# Package 'Giotto'

December 2, 2019

```
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      ggplot2 (>= 3.1.1),
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Imports Rtsne (>= 0.15),
      uwot (>= 0.0.0.9010),
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      FactoMineR (>= 1.34),
      factoextra (>= 1.0.5),
      cowplot (>= 0.9.4),
      grDevices,
      RColorBrewer,
      jackstraw (>= 1.3),
      dbscan (>= 1.1-3),
      ggalluvial (>= 0.9.1),
      scales (>= 1.0.0),
      ComplexHeatmap (>= 1.20.0),
      \frac{1}{2} qvalue (>= 2.14.1),
      1fa (>= 1.12.0),
      igraph (>= 1.2.4.1),
      plotly,
      reticulate,
      magrittr,
      limma,
      ggdendro,
      smfishHmrf,
      matrixStats (\geq 0.55.0),
      IRanges
```

Suggests knitr,

2 R topics documented:

```
rmarkdown,
MAST,
scran (>= 1.10.1),
png,
tiff
```

# biocViews

VignetteBuilder knitr

**Remotes** lambdamoses/smfishhmrf-r

# R topics documented:

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addCellMetadata

addCellMetadata

# Description

adds cell metadata to the giotto object

# Usage

```
addCellMetadata(
  gobject,
  new_metadata,
  by_column = FALSE,
  column_cell_ID = NULL
)
```

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#### **Arguments**

```
gobject giotto object

new_metadata new cell metadata to use (data.table, data.frame, ...)

by_column merge metadata based on cell_ID column in pDataDT (default = FALSE)

column_cell_ID column name of new metadata to use if by_column = TRUE
```

#### **Details**

You can add additional cell metadata in two manners: 1. Provide a data.table or data.frame with cell annotations in the same order as the cell\_ID column in pDataDT(gobject) 2. Provide a data.table or data.frame with cell annotations and specificy which column contains the cell IDs, these cell IDs need to match with the cell\_ID column in pDataDT(gobject)

#### Value

giotto object

## **Examples**

```
addCellMetadata(gobject)
```

addCellStatistics

addCellStatistics

#### **Description**

adds cells statistics to the giotto object

# Usage

```
addCellStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

8 addGeneMetadata

#### **Details**

This function will add the following statistics to cell metadata:

- nr\_genes: Denotes in how many genes are detected per cell
- perc\_genes: Denotes what percentage of genes is detected per cell
- total\_expr: Shows the total sum of gene expression per cell

#### Value

```
giotto object if return_gobject = TRUE
```

#### **Examples**

```
addCellStatistics(gobject)
```

addGeneMetadata

addGeneMetadata

# **Description**

adds gene metadata to the giotto object

## Usage

```
addGeneMetadata(gobject, new_metadata, by_column = F, column_gene_ID = NULL)
```

## **Arguments**

gobject giotto object

new\_metadata new metadata to use

by\_column merge metadata based on gene\_ID column in fDataDT column\_cell\_ID column name of new metadata to use if by\_column = TRUE

#### **Details**

You can add additional gene metadata in two manners: 1. Provide a data.table or data.frame with gene annotations in the same order as the gene\_ID column in fDataDT(gobject) 2. Provide a data.table or data.frame with gene annotations and specificy which column contains the gene IDs, these gene IDs need to match with the gene\_ID column in fDataDT(gobject)

# Value

giotto object

## **Examples**

addGeneMetadata(gobject)

addGeneStatistics 9

addGeneStatistics

addGeneStatistics

## **Description**

adds gene statistics to the giotto object

## Usage

```
addGeneStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

## **Arguments**

# **Details**

This function will add the following statistics to gene metadata:

- nr\_cells: Denotes in how many cells the gene is detected
- per\_cells: Denotes in what percentage of cells the gene is detected
- total\_expr: Shows the total sum of gene expression in all cells
- mean\_expr: Average gene expression in all cells
- mean\_expr\_det: Average gene expression in cells with detectable levels of the gene

# Value

```
giotto object if return_gobject = TRUE
```

```
addGeneStatistics(gobject)
```

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addHMRF addHMRF

## **Description**

Add selected results from doHMRF to the giotto object

#### Usage

```
addHMRF(gobject, HMRFoutput, k = NULL, betas_to_add = NULL, hmrf_name = NULL)
```

## **Arguments**

gobject giotto object

 $HMRF output \qquad \qquad HMRF output \ from \ doHMRF()$ 

k number of domains

betas\_to\_add results from different betas that you want to add

name specify a custom name

#### **Details**

Description ...

#### Value

giotto object

## **Examples**

addHMRF(gobject)

addNetworkLayout

addNetworkLayout

## **Description**

Add a network layout for a selected nearest neighbor network

# Usage

```
addNetworkLayout(
  gobject,
  nn_network_to_use = NULL,
  network_name = NULL,
  layout_type = c("drl"),
  options_list = NULL,
  layout_name = "layout",
  return_gobject = TRUE
)
```

addStatistics 11

#### **Arguments**

#### **Details**

Description of layouts and options.

#### Value

giotto object with updated layout for selected NN network

## **Examples**

```
addNetworkLayout(gobject)
```

addStatistics addStatistics

## **Description**

adds genes and cells statistics to the giotto object

# Usage

```
addStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

## **Arguments**

# Details

See addGeneStatistics and addCellStatistics

12 adjustGiottoMatrix

#### Value

```
giotto object if return_gobject = TRUE, else a list with results
```

#### **Examples**

```
addStatistics(gobject)
```

```
adjustGiottoMatrix adjustGiottoMatrix
```

# **Description**

normalize and/or scale expresion values of Giotto object

## Usage

```
adjustGiottoMatrix(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  batch_columns = NULL,
  covariate_columns = NULL,
  return_gobject = TRUE,
  update_slot = c("custom")
)
```

## **Arguments**

```
gobject giotto object
expression_values
expression values to use

batch_columns metadata columns that represent different batch (max = 2)
covariate_columns
metadata columns that represent covariates to regress out
return_gobject boolean: return giotto object (default = TRUE)
update_slot expression slot that will be updated (default = custom)
```

#### **Details**

This function implements the limma::removeBatchEffect function to remove known batch effects and to adjust expression values according to provided covariates.

# Value

```
giotto object
```

```
adjustGiottoMatrix(gobject)
```

aes\_string2

#### **Description**

makes sure aes\_string can also be used with names that start with numeric values

#### Usage

```
aes_string2(...)
```

 $all \textit{CellCellcommunications} Scores \\ all \textit{CellCellcommunications} Scores$ 

## **Description**

All Cell-Cell communication scores based on spatial expression of interacting cells

# Usage

```
allCellCellcommunicationsScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "cell_types",
  random_iter = 100,
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  min_observations = 2,
  verbose = c("a little", "a lot", "none")
)
```

```
giotto object to use
gobject
spatial_network_name
                  spatial network to use for identifying interacting cells
cluster_column cluster column with cell type information
                  number of iterations
random_iter
gene_set_1
                  first specific gene set from gene pairs
gene_set_2
                  second specific gene set from gene pairs
log2FC_addendum
                  addendum to add when calculating log2FC
min_observations
                  minimum number of interactions needed to be considered
verbose
                  verbose
```

#### **Details**

Details will follow.

#### Value

Cell-Cell communication scores for gene pairs based on spatial interaction

#### **Examples**

```
allCellCellcommunicationsScores(gobject)
```

```
all_plots_save_function 
 all_plots_save_function
```

# Description

Function to automatically save plots to directory of interest

# Usage

```
all_plots_save_function(
  gobject,
  plot_object,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  default_save_name = "giotto_plot",
  save_format = NULL,
  show_saved_plot = F,
  ncol = 1,
  nrow = 1,
  scale = 1,
  base_width = NULL,
  base_height = NULL,
  base_aspect_ratio = NULL,
  units = NULL,
  dpi = NULL,
  limitsize = TRUE,
)
```

```
gobject giotto object
plot_object object to plot
save_dir directory to save to
save_folder folder in save_dir to save to
save_name name of plot
```

annotateGiotto 15

```
save_format format (e.g. png, tiff, pdf, ...)
```

show\_saved\_plot

load & display the saved plot

ncol number of columns nrow number of rows

scale scale
base\_width width
base\_height height
base\_aspect\_ratio

aspect ratio

units units

dpi Plot resolution

limitsize When TRUE (the default), ggsave will not save images larger than 50x50 inches,

to prevent the common error of specifying dimensions in pixels.

... additional parameters to ggplot\_save\_function or general\_save\_function

#### See Also

```
Giotto::general_save_function
```

#### **Examples**

```
all_plots_save_function(gobject)
```

 $annotate {\tt Giotto}$ 

annotateGiotto

## **Description**

Converts cluster results into provided annotation.

# Usage

```
annotateGiotto(
  gobject,
  annotation_vector = NULL,
  cluster_column = NULL,
  name = "cell_types"
)
```

#### **Details**

You need to specifify which (cluster) column you want to annotate and you need to provide an annotation vector like this:

- 1. identify the cell type of each cluster
- 2. create a vector of these cell types, e.g. cell\_types = c('T-cell', 'B-cell', 'Stromal')
- 3. provide original cluster names to previous vector, e.g. names(cell\_types) = c(2, 1, 3)

#### Value

```
giotto object
```

## **Examples**

```
annotateGiotto(gobject)
```

```
annotate {\tt Spatial Network}
```

annotate Spatial Network

## **Description**

Annotate spatial network with cell metadata information.

## Usage

```
annotateSpatialNetwork(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column
)
```

# **Arguments**

## Value

annotated network in data.table format

```
annotateSpatialNetwork(gobject)
```

```
annotate\_spatlocs\_with\_spatgrid\_2D \\ annotate\_spatlocs\_with\_spatgrid\_2D
```

# Description

annotate spatial locations with 2D spatial grid information

## Usage

```
annotate_spatlocs_with_spatgrid_2D(spatloc, spatgrid)
```

## **Arguments**

spatloc spatial\_locs slot from giotto object

spatgrid selected spatial\_grid slot from giotto object

## Value

annotated spatial location data.table

## **Examples**

```
annotate_spatlocs_with_spatgrid_2D()
```

# **Description**

annotate spatial locations with 3D spatial grid information

# Usage

```
annotate_spatlocs_with_spatgrid_3D(spatloc, spatgrid)
```

# Arguments

spatloc spatial\_locs slot from giotto object

spatgrid selected spatial\_grid slot from giotto object

#### Value

annotated spatial location data.table

```
{\tt annotate\_spatlocs\_with\_spatgrid\_3D()}
```

18 binGetSpatialGenes

```
average_gene_gene_expression_in_groups

average_gene_expression_in_groups
```

# Description

calculate average expression per cluster

## Usage

```
average_gene_gene_expression_in_groups(
  gobject,
  cluster_column = "cell_types",
  gene_set_1,
  gene_set_2
)
```

## **Arguments**

```
gobject giotto object to use

cluster_column cluster column with cell type information

gene_set_1 first specific gene set from gene pairs

gene_set_2 second specific gene set from gene pairs
```

## **Details**

Details will follow.

## Value

data.table with average expression scores for each cluster

## **Examples**

```
average_gene_gene_expression_in_groups(gobject)
```

binGetSpatialGenes binGetSpatialGenes

# Description

Rapid computation of genes that are spatially clustered

binGetSpatialGenes 19

#### Usage

```
binGetSpatialGenes(
  gobject,
  bin_method = c("kmeans", "rank"),
  expression_values = c("normalized", "scaled", "custom"),
  spatial_network_name = "spatial_network",
  nstart = 3,
  iter_max = 10,
  percentage_rank = 10,
  do_fisher_test = F,
  community_expectation = 5,
  verbose = F
)
```

#### **Arguments**

```
gobject
                  giotto object
bin_method
                  method to binarize gene expression
expression_values
                  expression values to use
spatial_network_name
                  name of spatial network to use (default = 'spatial_network')
                  kmeans: nstart parameter
nstart
iter_max
                  kmeans: iter.max parameter
do_fisher_test perform fisher test
community_expectation
                  cell degree expectation in spatial communities
                  be verbose
verbose
rank_percentage
                  percentage of top cells for binarization
```

## **Details**

We provide two ways to identify spatial genes based on gene expression binarization. Both methods are identicial except for how binarization is performed.

- 1. binarize: Each gene is binarized (0 or 1) in each cell with **kmeans** (k = 2) or based on **rank** percentile
- 2. network: Alll cells are connected through a k-nearest neighbor network
- 3. contingency table: A contingency table is calculated based on all pairwise cell-cell interactions (0-0, 0-1, 1-0 or 1-1)
- 4. For each gene an odds-ratio (OR) and fisher.test (optional) is calculated

Additionally 2 other statistics are provided:

- Number of cells with high expression (binary = 1)
- total and ratio of highly connected cells: Cells with a connectivity higher than community\_expectation

20 calculateHVG

#### Value

```
data.table with results (see details)
```

#### **Examples**

binGetSpatialGenes(gobject)

calculateHVG

calculateHVG

## **Description**

compute highly variable genes

# Usage

```
calculateHVG(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  method = c("cov_groups", "cov_loess"),
  reverse_log_scale = FALSE,
  logbase = 2,
  expression_threshold = 0,
  nr_expression_groups = 20,
  zscore_threshold = 1.5,
  HVGname = "hvg",
  difference_in_cov = 0.1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "HVGplot",
  return_gobject = TRUE
)
```

```
gobject
                  giotto object
expression_values
                  expression values to use
method
                  method to calculate highly variable genes
reverse_log_scale
                  reverse log-scale of expression values (default = FALSE)
                  if reverse_log_scale is TRUE, which log base was used?
logbase
expression_threshold
                  expression threshold to consider a gene detected
nr_expression_groups
                  number of expression groups for cov_groups
zscore_threshold
                  zscore to select hvg for cov_groups
```

calculateMetaTable 21

#### **Details**

Currently we provide 2 ways to calculate highly variable genes: 1. high coeff of variance (COV) within groups:

First genes are binned (*nr\_expression\_groups*) into average expression groups and the COV for each gene is converted into a z-score within each bin. Genes with a z-score higher than the threshold (*zscore\_threshold*) are considered highly variable.

#### 2. high COV based on loess regression prediction:

A predicted COV is calculated for each gene using loess regression (COV~log(mean expression)) Genes that show a higher than predicted COV (*difference\_in\_cov*) are considered highly variable.

#### Value

giotto object highly variable genes appended to gene metadata (fDataDT)

## **Examples**

```
calculateHVG(gobject)
```

calculateMetaTable calculateMetaTable

#### **Description**

calculates the average gene expression for one or more (combined) annotation columns.

## Usage

```
calculateMetaTable(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metadata_cols = NULL,
  selected_genes = NULL
)
```

22 calculateMetaTableCells

## **Arguments**

```
gobject giotto object
expression_values
expression values to use
metadata_cols annotation columns found in pDataDT(gobject)
selected_genes subset of genes to use
```

## Value

data.table with average expression values for each gene per (combined) annotation

## **Examples**

```
calculateMetaTable(gobject)
```

```
{\tt calculateMetaTableCells}
```

calculateMetaTableCells

# Description

calculates the average metadata values for one or more (combined) annotation columns.

## Usage

```
calculateMetaTableCells(
  gobject,
  value_cols = NULL,
  metadata_cols = NULL,
  spat_enr_names = NULL
)
```

## **Arguments**

```
gobject giotto object
value_cols metadata or enrichment value columns to use
metadata_cols annotation columns found in pDataDT(gobject)
spat_enr_names which spatial enrichment results to include
```

## Value

data.table with average metadata values per (combined) annotation

```
calculateMetaTableCells(gobject)
```

# Description

Calculate spatial genes using distance matrix.

# Usage

```
calculate_spatial_genes_python(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metric = "euclidean",
  subset_genes = NULL,
  rbp_p = 0.95,
  examine_top = 0.3,
  python_path = NULL
)
```

# **Arguments**

```
gobject giotto object
expression_values
expression values to use

metric distance metric to use

subset_genes only run on this subset of genes
rbp_p fractional binarization threshold
examine_top top fraction to evaluate with silhouette
python_path specify specific path to python if required
```

## **Details**

Description of how we compute spatial pattern genes.

## Value

data.table with spatial scores

```
calculate_spatial_genes_python(gobject)
```

24 cellProximityBarplot

```
cellProximityBarplot cellProximityBarplot
```

## **Description**

Create barplot from cell-cell proximity scores

## Usage

```
cellProximityBarplot(
  gobject,
  CPscore,
  min_orig_ints = 5,
  min_sim_ints = 5,
  p_val = 0.05,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityBarplot"
)
```

## **Arguments**

```
gobject
                  giotto object
                  CPscore, output from cellProximityEnrichment()
CPscore
                  filter on minimum original cell-cell interactions
min_orig_ints
                  filter on minimum simulated cell-cell interactions
min_sim_ints
                  p-value
p_val
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

# **Details**

This function creates a barplot that shows the spatial proximity enrichment or depletion of cell type pairs.

#### Value

```
ggplot barplot
```

```
cellProximityBarplot(CPscore)
```

```
cellProximityEnrichment
```

cellProximityEnrichment

#### **Description**

Compute cell-cell interaction enrichment (observed vs expected)

#### Usage

```
cellProximityEnrichment(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column,
  number_of_simulations = 100
)
```

#### **Arguments**

## **Details**

Spatial proximity enrichment or depletion between pairs of cell types is calculated by calculating the observed over the expected frequency of cell-cell proximity interactions. The expected frequency is the average frequency calculated from a number of spatial network simulations. Each individual simulation is obtained by random permutations of the cell type labels of each node (cell) in the spatial network.

## Value

List of cell Proximity scores (CPscores) in data.table format. The first data.table (raw\_sim\_table) shows the raw observations of both the original and simulated networks. The second data.table (enrichm\_res) shows the enrichment results.

```
cellProximityEnrichment(gobject)
```

cellProximityHeatmap cellProximityHeatmap

## **Description**

Create heatmap from cell-cell proximity scores

# Usage

```
cellProximityHeatmap(
  gobject,
  CPscore,
  scale = T,
  order_cell_types = T,
  color_breaks = NULL,
  color_names = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityHeatmap")
```

## Arguments

```
giotto object
gobject
                  CPscore, output from cellProximityEnrichment()
CPscore
scale
                  scale cell-cell proximity interaction scores
order_cell_types
                  order cell types based on enrichment correlation
color_breaks
                  numerical vector of length 3 to represent min, mean and maximum
                  character color vector of length 3
color_names
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

#### **Details**

This function creates a heatmap that shows the spatial proximity enrichment or depletion of cell type pairs.

# Value

ggplot heatmap

cellProximityNetwork 27

#### **Examples**

```
cellProximityHeatmap(CPscore)
```

```
cell Proximity Network \qquad cell Proximity Network
```

# **Description**

Create network from cell-cell proximity scores

## Usage

```
cellProximityNetwork(
  gobject,
  CPscore,
  remove_self_edges = FALSE,
  self_loop_strength = 0.1,
  color_depletion = "lightgreen",
  color_enrichment = "red",
  rescale_edge_weights = TRUE,
  edge_weight_range_depletion = c(0.1, 1),
  edge_weight_range_enrichment = c(1, 5),
  layout = "Fruchterman",
  only_show_enrichment_edges = F,
  edge_width_range = c(0.1, 2),
  node_size = 4,
  node_text_size = 6,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityNetwork"
```

```
gobject
                  giotto object
CPscore
                  CPscore, output from cellProximityEnrichment()
remove_self_edges
                  remove enrichment/depletion edges with itself
self_loop_strength
                  size of self-loops
color_depletion
                  color for depleted cell-cell interactions
color_enrichment
                  color for enriched cell-cell interactions
rescale_edge_weights
                  rescale edge weights (boolean)
{\tt edge\_weight\_range\_depletion}
                  numerical vector of length 2 to rescale depleted edge weights
```

```
edge_weight_range_enrichment
```

numerical vector of length 2 to rescale enriched edge weights

layout algorithm to use to draw nodes and edges

only\_show\_enrichment\_edges

show only the enriched pairwise scores

edge\_width\_range

range of edge width

node\_size size of nodes

node\_text\_size size of node labels

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

## **Details**

This function creates a network that shows the spatial proximity enrichment or depletion of cell type pairs.

#### Value

igraph plot

# **Examples**

```
cellProximityNetwork(CPscore)
```

```
cellProximitySpatPlot cellProximitySpatPlot
```

## **Description**

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

# Usage

```
cellProximitySpatPlot(gobject, ...)
```

# **Arguments**

gobject giotto object

 ${\tt interaction\_name}$ 

cell-cell interaction name

cluster\_column cluster column with cell clusters

sdimx x-axis dimension name (default = 'sdimx') sdimy y-axis dimension name (default = 'sdimy')

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```
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
point_size_select
                  size of selected points
point_select_border_col
                  border color of selected points
point_select_border_stroke
                  stroke size of selected points
point_size_other
                  size of other points
point_other_border_col
                  border color of other points
point_other_border_stroke
                  stroke size of other points
                  show plots
show_plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

# **Details**

Description of parameters.

#### Value

ggplot

#### See Also

cellProximitySpatPlot2D and cellProximitySpatPlot3D for 3D

#### **Examples**

```
cellProximitySpatPlot(gobject)
```

```
cellProximitySpatPlot2D
```

cellProximitySpatPlot2D

# Description

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

# Usage

```
cellProximitySpatPlot2D(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 1,
  show_legend = T,
  point_size_select = 2,
  point_select_border_col = "black",
  point_select_border_stroke = 0.05,
  point_size_other = 1,
  point_alpha_other = 0.3,
  point_other_border_col = "lightgrey",
  point_other_border_stroke = 0.01,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximitySpatPlot2D"
)
```

cluster\_column cluster column with cell clusters x-axis dimension name (default = 'sdimx') sdimx sdimy y-axis dimension name (default = 'sdimy') cell\_color color for cells (see details) cell\_color\_code named vector with colors color\_as\_factor convert color column to factor show\_other\_cells decide if show cells not in network show\_network show underlying spatial network color of spatial network network\_color spatial\_network\_name name of spatial network to use show spatial grid show\_grid grid\_color color of spatial grid spatial\_grid\_name name of spatial grid to use coord\_fix\_ratio fix ratio between x and y-axis show\_legend show legend point\_size\_select size of selected points point\_select\_border\_col border color of selected points point\_select\_border\_stroke stroke size of selected points point\_size\_other size of other points point\_other\_border\_col border color of other points point\_other\_border\_stroke stroke size of other points show\_plot show plots return\_plot return ggplot object directly save the plot [boolean] save\_plot save\_param list of saving parameters from all\_plots\_save\_function() default\_save\_name default save name for saving, don't change, change save\_name in save\_param

## **Details**

Description of parameters.

#### Value

ggplot

# **Examples**

```
cellProximitySpatPlot2D(gobject)
```

```
cell Proximity SpatPlot 3D \\ cell Proximity SpatPlot 2D
```

# Description

Visualize 3D cell-cell interactions according to spatial coordinates in plotly mode

## Usage

```
cellProximitySpatPlot3D(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = T,
  show_network = T,
  show\_other\_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  point_size_select = 4,
  point_size_other = 2,
  point_alpha_other = 0.5,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximitySpatPlot3D",
)
```

#### **Arguments**

```
gobject
                  giotto object
interaction_name
                  cell-cell interaction name
cluster_column cluster column with cell clusters
                  x-axis dimension name (default = 'sdimx')
sdimx
sdimy
                  y-axis dimension name (default = 'sdimy')
sdimz
                  z-axis dimension name (default = 'sdimz')
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
show_legend
                  show legend
point_size_select
                  size of selected points
point_size_other
                  size of other points
                  show plots
show_plot
return_plot
                  return plotly object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

#### **Details**

Description of parameters.

#### Value

plotly

```
cellProximitySpatPlot3D(gobject)
```

34 cellProximityVisPlot

```
cellProximityVisPlot cellProximityVisPlot
```

# Description

Visualize cell-cell interactions according to spatial coordinates

# Usage

```
cellProximityVisPlot(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 1,
  show_legend = T,
  point_size_select = 2,
  point_select_border_col = "black",
  point_select_border_stroke = 0.05,
  point_size_other = 1,
  point_alpha_other = 0.3,
  point_other_border_col = "lightgrey",
  point_other_border_stroke = 0.01,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_{ticks} = NULL,
  plot_method = c("ggplot", "plotly"),
)
```

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sdimx x-axis dimension name (default = 'sdimx')
sdimy y-axis dimension name (default = 'sdimy')
sdimz z-axis dimension name (default = 'sdimz')

cell\_color color for cells (see details)

cell\_color\_code

named vector with colors

color\_as\_factor

convert color column to factor

show\_network show underlying spatial network

network\_color color of spatial network

spatial\_network\_name

name of spatial network to use

show\_grid show spatial grid grid\_color color of spatial grid

spatial\_grid\_name

name of spatial grid to use

coord\_fix\_ratio

fix ratio between x and y-axis

show\_legend show legend

point\_size\_select

size of selected points

point\_select\_border\_col

border color of selected points

point\_select\_border\_stroke

stroke size of selected points

point\_size\_other

size of other points

point\_other\_border\_col

border color of other points

point\_other\_border\_stroke

stroke size of other points

## **Details**

Description of parameters.

#### Value

ggplot or plotly

## **Examples**

cellProximityVisPlot(gobject)

```
cell Proximity VisPlot\_2D\_ggplot \\ cell Proximity VisPlot\_2D\_ggplot
```

#### **Description**

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

#### Usage

```
cellProximityVisPlot_2D_ggplot(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 1,
  show_legend = T,
  point_size_select = 2,
  point_select_border_col = "black",
  point_select_border_stroke = 0.05,
  point_size_other = 1,
  point_alpha_other = 0.3,
  point_other_border_col = "lightgrey",
  point_other_border_stroke = 0.01,
)
```

```
color_as_factor
```

convert color column to factor

show\_other\_cells

decide if show cells not in network

show\_network show underlying spatial network

network\_color color of spatial network

 $spatial\_network\_name$ 

name of spatial network to use

show\_grid show spatial grid

grid\_color color of spatial grid

spatial\_grid\_name

name of spatial grid to use

coord\_fix\_ratio

fix ratio between x and y-axis

show\_legend show legend

point\_size\_select

size of selected points

point\_select\_border\_col

border color of selected points

point\_select\_border\_stroke

stroke size of selected points

point\_size\_other

size of other points

point\_other\_border\_col

border color of other points

point\_other\_border\_stroke

stroke size of other points

# Details

Description of parameters.

# Value

ggplot

# **Examples**

cellProximityVisPlot\_2D\_ggplot(gobject)

```
cell Proximity VisPlot\_2D\_plotly \\ cell Proximity VisPlot\_2D\_plotly
```

# Description

Visualize 2D cell-cell interactions according to spatial coordinates in plotly mode

# Usage

```
cellProximityVisPlot_2D_plotly(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  point_size_select = 2,
  point_size_other = 1,
  point_alpha_other = 0.3,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
)
```

# **Arguments**

```
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
                  color of spatial grid
grid_color
spatial_grid_name
                  name of spatial grid to use
show_legend
                  show legend
point_size_select
                  size of selected points
coord_fix_ratio
                  fix ratio between x and y-axis
```

# **Details**

Description of parameters.

# Value

plotly

## **Examples**

```
cell Proximity VisPlot\_2D\_plotly (gobject)
```

# **Description**

Visualize 3D cell-cell interactions according to spatial coordinates in plotly mode

```
cellProximityVisPlot_3D_plotly(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
```

```
color_as_factor = T,
show_other_cells = F,
show_network = F,
show_other_network = F,
network_color = NULL,
spatial_network_name = "spatial_network",
show\_grid = F,
grid_color = NULL,
spatial_grid_name = "spatial_grid",
show_legend = T,
point_size_select = 2,
point_size_other = 1,
point_alpha_other = 0.5,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
z_{ticks} = NULL,
```

# **Arguments**

```
gobject
                  giotto object
interaction_name
                  cell-cell interaction name
cluster_column cluster column with cell clusters
sdimx
                  x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimy
                  z-axis dimension name (default = 'sdimz')
sdimz
cell color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
                  show underlying spatial network
show_network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
show_legend
                  show legend
point_size_select
                  size of selected points
coord_fix_ratio
                  fix ratio between x and y-axis
```

# **Details**

Description of parameters.

# Value

plotly

# **Examples**

```
cellProximityVisPlot_3D_plotly(gobject)
```

```
{\tt changeGiottoInstructions}
```

change Giot to Instructions

# **Description**

Function to change one or more instructions from giotto object

# Usage

```
changeGiottoInstructions(
  gobject,
  params = NULL,
  new_values = NULL,
  return_gobject = TRUE
)
```

# Arguments

```
gobject giotto object

params parameter(s) to change

new_values new value(s) for parameter(s)

return_gobject (boolean) return giotto object
```

#### Value

named vector with giotto instructions

# **Examples**

changeGiottoInstructions()

42 clusterCells

clusterCells

clusterCells

#### **Description**

cluster cells using a NN-network and community detection algorithms

```
clusterCells(
  gobject,
 cluster_method = c("leiden", "louvain_community", "louvain_multinet", "randomwalk",
    "sNNclust", "kmeans", "hierarchical"),
  name = "cluster_name",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  pyth_leid_resolution = 1,
  pyth_leid_weight_col = "weight",
 pyth_leid_part_type = c("RBConfigurationVertexPartition", "ModularityVertexPartition"),
  pyth_leid_init_memb = NULL,
  pyth_leid_iterations = 1000,
  pyth_louv_resolution = 1,
  pyth_louv_weight_col = NULL,
  python_louv_random = F,
  python_path = NULL,
  louvain_gamma = 1,
  louvain\_omega = 1,
  walk\_steps = 4,
  walk_clusters = 10,
  walk_weights = NA,
  sNNclust_k = 20,
  sNNclust_eps = 4,
  sNNclust_minPts = 16,
  borderPoints = TRUE,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
  dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  distance_method = c("original", "pearson", "spearman", "euclidean", "maximum",
    "manhattan", "canberra", "binary", "minkowski"),
  km_centers = 10,
  km_iter_max = 100,
  km_nstart = 1000,
  km_algorithm = "Hartigan-Wong",
 hc_agglomeration_method = c("ward.D2", "ward.D", "single", "complete", "average",
    "mcquitty", "median", "centroid"),
  hc_k = 10,
  hc_h = NULL
  return_gobject = TRUE,
  set_seed = T,
```

clusterCells 43

```
seed_number = 1234,
...
)
```

#### **Arguments**

```
gobject
                 giotto object
cluster_method community cluster method to use
                 name for new clustering result
name
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
pyth_leid_resolution
                 resolution for leiden
pyth_leid_weight_col
                 column to use for weights
pyth_leid_part_type
                 partition type to use
pyth_leid_init_memb
                 initial membership
pyth_leid_iterations
                 number of iterations
pyth_louv_resolution
                 resolution for louvain
pyth_louv_weight_col
                 python louvain param: weight column
python_louv_random
                 python louvain param: random
python_path
                 specify specific path to python if required
                 louvain param: gamma or resolution
louvain_gamma
louvain_omega
                 louvain param: omega
walk_steps
                 randomwalk: number of steps
walk_clusters
                 randomwalk: number of clusters
                 randomwalk: weight column
walk_weights
                 SNNclust: k neighbors to use
sNNclust_k
                 SNNclust: epsilon
sNNclust_eps
sNNclust_minPts
                 SNNclust: min points
borderPoints
                 SNNclust: border points
expression_values
                 expression values to use
genes_to_use
                 = NULL,
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
```

name of reduction 'pca',

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dimensions\_to\_use

dimensions to use

distance\_method

distance method

km\_centers kmeans centers km\_iter\_max kmeans iterations

km\_nstart kmeans random starting points

km\_algorithm kmeans algorithm

hc\_agglomeration\_method

hierarchical clustering method

hc\_k hierachical number of clusters

hc\_h hierarchical tree cutoff

return\_gobject boolean: return giotto object (default = TRUE)

set\_seed set seed

seed\_number number for seed
... additional parameters

#### **Details**

Description of different clustering methods.

#### Value

giotto object appended with new cluster

# **Examples**

clusterCells(gobject)

combineMetadata combineMetadata

# **Description**

This function combines the cell metedata with spatial enrichment results from createSpatialEnrich

#### Usage

```
combineMetadata(gobject, spat_enr_names = NULL)
```

# **Arguments**

gobject Giotto object

spat\_enr\_names names of spatial enrichment results

# Value

Extended cell metadata in data.table format.

#### **Examples**

combineMetadata(gobject)

```
{\tt convertSignListToMatrix}
```

convertSignListToMatrix

# Description

Function to convert list of signature genes (e.g. for cell types) into a binary matrix format that can be used with the PAGE enrichment option.

# Usage

```
convertSignListToMatrix(sign_names, sign_list)
```

# Arguments

```
sign_names vector with names for each provided gene signature sign_list list of gene signatures
```

# Value

matrix

# **Examples**

```
convertSignListToMatrix()
```

createGiottoInstructions

create Giot to Instructions

# Description

Function to set global instructions for giotto functions

```
createGiottoInstructions(
  python_path = NULL,
  show_plot = NULL,
  return_plot = NULL,
  save_plot = NULL,
  save_dir = NULL,
  plot_format = NULL,
  dpi = NULL,
  units = NULL,
  height = NULL,
  width = NULL
```

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# **Arguments**

path to python binary to use python\_path print plot to console, default = TRUE show\_plot return plot as object, default = TRUE return\_plot automatically save plot, dafault = FALSE save\_plot path to directory where to save plots save\_dir resolution for raster images dpi height of plots height width of plots width

#### Value

named vector with giotto instructions

# **Examples**

createGiottoInstructions()

# Description

Function to create a giotto object

```
createGiottoObject(
  raw_exprs,
  spatial_locs = NULL,
  norm_expr = NULL,
  norm_scaled_expr = NULL,
  custom_expr = NULL,
  cell_metadata = NULL,
  gene_metadata = NULL,
  spatial_network = NULL,
  spatial_network_name = NULL,
  spatial_grid = NULL,
  spatial_grid_name = NULL,
  spatial_enrichment = NULL,
  spatial_enrichment_name = NULL,
  dimension_reduction = NULL,
  nn_network = NULL,
  offset_file = NULL,
  instructions = NULL
)
```

createGiottoObject 47

## **Arguments**

matrix with raw expression counts [required] raw\_exprs data.table or data.frame with coordinates for cell centroids spatial\_locs normalized expression values norm\_expr norm\_scaled\_expr scaled expression values custom\_expr custom expression values cell\_metadata cell annotation metadata gene\_metadata gene annotation metadata spatial\_network list of spatial network(s) spatial\_network\_name list of spatial network name(s) list of spatial grid(s) spatial\_grid spatial\_grid\_name list of spatial grid name(s) spatial\_enrichment list of spatial enrichment score(s) for each spatial region spatial\_enrichment\_name list of spatial enrichment name(s) dimension\_reduction list of dimension reduction(s) list of nearest neighbor network(s) nn\_network file used to stitch fields together (optional) offset\_file list of instructions or output result from createGiottoInstructions instructions

# Details

[Requirements] To create a giotto object you need to provide at least a matrix with genes as row names and cells as column names. To include spatial information about cells (or regions) you need to provide a data.table or data.frame with coordinates for all spatial dimensions. This can be 2D (x and y) or 3D (x, y, x). The row order for the cell coordinates should be the same as the column order for the provided expression data.

[Instructions] Additionally an instruction file, generated manually or with createGiottoInstructions can be provided to instructions, if not a default instruction file will be created for the Giotto object.

[Multiple fields] In case a dataset consists of multiple fields, like seqFISH+ for example, an offset file can be provided to stitch the different fields together. stitchFieldCoordinates can be used to generate such an offset file.

[**Processed data**] Processed count data, such as normalized data, can be provided using one of the different expression slots (norm\_expr, norm\_scaled\_expr, custom\_expr).

[Metadata] Cell and gene metadata can be provided using the cell and gene metadata slots. This data can also be added afterwards using the addGeneMetadata or addCellMetadata functions.

[Other information] Additional information can be provided through the appropriate slots:

- · spatial networks
- · spatial girds
- spatial enrichments
- · dimensions reductions
- · nearest neighbours networks

48 createHeatmap\_DT

#### Value

```
giotto object
```

## **Examples**

```
createGiottoObject(raw_exprs, spatial_locs)
```

createHeatmap\_DT

createHeatmap\_DT

# **Description**

creates order for clusters

### Usage

```
createHeatmap_DT(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cluster_cor_method = "pearson",
  cluster_hclust_method = "ward.D",
  gene_order = c("custom", "correlation"),
  gene_custom_order = NULL,
  gene_cor_method = "pearson",
  gene_hclust_method = "complete"
)
```

# **Arguments**

```
gobject
                 giotto object
expression_values
                 expression values to use
genes
                 genes to use
cluster_column name of column to use for clusters
                 method to determine cluster order
cluster_order
cluster_custom_order
                 custom order for clusters
cluster_cor_method
                 method for cluster correlation
cluster_hclust_method
                 method for hierarchical clustering of clusters
gene_order
                 method to determine gene order
gene_custom_order
                 custom order for genes
gene_cor_method
                 method for gene correlation
gene_hclust_method
                 method for hierarchical clustering of genes
```

createNearestNetwork 49

#### **Details**

Creates input data.tables for plotHeatmap function.

#### Value

list

### **Examples**

```
createHeatmap_DT(gobject)
```

createNearestNetwork createNearestNetwork

# **Description**

create a nearest neighbour network based on previously computed dimension reductions

# Usage

```
createNearestNetwork(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  type = c("sNN", "kNN"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  genes_to_use = NULL,
  name = "sNN.pca",
  return_gobject = TRUE,
  k = 30,
  minimum_shared = 5,
  top_shared = 3,
  verbose = T,
  ...
)
```

# **Arguments**

50 createSpatialEnrich

```
genes_to_use if dim_reduction_to_use = NULL, which genes to use

name arbitrary name for NN network

return_gobject boolean: return giotto object (default = TRUE)

k number of k neighbors to use

minimum_shared minimum shared neighbors

top_shared keep at ...

verbose be verbose

... additional parameters
```

#### **Details**

Description of nearest neighbor network creation and filter steps.

#### Value

giotto object with updated NN network

# **Examples**

```
createNearestNetwork(gobject)
```

```
createSpatialEnrich createSpatialEnrich
```

# Description

Function to calculate gene signature enrichment scores per spatial position using a hypergeometric test.

```
createSpatialEnrich(
  gobject,
  enrich_method = c("PAGE", "rank", "hypergeometric"),
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore"),
  name = "PAGE",
  return_gobject = TRUE
)
```

createSpatialGrid 51

# **Arguments**

```
gobject
                  Giotto object
                  method for gene signature enrichment calculation
enrich_method
                  Matrix of signature genes for each cell type / process
sign_matrix
expression_values
                  expression values to use
reverse_log_scale
                  reverse expression values from log scale
                  log base to use if reverse_log_scale = TRUE
logbase
output_enrichment
                  how to return enrichment output
                  to give to spatial enrichment results, default = PAGE
return_gobject return giotto object
```

# **Details**

For details see the individual functions:

```
PAGE: PAGEEnrichPAGE: rankEnrichPAGE: hyperGeometricEnrich
```

# Value

Giotto object or enrichment results if return\_gobject = FALSE

## **Examples**

```
createSpatialEnrich(gobject)
```

# **Description**

Create a spatial grid.

```
createSpatialGrid(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  sdimz_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

#### **Arguments**

```
gobject giotto object

sdimx_stepsize stepsize along the x-axis

sdimy_stepsize stepsize along the y-axis

sdimz_stepsize stepsize along the z-axis

minimum_padding

minimum padding on the edges

name name for spatial grid (default = 'spatial_grid')

return_gobject boolean: return giotto object (default = TRUE)
```

#### **Details**

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

createSpatialGrid\_2D

#### Value

giotto object with updated spatial grid slot

#### **Examples**

```
createSpatialGrid(gobject)
```

```
createSpatialGrid\_2D createSpatialGrid\_2D
```

# **Description**

create a spatial grid for 2D spatial data.

#### Usage

```
createSpatialGrid_2D(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

#### **Arguments**

createSpatialGrid\_3D

#### **Details**

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

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#### Value

giotto object with updated spatial grid slot

# **Examples**

```
createSpatialGrid_2D(gobject)
```

```
createSpatialGrid_3D createSpatialGrid_3D
```

#### **Description**

Create a spatial grid for 3D spatial data.

#### Usage

```
createSpatialGrid_3D(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  sdimz_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

# **Arguments**

```
gobject giotto object

sdimx_stepsize stepsize along the x-axis

sdimy_stepsize stepsize along the y-axis

sdimz_stepsize stepsize along the z-axis

minimum_padding

minimum padding on the edges

name name for spatial grid (default = 'spatial_grid')

return_gobject boolean: return giotto object (default = TRUE)
```

# **Details**

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

#### Value

giotto object with updated spatial grid slot

54 createSpatialNetwork

#### **Examples**

```
createSpatialGrid_3D(gobject)
```

```
createSpatialNetwork createSpatialNetwork
```

# **Description**

Create a spatial network based on cell centroid physical distances.

## Usage

```
createSpatialNetwork(
  gobject,
  k = 4,
  dimensions = "all",
  maximum_distance = NULL,
  minimum_k = 0,
  name = "spatial_network",
  verbose = F,
  return_gobject = TRUE
)
```

#### **Arguments**

```
gobject giotto object
k number of nearest neighbors based on physical distance
dimensions which spatial dimensions to use (default = all)
maximum_distance
distance cuttof for nearest neighbors to consider
minimum_k minimum nearest neighbours if maximum_distance != NULL
name name for spatial network (default = 'spatial_network')
verbose verbose
return_gobject boolean: return giotto object (default = TRUE)
```

# **Details**

Creates a spatial network connecting single-cells based on their physical distance to each other. Number of neighbors can be determined by k, maximum distance from each cell with or without setting a minimum k for each cell.

**dimensions:** default = 'all' which takes all possible dimensions. Alternatively you can provide a character vector that specififies the spatial dimensions to use, e.g. c("sdimx', "sdimy") or a numerical vector, e.g. 2:3

**maximum\_distance:** to create a network based on maximum distance only, you also need to set k to a very high value, e.g. k = 100

#### Value

giotto object with updated spatial network slot

## **Examples**

```
createSpatialNetwork(gobject)
```

# Description

calculates average gene detection for a cell metadata factor (e.g. cluster)

# Usage

```
create_average_detection_DT(
  gobject,
  meta_data_name,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0
)
```

# **Arguments**

## Value

data.table with average gene epression values for each factor

# **Description**

calculates average gene expression for a cell metadata factor (e.g. cluster)

```
create_average_DT(
  gobject,
  meta_data_name,
  expression_values = c("normalized", "scaled", "custom")
)
```

# **Arguments**

```
gobject giotto object

meta_data_name name of metadata column to use
expression_values

which expression values to use
```

#### Value

data.table with average gene epression values for each factor

# Description

creates randomized cell ids within a selection of cell types

# Usage

```
create_cell_type_random_cell_IDs(
  gobject,
  cluster_column = "cell_types",
  needed_cell_types
)
```

# **Arguments**

```
gobject giotto object to use

cluster_column cluster column with cell type information

needed_cell_types

vector of cell type names for which a random id will be found
```

# Details

Details will follow.

#### Value

list of randomly sampled cell ids with same cell type composition

# **Examples**

```
create_cell_type_random_cell_IDs(gobject)
```

create\_cluster\_matrix 57

```
create_cluster_matrix create_cluster_matrix
```

# **Description**

creates aggregated matrix for a given clustering

# Usage

```
create_cluster_matrix(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  gene_subset = NULL
)
```

# **Examples**

```
create_cluster_matrix(gobject)
```

create\_dimObject

create\_dimObject

# **Description**

Creates an object that stores a dimension reduction output

# Usage

```
create_dimObject(
  name = "test",
  reduction_method = NULL,
  coordinates = NULL,
  misc = NULL
)
```

# **Arguments**

```
name arbitrary name for object
reduction_method
method used to reduce dimensions
coordinates accepts the coordinates after dimension reduction
misc any additional information will be added to this slot
```

#### Value

number of distinct colors

58 decide\_cluster\_order

```
decide_cluster_order
```

# **Description**

creates order for clusters

# Usage

```
decide_cluster_order(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cor_method = "pearson",
  hclust_method = "ward.D"
)
```

# **Arguments**

```
gobject giotto object
expression_values
expression values to use
genes genes to use
cluster_column name of column to use for clusters
cluster_order method to determine cluster order
cluster_custom_order
custom order for clusters

cor_method method for correlation
hclust_method method for hierarchical clustering
```

## **Details**

Calculates order for clusters.

#### Value

custom

# **Examples**

```
decide_cluster_order(gobject)
```

detectSpatialPatterns 59

detectSpatialPatterns detectSpatialPatterns

# Description

Identify spatial patterns through PCA on average expression in a spatial grid.

# Usage

```
detectSpatialPatterns(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  spatial_grid_name = "spatial_grid",
  min_cells_per_grid = 4,
  scale_unit = F,
  ncp = 100,
  show_plot = T,
  PC_zscore = 1.5
)
```

# Arguments

## **Details**

Steps to identify spatial patterns:

- 1. average gene expression for cells within a grid, see createSpatialGrid
- 2. perform PCA on the average grid expression profiles
- 3. convert variance of principlal components (PCs) to z-scores and select PCs based on a z-score threshold

# Value

```
spatial pattern object 'spatPatObj'
```

# **Examples**

```
detectSpatialPatterns(gobject)
```

60 dimCellPlot

dimCellPlot

dimCellPlot

# Description

Visualize cells according to dimension reduction coordinates

# Usage

```
dimCellPlot(gobject, ...)
```

## **Arguments**

```
gobject
                 giotto object
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
cell_annotation_values
                 numeric cell annotation columns
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
cell_color
                 color for cells (see details)
color_as_factor
                 convert color column to factor
cell_color_code
                 named vector with colors
cell_color_gradient
                 vector with 3 colors for numeric data
gradient_midpoint
                 midpoint for color gradient
gradient_limits
                 vector with lower and upper limits
select_cell_groups
                 select subset of cells/clusters based on cell_color parameter
                 select subset of cells based on cell IDs
select_cells
show_other_cells
                 display not selected cells
other_cell_color
                 color of not selected cells
other_point_size
                 size of not selected cells
```

dimCellPlot2D 61

```
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
                  title for plot, defaults to cell_color parameter
title
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

# **Details**

Description of parameters. For 3D plots see dimCellPlot2D

## Value

ggplot

# **Examples**

dimCellPlot(gobject)

dimCellPlot2D dimCellPlot2D

# **Description**

Visualize cells according to dimension reduction coordinates

62 dimCellPlot2D

```
dimCellPlot2D(
     gobject,
     dim_reduction_to_use = "umap",
     dim_reduction_name = "umap",
     dim1_to_use = 1,
     dim2_to_use = 2,
     spat_enr_names = NULL,
     cell_annotation_values,
     show_NN_network = F,
     nn_network_to_use = "sNN",
     network_name = "sNN.pca",
     cell_color_gradient = c("blue", "white", "red"),
     gradient_midpoint = NULL,
     gradient_limits = NULL,
     select_cell_groups = NULL,
     select_cells = NULL,
     show_other_cells = T,
     other_cell_color = "lightgrey",
     other_point_size = 0.5,
     show_cluster_center = F,
      show_center_label = T,
     center_point_size = 4,
     center_point_border_col = "black",
     center_point_border_stroke = 0.1,
     label_size = 4,
     label_fontface = "bold",
     edge_alpha = NULL,
     point_size = 1,
     point_border_col = "black",
     point_border_stroke = 0.1,
     show_legend = T,
     cow_n_col = 2,
     cow_rel_h = 1,
     cow_rel_w = 1,
     cow_align = "h",
     show_plot = NA,
     return_plot = NA,
     save_plot = NA,
     save_param = list(),
     default_save_name = "dimCellPlot2D"
Arguments
```

```
giotto object
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
```

spat\_enr\_names names of spatial enrichment results to include cell\_annotation\_values numeric cell annotation columns show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels edge\_alpha column to use for alpha of the edges point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_legend show legend show plot show\_plot return\_plot return ggplot object save\_plot directly save the plot [boolean] save\_param list of saving parameters from all\_plots\_save\_function() default\_save\_name default save name for saving, don't change, change save\_name in save\_param cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors title for plot, defaults to cell\_color parameter title

64 dimGenePlot

#### **Details**

Description of parameters. For 3D plots see dimPlot3D

#### Value

ggplot

#### **Examples**

dimCellPlot2D(gobject)

dimGenePlot

dimGenePlot

# **Description**

Visualize cells and gene expression according to dimension reduction coordinates

# Usage

```
dimGenePlot(gobject, ...)
```

# **Arguments**

```
giotto object
gobject
                 parameters for cowplot::save_plot()
expression_values
                 gene expression values to use
                 genes to show
genes
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
{\tt network\_name}
edge_alpha
                 column to use for alpha of the edges
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
                 size of point (cell)
point_size
point_border_col
                 color of border around points
point_border_stroke
```

stroke size of border around points

dimGenePlot2D 65

```
midpoint
                  size of point (cell)
                  cowplot param: how many columns
cow_n_col
                  cowplot param: relative height
cow_rel_h
                  cowplot param: relative width
cow_rel_w
                  cowplot param: how to align
cow_align
show_legend
                  show legend
show_plot
                  show plots
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

#### **Details**

Description of parameters.

### Value

ggplot

#### See Also

dimGenePlot3D

# **Examples**

```
dimGenePlot(gobject)
```

dimGenePlot2D dimGenePlot2D

# **Description**

Visualize cells and gene expression according to dimension reduction coordinates

```
dimGenePlot2D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
```

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```
network_color = "lightgray",
      edge_alpha = NULL,
      scale_alpha_with_expression = FALSE,
      point_size = 1,
      genes_high_color = "red",
      genes_mid_color = "white",
      genes_low_color = "blue",
      point_border_col = "black",
      point_border_stroke = 0.1,
      midpoint = 0,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_legend = T,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dimGenePlot2D"
    )
Arguments
   gobject
                     giotto object
    expression_values
                     gene expression values to use
                     genes to show
    genes
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
   dim2_to_use
                     dimension to use on y-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
    edge_alpha
                     column to use for alpha of the edges
    scale_alpha_with_expression
                     scale expression with ggplot alpha parameter
                     size of point (cell)
    point_size
   point_border_col
                     color of border around points
   point_border_stroke
                     stroke size of border around points
   midpoint
                     size of point (cell)
                     cowplot param: how many columns
    cow_n_col
```

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```
cowplot param: relative height
cow_rel_h
                  cowplot param: relative width
cow_rel_w
                  cowplot param: how to align
cow_align
show_legend
                  show legend
show_plot
                  show plots
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  parameters for cowplot::save_plot()
```

#### **Details**

Description of parameters.

#### Value

ggplot

#### See Also

dimGenePlot3D

# **Examples**

```
dimGenePlot2D(gobject)
```

dimGenePlot3D

dimGenePlot3D

# Description

Visualize cells and gene expression according to dimension reduction coordinates

```
dimGenePlot3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = 3,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
```

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```
network_color = "lightgray",
      cluster_column = NULL,
      select_cell_groups = NULL,
      select_cells = NULL,
      show_other_cells = T,
      other_cell_color = "lightgrey",
      other_point_size = 1,
      edge_alpha = NULL,
      point_size = 2,
      genes_high_color = NULL,
      genes_mid_color = "white",
      genes_low_color = "blue",
      show_legend = T,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dimGenePlot3D"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
                     genes to show
    genes
   dim_reduction_to_use
                     dimension reduction to use
   dim_reduction_name
                     dimension reduction name
   dim1_to_use
                     dimension to use on x-axis
   dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    show_NN_network
                     show underlying NN network
   nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
   network_name
    edge_alpha
                     column to use for alpha of the edges
    point_size
                     size of point (cell)
    show_legend
                     show legend
    show_plot
                     show plots
    return_plot
                     return ggplot object
    save_plot
                     directly save the plot [boolean]
                     list of saving parameters from all_plots_save_function()
    save_param
    default_save_name
                     default save name for saving, don't change, change save_name in save_param
                     parameters for cowplot::save_plot()
    . . .
```

dimPlot2D 69

#### **Details**

Description of parameters.

#### Value

ggplot

#### **Examples**

dimGenePlot3D(gobject)

dimPlot2D

dimPlot2D

# **Description**

Visualize cells according to dimension reduction coordinates Visualize cells according to dimension reduction coordinates

```
dimPlot2D(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 1,
```

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```
point_border_col = "black",
      point_border_stroke = 0.1,
      title = NULL,
      show_legend = T,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dimPlot2D"
    dimPlot(gobject, ...)
Arguments
    gobject
                     giotto object
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
                     dimension to use on y-axis
    dim2_to_use
    spat_enr_names names of spatial enrichment results to include
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_cluster_center
                     plot center of selected clusters
```

dimPlot3D 71

```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
title
                  title for plot, defaults to cell_color parameter
                  show legend
show_legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
```

default save name for saving, don't change, change save\_name in save\_param

# **Details**

```
Description of parameters. For 3D plots see dimPlot3D Description of parameters, see dimPlot2D. For 3D plots see dimPlot3D
```

#### Value

ggplot ggplot

# **Examples**

```
dimPlot2D(gobject)
dimPlot2D(gobject)
```

dimPlot3D dimPlot3D

# Description

Visualize cells according to dimension reduction coordinates

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#### Usage

```
dimPlot3D(
      gobject,
      dim_reduction_to_use = "umap",
      dim_reduction_name = "umap",
      dim1_to_use = 1,
      dim2_to_use = 2,
      dim3_to_use = 3,
      select_cell_groups = NULL,
      select_cells = NULL,
      show_other_cells = T,
      other_cell_color = "lightgrey",
      other_point_size = 2,
      show_NN_network = F,
      nn_network_to_use = "sNN",
      network_name = "sNN.pca",
      color_as_factor = T,
      cell_color = NULL,
      cell_color_code = NULL,
      show_cluster_center = F,
      show_center_label = T,
      center_point_size = 4,
      label_size = 4,
      edge_alpha = NULL,
      point_size = 3,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dim3D"
Arguments
    gobject
                    giotto object
    dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   dim3_to_use
                    dimension to use on z-axis
    select_cell_groups
                    select subset of cells/clusters based on cell_color parameter
                    select subset of cells based on cell IDs
    select_cells
    show_other_cells
                    display not selected cells
   other_cell_color
                    color of not selected cells
   other_point_size
```

size of not selected cells

dimPlot3D 73

show\_NN\_network

show underlying NN network

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

network\_name name of NN network to use, if show\_NN\_network = TRUE

color\_as\_factor

convert color column to factor

cell\_color color for cells (see details)

cell\_color\_code

named vector with colors

show\_cluster\_center

plot center of selected clusters

show\_center\_label

plot label of selected clusters

center\_point\_size

size of center points

label\_size size of labels

edge\_alpha column to use for alpha of the edges

point\_size size of point (cell)

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

show\_legend show legend

### **Details**

Description of parameters.

### Value

plotly

## **Examples**

dimPlot3D(gobject)

74 doHclust

## **Description**

shows direction of change

### Usage

```
direction_test(x, min_fdr = 0.05)
```

### **Examples**

```
direction_test_CPG()
```

doHclust

doHclust

## **Description**

cluster cells using hierarchical clustering algorithm

## Usage

```
doHclust(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
  dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  distance_method = c("pearson", "spearman", "original", "euclidean", "maximum",
  "manhattan", "canberra", "binary", "minkowski"),
agglomeration_method = c("ward.D2", "ward.D", "single", "complete", "average",
    "mcquitty", "median", "centroid"),
  k = 10,
  h = NULL
  name = "hclust",
  return_gobject = TRUE,
  set\_seed = T,
  seed_number = 1234
)
```

```
gobject giotto object
expression_values
expression values to use
genes_to_use subset of genes to use
```

doHMRF 75

```
dim_reduction_to_use
                  dimension reduction to use
dim_reduction_name
                  dimensions reduction name
dimensions_to_use
                  dimensions to use
distance_method
                  distance method
{\tt agglomeration\_method}
                  agglomeration method for hclust
k
                  number of final clusters
h
                  cut hierarchical tree at height = h
                  name for hierarchical clustering
name
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                  set seed
seed\_number
                  number for seed
                  additional parameters
```

### **Details**

Description on how to use Kmeans clustering method.

#### Value

giotto object appended with new cluster

### **Examples**

doHclust(gobject)

doHMRF

doHMRF

# **Description**

Run HMRF

```
doHMRF(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  spatial_network_name = "spatial_network",
  spatial_genes = NULL,
  spatial_dimensions = c("sdimx", "sdimy", "sdimz"),
  dim_reduction_to_use = NULL,
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "test",
  k = 10,
```

76 doHMRF

```
betas = c(0, 2, 50),
tolerance = 1e-10,
zscore = c("none", "rowcol", "colrow"),
numinit = 100,
python_path = NULL,
output_folder = NULL,
overwrite_output = TRUE
)
```

## **Arguments**

```
giotto object
gobject
expression_values
                 expression values to use
spatial_network_name
                 name of spatial network to use for HMRF
                 spatial genes to use for HMRF
spatial_genes
spatial_dimensions
                 select spatial dimensions to use, default is all possible dimensions
dim_reduction_to_use
                 use another dimension reduction set as input
dim_reduction_name
                 name of dimension reduction set to use
dimensions_to_use
                 number of dimensions to use as input
                 name of HMRF run
name
k
                 number of HMRF domains
betas
                 betas to test for
tolerance
                 tolerance
zscore
                 zscore
numinit
                 number of initializations
                 python path to use
python_path
output_folder
                 output folder to save results
overwrite_output
                 overwrite output folder
```

## **Details**

Description of HMRF parameters ...

### Value

Creates a directory with results that can be viewed with viewHMRFresults

```
doHMRF(gobject)
```

doKmeans 77

doKmeans doKmeans

## **Description**

cluster cells using kmeans algorithm

### Usage

```
doKmeans(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
  dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  distance_method = c("original", "pearson", "spearman", "euclidean", "maximum",
    "manhattan", "canberra", "binary", "minkowski"),
  centers = 10,
  iter_max = 100,
  nstart = 1000,
  algorithm = "Hartigan-Wong",
  name = "kmeans",
  return_gobject = TRUE,
  set_seed = T,
  seed_number = 1234
)
```

```
gobject
                 giotto object
expression_values
                 expression values to use
genes_to_use
                 subset of genes to use
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimensions reduction name
dimensions_to_use
                 dimensions to use
distance\_method
                 distance method
                 number of final clusters
centers
                 kmeans maximum iterations
iter_max
nstart
                 kmeans nstart
algorithm
                 kmeans algorithm
                 name for kmeans clustering
return_gobject boolean: return giotto object (default = TRUE)
```

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```
set_seed set seed
seed_number number for seed
... additional parameters
```

### **Details**

Description on how to use Kmeans clustering method.

#### Value

giotto object appended with new cluster

# **Examples**

```
doKmeans(gobject)
```

doLeidenCluster

doLeidenCluster

# Description

cluster cells using a NN-network and the Leiden community detection algorithm

## Usage

```
doLeidenCluster(
  gobject,
  name = "leiden_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = "weight",
  partition_type = c("RBConfigurationVertexPartition", "ModularityVertexPartition"),
  init_membership = NULL,
  n_iterations = 1000,
  return_gobject = TRUE,
  set_seed = T,
  seed_number = 1234,
  ...
)
```

doLeidenSubCluster 79

#### **Details**

Description of Leiden clustering method.

### Value

giotto object appended with new cluster

## **Examples**

```
doLeidenCluster(gobject)
```

 ${\tt doLeidenSubCluster} \qquad {\tt doLeidenSubCluster}$ 

## **Description**

subcluster cells using a NN-network and the Leiden algorithm

```
doLeidenSubCluster(
  gobject,
  name = "sub_pleiden_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 0.5,
  n_{iterations} = 500,
  python_path = NULL,
```

80 doLeidenSubCluster

```
nn_network_to_use = "sNN",
network_name = "sNN.pca",
return_gobject = TRUE,
verbose = T,
...
)
```

## **Arguments**

gobject giotto object

name name for new clustering result cluster\_column cluster column to subcluster selected\_clusters

only do subclustering on these clusters

hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

hvg\_mean\_expr\_det

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for createNearestNetwork

k\_neighbors number of k for createNearestNetwork

resolution resolution of Leiden clustering

n\_iterations number of iterations

python\_path specify specific path to python if required

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

network\_name name of NN network to use

return\_gobject boolean: return giotto object (default = TRUE)

verbose verbose

... additional parameters

## **Details**

Description of Leiden clustering method.

# Value

giotto object appended with new cluster

```
\\ do Leiden Sub Cluster (gobject)
```

doLouvainCluster 81

doLouvainCluster doLouvainCluster

## **Description**

cluster cells using a NN-network and the Louvain algorithm.

#### Usage

```
doLouvainCluster(
  gobject,
  version = c("community", "multinet"),
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = NULL,
  gamma = 1,
  omega = 1,
  louv_random = F,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
)
```

# Arguments

```
gobject
                 giotto object
                 implemented version of Louvain clustering to use
version
name
                 name for cluster
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
python_path
                 specify specific path to python if required
resolution
                 resolution
gamma
                 gamma
omega
                 omega
return_gobject boolean: return giotto object (default = TRUE)
                 set seed
set_seed
seed_number
                 number for seed
                 additional parameters
```

## **Details**

Louvain clustering using the community or multinet implementation of the louvain clustering algorithm.

### Value

giotto object appended with new cluster

## **Examples**

```
doLouvainCluster(gobject)
```

```
\begin{tabular}{ll} do Louvain Cluster\_community \\ & do Louvain Cluster\_community \\ \end{tabular}
```

# Description

cluster cells using a NN-network and the Louvain algorithm from the community module in Python

## Usage

```
doLouvainCluster_community(
   gobject,
   name = "louvain_clus",
   nn_network_to_use = "sNN",
   network_name = "sNN.pca",
   python_path = NULL,
   resolution = 1,
   weight_col = NULL,
   louv_random = F,
   return_gobject = TRUE,
   set_seed = F,
   seed_number = 1234,
   ...
)
```

```
gobject
                 giotto object
                 name for cluster
name
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
                 specify specific path to python if required
python_path
resolution
                 resolution
weight_col
                 weight column
louv_random
                 random
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                 set seed
                 number for seed
seed_number
                 additional parameters
```

### **Details**

Description of Leiden clustering method.

### Value

giotto object appended with new cluster

### **Examples**

```
doLouvainCluster_community(gobject)
```

```
{\tt doLouvainCluster\_multinet}
```

doLouvainCluster\_multinet

## **Description**

cluster cells using a NN-network and the Louvain algorithm from the multinet package in R.

# Usage

```
doLouvainCluster_multinet(
  gobject,
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  weight_col = NULL,
  gamma = 1,
  omega = 1,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

```
gobject
                 giotto object
                 name for cluster
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
                 gamma
gamma
omega
                 omega
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                 set seed
seed_number
                 number for seed
                 additional parameters
                 specify specific path to python if required
python_path
```

84 doLouvainSubCluster

### **Details**

See louvain algorithm from the multinet package in R.

### Value

giotto object appended with new cluster

### **Examples**

```
doLouvainCluster_multinet(gobject)
```

 ${\tt doLouvainSubCluster} \qquad {\tt doLouvainSubCluster}$ 

## Description

subcluster cells using a NN-network and the Louvain algorithm

## Usage

```
doLouvainSubCluster(
  gobject,
  name = "sub_louvain_clus",
  version = c("community", "multinet"),
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 0.5,
  gamma = 1,
  omega = 1,
  python_path = NULL,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T,
)
```

```
gobject giotto object

name name for new clustering result

version version of Louvain algorithm to use
```

doLouvainSubCluster 85

cluster\_column cluster column to subcluster

selected\_clusters

only do subclustering on these clusters

hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

hvg\_mean\_expr\_det

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for createNearestNetwork

k\_neighbors number of k for createNearestNetwork
resolution resolution for community algorithm

gamma gamma

omega omega

python\_path specify specific path to python if required

 $nn\_network\_to\_use$ 

type of NN network to use (kNN vs sNN)

 ${\tt network\_name} \quad \quad name \ of \ NN \ network \ to \ use$ 

return\_gobject boolean: return giotto object (default = TRUE)

verbose verbose

... additional parameters

### **Details**

Description of Louvain clustering method.

### Value

giotto object appended with new cluster

### **Examples**

doLouvainSubCluster(gobject)

```
\label{loss} do Louvain SubCluster\_community \\ do Louvain SubCluster\_community
```

## **Description**

subcluster cells using a NN-network and the Louvain community detection algorithm

## Usage

```
doLouvainSubCluster_community(
  gobject,
  name = "sub_louvain_comm_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 0.5,
  python_path = NULL,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T,
)
```

```
gobject
                  giotto object
                  name for new clustering result
name
cluster_column cluster column to subcluster
selected_clusters
                  only do subclustering on these clusters
                  parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                  threshold for detection in min percentage of cells
hvg_mean_expr_det
                  threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                  forces all genes to be HVG and to be used as input for PCA
                  minimum number of HVG, or all genes will be used as input for PCA
min_nr_of_hvg
                  parameters for runPCA
pca_param
```

```
nn_param
                 parameters for parameters for createNearestNetwork
                 number of k for createNearestNetwork
k_neighbors
resolution
                 resolution
python_path
                 specify specific path to python if required
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
return_gobject boolean: return giotto object (default = TRUE)
verbose
                 verbose
                 additional parameters
```

#### **Details**

Description of Leiden clustering method.

#### Value

giotto object appended with new cluster

## **Examples**

```
doLouvainSubCluster_community(gobject)
```

```
\label{lower_multinet} do Louvain SubCluster\_multinet \\ do Louvain SubCluster\_multinet
```

## **Description**

subcluster cells using a NN-network and the Louvain multinet detection algorithm

```
doLouvainSubCluster_multinet(
 gobject,
 name = "sub_louvain_mult_clus",
 cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 10,
 gamma = 1,
  omega = 1,
```

```
nn_network_to_use = "sNN",
network_name = "sNN.pca",
return_gobject = TRUE,
verbose = T,
...
)
```

## **Arguments**

gobject giotto object

name name for new clustering result cluster\_column cluster column to subcluster selected\_clusters

only do subclustering on these clusters

hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

hvg\_mean\_expr\_det

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for createNearestNetwork

k\_neighbors number of k for createNearestNetwork

gamma gamma omega omega nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

network\_name name of NN network to use

return\_gobject boolean: return giotto object (default = TRUE)

verbose verbose

... additional parameters

python\_path specify specific path to python if required

### **Details**

Description of Louvain clustering method.

# Value

giotto object appended with new cluster

```
{\tt doLouvainSubCluster\_multinet(gobject)}
```

doRandomWalkCluster 89

doRandomWalkCluster doRandomWalkCluster

## **Description**

Cluster cells using a random walk approach.

# Usage

```
doRandomWalkCluster(
  gobject,
  name = "random_walk_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  walk_steps = 4,
  walk_clusters = 10,
  walk_weights = NA,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

## **Arguments**

```
giotto object
gobject
name
                 name for cluster
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use
                 number of walking steps
walk_steps
walk_clusters
                 number of final clusters
walk_weights
                 cluster column defining the walk weights
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                 set seed
seed_number
                 number for seed
                 additional parameters
```

### **Details**

See random walk algorithm from the igraph package in R.

## Value

giotto object appended with new cluster

```
doRandomWalkCluster(gobject)
```

90 doSNNCluster

doSNNCluster

do SNN Cluster

## **Description**

Cluster cells using a SNN cluster approach.

## Usage

```
doSNNCluster(
  gobject,
  name = "sNN_clus",
  nn_network_to_use = "kNN",
  network_name = "kNN.pca",
  k = 20,
  eps = 4,
  minPts = 16,
  borderPoints = TRUE,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

## **Arguments**

gobject giotto object name name for cluster

nn\_network\_to\_use

type of NN network to use (only works on kNN)

 $network\_name$  name of kNN network to use

k Neighborhood size for nearest neighbor sparsification to create the shared NN

graph.

eps Two objects are only reachable from each other if they share at least eps nearest

neighbors.

minPts minimum number of points that share at least eps nearest neighbors for a point

to be considered a core points.

borderPoints should borderPoints be assigned to clusters like in DBSCAN?

return\_gobject boolean: return giotto object (default = TRUE)

set\_seed set seed

seed\_number number for seed

... additional parameters

## **Details**

See sNNclust algorithm from dbscan package

dt\_to\_matrix 91

### Value

giotto object appended with new cluster

# **Examples**

```
doSNNCluster(gobject)
```

dt\_to\_matrix

 $dt\_to\_matrix$ 

## **Description**

converts data.table to matrix

# Usage

```
dt_to_matrix(x)
```

# **Examples**

```
dt_to_matrix(x)
```

exportGiottoViewer

export Giot to Viewer

# Description

compute highly variable genes

```
exportGiottoViewer(
  gobject,
  output_directory = NULL,
  annotations,
  dim_reductions,
  dim_reduction_names,
  expression_values = c("normalized", "scaled", "custom"),
  dim_red_rounding = NULL,
  dim_red_rescale = c(-20, 20),
  expression_rounding = NULL,
  overwrite_dir = F,
  verbose = T
)
```

#### **Arguments**

```
gobject
                  giotto object
output_directory
                  directory where to save the files
                  giotto cell annotations to view
annotations
dim_reductions high level dimension reductions to view
dim_reduction_names
                  specific dimension reduction names
expression_values
                  expression values to use in Viewer
dim_red_rounding
                  numerical indicating how to round the coordinates
dim_red_rescale
                  numericals to rescale the coordinates
expression_rounding
                  numerical indicating how to round the expression data
overwrite_dir overwrite files in the directory if it already existed
verbose
                  be verbose
```

#### **Details**

Giotto Viewer expects the results from Giotto Analyzer in a specific format, which is provided by this function.

#### Value

writes the necessary output to use in Giotto Viewer

### **Examples**

```
exportGiottoViewer(gobject)
```

```
\label{eq:communication} exprOnlyCellCellcommunicationScores \\ exprOnlyCellCellcommunicationScores
```

### **Description**

Cell-Cell communication scores based on expression only

```
exprOnlyCellCellcommunicationScores(
  gobject,
  cluster_column = "cell_types",
  random_iter = 100,
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  verbose = T
)
```

extended\_gini\_fun 93

## **Arguments**

gobject giotto object to use

cluster\_column cluster column with cell type information

random\_iter number of iterations

gene\_set\_1 first specific gene set from gene pairs

gene\_set\_2 second specific gene set from gene pairs

log2FC\_addendum

addendum to add when calculating log2FC

verbose verbose

### **Details**

Details will follow.

## Value

Cell-Cell communication scores for gene pairs based on expression only

# **Examples**

exprOnlyCellCellcommunicationScores(gobject)

## **Description**

calculate gini coefficient on a minimum length vector

# Usage

```
extended_gini_fun(x, weights = rep(1, length = length(x)), minimum_length = 16)
```

# Value

gini coefficient

94 fDataDT

```
extractNearestNetwork extractNearestNetwork
```

# Description

Extracts a NN-network from a Giotto object as an igraph object

# Usage

```
extractNearestNetwork(
  gobject,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca"
)
```

# Arguments

```
\begin{array}{ccc} \text{gobject} & \text{giotto object} \\ \text{nn\_network\_to\_use} & & & \\ & & & & \\ & & & & \\ \text{NN or sNN} & \\ \text{network\_name} & & \text{name of NN network to be used} \end{array}
```

### Value

igraph object

# **Examples**

extractNearestNetwork(gobject)

 ${\sf fDataDT}$ 

fDataDT

# Description

show gene metadata

# Usage

```
fDataDT(gobject)
```

# Arguments

gobject giotto object

## Value

data.table with gene metadata

```
pDataDT(gobject)
```

filterCombinations 95

filterCombinations filterCombinations

### **Description**

Shows how many genes and cells are lost with combinations of thresholds.

#### Usage

```
filterCombinations(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_thresholds = c(1, 2),
  gene_det_in_min_cells = c(5, 50),
  min_det_genes_per_cell = c(200, 400),
  scale_x_axis = "identity",
  x_axis_offset = 0,
  scale_y_axis = "identity",
  y_axis_offset = 0,
  show_plot = TRUE
)
```

### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
expression_thresholds
                  all thresholds to consider a gene expressed
gene_det_in_min_cells
                  minimum number of cells that should express a gene to consider that gene fur-
                  ther
min_det_genes_per_cell
                  minimum number of expressed genes per cell to consider that cell further
scale_x_axis
                  ggplot transformation for x-axis (e.g. log2)
x_axis_offset x-axis offset to be used together with the scaling transformation
                  ggplot transformation for y-axis (e.g. log2)
scale_y_axis
y_axis_offset
                  y-axis offset to be used together with the scaling transformation
show_plot
                  show plot
```

## **Details**

Creates a scatterplot that visualizes the number of genes and cells that are lost with a specific combination of a gene and cell threshold given an arbitrary cutoff to call a gene expressed. This function can be used to make an informed decision at the filtering step with filterGiotto.

### Value

list of data.table and ggplot object

96 filterCPGscores

### **Examples**

```
filterCombinations(gobject)
```

filterCPGscores

filterCPGscores

## **Description**

visualize Cell Proximity Gene enrichment scores

# Usage

```
filterCPGscores(
   CPGscore,
   min_cells = 5,
   min_fdr = 0.05,
   min_spat_diff = 0.2,
   min_log2_fc = 0.5,
   keep_int_duplicates = TRUE,
   direction = c("both", "up", "down")
)
```

## **Arguments**

### **Details**

This function filters the output from getCellProximityGeneScores based on false-discovery rate, minimum absolute difference, minimum log fold-change and direction of change.

# Value

Gene to gene scores in data.table format

```
filterCPGscores(CPGscore)
```

filterDistributions 97

filterDistributions filterDistributions

## **Description**

show gene or cell distribution after filtering on expression threshold

# Usage

```
filterDistributions(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_threshold = 1,
  detection = c("genes", "cells"),
  plot_type = c("histogram", "violin"),
  nr_bins = 30,
  fill_color = "lightblue",
  scale_axis = "identity",
  axis_offset = 0,
  show_plot = TRUE
)
```

# Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
expression_threshold
                  threshold to consider a gene expressed
detection
                  consider genes or cells
plot_type
                  type of plot
nr_bins
                  number of bins for histogram plot
fill_color
                  fill color for plots
scale_axis
                  ggplot transformation for axis (e.g. log2)
                  offset to be used together with the scaling transformation
axis_offset
show_plot
                  show plot
```

## Value

ggplot object

```
filterDistributions(gobject)
```

98 filterGiotto

filterGiotto

filterGiotto

# Description

filter Giotto object based on expression threshold

## Usage

```
filterGiotto(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_threshold = 1,
  gene_det_in_min_cells = 100,
  min_det_genes_per_cell = 100,
  verbose = F
)
```

## **Arguments**

```
gobject giotto object

expression_values

expression values to use

expression_threshold

threshold to consider a gene expressed

gene_det_in_min_cells

minimum # of cells that need to express a gene

min_det_genes_per_cell

minimum # of genes that need to be detected in a cell

verbose

verbose
```

## **Details**

The function filterCombinations can be used to explore the effect of different parameter values.

### Value

giotto object

```
filterGiotto(gobject)
```

findGiniMarkers 99

findGiniMarkers

findGiniMarkers

### **Description**

Identify marker genes for selected clusters based on gini detection and expression scores.

### Usage

```
findGiniMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank_score = 1
)
```

### Arguments

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
subset_clusters
                  selection of clusters to compare
                  group 1 cluster IDs from cluster_column for pairwise comparison
group_1
                  group 2 cluster IDs from cluster_column for pairwise comparison
group_2
min_expr_gini_score
                  filter on minimum gini coefficient for expression
min_det_gini_score
                  filter minimum gini coefficient for detection
{\tt detection\_threshold}
                  detection threshold for gene expression
                  rank scores to include
rank_score
```

### **Details**

Description of parameters.

### Value

data.table with marker genes

```
find {\it GiniMarkers} (gobject)
```

## **Description**

Identify marker genes for all clusters based on gini detection and expression scores.

## Usage

```
findGiniMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  min_genes = 10,
  verbose = TRUE
)
```

# **Arguments**

```
giotto object
gobject
expression_values
                  gene expression values to use
cluster_column clusters to use
subset_clusters
                  selection of clusters to compare
min_expr_gini_score
                  filter on minimum gini coefficient on expression
min_det_gini_score
                  filter on minimum gini coefficient on detection
detection_threshold
                  detection threshold for gene expression
                  minimum genes to keep per cluster, overrides pval and logFC
min_genes
verbose
                  be verbose
```

#### **Details**

Description of parameters.

## Value

data.table with marker genes

```
findGiniMarkers_one_vs_all(gobject)
```

findMarkers 101

findMarkers findMarkers

### **Description**

Identify marker genes for selected clusters.

## Usage

```
findMarkers(
 gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
 method = c("scran", "gini", "mast"),
  subset_clusters = NULL,
 group_1 = NULL,
 group_2 = NULL,
 min_expr_gini_score = 0.5,
 min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank_score = 1,
  group_1_name = NULL,
  group_2_name = NULL,
  adjust_columns = NULL,
)
```

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
method
                  method to use to detect differentially expressed genes
subset_clusters
                  selection of clusters to compare
                  group 1 cluster IDs from cluster column for pairwise comparison
group_1
group_2
                  group 2 cluster IDs from cluster_column for pairwise comparison
min_expr_gini_score
                  gini: filter on minimum gini coefficient for expression
min_det_gini_score
                  gini: filter minimum gini coefficient for detection
detection_threshold
                  gini: detection threshold for gene expression
                  gini: rank scores to include
rank_score
                  mast: custom name for group_1 clusters
group_1_name
group_2_name
                  mast: custom name for group_2 clusters
adjust_columns mast: column in pDataDT to adjust for (e.g. detection rate)
                  additional parameters for the findMarkers function in scran or zlm function in
                  MAST
```

### **Details**

Wrapper for findScranMarkers, findGiniMarkers and FindMastMarkers.

### Value

data.table with marker genes

### **Examples**

```
findMarkers(gobject)
```

```
findMarkers_one_vs_all
```

findMarkers\_one\_vs\_all

# Description

Identify marker genes for all clusters.

# Usage

```
findMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  method = c("scran", "gini", "mast"),
  pval = 0.01,
  logFC = 0.5,
  min\_genes = 10,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank_score = 1,
  adjust_columns = NULL,
  verbose = TRUE,
)
```

findMastMarkers 103

```
logFC
                  scan & mast: filter on logFC
min_genes
                  minimum genes to keep per cluster, overrides pval and logFC
min_expr_gini_score
                  gini: filter on minimum gini coefficient for expression
min_det_gini_score
                  gini: filter minimum gini coefficient for detection
detection_threshold
                  gini: detection threshold for gene expression
                  gini: rank scores to include
rank_score
adjust_columns mast: column in pDataDT to adjust for (e.g. detection rate)
verbose
                  be verbose
                  additional parameters for the findMarkers function in scran or zlm function in
                  MAST
```

## **Details**

Wrapper for findScranMarkers\_one\_vs\_all, findGiniMarkers\_one\_vs\_all and FindMastMarkers\_one\_vs\_all.

#### Value

data.table with marker genes

### **Examples**

```
findMarkers_one_vs_all(gobject)
```

findMastMarkers

findMastMarkers

# Description

Identify marker genes for selected clusters based on the MAST package.

```
findMastMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  group_1 = NULL,
  group_1_name = NULL,
  group_2 = NULL,
  group_2_name = NULL,
  adjust_columns = NULL,
  ...
)
```

#### **Arguments**

```
gobject giotto object
expression_values
gene expression values to use

cluster_column clusters to use
group_1 group 1 cluster IDs from cluster_column for pairwise comparison
group_1_name custom name for group_1 clusters
group_2 group 2 cluster IDs from cluster_column for pairwise comparison
group_2_name custom name for group_2 clusters
adjust_columns column in pDataDT to adjust for (e.g. detection rate)
... additional parameters for the zlm function in MAST
```

#### Details

This is a minimal convenience wrapper around the MAST functions to detect differentially expressed genes.

### Value

data.table with marker genes

### **Examples**

```
findMastMarkers(gobject)
```

```
\label{lem:findMastMarkers_one_vs_all} find \textit{MastMarkers\_one\_vs\_all}
```

# Description

Identify marker genes for all clusters based on the MAST package.

```
findMastMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  adjust_columns = NULL,
  pval = 0.001,
  logFC = 1,
  min_genes = 10,
  verbose = TRUE,
  ...
)
```

findScranMarkers 105

# Arguments

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
subset_clusters
                  selection of clusters to compare
adjust_columns column in pDataDT to adjust for (e.g. detection rate)
pval
                  filter on minimal p-value
logFC
                  filter on logFC
                  minimum genes to keep per cluster, overrides pval and logFC
min_genes
verbose
                  be verbose
                  additional parameters for the zlm function in MAST
. . .
```

#### **Details**

This is a minimal convenience wrapper around the MAST functions to detect differentially expressed genes.

#### Value

data.table with marker genes

# **Examples**

```
findMastMarkers_one_vs_all(gobject)
```

findScranMarkers

findScranMarkers

## Description

Identify marker genes for selected clusters based on scran's implementation of findMarkers.

```
findScranMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  ...
)
```

## **Arguments**

```
gobject giotto object

expression_values

gene expression values to use

cluster_column clusters to use

subset_clusters

selection of clusters to compare

group_1 group 1 cluster IDs from cluster_column for pairwise comparison

group_2 group 2 cluster IDs from cluster_column for pairwise comparison

additional parameters for the findMarkers function in scran
```

### **Details**

This is a minimal convenience wrapper around the findMarkers function from the scran package.

### Value

data.table with marker genes

### **Examples**

```
findScranMarkers(gobject)
```

```
find Scran Markers\_one\_vs\_all \\ find Scran Markers\_one\_vs\_all
```

# Description

Identify marker genes for all clusters in a one vs all manner based on scran's implementation of findMarkers.

```
findScranMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  pval = 0.01,
  logFC = 0.5,
  min_genes = 10,
  verbose = TRUE,
  ...
)
```

find\_grid\_2D 107

## **Arguments**

gobject giotto object

expression\_values

gene expression values to use

cluster\_column clusters to use

subset\_clusters

subset of clusters to use

pval filter on minimal p-value

logFC filter on logFC

min\_genes minimum genes to keep per cluster, overrides pval and logFC

verbose be verbose

... additional parameters for the findMarkers function in scran

### **Details**

This is a minimal convenience wrapper around the findMarkers function from the scran package.

## Value

data.table with marker genes

# **Examples**

findScranMarkers\_one\_vs\_all(gobject)

find\_grid\_2D

 $find\_grid\_2D$ 

## **Description**

find grid location in 2D

# Usage

```
find_grid_2D(grid_DT, x_loc, y_loc)
```

find\_grid\_3D

find\_grid\_3D

# Description

find grid location in 3D

```
find_grid_3D(grid_DT, x_loc, y_loc, z_loc)
```

108 fish\_function

find\_grid\_x

 $find\_grid\_x$ 

# Description

find grid location on x-axis

# Usage

```
find_grid_x(grid_DT, x_loc)
```

find\_grid\_y

find\_grid\_y

# Description

find grid location on y-axis

# Usage

```
find_grid_y(grid_DT, y_loc)
```

find\_grid\_z

find\_grid\_z

# Description

find grid location on z-axis

# Usage

```
find_grid_z(grid_DT, z_loc)
```

fish\_function

fish\_function

# Description

perform fisher exact test

```
fish_function(x_to, x_from)
```

fish\_function2

fish\_function2

fish\_function2

## Description

perform fisher exact test

## Usage

```
fish_function2(A, B, C, D)
```

FSV\_show

FSV\_show

### **Description**

Visualize spatial varible genes caculated by spatial\_DE

## Usage

```
FSV_show(
  results,
  ms_results = NULL,
  size = c(4, 2, 1),
  color = c("blue", "green", "red"),
  sig_alpha = 0.5,
  unsig_alpha = 0.5
)
```

### **Arguments**

results results caculated by spatial\_DE
ms\_results ms\_results caculated by spatial\_DE
size indicate different levels of qval
color indicate different SV features
sig\_alpha transparency of significant genes
unsig\_alpha transparency of unsignificant genes

### **Details**

Description of parameters.

## Value

nothing

```
FSV_show(results)
```

110 GenePattern\_show

GenePattern\_show

GenePattern\_show

## Description

Visualize genes distribution patterns calculated by spatial\_AEH

## Usage

```
GenePattern_show(
  gobject = NULL,
  AEH_results = NULL,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_alpha = 1,
  low_color = "blue",
  mid_color = "white",
  high_color = "red",
  midpoint = 0
)
```

## **Arguments**

gobject giotto object results from spatial\_AEH AEH\_results sdimx x axis of spatial locus sdimy y axis of spatial locus point\_size size of points to indicate cells transparency of points to indicate cells point\_alpha low\_color color to indicate low score level color to indicate middle score level mid\_color high\_color color to indicate high score level point to set mid\_color midpoint

## Details

Description of parameters.

### Value

nothing

```
GenePattern_show(gobject,AEH_results)
```

general\_save\_function 111

```
general_save_function
```

## **Description**

Function to automatically save plots to directory of interest

### Usage

```
general_save_function(
   gobject,
   plot_object,
   save_dir = NULL,
   save_folder = NULL,
   save_name = NULL,
   default_save_name = "giotto_plot",
   save_format = c("png", "tiff", "pdf", "svg"),
   show_saved_plot = F,
   base_width = NULL,
   base_height = NULL,
   base_aspect_ratio = NULL,
   units = NULL,
   dpi = NULL,
   ...
)
```

## Arguments

```
gobject
                  giotto object
                  non-ggplot object to plot
plot_object
save_dir
                  directory to save to
save_folder
                  folder in save_dir to save to
                  name of plot
save_name
save_format
                  format (e.g. png, tiff, pdf, ...)
show_saved_plot
                  load & display the saved plot
base_width
                  width
base_height
                  height
base_aspect_ratio
                  aspect ratio
units
                  units
dpi
                  Plot resolution
```

```
general_save_function(gobject)
```

```
{\tt getCellProximityGeneScores} \\ {\tt getCellProximityGeneScores}
```

### **Description**

Compute cell-cell interaction enrichment (observed vs expected)

#### Usage

```
getCellProximityGeneScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "louvain_clus.1",
  selected_genes = NULL,
  expression_values = c("normalized", "scaled", "custom"),
  do_diff_test = TRUE,
  diff_test = c("t.test", "wilcox"),
  minimum_unique_cells = NA,
  fold_change_addendum = 0.1,
  in_two_directions = TRUE,
  exclude_selected_cells_from_test = F,
  verbose = T
)
```

```
gobject
                  giotto object
spatial\_network\_name
                  name of spatial network to use
cluster_column name of column to use for clusters
selected_genes selection of genes to perform calculations for
expression_values
                  expression values to use
do_diff_test
                  perform differential test
diff\_test
                  which differential expression test
minimum_unique_cells
                  minimum number of cells needed to proceed
fold_change_addendum
                  constant to add when calculating log2 fold-change
in_two_directions
                  shows enrichment in both directions: cell1-cell2, cell2-cell1
exclude_selected_cells_from_test
                  exclude certain cells from test
verbose
                  verbose
```

#### **Details**

Function to calculate if genes are differentially expressed in cell types when they interact (according to physical proximity) with other cell types. The results data.table contains the following columns:

- genes: All or selected list of tested genes
- cell\_expr\_1: average gene expression in cell type 1 from unified\_int cell-cell interaction
- cell\_expr\_2: average gene expression in cell type 2 from unified\_int cell-cell interaction
- comb\_expr: combined average gene expression in cell type 1 and 2 from unified\_int cell-cell interaction
- all\_cell\_expr\_1: average gene expression for all cells from cell type 1
- all\_cell\_expr\_2: average gene expression for all cells from cell type 2
- all comb expr: combined average gene expression for all cells from cell type 1 and 2
- pval\_1: p-value from test between interacting cells and all cells from cell type 1
- pval\_2: p-value from test between interacting cells and all cells from cell type 2
- cell\_type\_1: first cell type of cell-cell interaction
- cell type 2: second cell type of cell-cell interaction
- interaction: the cell-cell interaction, based on physical proximity
- nr\_1: number of cell type 1 in the unified cell-cell interaction
- nr\_2: number of cell type 2 in the unified cell-cell interaction
- all\_nr\_1: number of all cell type 1 in the whole dataset
- all\_nr\_2: number of all cell type 2 in the whole dataset
- diff\_spat: difference between comb\_expr and all\_comb\_expr
- diff\_spat\_1: difference between cell\_expr\_1 and all\_cell\_expr\_1
- diff\_spat\_2: difference between cell\_expr\_1 and all\_cell\_expr\_1
- log2fc spat 1: fold-change of diff spat 1
- log2fc\_spat\_2: fold-change of diff\_spat\_2
- log2fc\_spat: fold-change of diff\_spat
- type\_int: type of interaction
- unified\_int: interaction with alphabetically sorted cell type 1 and cell type 2
- unif\_int\_rank: 1 or 2
- fdr\_1: fdr from test between interacting cells and all cells from cell type 1
- fdr\_2: fdr from test between interacting cells and all cells from cell type 2

### Value

Cell Proximity Gene scores (CPGscores) in data.table format

### **Examples**

getCellProximityGeneScores(gobject)

114 getDendrogramSplits

```
getClusterSimilarity
getClusterSimilarity
```

### **Description**

Creates data.table with pairwise correlation scores between each cluster.

## Usage

```
getClusterSimilarity(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman")
)
```

### **Arguments**

```
gobject giotto object
expression_values
expression values to use
cluster_column name of column to use for clusters
cor correlation score to calculate distance
```

#### **Details**

Creates data.table with pairwise correlation scores between each cluster and the group size (# of cells) for each cluster. This information can be used together with mergeClusters to combine very similar or small clusters into bigger clusters.

## Value

data.table

### **Examples**

```
getClusterSimilarity(gobject)
```

```
getDendrogramSplits getDendrogramSplits
```

## Description

Split dendrogram at each node and keep the leave (label) information..

getDistinctColors 115

### Usage

```
getDendrogramSplits(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  h = NULL,
  h_color = "red",
  show_dend = TRUE,
  verbose = TRUE
)
```

## **Arguments**

```
giotto object
gobject
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
                  distance method to use for hierarchical clustering
distance
                  height of horizontal lines to plot
                  color of horizontal lines
h_color
show_dend
                  show dendrogram
verbose
                  be verbose
```

#### **Details**

Creates a data.table with three columns and each row represents a node in the dendrogram. For each node the height of the node is given together with the two subdendrograms. This information can be used to determine in a hierarchical manner differentially expressed marker genes at each node.

#### Value

data.table object

## **Examples**

```
getDendrogramSplits(gobject)
```

```
getDistinctColors
```

#### **Description**

Returns a number of distint colors based on the RGB scale

```
getDistinctColors(n)
```

#### **Arguments**

n

number of colors wanted

#### Value

number of distinct colors

```
getGeneToGeneScores
```

### **Description**

Compute gene-gene enrichment scores.

### Usage

```
getGeneToGeneScores(
   CPGscore,
   selected_genes = NULL,
   specific_genes_1 = NULL,
   specific_genes_2 = NULL,
   min_cells = 5,
   min_fdr = 0.05,
   min_spat_diff = 0.2,
   min_log2_fc = 0.5,
   direction = c("both", "up", "down"),
   fold_change_addendum = 0.1,
   verbose = TRUE
)
```

```
CPGscore
                  CPGscore, output from getCellProximityGeneScores()
selected_genes select subset of genes
specific_genes_1
                  specific source genes (see details)
specific_genes_2
                  specific target genes (see details)
                  min number of cells threshold
min_cells
                  spatial difference threshold
min_spat_diff
                  log2 fold-change threshold
min_log2_fc
direction
                  up or downregulation or both
fold_change_addendum
                  constant to add when calculating log2 fold-change
verbose
min_pval
                  p-value threshold
```

#### **Details**

Give more details ...

#### Value

Gene to gene scores in data.table format

#### **Examples**

```
getGeneToGeneScores(CPGscore)
```

### **Description**

creates unified cell-cell interaction names

#### Usage

```
get_cell_to_cell_sorted_name_conversion(all_cell_types)
```

### **Examples**

```
get_cell_to_cell_sorted_name_conversion()
```

## Description

Computes gene enrichment between all interactions

```
get_interaction_gene_enrichment(
   spatial_network,
   unified_int_col = "unified_int",
   source_col = "source_clus",
   source_IDs = "from",
   neighb_col = "neighb_clus",
   neighb_IDs = "to",
   expression_matrix,
   cell_annotation,
   annotation_ID = "uniq_ID",
   cell_type_col,
   do_diff_test = T,
   diff_test = c("t.test", "wilcox"),
```

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```
minimum_unique_cells = NA,
  exclude_selected_cells_from_test = T,
  verbose = T
)
```

## **Examples**

```
get_interaction_gene_enrichment()
```

### **Description**

Computes gene enrichment between specified interaction

## Usage

```
get_specific_interaction_gene_enrichment(
 sub_spatial_network,
 source_col = "source_clus",
 source_IDs = "from",
 neighb_col = "neighb_clus",
 neighb_IDs = "to",
 expression_matrix,
  interaction_name = "to_specify",
 cell_annotation,
 annotation_ID = "uniq_ID",
 cell_type_col,
 do_diff_test = T,
 diff_test = c("t.test", "wilcox"),
 minimum_unique_cells = NA,
 exclude_selected_cells_from_test = T
)
```

## **Examples**

```
get_specific_interaction_gene_enrichment()
```

```
ggplot_save_function ggplot_save_function
```

## Description

Function to automatically save plots to directory of interest

ggplot\_save\_function 119

to prevent the common error of specifying dimensions in pixels.

### Usage

```
ggplot_save_function(
  gobject,
  plot_object,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  default_save_name = "giotto_plot",
  save_format = NULL,
  show_saved_plot = F,
  ncol = 1,
  nrow = 1,
  scale = 1,
  base_width = NULL,
  base_height = NULL,
  base_aspect_ratio = NULL,
  units = NULL,
  dpi = NULL,
  limitsize = TRUE,
)
```

## **Arguments**

gobject

plot\_object ggplot object to plot save\_dir directory to save to save\_folder folder in save\_dir to save to save\_name name of plot format (e.g. png, tiff, pdf, ...) save\_format show\_saved\_plot load & display the saved plot ncol number of columns number of rows nrow scale scale base\_width width base\_height height base\_aspect\_ratio aspect ratio units units dpi Plot resolution limitsize When TRUE (the default), ggsave will not save images larger than 50x50 inches,

giotto object

## See Also

```
cowplot::save_plot
```

#### **Examples**

```
ggplot_save_function(gobject)
```

giotto-class

S4 giotto Class

#### **Description**

Framework of giotto object to store and work with spatial expression data

#### **Slots**

```
raw_exprs raw expression counts
norm_expr normalized expression counts
norm_scaled_expr normalized and scaled expression counts
custom_expr custom normalized counts
spatial_locs spatial location coordinates for cells
cell_metadata metadata for cells
gene_metadata metadata for genes
cell_ID unique cell IDs
gene_ID unique gene IDs
spatial_network spatial network in data.table/data.frame format
spatial_grid spatial grid in data.table/data.frame format
dimension_reduction slot to save dimension reduction coordinates
nn_network nearest neighbor network in igraph format
parameters slot to save parameters that have been used
instructions slot for global function instructions
offset_file offset file used to stitch together image fields
OS_platform Operating System to run Giotto analysis on
```

hyperGeometricEnrich hyperGeometricEnrich

## Description

Function to calculate gene signature enrichment scores per spatial position using a hypergeometric test.

```
hyperGeometricEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

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#### **Arguments**

# Details

The enrichment score is calculated based on the p-value from the hypergeometric test, -log10(p-value).

#### Value

data.table with enrichment results

## **Examples**

hyperGeometricEnrich(gobject)

iterCluster

iterCluster

#### **Description**

cluster cells iteratively

```
iterCluster(
 gobject,
 cluster_method = c("leiden", "louvain_community", "louvain_multinet"),
 nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
   = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 20,
 resolution = 1,
 gamma = 1,
 omega = 1,
 python_path = NULL,
```

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```
nn_network_to_use = "sNN",
network_name = "sNN.pca",
name = "iter_clus",
return_gobject = TRUE,
....
)
```

### **Arguments**

gobject giotto object

cluster\_method clustering algorithm to use
nr\_rounds number of iterative rounds
hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

hvg\_mean\_expr\_det

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for runPCA

k\_neighbors k for nn-network

resolution resolution gamma gamma omega omega

python\_path python path to use for Leiden clustering

 $nn\_network\_to\_use$ 

NN network to use

network\_name NN network name name of clustering

return\_gobject boolean: return giotto object (default = TRUE)

... additional parameters

### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

```
\verb|iterCluster(gobject)|
```

iterLeidenCluster 123

 $iter Leiden Cluster \hspace*{1.5cm} \textit{iter Leiden Cluster}$ 

## **Description**

cluster cells iteratively

#### Usage

```
iterLeidenCluster(
 gobject,
 name = "iter_clus",
 nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
   = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 20,
  resolution = 1,
 n_{iterations} = 1000,
 python_path = NULL,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 return_gobject = TRUE,
)
```

```
gobject
                  giotto object
name
                  name of clustering
nr_rounds
                  number of iterative rounds
hvg_param
                  parameters for calculateHVG
hvg_min_perc_cells
                  threshold for detection in min percentage of cells
hvg_mean_expr_det
                  threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                  forces all genes to be HVG and to be used as input for PCA
                  minimum number of HVG, or all genes will be used as input for PCA
min_nr_of_hvg
                  parameters for runPCA
pca_param
                  parameters for parameters for runPCA
nn_param
k_neighbors
                  k for nn-network
resolution
                  resolution for Leiden clustering
```

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```
n_iterations number of iterations for Leiden clustering

python_path python path to use for Leiden clustering

nn_network_to_use

NN network to use

network_name NN network name

return_gobject boolean: return giotto object (default = TRUE)

... additional parameters
```

#### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

#### **Examples**

```
iterLeidenCluster(gobject)
```

iterLouvainCluster

*iterLouvainCluster* 

### **Description**

cluster cells iteratively

```
iterLouvainCluster(
 gobject,
 version = c("community", "multinet"),
 nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
   = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 20,
 resolution = 1,
 gamma = 1,
 omega = 1,
 python_path = NULL,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 name = "iter_clus",
 return_gobject = TRUE,
)
```

iterLouvainCluster 125

### **Arguments**

gobject giotto object

version louvain clustering algorithm to use

nr\_rounds number of iterative rounds
hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

hvg\_mean\_expr\_det

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for runPCA

k\_neighbors k for nn-network

resolution resolution gamma gamma omega omega

python\_path python path to use for Leiden clustering

 $nn\_network\_to\_use$ 

NN network to use

network\_name NN network name name of clustering

return\_gobject boolean: return giotto object (default = TRUE)

... additional parameters

#### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

## **Examples**

iterLouvainCluster(gobject)

```
iter Louvain {\tt Cluster\_community} \\ iter Louvain {\tt Cluster\_community} \\
```

### **Description**

cluster cells iteratively

#### Usage

```
iterLouvainCluster_community(
 gobject,
 nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 20,
 resolution = 1,
 python_path = NULL,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 name = "iter_clus",
 return_gobject = TRUE,
)
```

```
gobject
                  giotto object
nr_rounds
                  number of iterative rounds
                  parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                  threshold for detection in min percentage of cells
hvg_mean_expr_det
                  threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                  forces all genes to be HVG and to be used as input for PCA
                  minimum number of HVG, or all genes will be used as input for PCA
min_nr_of_hvg
pca_param
                  parameters for runPCA
                  parameters for parameters for runPCA
nn_param
                  k for nn-network
k_neighbors
resolution
                  resolution for Leiden clustering
python_path
                  python path to use for Leiden clustering
```

#### **Details**

Description of iterative clustering.

#### Value

giotto object appended with new cluster

## **Examples**

```
iterLouvainCluster_community(gobject)
```

## Description

cluster cells iteratively

```
iterLouvainCluster_multinet(
  gobject,
  nr_rounds = 5,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 20,
  gamma = 1,
  omega = 1,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  name = "iter_clus",
  return_gobject = TRUE,
)
```

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#### **Arguments**

gobject giotto object

nr\_rounds number of iterative rounds hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

hvg\_mean\_expr\_det

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for runPCA

k\_neighbors k for nn-network

gamma gamma omega omega

nn\_network\_to\_use

NN network to use

network\_name NN network name name of clustering

return\_gobject boolean: return giotto object (default = TRUE)

... additional parameters

python\_path python path to use for Leiden clustering

### **Details**

Description of iterative clustering.

## Value

giotto object appended with new cluster

### **Examples**

iterLouvainCluster\_multinet(gobject)

kmeans\_binarize

kmeans\_binarize

#### **Description**

create binarized scores using kmeans

```
kmeans_binarize(x, nstart = 3, iter.max = 10)
```

loadHMRF

loadHMRF

loadHMRF

## Description

load previous HMRF

## Usage

```
loadHMRF(
  name_used = "test",
  output_folder_used,
  k_used = 10,
  betas_used,
  python_path_used
)
```

## **Arguments**

```
name_used name of HMRF that was run
output_folder_used
output folder that was used

k_used number of HMRF domains that was tested
betas_used betas that were tested
python_path_used
python path that was used
```

## **Details**

Description of HMRF parameters ...

#### Value

reloads a previous ran HMRF from doHRMF

## **Examples**

```
loadHMRF(gobject)
```

## Description

Simulate random network.

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#### Usage

```
make_simulated_network(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column,
  number_of_simulations = 100
)
```

### **Examples**

make\_simulated\_network(gobject)

mergeClusters

mergeClusters

### **Description**

Merge selected clusters based on pairwise correlation scores and size of cluster.

## Usage

```
mergeClusters(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  new_cluster_name = "merged_cluster",
  min_cor_score = 0.8,
  max_group_size = 20,
  force_min_group_size = 10,
  return_gobject = TRUE,
  verbose = TRUE
)
```

```
giotto object
gobject
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
new_cluster_name
                  new name for merged clusters
min_cor_score min correlation score to merge pairwise clusters
max_group_size max cluster size that can be merged
force_min_group_size
                  size of clusters that will be merged with their most similar neighbor(s)
return_gobject return giotto object
verbose
                  be verbose
```

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#### **Details**

Merge selected clusters based on pairwise correlation scores and size of cluster. To avoid large clusters to merge the max\_group\_size can be lowered. Small clusters can be forcibly merged with their most similar pairwise cluster by adjusting the force\_min\_group\_size parameter. Clusters smaller than this value will be merged independent on the provided min\_cor\_score value.

A giotto object is returned by default, if FALSE then the merging vector will be returned.

#### Value

Giotto object

## **Examples**

```
mergeClusters(gobject)
```

mygini\_fun

mygini\_fun

### **Description**

calculate gini coefficient

## Usage

```
mygini_fun(x, weights = rep(1, length(x)))
```

## Value

gini coefficient

nnDT\_to\_kNN

nnDT\_to\_kNN

### **Description**

Convert a nearest network data.table to a kNN object

## Usage

```
nnDT_to_kNN(nnDT)
```

### **Arguments**

nnDT

nearest neighbor network in data.table format

## Value

kNN object

normalizeGiotto

node\_clusters

node\_clusters

## Description

Merge selected clusters based on pairwise correlation scores and size of cluster.

## Usage

```
node_clusters(hclus_obj, verbose = TRUE)
```

## **Arguments**

```
hclus_obj hclus object
verbose be verbose
```

### Value

list of splitted dendrogram nodes from high to low node height

## **Examples**

```
node_clusters(hclus_obj)
```

normalizeGiotto

normalizeGiotto

## Description

normalize and/or scale expresion values of Giotto object

```
normalizeGiotto(
  gobject,
  norm_methods = c("standard", "osmFISH"),
  library_size_norm = TRUE,
  scalefactor = 6000,
  log_norm = TRUE,
  logbase = 2,
  scale_genes = T,
  scale_cells = T,
  scale_order = c("first_genes", "first_cells"),
  verbose = F
```

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### **Arguments**

gobject giotto object

norm\_methods normalization method to use

library\_size\_norm

normalize cells by library size

scalefactor scale factor to use after library size normalization

log\_norm transform values to log-scale

logbase log base to use to log normalize expression values

scale\_genes z-score genes over all cells scale\_cells z-score cells over all genes scale\_order order to scale genes and cells

verbose be verbose

#### **Details**

Currently there are two 'methods' to normalize your raw counts data.

A. The standard method follows the standard protocol which can be adjusted using the provided parameters and follows the following order:

- 1. Data normalization for total library size and scaling by a custom scale-factor.
- 2. Log transformation of data.
- 3. Z-scoring of data by genes and/or cells.
- B. The normalization method as provided by the osmFISH paper is also implemented:
  - 1. First normalize genes, for each gene divide the counts by the total gene count and multiply by the total number of genes.
  - 2. Next normalize cells, for each cell divide the normalized gene counts by the total counts per cell and multiply by the total number of cells.

This data will be saved in the Giotto slot for custom expression.

#### Value

giotto object

### **Examples**

 ${\tt normalizeGiotto(gobject)}$ 

OR\_function2 OR\_function2

#### **Description**

calculate odds-ratio

```
OR_function2(A, B, C, D)
```

PAGEEnrich

**PAGEEnrich** 

**PAGEEnrich** 

### **Description**

Function to calculate gene signature enrichment scores per spatial position using PAGE.

#### Usage

```
PAGEEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

## **Arguments**

```
gobject Giotto object

sign_matrix Matrix of signature genes for each cell type / process

expression_values

expression values to use

reverse_log_scale

reverse expression values from log scale

logbase log base to use if reverse_log_scale = TRUE

output_enrichment

how to return enrichment output
```

#### **Details**

The enrichment Z score is calculated by using method (PAGE) from Kim SY et al., BMC bioinformatics, 2005 as  $Z=((Sm\ \ mu)*m^(1/2))/delta$ . For each gene in each spot, mu is the fold change values versus the mean expression and delta is the standard deviation. Sm is the mean fold change value of a specific marker gene set and m is the size of a given marker gene set.

### Value

data.table with enrichment results

```
PAGEEnrich(gobject)
```

pDataDT

pDataDT

pDataDT

## Description

show cell metadata

## Usage

```
pDataDT(gobject)
```

## Arguments

gobject

giotto object

### Value

data.table with cell metadata

## **Examples**

```
pDataDT(gobject)
```

plotCPGscores

plotCPGscores

## Description

Create heatmap from cell-cell proximity scores

```
plotCPGscores(
   CPGscores,
   selected_interactions = NULL,
   selected_genes = NULL,
   detail_plot = T,
   simple_plot = F,
   simple_plot_facet = c("interaction", "genes"),
   facet_scales = "fixed",
   facet_ncol = length(selected_genes),
   facet_nrow = length(selected_interactions),
   show_plot = F
)
```

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#### **Arguments**

```
CPGscores
                  CPGscores, output from getCellProximityGeneScores()
selected_interactions
                  interactions to show
selected_genes genes to show
detail_plot
                  show detailed info in both interacting cell types
simple_plot
                  show a simplified plot
simple_plot_facet
                  facet on interactions or genes with simple plot
                  ggplot facet scales paramter
facet_scales
facet_ncol
                  ggplot facet ncol parameter
facet_nrow
                  ggplot facet nrow parameter
show_plot
                  show plot
```

#### **Details**

Give more details ...

#### Value

ggplot barplot

#### **Examples**

```
plotCPGscores(CPGscores)
```

plotGTGscores

plotGTGscores

### **Description**

Create heatmap from cell-cell proximity scores

```
plotGTGscores(
   gobject,
   GTGscore,
   selected_interactions = NULL,
   selected_gene_to_gene = NULL,
   detail_plot = T,
   simple_plot = F,
   simple_plot_facet = c("interaction", "genes"),
   facet_scales = "fixed",
   facet_ncol = length(selected_gene_to_gene),
   facet_nrow = length(selected_interactions),
   colors = c("blue", "red"),
   show_plot = NA,
   return_plot = NA,
```

plotGTGscores 137

```
save_plot = NA,
save_param = list(),
default_save_name = "plotGTGscores"
)
```

## Arguments

gobject giotto object GTGscore GTGscore, output from getGeneToGeneScores() selected\_interactions interactions to show show detailed info in both interacting cell types detail\_plot show a simplified plot simple\_plot simple\_plot\_facet facet on interactions or genes with simple plot facet\_scales ggplot facet scales paramter ggplot facet ncol parameter facet\_ncol facet\_nrow ggplot facet nrow parameter colors vector with 2 colors to represent respectively all and selected cells show\_plot show plots return\_plot return ggplot object directly save the plot [boolean] save\_plot list of saving parameters from all\_plots\_save\_function() save\_param

default save name for saving, don't change, change save\_name in save\_param

#### **Details**

Give more details ...

 ${\tt default\_save\_name}$ 

selected\_genes genes to show

## Value

ggplot barplot

```
plotGTGscores(GTGscore)
```

138 plotHeatmap

plotHeatmap

plotHeatmap

#### **Description**

creates order for clusters

#### Usage

```
plotHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cluster_color_code = NULL,
  cluster_cor_method = "pearson",
  cluster_hclust_method = "ward.D"
  gene_order = c("custom", "correlation"),
  gene_custom_order = NULL,
  gene_cor_method = "pearson",
  gene_hclust_method = "complete",
  show_values = c("rescaled", "z-scaled", "original"),
  size_vertical_lines = 1.1,
  gradient_colors = c("blue", "yellow", "red"),
  gene_label_selection = NULL,
  axis_text_y_size = NULL,
  legend_nrows = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotHeatmap"
)
```

```
gobject giotto object
expression_values
expression values to use
genes genes to use
cluster_column name of column to use for clusters
cluster_order method to determine cluster order
cluster_custom_order
custom order for clusters
cluster_color_code
color code for clusters
cluster_cor_method
method for cluster correlation
```

plotly\_axis\_scale\_2D 139

cluster\_hclust\_method

method for hierarchical clustering of clusters

gene\_order method to determine gene order

gene\_custom\_order

custom order for genes

gene\_cor\_method

method for gene correlation

gene\_hclust\_method

method for hierarchical clustering of genes

show\_values which values to show on heatmap

size\_vertical\_lines

sizes for vertical lines

gradient\_colors

colors for heatmap gradient

gene\_label\_selection

subset of genes to show on y-axis

axis\_text\_y\_size

size for y-axis text

legend\_nrows number of rows for the cluster legend

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

## **Details**

Creates heatmap for genes and clusters.

#### Value

ggplot

## **Examples**

plotHeatmap(gobject)

plotly\_axis\_scale\_2D plotly\_axis\_scale\_2D

## Description

adjust the axis scale in 3D plotly plot

plotly\_axis\_scale\_3D

### Usage

```
plotly_axis_scale_2D(
  cell_locations,
  sdimx = NULL,
  sdimy = NULL,
  mode = c("cube", "real", "custom"),
  custom_ratio = NULL
)
```

## Arguments

```
cell_locations spatial_loc in giotto object sdimx x axis of cell spatial location sdimy y axis of cell spatial location mode axis adjustment mode custom_ratio set the ratio artificially
```

#### Value

edges in spatial grid as data.table()

## **Examples**

```
plotly_axis_scale_2D(gobject)
```

```
plotly_axis_scale_3D plotly_axis_scale_3D
```

## Description

adjust the axis scale in 3D plotly plot

#### Usage

```
plotly_axis_scale_3D(
  cell_locations,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  mode = c("cube", "real", "custom"),
  custom_ratio = NULL
)
```

```
cell_locations spatial_loc in giotto object
sdimx x axis of cell spatial location
sdimy y axis of cell spatial location
sdimz z axis of cell spatial location
mode axis adjustment mode
custom_ratio set the ratio artificially
```

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#### Value

```
edges in spatial grid as data.table()
```

## **Examples**

```
plotly_axis_scale_3D(gobject)
```

plotly\_grid

plotly\_grid

### **Description**

provide grid segment to draw in plot\_ly()

## Usage

```
plotly_grid(
   spatial_grid,
   x_start = "x_start",
   y_start = "y_start",
   x_end = "x_end",
   y_end = "y_end"
)
```

## **Arguments**

```
spatial_grid spatial_grid in giotto object
```

## Value

edges in spatial grid as data.table()

### **Examples**

```
plotly_grid(gobject)
```

plotly\_network

plotly\_network

## Description

```
provide network segment to draw in 3D plot_ly()
```

#### Usage

```
plotly_network(
  network,
  x = "sdimx_begin",
  y = "sdimy_begin",
  z = "sdimz_begin",
  x_end = "sdimx_end",
  y_end = "sdimy_end",
  z_end = "sdimz_end")
```

### **Arguments**

gobject network in giotto object

#### Value

edges in network as data.table()

### **Examples**

```
plotly_network(gobject)
```

plotMetaDataCellsHeatmap

plot Meta Data Cells Heat map

## **Description**

creates order for clusters

```
plotMetaDataCellsHeatmap(
  gobject,
  metadata_cols = NULL,
  spat_enr_names = NULL,
  value_cols = NULL,
  first_meta_col = NULL,
  second_meta_col = NULL,
  show_values = c("zscores", "original", "zscores_rescaled"),
  custom_cluster_order = NULL,
  clus_cor_method = "pearson",
  clus_cluster_method = "complete",
  custom_values_order = NULL,
  values_cor_method = "pearson",
  values_cluster_method = "complete",
  midpoint = 0,
  x_{text_size} = 8,
  x_{text_angle} = 45,
  y_{text_size} = 8,
```

```
strip_text_size = 8,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "plotMetaDataCellsHeatmap"
)
```

## **Arguments**

```
gobject
                  giotto object
metadata_cols
                  annotation columns found in pDataDT(gobject)
spat_enr_names spatial enrichment results to include
value_cols
                  value columns to use
first_meta_col if more than 1 metadata column, select the x-axis factor
second_meta_col
                  if more than 1 metadata column, select the facetting factor
show_values
                  which values to show on heatmap
custom_cluster_order
                  custom cluster order (default = NULL)
clus_cor_method
                  correlation method for clusters
clus_cluster_method
                  hierarchical cluster method for the clusters
                  midpoint of show_values
midpoint
                  size of x-axis text
x_text_size
                  angle of x-axis text
x_text_angle
y_text_size
                  size of y-axis text
strip_text_size
                  size of strip text
                  show plot
show_plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
custom_gene_order
                  custom gene order (default = NULL)
gene_cor_method
                  correlation method for genes
gene_cluster_method
                  hierarchical cluster method for the genes
```

## **Details**

Creates heatmap for the average values of selected value columns in the different annotation groups

#### Value

ggplot or data.table

#### **Examples**

```
plotMetaDataCellsHeatmap(gobject)
```

plotMetaDataHeatmap

## Description

creates order for clusters

### Usage

```
plotMetaDataHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metadata_cols = NULL,
  selected_genes = NULL,
  first_meta_col = NULL,
  second_meta_col = NULL,
  show_values = c("zscores", "original", "zscores_rescaled"),
  custom_cluster_order = NULL,
  clus_cor_method = "pearson",
  clus_cluster_method = "complete",
  custom_gene_order = NULL,
  gene_cor_method = "pearson",
  gene_cluster_method = "complete",
  midpoint = 0,
  x_{text_size} = 10,
  x_{text_angle} = 45,
  y_{text_size} = 10,
  strip_text_size = 8,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotMetaDataHeatmap"
)
```

```
gobject giotto object
expression_values
expression values to use
metadata_cols annotation columns found in pDataDT(gobject)
selected_genes subset of genes to use
first_meta_col if more than 1 metadata column, select the x-axis factor
```

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 $second\_meta\_col$ 

if more than 1 metadata column, select the facetting factor

show\_values which values to show on heatmap

custom\_cluster\_order

custom cluster order (default = NULL)

clus\_cor\_method

correlation method for clusters

clus\_cluster\_method

hierarchical cluster method for the clusters

custom\_gene\_order

custom gene order (default = NULL)

gene\_cor\_method

correlation method for genes

gene\_cluster\_method

hierarchical cluster method for the genes

midpoint midpoint of show\_values

x\_text\_size size of x-axis text

x\_text\_angle angle of x-axis text

y\_text\_size size of y-axis text

strip\_text\_size

size of strip text

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

### **Details**

Creates heatmap for average the average expression of selected genes in the different annotation groups

#### Value

ggplot or data.table

## **Examples**

plotMetaDataHeatmap(gobject)

146 plotPCA

plotPCA plotPCA

#### **Description**

Short wrapper for PCA visualization

#### Usage

```
plotPCA(gobject, dim_reduction_name = "pca", default_save_name = "PCA", ...)
```

#### **Arguments**

```
gobject
                  giotto object
dim_reduction_name
                  dimension reduction name
default_save_name
                  default save name for saving, don't change, change save_name in save_param
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
                  name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
```

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```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
```

### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotPCA\_3D

#### Value

ggplot

## **Examples**

plotPCA(gobject)

plotPCA\_2D

plotPCA\_2D

## Description

Short wrapper for PCA visualization

```
plotPCA_2D(
  gobject,
  dim_reduction_name = "pca",
  default_save_name = "PCA_2D",
   ...
)
```

plotPCA\_2D

#### **Arguments**

giotto object gobject dim\_reduction\_name dimension reduction name default\_save\_name default save name for saving, don't change, change save\_name in save\_param dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis spat\_enr\_names names of spatial enrichment results to include show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name color for cells (see details) cell\_color color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select subset of cells based on cell IDs select\_cells show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels edge\_alpha column to use for alpha of the edges point\_size size of point (cell) point\_border\_col color of border around points

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```
point_border_stroke
```

stroke size of border around points

show\_legend show\_plot show\_plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

#### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotPCA\_3D

#### Value

ggplot

#### **Examples**

```
plotPCA_2D(gobject)
```

plotPCA\_3D

plotPCA\_3D

### **Description**

Visualize cells according to 3D PCA dimension reduction

## Usage

```
plotPCA_3D(
  gobject,
  dim_reduction_name = "pca",
  default_save_name = "PCA_3D",
   ...
)
```

### **Arguments**

plotPCA\_3D

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

network\_name name of NN network to use, if show\_NN\_network = TRUE

cell\_color color for cells (see details)

color\_as\_factor

convert color column to factor

cell\_color\_code

named vector with colors

select\_cell\_groups

select subset of cells/clusters based on cell\_color parameter

select\_cells select subset of cells based on cell IDs

show\_other\_cells

display not selected cells

other\_cell\_color

color of not selected cells

other\_point\_size

size of not selected cells

show\_cluster\_center

plot center of selected clusters

show\_center\_label

plot label of selected clusters

center\_point\_size

size of center points

label\_size size of labels

edge\_alpha column to use for alpha of the edges

point\_size size of point (cell)

show\_legend show legend

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

### **Details**

Description of parameters.

## Value

plotly

## **Examples**

plotPCA\_3D(gobject)

plotTSNE 151

*plotTSNE* plotTSNE

#### **Description**

Short wrapper for tSNE visualization

#### Usage

```
plotTSNE(gobject, dim_reduction_name = "tsne", default_save_name = "tSNE", ...)
```

#### **Arguments**

```
gobject
                  giotto object
dim_reduction_name
                  dimension reduction name
default_save_name
                  default save name for saving, don't change, change save_name in save_param
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
                  name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
```

plot center of selected clusters

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```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
```

### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotTSNE\_3D

#### Value

ggplot

## **Examples**

```
plotTSNE(gobject)
```

plotTSNE\_2D

plotTSNE\_2D

## Description

Short wrapper for tSNE visualization

```
plotTSNE_2D(
  gobject,
  dim_reduction_name = "tsne",
  default_save_name = "tSNE_2D",
   ...
)
```

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#### **Arguments**

giotto object gobject dim\_reduction\_name dimension reduction name default\_save\_name default save name for saving, don't change, change save\_name in save\_param dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis spat\_enr\_names names of spatial enrichment results to include show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select subset of cells based on cell IDs select\_cells show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels edge\_alpha column to use for alpha of the edges point\_size size of point (cell) point\_border\_col color of border around points

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```
point_border_stroke
```

stroke size of border around points

show\_legend show\_plot show\_plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotTSNE\_3D

### Value

ggplot

### **Examples**

```
plotTSNE_2D(gobject)
```

plotTSNE\_3D

plotTSNE\_3D

## Description

Visualize cells according to dimension reduction coordinates

#### Usage

```
plotTSNE_3D(
  gobject,
  dim_reduction_name = "tsne",
  default_save_name = "TSNE_3D",
  ...
)
```

## **Arguments**

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```
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
                  name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
edge_alpha
                  column to use for alpha of the edges
                  size of point (cell)
point_size
show_legend
                  show legend
                  show plot
show_plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
```

list of saving parameters from all\_plots\_save\_function()

### **Details**

Description of parameters.

## Value

plotly

save\_param

# Examples

```
plotTSNE_3D(gobject)
```

156 plotUMAP

plotUMAP plotUMAP

#### **Description**

Short wrapper for UMAP visualization

#### Usage

```
plotUMAP(gobject, dim_reduction_name = "umap", default_save_name = "UMAP", ...)
```

#### **Arguments**

```
gobject
                  giotto object
dim_reduction_name
                  dimension reduction name
default_save_name
                  default save name for saving, don't change, change save_name in save_param
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
                  name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
```

plot center of selected clusters

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```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
```

### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotUMAP\_3D

#### Value

ggplot

## **Examples**

```
plotUMAP(gobject)
```

plotUMAP\_2D

plotUMAP\_2D

## Description

Short wrapper for UMAP visualization

```
plotUMAP_2D(
  gobject,
  dim_reduction_name = "umap",
  default_save_name = "UMAP_2D",
   ...
)
```

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#### **Arguments**

giotto object gobject dim\_reduction\_name dimension reduction name default\_save\_name default save name for saving, don't change, change save\_name in save\_param dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis spat\_enr\_names names of spatial enrichment results to include show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name color for cells (see details) cell\_color color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select subset of cells based on cell IDs select\_cells show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels edge\_alpha column to use for alpha of the edges point\_size size of point (cell) point\_border\_col color of border around points

plotUMAP\_3D

```
point_border_stroke
```

stroke size of border around points

show\_legend show legend
show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

#### **Details**

Description of parameters, see dimPlot2D. For 3D plots see plotUMAP\_3D

#### Value

ggplot

#### **Examples**

```
plotUMAP_2D(gobject)
```

plotUMAP\_3D

plotUMAP\_3D

### **Description**

Visualize cells according to dimension reduction coordinates

## Usage

```
plotUMAP_3D(
  gobject,
  dim_reduction_name = "umap",
  default_save_name = "UMAP_3D",
   ...
)
```

### **Arguments**

plotUMAP\_3D

nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size size of not selected cells show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels edge\_alpha column to use for alpha of the edges size of point (cell) point\_size show\_legend show legend show plot show\_plot return\_plot return ggplot object

directly save the plot [boolean]

list of saving parameters from all\_plots\_save\_function()

### **Details**

save\_plot

save\_param

Description of parameters.

## Value

plotly

## **Examples**

plotUMAP\_3D(gobject)

## Description

Visualize cells in network layer according to dimension reduction coordinates

# Usage

```
plot_network_layer_ggplot(
   ggobject,
   annotated_network_DT,
   edge_alpha = NULL,
   show_legend = T
)
```

## Arguments

```
annotated\_network\_DT
```

annotated network data.table of selected cells

edge\_alpha alpha of network edges

show\_legend show legend gobject giotto object

### **Details**

Description of parameters.

### Value

ggplot

# **Examples**

```
plot_network_layer_ggplot(gobject)
```

## Description

Visualize cells in point layer according to dimension reduction coordinates

### Usage

plot\_point\_layer\_ggplot(

```
ggobject,
      annotated_DT_selected,
      annotated_DT_other,
      cell_color = NULL,
      color_as_factor = T,
      cell_color_code = NULL,
      cell_color_gradient = c("blue", "white", "red"),
      gradient_midpoint = 0,
      gradient_limits = NULL,
      select_cell_groups = NULL,
      select_cells = NULL,
      point_size = 1,
      point_border_col = "black",
      point_border_stroke = 0.1,
      show_cluster_center = F,
      show_center_label = T,
      center_point_size = 4,
      center_point_border_col = "black",
      center_point_border_stroke = 0.1,
      label_size = 4,
      label_fontface = "bold",
      edge_alpha = NULL,
      show_other_cells = T,
      other_cell_color = "lightgrey",
      other_point_size = 0.5,
      show_legend = T
    )
Arguments
    annotated_DT_selected
                     annotated data.table of selected cells
    annotated_DT_other
                     annotated data.table of not selected cells
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
                     size of point (cell)
    point_size
```

```
point_border_col
```

color of border around points

point\_border\_stroke

stroke size of border around points

show\_cluster\_center

plot center of selected clusters

show\_center\_label

plot label of selected clusters

 ${\tt center\_point\_size}$ 

size of center points

label\_size size of labels

label\_fontface font of labels

edge\_alpha column to use for alpha of the edges

show\_other\_cells

display not selected cells

other\_cell\_color

color of not selected cells

other\_point\_size

size of not selected cells

show\_legend show legend gobject giotto object

### **Details**

Description of parameters.

#### Value

ggplot

## **Examples**

plot\_point\_layer\_ggplot(gobject)

## Description

creat ggplot point layer for spatial coordinates

#### Usage

```
plot_spat_point_layer_ggplot(
  ggobject,
  sdimx = NULL,
  sdimy = NULL,
  cell_locations_metadata_selected,
  cell_locations_metadata_other,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 2,
  point_border_col = "lightgrey",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  show_legend = TRUE
)
```

### Arguments

```
sdimx
                  x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimv
cell_locations_metadata_selected
                  annotated location from selected cells
cell_locations_metadata_other
                  annotated location from non-selected cells
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
{\tt cell\_color\_gradient}
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
```

print.giotto 165

```
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
                  size of point (cell)
point_size
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
show_other_cells
                  display not selected cells
other_cell_color
                  color for not selected cells
other_point_size
                  point size for not selected cells
show_legend
                  show legend
gobject
                  giotto object
```

### **Details**

Description of parameters.

## Value

ggplot

# **Examples**

```
plot_spat_point_layer_ggplot(gobject)
```

print.giotto

print method for giotto class

## Description

print method for giotto class. Prints the chosen number of genes (rows) and cells (columns) from the raw count matrix. Also print the spatial locations for the chosen number of cells.

```
print.giotto(object, ...)
```

166 rankEnrich

#### **Arguments**

nr\_genes number of genes (rows) to print nr\_cells number of cells (columns) to print

rankEnrich

rankEnrich

### **Description**

Function to calculate gene signature enrichment scores per spatial position using a rank based approach.

### Usage

```
rankEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

#### **Arguments**

```
gobject Giotto object

sign_matrix Matrix of signature genes for each cell type / process

expression_values

expression values to use

reverse_log_scale

reverse expression values from log scale

logbase log base to use if reverse_log_scale = TRUE

output_enrichment

how to return enrichment output
```

#### **Details**

First a new rank is calculated as  $R = (R1*R2)^{n}(1/2)$ , where R1 is the rank of fold-change for each gene in each spot and R2 is the rank of each marker in each cell type. The Rank-Biased Precision is then calculated as:  $RBP = (1 - 0.99) * (0.99)^{n}(R - 1)$  and the final enrichment score is then calculated as the sum of top 100 RBPs.

#### Value

data.table with enrichment results

### **Examples**

```
rankEnrich(gobject)
```

rank\_binarize 167

rank\_binarize

rank\_binarize

# Description

create binarized scores using arbitrary rank of top genes

# Usage

```
rank_binarize(x, max_rank = 200)
```

readGiottoInstructions

read Giot to Instrunctions

# Description

Retrieves the instruction associated with the provided parameter

# Usage

```
readGiottoInstructions(giotto_instructions, param = NULL)
```

# Arguments

giotto\_instructions

 $giot to\ object\ or\ result\ from\ create Giot to Instructions()$ 

param

parameter to retrieve

## Value

specific parameter

# **Examples**

readGiottoInstrunctions()

168 removeGeneAnnotation

remove Cell Annotation remove Cell Annotation

## Description

removes cell annotation of giotto object

### Usage

```
removeCellAnnotation(gobject, columns = NULL, return_gobject = TRUE)
```

#### **Arguments**

gobject giotto object

columns names of columns to remove

return\_gobject boolean: return giotto object (default = TRUE)

#### **Details**

if return\_gobject = FALSE, it will return the cell metadata

### Value

giotto object

## **Examples**

removeCellAnnotation(gobject)

removeGeneAnnotation removeGeneAnnotation

### **Description**

removes gene annotation of giotto object

# Usage

```
removeGeneAnnotation(gobject, columns = NULL, return_gobject = TRUE)
```

### **Arguments**

gobject giotto object

columns names of columns to remove

return\_gobject boolean: return giotto object (default = TRUE)

### **Details**

if return\_gobject = FALSE, it will return the gene metadata

#### Value

giotto object

### **Examples**

removeGeneAnnotation(gobject)

```
replaceGiottoInstructions
```

replace Giot to Instructions

## **Description**

Function to replace all instructions from giotto object

## Usage

```
replaceGiottoInstructions(gobject, instructions = NULL)
```

### **Arguments**

```
gobject giotto object
instructions new instructions (e.g. result from createGiottoInstructions)
```

### Value

named vector with giotto instructions

### **Examples**

```
replaceGiottoInstructions()
```

runPCA

runPCA

## Description

run PCA

```
runPCA(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  name = "pca",
  genes_to_use = NULL,
  return_gobject = TRUE,
  scale_unit = F,
  ncp = 200,
  ...
)
```

170 runtSNE

### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
                  cells or genes
reduction
name
                  arbitrary name for PCA run
                  subset of genes to use for PCA
genes_to_use
return_gobject boolean: return giotto object (default = TRUE)
scale_unit
                  scale features before PCA
                  number of principal components to calculate
ncp
                  additional parameters for PCA
```

#### **Details**

. . .

Description of PCA steps...

### Value

giotto object with updated PCA dimension recuction

### **Examples**

```
runPCA(gobject)
```

runtSNE

runtSNE

## **Description**

run tSNE

```
runtSNE(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "tsne",
  genes_to_use = NULL,
  return_gobject = TRUE,
  dims = 2,
  perplexity = 30,
  theta = 0.5,
  do_PCA_first = F,
  set\_seed = T,
  seed_number = 1234,
)
```

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### **Arguments**

gobject giotto object

expression\_values

expression values to use

reduction cells or genes

dim\_reduction\_to\_use

use another dimension reduction set as input

dim\_reduction\_name

name of dimension reduction set to use

dimensions\_to\_use

number of dimensions to use as input

name arbitrary name for tSNE run

genes\_to\_use if dim\_reduction\_to\_use = NULL, which genes to use

 $\verb|return_gobject|| boolean: return giotto object (default = TRUE)$ 

dims tSNE param: number of dimensions to return

perplexity tSNE param: perplexity

theta tSNE param: theta

do\_PCA\_first tSNE param: do PCA before tSNE (default = FALSE)

set\_seed use of seed

seed\_number seed number to use

... additional tSNE parameters

#### **Details**

Description of tSNE steps and params ...

### Value

giotto object with updated tSNE dimension recuction

## **Examples**

runtSNE(gobject)

runUMAP runUMAP

## **Description**

run UMAP

172 runUMAP

## Usage

```
runUMAP(
      gobject,
      expression_values = c("normalized", "scaled", "custom"),
      reduction = c("cells", "genes"),
      dim_reduction_to_use = "pca",
      dim_reduction_name = "pca",
      dimensions_to_use = 1:10,
      name = "umap",
      genes_to_use = NULL,
      return_gobject = TRUE,
      n_neighbors = 40,
      n_{components} = 2,
      n_{epochs} = 400,
      min_dist = 0.01,
      n_{threads} = 1,
      spread = 5,
      set\_seed = T,
      seed_number = 1234,
    )
Arguments
   gobject
                    giotto object
    expression_values
                    expression values to use
    reduction
                    cells or genes
    dim_reduction_to_use
                    use another dimension reduction set as input
   dim_reduction_name
                    name of dimension reduction set to use
   dimensions_to_use
                    number of dimensions to use as input
                    arbitrary name for UMAP run
   name
                    if dim_reduction_to_use = NULL, which genes to use
   genes_to_use
    return_gobject boolean: return giotto object (default = TRUE)
   n_neighbors
                    UMAP param: number of neighbors
                    UMAP param: number of components
   n_components
   n_epochs
                    UMAP param: number of epochs
                    UMAP param: minimum distance
   min_dist
   n_threads
                    UMAP param: threads to use
                    UMAP param: spread
    spread
    set\_seed
                    use of seed
    seed_number
                    seed number to use
```

additional UMAP parameters

selectPatternGenes 173

#### **Details**

Description of UMAP steps...

#### Value

giotto object with updated UMAP dimension recuction

### **Examples**

```
runUMAP(gobject)
```

selectPatternGenes

selectPatternGenes

## Description

Select genes correlated with spatial patterns

## Usage

```
selectPatternGenes(
  spatPatObj,
  dimensions = 1:5,
  top_pos_genes = 10,
  top_neg_genes = 10,
  min_pos_cor = 0.5,
  min_neg_cor = -0.5,
  return_top_selection = FALSE
)
```

## Arguments

```
spatPatObj Output from detectSpatialPatterns
dimensions dimensions to identify correlated genes for.
top_pos_genes Top positively correlated genes.
top_neg_genes Top negatively correlated genes.
min_pos_cor Minimum positive correlation score to include a gene.
min_neg_cor Minimum negative correlation score to include a gene.
```

## **Details**

Description.

#### Value

Data.table with genes associated with selected dimension (PC).

## **Examples**

```
selectPatternGenes(gobject)
```

# Description

helper function to select expression values

### Usage

```
select_expression_values(gobject, values)
```

# Arguments

gobject giotto object

values expression values to extract

## Value

expression matrix

show, giotto-method show method for giotto class

## Description

show method for giotto class

## Usage

```
## S4 method for signature 'giotto'
show(object)
```

 $show {\tt ClusterDendrogram} \ \ \textit{show ClusterDendrogram}$ 

## **Description**

Creates dendrogram based on identified clusters

#### Usage

```
showClusterDendrogram(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  h = NULL,
  h_color = "red",
  rotate = FALSE,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showClusterDendrogram",
  ...
)
```

#### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
distance
                  distance method to use for hierarchical clustering
                  height of horizontal lines to plot
h
h_color
                  color of horizontal lines
rotate
                  rotate dendrogram 90 degrees
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  additional parameters for ggdendrogram()
```

#### **Details**

Correlation dendrogram of selected clustering.

#### Value

ggplot

### **Examples**

```
showClusterDendrogram(gobject)
```

176 showClusterHeatmap

showClusterHeatmap showClusterHeatmap

#### **Description**

Creates heatmap based on identified clusters

### Usage

```
showClusterHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = "all",
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showClusterHeatmap",
  ...
)
```

### **Arguments**

```
gobject
                  giotto object
expression_values
                  expression values to use
                  vector of genes to use, default to 'all'
genes
cluster_column name of column to use for clusters
cor
                  correlation score to calculate distance
                  distance method to use for hierarchical clustering
distance
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  additional parameters for the Heatmap function from ComplexHeatmap
```

#### **Details**

Correlation heatmap of selected clusters.

#### Value

ggplot

showCPGscores 177

#### **Examples**

showClusterHeatmap(gobject)

showCPGscores

showCPGscores

#### **Description**

visualize Cell Proximity Gene enrichment scores

### Usage

```
showCPGscores(
 gobject,
 CPGscore,
 method = c("volcano", "cell_barplot", "cell-cell", "cell_sankey"),
 min_cells = 5,
 min_fdr = 0.05,
 min_spat_diff = 0.2,
 min_{log2_fc} = 0.5,
 keep_int_duplicates = TRUE,
 direction = c("both", "up", "down"),
  cell_color_code = NULL,
  show_plot = NA,
 return_plot = NA,
  save_plot = NA,
  save_param = list(),
 default_save_name = "showCPGscores"
)
```

# Arguments

```
CPGscore, output from getCellProximityGeneScores()
CPGscore
                  visualization method
method
                  min number of cells threshold
min_cells
min_fdr
                  fdr threshold
                  spatial difference threshold
min_spat_diff
                  min log2 fold-change
min_log2_fc
keep_int_duplicates
                  keep both cell_A-cell_B and cell_B-cell_A
                  up or downregulation or both
direction
cell_color_code
                  color code for cell types
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

#### **Details**

Give more details ...

#### Value

Gene to gene scores in data.table format

## **Examples**

```
showCPGscores(CPGscore)
```

```
show Gene Expression Proximity Score \\ show Gene Expression Proximity Score
```

# Description

Create heatmap from cell-cell proximity scores

## Usage

```
showGeneExpressionProximityScore(
  scores,
  selected_gene,
  sort_column = "diff_spat"
)
```

# Arguments

```
scores CPscore, output from getAverageCellProximityGeneScores()
selected_gene gene to show
sort_column column name to use for sorting
```

### **Details**

Give more details ...

## Value

ggplot barplot

### **Examples**

showGeneExpressionProximityScore(scores)

showGiottoInstructions 179

```
showGiottoInstructions
```

showGiottoInstructions

## **Description**

Function to display all instructions from giotto object

### Usage

```
showGiottoInstructions(gobject)
```

# Arguments

```
gobject giotto object
```

#### Value

named vector with giotto instructions

### **Examples**

```
showGiottoInstructions()
```

 $\verb|showGTGscores||$ 

showGTGscores

### **Description**

visualize Cell Proximity Gene enrichment scores

```
showGTGscores(
  GTGscore,
  method = c("cell_barplot", "cell-cell", "cell_sankey"),
  min_cells = 5,
  min_pval = 0.05,
  min_spat_diff = 0.2,
  min_log2_fc = 0.5,
  direction = c("both", "up", "down"),
  cell_color_code = NULL,
  show_plot = T,
  specific_genes_1 = NULL,
  specific_genes_2 = NULL,
  first_cell_name = "ligand cell",
  second_cell_name = "receptor cell",
  return_DT = F
```

#### **Arguments**

```
visualization method
method
min_cells
                  min number of cells threshold
                  p-value threshold
min_pval
min_spat_diff
                  spatial difference threshold
                  log2 fold-change threshold
min_log2_fc
direction
                  up or downregulation or both
cell_color_code
                  color code for cell types
show_plot
                  print plot
specific_genes_1
                  subset of genes, matched with specific_genes_2
specific_genes_2
                  subset of genes, matched with specific_genes_1
first_cell_name
                  name for first cells
second_cell_name
                  name for second cells
                  CPGscore, output from getCellProximityGeneScores()
CPGscore
```

#### **Details**

Give more details ...

### Value

ggplot

### **Examples**

```
showGTGscores(CPGscore)
```

```
show Int {\it Expression Proximity Score} \\ show Int {\it Expression Proximity Score}
```

### **Description**

Create heatmap from cell-cell proximity scores

```
showIntExpressionProximityScore(
   scores,
   selected_interaction,
   sort_column = "diff_spat",
   show_enriched_n = 5,
   show_depleted_n = 5
)
```

showPattern 181

## **Arguments**

show top depleted interactions

### **Details**

Give more details ...

## Value

ggplot barplot

### **Examples**

showIntExpressionProximityScore(scores)

showPattern showPattern

## Description

```
create a spatial grid
show patterns for 2D spatial data
```

## Usage

```
showPattern(gobject, spatPatObj, ...)
showPattern(gobject, spatPatObj, ...)
```

## **Arguments**

gobject giotto object

spatPatObj Output from detectSpatialPatterns

dimension dimension to plot

trim Trim ends of the PC values.

background\_color

background color for plot

grid\_border\_color

color for grid

show\_legend show legend of ggplot

show\_plot show plot

showPattern2D

## **Details**

Description.

## Value

ggplot ggplot

## See Also

showPattern2D

## **Examples**

```
showPattern(gobject)
showPattern(gobject)
```

showPattern2D

showPattern2D

## Description

show patterns for 2D spatial data

```
showPattern2D(
  gobject,
  spatPatObj,
  dimension = 1,
  trim = c(0.02, 0.98),
  background_color = "white",
  grid_border_color = "grey",
  show_legend = T,
  point_size = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showPattern2D"
)
```

showPattern3D 183

## **Arguments**

gobject giotto object spatPatObj Output from detectSpatialPatterns dimension dimension to plot trim Trim ends of the PC values. background\_color background color for plot grid\_border\_color color for grid show\_legend show legend of ggplot  ${\sf show\_plot}$ show plot return\_plot return ggplot object directly save the plot [boolean] save\_plot save\_param list of saving parameters from all\_plots\_save\_function()

default save name for saving, don't change, change save\_name in save\_param

### Value

ggplot

## **Examples**

showPattern2D(gobject)

showPattern3D

default\_save\_name

showPattern3D

## **Description**

show patterns for 3D spatial data

```
showPattern3D(
  gobject,
  spatPatObj,
  dimension = 1,
  trim = c(0.02, 0.98),
  background_color = "white",
  grid_border_color = "grey",
  show_legend = T,
  point_size = 1,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
```

184 showPattern3D

```
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "showPattern3D"
)
```

giotto object

## **Arguments**

gobject

spatPatObj Output from detectSpatialPatterns dimension to plot dimension Trim ends of the PC values. trim background\_color background color for plot grid\_border\_color color for grid show\_legend show legend of plot point\_size adjust the point size scale the axis axis\_scale cutomize the scale of the axis custom\_ratio the tick number of x\_axis x\_ticks y\_ticks the tick number of y\_axis

show\_plot show plot

return\_plot return plot object

save\_plot directly save the plot [boolean]

 $save\_param \qquad list \ of \ saving \ parameters \ from \ all\_plots\_save\_function()$ 

the tick number of z\_axis

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

### Value

plotly

z\_ticks

# Examples

```
showPattern3D(gobject)
```

showPatternGenes 185

showPatternGenes

showPatternGenes

## **Description**

show genes correlated with spatial patterns

### Usage

```
showPatternGenes(
  gobject,
  spatPatObj,
  dimension = 1,
  top_pos_genes = 5,
  top_neg_genes = 5,
  point_size = 1,
  return_DT = FALSE,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showPatternGenes"
)
```

## **Arguments**

```
gobject
                  giotto object
spatPatObj
                  Output from detectSpatialPatterns
dimension
                  dimension to plot genes for.
top_pos_genes
                  Top positively correlated genes.
top_neg_genes
                  Top negatively correlated genes.
                  size of points
point_size
return_DT
                  if TRUE, it will return the data.table used to generate the plots
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

## Value

ggplot

## **Examples**

```
showPatternGenes(gobject)
```

showTopGeneToGene

```
showProcessingSteps showProcessingSteps
```

## Description

shows the sequential processing steps that were performed in a summarized format

## Usage

```
showProcessingSteps(gobject)
```

## **Arguments**

```
gobject giotto object
```

#### Value

list of processing steps and names

## **Examples**

```
showProcessingSteps(gobject)
```

showTopGeneToGene

*showTopGeneToGene* 

## Description

Show enriched/depleted gene-gene enrichments

## Usage

```
showTopGeneToGene(
  GTGscore,
  top_interactions = 10,
  direction = c("increased", "decreased"),
  complement_data = T,
  subset_cell_ints = NULL,
  subset_genes = NULL
)
```

## **Arguments**

signPCA 187

### **Details**

Give more details ...

### Value

ggplot barplot

### **Examples**

showTopGeneToGene(scores)

signPCA

signPCA

## Description

identify significant prinicipal components (PCs)

## Usage

```
signPCA(
  gobject,
  method = c("screeplot", "jackstraw"),
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  genes_to_use = NULL,
  scale_unit = T,
  ncp = 50,
  scree_labels = T,
  scree_ylim = c(0, 10),
  jack_iter = 10,
  jack_threshold = 0.01,
  jack_verbose = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "signPCA",
)
```

## **Arguments**

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ncp number of principal components to calculate
scree\_labels show labels on scree plot
scree\_ylim y-axis limits on scree plot

jack\_iter number of interations for jackstraw

jack\_threshold p-value threshold to call a PC significant

jack\_verbose show progress of jackstraw method

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

... additional parameters for PCA

#### **Details**

Two different methods can be used to assess the number of relevant or significant prinicipal components (PC's). Screeplot works by plotting the explained variance of each individual PC in a barplot and then you can identify which PC does not contribute anymore (= 'elbow method'). The Jackstraw method used the permutationPA function from the jackstraw packages. By systematically permuting genes it identifies robust, and thus significant, PCs.

### Value

ggplot object for scree method and maxtrix of p-values for jackstraw

### **Examples**

```
signPCA(gobject)
```

spatCellPlot spatCellPlot

## **Description**

Visualize cells according to spatial coordinates

```
spatCellPlot(gobject, ...)
```

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### **Arguments**

gobject giotto object sdimx x-axis dimension name (default = 'sdimx') sdimy y-axis dimension name (default = 'sdimy') spat\_enr\_names names of spatial enrichment results to include cell\_annotation\_values numeric cell annotation columns cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs size of point (cell) point\_size point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels show\_network show underlying spatial network spatial\_network\_name name of spatial network to use network\_color color of spatial network network\_alpha alpha of spatial network show\_grid show spatial grid spatial\_grid\_name name of spatial grid to use grid\_color color of spatial grid show\_other\_cells display not selected cells other\_cell\_color color of not selected cells other\_point\_size point size of not selected cells other\_cells\_alpha alpha of not selected cells

```
coord_fix_ratio

fix ratio between x and y-axis

show_legend show legend

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function()

default_save_name

default save name for saving, don't change, change save_name in save_param
```

### **Details**

Description of parameters.

#### Value

ggplot

## **Examples**

```
spatCellPlot(gobject)
```

spatCellPlot2D spatCellPlot2D

## Description

Visualize cells according to spatial coordinates

```
spatCellPlot2D(
 gobject,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_annotation_values,
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
 select_cell_groups = NULL,
 select_cells = NULL,
 point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
 center_point_size = 4,
 center_point_border_col = "black",
  center_point_border_stroke = 0.1,
```

```
label_size = 4,
  label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
  network_color = NULL,
 network_alpha = 1,
  show\_grid = F,
  spatial_grid_name = "spatial_grid",
  grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
  other_cells_alpha = 0.1,
 coord_fix_ratio = NULL,
  show_legend = T,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
 cow_align = "h",
  show_plot = NA,
 return_plot = NA,
 save_plot = NA,
  save_param = list(),
  default_save_name = "spatCellPlot2D"
)
```

## Arguments

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimy
spat_enr_names names of spatial enrichment results to include
cell_annotation_values
                  numeric cell annotation columns
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
                  size of point (cell)
point_size
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_cluster_center
                  plot center of selected clusters
```

```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
show_network
                  show underlying spatial network
spatial_network_name
                  name of spatial network to use
                  color of spatial network
network_color
                  alpha of spatial network
network_alpha
                  show spatial grid
show_grid
spatial_grid_name
                  name of spatial grid to use
grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  point size of not selected cells
other_cells_alpha
                  alpha of not selected cells
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
```

list of saving parameters from all\_plots\_save\_function()

default save name for saving, don't change, change save\_name in save\_param

### **Details**

Description of parameters.

### Value

ggplot

save\_param

default\_save\_name

## **Examples**

spatCellPlot2D(gobject)

spatDimCellPlot
 spatDimCellPlot

### **Description**

Visualize numerical features of cells according to spatial AND dimension reduction coordinates in 2D

## Usage

```
spatDimCellPlot(gobject, ...)
```

## **Arguments**

```
gobject
                  giotto object
plot_alignment direction to align plot
spat_enr_names names of spatial enrichment results to include
cell_annotation_values
                  numeric cell annotation columns
dim_reduction_to_use
                  dimension reduction to use
dim_reduction_name
                  dimension reduction name
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
sdimx
                  = spatial dimension to use on x-axis
sdimy
                  = spatial dimension to use on y-axis
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
dim_point_size size of points in dim. reduction space
dim_point_border_col
                  border color of points in dim. reduction space
dim_point_border_stroke
                  border stroke of points in dim. reduction space
spat_point_size
                  size of spatial points
spat_point_border_col
                  border color of spatial points
spat_point_border_stroke
                  border stroke of spatial points
```

dim\_show\_cluster\_center show the center of each cluster dim\_show\_center\_label provide a label for each cluster dim\_center\_point\_size size of the center point dim\_center\_point\_border\_col border color of center point dim\_center\_point\_border\_stroke stroke size of center point dim\_label\_size size of the center label dim\_label\_fontface font of the center label spat\_show\_cluster\_center show the center of each cluster spat\_show\_center\_label provide a label for each cluster spat\_center\_point\_size size of the center point spat\_label\_size size of the center label spat\_label\_fontface font of the center label show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) nn\_network\_name name of NN network to use, if show\_NN\_network = TRUE dim\_edge\_alpha column to use for alpha of the edges spat\_show\_network show spatial network spatial\_network\_name name of spatial network to use spat\_network\_color color of spatial network spat\_show\_grid show spatial grid spatial\_grid\_name name of spatial grid to use spat\_grid\_color color of spatial grid show\_other\_cells display not selected cells other\_cell\_color color of not selected cells dim\_other\_point\_size size of not selected dim cells spat\_other\_point\_size

size of not selected spat cells

```
spat_other_cells_alpha
                  alpha of not selected spat cells
coord_fix_ratio
                  ratio for coordinates
cow_n_col
                  cowplot param: how many columns
                  cowplot param: relative height
cow_rel_h
                  cowplot param: relative width
cow_rel_w
                  cowplot param: how to align
cow_align
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

### **Details**

Description of parameters.

### Value

ggplot

## **Examples**

```
spatDimCellPlot(gobject)
```

spatDimCellPlot2D

## **Description**

Visualize numerical features of cells according to spatial AND dimension reduction coordinates in 2D

```
spatDimCellPlot2D(
  gobject,
  plot_alignment = c("vertical", "horizontal"),
  spat_enr_names = NULL,
  cell_annotation_values,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
```

```
cell_color_gradient = c("blue", "white", "red"),
gradient_midpoint = NULL,
gradient_limits = NULL,
select_cell_groups = NULL,
select_cells = NULL,
dim_point_size = 1,
dim_point_border_col = "black",
dim_point_border_stroke = 0.1,
spat_point_size = 1,
spat_point_border_col = "black",
spat_point_border_stroke = 0.1,
dim_show_cluster_center = F,
dim_show_center_label = T,
dim_center_point_size = 4,
dim_center_point_border_col = "black",
dim_center_point_border_stroke = 0.1,
dim_label_size = 4,
dim_label_fontface = "bold",
spat_show_cluster_center = F,
spat_show_center_label = F,
spat_center_point_size = 4,
spat_center_point_border_col = "black",
spat_center_point_border_stroke = 0.1,
spat_label_size = 4,
spat_label_fontface = "bold",
show_NN_network = F,
nn_network_to_use = "sNN",
nn_network_name = "sNN.pca",
dim_edge_alpha = 0.5,
spat\_show\_network = F,
spatial_network_name = "spatial_network",
spat_network_color = "red",
spat_network_alpha = 0.5,
spat_show_grid = F,
spatial_grid_name = "spatial_grid",
spat_grid_color = "green",
show_other_cells = TRUE,
other_cell_color = "grey";
dim_other_point_size = 0.5,
spat_other_point_size = 0.5,
spat_other_cells_alpha = 0.5,
coord_fix_ratio = NULL,
cow_n_col = 2,
cow_rel_h = 1,
cow_rel_w = 1,
cow_align = "h"
show_legend = T,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spatDimCellPlot2D"
```

)

## Arguments

gobject giotto object plot\_alignment direction to align plot spat\_enr\_names names of spatial enrichment results to include cell\_annotation\_values numeric cell annotation columns  $\operatorname{dim\_reduction\_to\_use}$ dimension reduction to use dim\_reduction\_name dimension reduction name dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis sdimx = spatial dimension to use on x-axis sdimy = spatial dimension to use on y-axis cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs dim\_point\_size size of points in dim. reduction space dim\_point\_border\_col border color of points in dim. reduction space dim\_point\_border\_stroke border stroke of points in dim. reduction space spat\_point\_size size of spatial points spat\_point\_border\_col border color of spatial points spat\_point\_border\_stroke border stroke of spatial points dim\_show\_cluster\_center show the center of each cluster dim\_show\_center\_label provide a label for each cluster dim\_center\_point\_size size of the center point dim\_center\_point\_border\_col border color of center point dim\_center\_point\_border\_stroke stroke size of center point

dim\_label\_size size of the center label dim\_label\_fontface font of the center label spat\_show\_cluster\_center show the center of each cluster spat\_show\_center\_label provide a label for each cluster spat\_center\_point\_size size of the center point spat\_label\_size size of the center label spat\_label\_fontface font of the center label show\_NN\_network show underlying NN network  $nn\_network\_to\_use$ type of NN network to use (kNN vs sNN) nn\_network\_name name of NN network to use, if show\_NN\_network = TRUE dim\_edge\_alpha column to use for alpha of the edges spat\_show\_network show spatial network spatial\_network\_name name of spatial network to use spat\_network\_color color of spatial network spat\_show\_grid show spatial grid spatial\_grid\_name name of spatial grid to use spat\_grid\_color color of spatial grid show\_other\_cells display not selected cells other\_cell\_color color of not selected cells dim\_other\_point\_size size of not selected dim cells spat\_other\_point\_size size of not selected spat cells spat\_other\_cells\_alpha alpha of not selected spat cells coord\_fix\_ratio ratio for coordinates cowplot param: how many columns cow\_n\_col cow\_rel\_h cowplot param: relative height cowplot param: relative width cow\_rel\_w cow\_align cowplot param: how to align

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```
show_legend show legend show_plot show plot
```

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_param list of saving parameters from all\_plots\_save\_function()

default\_save\_name

default save name for saving, don't change, change save\_name in save\_param

## **Details**

Description of parameters.

#### Value

ggplot

## **Examples**

```
spatDimCellPlot2D(gobject)
```

spatDimGenePlot spatDimGenePlot

## Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

## Usage

```
spatDimGenePlot(gobject, ...)
```

## **Arguments**

```
gobject giotto object
```

 ${\tt expression\_values}$ 

gene expression values to use

plot\_alignment direction to align plot

genes genes to show

 ${\tt dim\_reduction\_to\_use}$ 

dimension reduction to use

dim\_reduction\_name

dimension reduction name

dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis dim\_point\_size dim reduction plot: point size

dim\_point\_border\_col

color of border around points

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```
dim_point_border_stroke
                 stroke size of border around points
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
edge_alpha_dim dim reduction plot: column to use for alpha of the edges
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
spatial_network_name
                 name of spatial network to use
spatial_grid_name
                 name of spatial grid to use
spatial_point_size
                 spatial plot: point size
spatial_point_border_col
                 color of border around points
spatial_point_border_stroke
                 stroke size of border around points
midpoint
                 size of point (cell)
point_size
                 size of point (cell)
                 cowplot param: how many columns
cow_n_col
cow_rel_h
                 cowplot param: relative height
                 cowplot param: relative width
cow_rel_w
                 cowplot param: how to align
cow_align
show_legend
                 show legend
show_plot
                 show plots
return_plot
                 return ggplot object
                 directly save the plot [boolean]
save_plot
save_param
                 list of saving parameters from all_plots_save_function()
default_save_name
                 default save name for saving, don't change, change save_name in save_param
```

## **Details**

Description of parameters.

#### Value

ggplot

### See Also

spatDimGenePlot3D

## **Examples**

spatDimGenePlot(gobject)

spatDimGenePlot2D 201

spatDimGenePlot2D

spatDimGenePlot2D

## **Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

## Usage

```
spatDimGenePlot2D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("vertical", "horizontal"),
 genes,
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  show_NN_network = F,
  show_spatial_network = F,
  show_spatial_grid = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
  edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
  spatial_network_name = "spatial_network",
  spatial_grid_name = "spatial_grid",
  spatial_point_size = 1,
  spatial_point_border_col = "black",
  spatial_point_border_stroke = 0.1,
 midpoint = 0,
 genes_high_color = "red",
  genes_mid_color = "white";
 genes_low_color = "blue",
 cow_n_col = 2,
 cow_rel_h = 1,
  cow_rel_w = 1,
 cow_align = "h",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
 save_plot = NA,
 save_param = list(),
  default_save_name = "spatDimGenePlot2D"
```

### **Arguments**

gobject giotto object

expression\_values gene expression values to use plot\_alignment direction to align plot genes genes to show dim\_reduction\_to\_use dimension reduction to use dim\_reduction\_name dimension reduction name dim1\_to\_use dimension to use on x-axis dimension to use on y-axis dim2\_to\_use point\_size size of point (cell) dim\_point\_border\_col color of border around points dim\_point\_border\_stroke stroke size of border around points show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name edge\_alpha\_dim dim reduction plot: column to use for alpha of the edges scale\_alpha\_with\_expression scale expression with ggplot alpha parameter spatial\_network\_name name of spatial network to use spatial\_grid\_name name of spatial grid to use spatial\_point\_size spatial plot: point size spatial\_point\_border\_col color of border around points spatial\_point\_border\_stroke stroke size of border around points midpoint size of point (cell) cowplot param: how many columns cow\_n\_col cow\_rel\_h cowplot param: relative height cowplot param: relative width cow\_rel\_w cowplot param: how to align cow\_align show\_legend show legend show plots show\_plot return ggplot object return\_plot save\_plot directly save the plot [boolean] save\_param list of saving parameters from all\_plots\_save\_function() default\_save\_name default save name for saving, don't change, change save\_name in save\_param dim\_point\_size dim reduction plot: point size

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### **Details**

Description of parameters.

#### Value

ggplot

## See Also

spatDimGenePlot3D

### **Examples**

```
spatDimGenePlot2D(gobject)
```

spatDimGenePlot3D

spatDimGenePlot3D

## **Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

```
spatDimGenePlot3D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("horizontal", "vertical"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 dim3_to_use = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
 genes,
 cluster_column = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1.5,
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 label_size = 16,
  genes_low_color = "blue",
 genes_mid_color = "white",
 genes_high_color = "red",
 dim_point_size = 3,
```

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```
nn_network_alpha = 0.5,
      show_spatial_network = F,
      spatial_network_name = "spatial_network",
      network_color = "lightgray",
      spatial_network_alpha = 0.5,
      show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_grid_alpha = 0.5,
      spatial_point_size = 3,
      legend_text_size = 12,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimGenePlot3D"
    )
Arguments
   gobject
                    giotto object
    expression_values
                    gene expression values to use
   plot_alignment direction to align plot
    dim_reduction_to_use
                    dimension reduction to use
    dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   dim3_to_use
                    dimension to use on z-axis
    genes
                    genes to show
    show_NN_network
                    show underlying NN network
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
                    name of NN network to use, if show_NN_network = TRUE
   network_name
   dim_point_size dim reduction plot: point size
    spatial_network_name
                    name of spatial network to use
    spatial_grid_name
                    name of spatial grid to use
    spatial_point_size
```

spatial plot: point size

```
show plots
show_plot
return_plot
                  return plotly object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
                  default save name for saving, don't change, change save_name in save_param
edge_alpha_dim dim reduction plot: column to use for alpha of the edges
scale_alpha_with_expression
                  scale expression with ggplot alpha parameter
                  size of point (cell)
point_size
show_legend
                  show legend
```

### **Details**

Description of parameters.

### Value

plotly

## **Examples**

spatDimGenePlot3D(gobject)

spatDimPlot spatDimPlot

## **Description**

Visualize cells according to spatial AND dimension reduction coordinates 2D

## Usage

```
spatDimPlot(gobject, ...)
```

## **Arguments**

spat\_enr\_names names of spatial enrichment results to include

cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs dim\_point\_size size of points in dim. reduction space dim\_point\_border\_col border color of points in dim. reduction space dim\_point\_border\_stroke border stroke of points in dim. reduction space spat\_point\_size size of spatial points spat\_point\_border\_col border color of spatial points spat\_point\_border\_stroke border stroke of spatial points dim\_show\_cluster\_center show the center of each cluster dim\_show\_center\_label provide a label for each cluster dim\_center\_point\_size size of the center point  ${\tt dim\_center\_point\_border\_col}$ border color of center point dim\_center\_point\_border\_stroke stroke size of center point dim\_label\_size size of the center label dim\_label\_fontface font of the center label spat\_show\_cluster\_center show the center of each cluster spat\_show\_center\_label provide a label for each cluster spat\_center\_point\_size size of the center point spat\_label\_size size of the center label spat\_label\_fontface

font of the center label

```
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
nn_network_alpha
                 column to use for alpha of the edges
show_spatial_network
                 show spatial network
spat_network_name
                 name of spatial network to use
spat_network_color
                 color of spatial network
show_spatial_grid
                 show spatial grid
spat_grid_name name of spatial grid to use
spat_grid_color
                 color of spatial grid
show_other_cells
                 display not selected cells
other_cell_color
                 color of not selected cells
dim_other_point_size
                 size of not selected dim cells
spat_other_point_size
                 size of not selected spat cells
spat_other_cells_alpha
                 alpha of not selected spat cells
dim_show_legend
                 show legend of dimension reduction plot
spat_show_legend
                 show legend of spatial plot
show_plot
                 show plot
return_plot
                 return ggplot object
save_plot
                 directly save the plot [boolean]
                 list of saving parameters from all_plots_save_function()
save_param
default_save_name
                 default save name for saving, don't change, change save_name in save_param
```

## **Details**

Description of parameters.

# Value

ggplot

### See Also

spatDimPlot2D and spatDimPlot3D for 3D visualization.

#### **Examples**

```
spatDimPlot(gobject)
```

spatDimPlot2D

spatDimPlot2D

### **Description**

Visualize cells according to spatial AND dimension reduction coordinates 2D

```
spatDimPlot2D(
 gobject,
 plot_alignment = c("vertical", "horizontal"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
  sdimx = "sdimx",
 sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_color = NULL,
  color_as_factor = T,
 cell_color_code = NULL,
 cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  dim_point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 dim_show_cluster_center = F,
 dim_show_center_label = T,
 dim_center_point_size = 4,
 dim_center_point_border_col = "black",
 dim_center_point_border_stroke = 0.1,
 dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
 nn_network_to_use = "sNN",
```

```
network_name = "sNN.pca",
      nn_network_alpha = 0.05,
      show_spatial_network = F,
      spat_network_name = "spatial_network",
      spat_network_color = "blue",
      spat_network_alpha = 0.5,
      show_spatial_grid = F,
      spat_grid_name = "spatial_grid",
      spat_grid_color = "blue",
      show_other_cells = T,
      other_cell_color = "lightgrey",
      dim_other_point_size = 1,
      spat_other_point_size = 1,
      spat_other_cells_alpha = 0.5,
      dim_show_legend = F,
      spat_show_legend = F,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimPlot2D"
    )
Arguments
    gobject
                     giotto object
   plot_alignment direction to align plot
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
   dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
                     = spatial dimension to use on x-axis
    sdimx
                     = spatial dimension to use on y-axis
    sdimy
    spat_enr_names names of spatial enrichment results to include
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
```

select subset of cells/clusters based on cell\_color parameter

select subset of cells based on cell IDs

select\_cells

dim\_point\_size size of points in dim. reduction space dim\_point\_border\_col border color of points in dim. reduction space dim\_point\_border\_stroke border stroke of points in dim. reduction space spat\_point\_size size of spatial points spat\_point\_border\_col border color of spatial points spat\_point\_border\_stroke border stroke of spatial points dim\_show\_cluster\_center show the center of each cluster dim\_show\_center\_label provide a label for each cluster dim\_center\_point\_size size of the center point dim\_center\_point\_border\_col border color of center point dim\_center\_point\_border\_stroke stroke size of center point dim\_label\_size size of the center label dim\_label\_fontface font of the center label spat\_show\_cluster\_center show the center of each cluster spat\_show\_center\_label provide a label for each cluster spat\_center\_point\_size size of the center point spat\_label\_size size of the center label spat\_label\_fontface font of the center label show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name nn\_network\_alpha column to use for alpha of the edges show\_spatial\_network show spatial network spat\_network\_name name of spatial network to use spat\_network\_color color of spatial network

```
show_spatial_grid
                  show spatial grid
spat_grid_name name of spatial grid to use
spat_grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
\dim_{-}other_{-}point_{-}size
                  size of not selected dim cells
spat_other_point_size
                  size of not selected spat cells
spat_other_cells_alpha
                  alpha of not selected spat cells
dim_show_legend
                  show legend of dimension reduction plot
spat_show_legend
                  show legend of spatial plot
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

## **Details**

Description of parameters.

## Value

ggplot

## See Also

 ${\tt spatDimPlot3D}$ 

## **Examples**

spatDimPlot2D(gobject)

spatDimPlot3D

spatDimPlot3D

## **Description**

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

```
spatDimPlot3D(
 gobject,
 plot_alignment = c("horizontal", "vertical"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
 dim3_to_use = 3,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
  show_cluster_center = F,
  show_center_label = T,
 center_point_size = 4,
 label_size = 16,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1.5,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
 dim_point_size = 3,
 nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
 network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
 x_ticks = NULL,
 y_ticks = NULL,
  z_ticks = NULL,
```

```
legend_text_size = 12,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimPlot3D"
Arguments
    gobject
                      giotto object
    plot_alignment direction to align plot
    dim_reduction_to_use
                      dimension reduction to use
    dim_reduction_name
                      dimension reduction name
    dim1_to_use
                      dimension to use on x-axis
    dim2_to_use
                      dimension to use on y-axis
                      dimension to use on z-axis
    dim3_to_use
    sdimx
                      = spatial dimension to use on x-axis
    sdimy
                      = spatial dimension to use on y-axis
                      = spatial dimension to use on z-axis
    sdimz
    show_NN_network
                      show underlying NN network
    nn_network_to_use
                      type of NN network to use (kNN vs sNN)
                      name of NN network to use, if show_NN_network = TRUE
    network_name
    show_cluster_center
                      show the center of each cluster
    show_center_label
                      provide a label for each cluster
    center_point_size
                      size of the center point
    label_size
                      size of the center label
    select_cell_groups
                      select subset of cells/clusters based on cell_color parameter
                      select subset of cells based on cell IDs
    select_cells
    show_other_cells
                      display not selected cells
    other_cell_color
                      color of not selected cells
    other_point_size
                      size of not selected cells
    cell_color
                      color for cells (see details)
    color_as_factor
                      convert color column to factor
    cell_color_code
                      named vector with colors
```

```
dim_point_size size of points in dim. reduction space
nn_network_alpha
                  column to use for alpha of the edges
show_spatial_network
                  show spatial network
spatial_network_name
                  name of spatial network to use
spatial_network_alpha
                  alpha of spatial network
show_spatial_grid
                  show spatial grid
spatial_grid_name
                  name of spatial grid to use
spatial_grid_color
                  color of spatial grid
spatial_point_size
                  size of spatial points
                  show plot
show_plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
dim_point_border_col
                  border color of points in dim. reduction space
dim_point_border_stroke
                  border stroke of points in dim. reduction space
spatial_network_color
                  color of spatial network
spatial_other_point_size
                  size of not selected spatial points
{\tt spatial\_other\_cells\_alpha}
                  alpha of not selected spatial points
dim_other_point_size
                  size of not selected dim. reduction points
show_legend
                  show legend
```

### **Details**

Description of parameters.

### Value

plotly

### **Examples**

spatDimPlot3D(gobject)

spatGenePlot 215

spatGenePlot	spatGenePlot

## Description

Visualize cells and gene expression according to spatial coordinates

### Usage

```
spatGenePlot(gobject, ...)
```

### **Arguments**

show\_legend

```
gobject
                  giotto object
                  parameters for cowplot::save_plot()
expression_values
                  gene expression values to use
                  genes to show
genes
genes_high_color
                  color represents high gene expression
genes_mid_color
                  color represents middle gene expression
genes_low_color
                  color represents low gene expression
show_network
                  show underlying spatial network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
midpoint
                  expression midpoint
scale_alpha_with_expression
                  scale expression with ggplot alpha parameter
                  size of point (cell)
point_size
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
                  cowplot param: how many columns
cow_n_col
                  cowplot param: relative height
cow_rel_h
cow_rel_w
                  cowplot param: relative width
cow_align
                  cowplot param: how to align
```

show legend

216 spatGenePlot2D

```
show_plot show plots

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function()

default_save_name

default save name for saving, don't change, change save_name in save_param
```

### **Details**

Description of parameters.

### Value

ggplot

### See Also

```
spatGenePlot3D and spatGenePlot2D
```

### **Examples**

```
spatGenePlot(gobject)
```

spatGenePlot2D spatGenePlot2D

## **Description**

Visualize cells and gene expression according to spatial coordinates

```
spatGenePlot2D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 genes_high_color = "darkred",
 genes_mid_color = "white",
 genes_low_color = "darkblue",
  show_network = F,
 network_color = NULL,
 spatial_network_name = "spatial_network",
 edge_alpha = NULL,
  show_grid = F,
 grid_color = NULL,
  spatial_grid_name = "spatial_grid",
 midpoint = 0,
  scale_alpha_with_expression = FALSE,
 point_size = 1,
 point_border_col = "black",
 point_border_stroke = 0.1,
```

spatGenePlot2D 217

```
show_legend = T,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatGenePlot2D"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
                     genes to show
    genes
    genes_high_color
                     color represents high gene expression
    genes_mid_color
                     color represents middle gene expression
    genes_low_color
                     color represents low gene expression
                     show underlying spatial network
    show_network
                     color of spatial network
    network_color
    spatial_network_name
                     name of spatial network to use
    show_grid
                     show spatial grid
    grid_color
                     color of spatial grid
    spatial_grid_name
                     name of spatial grid to use
                     expression midpoint
    midpoint
    scale_alpha_with_expression
                     scale expression with ggplot alpha parameter
    point_size
                     size of point (cell)
    point_border_col
                     color of border around points
    point_border_stroke
                     stroke size of border around points
    show_legend
                     show legend
    cow_n_col
                     cowplot param: how many columns
    cow_rel_h
                     cowplot param: relative height
    cow_rel_w
                     cowplot param: relative width
                     cowplot param: how to align
    cow_align
    show_plot
                     show plots
```

return ggplot object

return\_plot

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### **Details**

Description of parameters.

### Value

ggplot

### See Also

```
spatGenePlot3D
```

### **Examples**

```
spatGenePlot2D(gobject)
```

spatGenePlot3D

spatGenePlot3D

### **Description**

Visualize cells and gene expression according to spatial coordinates

```
spatGenePlot3D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 genes,
  show_network = F,
 network_color = NULL,
  spatial_network_name = "spatial_network",
 edge_alpha = NULL,
  show\_grid = F,
 cluster_column = NULL,
 select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
 genes_low_color = "blue",
  spatial_grid_name = "spatial_grid",
 point_size = 2,
```

spatGenePlot3D 219

```
show_legend = T,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatGenePlot3D"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
    genes
                     genes to show
                     show underlying spatial network
    show_network
                     color of spatial network
    network_color
    spatial_network_name
                     name of spatial network to use
                     show spatial grid
    show_grid
    genes_high_color
                     color represents high gene expression
    genes_mid_color
                     color represents middle gene expression
    genes_low_color
                     color represents low gene expression
    spatial_grid_name
                     name of spatial grid to use
    point_size
                     size of point (cell)
    show_legend
                     show legend
    show_plot
                     show plots
    return_plot
                     return ggplot object
    save_plot
                     directly save the plot [boolean]
                     list of saving parameters from all_plots_save_function()
    save_param
    default_save_name
                     default save name for saving, don't change, change save_name in save_param
    grid_color
                     color of spatial grid
    midpoint
                     expression midpoint
    scale_alpha_with_expression
                     scale expression with ggplot alpha parameter
```

parameters for cowplot::save\_plot()

220 Spatial\_AEH

### **Details**

Description of parameters.

## Value

ggplot

### **Examples**

```
spatGenePlot3D(gobject)
```

Spatial\_AEH

Spatial\_AEH

## Description

calculate automatic expression histology with spatialDE method

## Usage

```
Spatial_AEH(
  gobject = NULL,
  results = NULL,
  pattern_num = 5,
  1 = 1.05,
  show_AEH = T,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_alpha = 1,
  low_color = "blue"
  mid_color = "white",
  high_color = "red",
  midpoint = 0,
  python_path = NULL
)
```

## Arguments

gobject Giotto object

results output from spatial\_DE

pattern\_num the number of gene expression patterns

show\_AEH show AEH plot

python\_path specify specific path to python if required

## **Details**

Description.

Spatial\_DE 221

### Value

a list or a dataframe of SVs

## **Examples**

```
Spatial_DE(gobject)
```

Spatial\_DE

Spatial\_DE

## **Description**

calculate spatial varible genes with spatialDE method

# Usage

```
Spatial_DE(
  gobject = NULL,
  show_plot = T,
  size = c(4, 2, 1),
  color = c("blue", "green", "red"),
  sig_alpha = 0.5,
  unsig_alpha = 0.5,
  python_path = NULL
)
```

## **Arguments**

gobject Giotto object show\_plot show FSV plot

python\_path specify specific path to python if required

## **Details**

Description.

## Value

a list or a dataframe of SVs

# Examples

```
Spatial_DE(gobject)
```

222 spatPlot

spatPlot spatPlot

## **Description**

Visualize cells according to spatial coordinates

#### **Usage**

```
spatPlot(gobject, ...)
```

### **Arguments**

```
giotto object
gobject
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
spat_enr_names names of spatial enrichment results to include
                  color for cells (see details)
cell_color
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
show_network
                  show underlying spatial network
spatial_network_name
                  name of spatial network to use
```

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```
network_color
                color of spatial network
network_alpha alpha of spatial network
show_grid
                  show spatial grid
spatial_grid_name
                  name of spatial grid to use
grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  point size of not selected cells
other_cells_alpha
                  alpha of not selected cells
coord_fix_ratio
                  fix ratio between x and y-axis
title
                  title of plot
                  show legend
show_legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function()
default_save_name
```

default save name for saving, don't change, change save\_name in save\_param

## **Details**

Description of parameters.

### Value

ggplot

## See Also

spatPlot3D

## **Examples**

spatPlot(gobject)

224 spatPlot2D

spatPlot2D

spatPlot2D

## **Description**

Visualize cells according to spatial coordinates

```
spatPlot2D(
 gobject,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
 center_point_size = 4,
 center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network\_alpha = 1,
  show\_grid = F,
  spatial_grid_name = "spatial_grid",
  grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
 other_cells_alpha = 0.1,
  coord_fix_ratio = 0.6,
  title = NULL,
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
 default_save_name = "spatPlot2D"
```

spatPlot2D 225

)

### Arguments

gobject giotto object sdimx x-axis dimension name (default = 'sdimx') sdimy y-axis dimension name (default = 'sdimy') spat\_enr\_names names of spatial enrichment results to include cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors cell\_color\_gradient vector with 3 colors for numeric data gradient\_midpoint midpoint for color gradient gradient\_limits vector with lower and upper limits select\_cell\_groups select subset of cells/clusters based on cell\_color parameter select\_cells select subset of cells based on cell IDs point\_size size of point (cell) point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_cluster\_center plot center of selected clusters show\_center\_label plot label of selected clusters center\_point\_size size of center points label\_size size of labels label\_fontface font of labels show underlying spatial network show\_network spatial\_network\_name name of spatial network to use color of spatial network network\_color network\_alpha alpha of spatial network show\_grid show spatial grid spatial\_grid\_name name of spatial grid to use color of spatial grid grid\_color show\_other\_cells display not selected cells

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```
other_cell_color
                  color of not selected cells
other_point_size
                  point size of not selected cells
other_cells_alpha
                  alpha of not selected cells
coord_fix_ratio
                  fix ratio between x and y-axis
                  title of plot
title
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

#### **Details**

Description of parameters.

#### Value

ggplot

## See Also

```
spatPlot3D
```

## **Examples**

```
spatPlot2D(gobject)
```

spatPlot3D

spatPlot3D

## **Description**

Visualize cells according to spatial coordinates

```
spatPlot3D(
  gobject,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  point_size = 3,
  cell_color = NULL,
  cell_color_code = NULL,
```

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```
select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 0.5,
  show_network = F,
 network_color = NULL,
  network_alpha = 1,
 other_cell_alpha = 0.5,
  spatial_network_name = "spatial_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  title = "",
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
 x_{ticks} = NULL,
 y_ticks = NULL,
 z_ticks = NULL,
 show_plot = NA,
 return_plot = NA,
  save_plot = NA,
 save_param = list(),
 default_save_name = "spat3D"
)
```

# Arguments

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
                  z-axis dimension name (default = 'sdimy')
sdimz
                  size of point (cell)
point_size
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
                  show underlying spatial network
show_network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
```

```
color of spatial grid
grid_color
spatial_grid_name
                  name of spatial grid to use
                  title of plot
title
                  show legend
show_legend
axis_scale
                  the way to scale the axis
custom_ratio
                  customize the scale of the plot
                  set the number of ticks on the x-axis
x_ticks
y_ticks
                  set the number of ticks on the y-axis
z_ticks
                  set the number of ticks on the z-axis
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function()
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

### **Details**

Description of parameters.

## Value

ggplot

## **Examples**

```
spatPlot3D(gobject)
```

```
specific \textit{CellCellcommunicationScores} \\ specific \textit{CellCellcommunicationScores}
```

## **Description**

Specific Cell-Cell communication scores based on spatial expression of interacting cells

```
specificCellCellcommunicationScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "cell_types",
  random_iter = 100,
  cell_type_1 = "astrocyte",
  cell_type_2 = "endothelial",
  gene_set_1,
```

```
gene_set_2,
log2FC_addendum = 0.1,
min_observations = 2,
verbose = T
```

## **Arguments**

```
gobject
                  giotto object to use
spatial_network_name
                  spatial network to use for identifying interacting cells
cluster_column cluster column with cell type information
random_iter
                  number of iterations
cell_type_1
                  first cell type
cell_type_2
                  second cell type
                  first specific gene set from gene pairs
gene_set_1
gene_set_2
                  second specific gene set from gene pairs
log2FC\_addendum
                  addendum to add when calculating log2FC
min_observations
                  minimum number of interactions needed to be considered
verbose
                  verbose
```

#### **Details**

Details will follow.

### Value

Cell-Cell communication scores for gene pairs based on spatial interaction

## **Examples**

```
specificCellCellcommunicationScores(gobject)
```

## Description

Merge selected clusters based on pairwise correlation scores and size of cluster.

### Usage

```
split_dendrogram_in_two(dend)
```

### **Arguments**

dend dendrogram object

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#### Value

list of two dendrograms and height of node

## **Examples**

```
split_dendrogram_in_two(dend)
```

```
stitchFieldCoordinates
```

stitchFieldCoordinates

### **Description**

Helper function to stitch field coordinates together to form one complete picture

## Usage

```
stitchFieldCoordinates(
  location_file,
  offset_file,
  cumulate_offset_x = F,
  cumulate_offset_y = F,
  field_col = "Field of View",
  X_coord_col = "X",
  Y_coord_col = "Y",
  reverse_final_x = F,
  reverse_final_y = T
)
```

## **Arguments**

```
location_file location dataframe with X and Y coordinates
offset_file
                  dataframe that describes the offset for each field (see details)
cumulate_offset_x
                  (boolean) Do the x-axis offset values need to be cumulated?
cumulate_offset_y
                  (boolean) Do the y-axis offset values need to be cumulated?
field_col
                  column that indicates the field within the location_file
                  column that indicates the x coordinates
X_coord_col
Y_coord_col
                  column that indicates the x coordinates
reverse_final_x
                  (boolean) Do the final x coordinates need to be reversed?
reverse_final_y
                  (boolean) Do the final y coordinates need to be reversed?
```

subClusterCells 231

#### **Details**

Stitching of fields:

- 1. have cell locations: at least 3 columns: field, X, Y
- 2. create offset file: offset file has 3 columns: field, x\_offset, y\_offset
- 3. create new cell location file by stitching original cell locations with stitchFieldCoordinates
- 4. provide new cell location file to createGiottoObject

#### Value

Updated location dataframe with new X ['X\_final'] and Y ['Y\_final'] coordinates

### **Examples**

```
stitchFieldCoordinates(gobject)
```

subClusterCells

subClusterCells

## **Description**

subcluster cells

```
subClusterCells(
  gobject,
  name = "sub_clus",
  cluster_method = c("leiden", "louvain_community", "louvain_multinet"),
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 1,
  gamma = 1,
  omega = 1,
  python_path = NULL,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T,
)
```

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### **Arguments**

gobject giotto object

name name for new clustering result

cluster\_method clustering method to use

cluster\_column cluster column to subcluster

selected\_clusters

only do subclustering on these clusters

hvg\_param parameters for calculateHVG

hvg\_min\_perc\_cells

threshold for detection in min percentage of cells

 $hvg\_mean\_expr\_det$ 

threshold for mean expression level in cells with detection

use\_all\_genes\_as\_hvg

forces all genes to be HVG and to be used as input for PCA

min\_nr\_of\_hvg minimum number of HVG, or all genes will be used as input for PCA

pca\_param parameters for runPCA

nn\_param parameters for parameters for createNearestNetwork

k\_neighbors number of k for createNearestNetwork

resolution resolution gamma gamma omega omega

python\_path specify specific path to python if required

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

 ${\tt network\_name} \qquad {\tt name} \ of \ NN \ network \ to \ use$ 

return\_gobject boolean: return giotto object (default = TRUE)

verbose verbose

... additional parameters

### **Details**

Description of Louvain clustering method.

### Value

giotto object appended with new cluster

## **Examples**

subClusterCells(gobject)

subsetGiotto 233

subsetGiotto subsetGiotto

## Description

subsets Giotto object including previous analyses.

## Usage

```
subsetGiotto(gobject, cell_ids = NULL, gene_ids = NULL)
```

# Arguments

```
gobject giotto object
cell_ids cell IDs to keep
gene_ids gene IDs to keep
```

### Value

giotto object

# **Examples**

```
subsetGiotto(gobject)
```

viewHMRFresults

viewHMRFresults

## Description

View results from doHMRF.

## Usage

```
viewHMRFresults(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

## Arguments

```
gobject giotto object
```

 $\begin{array}{ll} \mbox{HMRF output} & \mbox{HMRF output from doHMRF} \\ \mbox{k} & \mbox{number of HMRF domains} \end{array}$ 

betas\_to\_view results from different betas that you want to view

... paramters to visPlot()

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### **Details**

Description ...

### Value

spatial plots with HMRF domains

### See Also

```
visPlot
```

### **Examples**

viewHMRFresults(gobject)

viewHMRFresults2D

viewHMRFresults2D

## Description

View results from doHMRF.

## Usage

```
viewHMRFresults2D(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

# Arguments

gobject giotto object

HMRF output from doHMRF k number of HMRF domains

betas\_to\_view results from different betas that you want to view

... paramters to visPlot()

## **Details**

Description ...

### Value

spatial plots with HMRF domains

## See Also

```
spatPlot2D
```

viewHMRFresults3D 235

### **Examples**

```
viewHMRFresults2D(gobject)
```

viewHMRFresults3D

viewHMRFresults3D

# Description

View results from doHMRF.

# Usage

```
viewHMRFresults3D(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

# Arguments

gobject giotto object

HMRF output from doHMRF k number of HMRF domains

betas\_to\_view results from different betas that you want to view

... paramters to visPlot()

## **Details**

Description ...

# Value

spatial plots with HMRF domains

### See Also

```
spatPlot3D
```

# **Examples**

```
viewHMRFresults3D(gobject)
```

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violinPlot

violinPlot

### **Description**

Creates heatmap based on identified clusters

## Usage

```
violinPlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column,
  cluster_custom_order = NULL,
  color_violin = c("genes", "cluster"),
  cluster_color_code = NULL,
  strip_position = c("top", "right", "left", "bottom"),
  strip\_text = 7,
  axis_text_x_size = 10,
  axis_text_y_size = 6,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "violinPlot"
)
```

# Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
                  genes to plot
genes
cluster_column name of column to use for clusters
cluster_custom_order
                  custom order of clusters
color_violin
                  color violin according to genes or clusters
cluster_color_code
                  color code for clusters
strip_position position of gene labels
strip_text
                  size of strip text
axis_text_x_size
                  size of x-axis text
axis_text_y_size
                  size of y-axis text
show_plot
                  show plot
return_plot
                  return ggplot object
```

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```
save_plot directly save the plot [boolean]
save_param list of saving parameters from all_plots_save_function()
default_save_name
    default save name for saving, don't change, change save_name in save_param
```

### **Details**

Correlation heatmap of clusters vs genes.

#### Value

ggplot

## **Examples**

```
violinPlot(gobject)
```

visDimGenePlot

visDimGenePlot

## **Description**

Visualize cells and gene expression according to dimension reduction coordinates

```
visDimGenePlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
```

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```
cow_align = "h",
show_legend = T,
plot_method = c("ggplot", "plotly"),
show_plots = F
```

# **Arguments**

```
gobject
                 giotto object
expression_values
                 gene expression values to use
                 genes to show
genes
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
dim3 to use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
edge_alpha
                 column to use for alpha of the edges
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
point_size
                 size of point (cell)
point_border_col
                 color of border around points
point_border_stroke
                 stroke size of border around points
midpoint
                 size of point (cell)
cow_n_col
                 cowplot param: how many columns
                 cowplot param: relative height
cow_rel_h
                 cowplot param: relative width
cow_rel_w
                 cowplot param: how to align
cow_align
show_legend
                 show legend
show_plots
                 show plots
```

### **Details**

Description of parameters.

## Value

ggplot

## **Examples**

```
visDimGenePlot(gobject)
```

### **Description**

Visualize cells and gene expression according to dimension reduction coordinates

### Usage

```
visDimGenePlot_2D_ggplot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = "red",
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_legend = T,
  show_plots = F
```

## **Arguments**

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
dim_reduction_to_use
dimension reduction to use
dim_reduction_name
dimension reduction name
dim1_to_use
dimension to use on x-axis
```

```
dim2_to_use
                 dimension to use on y-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
edge_alpha
                 column to use for alpha of the edges
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
                 size of point (cell)
point_size
point_border_col
                 color of border around points
point_border_stroke
                 stroke size of border around points
                 size of point (cell)
midpoint
cow_n_col
                 cowplot param: how many columns
                 cowplot param: relative height
cow_rel_h
                 cowplot param: relative width
cow_rel_w
cow_align
                 cowplot param: how to align
show_legend
                 show legend
show_plots
                 show plots
```

### **Details**

Description of parameters.

### Value

ggplot

## **Examples**

```
visDimGenePlot_2D_ggplot(gobject)
```

# Description

Visualize cells and gene expression according to dimension reduction coordinates

#### Usage

```
visDimGenePlot_3D_plotly(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = 3,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_legend = T,
  show_plots = F
)
```

## **Arguments**

```
gobject
                 giotto object
expression_values
                 gene expression values to use
genes
                 genes to show
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
edge_alpha
                 column to use for alpha of the edges
point_size
                 size of point (cell)
show_legend
                 show legend
                 show plots
show_plots
```

### **Details**

Description of parameters.

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### Value

ggplot

#### **Examples**

```
visDimGenePlot_3D_plotly(gobject)
```

visDimPlot

visDimPlot

### **Description**

Visualize cells according to dimension reduction coordinates

```
visDimPlot(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  plot_method = c("ggplot", "plotly"),
  show_legend = T,
  show_plot = F,
  return_plot = TRUE,
  save_plot = F,
  save_dir = NULL,
```

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```
save_folder = NULL,
      save_name = NULL.
      save_format = NULL,
      show_saved_plot = F,
    )
Arguments
                      giotto object
    gobject
    dim_reduction_to_use
                      dimension reduction to use
    {\tt dim\_reduction\_name}
                      dimension reduction name
    dim1_to_use
                      dimension to use on x-axis
    dim2_to_use
                      dimension to use on y-axis
                      dimension to use on z-axis
    dim3_to_use
    show_NN_network
                      show underlying NN network
    nn_network_to_use
                      type of NN network to use (kNN vs sNN)
                      name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color
                      color for cells (see details)
    color_as_factor
                      convert color column to factor
    cell_color_code
                      named vector with colors
    show_cluster_center
                      plot center of selected clusters
    show_center_label
                      plot label of selected clusters
    center_point_size
                      size of center points
    label_size
                      size of labels
    label_fontface font of labels
    edge_alpha
                      column to use for alpha of the edges
    point_size
                      size of point (cell)
    point_border_col
                      color of border around points
    point_border_stroke
                      stroke size of border around points
    show_legend
                      show legend
    show_plot
                      show plot
    return_plot
                      return ggplot object
                      directly save the plot [boolean]
    save_plot
```

directory to save the plot

save\_dir

### **Details**

Description of parameters.

### Value

ggplot or plotly

## **Examples**

```
visDimPlot(gobject)
```

```
visDimPlot_2D_ggplot visDimPlot_2D_ggplot
```

### **Description**

Visualize cells according to dimension reduction coordinates

```
visDimPlot_2D_ggplot(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
```

```
edge_alpha = NULL,
      point_size = 1,
      point_border_col = "black",
      point_border_stroke = 0.1,
      show_legend = T,
      show_plot = F,
      return_plot = TRUE,
      save_plot = F,
      save_dir = NULL,
      save_folder = NULL,
      save_name = NULL,
      save_format = NULL,
      show_saved_plot = F,
    )
Arguments
    gobject
                     giotto object
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
```

size of center points

#### **Details**

Description of parameters.

#### Value

ggplot

## **Examples**

```
visDimPlot_2D_ggplot(gobject)
```

```
visDimPlot_2D_plotly visDimPlot_2D_plotly
```

### **Description**

Visualize cells according to dimension reduction coordinates

```
visDimPlot_2D_plotly(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  color_as_factor = T,
  cell_color = NULL,
  cell_color_code = NULL,
  show_cluster_center = F,
  show_center_label = T,
```

```
center_point_size = 4,
label_size = 4,
edge_alpha = NULL,
point_size = 5
)
```

## **Arguments**

```
gobject
                 giotto object
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
                 dimension to use on x-axis
dim1_to_use
dim2_to_use
                 dimension to use on y-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
color_as_factor
                 convert color column to factor
cell_color
                 color for cells (see details)
cell_color_code
                 named vector with colors
show_cluster_center
                 plot center of selected clusters
show_center_label
                 plot label of selected clusters
center_point_size
                 size of center points
label_size
                 size of labels
edge_alpha
                 column to use for alpha of the edges
point_size
                 size of point (cell)
```

### **Details**

Description of parameters.

### Value

plotly

## Examples

```
visDimPlot_2D_plotly(gobject)
```

```
visDimPlot_3D_plotly
```

### **Description**

Visualize cells according to dimension reduction coordinates

## Usage

```
visDimPlot_3D_plotly(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = 3,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  color_as_factor = T,
  cell_color = NULL,
  cell_color_code = NULL,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 4,
  edge_alpha = NULL,
  point_size = 1
```

# **Arguments**

```
giotto object
gobject
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
```

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```
name of NN network to use, if show_NN_network = TRUE
network_name
color_as_factor
                  convert color column to factor
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
                  column to use for alpha of the edges
edge_alpha
point_size
                  size of point (cell)
```

### **Details**

Description of parameters.

### Value

plotly

### **Examples**

```
visDimPlot_3D_plotly(gobject)
```

visForceLayoutPlot visForceLayoutPlot

## Description

Visualize cells according to forced layout algorithm coordinates

```
visForceLayoutPlot(
  gobject,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  layout_name = "layout",
  dim1_to_use = 1,
  dim2_to_use = 2,
  show_NN_network = T,
  cell_color = NULL,
  color_as_factor = F,
  cell_color_code = NULL,
  edge_alpha = NULL,
  point_size = 1,
```

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```
point_border_col = "black",
point_border_stroke = 0.1,
show_legend = T,
show_plot = F,
return_plot = TRUE,
save_plot = F,
save_dir = NULL,
save_folder = NULL,
save_format = NULL,
show_saved_plot = F,
...
)
```

### Arguments

```
giotto object
gobject
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
network_name
                  NN network to use
                  name of layout to use
layout_name
dim1_to_use
                  dimension to use on x-axis
                  dimension to use on y-axis
dim2_to_use
show_NN_network
                  show underlying NN network
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
                  column to use for alpha of the edges
edge_alpha
                  size of point (cell)
point_size
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
                  show legend
show_legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  directory to save the plot
save_dir
save_folder
                  (optional) folder in directory to save the plot
                  name of plot
save_name
save_format
                  format of plot (e.g. tiff, png, pdf, ...)
show_saved_plot
                  load & display the saved plot
```

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### **Details**

Description of parameters.

#### Value

ggplot

### **Examples**

visForceLayoutPlot(gobject)

visGenePlot

visGenePlot

## Description

Visualize cells and gene expression according to spatial coordinates

```
visGenePlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  edge_alpha = NULL,
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  midpoint = 0,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  plot_method = c("ggplot", "plotly"),
  show_plots = F
```

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### **Arguments**

gobject giotto object expression\_values gene expression values to use genes to show genes genes\_high\_color color represents high gene expression genes\_mid\_color color represents middle gene expression genes\_low\_color color represents low gene expression show underlying spatial network show\_network network\_color color of spatial network spatial\_network\_name name of spatial network to use show spatial grid show\_grid grid\_color color of spatial grid spatial\_grid\_name name of spatial grid to use midpoint expression midpoint scale\_alpha\_with\_expression scale expression with ggplot alpha parameter size of point (cell) point\_size point\_border\_col color of border around points point\_border\_stroke stroke size of border around points show\_legend show legend cow\_n\_col cowplot param: how many columns cow\_rel\_h cowplot param: relative height cow\_rel\_w cowplot param: relative width cow\_align cowplot param: how to align three mode to adjust axis scale axis\_scale x\_ticks number of ticks on x axis number of ticks on y axis y\_ticks number of ticks on z axis z\_ticks plot\_method two methods of plot show plots show\_plots

### **Details**

Description of parameters.

## Value

ggplot or plotly

## **Examples**

```
visGenePlot(gobject)
```

```
\verb|visGenePlot_2D_ggplot| | \textit{visGenePlot}_2D\_ggplot|
```

# **Description**

Visualize cells and gene expression according to spatial coordinates

# Usage

```
visGenePlot_2D_ggplot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  genes_high_color = "darkred",
  genes_mid_color = "white",
  genes_low_color = "darkblue",
  show_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  edge_alpha = NULL,
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  midpoint = 0,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_plots = F
)
```

# Arguments

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
genes_high_color
color represents high gene expression
genes_mid_color
color represents middle gene expression
```

genes\_low\_color

color represents low gene expression

show\_network show underlying spatial network

network\_color color of spatial network

spatial\_network\_name

name of spatial network to use

show\_grid show spatial grid grid\_color color of spatial grid

spatial\_grid\_name

name of spatial grid to use

midpoint expression midpoint
scale\_alpha\_with\_expression

scale expression with ggplot alpha parameter

point\_size size of point (cell)

point\_border\_col

color of border around points

point\_border\_stroke

stroke size of border around points

show\_legend show legend

cow\_n\_colcowplot param: how many columnscow\_rel\_hcowplot param: relative heightcow\_rel\_wcowplot param: relative widthcow\_aligncowplot param: how to align

show\_plots show plots

# **Details**

Description of parameters.

#### Value

ggplot

# **Examples**

visGenePlot\_2D\_ggplot(gobject)

 ${\tt visGenePlot\_3D\_plotly} \ \ {\it visGenePlot\_3D\_plotly}$ 

# Description

Visualize cells and gene expression according to spatial coordinates

# Usage

```
visGenePlot_3D_plotly(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  show_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  edge_alpha = NULL,
  show\_grid = F,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  spatial_grid_name = "spatial_grid",
  point_size = 1,
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  show_plots = F
gobject
                giotto object
```

## **Arguments**

```
expression_values
                  gene expression values to use
genes
                  genes to show
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
genes_high_color
                  color represents high gene expression
genes_mid_color
                  color represents middle gene expression
genes_low_color
                  color represents low gene expression
spatial_grid_name
                  name of spatial grid to use
                  size of point (cell)
point_size
show_legend
                  show legend
axis_scale
                  three mode to adjust axis scale
x_ticks
                  number of ticks on x axis
y_ticks
                  number of ticks on y axis
```

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```
z_ticks number of ticks on z axis
show_plots show plots
grid_color color of spatial grid
cow_n_col cowplot param: how many columns
cow_rel_h cowplot param: relative height
cow_rel_w cowplot param: relative width
cow_align cowplot param: how to align
```

#### **Details**

Description of parameters.

#### Value

plotly

# **Examples**

```
visGenePlot_3D_plotly(gobject)
```

visPlot visPlot

# Description

Visualize cells according to spatial coordinates

```
visPlot(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  show_network = F,
  network_color = NULL,
  network_alpha = 1,
  other_cells_alpha = 0.1,
  spatial_network_name = "spatial_network",
  show\_grid = F,
```

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```
grid_color = NULL,
      grid_alpha = 1,
      spatial_grid_name = "spatial_grid",
      coord_fix_ratio = 0.6,
      title = "",
      show_legend = T,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_{ticks} = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      plot_method = c("ggplot", "plotly"),
      show_plot = F,
      return_plot = TRUE,
      save_plot = F,
      save_dir = NULL,
      save_folder = NULL,
      save_name = NULL,
      save_format = NULL,
      show_saved_plot = F,
    )
Arguments
   gobject
                     giotto object
    sdimx
                     x-axis dimension name (default = 'sdimx')
    sdimy
                     y-axis dimension name (default = 'sdimy')
    sdimz
                     z-axis dimension name (default = 'sdimz')
   point_size
                     size of point (cell)
   point_border_col
                     color of border around points
   point_border_stroke
                     stroke size of border around points
    cell_color
                     color for cells (see details)
    cell_color_code
                     named vector with colors
    color_as_factor
                     convert color column to factor
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
    show_other_cells
                     display not selected cells
   other_cell_color
                     color of not selected cells
```

show underlying spatial network

name of spatial network to use

color of spatial network

show\_network

network\_color

spatial\_network\_name

visPlot\_2D\_ggplot

```
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
title
                  title of plot
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_dir
                  directory to save the plot
                  (optional) folder in directory to save the plot
save_folder
                  name of plot
save_name
save_format
                  format of plot (e.g. tiff, png, pdf, ...)
show_saved_plot
                  load & display the saved plot
```

#### **Details**

Description of parameters.

## Value

ggplot

# **Examples**

```
visPlot(gobject)
```

```
visPlot_2D_ggplot
visPlot_2D_ggplot
```

# **Description**

Visualize cells according to spatial coordinates

```
visPlot_2D_ggplot(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  cell_color = NULL,
  cell_color_code = NULL,
```

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```
color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  show_network = F,
  network_color = NULL,
  network_alpha = 1,
  other_cells_alpha = 0.1,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 0.6,
  title = "",
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  show_plot = F,
  return_plot = TRUE,
  save_plot = F,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  save_format = NULL,
  show_saved_plot = F,
)
```

# **Arguments**

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
```

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show\_other\_cells

display not selected cells

other\_cell\_color

color of not selected cells

show\_network show underlying spatial network

network\_color color of spatial network

spatial\_network\_name

name of spatial network to use

show\_grid show spatial grid

grid\_color color of spatial grid

spatial\_grid\_name

name of spatial grid to use

coord\_fix\_ratio

fix ratio between x and y-axis

title title of plot

show\_legend show legend

show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_dir directory to save the plot

save\_folder (optional) folder in directory to save the plot

save\_name name of plot

save\_format format of plot (e.g. tiff, png, pdf, ...)

show\_saved\_plot

load & display the saved plot

# Details

Description of parameters.

#### Value

ggplot

## **Examples**

 ${\tt visPlot\_2D\_ggplot(gobject)}$ 

visPlot\_2D\_plotly 261

```
visPlot_2D_plotly
```

# **Description**

Visualize cells according to spatial coordinates

#### Usage

```
visPlot_2D_plotly(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_network = F,
  network_color = "lightgray",
  network_alpha = 1,
  other_cell_alpha = 0.5,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  grid_alpha = 1,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  show_plot = F
```

# **Arguments**

```
gobject giotto object

sdimx x-axis dimension name (default = 'sdimx')

sdimy y-axis dimension name (default = 'sdimy')

point_size size of point (cell)

cell_color color for cells (see details)

cell_color_code

named vector with colors

color_as_factor

convert color column to factor
```

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```
select_cell_groups
                  select a subset of the groups from cell_color
                  show underlying spatial network
show_network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
                  alpha of spatial grid
grid_alpha
spatial_grid_name
                  name of spatial grid to use
                  show legend
show_legend
show_plot
                  show plot
```

#### **Details**

Description of parameters.

#### Value

plotly

# **Examples**

```
visPlot_2D_plotly(gobject)
```

```
visPlot_3D_plotly
```

# Description

Visualize cells according to spatial coordinates

```
visPlot_3D_plotly(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  point_size = 3,
  cell_color = NULL,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_network = F,
```

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```
network_color = NULL,
network_alpha = 1,
other_cell_alpha = 0.5,
spatial_network_name = "spatial_network",
spatial_grid_name = "spatial_grid",
title = "",
show_legend = T,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
stow_plot = F
```

# Arguments

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
sdimz
                  z-axis dimension name (default = 'sdimz')
point_size
                  size of point (cell)
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
select_cell_groups
                  select a subset of the groups from cell_color
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
spatial_grid_name
                  name of spatial grid to use
                  title of plot
title
show_legend
                  show legend
show_plot
                  show plot
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
color_as_factor
                  convert color column to factor
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
coord_fix_ratio
                  fix ratio between x and y-axis
```

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#### **Details**

Description of parameters.

#### Value

ggplot

#### **Examples**

```
visPlot_3D_plotly(gobject)
```

visSpatDimGenePlot

visSpatDimGenePlot

#### **Description**

integration of visSpatDimGenePlot\_2D(ggplot) and visSpatDimGenePlot\_3D(plotly)

```
visSpatDimGenePlot(
 gobject,
 plot_method = c("ggplot", "plotly"),
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("horizontal", "vertical"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
 genes,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
 label_size = 16,
 genes_low_color = "blue",
 genes_mid_color = "white",
 genes_high_color = "red",
 dim_point_size = 3,
 nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
 network_color = "lightgray",
  spatial_network_alpha = 0.5,
```

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```
show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_grid_alpha = 0.5,
      spatial_point_size = 3,
      spatial_point_border_col = "black",
      spatial_point_border_stroke = 0.1,
      legend_text_size = 12,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
     x_ticks = NULL,
     y_ticks = NULL,
     z_ticks = NULL,
     midpoint = 0,
     point_size = 1,
      cow_n_col = 2,
      cow_rel_h = 1,
     cow_rel_w = 1,
     cow_align = "h",
     show_legend = T,
      show_plots = F
   )
Arguments
   gobject
                    giotto object
   expression_values
                    gene expression values to use
   plot_alignment direction to align plot
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   dim3_to_use
                    dimension to use on z-axis
   sdimx
                    x-axis dimension name (default = 'sdimx')
   sdimy
                    y-axis dimension name (default = 'sdimy')
    sdimz
                    z-axis dimension name (default = 'sdimz')
   genes
                    genes to show
   dim_point_border_col
                    color of border around points
   dim_point_border_stroke
                    stroke size of border around points
   show_NN_network
                    show underlying NN network
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
                    name of NN network to use, if show_NN_network = TRUE
   network_name
```

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edge\_alpha\_dim dim reduction plot: column to use for alpha of the edges scale\_alpha\_with\_expression

scale expression with ggplot alpha parameter

label\_size size for the label

genes\_low\_color

color to represent low expression of gene

genes\_high\_color

color to represent high expression of gene

dim\_point\_size dim reduction plot: point size

spatial\_network\_name

name of spatial network to use

spatial\_grid\_name

name of spatial grid to use

spatial\_point\_size

spatial plot: point size

spatial\_point\_border\_col

color of border around points

spatial\_point\_border\_stroke

stroke size of border around points

legend\_text\_size

the size of the text in legend

axis\_scale three modes to adjust axis scale ratio custom\_ratio set the axis scale ratio on custom

 $x_{ticks}$  number of ticks on x axis  $y_{ticks}$  number of ticks on y axis  $z_{ticks}$  number of ticks on z axis

midpoint size of point (cell)
point\_size size of point (cell)

cow\_n\_col cowplot param: how many columns
cow\_rel\_h cowplot param: relative height
cow\_rel\_w cowplot param: relative width
cow\_align cowplot param: how to align

show\_legend show legend show\_plot show plot

#### **Details**

Description of parameters.

#### Value

ggplot or plotly

# **Examples**

 $\verb|visSpatDimGenePlot(gobject)| \\$ 

visSpatDimGenePlot\_2D visSpatDimGenePlot\_2D

# **Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

# Usage

```
visSpatDimGenePlot_2D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("horizontal", "vertical"),
  genes,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  point_size = 1,
  dim_point_border_col = "black",
  dim_point_border_stroke = 0.1,
  show_NN_network = F,
  show_spatial_network = F,
  show_spatial_grid = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
  spatial_network_name = "spatial_network",
  spatial_grid_name = "spatial_grid",
  spatial_point_size = 1,
  spatial_point_border_col = "black",
  spatial_point_border_stroke = 0.1,
  midpoint = 0,
  genes_high_color = "red",
  genes_mid_color = "white";
  genes_low_color = "blue",
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  show_legend = T,
  show_plots = F
```

#### **Arguments**

gobject giotto object

expression\_values

gene expression values to use

plot\_alignment direction to align plot

genes genes to show

dim\_reduction\_to\_use

dimension reduction to use

dim\_reduction\_name

dimension reduction name

dim1\_to\_use dimension to use on x-axis dim2\_to\_use dimension to use on y-axis

point\_size size of point (cell)

dim\_point\_border\_col

color of border around points

dim\_point\_border\_stroke

stroke size of border around points

show\_NN\_network

show underlying NN network

nn\_network\_to\_use

type of NN network to use (kNN vs sNN)

 $network\_name \qquad name \ of \ NN \ network \ to \ use, \ if \ show\_NN\_network = TRUE$ 

edge\_alpha\_dim dim reduction plot: column to use for alpha of the edges

scale\_alpha\_with\_expression

scale expression with ggplot alpha parameter

spatial\_network\_name

name of spatial network to use

spatial\_grid\_name

name of spatial grid to use

spatial\_point\_size

spatial plot: point size

spatial\_point\_border\_col

color of border around points

 ${\tt spatial\_point\_border\_stroke}$ 

stroke size of border around points

midpoint size of point (cell)

cow\_n\_col cowplot param: how many columns cow\_rel\_h cowplot param: relative height

cow\_rel\_w cowplot param: relative width cow\_align cowplot param: how to align

show\_legend show legend

dim\_point\_size dim reduction plot: point size

show\_plot show plot

#### **Details**

Description of parameters.

#### Value

ggplot

#### **Examples**

```
visSpatDimGenePlot_2D(gobject)
```

```
visSpatDimGenePlot_3D visSpatDimGenePlot_3D
```

# **Description**

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

```
visSpatDimGenePlot_3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("horizontal", "vertical"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  genes,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  label_size = 16,
  genes_low_color = "blue",
  genes_mid_color = "white",
  genes_high_color = "red",
  dim_point_size = 3,
  nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  legend_text_size = 12,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
```

y\_ticks = NULL, z\_ticks = NULL

```
Arguments
   gobject
                     giotto object
    plot_alignment direction to align plot
   dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
                     dimension to use on x-axis
   dim1_to_use
   dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
   network_name
    genes_low_color
                     color represent high gene expression (see details)
    genes_high_color
                     color represent high gene expression (see details)
    nn_network_alpha
                     column to use for alpha of the edges
    show_spatial_network
                     show spatial network
    spatial_network_name
                     name of spatial network to use
    network_color color of spatial/nn network
    spatial_network_alpha
                     alpha of spatial network
    show_spatial_grid
                     show spatial grid
    spatial_grid_name
                     name of spatial grid to use
    spatial_grid_color
                     color of spatial grid
    spatial_grid_alpha
                     alpha of spatial grid
    legend_text_size
                     text size of legend
    show_legend
                     show legend
    show_plot
                     show plot
```

# **Details**

Description of parameters.

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#### Value

plotly

#### **Examples**

```
visSpatDimPlot_3D(gobject)
```

visSpatDimPlot

visSpatDimPlot

# **Description**

integration of visSpatDimPlot\_2D and visSpatDimPlot\_3D

```
visSpatDimPlot(
  gobject,
  plot_method = c("ggplot", "plotly"),
  plot_alignment = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = NULL,
  label_fontface = "bold",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  dim_point_size = 3,
  dim_point_border_col = "black",
  dim_point_border_stroke = 0.1,
  nn_network_alpha = NULL,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
```

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```
show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_grid_alpha = 0.5,
      spatial_point_size = 3,
      legend_text_size = 12,
      spatial_point_border_col = "black",
      spatial_point_border_stroke = 0.1,
      show_legend = T,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      show_plot = F
Arguments
    gobject
                     giotto object
    plot_alignment direction to align plot
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    nn_network_alpha
                     column to use for alpha of the edges
    show\_spatial\_network
                     show spatial network
```

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```
spatial_network_name
                 name of spatial network to use
spatial_network_alpha
                 alpha of spatial network
show_spatial_grid
                 show spatial grid
spatial_grid_name
                 name of spatial grid to use
spatial_grid_color
                 color of spatial grid
spatial_grid_alpha
                 alpha of spatial grid
legend_text_size
                 text size of legend
show_legend
                 show legend
show_plot
                 show plot
plot_mode
                 choose the mode to draw plot: ggplot or plotly
spatial_network_color
                 color of spatial network
```

#### **Details**

Description of parameters.

#### Value

ggplot or plotly

## **Examples**

```
visSpatDimPlot(gobject)
```

visSpatDimPlot\_2D
visSpatDimPlot\_2D

# **Description**

Visualize cells according to spatial AND dimension reduction coordinates in ggplot2 mode

```
visSpatDimPlot_2D(
  gobject,
  plot_alignment = c("vertical", "horizontal"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  sdimx = NULL,
  sdimy = NULL,
```

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```
nn_network_to_use = "sNN",
     network_name = "sNN.pca",
     show_cluster_center = F,
     show_center_label = T,
     center_point_size = 4,
     label_size = 4,
     label_fontface = "bold",
     cell_color = NULL,
     color_as_factor = T,
     cell_color_code = NULL,
     select_cell_groups = NULL,
     select_cells = NULL,
     show_other_cells = T,
     other_cell_color = "lightgrey",
     dim_plot_mode = NULL,
     dim_point_size = 1,
     dim_point_border_col = "black",
     dim_point_border_stroke = 0.1,
     nn_network_alpha = 0.05,
     show_spatial_network = F,
     spatial_network_name = "spatial_network",
      spatial_network_color = NULL,
      show_spatial_grid = F,
     spatial_grid_name = "spatial_grid",
     spatial_grid_color = NULL,
     spatial_point_size = 1,
     spatial_point_border_col = "black",
     spatial_point_border_stroke = 0.1,
     show_legend = T,
     show_plot = F,
     plot_method = "ggplot"
Arguments
                   giotto object
   gobject
   plot_alignment direction to align plot
   dim_reduction_to_use
                   dimension reduction to use
   dim_reduction_name
                   dimension reduction name
```

 $show_NN_network = F,$ 

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```
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
nn_network_alpha
                  column to use for alpha of the edges
show_spatial_network
                  show spatial network
spatial_network_name
                 name of spatial network to use
spatial_network_color
                  color of spatial network
```

show\_spatial\_grid

show spatial grid

spatial\_grid\_name

name of spatial grid to use

spatial\_grid\_color

color of spatial grid

show\_legend show legend
show\_plot show plot

return\_plot return ggplot object

save\_plot directly save the plot [boolean]

save\_dir directory to save the plot

save\_folder (optional) folder in directory to save the plot

save\_name name of plot

 $\texttt{save\_format} \qquad \text{format of plot (e.g. tiff, png, pdf, ...)}$ 

show\_saved\_plot

load & display the saved plot

## **Details**

Description of parameters.

#### Value

ggplot

# **Examples**

```
visSpatDimPlot_2D(gobject)
```

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visSpatDimPlot\_3D
visSpatDimPlot\_3D

# Description

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

# Usage

```
visSpatDimPlot_3D(
  gobject,
  plot_alignment = c("horizontal", "vertical"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 16,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  dim_point_size = 3,
  nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  legend_text_size = 12
```

#### **Arguments**

gobject giotto object

plot\_alignment direction to align plot dim\_reduction\_to\_use dimension reduction to use dim\_reduction\_name dimension reduction name dimension to use on x-axis dim1\_to\_use dimension to use on y-axis dim2\_to\_use dim3\_to\_use dimension to use on z-axis show\_NN\_network show underlying NN network nn\_network\_to\_use type of NN network to use (kNN vs sNN) name of NN network to use, if show\_NN\_network = TRUE network\_name cell\_color color for cells (see details) color\_as\_factor convert color column to factor cell\_color\_code named vector with colors nn\_network\_alpha column to use for alpha of the edges show\_spatial\_network show spatial network spatial\_network\_name name of spatial network to use spatial\_network\_alpha alpha of spatial network show\_spatial\_grid show spatial grid spatial\_grid\_name name of spatial grid to use spatial\_grid\_color color of spatial grid spatial\_grid\_alpha alpha of spatial grid legend\_text\_size text size of legend spatial\_network\_color color of spatial network show legend show\_legend show plot show\_plot

# **Details**

Description of parameters.

## Value

plotly

# Examples

```
visSpatDimPlot_3D(gobject)
```

writeHMRFresults

writeHMRFresults

## **Description**

write results from doHMRF to a data.table.

#### Usage

```
writeHMRFresults(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  print_command = F
)
```

# Arguments

gobject giotto object

HMRF output HMRF output from doHMRF

k k to write results for

betas\_to\_view results from different betas that you want to view

print\_command see the python command

# Value

data.table with HMRF results for each b and the selected k

# **Examples**

```
writeHMRFresults(gobject)
```

# **Description**

write out annotation data from a giotto object for the Viewer

```
write_giotto_viewer_annotation(
  annotation,
  annot_name = "test",
  output_directory = getwd()
)
```

#### **Arguments**

```
annotation annotation from the data.table from giotto object
annot_name name of the annotation
output_directory
directory where to save the files
```

# Value

write a .txt and .annot file for the selection annotation

# **Description**

write out dimensional reduction data from a giotto object for the Viewer

# Usage

```
write_giotto_viewer_dim_reduction(
  dim_reduction_cell,
  dim_red = NULL,
  dim_red_name = NULL,
  dim_red_rounding = NULL,
  dim_red_rescale = c(-20, 20),
  output_directory = getwd()
)
```

# **Arguments**

#### Value

write a .txt and .annot file for the selection annotation

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