **Examples**

```
mergeClusters(gobject)
```

mygini\_fun

mygini\_fun

# Description

calculate gini coefficient

# Usage

```
mygini_fun(x, weights = rep(1, length(x)))
```

# Value

gini coefficient

nnDT\_to\_kNN

nnDT\_to\_kNN

# Description

Convert a nearest network data.table to a kNN object

# Usage

```
nnDT_to_kNN(nnDT)
```

# Arguments

nnDT

nearest neighbor network in data.table format

# Value

kNN object

node\_clusters 143

node\_clusters

node\_clusters

# Description

Merge selected clusters based on pairwise correlation scores and size of cluster.

# Usage

```
node_clusters(hclus_obj, verbose = TRUE)
```

# **Arguments**

hclus\_obj hclus object verbose be verbose

### Value

list of splitted dendrogram nodes from high to low node height

# **Examples**

```
node_clusters(hclus_obj)
```

normalizeGiotto

normalizeGiotto

# Description

normalize and/or scale expresion values of Giotto object

```
normalizeGiotto(
  gobject,
  norm_methods = c("standard", "osmFISH"),
  library_size_norm = TRUE,
  scalefactor = 6000,
  log_norm = TRUE,
  logbase = 2,
  scale_genes = T,
  scale_cells = T,
  scale_order = c("first_genes", "first_cells"),
  verbose = F
)
```

OR\_function2

#### **Arguments**

gobject giotto object

norm\_methods normalization method to use

library\_size\_norm

normalize cells by library size

scalefactor scale factor to use after library size normalization

log\_norm transform values to log-scale

logbase log base to use to log normalize expression values

scale\_genes z-score genes over all cells scale\_cells z-score cells over all genes scale\_order order to scale genes and cells

verbose be verbose

#### **Details**

Currently there are two 'methods' to normalize your raw counts data.

A. The standard method follows the standard protocol which can be adjusted using the provided parameters and follows the following order:

- 1. Data normalization for total library size and scaling by a custom scale-factor.
- 2. Log transformation of data.
- 3. Z-scoring of data by genes and/or cells.
- B. The normalization method as provided by the osmFISH paper is also implemented:
  - 1. First normalize genes, for each gene divide the counts by the total gene count and multiply by the total number of genes.
  - 2. Next normalize cells, for each cell divide the normalized gene counts by the total counts per cell and multiply by the total number of cells.

This data will be saved in the Giotto slot for custom expression.

#### Value

giotto object

### **Examples**

normalizeGiotto(gobject)

OR\_function2 OR\_function2

#### **Description**

calculate odds-ratio

```
OR_function2(A, B, C, D)
```

PAGEEnrich 145

PAGEEnrich PAGEEnrich

# **Description**

Function to calculate gene signature enrichment scores per spatial position using PAGE.

### Usage

```
PAGEEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

# **Arguments**

```
gobject Giotto object

sign_matrix Matrix of signature genes for each cell type / process

expression_values

expression values to use

reverse_log_scale

reverse expression values from log scale

logbase log base to use if reverse_log_scale = TRUE

output_enrichment

how to return enrichment output
```

### **Details**

The enrichment Z score is calculated by using method (PAGE) from Kim SY et al., BMC bioinformatics, 2005 as  $Z=((Sm\ \ mu)*m^(1/2))/delta$ . For each gene in each spot, mu is the fold change values versus the mean expression and delta is the standard deviation. Sm is the mean fold change value of a specific marker gene set and m is the size of a given marker gene set.

# Value

data.table with enrichment results

# Examples

```
PAGEEnrich(gobject)
```

plotCPGscores

pDataDT

pDataDT

# Description

show cell metadata

# Usage

```
pDataDT(gobject)
```

# Arguments

gobject

giotto object

# Value

data.table with cell metadata

# **Examples**

```
pDataDT(gobject)
```

plotCPGscores

plotCPGscores

# Description

Create heatmap from cell-cell proximity scores

# Usage

```
plotCPGscores(
   CPGscores,
   selected_interactions = NULL,
   selected_genes = NULL,
   detail_plot = T,
   simple_plot = F,
   simple_plot_facet = c("interaction", "genes"),
   facet_scales = "fixed",
   facet_ncol = length(selected_genes),
   facet_nrow = length(selected_interactions),
   show_plot = F
)
```

plotGTGscores 147

# **Arguments**

```
CPGscores
                  CPGscores, output from getCellProximityGeneScores()
selected_interactions
                  interactions to show
selected_genes genes to show
detail_plot
                  show detailed info in both interacting cell types
                  show a simplified plot
simple_plot
simple_plot_facet
                  facet on interactions or genes with simple plot
                  ggplot facet scales paramter
facet_scales
facet_ncol
                  ggplot facet ncol parameter
facet_nrow
                  ggplot facet nrow parameter
show_plot
                  show plot
```

### **Details**

Give more details ...

### Value

ggplot barplot

# **Examples**

```
plotCPGscores(CPGscores)
```

plotGTGscores

plotGTGscores

# **Description**

Create heatmap from cell-cell proximity scores

# Usage

```
plotGTGscores(
  gobject,
  GTGscore,
  selected_interactions = NULL,
  selected_gene_to_gene = NULL,
  detail_plot = T,
  simple_plot = F,
  simple_plot_facet = c("interaction", "genes"),
  facet_scales = "fixed",
  facet_ncol = length(selected_gene_to_gene),
  facet_nrow = length(selected_interactions),
  colors = c("blue", "red"),
  show_plot = NA,
  return_plot = NA,
```

148 plotGTGscores

```
save_plot = NA,
save_param = list(),
default_save_name = "plotGTGscores"
)
```

# Arguments

gobject giotto object

GTGscore GTGscore, output from getGeneToGeneScores()

selected\_interactions

interactions to show

detail\_plot show detailed info in both interacting cell types

simple\_plot show a simplified plot

simple\_plot\_facet

facet on interactions or genes with simple plot

facet\_scales ggplot facet scales paramter
facet\_ncol ggplot facet ncol parameter
facet\_nrow ggplot facet nrow parameter

colors vector with 2 colors to represent respectively all and selected cells

show\_plot show plots

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

selected\_genes genes to show

### **Details**

Give more details ...

# Value

ggplot barplot

# **Examples**

```
plotGTGscores(GTGscore)
```

plotHeatmap 149

plotHeatmap

plotHeatmap

### **Description**

creates order for clusters

### Usage

```
plotHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cluster_color_code = NULL,
  cluster_cor_method = "pearson",
  cluster_hclust_method = "ward.D"
  gene_order = c("custom", "correlation"),
  gene_custom_order = NULL,
  gene_cor_method = "pearson",
  gene_hclust_method = "complete",
  show_values = c("rescaled", "z-scaled", "original"),
  size_vertical_lines = 1.1,
  gradient_colors = c("blue", "yellow", "red"),
  gene_label_selection = NULL,
  axis_text_y_size = NULL,
  legend_nrows = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotHeatmap"
)
```

```
gobject giotto object
expression_values
expression values to use
genes genes to use
cluster_column name of column to use for clusters
cluster_order method to determine cluster order
cluster_custom_order
custom order for clusters
cluster_color_code
color code for clusters
cluster_cor_method
method for cluster correlation
```

150 plotly\_axis\_scale\_2D

cluster\_hclust\_method

method for hierarchical clustering of clusters

gene\_order method to determine gene order

gene\_custom\_order

custom order for genes

gene\_cor\_method

method for gene correlation

gene\_hclust\_method

method for hierarchical clustering of genes

show\_values which values to show on heatmap

size\_vertical\_lines

sizes for vertical lines

gradient\_colors

colors for heatmap gradient

gene\_label\_selection

subset of genes to show on y-axis

axis\_text\_y\_size

size for y-axis text

legend\_nrows number of rows for the cluster legend

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

# **Details**

Creates heatmap for genes and clusters.

### Value

ggplot

# **Examples**

plotHeatmap(gobject)

plotly\_axis\_scale\_2D plotly\_axis\_scale\_2D

# Description

adjust the axis scale in 3D plotly plot

plotly\_axis\_scale\_3D 151

### Usage

```
plotly_axis_scale_2D(
  cell_locations,
  sdimx = NULL,
  sdimy = NULL,
  mode = c("cube", "real", "custom"),
  custom_ratio = NULL
)
```

# Arguments

```
cell_locations spatial_loc in giotto object sdimx x axis of cell spatial location sdimy y axis of cell spatial location mode axis adjustment mode custom_ratio set the ratio artificially
```

### Value

edges in spatial grid as data.table()

# **Examples**

```
plotly_axis_scale_2D(gobject)
```

```
plotly_axis_scale_3D plotly_axis_scale_3D
```

# Description

adjust the axis scale in 3D plotly plot

### Usage

```
plotly_axis_scale_3D(
  cell_locations,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  mode = c("cube", "real", "custom"),
  custom_ratio = NULL
)
```

```
cell_locations spatial_loc in giotto object
sdimx x axis of cell spatial location
sdimy y axis of cell spatial location
sdimz z axis of cell spatial location
mode axis adjustment mode
custom_ratio set the ratio artificially
```

plotly\_network

### Value

```
edges in spatial grid as data.table()
```

# **Examples**

```
plotly_axis_scale_3D(gobject)
```

plotly\_grid

plotly\_grid

# Description

```
provide grid segment to draw in plot_ly()
```

# Usage

```
plotly_grid(
   spatial_grid,
   x_start = "x_start",
   y_start = "y_start",
   x_end = "x_end",
   y_end = "y_end"
)
```

# **Arguments**

```
spatial_grid spatial_grid in giotto object
```

# Value

edges in spatial grid as data.table()

# **Examples**

```
plotly_grid(gobject)
```

plotly\_network

plotly\_network

# **Description**

```
provide network segment to draw in 3D plot_ly()
```

### Usage

```
plotly_network(
  network,
  x = "sdimx_begin",
  y = "sdimy_begin",
  z = "sdimz_begin",
  x_end = "sdimx_end",
  y_end = "sdimy_end",
  z_end = "sdimz_end"
)
```

# **Arguments**

gobject network in giotto object

### Value

edges in network as data.table()

### **Examples**

```
plotly_network(gobject)
```

plotMetaDataCellsHeatmap

plot Meta Data Cells Heat map

# **Description**

creates order for clusters

# Usage

```
plotMetaDataCellsHeatmap(
  gobject,
  metadata_cols = NULL,
  spat_enr_names = NULL,
  value_cols = NULL,
  first_meta_col = NULL,
  second_meta_col = NULL,
  show_values = c("zscores", "original", "zscores_rescaled"),
  custom_cluster_order = NULL,
  clus_cor_method = "pearson",
  clus_cluster_method = "complete",
  custom_values_order = NULL,
  values_cor_method = "pearson",
  values_cluster_method = "complete",
  midpoint = 0,
  x_{text_size} = 8,
  x_{text_angle} = 45,
  y_{text_size} = 8,
```

```
strip_text_size = 8,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "plotMetaDataCellsHeatmap")
```

# **Arguments**

```
gobject
                  giotto object
metadata_cols
                  annotation columns found in pDataDT(gobject)
spat_enr_names spatial enrichment results to include
value_cols
                  value columns to use
first_meta_col if more than 1 metadata column, select the x-axis factor
second_meta_col
                  if more than 1 metadata column, select the facetting factor
show_values
                  which values to show on heatmap
custom_cluster_order
                  custom cluster order (default = NULL)
clus_cor_method
                  correlation method for clusters
clus_cluster_method
                  hierarchical cluster method for the clusters
                  midpoint of show_values
midpoint
                  size of x-axis text
x_text_size
                  angle of x-axis text
x_text_angle
y_text_size
                  size of y-axis text
strip_text_size
                  size of strip text
                  show plot
show_plot
                  return ggplot object
return_plot
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
custom_gene_order
                  custom gene order (default = NULL)
gene_cor_method
                  correlation method for genes
gene_cluster_method
                  hierarchical cluster method for the genes
```

# Details

Creates heatmap for the average values of selected value columns in the different annotation groups

plotMetaDataHeatmap 155

### Value

ggplot or data.table

### **Examples**

```
plotMetaDataCellsHeatmap(gobject)
```

plotMetaDataHeatmap

# Description

creates order for clusters

# Usage

```
plotMetaDataHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metadata_cols = NULL,
  selected_genes = NULL,
  first_meta_col = NULL,
  second_meta_col = NULL,
  show_values = c("zscores", "original", "zscores_rescaled"),
  custom_cluster_order = NULL,
  clus_cor_method = "pearson",
  clus_cluster_method = "complete",
  custom_gene_order = NULL,
  gene_cor_method = "pearson",
  gene_cluster_method = "complete",
  midpoint = 0,
  x_{text_size} = 10,
  x_{text_angle} = 45,
  y_{text_size} = 10,
  strip_text_size = 8,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotMetaDataHeatmap"
)
```

```
gobject giotto object
expression_values
expression values to use

metadata_cols annotation columns found in pDataDT(gobject)
selected_genes subset of genes to use
first_meta_col if more than 1 metadata column, select the x-axis factor
```

```
second_meta_col
                  if more than 1 metadata column, select the facetting factor
show_values
                  which values to show on heatmap
custom_cluster_order
                  custom cluster order (default = NULL)
clus_cor_method
                  correlation method for clusters
clus_cluster_method
                  hierarchical cluster method for the clusters
custom_gene_order
                  custom gene order (default = NULL)
gene_cor_method
                  correlation method for genes
gene_cluster_method
                  hierarchical cluster method for the genes
midpoint
                  midpoint of show_values
                  size of x-axis text
x_text_size
x_text_angle
                  angle of x-axis text
y_text_size
                  size of y-axis text
strip_text_size
                  size of strip text
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

# **Details**

Creates heatmap for average the average expression of selected genes in the different annotation groups

### Value

ggplot or data.table

# **Examples**

plotMetaDataHeatmap(gobject)

plotPCA 157

plotPCA plotPCA

# **Description**

Short wrapper for PCA visualization

### Usage

```
plotPCA(gobject, dim_reduction_name = "pca", default_save_name = "PCA", ...)
```

```
gobject
                  giotto object
dim_reduction_name
                  dimension reduction name
default_save_name
                  default save name for saving, don't change, change save_name in save_param
groub_by
                  create multiple plots based on cell annotation column
group_by_subset
                  subset the group_by factor column
                  dimension to use on x-axis
dim1_to_use
                  dimension to use on y-axis
dim2_to_use
spat_enr_names names of spatial enrichment results to include
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
network_name
                  name of NN network to use, if show_NN_network = TRUE
                  color for cells (see details)
cell_color
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
```

158 plotPCA

```
other_point_size
```

size of not selected cells

show\_cluster\_center

plot center of selected clusters

show\_center\_label

plot label of selected clusters

center\_point\_size

size of center points

label\_size size of labels
label\_fontface font of labels

edge\_alpha column to use for alpha of the edges

point\_size size of point (cell)

point\_border\_col

color of border around points

point\_border\_stroke

stroke size of border around points

show\_legend show legend

title title for plot, defaults to cell\_color parameter

legend\_text size of legend text
axis\_text size of axis text
axis\_title size of axis title

cow\_n\_col cowplot param: how many columns

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

 $save\_param \qquad list of saving \ parameters \ from \ all\_plots\_save\_function()$ 

# **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotPCA\_3D

# Value

ggplot

### **Examples**

plotPCA(gobject)

plotPCA\_2D 159

plotPCA\_2D plotPCA\_2D

### **Description**

Short wrapper for PCA visualization

# Usage

```
plotPCA_2D(
  gobject,
  dim_reduction_name = "pca",
  default_save_name = "PCA_2D",
)
```

```
gobject
                 giotto object
dim_reduction_name
                 dimension reduction name
default_save_name
                 default save name for saving, don't change, change save_name in save_param
                 create multiple plots based on cell annotation column
groub_by
group_by_subset
                 subset the group_by factor column
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
                 color for cells (see details)
cell_color
color_as_factor
                 convert color column to factor
cell_color_code
                 named vector with colors
cell_color_gradient
                 vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                 select subset of cells/clusters based on cell_color parameter
```

plotPCA\_2D

```
select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
title
                  title for plot, defaults to cell_color parameter
show_legend
                  show legend
legend_text
                  size of legend text
axis_text
                  size of axis text
axis_title
                  size of axis title
                  cowplot param: how many columns
cow_n_col
cow_rel_h
                  cowplot param: relative height
                  cowplot param: relative width
cow_rel_w
cow_align
                  cowplot param: how to align
show_plot
                  show plot
                  return ggplot object
return_plot
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
```

### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotPCA\_3D

### Value

ggplot

# **Examples**

```
plotPCA_2D(gobject)
```

plotPCA\_3D 161

plotPCA\_3D plotPCA\_3D

### **Description**

Visualize cells according to 3D PCA dimension reduction

# Usage

```
plotPCA_3D(
  gobject,
  dim_reduction_name = "pca",
  default_save_name = "PCA_3D",
   ...
)
```

```
gobject
                  giotto object
dim_reduction_name
                  pca dimension reduction name
default_save_name
                  default save name for saving, ideally change save_name in save_param
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
dim3_to_use
                  dimension to use on z-axis
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
                  name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
```

162 plotTSNE

list of saving parameters from all\_plots\_save\_function()

```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
                  size of labels
label_size
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
```

**Details** 

save\_param

Description of parameters.

### Value

plotly

# **Examples**

```
plotPCA_3D(gobject)
```

plotTSNE plotTSNE

# Description

Short wrapper for tSNE visualization

# Usage

```
plotTSNE(gobject, dim_reduction_name = "tsne", default_save_name = "tSNE", ...)
```

plotTSNE 163

dim2\_to\_use dimension to use on y-axis spat\_enr\_names names of spatial enrichment results to include show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) network\_name name of NN network to use, if show\_NN\_network = TRUE cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels edge\_alpha column to use for alpha of the edges point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points title title for plot, defaults to cell\_color parameter show\_legend show legend legend\_text size of legend text axis\_text size of axis text axis\_title size of axis title

164 plotTSNE\_2D

```
cow_n_col
                  cowplot param: how many columns
                  cowplot param: relative height
cow_rel_h
                  cowplot param: relative width
cow_rel_w
cow_align
                  cowplot param: how to align
show_plot
                  show plot
                  return ggplot object
return_plot
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
```

# **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotTSNE\_3D

### Value

ggplot

### **Examples**

```
plotTSNE(gobject)
```

```
plotTSNE_2D
```

plotTSNE\_2D

# **Description**

Short wrapper for tSNE visualization

# Usage

```
plotTSNE_2D(
  gobject,
  dim_reduction_name = "tsne",
  default_save_name = "tSNE_2D",
  ...
)
```

plotTSNE\_2D 165

dim2\_to\_use dimension to use on y-axis spat\_enr\_names names of spatial enrichment results to include show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) network\_name name of NN network to use, if show\_NN\_network = TRUE cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels edge\_alpha column to use for alpha of the edges point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points title title for plot, defaults to cell\_color parameter show\_legend show legend size of legend text legend\_text axis\_text size of axis text axis\_title size of axis title

166 plotTSNE\_3D

```
cow_n_col
                  cowplot param: how many columns
                  cowplot param: relative height
cow_rel_h
                  cowplot param: relative width
cow_rel_w
                  cowplot param: how to align
cow_align
show_plot
                  show plot
                  return ggplot object
return_plot
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
```

### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotTSNE\_3D

### Value

ggplot

# **Examples**

```
plotTSNE_2D(gobject)
```

plotTSNE\_3D plotTSNE\_3D

# **Description**

Visualize cells according to dimension reduction coordinates

# Usage

```
plotTSNE_3D(
  gobject,
  dim_reduction_name = "tsne",
  default_save_name = "TSNE_3D",
  ...
)
```

plotTSNE\_3D 167

nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels edge\_alpha column to use for alpha of the edges size of point (cell) point\_size show\_legend show legend show plot show\_plot

# **Details**

Description of parameters.

return ggplot object

directly save the plot [boolean]

list of saving parameters from all\_plots\_save\_function()

return\_plot

save\_plot

save\_param

# Value

plotly

# **Examples**

```
plotTSNE_3D(gobject)
```

168 plotUMAP

plotUMAP plotUMAP

# **Description**

Short wrapper for UMAP visualization

### Usage

```
plotUMAP(gobject, dim_reduction_name = "umap", default_save_name = "UMAP", ...)
```

```
gobject
                  giotto object
dim_reduction_name
                  dimension reduction name
default_save_name
                  default save name for saving, don't change, change save_name in save_param
groub_by
                  create multiple plots based on cell annotation column
group_by_subset
                  subset the group_by factor column
                  dimension to use on x-axis
dim1_to_use
                  dimension to use on y-axis
dim2_to_use
spat_enr_names names of spatial enrichment results to include
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
network_name
                  name of NN network to use, if show_NN_network = TRUE
                  color for cells (see details)
cell_color
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
```

plotUMAP 169

```
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
title
                  title for plot, defaults to cell_color parameter
show_legend
                  show legend
legend_text
                  size of legend text
axis_text
                  size of axis text
                  size of axis title
axis_title
cow_n_col
                  cowplot param: how many columns
cow_rel_h
                  cowplot param: relative height
                  cowplot param: relative width
cow_rel_w
cow_align
                  cowplot param: how to align
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
```

# **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotUMAP\_3D

# Value

ggplot

### **Examples**

```
plotUMAP(gobject)
```

170 plotUMAP\_2D

plotUMAP\_2D

plotUMAP\_2D

### **Description**

Short wrapper for UMAP visualization

# Usage

```
plotUMAP_2D(
  gobject,
  dim_reduction_name = "umap",
  default_save_name = "UMAP_2D",
)
```

```
gobject
                 giotto object
dim_reduction_name
                 dimension reduction name
default_save_name
                 default save name for saving, don't change, change save_name in save_param
                 create multiple plots based on cell annotation column
groub_by
group_by_subset
                 subset the group_by factor column
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
                 color for cells (see details)
cell_color
color_as_factor
                 convert color column to factor
cell_color_code
                 named vector with colors
cell_color_gradient
                 vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                 select subset of cells/clusters based on cell_color parameter
```

plotUMAP\_2D 171

```
select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
title
                  title for plot, defaults to cell_color parameter
show_legend
                  show legend
legend_text
                  size of legend text
axis_text
                  size of axis text
axis_title
                  size of axis title
                  cowplot param: how many columns
cow_n_col
cow_rel_h
                  cowplot param: relative height
                  cowplot param: relative width
cow_rel_w
cow_align
                  cowplot param: how to align
show_plot
                  show plot
                  return ggplot object
return_plot
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
```

### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotUMAP\_3D

### Value

ggplot

# **Examples**

```
plotUMAP_2D(gobject)
```

172 plotUMAP\_3D

plotUMAP\_3D

plotUMAP\_3D

### **Description**

Visualize cells according to dimension reduction coordinates

# Usage

```
plotUMAP_3D(
  gobject,
  dim_reduction_name = "umap",
  default_save_name = "UMAP_3D",
   ...
)
```

```
gobject
                  giotto object
dim_reduction_name
                  umap dimension reduction name
default_save_name
                  default save name for saving, don't change, change save_name in save_param
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
dim3_to_use
                  dimension to use on z-axis
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
                  name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
```

```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
                  column to use for alpha of the edges
edge_alpha
point_size
                  size of point (cell)
show_legend
                  show legend
show_plot
                  show plot
                  return ggplot object
return_plot
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
```

### **Details**

Description of parameters.

# Value

plotly

### **Examples**

```
plotUMAP_3D(gobject)
```

# **Description**

Visualize cells in network layer according to dimension reduction coordinates

# Usage

```
plot_network_layer_ggplot(
   ggobject,
   annotated_network_DT,
   edge_alpha = NULL,
   show_legend = T
)
```

# **Arguments**

```
annotated_network_DT
```

annotated network data.table of selected cells

edge\_alpha alpha of network edges

show\_legend show legend gobject giotto object

### **Details**

Description of parameters.

### Value

ggplot

# **Examples**

```
plot_network_layer_ggplot(gobject)
```

# **Description**

Visualize cells in point layer according to dimension reduction coordinates

# Usage

```
plot_point_layer_ggplot(
  ggobject,
  annotated_DT_selected,
  annotated_DT_other,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = 0,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_legend = T
)
```

### **Arguments**

```
annotated_DT_selected
                  annotated data.table of selected cells
annotated_DT_other
                  annotated data.table of not selected cells
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
                  show legend
show_legend
                  giotto object
gobject
```

### **Details**

Description of parameters.

### Value

ggplot

### **Examples**

```
plot_point_layer_ggplot(gobject)
```

### **Description**

creat ggplot point layer for spatial coordinates

# Usage

```
plot_spat_point_layer_ggplot(
  ggobject,
  sdimx = NULL,
  sdimy = NULL,
  cell_locations_metadata_selected,
  cell_locations_metadata_other,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 2,
  point_border_col = "lightgrey",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  show_legend = TRUE
)
```

```
sdimx x-axis dimension name (default = 'sdimx')
sdimy y-axis dimension name (default = 'sdimy')
cell_locations_metadata_selected
annotated location from selected cells
```

```
cell_locations_metadata_other
                  annotated location from non-selected cells
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
                  size of labels
label_size
label_fontface font of labels
show_other_cells
                  display not selected cells
other_cell_color
                  color for not selected cells
other_point_size
                  point size for not selected cells
show_legend
                  show legend
                  giotto object
gobject
```

# **Details**

Description of parameters.

### Value

ggplot

# **Examples**

```
plot_spat_point_layer_ggplot(gobject)
```

178 rankEnrich

print.giotto

print method for giotto class

### **Description**

print method for giotto class. Prints the chosen number of genes (rows) and cells (columns) from the raw count matrix. Also print the spatial locations for the chosen number of cells.

# Usage

```
print.giotto(object, ...)
```

# **Arguments**

nr\_genes number of genes (rows) to print nr\_cells number of cells (columns) to print

rankEnrich

rankEnrich

# Description

Function to calculate gene signature enrichment scores per spatial position using a rank based approach.

# Usage

```
rankEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

rank\_binarize 179

### **Details**

First a new rank is calculated as  $R = (R1*R2)^{n}(1/2)$ , where R1 is the rank of fold-change for each gene in each spot and R2 is the rank of each marker in each cell type. The Rank-Biased Precision is then calculated as:  $RBP = (1 - 0.99) * (0.99)^{n}(R - 1)$  and the final enrichment score is then calculated as the sum of top 100 RBPs.

# Value

data.table with enrichment results

# **Examples**

```
rankEnrich(gobject)
```

rank\_binarize

rank\_binarize

# Description

create binarized scores using arbitrary rank of top genes

# Usage

```
rank\_binarize(x, max\_rank = 200)
```

 ${\tt readGiottoInstructions}$ 

read Giot to Instrunctions

# Description

Retrieves the instruction associated with the provided parameter

# Usage

```
readGiottoInstructions(giotto_instructions, param = NULL)
```

# **Arguments**

```
giotto_instructions
```

giotto object or result from createGiottoInstructions()

param

parameter to retrieve

### Value

specific parameter

### **Examples**

readGiottoInstrunctions()

180 removeGeneAnnotation

remove Cell Annotation remove Cell Annotation

# Description

removes cell annotation of giotto object

# Usage

```
removeCellAnnotation(gobject, columns = NULL, return_gobject = TRUE)
```

### **Arguments**

gobject giotto object

columns names of columns to remove

return\_gobject boolean: return giotto object (default = TRUE)

### **Details**

if return\_gobject = FALSE, it will return the cell metadata

# Value

giotto object

# **Examples**

removeCellAnnotation(gobject)

removeGeneAnnotation removeGeneAnnotation

# **Description**

removes gene annotation of giotto object

# Usage

```
removeGeneAnnotation(gobject, columns = NULL, return_gobject = TRUE)
```

# **Arguments**

gobject giotto object

columns names of columns to remove

return\_gobject boolean: return giotto object (default = TRUE)

# **Details**

if return\_gobject = FALSE, it will return the gene metadata

#### Value

giotto object

### **Examples**

```
removeGeneAnnotation(gobject)
```

```
replaceGiottoInstructions
```

replace Giot to Instructions

## **Description**

Function to replace all instructions from giotto object

### Usage

```
replaceGiottoInstructions(gobject, instructions = NULL)
```

### **Arguments**

```
gobject giotto object
instructions new instructions (e.g. result from createGiottoInstructions)
```

#### Value

named vector with giotto instructions

### **Examples**

```
replaceGiottoInstructions()
```

runPCA

runPCA

## Description

runs a Principal Component Analysis

```
runPCA(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  name = "pca",
  genes_to_use = NULL,
  return_gobject = TRUE,
  scale_unit = F,
  ncp = 200,
  ...
)
```

182 runtSNE

#### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
                  cells or genes
reduction
name
                  arbitrary name for PCA run
                  subset of genes to use for PCA
genes_to_use
return_gobject boolean: return giotto object (default = TRUE)
scale_unit
                  scale features before PCA
                  number of principal components to calculate
ncp
                  additional parameters for PCA (see details)
. . .
```

#### **Details**

See PCA for more information about other parameters.

### Value

giotto object with updated PCA dimension recuction

### **Examples**

```
runPCA(gobject)
```

runtSNE

runtSNE

## Description

run tSNE

```
runtSNE(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "tsne",
  genes_to_use = NULL,
  return_gobject = TRUE,
  dims = 2,
  perplexity = 30,
  theta = 0.5,
  do_PCA_first = F,
  set\_seed = T,
  seed_number = 1234,
)
```

runtSNE 183

### **Arguments**

gobject giotto object

expression\_values

expression values to use

reduction cells or genes

dim\_reduction\_to\_use

use another dimension reduction set as input

dim\_reduction\_name

name of dimension reduction set to use

dimensions\_to\_use

number of dimensions to use as input

name arbitrary name for tSNE run

genes\_to\_use if dim\_reduction\_to\_use = NULL, which genes to use

return\_gobject boolean: return giotto object (default = TRUE)

dims tSNE param: number of dimensions to return

perplexity tSNE param: perplexity

theta tSNE param: theta

do\_PCA\_first tSNE param: do PCA before tSNE (default = FALSE)

set\_seed use of seed

seed\_number seed number to use

... additional tSNE parameters

#### **Details**

See Rtsne for more information about these and other parameters.

- Input for tSNE dimension reduction can be another dimension reduction (default = 'pca')
- To use gene expression as input set dim\_reduction\_to\_use = NULL
- multiple tSNE results can be stored by changing the *name* of the analysis

#### Value

giotto object with updated tSNE dimension recuction

### **Examples**

runtSNE(gobject)

184 runUMAP

runUMAP

runUMAP

### **Description**

run UMAP

### Usage

```
runUMAP(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "umap",
  genes_to_use = NULL,
  return_gobject = TRUE,
  n_neighbors = 40,
  n_{components} = 2,
  n_{epochs} = 400,
  min_dist = 0.01,
  n_{threads} = 1,
  spread = 5,
  set\_seed = T,
  seed_number = 1234,
)
```

### **Arguments**

```
gobject
                 giotto object
expression_values
                 expression values to use
                 cells or genes
reduction
dim_reduction_to_use
                 use another dimension reduction set as input
dim_reduction_name
                 name of dimension reduction set to use
dimensions_to_use
                 number of dimensions to use as input
                 arbitrary name for UMAP run
name
                 if dim_reduction_to_use = NULL, which genes to use
genes_to_use
return_gobject boolean: return giotto object (default = TRUE)
n\_neighbors
                 UMAP param: number of neighbors
                 UMAP param: number of components
n_components
n_epochs
                 UMAP param: number of epochs
```

selectPatternGenes 185

```
min_dist UMAP param: minimum distance
n_threads UMAP param: threads to use
spread UMAP param: spread
set_seed use of seed
seed_number seed number to use
additional UMAP parameters
```

#### **Details**

See umap for more information about these and other parameters.

- Input for UMAP dimension reduction can be another dimension reduction (default = 'pca')
- To use gene expression as input set dim\_reduction\_to\_use = NULL
- multiple UMAP results can be stored by changing the *name* of the analysis

#### Value

giotto object with updated UMAP dimension recuction

#### **Examples**

```
runUMAP(gobject)
```

selectPatternGenes selectPatternGenes

### Description

Select genes correlated with spatial patterns

#### Usage

```
selectPatternGenes(
   spatPatObj,
   dimensions = 1:5,
   top_pos_genes = 10,
   top_neg_genes = 10,
   min_pos_cor = 0.5,
   min_neg_cor = -0.5,
   return_top_selection = FALSE
)
```

#### **Arguments**

spatPatObj

```
dimensions dimensions to identify correlated genes for.

top_pos_genes Top positively correlated genes.

top_neg_genes Top negatively correlated genes.

min_pos_cor Minimum positive correlation score to include a gene.

min_neg_cor Minimum negative correlation score to include a gene.
```

Output from detectSpatialPatterns

show,giotto-method

#### **Details**

Description.

#### Value

Data.table with genes associated with selected dimension (PC).

## **Examples**

```
selectPatternGenes(gobject)
```

```
select_expression_values
```

select\_expression\_values

## Description

helper function to select expression values

## Usage

```
select_expression_values(gobject, values)
```

## Arguments

gobject giotto object

values expression values to extract

## Value

expression matrix

show,giotto-method

show method for giotto class

## Description

show method for giotto class

```
## S4 method for signature 'giotto'
show(object)
```

 $show Cluster Dendrogram \quad show Cluster Dendrogram \quad$ 

#### **Description**

Creates dendrogram based on identified clusters

#### Usage

```
showClusterDendrogram(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  h = NULL,
  h_color = "red",
  rotate = FALSE,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showClusterDendrogram",
  ...
)
```

#### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
distance
                  distance method to use for hierarchical clustering
h
                  height of horizontal lines to plot
h_color
                  color of horizontal lines
                  rotate dendrogram 90 degrees
rotate
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  additional parameters for ggdendrogram()
```

### **Details**

Correlation dendrogram of selected clustering.

188 showClusterHeatmap

#### Value

ggplot

### **Examples**

showClusterDendrogram(gobject)

showClusterHeatmap

*showClusterHeatmap* 

#### **Description**

Creates heatmap based on identified clusters

### Usage

```
showClusterHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = "all",
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showClusterHeatmap",
  ...
)
```

## Arguments

```
giotto object
gobject
expression_values
                  expression values to use
                  vector of genes to use, default to 'all'
genes
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
distance
                  distance method to use for hierarchical clustering
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  additional parameters for the Heatmap function from ComplexHeatmap
. . .
```

showCPGscores 189

#### **Details**

Correlation heatmap of selected clusters.

#### Value

ggplot

### **Examples**

```
showClusterHeatmap(gobject)
```

showCPGscores

showCPGscores

### **Description**

visualize Cell Proximity Gene enrichment scores

## Usage

```
showCPGscores(
 gobject,
 CPGscore,
 method = c("volcano", "cell_barplot", "cell-cell", "cell_sankey"),
 min_cells = 5,
 min_fdr = 0.05,
 min_spat_diff = 0.2,
 min_log2_fc = 0.5,
 keep_int_duplicates = TRUE,
 direction = c("both", "up", "down"),
 cell_color_code = NULL,
  show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "showCPGscores"
)
```

## Arguments

```
CPGscore
                 CPGscore, output from getCellProximityGeneScores()
method
                 visualization method
                 min number of cells threshold
min_cells
min_fdr
                 fdr threshold
min_spat_diff
                 spatial difference threshold
                 min log2 fold-change
min_log2_fc
keep_int_duplicates
                 keep both cell_A-cell_B and cell_B-cell_A
direction
                 up or downregulation or both
```

```
cell_color_code
```

color code for cell types

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

#### **Details**

Give more details ...

## Value

Gene to gene scores in data.table format

## **Examples**

```
showCPGscores(CPGscore)
```

showGeneExpressionProximityScore

show Gene Expression Proximity Score

## **Description**

Create heatmap from cell-cell proximity scores

## Usage

```
showGeneExpressionProximityScore(
  scores,
  selected_gene,
  sort_column = "diff_spat"
)
```

## Arguments

scores CPscore, output from getAverageCellProximityGeneScores()

selected\_gene gene to show

sort\_column column name to use for sorting

#### **Details**

Give more details ...

#### Value

ggplot barplot

showGiottoInstructions 191

### **Examples**

```
showGeneExpressionProximityScore(scores)
```

```
{\tt showGiottoInstructions}
```

showGiottoInstructions

## Description

Function to display all instructions from giotto object

## Usage

```
showGiottoInstructions(gobject)
```

### **Arguments**

```
gobject giotto object
```

#### Value

named vector with giotto instructions

### **Examples**

```
showGiottoInstructions()
```

 $\verb|showGTGscores||$ 

showGTGscores

## Description

visualize Cell Proximity Gene enrichment scores

```
showGTGscores(
  GTGscore,
  method = c("cell_barplot", "cell-cell", "cell_sankey"),
  min_cells = 5,
  min_pval = 0.05,
  min_spat_diff = 0.2,
  min_log2_fc = 0.5,
  direction = c("both", "up", "down"),
  cell_color_code = NULL,
  show_plot = T,
  specific_genes_1 = NULL,
  specific_genes_2 = NULL,
  first_cell_name = "ligand cell",
  second_cell_name = "receptor cell",
  return_DT = F
)
```

#### **Arguments**

```
visualization method
method
min_cells
                  min number of cells threshold
                  p-value threshold
min_pval
min_spat_diff
                  spatial difference threshold
                  log2 fold-change threshold
min_log2_fc
direction
                  up or downregulation or both
cell_color_code
                  color code for cell types
show_plot
                  print plot
specific_genes_1
                  subset of genes, matched with specific_genes_2
specific_genes_2
                  subset of genes, matched with specific_genes_1
first_cell_name
                  name for first cells
second_cell_name
                  name for second cells
                  CPGscore, output from getCellProximityGeneScores()
CPGscore
```

#### **Details**

Give more details ...

### Value

ggplot

### **Examples**

```
showGTGscores(CPGscore)
```

```
show Int {\it Expression Proximity Score} \\ show Int {\it Expression Proximity Score}
```

## **Description**

Create heatmap from cell-cell proximity scores

```
showIntExpressionProximityScore(
   scores,
   selected_interaction,
   sort_column = "diff_spat",
   show_enriched_n = 5,
   show_depleted_n = 5
```

showPattern 193

### **Arguments**

scores scores, output from getAverageCellProximityGeneScores()
selected\_interaction
interaction to show
sort\_column column name to use for sorting
show\_enriched\_n
show top enriched interactions
show\_depleted\_n

show top depleted interactions

#### **Details**

Give more details ...

### Value

ggplot barplot

#### **Examples**

showIntExpressionProximityScore(scores)

showPattern showPattern

## Description

```
create a spatial grid
show patterns for 2D spatial data
```

### Usage

```
showPattern(gobject, spatPatObj, ...)
showPattern(gobject, spatPatObj, ...)
```

### **Arguments**

gobject giotto object

spatPatObj Output from detectSpatialPatterns

dimension dimension to plot

trim Trim ends of the PC values.

background\_color

background color for plot

grid\_border\_color

color for grid

show\_legend show legend of ggplot

show\_plot show plot

194 showPattern2D

### **Details**

Description.

### Value

ggplot ggplot

### See Also

showPattern2D

## **Examples**

```
showPattern(gobject)
showPattern(gobject)
```

showPattern2D

showPattern2D

## **Description**

show patterns for 2D spatial data

```
showPattern2D(
  gobject,
  spatPatObj,
  dimension = 1,
  trim = c(0.02, 0.98),
  background_color = "white",
  grid_border_color = "grey",
  show_legend = T,
  point_size = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showPattern2D"
)
```

showPattern3D 195

### **Arguments**

gobject giotto object spatPatObj Output from detectSpatialPatterns dimension dimension to plot trim Trim ends of the PC values. background\_color background color for plot grid\_border\_color color for grid show\_legend show legend of ggplot  ${\sf show\_plot}$ show plot return\_plot return ggplot object directly save the plot [boolean] save\_plot save\_param list of saving parameters from all\_plots\_save\_function()

default save name for saving, don't change, change save\_name in save\_param

#### Value

ggplot

default\_save\_name

## **Examples**

showPattern2D(gobject)

showPattern3D showPattern3D

### **Description**

show patterns for 3D spatial data

```
showPattern3D(
  gobject,
  spatPatObj,
  dimension = 1,
  trim = c(0.02, 0.98),
  background_color = "white",
  grid_border_color = "grey",
  show_legend = T,
  point_size = 1,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
```

196 showPattern3D

```
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "showPattern3D"
)
```

giotto object

show plot

return plot object

directly save the plot [boolean]

list of saving parameters from all\_plots\_save\_function()

default save name for saving, don't change, change save\_name in save\_param

### **Arguments**

gobject

```
spatPatObj
                  Output from detectSpatialPatterns
                  dimension to plot
dimension
                  Trim ends of the PC values.
trim
background_color
                  background color for plot
grid_border_color
                  color for grid
show_legend
                  show legend of plot
point_size
                  adjust the point size
                  scale the axis
axis_scale
                  cutomize the scale of the axis
custom_ratio
                  the tick number of x_axis
x_ticks
y_ticks
                  the tick number of y_axis
                  the tick number of z_axis
z_ticks
```

#### Value

plotly

show\_plot
return\_plot

save\_plot

save\_param

default\_save\_name

# Examples

```
showPattern3D(gobject)
```

showPatternGenes 197

showPatternGenes

showPatternGenes

### **Description**

show genes correlated with spatial patterns

#### Usage

```
showPatternGenes(
  gobject,
  spatPatObj,
  dimension = 1,
  top_pos_genes = 5,
  top_neg_genes = 5,
  point_size = 1,
  return_DT = FALSE,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showPatternGenes"
)
```

## Arguments

```
gobject
                  giotto object
spatPatObj
                  Output from detectSpatialPatterns
dimension
                  dimension to plot genes for.
top_pos_genes
                  Top positively correlated genes.
top_neg_genes
                  Top negatively correlated genes.
                  size of points
point_size
return_DT
                  if TRUE, it will return the data.table used to generate the plots
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

## Value

ggplot

### **Examples**

```
showPatternGenes(gobject)
```

showTopGeneToGene

```
showProcessingSteps showProcessingSteps
```

### **Description**

shows the sequential processing steps that were performed in a summarized format

### Usage

```
showProcessingSteps(gobject)
```

### **Arguments**

```
gobject giotto object
```

#### Value

list of processing steps and names

### **Examples**

```
showProcessingSteps(gobject)
```

showTopGeneToGene

*showTopGeneToGene* 

## Description

Show enriched/depleted gene-gene enrichments

### Usage

```
showTopGeneToGene(
  GTGscore,
  top_interactions = 10,
  direction = c("increased", "decreased"),
  complement_data = T,
  subset_cell_ints = NULL,
  subset_genes = NULL
)
```

### **Arguments**

signPCA 199

#### **Details**

Give more details ...

#### Value

ggplot barplot

#### **Examples**

showTopGeneToGene(scores)

signPCA

signPCA

## Description

identify significant prinicipal components (PCs)

## Usage

```
signPCA(
  gobject,
  method = c("screeplot", "jackstraw"),
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  genes_to_use = NULL,
  scale_unit = T,
  ncp = 50,
  scree_labels = T,
  scree_ylim = c(0, 10),
  jack_iter = 10,
  jack_threshold = 0.01,
  jack_verbose = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "signPCA",
)
```

## **Arguments**

200 spatCellPlot

```
number of principal components to calculate
ncp
scree_labels
                  show labels on scree plot
scree_ylim
                  y-axis limits on scree plot
jack_iter
                  number of interations for jackstraw
jack_threshold p-value threshold to call a PC significant
                  show progress of jackstraw method
jack_verbose
show_plot
                  show plot
                  return ggplot object
return_plot
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  additional parameters for PCA
```

#### **Details**

Two different methods can be used to assess the number of relevant or significant prinicipal components (PC's).

- 1. Screeplot works by plotting the explained variance of each individual PC in a barplot allowing you to identify which PC does not show a significant contribution anymore ( = 'elbow method').
- 2. The Jackstraw method uses the permutationPA function. By systematically permuting genes it identifies robust, and thus significant, PCs.

multiple PCA results can be stored by changing the name parameter

#### Value

ggplot object for scree method and maxtrix of p-values for jackstraw

### **Examples**

```
signPCA(gobject)
```

spatCellPlot spatCellPlot

### **Description**

Visualize cells according to spatial coordinates

```
spatCellPlot(
  gobject,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
  cell_annotation_values,
  cell_color_gradient = c("blue", "white", "red"),
```

spatCellPlot 201

```
gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
 center_point_size = 4,
 center_point_border_col = "black",
 center_point_border_stroke = 0.1,
  label_size = 4,
 label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
 show\_grid = F,
  spatial_grid_name = "spatial_grid",
 grid_color = NULL,
 show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
 other_cells_alpha = 0.1,
 coord_fix_ratio = NULL,
  show_legend = T,
  legend_text = 8,
  axis_text = 8,
 axis_title = 8,
 cow_n_col = 2,
  cow_rel_h = 1,
 cow_rel_w = 1,
 cow_align = "h",
 show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
  default_save_name = "spatCellPlot"
)
```

### **Arguments**

202 spatCellPlot

gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points size of labels label\_size label\_fontface font of labels show\_network show underlying spatial network spatial\_network\_name name of spatial network to use network\_color color of spatial network network\_alpha alpha of spatial network show\_grid show spatial grid spatial\_grid\_name name of spatial grid to use grid\_color color of spatial grid show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size point size of not selected cells other\_cells\_alpha alpha of not selected cells coord\_fix\_ratio fix ratio between x and y-axis show legend show\_legend legend\_text size of legend text axis\_text size of axis text axis\_title size of axis title show\_plot show plot return ggplot object return\_plot directly save the plot [boolean] save\_plot list of saving parameters from all\_plots\_save\_function() save\_param default\_save\_name default save name for saving, don't change, change save\_name in save\_param

#### **Details**

Description of parameters.

#### Value

ggplot

#### **Examples**

```
spatCellPlot(gobject)
```

spatCellPlot2D

spatCellPlot2D

### **Description**

Visualize cells according to spatial coordinates

```
spatCellPlot2D(
 gobject,
 sdimx = "sdimx",
 sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_annotation_values,
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
 select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
 label_fontface = "bold",
 show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
  show\_grid = F,
  spatial_grid_name = "spatial_grid",
  grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
```

```
other_cells_alpha = 0.1,
      coord_fix_ratio = NULL,
      show_legend = T,
      legend_text = 8,
      axis_text = 8,
      axis_title = 8,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatCellPlot2D"
    )
Arguments
    gobject
                     giotto object
                     x-axis dimension name (default = 'sdimx')
    sdimx
                     y-axis dimension name (default = 'sdimy')
    sdimy
    spat_enr_names names of spatial enrichment results to include
    cell_annotation_values
                     numeric cell annotation columns
    cell_color_gradient
                      vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell color parameter
                     select subset of cells based on cell IDs
    select_cells
    point_size
                     size of point (cell)
    point_border_col
                     color of border around points
    point_border_stroke
                     stroke size of border around points
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
                     size of center points
    label_size
                     size of labels
    label_fontface font of labels
```

show underlying spatial network

show\_network

spatial\_network\_name name of spatial network to use network\_color color of spatial network network\_alpha alpha of spatial network show\_grid show spatial grid spatial\_grid\_name name of spatial grid to use grid\_color color of spatial grid show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size point size of not selected cells other\_cells\_alpha alpha of not selected cells coord\_fix\_ratio fix ratio between x and y-axis show\_legend show legend legend\_text size of legend text axis\_text size of axis text size of axis title axis\_title show\_plot show plot return\_plot return ggplot object save\_plot directly save the plot [boolean] list of saving parameters from all\_plots\_save\_function() save\_param

default save name for saving, don't change, change save\_name in save\_param

### **Details**

Description of parameters.

default\_save\_name

### Value

ggplot

# Examples

spatCellPlot2D(gobject)

spatDimCellPlot

spatDimCellPlot

#### **Description**

Visualize numerical features of cells according to spatial AND dimension reduction coordinates in 2D

```
spatDimCellPlot(
 gobject,
 plot_alignment = c("vertical", "horizontal"),
  spat_enr_names = NULL,
  cell_annotation_values,
  dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 dim_point_size = 1,
  dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 dim_show_cluster_center = F,
 dim_show_center_label = T,
 dim_center_point_size = 4,
  dim_center_point_border_col = "black",
 dim_center_point_border_stroke = 0.1,
 dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_center_point_border_col = "black",
  spat_center_point_border_stroke = 0.1,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
 nn_network_to_use = "sNN",
 nn_network_name = "sNN.pca",
 dim_edge_alpha = 0.5,
  spat_show_network = F,
```

```
spatial_network_name = "spatial_network",
      spat_network_color = "red",
      spat_network_alpha = 0.5,
      spat_show_grid = F,
      spatial_grid_name = "spatial_grid",
      spat_grid_color = "green",
      show_other_cells = TRUE,
     other_cell_color = "grey"
     dim_other_point_size = 0.5,
      spat_other_point_size = 0.5,
      spat_other_cells_alpha = 0.5,
     coord_fix_ratio = NULL,
      cow_n_col = 2,
     cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_legend = T,
     legend_text = 8,
      axis_text = 8,
     axis_title = 8,
      show_plot = NA,
     return_plot = NA,
      save_plot = NA,
      save_param = list(),
     default_save_name = "spatDimCellPlot"
   )
Arguments
   gobject
                    giotto object
   plot_alignment direction to align plot
   spat_enr_names names of spatial enrichment results to include
   cell_annotation_values
                    numeric cell annotation columns
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
    sdimx
                    = spatial dimension to use on x-axis
   sdimy
                    = spatial dimension to use on y-axis
   cell_color_gradient
                    vector with 3 colors for numeric data
```

midpoint for color gradient

vector with lower and upper limits

gradient\_midpoint

gradient\_limits

select\_cell\_groups

select subset of cells/clusters based on cell\_color parameter

select subset of cells based on cell IDs

dim\_point\_size size of points in dim. reduction space

select\_cells

dim\_point\_border\_col border color of points in dim. reduction space dim\_point\_border\_stroke border stroke of points in dim. reduction space spat\_point\_size size of spatial points spat\_point\_border\_col border color of spatial points spat\_point\_border\_stroke border stroke of spatial points dim\_show\_cluster\_center show the center of each cluster dim\_show\_center\_label provide a label for each cluster dim\_center\_point\_size size of the center point dim\_center\_point\_border\_col border color of center point dim\_center\_point\_border\_stroke stroke size of center point dim\_label\_size size of the center label dim\_label\_fontface font of the center label spat\_show\_cluster\_center show the center of each cluster spat\_show\_center\_label provide a label for each cluster spat\_center\_point\_size size of the center point spat\_label\_size size of the center label spat\_label\_fontface font of the center label show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) nn\_network\_name name of NN network to use, if show\_NN\_network = TRUE dim\_edge\_alpha column to use for alpha of the edges spat\_show\_network show spatial network spatial\_network\_name name of spatial network to use spat\_network\_color color of spatial network

```
spat_show_grid show spatial grid
{\tt spatial\_grid\_name}
                  name of spatial grid to use
spat_grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
dim_other_point_size
                  size of not selected dim cells
spat_other_point_size
                  size of not selected spat cells
spat_other_cells_alpha
                  alpha of not selected spat cells
coord_fix_ratio
                  ratio for coordinates
                  cowplot param: how many columns
cow_n_col
                  cowplot param: relative height
cow_rel_h
                  cowplot param: relative width
cow_rel_w
cow_align
                  cowplot param: how to align
show_legend
                  show legend
legend_text
                  size of legend text
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

### **Details**

Description of parameters.

#### Value

ggplot

#### **Examples**

```
spatDimCellPlot(gobject)
```

spatDimCellPlot2D

spatDimCellPlot2D

#### **Description**

Visualize numerical features of cells according to spatial AND dimension reduction coordinates in 2D

```
spatDimCellPlot2D(
 gobject,
 plot_alignment = c("vertical", "horizontal"),
  spat_enr_names = NULL,
  cell_annotation_values,
  dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 dim_point_size = 1,
  dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 dim_show_cluster_center = F,
 dim_show_center_label = T,
 dim_center_point_size = 4,
  dim_center_point_border_col = "black",
 dim_center_point_border_stroke = 0.1,
 dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_center_point_border_col = "black",
  spat_center_point_border_stroke = 0.1,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
 nn_network_to_use = "sNN",
 nn_network_name = "sNN.pca",
 dim_edge_alpha = 0.5,
  spat_show_network = F,
```

```
spatial_network_name = "spatial_network",
      spat_network_color = "red",
      spat_network_alpha = 0.5,
      spat_show_grid = F,
      spatial_grid_name = "spatial_grid",
      spat_grid_color = "green",
      show_other_cells = TRUE,
     other_cell_color = "grey"
     dim_other_point_size = 0.5,
      spat_other_point_size = 0.5,
      spat_other_cells_alpha = 0.5,
     coord_fix_ratio = NULL,
      cow_n_col = 2,
     cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_legend = T,
     legend_text = 8,
      axis_text = 8,
     axis_title = 8,
      show_plot = NA,
     return_plot = NA,
      save_plot = NA,
      save_param = list(),
     default_save_name = "spatDimCellPlot2D"
   )
Arguments
   gobject
                    giotto object
   plot_alignment direction to align plot
   spat_enr_names names of spatial enrichment results to include
   cell_annotation_values
                    numeric cell annotation columns
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
    sdimx
                    = spatial dimension to use on x-axis
   sdimy
                    = spatial dimension to use on y-axis
   cell_color_gradient
                    vector with 3 colors for numeric data
   gradient_midpoint
                    midpoint for color gradient
   gradient_limits
```

vector with lower and upper limits

select subset of cells/clusters based on cell\_color parameter

select\_cell\_groups

select subset of cells based on cell IDs

select\_cells

dim\_point\_size size of points in dim. reduction space dim\_point\_border\_col border color of points in dim. reduction space dim\_point\_border\_stroke border stroke of points in dim. reduction space spat\_point\_size size of spatial points spat\_point\_border\_col border color of spatial points spat\_point\_border\_stroke border stroke of spatial points dim\_show\_cluster\_center show the center of each cluster dim\_show\_center\_label provide a label for each cluster dim\_center\_point\_size size of the center point dim\_center\_point\_border\_col border color of center point dim\_center\_point\_border\_stroke stroke size of center point dim\_label\_size size of the center label dim\_label\_fontface font of the center label spat\_show\_cluster\_center show the center of each cluster spat\_show\_center\_label provide a label for each cluster spat\_center\_point\_size size of the center point spat\_label\_size size of the center label spat\_label\_fontface font of the center label show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) nn\_network\_name name of NN network to use, if show\_NN\_network = TRUE dim\_edge\_alpha column to use for alpha of the edges spat\_show\_network show spatial network spatial\_network\_name name of spatial network to use spat\_network\_color color of spatial network

```
spat_show_grid show spatial grid
spatial_grid_name
                  name of spatial grid to use
spat_grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
dim_other_point_size
                  size of not selected dim cells
spat_other_point_size
                  size of not selected spat cells
spat_other_cells_alpha
                  alpha of not selected spat cells
coord_fix_ratio
                  ratio for coordinates
                  cowplot param: how many columns
cow_n_col
                  cowplot param: relative height
cow_rel_h
                  cowplot param: relative width
cow_rel_w
cow_align
                  cowplot param: how to align
show_legend
                  show legend
legend_text
                  size of legend text
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

### **Details**

Description of parameters.

#### Value

ggplot

#### **Examples**

```
spatDimCellPlot2D(gobject)
```

214 spatDimGenePlot

spatDimGenePlot

spatDimGenePlot

### **Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

### Usage

```
spatDimGenePlot(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("vertical", "horizontal"),
 genes,
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  show_NN_network = F,
  show_spatial_network = F,
  show_spatial_grid = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
  edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
  spatial_network_name = "spatial_network",
  spatial_grid_name = "spatial_grid",
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 midpoint = 0,
 genes_high_color = "red",
  genes_mid_color = "white";
 genes_low_color = "blue",
 cow_n_col = 2,
 cow_rel_h = 1,
  cow_rel_w = 1,
 cow_align = "h",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "spatDimGenePlot"
```

#### **Arguments**

gobject giotto object

215

expression\_values gene expression values to use plot\_alignment direction to align plot genes genes to show dim\_reduction\_to\_use dimension reduction to use dim\_reduction\_name dimension reduction name dim1\_to\_use dimension to use on x-axis dimension to use on y-axis dim2\_to\_use point\_size size of point (cell) dim\_point\_border\_col color of border around points dim\_point\_border\_stroke stroke size of border around points show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name edge\_alpha\_dim dim reduction plot: column to use for alpha of the edges scale\_alpha\_with\_expression scale expression with ggplot alpha parameter spatial\_network\_name name of spatial network to use spatial\_grid\_name name of spatial grid to use spat\_point\_size spatial plot: point size spat\_point\_border\_col color of border around points  $spat\_point\_border\_stroke$ stroke size of border around points midpoint size of point (cell) cowplot param: how many columns cow\_n\_col cow\_rel\_h cowplot param: relative height cowplot param: relative width cow\_rel\_w cowplot param: how to align cow\_align show\_legend show legend show plots show\_plot return ggplot object return\_plot save\_plot directly save the plot [boolean] save\_param list of saving parameters from all\_plots\_save\_function() default\_save\_name default save name for saving, don't change, change save\_name in save\_param

dim\_point\_size dim reduction plot: point size

216 spatDimGenePlot2D

#### **Details**

Description of parameters.

#### Value

ggplot

#### See Also

spatDimGenePlot3D

#### **Examples**

```
spatDimGenePlot(gobject)
```

spatDimGenePlot2D

spatDimGenePlot2D

### **Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

```
spatDimGenePlot2D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("vertical", "horizontal"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  show_NN_network = F,
  show_spatial_network = F,
  show_spatial_grid = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 edge_alpha_dim = NULL,
 scale_alpha_with_expression = FALSE,
  spatial_network_name = "spatial_network",
 spatial_grid_name = "spatial_grid",
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 midpoint = 0,
 genes_high_color = "red",
  genes_mid_color = "white",
```

spatDimGenePlot2D 217

```
genes_low_color = "blue",
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_legend = T,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimGenePlot2D"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
    plot_alignment direction to align plot
    genes
                     genes to show
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    point_size
                     size of point (cell)
    dim_point_border_col
                     color of border around points
    dim_point_border_stroke
                     stroke size of border around points
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
    edge_alpha_dim dim reduction plot: column to use for alpha of the edges
    scale_alpha_with_expression
                     scale expression with ggplot alpha parameter
    spatial_network_name
                     name of spatial network to use
    spatial_grid_name
                     name of spatial grid to use
    spat_point_size
                     spatial plot: point size
    spat_point_border_col
                     color of border around points
    spat_point_border_stroke
                     stroke size of border around points
```

218 spatDimGenePlot3D

```
size of point (cell)
midpoint
                  cowplot param: how many columns
cow_n_col
cow_rel_h
                  cowplot param: relative height
cow_rel_w
                  cowplot param: relative width
cow_align
                  cowplot param: how to align
show_legend
                  show legend
show_plot
                  show plots
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
dim_point_size dim reduction plot: point size
```

#### **Details**

Description of parameters.

#### Value

ggplot

#### See Also

```
spatDimGenePlot3D
```

#### **Examples**

```
spatDimGenePlot2D(gobject)
```

```
spatDimGenePlot3D
```

## Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

```
spatDimGenePlot3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("horizontal", "vertical"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = NULL,
  sdimx = "sdimx",
```

spatDimGenePlot3D 219

```
sdimz = "sdimz",
     genes,
      cluster_column = NULL,
      select_cell_groups = NULL,
      select_cells = NULL,
      show_other_cells = T,
     other_cell_color = "lightgrey",
     other_point_size = 1.5,
      show_NN_network = F,
     nn_network_to_use = "sNN",
     network_name = "sNN.pca",
     label_size = 16,
     genes_low_color = "blue",
     genes_mid_color = "white",
      genes_high_color = "red",
     dim_point_size = 3,
     nn_network_alpha = 0.5,
      show_spatial_network = F,
      spatial_network_name = "spatial_network",
     network_color = "lightgray",
      spatial_network_alpha = 0.5,
      show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_grid_alpha = 0.5,
      spatial_point_size = 3,
      legend_text_size = 12,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
     x_{ticks} = NULL,
     y_ticks = NULL,
     z_{ticks} = NULL,
     show_plot = NA,
     return_plot = NA,
     save_plot = NA,
     save_param = list(),
     default_save_name = "spatDimGenePlot3D"
   )
Arguments
   gobject
                    giotto object
   expression_values
                    gene expression values to use
   plot_alignment direction to align plot
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
                    dimension to use on x-axis
   dim1_to_use
   dim2_to_use
                    dimension to use on y-axis
```

sdimy = "sdimy",

220 spatDimGenePlot3D

dim3\_to\_use dimension to use on z-axis

genes genes to show

show\_NN\_network

show underlying NN network

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

dim\_point\_size dim reduction plot: point size

spatial\_network\_name

name of spatial network to use

spatial\_grid\_name

name of spatial grid to use

spatial\_point\_size

spatial plot: point size

show\_plot show plots

return\_plot return plotly object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

edge\_alpha\_dim dim reduction plot: column to use for alpha of the edges

scale\_alpha\_with\_expression

scale expression with ggplot alpha parameter

point\_size size of point (cell)

show\_legend show legend

# Details

Description of parameters.

## Value

plotly

## **Examples**

spatDimGenePlot3D(gobject)

spatDimPlot

spatDimPlot

#### **Description**

Visualize cells according to spatial AND dimension reduction coordinates 2D

```
spatDimPlot(
 gobject,
 plot_alignment = c("vertical", "horizontal"),
  dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_color = NULL,
  color_as_factor = T,
 cell_color_code = NULL,
 cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 dim_point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 dim_show_cluster_center = F,
 dim_show_center_label = T,
 dim_center_point_size = 4,
 dim_center_point_border_col = "black",
 dim_center_point_border_stroke = 0.1,
 dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
  nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 nn_network_alpha = 0.05,
  show_spatial_network = F,
  spat_network_name = "spatial_network",
```

```
spat_network_color = "blue",
      spat_network_alpha = 0.5,
      show_spatial_grid = F,
      spat_grid_name = "spatial_grid",
      spat_grid_color = "blue",
      show_other_cells = T,
      other_cell_color = "lightgrey",
      dim_other_point_size = 1,
      spat_other_point_size = 1,
      spat_other_cells_alpha = 0.5,
      dim_show_legend = F,
      spat_show_legend = F,
      legend_text = 8,
      axis_text = 8,
      axis_title = 8,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimPlot"
    )
Arguments
                     giotto object
   gobject
   plot_alignment direction to align plot
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
   dim1_to_use
                     dimension to use on x-axis
   dim2_to_use
                     dimension to use on y-axis
                     = spatial dimension to use on x-axis
    sdimx
                     = spatial dimension to use on y-axis
    sdimy
    spat_enr_names names of spatial enrichment results to include
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
```

select subset of cells based on cell IDs

select\_cells

dim\_point\_size size of points in dim. reduction space

dim\_point\_border\_col

border color of points in dim. reduction space dim\_point\_border\_stroke border stroke of points in dim. reduction space spat\_point\_size size of spatial points spat\_point\_border\_col border color of spatial points spat\_point\_border\_stroke border stroke of spatial points dim\_show\_cluster\_center show the center of each cluster dim\_show\_center\_label provide a label for each cluster  ${\tt dim\_center\_point\_size}$ size of the center point dim\_center\_point\_border\_col border color of center point dim\_center\_point\_border\_stroke stroke size of center point dim\_label\_size size of the center label dim\_label\_fontface font of the center label spat\_show\_cluster\_center show the center of each cluster spat\_show\_center\_label provide a label for each cluster spat\_center\_point\_size size of the center point spat\_label\_size size of the center label spat\_label\_fontface font of the center label show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) network\_name name of NN network to use, if show\_NN\_network = TRUE nn\_network\_alpha column to use for alpha of the edges show\_spatial\_network show spatial network spat\_network\_name name of spatial network to use spat\_network\_color color of spatial network show\_spatial\_grid show spatial grid

```
spat_grid_name name of spatial grid to use
spat_grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
{\tt dim\_other\_point\_size}
                  size of not selected dim cells
spat_other_point_size
                  size of not selected spat cells
spat_other_cells_alpha
                  alpha of not selected spat cells
dim_show_legend
                  show legend of dimension reduction plot
spat_show_legend
                  show legend of spatial plot
legend_text
                  size of legend text
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

## **Details**

Description of parameters.

## Value

ggplot

## See Also

spatDimPlot2D and spatDimPlot3D for 3D visualization.

## **Examples**

```
spatDimPlot(gobject)
```

spatDimPlot2D

spatDimPlot2D

#### **Description**

Visualize cells according to spatial AND dimension reduction coordinates 2D

```
spatDimPlot2D(
 gobject,
 plot_alignment = c("vertical", "horizontal"),
  dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_color = NULL,
  color_as_factor = T,
 cell_color_code = NULL,
 cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 dim_point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 dim_show_cluster_center = F,
 dim_show_center_label = T,
 dim_center_point_size = 4,
 dim_center_point_border_col = "black",
 dim_center_point_border_stroke = 0.1,
 dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
  nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 nn_network_alpha = 0.05,
  show_spatial_network = F,
  spat_network_name = "spatial_network",
```

```
spat_network_color = "blue",
      spat_network_alpha = 0.5,
      show_spatial_grid = F,
      spat_grid_name = "spatial_grid",
      spat_grid_color = "blue",
      show_other_cells = T,
      other_cell_color = "lightgrey",
      dim_other_point_size = 1,
      spat_other_point_size = 1,
      spat_other_cells_alpha = 0.5,
      dim_show_legend = F,
      spat_show_legend = F,
      legend_text = 8,
      axis_text = 8,
      axis_title = 8,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimPlot2D"
    )
Arguments
                     giotto object
   gobject
   plot_alignment direction to align plot
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
   dim1_to_use
                     dimension to use on x-axis
   dim2_to_use
                     dimension to use on y-axis
                     = spatial dimension to use on x-axis
    sdimx
                     = spatial dimension to use on y-axis
    sdimy
    spat_enr_names names of spatial enrichment results to include
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
```

dim\_point\_size size of points in dim. reduction space

dim\_point\_border\_col

border color of points in dim. reduction space dim\_point\_border\_stroke border stroke of points in dim. reduction space spat\_point\_size size of spatial points spat\_point\_border\_col border color of spatial points spat\_point\_border\_stroke border stroke of spatial points dim\_show\_cluster\_center show the center of each cluster dim\_show\_center\_label provide a label for each cluster  ${\tt dim\_center\_point\_size}$ size of the center point dim\_center\_point\_border\_col border color of center point dim\_center\_point\_border\_stroke stroke size of center point dim\_label\_size size of the center label dim\_label\_fontface font of the center label spat\_show\_cluster\_center show the center of each cluster spat\_show\_center\_label provide a label for each cluster spat\_center\_point\_size size of the center point spat\_label\_size size of the center label spat\_label\_fontface font of the center label show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) network\_name name of NN network to use, if show\_NN\_network = TRUE nn\_network\_alpha column to use for alpha of the edges show\_spatial\_network show spatial network spat\_network\_name name of spatial network to use spat\_network\_color color of spatial network show\_spatial\_grid show spatial grid

```
spat_grid_name name of spatial grid to use
spat_grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
{\tt dim\_other\_point\_size}
                  size of not selected dim cells
spat_other_point_size
                  size of not selected spat cells
spat_other_cells_alpha
                  alpha of not selected spat cells
dim_show_legend
                  show legend of dimension reduction plot
spat_show_legend
                  show legend of spatial plot
legend_text
                  size of legend text
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

## **Details**

Description of parameters.

## Value

ggplot

## See Also

spatDimPlot3D

## **Examples**

spatDimPlot2D(gobject)

spatDimPlot3D

spatDimPlot3D

#### **Description**

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

```
spatDimPlot3D(
 gobject,
 plot_alignment = c("horizontal", "vertical"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
 dim3_to_use = 3,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
  show_cluster_center = F,
  show_center_label = T,
 center_point_size = 4,
 label_size = 16,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1.5,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
 dim_point_size = 3,
 nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
 network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
 x_ticks = NULL,
 y_ticks = NULL,
  z_ticks = NULL,
```

```
legend_text_size = 12,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimPlot3D"
Arguments
    gobject
                      giotto object
    plot_alignment direction to align plot
    dim_reduction_to_use
                      dimension reduction to use
    dim_reduction_name
                      dimension reduction name
    dim1_to_use
                      dimension to use on x-axis
    dim2_to_use
                      dimension to use on y-axis
                      dimension to use on z-axis
    dim3_to_use
    sdimx
                      = spatial dimension to use on x-axis
    sdimy
                      = spatial dimension to use on y-axis
                      = spatial dimension to use on z-axis
    sdimz
    show_NN_network
                      show underlying NN network
    nn_network_to_use
                      type of NN network to use (kNN vs sNN)
                      name of NN network to use, if show_NN_network = TRUE
    network_name
    show_cluster_center
                      show the center of each cluster
    show_center_label
                      provide a label for each cluster
    center_point_size
                      size of the center point
    label_size
                      size of the center label
    select_cell_groups
                      select subset of cells/clusters based on cell_color parameter
                      select subset of cells based on cell IDs
    select_cells
    show_other_cells
                      display not selected cells
    other_cell_color
                      color of not selected cells
    other_point_size
                      size of not selected cells
    cell_color
                      color for cells (see details)
    color_as_factor
                      convert color column to factor
    cell_color_code
                      named vector with colors
```

dim\_point\_size size of points in dim. reduction space nn\_network\_alpha column to use for alpha of the edges show\_spatial\_network show spatial network spatial\_network\_name name of spatial network to use spatial\_network\_alpha alpha of spatial network show\_spatial\_grid show spatial grid spatial\_grid\_name name of spatial grid to use spatial\_grid\_color color of spatial grid spatial\_point\_size size of spatial points show plot show\_plot return\_plot return ggplot object directly save the plot [boolean] save\_plot list of saving parameters from all\_plots\_save\_function() save\_param default\_save\_name default save name for saving, don't change, change save\_name in save\_param dim\_point\_border\_col border color of points in dim. reduction space dim\_point\_border\_stroke border stroke of points in dim. reduction space spatial\_network\_color color of spatial network spatial\_other\_point\_size size of not selected spatial points  ${\tt spatial\_other\_cells\_alpha}$ alpha of not selected spatial points dim\_other\_point\_size size of not selected dim. reduction points show\_legend show legend

#### **Details**

Description of parameters.

#### Value

plotly

#### **Examples**

spatDimPlot3D(gobject)

232 spatGenePlot

spatGenePlot

spatGenePlot

## **Description**

Visualize cells and gene expression according to spatial coordinates

## Usage

```
spatGenePlot(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 genes,
 genes_high_color = "darkred",
  genes_mid_color = "white",
 genes_low_color = "darkblue",
 show_network = F,
 network_color = NULL,
 spatial_network_name = "spatial_network",
 edge_alpha = NULL,
  show\_grid = F,
 grid_color = NULL,
  spatial_grid_name = "spatial_grid",
 midpoint = 0,
  scale_alpha_with_expression = FALSE,
 point_size = 1,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_legend = T,
  cow_n_col = 2,
 cow_rel_h = 1,
 cow_rel_w = 1,
 cow_align = "h",
 show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "spatGenePlot"
)
```

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
genes_high_color
color represents high gene expression
genes_mid_color
color represents middle gene expression
```

spatGenePlot 233

```
genes_low_color
                  color represents low gene expression
show_network
                  show underlying spatial network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
midpoint
                  expression midpoint
scale_alpha_with_expression
                  scale expression with ggplot alpha parameter
                  size of point (cell)
point_size
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_legend
                  show legend
cow_n_col
                  cowplot param: how many columns
cow_rel_h
                  cowplot param: relative height
cow_rel_w
                  cowplot param: relative width
cow_align
                  cowplot param: how to align
show_plot
                  show plots
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  parameters for cowplot::save_plot()
. . .
```

#### **Details**

Description of parameters.

#### Value

ggplot

## See Also

 ${\tt spatGenePlot3D} \ and \ {\tt spatGenePlot2D}$ 

## **Examples**

```
spatGenePlot(gobject)
```

234 spatGenePlot2D

spatGenePlot2D

spatGenePlot2D

#### **Description**

Visualize cells and gene expression according to spatial coordinates

#### Usage

```
spatGenePlot2D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 genes,
 genes_high_color = "darkred",
  genes_mid_color = "white",
 genes_low_color = "darkblue",
 show_network = F,
 network_color = NULL,
 spatial_network_name = "spatial_network",
 edge_alpha = NULL,
  show\_grid = F,
 grid_color = NULL,
  spatial_grid_name = "spatial_grid",
 midpoint = 0,
  scale_alpha_with_expression = FALSE,
 point_size = 1,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_legend = T,
  cow_n_col = 2,
 cow_rel_h = 1,
 cow_rel_w = 1,
 cow_align = "h",
 show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "spatGenePlot2D"
)
```

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
genes_high_color
color represents high gene expression
genes_mid_color
color represents middle gene expression
```

spatGenePlot2D 235

```
genes_low_color
                  color represents low gene expression
show_network
                  show underlying spatial network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
midpoint
                  expression midpoint
scale_alpha_with_expression
                  scale expression with ggplot alpha parameter
                  size of point (cell)
point_size
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_legend
                  show legend
cow_n_col
                  cowplot param: how many columns
cow_rel_h
                  cowplot param: relative height
cow_rel_w
                  cowplot param: relative width
cow_align
                  cowplot param: how to align
show_plot
                  show plots
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  parameters for cowplot::save_plot()
. . .
```

#### **Details**

Description of parameters.

#### Value

ggplot

## See Also

 ${\tt spatGenePlot3D}$ 

## **Examples**

spatGenePlot2D(gobject)

236 spatGenePlot3D

spatGenePlot3D spatGenePlot3D

#### **Description**

Visualize cells and gene expression according to spatial coordinates

#### Usage

```
spatGenePlot3D(
 gobject,
  expression_values = c("normalized", "scaled", "custom"),
 genes,
 show_network = F,
 network_color = NULL,
  spatial_network_name = "spatial_network",
 edge_alpha = NULL,
 show\_grid = F,
 cluster_column = NULL,
  select_cell_groups = NULL,
 select_cells = NULL,
 show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
 genes_high_color = NULL,
 genes_mid_color = "white",
 genes_low_color = "blue",
  spatial_grid_name = "spatial_grid",
 point_size = 2,
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
 x_ticks = NULL,
 y_ticks = NULL,
 z_ticks = NULL,
 show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "spatGenePlot3D"
)
```

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
show_network show underlying spatial network
network_color color of spatial network
```

Spatial\_AEH 237

genes\_high\_color

color represents high gene expression

genes\_mid\_color

color represents middle gene expression

genes\_low\_color

color represents low gene expression

spatial\_grid\_name

name of spatial grid to use

point\_size size of point (cell)
show\_legend show\_plot show plots

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

grid\_color color of spatial grid midpoint expression midpoint scale\_alpha\_with\_expression

scale expression with ggplot alpha parameter

... parameters for cowplot::save\_plot()

#### **Details**

Description of parameters.

# Value

ggplot

## **Examples**

spatGenePlot3D(gobject)

Spatial\_AEH Spatial\_AEH

## Description

calculate automatic expression histology with spatialDE method

Spatial\_DE

## Usage

```
Spatial_AEH(
  gobject = NULL,
  results = NULL,
  pattern_num = 5,
  1 = 1.05,
  show\_AEH = T,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_alpha = 1,
  low_color = "blue",
  mid_color = "white",
  high_color = "red",
  midpoint = 0,
  python_path = NULL
)
```

# Arguments

gobject Giotto object
results output from spatial\_DE
pattern\_num the number of gene expression patterns
show\_AEH show AEH plot

python\_path specify specific path to python if required

# Details

Description.

#### Value

a list or a dataframe of SVs

## **Examples**

```
Spatial_DE(gobject)
```

Spatial\_DE Spatial\_DE

## Description

calculate spatial varible genes with spatialDE method

spatPlot 239

## Usage

```
Spatial_DE(
  gobject = NULL,
  show_plot = T,
  size = c(4, 2, 1),
  color = c("blue", "green", "red"),
  sig_alpha = 0.5,
  unsig_alpha = 0.5,
  python_path = NULL
)
```

#### **Arguments**

gobject Giotto object
show\_plot show FSV plot
python\_path specific path to python if required

## **Details**

Description.

#### Value

a list or a dataframe of SVs

## **Examples**

```
Spatial_DE(gobject)
```

spatPlot

spatPlot

## Description

Visualize cells according to spatial coordinates

```
spatPlot(
  gobject,
  group_by = NULL,
  group_by_subset = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
```

240 spatPlot

```
select_cell_groups = NULL,
select_cells = NULL,
point_size = 3,
point_border_col = "black",
point_border_stroke = 0.1,
show_cluster_center = F,
show_center_label = F,
center_point_size = 4,
center_point_border_col = "black",
center_point_border_stroke = 0.1,
label_size = 4,
label_fontface = "bold",
show_network = F,
spatial_network_name = "spatial_network",
network_color = NULL,
network\_alpha = 1,
show_grid = F,
spatial_grid_name = "spatial_grid",
grid_color = NULL,
show_other_cells = T,
other_cell_color = "lightgrey",
other_point_size = 1,
other_cells_alpha = 0.1,
coord_fix_ratio = NULL,
title = NULL,
show_legend = T,
legend_text = 8,
axis_text = 8,
axis_title = 8,
cow_n_col = 2,
cow_rel_h = 1,
cow_rel_w = 1,
cow_align = "h",
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spatPlot"
```

```
gobject
                  giotto object
group_by_subset
                  subset the group_by factor column
                  x-axis dimension name (default = 'sdimx')
sdimx
sdimy
                  y-axis dimension name (default = 'sdimy')
spat_enr_names names of spatial enrichment results to include
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
```

spatPlot 241

cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select subset of cells based on cell IDs select\_cells point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels show\_network show underlying spatial network spatial\_network\_name name of spatial network to use network\_color color of spatial network network\_alpha alpha of spatial network show\_grid show spatial grid spatial\_grid\_name name of spatial grid to use grid\_color color of spatial grid show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size point size of not selected cells other\_cells\_alpha alpha of not selected cells coord\_fix\_ratio fix ratio between x and y-axis title title of plot show\_legend show legend

legend\_text

size of legend text

242 spatPlot2D

```
size of axis text
axis_text
                  size of axis title
axis_title
cow_n_col
                  cowplot param: how many columns
cow_rel_h
                  cowplot param: relative height
cow_rel_w
                  cowplot param: relative width
cow_align
                  cowplot param: how to align
show_plot
                  show plot
                  return ggplot object
return_plot
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
groub_by
                  create multiple plots based on cell annotation column
```

#### **Details**

Description of parameters.

## Value

ggplot

#### See Also

```
spatPlot3D
```

## **Examples**

```
spatPlot(gobject)
```

spatPlot2D

## Description

Visualize cells according to spatial coordinates

spatPlot2D

```
spatPlot2D(
  gobject,
  group_by = NULL,
  group_by_subset = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
```

spatPlot2D 243

```
cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 point_size = 3,
 point_border_col = "black",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
  center_point_size = 4,
 center_point_border_col = "black",
  center_point_border_stroke = 0.1,
 label_size = 4,
  label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
  show\_grid = F,
  spatial_grid_name = "spatial_grid",
 grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
 other_cells_alpha = 0.1,
  coord_fix_ratio = NULL,
  title = NULL,
  show_legend = T,
 legend_text = 8,
 axis_text = 8,
 axis_title = 8,
 cow_n_col = 2,
 cow_rel_h = 1,
 cow_rel_w = 1,
 cow_align = "h",
 show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "spatPlot2D"
)
```

244 spatPlot2D

color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select subset of cells based on cell IDs select\_cells point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels show\_network show underlying spatial network spatial\_network\_name name of spatial network to use network\_color color of spatial network network\_alpha alpha of spatial network show\_grid show spatial grid spatial\_grid\_name name of spatial grid to use color of spatial grid grid\_color show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size point size of not selected cells other\_cells\_alpha alpha of not selected cells coord\_fix\_ratio

fix ratio between x and y-axis

title of plot

title

spatPlot2D\_single 245

```
show_legend
                  show legend
legend_text
                  size of legend text
axis\_text
                  size of axis text
axis_title
                  size of axis title
                  cowplot param: how many columns
cow_n_col
cow_rel_h
                  cowplot param: relative height
cow_rel_w
                  cowplot param: relative width
                  cowplot param: how to align
cow_align
{\sf show\_plot}
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
groub_by
                  create multiple plots based on cell annotation column
```

#### **Details**

Description of parameters.

## Value

ggplot

## See Also

spatPlot3D

## **Examples**

spatPlot2D(gobject)

## **Description**

Visualize cells according to spatial coordinates

246 spatPlot2D\_single

#### Usage

```
spatPlot2D_single(
 gobject,
 sdimx = "sdimx",
 sdimy = "sdimy",
  spat_enr_names = NULL,
  cell_color = NULL,
 color_as_factor = T,
 cell_color_code = NULL,
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
 center_point_size = 4,
 center_point_border_col = "black",
  center_point_border_stroke = 0.1,
 label_size = 4,
 label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
  show\_grid = F,
  spatial_grid_name = "spatial_grid",
 grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
 other_cells_alpha = 0.1,
  coord_fix_ratio = NULL,
  title = NULL,
  show_legend = T,
 legend_text = 8,
 axis_text = 8,
 axis_title = 8,
 show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "spatPlot2D_single"
)
```

```
gobject giotto object
sdimx x-axis dimension name (default = 'sdimx')
```

spatPlot2D\_single 247

sdimy y-axis dimension name (default = 'sdimy') spat\_enr\_names names of spatial enrichment results to include color for cells (see details) cell\_color color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell color parameter select subset of cells based on cell IDs select\_cells size of point (cell) point\_size point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels show\_network show underlying spatial network spatial\_network\_name name of spatial network to use network\_color color of spatial network network\_alpha alpha of spatial network show\_grid show spatial grid  ${\tt spatial\_grid\_name}$ name of spatial grid to use color of spatial grid grid\_color show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size point size of not selected cells other\_cells\_alpha alpha of not selected cells

248 spatPlot3D

```
coord_fix_ratio
                  fix ratio between x and y-axis
                  title of plot
title
                  show legend
show_legend
                  size of legend text
legend_text
axis_text
                  size of axis text
                  size of axis title
axis_title
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

## **Details**

Description of parameters.

#### Value

ggplot

#### See Also

spatPlot3D

## **Examples**

```
spatPlot2D_single(gobject)
```

spatPlot3D spatPlot3D

## Description

Visualize cells according to spatial coordinates

```
spatPlot3D(
  gobject,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  point_size = 3,
  cell_color = NULL,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
```

spatPlot3D 249

```
show_other_cells = T,
other_cell_color = "lightgrey",
other_point_size = 0.5,
show_network = F,
network_color = NULL,
network_alpha = 1,
other_cell_alpha = 0.5,
spatial_network_name = "spatial_network",
show\_grid = F,
grid_color = NULL,
spatial_grid_name = "spatial_grid",
title = "",
show_legend = T,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
z_{ticks} = NULL,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spat3D"
```

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimy
sdimz
                  z-axis dimension name (default = 'sdimy')
point_size
                  size of point (cell)
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
show_network
                  show underlying spatial network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
```

```
spatial_grid_name
                  name of spatial grid to use
title
                  title of plot
show_legend
                  show legend
axis_scale
                  the way to scale the axis
custom_ratio
                  customize the scale of the plot
                  set the number of ticks on the x-axis
x_ticks
                  set the number of ticks on the y-axis
y_ticks
z_ticks
                  set the number of ticks on the z-axis
                  show plot
show_plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

#### **Details**

Description of parameters.

#### Value

ggplot

## **Examples**

```
spatPlot3D(gobject)
```

```
specific Cell Cell communication Scores\\ specific Cell Cell communication Scores
```

## **Description**

Specific Cell-Cell communication scores based on spatial expression of interacting cells

```
specificCellCellcommunicationScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "cell_types",
  random_iter = 100,
  cell_type_1 = "astrocyte",
  cell_type_2 = "endothelial",
  gene_set_1,
  gene_set_2,
```

```
log2FC_addendum = 0.1,
min_observations = 2,
verbose = T
)
```

## Arguments

```
gobject
                  giotto object to use
spatial_network_name
                  spatial network to use for identifying interacting cells
cluster_column cluster column with cell type information
                  number of iterations
random_iter
cell_type_1
                  first cell type
cell_type_2
                  second cell type
                  first specific gene set from gene pairs
gene_set_1
                  second specific gene set from gene pairs
gene_set_2
log2FC_addendum
                  addendum to add when calculating log2FC
min_observations
                  minimum number of interactions needed to be considered
verbose
                  verbose
```

## **Details**

Details will follow.

#### Value

Cell-Cell communication scores for gene pairs based on spatial interaction

## **Examples**

```
{\tt specific Cell Cell communication Scores (gobject)}
```

# Description

Merge selected clusters based on pairwise correlation scores and size of cluster.

## Usage

```
split_dendrogram_in_two(dend)
```

#### **Arguments**

dend dendrogram object

252 stitchFieldCoordinates

#### Value

list of two dendrograms and height of node

## **Examples**

```
split_dendrogram_in_two(dend)
```

```
stitchFieldCoordinates
```

stitchFieldCoordinates

## **Description**

Helper function to stitch field coordinates together to form one complete picture

## Usage

```
stitchFieldCoordinates(
  location_file,
  offset_file,
  cumulate_offset_x = F,
  cumulate_offset_y = F,
  field_col = "Field of View",
  X_coord_col = "X",
  Y_coord_col = "Y",
  reverse_final_x = F,
  reverse_final_y = T
)
```

```
location_file location dataframe with X and Y coordinates
offset_file
                  dataframe that describes the offset for each field (see details)
cumulate_offset_x
                  (boolean) Do the x-axis offset values need to be cumulated?
cumulate_offset_y
                  (boolean) Do the y-axis offset values need to be cumulated?
field_col
                  column that indicates the field within the location_file
                  column that indicates the x coordinates
X_coord_col
Y_coord_col
                  column that indicates the x coordinates
reverse_final_x
                  (boolean) Do the final x coordinates need to be reversed?
reverse_final_y
                  (boolean) Do the final y coordinates need to be reversed?
```

subClusterCells 253

#### **Details**

Stitching of fields:

- 1. have cell locations: at least 3 columns: field, X, Y
- 2. create offset file: offset file has 3 columns: field, x\_offset, y\_offset
- 3. create new cell location file by stitching original cell locations with stitchFieldCoordinates
- 4. provide new cell location file to createGiottoObject

#### Value

Updated location dataframe with new X ['X\_final'] and Y ['Y\_final'] coordinates

#### **Examples**

```
stitchFieldCoordinates(gobject)
```

subClusterCells

subClusterCells

## **Description**

subcluster cells

```
subClusterCells(
  gobject,
  name = "sub_clus",
  cluster_method = c("leiden", "louvain_community", "louvain_multinet"),
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 1,
  gamma = 1,
  omega = 1,
  python_path = NULL,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T,
)
```

254 subClusterCells

#### **Arguments**

gobject giotto object

name name for new clustering result

cluster\_method clustering method to use

cluster\_column cluster column to subcluster

selected\_clusters

only do subclustering on these clusters

hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

 $hvg\_mean\_expr\_det$ 

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for createNearestNetwork

k\_neighbors number of k for createNearestNetwork

resolution resolution gamma gamma omega omega

python\_path specify specific path to python if required

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

 ${\tt network\_name} \qquad {\tt name} \ of \ NN \ network \ to \ use$ 

return\_gobject boolean: return giotto object (default = TRUE)

verbose verbose

... additional parameters

### **Details**

Description of Louvain clustering method.

#### Value

giotto object appended with new cluster

## **Examples**

subClusterCells(gobject)

subsetGiotto 255

subsetGiotto

subsetGiot to

# Description

subsets Giotto object including previous analyses.

# Usage

```
subsetGiotto(gobject, cell_ids = NULL, gene_ids = NULL, verbose = FALSE)
```

# Arguments

```
gobject giotto object
cell_ids cell IDs to keep
gene_ids gene IDs to keep
verbose be verbose
```

#### Value

giotto object

## **Examples**

```
subsetGiotto(gobject)
```

 ${\tt subsetGiottoLocs}$ 

subsetGiottoLocs

## **Description**

subsets Giotto object based on spatial locations

```
subsetGiottoLocs(
  gobject,
  x_max = NULL,
  x_min = NULL,
  y_max = NULL,
  y_min = NULL,
  z_max = NULL,
  z_min = NULL,
  return_gobject = T,
  verbose = FALSE
)
```

256 viewHMRFresults

## **Arguments**

gobject	giotto object
x_max	maximum x-coordinate
x_min	minimum x-coordinate
y_max	maximum y-coordinate
y_min	minimum y-coordinate
z_max	maximum z-coordinate
z_min	minimum z-coordinate
return_gobject	return Giotto object

## **Details**

if return\_gobject = FALSE, then a filtered combined metadata data.table will be returned

# Value

giotto object

# **Examples**

```
subsetGiottoLocs(gobject)
```

viewHMRFresults

viewHMRFresults

# Description

View results from doHMRF.

# Usage

```
viewHMRFresults(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

# **Arguments**

```
gobject giotto object

HMRFoutput HMRF output from doHMRF

k number of HMRF domains

betas_to_view results from different betas that you want to view
```

... paramters to visPlot()

viewHMRFresults2D 257

#### **Details**

Description ...

## Value

spatial plots with HMRF domains

## See Also

```
visPlot
```

#### **Examples**

viewHMRFresults(gobject)

viewHMRFresults2D

viewHMRFresults2D

# Description

View results from doHMRF.

# Usage

```
viewHMRFresults2D(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

# Arguments

gobject giotto object

HMRF output from doHMRF k number of HMRF domains

 $\verb|betas_to_view| \quad \textit{results from different betas that you want to view}$ 

... paramters to visPlot()

# **Details**

Description ...

### Value

spatial plots with HMRF domains

## See Also

```
spatPlot2D
```

258 viewHMRFresults3D

## **Examples**

```
viewHMRFresults2D(gobject)
```

viewHMRFresults3D

viewHMRFresults3D

# Description

View results from doHMRF.

# Usage

```
viewHMRFresults3D(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

# Arguments

```
gobject giotto object
```

HMRF output from doHMRF k number of HMRF domains

betas\_to\_view results from different betas that you want to view

... paramters to visPlot()

## **Details**

Description ...

# Value

spatial plots with HMRF domains

### See Also

```
spatPlot3D
```

# **Examples**

```
viewHMRFresults3D(gobject)
```

violinPlot 259

violinPlot

violinPlot

## **Description**

Creates heatmap based on identified clusters

## Usage

```
violinPlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column,
  cluster_custom_order = NULL,
  color_violin = c("genes", "cluster"),
  cluster_color_code = NULL,
  strip_position = c("top", "right", "left", "bottom"),
  strip\_text = 7,
  axis_text_x_size = 10,
  axis_text_y_size = 6,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "violinPlot"
)
```

```
gobject
                  giotto object
expression_values
                  expression values to use
                  genes to plot
genes
cluster_column name of column to use for clusters
cluster_custom_order
                  custom order of clusters
color_violin
                  color violin according to genes or clusters
cluster_color_code
                  color code for clusters
strip_position position of gene labels
strip_text
                  size of strip text
axis_text_x_size
                  size of x-axis text
axis_text_y_size
                  size of y-axis text
show_plot
                  show plot
return_plot
                  return ggplot object
```

260 visDimGenePlot

```
save_plot directly save the plot [boolean]
save_param list of saving parameters from all_plots_save_function()
default_save_name
    default save name for saving, don't change, change save_name in save_param
```

#### **Details**

Correlation heatmap of clusters vs genes.

#### Value

ggplot

# **Examples**

```
violinPlot(gobject)
```

visDimGenePlot

visDimGenePlot

## **Description**

Visualize cells and gene expression according to dimension reduction coordinates

```
visDimGenePlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
```

visDimGenePlot 261

```
cow_align = "h",
show_legend = T,
plot_method = c("ggplot", "plotly"),
show_plots = F
)
```

# Arguments

```
gobject
                 giotto object
expression_values
                 gene expression values to use
                 genes to show
genes
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
dim3 to use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
edge_alpha
                 column to use for alpha of the edges
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
point_size
                 size of point (cell)
point_border_col
                 color of border around points
point_border_stroke
                 stroke size of border around points
midpoint
                 size of point (cell)
cow_n_col
                 cowplot param: how many columns
                 cowplot param: relative height
cow_rel_h
                 cowplot param: relative width
cow_rel_w
                 cowplot param: how to align
cow_align
show_legend
                 show legend
show_plots
                 show plots
```

### **Details**

Description of parameters.

## Value

ggplot

# **Examples**

```
visDimGenePlot(gobject)
```

## **Description**

Visualize cells and gene expression according to dimension reduction coordinates

## Usage

```
visDimGenePlot_2D_ggplot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = "red",
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_legend = T,
  show_plots = F
```

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
dim_reduction_to_use
dimension reduction to use
dim_reduction_name
dimension reduction name
dim1_to_use
dimension to use on x-axis
```

```
dim2_to_use
                 dimension to use on y-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
edge_alpha
                 column to use for alpha of the edges
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
                 size of point (cell)
point_size
point_border_col
                 color of border around points
point_border_stroke
                 stroke size of border around points
                 size of point (cell)
midpoint
cow_n_col
                 cowplot param: how many columns
                 cowplot param: relative height
cow_rel_h
                 cowplot param: relative width
cow_rel_w
cow_align
                 cowplot param: how to align
show_legend
                 show legend
show_plots
                 show plots
```

### **Details**

Description of parameters.

## Value

ggplot

## **Examples**

```
visDimGenePlot_2D_ggplot(gobject)
```

## **Description**

Visualize cells and gene expression according to dimension reduction coordinates

#### Usage

```
visDimGenePlot_3D_plotly(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = 3,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_legend = T,
  show_plots = F
)
```

## **Arguments**

```
gobject
                 giotto object
expression_values
                 gene expression values to use
genes
                 genes to show
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
edge_alpha
                 column to use for alpha of the edges
point_size
                 size of point (cell)
show_legend
                 show legend
                 show plots
show_plots
```

### **Details**

Description of parameters.

visDimPlot 265

#### Value

ggplot

#### **Examples**

```
visDimGenePlot_3D_plotly(gobject)
```

visDimPlot

visDimPlot

### **Description**

Visualize cells according to dimension reduction coordinates

```
visDimPlot(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  plot_method = c("ggplot", "plotly"),
  show_legend = T,
  show_plot = F,
  return_plot = TRUE,
  save_plot = F,
  save_dir = NULL,
```

266 visDimPlot

```
save_folder = NULL,
      save_name = NULL,
      save_format = NULL,
      show_saved_plot = F,
    )
Arguments
    gobject
                     giotto object
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
                     dimension to use on z-axis
    dim3_to_use
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
                     size of center points
    label_size
                     size of labels
    label_fontface font of labels
    edge_alpha
                     column to use for alpha of the edges
    point_size
                     size of point (cell)
    point_border_col
                     color of border around points
    point_border_stroke
                     stroke size of border around points
    show_legend
                     show legend
    show_plot
                     show plot
    return_plot
                     return ggplot object
```

directly save the plot [boolean]

directory to save the plot

save\_plot
save\_dir

visDimPlot\_2D\_ggplot 267

#### **Details**

Description of parameters.

#### Value

ggplot or plotly

## **Examples**

```
visDimPlot(gobject)
```

```
visDimPlot_2D_ggplot visDimPlot_2D_ggplot
```

## **Description**

Visualize cells according to dimension reduction coordinates

```
visDimPlot_2D_ggplot(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
```

```
edge_alpha = NULL,
      point_size = 1,
      point_border_col = "black",
      point_border_stroke = 0.1,
      show_legend = T,
      show_plot = F,
      return_plot = TRUE,
      save_plot = F,
      save_dir = NULL,
      save_folder = NULL,
      save_name = NULL,
      save_format = NULL,
      show_saved_plot = F,
    )
Arguments
    gobject
                     giotto object
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
                     color for cells (see details)
    cell_color
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
```

size of center points

visDimPlot\_2D\_plotly 269

#### **Details**

Description of parameters.

#### Value

ggplot

## **Examples**

```
visDimPlot_2D_ggplot(gobject)
```

```
visDimPlot_2D_plotly visDimPlot_2D_plotly
```

## **Description**

Visualize cells according to dimension reduction coordinates

```
visDimPlot_2D_plotly(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  color_as_factor = T,
  cell_color = NULL,
  cell_color_code = NULL,
  show_cluster_center = F,
  show_center_label = T,
```

```
center_point_size = 4,
label_size = 4,
edge_alpha = NULL,
point_size = 5
)
```

## **Arguments**

```
gobject
                 giotto object
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
                 dimension to use on x-axis
dim1_to_use
dim2_to_use
                 dimension to use on y-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
color_as_factor
                 convert color column to factor
cell_color
                 color for cells (see details)
cell_color_code
                 named vector with colors
show_cluster_center
                 plot center of selected clusters
show_center_label
                 plot label of selected clusters
center_point_size
                 size of center points
label_size
                 size of labels
edge_alpha
                 column to use for alpha of the edges
point_size
                 size of point (cell)
```

#### **Details**

Description of parameters.

### Value

plotly

# Examples

```
\verb|visDimPlot_2D_plotly(gobject)| \\
```

```
visDimPlot_3D_plotly
```

## **Description**

Visualize cells according to dimension reduction coordinates

### Usage

```
visDimPlot_3D_plotly(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = 3,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  color_as_factor = T,
  cell_color = NULL,
  cell_color_code = NULL,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 4,
  edge_alpha = NULL,
  point_size = 1
```

```
giotto object
gobject
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
                 dimension to use on y-axis
dim2_to_use
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
```

272 visForceLayoutPlot

```
name of NN network to use, if show_NN_network = TRUE
network_name
color_as_factor
                  convert color column to factor
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
                  column to use for alpha of the edges
edge_alpha
point_size
                  size of point (cell)
```

#### **Details**

Description of parameters.

#### Value

plotly

## **Examples**

```
visDimPlot_3D_plotly(gobject)
```

visForceLayoutPlot visForceLayoutPlot

# Description

Visualize cells according to forced layout algorithm coordinates

```
visForceLayoutPlot(
  gobject,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  layout_name = "layout",
  dim1_to_use = 1,
  dim2_to_use = 2,
  show_NN_network = T,
  cell_color = NULL,
  color_as_factor = F,
  cell_color_code = NULL,
  edge_alpha = NULL,
  point_size = 1,
```

visForceLayoutPlot 273

```
point_border_col = "black",
point_border_stroke = 0.1,
show_legend = T,
show_plot = F,
return_plot = TRUE,
save_plot = F,
save_dir = NULL,
save_folder = NULL,
save_format = NULL,
show_saved_plot = F,
...
)
```

```
gobject
                  giotto object
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
network_name
                  NN network to use
                  name of layout to use
layout_name
dim1_to_use
                  dimension to use on x-axis
                  dimension to use on y-axis
dim2_to_use
show_NN_network
                  show underlying NN network
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
                  column to use for alpha of the edges
edge_alpha
                  size of point (cell)
point_size
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
                  show legend
show_legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  directory to save the plot
save_dir
save_folder
                  (optional) folder in directory to save the plot
                  name of plot
save_name
save_format
                  format of plot (e.g. tiff, png, pdf, ...)
show_saved_plot
                  load & display the saved plot
```

274 visGenePlot

#### **Details**

Description of parameters.

#### Value

ggplot

#### **Examples**

visForceLayoutPlot(gobject)

visGenePlot

visGenePlot

## **Description**

Visualize cells and gene expression according to spatial coordinates

```
visGenePlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  edge_alpha = NULL,
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  midpoint = 0,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  plot_method = c("ggplot", "plotly"),
  show_plots = F
```

visGenePlot 275

### **Arguments**

gobject giotto object

expression\_values

gene expression values to use

genes genes to show

genes\_high\_color

color represents high gene expression

genes\_mid\_color

color represents middle gene expression

genes\_low\_color

color represents low gene expression

show\_network show underlying spatial network

network\_color color of spatial network

spatial\_network\_name

name of spatial network to use

show\_grid show spatial grid grid\_color color of spatial grid

spatial\_grid\_name

name of spatial grid to use

midpoint expression midpoint
scale\_alpha\_with\_expression

scale expression with ggplot alpha parameter

point\_size size of point (cell)

point\_border\_col

color of border around points

point\_border\_stroke

stroke size of border around points

show\_legend show legend

cow\_n\_col cowplot param: how many columns cow\_rel\_h cowplot param: relative height cow\_rel\_w cowplot param: relative width cow\_align cowplot param: how to align three mode to adjust axis scale axis\_scale x\_ticks number of ticks on x axis number of ticks on y axis y\_ticks number of ticks on z axis z\_ticks plot\_method two methods of plot

show\_plots show plots

## **Details**

Description of parameters.

### Value

ggplot or plotly

### **Examples**

```
visGenePlot(gobject)
```

```
{\tt visGenePlot\_2D\_ggplot} \ \ \textit{visGenePlot\_2D\_ggplot}
```

## **Description**

Visualize cells and gene expression according to spatial coordinates

### Usage

```
visGenePlot_2D_ggplot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  genes_high_color = "darkred",
  genes_mid_color = "white",
  genes_low_color = "darkblue",
  show_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  edge_alpha = NULL,
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  midpoint = 0,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_plots = F
)
```

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
genes_high_color
color represents high gene expression
genes_mid_color
color represents middle gene expression
```

visGenePlot\_3D\_plotly

277

genes\_low\_color

color represents low gene expression

show\_network show underlying spatial network

network\_color color of spatial network

spatial\_network\_name

name of spatial network to use

show\_grid show spatial grid grid\_color color of spatial grid

spatial\_grid\_name

name of spatial grid to use

midpoint expression midpoint

scale\_alpha\_with\_expression

scale expression with ggplot alpha parameter

point\_size size of point (cell)

point\_border\_col

color of border around points

point\_border\_stroke

stroke size of border around points

show\_legend show legend

cow\_n\_colcowplot param: how many columnscow\_rel\_hcowplot param: relative heightcow\_rel\_wcowplot param: relative widthcow\_aligncowplot param: how to align

show\_plots show plots

# Details

Description of parameters.

### Value

ggplot

## **Examples**

visGenePlot\_2D\_ggplot(gobject)

 ${\tt visGenePlot\_3D\_plotly} \ \ {\it visGenePlot\_3D\_plotly}$ 

# Description

Visualize cells and gene expression according to spatial coordinates

### Usage

spatial\_grid\_name

point\_size show\_legend

axis\_scale

x\_ticks

y\_ticks

```
visGenePlot_3D_plotly(
      gobject,
      expression_values = c("normalized", "scaled", "custom"),
      genes,
      show_network = F,
      network_color = NULL,
      spatial_network_name = "spatial_network",
      edge_alpha = NULL,
      show\_grid = F,
      genes_high_color = NULL,
      genes_mid_color = "white",
      genes_low_color = "blue",
      spatial_grid_name = "spatial_grid",
      point_size = 1,
      show_legend = T,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      show_plots = F
Arguments
   gobject
                    giotto object
    expression_values
                    gene expression values to use
    genes
                    genes to show
                    show underlying spatial network
    show_network
   network_color
                    color of spatial network
    spatial_network_name
                    name of spatial network to use
    show_grid
                    show spatial grid
    genes_high_color
                    color represents high gene expression
   genes_mid_color
                    color represents middle gene expression
    genes_low_color
                    color represents low gene expression
```

name of spatial grid to use

three mode to adjust axis scale

number of ticks on x axis

number of ticks on y axis

size of point (cell)

show legend

visPlot 279

#### **Details**

Description of parameters.

#### Value

plotly

## **Examples**

```
visGenePlot_3D_plotly(gobject)
```

visPlot visPlot

# Description

Visualize cells according to spatial coordinates

```
visPlot(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  show_network = F,
  network_color = NULL,
  network_alpha = 1,
  other_cell_alpha = 0.1,
  spatial_network_name = "spatial_network",
  show\_grid = F,
```

280 visPlot

```
grid_color = NULL,
 grid_alpha = 1,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 0.6,
  title = "",
  show_legend = T,
 axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
 y_ticks = NULL,
  z_ticks = NULL,
 plot_method = c("ggplot", "plotly"),
  show_plot = F,
  return_plot = TRUE,
  save_plot = F,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  save_format = NULL,
 show_saved_plot = F,
)
```

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
sdimz
                  z-axis dimension name (default = 'sdimz')
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
show_network
                  show underlying spatial network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
```

visPlot\_2D\_ggplot 281

```
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
title
                  title of plot
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_dir
                  directory to save the plot
                  (optional) folder in directory to save the plot
save_folder
                  name of plot
save_name
save_format
                  format of plot (e.g. tiff, png, pdf, ...)
show_saved_plot
                  load & display the saved plot
```

#### **Details**

Description of parameters.

### Value

ggplot

# **Examples**

visPlot(gobject)

## **Description**

Visualize cells according to spatial coordinates

```
visPlot_2D_ggplot(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  cell_color = NULL,
  cell_color_code = NULL,
```

282 visPlot\_2D\_ggplot

```
color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  show_network = F,
  network_color = NULL,
  network_alpha = 1,
  other_cells_alpha = 0.1,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 0.6,
  title = "",
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  show_plot = F,
  return_plot = TRUE,
  save_plot = F,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  save_format = NULL,
  show_saved_plot = F,
)
```

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
```

visPlot\_2D\_ggplot 283

show\_other\_cells

display not selected cells

other\_cell\_color

color of not selected cells

show\_network show underlying spatial network

network\_color color of spatial network

spatial\_network\_name

name of spatial network to use

show\_grid show spatial grid

grid\_color color of spatial grid

spatial\_grid\_name

name of spatial grid to use

coord\_fix\_ratio

fix ratio between x and y-axis

title title of plot

show\_legend show legend

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_dir directory to save the plot

save\_folder (optional) folder in directory to save the plot

save\_name name of plot

save\_format format of plot (e.g. tiff, png, pdf, ...)

show\_saved\_plot

load & display the saved plot

# **Details**

Description of parameters.

#### Value

ggplot

### **Examples**

 ${\tt visPlot\_2D\_ggplot(gobject)}$ 

284 visPlot\_2D\_plotly

```
visPlot_2D_plotly
```

#### **Description**

Visualize cells according to spatial coordinates

#### Usage

```
visPlot_2D_plotly(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_network = F,
  network_color = "lightgray",
  network_alpha = 1,
  other_cell_alpha = 0.5,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  grid_alpha = 1,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  show_plot = F
```

```
gobject giotto object

sdimx x-axis dimension name (default = 'sdimx')

sdimy y-axis dimension name (default = 'sdimy')

point_size size of point (cell)

cell_color color for cells (see details)

cell_color_code

named vector with colors

color_as_factor

convert color column to factor
```

visPlot\_3D\_plotly 285

```
select_cell_groups
                  select a subset of the groups from cell_color
                  show underlying spatial network
show_network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
                  alpha of spatial grid
grid_alpha
spatial_grid_name
                  name of spatial grid to use
                  show legend
show_legend
show_plot
                  show plot
```

#### **Details**

Description of parameters.

#### Value

plotly

# **Examples**

```
visPlot_2D_plotly(gobject)
```

```
visPlot_3D_plotly
```

# Description

Visualize cells according to spatial coordinates

```
visPlot_3D_plotly(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  point_size = 3,
  cell_color = NULL,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_network = F,
```

286 visPlot\_3D\_plotly

```
network_color = NULL,
network_alpha = 1,
other_cell_alpha = 0.5,
spatial_network_name = "spatial_network",
spatial_grid_name = "spatial_grid",
title = "",
show_legend = T,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
sticks = NULL,
show_plot = F
```

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
sdimz
                  z-axis dimension name (default = 'sdimz')
point_size
                  size of point (cell)
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
select_cell_groups
                  select a subset of the groups from cell_color
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
spatial_grid_name
                  name of spatial grid to use
                  title of plot
title
show_legend
                  show legend
show_plot
                  show plot
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
color_as_factor
                  convert color column to factor
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
coord_fix_ratio
                  fix ratio between x and y-axis
```

visSpatDimGenePlot 287

#### **Details**

Description of parameters.

#### Value

ggplot

### **Examples**

```
visPlot_3D_plotly(gobject)
```

visSpatDimGenePlot

visSpatDimGenePlot

#### **Description**

integration of visSpatDimGenePlot\_2D(ggplot) and visSpatDimGenePlot\_3D(plotly)

```
visSpatDimGenePlot(
 gobject,
 plot_method = c("ggplot", "plotly"),
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("horizontal", "vertical"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
 genes,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
 label_size = 16,
 genes_low_color = "blue",
 genes_mid_color = "white",
 genes_high_color = "red",
 dim_point_size = 3,
 nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
 network_color = "lightgray",
  spatial_network_alpha = 0.5,
```

288 visSpatDimGenePlot

```
show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_grid_alpha = 0.5,
      spatial_point_size = 3,
      spatial_point_border_col = "black",
      spatial_point_border_stroke = 0.1,
      legend_text_size = 12,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
     x_ticks = NULL,
     y_ticks = NULL,
     z_ticks = NULL,
     midpoint = 0,
     point_size = 1,
      cow_n_col = 2,
      cow_rel_h = 1,
     cow_rel_w = 1,
     cow_align = "h",
     show_legend = T,
      show_plots = F
   )
Arguments
   gobject
                    giotto object
   expression_values
                    gene expression values to use
   plot_alignment direction to align plot
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   dim3_to_use
                    dimension to use on z-axis
   sdimx
                    x-axis dimension name (default = 'sdimx')
   sdimy
                    y-axis dimension name (default = 'sdimy')
    sdimz
                    z-axis dimension name (default = 'sdimz')
   genes
                    genes to show
   dim_point_border_col
                    color of border around points
   dim_point_border_stroke
                    stroke size of border around points
   show_NN_network
                    show underlying NN network
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
                    name of NN network to use, if show_NN_network = TRUE
   network_name
```

visSpatDimGenePlot 289

edge\_alpha\_dim dim reduction plot: column to use for alpha of the edges scale\_alpha\_with\_expression scale expression with ggplot alpha parameter

label\_size size for the label

genes\_low\_color

color to represent low expression of gene

genes\_high\_color

color to represent high expression of gene

dim\_point\_size dim reduction plot: point size

spatial\_network\_name

name of spatial network to use

spatial\_grid\_name

name of spatial grid to use

spatial\_point\_size

spatial plot: point size

spatial\_point\_border\_col

color of border around points

spatial\_point\_border\_stroke

stroke size of border around points

legend\_text\_size

the size of the text in legend

axis\_scale three modes to adjust axis scale ratio custom\_ratio set the axis scale ratio on custom

 $x_{ticks}$  number of ticks on x axis  $y_{ticks}$  number of ticks on y axis  $z_{ticks}$  number of ticks on z axis

midpoint size of point (cell)
point\_size size of point (cell)

cow\_n\_col cowplot param: how many columns
cow\_rel\_h cowplot param: relative height
cow\_rel\_w cowplot param: relative width
cow\_align cowplot param: how to align

show\_legend show legend
show\_plot show plot

#### **Details**

Description of parameters.

#### Value

ggplot or plotly

## **Examples**

 $\verb|visSpatDimGenePlot(gobject)| \\$ 

visSpatDimGenePlot\_2D visSpatDimGenePlot\_2D

## **Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

## Usage

```
visSpatDimGenePlot_2D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("horizontal", "vertical"),
  genes,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  point_size = 1,
  dim_point_border_col = "black",
  dim_point_border_stroke = 0.1,
  show_NN_network = F,
  show_spatial_network = F,
  show_spatial_grid = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
  spatial_network_name = "spatial_network",
  spatial_grid_name = "spatial_grid",
  spatial_point_size = 1,
  spatial_point_border_col = "black",
  spatial_point_border_stroke = 0.1,
  midpoint = 0,
  genes_high_color = "red",
  genes_mid_color = "white";
  genes_low_color = "blue",
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  show_legend = T,
  show_plots = F
```

#### **Arguments**

gobject giotto object

expression\_values

gene expression values to use

plot\_alignment direction to align plot

genes genes to show

dim\_reduction\_to\_use

dimension reduction to use

dim\_reduction\_name

dimension reduction name

dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis

point\_size size of point (cell)

dim\_point\_border\_col

color of border around points

dim\_point\_border\_stroke

stroke size of border around points

show\_NN\_network

show underlying NN network

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

 $network\_name \qquad name \ of \ NN \ network \ to \ use, \ if \ show\_NN\_network = TRUE$ 

 ${\tt edge\_alpha\_dim} \ \ dim \ reduction \ plot: \ column \ to \ use \ for \ alpha \ of \ the \ edges$ 

scale\_alpha\_with\_expression

scale expression with ggplot alpha parameter

spatial\_network\_name

name of spatial network to use

spatial\_grid\_name

name of spatial grid to use

spatial\_point\_size

spatial plot: point size

spatial\_point\_border\_col

color of border around points

 ${\tt spatial\_point\_border\_stroke}$ 

stroke size of border around points

midpoint size of point (cell)

cow\_n\_col cowplot param: how many columns

cow\_rel\_h cowplot param: relative height cow\_rel\_w cowplot param: relative width cow\_align cowplot param: how to align

show\_legend show legend

dim\_point\_size dim reduction plot: point size

show\_plot show plot

#### **Details**

Description of parameters.

#### Value

ggplot

#### **Examples**

```
visSpatDimGenePlot_2D(gobject)
```

```
visSpatDimGenePlot_3D visSpatDimGenePlot_3D
```

## **Description**

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

```
visSpatDimGenePlot_3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("horizontal", "vertical"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  genes,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  label_size = 16,
  genes_low_color = "blue",
  genes_mid_color = "white",
  genes_high_color = "red",
  dim_point_size = 3,
  nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  legend_text_size = 12,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
```

y\_ticks = NULL, z\_ticks = NULL

```
Arguments
   gobject
                     giotto object
    plot_alignment direction to align plot
   dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
                     dimension to use on x-axis
   dim1_to_use
   dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
   network_name
    genes_low_color
                     color represent high gene expression (see details)
    genes_high_color
                     color represent high gene expression (see details)
    nn_network_alpha
                     column to use for alpha of the edges
    show_spatial_network
                     show spatial network
    spatial_network_name
                     name of spatial network to use
    network_color color of spatial/nn network
    spatial_network_alpha
                     alpha of spatial network
    show_spatial_grid
                     show spatial grid
    spatial_grid_name
                     name of spatial grid to use
    spatial_grid_color
                     color of spatial grid
```

## **Details**

Description of parameters.

spatial\_grid\_alpha

legend\_text\_size

show\_legend

show\_plot

alpha of spatial grid

text size of legend

show legend show plot

294 visSpatDimPlot

#### Value

plotly

#### **Examples**

```
visSpatDimPlot_3D(gobject)
```

visSpatDimPlot

visSpatDimPlot

## **Description**

integration of visSpatDimPlot\_2D and visSpatDimPlot\_3D

```
visSpatDimPlot(
  gobject,
  plot_method = c("ggplot", "plotly"),
  plot_alignment = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = NULL,
  label_fontface = "bold",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  dim_point_size = 3,
  dim_point_border_col = "black",
  dim_point_border_stroke = 0.1,
  nn_network_alpha = NULL,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
```

visSpatDimPlot 295

```
show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_grid_alpha = 0.5,
      spatial_point_size = 3,
      legend_text_size = 12,
      spatial_point_border_col = "black",
      spatial_point_border_stroke = 0.1,
      show_legend = T,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      show_plot = F
Arguments
    gobject
                     giotto object
    plot_alignment direction to align plot
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    nn_network_alpha
                     column to use for alpha of the edges
    show\_spatial\_network
                     show spatial network
```

296 visSpatDimPlot\_2D

```
spatial_network_name
                 name of spatial network to use
spatial_network_alpha
                 alpha of spatial network
show_spatial_grid
                 show spatial grid
spatial_grid_name
                 name of spatial grid to use
spatial_grid_color
                 color of spatial grid
spatial_grid_alpha
                 alpha of spatial grid
legend_text_size
                 text size of legend
show_legend
                 show legend
show_plot
                 show plot
plot_mode
                 choose the mode to draw plot: ggplot or plotly
spatial_network_color
                 color of spatial network
```

#### **Details**

Description of parameters.

#### Value

ggplot or plotly

#### **Examples**

```
visSpatDimPlot(gobject)
```

```
visSpatDimPlot_2D
visSpatDimPlot_2D
```

## **Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot2 mode

```
visSpatDimPlot_2D(
  gobject,
  plot_alignment = c("vertical", "horizontal"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  sdimx = NULL,
  sdimy = NULL,
```

visSpatDimPlot\_2D 297

 $show_NN_network = F,$ 

cell\_color

```
nn_network_to_use = "sNN",
     network_name = "sNN.pca",
      show\_cluster\_center = F,
      show_center_label = T,
      center_point_size = 4,
      label_size = 4,
      label_fontface = "bold",
      cell_color = NULL,
      color_as_factor = T,
      cell_color_code = NULL,
      select_cell_groups = NULL,
      select_cells = NULL,
      show_other_cells = T,
     other_cell_color = "lightgrey",
      dim_plot_mode = NULL,
     dim_point_size = 1,
     dim_point_border_col = "black",
     dim_point_border_stroke = 0.1,
     nn_network_alpha = 0.05,
      show_spatial_network = F,
      spatial_network_name = "spatial_network",
      spatial_network_color = NULL,
      show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_point_size = 1,
      spatial_point_border_col = "black",
      spatial_point_border_stroke = 0.1,
      show_legend = T,
      show_plot = F,
     plot_method = "ggplot"
Arguments
   gobject
                    giotto object
   plot_alignment direction to align plot
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   show_NN_network
                    show underlying NN network
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
                    name of NN network to use, if show_NN_network = TRUE
   network_name
```

color for cells (see details)

298 visSpatDimPlot\_2D

```
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
nn_network_alpha
                  column to use for alpha of the edges
show_spatial_network
                  show spatial network
spatial_network_name
                  name of spatial network to use
spatial_network_color
                  color of spatial network
show_spatial_grid
                  show spatial grid
spatial_grid_name
                  name of spatial grid to use
spatial_grid_color
                  color of spatial grid
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_dir
                  directory to save the plot
                  (optional) folder in directory to save the plot
save_folder
                  name of plot
save_name
save_format
                  format of plot (e.g. tiff, png, pdf, ...)
show_saved_plot
                  load & display the saved plot
```

#### **Details**

Description of parameters.

#### Value

ggplot

## **Examples**

```
visSpatDimPlot_2D(gobject)
```

visSpatDimPlot\_3D 299

visSpatDimPlot\_3D

visSpatDimPlot\_3D

# Description

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

## Usage

```
visSpatDimPlot_3D(
  gobject,
  plot_alignment = c("horizontal", "vertical"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 16,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  dim_point_size = 3,
  nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  legend_text_size = 12
```

#### **Arguments**

gobject giotto object

300 visSpatDimPlot\_3D

```
plot_alignment direction to align plot
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
                 dimension to use on x-axis
dim1_to_use
                 dimension to use on y-axis
dim2_to_use
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                 color for cells (see details)
color_as_factor
                 convert color column to factor
cell_color_code
                 named vector with colors
nn_network_alpha
                 column to use for alpha of the edges
show_spatial_network
                 show spatial network
spatial_network_name
                 name of spatial network to use
spatial_network_alpha
                 alpha of spatial network
show_spatial_grid
                 show spatial grid
spatial_grid_name
                 name of spatial grid to use
spatial_grid_color
                 color of spatial grid
spatial_grid_alpha
                 alpha of spatial grid
legend_text_size
                 text size of legend
spatial_network_color
                 color of spatial network
                 show legend
show_legend
                 show plot
show_plot
```

## **Details**

Description of parameters.

#### Value

plotly

writeHMRFresults 301

## **Examples**

```
visSpatDimPlot_3D(gobject)
```

writeHMRFresults

writeHMRFresults

## **Description**

write results from doHMRF to a data.table.

#### Usage

```
writeHMRFresults(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  print_command = F
)
```

# Arguments

gobject giotto object

HMRF output HMRF output from doHMRF

k k to write results for

betas\_to\_view results from different betas that you want to view

print\_command see the python command

# Value

data.table with HMRF results for each b and the selected k

# **Examples**

```
writeHMRFresults(gobject)
```

## **Description**

write out factor-like annotation data from a giotto object for the Viewer

```
write_giotto_viewer_annotation(
  annotation,
  annot_name = "test",
  output_directory = getwd()
)
```

#### **Arguments**

```
annotation annotation from the data.table from giotto object
annot_name name of the annotation
output_directory
directory where to save the files
```

#### Value

write a .txt and .annot file for the selection annotation

#### **Description**

write out dimensional reduction data from a giotto object for the Viewer

## Usage

```
write_giotto_viewer_dim_reduction(
  dim_reduction_cell,
  dim_red = NULL,
  dim_red_name = NULL,
  dim_red_rounding = NULL,
  dim_red_rescale = c(-20, 20),
  output_directory = getwd()
)
```

## **Arguments**

```
dim_reduction_cell

dimension reduction slot from giotto object

dim_red high level name of dimension reduction

dim_red_name specific name of dimension reduction to use

dim_red_rounding

numerical indicating how to round the coordinates

dim_red_rescale

numericals to rescale the coordinates

output_directory

directory where to save the files
```

#### Value

write a .txt and .annot file for the selection annotation

```
write\_giotto\_viewer\_numeric\_annotation \\ write\_giotto\_viewer\_numeric\_annotation
```

# Description

write out numeric annotation data from a giotto object for the Viewer

# Usage

```
write_giotto_viewer_numeric_annotation(
  annotation,
  annot_name = "test",
  output_directory = getwd()
)
```

# **Arguments**

```
annotation annotation from the data.table from giotto object
annot_name name of the annotation
output_directory
directory where to save the files
```

# Value

write a .txt and .annot file for the selection annotation

# Index

*Topic <b>giotto</b> ,	cellProximityVisPlot_2D_plotly, 38
giotto-class, 129	cellProximityVisPlot_3D_plotly, 40
print.giotto, 178	changeGiottoInstructions, 41
show, giotto-method, 186	clusterCells, 42
*Topic <b>giotto</b>	combineMetadata, 45
createGiottoObject, 47	convertEnsemblToGeneSymbol, 45
*Topic <b>object</b>	cowplot::save_plot, 129
giotto-class, 129	create_average_detection_DT, 56
print.giotto, 178	create_average_DT, 56
show, giotto-method, 186	create_cell_type_random_cell_IDs, 57
snow, grocco mechod, 100	create_cluster_matrix, 58
addCellMetadata, 7, 48	create_dimObject,58
addCellStatistics, 7, 12	createGiottoInstructions, 46, 48
addGeneMetadata, 8, 48	createGiottoObject, 47, 253
addGeneStatistics, 9, 12	createHeatmap_DT, 49
addHMRF, 10	createNearestNetwork, 50
addNetworkLayout, 11	createSpatialEnrich, 51, 101
addStatistics, 12	createSpatialGrid, 52
adjustGiottoMatrix, 12	createSpatialGrid_2D, 53
aes_string2, 13	createSpatialGrid_3D, 54
all_plots_save_function, 15	createSpatialNetwork, 55
allCellCellcommunicationsScores, 14	
<pre>annotate_spatlocs_with_spatgrid_2D, 17</pre>	decide_cluster_order,59
<pre>annotate_spatlocs_with_spatgrid_3D, 18</pre>	detectSpatialPatterns, 60
annotateGiotto, 16	dimCellPlot, 61
annotateSpatialNetwork, 17	dimCellPlot2D, 63, 63
<pre>average_gene_gene_expression_in_groups,</pre>	dimGenePlot, 66
18	dimGenePlot2D, 68
	dimGenePlot3D, <i>68</i> , <i>70</i> , <i>70</i>
binGetSpatialGenes, 19	dimPlot, 72
	dimPlot2D, 74, 75, 158, 160, 164, 166, 169,
<pre>calculate_spatial_genes_python, 23</pre>	171
calculateHVG, 20	dimPlot2D_single, 78
calculateMetaTable, 22	dimPlot3D, 66, 74, 77, 80, 80
calculateMetaTableCells, 23	<pre>direction_test (direction_test_CPG), 82</pre>
cellProximityBarplot, 24	direction_test_CPG,82
cellProximityEnrichment, 25	doHclust, 83
cellProximityHeatmap, 26	doHMRF, 84
cellProximityNetwork, 27	doKmeans, 85
cellProximitySpatPlot, 29	doLeidenCluster,87
cellProximitySpatPlot2D, 30, 30	doLeidenSubCluster, 88
cellProximitySpatPlot3D, 30, 32	doLouvainCluster,89
cellProximityVisPlot, 34	${\tt doLouvainCluster\_community}, 90$
<pre>cellProximityVisPlot_2D_ggplot, 36</pre>	doLouvainCluster_multinet, 92

INDEX 305

doLouvainSubCluster, 93	iterCluster, 131
doLouvainSubCluster_community, 94	iterLeidenCluster, 132
doLouvainSubCluster_multinet, 96	iterLouvainCluster, 134
doRandomWalkCluster, 97	<pre>iterLouvainCluster_community, 135</pre>
doSNNCluster, 98	<pre>iterLouvainCluster_multinet, 137</pre>
dt_to_matrix, 99	_ · · · · · · · · · · · · · · · · · · ·
ao_ooao. 27,97	kmeans_binarize, 138
exportGiottoViewer, 100	_ ,
exprOnlyCellCellcommunicationScores,	loadHMRF, 138
101	
extended_gini_fun, 102	make_simulated_network, 140
extractNearestNetwork, 102	makeSignMatrixPAGE, 139
, , , , , , , , , , , , , , , , , , , ,	makeSignMatrixRank, 140
fDataDT, 103	mergeClusters, 141
filterCombinations, 103, 107	mygini_fun, 142
filterCPGscores, 104	36 – /
filterDistributions, 105	nnDT_to_kNN, 142
filterGiotto, 106	node_clusters, 143
find_grid_2D, 116	normalizeGiotto, 143
find_grid_3D, 116	
find_grid_x, 116	OR_function2, 144
find_grid_y, 116	
find_grid_z, 117	PAGEEnrich, <i>52</i> , <i>139</i> , 145
	PCA, 182
findGiniMarkers, 107	pDataDT, 146
findGiniMarkers_one_vs_all, 108	permutationPA, 200
findMarkers, 109	plot_network_layer_ggplot, 173
findMarkers_one_vs_all, 110	plot_point_layer_ggplot, 174
findMastMarkers, 112	plot_spat_point_layer_ggplot, 176
findMastMarkers_one_vs_all, 113	plotCPGscores, 146
findScranMarkers, 114	plotGTGscores, 147
findScranMarkers_one_vs_all, 115	plotHeatmap, 149
fish_function, 117	plotly_axis_scale_2D, 150
fish_function2, 117	plotly_axis_scale_3D, 151
FSV_show, 118	
0 0 110	plotly_grid, 152
GenePattern_show, 119	plotly_network, 152
general_save_function, 120	plotMetaDataCellsHeatmap, 153
get10Xmatrix, 121	plotMetaDataHeatmap, 155
<pre>get_cell_to_cell_sorted_name_conversion,</pre>	plotPCA, 157
126	plotPCA_2D, 159
<pre>get_interaction_gene_enrichment, 127</pre>	plotPCA_3D, 158, 160, 161
<pre>get_specific_interaction_gene_enrichment,</pre>	plotTSNE, 162
127	plotTSNE_2D, 164
<pre>getCellProximityGeneScores, 121</pre>	plotTSNE_3D, <i>164</i> , <i>166</i> , 166
getClusterSimilarity, 123	plotUMAP, 168
getDendrogramSplits, 124	plotUMAP_2D, 170
getDistinctColors, 125	plotUMAP_3D, <i>169</i> , <i>171</i> , 172
getGeneToGeneScores, 125	print.giotto, 178
ggplot_save_function, 128	
giotto (giotto-class), 129	rank_binarize, 179
giotto-class, 129	rankEnrich, <i>52</i> , <i>140</i> , 178
Giotto::general_save_function, 16	readGiottoInstructions, 179
ofoctogeneral_save_runction, 10	removeCellAnnotation, 180
hyperGeometricEnrich, 52, 130	removeGeneAnnotation, 180
	*

306 INDEX

replaceGiottoInstructions, 181	viewHMRFresults2D, 257
Rtsne, <i>183</i>	viewHMRFresults3D, 258
runPCA, 181	violinPlot, 259
runtSNE, 182	visDimGenePlot, 260
runUMAP, 184	visDimGenePlot_2D_ggplot, 262
	visDimGenePlot_3D_plotly, 263
select_expression_values, 186	visDimPlot, 265
selectPatternGenes, 185	visDimPlot_2D_ggplot, 267
show, giotto-method, 186	visDimPlot_2D_plotly, 269
showClusterDendrogram, 187	visDimPlot_3D_plotly, 271
showClusterHeatmap, 188	visForceLayoutPlot, 272
showCPGscores, 189	visGenePlot, 274
showGeneExpressionProximityScore, 190	visGenePlot_2D_ggplot, 276
showGiottoInstructions, 191	visGenePlot_3D_plotly, 277
showGTGscores, 191	visPlot, 257, 279
showIntExpressionProximityScore, 192	visPlot_2D_ggplot, 281
showPattern, 193	visPlot_2D_plotly, 284
showPattern2D, <i>194</i> , 194	visPlot_3D_plotly, 285
showPattern3D, 195	visSpatDimGenePlot, 287
showPatternGenes, 197	visSpatDimGenePlot_2D, 290
showProcessingSteps, 198	visSpatDimGenePlot_3D, 292
showTopGeneToGene, 198	visSpatDimPlot, 294
signPCA, 199	visSpatDimPlot_2D, 296
spatCellPlot, 200	visSpatDimPlot_3D, 299
spatCellPlot2D, 203	V135patb1iii 10t_5b, 277
spatDimCellPlot, 206	write_giotto_viewer_annotation, 301
spatDimCellPlot2D, 210	write_giotto_viewer_dim_reduction, 302
spatDimGenePlot, 214	write_giotto_viewer_numeric_annotation,
spatDimGenePlot2D, 216	303
spatDimGenePlot3D, 216, 218, 218	writeHMRFresults, 301
spatDimPlot, 221	
spatDimPlot2D, 224, 225	
spatDimPlot3D, 224, 228, 229	
spatGenePlot, 232	
spatGenePlot2D, 233, 234	
spatGenePlot3D, 233, 235, 236	
Spatial_AEH, 237	
Spatial_DE, 238	
spatPlot, 239	
spatr 10t, 257 spatrlot2D, 242, 257	
spatrlot2D_single, 245	
spatr 10t2b_single, 243 spatPlot3D, 242, 245, 248, 248, 258	
specificCellCellcommunicationScores,	
250	
<pre>split_dendrogram_in_two, 251</pre>	
stitchFieldCoordinates, 48, 252	
subClusterCells, 253	
subsetGiotto, 255	
subsetGiottoLocs, 255	
umap, <i>185</i>	
•	
viewHMRFresults, 256	