

# Package ‘Giotto’

July 21, 2020

**Title** Spatial single-cell transcriptomics toolbox

**Version** 0.3.6.9013

**Description** Toolbox to process, analyze and visualize spatial single-cell expression data

**License** MIT + file LICENSE

**Encoding** UTF-8

**LazyData** true

**URL** <https://rubd.github.io/Giotto/>, <https://github.com/RubD/Giotto>

**BugReports** <https://github.com/RubD/Giotto/issues>

**RoxygenNote** 7.1.1

**Depends** base (>= 3.5.1),  
utils (>= 3.5.1),  
R (>= 3.5.1)

**Imports** data.table (>= 1.12.2),  
deldir,  
ggplot2 (>= 3.1.1),  
Matrix,  
magick,  
matrixStats (>= 0.55.0),  
methods,  
uwot (>= 0.0.0.9010),  
cowplot (>= 0.9.4),  
grDevices,  
graphics,  
RColorBrewer (>= 1.1-2),  
dbscan (>= 1.1-3),  
farver (>= 2.0.3),  
ggalluvial (>= 0.9.1),  
scales (>= 1.0.0),  
ComplexHeatmap (>= 1.20.0),  
qvalue (>= 2.14.1),  
lfa (>= 1.12.0),  
igraph (>= 1.2.4.1),  
irlba,  
plotly,  
parallel,  
reticulate (>= 1.14),

magrittr,  
 limma,  
 gg dendro,  
 smfishHmrf,  
 devtools,  
 reshape2,  
 ggraph,  
 Rcpp,  
 Rfast,  
 Rtsne ( $\geq 0.15$ ),  
 rlang ( $\geq 0.4.3$ ),  
 R.utils,  
 fitdistrplus,  
 quadprog

**Suggests** knitr,  
 rmarkdown,  
 MAST,  
 scan ( $\geq 1.10.1$ ),  
 png,  
 FactoMineR,  
 tiff,  
 biomaRt,  
 trendsceek,  
 multinet ( $\geq 3.0.2$ ),  
 RTriangle ( $\geq 1.6-0.10$ )

**biocViews**

**VignetteBuilder** knitr

**LinkingTo** Rcpp,  
 RcppArmadillo

**Remotes** lambdamoses/smfishhmrf-r

## R topics documented:

addCellIntMetadata . . . . .	7
addCellMetadata . . . . .	8
addCellStatistics . . . . .	9
addGeneMetadata . . . . .	10
addGenesPerc . . . . .	10
addGeneStatistics . . . . .	11
addGiottoImage . . . . .	12
addGiottoImageToSpatPlot . . . . .	12
addHMRF . . . . .	13
addNetworkLayout . . . . .	14
addStatistics . . . . .	15
adjustGiottoMatrix . . . . .	15
all_plots_save_function . . . . .	16
annotateGiotto . . . . .	18
annotateSpatialGrid . . . . .	19
annotateSpatialNetwork . . . . .	19
average_gene_gene_expression_in_groups . . . . .	20

binSpect . . . . .	21
calculateHVG . . . . .	22
calculateMetaTable . . . . .	24
calculateMetaTableCells . . . . .	25
cellProximityBarplot . . . . .	25
cellProximityEnrichment . . . . .	26
cellProximityHeatmap . . . . .	27
cellProximityNetwork . . . . .	28
cellProximitySpatPlot . . . . .	30
cellProximitySpatPlot2D . . . . .	31
cellProximitySpatPlot3D . . . . .	33
cellProximityVisPlot . . . . .	35
changeGiottoInstructions . . . . .	38
changeImageBg . . . . .	38
clusterCells . . . . .	39
clusterSpatialCorGenes . . . . .	42
combCCcom . . . . .	42
combineCellProximityGenes . . . . .	43
combineCPG . . . . .	44
combineMetadata . . . . .	45
convertEnsemblToGeneSymbol . . . . .	46
convert_mgImage_to_array_DT . . . . .	47
createCrossSection . . . . .	47
createGiottoImage . . . . .	49
createGiottoInstructions . . . . .	50
createGiottoObject . . . . .	51
createGiottoVisiumObject . . . . .	53
createHeatmap_DT . . . . .	54
createMetagenes . . . . .	55
createNearestNetwork . . . . .	56
createSpatialDefaultGrid . . . . .	57
createSpatialDelaunayNetwork . . . . .	58
createSpatialEnrich . . . . .	60
createSpatialGrid . . . . .	60
createSpatialKNNnetwork . . . . .	61
createSpatialNetwork . . . . .	62
create_average_detection_DT . . . . .	64
create_average_DT . . . . .	64
create_cell_type_random_cell_IDs . . . . .	65
create_crossSection_object . . . . .	66
create_screepplot . . . . .	67
crossSectionGenePlot . . . . .	67
crossSectionGenePlot3D . . . . .	68
crossSectionPlot . . . . .	69
crossSectionPlot3D . . . . .	70
decide_cluster_order . . . . .	71
detectSpatialCorGenes . . . . .	72
detectSpatialPatterns . . . . .	73
dimCellPlot . . . . .	74
dimCellPlot2D . . . . .	76
dimGenePlot . . . . .	79
dimGenePlot2D . . . . .	80

dimGenePlot3D . . . . .	83
dimPlot . . . . .	85
dimPlot2D . . . . .	86
dimPlot3D . . . . .	90
doHclust . . . . .	92
doHMRF . . . . .	93
doKmeans . . . . .	95
doLeidenCluster . . . . .	96
doLeidenSubCluster . . . . .	97
doLouvainCluster . . . . .	99
doLouvainCluster_community . . . . .	100
doLouvainCluster_multinet . . . . .	101
doLouvainSubCluster . . . . .	102
doLouvainSubCluster_community . . . . .	104
doLouvainSubCluster_multinet . . . . .	106
doRandomWalkCluster . . . . .	108
doSNNCluster . . . . .	109
estimateImageBg . . . . .	110
exportGiottoViewer . . . . .	110
exprCellCellcom . . . . .	111
fDataDT . . . . .	113
filterCellProximityGenes . . . . .	113
filterCombinations . . . . .	114
filterCPG . . . . .	115
filterDistributions . . . . .	116
filterGiotto . . . . .	118
findCellProximityGenes . . . . .	119
findCPG . . . . .	120
findGiniMarkers . . . . .	122
findGiniMarkers_one_vs_all . . . . .	124
findMarkers . . . . .	125
findMarkers_one_vs_all . . . . .	126
findMastMarkers . . . . .	128
findMastMarkers_one_vs_all . . . . .	129
findNetworkNeighbors . . . . .	130
findScranMarkers . . . . .	130
findScranMarkers_one_vs_all . . . . .	131
get10Xmatrix . . . . .	132
getClusterSimilarity . . . . .	133
getDendrogramSplits . . . . .	134
getDistinctColors . . . . .	135
getGiottoImage . . . . .	135
get_os . . . . .	136
giotto-class . . . . .	136
heatmapSpatialCorGenes . . . . .	137
hyperGeometricEnrich . . . . .	138
insertCrossSectionGenePlot3D . . . . .	139
insertCrossSectionSpatPlot3D . . . . .	140
jackstrawPlot . . . . .	141
loadHMRF . . . . .	143
makeSignMatrixPAGE . . . . .	144
makeSignMatrixRank . . . . .	144

mergeClusters . . . . .	145
node_clusters . . . . .	146
normalizeGiotto . . . . .	147
PAGEEnrich . . . . .	148
pDataDT . . . . .	149
plotCCcomDotplot . . . . .	149
plotCCcomHeatmap . . . . .	150
plotCellProximityGenes . . . . .	151
plotCombineCCcom . . . . .	153
plotCombineCellCellCommunication . . . . .	154
plotCombineCellProximityGenes . . . . .	155
plotCombineCPG . . . . .	157
plotCPG . . . . .	158
plotGiottoImage . . . . .	159
plotHeatmap . . . . .	160
plotICG . . . . .	162
plotInteractionChangedGenes . . . . .	163
plotMetaDataCellsHeatmap . . . . .	164
plotMetaDataHeatmap . . . . .	166
plotPCA . . . . .	168
plotPCA_2D . . . . .	169
plotPCA_3D . . . . .	171
plotRankSpatvsExpr . . . . .	173
plotRecovery . . . . .	174
plotRecovery_sub . . . . .	175
plotStatDelaunayNetwork . . . . .	175
plotTSNE . . . . .	176
plotTSNE_2D . . . . .	178
plotTSNE_3D . . . . .	180
plotUMAP . . . . .	182
plotUMAP_2D . . . . .	184
plotUMAP_3D . . . . .	186
print.giotto . . . . .	187
rankEnrich . . . . .	188
rankSpatialCorGroups . . . . .	188
readExprMatrix . . . . .	189
readGiottoInstructions . . . . .	190
removeCellAnnotation . . . . .	190
removeGeneAnnotation . . . . .	191
replaceGiottoInstructions . . . . .	191
runDWLSDeconv . . . . .	192
runHyperGeometricEnrich . . . . .	193
runPAGEEnrich . . . . .	194
runPCA . . . . .	195
runRankEnrich . . . . .	196
runSpatialDeconv . . . . .	197
runSpatialEnrich . . . . .	198
runtSNE . . . . .	199
runUMAP . . . . .	201
screePlot . . . . .	203
selectPatternGenes . . . . .	205
select_expression_values . . . . .	206

show,giotto-method . . . . .	206
showClusterDendrogram . . . . .	206
showClusterHeatmap . . . . .	208
showGiottoImageNames . . . . .	209
showGiottoInstructions . . . . .	209
showGrids . . . . .	210
showNetworks . . . . .	210
showPattern . . . . .	211
showPattern2D . . . . .	212
showPattern3D . . . . .	213
showPatternGenes . . . . .	214
showProcessingSteps . . . . .	215
showSaveParameters . . . . .	216
showSpatialCorGenes . . . . .	216
signPCA . . . . .	217
silhouetteRank . . . . .	219
spatCellCellcom . . . . .	220
spatCellPlot . . . . .	221
spatCellPlot2D . . . . .	223
spatDimCellPlot . . . . .	226
spatDimCellPlot2D . . . . .	228
spatDimGenePlot . . . . .	233
spatDimGenePlot2D . . . . .	235
spatDimGenePlot3D . . . . .	239
spatDimPlot . . . . .	242
spatDimPlot2D . . . . .	244
spatDimPlot3D . . . . .	249
spatGenePlot . . . . .	252
spatGenePlot2D . . . . .	254
spatGenePlot3D . . . . .	256
spatialAEH . . . . .	259
spatialDE . . . . .	260
spatNetwDistributions . . . . .	261
spatNetwDistributionsDistance . . . . .	262
spatNetwDistributionsKneighbors . . . . .	263
spatPlot . . . . .	264
spatPlot2D . . . . .	266
spatPlot3D . . . . .	269
specificCellCellcommunicationScores . . . . .	272
split_dendrogram_in_two . . . . .	273
stitchFieldCoordinates . . . . .	273
stitchTileCoordinates . . . . .	275
subClusterCells . . . . .	275
subsetGiotto . . . . .	277
subsetGiottoLocs . . . . .	278
trendSceek . . . . .	279
updateGiottoImage . . . . .	280
viewHMRResults . . . . .	280
viewHMRResults2D . . . . .	281
viewHMRResults3D . . . . .	282
violinPlot . . . . .	283
writeHMRResults . . . . .	284

write\_giotto\_viewer\_annotation . . . . . 285

write\_giotto\_viewer\_dim\_reduction . . . . . 285

write\_giotto\_viewer\_numeric\_annotation . . . . . 286

**Index** 287

---

addCellIntMetadata	<i>addCellIntMetadata</i>
--------------------	---------------------------

---

**Description**

Creates an additional metadata column with information about interacting and non-interacting cell types of the selected cell-cell interaction.

**Usage**

```
addCellIntMetadata(  
  gobject,  
  spatial_network = "spatial_network",  
  cluster_column,  
  cell_interaction,  
  name = "select_int",  
  return_gobject = TRUE  
)
```

**Arguments**

- gobject            giotto object
- spatial\_network            name of spatial network to use
- cluster\_column   column of cell types
- cell\_interaction            cell-cell interaction to use
- name                name for the new metadata column
- return\_gobject   return an updated giotto object

**Details**

This function will create an additional metadata column which selects interacting cell types for a specific cell-cell interaction. For example, if you want to color interacting astrocytes and oligodendrocytes it will create a new metadata column with the values "select\_astrocytes", "select\_oligodendrocytes", "other\_astrocytes", "other\_oligodendrocytes" and "other". Where "other" is all other cell types found within the selected cell type column.

**Value**

Giotto object

**Examples**

```
addCellIntMetadata(gobject)
```

---

addCellMetadata	<i>addCellMetadata</i>
-----------------	------------------------

---

## Description

adds cell metadata to the giotto object

## Usage

```
addCellMetadata(
  gobject,
  new_metadata,
  vector_name = NULL,
  by_column = FALSE,
  column_cell_ID = NULL
)
```

## Arguments

<code>gobject</code>	giotto object
<code>new_metadata</code>	new cell metadata to use (data.table, data.frame, ...)
<code>vector_name</code>	(optional) custom name if you provide a single vector
<code>by_column</code>	merge metadata based on cell_ID column in pDataDT (default = FALSE)
<code>column_cell_ID</code>	column name of new metadata to use if by_column = TRUE

## Details

You can add additional cell metadata in two manners:

- 1. Provide a data.table or data.frame with cell annotations in the same order as the cell\_ID column in pDataDT(gobject)
- 2. Provide a data.table or data.frame with cell annotations and specify which column contains the cell IDs, these cell IDs need to match with the cell\_ID column in pDataDT(gobject)

## Value

giotto object

## Examples

```
addCellMetadata(gobject)
```



---

addCellStatistics	<i>addCellStatistics</i>
-------------------	--------------------------

---

## Description

adds cells statistics to the giotto object

## Usage

```
addCellStatistics(  
  gobject,  
  expression_values = c("normalized", "scaled", "custom"),  
  detection_threshold = 0,  
  return_gobject = TRUE  
)
```

## Arguments

gobject	giotto object
expression_values	expression values to use
detection_threshold	detection threshold to consider a gene detected
return_gobject	boolean: return giotto object (default = TRUE)

## Details

This function will add the following statistics to cell metadata:

- `nr_genes`: Denotes in how many genes are detected per cell
- `perc_genes`: Denotes what percentage of genes is detected per cell
- `total_expr`: Shows the total sum of gene expression per cell

## Value

giotto object if `return_gobject = TRUE`

## Examples

```
addCellStatistics(gobject)
```

---

addGeneMetadata	<i>addGeneMetadata</i>
-----------------	------------------------

---

### Description

adds gene metadata to the giotto object

### Usage

```
addGeneMetadata(gobject, new_metadata, by_column = F, column_gene_ID = NULL)
```

### Arguments

gobject	giotto object
new_metadata	new metadata to use
by_column	merge metadata based on gene_ID column in fDataDT
column_gene_ID	column name of new metadata to use if by_column = TRUE

### Details

You can add additional gene metadata in two manners: 1. Provide a data.table or data.frame with gene annotations in the same order as the gene\_ID column in fDataDT(gobject) 2. Provide a data.table or data.frame with gene annotations and specify which column contains the gene IDs, these gene IDs need to match with the gene\_ID column in fDataDT(gobject)

### Value

giotto object

### Examples

```
addGeneMetadata(gobject)
```

---

addGenesPerc	<i>addGenesPerc</i>
--------------	---------------------

---

### Description

calculates the total percentage of (normalized) counts for a subset of selected genes

### Usage

```
addGenesPerc(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  vector_name = "gene_perc",
  return_gobject = TRUE
)
```

**Arguments**

gobject            giotto object  
 expression\_values       expression values to use  
 genes              vector of selected genes  
 vector\_name        column name as seen in pDataDT()  
 return\_gobject    boolean: return giotto object (default = TRUE)

**Value**

giotto object if return\_gobject = TRUE, else a vector with

**Examples**

```
addGenesPerc(gobject)
```

---

addGeneStatistics	<i>addGeneStatistics</i>
-------------------	--------------------------

---

**Description**

adds gene statistics to the giotto object

**Usage**

```
addGeneStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

**Arguments**

gobject            giotto object  
 expression\_values       expression values to use  
 detection\_threshold       detection threshold to consider a gene detected  
 return\_gobject    boolean: return giotto object (default = TRUE)

**Details**

This function will add the following statistics to gene metadata:

- nr\_cells: Denotes in how many cells the gene is detected
- per\_cells: Denotes in what percentage of cells the gene is detected
- total\_expr: Shows the total sum of gene expression in all cells
- mean\_expr: Average gene expression in all cells
- mean\_expr\_det: Average gene expression in cells with detectable levels of the gene

**Value**

giotto object if return\_gobject = TRUE

**Examples**

```
addGeneStatistics(gobject)
```

---

```
addGiottoImage
```

```
addGiottoImage
```

---

**Description**

Adds giotto image objects to your giotto object

**Usage**

```
addGiottoImage(gobject, images)
```

**Arguments**

gobject            giotto object

images            list of giotto image objects, see [createGiottoImage](#)

**Value**

an updated Giotto object with access to the list of images

**Examples**

```
addGiottoImage(mg_object)
```

---

```
addGiottoImageToSpatPlot
```

```
addGiottoImageToSpatPlot
```

---

**Description**

Add a giotto image to a spatial ggplot object post creation

**Usage**

```
addGiottoImageToSpatPlot(spatpl = NULL, gimage = NULL)
```

**Arguments**

spatpl            a spatial ggplot object

gimage            a giotto image, see [createGiottoImage](#)

**Value**

an updated spatial ggplot object

**Examples**

```
addGiottoImageToSpatPlot(mg_object)
```

---

addHMRF	<i>addHMRF</i>
---------	----------------

---

**Description**

Add selected results from doHMRF to the giotto object

**Usage**

```
addHMRF(gobject, HMRFOutput, k = NULL, betas_to_add = NULL, hmrf_name = NULL)
```

**Arguments**

gobject	giotto object
HMRFOutput	HMRF output from doHMRF()
k	number of domains
betas_to_add	results from different betas that you want to add
hmrf_name	specify a custom name

**Details**

Description ...

**Value**

giotto object

**Examples**

```
addHMRF(gobject)
```

---

addNetworkLayout	<i>addNetworkLayout</i>
------------------	-------------------------

---

## Description

Add a network layout for a selected nearest neighbor network

## Usage

```
addNetworkLayout(
  gobject,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  layout_type = c("drl"),
  options_list = NULL,
  layout_name = "layout",
  return_gobject = TRUE
)
```

## Arguments

gobject	giotto object
nn_network_to_use	kNN or sNN
network_name	name of NN network to be used
layout_type	layout algorithm to use
options_list	list of options for selected layout
layout_name	name for layout
return_gobject	boolean: return giotto object (default = TRUE)

## Details

This function creates layout coordinates based on the provided kNN or sNN. Currently only the force-directed graph layout "drl", see [layout\\_with\\_drl](#), is implemented. This provides an alternative to tSNE or UMAP based visualizations.

## Value

giotto object with updated layout for selected NN network

## Examples

```
addNetworkLayout(gobject)
```

---

addStatistics

*addStatistics*


---

## Description

adds genes and cells statistics to the giotto object

## Usage

```
addStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

## Arguments

gobject            giotto object

expression\_values  
                  expression values to use

detection\_threshold  
                  detection threshold to consider a gene detected

return\_gobject    boolean: return giotto object (default = TRUE)

## Details

See [addGeneStatistics](#) and [addCellStatistics](#)

## Value

giotto object if return\_gobject = TRUE, else a list with results

## Examples

```
addStatistics(gobject)
```

---

adjustGiottoMatrix

*adjustGiottoMatrix*


---

## Description

normalize and/or scale expresion values of Giotto object

**Usage**

```
adjustGiottoMatrix(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  batch_columns = NULL,
  covariate_columns = NULL,
  return_gobject = TRUE,
  update_slot = c("custom")
)
```

**Arguments**

gobject	giotto object
expression_values	expression values to use
batch_columns	metadata columns that represent different batch (max = 2)
covariate_columns	metadata columns that represent covariates to regress out
return_gobject	boolean: return giotto object (default = TRUE)
update_slot	expression slot that will be updated (default = custom)

**Details**

This function implements the [removeBatchEffect](#) function to remove known batch effects and to adjust expression values according to provided covariates.

**Value**

giotto object

**Examples**

```
adjustGiottoMatrix(gobject)
```

---

```
all_plots_save_function
```

```
all_plots_save_function
```

---

**Description**

Function to automatically save plots to directory of interest

**Usage**

```
all_plots_save_function(
  gobject,
  plot_object,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
```



```

    default_save_name = "giotto_plot",
    save_format = NULL,
    show_saved_plot = F,
    ncol = 1,
    nrow = 1,
    scale = 1,
    base_width = NULL,
    base_height = NULL,
    base_aspect_ratio = NULL,
    units = NULL,
    dpi = NULL,
    limitsize = TRUE,
    ...
)

```

### Arguments

<code>gobject</code>	giotto object
<code>plot_object</code>	object to plot
<code>save_dir</code>	directory to save to
<code>save_folder</code>	folder in <code>save_dir</code> to save to
<code>save_name</code>	name of plot
<code>default_save_name</code>	default name to save a plot
<code>save_format</code>	format (e.g. png, tiff, pdf, ...)
<code>show_saved_plot</code>	load & display the saved plot
<code>ncol</code>	number of columns
<code>nrow</code>	number of rows
<code>scale</code>	scale
<code>base_width</code>	width
<code>base_height</code>	height
<code>base_aspect_ratio</code>	aspect ratio
<code>units</code>	units
<code>dpi</code>	Plot resolution
<code>limitsize</code>	When TRUE (the default), ggsave will not save images larger than 50x50 inches, to prevent the common error of specifying dimensions in pixels.
<code>...</code>	additional parameters to <code>ggplot_save_function</code> or <code>general_save_function</code>

### See Also

[general\\_save\\_function](#)

### Examples

```
all_plots_save_function(gobject)
```

---

annotateGiotto	<i>annotateGiotto</i>
----------------	-----------------------

---

## Description

Converts cluster results into provided annotation.

## Usage

```
annotateGiotto(
  gobject,
  annotation_vector = NULL,
  cluster_column = NULL,
  name = "cell_types"
)
```

## Arguments

<code>gobject</code>	giotto object
<code>annotation_vector</code>	named annotation vector (names = cluster ids)
<code>cluster_column</code>	cluster column to convert to annotation names
<code>name</code>	new name for annotation column

## Details

You need to specify which (cluster) column you want to annotate and you need to provide an annotation vector like this:

- 1. identify the cell type of each cluster
- 2. create a vector of these cell types, e.g. `cell_types = c('T-cell', 'B-cell', 'Stromal')`
- 3. provide original cluster names to previous vector, e.g. `names(cell_types) = c(2, 1, 3)`

## Value

giotto object

## Examples

```
annotateGiotto(gobject)
```

---

annotateSpatialGrid     *annotateSpatialGrid*

---

### Description

annotate spatial grid with cell ID and cell metadata (optional)

### Usage

```
annotateSpatialGrid(
  gobject,
  spatial_grid_name = "spatial_grid",
  cluster_columns = NULL
)
```

### Arguments

gobject                Giotto object  
 spatial\_grid\_name        name of spatial grid, see [showGrids](#)  
 cluster\_columns        names of cell metadata, see [pDataDT](#)

### Value

annotated spatial grid data.table

### Examples

```
annotateSpatialGrid()
```

---

annotateSpatialNetwork  
                              *annotateSpatialNetwork*

---

### Description

Annotate spatial network with cell metadata information.

### Usage

```
annotateSpatialNetwork(
  gobject,
  spatial_network_name = "Delaunay_network",
  cluster_column,
  create_full_network = F
)
```

**Arguments**

gobject            giotto object  
 spatial\_network\_name    name of spatial network to use  
 cluster\_column    name of column to use for clusters  
 create\_full\_network    convert from reduced to full network representation

**Value**

annotated network in data.table format

**Examples**

```
annotateSpatialNetwork(gobject)
```

---

```
average_gene_gene_expression_in_groups
      average_gene_gene_expression_in_groups
```

---

**Description**

calculate average expression per cluster

**Usage**

```
average_gene_gene_expression_in_groups(
  gobject,
  cluster_column = "cell_types",
  gene_set_1,
  gene_set_2
)
```

**Arguments**

gobject            giotto object to use  
 cluster\_column    cluster column with cell type information  
 gene\_set\_1        first specific gene set from gene pairs  
 gene\_set\_2        second specific gene set from gene pairs

**Details**

Details will follow soon.

**Value**

data.table with average expression scores for each cluster

**Examples**

```
average_gene_gene_expression_in_groups(gobject)
```

---

binSpect	<i>binSpect</i>
----------	-----------------

---

## Description

Previously: binGetSpatialGenes. BinSpect (Binary Spatial Extraction of genes) is a fast computational method that identifies genes with a spatially coherent expression pattern.

## Usage

```
binSpect(
  gobject,
  bin_method = c("kmeans", "rank"),
  expression_values = c("normalized", "scaled", "custom"),
  subset_genes = NULL,
  spatial_network_name = "Delaunay_network",
  nstart = 3,
  iter_max = 10,
  percentage_rank = 30,
  do_fisher_test = TRUE,
  calc_hub = FALSE,
  hub_min_int = 3,
  get_av_expr = TRUE,
  get_high_expr = TRUE,
  do_parallel = TRUE,
  cores = NA,
  verbose = T
)
```

## Arguments

<code>gobject</code>	giotto object
<code>bin_method</code>	method to binarize gene expression
<code>expression_values</code>	expression values to use
<code>subset_genes</code>	only select a subset of genes to test
<code>spatial_network_name</code>	name of spatial network to use (default = 'spatial_network')
<code>nstart</code>	kmeans: nstart parameter
<code>iter_max</code>	kmeans: iter.max parameter
<code>percentage_rank</code>	percentage of top cells for binarization
<code>do_fisher_test</code>	perform fisher test
<code>calc_hub</code>	calculate the number of hub cells
<code>hub_min_int</code>	minimum number of cell-cell interactions for a hub cell
<code>get_av_expr</code>	calculate the average expression per gene of the high expressing cells
<code>get_high_expr</code>	calculate the number of high expressing cells per gene

do_parallel	run calculations in parallel with mclapply
cores	number of cores to use if do_parallel = TRUE
verbose	be verbose

### Details

We provide two ways to identify spatial genes based on gene expression binarization. Both methods are identical except for how binarization is performed.

- 1. binarize: Each gene is binarized (0 or 1) in each cell with **kmeans** ( $k = 2$ ) or based on **rank** percentile
- 2. network: All cells are connected through a spatial network based on the physical coordinates
- 3. contingency table: A contingency table is calculated based on all edges of neighboring cells and the binarized expression (0-0, 0-1, 1-0 or 1-1)
- 4. For each gene an odds-ratio (OR) and fisher.test (optional) is calculated

Other statistics are provided (optional):

- Number of cells with high expression (binary = 1)
- Average expression of each gene within high expressing cells
- Number of hub cells, these are high expressing cells that have a user defined number of high expressing neighbors

By selecting a subset of likely spatial genes (e.g. soft thresholding highly variable genes) or using multiple cores can accelerate the speed.

### Value

data.table with results (see details)

### Examples

```
binSpect(gobject)
```

---

calculateHVG	<i>calculateHVG</i>
--------------	---------------------

---

### Description

compute highly variable genes

### Usage

```
calculateHVG(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  method = c("cov_groups", "cov_loess"),
  reverse_log_scale = FALSE,
  logbase = 2,
  expression_threshold = 0,
```

```

nr_expression_groups = 20,
zscore_threshold = 1.5,
HVGname = "hvg",
difference_in_cov = 0.1,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "HVGplot",
return_gobject = TRUE
)

```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>method</code>	method to calculate highly variable genes
<code>reverse_log_scale</code>	reverse log-scale of expression values (default = FALSE)
<code>logbase</code>	if <code>reverse_log_scale</code> is TRUE, which log base was used?
<code>expression_threshold</code>	expression threshold to consider a gene detected
<code>nr_expression_groups</code>	number of expression groups for <code>cov_groups</code>
<code>zscore_threshold</code>	zscore to select hvg for <code>cov_groups</code>
<code>HVGname</code>	name for highly variable genes in cell metadata
<code>difference_in_cov</code>	minimum difference in coefficient of variance required
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)

## Details

Currently we provide 2 ways to calculate highly variable genes: **1. high coeff of variance (COV) within groups:**

First genes are binned (*nr\_expression\_groups*) into average expression groups and the COV for each gene is converted into a z-score within each bin. Genes with a z-score higher than the threshold (*zscore\_threshold*) are considered highly variable.

### 2. high COV based on loess regression prediction:

A predicted COV is calculated for each gene using loess regression ( $COV \sim \log(\text{mean expression})$ ). Genes that show a higher than predicted COV (*difference\_in\_cov*) are considered highly variable.

**Value**

giotto object highly variable genes appended to gene metadata (fDataDT)

**Examples**

```
# 1. create giotto object
expr_path = system.file("extdata", "seqfish_field_expr.txt", package = 'Giotto')
loc_path = system.file("extdata", "seqfish_field_locs.txt", package = 'Giotto')
VC_small <- createGiottoObject(raw_exprs = expr_path, spatial_locs = loc_path)

# 2. normalize giotto
VC_small <- normalizeGiotto(gobject = VC_small, scalefactor = 6000)
VC_small <- addStatistics(gobject = VC_small)

# 3. highly variable genes detection
VC_small <- calculateHVG(gobject = VC_small)
```

---

calculateMetaTable	<i>calculateMetaTable</i>
--------------------	---------------------------

---

**Description**

calculates the average gene expression for one or more (combined) annotation columns.

**Usage**

```
calculateMetaTable(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metadata_cols = NULL,
  selected_genes = NULL
)
```

**Arguments**

gobject	giotto object
expression_values	expression values to use
metadata_cols	annotation columns found in pDataDT(gobject)
selected_genes	subset of genes to use

**Value**

data.table with average expression values for each gene per (combined) annotation

**Examples**

```
calculateMetaTable(gobject)
```



---

```
calculateMetaTableCells
      calculateMetaTableCells
```

---

**Description**

calculates the average metadata values for one or more (combined) annotation columns.

**Usage**

```
calculateMetaTableCells(
  gobject,
  value_cols = NULL,
  metadata_cols = NULL,
  spat_enr_names = NULL
)
```

**Arguments**

```
gobject      giotto object
value_cols   metadata or enrichment value columns to use
metadata_cols annotation columns found in pDataDT(gobject)
spat_enr_names which spatial enrichment results to include
```

**Value**

data.table with average metadata values per (combined) annotation

**Examples**

```
calculateMetaTableCells(gobject)
```

---

```
cellProximityBarplot  cellProximityBarplot
```

---

**Description**

Create barplot from cell-cell proximity scores

**Usage**

```
cellProximityBarplot(
  gobject,
  CPscore,
  min_orig_ints = 5,
  min_sim_ints = 5,
  p_val = 0.05,
  show_plot = NA,
  return_plot = NA,
```

```

    save_plot = NA,
    save_param = list(),
    default_save_name = "cellProximityBarplot"
  )

```

### Arguments

<code>gobject</code>	giotto object
<code>CPscore</code>	CPscore, output from <code>cellProximityEnrichment()</code>
<code>min_orig_ints</code>	filter on minimum original cell-cell interactions
<code>min_sim_ints</code>	filter on minimum simulated cell-cell interactions
<code>p_val</code>	p-value
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

### Details

This function creates a barplot that shows the spatial proximity enrichment or depletion of cell type pairs.

### Value

ggplot barplot

### Examples

```
cellProximityBarplot(CPscore)
```

---

```
cellProximityEnrichment
```

*cellProximityEnrichment*

---

### Description

Compute cell-cell interaction enrichment (observed vs expected)

### Usage

```

cellProximityEnrichment(
  gobject,
  spatial_network_name = "Delaunay_network",
  cluster_column,
  number_of_simulations = 1000,
  adjust_method = c("none", "fdr", "bonferroni", "BH", "holm", "hochberg", "hommel",
    "BY")
)

```

**Arguments**

**gobject**                giotto object  
**spatial\_network\_name**  
                          name of spatial network to use  
**cluster\_column**    name of column to use for clusters  
**number\_of\_simulations**  
                          number of simulations to create expected observations  
**adjust\_method**    method to adjust p.values

**Details**

Spatial proximity enrichment or depletion between pairs of cell types is calculated by calculating the observed over the expected frequency of cell-cell proximity interactions. The expected frequency is the average frequency calculated from a number of spatial network simulations. Each individual simulation is obtained by reshuffling the cell type labels of each node (cell) in the spatial network.

**Value**

List of cell Proximity scores (CPscores) in data.table format. The first data.table (raw\_sim\_table) shows the raw observations of both the original and simulated networks. The second data.table (enrichm\_res) shows the enrichment results.

**Examples**

```
cellProximityEnrichment(gobject)
```

---

```
cellProximityHeatmap    cellProximityHeatmap
```

---

**Description**

Create heatmap from cell-cell proximity scores

**Usage**

```

cellProximityHeatmap(
  gobject,
  CPscore,
  scale = T,
  order_cell_types = T,
  color_breaks = NULL,
  color_names = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityHeatmap"
)

```

**Arguments**

<code>gobject</code>	giotto object
<code>CPscore</code>	CPscore, output from <code>cellProximityEnrichment()</code>
<code>scale</code>	scale cell-cell proximity interaction scores
<code>order_cell_types</code>	order cell types based on enrichment correlation
<code>color_breaks</code>	numerical vector of length 3 to represent min, mean and maximum
<code>color_names</code>	character color vector of length 3
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

**Details**

This function creates a heatmap that shows the spatial proximity enrichment or depletion of cell type pairs.

**Value**

ggplot heatmap

**Examples**

```
cellProximityHeatmap(CPscore)
```

---

```
cellProximityNetwork  cellProximityNetwork
```

---

**Description**

Create network from cell-cell proximity scores

**Usage**

```
cellProximityNetwork(
  gobject,
  CPscore,
  remove_self_edges = FALSE,
  self_loop_strength = 0.1,
  color_depletion = "lightgreen",
  color_enrichment = "red",
  rescale_edge_weights = TRUE,
  edge_weight_range_depletion = c(0.1, 1),
  edge_weight_range_enrichment = c(1, 5),
  layout = c("Fruchterman", "DrL", "Kamada-Kawai"),
```

```

    only_show_enrichment_edges = F,
    edge_width_range = c(0.1, 2),
    node_size = 4,
    node_text_size = 6,
    show_plot = NA,
    return_plot = NA,
    save_plot = NA,
    save_param = list(),
    default_save_name = "cellProximityNetwork"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>CPscore</code>	CPscore, output from <code>cellProximityEnrichment()</code>
<code>remove_self_edges</code>	remove enrichment/depletion edges with itself
<code>self_loop_strength</code>	size of self-loops
<code>color_depletion</code>	color for depleted cell-cell interactions
<code>color_enrichment</code>	color for enriched cell-cell interactions
<code>rescale_edge_weights</code>	rescale edge weights (boolean)
<code>edge_weight_range_depletion</code>	numerical vector of length 2 to rescale depleted edge weights
<code>edge_weight_range_enrichment</code>	numerical vector of length 2 to rescale enriched edge weights
<code>layout</code>	layout algorithm to use to draw nodes and edges
<code>only_show_enrichment_edges</code>	show only the enriched pairwise scores
<code>edge_width_range</code>	range of edge width
<code>node_size</code>	size of nodes
<code>node_text_size</code>	size of node labels
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

### Details

This function creates a network that shows the spatial proximity enrichment or depletion of cell type pairs.

**Value**

igraph plot

**Examples**

```
cellProximityNetwork(CPscore)
```

---

```
cellProximitySpatPlot  cellProximitySpatPlot
```

---

**Description**

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

**Usage**

```
cellProximitySpatPlot(gobject, ...)
```

**Arguments**

<code>gobject</code>	giotto object
<code>...</code>	Arguments passed on to <a href="#">cellProximitySpatPlot2D</a>
	<code>interaction_name</code> cell-cell interaction name
	<code>cluster_column</code> cluster column with cell clusters
	<code>sdimx</code> x-axis dimension name (default = 'sdimx')
	<code>sdimy</code> y-axis dimension name (default = 'sdimy')
	<code>cell_color</code> color for cells (see details)
	<code>cell_color_code</code> named vector with colors
	<code>color_as_factor</code> convert color column to factor
	<code>show_other_cells</code> decide if show cells not in network
	<code>show_network</code> show spatial network of selected cells
	<code>show_other_network</code> show spatial network of not selected cells
	<code>network_color</code> color of spatial network
	<code>spatial_network_name</code> name of spatial network to use
	<code>show_grid</code> show spatial grid
	<code>grid_color</code> color of spatial grid
	<code>spatial_grid_name</code> name of spatial grid to use
	<code>coord_fix_ratio</code> fix ratio between x and y-axis
	<code>show_legend</code> show legend
	<code>point_size_select</code> size of selected points
	<code>point_select_border_col</code> border color of selected points
	<code>point_select_border_stroke</code> stroke size of selected points
	<code>point_size_other</code> size of other points
	<code>point_alpha_other</code> opacity of other points
	<code>point_other_border_col</code> border color of other points
	<code>point_other_border_stroke</code> stroke size of other points
	<code>show_plot</code> show plots

return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters from [all\\_plots\\_save\\_function](#)  
 default\_save\_name default save name for saving, don't change, change save\_name  
 in save\_param

## Details

Description of parameters.

## Value

ggplot

## See Also

[cellProximitySpatPlot2D](#) and [cellProximitySpatPlot3D](#) for 3D

## Examples

```
cellProximitySpatPlot(gobject)
```

---

cellProximitySpatPlot2D

*cellProximitySpatPlot2D*

---

## Description

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

## Usage

```
cellProximitySpatPlot2D(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "Delaunay_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 1,
  show_legend = T,
```

```

point_size_select = 2,
point_select_border_col = "black",
point_select_border_stroke = 0.05,
point_size_other = 1,
point_alpha_other = 0.3,
point_other_border_col = "lightgrey",
point_other_border_stroke = 0.01,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "cellProximitySpatPlot2D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>interaction_name</code>	cell-cell interaction name
<code>cluster_column</code>	cluster column with cell clusters
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>cell_color</code>	color for cells (see details)
<code>cell_color_code</code>	named vector with colors
<code>color_as_factor</code>	convert color column to factor
<code>show_other_cells</code>	decide if show cells not in network
<code>show_network</code>	show spatial network of selected cells
<code>show_other_network</code>	show spatial network of not selected cells
<code>network_color</code>	color of spatial network
<code>spatial_network_name</code>	name of spatial network to use
<code>show_grid</code>	show spatial grid
<code>grid_color</code>	color of spatial grid
<code>spatial_grid_name</code>	name of spatial grid to use
<code>coord_fix_ratio</code>	fix ratio between x and y-axis
<code>show_legend</code>	show legend
<code>point_size_select</code>	size of selected points
<code>point_select_border_col</code>	border color of selected points
<code>point_select_border_stroke</code>	stroke size of selected points



point_size_other	size of other points
point_alpha_other	opacity of other points
point_other_border_col	border color of other points
point_other_border_stroke	stroke size of other points
show_plot	show plots
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from <a href="#">all_plots_save_function</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

**Details**

Description of parameters.

**Value**

ggplot

**Examples**

```
cellProximitySpatPlot2D(gobject)
```

---

```
cellProximitySpatPlot3D
      cellProximitySpatPlot2D
```

---

**Description**

Visualize 3D cell-cell interactions according to spatial coordinates in plotly mode

**Usage**

```
cellProximitySpatPlot3D(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = T,
  show_network = T,
  show_other_network = F,
```

```

network_color = NULL,
spatial_network_name = "Delaunay_network",
show_grid = F,
grid_color = NULL,
spatial_grid_name = "spatial_grid",
show_legend = T,
point_size_select = 4,
point_size_other = 2,
point_alpha_other = 0.5,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
z_ticks = NULL,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "cellProximitySpatPlot3D",
...
)

```

### Arguments

<code>gobject</code>	giotto object
<code>interaction_name</code>	cell-cell interaction name
<code>cluster_column</code>	cluster column with cell clusters
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>sdimz</code>	z-axis dimension name (default = 'sdimz')
<code>cell_color</code>	color for cells (see details)
<code>cell_color_code</code>	named vector with colors
<code>color_as_factor</code>	convert color column to factor
<code>show_other_cells</code>	decide if show cells not in network
<code>show_network</code>	show spatial network of selected cells
<code>show_other_network</code>	show spatial network of not selected cells
<code>network_color</code>	color of spatial network
<code>spatial_network_name</code>	name of spatial network to use
<code>show_grid</code>	show spatial grid
<code>grid_color</code>	color of spatial grid
<code>spatial_grid_name</code>	name of spatial grid to use
<code>show_legend</code>	show legend

point_size_select	size of selected points
point_size_other	size of other points
point_alpha_other	opacity of other points
axis_scale	scale of axis
custom_ratio	custom ratio of axes
x_ticks	ticks on x-axis
y_ticks	ticks on y-axis
z_ticks	ticks on z-axis
show_plot	show plots
return_plot	return plotly object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from <a href="#">all_plots_save_function</a>
default_save_name	default save name for saving, don't change, change save_name in save_param
...	additional parameters

**Details**

Description of parameters.

**Value**

plotly

**Examples**

```
cellProximitySpatPlot3D(gobject)
```

---

cellProximityVisPlot	<i>cellProximityVisPlot</i>
----------------------	-----------------------------

---

**Description**

Visualize cell-cell interactions according to spatial coordinates

**Usage**

```
cellProximityVisPlot(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  cell_color = NULL,
```

```

cell_color_code = NULL,
color_as_factor = T,
show_other_cells = F,
show_network = F,
show_other_network = F,
network_color = NULL,
spatial_network_name = "Delaunay_network",
show_grid = F,
grid_color = NULL,
spatial_grid_name = "spatial_grid",
coord_fix_ratio = 1,
show_legend = T,
point_size_select = 2,
point_select_border_col = "black",
point_select_border_stroke = 0.05,
point_size_other = 1,
point_alpha_other = 0.3,
point_other_border_col = "lightgrey",
point_other_border_stroke = 0.01,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
z_ticks = NULL,
plot_method = c("ggplot", "plotly"),
...
)

```

### Arguments

<code>gobject</code>	giotto object
<code>interaction_name</code>	cell-cell interaction name
<code>cluster_column</code>	cluster column with cell clusters
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>sdimz</code>	z-axis dimension name (default = 'sdimz')
<code>cell_color</code>	color for cells (see details)
<code>cell_color_code</code>	named vector with colors
<code>color_as_factor</code>	convert color column to factor
<code>show_other_cells</code>	show not selected cells
<code>show_network</code>	show underlying spatial network
<code>show_other_network</code>	show underlying spatial network of other cells
<code>network_color</code>	color of spatial network
<code>spatial_network_name</code>	name of spatial network to use

show_grid	show spatial grid
grid_color	color of spatial grid
spatial_grid_name	name of spatial grid to use
coord_fix_ratio	fix ratio between x and y-axis
show_legend	show legend
point_size_select	size of selected points
point_select_border_col	border color of selected points
point_select_border_stroke	stroke size of selected points
point_size_other	size of other points
point_alpha_other	alpha of other points
point_other_border_col	border color of other points
point_other_border_stroke	stroke size of other points
axis_scale	scale of axis
custom_ratio	custom ratio of scales
x_ticks	x ticks
y_ticks	y ticks
z_ticks	z ticks
plot_method	method to plot
...	additional parameters

## Details

Description of parameters.

## Value

ggplot or plotly

## Examples

```
cellProximityVisPlot(gobject)
```

---

changeGiottoInstructions	<i>changeGiottoInstructions</i>
--------------------------	---------------------------------

---

**Description**

Function to change one or more instructions from giotto object

**Usage**

```
changeGiottoInstructions(
  gobject,
  params = NULL,
  new_values = NULL,
  return_gobject = TRUE
)
```

**Arguments**

gobject	giotto object
params	parameter(s) to change
new_values	new value(s) for parameter(s)
return_gobject	(boolean) return giotto object

**Value**

giotto object with one or more changed instructions

**Examples**

```
changeGiottoInstructions()
```

---

changeImageBg	<i>changeImageBg</i>
---------------	----------------------

---

**Description**

Function to change the background color of a magick image plot to another color

**Usage**

```
changeImageBg(
  mg_object,
  bg_color,
  perc_range = 10,
  new_color = "#FFFFFF",
  new_name = NULL
)
```

**Arguments**

mg_object	magick image or giotto image object
bg_color	estimated current background color
perc_range	range around estimated background color to include (percentage)
new_color	new background color
new_name	change name of Giotto image

**Value**

magick image or giotto image object with updated background color

**Examples**

```
changeImageBg(mg_object)
```

---

clusterCells	<i>clusterCells</i>
--------------	---------------------

---

**Description**

cluster cells using a variety of different methods

**Usage**

```
clusterCells(
  gobject,
  cluster_method = c("leiden", "louvain_community", "louvain_multinet", "randomwalk",
    "sNNclust", "kmeans", "hierarchical"),
  name = "cluster_name",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  pyth_leid_resolution = 1,
  pyth_leid_weight_col = "weight",
  pyth_leid_part_type = c("RBConfigurationVertexPartition",
    "ModularityVertexPartition"),
  pyth_leid_init_memb = NULL,
  pyth_leid_iterations = 1000,
  pyth_louv_resolution = 1,
  pyth_louv_weight_col = NULL,
  python_louv_random = F,
  python_path = NULL,
  louvain_gamma = 1,
  louvain_omega = 1,
  walk_steps = 4,
  walk_clusters = 10,
  walk_weights = NA,
  sNNclust_k = 20,
  sNNclust_eps = 4,
  sNNclust_minPts = 16,
  borderPoints = TRUE,
```

```

expression_values = c("normalized", "scaled", "custom"),
genes_to_use = NULL,
dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
dim_reduction_name = "pca",
dimensions_to_use = 1:10,
distance_method = c("original", "pearson", "spearman", "euclidean", "maximum",
    "manhattan", "canberra", "binary", "minkowski"),
km_centers = 10,
km_iter_max = 100,
km_nstart = 1000,
km_algorithm = "Hartigan-Wong",
hc_agglomeration_method = c("ward.D2", "ward.D", "single", "complete", "average",
    "mcquitty", "median", "centroid"),
hc_k = 10,
hc_h = NULL,
return_gobject = TRUE,
set_seed = T,
seed_number = 1234
)

```

### Arguments

<code>gobject</code>	giotto object
<code>cluster_method</code>	community cluster method to use
<code>name</code>	name for new clustering result
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use
<code>pyth_leid_resolution</code>	resolution for leiden
<code>pyth_leid_weight_col</code>	column to use for weights
<code>pyth_leid_part_type</code>	partition type to use
<code>pyth_leid_init_memb</code>	initial membership
<code>pyth_leid_iterations</code>	number of iterations
<code>pyth_louv_resolution</code>	resolution for louvain
<code>pyth_louv_weight_col</code>	python louvain param: weight column
<code>python_louv_random</code>	python louvain param: random
<code>python_path</code>	specify specific path to python if required
<code>louvain_gamma</code>	louvain param: gamma or resolution
<code>louvain_omega</code>	louvain param: omega
<code>walk_steps</code>	randomwalk: number of steps
<code>walk_clusters</code>	randomwalk: number of clusters



walk_weights	randomwalk: weight column
sNNclust_k	SNNclust: k neighbors to use
sNNclust_eps	SNNclust: epsilon
sNNclust_minPts	SNNclust: min points
borderPoints	SNNclust: border points
expression_values	expression values to use
genes_to_use	= NULL,
dim_reduction_to_use	dimension reduction to use
dim_reduction_name	name of reduction 'pca',
dimensions_to_use	dimensions to use
distance_method	distance method
km_centers	kmeans centers
km_iter_max	kmeans iterations
km_nstart	kmeans random starting points
km_algorithm	kmeans algorithm
hc_agglomeration_method	hierarchical clustering method
hc_k	hierachical number of clusters
hc_h	hierarchical tree cutoff
return_gobject	boolean: return giotto object (default = TRUE)
set_seed	set seed
seed_number	number for seed

## Details

Wrapper for the different clustering methods.

## Value

giotto object with new clusters appended to cell metadata

## See Also

[doLeidenCluster](#), [doLouvainCluster\\_community](#), [doLouvainCluster\\_multinet](#), [doLouvainCluster](#), [doRandomWalkCluster](#), [doSNNCluster](#), [doKmeans](#), [doHclust](#)

## Examples

```
clusterCells(gobject)
```

---

clusterSpatialCorGenes	<i>clusterSpatialCorGenes</i>
------------------------	-------------------------------

---

### Description

Cluster based on spatially correlated genes

### Usage

```
clusterSpatialCorGenes(
  spatCorObject,
  name = "spat_clus",
  hclust_method = "ward.D",
  k = 10,
  return_obj = TRUE
)
```

### Arguments

spatCorObject	spatial correlation object
name	name for spatial clustering results
hclust_method	method for hierarchical clustering
k	number of clusters to extract
return_obj	return spatial correlation object (spatCorObject)

### Value

spatCorObject or cluster results

### Examples

```
clusterSpatialCorGenes(gobject)
```

---

combCCcom	<i>combCCcom</i>
-----------	------------------

---

### Description

Combine spatial and expression based cell-cell communication data.tables

**Usage**

```
combCCcom(
  spatialCC,
  exprCC,
  min_lig_nr = 3,
  min_rec_nr = 3,
  min_padj_value = 1,
  min_log2fc = 0,
  min_av_diff = 0
)
```

**Arguments**

spatialCC	spatial cell-cell communication scores
exprCC	expression cell-cell communication scores
min_lig_nr	minimum number of ligand cells
min_rec_nr	minimum number of receptor cells
min_padj_value	minimum adjusted p-value
min_log2fc	minimum log2 fold-change
min_av_diff	minimum average expression difference

**Value**

combined data.table with spatial and expression communication data

**Examples**

```
combCCcom(gobject)
```

---

```
combineCellProximityGenes
      combineCellProximityGenes
```

---

**Description**

Combine CPG scores in a pairwise manner.

**Usage**

```
combineCellProximityGenes(
  cpgObject,
  selected_ints = NULL,
  selected_genes = NULL,
  specific_genes_1 = NULL,
  specific_genes_2 = NULL,
  min_cells = 5,
  min_int_cells = 3,
  min_fdr = 0.05,
  min_spat_diff = 0,
```

```

    min_log2_fc = 0.5,
    do_parallel = TRUE,
    cores = NA,
    verbose = T
  )

```

### Arguments

<code>cpgObject</code>	cell proximity gene score object
<code>selected_ints</code>	subset of selected cell-cell interactions (optional)
<code>selected_genes</code>	subset of selected genes (optional)
<code>specific_genes_1</code>	specific geneset combo (need to position match <code>specific_genes_2</code> )
<code>specific_genes_2</code>	specific geneset combo (need to position match <code>specific_genes_1</code> )
<code>min_cells</code>	minimum number of target cell type
<code>min_int_cells</code>	minimum number of interacting cell type
<code>min_fdr</code>	minimum adjusted p-value
<code>min_spat_diff</code>	minimum absolute spatial expression difference
<code>min_log2_fc</code>	minimum absolute log2 fold-change
<code>do_parallel</code>	run calculations in parallel with <code>mclapply</code>
<code>cores</code>	number of cores to use if <code>do_parallel = TRUE</code>
<code>verbose</code>	verbose

### Value

`cpgObject` that contains the filtered differential gene scores

### Examples

```
combineCellProximityGenes(gobject)
```

---

combineCPG

*combineCPG*

---

### Description

Combine CPG scores in a pairwise manner.

### Usage

```

combineCPG(
  cpgObject,
  selected_ints = NULL,
  selected_genes = NULL,
  specific_genes_1 = NULL,
  specific_genes_2 = NULL,
  min_cells = 5,

```

```

    min_int_cells = 3,
    min_fdr = 0.05,
    min_spat_diff = 0,
    min_log2_fc = 0.5,
    do_parallel = TRUE,
    cores = NA,
    verbose = T
  )

```

### Arguments

cpgObject	cell proximity gene score object
selected_ints	subset of selected cell-cell interactions (optional)
selected_genes	subset of selected genes (optional)
specific_genes_1	specific geneset combo (need to position match specific_genes_2)
specific_genes_2	specific geneset combo (need to position match specific_genes_1)
min_cells	minimum number of target cell type
min_int_cells	minimum number of interacting cell type
min_fdr	minimum adjusted p-value
min_spat_diff	minimum absolute spatial expression difference
min_log2_fc	minimum absolute log2 fold-change
do_parallel	run calculations in parallel with mclapply
cores	number of cores to use if do_parallel = TRUE
verbose	verbose

### Value

cpgObject that contains the filtered differential gene scores

### Examples

```
combineCPG(gobject)
```

---

combineMetadata	<i>combineMetadata</i>
-----------------	------------------------

---

### Description

This function combines the cell metadata with spatial locations and enrichment results from createSpatialEnrich

### Usage

```
combineMetadata(gobject, spat_enr_names = NULL)
```

**Arguments**

`gobject`                Giotto object  
`spat_enr_names`    names of spatial enrichment results to include

**Value**

Extended cell metadata in data.table format.

**Examples**

```
combineMetadata(gobject)
```

---

`convertEnsemblToGeneSymbol`  
*convertEnsemblToGeneSymbol*

---

**Description**

This function convert ensembl gene IDs from a matrix to official gene symbols

**Usage**

```
convertEnsemblToGeneSymbol(matrix, species = c("mouse", "human"))
```

**Arguments**

`matrix`                an expression matrix with ensembl gene IDs as rownames  
`species`                species to use for gene symbol conversion

**Details**

This function requires that the biomaRt library is installed

**Value**

expression matrix with gene symbols as rownames

**Examples**

```
convertEnsemblToGeneSymbol(matrix)
```

---

```
convert_mgImage_to_array_DT
      convert_mgImage_to_array_DT
```

---

**Description**

converts a magick image object to a data.table

**Usage**

```
convert_mgImage_to_array_DT(mg_object)
```

**Arguments**

mg\_object            magick image or Giotto image object

**Value**

data.table with image pixel information

---

```
createCrossSection    createCrossSection
```

---

**Description**

Create a virtual 2D cross section.

**Usage**

```
createCrossSection(
  gobject,
  name = "cross_section",
  spatial_network_name = "Delaunay_network",
  thickness_unit = c("cell", "natural"),
  slice_thickness = 2,
  cell_distance_estimate_method = "mean",
  extend_ratio = 0.2,
  method = c("equation", "3 points", "point and norm vector",
    "point and two plane vectors"),
  equation = NULL,
  point1 = NULL,
  point2 = NULL,
  point3 = NULL,
  normVector = NULL,
  planeVector1 = NULL,
  planeVector2 = NULL,
  mesh_grid_n = 20,
  return_gobject = TRUE
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>name</code>	name of cross section object. (default = <code>cross_sectino</code> )
<code>spatial_network_name</code>	name of spatial network object. (default = <code>Delaunay_network</code> )
<code>thickness_unit</code>	unit of the virtual section thickness. If "cell", average size of the observed cells is used as length unit. If "natural", the unit of cell location coordinates is used. (default = <code>cell</code> )
<code>slice_thickness</code>	thickness of slice. default = 2
<code>cell_distance_estimate_method</code>	method to estimate average distance between neighboring cells. (default = <code>mean</code> )
<code>extend_ratio</code>	deciding the span of the cross section meshgrid, as a ratio of extension compared to the borders of the virtual tissue section. (default = 0.2)
<code>method</code>	method to define the cross section plane. If <code>equation</code> , the plane is defined by a four element numerical vector ( <code>equation</code> ) in the form of $c(A,B,C,D)$ , corresponding to a plane with equation $Ax+By+Cz=D$ . If <code>3 points</code> , the plane is defined by the coordinates of 3 points, as given by <code>point1</code> , <code>point2</code> , and <code>point3</code> . If <code>point and norm vector</code> , the plane is defined by the coordinates of one point ( <code>point1</code> ) in the plane and the coordinates of one norm vector ( <code>normVector</code> ) to the plane. If <code>point and two plane vector</code> , the plane is defined by the coordinates of one point ( <code>point1</code> ) in the plane and the coordinates of two vectors ( <code>planeVector1</code> , <code>planeVector2</code> ) in the plane. (default = <code>equation</code> )
<code>equation</code>	equation required by method "equation". equations needs to be a numerical vector of length 4, in the form of $c(A,B,C,D)$ , which defines plane $Ax+By+Cz=D$ .
<code>point1</code>	coordinates of the first point required by method "3 points", "point and norm vector", and "point and two plane vectors".
<code>point2</code>	coordinates of the second point required by method "3 points"
<code>point3</code>	coordinates of the third point required by method "3 points"
<code>normVector</code>	coordinates of the norm vector required by method "point and norm vector"
<code>planeVector1</code>	coordinates of the first plane vector required by method "point and two plane vectors"
<code>planeVector2</code>	coordinates of the second plane vector required by method "point and two plane vectors"
<code>mesh_grid_n</code>	number of meshgrid lines to generate along both directions for the cross section plane.
<code>return_gobject</code>	boolean: return giotto object (default = <code>TRUE</code> )

**Details**

Creates a virtual 2D cross section object for a given spatial network object. The users need to provide the definition of the cross section plane (see method).

**Value**

giotto object with updated spatial network slot



---

createGiottoImage	<i>createGiottoImage</i>
-------------------	--------------------------

---

## Description

Creates a giotto image that can be added to a Giotto object and/or used to add an image to the spatial plotting functions

## Usage

```
createGiottoImage(  
  gobject = NULL,  
  spatial_locs = NULL,  
  mg_object,  
  name = "image",  
  xmax_adj = 0,  
  xmin_adj = 0,  
  ymax_adj = 0,  
  ymin_adj = 0  
)
```

## Arguments

<code>gobject</code>	giotto object
<code>spatial_locs</code>	spatial locations (alternative if <code>gobject = NULL</code> )
<code>mg_object</code>	magick image object
<code>name</code>	name for the image
<code>xmax_adj</code>	adjustment of the maximum x-value to align the image
<code>xmin_adj</code>	adjustment of the minimum x-value to align the image
<code>ymax_adj</code>	adjustment of the maximum y-value to align the image
<code>ymin_adj</code>	adjustment of the minimum y-value to align the image

## Value

a giotto image object

## Examples

```
createGiottoImage(mg_object)
```

---

```
createGiottoInstructions
```

*createGiottoInstructions*

---

## Description

Function to set global instructions for giotto functions

## Usage

```
createGiottoInstructions(  
  python_path = NULL,  
  show_plot = NULL,  
  return_plot = NULL,  
  save_plot = NULL,  
  save_dir = NULL,  
  plot_format = NULL,  
  dpi = NULL,  
  units = NULL,  
  height = NULL,  
  width = NULL  
)
```

## Arguments

python_path	path to python binary to use
show_plot	print plot to console, default = TRUE
return_plot	return plot as object, default = TRUE
save_plot	automatically save plot, default = FALSE
save_dir	path to directory where to save plots
plot_format	format of plots (defaults to png)
dpi	resolution for raster images
units	units of format (defaults to in)
height	height of plots
width	width of plots

## Value

named vector with giotto instructions

## See Also

More online information can be found here [https://rubd.github.io/Giotto\\_site/articles/instructions\\_and\\_plotting.html](https://rubd.github.io/Giotto_site/articles/instructions_and_plotting.html)

## Examples

```
createGiottoInstructions()
```

---

createGiottoObject	<i>create Giotto object</i>
--------------------	-----------------------------

---

## Description

Function to create a giotto object

## Usage

```
createGiottoObject(
  raw_exprs,
  spatial_locs = NULL,
  norm_expr = NULL,
  norm_scaled_expr = NULL,
  custom_expr = NULL,
  cell_metadata = NULL,
  gene_metadata = NULL,
  spatial_network = NULL,
  spatial_network_name = NULL,
  spatial_grid = NULL,
  spatial_grid_name = NULL,
  spatial_enrichment = NULL,
  spatial_enrichment_name = NULL,
  dimension_reduction = NULL,
  nn_network = NULL,
  images = NULL,
  offset_file = NULL,
  instructions = NULL,
  cores = NA
)
```

## Arguments

raw_exprs	matrix with raw expression counts [required]
spatial_locs	data.table or data.frame with coordinates for cell centroids
norm_expr	normalized expression values
norm_scaled_expr	scaled expression values
custom_expr	custom expression values
cell_metadata	cell annotation metadata
gene_metadata	gene annotation metadata
spatial_network	list of spatial network(s)
spatial_network_name	list of spatial network name(s)
spatial_grid	list of spatial grid(s)
spatial_grid_name	list of spatial grid name(s)

spatial_enrichment	list of spatial enrichment score(s) for each spatial region
spatial_enrichment_name	list of spatial enrichment name(s)
dimension_reduction	list of dimension reduction(s)
nn_network	list of nearest neighbor network(s)
images	list of images
offset_file	file used to stitch fields together (optional)
instructions	list of instructions or output result from <a href="#">createGiottoInstructions</a>
cores	how many cores or threads to use to read data if paths are provided

## Details

**[Requirements]** To create a giotto object you need to provide at least a matrix with genes as row names and cells as column names. This matrix can be provided as a base matrix, sparse Matrix, data.frame, data.table or as a path to any of those. To include spatial information about cells (or regions) you need to provide a matrix, data.table or data.frame (or path to them) with coordinates for all spatial dimensions. This can be 2D (x and y) or 3D (x, y, x). The row order for the cell coordinates should be the same as the column order for the provided expression data.

**[Instructions]** Additionally an instruction file, generated manually or with [createGiottoInstructions](#) can be provided to instructions, if not a default instruction file will be created for the Giotto object.

**[Multiple fields]** In case a dataset consists of multiple fields, like seqFISH+ for example, an offset file can be provided to stitch the different fields together. [stitchFieldCoordinates](#) can be used to generate such an offset file.

**[Processed data]** Processed count data, such as normalized data, can be provided using one of the different expression slots (norm\_expr, norm\_scaled\_expr, custom\_expr).

**[Metadata]** Cell and gene metadata can be provided using the cell and gene metadata slots. This data can also be added afterwards using the [addGeneMetadata](#) or [addCellMetadata](#) functions.

**[Other information]** Additional information can be provided through the appropriate slots:

- spatial networks
- spatial girds
- spatial enrichments
- dimensions reductions
- nearest neighbours networks
- images

## Value

giotto object

## Examples

```
createGiottoObject(raw_exprs, spatial_locs)
```

---

```
createGiottoVisiumObject
      createGiottoVisiumObject
```

---

## Description

creates Giotto object directly from a 10X visium folder

## Usage

```
createGiottoVisiumObject(
  visium_dir = NULL,
  expr_data = c("raw", "filter"),
  gene_column_index = 1,
  png_name = NULL,
  xmax_adj = 0,
  xmin_adj = 0,
  ymax_adj = 0,
  ymin_adj = 0,
  instructions = NULL,
  cores = NA
)
```

## Arguments

visium_dir	path to the 10X visium directory [required]
expr_data	raw or filtered data (see details)
gene_column_index	which column index to select (see details)
png_name	select name of png to use (see details)
xmax_adj	adjustment of the maximum x-value to align the image
xmin_adj	adjustment of the minimum x-value to align the image
ymax_adj	adjustment of the maximum y-value to align the image
ymin_adj	adjustment of the minimum y-value to align the image
instructions	list of instructions or output result from <a href="#">createGiottoInstructions</a>
cores	how many cores or threads to use to read data if paths are provided

## Details

- `expr_data`: raw will take expression data from `raw_feature_bc_matrix` and filter from `filtered_feature_bc_matrix`
- `gene_column_index`: which gene identifiers (names) to use if there are multiple columns (e.g. ensemble and gene symbol)
- `png_name`: by default the first png will be selected, provide the png name to override this (e.g. myimage.png)

## Value

giotto object

**Examples**

```
createGiottoVisiumObject(visium_dir)
```

---

```
createHeatmap_DT
```

```
createHeatmap_DT
```

---

**Description**

creates order for clusters

**Usage**

```
createHeatmap_DT(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cluster_cor_method = "pearson",
  cluster_hclust_method = "ward.D",
  gene_order = c("correlation", "custom"),
  gene_custom_order = NULL,
  gene_cor_method = "pearson",
  gene_hclust_method = "complete"
)
```

**Arguments**

gobject	giotto object
expression_values	expression values to use
genes	genes to use
cluster_column	name of column to use for clusters
cluster_order	method to determine cluster order
cluster_custom_order	custom order for clusters
cluster_cor_method	method for cluster correlation
cluster_hclust_method	method for hierarchical clustering of clusters
gene_order	method to determine gene order
gene_custom_order	custom order for genes
gene_cor_method	method for gene correlation
gene_hclust_method	method for hierarchical clustering of genes

**Details**

Creates input data.tables for plotHeatmap function.

**Value**

list

**Examples**

```
createHeatmap_DT(gobject)
```

---

createMetagenes	<i>createMetagenes</i>
-----------------	------------------------

---

**Description**

This function creates an average metagene for gene clusters.

**Usage**

```
createMetagenes(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  gene_clusters,
  name = "metagene",
  return_gobject = TRUE
)
```

**Arguments**

gobject	Giotto object
expression_values	expression values to use
gene_clusters	numerical vector with genes as names
name	name of the metagene results
return_gobject	return giotto object

**Details**

An example for the 'gene\_clusters' could be like this: cluster\_vector = c(1, 1, 2, 2); names(cluster\_vector) = c('geneA', 'geneB', 'geneC', 'geneD')

**Value**

giotto object

**Examples**

```
createMetagenes(gobject)
```

---

createNearestNetwork    *createNearestNetwork*


---

## Description

create a nearest neighbour (NN) network

## Usage

```
createNearestNetwork(
  gobject,
  type = c("sNN", "kNN"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  genes_to_use = NULL,
  expression_values = c("normalized", "scaled", "custom"),
  name = "sNN.pca",
  return_gobject = TRUE,
  k = 30,
  minimum_shared = 5,
  top_shared = 3,
  verbose = T,
  ...
)
```

## Arguments

<code>gobject</code>	giotto object
<code>type</code>	sNN or kNN
<code>dim_reduction_to_use</code>	dimension reduction method to use
<code>dim_reduction_name</code>	name of dimension reduction set to use
<code>dimensions_to_use</code>	number of dimensions to use as input
<code>genes_to_use</code>	if <code>dim_reduction_to_use = NULL</code> , which genes to use
<code>expression_values</code>	expression values to use
<code>name</code>	arbitrary name for NN network
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>k</code>	number of k neighbors to use
<code>minimum_shared</code>	minimum shared neighbors
<code>top_shared</code>	keep at ...
<code>verbose</code>	be verbose
<code>...</code>	additional parameters for kNN and sNN functions from dbscan



## Details

This function creates a k-nearest neighbour (kNN) or shared nearest neighbour (sNN) network based on the provided dimension reduction space. To run it directly on the gene expression matrix set *dim\_reduction\_to\_use* = *NULL*.

See also [kNN](#) and [sNN](#) for more information about how the networks are created.

Output for kNN:

- from: cell\_ID for source cell
- to: cell\_ID for target cell
- distance: distance between cells
- weight:  $\text{weight} = 1/(1 + \text{distance})$

Output for sNN:

- from: cell\_ID for source cell
- to: cell\_ID for target cell
- distance: distance between cells
- weight:  $1/(1 + \text{distance})$
- shared: number of shared neighbours
- rank: ranking of pairwise cell neighbours

For sNN networks two additional parameters can be set:

- minimum\_shared: minimum number of shared neighbours needed
- top\_shared: keep this number of the top shared neighbours, irrespective of minimum\_shared setting

## Value

giotto object with updated NN network

## Examples

```
createNearestNetwork(gobject)
```

---

```
createSpatialDefaultGrid  
createSpatialDefaultGrid
```

---

## Description

Create a spatial grid using the default method

**Usage**

```
createSpatialDefaultGrid(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  sdimz_stepsize = NULL,
  minimum_padding = 1,
  name = NULL,
  return_gobject = TRUE
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>sdimx_stepsize</code>	stepsize along the x-axis
<code>sdimy_stepsize</code>	stepsize along the y-axis
<code>sdimz_stepsize</code>	stepsize along the z-axis
<code>minimum_padding</code>	minimum padding on the edges
<code>name</code>	name for spatial grid (default = 'spatial_grid')
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)

**Details**

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

**Value**

giotto object with updated spatial grid slot

---

```
createSpatialDelaunayNetwork
      createSpatialDelaunayNetwork
```

---

**Description**

Create a spatial Delaunay network based on cell centroid physical distances.

**Usage**

```
createSpatialDelaunayNetwork(
  gobject,
  method = c("deldir", "delaunayn_geometry", "RTriangle"),
  dimensions = "all",
  name = "Delaunay_network",
  maximum_distance = "auto",
  minimum_k = 0,
  options = "Pp",
```

```

Y = TRUE,
j = TRUE,
S = 0,
verbose = T,
return_gobject = TRUE,
...
)

```

## Arguments

gobject	giotto object
method	package to use to create a Delaunay network
dimensions	which spatial dimensions to use (default = all)
name	name for spatial network (default = 'delaunay_network')
maximum_distance	distance cutoff for Delaunay neighbors to consider. If "auto", "upper whisker" value of the distance vector between neighbors is used; see the boxplotgraphics documentation for more details.(default = "auto")
minimum_k	minimum number of neighbours if maximum_distance != NULL
options	(geometry) String containing extra control options for the underlying Qhull command; see the Qhull documentation (../doc/qhull/html/qdelaun.html) for the available options. (default = 'Pp', do not report precision problems)
Y	(RTriangle) If TRUE prohibits the insertion of Steiner points on the mesh boundary.
j	(RTriangle) If TRUE jettisons vertices that are not part of the final triangulation from the output.
S	(RTriangle) Specifies the maximum number of added Steiner points.
verbose	verbose
return_gobject	boolean: return giotto object (default = TRUE)
...	Other additional parameters

## Details

Creates a spatial Delaunay network as explained in [delaunayn](#) (default), [deldir](#), or [triangulate](#).

## Value

giotto object with updated spatial network slot

## Examples

```
createSpatialDelaunayNetwork(gobject)
```

---

createSpatialEnrich	<i>createSpatialEnrich</i>
---------------------	----------------------------

---

### Description

Function to calculate gene signature enrichment scores per spatial position using an enrichment test.

### Usage

```
createSpatialEnrich(...)
```

### Arguments

...	Arguments passed on to <a href="#">runSpatialEnrich</a>
gobject	Giotto object
enrich_method	method for gene signature enrichment calculation
sign_matrix	Matrix of signature genes for each cell type / process
expression_values	expression values to use
reverse_log_scale	reverse expression values from log scale
logbase	log base to use if reverse_log_scale = TRUE
p_value	calculate p-value (default = FALSE)
n_times	(page/rank) number of permutation iterations to calculate p-value
top_percentage	(hyper) percentage of cells that will be considered to have gene expression with matrix binarization
output_enrichment	how to return enrichment output
name	to give to spatial enrichment results, default = PAGE
return_gobject	return giotto object

### See Also

[runSpatialEnrich](#)

---

createSpatialGrid	<i>createSpatialGrid</i>
-------------------	--------------------------

---

### Description

Create a spatial grid using the default method

### Usage

```
createSpatialGrid(
  gobject,
  name = NULL,
  method = c("default"),
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  sdimz_stepsize = NULL,
  minimum_padding = 1,
  return_gobject = TRUE
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>name</code>	name for spatial grid
<code>method</code>	method to create a spatial grid
<code>sdimx_stepsize</code>	stepsize along the x-axis
<code>sdimy_stepsize</code>	stepsize along the y-axis
<code>sdimz_stepsize</code>	stepsize along the z-axis
<code>minimum_padding</code>	minimum padding on the edges
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)

**Details**

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

- default method: [createSpatialDefaultGrid](#)

**Value**

giotto object with updated spatial grid slot

---

```
createSpatialKNNnetwork
      createSpatialKNNnetwork
```

---

**Description**

Create a spatial knn network.

**Usage**

```
createSpatialKNNnetwork(
  gobject,
  method = "dbscan",
  dimensions = "all",
  name = "knn_network",
  k = 4,
  maximum_distance = NULL,
  minimum_k = 0,
  verbose = F,
  return_gobject = TRUE,
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>method</code>	method to create kNN network
<code>dimensions</code>	which spatial dimensions to use (default = all)
<code>name</code>	name for spatial network (default = 'spatial_network')
<code>k</code>	number of nearest neighbors based on physical distance
<code>maximum_distance</code>	distance cutoff for nearest neighbors to consider for kNN network
<code>minimum_k</code>	minimum nearest neighbours if <code>maximum_distance</code> != NULL
<code>verbose</code>	verbose
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>...</code>	additional arguments to the selected method function

**Value**

giotto object with updated spatial network slot

**dimensions:** default = 'all' which takes all possible dimensions. Alternatively you can provide a character vector that specifies the spatial dimensions to use, e.g. `c("sdimx", "sdimy")` or a numerical vector, e.g. `2:3`

**maximum\_distance:** to create a network based on maximum distance only, you also need to set `k` to a very high value, e.g. `k = 100`

**Examples**

```
createSpatialKNNnetwork(gobject)
```

---

```
createSpatialNetwork    createSpatialNetwork
```

---

**Description**

Create a spatial network based on cell centroid physical distances.

**Usage**

```
createSpatialNetwork(
  gobject,
  name = NULL,
  dimensions = "all",
  method = c("Delaunay", "kNN"),
  delaunay_method = c("deldir", "delaunayn_geometry", "RTriangle"),
  maximum_distance_delaunay = "auto",
  options = "Pp",
  Y = TRUE,
  j = TRUE,
  S = 0,
  minimum_k = 0,
  knn_method = "dbscan",
```

```

    k = 4,
    maximum_distance_knn = NULL,
    verbose = F,
    return_gobject = TRUE,
    ...
)

```

### Arguments

<code>gobject</code>	giotto object
<code>name</code>	name for spatial network (default = 'spatial_network')
<code>dimensions</code>	which spatial dimensions to use (default = all)
<code>method</code>	which method to use to create a spatial network. (default = Delaunay)
<code>delaunay_method</code>	Delaunay method to use
<code>maximum_distance_delaunay</code>	distance cutoff for nearest neighbors to consider for Delaunay network
<code>options</code>	(geometry) String containing extra control options for the underlying Qhull command; see the Qhull documentation ( <a href="#">../doc/qhull/html/qdelaun.html</a> ) for the available options. (default = 'Pp', do not report precision problems)
<code>Y</code>	(RTriangle) If TRUE prohibits the insertion of Steiner points on the mesh boundary.
<code>j</code>	(RTriangle) If TRUE jettisons vertices that are not part of the final triangulation from the output.
<code>S</code>	(RTriangle) Specifies the maximum number of added Steiner points.
<code>minimum_k</code>	minimum nearest neighbours if <code>maximum_distance</code> != NULL
<code>knn_method</code>	method to create kNN network
<code>k</code>	number of nearest neighbors based on physical distance
<code>maximum_distance_knn</code>	distance cutoff for nearest neighbors to consider for kNN network
<code>verbose</code>	verbose
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>...</code>	Additional parameters for the selected function

### Details

Creates a spatial network connecting single-cells based on their physical distance to each other. For Delaunay method, neighbors will be decided by delaunay triangulation and a maximum distance criteria. For kNN method, number of neighbors can be determined by `k`, or maximum distance from each cell with or without setting a minimum `k` for each cell.

**dimensions:** default = 'all' which takes all possible dimensions. Alternatively you can provide a character vector that specifies the spatial dimensions to use, e.g. `c("sdimx", "sdimy")` or a numerical vector, e.g. `2:3`

### Value

giotto object with updated spatial network slot

**Examples**

```
createSpatialNetwork(gobject)
```

---

```
create_average_detection_DT
      create_average_detection_DT
```

---

**Description**

calculates average gene detection for a cell metadata factor (e.g. cluster)

**Usage**

```
create_average_detection_DT(
  gobject,
  meta_data_name,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0
)
```

**Arguments**

```
gobject          giotto object
meta_data_name   name of metadata column to use
expression_values
                  which expression values to use
detection_threshold
                  detection threshold to consider a gene detected
```

**Value**

data.table with average gene expression values for each factor

---

```
create_average_DT      create_average_DT
```

---

**Description**

calculates average gene expression for a cell metadata factor (e.g. cluster)

**Usage**

```
create_average_DT(
  gobject,
  meta_data_name,
  expression_values = c("normalized", "scaled", "custom")
)
```



**Arguments**

gobject                giotto object

meta\_data\_name   name of metadata column to use

expression\_values        which expression values to use

**Value**

data.table with average gene expression values for each factor

---

```
create_cell_type_random_cell_IDs
      create_cell_type_random_cell_IDs
```

---

**Description**

creates randomized cell ids within a selection of cell types

**Usage**

```
create_cell_type_random_cell_IDs(
  gobject,
  cluster_column = "cell_types",
  needed_cell_types
)
```

**Arguments**

gobject                giotto object to use

cluster\_column   cluster column with cell type information

needed\_cell\_types        vector of cell type names for which a random id will be found

**Details**

Details will follow.

**Value**

list of randomly sampled cell ids with same cell type composition

**Examples**

```
create_cell_type_random_cell_IDs(gobject)
```

---

```
create_crossSection_object
    create_crossSection_object
```

---

## Description

create a crossSection object

## Usage

```
create_crossSection_object(
    name = NULL,
    method = NULL,
    thickness_unit = NULL,
    slice_thickness = NULL,
    cell_distance_estimate_method = NULL,
    extend_ratio = NULL,
    plane_equation = NULL,
    mesh_grid_n = NULL,
    mesh_obj = NULL,
    cell_subset = NULL,
    cell_subset_spatial_locations = NULL,
    cell_subset_projection_locations = NULL,
    cell_subset_projection_PCA = NULL,
    cell_subset_projection_coords = NULL
)
```

## Arguments

name	name of cross section object. (default = cross_sectino)
method	method to define the cross section plane.
thickness_unit	unit of the virtual section thickness. If "cell", average size of the observed cells is used as length unit. If "natural", the unit of cell location coordinates is used.(default = cell)
slice_thickness	thickness of slice
cell_distance_estimate_method	method to estimate average distance between neighboring cells. (default = mean)
extend_ratio	deciding the span of the cross section meshgrid, as a ratio of extension compared to the borders of the vitural tissue section. (default = 0.2)
plane_equation	a numerical vector of length 4, in the form of c(A,B,C,D), which defines plane $Ax+By+Cz=D$ .
mesh_grid_n	numner of meshgrid lines to generate along both directions for the cross section plane.
mesh_obj	object that stores the cross section meshgrid information.
cell_subset	cells selected by the cross section
cell_subset_spatial_locations	locations of cells selected by the cross section

cell\_subset\_projection\_locations  
3D projection coordinates of selected cells onto the cross section plane

cell\_subset\_projection\_PCA  
pca of projection coordinates

cell\_subset\_projection\_coords  
2D PCA coordinates of selected cells in the cross section plane

---

create_screepLOT	<i>create_screepLOT</i>
------------------	-------------------------

---

## Description

create screepLOT with ggplot

## Usage

```
create_screepLOT(pca_obj, ncp = 20, ylim = c(0, 20))
```

## Arguments

pca_obj	pca dimension reduction object
ncp	number of principal components to calculate
ylim	y-axis limits on scree plot

## Value

ggplot

---

crossSectionGenePlot	<i>crossSectionGenePlot</i>
----------------------	-----------------------------

---

## Description

Visualize cells and gene expression in a virtual cross section according to spatial coordinates

## Usage

```
crossSectionGenePlot(
  gobject = NULL,
  crossSection_obj = NULL,
  name = NULL,
  spatial_network_name = "Delaunay_network",
  default_save_name = "crossSectionGenePlot",
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>crossSection_obj</code>	crossSection object
<code>name</code>	name of virtual cross section to use
<code>spatial_network_name</code>	name of spatial network to use
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>
<code>...</code>	parameters for <code>spatGenePlot2D</code>

**Details**

Description of parameters.

**Value**

ggplot

**See Also**

[spatGenePlot3D](#) and [spatGenePlot2D](#)

---

`crossSectionGenePlot3D`

*crossSectionGenePlot3D*

---

**Description**

Visualize cells and gene expression in a virtual cross section according to spatial coordinates

**Usage**

```
crossSectionGenePlot3D(
  gobject,
  crossSection_obj = NULL,
  name = NULL,
  spatial_network_name = "Delaunay_network",
  other_cell_color = alpha("lightgrey", 0),
  default_save_name = "crossSectionGenePlot3D",
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>crossSection_obj</code>	cross section object as alternative input. default = NULL.
<code>name</code>	name of virtual cross section to use

```

    spatial_network_name
                        name of spatial network to use
    other_cell_color
                        color of cells outside the cross section. default = transparent.
    default_save_name
                        default save name for saving, don't change, change save_name in save_param
    ...
                        parameters for spatGenePlot3D

```

### Details

Description of parameters.

### Value

ggplot

### Examples

```
crossSectionGenePlot3D(gobject)
```

---

crossSectionPlot	<i>crossSectionPlot</i>
------------------	-------------------------

---

### Description

Visualize cells in a virtual cross section according to spatial coordinates

### Usage

```

crossSectionPlot(
  gobject,
  crossSection_obj = NULL,
  name = NULL,
  spatial_network_name = "Delaunay_network",
  default_save_name = "crossSectionPlot",
  ...
)

```

### Arguments

```

gobject          giotto object
crossSection_obj
                  cross section object as alternative input. default = NULL.
name              name of virtual cross section to use
spatial_network_name
                  name of spatial network to use
default_save_name
                  default save name for saving, don't change, change save_name in save_param
...
                  parameters for spatPlot2D

```

**Details**

Description of parameters.

**Value**

ggplot

**See Also**

[crossSectionPlot](#)

---

crossSectionPlot3D	<i>crossSectionPlot3D</i>
--------------------	---------------------------

---

**Description**

Visualize cells in a virtual cross section according to spatial coordinates

**Usage**

```
crossSectionPlot3D(
  gobject,
  crossSection_obj = NULL,
  name = NULL,
  spatial_network_name = "Delaunay_network",
  show_other_cells = T,
  other_cell_color = alpha("lightgrey", 0),
  default_save_name = "crossSection3D",
  ...
)
```

**Arguments**

gobject	giotto object
crossSection_obj	cross section object as alternative input. default = NULL.
name	name of virtual cross section to use
spatial_network_name	name of spatial network to use
show_other_cells	display not selected cells
other_cell_color	color of cells outside the cross section. default = transparent.
default_save_name	default save name for saving, don't change, change save_name in save_param
...	parameters for spatPlot3D

**Details**

Description of parameters.

**Value**

ggplot

**Examples**

```
crossSectionPlot3D(gobject)
```

---

decide_cluster_order	<i>decide_cluster_order</i>
----------------------	-----------------------------

---

**Description**

creates order for clusters

**Usage**

```
decide_cluster_order(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cor_method = "pearson",
  hclust_method = "ward.D"
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>genes</code>	genes to use
<code>cluster_column</code>	name of column to use for clusters
<code>cluster_order</code>	method to determine cluster order
<code>cluster_custom_order</code>	custom order for clusters
<code>cor_method</code>	method for correlation
<code>hclust_method</code>	method for hierarchical clustering

**Details**

Calculates order for clusters.

**Value**

custom

**Examples**

```
decide_cluster_order(gobject)
```

---

detectSpatialCorGenes    *detectSpatialCorGenes*

---

## Description

Detect genes that are spatially correlated

## Usage

```
detectSpatialCorGenes(
  gobject,
  method = c("grid", "network"),
  expression_values = c("normalized", "scaled", "custom"),
  subset_genes = NULL,
  spatial_network_name = "Delaunay_network",
  network_smoothing = NULL,
  spatial_grid_name = "spatial_grid",
  min_cells_per_grid = 4,
  cor_method = c("pearson", "kendall", "spearman")
)
```

## Arguments

gobject	giotto object
method	method to use for spatial averaging
expression_values	gene expression values to use
subset_genes	subset of genes to use
spatial_network_name	name of spatial network to use
network_smoothing	smoothing factor between 0 and 1 (default: automatic)
spatial_grid_name	name of spatial grid to use
min_cells_per_grid	minimum number of cells to consider a grid
cor_method	correlation method

## Details

For method = network, it expects a fully connected spatial network. You can make sure to create a fully connected network by setting minimal\_k > 0 in the [createSpatialNetwork](#) function.

- 1. grid-averaging: average gene expression values within a predefined spatial grid
- 2. network-averaging: smoothens the gene expression matrix by averaging the expression within one cell by using the neighbours within the predefined spatial network. b is a smoothening factor that defaults to  $1 - 1/k$ , where k is the median number of k-neighbors in the selected spatial network. Setting b = 0 means no smoothing and b = 1 means no contribution from its own expression.

The spatCorObject can be further explored with showSpatialCorGenes()



**Value**

returns a spatial correlation object: "spatCorObject"

**See Also**

[showSpatialCorGenes](#)

**Examples**

```
detectSpatialCorGenes(gobject)
```

---

detectSpatialPatterns *detectSpatialPatterns*

---

**Description**

Identify spatial patterns through PCA on average expression in a spatial grid.

**Usage**

```
detectSpatialPatterns(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  spatial_grid_name = "spatial_grid",
  min_cells_per_grid = 4,
  scale_unit = F,
  ncp = 100,
  show_plot = T,
  PC_zscore = 1.5
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>spatial_grid_name</code>	name of spatial grid to use (default = 'spatial_grid')
<code>min_cells_per_grid</code>	minimum number of cells in a grid to be considered
<code>scale_unit</code>	scale features
<code>ncp</code>	number of principal components to calculate
<code>show_plot</code>	show plots
<code>PC_zscore</code>	minimum z-score of variance explained by a PC

Details

Steps to identify spatial patterns:

- 1. average gene expression for cells within a grid, see createSpatialGrid
- 2. perform PCA on the average grid expression profiles
- 3. convert variance of principal components (PCs) to z-scores and select PCs based on a z-score threshold

Value

spatial pattern object 'spatPatObj'

Examples

```
detectSpatialPatterns(gobject)
```

---

dimCellPlot	<i>dimCellPlot</i>
-------------	--------------------

---

Description

Visualize cells according to dimension reduction coordinates

Usage

```
dimCellPlot(gobject, ...)
```

Arguments

gobject	giotto object
...	Arguments passed on to <a href="#">dimCellPlot2D</a>
	dim_reduction_to_use dimension reduction to use
	dim_reduction_name dimension reduction name
	dim1_to_use dimension to use on x-axis
	dim2_to_use dimension to use on y-axis
	spat_enr_names names of spatial enrichment results to include
	cell_annotation_values numeric cell annotation columns
	show_NN_network show underlying NN network
	nn_network_to_use type of NN network to use (kNN vs sNN)
	network_name name of NN network to use, if show_NN_network = TRUE
	cell_color_code named vector with colors for cell annotation values
	cell_color_gradient vector with 3 colors for numeric data
	gradient_midpoint midpoint for color gradient
	gradient_limits vector with lower and upper limits
	select_cell_groups select subset of cells/clusters based on cell_color parameter
	select_cells select subset of cells based on cell IDs
	show_other_cells display not selected cells

other\_cell\_color color of not selected cells  
 other\_point\_size size of not selected cells  
 show\_cluster\_center plot center of selected clusters  
 show\_center\_label plot label of selected clusters  
 center\_point\_size size of center points  
 center\_point\_border\_col border color of center points  
 center\_point\_border\_stroke border stroke size of center points  
 label\_size size of labels  
 label\_fontface font of labels  
 edge\_alpha column to use for alpha of the edges  
 point\_shape point with border or not (border or no\_border)  
 point\_size size of point (cell)  
 point\_alpha transparency of dim. reduction points  
 point\_border\_col color of border around points  
 point\_border\_stroke stroke size of border around points  
 show\_legend show legend  
 legend\_text size of legend text  
 legend\_symbol\_size size of legend symbols  
 background\_color color of plot background  
 axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plot  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)  
 default\_save\_name default save name for saving, don't change, change save\_name  
 in save\_param

## Details

Description of parameters. For 3D plots see [dimCellPlot2D](#)

## Value

ggplot

## See Also

Other dimension reduction cell annotation visualizations: [dimCellPlot2D\(\)](#)

## Examples

```
dimCellPlot(gobject)
```

dimCellPlot2D

*dimCellPlot2D***Description**

Visualize cells according to dimension reduction coordinates

**Usage**

```
dimCellPlot2D(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  spat_enr_names = NULL,
  cell_annotation_values = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_shape = c("border", "no_border"),
  point_size = 1,
  point_alpha = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  legend_text = 8,
  legend_symbol_size = 1,
  background_color = "white",
  axis_text = 8,
  axis_title = 8,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
```

```

    cow_align = "h",
    show_plot = NA,
    return_plot = NA,
    save_plot = NA,
    save_param = list(),
    default_save_name = "dimCellPlot2D"
)

```

## Arguments

<code>gobject</code>	giotto object
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>cell_annotation_values</code>	numeric cell annotation columns
<code>show_NN_network</code>	show underlying NN network
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use, if <code>show_NN_network = TRUE</code>
<code>cell_color_code</code>	named vector with colors for cell annotation values
<code>cell_color_gradient</code>	vector with 3 colors for numeric data
<code>gradient_midpoint</code>	midpoint for color gradient
<code>gradient_limits</code>	vector with lower and upper limits
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs
<code>show_other_cells</code>	display not selected cells
<code>other_cell_color</code>	color of not selected cells
<code>other_point_size</code>	size of not selected cells
<code>show_cluster_center</code>	plot center of selected clusters
<code>show_center_label</code>	plot label of selected clusters
<code>center_point_size</code>	size of center points

center_point_border_col	border color of center points
center_point_border_stroke	border stroke size of center points
label_size	size of labels
label_fontface	font of labels
edge_alpha	column to use for alpha of the edges
point_shape	point with border or not (border or no_border)
point_size	size of point (cell)
point_alpha	transparency of dim. reduction points
point_border_col	color of border around points
point_border_stroke	stroke size of border around points
show_legend	show legend
legend_text	size of legend text
legend_symbol_size	size of legend symbols
background_color	color of plot background
axis_text	size of axis text
axis_title	size of axis title
cow_n_col	cowplot param: how many columns
cow_rel_h	cowplot param: relative height
cow_rel_w	cowplot param: relative width
cow_align	cowplot param: how to align
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

## Details

Description of parameters. For 3D plots see [dimPlot3D](#)

## Value

ggplot

## See Also

Other dimension reduction cell annotation visualizations: [dimCellPlot\(\)](#)

## Examples

```
dimCellPlot2D(gobject)
```

dimGenePlot

*dimGenePlot***Description**

Visualize gene expression according to dimension reduction coordinates

**Usage**

```
dimGenePlot(...)
```

**Arguments**

```
...           Arguments passed on to dimGenePlot2D
gobject      giotto object
expression_values  gene expression values to use
genes        genes to show
dim_reduction_to_use  dimension reduction to use
dim_reduction_name  dimension reduction name
dim1_to_use    dimension to use on x-axis
dim2_to_use    dimension to use on y-axis
show_NN_network  show underlying NN network
nn_network_to_use  type of NN network to use (kNN vs sNN)
network_name    name of NN network to use, if show_NN_network = TRUE
network_color   color of NN network
edge_alpha      column to use for alpha of the edges
scale_alpha_with_expression  scale expression with ggplot alpha parameter
point_shape     point with border or not (border or no_border)
point_size      size of point (cell)
point_alpha     transparency of points
cell_color_gradient  vector with 3 colors for numeric data
gradient_midpoint  midpoint for color gradient
gradient_limits   vector with lower and upper limits
point_border_col  color of border around points
point_border_stroke  stroke size of border around points
show_legend      show legend
legend_text      size of legend text
background_color  color of plot background
axis_text        size of axis text
axis_title       size of axis title
cow_n_col        cowplot param: how many columns
cow_rel_h        cowplot param: relative height
cow_rel_w        cowplot param: relative width
cow_align        cowplot param: how to align
show_plot        show plots
return_plot      return ggplot object
```

save\_plot directly save the plot [boolean]  
save\_param list of saving parameters, see [showSaveParameters](#)  
default\_save\_name default save name for saving, don't change, change save\_name  
in save\_param

Details

Description of parameters.

Value

ggplot

See Also

[dimGenePlot3D](#)  
Other dimension reduction gene expression visualizations: [dimGenePlot2D\(\)](#), [dimGenePlot3D\(\)](#)

Examples

dimGenePlot(gobject)

---

dimGenePlot2D	<i>dimGenePlot2D</i>
---------------	----------------------

---

Description

Visualize gene expression according to dimension reduction coordinates

Usage

```
dimGenePlot2D(  
  gobject,  
  expression_values = c("normalized", "scaled", "custom"),  
  genes = NULL,  
  dim_reduction_to_use = "umap",  
  dim_reduction_name = "umap",  
  dim1_to_use = 1,  
  dim2_to_use = 2,  
  show_NN_network = F,  
  nn_network_to_use = "sNN",  
  network_name = "sNN.pca",  
  network_color = "lightgray",  
  edge_alpha = NULL,  
  scale_alpha_with_expression = FALSE,  
  point_shape = c("border", "no_border"),  
  point_size = 1,  
  point_alpha = 1,  
  cell_color_gradient = c("blue", "white", "red"),  
  gradient_midpoint = NULL,  
  gradient_limits = NULL,
```



```

    point_border_col = "black",
    point_border_stroke = 0.1,
    show_legend = T,
    legend_text = 8,
    background_color = "white",
    axis_text = 8,
    axis_title = 8,
    cow_n_col = 2,
    cow_rel_h = 1,
    cow_rel_w = 1,
    cow_align = "h",
    show_plot = NA,
    return_plot = NA,
    save_plot = NA,
    save_param = list(),
    default_save_name = "dimGenePlot2D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>genes</code>	genes to show
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>show_NN_network</code>	show underlying NN network
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use, if <code>show_NN_network = TRUE</code>
<code>network_color</code>	color of NN network
<code>edge_alpha</code>	column to use for alpha of the edges
<code>scale_alpha_with_expression</code>	scale expression with ggplot alpha parameter
<code>point_shape</code>	point with border or not (border or no_border)
<code>point_size</code>	size of point (cell)
<code>point_alpha</code>	transparency of points
<code>cell_color_gradient</code>	vector with 3 colors for numeric data
<code>gradient_midpoint</code>	midpoint for color gradient
<code>gradient_limits</code>	vector with lower and upper limits

point_border_col	color of border around points
point_border_stroke	stroke size of border around points
show_legend	show legend
legend_text	size of legend text
background_color	color of plot background
axis_text	size of axis text
axis_title	size of axis title
cow_n_col	cowplot param: how many columns
cow_rel_h	cowplot param: relative height
cow_rel_w	cowplot param: relative width
cow_align	cowplot param: how to align
show_plot	show plots
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

## Details

Description of parameters.

## Value

ggplot

## See Also

[dimGenePlot3D](#)

Other dimension reduction gene expression visualizations: [dimGenePlot3D\(\)](#), [dimGenePlot\(\)](#)

## Examples

```
dimGenePlot2D(gobject)
```

---

dimGenePlot3D	<i>dimGenePlot3D</i>
---------------	----------------------

---

## Description

Visualize cells and gene expression according to dimension reduction coordinates

## Usage

```
dimGenePlot3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = 3,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  cluster_column = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  edge_alpha = NULL,
  point_size = 2,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dimGenePlot3D"
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>genes</code>	genes to show
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name

dim1_to_use	dimension to use on x-axis
dim2_to_use	dimension to use on y-axis
dim3_to_use	dimension to use on z-axis
show_NN_network	show underlying NN network
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use, if show_NN_network = TRUE
network_color	color of NN network
cluster_column	cluster column to select groups
select_cell_groups	select subset of cells/clusters based on cell_color parameter
select_cells	select subset of cells based on cell IDs
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	size of not selected cells
edge_alpha	column to use for alpha of the edges
point_size	size of point (cell)
genes_high_color	color for high expression levels
genes_mid_color	color for medium expression levels
genes_low_color	color for low expression levels
show_legend	show legend
show_plot	show plots
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

## Details

Description of parameters.

## Value

ggplot

## See Also

Other dimension reduction gene expression visualizations: [dimGenePlot2D\(\)](#), [dimGenePlot\(\)](#)

## Examples

```
dimGenePlot3D(gobject)
```

dimPlot

*dimPlot***Description**

Visualize cells according to dimension reduction coordinates

**Usage**

```
dimPlot(...)
```

**Arguments**

```
...      Arguments passed on to dimPlot2D
gobject  giotto object
group_by create multiple plots based on cell annotation column
group_by_subset subset the group_by factor column
dim_reduction_to_use dimension reduction to use
dim_reduction_name dimension reduction name
dim1_to_use dimension to use on x-axis
dim2_to_use dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
show_NN_network show underlying NN network
nn_network_to_use type of NN network to use (kNN vs sNN)
network_name name of NN network to use, if show_NN_network = TRUE
cell_color color for cells (see details)
color_as_factor convert color column to factor
cell_color_code named vector with colors
cell_color_gradient vector with 3 colors for numeric data
gradient_midpoint midpoint for color gradient
gradient_limits vector with lower and upper limits
select_cell_groups select subset of cells/clusters based on cell_color parameter
select_cells select subset of cells based on cell IDs
show_other_cells display not selected cells
other_cell_color color of not selected cells
other_point_size size of not selected cells
show_cluster_center plot center of selected clusters
show_center_label plot label of selected clusters
center_point_size size of center points
center_point_border_col border color of center points
center_point_border_stroke border stroke size of center points
label_size size of labels
label_fontface font of labels
edge_alpha column to use for alpha of the edges
point_shape point with border or not (border or no_border)
```

point\_size size of point (cell)  
 point\_alpha transparency of point  
 point\_border\_col color of border around points  
 point\_border\_stroke stroke size of border around points  
 title title for plot, defaults to cell\_color parameter  
 show\_legend show legend  
 legend\_text size of legend text  
 legend\_symbol\_size size of legend symbols  
 background\_color color of plot background  
 axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plot  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)  
 default\_save\_name default save name for saving, don't change, change save\_name in save\_param

## Details

Description of parameters, see [dimPlot2D](#). For 3D plots see [dimPlot3D](#)

## Value

ggplot

## See Also

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

## Examples

```
dimPlot(gobject)
```

---

dimPlot2D

*dimPlot2D*

---

## Description

Visualize cells according to dimension reduction coordinates

**Usage**

```
dimPlot2D(  
  gobject,  
  group_by = NULL,  
  group_by_subset = NULL,  
  dim_reduction_to_use = "umap",  
  dim_reduction_name = "umap",  
  dim1_to_use = 1,  
  dim2_to_use = 2,  
  spat_enr_names = NULL,  
  show_NN_network = F,  
  nn_network_to_use = "sNN",  
  network_name = "sNN.pca",  
  cell_color = NULL,  
  color_as_factor = T,  
  cell_color_code = NULL,  
  cell_color_gradient = c("blue", "white", "red"),  
  gradient_midpoint = NULL,  
  gradient_limits = NULL,  
  select_cell_groups = NULL,  
  select_cells = NULL,  
  show_other_cells = T,  
  other_cell_color = "lightgrey",  
  other_point_size = 0.5,  
  show_cluster_center = F,  
  show_center_label = T,  
  center_point_size = 4,  
  center_point_border_col = "black",  
  center_point_border_stroke = 0.1,  
  label_size = 4,  
  label_fontface = "bold",  
  edge_alpha = NULL,  
  point_shape = c("border", "no_border"),  
  point_size = 1,  
  point_alpha = 1,  
  point_border_col = "black",  
  point_border_stroke = 0.1,  
  title = NULL,  
  show_legend = T,  
  legend_text = 8,  
  legend_symbol_size = 1,  
  background_color = "white",  
  axis_text = 8,  
  axis_title = 8,  
  cow_n_col = 2,  
  cow_rel_h = 1,  
  cow_rel_w = 1,  
  cow_align = "h",  
  show_plot = NA,  
  return_plot = NA,  
  save_plot = NA,  
  save_param = list(),
```

```

    default_save_name = "dimPlot2D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>group_by</code>	create multiple plots based on cell annotation column
<code>group_by_subset</code>	subset the <code>group_by</code> factor column
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>show_NN_network</code>	show underlying NN network
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use, if <code>show_NN_network</code> = TRUE
<code>cell_color</code>	color for cells (see details)
<code>color_as_factor</code>	convert color column to factor
<code>cell_color_code</code>	named vector with colors
<code>cell_color_gradient</code>	vector with 3 colors for numeric data
<code>gradient_midpoint</code>	midpoint for color gradient
<code>gradient_limits</code>	vector with lower and upper limits
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs
<code>show_other_cells</code>	display not selected cells
<code>other_cell_color</code>	color of not selected cells
<code>other_point_size</code>	size of not selected cells
<code>show_cluster_center</code>	plot center of selected clusters
<code>show_center_label</code>	plot label of selected clusters
<code>center_point_size</code>	size of center points



center_point_border_col	border color of center points
center_point_border_stroke	border stroke size of center points
label_size	size of labels
label_fontface	font of labels
edge_alpha	column to use for alpha of the edges
point_shape	point with border or not (border or no_border)
point_size	size of point (cell)
point_alpha	transparency of point
point_border_col	color of border around points
point_border_stroke	stroke size of border around points
title	title for plot, defaults to cell_color parameter
show_legend	show legend
legend_text	size of legend text
legend_symbol_size	size of legend symbols
background_color	color of plot background
axis_text	size of axis text
axis_title	size of axis title
cow_n_col	cowplot param: how many columns
cow_rel_h	cowplot param: relative height
cow_rel_w	cowplot param: relative width
cow_align	cowplot param: how to align
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

## Details

Description of parameters. For 3D plots see [dimPlot3D](#)

## Value

ggplot

## See Also

Other reduced dimension visualizations: [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

## Examples

```
dimPlot2D(gobject)
```

---

dimPlot3D

*dimPlot3D*


---

## Description

Visualize cells according to dimension reduction coordinates

## Usage

```
dimPlot3D(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = 3,
  spat_enr_names = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  color_as_factor = T,
  cell_color = NULL,
  cell_color_code = NULL,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 4,
  edge_alpha = NULL,
  point_size = 3,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dim3D"
)
```

## Arguments

<code>gobject</code>	giotto object
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis

dim3_to_use	dimension to use on z-axis
spat_enr_names	names of spatial enrichment results to include
select_cell_groups	select subset of cells/clusters based on cell_color parameter
select_cells	select subset of cells based on cell IDs
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	size of not selected cells
show_NN_network	show underlying NN network
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use, if show_NN_network = TRUE
color_as_factor	convert color column to factor
cell_color	color for cells (see details)
cell_color_code	named vector with colors
show_cluster_center	plot center of selected clusters
show_center_label	plot label of selected clusters
center_point_size	size of center points
label_size	size of labels
edge_alpha	column to use for alpha of the edges
point_size	size of point (cell)
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

## Details

Description of parameters.

## Value

plotly

## See Also

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

**Examples**

```
dimPlot3D(gobject)
```

---

doHclust	<i>doHclust</i>
----------	-----------------

---

**Description**

cluster cells using hierarchical clustering algorithm

**Usage**

```
doHclust(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
  dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  distance_method = c("pearson", "spearman", "original", "euclidean", "maximum",
    "manhattan", "canberra", "binary", "minkowski"),
  agglomeration_method = c("ward.D2", "ward.D", "single", "complete", "average",
    "mcquitty", "median", "centroid"),
  k = 10,
  h = NULL,
  name = "hclust",
  return_gobject = TRUE,
  set_seed = T,
  seed_number = 1234
)
```

**Arguments**

gobject	giotto object
expression_values	expression values to use
genes_to_use	subset of genes to use
dim_reduction_to_use	dimension reduction to use
dim_reduction_name	dimensions reduction name
dimensions_to_use	dimensions to use
distance_method	distance method
agglomeration_method	agglomeration method for hclust
k	number of final clusters

h	cut hierarchical tree at height = h
name	name for hierarchical clustering
return_gobject	boolean: return giotto object (default = TRUE)
set_seed	set seed
seed_number	number for seed

### Details

Description on how to use Kmeans clustering method.

### Value

giotto object with new clusters appended to cell metadata

### See Also

[hclust](#)

### Examples

```
doHclust(gobject)
```

---

doHMRF

*doHMRF*


---

### Description

Run HMRF

### Usage

```
doHMRF(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  spatial_network_name = "Delaunay_network",
  spatial_genes = NULL,
  spatial_dimensions = c("sdimx", "sdimy", "sdimz"),
  dim_reduction_to_use = NULL,
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "test",
  k = 10,
  betas = c(0, 2, 50),
  tolerance = 1e-10,
  zscore = c("none", "rowcol", "colrow"),
  numinit = 100,
  python_path = NULL,
  output_folder = NULL,
  overwrite_output = TRUE
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>spatial_network_name</code>	name of spatial network to use for HMRF
<code>spatial_genes</code>	spatial genes to use for HMRF
<code>spatial_dimensions</code>	select spatial dimensions to use, default is all possible dimensions
<code>dim_reduction_to_use</code>	use another dimension reduction set as input
<code>dim_reduction_name</code>	name of dimension reduction set to use
<code>dimensions_to_use</code>	number of dimensions to use as input
<code>name</code>	name of HMRF run
<code>k</code>	number of HMRF domains
<code>betas</code>	betas to test for
<code>tolerance</code>	tolerance
<code>zscore</code>	zscore
<code>numinit</code>	number of initializations
<code>python_path</code>	python path to use
<code>output_folder</code>	output folder to save results
<code>overwrite_output</code>	overwrite output folder

**Details**

Description of HMRF parameters ...

**Value**

Creates a directory with results that can be viewed with `viewHMRResults`

**Examples**

```
doHMRF(gobject)
```

doKmeans

*doKmeans***Description**

cluster cells using kmeans algorithm

**Usage**

```
doKmeans(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
  dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  distance_method = c("original", "pearson", "spearman", "euclidean", "maximum",
    "manhattan", "canberra", "binary", "minkowski"),
  centers = 10,
  iter_max = 100,
  nstart = 1000,
  algorithm = "Hartigan-Wong",
  name = "kmeans",
  return_gobject = TRUE,
  set_seed = T,
  seed_number = 1234
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>genes_to_use</code>	subset of genes to use
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimensions reduction name
<code>dimensions_to_use</code>	dimensions to use
<code>distance_method</code>	distance method
<code>centers</code>	number of final clusters
<code>iter_max</code>	kmeans maximum iterations
<code>nstart</code>	kmeans nstart
<code>algorithm</code>	kmeans algorithm
<code>name</code>	name for kmeans clustering
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>set_seed</code>	set seed
<code>seed_number</code>	number for seed

**Details**

Description on how to use Kmeans clustering method.

**Value**

giotto object with new clusters appended to cell metadata

**See Also**

[kmeans](#)

**Examples**

```
doKmeans(gobject)
```

---

doLeidenCluster	<i>doLeidenCluster</i>
-----------------	------------------------

---

**Description**

cluster cells using a NN-network and the Leiden community detection algorithm

**Usage**

```
doLeidenCluster(
  gobject,
  name = "leiden_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = "weight",
  partition_type = c("RBConfigurationVertexPartition", "ModularityVertexPartition"),
  init_membership = NULL,
  n_iterations = 1000,
  return_gobject = TRUE,
  set_seed = T,
  seed_number = 1234
)
```

**Arguments**

gobject	giotto object
name	name for cluster
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use
python_path	specify specific path to python if required
resolution	resolution
weight_col	weight column to use for edges



partition_type	The type of partition to use for optimisation.
init_membership	initial membership of cells for the partition
n_iterations	number of iterations to run the Leiden algorithm. If the number of iterations is negative, the Leiden algorithm is run until an iteration in which there was no improvement.
return_gobject	boolean: return giotto object (default = TRUE)
set_seed	set seed
seed_number	number for seed

## Details

This function is a wrapper for the Leiden algorithm implemented in python, which can detect communities in graphs of millions of nodes (cells), as long as they can fit in memory. See the <https://github.com/vtraag/leidenalg> github page or the <https://leidenalg.readthedocs.io/en/stable/index.html> readthedocs page for more information.

Partition types available and information:

- **RBCConfigurationVertexPartition**: Implements Reichardt and Bornholdt's Potts model with a configuration null model. This quality function is well-defined only for positive edge weights. This quality function uses a linear resolution parameter.
- **ModularityVertexPartition**: Implements modularity. This quality function is well-defined only for positive edge weights. It does *not* use the resolution parameter

Set *weight\_col* = *NULL* to give equal weight (=1) to each edge.

## Value

giotto object with new clusters appended to cell metadata

## Examples

```
doLeidenCluster(gobject)
```

---

doLeidenSubCluster	<i>doLeidenSubCluster</i>
--------------------	---------------------------

---

## Description

Further subcluster cells using a NN-network and the Leiden algorithm

## Usage

```
doLeidenSubCluster(
  gobject,
  name = "sub_pleiden_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
  hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
```

```

    hvg_min_perc_cells = 5,
    hvg_mean_expr_det = 1,
    use_all_genes_as_hvg = FALSE,
    min_nr_of_hvg = 5,
    pca_param = list(expression_values = "normalized", scale_unit = T),
    nn_param = list(dimensions_to_use = 1:20),
    k_neighbors = 10,
    resolution = 0.5,
    n_iterations = 500,
    python_path = NULL,
    nn_network_to_use = "sNN",
    network_name = "sNN.pca",
    return_gobject = TRUE,
    verbose = T
)

```

### Arguments

<code>gobject</code>	giotto object
<code>name</code>	name for new clustering result
<code>cluster_column</code>	cluster column to subcluster
<code>selected_clusters</code>	only do subclustering on these clusters
<code>hvg_param</code>	parameters for calculateHVG
<code>hvg_min_perc_cells</code>	threshold for detection in min percentage of cells
<code>hvg_mean_expr_det</code>	threshold for mean expression level in cells with detection
<code>use_all_genes_as_hvg</code>	forces all genes to be HVG and to be used as input for PCA
<code>min_nr_of_hvg</code>	minimum number of HVG, or all genes will be used as input for PCA
<code>pca_param</code>	parameters for runPCA
<code>nn_param</code>	parameters for parameters for createNearestNetwork
<code>k_neighbors</code>	number of k for createNearestNetwork
<code>resolution</code>	resolution of Leiden clustering
<code>n_iterations</code>	number of iterations to run the Leiden algorithm.
<code>python_path</code>	specify specific path to python if required
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>verbose</code>	verbose

### Details

This function performs subclustering using the Leiden algorithm on selected clusters. The systematic steps are:

- 1. subset Giotto object

- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do Leiden clustering

**Value**

giotto object with new subclusters appended to cell metadata

**See Also**

[doLeidenCluster](#)

**Examples**

```
doLeidenSubCluster(gobject)
```

---

doLouvainCluster	<i>doLouvainCluster</i>
------------------	-------------------------

---

**Description**

cluster cells using a NN-network and the Louvain algorithm.

**Usage**

```
doLouvainCluster(
  gobject,
  version = c("community", "multinet"),
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = NULL,
  gamma = 1,
  omega = 1,
  louv_random = F,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

**Arguments**

gobject	giotto object
version	implemented version of Louvain clustering to use
name	name for cluster
nn_network_to_use	type of NN network to use (kNN vs sNN)

network_name	name of NN network to use
python_path	[community] specify specific path to python if required
resolution	[community] resolution
weight_col	weight column name
gamma	[multinet] Resolution parameter for modularity in the generalized louvain method.
omega	[multinet] Inter-layer weight parameter in the generalized louvain method
louv_random	[community] Will randomize the node evaluation order and the community evaluation order to get different partitions at each call
return_gobject	boolean: return giotto object (default = TRUE)
set_seed	set seed
seed_number	number for seed
...	additional parameters

### Details

Louvain clustering using the community or multinet implementation of the louvain clustering algorithm.

### Value

giotto object with new clusters appended to cell metadata

### See Also

[doLouvainCluster\\_community](#) and [doLouvainCluster\\_multinet](#)

### Examples

```
doLouvainCluster(gobject)
```

---

```
doLouvainCluster_community
doLouvainCluster_community
```

---

### Description

cluster cells using a NN-network and the Louvain algorithm from the community module in Python

### Usage

```
doLouvainCluster_community(
  gobject,
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = NULL,
  louv_random = F,
```

```

    return_gobject = TRUE,
    set_seed = F,
    seed_number = 1234
)

```

### Arguments

<code>gobject</code>	giotto object
<code>name</code>	name for cluster
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use
<code>python_path</code>	specify specific path to python if required
<code>resolution</code>	resolution
<code>weight_col</code>	weight column to use for edges
<code>louv_random</code>	Will randomize the node evaluation order and the community evaluation order to get different partitions at each call
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>set_seed</code>	set seed
<code>seed_number</code>	number for seed

### Details

This function is a wrapper for the Louvain algorithm implemented in Python, which can detect communities in graphs of nodes (cells). See the <https://python-louvain.readthedocs.io/en/latest/index.html> readthedocs page for more information.

Set `weight_col = NULL` to give equal weight (=1) to each edge.

### Value

giotto object with new clusters appended to cell metadata

### Examples

```
doLouvainCluster_community(gobject)
```

---

```
doLouvainCluster_multinet
doLouvainCluster_multinet
```

---

### Description

cluster cells using a NN-network and the Louvain algorithm from the multinet package in R.

**Usage**

```
doLouvainCluster_multinet(
  gobject,
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  gamma = 1,
  omega = 1,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234
)
```

**Arguments**

gobject	giotto object
name	name for cluster
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use
gamma	Resolution parameter for modularity in the generalized louvain method.
omega	Inter-layer weight parameter in the generalized louvain method.
return_gobject	boolean: return giotto object (default = TRUE)
set_seed	set seed
seed_number	number for seed

**Details**

See [glouvain\\_ml](#) from the multinet package in R for more information.

**Value**

giotto object with new clusters appended to cell metadata

**Examples**

```
doLouvainCluster_multinet(gobject)
```

---

doLouvainSubCluster	<i>doLouvainSubCluster</i>
---------------------	----------------------------

---

**Description**

subcluster cells using a NN-network and the Louvain algorithm

**Usage**

```
doLouvainSubCluster(
  gobject,
  name = "sub_louvain_clus",
  version = c("community", "multinet"),
  cluster_column = NULL,
  selected_clusters = NULL,
  hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 0.5,
  gamma = 1,
  omega = 1,
  python_path = NULL,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>name</code>	name for new clustering result
<code>version</code>	version of Louvain algorithm to use
<code>cluster_column</code>	cluster column to subcluster
<code>selected_clusters</code>	only do subclustering on these clusters
<code>hvg_param</code>	parameters for calculateHVG
<code>hvg_min_perc_cells</code>	threshold for detection in min percentage of cells
<code>hvg_mean_expr_det</code>	threshold for mean expression level in cells with detection
<code>use_all_genes_as_hvg</code>	forces all genes to be HVG and to be used as input for PCA
<code>min_nr_of_hvg</code>	minimum number of HVG, or all genes will be used as input for PCA
<code>pca_param</code>	parameters for runPCA
<code>nn_param</code>	parameters for parameters for createNearestNetwork
<code>k_neighbors</code>	number of k for createNearestNetwork
<code>resolution</code>	resolution for community algorithm
<code>gamma</code>	gamma
<code>omega</code>	omega

python_path	specify specific path to python if required
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use
return_gobject	boolean: return giotto object (default = TRUE)
verbose	verbose

### Details

This function performs subclustering using the Louvain algorithm on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do Louvain clustering

### Value

giotto object with new subclusters appended to cell metadata

### See Also

[doLouvainCluster\\_multinet](#) and [doLouvainCluster\\_community](#)

### Examples

```
doLouvainSubCluster(gobject)
```

---

```
doLouvainSubCluster_community
doLouvainSubCluster_community
```

---

### Description

subcluster cells using a NN-network and the Louvain community detection algorithm

### Usage

```
doLouvainSubCluster_community(
  gobject,
  name = "sub_louvain_comm_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
  hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
```



```

min_nr_of_hvg = 5,
pca_param = list(expression_values = "normalized", scale_unit = T),
nn_param = list(dimensions_to_use = 1:20),
k_neighbors = 10,
resolution = 0.5,
python_path = NULL,
nn_network_to_use = "sNN",
network_name = "sNN.pca",
return_gobject = TRUE,
verbose = T
)

```

### Arguments

<code>gobject</code>	giotto object
<code>name</code>	name for new clustering result
<code>cluster_column</code>	cluster column to subcluster
<code>selected_clusters</code>	only do subclustering on these clusters
<code>hvg_param</code>	parameters for calculateHVG
<code>hvg_min_perc_cells</code>	threshold for detection in min percentage of cells
<code>hvg_mean_expr_det</code>	threshold for mean expression level in cells with detection
<code>use_all_genes_as_hvg</code>	forces all genes to be HVG and to be used as input for PCA
<code>min_nr_of_hvg</code>	minimum number of HVG, or all genes will be used as input for PCA
<code>pca_param</code>	parameters for runPCA
<code>nn_param</code>	parameters for parameters for createNearestNetwork
<code>k_neighbors</code>	number of k for createNearestNetwork
<code>resolution</code>	resolution
<code>python_path</code>	specify specific path to python if required
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>verbose</code>	verbose

### Details

This function performs subclustering using the Louvain community algorithm on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do Louvain community clustering

**Value**

giotto object with new subclusters appended to cell metadata

**See Also**

[doLouvainCluster\\_community](#)

**Examples**

```
doLouvainSubCluster_community(gobject)
```

---

```
doLouvainSubCluster_multinet
```

```
doLouvainSubCluster_multinet
```

---

**Description**

subcluster cells using a NN-network and the Louvain multinet detection algorithm

**Usage**

```
doLouvainSubCluster_multinet(
  gobject,
  name = "sub_louvain_mult_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
  hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  gamma = 1,
  omega = 1,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>name</code>	name for new clustering result
<code>cluster_column</code>	cluster column to subcluster
<code>selected_clusters</code>	only do subclustering on these clusters

hvg_param	parameters for calculateHVG
hvg_min_perc_cells	threshold for detection in min percentage of cells
hvg_mean_expr_det	threshold for mean expression level in cells with detection
use_all_genes_as_hvg	forces all genes to be HVG and to be used as input for PCA
min_nr_of_hvg	minimum number of HVG, or all genes will be used as input for PCA
pca_param	parameters for runPCA
nn_param	parameters for parameters for createNearestNetwork
k_neighbors	number of k for createNearestNetwork
gamma	gamma
omega	omega
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use
return_gobject	boolean: return giotto object (default = TRUE)
verbose	verbose

## Details

This function performs subclustering using the Louvain multinet algorithm on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do Louvain multinet clustering

## Value

giotto object with new subclusters appended to cell metadata

## See Also

[doLouvainCluster\\_multinet](#)

## Examples

```
doLouvainSubCluster_multinet(gobject)
```

---

doRandomWalkCluster     *doRandomWalkCluster*


---

## Description

Cluster cells using a random walk approach.

## Usage

```
doRandomWalkCluster(
  gobject,
  name = "random_walk_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  walk_steps = 4,
  walk_clusters = 10,
  walk_weights = NA,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234
)
```

## Arguments

gobject	giotto object
name	name for cluster
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use
walk_steps	number of walking steps
walk_clusters	number of final clusters
walk_weights	cluster column defining the walk weights
return_gobject	boolean: return giotto object (default = TRUE)
set_seed	set seed
seed_number	number for seed

## Details

See [cluster\\_walktrap](#) function from the igraph package in R for more information.

## Value

giotto object with new clusters appended to cell metadata

## Examples

```
doRandomWalkCluster(gobject)
```

---

doSNNCluster	<i>doSNNCluster</i>
--------------	---------------------

---

## Description

Cluster cells using a SNN cluster approach.

## Usage

```
doSNNCluster(
  gobject,
  name = "sNN_clus",
  nn_network_to_use = "kNN",
  network_name = "kNN.pca",
  k = 20,
  eps = 4,
  minPts = 16,
  borderPoints = TRUE,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234
)
```

## Arguments

gobject	giotto object
name	name for cluster
nn_network_to_use	type of NN network to use (only works on kNN)
network_name	name of kNN network to use
k	Neighborhood size for nearest neighbor sparsification to create the shared NN graph.
eps	Two objects are only reachable from each other if they share at least eps nearest neighbors.
minPts	minimum number of points that share at least eps nearest neighbors for a point to be considered a core points.
borderPoints	should borderPoints be assigned to clusters like in DBSCAN?
return_gobject	boolean: return giotto object (default = TRUE)
set_seed	set seed
seed_number	number for seed

## Details

See [sNNclust](#) from dbscan package

## Value

giotto object with new clusters appended to cell metadata

**Examples**

```
doSNNCluster(gobject)
```

---

estimateImageBg	<i>estimateImageBg</i>
-----------------	------------------------

---

**Description**

helps to estimate which color is the background color of your plot

**Usage**

```
estimateImageBg(mg_object, top_color_range = 1:50)
```

**Arguments**

mg_object	magick image or Giotto image object
top_color_range	top possible background colors to return

**Value**

vector of pixel color frequencies and an associated barplot

**Examples**

```
estimateImageBg(mg_object)
```

---

exportGiottoViewer	<i>exportGiottoViewer</i>
--------------------	---------------------------

---

**Description**

compute highly variable genes

**Usage**

```
exportGiottoViewer(
  gobject,
  output_directory = NULL,
  spat_enr_names = NULL,
  factor_annotations = NULL,
  numeric_annotations = NULL,
  dim_reductions,
  dim_reduction_names,
  expression_values = c("scaled", "normalized", "custom"),
  dim_red_rounding = NULL,
  dim_red_rescale = c(-20, 20),
  expression_rounding = 2,
  overwrite_dir = T,
  verbose = T
)
```

**Arguments**

**gobject**                giotto object  
**output\_directory**        directory where to save the files  
**spat\_enr\_names**    spatial enrichment results to include for annotations  
**factor\_annotations**        giotto cell annotations to view as factor  
**numeric\_annotations**        giotto cell annotations to view as numeric  
**dim\_reductions**    high level dimension reductions to view  
**dim\_reduction\_names**        specific dimension reduction names  
**expression\_values**        expression values to use in Viewer  
**dim\_red\_rounding**        numerical indicating how to round the coordinates  
**dim\_red\_rescale**        numerals to rescale the coordinates  
**expression\_rounding**        numerical indicating how to round the expression data  
**overwrite\_dir**    overwrite files in the directory if it already existed  
**verbose**            be verbose

**Details**

Giotto Viewer expects the results from Giotto Analyzer in a specific format, which is provided by this function. To include enrichment results from [createSpatialEnrich](#) include the provided spatial enrichment name (default PAGE or rank) and add the gene signature names (.e.g cell types) to the numeric annotations parameter.

**Value**

writes the necessary output to use in Giotto Viewer

**Examples**

```
exportGiottoViewer(gobject)
```

---

 exprCellCellcom

*exprCellCellcom*


---

**Description**

Cell-Cell communication scores based on expression only

**Usage**

```

exprCellCellcom(
  gobject,
  cluster_column = "cell_types",
  random_iter = 1000,
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  adjust_method = c("fdr", "bonferroni", "BH", "holm", "hochberg", "hommel", "BY",
    "none"),
  adjust_target = c("genes", "cells"),
  verbose = T
)

```

**Arguments**

<code>gobject</code>	giotto object to use
<code>cluster_column</code>	cluster column with cell type information
<code>random_iter</code>	number of iterations
<code>gene_set_1</code>	first specific gene set from gene pairs
<code>gene_set_2</code>	second specific gene set from gene pairs
<code>log2FC_addendum</code>	addendum to add when calculating log2FC
<code>adjust_method</code>	which method to adjust p-values
<code>adjust_target</code>	adjust multiple hypotheses at the cell or gene level
<code>verbose</code>	verbose

**Details**

Statistical framework to identify if pairs of genes (such as ligand-receptor combinations) are expressed at higher levels than expected based on a reshuffled null distribution of gene expression values, without considering the spatial position of cells. More details will follow soon.

**Value**

Cell-Cell communication scores for gene pairs based on expression only

**Examples**

```
exprCellCellcom(gobject)
```



fDataDT

*fDataDT***Description**

show gene metadata

**Usage**

fDataDT(gobject)

**Arguments**

gobject                  giotto object

**Value**

data.table with gene metadata

**Examples**

pDataDT(gobject)

filterCellProximityGenes

*filterCellProximityGenes***Description**

Filter cell proximity gene scores.

**Usage**

```
filterCellProximityGenes(
  cpqObject,
  min_cells = 4,
  min_cells_expr = 1,
  min_int_cells = 4,
  min_int_cells_expr = 1,
  min_fdr = 0.1,
  min_spat_diff = 0.2,
  min_log2_fc = 0.2,
  min_zscore = 2,
  zscores_column = c("cell_type", "genes"),
  direction = c("both", "up", "down")
)
```

**Arguments**

cpgObject	cell proximity gene score object
min_cells	minimum number of source cell type
min_cells_expr	minimum expression level for source cell type
min_int_cells	minimum number of interacting neighbor cell type
min_int_cells_expr	minimum expression level for interacting neighbor cell type
min_fdr	minimum adjusted p-value
min_spat_diff	minimum absolute spatial expression difference
min_log2_fc	minimum log2 fold-change
min_zscore	minimum z-score change
zscores_column	calculate z-scores over cell types or genes
direction	differential expression directions to keep

**Value**

cpgObject that contains the filtered differential gene scores

**Examples**

```
filterCellProximityGenes(gobject)
```

---

filterCombinations	<i>filterCombinations</i>
--------------------	---------------------------

---

**Description**

Shows how many genes and cells are lost with combinations of thresholds.

**Usage**

```
filterCombinations(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_thresholds = c(1, 2),
  gene_det_in_min_cells = c(5, 50),
  min_det_genes_per_cell = c(200, 400),
  scale_x_axis = "identity",
  x_axis_offset = 0,
  scale_y_axis = "identity",
  y_axis_offset = 0,
  show_plot = TRUE,
  return_plot = FALSE,
  save_plot = NA,
  save_param = list(),
  default_save_name = "filterCombinations"
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>expression_thresholds</code>	all thresholds to consider a gene expressed
<code>gene_det_in_min_cells</code>	minimum number of cells that should express a gene to consider that gene further
<code>min_det_genes_per_cell</code>	minimum number of expressed genes per cell to consider that cell further
<code>scale_x_axis</code>	ggplot transformation for x-axis (e.g. log2)
<code>x_axis_offset</code>	x-axis offset to be used together with the scaling transformation
<code>scale_y_axis</code>	ggplot transformation for y-axis (e.g. log2)
<code>y_axis_offset</code>	y-axis offset to be used together with the scaling transformation
<code>show_plot</code>	show plot
<code>return_plot</code>	return only ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change save_name in save_param

**Details**

Creates a scatterplot that visualizes the number of genes and cells that are lost with a specific combination of a gene and cell threshold given an arbitrary cutoff to call a gene expressed. This function can be used to make an informed decision at the filtering step with filterGiotto.

**Value**

list of data.table and ggplot object

**Examples**

```
filterCombinations(gobject)
```

---

filterCPG

*filterCPG*


---

**Description**

Filter cell proximity gene scores.

**Usage**

```
filterCPG(
  cpgObject,
  min_cells = 4,
  min_cells_expr = 1,
  min_int_cells = 4,
  min_int_cells_expr = 1,
  min_fdr = 0.1,
  min_spat_diff = 0.2,
  min_log2_fc = 0.2,
  min_zscore = 2,
  zscores_column = c("cell_type", "genes"),
  direction = c("both", "up", "down")
)
```

**Arguments**

cpgObject	cell proximity gene score object
min_cells	minimum number of source cell type
min_cells_expr	minimum expression level for source cell type
min_int_cells	minimum number of interacting neighbor cell type
min_int_cells_expr	minimum expression level for interacting neighbor cell type
min_fdr	minimum adjusted p-value
min_spat_diff	minimum absolute spatial expression difference
min_log2_fc	minimum log2 fold-change
min_zscore	minimum z-score change
zscores_column	calculate z-scores over cell types or genes
direction	differential expression directions to keep

**Value**

cpgObject that contains the filtered differential gene scores

**Examples**

```
filterCPG(gobject)
```

---

filterDistributions	<i>filterDistributions</i>
---------------------	----------------------------

---

**Description**

show gene or cell distribution after filtering on expression threshold

**Usage**

```
filterDistributions(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_threshold = 1,
  detection = c("genes", "cells"),
  plot_type = c("histogram", "violin"),
  nr_bins = 30,
  fill_color = "lightblue",
  scale_axis = "identity",
  axis_offset = 0,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "filterDistributions"
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>expression_threshold</code>	threshold to consider a gene expressed
<code>detection</code>	consider genes or cells
<code>plot_type</code>	type of plot
<code>nr_bins</code>	number of bins for histogram plot
<code>fill_color</code>	fill color for plots
<code>scale_axis</code>	ggplot transformation for axis (e.g. log2)
<code>axis_offset</code>	offset to be used together with the scaling transformation
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

**Value**

ggplot object

**Examples**

```
filterDistributions(gobject)
```

---

filterGiotto

*filterGiotto*


---

## Description

filter Giotto object based on expression threshold

## Usage

```
filterGiotto(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_threshold = 1,
  gene_det_in_min_cells = 100,
  min_det_genes_per_cell = 100,
  verbose = F
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>expression_threshold</code>	threshold to consider a gene expressed
<code>gene_det_in_min_cells</code>	minimum # of cells that need to express a gene
<code>min_det_genes_per_cell</code>	minimum # of genes that need to be detected in a cell
<code>verbose</code>	verbose

## Details

The function [filterCombinations](#) can be used to explore the effect of different parameter values.

## Value

giotto object

## Examples

```
filterGiotto(gobject)
```

---

```
findCellProximityGenes
      findCellProximityGenes
```

---

## Description

Identifies genes that are differentially expressed due to proximity to other cell types.

## Usage

```
findCellProximityGenes(
  gobject,
  expression_values = "normalized",
  selected_genes = NULL,
  cluster_column,
  spatial_network_name = "Delaunay_network",
  minimum_unique_cells = 1,
  minimum_unique_int_cells = 1,
  diff_test = c("permutation", "limma", "t.test", "wilcox"),
  mean_method = c("arithmetic", "geometric"),
  offset = 0.1,
  adjust_method = c("bonferroni", "BH", "holm", "hochberg", "hommel", "BY", "fdr",
    "none"),
  nr_permutations = 1000,
  exclude_selected_cells_from_test = T,
  do_parallel = TRUE,
  cores = NA
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>selected_genes</code>	subset of selected genes (optional)
<code>cluster_column</code>	name of column to use for cell types
<code>spatial_network_name</code>	name of spatial network to use
<code>minimum_unique_cells</code>	minimum number of target cells required
<code>minimum_unique_int_cells</code>	minimum number of interacting cells required
<code>diff_test</code>	which differential expression test
<code>mean_method</code>	method to use to calculate the mean
<code>offset</code>	offset value to use when calculating log2 ratio
<code>adjust_method</code>	which method to adjust p-values
<code>nr_permutations</code>	number of permutations if <code>diff_test = permutation</code>

exclude_selected_cells_from_test	exclude interacting cells other cells
do_parallel	run calculations in parallel with mclapply
cores	number of cores to use if do_parallel = TRUE

## Details

Function to calculate if genes are differentially expressed in cell types when they interact (approximated by physical proximity) with other cell types. The results data.table in the cpgObject contains - at least - the following columns:

- genes: All or selected list of tested genes
- sel: average gene expression in the interacting cells from the target cell type
- other: average gene expression in the NOT-interacting cells from the target cell type
- log2fc: log2 fold-change between sel and other
- diff: spatial expression difference between sel and other
- p.value: associated p-value
- p.adj: adjusted p-value
- cell\_type: target cell type
- int\_cell\_type: interacting cell type
- nr\_select: number of cells for selected target cell type
- int\_nr\_select: number of cells for interacting cell type
- nr\_other: number of other cells of selected target cell type
- int\_nr\_other: number of other cells for interacting cell type
- unif\_int: cell-cell interaction

## Value

cpgObject that contains the differential gene scores

## Examples

```
findCellProximityGenes(gobject)
```

---

findCPG

*findCPG*


---

## Description

Identifies genes that are differentially expressed due to proximity to other cell types.



**Usage**

```
findCPG(
  gobject,
  expression_values = "normalized",
  selected_genes = NULL,
  cluster_column,
  spatial_network_name = "Delaunay_network",
  minimum_unique_cells = 1,
  minimum_unique_int_cells = 1,
  diff_test = c("permutation", "limma", "t.test", "wilcox"),
  mean_method = c("arithmetic", "geometric"),
  offset = 0.1,
  adjust_method = c("bonferroni", "BH", "holm", "hochberg", "hommel", "BY", "fdr",
    "none"),
  nr_permutations = 100,
  exclude_selected_cells_from_test = T,
  do_parallel = TRUE,
  cores = NA
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>selected_genes</code>	subset of selected genes (optional)
<code>cluster_column</code>	name of column to use for cell types
<code>spatial_network_name</code>	name of spatial network to use
<code>minimum_unique_cells</code>	minimum number of target cells required
<code>minimum_unique_int_cells</code>	minimum number of interacting cells required
<code>diff_test</code>	which differential expression test
<code>mean_method</code>	method to use to calculate the mean
<code>offset</code>	offset value to use when calculating log2 ratio
<code>adjust_method</code>	which method to adjust p-values
<code>nr_permutations</code>	number of permutations if <code>diff_test = permutation</code>
<code>exclude_selected_cells_from_test</code>	exclude interacting cells other cells
<code>do_parallel</code>	run calculations in parallel with <code>mclapply</code>
<code>cores</code>	number of cores to use if <code>do_parallel = TRUE</code>

**Details**

Function to calculate if genes are differentially expressed in cell types when they interact (approximated by physical proximity) with other cell types. The results data.table in the `cpgObject` contains - at least - the following columns:

- genes: All or selected list of tested genes
- sel: average gene expression in the interacting cells from the target cell type
- other: average gene expression in the NOT-interacting cells from the target cell type
- log2fc: log2 fold-change between sel and other
- diff: spatial expression difference between sel and other
- p.value: associated p-value
- p.adj: adjusted p-value
- cell\_type: target cell type
- int\_cell\_type: interacting cell type
- nr\_select: number of cells for selected target cell type
- int\_nr\_select: number of cells for interacting cell type
- nr\_other: number of other cells of selected target cell type
- int\_nr\_other: number of other cells for interacting cell type
- unif\_int: cell-cell interaction

### Value

cpgObject that contains the differential gene scores

### Examples

```
findCPG(gobject)
```

---

findGiniMarkers

*findGiniMarkers*

---

### Description

Identify marker genes for selected clusters based on gini detection and expression scores.

### Usage

```
findGiniMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  min_expr_gini_score = 0.2,
  min_det_gini_score = 0.2,
  detection_threshold = 0,
  rank_score = 1,
  min_genes = 5
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>cluster_column</code>	clusters to use
<code>subset_clusters</code>	selection of clusters to compare
<code>group_1</code>	group 1 cluster IDs from <code>cluster_column</code> for pairwise comparison
<code>group_2</code>	group 2 cluster IDs from <code>cluster_column</code> for pairwise comparison
<code>min_expr_gini_score</code>	filter on minimum gini coefficient for expression
<code>min_det_gini_score</code>	filter on minimum gini coefficient for detection
<code>detection_threshold</code>	detection threshold for gene expression
<code>rank_score</code>	rank scores for both detection and expression to include
<code>min_genes</code>	minimum number of top genes to return

**Details**

Detection of marker genes using the [https://en.wikipedia.org/wiki/Gini\\_coefficient](https://en.wikipedia.org/wiki/Gini_coefficient) gini coefficient is based on the following steps/principles per gene:

- 1. calculate average expression per cluster
- 2. calculate detection fraction per cluster
- 3. calculate gini-coefficient for av. expression values over all clusters
- 4. calculate gini-coefficient for detection fractions over all clusters
- 5. convert gini-scores to rank scores
- 6. for each gene create combined score = detection rank x expression rank x expr gini-coefficient x detection gini-coefficient
- 7. for each gene sort on expression and detection rank and combined score

As a results "top gini" genes are genes that are very selectively expressed in a specific cluster, however not always expressed in all cells of that cluster. In other words highly specific, but not necessarily sensitive at the single-cell level.

To perform differential expression between cluster groups you need to specify cluster IDs to the parameters `group_1` and `group_2`.

**Value**

data.table with marker genes

**Examples**

```
findGiniMarkers(gobject)
```

---

```
findGiniMarkers_one_vs_all
      findGiniMarkers_one_vs_all
```

---

### Description

Identify marker genes for all clusters in a one vs all manner based on gini detection and expression scores.

### Usage

```
findGiniMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank_score = 1,
  min_genes = 4,
  verbose = TRUE
)
```

### Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>cluster_column</code>	clusters to use
<code>subset_clusters</code>	selection of clusters to compare
<code>min_expr_gini_score</code>	filter on minimum gini coefficient on expression
<code>min_det_gini_score</code>	filter on minimum gini coefficient on detection
<code>detection_threshold</code>	detection threshold for gene expression
<code>rank_score</code>	rank scores for both detection and expression to include
<code>min_genes</code>	minimum number of top genes to return
<code>verbose</code>	be verbose

### Value

data.table with marker genes

### See Also

[findGiniMarkers](#)

Examples

```
findGiniMarkers_one_vs_all(gobject)
```

---

findMarkers	<i>findMarkers</i>
-------------	--------------------

---

Description

Identify marker genes for selected clusters.

Usage

```
findMarkers(  
  gobject,  
  expression_values = c("normalized", "scaled", "custom"),  
  cluster_column = NULL,  
  method = c("scrn", "gini", "mast"),  
  subset_clusters = NULL,  
  group_1 = NULL,  
  group_2 = NULL,  
  min_expr_gini_score = 0.5,  
  min_det_gini_score = 0.5,  
  detection_threshold = 0,  
  rank_score = 1,  
  min_genes = 4,  
  group_1_name = NULL,  
  group_2_name = NULL,  
  adjust_columns = NULL,  
  ...  
)
```

Arguments

gobject	giotto object
expression_values	gene expression values to use
cluster_column	clusters to use
method	method to use to detect differentially expressed genes
subset_clusters	selection of clusters to compare
group_1	group 1 cluster IDs from cluster_column for pairwise comparison
group_2	group 2 cluster IDs from cluster_column for pairwise comparison
min_expr_gini_score	gini: filter on minimum gini coefficient for expression
min_det_gini_score	gini: filter minimum gini coefficient for detection
detection_threshold	gini: detection threshold for gene expression

rank_score	gini: rank scores to include
min_genes	minimum number of top genes to return (for gini)
group_1_name	mast: custom name for group_1 clusters
group_2_name	mast: custom name for group_2 clusters
adjust_columns	mast: column in pDataDT to adjust for (e.g. detection rate)
...	additional parameters for the findMarkers function in scran or zlm function in MAST

### Details

Wrapper for all individual functions to detect marker genes for clusters.

### Value

data.table with marker genes

### See Also

[findScranMarkers](#), [findGiniMarkers](#) and [findMastMarkers](#)

### Examples

```
findMarkers(gobject)
```

---

```
findMarkers_one_vs_all
```

```
findMarkers_one_vs_all
```

---

### Description

Identify marker genes for all clusters in a one vs all manner.

### Usage

```
findMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  method = c("scran", "gini", "mast"),
  pval = 0.01,
  logFC = 0.5,
  min_genes = 10,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank_score = 1,
  adjust_columns = NULL,
  verbose = TRUE,
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>cluster_column</code>	clusters to use
<code>subset_clusters</code>	selection of clusters to compare
<code>method</code>	method to use to detect differentially expressed genes
<code>pval</code>	scan & mast: filter on minimal p-value
<code>logFC</code>	scan & mast: filter on logFC
<code>min_genes</code>	minimum genes to keep per cluster, overrides pval and logFC
<code>min_expr_gini_score</code>	gini: filter on minimum gini coefficient for expression
<code>min_det_gini_score</code>	gini: filter minimum gini coefficient for detection
<code>detection_threshold</code>	gini: detection threshold for gene expression
<code>rank_score</code>	gini: rank scores to include
<code>adjust_columns</code>	mast: column in pDataDT to adjust for (e.g. detection rate)
<code>verbose</code>	be verbose
<code>...</code>	additional parameters for the findMarkers function in scan or zlm function in MAST

**Details**

Wrapper for all one vs all functions to detect marker genes for clusters.

**Value**

data.table with marker genes

**See Also**

[findScranMarkers\\_one\\_vs\\_all](#), [findGiniMarkers\\_one\\_vs\\_all](#) and [findMastMarkers\\_one\\_vs\\_all](#)

**Examples**

```
findMarkers_one_vs_all(gobject)
```

---

findMastMarkers	<i>findMastMarkers</i>
-----------------	------------------------

---

## Description

Identify marker genes for selected clusters based on the MAST package.

## Usage

```
findMastMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  group_1 = NULL,
  group_1_name = NULL,
  group_2 = NULL,
  group_2_name = NULL,
  adjust_columns = NULL,
  ...
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>cluster_column</code>	clusters to use
<code>group_1</code>	group 1 cluster IDs from <code>cluster_column</code> for pairwise comparison
<code>group_1_name</code>	custom name for <code>group_1</code> clusters
<code>group_2</code>	group 2 cluster IDs from <code>cluster_column</code> for pairwise comparison
<code>group_2_name</code>	custom name for <code>group_2</code> clusters
<code>adjust_columns</code>	column in <code>pDataDT</code> to adjust for (e.g. detection rate)
<code>...</code>	additional parameters for the <code>zlm</code> function in MAST

## Details

This is a minimal convenience wrapper around the [zlm](#) from the MAST package to detect differentially expressed genes.

## Value

data.table with marker genes

## Examples

```
findMastMarkers(gobject)
```



---

```
findMastMarkers_one_vs_all  
  findMastMarkers_one_vs_all
```

---

## Description

Identify marker genes for all clusters in a one vs all manner based on the MAST package.

## Usage

```
findMastMarkers_one_vs_all(  
  gobject,  
  expression_values = c("normalized", "scaled", "custom"),  
  cluster_column,  
  subset_clusters = NULL,  
  adjust_columns = NULL,  
  pval = 0.001,  
  logFC = 1,  
  min_genes = 10,  
  verbose = TRUE,  
  ...  
)
```

## Arguments

gobject	giotto object
expression_values	gene expression values to use
cluster_column	clusters to use
subset_clusters	selection of clusters to compare
adjust_columns	column in pDataDT to adjust for (e.g. detection rate)
pval	filter on minimal p-value
logFC	filter on logFC
min_genes	minimum genes to keep per cluster, overrides pval and logFC
verbose	be verbose
...	additional parameters for the zlm function in MAST

## Value

data.table with marker genes

## See Also

[findMastMarkers](#)

## Examples

```
findMastMarkers_one_vs_all(gobject)
```

---

findNetworkNeighbors    *findNetworkNeighbors*


---

### Description

Find the spatial neighbors for a selected group of cells within the selected spatial network.

### Usage

```
findNetworkNeighbors(
  gobject,
  spatial_network_name,
  source_cell_ids = NULL,
  name = "nb_cells"
)
```

### Arguments

gobject	Giotto object
spatial_network_name	name of spatial network
source_cell_ids	cell ids for which you want to know the spatial neighbors
name	name of the results

### Value

data.table

### Examples

```
findNetworkNeighbors(gobject)
```

---

findScranMarkers    *findScranMarkers*


---

### Description

Identify marker genes for all or selected clusters based on scran's implementation of findMarkers.

### Usage

```
findScranMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>cluster_column</code>	clusters to use
<code>subset_clusters</code>	selection of clusters to compare
<code>group_1</code>	group 1 cluster IDs from <code>cluster_column</code> for pairwise comparison
<code>group_2</code>	group 2 cluster IDs from <code>cluster_column</code> for pairwise comparison
<code>...</code>	additional parameters for the <code>findMarkers</code> function in <code>scrn</code>

**Details**

This is a minimal convenience wrapper around the [findMarkers](#) function from the `scrn` package.

To perform differential expression between cluster groups you need to specify cluster IDs to the parameters `group_1` and `group_2`.

**Value**

data.table with marker genes

**Examples**

```
findScranMarkers(gobject)
```

---

```
findScranMarkers_one_vs_all
  findScranMarkers_one_vs_all
```

---

**Description**

Identify marker genes for all clusters in a one vs all manner based on `scrn`'s implementation of `findMarkers`.

**Usage**

```
findScranMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  pval = 0.01,
  logFC = 0.5,
  min_genes = 10,
  verbose = TRUE,
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>cluster_column</code>	clusters to use
<code>subset_clusters</code>	subset of clusters to use
<code>pval</code>	filter on minimal p-value
<code>logFC</code>	filter on logFC
<code>min_genes</code>	minimum genes to keep per cluster, overrides pval and logFC
<code>verbose</code>	be verbose
<code>...</code>	additional parameters for the findMarkers function in scan

**Value**

data.table with marker genes

**See Also**

[findScranMarkers](#)

**Examples**

```
findScranMarkers_one_vs_all(gobject)
```

---

<code>get10Xmatrix</code>	<i>get10Xmatrix</i>
---------------------------	---------------------

---

**Description**

This function creates an expression matrix from a 10X structured folder

**Usage**

```
get10Xmatrix(path_to_data, gene_column_index = 1)
```

**Arguments**

<code>path_to_data</code>	path to the 10X folder
<code>gene_column_index</code>	which column from the features or genes .tsv file to use for row ids

**Details**

A typical 10X folder is named `raw_feature_bc_matrix` or `raw_feature_bc_matrix` and it has 3 files:

- `barcodes.tsv.gz`
- `features.tsv.gz` or `genes.tsv.gz`
- `matrix.mtx.gz`

By default the first column of the features or genes .tsv file will be used, however if multiple annotations are provided (e.g. ensembl gene ids and gene symbols) the user can select another column.

**Value**

sparse expression matrix from 10X

**Examples**

```
get10Xmatrix(path_to_data)
```

---

```
getClusterSimilarity  getClusterSimilarity
```

---

**Description**

Creates data.table with pairwise correlation scores between each cluster.

**Usage**

```
getClusterSimilarity(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman")
)
```

**Arguments**

gobject	giotto object
expression_values	expression values to use
cluster_column	name of column to use for clusters
cor	correlation score to calculate distance

**Details**

Creates data.table with pairwise correlation scores between each cluster and the group size (# of cells) for each cluster. This information can be used together with mergeClusters to combine very similar or small clusters into bigger clusters.

**Value**

data.table

**Examples**

```
getClusterSimilarity(gobject)
```

---

getDendrogramSplits	<i>getDendrogramSplits</i>
---------------------	----------------------------

---

## Description

Split dendrogram at each node and keep the leave (label) information..

## Usage

```
getDendrogramSplits(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  h = NULL,
  h_color = "red",
  show_dend = TRUE,
  verbose = TRUE
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>cluster_column</code>	name of column to use for clusters
<code>cor</code>	correlation score to calculate distance
<code>distance</code>	distance method to use for hierarchical clustering
<code>h</code>	height of horizontal lines to plot
<code>h_color</code>	color of horizontal lines
<code>show_dend</code>	show dendrogram
<code>verbose</code>	be verbose

## Details

Creates a `data.table` with three columns and each row represents a node in the dendrogram. For each node the height of the node is given together with the two subdendrograms. This information can be used to determine in a hierarchical manner differentially expressed marker genes at each node.

## Value

`data.table` object

## Examples

```
getDendrogramSplits(gobject)
```

---

getDistinctColors	<i>getDistinctColors</i>
-------------------	--------------------------

---

**Description**

Returns a number of distinct colors based on the RGB scale

**Usage**

```
getDistinctColors(n)
```

**Arguments**

n	number of colors wanted
---	-------------------------

**Value**

number of distinct colors

---

getGiottoImage	<i>getGiottoImage</i>
----------------	-----------------------

---

**Description**

get get a giotto image from a giotto object

**Usage**

```
getGiottoImage(gobject, image_name)
```

**Arguments**

gobject	giotto object
image_name	name of giotto image <a href="#">showGiottoImageNames</a>

**Value**

a giotto image

**Examples**

```
getGiottoImage(gobject)
```

---

<code>get_os</code>	<i>get_os</i>
---------------------	---------------

---

**Description**

return the type of operating system, see <https://conjugateprior.org/2015/06/identifying-the-os-from-r/>

**Usage**

```
get_os()
```

**Value**

character osx, linux or windows

---

<code>giotto-class</code>	<i>S4 giotto Class</i>
---------------------------	------------------------

---

**Description**

Framework of giotto object to store and work with spatial expression data

**Slots**

`raw_exprs` raw expression counts

`norm_expr` normalized expression counts

`norm_scaled_expr` normalized and scaled expression counts

`custom_expr` custom normalized counts

`spatial_locs` spatial location coordinates for cells

`cell_metadata` metadata for cells

`gene_metadata` metadata for genes

`cell_ID` unique cell IDs

`gene_ID` unique gene IDs

`spatial_network` spatial network in data.table/data.frame format

`spatial_grid` spatial grid in data.table/data.frame format

`spatial_enrichment` slot to save spatial enrichment-like results

`dimension_reduction` slot to save dimension reduction coordinates

`nn_network` nearest neighbor network in igraph format

`images` slot to store giotto images

`parameters` slot to save parameters that have been used

`instructions` slot for global function instructions

`offset_file` offset file used to stitch together image fields

`OS_platform` Operating System to run Giotto analysis on



---

heatmSpatialCorGenes    *heatmSpatialCorGenes*


---

## Description

Create heatmap of spatially correlated genes

## Usage

```
heatmSpatialCorGenes(
  gobject,
  spatCorObject,
  use_clus_name = NULL,
  show_cluster_annot = TRUE,
  show_row_dend = T,
  show_column_dend = F,
  show_row_names = F,
  show_column_names = F,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "heatmSpatialCorGenes",
  ...
)
```

## Arguments

<code>gobject</code>	giotto object
<code>spatCorObject</code>	spatial correlation object
<code>use_clus_name</code>	name of clusters to visualize (from <code>clusterSpatialCorGenes()</code> )
<code>show_cluster_annot</code>	show cluster annotation on top of heatmap
<code>show_row_dend</code>	show row dendrogram
<code>show_column_dend</code>	show column dendrogram
<code>show_row_names</code>	show row names
<code>show_column_names</code>	show column names
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters, see <a href="#">showSaveParameters</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>
<code>...</code>	additional parameters to the <a href="#">Heatmap</a> function from <code>ComplexHeatmap</code>

**Value**

Heatmap generated by ComplexHeatmap

**Examples**

```
heatmSpatialCorGenes(gobject)
```

---

hyperGeometricEnrich	<i>hyperGeometricEnrich</i>
----------------------	-----------------------------

---

**Description**

Function to calculate gene signature enrichment scores per spatial position using a hypergeometric test.

**Usage**

```
hyperGeometricEnrich(...)
```

**Arguments**

...	Arguments passed on to <a href="#">runHyperGeometricEnrich</a>
gobject	Giotto object
sign_matrix	Matrix of signature genes for each cell type / process
expression_values	expression values to use
reverse_log_scale	reverse expression values from log scale
logbase	log base to use if reverse_log_scale = TRUE
top_percentage	percentage of cells that will be considered to have gene expression with matrix binarization
output_enrichment	how to return enrichment output
p_value	calculate p-values (boolean, default = FALSE)
name	to give to spatial enrichment results, default = rank
return_gobject	return giotto object

**See Also**

[runHyperGeometricEnrich](#)

---

```
insertCrossSectionGenePlot3D
      insertCrossSectionGenePlot3D
```

---

## Description

Visualize cells and gene expression in a virtual cross section according to spatial coordinates

## Usage

```
insertCrossSectionGenePlot3D(
  gobject,
  crossSection_obj = NULL,
  name = NULL,
  spatial_network_name = "Delaunay_network",
  mesh_grid_color = "#1f77b4",
  mesh_grid_width = 3,
  mesh_grid_style = "dot",
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  show_other_cells = F,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "spatGenePlot3D_with_cross_section",
  ...
)
```

## Arguments

<code>gobject</code>	giotto object
<code>crossSection_obj</code>	cross section object as alternative input. default = NULL.
<code>name</code>	name of virtual cross section to use
<code>spatial_network_name</code>	name of spatial network to use
<code>mesh_grid_color</code>	color for the meshgrid lines
<code>mesh_grid_width</code>	width for the meshgrid lines
<code>mesh_grid_style</code>	style for the meshgrid lines
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>sdimz</code>	z-axis dimension name (default = 'sdimy')

show_other_cells	display not selected cells
axis_scale	axis_scale
custom_ratio	custom_ratio
show_plot	show plots
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from <a href="#">all_plots_save_function</a>
default_save_name	default save name for saving, don't change, change save_name in save_param
...	parameters for spatGenePlot3D

**Details**

Description of parameters.

**Value**

ggplot

**Examples**

```
insertCrossSectionGenePlot3D(gobject)
```

---

```
insertCrossSectionSpatPlot3D
      insertCrossSectionSpatPlot3D
```

---

**Description**

Visualize the meshgrid lines of cross section together with cells

**Usage**

```
insertCrossSectionSpatPlot3D(
  gobject,
  crossSection_obj = NULL,
  name = NULL,
  spatial_network_name = "Delaunay_network",
  mesh_grid_color = "#1f77b4",
  mesh_grid_width = 3,
  mesh_grid_style = "dot",
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  show_other_cells = F,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  default_save_name = "spat3D_with_cross_section",
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>crossSection_obj</code>	cross section object as alternative input. default = NULL.
<code>name</code>	name of virtual cross section to use
<code>spatial_network_name</code>	name of spatial network to use
<code>mesh_grid_color</code>	color for the meshgrid lines
<code>mesh_grid_width</code>	width for the meshgrid lines
<code>mesh_grid_style</code>	style for the meshgrid lines
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>sdimz</code>	z-axis dimension name (default = 'sdimy')
<code>show_other_cells</code>	display not selected cells
<code>axis_scale</code>	axis_scale
<code>custom_ratio</code>	custom_ratio
<code>default_save_name</code>	default save name for saving, don't change, change save_name in save_param
<code>...</code>	parameters for spatPlot3D

**Details**

Description of parameters.

**Value**

ggplot

**Examples**

```
insertCrossSectionSpatPlot3D(gobject)
```

---

jackstrawPlot	<i>jackstrawPlot</i>
---------------	----------------------

---

**Description**

identify significant principal components (PCs)

**Usage**

```
jackstrawPlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  genes_to_use = NULL,
  center = FALSE,
  scale_unit = FALSE,
  ncp = 20,
  ylim = c(0, 1),
  iter = 10,
  threshold = 0.01,
  verbose = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "jackstrawPlot"
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>reduction</code>	cells or genes
<code>genes_to_use</code>	subset of genes to use for PCA
<code>center</code>	center data before PCA
<code>scale_unit</code>	scale features before PCA
<code>ncp</code>	number of principal components to calculate
<code>ylim</code>	y-axis limits on jackstraw plot
<code>iter</code>	number of iterations for jackstraw
<code>threshold</code>	p-value threshold to call a PC significant
<code>verbose</code>	show progress of jackstraw method
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <code>all_plots_save_function()</code>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

**Details**

The Jackstraw method uses the [permutationPA](#) function. By systematically permuting genes it identifies robust, and thus significant, PCs.

**Value**

ggplot object for jackstraw method

**Examples**

```
jackstrawPlot(gobject)
```

---

loadHMRF	<i>loadHMRF</i>
----------	-----------------

---

**Description**

load previous HMRF

**Usage**

```
loadHMRF(  
  name_used = "test",  
  output_folder_used,  
  k_used = 10,  
  betas_used,  
  python_path_used  
)
```

**Arguments**

name_used	name of HMRF that was run
output_folder_used	output folder that was used
k_used	number of HMRF domains that was tested
betas_used	betas that were tested
python_path_used	python path that was used

**Details**

Description of HMRF parameters ...

**Value**

reloads a previous ran HMRF from doHRMF

**Examples**

```
loadHMRF(gobject)
```

---

makeSignMatrixPAGE	<i>makeSignMatrixPAGE</i>
--------------------	---------------------------

---

### Description

Function to convert a list of signature genes (e.g. for cell types or processes) into a binary matrix format that can be used with the PAGE enrichment option. Each cell type or process should have a vector of cell-type or process specific genes. These vectors need to be combined into a list (sign\_list). The names of the cell types or processes that are provided in the list need to be given (sign\_names).

### Usage

```
makeSignMatrixPAGE(sign_names, sign_list)
```

### Arguments

sign_names	vector with names for each provided gene signature
sign_list	list of genes (signature)

### Value

matrix

### See Also

[PAGEEnrich](#)

### Examples

```
makeSignMatrixPAGE()
```

---

makeSignMatrixRank	<i>makeSignMatrixRank</i>
--------------------	---------------------------

---

### Description

Function to convert a single-cell count matrix and a corresponding single-cell cluster vector into a rank matrix that can be used with the Rank enrichment option.

### Usage

```
makeSignMatrixRank(sc_matrix, sc_cluster_ids, gobject = NULL)
```

### Arguments

sc_matrix	matrix of single-cell RNAseq expression data
sc_cluster_ids	vector of cluster ids
gobject	if giotto object is given then only genes present in both datasets will be considered



**Value**

matrix

**See Also**[rankEnrich](#)**Examples**

```
makeSignMatrixRank()
```

---

mergeClusters	<i>mergeClusters</i>
---------------	----------------------

---

**Description**

Merge selected clusters based on pairwise correlation scores and size of cluster.

**Usage**

```
mergeClusters(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  new_cluster_name = "merged_cluster",
  min_cor_score = 0.8,
  max_group_size = 20,
  force_min_group_size = 10,
  max_sim_clusters = 10,
  return_gobject = TRUE,
  verbose = TRUE
)
```

**Arguments**

gobject	giotto object
expression_values	expression values to use
cluster_column	name of column to use for clusters
cor	correlation score to calculate distance
new_cluster_name	new name for merged clusters
min_cor_score	min correlation score to merge pairwise clusters
max_group_size	max cluster size that can be merged
force_min_group_size	size of clusters that will be merged with their most similar neighbor(s)
max_sim_clusters	maximum number of clusters to potentially merge to reach force_min_group_size
return_gobject	return giotto object
verbose	be verbose

**Details**

Merge selected clusters based on pairwise correlation scores and size of cluster. To avoid large clusters to merge the `max_group_size` can be lowered. Small clusters can be forcibly merged with their most similar pairwise cluster by adjusting the `force_min_group_size` parameter. Clusters smaller than this value will be merged independent on the provided `min_cor_score` value. The `force_min_group_size` might not always be reached if clusters have already been merged before. A giotto object is returned by default, if FALSE then the merging vector will be returned.

**Value**

Giotto object

**Examples**

```
mergeClusters(gobject)
```

---

<code>node_clusters</code>	<i>node_clusters</i>
----------------------------	----------------------

---

**Description**

Merge selected clusters based on pairwise correlation scores and size of cluster.

**Usage**

```
node_clusters(hclus_obj, verbose = TRUE)
```

**Arguments**

<code>hclus_obj</code>	hclus object
<code>verbose</code>	be verbose

**Value**

list of splitted dendrogram nodes from high to low node height

**Examples**

```
node_clusters(hclus_obj)
```

---

normalizeGiotto	<i>normalizeGiotto</i>
-----------------	------------------------

---

## Description

fast normalize and/or scale expression values of Giotto object

## Usage

```
normalizeGiotto(
  gobject,
  norm_methods = c("standard", "osmFISH"),
  library_size_norm = TRUE,
  scalefactor = 6000,
  log_norm = TRUE,
  log_offset = 1,
  logbase = 2,
  scale_genes = T,
  scale_cells = T,
  scale_order = c("first_genes", "first_cells"),
  verbose = F
)
```

## Arguments

gobject	giotto object
norm_methods	normalization method to use
library_size_norm	normalize cells by library size
scalefactor	scale factor to use after library size normalization
log_norm	transform values to log-scale
log_offset	offset value to add to expression matrix, default = 1
logbase	log base to use to log normalize expression values
scale_genes	z-score genes over all cells
scale_cells	z-score cells over all genes
scale_order	order to scale genes and cells
verbose	be verbose

## Details

Currently there are two 'methods' to normalize your raw counts data.

A. The standard method follows the standard protocol which can be adjusted using the provided parameters and follows the following order:

- 1. Data normalization for total library size and scaling by a custom scale-factor.
- 2. Log transformation of data.
- 3. Z-scoring of data by genes and/or cells.

B. The normalization method as provided by the osmFISH paper is also implemented:

- 1. First normalize genes, for each gene divide the counts by the total gene count and multiply by the total number of genes.
- 2. Next normalize cells, for each cell divide the normalized gene counts by the total counts per cell and multiply by the total number of cells.

This data will be saved in the Giotto slot for custom expression.

**Value**

giotto object

**Examples**

```
normalizeGiotto(gobject)
```

---

PAGEEnrich	<i>runPAGEEnrich</i>
------------	----------------------

---

**Description**

Function to calculate gene signature enrichment scores per spatial position using PAGE.

**Usage**

```
PAGEEnrich(...)
```

**Arguments**

```
... Arguments passed on to runPAGEEnrich
gobject Giotto object
sign_matrix Matrix of signature genes for each cell type / process
expression_values expression values to use
reverse_log_scale reverse expression values from log scale
logbase log base to use if reverse_log_scale = TRUE
output_enrichment how to return enrichment output
p_value calculate p-values (boolean, default = FALSE)
n_times number of permutations to calculate for p_value
name to give to spatial enrichment results, default = PAGE
return_gobject return giotto object
```

**See Also**

[runPAGEEnrich](#)

---

pDataDT	<i>pDataDT</i>
---------	----------------

---

**Description**

show cell metadata

**Usage**

pDataDT(gobject)

**Arguments**

gobject                  giotto object

**Value**

data.table with cell metadata

**Examples**

pDataDT(gobject)

---

plotCCcomDotplot	<i>plotCCcomDotplot</i>
------------------	-------------------------

---

**Description**

Plots dotplot for ligand-receptor communication scores in cell-cell interactions

**Usage**

```
plotCCcomDotplot(  
  gobject,  
  comScores,  
  selected_LR = NULL,  
  selected_cell_LR = NULL,  
  show_LR_names = TRUE,  
  show_cell_LR_names = TRUE,  
  cluster_on = c("PI", "LR_expr", "log2fc"),  
  cor_method = c("pearson", "kendall", "spearman"),  
  aggl_method = c("ward.D", "ward.D2", "single", "complete", "average", "mcquitty",  
    "median", "centroid"),  
  show_plot = NA,  
  return_plot = NA,  
  save_plot = NA,  
  save_param = list(),  
  default_save_name = "plotCCcomDotplot"  
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>comScores</code>	communication scores from <a href="#">exprCellCellcom</a> or <a href="#">spatCellCellcom</a>
<code>selected_LR</code>	selected ligand-receptor combinations
<code>selected_cell_LR</code>	selected cell-cell combinations for ligand-receptor combinations
<code>show_LR_names</code>	show ligand-receptor names
<code>show_cell_LR_names</code>	show cell-cell names
<code>cluster_on</code>	values to use for clustering of cell-cell and ligand-receptor pairs
<code>cor_method</code>	correlation method used for clustering
<code>aggl_method</code>	agglomeration method used by hclust
<code>show_plot</code>	show plots
<code>return_plot</code>	return plotting object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

**Value**

ggplot

**Examples**

```
plotCCcomDotplot(CPGscores)
```

---

<code>plotCCcomHeatmap</code>	<i>plotCCcomHeatmap</i>
-------------------------------	-------------------------

---

**Description**

Plots heatmap for ligand-receptor communication scores in cell-cell interactions

**Usage**

```
plotCCcomHeatmap(
  gobject,
  comScores,
  selected_LR = NULL,
  selected_cell_LR = NULL,
  show_LR_names = TRUE,
  show_cell_LR_names = TRUE,
  show = c("PI", "LR_expr", "log2fc"),
  cor_method = c("pearson", "kendall", "spearman"),
  aggl_method = c("ward.D", "ward.D2", "single", "complete", "average", "mcquitty",
    "median", "centroid"),
```

```

    show_plot = NA,
    return_plot = NA,
    save_plot = NA,
    save_param = list(),
    default_save_name = "plotCCcomHeatmap"
  )

```

### Arguments

<code>gobject</code>	giotto object
<code>comScores</code>	communication scores from <a href="#">exprCellCellcom</a> or <a href="#">spatCellCellcom</a>
<code>selected_LR</code>	selected ligand-receptor combinations
<code>selected_cell_LR</code>	selected cell-cell combinations for ligand-receptor combinations
<code>show_LR_names</code>	show ligand-receptor names
<code>show_cell_LR_names</code>	show cell-cell names
<code>show</code>	values to show on heatmap
<code>cor_method</code>	correlation method used for clustering
<code>aggl_method</code>	agglomeration method used by hclust
<code>show_plot</code>	show plots
<code>return_plot</code>	return plotting object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change save_name in save_param

### Value

ggplot

### Examples

```
plotCCcomHeatmap(CPGscores)
```

---

plotCellProximityGenes

*plotCellProximityGenes*

---

### Description

Create visualization for cell proximity gene scores

**Usage**

```
plotCellProximityGenes(
  gobject,
  cpqObject,
  method = c("volcano", "cell_barplot", "cell-cell", "cell_sankey", "heatmap",
    "dotplot"),
  min_cells = 4,
  min_cells_expr = 1,
  min_int_cells = 4,
  min_int_cells_expr = 1,
  min_fdr = 0.1,
  min_spat_diff = 0.2,
  min_log2_fc = 0.2,
  min_zscore = 2,
  zscores_column = c("cell_type", "genes"),
  direction = c("both", "up", "down"),
  cell_color_code = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotCellProximityGenes"
)
```

**Arguments**

gobject	giotto object
cpqObject	cell proximity gene score object
method	plotting method to use
min_cells	minimum number of source cell type
min_cells_expr	minimum expression level for source cell type
min_int_cells	minimum number of interacting neighbor cell type
min_int_cells_expr	minimum expression level for interacting neighbor cell type
min_fdr	minimum adjusted p-value
min_spat_diff	minimum absolute spatial expression difference
min_log2_fc	minimum log2 fold-change
min_zscore	minimum z-score change
zscores_column	calculate z-scores over cell types or genes
direction	differential expression directions to keep
cell_color_code	vector of colors with cell types as names
show_plot	show plots
return_plot	return plotting object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from <a href="#">all_plots_save_function</a>
default_save_name	default save name for saving, don't change, change save_name in save_param



**Value**

plot

**Examples**

```
plotCellProximityGenes(CPGscores)
```

---

plotCombineCCcom

*plotCombineCCcom*


---

**Description**

Create visualization for combined (pairwise) cell proximity gene scores

**Usage**

```
plotCombineCCcom(
  gobject,
  combCCcom,
  selected_LR = NULL,
  selected_cell_LR = NULL,
  detail_plot = T,
  simple_plot = F,
  simple_plot_facet = c("interaction", "genes"),
  facet_scales = "fixed",
  facet_ncol = length(selected_LR),
  facet_nrow = length(selected_cell_LR),
  colors = c("#9932CC", "#FF8C00"),
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotCombineCCcom"
)
```

**Arguments**

gobject	giotto object
combCCcom	combined communication scores, output from combCCcom()
selected_LR	selected ligand-receptor pair
selected_cell_LR	selected cell-cell interaction pair for ligand-receptor pair
detail_plot	show detailed info in both interacting cell types
simple_plot	show a simplified plot
simple_plot_facet	facet on interactions or genes with simple plot
facet_scales	ggplot facet scales paramter
facet_ncol	ggplot facet ncol parameter

facet_nrow	ggplot facet nrow parameter
colors	vector with two colors to use
show_plot	show plots
return_plot	return plotting object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from <a href="#">all_plots_save_function</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

**Value**

ggplot

**Examples**

```
plotCombineCCcom(CPGscores)
```

---

```
plotCombineCellCellCommunication
      plotCombineCellCellCommunication
```

---

**Description**

Create visualization for combined (pairwise) cell proximity gene scores

**Usage**

```
plotCombineCellCellCommunication(
  gobject,
  combCCcom,
  selected_LR = NULL,
  selected_cell_LR = NULL,
  detail_plot = T,
  simple_plot = F,
  simple_plot_facet = c("interaction", "genes"),
  facet_scales = "fixed",
  facet_ncol = length(selected_LR),
  facet_nrow = length(selected_cell_LR),
  colors = c("#9932CC", "#FF8C00"),
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotCombineCellCellCommunication"
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>combCCcom</code>	combined communication scores, output from <code>combCCcom()</code>
<code>selected_LR</code>	selected ligand-receptor pair
<code>selected_cell_LR</code>	selected cell-cell interaction pair for ligand-receptor pair
<code>detail_plot</code>	show detailed info in both interacting cell types
<code>simple_plot</code>	show a simplified plot
<code>simple_plot_facet</code>	facet on interactions or genes with simple plot
<code>facet_scales</code>	ggplot facet scales parameter
<code>facet_ncol</code>	ggplot facet ncol parameter
<code>facet_nrow</code>	ggplot facet nrow parameter
<code>colors</code>	vector with two colors to use
<code>show_plot</code>	show plots
<code>return_plot</code>	return plotting object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

**Value**

ggplot

**Examples**

```
plotCombineCellCellCommunication(CPGscores)
```

---

```
plotCombineCellProximityGenes
```

```
plotCombineCellProximityGenes
```

---

**Description**

Create visualization for combined (pairwise) cell proximity gene scores

**Usage**

```
plotCombineCellProximityGenes(
  gobject,
  combCpgObject,
  selected_interactions = NULL,
  selected_gene_to_gene = NULL,
  detail_plot = T,
  simple_plot = F,
```

```

simple_plot_facet = c("interaction", "genes"),
facet_scales = "fixed",
facet_ncol = length(selected_gene_to_gene),
facet_nrow = length(selected_interactions),
colors = c("#9932CC", "#FF8C00"),
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "plotCombineCPG"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>combCpgObject</code>	CPGscores, output from <code>combineCellProximityGenes()</code>
<code>selected_interactions</code>	interactions to show
<code>selected_gene_to_gene</code>	pairwise gene combinations to show
<code>detail_plot</code>	show detailed info in both interacting cell types
<code>simple_plot</code>	show a simplified plot
<code>simple_plot_facet</code>	facet on interactions or genes with simple plot
<code>facet_scales</code>	ggplot facet scales paramter
<code>facet_ncol</code>	ggplot facet ncol parameter
<code>facet_nrow</code>	ggplot facet nrow parameter
<code>colors</code>	vector with two colors to use
<code>show_plot</code>	show plots
<code>return_plot</code>	return plotting object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

### Value

ggplot

### Examples

```
plotCombineCellProximityGenes(CPGscores)
```

---

plotCombineCPG	<i>plotCombineCPG</i>
----------------	-----------------------

---

## Description

Create visualization for combined (pairwise) cell proximity gene scores

## Usage

```
plotCombineCPG(
  gobject,
  combCpgObject,
  selected_interactions = NULL,
  selected_gene_to_gene = NULL,
  detail_plot = T,
  simple_plot = F,
  simple_plot_facet = c("interaction", "genes"),
  facet_scales = "fixed",
  facet_ncol = length(selected_gene_to_gene),
  facet_nrow = length(selected_interactions),
  colors = c("#9932CC", "#FF8C00"),
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotCombineCPG"
)
```

## Arguments

<code>gobject</code>	giotto object
<code>combCpgObject</code>	CPGscores, output from <code>combineCellProximityGenes()</code>
<code>selected_interactions</code>	interactions to show
<code>selected_gene_to_gene</code>	pairwise gene combinations to show
<code>detail_plot</code>	show detailed info in both interacting cell types
<code>simple_plot</code>	show a simplified plot
<code>simple_plot_facet</code>	facet on interactions or genes with simple plot
<code>facet_scales</code>	ggplot facet scales paramter
<code>facet_ncol</code>	ggplot facet ncol parameter
<code>facet_nrow</code>	ggplot facet nrow parameter
<code>colors</code>	vector with two colors to use
<code>show_plot</code>	show plots
<code>return_plot</code>	return plotting object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

Value

ggplot

Examples

```
plotCombineCPG(CPGscores)
```

---

plotCPG	<i>plotCPG</i>
---------	----------------

---

Description

Create visualization for cell proximity gene scores

Usage

```
plotCPG(  
  gobject,  
  cpgObject,  
  method = c("volcano", "cell_barplot", "cell-cell", "cell_sankey", "heatmap",  
             "dotplot"),  
  min_cells = 5,  
  min_cells_expr = 1,  
  min_int_cells = 3,  
  min_int_cells_expr = 1,  
  min_fdr = 0.05,  
  min_spat_diff = 0.2,  
  min_log2_fc = 0.2,  
  min_zscore = 2,  
  zscores_column = c("cell_type", "genes"),  
  direction = c("both", "up", "down"),  
  cell_color_code = NULL,  
  show_plot = NA,  
  return_plot = NA,  
  save_plot = NA,  
  save_param = list(),  
  default_save_name = "plotCPG"  
)
```

Arguments

- |                    |   |
|--------------------|---|
| gobject            | giotto object   |
| cpgObject          | cell proximity gene score object                            |
| method             | plotting method to use                                      |
| min_cells          | minimum number of source cell type                          |
| min_cells_expr     | minimum expression level for source cell type               |
| min_int_cells      | minimum number of interacting neighbor cell type            |
| min_int_cells_expr | minimum expression level for interacting neighbor cell type |

min_fdr	minimum adjusted p-value
min_spat_diff	minimum absolute spatial expression difference
min_log2_fc	minimum log2 fold-change
min_zscore	minimum z-score change
zscores_column	calculate z-scores over cell types or genes
direction	differential expression directions to keep
cell_color_code	vector of colors with cell types as names
show_plot	show plots
return_plot	return plotting object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from <a href="#">all_plots_save_function</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

**Value**

plot

**Examples**

```
plotCPG(CPGscores)
```

---

plotGiottoImage	<i>plotGiottoImage</i>
-----------------	------------------------

---

**Description**

get plot a giotto image from a giotto object

**Usage**

```
plotGiottoImage(gobject, image_name)
```

**Arguments**

gobject	giotto object
image_name	name of giotto image <a href="#">showGiottoImageNames</a>

**Value**

plot

**Examples**

```
plotGiottoImage(gobject)
```

---

plotHeatmap

*plotHeatmap*


---

## Description

Creates heatmap for genes and clusters.

## Usage

```
plotHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cluster_color_code = NULL,
  cluster_cor_method = "pearson",
  cluster_hclust_method = "ward.D",
  gene_order = c("correlation", "custom"),
  gene_custom_order = NULL,
  gene_cor_method = "pearson",
  gene_hclust_method = "complete",
  show_values = c("rescaled", "z-scaled", "original"),
  size_vertical_lines = 1.1,
  gradient_colors = c("blue", "yellow", "red"),
  gene_label_selection = NULL,
  axis_text_y_size = NULL,
  legend_nrows = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotHeatmap"
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>genes</code>	genes to use
<code>cluster_column</code>	name of column to use for clusters
<code>cluster_order</code>	method to determine cluster order
<code>cluster_custom_order</code>	custom order for clusters
<code>cluster_color_code</code>	color code for clusters
<code>cluster_cor_method</code>	method for cluster correlation



cluster_hclust_method	method for hierarchical clustering of clusters
gene_order	method to determine gene order
gene_custom_order	custom order for genes
gene_cor_method	method for gene correlation
gene_hclust_method	method for hierarchical clustering of genes
show_values	which values to show on heatmap
size_vertical_lines	sizes for vertical lines
gradient_colors	colors for heatmap gradient
gene_label_selection	subset of genes to show on y-axis
axis_text_y_size	size for y-axis text
legend_nrows	number of rows for the cluster legend
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name

## Details

If you want to display many genes there are 2 ways to proceed:

- 1. set axis\_text\_y\_size to a really small value and show all genes
- 2. provide a subset of genes to display to gene\_label\_selection

## Value

ggplot

## Examples

```
## Not run:
plotHeatmap(gobject)

## End(Not run)
```

---

plotICG

*plotICG*


---

## Description

Create barplot to visualize interaction changed genes

## Usage

```
plotICG(
  gobject,
  cpgObject,
  source_type,
  source_markers,
  ICG_genes,
  cell_color_code = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotICG"
)
```

## Arguments

<code>gobject</code>	giotto object
<code>cpgObject</code>	cell proximity gene score object
<code>source_type</code>	cell type of the source cell
<code>source_markers</code>	markers for the source cell type
<code>ICG_genes</code>	named character vector of ICG genes
<code>cell_color_code</code>	cell color code for the interacting cell types
<code>show_plot</code>	show plots
<code>return_plot</code>	return plotting object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

## Value

plot

## Examples

```
plotICG(CPGscores)
```

---

```
plotInteractionChangedGenes
      plotInteractionChangedGenes
```

---

## Description

Create barplot to visualize interaction changed genes

## Usage

```
plotInteractionChangedGenes(
  gobject,
  cpgObject,
  source_type,
  source_markers,
  ICG_genes,
  cell_color_code = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotInteractionChangedGenes"
)
```

## Arguments

<code>gobject</code>	giotto object
<code>cpgObject</code>	cell proximity gene score object
<code>source_type</code>	cell type of the source cell
<code>source_markers</code>	markers for the source cell type
<code>ICG_genes</code>	named character vector of ICG genes
<code>cell_color_code</code>	cell color code for the interacting cell types
<code>show_plot</code>	show plots
<code>return_plot</code>	return plotting object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

## Value

plot

## Examples

```
plotInteractionChangedGenes(CPGscores)
```

---

plotMetaDataCellsHeatmap

*plotMetaDataCellsHeatmap*


---

## Description

Creates heatmap for numeric cell metadata within aggregated clusters.

## Usage

```
plotMetaDataCellsHeatmap(
  gobject,
  metadata_cols = NULL,
  spat_enr_names = NULL,
  value_cols = NULL,
  first_meta_col = NULL,
  second_meta_col = NULL,
  show_values = c("zscores", "original", "zscores_rescaled"),
  custom_cluster_order = NULL,
  clus_cor_method = "pearson",
  clus_cluster_method = "complete",
  custom_values_order = NULL,
  values_cor_method = "pearson",
  values_cluster_method = "complete",
  midpoint = 0,
  x_text_size = 8,
  x_text_angle = 45,
  y_text_size = 8,
  strip_text_size = 8,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotMetaDataCellsHeatmap"
)
```

## Arguments

gobject	giotto object
metadata_cols	annotation columns found in pDataDT(gobject)
spat_enr_names	spatial enrichment results to include
value_cols	value columns to use
first_meta_col	if more than 1 metadata column, select the x-axis factor
second_meta_col	if more than 1 metadata column, select the facetting factor
show_values	which values to show on heatmap
custom_cluster_order	custom cluster order (default = NULL)

clus_cor_method	correlation method for clusters
clus_cluster_method	hierarchical cluster method for the clusters
custom_values_order	custom values order (default = NULL)
values_cor_method	correlation method for values
values_cluster_method	hierarchical cluster method for the values
midpoint	midpoint of show_values
x_text_size	size of x-axis text
x_text_angle	angle of x-axis text
y_text_size	size of y-axis text
strip_text_size	size of strip text
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

## Details

Creates heatmap for the average values of selected value columns in the different annotation groups.

## Value

ggplot or data.table

## See Also

[plotMetaDataHeatmap](#) for gene expression instead of numeric cell annotation data.

## Examples

```
plotMetaDataCellsHeatmap(gobject)
```

---

plotMetaDataHeatmap	<i>plotMetaDataHeatmap</i>
---------------------	----------------------------

---

## Description

Creates heatmap for genes within aggregated clusters.

## Usage

```
plotMetaDataHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metadata_cols = NULL,
  selected_genes = NULL,
  first_meta_col = NULL,
  second_meta_col = NULL,
  show_values = c("zscores", "original", "zscores_rescaled"),
  custom_cluster_order = NULL,
  clus_cor_method = "pearson",
  clus_cluster_method = "complete",
  custom_gene_order = NULL,
  gene_cor_method = "pearson",
  gene_cluster_method = "complete",
  gradient_color = c("blue", "white", "red"),
  gradient_midpoint = 0,
  gradient_limits = NULL,
  x_text_size = 10,
  x_text_angle = 45,
  y_text_size = 10,
  strip_text_size = 8,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotMetaDataHeatmap"
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>metadata_cols</code>	annotation columns found in <code>pDataDT(gobject)</code>
<code>selected_genes</code>	subset of genes to use
<code>first_meta_col</code>	if more than 1 metadata column, select the x-axis factor
<code>second_meta_col</code>	if more than 1 metadata column, select the facetting factor
<code>show_values</code>	which values to show on heatmap
<code>custom_cluster_order</code>	custom cluster order (default = NULL)

clus_cor_method	correlation method for clusters
clus_cluster_method	hierarchical cluster method for the clusters
custom_gene_order	custom gene order (default = NULL)
gene_cor_method	correlation method for genes
gene_cluster_method	hierarchical cluster method for the genes
gradient_color	vector with 3 colors for numeric data
gradient_midpoint	midpoint for color gradient
gradient_limits	vector with lower and upper limits
x_text_size	size of x-axis text
x_text_angle	angle of x-axis text
y_text_size	size of y-axis text
strip_text_size	size of strip text
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name

## Details

Creates heatmap for the average expression of selected genes in the different annotation/cluster groups. Calculation of cluster or gene order is done on the provided expression values, but visualization is by default on the z-scores. Other options are the original values or z-scores rescaled per gene (-1 to 1).

## Value

ggplot or data.table

## See Also

[plotMetaDataCellsHeatmap](#) for numeric cell annotation instead of gene expression.

## Examples

```
plotMetaDataHeatmap(gobject)
```

plotPCA

*plotPCA***Description**

Short wrapper for PCA visualization

**Usage**

```
plotPCA(gobject, dim_reduction_name = "pca", default_save_name = "PCA", ...)
```

**Arguments**

gobject	giotto object
dim_reduction_name	name of PCA
default_save_name	default save name of PCA plot
...	Arguments passed on to <a href="#">dimPlot2D</a>
group_by	create multiple plots based on cell annotation column
group_by_subset	subset the group_by factor column
dim1_to_use	dimension to use on x-axis
dim2_to_use	dimension to use on y-axis
spat_enr_names	names of spatial enrichment results to include
show_NN_network	show underlying NN network
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use, if show_NN_network = TRUE
cell_color	color for cells (see details)
color_as_factor	convert color column to factor
cell_color_code	named vector with colors
cell_color_gradient	vector with 3 colors for numeric data
gradient_midpoint	midpoint for color gradient
gradient_limits	vector with lower and upper limits
select_cell_groups	select subset of cells/clusters based on cell_color parameter
select_cells	select subset of cells based on cell IDs
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	size of not selected cells
show_cluster_center	plot center of selected clusters
show_center_label	plot label of selected clusters
center_point_size	size of center points
center_point_border_col	border color of center points
center_point_border_stroke	border stroke size of center points
label_size	size of labels
label_fontface	font of labels



edge\_alpha column to use for alpha of the edges  
 point\_shape point with border or not (border or no\_border)  
 point\_size size of point (cell)  
 point\_alpha transparency of point  
 point\_border\_col color of border around points  
 point\_border\_stroke stroke size of border around points  
 title title for plot, defaults to cell\_color parameter  
 show\_legend show legend  
 legend\_text size of legend text  
 legend\_symbol\_size size of legend symbols  
 background\_color color of plot background  
 axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plot  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)

## Details

Description of parameters, see [dimPlot2D](#). For 3D plots see [plotPCA\\_3D](#)

## Value

ggplot

## See Also

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

## Examples

```
plotPCA(gobject)
```

---

plotPCA\_2D

*plotPCA\_2D*

---

## Description

Short wrapper for PCA visualization

**Usage**

```
plotPCA_2D(
  gobject,
  dim_reduction_name = "pca",
  default_save_name = "PCA_2D",
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>dim_reduction_name</code>	name of PCA
<code>default_save_name</code>	default save name of PCA plot
<code>...</code>	Arguments passed on to <a href="#">dimPlot2D</a>
<code>group_by</code>	create multiple plots based on cell annotation column
<code>group_by_subset</code>	subset the <code>group_by</code> factor column
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>show_NN_network</code>	show underlying NN network
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use, if <code>show_NN_network = TRUE</code>
<code>cell_color</code>	color for cells (see details)
<code>color_as_factor</code>	convert color column to factor
<code>cell_color_code</code>	named vector with colors
<code>cell_color_gradient</code>	vector with 3 colors for numeric data
<code>gradient_midpoint</code>	midpoint for color gradient
<code>gradient_limits</code>	vector with lower and upper limits
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs
<code>show_other_cells</code>	display not selected cells
<code>other_cell_color</code>	color of not selected cells
<code>other_point_size</code>	size of not selected cells
<code>show_cluster_center</code>	plot center of selected clusters
<code>show_center_label</code>	plot label of selected clusters
<code>center_point_size</code>	size of center points
<code>center_point_border_col</code>	border color of center points
<code>center_point_border_stroke</code>	border stroke size of center points
<code>label_size</code>	size of labels
<code>label_fontface</code>	font of labels
<code>edge_alpha</code>	column to use for alpha of the edges
<code>point_shape</code>	point with border or not (border or no_border)
<code>point_size</code>	size of point (cell)
<code>point_alpha</code>	transparency of point

point\_border\_col color of border around points  
 point\_border\_stroke stroke size of border around points  
 title title for plot, defaults to cell\_color parameter  
 show\_legend show legend  
 legend\_text size of legend text  
 legend\_symbol\_size size of legend symbols  
 background\_color color of plot background  
 axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plot  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)

### Details

Description of parameters, see [dimPlot2D](#). For 3D plots see [plotPCA\\_3D](#)

### Value

ggplot

### See Also

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

### Examples

```
plotPCA_2D(gobject)
```

---

plotPCA\_3D

*plotPCA\_3D*

---

### Description

Visualize cells according to 3D PCA dimension reduction

### Usage

```
plotPCA_3D(
  gobject,
  dim_reduction_name = "pca",
  default_save_name = "PCA_3D",
  ...
)
```

**Arguments**

gobject            giotto object  
 dim\_reduction\_name            name of PCA  
 default\_save\_name            default save name of PCA plot  
 ...            Arguments passed on to [dimPlot3D](#)  
           dim1\_to\_use   dimension to use on x-axis  
           dim2\_to\_use   dimension to use on y-axis  
           dim3\_to\_use   dimension to use on z-axis  
           spat\_enr\_names   names of spatial enrichment results to include  
           show\_NN\_network   show underlying NN network  
           nn\_network\_to\_use   type of NN network to use (kNN vs sNN)  
           network\_name   name of NN network to use, if show\_NN\_network = TRUE  
           cell\_color   color for cells (see details)  
           color\_as\_factor   convert color column to factor  
           cell\_color\_code   named vector with colors  
           select\_cell\_groups   select subset of cells/clusters based on cell\_color parameter  
           select\_cells   select subset of cells based on cell IDs  
           show\_other\_cells   display not selected cells  
           other\_cell\_color   color of not selected cells  
           other\_point\_size   size of not selected cells  
           show\_cluster\_center   plot center of selected clusters  
           show\_center\_label   plot label of selected clusters  
           center\_point\_size   size of center points  
           label\_size   size of labels  
           edge\_alpha   column to use for alpha of the edges  
           point\_size   size of point (cell)  
           show\_plot   show plot  
           return\_plot   return ggplot object  
           save\_plot   directly save the plot [boolean]  
           save\_param   list of saving parameters, see [showSaveParameters](#)

**Details**

Description of parameters.

**Value**

plotly

**See Also**

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

**Examples**

```
plotPCA_3D(gobject)
```

---

plotRankSpatvsExpr	<i>plotRankSpatvsExpr</i>
--------------------	---------------------------

---

## Description

Plots dotplot to compare ligand-receptor rankings from spatial and expression information

## Usage

```
plotRankSpatvsExpr(
  gobject,
  combCC,
  expr_rnk_column = "LR_expr_rnk",
  spat_rnk_column = "LR_spat_rnk",
  midpoint = 10,
  size_range = c(0.01, 1.5),
  xlims = NULL,
  ylims = NULL,
  selected_ranks = c(1, 10, 20),
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotRankSpatvsExpr"
)
```

## Arguments

gobject	giotto object
combCC	combined communication scores from <a href="#">combCCcom</a>
expr_rnk_column	column with expression rank information to use
spat_rnk_column	column with spatial rank information to use
midpoint	midpoint of colors
size_range	size ranges of dotplot
xlims	x-limits, numerical vector of 2
ylims	y-limits, numerical vector of 2
selected_ranks	numerical vector, will be used to print out the percentage of top spatial ranks are recovered
show_plot	show plots
return_plot	return plotting object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from <a href="#">all_plots_save_function</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

Value

ggplot

Examples

plotRankSpatvsExpr(CPGscores)

---

plotRecovery	<i>plotRecovery</i>
--------------	---------------------

---

Description

Plots recovery plot to compare ligand-receptor rankings from spatial and expression information

Usage

```
plotRecovery(  
  gobject,  
  combCC,  
  expr_rnk_column = "exprPI_rnk",  
  spat_rnk_column = "spatPI_rnk",  
  ground_truth = c("spatial", "expression"),  
  show_plot = NA,  
  return_plot = NA,  
  save_plot = NA,  
  save_param = list(),  
  default_save_name = "plotRecovery"  
)
```

Arguments

gobject	giotto object
combCC	combined communication scores from <a href="#">combCCcom</a>
expr_rnk_column	column with expression rank information to use
spat_rnk_column	column with spatial rank information to use
ground_truth	what to consider as ground truth (default: spatial)
show_plot	show plots
return_plot	return plotting object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from <a href="#">all_plots_save_function</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

Value

ggplot

**Examples**

```
plotRecovery(CPGscores)
```

---

plotRecovery_sub	<i>plotRecovery_sub</i>
------------------	-------------------------

---

**Description**

Plots recovery plot to compare ligand-receptor rankings from spatial and expression information

**Usage**

```
plotRecovery_sub(combCC, first_col = "LR_expr_rnk", second_col = "LR_spat_rnk")
```

**Arguments**

combCC	combined communication scores from <a href="#">combCCcom</a>
first_col	first column to use
second_col	second column to use

**Examples**

```
plotRecovery_sub(CPGscores)
```

---

plotStatDelaunayNetwork	<i>plotStatDelaunayNetwork</i>
-------------------------	--------------------------------

---

**Description**

Plots network statistics for a Delaunay network..

**Usage**

```
plotStatDelaunayNetwork(
  gobject,
  method = c("deldir", "delaunayn_geometry", "RTriangle"),
  dimensions = "all",
  maximum_distance = "auto",
  minimum_k = 0,
  options = "Pp",
  Y = TRUE,
  j = TRUE,
  S = 0,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotStatDelaunayNetwork",
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>method</code>	package to use to create a Delaunay network
<code>dimensions</code>	which spatial dimensions to use (maximum 2 dimensions)
<code>maximum_distance</code>	distance cutoff for Delaunay neighbors to consider
<code>minimum_k</code>	minimum neighbours if <code>maximum_distance</code> != NULL
<code>options</code>	(geometry) String containing extra control options for the underlying Qhull command; see the Qhull documentation ( <a href="http://doc/qhull/html/qdelaun.html">../doc/qhull/html/qdelaun.html</a> ) for the available options. (default = 'Pp', do not report precision problems)
<code>Y</code>	(RTriangle) If TRUE prohibits the insertion of Steiner points on the mesh boundary.
<code>j</code>	(RTriangle) If TRUE jettisons vertices that are not part of the final triangulation from the output.
<code>S</code>	(RTriangle) Specifies the maximum number of added Steiner points.
<code>show_plot</code>	show plots
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters, see <a href="#">showSaveParameters</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>
<code>...</code>	Other parameters

**Value**

giotto object with updated spatial network slot

**Examples**

```
plotStatDelaunayNetwork(gobject)
```

---

plotTSNE

*plotTSNE*


---

**Description**

Short wrapper for tSNE visualization

**Usage**

```
plotTSNE(gobject, dim_reduction_name = "tsne", default_save_name = "tSNE", ...)
```



**Arguments**

**gobject** giotto object  
**dim\_reduction\_name** name of TSNE  
**default\_save\_name** default save name of TSNE plot  
**...** Arguments passed on to [dimPlot2D](#)  
**group\_by** create multiple plots based on cell annotation column  
**group\_by\_subset** subset the group\_by factor column  
**dim1\_to\_use** dimension to use on x-axis  
**dim2\_to\_use** dimension to use on y-axis  
**spat\_enr\_names** names of spatial enrichment results to include  
**show\_NN\_network** show underlying NN network  
**nn\_network\_to\_use** type of NN network to use (kNN vs sNN)  
**network\_name** name of NN network to use, if show\_NN\_network = TRUE  
**cell\_color** color for cells (see details)  
**color\_as\_factor** convert color column to factor  
**cell\_color\_code** named vector with colors  
**cell\_color\_gradient** vector with 3 colors for numeric data  
**gradient\_midpoint** midpoint for color gradient  
**gradient\_limits** vector with lower and upper limits  
**select\_cell\_groups** select subset of cells/clusters based on cell\_color parameter  
**select\_cells** select subset of cells based on cell IDs  
**show\_other\_cells** display not selected cells  
**other\_cell\_color** color of not selected cells  
**other\_point\_size** size of not selected cells  
**show\_cluster\_center** plot center of selected clusters  
**show\_center\_label** plot label of selected clusters  
**center\_point\_size** size of center points  
**center\_point\_border\_col** border color of center points  
**center\_point\_border\_stroke** border stroke size of center points  
**label\_size** size of labels  
**label\_fontface** font of labels  
**edge\_alpha** column to use for alpha of the edges  
**point\_shape** point with border or not (border or no\_border)  
**point\_size** size of point (cell)  
**point\_alpha** transparency of point  
**point\_border\_col** color of border around points  
**point\_border\_stroke** stroke size of border around points  
**title** title for plot, defaults to cell\_color parameter  
**show\_legend** show legend  
**legend\_text** size of legend text  
**legend\_symbol\_size** size of legend symbols  
**background\_color** color of plot background

axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plot  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)

## Details

Description of parameters, see [dimPlot2D](#). For 3D plots see [plotTSNE\\_3D](#)

## Value

ggplot

## See Also

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

## Examples

```
plotTSNE(gobject)
```

---

plotTSNE\_2D

*plotTSNE\_2D*


---

## Description

Short wrapper for tSNE visualization

## Usage

```

plotTSNE_2D(
  gobject,
  dim_reduction_name = "tsne",
  default_save_name = "tSNE_2D",
  ...
)
```

**Arguments**

**gobject** giotto object  
**dim\_reduction\_name** name of TSNE  
**default\_save\_name** default save name of TSNE plot  
**...** Arguments passed on to [dimPlot2D](#)  
**group\_by** create multiple plots based on cell annotation column  
**group\_by\_subset** subset the group\_by factor column  
**dim1\_to\_use** dimension to use on x-axis  
**dim2\_to\_use** dimension to use on y-axis  
**spat\_enr\_names** names of spatial enrichment results to include  
**show\_NN\_network** show underlying NN network  
**nn\_network\_to\_use** type of NN network to use (kNN vs sNN)  
**network\_name** name of NN network to use, if show\_NN\_network = TRUE  
**cell\_color** color for cells (see details)  
**color\_as\_factor** convert color column to factor  
**cell\_color\_code** named vector with colors  
**cell\_color\_gradient** vector with 3 colors for numeric data  
**gradient\_midpoint** midpoint for color gradient  
**gradient\_limits** vector with lower and upper limits  
**select\_cell\_groups** select subset of cells/clusters based on cell\_color parameter  
**select\_cells** select subset of cells based on cell IDs  
**show\_other\_cells** display not selected cells  
**other\_cell\_color** color of not selected cells  
**other\_point\_size** size of not selected cells  
**show\_cluster\_center** plot center of selected clusters  
**show\_center\_label** plot label of selected clusters  
**center\_point\_size** size of center points  
**center\_point\_border\_col** border color of center points  
**center\_point\_border\_stroke** border stroke size of center points  
**label\_size** size of labels  
**label\_fontface** font of labels  
**edge\_alpha** column to use for alpha of the edges  
**point\_shape** point with border or not (border or no\_border)  
**point\_size** size of point (cell)  
**point\_alpha** transparency of point  
**point\_border\_col** color of border around points  
**point\_border\_stroke** stroke size of border around points  
**title** title for plot, defaults to cell\_color parameter  
**show\_legend** show legend  
**legend\_text** size of legend text  
**legend\_symbol\_size** size of legend symbols  
**background\_color** color of plot background

axis\_text size of axis text  
axis\_title size of axis title  
cow\_n\_col cowplot param: how many columns  
cow\_rel\_h cowplot param: relative height  
cow\_rel\_w cowplot param: relative width  
cow\_align cowplot param: how to align  
show\_plot show plot  
return\_plot return ggplot object  
save\_plot directly save the plot [boolean]  
save\_param list of saving parameters, see [showSaveParameters](#)

**Details**

Description of parameters, see [dimPlot2D](#). For 3D plots see [plotTSNE\\_3D](#)

**Value**

ggplot

**See Also**

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

**Examples**

```
plotTSNE_2D(gobject)
```

---

plotTSNE_3D	<i>plotTSNE_3D</i>
-------------	--------------------

---

**Description**

Visualize cells according to dimension reduction coordinates

**Usage**

```
plotTSNE_3D(  
  gobject,  
  dim_reduction_name = "tsne",  
  default_save_name = "TSNE_3D",  
  ...  
)
```

**Arguments**

**gobject**            giotto object  
**dim\_reduction\_name**            name of TSNE  
**default\_save\_name**            default save name of TSNE plot  
**...**            Arguments passed on to [dimPlot3D](#)  
**dim1\_to\_use**    dimension to use on x-axis  
**dim2\_to\_use**    dimension to use on y-axis  
**dim3\_to\_use**    dimension to use on z-axis  
**spat\_enr\_names**    names of spatial enrichment results to include  
**show\_NN\_network**    show underlying NN network  
**nn\_network\_to\_use**    type of NN network to use (kNN vs sNN)  
**network\_name**    name of NN network to use, if **show\_NN\_network** = TRUE  
**cell\_color**    color for cells (see details)  
**color\_as\_factor**    convert color column to factor  
**cell\_color\_code**    named vector with colors  
**select\_cell\_groups**    select subset of cells/clusters based on **cell\_color** parameter  
**select\_cells**    select subset of cells based on cell IDs  
**show\_other\_cells**    display not selected cells  
**other\_cell\_color**    color of not selected cells  
**other\_point\_size**    size of not selected cells  
**show\_cluster\_center**    plot center of selected clusters  
**show\_center\_label**    plot label of selected clusters  
**center\_point\_size**    size of center points  
**label\_size**    size of labels  
**edge\_alpha**    column to use for alpha of the edges  
**point\_size**    size of point (cell)  
**show\_plot**    show plot  
**return\_plot**    return ggplot object  
**save\_plot**    directly save the plot [boolean]  
**save\_param**    list of saving parameters, see [showSaveParameters](#)

**Details**

Description of parameters.

**Value**

plotly

**See Also**

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

**Examples**

```
plotTSNE_3D(gobject)
```

---

plotUMAP	<i>plotUMAP</i>
----------	-----------------

---

**Description**

Short wrapper for UMAP visualization

**Usage**

```
plotUMAP(gobject, dim_reduction_name = "umap", default_save_name = "UMAP", ...)
```

**Arguments**

gobject	giotto object
dim_reduction_name	name of UMAP
default_save_name	default save name of UMAP plot
...	Arguments passed on to <a href="#">dimPlot2D</a>
group_by	create multiple plots based on cell annotation column
group_by_subset	subset the group_by factor column
dim1_to_use	dimension to use on x-axis
dim2_to_use	dimension to use on y-axis
spat_enr_names	names of spatial enrichment results to include
show_NN_network	show underlying NN network
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use, if show_NN_network = TRUE
cell_color	color for cells (see details)
color_as_factor	convert color column to factor
cell_color_code	named vector with colors
cell_color_gradient	vector with 3 colors for numeric data
gradient_midpoint	midpoint for color gradient
gradient_limits	vector with lower and upper limits
select_cell_groups	select subset of cells/clusters based on cell_color parameter
select_cells	select subset of cells based on cell IDs
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	size of not selected cells
show_cluster_center	plot center of selected clusters
show_center_label	plot label of selected clusters
center_point_size	size of center points

`center_point_border_col` border color of center points  
`center_point_border_stroke` border stroke size of center points  
`label_size` size of labels  
`label_fontface` font of labels  
`edge_alpha` column to use for alpha of the edges  
`point_shape` point with border or not (border or no\_border)  
`point_size` size of point (cell)  
`point_alpha` transparency of point  
`point_border_col` color of border around points  
`point_border_stroke` stroke size of border around points  
`title` title for plot, defaults to `cell_color` parameter  
`show_legend` show legend  
`legend_text` size of legend text  
`legend_symbol_size` size of legend symbols  
`background_color` color of plot background  
`axis_text` size of axis text  
`axis_title` size of axis title  
`cow_n_col` cowplot param: how many columns  
`cow_rel_h` cowplot param: relative height  
`cow_rel_w` cowplot param: relative width  
`cow_align` cowplot param: how to align  
`show_plot` show plot  
`return_plot` return ggplot object  
`save_plot` directly save the plot [boolean]  
`save_param` list of saving parameters, see [showSaveParameters](#)

## Details

Description of parameters, see [dimPlot2D](#). For 3D plots see [plotUMAP\\_3D](#)

## Value

ggplot

## See Also

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\\_3D\(\)](#)

## Examples

```
plotUMAP(gobject)
```

---

plotUMAP\_2D

*plotUMAP\_2D*


---

## Description

Short wrapper for UMAP visualization

## Usage

```
plotUMAP_2D(
  gobject,
  dim_reduction_name = "umap",
  default_save_name = "UMAP_2D",
  ...
)
```

## Arguments

gobject	giotto object
dim_reduction_name	name of UMAP
default_save_name	default save name of UMAP plot
...	Arguments passed on to <a href="#">dimPlot2D</a>
group_by	create multiple plots based on cell annotation column
group_by_subset	subset the group_by factor column
dim1_to_use	dimension to use on x-axis
dim2_to_use	dimension to use on y-axis
spat_enr_names	names of spatial enrichment results to include
show_NN_network	show underlying NN network
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use, if show_NN_network = TRUE
cell_color	color for cells (see details)
color_as_factor	convert color column to factor
cell_color_code	named vector with colors
cell_color_gradient	vector with 3 colors for numeric data
gradient_midpoint	midpoint for color gradient
gradient_limits	vector with lower and upper limits
select_cell_groups	select subset of cells/clusters based on cell_color parameter
select_cells	select subset of cells based on cell IDs
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	size of not selected cells
show_cluster_center	plot center of selected clusters
show_center_label	plot label of selected clusters
center_point_size	size of center points



center\_point\_border\_col border color of center points  
 center\_point\_border\_stroke border stroke size of center points  
 label\_size size of labels  
 label\_fontface font of labels  
 edge\_alpha column to use for alpha of the edges  
 point\_shape point with border or not (border or no\_border)  
 point\_size size of point (cell)  
 point\_alpha transparency of point  
 point\_border\_col color of border around points  
 point\_border\_stroke stroke size of border around points  
 title title for plot, defaults to cell\_color parameter  
 show\_legend show legend  
 legend\_text size of legend text  
 legend\_symbol\_size size of legend symbols  
 background\_color color of plot background  
 axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plot  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)

## Details

Description of parameters, see [dimPlot2D](#). For 3D plots see [plotUMAP\\_3D](#)

## Value

ggplot

## See Also

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_3D\(\)](#), [plotUMAP\(\)](#)

## Examples

```
plotUMAP_2D(gobject)
```

plotUMAP\_3D

*plotUMAP\_3D***Description**

Visualize cells according to dimension reduction coordinates

**Usage**

```
plotUMAP_3D(
  gobject,
  dim_reduction_name = "umap",
  default_save_name = "UMAP_3D",
  ...
)
```

**Arguments**

gobject	giotto object
dim_reduction_name	name of UMAP
default_save_name	default save name of UMAP plot
...	Arguments passed on to <a href="#">dimPlot3D</a>
dim1_to_use	dimension to use on x-axis
dim2_to_use	dimension to use on y-axis
dim3_to_use	dimension to use on z-axis
spat_enr_names	names of spatial enrichment results to include
show_NN_network	show underlying NN network
nn_network_to_use	type of NN network to use (kNN vs sNN)
network_name	name of NN network to use, if show_NN_network = TRUE
cell_color	color for cells (see details)
color_as_factor	convert color column to factor
cell_color_code	named vector with colors
select_cell_groups	select subset of cells/clusters based on cell_color parameter
select_cells	select subset of cells based on cell IDs
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	size of not selected cells
show_cluster_center	plot center of selected clusters
show_center_label	plot label of selected clusters
center_point_size	size of center points
label_size	size of labels
edge_alpha	column to use for alpha of the edges
point_size	size of point (cell)
show_plot	show plot

return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)

## Details

Description of parameters.

## Value

plotly

## See Also

Other reduced dimension visualizations: [dimPlot2D\(\)](#), [dimPlot3D\(\)](#), [dimPlot\(\)](#), [plotPCA\\_2D\(\)](#), [plotPCA\\_3D\(\)](#), [plotPCA\(\)](#), [plotTSNE\\_2D\(\)](#), [plotTSNE\\_3D\(\)](#), [plotTSNE\(\)](#), [plotUMAP\\_2D\(\)](#), [plotUMAP\(\)](#)

## Examples

```
plotUMAP_3D(gobject)
```

---

print.giotto	<i>print method for giotto class</i>
--------------	--------------------------------------

---

## Description

print method for giotto class. Prints the chosen number of genes (rows) and cells (columns) from the raw count matrix. Also print the spatial locations for the chosen number of cells.

## Usage

```
print.giotto(object, ...)
```

## Arguments

nr_genes	number of genes (rows) to print
nr_cells	number of cells (columns) to print

---

rankEnrich	<i>rankEnrich</i>
------------	-------------------

---

**Description**

Function to calculate gene signature enrichment scores per spatial position using a rank based approach.

**Usage**

```
rankEnrich(...)
```

**Arguments**

```
...           Arguments passed on to runRankEnrich
gobject       Giotto object
sign_matrix   Matrix of signature genes for each cell type / process
expression_values expression values to use
reverse_log_scale reverse expression values from log scale
logbase       log base to use if reverse_log_scale = TRUE
output_enrichment how to return enrichment output
p_value       calculate p-values (boolean, default = FALSE)
n_times       number of permutations to calculate for p_value
name          to give to spatial enrichment results, default = rank
return_gobject return giotto object
```

**See Also**

[runRankEnrich](#)

---

rankSpatialCorGroups	<i>rankSpatialCorGroups</i>
----------------------	-----------------------------

---

**Description**

Rank spatial correlated clusters according to correlation structure

**Usage**

```
rankSpatialCorGroups(
  gobject,
  spatCorObject,
  use_clus_name = NULL,
  show_plot = NA,
  return_plot = FALSE,
  save_plot = NA,
  save_param = list(),
  default_save_name = "rankSpatialCorGroups"
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>spatCorObject</code>	spatial correlation object
<code>use_clus_name</code>	name of clusters to visualize (from <code>clusterSpatialCorGenes()</code> )
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters, see <a href="#">showSaveParameters</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

**Value**

data.table with positive (within group) and negative (outside group) scores

**Examples**

```
rankSpatialCorGroups(gobject)
```

---

<code>readExprMatrix</code>	<i>readExprMatrix</i>
-----------------------------	-----------------------

---

**Description**

Function to read an expression matrix into a sparse matrix.

**Usage**

```
readExprMatrix(path, cores = NA, transpose = FALSE)
```

**Arguments**

<code>path</code>	path to the expression matrix
<code>cores</code>	number of cores to use
<code>transpose</code>	transpose matrix

**Details**

The expression matrix needs to have both unique column names and row names

**Value**

sparse matrix

**Examples**

```
readExprMatrix()
```

---

readGiottoInstructions
<i>readGiottoInstructions</i>

---

**Description**

Retrieves the instruction associated with the provided parameter

**Usage**

readGiottoInstructions(giotto\_instructions, param = NULL)

**Arguments**

giotto_instructions	giotto object or result from createGiottoInstructions()
param	parameter to retrieve

**Value**

specific parameter

**Examples**

readGiottoInstructions()

---

removeCellAnnotation	<i>removeCellAnnotation</i>
----------------------	-----------------------------

---

**Description**

removes cell annotation of giotto object

**Usage**

removeCellAnnotation(gobject, columns = NULL, return\_gobject = TRUE)

**Arguments**

gobject	giotto object
columns	names of columns to remove
return_gobject	boolean: return giotto object (default = TRUE)

**Details**

if return\_gobject = FALSE, it will return the cell metadata

**Value**

giotto object

**Examples**

```
removeCellAnnotation(gobject)
```

---

```
removeGeneAnnotation    removeGeneAnnotation
```

---

**Description**

removes gene annotation of giotto object

**Usage**

```
removeGeneAnnotation(gobject, columns = NULL, return_gobject = TRUE)
```

**Arguments**

gobject	giotto object
columns	names of columns to remove
return_gobject	boolean: return giotto object (default = TRUE)

**Details**

if return\_gobject = FALSE, it will return the gene metadata

**Value**

giotto object

**Examples**

```
removeGeneAnnotation(gobject)
```

---

```
replaceGiottoInstructions  
                          replaceGiottoInstructions
```

---

**Description**

Function to replace all instructions from giotto object

**Usage**

```
replaceGiottoInstructions(gobject, instructions = NULL)
```

**Arguments**

gobject	giotto object
instructions	new instructions (e.g. result from createGiottoInstructions)

**Value**

giotto object with replaces instructions

**Examples**

```
replaceGiottoInstructions()
```

---

runDWLSDeconv	<i>runDWLSDeconv</i>
---------------	----------------------

---

**Description**

Function to perform DWLS deconvolution based on single cell expression data

**Usage**

```
runDWLSDeconv(  
  gobject,  
  expression_values = c("normalized"),  
  logbase = 2,  
  cluster_column = "leiden_clus",  
  sign_matrix,  
  n_cell = 50,  
  cutoff = 2,  
  name = NULL,  
  return_gobject = TRUE  
)
```

**Arguments**

gobject	giotto object
expression_values	expression values to use
logbase	base used for log normalization
cluster_column	name of cluster column
sign_matrix	sig matrix for deconvolution
n_cell	number of cells per spot
cutoff	cut off (default = 2)
name	name to give to spatial deconvolution results, default = DWLS
return_gobject	return giotto object

**Value**

giotto object or deconvolution results



---

```
runHyperGeometricEnrich
      runHyperGeometricEnrich
```

---

## Description

Function to calculate gene signature enrichment scores per spatial position using a hypergeometric test.

## Usage

```
runHyperGeometricEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  top_percentage = 5,
  output_enrichment = c("original", "zscore"),
  p_value = FALSE,
  name = NULL,
  return_gobject = TRUE
)
```

## Arguments

<code>gobject</code>	Giotto object
<code>sign_matrix</code>	Matrix of signature genes for each cell type / process
<code>expression_values</code>	expression values to use
<code>reverse_log_scale</code>	reverse expression values from log scale
<code>logbase</code>	log base to use if <code>reverse_log_scale = TRUE</code>
<code>top_percentage</code>	percentage of cells that will be considered to have gene expression with matrix binarization
<code>output_enrichment</code>	how to return enrichment output
<code>p_value</code>	calculate p-values (boolean, default = FALSE)
<code>name</code>	to give to spatial enrichment results, default = rank
<code>return_gobject</code>	return giotto object

## Details

The enrichment score is calculated based on the p-value from the hypergeometric test,  $-\log_{10}(p\text{-value})$ .

## Value

data.table with enrichment results

---

runPAGEEnrich	<i>runPAGEEnrich</i>
---------------	----------------------

---

## Description

Function to calculate gene signature enrichment scores per spatial position using PAGE.

## Usage

```
runPAGEEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore"),
  p_value = FALSE,
  n_times = 1000,
  name = NULL,
  return_gobject = TRUE
)
```

## Arguments

<code>gobject</code>	Giotto object
<code>sign_matrix</code>	Matrix of signature genes for each cell type / process
<code>expression_values</code>	expression values to use
<code>reverse_log_scale</code>	reverse expression values from log scale
<code>logbase</code>	log base to use if <code>reverse_log_scale = TRUE</code>
<code>output_enrichment</code>	how to return enrichment output
<code>p_value</code>	calculate p-values (boolean, default = FALSE)
<code>n_times</code>	number of permutations to calculate for <code>p_value</code>
<code>name</code>	to give to spatial enrichment results, default = PAGE
<code>return_gobject</code>	return giotto object

## Details

`sign_matrix`: a binary matrix with genes as row names and cell-types as column names. Alternatively a list of signature genes can be provided to `makeSignMatrixPAGE`, which will create the matrix for you.

The enrichment Z score is calculated by using method (PAGE) from Kim SY et al., BMC bioinformatics, 2005 as  $Z = ((Sm \sim \mu) * m^{(1/2)}) / \delta$ . For each gene in each spot,  $\mu$  is the fold change values versus the mean expression and  $\delta$  is the standard deviation.  $Sm$  is the mean fold change value of a specific marker gene set and  $m$  is the size of a given marker gene set.

**Value**

data.table with enrichment results

**See Also**

[makeSignMatrixPAGE](#)

---

runPCA	<i>runPCA</i>
--------	---------------

---

**Description**

runs a Principal Component Analysis

**Usage**

```
runPCA(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  name = "pca",
  genes_to_use = "hvg",
  return_gobject = TRUE,
  center = F,
  scale_unit = F,
  ncp = 100,
  method = c("irlba", "factominer"),
  rev = FALSE,
  verbose = TRUE,
  ...
)
```

**Arguments**

gobject	giotto object
expression_values	expression values to use
reduction	cells or genes
name	arbitrary name for PCA run
genes_to_use	subset of genes to use for PCA
return_gobject	boolean: return giotto object (default = TRUE)
center	center data first (default = FALSE)
scale_unit	scale features before PCA (default = FALSE)
ncp	number of principal components to calculate
method	which implementation to use
rev	do a reverse PCA
verbose	verbosity of the function
...	additional parameters for PCA (see details)

## Details

See [prcomp\\_irlba](#) and [PCA](#) for more information about other parameters.

- `genes_to_use = NULL`: will use all genes from the selected matrix
- `genes_to_use = <hvg name>`: can be used to select a column name of highly variable genes, created by (see [calculateHVG](#))
- `genes_to_use = c('geneA', 'geneB', ...)`: will use all manually provided genes

## Value

giotto object with updated PCA dimension reduction

## Examples

```
# 1. create giotto object
expr_path = system.file("extdata", "seqfish_field_expr.txt", package = 'Giotto')
loc_path = system.file("extdata", "seqfish_field_locs.txt", package = 'Giotto')
VC_small <- createGiottoObject(raw_exprs = expr_path, spatial_locs = loc_path)

# 2. normalize giotto
VC_small <- normalizeGiotto(gobject = VC_small, scalefactor = 6000)
VC_small <- addStatistics(gobject = VC_small)

# 3. dimension reduction
VC_small <- calculateHVG(gobject = VC_small)
VC_small <- runPCA(gobject = VC_small)
plotPCA(VC_small)
```

---

runRankEnrich

*runRankEnrich*


---

## Description

Function to calculate gene signature enrichment scores per spatial position using a rank based approach.

## Usage

```
runRankEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore"),
  p_value = FALSE,
  n_times = 1000,
  name = NULL,
  return_gobject = TRUE
)
```

**Arguments**

<code>gobject</code>	Giotto object
<code>sign_matrix</code>	Matrix of signature genes for each cell type / process
<code>expression_values</code>	expression values to use
<code>reverse_log_scale</code>	reverse expression values from log scale
<code>logbase</code>	log base to use if <code>reverse_log_scale = TRUE</code>
<code>output_enrichment</code>	how to return enrichment output
<code>p_value</code>	calculate p-values (boolean, default = FALSE)
<code>n_times</code>	number of permutations to calculate for <code>p_value</code>
<code>name</code>	to give to spatial enrichment results, default = rank
<code>return_gobject</code>	return giotto object

**Details**

`sign_matrix`: a rank-fold matrix with genes as row names and cell-types as column names. Alternatively a scRNA-seq matrix and vector with clusters can be provided to `makeSignMatrixRank`, which will create the matrix for you.

First a new rank is calculated as  $R = (R1 * R2)^{(1/2)}$ , where  $R1$  is the rank of fold-change for each gene in each spot and  $R2$  is the rank of each marker in each cell type. The Rank-Biased Precision is then calculated as:  $RBP = (1 - 0.99) * (0.99)^{(R - 1)}$  and the final enrichment score is then calculated as the sum of top 100 RBPs.

**Value**

data.table with enrichment results

**See Also**

[makeSignMatrixRank](#)

---

runSpatialDeconv

*runSpatialDeconv*


---

**Description**

Function to perform deconvolution based on single cell expression data

**Usage**

```
runSpatialDeconv(
  gobject,
  deconv_method = c("DWLS"),
  expression_values = c("normalized"),
  logbase = 2,
  cluster_column = "leiden_clus",
  sign_matrix,
  n_cell = 50,
  cutoff = 2,
  name = NULL,
  return_gobject = TRUE
)
```

**Arguments**

gobject	giotto object
deconv_method	method to use for deconvolution
expression_values	expression values to use
logbase	base used for log normalization
cluster_column	name of cluster column
sign_matrix	signature matrix for deconvolution
n_cell	number of cells per spot
cutoff	cut off (default = 2)
name	name to give to spatial deconvolution results
return_gobject	return giotto object

**Value**

giotto object or deconvolution results

---

runSpatialEnrich	<i>runSpatialEnrich</i>
------------------	-------------------------

---

**Description**

Function to calculate gene signature enrichment scores per spatial position using an enrichment test.

**Usage**

```
runSpatialEnrich(
  gobject,
  enrich_method = c("PAGE", "rank", "hypergeometric"),
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
```

```

    p_value = FALSE,
    n_times = 1000,
    top_percentage = 5,
    output_enrichment = c("original", "zscore"),
    name = NULL,
    return_gobject = TRUE
  )

```

### Arguments

<code>gobject</code>	Giotto object
<code>enrich_method</code>	method for gene signature enrichment calculation
<code>sign_matrix</code>	Matrix of signature genes for each cell type / process
<code>expression_values</code>	expression values to use
<code>reverse_log_scale</code>	reverse expression values from log scale
<code>logbase</code>	log base to use if <code>reverse_log_scale = TRUE</code>
<code>p_value</code>	calculate p-value (default = FALSE)
<code>n_times</code>	(page/rank) number of permutation iterations to calculate p-value
<code>top_percentage</code>	(hyper) percentage of cells that will be considered to have gene expression with matrix binarization
<code>output_enrichment</code>	how to return enrichment output
<code>name</code>	to give to spatial enrichment results, default = PAGE
<code>return_gobject</code>	return giotto object

### Details

For details see the individual functions:

- PAGE: [runPAGEEnrich](#)
- Rank: [runRankEnrich](#)
- Hypergeometric: [runHyperGeometricEnrich](#)

### Value

Giotto object or enrichment results if `return_gobject = FALSE`

---

runtSNE

*runtSNE*

---

### Description

run tSNE

**Usage**

```

runtSNE(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "tsne",
  genes_to_use = NULL,
  return_gobject = TRUE,
  dims = 2,
  perplexity = 30,
  theta = 0.5,
  do_PCA_first = F,
  set_seed = T,
  seed_number = 1234,
  verbose = TRUE,
  ...
)

```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>reduction</code>	cells or genes
<code>dim_reduction_to_use</code>	use another dimension reduction set as input
<code>dim_reduction_name</code>	name of dimension reduction set to use
<code>dimensions_to_use</code>	number of dimensions to use as input
<code>name</code>	arbitrary name for tSNE run
<code>genes_to_use</code>	if <code>dim_reduction_to_use = NULL</code> , which genes to use
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>dims</code>	tSNE param: number of dimensions to return
<code>perplexity</code>	tSNE param: perplexity
<code>theta</code>	tSNE param: theta
<code>do_PCA_first</code>	tSNE param: do PCA before tSNE (default = FALSE)
<code>set_seed</code>	use of seed
<code>seed_number</code>	seed number to use
<code>verbose</code>	verbosity of the function
<code>...</code>	additional tSNE parameters



## Details

See [Rtsne](#) for more information about these and other parameters.

- Input for tSNE dimension reduction can be another dimension reduction (default = 'pca')
- To use gene expression as input set `dim_reduction_to_use = NULL`
- If `dim_reduction_to_use = NULL`, `genes_to_use` can be used to select a column name of highly variable genes (see [calculateHVG](#)) or simply provide a vector of genes
- multiple tSNE results can be stored by changing the *name* of the analysis

## Value

giotto object with updated tSNE dimension reduction

## Examples

```
# 1. create giotto object
expr_path = system.file("extdata", "seqfish_field_expr.txt", package = 'Giotto')
loc_path = system.file("extdata", "seqfish_field_locs.txt", package = 'Giotto')
VC_small <- createGiottoObject(raw_exprs = expr_path, spatial_locs = loc_path)

# 2. normalize giotto
VC_small <- normalizeGiotto(gobject = VC_small, scalefactor = 6000)
VC_small <- addStatistics(gobject = VC_small)

# 3. dimension reduction
VC_small <- calculateHVG(gobject = VC_small)
VC_small <- runPCA(gobject = VC_small)
VC_small <- runTSNE(VC_small, dimensions_to_use = 1:5, n_threads = 2)
plotTSNE(gobject = VC_small)
```

---

runUMAP

---

*runUMAP*


---

## Description

run UMAP

## Usage

```
runUMAP(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "umap",
  genes_to_use = NULL,
  return_gobject = TRUE,
  n_neighbors = 40,
```

```

    n_components = 2,
    n_epochs = 400,
    min_dist = 0.01,
    n_threads = 1,
    spread = 5,
    set_seed = T,
    seed_number = 1234,
    verbose = T,
    ...
)

```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>reduction</code>	cells or genes
<code>dim_reduction_to_use</code>	use another dimension reduction set as input
<code>dim_reduction_name</code>	name of dimension reduction set to use
<code>dimensions_to_use</code>	number of dimensions to use as input
<code>name</code>	arbitrary name for UMAP run
<code>genes_to_use</code>	if <code>dim_reduction_to_use = NULL</code> , which genes to use
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>n_neighbors</code>	UMAP param: number of neighbors
<code>n_components</code>	UMAP param: number of components
<code>n_epochs</code>	UMAP param: number of epochs
<code>min_dist</code>	UMAP param: minimum distance
<code>n_threads</code>	UMAP param: threads to use
<code>spread</code>	UMAP param: spread
<code>set_seed</code>	use of seed
<code>seed_number</code>	seed number to use
<code>verbose</code>	verbosity of function
<code>...</code>	additional UMAP parameters

## Details

See [umap](#) for more information about these and other parameters.

- Input for UMAP dimension reduction can be another dimension reduction (default = 'pca')
- To use gene expression as input set `dim_reduction_to_use = NULL`
- If `dim_reduction_to_use = NULL`, `genes_to_use` can be used to select a column name of highly variable genes (see [calculateHVG](#)) or simply provide a vector of genes
- multiple UMAP results can be stored by changing the *name* of the analysis

## Value

giotto object with updated UMAP dimension reduction

## Examples

```
# 1. create giotto object
expr_path = system.file("extdata", "seqfish_field_expr.txt", package = 'Giotto')
loc_path = system.file("extdata", "seqfish_field_locs.txt", package = 'Giotto')
VC_small <- createGiottoObject(raw_exprs = expr_path, spatial_locs = loc_path)

# 2. normalize giotto
VC_small <- normalizeGiotto(gobject = VC_small, scalefactor = 6000)
VC_small <- addStatistics(gobject = VC_small)

# 3. dimension reduction
VC_small <- calculateHVG(gobject = VC_small)
VC_small <- runPCA(gobject = VC_small)
VC_small <- runUMAP(VC_small, dimensions_to_use = 1:5, n_threads = 2)
plotUMAP(gobject = VC_small)
```

---

screePlot

*screePlot*

---

## Description

identify significant principal components (PCs) using an screeplot (a.k.a. elbowplot)

## Usage

```
screePlot(
  gobject,
  name = "pca",
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  method = c("irlba", "factominer"),
  rev = FALSE,
  genes_to_use = NULL,
  center = F,
  scale_unit = F,
  ncp = 100,
  ylim = c(0, 20),
  verbose = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "screePlot",
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>name</code>	name of PCA object if available
<code>expression_values</code>	expression values to use
<code>reduction</code>	cells or genes
<code>method</code>	which implementation to use
<code>rev</code>	do a reverse PCA
<code>genes_to_use</code>	subset of genes to use for PCA
<code>center</code>	center data before PCA
<code>scale_unit</code>	scale features before PCA
<code>ncp</code>	number of principal components to calculate
<code>ylim</code>	y-axis limits on scree plot
<code>verbose</code>	verbosity
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <code>all_plots_save_function()</code>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>
<code>...</code>	additional arguments to <code>pca</code> function, see <a href="#">runPCA</a>

**Details**

Screeplot works by plotting the explained variance of each individual PC in a barplot allowing you to identify which PC provides a significant contribution (a.k.a 'elbow method').

Screeplot will use an available `pca` object, based on the parameter 'name', or it will create it if it's not available (see [runPCA](#))

**Value**

ggplot object for scree method

**Examples**

```
screePlot(gobject)
```

---

selectPatternGenes	<i>selectPatternGenes</i>
--------------------	---------------------------

---

## Description

Select genes correlated with spatial patterns

## Usage

```
selectPatternGenes(  
  spatPatObj,  
  dimensions = 1:5,  
  top_pos_genes = 10,  
  top_neg_genes = 10,  
  min_pos_cor = 0.5,  
  min_neg_cor = -0.5,  
  return_top_selection = FALSE  
)
```

## Arguments

spatPatObj	Output from detectSpatialPatterns
dimensions	dimensions to identify correlated genes for.
top_pos_genes	Top positively correlated genes.
top_neg_genes	Top negatively correlated genes.
min_pos_cor	Minimum positive correlation score to include a gene.
min_neg_cor	Minimum negative correlation score to include a gene.
return_top_selection	only return selection based on correlation criteria (boolean)

## Details

Description.

## Value

Data.table with genes associated with selected dimension (PC).

## Examples

```
selectPatternGenes(gobject)
```

---

```
select_expression_values
      select_expression_values
```

---

**Description**

helper function to select expression values

**Usage**

```
select_expression_values(gobject, values)
```

**Arguments**

<code>gobject</code>	giotto object
<code>values</code>	expression values to extract

**Value**

expression matrix

---

```
show,giotto-method      show method for giotto class
```

---

**Description**

show method for giotto class

**Usage**

```
## S4 method for signature 'giotto'
show(object)
```

---

```
showClusterDendrogram  showClusterDendrogram
```

---

**Description**

Creates dendrogram for selected clusters.

**Usage**

```
showClusterDendrogram(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  h = NULL,
  h_color = "red",
  rotate = FALSE,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showClusterDendrogram",
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>cluster_column</code>	name of column to use for clusters
<code>cor</code>	correlation score to calculate distance
<code>distance</code>	distance method to use for hierarchical clustering
<code>h</code>	height of horizontal lines to plot
<code>h_color</code>	color of horizontal lines
<code>rotate</code>	rotate dendrogram 90 degrees
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters, see <a href="#">showSaveParameters</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>
<code>...</code>	additional parameters for <code>ggdendrogram()</code>

**Details**

Expression correlation dendrogram for selected clusters.

**Value**

ggplot

**Examples**

```
showClusterDendrogram(gobject)
```

---

showClusterHeatmap	<i>showClusterHeatmap</i>
--------------------	---------------------------

---

## Description

Creates heatmap based on identified clusters

## Usage

```
showClusterHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = "all",
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showClusterHeatmap",
  ...
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>genes</code>	vector of genes to use, default to 'all'
<code>cluster_column</code>	name of column to use for clusters
<code>cor</code>	correlation score to calculate distance
<code>distance</code>	distance method to use for hierarchical clustering
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters, see <a href="#">showSaveParameters</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>
<code>...</code>	additional parameters for the Heatmap function from ComplexHeatmap

## Details

Correlation heatmap of selected clusters.

## Value

ggplot



**Examples**

```
showClusterHeatmap(gobject)
```

---

```
showGiottoImageNames    showGiottoImageNames
```

---

**Description**

Prints the available giotto images that are attached to the Giotto object

**Usage**

```
showGiottoImageNames(gobject, verbose = TRUE)
```

**Arguments**

gobject	a giotto object
verbose	verbosity of function

**Value**

a vector of giotto image names attached to the giotto object

**Examples**

```
showGiottoImageNames(gobject)
```

---

```
showGiottoInstructions  
                      showGiottoInstructions
```

---

**Description**

Function to display all instructions from giotto object

**Usage**

```
showGiottoInstructions(gobject)
```

**Arguments**

gobject	giotto object
---------	---------------

**Value**

named vector with giotto instructions

**Examples**

```
showGiottoInstructions()
```

---

`showGrids`*showGrids*

---

**Description**

Prints the available spatial grids that are attached to the Giotto object

**Usage**

```
showGrids(gobject, verbose = TRUE)
```

**Arguments**

<code>gobject</code>	a giotto object
<code>verbose</code>	verbosity of function#'

**Value**

vector

**Examples**

```
showGrids()
```

---

`showNetworks`*showNetworks*

---

**Description**

Prints the available spatial networks that are attached to the Giotto object

**Usage**

```
showNetworks(gobject, verbose = TRUE)
```

**Arguments**

<code>gobject</code>	a giotto object
<code>verbose</code>	verbosity of function#'

**Value**

vector

**Examples**

```
showNetworks()
```

---

showPattern	<i>showPattern</i>
-------------	--------------------

---

## Description

show patterns for 2D spatial data

## Usage

```
showPattern(gobject, spatPatObj, ...)
```

## Arguments

gobject	giotto object
spatPatObj	Output from detectSpatialPatterns
...	Arguments passed on to <a href="#">showPattern2D</a>
dimension	dimension to plot
trim	Trim ends of the PC values.
background_color	background color for plot
grid_border_color	color for grid
show_legend	show legend of ggplot
point_size	size of points
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

## Value

ggplot

## See Also

[showPattern2D](#)

## Examples

```
showPattern(gobject)
```

showPattern2D

*showPattern2D***Description**

show patterns for 2D spatial data

**Usage**

```
showPattern2D(
  gobject,
  spatPatObj,
  dimension = 1,
  trim = c(0.02, 0.98),
  background_color = "white",
  grid_border_color = "grey",
  show_legend = T,
  point_size = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showPattern2D"
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>spatPatObj</code>	Output from <code>detectSpatialPatterns</code>
<code>dimension</code>	dimension to plot
<code>trim</code>	Trim ends of the PC values.
<code>background_color</code>	background color for plot
<code>grid_border_color</code>	color for grid
<code>show_legend</code>	show legend of ggplot
<code>point_size</code>	size of points
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters, see <a href="#">showSaveParameters</a>
<code>default_save_name</code>	default save name for saving, don't change, change <code>save_name</code> in <code>save_param</code>

**Value**

ggplot

Examples

```
showPattern2D(gobject)
```

---

showPattern3D	<i>showPattern3D</i>
---------------	----------------------

---

Description

show patterns for 3D spatial data

Usage

```
showPattern3D(  
  gobject,  
  spatPatObj,  
  dimension = 1,  
  trim = c(0.02, 0.98),  
  background_color = "white",  
  grid_border_color = "grey",  
  show_legend = T,  
  point_size = 1,  
  axis_scale = c("cube", "real", "custom"),  
  custom_ratio = NULL,  
  x_ticks = NULL,  
  y_ticks = NULL,  
  z_ticks = NULL,  
  show_plot = NA,  
  return_plot = NA,  
  save_plot = NA,  
  save_param = list(),  
  default_save_name = "showPattern3D"  
)
```

Arguments

gobject	giotto object
spatPatObj	Output from detectSpatialPatterns
dimension	dimension to plot
trim	Trim ends of the PC values.
background_color	background color for plot
grid_border_color	color for grid
show_legend	show legend of plot
point_size	adjust the point size
axis_scale	scale the axis
custom_ratio	customize the scale of the axis
x_ticks	the tick number of x_axis

y_ticks	the tick number of y_axis
z_ticks	the tick number of z_axis
show_plot	show plot
return_plot	return plot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

**Value**

plotly

**Examples**

```
showPattern3D(gobject)
```

---

showPatternGenes	<i>showPatternGenes</i>
------------------	-------------------------

---

**Description**

show genes correlated with spatial patterns

**Usage**

```
showPatternGenes(  
  gobject,  
  spatPatObj,  
  dimension = 1,  
  top_pos_genes = 5,  
  top_neg_genes = 5,  
  point_size = 1,  
  return_DT = FALSE,  
  show_plot = NA,  
  return_plot = NA,  
  save_plot = NA,  
  save_param = list(),  
  default_save_name = "showPatternGenes"  
)
```

**Arguments**

gobject	giotto object
spatPatObj	Output from detectSpatialPatterns
dimension	dimension to plot genes for.
top_pos_genes	Top positively correlated genes.
top_neg_genes	Top negatively correlated genes.
point_size	size of points

return_DT	if TRUE, it will return the data.table used to generate the plots
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

**Value**

ggplot

**Examples**

```
showPatternGenes(gobject)
```

---

showProcessingSteps	<i>showProcessingSteps</i>
---------------------	----------------------------

---

**Description**

shows the sequential processing steps that were performed in a summarized format

**Usage**

```
showProcessingSteps(gobject)
```

**Arguments**

gobject                  giotto object

**Value**

list of processing steps and names

**Examples**

```
showProcessingSteps(gobject)
```

---

showSaveParameters	<i>showSaveParameters</i>
--------------------	---------------------------

---

**Description**

Description of Giotto saving options, links to [all\\_plots\\_save\\_function](#)

**Usage**

```
showSaveParameters()
```

**Value**

Instruction on how to use the automatic plot saving options within Giotto

**Examples**

```
showSaveParameters()
```

---

showSpatialCorGenes	<i>showSpatialCorGenes</i>
---------------------	----------------------------

---

**Description**

Shows and filters spatially correlated genes

**Usage**

```
showSpatialCorGenes(
  spatCorObject,
  use_clus_name = NULL,
  selected_clusters = NULL,
  genes = NULL,
  min_spat_cor = 0.5,
  min_expr_cor = NULL,
  min_cor_diff = NULL,
  min_rank_diff = NULL,
  show_top_genes = NULL
)
```

**Arguments**

spatCorObject	spatial correlation object
use_clus_name	cluster information to show
selected_clusters	subset of clusters to show
genes	subset of genes to show
min_spat_cor	filter on minimum spatial correlation
min_expr_cor	filter on minimum single-cell expression correlation



min\_cor\_diff     filter on minimum correlation difference (spatial vs expression)  
 min\_rank\_diff   filter on minimum correlation rank difference (spatial vs expression)  
 show\_top\_genes   show top genes per gene

**Value**

data.table with filtered information

**Examples**

```
showSpatialCorGenes(gobject)
```

---

signPCA	<i>signPCA</i>
---------	----------------

---

**Description**

identify significant principal components (PCs)

**Usage**

```
signPCA(
  gobject,
  name = "pca",
  method = c("screeplot", "jackstraw"),
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  pca_method = c("irlba", "factominer"),
  rev = FALSE,
  genes_to_use = NULL,
  center = T,
  scale_unit = T,
  ncp = 50,
  scree_ylim = c(0, 10),
  jack_iter = 10,
  jack_threshold = 0.01,
  jack_ylim = c(0, 1),
  verbose = TRUE,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "signPCA"
)
```

**Arguments**

gobject             giotto object  
 name                name of PCA object if available  
 method              method to use to identify significant PCs

expression_values	expression values to use
reduction	cells or genes
pca_method	which implementation to use
rev	do a reverse PCA
genes_to_use	subset of genes to use for PCA
center	center data before PCA
scale_unit	scale features before PCA
ncp	number of principal components to calculate
scree_ylim	y-axis limits on scree plot
jack_iter	number of iterations for jackstraw
jack_threshold	p-value threshold to call a PC significant
jack_ylim	y-axis limits on jackstraw plot
verbose	verbosity
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from all_plots_save_function()
default_save_name	default save name for saving, don't change, change save_name in save_param

## Details

Two different methods can be used to assess the number of relevant or significant principal components (PC's).

1. Screeplot works by plotting the explained variance of each individual PC in a barplot allowing you to identify which PC provides a significant contribution (a.k.a. 'elbow method').
2. The Jackstraw method uses the [permutationPA](#) function. By systematically permuting genes it identifies robust, and thus significant, PCs.

## Value

ggplot object for scree method and maxtrix of p-values for jackstraw

## Examples

```
signPCA(gobject)
```

---

silhouetteRank	<i>silhouetteRank</i>
----------------	-----------------------

---

## Description

Previously: `calculate_spatial_genes_python`. This method computes a silhouette score per gene based on the spatial distribution of two partitions of cells (expressed L1, and non-expressed L0). Here, rather than L2 Euclidean norm, it uses a rank-transformed, exponentially weighted function to represent the local physical distance between two cells.

## Usage

```
silhouetteRank(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metric = "euclidean",
  subset_genes = NULL,
  rbp_p = 0.95,
  examine_top = 0.3,
  python_path = NULL
)
```

## Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	expression values to use
<code>metric</code>	distance metric to use
<code>subset_genes</code>	only run on this subset of genes
<code>rbp_p</code>	fractional binarization threshold
<code>examine_top</code>	top fraction to evaluate with silhouette
<code>python_path</code>	specify specific path to python if required

## Value

data.table with spatial scores

## Examples

```
silhouetteRank(gobject)
```

---

spatCellCellcom	<i>spatCellCellcom</i>
-----------------	------------------------

---

## Description

Spatial Cell-Cell communication scores based on spatial expression of interacting cells

## Usage

```
spatCellCellcom(
  gobject,
  spatial_network_name = "Delaunay_network",
  cluster_column = "cell_types",
  random_iter = 1000,
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  min_observations = 2,
  adjust_method = c("fdr", "bonferroni", "BH", "holm", "hochberg", "hommel", "BY",
    "none"),
  adjust_target = c("genes", "cells"),
  do_parallel = TRUE,
  cores = NA,
  verbose = c("a little", "a lot", "none")
)
```

## Arguments

<code>gobject</code>	giotto object to use
<code>spatial_network_name</code>	spatial network to use for identifying interacting cells
<code>cluster_column</code>	cluster column with cell type information
<code>random_iter</code>	number of iterations
<code>gene_set_1</code>	first specific gene set from gene pairs
<code>gene_set_2</code>	second specific gene set from gene pairs
<code>log2FC_addendum</code>	addendum to add when calculating log2FC
<code>min_observations</code>	minimum number of interactions needed to be considered
<code>adjust_method</code>	which method to adjust p-values
<code>adjust_target</code>	adjust multiple hypotheses at the cell or gene level
<code>do_parallel</code>	run calculations in parallel with mclapply
<code>cores</code>	number of cores to use if <code>do_parallel = TRUE</code>
<code>verbose</code>	verbose

## Details

Statistical framework to identify if pairs of genes (such as ligand-receptor combinations) are expressed at higher levels than expected based on a reshuffled null distribution of gene expression values in cells that are spatially in proximity to eachother.. More details will follow soon.

## Value

Cell-Cell communication scores for gene pairs based on spatial interaction

## Examples

```
spatCellCellcom(gobject)
```

---

spatCellPlot	<i>spatCellPlot</i>
--------------	---------------------

---

## Description

Visualize cells according to spatial coordinates

## Usage

```
spatCellPlot(...)
```

## Arguments

```
...           Arguments passed on to spatCellPlot2D
gobject      giotto object
show_image   show a tissue background image
gimage       a giotto image
image_name   name of a giotto image
sdimx        x-axis dimension name (default = 'sdimx')
sdimy        y-axis dimension name (default = 'sdimy')
spat_enr_names names of spatial enrichment results to include
cell_annotation_values numeric cell annotation columns
cell_color_gradient vector with 3 colors for numeric data
gradient_midpoint midpoint for color gradient
gradient_limits vector with lower and upper limits
select_cell_groups select subset of cells/clusters based on cell_color parameter
select_cells select subset of cells based on cell IDs
point_shape  shape of points (border, no_border or voronoi)
point_size   size of point (cell)
point_alpha  transparency of spatial points
point_border_col color of border around points
point_border_stroke stroke size of border around points
show_cluster_center plot center of selected clusters
show_center_label  plot label of selected clusters
center_point_size  size of center points
center_point_border_col border color of center points
center_point_border_stroke border stroke size of center points
label_size        size of labels
label_fontface     font of labels
```

show\_network show underlying spatial network  
 spatial\_network\_name name of spatial network to use  
 network\_color color of spatial network  
 network\_alpha alpha of spatial network  
 show\_grid show spatial grid  
 spatial\_grid\_name name of spatial grid to use  
 grid\_color color of spatial grid  
 show\_other\_cells display not selected cells  
 other\_cell\_color color of not selected cells  
 other\_point\_size point size of not selected cells  
 other\_cells\_alpha alpha of not selected cells  
 coord\_fix\_ratio fix ratio between x and y-axis  
 show\_legend show legend  
 legend\_text size of legend text  
 legend\_symbol\_size size of legend symbols  
 background\_color color of plot background  
 vor\_border\_color border color for voronoi plot  
 vor\_max\_radius maximum radius for voronoi 'cells'  
 vor\_alpha transparency of voronoi 'cells'  
 axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plot  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)  
 default\_save\_name default save name for saving, don't change, change save\_name  
 in save\_param

## Details

Description of parameters.

## Value

ggplot

## See Also

Other spatial cell annotation visualizations: [spatCellPlot2D\(\)](#)

## Examples

```
spatCellPlot(gobject)
```

---

spatCellPlot2D

*spatCellPlot2D*


---

## Description

Visualize cells according to spatial coordinates

## Usage

```
spatCellPlot2D(
  gobject,
  show_image = F,
  gimage = NULL,
  image_name = "image",
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
  cell_annotation_values = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_shape = c("border", "no_border", "voronoi"),
  point_size = 3,
  point_alpha = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  show_network = F,
  spatial_network_name = "Delaunay_network",
  network_color = NULL,
  network_alpha = 1,
  show_grid = F,
  spatial_grid_name = "spatial_grid",
  grid_color = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  other_cells_alpha = 0.1,
  coord_fix_ratio = NULL,
  show_legend = T,
  legend_text = 8,
  legend_symbol_size = 1,
  background_color = "white",
```

```

    vor_border_color = "white",
    vor_max_radius = 200,
    vor_alpha = 1,
    axis_text = 8,
    axis_title = 8,
    cow_n_col = 2,
    cow_rel_h = 1,
    cow_rel_w = 1,
    cow_align = "h",
    show_plot = NA,
    return_plot = NA,
    save_plot = NA,
    save_param = list(),
    default_save_name = "spatCellPlot2D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>show_image</code>	show a tissue background image
<code>gimage</code>	a giotto image
<code>image_name</code>	name of a giotto image
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>cell_annotation_values</code>	numeric cell annotation columns
<code>cell_color_gradient</code>	vector with 3 colors for numeric data
<code>gradient_midpoint</code>	midpoint for color gradient
<code>gradient_limits</code>	vector with lower and upper limits
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs
<code>point_shape</code>	shape of points (border, no_border or voronoi)
<code>point_size</code>	size of point (cell)
<code>point_alpha</code>	transparency of spatial points
<code>point_border_col</code>	color of border around points
<code>point_border_stroke</code>	stroke size of border around points
<code>show_cluster_center</code>	plot center of selected clusters
<code>show_center_label</code>	plot label of selected clusters



<code>center_point_size</code>	size of center points
<code>center_point_border_col</code>	border color of center points
<code>center_point_border_stroke</code>	border stroke size of center points
<code>label_size</code>	size of labels
<code>label_fontface</code>	font of labels
<code>show_network</code>	show underlying spatial network
<code>spatial_network_name</code>	name of spatial network to use
<code>network_color</code>	color of spatial network
<code>network_alpha</code>	alpha of spatial network
<code>show_grid</code>	show spatial grid
<code>spatial_grid_name</code>	name of spatial grid to use
<code>grid_color</code>	color of spatial grid
<code>show_other_cells</code>	display not selected cells
<code>other_cell_color</code>	color of not selected cells
<code>other_point_size</code>	point size of not selected cells
<code>other_cells_alpha</code>	alpha of not selected cells
<code>coord_fix_ratio</code>	fix ratio between x and y-axis
<code>show_legend</code>	show legend
<code>legend_text</code>	size of legend text
<code>legend_symbol_size</code>	size of legend symbols
<code>background_color</code>	color of plot background
<code>vor_border_color</code>	border color for voronoi plot
<code>vor_max_radius</code>	maximum radius for voronoi 'cells'
<code>vor_alpha</code>	transparency of voronoi 'cells'
<code>axis_text</code>	size of axis text
<code>axis_title</code>	size of axis title
<code>cow_n_col</code>	cowplot param: how many columns
<code>cow_rel_h</code>	cowplot param: relative height
<code>cow_rel_w</code>	cowplot param: relative width
<code>cow_align</code>	cowplot param: how to align
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object

save\_plot            directly save the plot [boolean]  
save\_param          list of saving parameters, see [showSaveParameters](#)  
default\_save\_name       default save name for saving, don't change, change save\_name in save\_param

**Details**

Description of parameters.

**Value**

ggplot

**See Also**

Other spatial cell annotation visualizations: [spatCellPlot\(\)](#)

**Examples**

spatCellPlot2D(gobject)

---

spatDimCellPlot	<i>spatDimCellPlot</i>
-----------------	------------------------

---

**Description**

Visualize numerical features of cells according to spatial AND dimension reduction coordinates in 2D

**Usage**

spatDimCellPlot(...)

**Arguments**

...                    Arguments passed on to [spatDimCellPlot2D](#)  
gobject   giotto object  
show\_image   show a tissue background image  
gimage   a giotto image  
image\_name   name of a giotto image  
plot\_alignment   direction to align plot  
spat\_enr\_names   names of spatial enrichment results to include  
cell\_annotation\_values   numeric cell annotation columns  
dim\_reduction\_to\_use   dimension reduction to use  
dim\_reduction\_name   dimension reduction name  
dim1\_to\_use   dimension to use on x-axis  
dim2\_to\_use   dimension to use on y-axis  
sdimx   = spatial dimension to use on x-axis  
sdimy   = spatial dimension to use on y-axis

cell\_color\_gradient vector with 3 colors for numeric data  
 gradient\_midpoint midpoint for color gradient  
 gradient\_limits vector with lower and upper limits  
 select\_cell\_groups select subset of cells/clusters based on cell\_color parameter  
 select\_cells select subset of cells based on cell IDs  
 dim\_point\_shape dim reduction points with border or not (border or no\_border)  
 dim\_point\_size size of points in dim. reduction space  
 dim\_point\_alpha transparency of dim. reduction points  
 dim\_point\_border\_col border color of points in dim. reduction space  
 dim\_point\_border\_stroke border stroke of points in dim. reduction space  
 spat\_point\_shape shape of points (border, no\_border or voronoi)  
 spat\_point\_size size of spatial points  
 spat\_point\_alpha transparency of spatial points  
 spat\_point\_border\_col border color of spatial points  
 spat\_point\_border\_stroke border stroke of spatial points  
 dim\_show\_cluster\_center show the center of each cluster  
 dim\_show\_center\_label provide a label for each cluster  
 dim\_center\_point\_size size of the center point  
 dim\_center\_point\_border\_col border color of center point  
 dim\_center\_point\_border\_stroke stroke size of center point  
 dim\_label\_size size of the center label  
 dim\_label\_fontface font of the center label  
 spat\_show\_cluster\_center show the center of each cluster  
 spat\_show\_center\_label provide a label for each cluster  
 spat\_center\_point\_size size of the spatial center points  
 spat\_center\_point\_border\_col border color of the spatial center points  
 spat\_center\_point\_border\_stroke stroke size of the spatial center points  
 spat\_label\_size size of the center label  
 spat\_label\_fontface font of the center label  
 show\_NN\_network show underlying NN network  
 nn\_network\_to\_use type of NN network to use (kNN vs sNN)  
 nn\_network\_name name of NN network to use, if show\_NN\_network = TRUE  
 dim\_edge\_alpha column to use for alpha of the edges  
 spat\_show\_network show spatial network  
 spatial\_network\_name name of spatial network to use  
 spat\_network\_color color of spatial network  
 spat\_network\_alpha alpha of spatial network  
 spat\_show\_grid show spatial grid  
 spatial\_grid\_name name of spatial grid to use  
 spat\_grid\_color color of spatial grid  
 show\_other\_cells display not selected cells  
 other\_cell\_color color of not selected cells  
 dim\_other\_point\_size size of not selected dim cells  
 spat\_other\_point\_size size of not selected spat cells  
 spat\_other\_cells\_alpha alpha of not selected spat cells

coord\_fix\_ratio ratio for coordinates  
cow\_n\_col cowplot param: how many columns  
cow\_rel\_h cowplot param: relative height  
cow\_rel\_w cowplot param: relative width  
cow\_align cowplot param: how to align  
show\_legend show legend  
legend\_text size of legend text  
legend\_symbol\_size size of legend symbols  
dim\_background\_color background color of points in dim. reduction space  
spat\_background\_color background color of spatial points  
vor\_border\_color border color for voronoi plot  
vor\_max\_radius maximum radius for voronoi 'cells'  
vor\_alpha transparency of voronoi 'cells'  
axis\_text size of axis text  
axis\_title size of axis title  
show\_plot show plot  
return\_plot return ggplot object  
save\_plot directly save the plot [boolean]  
save\_param list of saving parameters, see [showSaveParameters](#)  
default\_save\_name default save name for saving, don't change, change save\_name in save\_param

**Details**

Description of parameters.

**Value**

ggplot

**See Also**

Other spatial and dimension reduction cell annotation visualizations: [spatDimCellPlot2D\(\)](#)

**Examples**

spatDimCellPlot(gobject)

---

spatDimCellPlot2D	<i>spatDimCellPlot2D</i>
-------------------	--------------------------

---

**Description**

Visualize numerical features of cells according to spatial AND dimension reduction coordinates in 2D

**Usage**

```
spatDimCellPlot2D(  
  gobject,  
  show_image = F,  
  gimage = NULL,  
  image_name = "image",  
  plot_alignment = c("vertical", "horizontal"),  
  spat_enr_names = NULL,  
  cell_annotation_values = NULL,  
  dim_reduction_to_use = "umap",  
  dim_reduction_name = "umap",  
  dim1_to_use = 1,  
  dim2_to_use = 2,  
  sdimx = "sdimx",  
  sdimy = "sdimy",  
  cell_color_gradient = c("blue", "white", "red"),  
  gradient_midpoint = NULL,  
  gradient_limits = NULL,  
  select_cell_groups = NULL,  
  select_cells = NULL,  
  dim_point_shape = c("border", "no_border"),  
  dim_point_size = 1,  
  dim_point_alpha = 1,  
  dim_point_border_col = "black",  
  dim_point_border_stroke = 0.1,  
  spat_point_shape = c("border", "no_border", "voronoi"),  
  spat_point_size = 1,  
  spat_point_alpha = 1,  
  spat_point_border_col = "black",  
  spat_point_border_stroke = 0.1,  
  dim_show_cluster_center = F,  
  dim_show_center_label = T,  
  dim_center_point_size = 4,  
  dim_center_point_border_col = "black",  
  dim_center_point_border_stroke = 0.1,  
  dim_label_size = 4,  
  dim_label_fontface = "bold",  
  spat_show_cluster_center = F,  
  spat_show_center_label = F,  
  spat_center_point_size = 4,  
  spat_center_point_border_col = "black",  
  spat_center_point_border_stroke = 0.1,  
  spat_label_size = 4,  
  spat_label_fontface = "bold",  
  show_NN_network = F,  
  nn_network_to_use = "sNN",  
  nn_network_name = "sNN.pca",  
  dim_edge_alpha = 0.5,  
  spat_show_network = F,  
  spatial_network_name = "Delaunay_network",  
  spat_network_color = "red",  
  spat_network_alpha = 0.5,
```

```

    spat_show_grid = F,
    spatial_grid_name = "spatial_grid",
    spat_grid_color = "green",
    show_other_cells = TRUE,
    other_cell_color = "grey",
    dim_other_point_size = 0.5,
    spat_other_point_size = 0.5,
    spat_other_cells_alpha = 0.5,
    show_legend = T,
    legend_text = 8,
    legend_symbol_size = 1,
    dim_background_color = "white",
    spat_background_color = "white",
    vor_border_color = "white",
    vor_max_radius = 200,
    vor_alpha = 1,
    axis_text = 8,
    axis_title = 8,
    coord_fix_ratio = NULL,
    cow_n_col = 2,
    cow_rel_h = 1,
    cow_rel_w = 1,
    cow_align = "h",
    show_plot = NA,
    return_plot = NA,
    save_plot = NA,
    save_param = list(),
    default_save_name = "spatDimCellPlot2D"
  )

```

### Arguments

<code>gobject</code>	giotto object
<code>show_image</code>	show a tissue background image
<code>gimage</code>	a giotto image
<code>image_name</code>	name of a giotto image
<code>plot_alignment</code>	direction to align plot
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>cell_annotation_values</code>	numeric cell annotation columns
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>sdimx</code>	= spatial dimension to use on x-axis
<code>sdimy</code>	= spatial dimension to use on y-axis
<code>cell_color_gradient</code>	vector with 3 colors for numeric data

gradient\_midpoint      midpoint for color gradient  
 gradient\_limits      vector with lower and upper limits  
 select\_cell\_groups      select subset of cells/clusters based on cell\_color parameter  
 select\_cells      select subset of cells based on cell IDs  
 dim\_point\_shape      dim reduction points with border or not (border or no\_border)  
 dim\_point\_size      size of points in dim. reduction space  
 dim\_point\_alpha      transparency of dim. reduction points  
 dim\_point\_border\_col      border color of points in dim. reduction space  
 dim\_point\_border\_stroke      border stroke of points in dim. reduction space  
 spat\_point\_shape      shape of points (border, no\_border or voronoi)  
 spat\_point\_size      size of spatial points  
 spat\_point\_alpha      transparency of spatial points  
 spat\_point\_border\_col      border color of spatial points  
 spat\_point\_border\_stroke      border stroke of spatial points  
 dim\_show\_cluster\_center      show the center of each cluster  
 dim\_show\_center\_label      provide a label for each cluster  
 dim\_center\_point\_size      size of the center point  
 dim\_center\_point\_border\_col      border color of center point  
 dim\_center\_point\_border\_stroke      stroke size of center point  
 dim\_label\_size      size of the center label  
 dim\_label\_fontface      font of the center label  
 spat\_show\_cluster\_center      show the center of each cluster  
 spat\_show\_center\_label      provide a label for each cluster  
 spat\_center\_point\_size      size of the spatial center points  
 spat\_center\_point\_border\_col      border color of the spatial center points

```

spat_center_point_border_stroke
    stroke size of the spatial center points
spat_label_size
    size of the center label
spat_label_fontface
    font of the center label
show_NN_network
    show underlying NN network
nn_network_to_use
    type of NN network to use (kNN vs sNN)
nn_network_name
    name of NN network to use, if show_NN_network = TRUE
dim_edge_alpha
    column to use for alpha of the edges
spat_show_network
    show spatial network
spatial_network_name
    name of spatial network to use
spat_network_color
    color of spatial network
spat_network_alpha
    alpha of spatial network
spat_show_grid
    show spatial grid
spatial_grid_name
    name of spatial grid to use
spat_grid_color
    color of spatial grid
show_other_cells
    display not selected cells
other_cell_color
    color of not selected cells
dim_other_point_size
    size of not selected dim cells
spat_other_point_size
    size of not selected spat cells
spat_other_cells_alpha
    alpha of not selected spat cells
show_legend
    show legend
legend_text
    size of legend text
legend_symbol_size
    size of legend symbols
dim_background_color
    background color of points in dim. reduction space
spat_background_color
    background color of spatial points
vor_border_color
    border color for voronoi plot
vor_max_radius
    maximum radius for voronoi 'cells'

```



vor_alpha	transparancy of voronoi 'cells'
axis_text	size of axis text
axis_title	size of axis title
coord_fix_ratio	ratio for coordinates
cow_n_col	cowplot param: how many columns
cow_rel_h	cowplot param: relative height
cow_rel_w	cowplot param: relative width
cow_align	cowplot param: how to align
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

Details

Description of parameters.

Value

ggplot

See Also

Other spatial and dimension reduction cell annotation visualizations: [spatDimCellPlot\(\)](#)

Examples

```
spatDimCellPlot2D(gobject)
```

---

spatDimGenePlot	<i>spatDimGenePlot</i>
-----------------	------------------------

---

Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

Usage

```
spatDimGenePlot(...)
```

**Arguments**

... Arguments passed on to [spatDimGenePlot2D](#)

`gobject` giotto object

`show_image` show a tissue background image

`gimage` a giotto image

`image_name` name of a giotto image

`expression_values` gene expression values to use

`plot_alignment` direction to align plot

`genes` genes to show

`dim_reduction_to_use` dimension reduction to use

`dim_reduction_name` dimension reduction name

`dim1_to_use` dimension to use on x-axis

`dim2_to_use` dimension to use on y-axis

`dim_point_shape` dim reduction points with border or not (border or no\_border)

`dim_point_size` dim reduction plot: point size

`dim_point_alpha` transparency of dim. reduction points

`dim_point_border_col` color of border around points

`dim_point_border_stroke` stroke size of border around points

`show_NN_network` show underlying NN network

`show_spatial_network` show underlying spatial network

`nn_network_to_use` type of NN network to use (kNN vs sNN)

`network_name` name of NN network to use, if `show_NN_network = TRUE`

`dim_network_color` color of NN network

`dim_edge_alpha` dim reduction plot: column to use for alpha of the edges

`scale_alpha_with_expression` scale expression with ggplot alpha parameter

`sdimx` spatial x-axis dimension name (default = 'sdimx')

`sdimy` spatial y-axis dimension name (default = 'sdimy')

`spatial_network_name` name of spatial network to use

`spatial_network_color` color of spatial network

`show_spatial_grid` show spatial grid

`grid_color` color of spatial grid

`spatial_grid_name` name of spatial grid to use

`spat_point_shape` spatial points with border or not (border or no\_border)

`spat_point_size` spatial plot: point size

`spat_point_alpha` transparency of spatial points

`spat_point_border_col` color of border around points

`spat_point_border_stroke` stroke size of border around points

`spat_edge_alpha` edge alpha

`cell_color_gradient` vector with 3 colors for numeric data

`gradient_midpoint` midpoint for color gradient

`gradient_limits` vector with lower and upper limits

`show_legend` show legend

`legend_text` size of legend text

`dim_background_color` color of plot background for dimension plot

`spat_background_color` color of plot background for spatial plot

vor\_border\_color border color for voronoi plot  
 vor\_max\_radius maximum radius for voronoi 'cells'  
 vor\_alpha transparency of voronoi 'cells'  
 axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plots  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)  
 default\_save\_name default save name for saving, don't change, change save\_name in save\_param

## Details

Description of parameters.

## Value

ggplot

## See Also

[spatDimGenePlot3D](#)

Other spatial and dimension reduction gene expression visualizations: [spatDimGenePlot2D\(\)](#), [spatDimGenePlot3D\(\)](#)

## Examples

```
spatDimGenePlot(gobject)
```

---

spatDimGenePlot2D	<i>spatDimGenePlot2D</i>
-------------------	--------------------------

---

## Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

## Usage

```

spatDimGenePlot2D(
  gobject,
  show_image = F,
  gimage = NULL,
  image_name = "image",
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("vertical", "horizontal"),

```

```

genes,
dim_reduction_to_use = "umap",
dim_reduction_name = "umap",
dim1_to_use = 1,
dim2_to_use = 2,
dim_point_shape = c("border", "no_border"),
dim_point_size = 1,
dim_point_alpha = 1,
dim_point_border_col = "black",
dim_point_border_stroke = 0.1,
show_NN_network = F,
show_spatial_network = F,
dim_network_color = "gray",
nn_network_to_use = "sNN",
network_name = "sNN.pca",
dim_edge_alpha = NULL,
scale_alpha_with_expression = FALSE,
sdmx = "sdmx",
sdmy = "sdmy",
spatial_network_name = "Delaunay_network",
spatial_network_color = NULL,
show_spatial_grid = F,
grid_color = NULL,
spatial_grid_name = "spatial_grid",
spat_point_shape = c("border", "no_border", "voronoi"),
spat_point_size = 1,
spat_point_alpha = 1,
spat_point_border_col = "black",
spat_point_border_stroke = 0.1,
spat_edge_alpha = NULL,
cell_color_gradient = c("blue", "white", "red"),
gradient_midpoint = NULL,
gradient_limits = NULL,
cow_n_col = 2,
cow_rel_h = 1,
cow_rel_w = 1,
cow_align = "h",
show_legend = T,
legend_text = 8,
dim_background_color = "white",
spat_background_color = "white",
vor_border_color = "white",
vor_max_radius = 200,
vor_alpha = 1,
axis_text = 8,
axis_title = 8,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spatDimGenePlot2D"
)

```

**Arguments**

<code>gobject</code>	giotto object
<code>show_image</code>	show a tissue background image
<code>gimage</code>	a giotto image
<code>image_name</code>	name of a giotto image
<code>expression_values</code>	gene expression values to use
<code>plot_alignment</code>	direction to align plot
<code>genes</code>	genes to show
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>dim_point_shape</code>	dim reduction points with border or not (border or no_border)
<code>dim_point_size</code>	dim reduction plot: point size
<code>dim_point_alpha</code>	transparency of dim. reduction points
<code>dim_point_border_col</code>	color of border around points
<code>dim_point_border_stroke</code>	stroke size of border around points
<code>show_NN_network</code>	show underlying NN network
<code>show_spatial_network</code>	show underlying spatial network
<code>dim_network_color</code>	color of NN network
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use, if <code>show_NN_network = TRUE</code>
<code>dim_edge_alpha</code>	dim reduction plot: column to use for alpha of the edges
<code>scale_alpha_with_expression</code>	scale expression with ggplot alpha parameter
<code>sdimx</code>	spatial x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	spatial y-axis dimension name (default = 'sdimy')
<code>spatial_network_name</code>	name of spatial network to use
<code>spatial_network_color</code>	color of spatial network
<code>show_spatial_grid</code>	show spatial grid
<code>grid_color</code>	color of spatial grid

spatial_grid_name	name of spatial grid to use
spat_point_shape	spatial points with border or not (border or no_border)
spat_point_size	spatial plot: point size
spat_point_alpha	transparancy of spatial points
spat_point_border_col	color of border around points
spat_point_border_stroke	stroke size of border around points
spat_edge_alpha	edge alpha
cell_color_gradient	vector with 3 colors for numeric data
gradient_midpoint	midpoint for color gradient
gradient_limits	vector with lower and upper limits
cow_n_col	cowplot param: how many columns
cow_rel_h	cowplot param: relative height
cow_rel_w	cowplot param: relative width
cow_align	cowplot param: how to align
show_legend	show legend
legend_text	size of legend text
dim_background_color	color of plot background for dimension plot
spat_background_color	color of plot background for spatial plot
vor_border_color	border color for voronoi plot
vor_max_radius	maximum radius for voronoi 'cells'
vor_alpha	transparancy of voronoi 'cells'
axis_text	size of axis text
axis_title	size of axis title
show_plot	show plots
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

## Details

Description of parameters.

**Value**

ggplot

**See Also**

[spatDimGenePlot3D](#)

Other spatial and dimension reduction gene expression visualizations: [spatDimGenePlot3D\(\)](#), [spatDimGenePlot\(\)](#)

**Examples**

```
spatDimGenePlot2D(gobject)
```

---

spatDimGenePlot3D	<i>spatDimGenePlot3D</i>
-------------------	--------------------------

---

**Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

**Usage**

```
spatDimGenePlot3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("horizontal", "vertical"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  genes,
  cluster_column = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1.5,
  show_NN_network = FALSE,
  nn_network_to_use = "sNN",
  nn_network_color = "lightgrey",
  nn_network_alpha = 0.5,
  network_name = "sNN.pca",
  label_size = 16,
  genes_low_color = "blue",
  genes_mid_color = "white",
  genes_high_color = "red",
  dim_point_size = 3,
```

```

show_spatial_network = FALSE,
spatial_network_name = "Delaunay_network",
spatial_network_color = "lightgray",
spatial_network_alpha = 0.5,
show_spatial_grid = FALSE,
spatial_grid_name = "spatial_grid",
spatial_grid_color = NULL,
spatial_grid_alpha = 0.5,
spatial_point_size = 3,
legend_text_size = 12,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
z_ticks = NULL,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spatDimGenePlot3D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>plot_alignment</code>	direction to align plot
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>dim3_to_use</code>	dimension to use on z-axis
<code>sdimx</code>	spatial dimension to use on x-axis
<code>sdimy</code>	spatial dimension to use on y-axis
<code>sdimz</code>	spatial dimension to use on z-axis
<code>genes</code>	genes to show
<code>cluster_column</code>	cluster column to select groups
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs
<code>show_other_cells</code>	display not selected cells
<code>other_cell_color</code>	color of not selected cells
<code>other_point_size</code>	size of not selected cells



show_NN_network	show underlying NN network
nn_network_to_use	type of NN network to use (kNN vs sNN)
nn_network_color	color of NN network
nn_network_alpha	alpha of NN network
network_name	name of NN network to use, if show_NN_network = TRUE
label_size	size of labels
genes_low_color	color for low expression levels
genes_mid_color	color for medium expression levels
genes_high_color	color for high expression levels
dim_point_size	dim reduction plot: point size
show_spatial_network	show spatial network (boolean)
spatial_network_name	name of spatial network to use
spatial_network_color	color of spatial network
spatial_network_alpha	alpha of spatial network
show_spatial_grid	show spatial grid (boolean)
spatial_grid_name	name of spatial grid to use
spatial_grid_color	color of spatial grid
spatial_grid_alpha	alpha of spatial grid
spatial_point_size	spatial plot: point size
legend_text_size	size of legend
axis_scale	the way to scale the axis
custom_ratio	customize the scale of the plot
x_ticks	set the number of ticks on the x-axis
y_ticks	set the number of ticks on the y-axis
z_ticks	set the number of ticks on the z-axis
show_plot	show plots
return_plot	return plotly object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param
edge_alpha_dim	dim reduction plot: column to use for alpha of the edges

Details

Description of parameters.

Value

plotly

See Also

Other spatial and dimension reduction gene expression visualizations: [spatDimGenePlot2D\(\)](#), [spatDimGenePlot\(\)](#)

---

spatDimPlot	<i>spatDimPlot</i>
-------------	--------------------

---

Description

Visualize cells according to spatial AND dimension reduction coordinates 2D

Usage

spatDimPlot(...)

Arguments

... Arguments passed on to [spatDimPlot2D](#)

`gobject` giotto object

`show_image` show a tissue background image

`gimage` a giotto image

`image_name` name of a giotto image

`plot_alignment` direction to align plot

`dim_reduction_to_use` dimension reduction to use

`dim_reduction_name` dimension reduction name

`dim1_to_use` dimension to use on x-axis

`dim2_to_use` dimension to use on y-axis

`sdimx` = spatial dimension to use on x-axis

`sdimy` = spatial dimension to use on y-axis

`spat_enr_names` names of spatial enrichment results to include

`cell_color` color for cells (see details)

`color_as_factor` convert color column to factor

`cell_color_code` named vector with colors

`cell_color_gradient` vector with 3 colors for numeric data

`gradient_midpoint` midpoint for color gradient

`gradient_limits` vector with lower and upper limits

`select_cell_groups` select subset of cells/clusters based on `cell_color` parameter

`select_cells` select subset of cells based on cell IDs

`dim_point_shape` point with border or not (border or `no_border`)

dim\_point\_size size of points in dim. reduction space  
 dim\_point\_alpha transparency of point in dim. reduction space  
 dim\_point\_border\_col border color of points in dim. reduction space  
 dim\_point\_border\_stroke border stroke of points in dim. reduction space  
 spat\_point\_shape shape of points (border, no\_border or voronoi)  
 spat\_point\_size size of spatial points  
 spat\_point\_alpha transparency of spatial points  
 spat\_point\_border\_col border color of spatial points  
 spat\_point\_border\_stroke border stroke of spatial points  
 dim\_show\_cluster\_center show the center of each cluster  
 dim\_show\_center\_label provide a label for each cluster  
 dim\_center\_point\_size size of the center point  
 dim\_center\_point\_border\_col border color of center point  
 dim\_center\_point\_border\_stroke stroke size of center point  
 dim\_label\_size size of the center label  
 dim\_label\_fontface font of the center label  
 spat\_show\_cluster\_center show the center of each cluster  
 spat\_show\_center\_label provide a label for each cluster  
 spat\_center\_point\_size size of the center point  
 spat\_center\_point\_border\_col border color of spatial center points  
 spat\_center\_point\_border\_stroke border strike size of spatial center points  
 spat\_label\_size size of the center label  
 spat\_label\_fontface font of the center label  
 show\_NN\_network show underlying NN network  
 nn\_network\_to\_use type of NN network to use (kNN vs sNN)  
 network\_name name of NN network to use, if show\_NN\_network = TRUE  
 nn\_network\_alpha column to use for alpha of the edges  
 show\_spatial\_network show spatial network  
 spat\_network\_name name of spatial network to use  
 spat\_network\_color color of spatial network  
 spat\_network\_alpha alpha of spatial network  
 show\_spatial\_grid show spatial grid  
 spat\_grid\_name name of spatial grid to use  
 spat\_grid\_color color of spatial grid  
 show\_other\_cells display not selected cells  
 other\_cell\_color color of not selected cells  
 dim\_other\_point\_size size of not selected dim cells  
 spat\_other\_point\_size size of not selected spat cells  
 spat\_other\_cells\_alpha alpha of not selected spat cells  
 dim\_show\_legend show legend of dimension reduction plot  
 spat\_show\_legend show legend of spatial plot  
 legend\_text size of legend text  
 legend\_symbol\_size size of legend symbols  
 dim\_background\_color background color of points in dim. reduction space  
 spat\_background\_color background color of spatial points

vor\_border\_color border color for voronoi plot  
vor\_max\_radius maximum radius for voronoi 'cells'  
vor\_alpha transparency of voronoi 'cells'  
axis\_text size of axis text  
axis\_title size of axis title  
show\_plot show plot  
return\_plot return ggplot object  
save\_plot directly save the plot [boolean]  
save\_param list of saving parameters, see [showSaveParameters](#)  
default\_save\_name default save name for saving, don't change, change save\_name in save\_param

**Details**

Description of parameters.

**Value**

ggplot

**See Also**

[spatDimPlot2D](#) and [spatDimPlot3D](#) for 3D visualization.  
Other spatial and dimension reduction visualizations: [spatDimPlot2D\(\)](#), [spatDimPlot3D\(\)](#)

**Examples**

spatDimPlot(gobject)

---

spatDimPlot2D	<i>spatDimPlot2D</i>
---------------	----------------------

---

**Description**

Visualize cells according to spatial AND dimension reduction coordinates 2D

**Usage**

```
spatDimPlot2D(  
  gobject,  
  show_image = F,  
  gimage = NULL,  
  image_name = "image",  
  plot_alignment = c("vertical", "horizontal"),  
  dim_reduction_to_use = "umap",  
  dim_reduction_name = "umap",  
  dim1_to_use = 1,  
  dim2_to_use = 2,  
  sdinx = "sdinx",  
  sdimy = "sdimy",
```

```
spat_enr_names = NULL,
cell_color = NULL,
color_as_factor = T,
cell_color_code = NULL,
cell_color_gradient = c("blue", "white", "red"),
gradient_midpoint = NULL,
gradient_limits = NULL,
select_cell_groups = NULL,
select_cells = NULL,
dim_point_shape = c("border", "no_border"),
dim_point_size = 1,
dim_point_alpha = 1,
dim_point_border_col = "black",
dim_point_border_stroke = 0.1,
spat_point_shape = c("border", "no_border", "voronoi"),
spat_point_size = 1,
spat_point_alpha = 1,
spat_point_border_col = "black",
spat_point_border_stroke = 0.1,
dim_show_cluster_center = F,
dim_show_center_label = T,
dim_center_point_size = 4,
dim_center_point_border_col = "black",
dim_center_point_border_stroke = 0.1,
dim_label_size = 4,
dim_label_fontface = "bold",
spat_show_cluster_center = F,
spat_show_center_label = F,
spat_center_point_size = 4,
spat_center_point_border_col = "blue",
spat_center_point_border_stroke = 0.1,
spat_label_size = 4,
spat_label_fontface = "bold",
show_NN_network = F,
nn_network_to_use = "sNN",
network_name = "sNN.pca",
nn_network_alpha = 0.05,
show_spatial_network = F,
spat_network_name = "Delaunay_network",
spat_network_color = "blue",
spat_network_alpha = 0.5,
show_spatial_grid = F,
spat_grid_name = "spatial_grid",
spat_grid_color = "blue",
show_other_cells = T,
other_cell_color = "lightgrey",
dim_other_point_size = 1,
spat_other_point_size = 1,
spat_other_cells_alpha = 0.5,
dim_show_legend = F,
spat_show_legend = F,
legend_text = 8,
```

```

    legend_symbol_size = 1,
    dim_background_color = "white",
    spat_background_color = "white",
    vor_border_color = "white",
    vor_max_radius = 200,
    vor_alpha = 1,
    axis_text = 8,
    axis_title = 8,
    show_plot = NA,
    return_plot = NA,
    save_plot = NA,
    save_param = list(),
    default_save_name = "spatDimPlot2D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>show_image</code>	show a tissue background image
<code>gimage</code>	a giotto image
<code>image_name</code>	name of a giotto image
<code>plot_alignment</code>	direction to align plot
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>sdimx</code>	= spatial dimension to use on x-axis
<code>sdimy</code>	= spatial dimension to use on y-axis
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>cell_color</code>	color for cells (see details)
<code>color_as_factor</code>	convert color column to factor
<code>cell_color_code</code>	named vector with colors
<code>cell_color_gradient</code>	vector with 3 colors for numeric data
<code>gradient_midpoint</code>	midpoint for color gradient
<code>gradient_limits</code>	vector with lower and upper limits
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs
<code>dim_point_shape</code>	point with border or not ( <code>border</code> or <code>no_border</code> )
<code>dim_point_size</code>	size of points in dim. reduction space

```

dim_point_alpha
    transparency of point in dim. reduction space
dim_point_border_col
    border color of points in dim. reduction space
dim_point_border_stroke
    border stroke of points in dim. reduction space
spat_point_shape
    shape of points (border, no_border or voronoi)
spat_point_size
    size of spatial points
spat_point_alpha
    transparency of spatial points
spat_point_border_col
    border color of spatial points
spat_point_border_stroke
    border stroke of spatial points
dim_show_cluster_center
    show the center of each cluster
dim_show_center_label
    provide a label for each cluster
dim_center_point_size
    size of the center point
dim_center_point_border_col
    border color of center point
dim_center_point_border_stroke
    stroke size of center point
dim_label_size
    size of the center label
dim_label_fontface
    font of the center label
spat_show_cluster_center
    show the center of each cluster
spat_show_center_label
    provide a label for each cluster
spat_center_point_size
    size of the center point
spat_center_point_border_col
    border color of spatial center points
spat_center_point_border_stroke
    border strike size of spatial center points
spat_label_size
    size of the center label
spat_label_fontface
    font of the center label
show_NN_network
    show underlying NN network
nn_network_to_use
    type of NN network to use (kNN vs sNN)
network_name
    name of NN network to use, if show_NN_network = TRUE

```

nn_network_alpha	column to use for alpha of the edges
show_spatial_network	show spatial network
spat_network_name	name of spatial network to use
spat_network_color	color of spatial network
spat_network_alpha	alpha of spatial network
show_spatial_grid	show spatial grid
spat_grid_name	name of spatial grid to use
spat_grid_color	color of spatial grid
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
dim_other_point_size	size of not selected dim cells
spat_other_point_size	size of not selected spat cells
spat_other_cells_alpha	alpha of not selected spat cells
dim_show_legend	show legend of dimension reduction plot
spat_show_legend	show legend of spatial plot
legend_text	size of legend text
legend_symbol_size	size of legend symbols
dim_background_color	background color of points in dim. reduction space
spat_background_color	background color of spatial points
vor_border_color	border color for voronoi plot
vor_max_radius	maximum radius for voronoi 'cells'
vor_alpha	transparency of voronoi 'cells'
axis_text	size of axis text
axis_title	size of axis title
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param



Details

Description of parameters.

Value

ggplot

See Also

[spatDimPlot3D](#)

Other spatial and dimension reduction visualizations: [spatDimPlot3D\(\)](#), [spatDimPlot\(\)](#)

Examples

```
spatDimPlot2D(gobject)
```

---

spatDimPlot3D	<i>spatDimPlot3D</i>
---------------	----------------------

---

Description

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

Usage

```
spatDimPlot3D(  
  gobject,  
  plot_alignment = c("horizontal", "vertical"),  
  dim_reduction_to_use = "umap",  
  dim_reduction_name = "umap",  
  dim1_to_use = 1,  
  dim2_to_use = 2,  
  dim3_to_use = 3,  
  sdimx = "sdimx",  
  sdimy = "sdimy",  
  sdimz = "sdimz",  
  spat_enr_names = NULL,  
  show_NN_network = FALSE,  
  nn_network_to_use = "sNN",  
  network_name = "sNN.pca",  
  nn_network_color = "lightgray",  
  nn_network_alpha = 0.5,  
  show_cluster_center = F,  
  show_center_label = T,  
  center_point_size = 4,  
  label_size = 16,  
  select_cell_groups = NULL,  
  select_cells = NULL,  
  show_other_cells = T,  
  other_cell_color = "lightgrey",  
  other_point_size = 1.5,
```

```

cell_color = NULL,
color_as_factor = T,
cell_color_code = NULL,
dim_point_size = 3,
show_spatial_network = F,
spatial_network_name = "Delaunay_network",
spatial_network_color = "lightgray",
spatial_network_alpha = 0.5,
show_spatial_grid = F,
spatial_grid_name = "spatial_grid",
spatial_grid_color = NULL,
spatial_grid_alpha = 0.5,
spatial_point_size = 3,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
z_ticks = NULL,
legend_text_size = 12,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spatDimPlot3D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>plot_alignment</code>	direction to align plot
<code>dim_reduction_to_use</code>	dimension reduction to use
<code>dim_reduction_name</code>	dimension reduction name
<code>dim1_to_use</code>	dimension to use on x-axis
<code>dim2_to_use</code>	dimension to use on y-axis
<code>dim3_to_use</code>	dimension to use on z-axis
<code>sdimx</code>	= spatial dimension to use on x-axis
<code>sdimy</code>	= spatial dimension to use on y-axis
<code>sdimz</code>	= spatial dimension to use on z-axis
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>show_NN_network</code>	show underlying NN network
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use, if <code>show_NN_network = TRUE</code>
<code>nn_network_color</code>	color of nn network
<code>nn_network_alpha</code>	column to use for alpha of the edges

show_cluster_center	show the center of each cluster
show_center_label	provide a label for each cluster
center_point_size	size of the center point
label_size	size of the center label
select_cell_groups	select subset of cells/clusters based on cell_color parameter
select_cells	select subset of cells based on cell IDs
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	size of not selected cells
cell_color	color for cells (see details)
color_as_factor	convert color column to factor
cell_color_code	named vector with colors
dim_point_size	size of points in dim. reduction space
show_spatial_network	show spatial network
spatial_network_name	name of spatial network to use
spatial_network_color	color of spatial network
spatial_network_alpha	alpha of spatial network
show_spatial_grid	show spatial grid
spatial_grid_name	name of spatial grid to use
spatial_grid_color	color of spatial grid
spatial_point_size	size of spatial points
axis_scale	the way to scale the axis
custom_ratio	customize the scale of the plot
x_ticks	set the number of ticks on the x-axis
y_ticks	set the number of ticks on the y-axis
z_ticks	set the number of ticks on the z-axis
legend_text_size	size of legend
show_plot	show plot
return_plot	return ggplot object

save\_plot            directly save the plot [boolean]  
save\_param          list of saving parameters, see [showSaveParameters](#)  
default\_save\_name        default save name for saving, don't change, change save\_name in save\_param

**Details**

Description of parameters.

**Value**

plotly

**See Also**

Other spatial and dimension reduction visualizations: [spatDimPlot2D\(\)](#), [spatDimPlot\(\)](#)

**Examples**

spatDimPlot3D(gobject)

---

spatGenePlot	<i>spatGenePlot</i>
--------------	---------------------

---

**Description**

Visualize cells and gene expression according to spatial coordinates

**Usage**

spatGenePlot(...)

**Arguments**

...            Arguments passed on to [spatGenePlot2D](#)  
gobject    giotto object  
show\_image    show a tissue background image  
gimage    a giotto image  
image\_name    name of a giotto image  
sdimx    x-axis dimension name (default = 'sdimx')  
sdimy    y-axis dimension name (default = 'sdimy')  
expression\_values    gene expression values to use  
genes    genes to show  
cell\_color\_gradient    vector with 3 colors for numeric data  
gradient\_midpoint    midpoint for color gradient  
gradient\_limits    vector with lower and upper limits  
show\_network    show underlying spatial network  
network\_color    color of spatial network  
spatial\_network\_name    name of spatial network to use

edge\_alpha alpha of edge  
 show\_grid show spatial grid  
 grid\_color color of spatial grid  
 spatial\_grid\_name name of spatial grid to use  
 midpoint expression midpoint  
 scale\_alpha\_with\_expression scale expression with ggplot alpha parameter  
 point\_shape shape of points (border, no\_border or voronoi)  
 point\_size size of point (cell)  
 point\_alpha transparency of points  
 point\_border\_col color of border around points  
 point\_border\_stroke stroke size of border around points  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_legend show legend  
 legend\_text size of legend text  
 background\_color color of plot background  
 vor\_border\_color border color for voronoi plot  
 vor\_max\_radius maximum radius for voronoi 'cells'  
 vor\_alpha transparency of voronoi 'cells'  
 axis\_text size of axis text  
 axis\_title size of axis title  
 show\_plot show plots  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)  
 default\_save\_name default save name for saving, don't change, change save\_name in save\_param

## Details

Description of parameters.

## Value

ggplot

## See Also

[spatGenePlot3D](#) and [spatGenePlot2D](#)

Other spatial gene expression visualizations: [spatGenePlot2D\(\)](#), [spatGenePlot3D\(\)](#)

## Examples

```
spatGenePlot(gobject)
```

---

spatGenePlot2D

*spatGenePlot2D*


---

## Description

Visualize cells and gene expression according to spatial coordinates

## Usage

```
spatGenePlot2D(
  gobject,
  show_image = F,
  gimage = NULL,
  image_name = "image",
  sdimx = "sdimx",
  sdimy = "sdimy",
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  show_network = F,
  network_color = NULL,
  spatial_network_name = "Delaunay_network",
  edge_alpha = NULL,
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  midpoint = 0,
  scale_alpha_with_expression = FALSE,
  point_shape = c("border", "no_border", "voronoi"),
  point_size = 1,
  point_alpha = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  legend_text = 8,
  background_color = "white",
  vor_border_color = "white",
  vor_alpha = 1,
  vor_max_radius = 200,
  axis_text = 8,
  axis_title = 8,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
```

```

    default_save_name = "spatGenePlot2D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>show_image</code>	show a tissue background image
<code>gimage</code>	a giotto image
<code>image_name</code>	name of a giotto image
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>expression_values</code>	gene expression values to use
<code>genes</code>	genes to show
<code>cell_color_gradient</code>	vector with 3 colors for numeric data
<code>gradient_midpoint</code>	midpoint for color gradient
<code>gradient_limits</code>	vector with lower and upper limits
<code>show_network</code>	show underlying spatial network
<code>network_color</code>	color of spatial network
<code>spatial_network_name</code>	name of spatial network to use
<code>edge_alpha</code>	alpha of edge
<code>show_grid</code>	show spatial grid
<code>grid_color</code>	color of spatial grid
<code>spatial_grid_name</code>	name of spatial grid to use
<code>midpoint</code>	expression midpoint
<code>scale_alpha_with_expression</code>	scale expression with ggplot alpha parameter
<code>point_shape</code>	shape of points (border, no_border or voronoi)
<code>point_size</code>	size of point (cell)
<code>point_alpha</code>	transparency of points
<code>point_border_col</code>	color of border around points
<code>point_border_stroke</code>	stroke size of border around points
<code>show_legend</code>	show legend
<code>legend_text</code>	size of legend text
<code>background_color</code>	color of plot background
<code>vor_border_color</code>	border color for voronoi plot

vor_alpha	transparency of voronoi 'cells'
vor_max_radius	maximum radius for voronoi 'cells'
axis_text	size of axis text
axis_title	size of axis title
cow_n_col	cowplot param: how many columns
cow_rel_h	cowplot param: relative height
cow_rel_w	cowplot param: relative width
cow_align	cowplot param: how to align
show_plot	show plots
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

**Details**

Description of parameters.

**Value**

ggplot

**See Also**

[spatGenePlot3D](#)  
Other spatial gene expression visualizations: [spatGenePlot3D\(\)](#), [spatGenePlot\(\)](#)

**Examples**

spatGenePlot2D(gobject)

---

spatGenePlot3D	<i>spatGenePlot3D</i>
----------------	-----------------------

---

**Description**

Visualize cells and gene expression according to spatial coordinates



**Usage**

```

spatGenePlot3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  show_network = FALSE,
  network_color = NULL,
  spatial_network_name = "Delaunay_network",
  edge_alpha = NULL,
  cluster_column = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_grid = FALSE,
  spatial_grid_name = "spatial_grid",
  point_size = 2,
  show_legend = TRUE,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "spatGenePlot3D"
)

```

**Arguments**

<code>gobject</code>	giotto object
<code>expression_values</code>	gene expression values to use
<code>genes</code>	genes to show
<code>show_network</code>	show underlying spatial network
<code>network_color</code>	color of spatial network
<code>spatial_network_name</code>	name of spatial network to use
<code>edge_alpha</code>	alpha of edges
<code>cluster_column</code>	cluster column to select groups
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs

show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	size of not selected cells
genes_high_color	color represents high gene expression
genes_mid_color	color represents middle gene expression
genes_low_color	color represents low gene expression
show_grid	show spatial grid
spatial_grid_name	name of spatial grid to use
point_size	size of point (cell)
show_legend	show legend
axis_scale	the way to scale the axis
custom_ratio	customize the scale of the plot
x_ticks	set the number of ticks on the x-axis
y_ticks	set the number of ticks on the y-axis
z_ticks	set the number of ticks on the z-axis
show_plot	show plots
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param
grid_color	color of spatial grid

## Details

Description of parameters.

## Value

ggplot

## See Also

Other spatial gene expression visualizations: [spatGenePlot2D\(\)](#), [spatGenePlot\(\)](#)

## Examples

```
spatGenePlot3D(gobject)
```

---

spatialAEH	<i>spatialAEH</i>
------------	-------------------

---

**Description**

Compute spatial variable genes with spatialDE method

**Usage**

```
spatialAEH(  
  gobject = NULL,  
  SpatialDE_results = NULL,  
  name_pattern = "AEH_patterns",  
  expression_values = c("raw", "normalized", "scaled", "custom"),  
  pattern_num = 6,  
  l = 1.05,  
  python_path = NULL,  
  return_gobject = TRUE  
)
```

**Arguments**

- gobject            Giotto object
- SpatialDE\_results       results of [SpatialDE](#) function
- name\_pattern      name for the computed spatial patterns
- expression\_values    gene expression values to use
- pattern\_num       number of spatial patterns to look for
- l                  lengthscale
- python\_path       specify specific path to python if required
- return\_gobject    show plot

**Details**

This function is a wrapper for the SpatialAEH method implemented in the ...

**Value**

An updated giotto object

spatialDE

*spatialDE***Description**

Compute spatial variable genes with spatialDE method

**Usage**

```
spatialDE(
  gobject = NULL,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  size = c(4, 2, 1),
  color = c("blue", "green", "red"),
  sig_alpha = 0.5,
  unsig_alpha = 0.5,
  python_path = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "SpatialDE"
)
```

**Arguments**

<code>gobject</code>	Giotto object
<code>expression_values</code>	gene expression values to use
<code>size</code>	size of plot
<code>color</code>	low/medium/high color scheme for plot
<code>sig_alpha</code>	alpha value for significance
<code>unsig_alpha</code>	alpha value for unsignificance
<code>python_path</code>	specify specific path to python if required
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters, see <a href="#">showSaveParameters</a>
<code>default_save_name</code>	default save name for saving, don't change, change save_name in save_param

**Details**

This function is a wrapper for the SpatialDE method implemented in the ...

**Value**

a list of data.frames with results and plot (optional)

---

spatNetwDistributions    *spatNetwDistributionsDistance*


---

## Description

This function return histograms displaying the distance distribution for each spatial k-neighbor

## Usage

```
spatNetwDistributions(
  gobject,
  spatial_network_name = "spatial_network",
  distribution = c("distance", "k_neighbors"),
  hist_bins = 30,
  test_distance_limit = NULL,
  ncol = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "spatNetwDistributions"
)
```

## Arguments

<code>gobject</code>	Giotto object
<code>spatial_network_name</code>	name of spatial network
<code>distribution</code>	show the distribution of cell-to-cell distance or number of k neighbors
<code>hist_bins</code>	number of binds to use for the histogram
<code>test_distance_limit</code>	effect of different distance threshold on k-neighbors
<code>ncol</code>	number of columns to visualize the histograms in
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, alternatively change <code>save_name</code> in <code>save_param</code>

## Details

The **distance** option shows the spatial distance distribution for each nearest neighbor rank (1st, 2nd, 3th, ... neighbor). With this option the user can also test the effect of a distance limit on the spatial network. This distance limit can be used to remove neighbor cells that are considered to far away. The **k\_neighbors** option shows the number of k neighbors distribution over all cells.

**Value**

ggplot plot

**Examples**

```
spatNetwDistributionsDistance(gobject)
```

---

```
spatNetwDistributionsDistance
```

```
spatNetwDistributionsDistance
```

---

**Description**

This function return histograms displaying the distance distribution for each spatial k-neighbor

**Usage**

```
spatNetwDistributionsDistance(
  gobject,
  spatial_network_name = "spatial_network",
  hist_bins = 30,
  test_distance_limit = NULL,
  ncol = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "spatNetwDistributionsDistance"
)
```

**Arguments**

<code>gobject</code>	Giotto object
<code>spatial_network_name</code>	name of spatial network
<code>hist_bins</code>	number of binds to use for the histogram
<code>test_distance_limit</code>	effect of different distance threshold on k-neighbors
<code>ncol</code>	number of columns to visualize the histograms in
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, alternatively change <code>save_name</code> in <code>save_param</code>

**Value**

ggplot plot

## Examples

```
spatNetwDistributionsDistance(gobject)
```

---

```
spatNetwDistributionsKneighbors  
    spatNetwDistributionsKneighbors
```

---

## Description

This function returns a histogram displaying the number of k-neighbors distribution for each cell

## Usage

```
spatNetwDistributionsKneighbors(  
  gobject,  
  spatial_network_name = "spatial_network",  
  hist_bins = 30,  
  show_plot = NA,  
  return_plot = NA,  
  save_plot = NA,  
  save_param = list(),  
  default_save_name = "spatNetwDistributionsKneighbors"  
)
```

## Arguments

<code>gobject</code>	Giotto object
<code>spatial_network_name</code>	name of spatial network
<code>hist_bins</code>	number of binds to use for the histogram
<code>show_plot</code>	show plot
<code>return_plot</code>	return ggplot object
<code>save_plot</code>	directly save the plot [boolean]
<code>save_param</code>	list of saving parameters from <a href="#">all_plots_save_function</a>
<code>default_save_name</code>	default save name for saving, alternatively change <code>save_name</code> in <code>save_param</code>

## Value

ggplot plot

## Examples

```
spatNetwDistributionsKneighbors(gobject)
```

---

spatPlot

*spatPlot*


---

## Description

Visualize cells according to spatial coordinates

## Usage

```
spatPlot(...)
```

## Arguments

```
... Arguments passed on to spatPlot2D
gobject giotto object
show_image show a tissue background image
gimage a giotto image
image_name name of a giotto image
group_by create multiple plots based on cell annotation column
group_by_subset subset the group_by factor column
sdimx x-axis dimension name (default = 'sdimx')
sdimy y-axis dimension name (default = 'sdimy')
spat_enr_names names of spatial enrichment results to include
cell_color color for cells (see details)
color_as_factor convert color column to factor
cell_color_code named vector with colors
cell_color_gradient vector with 3 colors for numeric data
gradient_midpoint midpoint for color gradient
gradient_limits vector with lower and upper limits
select_cell_groups select subset of cells/clusters based on cell_color parameter
select_cells select subset of cells based on cell IDs
point_shape shape of points (border, no_border or voronoi)
point_size size of point (cell)
point_alpha transparency of point
point_border_col color of border around points
point_border_stroke stroke size of border around points
show_cluster_center plot center of selected clusters
show_center_label plot label of selected clusters
center_point_size size of center points
center_point_border_col border color of center points
center_point_border_stroke border stroke size of center points
label_size size of labels
label_fontface font of labels
show_network show underlying spatial network
spatial_network_name name of spatial network to use
```



network\_color color of spatial network  
 network\_alpha alpha of spatial network  
 show\_grid show spatial grid  
 spatial\_grid\_name name of spatial grid to use  
 grid\_color color of spatial grid  
 show\_other\_cells display not selected cells  
 other\_cell\_color color of not selected cells  
 other\_point\_size point size of not selected cells  
 other\_cells\_alpha alpha of not selected cells  
 coord\_fix\_ratio fix ratio between x and y-axis  
 title title of plot  
 show\_legend show legend  
 legend\_text size of legend text  
 legend\_symbol\_size size of legend symbols  
 background\_color color of plot background  
 vor\_border\_color border color for voronoi plot  
 vor\_max\_radius maximum radius for voronoi 'cells'  
 vor\_alpha transparency of voronoi 'cells'  
 axis\_text size of axis text  
 axis\_title size of axis title  
 cow\_n\_col cowplot param: how many columns  
 cow\_rel\_h cowplot param: relative height  
 cow\_rel\_w cowplot param: relative width  
 cow\_align cowplot param: how to align  
 show\_plot show plot  
 return\_plot return ggplot object  
 save\_plot directly save the plot [boolean]  
 save\_param list of saving parameters, see [showSaveParameters](#)  
 default\_save\_name default save name for saving, don't change, change save\_name  
 in save\_param

## Details

Description of parameters.

## Value

ggplot

## See Also

[spatPlot3D](#)

Other spatial visualizations: [spatPlot2D\(\)](#), [spatPlot3D\(\)](#)

## Examples

```
spatPlot(gobject)
```

---

spatPlot2D

*spatPlot2D*


---

## Description

Visualize cells according to spatial coordinates

## Usage

```
spatPlot2D(
  gobject,
  show_image = F,
  gimage = NULL,
  image_name = "image",
  group_by = NULL,
  group_by_subset = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_shape = c("border", "no_border", "voronoi"),
  point_size = 3,
  point_alpha = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  show_network = F,
  spatial_network_name = "Delaunay_network",
  network_color = NULL,
  network_alpha = 1,
  show_grid = F,
  spatial_grid_name = "spatial_grid",
  grid_color = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  other_cells_alpha = 0.1,
  coord_fix_ratio = NULL,
```

```

    title = NULL,
    show_legend = T,
    legend_text = 8,
    legend_symbol_size = 1,
    background_color = "white",
    vor_border_color = "white",
    vor_max_radius = 200,
    vor_alpha = 1,
    axis_text = 8,
    axis_title = 8,
    cow_n_col = 2,
    cow_rel_h = 1,
    cow_rel_w = 1,
    cow_align = "h",
    show_plot = NA,
    return_plot = NA,
    save_plot = NA,
    save_param = list(),
    default_save_name = "spatPlot2D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>show_image</code>	show a tissue background image
<code>gimage</code>	a giotto image
<code>image_name</code>	name of a giotto image
<code>group_by</code>	create multiple plots based on cell annotation column
<code>group_by_subset</code>	subset the <code>group_by</code> factor column
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>cell_color</code>	color for cells (see details)
<code>color_as_factor</code>	convert color column to factor
<code>cell_color_code</code>	named vector with colors
<code>cell_color_gradient</code>	vector with 3 colors for numeric data
<code>gradient_midpoint</code>	midpoint for color gradient
<code>gradient_limits</code>	vector with lower and upper limits
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs
<code>point_shape</code>	shape of points (border, no_border or voronoi)
<code>point_size</code>	size of point (cell)

point_alpha	transparancy of point
point_border_col	color of border around points
point_border_stroke	stroke size of border around points
show_cluster_center	plot center of selected clusters
show_center_label	plot label of selected clusters
center_point_size	size of center points
center_point_border_col	border color of center points
center_point_border_stroke	border stroke size of center points
label_size	size of labels
label_fontface	font of labels
show_network	show underlying spatial network
spatial_network_name	name of spatial network to use
network_color	color of spatial network
network_alpha	alpha of spatial network
show_grid	show spatial grid
spatial_grid_name	name of spatial grid to use
grid_color	color of spatial grid
show_other_cells	display not selected cells
other_cell_color	color of not selected cells
other_point_size	point size of not selected cells
other_cells_alpha	alpha of not selected cells
coord_fix_ratio	fix ratio between x and y-axis
title	title of plot
show_legend	show legend
legend_text	size of legend text
legend_symbol_size	size of legend symbols
background_color	color of plot background
vor_border_color	border color for voronoi plot
vor_max_radius	maximum radius for voronoi 'cells'

vor_alpha	transparancy of voronoi 'cells'
axis_text	size of axis text
axis_title	size of axis title
cow_n_col	cowplot param: how many columns
cow_rel_h	cowplot param: relative height
cow_rel_w	cowplot param: relative width
cow_align	cowplot param: how to align
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

Details

Description of parameters.

Value

ggplot

See Also

[spatPlot3D](#)  
Other spatial visualizations: [spatPlot3D\(\)](#), [spatPlot\(\)](#)

Examples

spatPlot2D(gobject)

---

spatPlot3D	<i>spatPlot3D</i>
------------	-------------------

---

Description

Visualize cells according to spatial coordinates

Usage

```
spatPlot3D(  
  gobject,  
  sdimx = "sdimx",  
  sdimy = "sdimy",  
  sdimz = "sdimz",  
  spat_enr_names = NULL,  
  point_size = 3,  
  cell_color = NULL,
```

```

cell_color_code = NULL,
select_cell_groups = NULL,
select_cells = NULL,
show_other_cells = T,
other_cell_color = "lightgrey",
other_point_size = 0.5,
other_cell_alpha = 0.5,
show_network = F,
spatial_network_name = "Delaunay_network",
network_color = NULL,
network_alpha = 1,
show_grid = F,
spatial_grid_name = "spatial_grid",
grid_color = NULL,
grid_alpha = 1,
title = "",
show_legend = T,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
z_ticks = NULL,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spat3D"
)

```

### Arguments

<code>gobject</code>	giotto object
<code>sdimx</code>	x-axis dimension name (default = 'sdimx')
<code>sdimy</code>	y-axis dimension name (default = 'sdimy')
<code>sdimz</code>	z-axis dimension name (default = 'sdimy')
<code>spat_enr_names</code>	names of spatial enrichment results to include
<code>point_size</code>	size of point (cell)
<code>cell_color</code>	color for cells (see details)
<code>cell_color_code</code>	named vector with colors
<code>select_cell_groups</code>	select subset of cells/clusters based on <code>cell_color</code> parameter
<code>select_cells</code>	select subset of cells based on cell IDs
<code>show_other_cells</code>	display not selected cells
<code>other_cell_color</code>	color of not selected cells
<code>other_point_size</code>	size of not selected cells

other_cell_alpha	alpha of not selected cells
show_network	show underlying spatial network
spatial_network_name	name of spatial network to use
network_color	color of spatial network
network_alpha	opacity of spatial network
show_grid	show spatial grid
spatial_grid_name	name of spatial grid to use
grid_color	color of spatial grid
grid_alpha	opacity of spatial grid
title	title of plot
show_legend	show legend
axis_scale	the way to scale the axis
custom_ratio	customize the scale of the plot
x_ticks	set the number of ticks on the x-axis
y_ticks	set the number of ticks on the y-axis
z_ticks	set the number of ticks on the z-axis
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

**Value**

ggplot

**See Also**

Other spatial visualizations: [spatPlot2D\(\)](#), [spatPlot\(\)](#)

**Examples**

```
spatPlot3D(gobject)
```

---

```
specificCellCellcommunicationScores
      specificCellCellcommunicationScores
```

---

## Description

Specific Cell-Cell communication scores based on spatial expression of interacting cells

## Usage

```
specificCellCellcommunicationScores(
  gobject,
  spatial_network_name = "Delaunay_network",
  cluster_column = "cell_types",
  random_iter = 100,
  cell_type_1 = "astrocyte",
  cell_type_2 = "endothelial",
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  min_observations = 2,
  adjust_method = c("fdr", "bonferroni", "BH", "holm", "hochberg", "hommel", "BY",
    "none"),
  adjust_target = c("genes", "cells"),
  verbose = T
)
```

## Arguments

<code>gobject</code>	giotto object to use
<code>spatial_network_name</code>	spatial network to use for identifying interacting cells
<code>cluster_column</code>	cluster column with cell type information
<code>random_iter</code>	number of iterations
<code>cell_type_1</code>	first cell type
<code>cell_type_2</code>	second cell type
<code>gene_set_1</code>	first specific gene set from gene pairs
<code>gene_set_2</code>	second specific gene set from gene pairs
<code>log2FC_addendum</code>	addendum to add when calculating log2FC
<code>min_observations</code>	minimum number of interactions needed to be considered
<code>adjust_method</code>	which method to adjust p-values
<code>adjust_target</code>	adjust multiple hypotheses at the cell or gene level
<code>verbose</code>	verbose



**Details**

Statistical framework to identify if pairs of genes (such as ligand-receptor combinations) are expressed at higher levels than expected based on a reshuffled null distribution of gene expression values in cells that are spatially in proximity to eachother.. More details will follow soon.

**Value**

Cell-Cell communication scores for gene pairs based on spatial interaction

**Examples**

```
specificCellCellcommunicationScores(gobject)
```

---

```
split_dendrogram_in_two  
      split_dendrogram_in_two
```

---

**Description**

Merge selected clusters based on pairwise correlation scores and size of cluster.

**Usage**

```
split_dendrogram_in_two(dend)
```

**Arguments**

dend	dendrogram object
------	-------------------

**Value**

list of two dendrograms and height of node

**Examples**

```
split_dendrogram_in_two(dend)
```

---

```
stitchFieldCoordinates  
      stitchFieldCoordinates
```

---

**Description**

Helper function to stitch field coordinates together to form one complete picture

**Usage**

```

stitchFieldCoordinates(
  location_file,
  offset_file,
  cumulate_offset_x = F,
  cumulate_offset_y = F,
  field_col = "Field of View",
  X_coord_col = "X",
  Y_coord_col = "Y",
  reverse_final_x = F,
  reverse_final_y = T
)

```

**Arguments**

location\_file    location dataframe with X and Y coordinates

offset\_file      dataframe that describes the offset for each field (see details)

cumulate\_offset\_x  
                  (boolean) Do the x-axis offset values need to be cumulated?

cumulate\_offset\_y  
                  (boolean) Do the y-axis offset values need to be cumulated?

field\_col        column that indicates the field within the location\_file

X\_coord\_col     column that indicates the x coordinates

Y\_coord\_col     column that indicates the x coordinates

reverse\_final\_x  
                  (boolean) Do the final x coordinates need to be reversed?

reverse\_final\_y  
                  (boolean) Do the final y coordinates need to be reversed?

**Details**

Stitching of fields:

- 1. have cell locations: at least 3 columns: field, X, Y
- 2. create offset file: offset file has 3 columns: field, x\_offset, y\_offset
- 3. create new cell location file by stitching original cell locations with stitchFieldCoordinates
- 4. provide new cell location file to [createGiottoObject](#)

**Value**

Updated location dataframe with new X ['X\_final'] and Y ['Y\_final'] coordinates

**Examples**

```
stitchFieldCoordinates(gobject)
```

---

stitchTileCoordinates	<i>stitchTileCoordinates</i>
-----------------------	------------------------------

---

**Description**

Helper function to stitch tile coordinates together to form one complete picture

**Usage**

```
stitchTileCoordinates(location_file, Xtilespan, Ytilespan)
```

**Arguments**

- location\_file    location dataframe with X and Y coordinates
- Xtilespan        numerical value specifying the width of each tile
- Ytilespan        numerical value specifying the height of each tile

**Details**

...

**Examples**

```
stitchTileCoordinates(gobject)
```

---

subClusterCells	<i>subClusterCells</i>
-----------------	------------------------

---

**Description**

subcluster cells

**Usage**

```
subClusterCells(  
  gobject,  
  name = "sub_clus",  
  cluster_method = c("leiden", "louvain_community", "louvain_multinet"),  
  cluster_column = NULL,  
  selected_clusters = NULL,  
  hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values  
    = "normalized"),  
  hvg_min_perc_cells = 5,  
  hvg_mean_expr_det = 1,  
  use_all_genes_as_hvg = FALSE,  
  min_nr_of_hvg = 5,  
  pca_param = list(expression_values = "normalized", scale_unit = T),  
  nn_param = list(dimensions_to_use = 1:20),  
  k_neighbors = 10,  
  resolution = 1,
```

```

    n_iterations = 1000,
    gamma = 1,
    omega = 1,
    python_path = NULL,
    nn_network_to_use = "sNN",
    network_name = "sNN.pca",
    return_gobject = TRUE,
    verbose = T
)

```

### Arguments

<code>gobject</code>	giotto object
<code>name</code>	name for new clustering result
<code>cluster_method</code>	clustering method to use
<code>cluster_column</code>	cluster column to subcluster
<code>selected_clusters</code>	only do subclustering on these clusters
<code>hvg_param</code>	parameters for calculateHVG
<code>hvg_min_perc_cells</code>	threshold for detection in min percentage of cells
<code>hvg_mean_expr_det</code>	threshold for mean expression level in cells with detection
<code>use_all_genes_as_hvg</code>	forces all genes to be HVG and to be used as input for PCA
<code>min_nr_of_hvg</code>	minimum number of HVG, or all genes will be used as input for PCA
<code>pca_param</code>	parameters for runPCA
<code>nn_param</code>	parameters for parameters for createNearestNetwork
<code>k_neighbors</code>	number of k for createNearestNetwork
<code>resolution</code>	resolution
<code>n_iterations</code>	number of iterations to run the Leiden algorithm.
<code>gamma</code>	gamma
<code>omega</code>	omega
<code>python_path</code>	specify specific path to python if required
<code>nn_network_to_use</code>	type of NN network to use (kNN vs sNN)
<code>network_name</code>	name of NN network to use
<code>return_gobject</code>	boolean: return giotto object (default = TRUE)
<code>verbose</code>	verbose

### Details

This function performs subclustering on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do clustering

**Value**

giotto object with new subclusters appended to cell metadata

**See Also**

[doLouvainCluster\\_multinet](#), [doLouvainCluster\\_community](#) and [@seealso doLeidenCluster](#)

**Examples**

```
subClusterCells(gobject)
```

---

subsetGiotto	<i>subsetGiotto</i>
--------------	---------------------

---

**Description**

subsets Giotto object including previous analyses.

**Usage**

```
subsetGiotto(gobject, cell_ids = NULL, gene_ids = NULL, verbose = FALSE)
```

**Arguments**

gobject	giotto object
cell_ids	cell IDs to keep
gene_ids	gene IDs to keep
verbose	be verbose

**Value**

giotto object

**Examples**

```
subsetGiotto(gobject)
```

---

subsetGiottoLocs	<i>subsetGiottoLocs</i>
------------------	-------------------------

---

### Description

subsets Giotto object based on spatial locations

### Usage

```
subsetGiottoLocs(  
  gobject,  
  x_max = NULL,  
  x_min = NULL,  
  y_max = NULL,  
  y_min = NULL,  
  z_max = NULL,  
  z_min = NULL,  
  return_gobject = T,  
  verbose = FALSE  
)
```

### Arguments

<code>gobject</code>	giotto object
<code>x_max</code>	maximum x-coordinate
<code>x_min</code>	minimum x-coordinate
<code>y_max</code>	maximum y-coordinate
<code>y_min</code>	minimum y-coordinate
<code>z_max</code>	maximum z-coordinate
<code>z_min</code>	minimum z-coordinate
<code>return_gobject</code>	return Giotto object
<code>verbose</code>	be verbose

### Details

if `return_gobject = FALSE`, then a filtered combined metadata `data.table` will be returned

### Value

giotto object

### Examples

```
subsetGiottoLocs(gobject)
```

---

trendSceek	<i>trendSceek</i>
------------	-------------------

---

## Description

Compute spatial variable genes with trendsceek method

## Usage

```
trendSceek(  
  gobject,  
  expression_values = c("normalized", "raw"),  
  subset_genes = NULL,  
  nrand = 100,  
  ncores = 8,  
  ...  
)
```

## Arguments

<code>gobject</code>	Giotto object
<code>expression_values</code>	gene expression values to use
<code>subset_genes</code>	subset of genes to run trendsceek on
<code>nrand</code>	An integer specifying the number of random resamplings of the mark distribution as to create the null-distribution.
<code>ncores</code>	An integer specifying the number of cores to be used by BiocParallel
<code>...</code>	Additional parameters to the <a href="#">trendsceek_test</a> function

## Details

This function is a wrapper for the `trendsceek_test` method implemented in the `trendsceek` package

## Value

data.frame with trendsceek spatial genes results

## Examples

```
trendSceek(gobject)
```

---

updateGiottoImage	<i>updateGiottoImage</i>
-------------------	--------------------------

---

### Description

Updates the boundaries of a giotto image attached to a giotto object

### Usage

```
updateGiottoImage(
  gobject,
  image_name,
  xmax_adj = 0,
  xmin_adj = 0,
  ymax_adj = 0,
  ymin_adj = 0,
  return_gobject = TRUE
)
```

### Arguments

gobject	giotto object
image_name	spatial locations
xmax_adj	adjustment of the maximum x-value to align the image
xmin_adj	adjustment of the minimum x-value to align the image
ymax_adj	adjustment of the maximum y-value to align the image
ymin_adj	adjustment of the minimum y-value to align the image
return_gobject	return a giotto object

### Value

a giotto object or an updated giotto image if return\_gobject = F

### Examples

```
updateGiottoImage(gobject)
```

---

viewHMRResults	<i>viewHMRResults</i>
----------------	-----------------------

---

### Description

View results from doHMRF.



**Usage**

```
viewHMRResults(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = FALSE,
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>HMRFoutput</code>	HMRF output from doHMRF
<code>k</code>	number of HMRF domains
<code>betas_to_view</code>	results from different betas that you want to view
<code>third_dim</code>	3D data (boolean)
<code>...</code>	additional paramters (see details)

**Value**

spatial plots with HMRF domains

**See Also**

[spatPlot2D](#) and [spatPlot3D](#)

---

viewHMRResults2D	<i>viewHMRResults2D</i>
------------------	-------------------------

---

**Description**

View results from doHMRF.

**Usage**

```
viewHMRResults2D(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>HMRFoutput</code>	HMRF output from doHMRF
<code>k</code>	number of HMRF domains
<code>betas_to_view</code>	results from different betas that you want to view
<code>...</code>	paramters to visPlot()

**Details**

Description ...

**Value**

spatial plots with HMRF domains

**See Also**

[spatPlot2D](#)

**Examples**

```
viewHMRFresults2D(gobject)
```

---

<code>viewHMRFresults3D</code>	<i>viewHMRFresults3D</i>
--------------------------------	--------------------------

---

**Description**

View results from doHMRF.

**Usage**

```
viewHMRFresults3D(  
  gobject,  
  HMRFoutput,  
  k = NULL,  
  betas_to_view = NULL,  
  third_dim = NULL,  
  ...  
)
```

**Arguments**

<code>gobject</code>	giotto object
<code>HMRFoutput</code>	HMRF output from doHMRF
<code>k</code>	number of HMRF domains
<code>betas_to_view</code>	results from different betas that you want to view
<code>...</code>	paramters to visPlot()

Details

Description ...

Value

spatial plots with HMRF domains

See Also

[spatPlot3D](#)

Examples

viewHMRFresults3D(gobject)

---

violinPlot	<i>violinPlot</i>
------------	-------------------

---

Description

Creates violinplot for selected clusters

Usage

```
violinPlot(  
  gobject,  
  expression_values = c("normalized", "scaled", "custom"),  
  genes,  
  cluster_column,  
  cluster_custom_order = NULL,  
  color_violin = c("genes", "cluster"),  
  cluster_color_code = NULL,  
  strip_position = c("top", "right", "left", "bottom"),  
  strip_text = 7,  
  axis_text_x_size = 10,  
  axis_text_y_size = 6,  
  show_plot = NA,  
  return_plot = NA,  
  save_plot = NA,  
  save_param = list(),  
  default_save_name = "violinPlot"  
)
```

Arguments

gobject	giotto object
expression_values	expression values to use
genes	genes to plot
cluster_column	name of column to use for clusters

cluster_custom_order	custom order of clusters
color_violin	color violin according to genes or clusters
cluster_color_code	color code for clusters
strip_position	position of gene labels
strip_text	size of strip text
axis_text_x_size	size of x-axis text
axis_text_y_size	size of y-axis text
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters, see <a href="#">showSaveParameters</a>
default_save_name	default save name for saving, don't change, change save_name in save_param

Value

ggplot

Examples

violinPlot(gobject)

---

writeHMRResults	<i>writeHMRResults</i>
-----------------	------------------------

---

Description

write results from doHMRF to a data.table.

Usage

```
writeHMRResults(  
  gobject,  
  HMRFoutput,  
  k = NULL,  
  betas_to_view = NULL,  
  print_command = F  
)
```

Arguments

gobject	giotto object
HMRFoutput	HMRF output from doHMRF
k	k to write results for
betas_to_view	results from different betas that you want to view
print_command	see the python command

**Value**

data.table with HMRF results for each b and the selected k

**Examples**

```
writeHMRFresults(gobject)
```

---

```
write_giotto_viewer_annotation
      write_giotto_viewer_annotation
```

---

**Description**

write out factor-like annotation data from a giotto object for the Viewer

**Usage**

```
write_giotto_viewer_annotation(
  annotation,
  annot_name = "test",
  output_directory = getwd()
)
```

**Arguments**

annotation	annotation from the data.table from giotto object
annot_name	name of the annotation
output_directory	directory where to save the files

**Value**

write a .txt and .annot file for the selection annotation

---

```
write_giotto_viewer_dim_reduction
      write_giotto_viewer_dim_reduction
```

---

**Description**

write out dimensional reduction data from a giotto object for the Viewer

**Usage**

```
write_giotto_viewer_dim_reduction(
  dim_reduction_cell,
  dim_red = NULL,
  dim_red_name = NULL,
  dim_red_rounding = NULL,
  dim_red_rescale = c(-20, 20),
  output_directory = getwd()
)
```

**Arguments**

dim_reduction_cell	dimension reduction slot from giotto object
dim_red	high level name of dimension reduction
dim_red_name	specific name of dimension reduction to use
dim_red_rounding	numerical indicating how to round the coordinates
dim_red_rescale	numericals to rescale the coordinates
output_directory	directory where to save the files

**Value**

write a .txt and .annot file for the selection annotation

---

```
write_giotto_viewer_numeric_annotation
      write_giotto_viewer_numeric_annotation
```

---

**Description**

write out numeric annotation data from a giotto object for the Viewer

**Usage**

```
write_giotto_viewer_numeric_annotation(
  annotation,
  annot_name = "test",
  output_directory = getwd()
)
```

**Arguments**

annotation	annotation from the data.table from giotto object
annot_name	name of the annotation
output_directory	directory where to save the files

**Value**

write a .txt and .annot file for the selection annotation

# Index

- \*Topic **giotto**,
  - giotto-class, [136](#)
  - print.giotto, [187](#)
  - show,giotto-method, [206](#)
- \*Topic **giotto**
  - createGiottoObject, [51](#)
- \*Topic **object**
  - giotto-class, [136](#)
  - print.giotto, [187](#)
  - show,giotto-method, [206](#)
- [addCellIntMetadata, 7](#)
- [addCellMetadata, 8, 52](#)
- [addCellStatistics, 9, 15](#)
- [addGeneMetadata, 10, 52](#)
- [addGenesPerc, 10](#)
- [addGeneStatistics, 11, 15](#)
- [addGiottoImage, 12](#)
- [addGiottoImageToSpatPlot, 12](#)
- [addHMRF, 13](#)
- [addNetworkLayout, 14](#)
- [addStatistics, 15](#)
- [adjustGiottoMatrix, 15](#)
- [all\\_plots\\_save\\_function, 16, 23, 26, 28, 29, 31, 33, 35, 115, 117, 140, 150–152, 154–157, 159, 162, 163, 173, 174, 216, 261–263](#)
- [annotateGiotto, 18](#)
- [annotateSpatialGrid, 19](#)
- [annotateSpatialNetwork, 19](#)
- [average\\_gene\\_gene\\_expression\\_in\\_groups, 20](#)
- [binSpect, 21](#)
- [calculateHVG, 22, 196, 201, 202](#)
- [calculateMetaTable, 24](#)
- [calculateMetaTableCells, 25](#)
- [cellProximityBarplot, 25](#)
- [cellProximityEnrichment, 26](#)
- [cellProximityHeatmap, 27](#)
- [cellProximityNetwork, 28](#)
- [cellProximitySpatPlot, 30](#)
- [cellProximitySpatPlot2D, 30, 31, 31](#)
- [cellProximitySpatPlot3D, 31, 33](#)
- [cellProximityVisPlot, 35](#)
- [changeGiottoInstructions, 38](#)
- [changeImageBg, 38](#)
- [cluster\\_walktrap, 108](#)
- [clusterCells, 39](#)
- [clusterSpatialCorGenes, 42](#)
- [combCCcom, 42, 173–175](#)
- [combineCellProximityGenes, 43](#)
- [combineCPG, 44](#)
- [combineMetadata, 45](#)
- [convert\\_mgImage\\_to\\_array\\_DT, 47](#)
- [convertEnsemblToGeneSymbol, 46](#)
- [create\\_average\\_detection\\_DT, 64](#)
- [create\\_average\\_DT, 64](#)
- [create\\_cell\\_type\\_random\\_cell\\_IDs, 65](#)
- [create\\_crossSection\\_object, 66](#)
- [create\\_screepilot, 67](#)
- [createCrossSection, 47](#)
- [createGiottoImage, 12, 49](#)
- [createGiottoInstructions, 50, 52, 53](#)
- [createGiottoObject, 51, 274](#)
- [createGiottoVisiumObject, 53](#)
- [createHeatmap\\_DT, 54](#)
- [createMetagenes, 55](#)
- [createNearestNetwork, 56](#)
- [createSpatialDefaultGrid, 57, 61](#)
- [createSpatialDelaunayNetwork, 58](#)
- [createSpatialEnrich, 60, 111](#)
- [createSpatialGrid, 60](#)
- [createSpatialKNNnetwork, 61](#)
- [createSpatialNetwork, 62, 72](#)
- [crossSectionGenePlot, 67](#)
- [crossSectionGenePlot3D, 68](#)
- [crossSectionPlot, 69, 70](#)
- [crossSectionPlot3D, 70](#)
- [decide\\_cluster\\_order, 71](#)
- [delaunayn, 59](#)
- [deldir, 59](#)
- [detectSpatialCorGenes, 72](#)
- [detectSpatialPatterns, 73](#)
- [dimCellPlot, 74, 78](#)
- [dimCellPlot2D, 74, 75, 76](#)

- dimGenePlot, [79](#), [82](#), [84](#)
- dimGenePlot2D, [79](#), [80](#), [80](#), [84](#)
- dimGenePlot3D, [80](#), [82](#), [83](#)
- dimPlot, [85](#), [89](#), [91](#), [169](#), [171](#), [172](#), [178](#), [180](#), [181](#), [183](#), [185](#), [187](#)
- dimPlot2D, [85](#), [86](#), [86](#), [91](#), [168–172](#), [177–185](#), [187](#)
- dimPlot3D, [78](#), [86](#), [89](#), [90](#), [169](#), [171](#), [172](#), [178](#), [180](#), [181](#), [183](#), [185–187](#)
- doHclust, [41](#), [92](#)
- doHMRF, [93](#)
- doKmeans, [41](#), [95](#)
- doLeidenCluster, [41](#), [96](#), [99](#), [277](#)
- doLeidenSubCluster, [97](#)
- doLouvainCluster, [41](#), [99](#)
- doLouvainCluster\_community, [41](#), [100](#), [100](#), [104](#), [106](#), [277](#)
- doLouvainCluster\_multinet, [41](#), [100](#), [101](#), [104](#), [107](#), [277](#)
- doLouvainSubCluster, [102](#)
- doLouvainSubCluster\_community, [104](#)
- doLouvainSubCluster\_multinet, [106](#)
- doRandomWalkCluster, [41](#), [108](#)
- doSNNCluster, [41](#), [109](#)
  
- estimateImageBg, [110](#)
- exportGiottoViewer, [110](#)
- exprCellCellcom, [111](#), [150](#), [151](#)
  
- fDataDT, [113](#)
- filterCellProximityGenes, [113](#)
- filterCombinations, [114](#), [118](#)
- filterCPG, [115](#)
- filterDistributions, [116](#)
- filterGiotto, [118](#)
- findCellProximityGenes, [119](#)
- findCPG, [120](#)
- findGiniMarkers, [122](#), [124](#), [126](#)
- findGiniMarkers\_one\_vs\_all, [124](#), [127](#)
- findMarkers, [125](#), [131](#)
- findMarkers\_one\_vs\_all, [126](#)
- findMastMarkers, [126](#), [128](#), [129](#)
- findMastMarkers\_one\_vs\_all, [127](#), [129](#)
- findNetworkNeighbors, [130](#)
- findScranMarkers, [126](#), [130](#), [132](#)
- findScranMarkers\_one\_vs\_all, [127](#), [131](#)
  
- general\_save\_function, [17](#)
- get10Xmatrix, [132](#)
- get\_os, [136](#)
- getClusterSimilarity, [133](#)
- getDendrogramSplits, [134](#)
- getDistinctColors, [135](#)
- getGiottoImage, [135](#)
- giotto (giotto-class), [136](#)
- giotto-class, [136](#)
- glouvain\_ml, [102](#)
  
- hclust, [93](#)
- Heatmap, [137](#)
- heatmSpatialCorGenes, [137](#)
- hyperGeometricEnrich, [138](#)
  
- insertCrossSectionGenePlot3D, [139](#)
- insertCrossSectionSpatPlot3D, [140](#)
  
- jackstrawPlot, [141](#)
  
- kmeans, [96](#)
- kNN, [57](#)
  
- layout\_with\_drl, [14](#)
- loadHMRF, [143](#)
  
- makeSignMatrixPAGE, [144](#), [195](#)
- makeSignMatrixRank, [144](#), [197](#)
- mergeClusters, [145](#)
  
- node\_clusters, [146](#)
- normalizeGiotto, [147](#)
  
- PAGEEnrich, [144](#), [148](#)
- PCA, [196](#)
- pDataDT, [19](#), [149](#)
- permutationPA, [142](#), [218](#)
- plotCCcomDotplot, [149](#)
- plotCCcomHeatmap, [150](#)
- plotCellProximityGenes, [151](#)
- plotCombineCCcom, [153](#)
- plotCombineCellCellCommunication, [154](#)
- plotCombineCellProximityGenes, [155](#)
- plotCombineCPG, [157](#)
- plotCPG, [158](#)
- plotGiottoImage, [159](#)
- plotHeatmap, [160](#)
- plotICG, [162](#)
- plotInteractionChangedGenes, [163](#)
- plotMetaDataCellsHeatmap, [164](#), [167](#)
- plotMetaDataHeatmap, [165](#), [166](#)
- plotPCA, [86](#), [89](#), [91](#), [168](#), [171](#), [172](#), [178](#), [180](#), [181](#), [183](#), [185](#), [187](#)
- plotPCA\_2D, [86](#), [89](#), [91](#), [169](#), [169](#), [172](#), [178](#), [180](#), [181](#), [183](#), [185](#), [187](#)
- plotPCA\_3D, [86](#), [89](#), [91](#), [169](#), [171](#), [171](#), [178](#), [180](#), [181](#), [183](#), [185](#), [187](#)
- plotRankSpatvsExpr, [173](#)
- plotRecovery, [174](#)



- plotRecovery\_sub, 175
- plotStatDelaunayNetwork, 175
- plotTSNE, 86, 89, 91, 169, 171, 172, 176, 180, 181, 183, 185, 187
- plotTSNE\_2D, 86, 89, 91, 169, 171, 172, 178, 178, 181, 183, 185, 187
- plotTSNE\_3D, 86, 89, 91, 169, 171, 172, 178, 180, 180, 183, 185, 187
- plotUMAP, 86, 89, 91, 169, 171, 172, 178, 180, 181, 182, 185, 187
- plotUMAP\_2D, 86, 89, 91, 169, 171, 172, 178, 180, 181, 183, 184, 187
- plotUMAP\_3D, 86, 89, 91, 169, 171, 172, 178, 180, 181, 183, 185, 186
- prcomp\_irlba, 196
- print.giotto, 187
- rankEnrich, 145, 188
- rankSpatialCorGroups, 188
- readExprMatrix, 189
- readGiottoInstructions, 190
- removeBatchEffect, 16
- removeCellAnnotation, 190
- removeGeneAnnotation, 191
- replaceGiottoInstructions, 191
- Rtsne, 201
- runDWLSDeconv, 192
- runHyperGeometricEnrich, 138, 193, 199
- runPAGEEnrich, 148, 194, 199
- runPCA, 195, 204
- runRankEnrich, 188, 196, 199
- runSpatialDeconv, 197
- runSpatialEnrich, 60, 198
- runTSNE, 199
- runUMAP, 201
- screePlot, 203
- select\_expression\_values, 206
- selectPatternGenes, 205
- show, giotto-method, 206
- showClusterDendrogram, 206
- showClusterHeatmap, 208
- showGiottoImageNames, 135, 159, 209
- showGiottoInstructions, 209
- showGrids, 19, 210
- showNetworks, 210
- showPattern, 211
- showPattern2D, 211, 212
- showPattern3D, 213
- showPatternGenes, 214
- showProcessingSteps, 215
- showSaveParameters, 75, 78, 80, 82, 84, 86, 89, 91, 137, 161, 165, 167, 169, 171, 172, 176, 178, 180, 181, 183, 185, 187, 187, 189, 207, 208, 211, 212, 214, 215, 216, 222, 226, 228, 233, 235, 238, 241, 244, 248, 252, 253, 256, 258, 260, 265, 269, 271, 284
- showSpatialCorGenes, 73, 216
- signPCA, 217
- silhouetteRank, 219
- sNN, 57
- sNNclust, 109
- spatCellCellcom, 150, 151, 220
- spatCellPlot, 221, 226
- spatCellPlot2D, 221, 222, 223
- spatDimCellPlot, 226, 233
- spatDimCellPlot2D, 226, 228, 228
- spatDimGenePlot, 233, 239, 242
- spatDimGenePlot2D, 234, 235, 235, 242
- spatDimGenePlot3D, 235, 239, 239
- spatDimPlot, 242, 249, 252
- spatDimPlot2D, 242, 244, 244, 252
- spatDimPlot3D, 244, 249, 249
- spatGenePlot, 252, 256, 258
- spatGenePlot2D, 68, 252, 253, 254, 258
- spatGenePlot3D, 68, 253, 256, 256
- spatialAEH, 259
- SpatialDE, 259
- spatialDE, 260
- spatNetwDistributions, 261
- spatNetwDistributionsDistance, 262
- spatNetwDistributionsKneighbors, 263
- spatPlot, 264, 269, 271
- spatPlot2D, 264, 265, 266, 271, 281, 282
- spatPlot3D, 265, 269, 269, 281, 283
- specificCellCellcommunicationScores, 272
- split\_dendrogram\_in\_two, 273
- stitchFieldCoordinates, 52, 273
- stitchTileCoordinates, 275
- subClusterCells, 275
- subsetGiotto, 277
- subsetGiottoLocs, 278
- trendSceek, 279
- trendsceek\_test, 279
- triangulate, 59
- umap, 202
- updateGiottoImage, 280
- viewHMRFresults, 280
- viewHMRFresults2D, 281
- viewHMRFresults3D, 282
- violinPlot, 283

`write_giotto_viewer_annotation`, [285](#)  
`write_giotto_viewer_dim_reduction`, [285](#)  
`write_giotto_viewer_numeric_annotation`,  
    [286](#)  
`writeHMRResults`, [284](#)  
  
`zlm`, [128](#)