Package 'Giotto'

January 14, 2020

```
Title Spatial single-cell transcriptomics pipeline.
Version 0.1.4
Description Pipeline to process, analyze and visualize (spatial) single-cell expression data.
License MIT + file LICENSE
Encoding UTF-8
LazyData true
RoxygenNote 7.0.1
Depends data.table (>= 1.12.2),
      ggplot2 (>= 3.1.1),
      base (>= 3.5.1),
      utils (>= 3.5.1),
      R (>= 3.5.1)
Imports Rtsne (>= 0.15),
      uwot (>= 0.0.0.9010),
      multinet (>= 3.0.2),
      FactoMineR (>= 1.34),
      factoextra (>= 1.0.5),
      cowplot (>= 0.9.4),
      grDevices,
      RColorBrewer (>= 1.1-2),
      jackstraw (>= 1.3),
      dbscan (>= 1.1-3),
      ggalluvial (>= 0.9.1),
      scales (>= 1.0.0),
      ComplexHeatmap (>= 1.20.0),
      qvalue (>= 2.14.1),
      lfa (>= 1.12.0),
      igraph (>= 1.2.4.1),
      plotly,
      reticulate,
      magrittr,
      limma,
      ggdendro,
      smfishHmrf,
      matrixStats (>= 0.55.0),
      IRanges,
      devtools,
      reshape2,
```

2 R topics documented:

```
ggraph

Suggests knitr,
rmarkdown,
MAST,
scran (>= 1.10.1),
png,
tiff,
biomaRt

biocViews

VignetteBuilder knitr

Remotes lambdamoses/smfishhmrf-r
```

R topics documented:

addCellMetadata
addCellStatistics
addGeneMetadata
addGeneStatistics
addHMRF
addNetworkLayout
addStatistics
adjustGiottoMatrix
aes_string2
allCellCellcommunicationsScores
all_plots_save_function
annotateGiotto
annotateSpatialNetwork
annotate_spatlocs_with_spatgrid_2D
annotate_spatlocs_with_spatgrid_3D
average_gene_gene_expression_in_groups
binGetSpatialGenes
calculateHVG
calculateMetaTable
calculateMetaTableCells
calculate_spatial_genes_python
cellProximityBarplot
cellProximityEnrichment
cellProximityHeatmap
cellProximityNetwork
cellProximitySpatPlot
cellProximitySpatPlot2D
cellProximitySpatPlot3D
cellProximityVisPlot
cellProximityVisPlot_2D_ggplot
cellProximityVisPlot_2D_plotly
cellProximityVisPlot_3D_plotly
changeGiottoInstructions
clusterCells
clusterSpatialCorGenes
combineMetadata

convertEnsemblToGeneSymbol
createGiottoInstructions
createGiottoObject
createHeatmap_DT
createMetagenes
createNearestNetwork
createSpatialEnrich
createSpatialGrid
createSpatialGrid_2D
createSpatialGrid_3D
createSpatialNetwork
create_average_detection_DT 58
create_average_DT
create_cell_type_random_cell_IDs
create_cluster_matrix
create_dimObject
decide_cluster_order
detectSpatialCorGenes
detectSpatialPatterns
dimCellPlot
dimCellPlot2D
dimGenePlot
dimGenePlot2D
dimGenePlot3D
dimPlot
dimPlot2D
dimPlot2D_single
dimPlot3D
direction_test_CPG
doHclust
doHMRF
doKmeans
doLeidenCluster
doLeidenSubCluster
doLouvainCluster
doLouvainCluster_community
doLouvainCluster_multinet
doLouvainSubCluster
doLouvainSubCluster_community
doLouvainSubCluster_multinet
doRandomWalkCluster
doSNNCluster
do_spatial_grid_averaging
do_spatial_knn_smoothing
dt to matrix
exportGiottoViewer
exprOnlyCellCellcommunicationScores
extended_gini_fun
extractNearestNetwork
fDataDT
filterCombinations
filterCPGscores 112

filterDistributions	
filterGiotto	
findGiniMarkers	
findGiniMarkers_one_vs_all	
findMarkers	
findMarkers_one_vs_all	
findMastMarkers	121
findMastMarkers_one_vs_all	122
findScranMarkers	123
findScranMarkers_one_vs_all	124
find_grid_2D	125
find_grid_3D	125
find_grid_x	125
find_grid_y	125
find_grid_z	126
fish_function	126
fish_function2	126
FSV_show	127
GenePattern_show	128
general_save_function	129
get 10X matrix	
getCellProximityGeneScores	
getClusterSimilarity	
getDendrogramSplits	
getDistinctColors	
getGeneToGeneScores	
get_cell_to_cell_sorted_name_conversion	
get_interaction_gene_enrichment	
get_specific_interaction_gene_enrichment	
ggplot_save_function	
giotto-class	
heatmSpatialCorGenes	
hyperGeometricEnrich	
kmeans binarize	
loadHMRF	
makeSignMatrixPAGE	
makeSignMatrixRank	
make_simulated_network	
mergeClusters	
mygini_fun	
nnDT_to_kNN	
node clusters	
normalizeGiotto	
OR function2	
PAGEEnrich	
pDataDT	
plotCPGscores	
plotGTGscores	
plotHeatmap	
plotIv_axis_scale_2D	
plotly_axis_scale_2D	
plotly_grid	
prouy_griu	133

plotly_network	
plotMetaDataCellsHeatmap	
plotMetaDataHeatmap	
plotPCA	
plotPCA_2D	
plotPCA_3D	
plotTSNE	. 165
plotTSNE_2D	. 167
plotTSNE_3D	. 170
plotUMAP	. 171
plotUMAP_2D	. 173
plotUMAP_3D	. 175
plot_network_layer_ggplot	. 177
plot_point_layer_ggplot	
plot_point_layer_ggplot_noFILL	
plot_spat_point_layer_ggplot	
plot_spat_point_layer_ggplot_noFILL	
print.giotto	
rankEnrich	
rankEnrichDT	
rankSpatialCorGroups	
rank_binarize	
readGiottoInstructions	
removeCellAnnotation	
removeGeneAnnotation	
replaceGiottoInstructions	
runPCA	
runtSNE	
runUMAP	
selectPatternGenes	
select_expression_values	
show,giotto-method	
showClusterDendrogram	
showClusterHeatmap	
showCPGscores	
showGeneExpressionProximityScore	
showGiottoInstructions	
showGTGscores	
showIntExpressionProximityScore	
showPattern	
showPattern2D	
showPattern3D	
showPatternGenes	
showProcessingSteps	
showSpatialCorGenes	. 208
showTopGeneToGene	. 209
signPCA	. 210
spatCellPlot	
spatCellPlot2D	. 214
spatDimCellPlot	. 217
spatDimCellPlot2D	. 221
spatDimGenePlot	. 225

spatDimGenePlot2D	
spatDimGenePlot3D	
spatDimPlot	
spatDimPlot2D	
spatDimPlot3D	
spatGenePlot	. 243
spatGenePlot2D	. 245
spatGenePlot3D	. 247
Spatial_AEH	. 249
Spatial_DE	. 250
spatNetwDistributions	. 251
spatNetwDistributionsDistance	. 252
spatNetwDistributionsKneighbors	. 253
spatPlot	
spatPlot2D	. 257
spatPlot2D_single	
spatPlot3D	
specificCellCellcommunicationScores	
split_dendrogram_in_two	
stitchFieldCoordinates	
subClusterCells	
subsetGiotto	
subsetGiottoLocs	
trendSceek	
viewHMRFresults	
viewHMRFresults2D	
viewHMRFresults3D	
violinPlot	
visDimGenePlot	
visDimGenePlot_2D_ggplot	
visDimGenePlot_3D_plotly	
visDimPlot	
visDimPlot_2D_ggplot	
visDimPlot_2D_plotly	
visDimPlot_3D_plotly	
visForceLayoutPlot	
visGenePlot	
visGenePlot_2D_ggplot	
visGenePlot_3D_plotly	
visPlot	
visPlot_2D_ggplot	
visPlot_2D_ggptot visPlot_2D_plotly visPlotly visPlot_2D_plotly visPlot_2D_plotly visPlot_2D_plotly visPlotly visPlot_2D_plotly visPlot_2D_plotly visPlot_2D_plotly visPlotly visPlot_2D_plotly visPlot_2D_plotly visPlotly visPlot_2D_plotly visPlotly visPlotl	
visPlot_3D_plotly	
e · · · · · · · · · · · · · · · · · · ·	
visSpatDimGenePlot	
visSpatDimGenePlot_3D	
visSpatDimPlot	
visSpatDimPlot_2D	
visSpatDimPlot_3D	
writeHMRFresults	
write_giotto_viewer_annotation	
write_giotto_viewer_dim_reduction	. 218

addCellMetadata 7

Index 320

addCellMetadata addCellMetadata

Description

adds cell metadata to the giotto object

Usage

```
addCellMetadata(
  gobject,
  new_metadata,
  by_column = FALSE,
  column_cell_ID = NULL
)
```

Arguments

gobject giotto object

new_metadata new cell metadata to use (data.table, data.frame, ...)

by_column merge metadata based on cell_ID column in pDataDT (default = FALSE)

column_cell_ID column name of new metadata to use if by_column = TRUE

Details

You can add additional cell metadata in two manners: 1. Provide a data.table or data.frame with cell annotations in the same order as the cell_ID column in pDataDT(gobject) 2. Provide a data.table or data.frame with cell annotations and specificy which column contains the cell IDs, these cell IDs need to match with the cell_ID column in pDataDT(gobject)

Value

giotto object

Examples

```
addCellMetadata(gobject)
```

8 addCellStatistics

 ${\tt addCellStatistics}$

addCellStatistics

Description

adds cells statistics to the giotto object

Usage

```
addCellStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

Arguments

Details

This function will add the following statistics to cell metadata:

- nr_genes: Denotes in how many genes are detected per cell
- perc_genes: Denotes what percentage of genes is detected per cell
- total_expr: Shows the total sum of gene expression per cell

Value

```
giotto object if return_gobject = TRUE
```

Examples

```
addCellStatistics(gobject)
```

addGeneMetadata 9

addGeneMetadata

addGeneMetadata

Description

adds gene metadata to the giotto object

Usage

```
addGeneMetadata(gobject, new_metadata, by_column = F, column_gene_ID = NULL)
```

Arguments

```
gobject giotto object
```

new_metadata new metadata to use

by_column merge metadata based on gene_ID column in fDataDT column_cell_ID column name of new metadata to use if by_column = TRUE

Details

You can add additional gene metadata in two manners: 1. Provide a data.table or data.frame with gene annotations in the same order as the gene_ID column in fDataDT(gobject) 2. Provide a data.table or data.frame with gene annotations and specificy which column contains the gene IDs, these gene IDs need to match with the gene_ID column in fDataDT(gobject)

Value

giotto object

Examples

addGeneMetadata(gobject)

addGeneStatistics

addGeneStatistics

Description

adds gene statistics to the giotto object

```
addGeneStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

10 addHMRF

Arguments

Details

This function will add the following statistics to gene metadata:

- nr_cells: Denotes in how many cells the gene is detected
- per_cells: Denotes in what percentage of cells the gene is detected
- total_expr: Shows the total sum of gene expression in all cells
- mean_expr: Average gene expression in all cells
- mean_expr_det: Average gene expression in cells with detectable levels of the gene

Value

```
giotto object if return_gobject = TRUE
```

Examples

```
addGeneStatistics(gobject)
```

addHMRF

Description

Add selected results from doHMRF to the giotto object

addHMRF

Usage

```
addHMRF(gobject, HMRFoutput, k = NULL, betas_to_add = NULL, hmrf_name = NULL)
```

Arguments

gobject giotto object

HMRF output from doHMRF()

k number of domains

betas_to_add results from different betas that you want to add

name specify a custom name

Details

Description ...

addNetworkLayout 11

Value

```
giotto object
```

Examples

```
addHMRF(gobject)
```

addNetworkLayout

addNetworkLayout

Description

Add a network layout for a selected nearest neighbor network

Usage

```
addNetworkLayout(
  gobject,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  layout_type = c("drl"),
  options_list = NULL,
  layout_name = "layout",
  return_gobject = TRUE
)
```

Arguments

Details

This function creates layout coordinates based on the provided kNN or sNN. Currently only the force-directed graph layout "drl", see layout_with_drl, is implemented. This provides an alternative to tSNE or UMAP based visualizations.

Value

giotto object with updated layout for selected NN network

Examples

```
addNetworkLayout(gobject)
```

12 adjustGiottoMatrix

addStatistics

addStatistics

Description

adds genes and cells statistics to the giotto object

Usage

```
addStatistics(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0,
  return_gobject = TRUE
)
```

Arguments

Details

See addGeneStatistics and addCellStatistics

Value

```
giotto object if return_gobject = TRUE, else a list with results
```

Examples

```
addStatistics(gobject)
```

adjustGiottoMatrix adjustGiottoMatrix

Description

normalize and/or scale expresion values of Giotto object

aes_string2

Usage

```
adjustGiottoMatrix(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  batch_columns = NULL,
  covariate_columns = NULL,
  return_gobject = TRUE,
  update_slot = c("custom")
)
```

Arguments

```
gobject giotto object

expression_values

expression values to use

batch_columns metadata columns that represent different batch (max = 2)

covariate_columns

metadata columns that represent covariates to regress out

return_gobject boolean: return giotto object (default = TRUE)

update_slot expression slot that will be updated (default = custom)
```

Details

This function implements the limma::removeBatchEffect function to remove known batch effects and to adjust expression values according to provided covariates.

Value

giotto object

Examples

```
adjustGiottoMatrix(gobject)
```

```
aes_string2 aes_string2
```

Description

makes sure aes_string can also be used with names that start with numeric values

```
aes_string2(...)
```

```
all {\tt Cell Cell communications Scores} \\ all {\tt Cell Cell communications Scores}
```

Description

All Cell-Cell communication scores based on spatial expression of interacting cells

Usage

```
allCellCellcommunicationsScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "cell_types",
  random_iter = 100,
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  min_observations = 2,
  verbose = c("a little", "a lot", "none")
)
```

Arguments

```
gobject
                  giotto object to use
spatial_network_name
                  spatial network to use for identifying interacting cells
cluster_column cluster column with cell type information
random\_iter
                  number of iterations
                  first specific gene set from gene pairs
gene_set_1
                  second specific gene set from gene pairs
gene_set_2
log2FC_addendum
                  addendum to add when calculating log2FC
min_observations
                  minimum number of interactions needed to be considered
                  verbose
verbose
```

Details

Statistical framework to identify if pairs of genes (such as ligand-receptor combinations) are expressed at higher levels than expected based on a reshuffled null distribution of gene expression values in cells that are spatially in proximity to eachother. More details will follow soon.

Value

Cell-Cell communication scores for gene pairs based on spatial interaction

Examples

```
allCellCellcommunicationsScores(gobject)
```

```
all_plots_save_function 
 all_plots_save_function
```

Description

Function to automatically save plots to directory of interest

Usage

```
all_plots_save_function(
  gobject,
  plot_object,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  default_save_name = "giotto_plot",
  save_format = NULL,
  show_saved_plot = F,
  ncol = 1,
  nrow = 1,
  scale = 1,
  base_width = NULL,
  base_height = NULL,
  base_aspect_ratio = NULL,
  units = NULL,
  dpi = NULL,
  limitsize = TRUE,
)
```

```
gobject
                  giotto object
                  object to plot
plot_object
save_dir
                  directory to save to
save_folder
                  folder in save_dir to save to
save_name
                  name of plot
save_format
                  format (e.g. png, tiff, pdf, ...)
show_saved_plot
                  load & display the saved plot
ncol
                  number of columns
                  number of rows
nrow
scale
                  scale
base_width
                  width
base_height
                  height
{\tt base\_aspect\_ratio}
                  aspect ratio
```

16 annotateGiotto

```
units units

dpi Plot resolution

limitsize When TRUE (the default), ggsave will not save images larger than 50x50 inches, to prevent the common error of specifying dimensions in pixels.
```

additional parameters to ggplot_save_function or general_save_function

See Also

```
general_save_function
```

Examples

```
all_plots_save_function(gobject)
```

annotateGiotto

annotateGiotto

Description

Converts cluster results into provided annotation.

Usage

```
annotateGiotto(
  gobject,
  annotation_vector = NULL,
  cluster_column = NULL,
  name = "cell_types"
)
```

Arguments

Details

You need to specifify which (cluster) column you want to annotate and you need to provide an annotation vector like this:

- 1. identify the cell type of each cluster
- 2. create a vector of these cell types, e.g. cell_types = c('T-cell', 'B-cell', 'Stromal')
- 3. provide original cluster names to previous vector, e.g. names(cell_types) = c(2, 1, 3)

Value

giotto object

Examples

```
annotateGiotto(gobject)
```

 $annotate {\tt Spatial Network}$

annotateSpatialNetwork

Description

Annotate spatial network with cell metadata information.

Usage

```
annotateSpatialNetwork(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column
)
```

Arguments

Value

annotated network in data.table format

Examples

```
annotateSpatialNetwork(gobject)
```

```
annotate\_spatlocs\_with\_spatgrid\_2D \\ annotate\_spatlocs\_with\_spatgrid\_2D
```

Description

annotate spatial locations with 2D spatial grid information

Usage

```
annotate_spatlocs_with_spatgrid_2D(spatloc, spatgrid)
```

```
spatloc spatial_locs slot from giotto object spatgrid selected spatial_grid slot from giotto object
```

Value

annotated spatial location data.table

Examples

```
annotate_spatlocs_with_spatgrid_2D()
```

```
annotate\_spatlocs\_with\_spatgrid\_3D \\ annotate\_spatlocs\_with\_spatgrid\_3D
```

Description

annotate spatial locations with 3D spatial grid information

Usage

```
annotate_spatlocs_with_spatgrid_3D(spatloc, spatgrid)
```

Arguments

```
spatloc spatial_locs slot from giotto object spatgrid selected spatial_grid slot from giotto object
```

Value

annotated spatial location data.table

Examples

```
annotate_spatlocs_with_spatgrid_3D()
```

```
average_gene_gene_expression_in_groups

average_gene_gene_expression_in_groups
```

Description

calculate average expression per cluster

```
average_gene_gene_expression_in_groups(
  gobject,
  cluster_column = "cell_types",
  gene_set_1,
  gene_set_2
)
```

binGetSpatialGenes 19

Arguments

```
gobject giotto object to use
cluster_column cluster column with cell type information
gene_set_1 first specific gene set from gene pairs
gene_set_2 second specific gene set from gene pairs
```

Details

Details will follow soon.

Value

data.table with average expression scores for each cluster

Examples

```
average_gene_gene_expression_in_groups(gobject)
```

binGetSpatialGenes

binGetSpatialGenes

Description

Rapid computation of genes that are spatially clustered

Usage

```
binGetSpatialGenes(
  gobject,
  bin_method = c("kmeans", "rank"),
  expression_values = c("normalized", "scaled", "custom"),
  subset_genes = NULL,
  spatial_network_name = "spatial_network",
  nstart = 3,
  iter_max = 10,
  percentage_rank = 10,
  do_fisher_test = F,
  community_expectation = 5,
  verbose = F
)
```

20 calculateHVG

nstart kmeans: nstart parameter iter_max kmeans: iter.max parameter

do_fisher_test perform fisher test

community_expectation

cell degree expectation in spatial communities

verbose be verbose

rank_percentage

percentage of top cells for binarization

Details

We provide two ways to identify spatial genes based on gene expression binarization. Both methods are identicial except for how binarization is performed.

- 1. binarize: Each gene is binarized (0 or 1) in each cell with **kmeans** (k = 2) or based on **rank** percentile
- 2. network: Alll cells are connected through a k-nearest neighbor network
- 3. contingency table: A contingency table is calculated based on all pairwise cell-cell interactions (0-0, 0-1, 1-0 or 1-1)
- 4. For each gene an odds-ratio (OR) and fisher.test (optional) is calculated

Additionally 2 other statistics are provided:

- Number of cells with high expression (binary = 1)
- total and ratio of highly connected cells: Cells with a connectivity higher than community_expectation

By selecting a subset of likely spatial genes (e.g. highly variable genes) the function will be much faster.

Value

data.table with results (see details)

Examples

binGetSpatialGenes(gobject)

calculateHVG calculateHVG

Description

compute highly variable genes

calculateHVG 21

Usage

```
calculateHVG(
      gobject,
      expression_values = c("normalized", "scaled", "custom"),
      method = c("cov_groups", "cov_loess"),
      reverse_log_scale = FALSE,
      logbase = 2,
      expression_threshold = 0,
      nr_expression_groups = 20,
      zscore_threshold = 1.5,
      HVGname = "hvg",
      difference_in_cov = 0.1,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "HVGplot",
      return_gobject = TRUE
    )
Arguments
    gobject
                     giotto object
    expression_values
                     expression values to use
                     method to calculate highly variable genes
    method
    reverse_log_scale
                     reverse log-scale of expression values (default = FALSE)
    logbase
                     if reverse_log_scale is TRUE, which log base was used?
    expression_threshold
                     expression threshold to consider a gene detected
    nr_expression_groups
                     number of expression groups for cov_groups
    zscore_threshold
                     zscore to select hvg for cov_groups
    HVGname
                     name for highly variable genes in cell metadata
    difference_in_cov
                     minimum difference in coefficient of variance required
    show_plot
                     show plot
                     return ggplot object
    return_plot
                     directly save the plot [boolean]
    save_plot
                     list of saving parameters from all_plots_save_function
    save_param
    default_save_name
                     default save name for saving, don't change, change save_name in save_param
```

return_gobject boolean: return giotto object (default = TRUE)

22 calculateMetaTable

Details

Currently we provide 2 ways to calculate highly variable genes: 1. high coeff of variance (COV) within groups:

First genes are binned (*nr_expression_groups*) into average expression groups and the COV for each gene is converted into a z-score within each bin. Genes with a z-score higher than the threshold (*zscore_threshold*) are considered highly variable.

2. high COV based on loess regression prediction:

A predicted COV is calculated for each gene using loess regression (COV~log(mean expression)) Genes that show a higher than predicted COV (*difference_in_cov*) are considered highly variable.

Value

giotto object highly variable genes appended to gene metadata (fDataDT)

Examples

```
calculateHVG(gobject)
```

calculateMetaTable

calculateMetaTable

Description

calculates the average gene expression for one or more (combined) annotation columns.

Usage

```
calculateMetaTable(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metadata_cols = NULL,
  selected_genes = NULL
)
```

Arguments

```
gobject giotto object
expression_values
expression values to use
metadata_cols annotation columns found in pDataDT(gobject)
selected_genes subset of genes to use
```

Value

data.table with average expression values for each gene per (combined) annotation

Examples

```
calculateMetaTable(gobject)
```

calculateMetaTableCells 23

```
calculateMetaTableCells
```

calculateMetaTableCells

Description

calculates the average metadata values for one or more (combined) annotation columns.

Usage

```
calculateMetaTableCells(
  gobject,
  value_cols = NULL,
  metadata_cols = NULL,
  spat_enr_names = NULL
)
```

Arguments

```
gobject giotto object
value_cols metadata or enrichment value columns to use
metadata_cols annotation columns found in pDataDT(gobject)
spat_enr_names which spatial enrichment results to include
```

Value

data.table with average metadata values per (combined) annotation

Examples

```
calculateMetaTableCells(gobject)
```

```
calculate\_spatial\_genes\_python \\ calculate\_spatial\_genes\_python
```

Description

Calculate spatial genes using distance matrix.

```
calculate_spatial_genes_python(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metric = "euclidean",
  subset_genes = NULL,
  rbp_p = 0.95,
  examine_top = 0.3,
  python_path = NULL
)
```

Arguments

```
gobject giotto object
expression_values
expression values to use

metric distance metric to use
subset_genes only run on this subset of genes
rbp_p fractional binarization threshold
examine_top top fraction to evaluate with silhouette
python_path specify specific path to python if required
```

Details

Description of how we compute spatial pattern genes.

Value

data.table with spatial scores

Examples

```
calculate_spatial_genes_python(gobject)
```

```
cellProximityBarplot cellProximityBarplot
```

Description

Create barplot from cell-cell proximity scores

```
cellProximityBarplot(
  gobject,
  CPscore,
  min_orig_ints = 5,
  min_sim_ints = 5,
  p_val = 0.05,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityBarplot"
)
```

Arguments

```
gobject
                  giotto object
                  CPscore, output from cellProximityEnrichment()
CPscore
                  filter on minimum original cell-cell interactions
min_orig_ints
min_sim_ints
                  filter on minimum simulated cell-cell interactions
p_val
                  p-value
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function
```

default_save_name

default save name for saving, don't change, change save_name in save_param

Details

This function creates a barplot that shows the spatial proximity enrichment or depletion of cell type pairs.

Value

ggplot barplot

Examples

```
cellProximityBarplot(CPscore)
```

```
{\tt cellProximityEnrichment}
```

cellProximityEnrichment

Description

Compute cell-cell interaction enrichment (observed vs expected)

Usage

```
cellProximityEnrichment(
 gobject,
  spatial_network_name = "spatial_network",
 cluster_column,
  number_of_simulations = 100
)
```

```
gobject
                 giotto object
spatial_network_name
                 name of spatial network to use
cluster_column name of column to use for clusters
number_of_simulations
                 number of simulations to create expected observations
```

Details

Spatial proximity enrichment or depletion between pairs of cell types is calculated by calculating the observed over the expected frequency of cell-cell proximity interactions. The expected frequency is the average frequency calculated from a number of spatial network simulations. Each individual simulation is obtained by reshuffling the cell type labels of each node (cell) in the spatial network.

Value

List of cell Proximity scores (CPscores) in data.table format. The first data.table (raw_sim_table) shows the raw observations of both the original and simulated networks. The second data.table (enrichm_res) shows the enrichment results.

Examples

```
cellProximityEnrichment(gobject)
```

```
cellProximityHeatmap cellProximityHeatmap
```

Description

Create heatmap from cell-cell proximity scores

Usage

```
cellProximityHeatmap(
  gobject,
  CPscore,
  scale = T,
  order_cell_types = T,
  color_breaks = NULL,
  color_names = NULL,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityHeatmap")
```

```
gobject giotto object

CPscore CPscore, output from cellProximityEnrichment()

scale scale cell-cell proximity interaction scores

order_cell_types

order cell types based on enrichment correlation

color_breaks numerical vector of length 3 to represent min, mean and maximum

color_names character color vector of length 3

show_plot show plot
```

cellProximityNetwork 27

Details

This function creates a heatmap that shows the spatial proximity enrichment or depletion of cell type pairs.

Value

ggplot heatmap

Examples

```
cellProximityHeatmap(CPscore)
```

cellProximityNetwork cellProximityNetwork

Description

Create network from cell-cell proximity scores

```
cellProximityNetwork(
  gobject,
  CPscore,
  remove_self_edges = FALSE,
  self_loop_strength = 0.1,
  color_depletion = "lightgreen",
  color_enrichment = "red",
  rescale_edge_weights = TRUE,
  edge_weight_range_depletion = c(0.1, 1),
  edge_weight_range_enrichment = c(1, 5),
  layout = c("Fruchterman", "DrL", "Kamada-Kawai"),
  only_show_enrichment_edges = F,
  edge_width_range = c(0.1, 2),
  node_size = 4,
  node_text_size = 6,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "cellProximityNetwork"
)
```

Arguments

gobject giotto object **CPscore** CPscore, output from cellProximityEnrichment() remove_self_edges remove enrichment/depletion edges with itself $self_loop_strength$ size of self-loops color_depletion color for depleted cell-cell interactions color_enrichment color for enriched cell-cell interactions rescale_edge_weights rescale edge weights (boolean) ${\tt edge_weight_range_depletion}$ numerical vector of length 2 to rescale depleted edge weights edge_weight_range_enrichment numerical vector of length 2 to rescale enriched edge weights layout layout algorithm to use to draw nodes and edges only_show_enrichment_edges show only the enriched pairwise scores edge_width_range range of edge width size of nodes node_size node_text_size size of node labels show_plot show plot return_plot return ggplot object save_plot directly save the plot [boolean] save_param list of saving parameters from all_plots_save_function

Details

This function creates a network that shows the spatial proximity enrichment or depletion of cell type pairs.

default save name for saving, don't change, change save_name in save_param

Value

igraph plot

default_save_name

Examples

 ${\tt cellProximityNetwork(CPscore)}$

cellProximitySpatPlot 29

cellProximitySpatPlot cellProximitySpatPlot

Description

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

Usage

```
cellProximitySpatPlot(gobject, ...)
```

```
gobject
                  giotto object
interaction_name
                  cell-cell interaction name
cluster_column cluster column with cell clusters
sdimx
                  x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimy
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
point_size_select
                  size of selected points
point_select_border_col
                  border color of selected points
point_select_border_stroke
                  stroke size of selected points
point_size_other
                  size of other points
```

Details

Description of parameters.

Value

ggplot

See Also

cellProximitySpatPlot2D and cellProximitySpatPlot3D for 3D

Examples

```
cellProximitySpatPlot(gobject)
```

```
cellProximitySpatPlot2D
```

cell Proximity Spat Plot 2D

Description

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

```
cellProximitySpatPlot2D(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
```

```
spatial_network_name = "spatial_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 1,
  show_legend = T,
 point_size_select = 2,
  point_select_border_col = "black",
  point_select_border_stroke = 0.05,
 point_size_other = 1,
 point_alpha_other = 0.3,
  point_other_border_col = "lightgrey",
 point_other_border_stroke = 0.01,
  show_plot = NA,
 return_plot = NA,
  save_plot = NA,
  save_param = list(),
 default_save_name = "cellProximitySpatPlot2D"
)
```

```
giotto object
gobject
interaction_name
                  cell-cell interaction name
cluster_column cluster column with cell clusters
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
                  color for cells (see details)
cell_color
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
point_size_select
                  size of selected points
```

```
point_select_border_col
                 border color of selected points
point_select_border_stroke
                 stroke size of selected points
point_size_other
                 size of other points
point_other_border_col
                 border color of other points
point_other_border_stroke
                 stroke size of other points
show_plot
                 show plots
return_plot
                 return ggplot object
                 directly save the plot [boolean]
save_plot
save_param
                 list of saving parameters from all_plots_save_function
default_save_name
                 default save name for saving, don't change, change save_name in save_param
```

Details

Description of parameters.

Value

ggplot

Examples

```
cellProximitySpatPlot2D(gobject)
```

```
cell Proximity Spat Plot 3D \\ cell Proximity Spat Plot 2D
```

Description

Visualize 3D cell-cell interactions according to spatial coordinates in plotly mode

```
cellProximitySpatPlot3D(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = T,
```

```
show_network = T,
      show_other_network = F,
     network_color = NULL,
      spatial_network_name = "spatial_network",
      show_grid = F,
     grid_color = NULL,
      spatial_grid_name = "spatial_grid",
      show_legend = T,
      point_size_select = 4,
     point_size_other = 2,
     point_alpha_other = 0.5,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
     x_ticks = NULL,
     y_ticks = NULL,
      z_ticks = NULL,
      show_plot = NA,
     return_plot = NA,
      save_plot = NA,
     save_param = list(),
     default_save_name = "cellProximitySpatPlot3D",
   )
Arguments
   gobject
                    giotto object
   interaction_name
                    cell-cell interaction name
   cluster_column cluster column with cell clusters
   sdimx
```

```
x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimy
                  z-axis dimension name (default = 'sdimz')
sdimz
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
                  color of spatial grid
grid_color
spatial_grid_name
                  name of spatial grid to use
show_legend
                  show legend
```

cellProximityVisPlot

```
point_size_select
size of selected points

point_size_other
size of other points

show_plot show plots

return_plot return plotly object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function

default_save_name
default save name for saving, don't change, change save_name in save_param
```

Details

Description of parameters.

Value

plotly

Examples

```
cellProximitySpatPlot3D(gobject)
```

```
cellProximityVisPlot cellProximityVisPlot
```

Description

Visualize cell-cell interactions according to spatial coordinates

```
cellProximityVisPlot(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
```

cellProximityVisPlot 35

```
coord_fix_ratio = 1,
      show_legend = T,
      point_size_select = 2,
      point_select_border_col = "black",
      point_select_border_stroke = 0.05,
      point_size_other = 1,
      point_alpha_other = 0.3,
      point_other_border_col = "lightgrey",
      point_other_border_stroke = 0.01,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      plot_method = c("ggplot", "plotly"),
    )
Arguments
   gobject
                     giotto object
    interaction_name
                     cell-cell interaction name
    cluster_column cluster column with cell clusters
                     x-axis dimension name (default = 'sdimx')
    sdimx
                     y-axis dimension name (default = 'sdimy')
    sdimy
    sdimz
                     z-axis dimension name (default = 'sdimz')
    cell_color
                     color for cells (see details)
    cell_color_code
                     named vector with colors
    color_as_factor
                     convert color column to factor
                     show underlying spatial network
    show_network
    network_color
                     color of spatial network
    spatial_network_name
                     name of spatial network to use
    show_grid
                     show spatial grid
    grid_color
                     color of spatial grid
    spatial_grid_name
                     name of spatial grid to use
    coord_fix_ratio
                     fix ratio between x and y-axis
    show\_legend
                     show legend
   point_size_select
                     size of selected points
   {\tt point\_select\_border\_col}
```

border color of selected points

Details

Description of parameters.

Value

ggplot or plotly

Examples

```
cellProximityVisPlot(gobject)
```

```
cell Proximity VisPlot\_2D\_ggplot \\ cell Proximity VisPlot\_2D\_ggplot
```

Description

Visualize 2D cell-cell interactions according to spatial coordinates in ggplot mode

```
cellProximityVisPlot_2D_ggplot(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 1,
  show_legend = T,
  point_size_select = 2,
```

```
point_select_border_col = "black",
 point_select_border_stroke = 0.05,
 point_size_other = 1,
 point_alpha_other = 0.3,
 point_other_border_col = "lightgrey",
 point_other_border_stroke = 0.01,
)
```

Arguments

```
gobject
                  giotto object
interaction_name
                  cell-cell interaction name
cluster_column cluster column with cell clusters
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
point_size_select
                  size of selected points
point_select_border_col
                  border color of selected points
point_select_border_stroke
                  stroke size of selected points
point_size_other
                  size of other points
point_other_border_col
                  border color of other points
\verb"point_other_border_stroke"
                  stroke size of other points
```

Details

Description of parameters.

Value

ggplot

Examples

```
cellProximityVisPlot_2D_ggplot(gobject)
```

```
cell Proximity VisPlot\_2D\_plotly \\ cell Proximity VisPlot\_2D\_plotly
```

Description

Visualize 2D cell-cell interactions according to spatial coordinates in plotly mode

```
cellProximityVisPlot_2D_plotly(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  point_size_select = 2,
  point_size_other = 1,
  point_alpha_other = 0.3,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
 y_ticks = NULL,
)
```

Arguments

gobject giotto object

interaction_name

cell-cell interaction name

cluster_column cluster column with cell clusters

sdimx x-axis dimension name (default = 'sdimx')

sdimy y-axis dimension name (default = 'sdimy')

cell_color color for cells (see details)

cell_color_code

named vector with colors

color_as_factor

convert color column to factor

show_other_cells

decide if show cells not in network

show_network show underlying spatial network

network_color color of spatial network

spatial_network_name

name of spatial network to use

show_grid show spatial grid

grid_color color of spatial grid

spatial_grid_name

name of spatial grid to use

show_legend show legend

point_size_select

size of selected points

coord_fix_ratio

fix ratio between x and y-axis

Details

Description of parameters.

Value

plotly

Examples

cellProximityVisPlot_2D_plotly(gobject)

```
cell Proximity VisPlot\_3D\_plotly \\ cell Proximity VisPlot\_3D\_plotly
```

Description

Visualize 3D cell-cell interactions according to spatial coordinates in plotly mode

Usage

```
cellProximityVisPlot_3D_plotly(
  gobject,
  interaction_name = NULL,
  cluster_column = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  show_other_cells = F,
  show_network = F,
  show_other_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  point_size_select = 2,
  point_size_other = 1,
  point_alpha_other = 0.5,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
)
```

Arguments

```
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
show_other_cells
                  decide if show cells not in network
show_network
                  show underlying spatial network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
                  show legend
show_legend
point_size_select
                  size of selected points
coord_fix_ratio
                  fix ratio between x and y-axis
```

Details

Description of parameters.

Value

plotly

Examples

```
cellProximityVisPlot_3D_plotly(gobject)
```

 ${\tt change} {\tt GiottoInstructions}$

change Giot to Instructions

Description

Function to change one or more instructions from giotto object

```
changeGiottoInstructions(
  gobject,
  params = NULL,
  new_values = NULL,
  return_gobject = TRUE
)
```

42 clusterCells

Arguments

```
gobject giotto object
params parameter(s) to change
new_values new value(s) for parameter(s)
return_gobject (boolean) return giotto object
```

Value

named vector with giotto instructions

Examples

changeGiottoInstructions()

clusterCells

clusterCells

Description

cluster cells using a variety of different methods

```
clusterCells(
  gobject,
 cluster_method = c("leiden", "louvain_community", "louvain_multinet", "randomwalk",
    "sNNclust", "kmeans", "hierarchical"),
  name = "cluster_name",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  pyth_leid_resolution = 1,
  pyth_leid_weight_col = "weight",
 pyth_leid_part_type = c("RBConfigurationVertexPartition", "ModularityVertexPartition"),
  pyth_leid_init_memb = NULL,
  pyth_leid_iterations = 1000,
  pyth_louv_resolution = 1,
  pyth_louv_weight_col = NULL,
  python_louv_random = F,
  python_path = NULL,
  louvain_gamma = 1,
  louvain_omega = 1,
  walk\_steps = 4,
  walk_clusters = 10,
  walk_weights = NA,
  sNNclust_k = 20,
  sNNclust_eps = 4,
  sNNclust_minPts = 16,
  borderPoints = TRUE,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
```

clusterCells 43

```
dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
      dim_reduction_name = "pca",
      dimensions_to_use = 1:10,
      distance_method = c("original", "pearson", "spearman", "euclidean", "maximum",
        "manhattan", "canberra", "binary", "minkowski"),
      km_centers = 10,
      km_iter_max = 100,
      km_nstart = 1000,
      km_algorithm = "Hartigan-Wong",
     hc_agglomeration_method = c("ward.D2", "ward.D", "single", "complete", "average",
        "mcquitty", "median", "centroid"),
      hc_k = 10,
      hc_h = NULL
      return_gobject = TRUE,
      set_seed = T,
      seed_number = 1234
Arguments
   gobject
                    giotto object
    cluster_method community cluster method to use
                    name for new clustering result
    nn_network_to_use
                    type of NN network to use (kNN vs sNN)
    network_name
                    name of NN network to use
   pyth_leid_resolution
                    resolution for leiden
    pyth_leid_weight_col
                    column to use for weights
   pyth_leid_part_type
                    partition type to use
   pyth_leid_init_memb
                    initial membership
    pyth_leid_iterations
                    number of iterations
    pyth_louv_resolution
                    resolution for louvain
   pyth_louv_weight_col
                    python louvain param: weight column
    python_louv_random
                    python louvain param: random
    python_path
                    specify specific path to python if required
    louvain_gamma
                    louvain param: gamma or resolution
                    louvain param: omega
    louvain_omega
                    randomwalk: number of steps
    walk_steps
                    randomwalk: number of clusters
   walk_clusters
                    randomwalk: weight column
    walk_weights
    sNNclust_k
                    SNNclust: k neighbors to use
```

44 clusterCells

sNNclust_eps SNNclust: epsilon

sNNclust_minPts

SNNclust: min points

borderPoints SNNclust: border points

expression_values

expression values to use

genes_to_use = NULL,
dim_reduction_to_use

dimension reduction to use

dim_reduction_name

name of reduction 'pca',

dimensions_to_use

dimensions to use

distance_method

distance method

km_centers kmeans centers km_iter_max kmeans iterations

km_nstart kmeans random starting points

 ${\tt km_algorithm} \quad \ \, {\tt kmeans} \; {\tt algorithm} \\$

hc_agglomeration_method

hierarchical clustering method

hc_k hierachical number of clusters

hc_h hierarchical tree cutoff

 $\verb|return_gobject|| boolean: return giotto object (default = TRUE)$

set_seed set seed

seed_number number for seed

Details

Wrapper for the different clustering methods.

Value

giotto object with new clusters appended to cell metadata

See Also

 $\label{lem:cluster_doLouvainCluster_multinet} do Louvain Cluster_community, do Louvain Cluster_multinet, do Louvain Cluster, do Random Walk Cluster, do SNN Cluster, do Kmeans, do H clust Cluster, do Louvain Cluster, do Louva$

Examples

clusterCells(gobject)

clusterSpatialCorGenes

```
clusterSpatialCorGenes
```

clusterSpatialCorGenes

Description

Cluster based on spatially correlated genes

Usage

```
clusterSpatialCorGenes(
  spatCorObject,
  name = "spat_clus",
  hclust_method = "ward.D",
  k = 10,
  return_obj = TRUE
)
```

Arguments

```
spatCorObject spatial correlation object
```

name name for spatial clustering results
hclust_method method for hierarchical clustering
k number of clusters to extract

return_obj return spatial correlation object (spatCorObject)

Value

spatCorObject or cluster results

Examples

```
clusterSpatialCorGenes(gobject)
```

combineMetadata

combineMetadata

Description

This function combines the cell metadata with spatial locations and enrichment results from createSpatialEnrich

Usage

```
combineMetadata(gobject, spat_enr_names = NULL)
```

Arguments

```
gobject Giotto object
spat_enr_names names of spatial enrichment results to include
```

46 createGiottoInstructions

Value

Extended cell metadata in data.table format.

Examples

```
combineMetadata(gobject)
```

convertEnsemblToGeneSymbol

convert Ensembl To Gene Symbol

Description

This function convert ensembl gene IDs from a matrix to official gene symbols

Usage

```
convertEnsemblToGeneSymbol(matrix, species = c("mouse", "human"))
```

Arguments

matrix an expression matrix with ensembl gene IDs as rownames

species species to use for gene symbol conversion

Details

This function requires that the biomaRt library is installed

Value

expression matrix with gene symbols as rownames

Examples

```
convertEnsemblToGeneSymbol(matrix)
```

create Giotto Instructions

createGiottoInstructions

Description

Function to set global instructions for giotto functions

createGiottoObject 47

Usage

```
createGiottoInstructions(
   python_path = NULL,
   show_plot = NULL,
   return_plot = NULL,
   save_plot = NULL,
   save_dir = NULL,
   plot_format = NULL,
   dpi = NULL,
   units = NULL,
   height = NULL,
   width = NULL
```

Arguments

path to python binary to use python_path show_plot print plot to console, default = TRUE return plot as object, default = TRUE return_plot save_plot automatically save plot, dafault = FALSE path to directory where to save plots save_dir dpi resolution for raster images height height of plots width width of plots

Value

named vector with giotto instructions

Examples

```
createGiottoInstructions()
```

Description

Function to create a giotto object

```
createGiottoObject(
  raw_exprs,
  spatial_locs = NULL,
  norm_expr = NULL,
  norm_scaled_expr = NULL,
  custom_expr = NULL,
  cell_metadata = NULL,
```

48 createGiottoObject

```
gene_metadata = NULL,
spatial_network = NULL,
spatial_network_name = NULL,
spatial_grid = NULL,
spatial_grid_name = NULL,
spatial_enrichment = NULL,
spatial_enrichment_name = NULL,
dimension_reduction = NULL,
nn_network = NULL,
offset_file = NULL,
instructions = NULL
```

Arguments

```
raw_exprs
                  matrix with raw expression counts [required]
                  data.table or data.frame with coordinates for cell centroids
spatial_locs
                  normalized expression values
norm_expr
norm_scaled_expr
                  scaled expression values
custom_expr
                  custom expression values
cell_metadata
                  cell annotation metadata
gene_metadata
                  gene annotation metadata
spatial_network
                  list of spatial network(s)
spatial_network_name
                  list of spatial network name(s)
                  list of spatial grid(s)
spatial_grid
spatial_grid_name
                  list of spatial grid name(s)
spatial_enrichment
                  list of spatial enrichment score(s) for each spatial region
spatial_enrichment_name
                  list of spatial enrichment name(s)
dimension_reduction
                  list of dimension reduction(s)
                  list of nearest neighbor network(s)
nn_network
                  file used to stitch fields together (optional)
offset_file
instructions
                  list of instructions or output result from createGiottoInstructions
```

Details

[Requirements] To create a giotto object you need to provide at least a matrix with genes as row names and cells as column names. To include spatial information about cells (or regions) you need to provide a data.table or data.frame with coordinates for all spatial dimensions. This can be 2D (x and y) or 3D (x, y, x). The row order for the cell coordinates should be the same as the column order for the provided expression data.

[Instructions] Additionally an instruction file, generated manually or with createGiottoInstructions can be provided to instructions, if not a default instruction file will be created for the Giotto object.

createHeatmap_DT 49

[Multiple fields] In case a dataset consists of multiple fields, like seqFISH+ for example, an offset file can be provided to stitch the different fields together. stitchFieldCoordinates can be used to generate such an offset file.

[**Processed data**] Processed count data, such as normalized data, can be provided using one of the different expression slots (norm_expr, norm_scaled_expr, custom_expr).

[Metadata] Cell and gene metadata can be provided using the cell and gene metadata slots. This data can also be added afterwards using the addGeneMetadata or addCellMetadata functions.

[Other information] Additional information can be provided through the appropriate slots:

- · spatial networks
- · spatial girds
- · spatial enrichments
- · dimensions reductions
- · nearest neighbours networks

Value

giotto object

Examples

```
createGiottoObject(raw_exprs, spatial_locs)
```

createHeatmap_DT

createHeatmap_DT

Description

creates order for clusters

```
createHeatmap_DT(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cluster_cor_method = "pearson",
  cluster_hclust_method = "ward.D",
  gene_order = c("custom", "correlation"),
  gene_custom_order = NULL,
  gene_cor_method = "pearson",
  gene_hclust_method = "complete"
)
```

50 createMetagenes

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
genes
                  genes to use
cluster_column name of column to use for clusters
cluster_order method to determine cluster order
cluster_custom_order
                  custom order for clusters
cluster_cor_method
                  method for cluster correlation
{\tt cluster\_hclust\_method}
                  method for hierarchical clustering of clusters
gene_order
                  method to determine gene order
gene_custom_order
                  custom order for genes
gene_cor_method
                  method for gene correlation
gene_hclust_method
                  method for hierarchical clustering of genes
```

Details

Creates input data.tables for plotHeatmap function.

Value

list

Examples

```
createHeatmap_DT(gobject)
```

createMetagenes

createMetagenes

Description

This function creates an average metagene for gene clusters.

```
createMetagenes(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  gene_clusters,
  name = "metagene",
  return_gobject = TRUE
)
```

createNearestNetwork 51

Arguments

Details

```
An example for the 'gene_clusters' could be like this: cluster_vector = c(1, 1, 2, 2); names(cluster_vector) = c('geneA', 'geneB', 'geneC', 'geneD')
```

Value

giotto object

Examples

```
createMetagenes(gobject)
```

createNearestNetwork createNearestNetwork

Description

create a nearest neighbour (NN) network

```
createNearestNetwork(
  gobject,
  type = c("sNN", "kNN"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  genes_to_use = NULL,
  expression_values = c("normalized", "scaled", "custom"),
  name = "sNN.pca",
  return_gobject = TRUE,
  k = 30,
  minimum_shared = 5,
  top_shared = 3,
  verbose = T,
  ...
)
```

52 createNearestNetwork

Arguments

gobject giotto object type sNN or kNN

dim_reduction_to_use

dimension reduction method to use

dim_reduction_name

name of dimension reduction set to use

dimensions_to_use

number of dimensions to use as input

genes_to_use if dim_reduction_to_use = NULL, which genes to use

expression_values

expression values to use

name arbitrary name for NN network

return_gobject boolean: return giotto object (default = TRUE)

k number of k neighbors to use minimum_shared minimum shared neighbors

top_shared keep at ...
verbose be verbose

additional parameters for kNN and sNN functions from dbscan

Details

This function creates a k-nearest neighbour (kNN) or shared nearest neighbour (sNN) network based on the provided dimension reduction space. To run it directly on the gene expression matrix set $dim_reduction_to_use = NULL$.

See also kNN and sNN for more information about how the networks are created.

Output for kNN:

• from: cell ID for source cell

• to: cell_ID for target cell

• distance: distance between cells

• weight: weight = 1/(1 + distance)

Output for sNN:

• from: cell_ID for source cell

• to: cell_ID for target cell

• distance: distance between cells

• weight: 1/(1 + distance)

• shared: number of shared neighbours

• rank: ranking of pairwise cell neighbours

For sNN networks two additional parameters can be set:

- minimum_shared: minimum number of shared neighbours needed
- top_shared: keep this number of the top shared neighbours, irrespective of minimum_shared setting

createSpatialEnrich 53

Value

giotto object with updated NN network

Examples

```
createNearestNetwork(gobject)
```

 $create Spatial Enrich \qquad \textit{create Spatial Enrich}$

Description

Function to calculate gene signature enrichment scores per spatial position using a hypergeometric test.

Usage

```
createSpatialEnrich(
  gobject,
  enrich_method = c("PAGE", "rank", "rankDT", "hypergeometric"),
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore"),
  name = "PAGE",
  return_gobject = TRUE
)
```

Arguments

```
Giotto object
gobject
enrich_method method for gene signature enrichment calculation
sign_matrix
                  Matrix of signature genes for each cell type / process
expression_values
                  expression values to use
reverse_log_scale
                  reverse expression values from log scale
logbase
                  log base to use if reverse_log_scale = TRUE
output_enrichment
                  how to return enrichment output
                  to give to spatial enrichment results, default = PAGE
name
return_gobject return giotto object
```

Details

For details see the individual functions:

```
PAGE: PAGEEnrichrank: rankEnrichrankDT: rankEnrichDT
```

• hypergeometric: hyperGeometricEnrich

54 createSpatialGrid

Value

Giotto object or enrichment results if return_gobject = FALSE

Examples

```
createSpatialEnrich(gobject)
```

createSpatialGrid

createSpatialGrid

Description

Create a spatial grid.

Usage

```
createSpatialGrid(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  sdimz_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

Arguments

Details

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

Value

giotto object with updated spatial grid slot

```
{\tt createSpatialGrid(gobject)}
```

createSpatialGrid_2D 55

```
createSpatialGrid\_2D createSpatialGrid\_2D
```

Description

create a spatial grid for 2D spatial data.

Usage

```
createSpatialGrid_2D(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

Arguments

Details

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

Value

giotto object with updated spatial grid slot

```
createSpatialGrid_2D(gobject)
```

```
createSpatialGrid\_3D \quad \textit{createSpatialGrid\_3D}
```

Description

Create a spatial grid for 3D spatial data.

Usage

```
createSpatialGrid_3D(
  gobject,
  sdimx_stepsize = NULL,
  sdimy_stepsize = NULL,
  sdimz_stepsize = NULL,
  minimum_padding = 1,
  name = "spatial_grid",
  return_gobject = TRUE
)
```

Arguments

```
gobject giotto object

sdimx_stepsize stepsize along the x-axis

sdimy_stepsize stepsize along the y-axis

sdimz_stepsize stepsize along the z-axis

minimum_padding

minimum padding on the edges

name name for spatial grid (default = 'spatial_grid')

return_gobject boolean: return giotto object (default = TRUE)
```

Details

Creates a spatial grid with defined x, y (and z) dimensions. The dimension units are based on the provided spatial location units.

Value

giotto object with updated spatial grid slot

```
createSpatialGrid_3D(gobject)
```

createSpatialNetwork 57

```
createSpatialNetwork createSpatialNetwork
```

Description

Create a spatial network based on cell centroid physical distances.

Usage

```
createSpatialNetwork(
  gobject,
  k = 4,
  dimensions = "all",
  maximum_distance = NULL,
  minimum_k = 0,
  name = "spatial_network",
  verbose = F,
  return_gobject = TRUE
)
```

Arguments

```
gobject giotto object
k number of nearest neighbors based on physical distance
dimensions which spatial dimensions to use (default = all)
maximum_distance
distance cuttof for nearest neighbors to consider
minimum_k minimum nearest neighbours if maximum_distance != NULL
name name for spatial network (default = 'spatial_network')
verbose verbose
```

return_gobject boolean: return giotto object (default = TRUE)

Details

Creates a spatial network connecting single-cells based on their physical distance to each other. Number of neighbors can be determined by k, maximum distance from each cell with or without setting a minimum k for each cell.

dimensions: default = 'all' which takes all possible dimensions. Alternatively you can provide a character vector that specififies the spatial dimensions to use, e.g. c("sdimx', "sdimy") or a numerical vector, e.g. 2:3

maximum_distance: to create a network based on maximum distance only, you also need to set k to a very high value, e.g. k = 100

Value

giotto object with updated spatial network slot

```
createSpatialNetwork(gobject)
```

58 create_average_DT

```
create\_average\_detection\_DT \\ create\_average\_detection\_DT
```

Description

calculates average gene detection for a cell metadata factor (e.g. cluster)

Usage

```
create_average_detection_DT(
  gobject,
  meta_data_name,
  expression_values = c("normalized", "scaled", "custom"),
  detection_threshold = 0
)
```

Arguments

```
gobject giotto object

meta_data_name name of metadata column to use
expression_values

which expression values to use

detection_threshold

detection threshold to consider a gene detected
```

Value

data.table with average gene epression values for each factor

Description

calculates average gene expression for a cell metadata factor (e.g. cluster)

Usage

```
create_average_DT(
  gobject,
  meta_data_name,
  expression_values = c("normalized", "scaled", "custom")
)
```

Arguments

```
gobject giotto object

meta_data_name name of metadata column to use
expression_values

which expression values to use
```

Value

data.table with average gene epression values for each factor

Description

creates randomized cell ids within a selection of cell types

Usage

```
create_cell_type_random_cell_IDs(
  gobject,
  cluster_column = "cell_types",
  needed_cell_types
)
```

Arguments

```
gobject giotto object to use

cluster_column cluster column with cell type information

needed_cell_types

vector of cell type names for which a random id will be found
```

Details

Details will follow.

Value

list of randomly sampled cell ids with same cell type composition

```
create_cell_type_random_cell_IDs(gobject)
```

60 create_dimObject

```
create_cluster_matrix create_cluster_matrix
```

Description

creates aggregated matrix for a given clustering

Usage

```
create_cluster_matrix(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  gene_subset = NULL
)
```

Examples

```
create_cluster_matrix(gobject)
```

create_dimObject

create_dimObject

Description

Creates an object that stores a dimension reduction output

Usage

```
create_dimObject(
  name = "test",
  reduction_method = NULL,
  coordinates = NULL,
  misc = NULL,
  my_rownames = NULL
)
```

Arguments

```
name arbitrary name for object
reduction_method
method used to reduce dimensions
coordinates accepts the coordinates after dimension reduction
misc any additional information will be added to this slot
```

Value

number of distinct colors

decide_cluster_order 61

```
decide_cluster_order
```

Description

creates order for clusters

Usage

```
decide_cluster_order(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cor_method = "pearson",
  hclust_method = "ward.D"
)
```

Arguments

```
gobject giotto object
expression_values
expression values to use
genes genes to use
cluster_column name of column to use for clusters
cluster_order method to determine cluster order
cluster_custom_order
custom order for clusters

cor_method method for correlation
hclust_method method for hierarchical clustering
```

Details

Calculates order for clusters.

Value

custom

```
decide_cluster_order(gobject)
```

detectSpatialCorGenes detectSpatialCorGenes

Description

Detect genes that are spatially correlated

Usage

62

```
detectSpatialCorGenes(
  gobject,
  method = c("grid", "network"),
  expression_values = c("normalized", "scaled", "custom"),
  subset_genes = NULL,
  spatial_network_name = "spatial_network",
  network_smoothing = NULL,
  spatial_grid_name = "spatial_grid",
  min_cells_per_grid = 4,
  cor_method = c("pearson", "kendall", "spearman")
)
```

Arguments

```
gobject
                  giotto object
method
                  method to use for spatial averaging
expression_values
                  gene expression values to use
subset_genes
                  subset of genes to use
spatial_network_name
                  name of spatial network to use
network_smoothing
                  smoothing factor beteen 0 and 1 (default: automatic)
spatial_grid_name
                  name of spatial grid to use
min_cells_per_grid
                  minimum number of cells to consider a grid
b
                  smoothing factor beteen 0 and 1 (default: automatic)
```

Details

For method = network, it expects a fully connected spatial network. You can make sure to create a fully connected network by setting minimal_k > 0 in the createSpatialNetwork function.

- 1. grid-averaging: average gene expression values within a predefined spatial grid
- 2. network-averaging: smoothens the gene expression matrix by averaging the expression within one cell by using the neighbours within the predefined spatial network. b is a smoothening factor that defaults to 1 1/k, where k is the median number of k-neighbors in the selected spatial network. Setting b = 0 means no smoothing and b = 1 means no contribution from its own expression.

The spatCorObject can be further explored with showSpatialCorGenes()

detectSpatialPatterns 63

Value

```
returns a spatial correlation object: "spatCorObject"
```

See Also

```
showSpatialCorGenes
```

Examples

```
detectSpatialCorGenes(gobject)
```

```
detectSpatialPatterns detectSpatialPatterns
```

Description

Identify spatial patterns through PCA on average expression in a spatial grid.

Usage

```
detectSpatialPatterns(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  spatial_grid_name = "spatial_grid",
  min_cells_per_grid = 4,
  scale_unit = F,
  ncp = 100,
  show_plot = T,
  PC_zscore = 1.5
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
spatial_grid_name
                  name of spatial grid to use (default = 'spatial_grid')
min_cells_per_grid
                  minimum number of cells in a grid to be considered
                  scale features
scale_unit
                  number of principal components to calculate
ncp
show_plot
                  show plots
PC_zscore
                  minimum z-score of variance explained by a PC
```

64 dimCellPlot

Details

Steps to identify spatial patterns:

- 1. average gene expression for cells within a grid, see createSpatialGrid
- 2. perform PCA on the average grid expression profiles
- 3. convert variance of principlal components (PCs) to z-scores and select PCs based on a z-score threshold

Value

```
spatial pattern object 'spatPatObj'
```

Examples

```
detectSpatialPatterns(gobject)
```

dimCellPlot

dimCellPlot

Description

Visualize cells according to dimension reduction coordinates

```
dimCellPlot(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  cell_annotation_values = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
```

dimCellPlot 65

```
edge_alpha = NULL,
      point_size = 1,
      point_border_col = "black",
      point_border_stroke = 0.1,
      show_legend = T,
      legend_text = 8,
      legend_symbol_size = 1,
      background_color = "white",
      axis_text = 8,
      axis_title = 8,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dimCellPlot"
    )
Arguments
                     giotto object
    gobject
    dim_reduction_to_use
                     dimension reduction to use
   dim_reduction_name
                     dimension reduction name
   dim1_to_use
                     dimension to use on x-axis
                     dimension to use on y-axis
    dim2_to_use
    spat_enr_names names of spatial enrichment results to include
    cell_annotation_values
                     numeric cell annotation columns
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
```

dimCellPlot

```
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_legend
                  show legend
legend_text
                  size of legend text
legend_symbol_size
                  size of legend symbols
background_color
                  color of plot background
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  color for cells (see details)
cell_color
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
                  title for plot, defaults to cell_color parameter
title
```

Details

Description of parameters. For 3D plots see dimCellPlot2D

Value

ggplot

```
dimCellPlot(gobject)
```

dimCellPlot2D 67

dimCellPlot2D

dimCellPlot2D

Description

Visualize cells according to dimension reduction coordinates

```
dimCellPlot2D(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  cell_annotation_values = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show\_center\_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  legend_text = 8,
  legend_symbol_size = 1,
  background_color = "white",
  axis_text = 8,
  axis_title = 8,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_plot = NA,
  return_plot = NA,
```

dimCellPlot2D

```
save_plot = NA,
      save_param = list(),
      default_save_name = "dimCellPlot2D"
    )
Arguments
    gobject
                      giotto object
    dim_reduction_to_use
                      dimension reduction to use
    dim_reduction_name
                      dimension reduction name
    dim1_to_use
                      dimension to use on x-axis
    dim2_to_use
                      dimension to use on y-axis
    spat_enr_names names of spatial enrichment results to include
    cell_annotation_values
                      numeric cell annotation columns
    show_NN_network
                      show underlying NN network
    nn_network_to_use
                      type of NN network to use (kNN vs sNN)
                      name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color_gradient
                      vector with 3 colors for numeric data
    gradient_midpoint
                      midpoint for color gradient
    gradient_limits
                      vector with lower and upper limits
    select_cell_groups
                      select subset of cells/clusters based on cell color parameter
    select_cells
                      select subset of cells based on cell IDs
    show_other_cells
                      display not selected cells
    other_cell_color
                      color of not selected cells
    other_point_size
                      size of not selected cells
    show_cluster_center
                      plot center of selected clusters
    show_center_label
                      plot label of selected clusters
    center_point_size
                      size of center points
    label_size
                      size of labels
    label_fontface font of labels
    edge_alpha
                      column to use for alpha of the edges
    point_size
                      size of point (cell)
```

dimGenePlot 69

```
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_legend
                  show legend
                  size of legend text
legend_text
legend_symbol_size
                  size of legend symbols
background_color
                  color of plot background
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function
default_save_name
                  default save name for saving, don't change, change save_name in save_param
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
title
                  title for plot, defaults to cell_color parameter
```

Details

Description of parameters. For 3D plots see dimPlot3D

Value

ggplot

Examples

dimCellPlot2D(gobject)

dimGenePlot dimGenePlot

Description

Visualize cells and gene expression according to dimension reduction coordinates

70 dimGenePlot

Usage

```
dimGenePlot(
     gobject,
     expression_values = c("normalized", "scaled", "custom"),
     genes = NULL,
     dim_reduction_to_use = "umap",
     dim_reduction_name = "umap",
     dim1_to_use = 1,
     dim2\_to\_use = 2,
     show_NN_network = F,
     nn_network_to_use = "sNN",
     network_name = "sNN.pca",
     network_color = "lightgray",
     edge_alpha = NULL,
     scale_alpha_with_expression = FALSE,
     point_size = 1,
     genes_high_color = "red",
     genes_mid_color = "white",
     genes_low_color = "blue",
     point_border_col = "black",
     point_border_stroke = 0.1,
     midpoint = 0,
     cow_n_col = 2,
     cow_rel_h = 1,
     cow_rel_w = 1,
     cow_align = "h",
     show_legend = T,
     show_plot = NA,
     return_plot = NA,
     save_plot = NA,
     save_param = list(),
     default_save_name = "dimGenePlot"
   )
Arguments
   gobject
                    giotto object
   expression_values
                    gene expression values to use
                    genes to show
   genes
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   show_NN_network
                    show underlying NN network
   nn_network_to_use
```

type of NN network to use (kNN vs sNN)

dimGenePlot 71

```
name of NN network to use, if show_NN_network = TRUE
network_name
edge_alpha
                 column to use for alpha of the edges
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
                 size of point (cell)
point_size
point_border_col
                 color of border around points
point_border_stroke
                 stroke size of border around points
midpoint
                 size of point (cell)
cow_n_col
                 cowplot param: how many columns
cow_rel_h
                 cowplot param: relative height
                 cowplot param: relative width
cow_rel_w
cow_align
                 cowplot param: how to align
show_legend
                 show legend
show_plot
                 show plots
return_plot
                 return ggplot object
save_plot
                 directly save the plot [boolean]
                 list of saving parameters from all_plots_save_function
save_param
default_save_name
                 default save name for saving, don't change, change save_name in save_param
                 parameters for cowplot::save_plot()
```

Details

Description of parameters.

Value

ggplot

See Also

dimGenePlot3D

Examples

dimGenePlot(gobject)

72 dimGenePlot2D

dimGenePlot2D

dimGenePlot2D

Description

Visualize cells and gene expression according to dimension reduction coordinates

Usage

```
dimGenePlot2D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = "red",
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dimGenePlot2D"
)
```

Arguments

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
dim_reduction_to_use
dimension reduction to use
```

dimGenePlot2D 73

```
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
                 column to use for alpha of the edges
edge_alpha
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
point_size
                 size of point (cell)
point_border_col
                 color of border around points
point_border_stroke
                 stroke size of border around points
midpoint
                 size of point (cell)
cow_n_col
                 cowplot param: how many columns
cow_rel_h
                 cowplot param: relative height
                 cowplot param: relative width
cow_rel_w
cow_align
                 cowplot param: how to align
show_legend
                 show legend
show_plot
                 show plots
return_plot
                 return ggplot object
save_plot
                 directly save the plot [boolean]
save_param
                 list of saving parameters from all_plots_save_function
default_save_name
                 default save name for saving, don't change, change save_name in save_param
                 parameters for cowplot::save_plot()
. . .
```

Details

Description of parameters.

Value

ggplot

See Also

dimGenePlot3D

Examples

dimGenePlot2D(gobject)

74 dimGenePlot3D

dimGenePlot3D

dimGenePlot3D

Description

Visualize cells and gene expression according to dimension reduction coordinates

Usage

```
dimGenePlot3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = 3,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  cluster_column = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  edge_alpha = NULL,
  point_size = 2,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dimGenePlot3D"
)
```

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
dim_reduction_to_use
dim_neduction_name
dimension reduction name
```

dimPlot 75

```
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
edge_alpha
                 column to use for alpha of the edges
point_size
                 size of point (cell)
show_legend
                 show legend
show_plot
                 show plots
return_plot
                 return ggplot object
save_plot
                 directly save the plot [boolean]
                 list of saving parameters from all_plots_save_function
save_param
default_save_name
                 default save name for saving, don't change, change save_name in save_param
                 parameters for cowplot::save_plot()
. . .
```

Details

Description of parameters.

Value

ggplot

Examples

```
dimGenePlot3D(gobject)
```

dimPlot dimPlot

Description

Visualize cells according to dimension reduction coordinates

```
dimPlot(
  gobject,
  group_by = NULL,
  group_by_subset = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
```

76 dimPlot

```
nn_network_to_use = "sNN",
     network_name = "sNN.pca",
     cell_color = NULL,
     color_as_factor = T,
     cell_color_code = NULL,
     cell_color_gradient = c("blue", "white", "red"),
     gradient_midpoint = NULL,
     gradient_limits = NULL,
     select_cell_groups = NULL,
     select_cells = NULL,
     show_other_cells = T,
     other_cell_color = "lightgrey",
     other_point_size = 0.5,
     show_cluster_center = F,
     show_center_label = T,
     center_point_size = 4,
     center_point_border_col = "black",
     center_point_border_stroke = 0.1,
     label_size = 4,
     label_fontface = "bold",
     edge_alpha = NULL,
     point_shape = c("border", "no_border"),
     point_size = 1,
     point_border_col = "black",
     point_border_stroke = 0.1,
     show_legend = T,
     legend_text = 8,
     legend_symbol_size = 1,
     background_color = "white",
     axis_text = 8,
     axis_title = 8,
     title = NULL,
     cow_n_col = 2,
     cow_rel_h = 1,
     cow_rel_w = 1,
     cow_align = "h",
     show_plot = NA,
     return_plot = NA,
     save_plot = NA,
     save_param = list(),
     default_save_name = "dimPlot"
   )
Arguments
   gobject
                   giotto object
   group_by_subset
                   subset the group_by factor column
   dim_reduction_to_use
                   dimension reduction to use
```

spat_enr_names = NULL, show_NN_network = F, dim_reduction_name dimension reduction name dim1_to_use dimension to use on x-axis dim2_to_use dimension to use on y-axis spat_enr_names names of spatial enrichment results to include show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) network_name name of NN network to use, if show_NN_network = TRUE cell_color color for cells (see details) color_as_factor convert color column to factor cell_color_code named vector with colors cell_color_gradient vector with 3 colors for numeric data gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size size of not selected cells show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points label_size size of labels label_fontface font of labels edge_alpha column to use for alpha of the edges point_shape point with border or not (border or no_border) size of point (cell) point_size point_border_col color of border around points point_border_stroke stroke size of border around points

show_legend

show legend

legend_text size of legend text
legend_symbol_size

size of legend symbols

background_color

color of plot background

axis_text size of axis text
axis_title size of axis title

title title for plot, defaults to cell_color parameter

cow_n_col cowplot param: how many columns
cow_rel_h cowplot param: relative height
cow_rel_w cowplot param: relative width
cow_align cowplot param: how to align

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function

default_save_name

default save name for saving, don't change, change save_name in save_param

groub_by create multiple plots based on cell annotation column

Details

Description of parameters, see dimPlot2D. For 3D plots see dimPlot3D

Value

ggplot

Examples

dimPlot(gobject)

dimPlot2D dimPlot2D

Description

Visualize cells according to dimension reduction coordinates

```
dimPlot2D(
  gobject,
  group_by = NULL,
  group_by_subset = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_shape = c("border", "no_border"),
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  title = NULL,
  show_legend = T,
  legend_text = 8,
  legend_symbol_size = 1,
  background_color = "white",
  axis_text = 8,
  axis_title = 8,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "dimPlot2D"
```

)

Arguments

gobject giotto object group_by_subset subset the group_by factor column dim_reduction_to_use dimension reduction to use dim_reduction_name dimension reduction name dim1_to_use dimension to use on x-axis dim2_to_use dimension to use on y-axis spat_enr_names names of spatial enrichment results to include show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) network_name name of NN network to use, if show_NN_network = TRUE cell_color color for cells (see details) color_as_factor convert color column to factor cell_color_code named vector with colors cell_color_gradient vector with 3 colors for numeric data gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size size of not selected cells show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points size of labels label_size label_fontface font of labels edge_alpha column to use for alpha of the edges

```
point with border or not (border or no_border)
point_shape
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
title
                  title for plot, defaults to cell_color parameter
show_legend
                  show legend
legend_text
                  size of legend text
legend_symbol_size
                  size of legend symbols
background_color
                  color of plot background
axis_text
                  size of axis text
axis_title
                  size of axis title
cow_n_col
                  cowplot param: how many columns
cow_rel_h
                  cowplot param: relative height
cow_rel_w
                  cowplot param: relative width
cow_align
                  cowplot param: how to align
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
groub_by
                  create multiple plots based on cell annotation column
```

Details

Description of parameters. For 3D plots see dimPlot3D

Value

ggplot

Examples

```
dimPlot2D(gobject)
```

82 dimPlot2D_single

dimPlot2D_single

dimPlot2D_single

Description

Visualize cells according to dimension reduction coordinates

```
dimPlot2D_single(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  spat_enr_names = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_shape = c("border", "no_border"),
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  title = NULL,
  show_legend = T,
  legend_text = 8,
  legend_symbol_size = 1,
  background_color = "white",
  axis_text = 8,
  axis_title = 8,
  show_plot = NA,
  return_plot = NA,
```

dimPlot2D_single 83

```
save_plot = NA,
      save_param = list(),
      default_save_name = "dimPlot2D_single"
    )
Arguments
                     giotto object
    gobject
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    spat_enr_names names of spatial enrichment results to include
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color
                     color for cells (see details)
    color_as_factor
                      convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
                     size of center points
    label_size
                     size of labels
```

label_fontface font of labels

84 dimPlot3D

column to use for alpha of the edges edge_alpha point_shape point with border or not (border or no_border) point_size size of point (cell) point_border_col color of border around points point_border_stroke stroke size of border around points title title for plot, defaults to cell_color parameter show_legend show legend legend_text size of legend text legend_symbol_size size of legend symbols background_color color of plot background axis_text size of axis text axis_title size of axis title show_plot show plot

directly save the plot [boolean]

return ggplot object

save_param list of saving parameters from all_plots_save_function

 ${\tt default_save_name}$

return_plot

save_plot

default save name for saving, don't change, change save_name in save_param

Details

Description of parameters. For 3D plots see dimPlot3D

Value

ggplot

Examples

dimPlot2D_single(gobject)

dimPlot3D dimPlot3D

Description

Visualize cells according to dimension reduction coordinates

dimPlot3D 85

Usage

```
dimPlot3D(
      gobject,
      dim_reduction_to_use = "umap",
      dim_reduction_name = "umap",
      dim1_to_use = 1,
      dim2_to_use = 2,
      dim3_to_use = 3,
      select_cell_groups = NULL,
      select_cells = NULL,
      show_other_cells = T,
      other_cell_color = "lightgrey",
      other_point_size = 2,
      show_NN_network = F,
      nn_network_to_use = "sNN",
      network_name = "sNN.pca",
      color_as_factor = T,
      cell_color = NULL,
      cell_color_code = NULL,
      show_cluster_center = F,
      show_center_label = T,
      center_point_size = 4,
      label_size = 4,
      edge_alpha = NULL,
      point_size = 3,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "dim3D"
Arguments
    gobject
                    giotto object
    dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   dim3_to_use
                    dimension to use on z-axis
    select_cell_groups
                    select subset of cells/clusters based on cell_color parameter
                    select subset of cells based on cell IDs
    select_cells
    show_other_cells
                    display not selected cells
   other_cell_color
                    color of not selected cells
   other_point_size
```

size of not selected cells

86 dimPlot3D

show_NN_network

show underlying NN network

nn_network_to_use

type of NN network to use (kNN vs sNN)

network_name name of NN network to use, if show_NN_network = TRUE

color_as_factor

convert color column to factor

cell_color color for cells (see details)

cell_color_code

named vector with colors

 ${\tt show_cluster_center}$

plot center of selected clusters

show_center_label

plot label of selected clusters

center_point_size

size of center points

label_size size of labels

edge_alpha column to use for alpha of the edges

point_size size of point (cell)

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param
list of saving parameters from all_plots_save_function

default_save_name

default save name for saving, don't change, change save_name in save_param

show_legend show legend

Details

Description of parameters.

Value

plotly

Examples

dimPlot3D(gobject)

direction_test_CPG 87

```
direction_test_CPG direction_test_CPG
```

Description

shows direction of change

Usage

```
direction_test(x, min_fdr = 0.05)
```

Examples

```
direction_test_CPG()
```

doHclust

doHclust

Description

cluster cells using hierarchical clustering algorithm

Usage

```
doHclust(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
  dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  distance_method = c("pearson", "spearman", "original", "euclidean", "maximum",
  "manhattan", "canberra", "binary", "minkowski"),
agglomeration_method = c("ward.D2", "ward.D", "single", "complete", "average",
    "mcquitty", "median", "centroid"),
  k = 10,
  h = NULL
  name = "hclust",
  return_gobject = TRUE,
  set\_seed = T,
  seed_number = 1234
)
```

```
gobject giotto object
expression_values
expression values to use
genes_to_use subset of genes to use
```

88 doHMRF

```
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimensions reduction name
dimensions_to_use
                 dimensions to use
distance_method
                 distance method
agglomeration_method
                 agglomeration method for hclust
k
                 number of final clusters
                 cut hierarchical tree at height = h
h
                 name for hierarchical clustering
name
return_gobject boolean: return giotto object (default = TRUE)
                 set seed
set_seed
seed_number
                 number for seed
```

Details

Description on how to use Kmeans clustering method.

Value

giotto object with new clusters appended to cell metadata

See Also

hclust

Examples

doHclust(gobject)

doHMRF

doHMRF

Description

Run HMRF

```
doHMRF(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  spatial_network_name = "spatial_network",
  spatial_genes = NULL,
  spatial_dimensions = c("sdimx", "sdimy", "sdimz"),
  dim_reduction_to_use = NULL,
  dim_reduction_name = "pca",
```

doHMRF 89

```
dimensions_to_use = 1:10,
      name = "test",
      k = 10,
      betas = c(0, 2, 50),
      tolerance = 1e-10,
      zscore = c("none", "rowcol", "colrow"),
      numinit = 100,
      python_path = NULL,
      output_folder = NULL,
      overwrite_output = TRUE
    )
Arguments
                     giotto object
   gobject
    expression_values
                     expression values to use
    spatial_network_name
                     name of spatial network to use for HMRF
    spatial_genes
                     spatial genes to use for HMRF
    spatial_dimensions
                     select spatial dimensions to use, default is all possible dimensions
    dim_reduction_to_use
                     use another dimension reduction set as input
    dim_reduction_name
                     name of dimension reduction set to use
   dimensions_to_use
                     number of dimensions to use as input
                     name of HMRF run
   name
                     number of HMRF domains
   k
   betas
                     betas to test for
    tolerance
                     tolerance
```

tolerance tolerance zscore zscore

numinit number of initializations

python_path python path to use

output_folder output folder to save results

overwrite_output

overwrite output folder

Details

Description of HMRF parameters ...

Value

Creates a directory with results that can be viewed with viewHMRFresults

Examples

```
doHMRF(gobject)
```

90 doKmeans

doKmeans

doKmeans

Description

cluster cells using kmeans algorithm

Usage

```
doKmeans(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_to_use = NULL,
  dim_reduction_to_use = c("cells", "pca", "umap", "tsne"),
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  distance_method = c("original", "pearson", "spearman", "euclidean", "maximum",
    "manhattan", "canberra", "binary", "minkowski"),
  centers = 10,
  iter_max = 100,
  nstart = 1000,
  algorithm = "Hartigan-Wong",
  name = "kmeans",
  return_gobject = TRUE,
  set_seed = T,
  seed_number = 1234
)
```

```
gobject
                 giotto object
expression_values
                 expression values to use
                 subset of genes to use
genes_to_use
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimensions reduction name
dimensions_to_use
                 dimensions to use
distance_method
                 distance method
                 number of final clusters
centers
                 kmeans maximum iterations
iter_max
nstart
                 kmeans nstart
                 kmeans algorithm
algorithm
name
                 name for kmeans clustering
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                 set seed
seed_number
                 number for seed
```

doLeidenCluster 91

Details

Description on how to use Kmeans clustering method.

Value

giotto object with new clusters appended to cell metadata

See Also

kmeans

Examples

```
doKmeans(gobject)
```

doLeidenCluster

doLeidenCluster

Description

cluster cells using a NN-network and the Leiden community detection algorithm

Usage

```
doLeidenCluster(
  gobject,
  name = "leiden_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = "weight",
  partition_type = c("RBConfigurationVertexPartition", "ModularityVertexPartition"),
  init_membership = NULL,
  n_iterations = 1000,
  return_gobject = TRUE,
  set_seed = T,
  seed_number = 1234,
  ...
)
```

92 doLeidenSubCluster

weight_col weight column to use for edges

partition_type The type of partition to use for optimisation.

init_membership

initial membership of cells for the partition

n_iterations number of interations to run the Leiden algorithm. If the number of iterations

is negative, the Leiden algorithm is run until an iteration in which there was no

improvement.

return_gobject boolean: return giotto object (default = TRUE)

set_seed set seed

seed_number number for seed

Details

This function is a wrapper for the Leiden algorithm implemented in python, which can detect communities in graphs of millions of nodes (cells), as long as they can fit in memory. See the https://github.com/vtraag/leidenalgleidenalg github page or the https://leidenalg.readthedocs.io/en/stable/index.htmlreadthedocs page for more information.

Partition types available and information:

- RBConfigurationVertexPartition: Implements Reichardt and Bornholdt's Potts model with a configuration null model. This quality function is well-defined only for positive edge weights. This quality function uses a linear resolution parameter.
- Modularity Vertex Partition: Implements modularity. This quality function is well-defined only for positive edge weights. It does *not* use the resolution parameter

Set $weight_col = NULL$ to give equal weight (=1) to each edge.

Value

giotto object with new clusters appended to cell metadata

Examples

doLeidenCluster(gobject)

doLeidenSubCluster

doLeidenSubCluster

Description

Further subcluster cells using a NN-network and the Leiden algorithm

doLeidenSubCluster 93

Usage

n_iterations

network_name

verbose

nn_network_to_use

python_path

```
doLeidenSubCluster(
      gobject,
      name = "sub_pleiden_clus",
      cluster_column = NULL,
      selected_clusters = NULL,
     hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
        = "normalized"),
     hvg_min_perc_cells = 5,
     hvg_mean_expr_det = 1,
      use_all_genes_as_hvg = FALSE,
     min_nr_of_hvg = 5,
     pca_param = list(expression_values = "normalized", scale_unit = T),
     nn_param = list(dimensions_to_use = 1:20),
     k_neighbors = 10,
      resolution = 0.5,
     n_{iterations} = 500,
      python_path = NULL,
      nn_network_to_use = "sNN",
     network_name = "sNN.pca",
     return_gobject = TRUE,
      verbose = T
Arguments
   gobject
                    giotto object
   name
                    name for new clustering result
   cluster_column cluster column to subcluster
    selected_clusters
                    only do subclustering on these clusters
   hvg_param
                    parameters for calculateHVG
   hvg_min_perc_cells
                    threshold for detection in min percentage of cells
   hvg_mean_expr_det
                    threshold for mean expression level in cells with detection
   use_all_genes_as_hvg
                    forces all genes to be HVG and to be used as input for PCA
                    minimum number of HVG, or all genes will be used as input for PCA
   min_nr_of_hvg
                    parameters for runPCA
   pca_param
   nn_param
                    parameters for parameters for createNearestNetwork
                    number of k for createNearestNetwork
   k_neighbors
                    resolution of Leiden clustering
   resolution
```

number of interations to run the Leiden algorithm.

specify specific path to python if required

type of NN network to use (kNN vs sNN)

name of NN network to use

return_gobject boolean: return giotto object (default = TRUE)

verbose

94 doLouvainCluster

Details

This function performs subclustering using the Leiden algorithm on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do Leiden clustering

Value

giotto object with new subclusters appended to cell metadata

See Also

```
doLeidenCluster
```

Examples

```
doLeidenSubCluster(gobject)
```

doLouvainCluster

doLouvainCluster

Description

cluster cells using a NN-network and the Louvain algorithm.

```
doLouvainCluster(
  gobject,
  version = c("community", "multinet"),
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  python_path = NULL,
  resolution = 1,
  weight_col = NULL,
  gamma = 1,
  omega = 1,
  louv_random = F,
  return_gobject = TRUE,
  set\_seed = F,
  seed_number = 1234,
)
```

Arguments

gobject giotto object

version implemented version of Louvain clustering to use

name name for cluster

nn_network_to_use

type of NN network to use (kNN vs sNN)

network_name name of NN network to use

python_path [community] specify specific path to python if required

resolution [community] resolution

gamma [multinet] Resolution parameter for modularity in the generalized louvain method.

omega [multinet] Inter-layer weight parameter in the generalized louvain method.

return_gobject boolean: return giotto object (default = TRUE)

set_seed set seed

seed_number number for seed

Details

Louvain clustering using the community or multinet implementation of the louvain clustering algorithm.

Value

giotto object with new clusters appended to cell metadata

See Also

doLouvainCluster_community and doLouvainCluster_multinet

Examples

doLouvainCluster(gobject)

doLouvainCluster_community

 $doLouvainCluster_community$

Description

cluster cells using a NN-network and the Louvain algorithm from the community module in Python

Usage

```
doLouvainCluster_community(
   gobject,
   name = "louvain_clus",
   nn_network_to_use = "sNN",
   network_name = "sNN.pca",
   python_path = NULL,
   resolution = 1,
   weight_col = NULL,
   louv_random = F,
   return_gobject = TRUE,
   set_seed = F,
   seed_number = 1234,
   ...
)
```

Arguments

gobject giotto object
name name for cluster
nn_network_to_use

type of NN network to use (kNN vs sNN)

network_name name of NN network to use

python_path specify specific path to python if required

resolution resolution

weight_col weight column to use for edges

louv_random Will randomize the node evaluation order and the community evaluation order

to get different partitions at each call

return_gobject boolean: return giotto object (default = TRUE)

set_seed set seed

seed_number number for seed

Details

This function is a wrapper for the Louvain algorithm implemented in Python, which can detect communities in graphs of nodes (cells). See the https://python-louvain.readthedocs.io/en/latest/index.htmlreadthedocs page for more information.

Set $weight_col = NULL$ to give equal weight (=1) to each edge.

Value

giotto object with new clusters appended to cell metadata

Examples

```
doLouvainCluster_community(gobject)
```

```
\label{lower} do Louvain Cluster\_multinet \\ do Louvain Cluster\_multinet
```

Description

cluster cells using a NN-network and the Louvain algorithm from the multinet package in R.

Usage

```
doLouvainCluster_multinet(
  gobject,
  name = "louvain_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  gamma = 1,
  omega = 1,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

Arguments

```
gobject
                  giotto object
                  name for cluster
name
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
                  name of NN network to use
network_name
                  Resolution parameter for modularity in the generalized louvain method.
gamma
                  Inter-layer weight parameter in the generalized louvain method.
omega
return_gobject boolean: return giotto object (default = TRUE)
set_seed
                  set seed
seed_number
                  number for seed
```

Details

See $glouvain_ml$ from the multinet package in R for more information.

Value

giotto object with new clusters appended to cell metadata

Examples

```
doLouvainCluster_multinet(gobject)
```

98 doLouvainSubCluster

 ${\tt doLouvainSubCluster} \qquad {\tt doLouvainSubCluster}$

Description

subcluster cells using a NN-network and the Louvain algorithm

Usage

```
doLouvainSubCluster(
  gobject,
  name = "sub_louvain_clus",
  version = c("community", "multinet"),
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
  hvg_min_perc_cells = 5,
  hvg_mean_expr_det = 1,
  use_all_genes_as_hvg = FALSE,
  min_nr_of_hvg = 5,
  pca_param = list(expression_values = "normalized", scale_unit = T),
  nn_param = list(dimensions_to_use = 1:20),
  k_neighbors = 10,
  resolution = 0.5,
  gamma = 1,
  omega = 1,
  python_path = NULL,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  return_gobject = TRUE,
  verbose = T
)
```

```
gobject
                  giotto object
name
                  name for new clustering result
                  version of Louvain algorithm to use
version
cluster_column cluster column to subcluster
selected_clusters
                  only do subclustering on these clusters
                  parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                  threshold for detection in min percentage of cells
hvg_mean_expr_det
                  threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                  forces all genes to be HVG and to be used as input for PCA
```

doLouvainSubCluster 99

min_nr_of_hvg minimum number of HVG, or all genes will be used as input for PCA

pca_param parameters for runPCA

nn_param parameters for parameters for createNearestNetwork

 $k_neighbors \qquad number \ of \ k \ for \ createNearestNetwork$

resolution resolution for community algorithm

gamma gamma omega omega

python_path specify specific path to python if required

nn_network_to_use

type of NN network to use (kNN vs sNN)

network_name name of NN network to use

return_gobject boolean: return giotto object (default = TRUE)

verbose verbose

Details

This function performs subclustering using the Louvain algorithm on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do Louvain clustering

Value

giotto object with new subclusters appended to cell metadata

See Also

doLouvainCluster_multinet and doLouvainCluster_community

Examples

doLouvainSubCluster(gobject)

```
\label{lower_community} do Louvain SubCluster\_community \\ do Louvain SubCluster\_community
```

Description

subcluster cells using a NN-network and the Louvain community detection algorithm

Usage

```
doLouvainSubCluster_community(
  gobject,
 name = "sub_louvain_comm_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
   = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 10,
  resolution = 0.5,
 python_path = NULL,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 return_gobject = TRUE,
  verbose = T
)
```

```
gobject
                  giotto object
                  name for new clustering result
name
cluster_column cluster column to subcluster
selected_clusters
                  only do subclustering on these clusters
                  parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                  threshold for detection in min percentage of cells
hvg_mean_expr_det
                  threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                  forces all genes to be HVG and to be used as input for PCA
                  minimum number of HVG, or all genes will be used as input for PCA
min_nr_of_hvg
                  parameters for runPCA
pca_param
```

nn_param parameters for parameters for createNearestNetwork

k_neighbors number of k for createNearestNetwork

resolution resolution

python_path specify specific path to python if required

nn_network_to_use

type of NN network to use (kNN vs sNN)

network_name name of NN network to use

return_gobject boolean: return giotto object (default = TRUE)

verbose verbose

Details

This function performs subclustering using the Louvain community algorithm on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do Louvain community clustering

Value

giotto object with new subclusters appended to cell metadata

See Also

```
doLouvainCluster_community
```

Examples

doLouvainSubCluster_community(gobject)

doLouvainSubCluster_multinet

doLouvainSubCluster_multinet

Description

subcluster cells using a NN-network and the Louvain multinet detection algorithm

Usage

```
doLouvainSubCluster_multinet(
  gobject,
  name = "sub_louvain_mult_clus",
  cluster_column = NULL,
  selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
    = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 10,
  gamma = 1,
 omega = 1,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 return_gobject = TRUE,
  verbose = T
               giotto object
```

```
gobject
name
                 name for new clustering result
cluster_column cluster column to subcluster
selected_clusters
                 only do subclustering on these clusters
                 parameters for calculateHVG
hvg_param
hvg_min_perc_cells
                 threshold for detection in min percentage of cells
hvg_mean_expr_det
                 threshold for mean expression level in cells with detection
use_all_genes_as_hvg
                 forces all genes to be HVG and to be used as input for PCA
                 minimum number of HVG, or all genes will be used as input for PCA
min_nr_of_hvg
pca_param
                 parameters for runPCA
nn_param
                 parameters for parameters for createNearestNetwork
                 number of k for createNearestNetwork
k_neighbors
gamma
                 gamma
                 omega
omega
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use
return_gobject boolean: return giotto object (default = TRUE)
                  verbose
verbose
                 specify specific path to python if required
python_path
```

doRandomWalkCluster 103

Details

This function performs subclustering using the Louvain multinet algorithm on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do Louvain multinet clustering

Value

giotto object with new subclusters appended to cell metadata

See Also

```
doLouvainCluster_multinet
```

Examples

```
doLouvainSubCluster_multinet(gobject)
```

doRandomWalkCluster

doRandomWalkCluster

Description

Cluster cells using a random walk approach.

```
doRandomWalkCluster(
  gobject,
  name = "random_walk_clus",
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  walk_steps = 4,
  walk_clusters = 10,
  walk_weights = NA,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

doSNNCluster

Arguments

```
gobject
                 giotto object
                 name for cluster
name
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use
network_name
walk_steps
                 number of walking steps
walk_clusters
                 number of final clusters
walk_weights
                 cluster column defining the walk weights
return_gobject boolean: return giotto object (default = TRUE)
set\_seed
                 set seed
seed_number
                 number for seed
```

Details

See cluster_walktrap function from the igraph package in R for more information.

Value

giotto object with new clusters appended to cell metadata

Examples

```
doRandomWalkCluster(gobject)
```

doSNNCluster

doSNNCluster

Description

Cluster cells using a SNN cluster approach.

```
doSNNCluster(
  gobject,
  name = "sNN_clus",
  nn_network_to_use = "kNN",
  network_name = "kNN.pca",
  k = 20,
  eps = 4,
  minPts = 16,
  borderPoints = TRUE,
  return_gobject = TRUE,
  set_seed = F,
  seed_number = 1234,
  ...
)
```

Arguments

gobject giotto object name name for cluster

nn_network_to_use

type of NN network to use (only works on kNN)

 $network_name$ name of kNN network to use

k Neighborhood size for nearest neighbor sparsification to create the shared NN

graph.

eps Two objects are only reachable from each other if they share at least eps nearest

neighbors.

minPts minimum number of points that share at least eps nearest neighbors for a point

to be considered a core points.

borderPoints should borderPoints be assigned to clusters like in DBSCAN?

return_gobject boolean: return giotto object (default = TRUE)

set_seed set seed

seed_number number for seed

Details

See sNNclust from dbscan package

Value

giotto object with new clusters appended to cell metadata

Examples

```
doSNNCluster(gobject)
```

```
do_spatial_grid_averaging
```

 $do_spatial_grid_averaging$

Description

smooth gene expression over a defined spatial grid

```
do_spatial_grid_averaging(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  subset_genes = NULL,
  spatial_grid_name = "spatial_grid",
  min_cells_per_grid = 4
)
```

Arguments

```
gobject giotto object
expression_values
gene expression values to use
subset_genes subset of genes to use
spatial_grid_name
name of spatial grid to use
min_cells_per_grid
minimum number of cells to consider a grid
```

Value

matrix with smoothened gene expression values based on spatial grid

Examples

```
do_spatial_grid_averaging(gobject)
```

```
\begin{tabular}{ll} $do\_spatial\_knn\_smoothing \\ $do\_spatial\_knn\_smoothing \\ \end{tabular}
```

Description

smooth gene expression over a kNN spatial network

Usage

```
do_spatial_knn_smoothing(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  subset_genes = NULL,
  spatial_network_name = "spatial_network",
  b = NULL
)
```

```
gobject giotto object

expression_values

gene expression values to use

subset_genes subset of genes to use

spatial_network_name

name of spatial network to use

b smoothing factor beteen 0 and 1 (default: automatic)
```

dt_to_matrix 107

Details

This function will smoothen the gene expression values per cell according to its neighbors in the selected spatial network.

b is a smoothening factor that defaults to 1 - 1/k, where k is the median number of k-neighbors in the selected spatial network. Setting b=0 means no smoothing and b=1 means no contribution from its own expression.

Value

matrix with smoothened gene expression values based on kNN spatial network

Examples

```
do_spatial_knn_smoothing(gobject)
```

```
dt_to_matrix
```

dt_to_matrix

Description

converts data.table to matrix

Usage

```
dt_to_matrix(x)
```

Examples

```
dt_to_matrix(x)
```

 ${\tt exportGiottoViewer}$

exportGiottoViewer

Description

compute highly variable genes

```
exportGiottoViewer(
  gobject,
  output_directory = NULL,
  spat_enr_names = NULL,
  factor_annotations = NULL,
  numeric_annotations = NULL,
  dim_reductions,
  dim_reduction_names,
  expression_values = c("scaled", "normalized", "custom"),
  dim_red_rounding = NULL,
  dim_red_rescale = c(-20, 20),
```

108 exportGiottoViewer

```
expression_rounding = 2,
  overwrite_dir = T,
  verbose = T
)
```

Arguments

```
gobject
                  giotto object
output_directory
                  directory where to save the files
spat_enr_names spatial enrichment results to include for annotations
factor_annotations
                  giotto cell annotations to view as factor
numeric_annotations
                  giotto cell annotations to view as numeric
dim_reductions high level dimension reductions to view
dim_reduction_names
                  specific dimension reduction names
expression_values
                  expression values to use in Viewer
dim_red_rounding
                  numerical indicating how to round the coordinates
dim_red_rescale
                  numericals to rescale the coordinates
expression_rounding
                  numerical indicating how to round the expression data
                  overwrite files in the directory if it already existed
overwrite_dir
                  be verbose
verbose
```

Details

Giotto Viewer expects the results from Giotto Analyzer in a specific format, which is provided by this function. To include enrichment results from <code>createSpatialEnrich</code> include the provided spatial enrichment name (default PAGE or rank) and add the gene signature names (.e.g cell types) to the numeric annotations parameter.

Value

writes the necessary output to use in Giotto Viewer

Examples

```
exportGiottoViewer(gobject)
```

```
\label{eq:constraint} exprOnlyCellCellcommunicationScores \\ exprOnlyCellCellcommunicationScores
```

Description

Cell-Cell communication scores based on expression only

Usage

```
exprOnlyCellCellcommunicationScores(
  gobject,
  cluster_column = "cell_types",
  random_iter = 100,
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  verbose = T
)
```

Arguments

```
gobject giotto object to use

cluster_column cluster column with cell type information

random_iter number of iterations

gene_set_1 first specific gene set from gene pairs

gene_set_2 second specific gene set from gene pairs

log2FC_addendum

addendum to add when calculating log2FC

verbose verbose
```

Details

Statistical framework to identify if pairs of genes (such as ligand-receptor combinations) are expressed at higher levels than expected based on a reshuffled null distribution of gene expression values, without considering the spatial position of cells. More details will follow soon.

Value

Cell-Cell communication scores for gene pairs based on expression only

```
exprOnlyCellCellcommunicationScores(gobject)
```

110 extractNearestNetwork

```
extended_gini_fun extended_gini_fun
```

Description

calculate gini coefficient on a minimum length vector

Usage

```
extended_gini_fun(x, weights = rep(1, length = length(x)), minimum_length = 16)
```

Value

gini coefficient

```
extractNearestNetwork extractNearestNetwork
```

Description

Extracts a NN-network from a Giotto object

Usage

```
extractNearestNetwork(
  gobject,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  output = c("igraph", "data.table")
)
```

Arguments

Value

igraph or data.table object

```
extractNearestNetwork(gobject)
```

fDataDT

fDataDT

fDataDT

Description

show gene metadata

Usage

```
fDataDT(gobject)
```

Arguments

gobject

giotto object

Value

data.table with gene metadata

Examples

```
pDataDT(gobject)
```

filterCombinations

filterCombinations

Description

Shows how many genes and cells are lost with combinations of thresholds.

```
filterCombinations(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_thresholds = c(1, 2),
  gene_det_in_min_cells = c(5, 50),
  min_det_genes_per_cell = c(200, 400),
  scale_x_axis = "identity",
  x_axis_offset = 0,
  scale_y_axis = "identity",
  y_axis_offset = 0,
  show_plot = TRUE
)
```

112 filterCPGscores

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
expression_thresholds
                  all thresholds to consider a gene expressed
gene_det_in_min_cells
                  minimum number of cells that should express a gene to consider that gene fur-
min_det_genes_per_cell
                  minimum number of expressed genes per cell to consider that cell further
                  ggplot transformation for x-axis (e.g. log2)
scale_x_axis
x_axis_offset x-axis offset to be used together with the scaling transformation
scale_y_axis
                  ggplot transformation for y-axis (e.g. log2)
y_axis_offset y-axis offset to be used together with the scaling transformation
show_plot
                  show plot
```

Details

Creates a scatterplot that visualizes the number of genes and cells that are lost with a specific combination of a gene and cell threshold given an arbitrary cutoff to call a gene expressed. This function can be used to make an informed decision at the filtering step with filterGiotto.

Value

list of data.table and ggplot object

Examples

```
filterCombinations(gobject)
```

 ${\tt filter CPG scores}$

filterCPGscores

Description

visualize Cell Proximity Gene enrichment scores

```
filterCPGscores(
   CPGscore,
   min_cells = 5,
   min_fdr = 0.05,
   min_spat_diff = 0.2,
   min_log2_fc = 0.5,
   keep_int_duplicates = TRUE,
   direction = c("both", "up", "down")
)
```

filterDistributions 113

Arguments

Details

This function filters the output from getCellProximityGeneScores based on false-discovery rate, minimum absolute difference, minimum log fold-change and direction of change.

Value

Gene to gene scores in data.table format

Examples

```
filterCPGscores(CPGscore)
```

```
filterDistributions filterDistributions
```

Description

show gene or cell distribution after filtering on expression threshold

```
filterDistributions(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_threshold = 1,
  detection = c("genes", "cells"),
  plot_type = c("histogram", "violin"),
  nr_bins = 30,
  fill_color = "lightblue",
  scale_axis = "identity",
  axis_offset = 0,
  show_plot = TRUE
)
```

114 filterGiotto

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
expression\_threshold
                  threshold to consider a gene expressed
detection
                  consider genes or cells
                  type of plot
plot_type
nr_bins
                  number of bins for histogram plot
fill_color
                  fill color for plots
scale_axis
                  ggplot transformation for axis (e.g. log2)
axis_offset
                  offset to be used together with the scaling transformation
show_plot
                  show plot
```

Value

ggplot object

Examples

filterDistributions(gobject)

filterGiotto filterGiotto

Description

filter Giotto object based on expression threshold

Usage

```
filterGiotto(
  gobject,
  expression_values = c("raw", "normalized", "scaled", "custom"),
  expression_threshold = 1,
  gene_det_in_min_cells = 100,
  min_det_genes_per_cell = 100,
  verbose = F
)
```

Arguments

```
gobject giotto object
expression_values
expression values to use
expression_threshold
threshold to consider a gene expressed
```

findGiniMarkers 115

Details

The function filterCombinations can be used to explore the effect of different parameter values.

Value

giotto object

Examples

```
filterGiotto(gobject)
```

findGiniMarkers

findGiniMarkers

Description

Identify marker genes for selected clusters based on gini detection and expression scores.

Usage

```
findGiniMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  min_expr_gini_score = 0.2,
  min_det_gini_score = 0.2,
  detection_threshold = 0,
  rank_score = 1,
  min_genes = 5
)
```

Arguments

```
gobject giotto object
expression_values
gene expression values to use
cluster_column clusters to use
subset_clusters
selection of clusters to compare
group_1 group 1 cluster IDs from cluster_column for pairwise comparison
```

Details

Detection of marker genes using the <a href="https://en.wikipedia.org/wiki/Gini_coefficientginic

- 1. calculate average expression per cluster
- 2. calculate detection fraction per cluster
- 3. calculate gini-coefficient for av. expression values over all clusters
- 4. calculate gini-coefficient for detection fractions over all clusters
- 5. convert gini-scores to rank scores
- 6. for each gene create combined score = detection rank x expression rank x expr gini-coefficient x detection gini-coefficient
- 7. for each gene sort on expression and detection rank and combined score

As a results "top gini" genes are genes that are very selectivily expressed in a specific cluster, however not always expressed in all cells of that cluster. In other words highly specific, but not necessarily sensitive at the single-cell level.

To perform differential expression between cluster groups you need to specificy cluster IDs to the parameters *group_1* and *group_2*.

Value

data.table with marker genes

Examples

findGiniMarkers(gobject)

Description

Identify marker genes for all clusters in a one vs all manner based on gini detection and expression scores.

Usage

```
findGiniMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank_score = 1,
  min_genes = 4,
  verbose = TRUE
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
subset_clusters
                  selection of clusters to compare
min_expr_gini_score
                  filter on minimum gini coefficient on expression
min_det_gini_score
                  filter on minimum gini coefficient on detection
detection_threshold
                  detection threshold for gene expression
rank_score
                 rank scores for both detection and expression to include
min_genes
                  minimum number of top genes to return
verbose
                  be verbose
```

Value

data.table with marker genes

See Also

findGiniMarkers

```
findGiniMarkers_one_vs_all(gobject)
```

118 findMarkers

findMarkers

findMarkers

Description

Identify marker genes for selected clusters.

Usage

```
findMarkers(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
 method = c("scran", "gini", "mast"),
  subset_clusters = NULL,
  group_1 = NULL,
 group_2 = NULL,
 min_expr_gini_score = 0.5,
 min_det_gini_score = 0.5,
 detection_threshold = 0,
 rank_score = 1,
 min\_genes = 4,
 group_1_name = NULL,
  group_2_name = NULL,
 adjust_columns = NULL,
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
method
                  method to use to detect differentially expressed genes
subset_clusters
                  selection of clusters to compare
                  group 1 cluster IDs from cluster_column for pairwise comparison
group_1
                  group 2 cluster IDs from cluster_column for pairwise comparison
group_2
min_expr_gini_score
                  gini: filter on minimum gini coefficient for expression
min_det_gini_score
                  gini: filter minimum gini coefficient for detection
{\tt detection\_threshold}
                  gini: detection threshold for gene expression
                  gini: rank scores to include
rank_score
                  minimum number of top genes to return (for gini)
min_genes
                  mast: custom name for group_1 clusters
group_1_name
```

findMarkers_one_vs_all 119

```
group_2_name mast: custom name for group_2 clusters

adjust_columns mast: column in pDataDT to adjust for (e.g. detection rate)

additional parameters for the findMarkers function in scran or zlm function in MAST
```

Details

Wrapper for all individual functions to detect marker genes for clusters.

Value

data.table with marker genes

See Also

findScranMarkers, findGiniMarkers and findMastMarkers

Examples

```
findMarkers(gobject)
```

Description

Identify marker genes for all clusters in a one vs all manner.

```
findMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  method = c("scran", "gini", "mast"),
  pval = 0.01,
  logFC = 0.5,
  min_genes = 10,
  min_expr_gini_score = 0.5,
  min_det_gini_score = 0.5,
  detection_threshold = 0,
  rank_score = 1,
  adjust_columns = NULL,
  verbose = TRUE,
)
```

Arguments

gobject giotto object

expression_values

gene expression values to use

cluster_column clusters to use

subset_clusters

selection of clusters to compare

method method to use to detect differentially expressed genes

pval scran & mast: filter on minimal p-value

logFC scan & mast: filter on logFC

min_genes minimum genes to keep per cluster, overrides pval and logFC

min_expr_gini_score

gini: filter on minimum gini coefficient for expression

min_det_gini_score

gini: filter minimum gini coefficient for detection

detection_threshold

gini: detection threshold for gene expression

rank_score gini: rank scores to include

adjust_columns mast: column in pDataDT to adjust for (e.g. detection rate)

verbose be verbose

... additional parameters for the findMarkers function in scran or zlm function in

MAST

Details

Wrapper for all one vs all functions to detect marker genes for clusters.

Value

data.table with marker genes

See Also

findScranMarkers_one_vs_all, findGiniMarkers_one_vs_all and findMastMarkers_one_vs_all

```
{\tt findMarkers\_one\_vs\_all(gobject)}
```

findMastMarkers 121

findMastMarkers

findMastMarkers

Description

Identify marker genes for selected clusters based on the MAST package.

Usage

```
findMastMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  group_1 = NULL,
  group_1_name = NULL,
  group_2 = NULL,
  group_2_name = NULL,
  adjust_columns = NULL,
  ...
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
                  group 1 cluster IDs from cluster_column for pairwise comparison
group_1
                  custom name for group_1 clusters
group_1_name
                  group 2 cluster IDs from cluster_column for pairwise comparison
group_2
group_2_name
                  custom name for group_2 clusters
adjust_columns column in pDataDT to adjust for (e.g. detection rate)
                  additional parameters for the zlm function in MAST
. . .
```

Details

This is a minimal convenience wrapper around the zlm from the MAST package to detect differentially expressed genes.

Value

data.table with marker genes

```
findMastMarkers(gobject)
```

Description

Identify marker genes for all clusters in a one vs all manner based on the MAST package.

Usage

```
findMastMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  adjust_columns = NULL,
  pval = 0.001,
  logFC = 1,
  min_genes = 10,
  verbose = TRUE,
  ...
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
subset_clusters
                  selection of clusters to compare
adjust_columns column in pDataDT to adjust for (e.g. detection rate)
                  filter on minimal p-value
pval
logFC
                  filter on logFC
min_genes
                  minimum genes to keep per cluster, overrides pval and logFC
                  be verbose
verbose
                  additional parameters for the zlm function in MAST
```

Value

data.table with marker genes

See Also

findMastMarkers

```
findMastMarkers_one_vs_all(gobject)
```

findScranMarkers 123

findScranMarkers findScranMarkers

Description

Identify marker genes for all or selected clusters based on scran's implementation of findMarkers.

Usage

```
findScranMarkers(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  group_1 = NULL,
  group_2 = NULL,
  ...
)
```

Arguments

Details

This is a minimal convenience wrapper around the findMarkers function from the scran package.

To perform differential expression between cluster groups you need to specificy cluster IDs to the parameters *group_1* and *group_2*.

Value

data.table with marker genes

```
findScranMarkers(gobject)
```

```
\label{lem:cone_vs_all} find Scran Markers\_one\_vs\_all \\ find Scran Markers\_one\_vs\_all
```

Description

Identify marker genes for all clusters in a one vs all manner based on scran's implementation of findMarkers.

Usage

```
findScranMarkers_one_vs_all(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  subset_clusters = NULL,
  pval = 0.01,
  logFC = 0.5,
  min_genes = 10,
  verbose = TRUE,
  ...
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  gene expression values to use
cluster_column clusters to use
subset_clusters
                  subset of clusters to use
pval
                  filter on minimal p-value
logFC
                  filter on logFC
                  minimum genes to keep per cluster, overrides pval and logFC
min_genes
verbose
                  be verbose
                  additional parameters for the findMarkers function in scran
```

Value

data.table with marker genes

See Also

findScranMarkers

```
findScranMarkers_one_vs_all(gobject)
```

find_grid_2D 125

find_grid_2D

 $find_grid_2D$

Description

find grid location in 2D

Usage

```
find_grid_2D(grid_DT, x_loc, y_loc)
```

 $find_grid_3D$

find_grid_3D

Description

find grid location in 3D

Usage

```
find_grid_3D(grid_DT, x_loc, y_loc, z_loc)
```

find_grid_x

find_grid_x

Description

find grid location on x-axis

Usage

```
find_grid_x(grid_DT, x_loc)
```

find_grid_y

 $find_grid_y$

Description

find grid location on y-axis

```
find_grid_y(grid_DT, y_loc)
```

fish_function2

find_grid_z

 $find_grid_z$

Description

find grid location on z-axis

Usage

```
find_grid_z(grid_DT, z_loc)
```

fish_function

fish_function

Description

perform fisher exact test

Usage

```
fish_function(x_to, x_from)
```

fish_function2

 $fish_function2$

Description

perform fisher exact test

```
fish_function2(A, B, C, D)
```

FSV_show 127

FSV_show FSV_show

Description

Visualize spatial varible genes caculated by spatial_DE

Usage

```
FSV_show(
  results,
  ms_results = NULL,
  size = c(4, 2, 1),
  color = c("blue", "green", "red"),
  sig_alpha = 0.5,
  unsig_alpha = 0.5
)
```

Arguments

results results caculated by spatial_DE

ms_results ms_results caculated by spatial_DE

size indicate different levels of qval

color indicate different SV features

sig_alpha transparency of significant genes

unsig_alpha transparency of unsignificant genes

Details

Description of parameters.

Value

nothing

```
FSV_show(results)
```

128 GenePattern_show

GenePattern_show

GenePattern_show

Description

Visualize genes distribution patterns calculated by spatial_AEH

Usage

```
GenePattern_show(
  gobject = NULL,
  AEH_results = NULL,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_alpha = 1,
  low_color = "blue",
  mid_color = "white",
  high_color = "red",
  midpoint = 0
)
```

Arguments

```
gobject
                  giotto object
                  results from spatial_AEH
AEH_results
sdimx
                  x axis of spatial locus
sdimy
                  y axis of spatial locus
point_size
                  size of points to indicate cells
point_alpha
                  transparency of points to indicate cells
low_color
                  color to indicate low score level
                  color to indicate middle score level
mid_color
high_color
                  color to indicate high score level
                  point to set mid_color
midpoint
```

Details

Description of parameters.

Value

nothing

```
GenePattern_show(gobject,AEH_results)
```

general_save_function 129

```
general_save_function general_save_function
```

Description

Function to automatically save plots to directory of interest

Usage

```
general_save_function(
   gobject,
   plot_object,
   save_dir = NULL,
   save_folder = NULL,
   save_name = NULL,
   default_save_name = "giotto_plot",
   save_format = c("png", "tiff", "pdf", "svg"),
   show_saved_plot = F,
   base_width = NULL,
   base_height = NULL,
   base_aspect_ratio = NULL,
   units = NULL,
   dpi = NULL,
   ...
)
```

Arguments

```
gobject
                  giotto object
                  non-ggplot object to plot
plot_object
save_dir
                  directory to save to
save_folder
                  folder in save_dir to save to
                  name of plot
save_name
save_format
                  format (e.g. png, tiff, pdf, ...)
show_saved_plot
                  load & display the saved plot
base_width
                  width
base_height
                  height
base_aspect_ratio
                  aspect ratio
units
                  units
dpi
                  Plot resolution
```

```
general_save_function(gobject)
```

get10Xmatrix

get10Xmatrix

Description

This function creates an expression matrix from a 10X structured folder

Usage

```
get10Xmatrix(path_to_data)
```

Arguments

```
path_to_data path to the 10X folder
```

Details

A typical 10X folder is named raw_feature_bc_matrix or raw_feature_bc_matrix. It has 3 files:

- barcodes
- features.tsv.gz
- matrix.mtx.gz

Value

expression matrix from 10X

Examples

```
get10Xmatrix(10Xmatrix)
```

```
{\it get Cell Proximity Gene Scores} \\ {\it get Cell Proximity Gene Scores}
```

Description

Compute cell-cell interaction enrichment (observed vs expected)

```
getCellProximityGeneScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "louvain_clus.1",
  selected_genes = NULL,
  expression_values = c("normalized", "scaled", "custom"),
  do_diff_test = TRUE,
  diff_test = c("t.test", "wilcox"),
  minimum_unique_cells = NA,
```

```
fold_change_addendum = 0.1,
  in_two_directions = TRUE,
  exclude_selected_cells_from_test = F,
  verbose = T
)
```

Arguments

```
gobject
                  giotto object
spatial_network_name
                  name of spatial network to use
cluster_column name of column to use for clusters
selected_genes selection of genes to perform calculations for
expression_values
                  expression values to use
do_diff_test
                  perform differential test
diff_test
                  which differential expression test
minimum_unique_cells
                  minimum number of cells needed to proceed
fold_change_addendum
                  constant to add when calculating log2 fold-change
in_two_directions
                  shows enrichment in both directions: cell1-cell2, cell2-cell1
exclude_selected_cells_from_test
                  exclude certain cells from test
verbose
                  verbose
```

Details

Function to calculate if genes are differentially expressed in cell types when they interact (according to physical proximity) with other cell types. The results data.table contains the following columns:

- genes: All or selected list of tested genes
- cell_expr_1: average gene expression in cell type 1 from unified_int cell-cell interaction
- cell_expr_2: average gene expression in cell type 2 from unified_int cell-cell interaction
- comb_expr: combined average gene expression in cell type 1 and 2 from unified_int cell-cell interaction
- all_cell_expr_1: average gene expression for all cells from cell type 1
- all_cell_expr_2: average gene expression for all cells from cell type 2
- all_comb_expr: combined average gene expression for all cells from cell type 1 and 2
- pval_1: p-value from test between interacting cells and all cells from cell type 1
- pval_2: p-value from test between interacting cells and all cells from cell type 2
- cell_type_1: first cell type of cell-cell interaction
- cell_type_2: second cell type of cell-cell interaction
- interaction: the cell-cell interaction, based on physical proximity
- nr_1: number of cell type 1 in the unified cell-cell interaction
- nr_2: number of cell type 2 in the unified cell-cell interaction

132 getClusterSimilarity

- all_nr_1: number of all cell type 1 in the whole dataset
- all_nr_2: number of all cell type 2 in the whole dataset
- diff_spat: difference between comb_expr and all_comb_expr
- diff_spat_1: difference between cell_expr_1 and all_cell_expr_1
- diff_spat_2: difference between cell_expr_1 and all_cell_expr_1
- log2fc_spat_1: fold-change of diff_spat_1
- log2fc_spat_2: fold-change of diff_spat_2
- log2fc_spat: fold-change of diff_spat
- type_int: type of interaction
- unified_int: interaction with alphabetically sorted cell type 1 and cell type 2
- unif_int_rank: 1 or 2
- fdr_1: fdr from test between interacting cells and all cells from cell type 1
- fdr_2: fdr from test between interacting cells and all cells from cell type 2

Value

Cell Proximity Gene scores (CPGscores) in data.table format

Examples

```
getCellProximityGeneScores(gobject)
```

```
getClusterSimilarity
```

Description

Creates data.table with pairwise correlation scores between each cluster.

Usage

```
getClusterSimilarity(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman")
)
```

Arguments

```
gobject giotto object
expression_values
expression values to use
cluster_column name of column to use for clusters
cor correlation score to calculate distance
```

getDendrogramSplits 133

Details

Creates data.table with pairwise correlation scores between each cluster and the group size (# of cells) for each cluster. This information can be used together with mergeClusters to combine very similar or small clusters into bigger clusters.

Value

data.table

Examples

```
getClusterSimilarity(gobject)
```

```
getDendrogramSplits getDendrogramSplits
```

Description

Split dendrogram at each node and keep the leave (label) information..

Usage

```
getDendrogramSplits(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  h = NULL,
  h_color = "red",
  show_dend = TRUE,
  verbose = TRUE
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
distance
                  distance method to use for hierarchical clustering
                  height of horizontal lines to plot
h
h_color
                  color of horizontal lines
                  show dendrogram
show_dend
verbose
                  be verbose
```

Details

Creates a data.table with three columns and each row represents a node in the dendrogram. For each node the height of the node is given together with the two subdendrograms. This information can be used to determine in a hierarchical manner differentially expressed marker genes at each node.

Value

data.table object

Examples

```
getDendrogramSplits(gobject)
```

getDistinctColors

getDistinctColors

Description

Returns a number of distint colors based on the RGB scale

Usage

```
getDistinctColors(n)
```

Arguments

n

number of colors wanted

Value

number of distinct colors

 ${\tt getGeneToGeneScores}$

getGeneToGeneScores

Description

Compute gene-gene enrichment scores.

```
getGeneToGeneScores(
   CPGscore,
   selected_genes = NULL,
   specific_genes_1 = NULL,
   specific_genes_2 = NULL,
   min_cells = 5,
   min_fdr = 0.05,
   min_spat_diff = 0.2,
   min_log2_fc = 0.5,
```

```
direction = c("both", "up", "down"),
fold_change_addendum = 0.1,
  verbose = TRUE
)
```

Arguments

```
CPGscore
                  CPGscore, output from getCellProximityGeneScores()
selected_genes select subset of genes
specific_genes_1
                  specific source genes (see details)
specific_genes_2
                  specific target genes (see details)
                  min number of cells threshold
min_cells
                  spatial difference threshold
min_spat_diff
min_log2_fc
                  log2 fold-change threshold
direction
                  up or downregulation or both
fold_change_addendum
                  constant to add when calculating log2 fold-change
verbose
                  verbose
min_pval
                  p-value threshold
```

Details

This converts the single gene cell proximity scores into pairwise combinations of genes, which allows you to determine if 2 genes are differentially expressed in interacting cell types.

Value

Gene to gene scores in data.table format

Examples

```
getGeneToGeneScores(CPGscore)
```

Description

creates unified cell-cell interaction names

Usage

```
get_cell_to_cell_sorted_name_conversion(all_cell_types)
```

```
get_cell_to_cell_sorted_name_conversion()
```

```
{\it get\_interaction\_gene\_enrichment} \\ {\it get\_interaction\_gene\_enrichment}
```

Description

Computes gene enrichment between all interactions

Usage

```
get_interaction_gene_enrichment(
  spatial_network,
  unified_int_col = "unified_int",
  source_col = "source_clus",
  source_IDs = "from",
  neighb_col = "neighb_clus",
  neighb_IDs = "to",
  expression_matrix,
  cell_annotation,
  annotation_ID = "uniq_ID",
  cell_type_col,
  do_diff_test = T,
  diff_test = c("t.test", "wilcox"),
  minimum_unique_cells = NA,
  exclude_selected_cells_from_test = T,
  verbose = T
)
```

Examples

```
get_interaction_gene_enrichment()
```

```
{\tt get\_specific\_interaction\_gene\_enrichment} \\ {\tt get\_specific\_interaction\_gene\_enrichment}
```

Description

Computes gene enrichment between specified interaction

```
get_specific_interaction_gene_enrichment(
  sub_spatial_network,
  source_col = "source_clus",
  source_IDs = "from",
  neighb_col = "neighb_clus",
  neighb_IDs = "to",
  expression_matrix,
```

ggplot_save_function 137

```
interaction_name = "to_specify",
  cell_annotation,
  annotation_ID = "uniq_ID",
  cell_type_col,
  do_diff_test = T,
  diff_test = c("t.test", "wilcox"),
  minimum_unique_cells = NA,
  exclude_selected_cells_from_test = T
)
```

Examples

get_specific_interaction_gene_enrichment()

```
ggplot_save_function ggplot_save_function
```

Description

Function to automatically save plots to directory of interest

Usage

```
ggplot_save_function(
  gobject,
  plot_object,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  default_save_name = "giotto_plot",
  save_format = NULL,
  show_saved_plot = F,
  ncol = 1,
  nrow = 1,
  scale = 1,
  base_width = NULL,
  base_height = NULL,
  base_aspect_ratio = NULL,
  units = NULL,
  dpi = NULL,
  limitsize = TRUE,
)
```

Arguments

```
gobject giotto object
plot_object ggplot object to plot
save_dir directory to save to
save_folder folder in save_dir to save to
```

138 giotto-class

save_name name of plot

save_format (e.g. png, tiff, pdf, ...)

show_saved_plot

load & display the saved plot

ncol number of columns nrow number of rows

scale scale
base_width width
base_height height
base_aspect_ratio

aspect ratio

units units

dpi Plot resolution

limitsize When TRUE (the default), ggsave will not save images larger than 50x50 inches,

to prevent the common error of specifying dimensions in pixels.

See Also

```
cowplot::save_plot
```

Examples

```
ggplot_save_function(gobject)
```

giotto-class S4 giotto Class

Description

Framework of giotto object to store and work with spatial expression data

Slots

```
raw_exprs raw expression counts

norm_expr normalized expression counts

norm_scaled_expr normalized and scaled expression counts

custom_expr custom normalized counts

spatial_locs spatial location coordinates for cells

cell_metadata metadata for cells

gene_metadata metadata for genes

cell_ID unique cell IDs

gene_ID unique gene IDs

spatial_network spatial network in data.table/data.frame format

spatial_grid spatial grid in data.table/data.frame format

dimension_reduction slot to save dimension reduction coordinates
```

heatmSpatialCorGenes 139

```
nn_network nearest neighbor network in igraph format parameters slot to save parameters that have been used instructions slot for global function instructions offset_file offset file used to stitch together image fields OS_platform Operating System to run Giotto analysis on
```

heatmSpatialCorGenes heatmSpatialCorGenes

Description

Create heatmap of spatially correlated genes

Usage

```
heatmSpatialCorGenes(
  gobject,
  spatCorObject,
  use_clus_name = NULL,
  show_cluster_annot = TRUE,
  show_row_dend = T,
  show_column_dend = F,
  show_row_names = F,
  show_column_names = F,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "heatmSpatialCorGenes",
  ...
)
```

Arguments

```
gobject
                 giotto object
                 spatial correlation object
spatCorObject
                 name of clusters to visualize (from clusterSpatialCorGenes())
use_clus_name
show_cluster_annot
                 show cluster annotation on top of heatmap
show_row_dend
                 show row dendrogram
show_column_dend
                 show column dendrogram
show_row_names show row names
show_column_names
                 show column names
show_plot
                 show plot
return_plot
                 return ggplot object
```

Value

Heatmap generated by ComplexHeatmap

Examples

```
heatmSpatialCorGenes(gobject)
```

```
hyperGeometricEnrich hyperGeometricEnrich
```

Description

Function to calculate gene signature enrichment scores per spatial position using a hypergeometric test.

Usage

```
hyperGeometricEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

Arguments

Details

The enrichment score is calculated based on the p-value from the hypergeometric test, -log10(p-value).

kmeans_binarize 141

Value

data.table with enrichment results

Examples

hyperGeometricEnrich(gobject)

kmeans_binarize

kmeans_binarize

Description

create binarized scores using kmeans

Usage

```
kmeans_binarize(x, nstart = 3, iter.max = 10)
```

loadHMRF

loadHMRF

Description

load previous HMRF

Usage

```
loadHMRF(
  name_used = "test",
  output_folder_used,
  k_used = 10,
  betas_used,
  python_path_used
)
```

Arguments

```
name_used name of HMRF that was run
output_folder_used
output folder that was used
k_used number of HMRF domains that was tested
betas_used betas that were tested
python_path_used
python path that was used
```

Details

Description of HMRF parameters ...

142 makeSignMatrixRank

Value

reloads a previous ran HMRF from doHRMF

Examples

loadHMRF(gobject)

make Sign Matrix PAGE

make Sign Matrix PAGE

Description

Function to convert a list of signature genes (e.g. for cell types or processes) into a binary matrix format that can be used with the PAGE enrichment option.

Usage

```
makeSignMatrixPAGE(sign_names, sign_list)
```

Arguments

sign_names vector with names for each provided gene signature

sign_list list of genes (signature)

Value

matrix

See Also

PAGEEnrich

Examples

makeSignMatrixPAGE()

 ${\tt make Sign Matrix Rank}$

makeSignMatrixRank

Description

Function to convert a single-cell count matrix and a corresponding single-cell cluster vector into a rank matrix that can be used with the Rank enrichment option.

```
makeSignMatrixRank(sc_matrix, sc_cluster_ids, gobject = NULL)
```

Arguments

```
sc_matrix matrix of single-cell RNAseq expression data
sc_cluster_ids vector of cluster ids
gobject if giotto object is given then only genes present in both datasets will be considered
```

Value

matrix

See Also

rankEnrich

Examples

```
makeSignMatrixRank()
```

```
make\_simulated\_network \\ make\_simulated\_network
```

Description

Simulate random network.

Usage

```
make_simulated_network(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column,
  number_of_simulations = 100
)
```

```
{\tt make\_simulated\_network(gobject)}
```

144 mergeClusters

mergeClusters

mergeClusters

Description

Merge selected clusters based on pairwise correlation scores and size of cluster.

Usage

```
mergeClusters(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  new_cluster_name = "merged_cluster",
  min_cor_score = 0.8,
  max_group_size = 20,
  force_min_group_size = 10,
  return_gobject = TRUE,
  verbose = TRUE
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
new_cluster_name
                  new name for merged clusters
min_cor_score min correlation score to merge pairwise clusters
max_group_size max cluster size that can be merged
force_min_group_size
                  size of clusters that will be merged with their most similar neighbor(s)
return_gobject return giotto object
verbose
                  be verbose
```

Details

Merge selected clusters based on pairwise correlation scores and size of cluster. To avoid large clusters to merge the max_group_size can be lowered. Small clusters can be forcibly merged with their most similar pairwise cluster by adjusting the force_min_group_size parameter. Clusters smaller than this value will be merged independent on the provided min_cor_score value.

A giotto object is returned by default, if FALSE then the merging vector will be returned.

Value

Giotto object

mygini_fun 145

Examples

mergeClusters(gobject)

mygini_fun

mygini_fun

Description

calculate gini coefficient

Usage

```
mygini_fun(x, weights = rep(1, length(x)))
```

Value

gini coefficient

nnDT_to_kNN

 $nnDT_to_kNN$

Description

Convert a nearest network data.table to a kNN object

Usage

```
nnDT_to_kNN(nnDT)
```

Arguments

nnDT

nearest neighbor network in data.table format

Value

kNN object

146 normalizeGiotto

node_clusters

node_clusters

Description

Merge selected clusters based on pairwise correlation scores and size of cluster.

Usage

```
node_clusters(hclus_obj, verbose = TRUE)
```

Arguments

```
hclus_obj hclus object
verbose be verbose
```

Value

list of splitted dendrogram nodes from high to low node height

Examples

```
node_clusters(hclus_obj)
```

normalizeGiotto

normalizeGiotto

Description

normalize and/or scale expresion values of Giotto object

```
normalizeGiotto(
  gobject,
  norm_methods = c("standard", "osmFISH"),
  library_size_norm = TRUE,
  scalefactor = 6000,
  log_norm = TRUE,
  logbase = 2,
  scale_genes = T,
  scale_cells = T,
  scale_order = c("first_genes", "first_cells"),
  verbose = F
)
```

OR_function2 147

Arguments

gobject giotto object

norm_methods normalization method to use

library_size_norm

normalize cells by library size

scalefactor scale factor to use after library size normalization

log_norm transform values to log-scale

logbase log base to use to log normalize expression values

scale_genes z-score genes over all cells scale_cells z-score cells over all genes scale_order order to scale genes and cells

verbose be verbose

Details

Currently there are two 'methods' to normalize your raw counts data.

A. The standard method follows the standard protocol which can be adjusted using the provided parameters and follows the following order:

- 1. Data normalization for total library size and scaling by a custom scale-factor.
- 2. Log transformation of data.
- 3. Z-scoring of data by genes and/or cells.
- B. The normalization method as provided by the osmFISH paper is also implemented:
 - 1. First normalize genes, for each gene divide the counts by the total gene count and multiply by the total number of genes.
 - 2. Next normalize cells, for each cell divide the normalized gene counts by the total counts per cell and multiply by the total number of cells.

This data will be saved in the Giotto slot for custom expression.

Value

giotto object

Examples

normalizeGiotto(gobject)

OR_function2 OR_function2

Description

calculate odds-ratio

```
OR_function2(A, B, C, D)
```

148 PAGEEnrich

PAGEEnrich

PAGEEnrich

Description

Function to calculate gene signature enrichment scores per spatial position using PAGE.

Usage

```
PAGEEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

Arguments

Details

sign_matrix: a binary matrix with genes as row names and cell-types as column names. Alternatively a list of signature genes can be provided to makeSignMatrixPAGE, which will create the matrix for you.

The enrichment Z score is calculated by using method (PAGE) from Kim SY et al., BMC bioinformatics, 2005 as $Z=((Sm\ mu)*m^(1/2))/delta$. For each gene in each spot, mu is the fold change values versus the mean expression and delta is the standard deviation. Sm is the mean fold change value of a specific marker gene set and m is the size of a given marker gene set.

Value

data.table with enrichment results

See Also

makeSignMatrixPAGE

Examples

```
PAGEEnrich(gobject)
```

pDataDT 149

pDataDT

pDataDT

Description

show cell metadata

Usage

```
pDataDT(gobject)
```

Arguments

gobject

giotto object

Value

data.table with cell metadata

Examples

```
pDataDT(gobject)
```

plotCPGscores

plotCPGscores

Description

Create heatmap from cell-cell proximity scores

```
plotCPGscores(
   CPGscores,
   selected_interactions = NULL,
   selected_genes = NULL,
   detail_plot = T,
   simple_plot = F,
   simple_plot_facet = c("interaction", "genes"),
   facet_scales = "fixed",
   facet_ncol = length(selected_genes),
   facet_nrow = length(selected_interactions),
   show_plot = F
)
```

150 plotGTGscores

Arguments

```
CPGscores
                  CPGscores, output from getCellProximityGeneScores()
selected_interactions
                  interactions to show
selected_genes genes to show
detail_plot
                  show detailed info in both interacting cell types
simple_plot
                  show a simplified plot
simple_plot_facet
                  facet on interactions or genes with simple plot
                  ggplot facet scales paramter
facet_scales
facet_ncol
                  ggplot facet ncol parameter
facet_nrow
                  ggplot facet nrow parameter
show_plot
                  show plot
```

Details

Give more details ...

Value

ggplot barplot

Examples

```
plotCPGscores(CPGscores)
```

plotGTGscores

plotGTGscores

Description

Create heatmap from cell-cell proximity scores

```
plotGTGscores(
   gobject,
   GTGscore,
   selected_interactions = NULL,
   selected_gene_to_gene = NULL,
   detail_plot = T,
   simple_plot = F,
   simple_plot_facet = c("interaction", "genes"),
   facet_scales = "fixed",
   facet_ncol = length(selected_gene_to_gene),
   facet_nrow = length(selected_interactions),
   colors = c("blue", "red"),
   show_plot = NA,
   return_plot = NA,
```

plotGTGscores 151

```
save_plot = NA,
save_param = list(),
default_save_name = "plotGTGscores"
)
```

Arguments

gobject giotto object GTGscore GTGscore, output from getGeneToGeneScores() selected_interactions interactions to show show detailed info in both interacting cell types detail_plot show a simplified plot simple_plot simple_plot_facet facet on interactions or genes with simple plot facet_scales ggplot facet scales paramter ggplot facet ncol parameter facet_ncol facet_nrow ggplot facet nrow parameter colors vector with 2 colors to represent respectively all and selected cells show_plot show plots return_plot return ggplot object directly save the plot [boolean] save_plot list of saving parameters from all_plots_save_function save_param ${\tt default_save_name}$ default save name for saving, don't change, change save_name in save_param

Details

Give more details ...

Value

ggplot barplot

Examples

```
plotGTGscores(GTGscore)
```

selected_genes genes to show

152 plotHeatmap

plotHeatmap

plotHeatmap

Description

Creates heatmap for genes and clusters.

Usage

```
plotHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column = NULL,
  cluster_order = c("size", "correlation", "custom"),
  cluster_custom_order = NULL,
  cluster_color_code = NULL,
  cluster_cor_method = "pearson",
  cluster_hclust_method = "ward.D"
  gene_order = c("custom", "correlation"),
  gene_custom_order = NULL,
  gene_cor_method = "pearson",
  gene_hclust_method = "complete",
  show_values = c("rescaled", "z-scaled", "original"),
  size_vertical_lines = 1.1,
  gradient_colors = c("blue", "yellow", "red"),
  gene_label_selection = NULL,
  axis_text_y_size = NULL,
  legend_nrows = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "plotHeatmap"
)
```

```
gobject giotto object
expression_values
expression values to use
genes genes to use
cluster_column name of column to use for clusters
cluster_order method to determine cluster order
cluster_custom_order
custom order for clusters
cluster_color_code
color code for clusters
cluster_cor_method
method for cluster correlation
```

plotHeatmap 153

```
cluster_hclust_method
                 method for hierarchical clustering of clusters
gene_order
                 method to determine gene order
gene_custom_order
                 custom order for genes
gene_cor_method
                 method for gene correlation
gene_hclust_method
                 method for hierarchical clustering of genes
show_values
                 which values to show on heatmap
size_vertical_lines
                 sizes for vertical lines
gradient_colors
                 colors for heatmap gradient
gene_label_selection
                 subset of genes to show on y-axis
axis_text_y_size
                 size for y-axis text
legend_nrows
                 number of rows for the cluster legend
show_plot
                 show plot
return_plot
                 return ggplot object
                 directly save the plot [boolean]
save_plot
                 list of saving parameters from all_plots_save_function
save_param
default_save_name
                 default save name
```

Details

If you want to display many genes there are 2 ways to proceed:

- 1. set axis_text_y_size to a really small value and show all genes
- 2. provide a subset of genes to display to gene_label_selection

Value

ggplot

Examples

```
plotHeatmap(gobject)
```

154 plotly_axis_scale_3D

```
plotly_axis_scale_2D plotly_axis_scale_2D
```

Description

adjust the axis scale in 3D plotly plot

Usage

```
plotly_axis_scale_2D(
  cell_locations,
  sdimx = NULL,
  sdimy = NULL,
  mode = c("cube", "real", "custom"),
  custom_ratio = NULL
)
```

Arguments

```
cell_locations spatial_loc in giotto object sdimx x axis of cell spatial location sdimy y axis of cell spatial location mode axis adjustment mode custom_ratio set the ratio artificially
```

Value

edges in spatial grid as data.table()

Examples

```
plotly_axis_scale_2D(gobject)
```

```
plotly_axis_scale_3D plotly_axis_scale_3D
```

Description

adjust the axis scale in 3D plotly plot

```
plotly_axis_scale_3D(
  cell_locations,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  mode = c("cube", "real", "custom"),
  custom_ratio = NULL
)
```

plotly_grid 155

Arguments

Value

edges in spatial grid as data.table()

Examples

```
plotly_axis_scale_3D(gobject)
```

plotly_grid

plotly_grid

Description

provide grid segment to draw in plot_ly()

Usage

```
plotly_grid(
   spatial_grid,
   x_start = "x_start",
   y_start = "y_start",
   x_end = "x_end",
   y_end = "y_end"
)
```

Arguments

```
spatial_grid spatial_grid in giotto object
```

Value

edges in spatial grid as data.table()

Examples

```
plotly_grid(gobject)
```

plotly_network

plotly_network

Description

provide network segment to draw in 3D plot_ly()

Usage

```
plotly_network(
  network,
  x = "sdimx_begin",
  y = "sdimy_begin",
  z = "sdimz_begin",
  x_end = "sdimx_end",
  y_end = "sdimy_end",
  z_end = "sdimz_end"
```

Arguments

gobject network in giotto object

Value

edges in network as data.table()

Examples

```
plotly_network(gobject)
```

```
plotMetaDataCellsHeatmap
```

plotMetaDataCellsHeatmap

Description

Creates heatmap for numeric cell metadata within aggregated clusters.

```
plotMetaDataCellsHeatmap(
  gobject,
  metadata_cols = NULL,
  spat_enr_names = NULL,
  value_cols = NULL,
  first_meta_col = NULL,
  second_meta_col = NULL,
  show_values = c("zscores", "original", "zscores_rescaled"),
  custom_cluster_order = NULL,
```

```
clus_cor_method = "pearson",
 clus_cluster_method = "complete",
  custom_values_order = NULL,
 values_cor_method = "pearson",
  values_cluster_method = "complete",
 midpoint = 0,
 x_{text_size} = 8,
  x_{text_angle} = 45,
 y_text_size = 8,
  strip_text_size = 8,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
 save_param = list(),
 default_save_name = "plotMetaDataCellsHeatmap"
)
```

```
gobject
                  giotto object
                  annotation columns found in pDataDT(gobject)
metadata_cols
spat_enr_names spatial enrichment results to include
value_cols
                  value columns to use
first_meta_col if more than 1 metadata column, select the x-axis factor
second_meta_col
                  if more than 1 metadata column, select the facetting factor
show_values
                  which values to show on heatmap
custom_cluster_order
                  custom cluster order (default = NULL)
clus_cor_method
                  correlation method for clusters
clus\_cluster\_method
                  hierarchical cluster method for the clusters
                  midpoint of show_values
midpoint
                  size of x-axis text
x_text_size
x_text_angle
                  angle of x-axis text
y_text_size
                  size of y-axis text
strip_text_size
                  size of strip text
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
custom_gene_order
                  custom gene order (default = NULL)
```

Details

Creates heatmap for the average values of selected value columns in the different annotation groups.

Value

ggplot or data.table

See Also

plotMetaDataHeatmap for gene expression instead of numeric cell annotation data.

Examples

```
plotMetaDataCellsHeatmap(gobject)
```

```
plotMetaDataHeatmap \hspace{3cm} plotMetaDataHeatmap
```

Description

Creates heatmap for genes within aggregated clusters.

```
plotMetaDataHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  metadata_cols = NULL,
  selected_genes = NULL,
  first_meta_col = NULL,
  second_meta_col = NULL,
  show_values = c("zscores", "original", "zscores_rescaled"),
  custom_cluster_order = NULL,
  clus_cor_method = "pearson",
  clus_cluster_method = "complete",
  custom_gene_order = NULL,
  gene_cor_method = "pearson",
  gene_cluster_method = "complete",
  midpoint = 0,
  x_{text_size} = 10,
  x_{text_angle} = 45,
  y_{text_size} = 10,
  strip_text_size = 8,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
```

plotMetaDataHeatmap 159

```
save_param = list(),
default_save_name = "plotMetaDataHeatmap"
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
                 annotation columns found in pDataDT(gobject)
metadata_cols
selected_genes subset of genes to use
first_meta_col if more than 1 metadata column, select the x-axis factor
second_meta_col
                  if more than 1 metadata column, select the facetting factor
show_values
                  which values to show on heatmap
custom_cluster_order
                  custom cluster order (default = NULL)
clus_cor_method
                  correlation method for clusters
clus_cluster_method
                  hierarchical cluster method for the clusters
custom_gene_order
                  custom gene order (default = NULL)
gene_cor_method
                  correlation method for genes
gene_cluster_method
                  hierarchical cluster method for the genes
                  midpoint of show_values
midpoint
                  size of x-axis text
x_text_size
x_text_angle
                  angle of x-axis text
                  size of y-axis text
y_text_size
strip_text_size
                  size of strip text
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name
```

Details

Creates heatmap for the average expression of selected genes in the different annotation/cluster groups. Calculation of cluster or gene order is done on the provided expression values, but visualization is by default on the z-scores. Other options are the original values or z-scores rescaled per gene (-1 to 1).

160 plotPCA

Value

ggplot or data.table

See Also

plotMetaDataCellsHeatmap for numeric cell annotation instead of gene expression.

Examples

```
plotMetaDataHeatmap(gobject)
```

plotPCA

plotPCA

Description

Short wrapper for PCA visualization

giotto object

Usage

```
plotPCA(gobject, dim_reduction_name = "pca", default_save_name = "PCA", ...)
```

Arguments

gobject

```
dim_reduction_name
                 dimension reduction name
default_save_name
                 default save name for saving, don't change, change save_name in save_param
                 create multiple plots based on cell annotation column
groub_by
group_by_subset
                 subset the group_by factor column
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                 color for cells (see details)
color_as_factor
                 convert color column to factor
cell_color_code
                 named vector with colors
cell_color_gradient
                 vector with 3 colors for numeric data
```

gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size size of not selected cells show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points size of labels label_size label_fontface font of labels edge_alpha column to use for alpha of the edges point with border or not (border or no_border) point_shape point_size size of point (cell) point_border_col color of border around points point_border_stroke stroke size of border around points show_legend show legend title for plot, defaults to cell_color parameter title legend_text size of legend text legend_symbol_size size of legend symbols background_color color of plot background axis_text size of axis text axis_title size of axis title cow_n_col cowplot param: how many columns cow_rel_h cowplot param: relative height cow_rel_w cowplot param: relative width cow_align cowplot param: how to align show_plot show plot return_plot return ggplot object save_plot directly save the plot [boolean] list of saving parameters from all_plots_save_function save_param

plotPCA_2D

Details

Description of parameters, see dimPlot2D. For 3D plots see plotPCA_3D

Value

ggplot

Examples

```
plotPCA(gobject)
```

plotPCA_2D

plotPCA_2D

Description

Short wrapper for PCA visualization

Usage

```
plotPCA_2D(
  gobject,
  dim_reduction_name = "pca",
  default_save_name = "PCA_2D",
   ...
)
```

```
gobject
                 giotto object
dim_reduction_name
                 dimension reduction name
default_save_name
                 default save name for saving, don't change, change save_name in save_param
groub_by
                 create multiple plots based on cell annotation column
group_by_subset
                 subset the group_by factor column
dim1_to_use
                 dimension to use on x-axis
dim2\_to\_use
                 dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                 color for cells (see details)
color_as_factor
                 convert color column to factor
```

plotPCA_2D 163

cell_color_code named vector with colors cell_color_gradient vector with 3 colors for numeric data gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select_cells select subset of cells based on cell IDs show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size size of not selected cells show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points label_size size of labels label_fontface font of labels edge_alpha column to use for alpha of the edges point_shape point with border or not (border or no_border) point_size size of point (cell) point_border_col color of border around points point_border_stroke stroke size of border around points title title for plot, defaults to cell_color parameter show_legend show legend legend_text size of legend text legend_symbol_size size of legend symbols background_color color of plot background axis_text size of axis text size of axis title axis_title cowplot param: how many columns cow_n_col cowplot param: relative height cow_rel_h cowplot param: relative width cow_rel_w cowplot param: how to align cow_align show_plot show plot return ggplot object return_plot directly save the plot [boolean] save_plot list of saving parameters from all_plots_save_function save_param

plotPCA_3D

Details

Description of parameters, see dimPlot2D. For 3D plots see plotPCA_3D

Value

ggplot

Examples

```
plotPCA_2D(gobject)
```

plotPCA_3D

plotPCA_3D

Description

Visualize cells according to 3D PCA dimension reduction

Usage

```
plotPCA_3D(
  gobject,
  dim_reduction_name = "pca",
  default_save_name = "PCA_3D",
   ...
)
```

```
gobject
                 giotto object
dim_reduction_name
                 pca dimension reduction name
default_save_name
                 default save name for saving, ideally change save_name in save_param
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
                 dimension to use on z-axis
dim3_to_use
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                 color for cells (see details)
color_as_factor
                 convert color column to factor
cell_color_code
                 named vector with colors
select_cell_groups
                 select subset of cells/clusters based on cell_color parameter
```

plotTSNE 165

```
select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
edge_alpha
                  column to use for alpha of the edges
                  size of point (cell)
point_size
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
```

directly save the plot [boolean]

list of saving parameters from all_plots_save_function

Details

Description of parameters.

Value

plotly

save_plot
save_param

Examples

plotPCA_3D(gobject)

plotTSNE plotTSNE

Description

Short wrapper for tSNE visualization

```
plotTSNE(gobject, dim_reduction_name = "tsne", default_save_name = "tSNE", ...)
```

166 plotTSNE

Arguments

giotto object gobject dim_reduction_name dimension reduction name default_save_name default save name for saving, don't change, change save_name in save_param create multiple plots based on cell annotation column groub_by group_by_subset subset the group_by factor column dim1_to_use dimension to use on x-axis dim2_to_use dimension to use on y-axis spat_enr_names names of spatial enrichment results to include show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) network_name name of NN network to use, if show_NN_network = TRUE cell_color color for cells (see details) color_as_factor convert color column to factor cell_color_code named vector with colors cell_color_gradient vector with 3 colors for numeric data gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size size of not selected cells show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points label_size size of labels label_fontface font of labels edge_alpha column to use for alpha of the edges

plotTSNE_2D 167

point_size size of point (cell)

point_border_col

color of border around points

 ${\tt point_border_stroke}$

stroke size of border around points

title title for plot, defaults to cell_color parameter

show_legend show legend

legend_text size of legend text

 ${\tt legend_symbol_size}$

size of legend symbols

background_color

color of plot background

axis_text size of axis text
axis_title size of axis title

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function

Details

Description of parameters, see dimPlot2D. For 3D plots see plotTSNE_3D

Value

ggplot

Examples

plotTSNE(gobject)

plotTSNE_2D plotTSNE_2D

Description

Short wrapper for tSNE visualization

plotTSNE_2D

```
Usage
```

```
plotTSNE_2D(
      gobject,
      dim_reduction_name = "tsne",
      default_save_name = "tSNE_2D",
Arguments
    gobject
                      giotto object
    dim_reduction_name
                      dimension reduction name
    default_save_name
                      default save name for saving, don't change, change save_name in save_param
                      create multiple plots based on cell annotation column
    groub_by
    group_by_subset
                      subset the group_by factor column
    dim1_to_use
                      dimension to use on x-axis
                      dimension to use on y-axis
    dim2_to_use
    spat_enr_names names of spatial enrichment results to include
    show_NN_network
                      show underlying NN network
    nn_network_to_use
                      type of NN network to use (kNN vs sNN)
                      name of NN network to use, if show_NN_network = TRUE
    network_name
                      color for cells (see details)
    cell_color
    color_as_factor
                      convert color column to factor
    cell_color_code
                      named vector with colors
    cell_color_gradient
                      vector with 3 colors for numeric data
    gradient_midpoint
                      midpoint for color gradient
    gradient_limits
                      vector with lower and upper limits
    select_cell_groups
                      select subset of cells/clusters based on cell color parameter
    select_cells
                      select subset of cells based on cell IDs
    show_other_cells
                      display not selected cells
    other_cell_color
                      color of not selected cells
    other_point_size
                      size of not selected cells
    show_cluster_center
```

plot center of selected clusters

plotTSNE_2D 169

```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
edge_alpha
                  column to use for alpha of the edges
point_shape
                  point with border or not (border or no_border)
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
title
                  title for plot, defaults to cell_color parameter
show_legend
                  show legend
legend_text
                  size of legend text
legend_symbol_size
                  size of legend symbols
background_color
                  color of plot background
                  size of axis text
axis_text
axis_title
                  size of axis title
                  cowplot param: how many columns
cow_n_col
cow_rel_h
                  cowplot param: relative height
                  cowplot param: relative width
cow_rel_w
cow_align
                  cowplot param: how to align
                  show plot
show_plot
```

Details

Description of parameters, see dimPlot2D. For 3D plots see plotTSNE_3D

list of saving parameters from all_plots_save_function

directly save the plot [boolean]

return ggplot object

Value

ggplot

return_plot

save_plot
save_param

Examples

```
plotTSNE_2D(gobject)
```

170 plotTSNE_3D

plotTSNE_3D

plotTSNE_3D

Description

Visualize cells according to dimension reduction coordinates

Usage

```
plotTSNE_3D(
  gobject,
  dim_reduction_name = "tsne",
  default_save_name = "TSNE_3D",
   ...
)
```

```
gobject
                  giotto object
dim_reduction_name
                  tsne dimension reduction name
default_save_name
                  default save name for saving, don't change, change save_name in save_param
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
dim3_to_use
                  dimension to use on z-axis
show_NN_network
                  show underlying NN network
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
                  name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  size of not selected cells
show_cluster_center
                  plot center of selected clusters
```

plotUMAP 171

list of saving parameters from all_plots_save_function

```
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
                  size of labels
label_size
edge_alpha
                  column to use for alpha of the edges
point_size
                  size of point (cell)
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
```

Details

Description of parameters.

Value

plotly

save_param

Examples

```
plotTSNE_3D(gobject)
```

plotUMAP plotUMAP

Description

Short wrapper for UMAP visualization

Usage

```
plotUMAP(gobject, dim_reduction_name = "umap", default_save_name = "UMAP", ...)
```

172 plotUMAP

dim2_to_use dimension to use on y-axis spat_enr_names names of spatial enrichment results to include show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) network_name name of NN network to use, if show_NN_network = TRUE cell_color color for cells (see details) color_as_factor convert color column to factor cell_color_code named vector with colors cell_color_gradient vector with 3 colors for numeric data gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size size of not selected cells show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points label_size size of labels label_fontface font of labels edge_alpha column to use for alpha of the edges point with border or not (border or no_border) point_shape point_size size of point (cell) point_border_col color of border around points point_border_stroke stroke size of border around points title for plot, defaults to cell_color parameter title show legend show_legend legend_text size of legend text legend_symbol_size size of legend symbols

plotUMAP_2D 173

```
background_color
```

```
color of plot background
```

axis_text	size of axis text
axis_title	size of axis title
cow_n_col	cowplot param: how many columns
cow_rel_h	cowplot param: relative height
cow_rel_w	cowplot param: relative width
cow_align	cowplot param: how to align
show_plot	show plot
return_plot	return ggplot object
save_plot	directly save the plot [boolean]
save_param	list of saving parameters from all_plots_save_function

Details

Description of parameters, see dimPlot2D. For 3D plots see plotUMAP_3D

Value

ggplot

Examples

```
plotUMAP(gobject)
```

plotUMAP_2D

 $plotUMAP_2D$

Description

Short wrapper for UMAP visualization

```
plotUMAP_2D(
  gobject,
  dim_reduction_name = "umap",
  default_save_name = "UMAP_2D",
   ...
)
```

174 plotUMAP_2D

Arguments

giotto object gobject dim_reduction_name dimension reduction name default_save_name default save name for saving, don't change, change save_name in save_param create multiple plots based on cell annotation column groub_by group_by_subset subset the group_by factor column dim1_to_use dimension to use on x-axis dim2_to_use dimension to use on y-axis spat_enr_names names of spatial enrichment results to include show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) network_name name of NN network to use, if show_NN_network = TRUE cell_color color for cells (see details) color_as_factor convert color column to factor cell_color_code named vector with colors cell_color_gradient vector with 3 colors for numeric data gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select_cells select subset of cells based on cell IDs show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size size of not selected cells show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points label_size size of labels label_fontface font of labels edge_alpha column to use for alpha of the edges

plotUMAP_3D 175

point_shape point with border or not (border or no_border)

point_size size of point (cell)

point_border_col

color of border around points

 ${\tt point_border_stroke}$

stroke size of border around points

title title for plot, defaults to cell_color parameter

show_legend show legend

 ${\tt legend_symbol_size}$

size of legend symbols

background_color

color of plot background

axis_text size of axis text
axis_title size of axis title

cow_n_col cowplot param: how many columns
cow_rel_h cowplot param: relative height
cow_rel_w cowplot param: relative width
cow_align cowplot param: how to align

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function

Details

Description of parameters, see dimPlot2D. For 3D plots see plotUMAP_3D

Value

ggplot

Examples

plotUMAP_2D(gobject)

plotUMAP_3D plotUMAP_3D

Description

Visualize cells according to dimension reduction coordinates

176 plotUMAP_3D

```
Usage
```

label_size

edge_alpha

point_size

show_legend

size of labels

show legend

size of point (cell)

column to use for alpha of the edges

```
plotUMAP_3D(
      gobject,
      dim_reduction_name = "umap",
      default_save_name = "UMAP_3D",
Arguments
    gobject
                      giotto object
    dim_reduction_name
                      umap dimension reduction name
    default_save_name
                      default save name for saving, don't change, change save_name in save_param
    dim1_to_use
                      dimension to use on x-axis
    dim2_to_use
                      dimension to use on y-axis
                      dimension to use on z-axis
    dim3_to_use
    show_NN_network
                      show underlying NN network
    nn_network_to_use
                      type of NN network to use (kNN vs sNN)
    network_name
                      name of NN network to use, if show_NN_network = TRUE
    cell_color
                      color for cells (see details)
    color_as_factor
                      convert color column to factor
    cell_color_code
                      named vector with colors
    select_cell_groups
                      select subset of cells/clusters based on cell_color parameter
                      select subset of cells based on cell IDs
    select_cells
    show_other_cells
                      display not selected cells
    other_cell_color
                      color of not selected cells
    other_point_size
                      size of not selected cells
    show_cluster_center
                      plot center of selected clusters
    show_center_label
                      plot label of selected clusters
    center_point_size
                      size of center points
```

```
show_plot show plot
```

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function

Details

Description of parameters.

Value

plotly

Examples

```
plotUMAP_3D(gobject)
```

```
plot\_network\_layer\_ggplot \\ plot\_network\_layer\_ggplot
```

Description

Visualize cells in network layer according to dimension reduction coordinates

Usage

```
plot_network_layer_ggplot(
   ggobject,
   annotated_network_DT,
   edge_alpha = NULL,
   show_legend = T
)
```

Arguments

```
annotated\_network\_DT
```

annotated network data.table of selected cells

edge_alpha alpha of network edges

show_legend show legend gobject giotto object

Details

Description of parameters.

Value

ggplot

Examples

```
plot_network_layer_ggplot(gobject)
```

Description

Visualize cells in point layer according to dimension reduction coordinates

Usage

```
plot_point_layer_ggplot(
  ggobject,
  annotated_DT_selected,
  annotated_DT_other,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = 0,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_legend = T
```

```
annotated_DT_selected
annotated data.table of selected cells
annotated_DT_other
annotated data.table of not selected cells
cell_color color for cells (see details)
color_as_factor
convert color column to factor
```

```
cell_color_code
```

named vector with colors

cell_color_gradient

vector with 3 colors for numeric data

gradient_midpoint

midpoint for color gradient

gradient_limits

vector with lower and upper limits

select_cell_groups

select subset of cells/clusters based on cell_color parameter

select_cells select subset of cells based on cell IDs

point_size size of point (cell)

point_border_col

color of border around points

point_border_stroke

stroke size of border around points

show_cluster_center

plot center of selected clusters

show_center_label

plot label of selected clusters

center_point_size

size of center points

label_size size of labels

label_fontface font of labels

edge_alpha column to use for alpha of the edges

show_other_cells

display not selected cells

other_cell_color

color of not selected cells

other_point_size

size of not selected cells

show_legend show legend gobject giotto object

Details

Description of parameters.

Value

ggplot

Examples

```
plot_point_layer_ggplot(gobject)
```

```
plot\_point\_layer\_ggplot\_noFILL \\ plot\_point\_layer\_ggplot\_noFILL
```

Description

Visualize cells in point layer according to dimension reduction coordinates without borders

Usage

```
plot_point_layer_ggplot_noFILL(
  ggobject,
  annotated_DT_selected,
  annotated_DT_other,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = 0,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 1,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_legend = T
```

```
annotated_DT_selected
annotated data.table of selected cells
annotated_DT_other
annotated data.table of not selected cells
cell_color color for cells (see details)
color_as_factor
convert color column to factor
cell_color_code
named vector with colors
cell_color_gradient
vector with 3 colors for numeric data
gradient_midpoint
midpoint for color gradient
```

select subset of cells/clusters based on cell_color parameter

select_cells select subset of cells based on cell IDs

point_size size of point (cell)

show_cluster_center

plot center of selected clusters

vector with lower and upper limits

show_center_label

plot label of selected clusters

center_point_size

size of center points

label_size size of labels
label_fontface font of labels

edge_alpha column to use for alpha of the edges

show_other_cells

display not selected cells

other_cell_color

color of not selected cells

other_point_size

size of not selected cells

show_legend show legend gobject giotto object

Details

Description of parameters.

Value

ggplot

Examples

```
plot_point_layer_ggplot_noFILL(gobject)
```

Description

creat ggplot point layer for spatial coordinates

Usage

```
plot_spat_point_layer_ggplot(
  ggobject,
  sdimx = NULL,
  sdimy = NULL,
  cell_locations_metadata_selected,
  cell_locations_metadata_other,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 2,
  point_border_col = "lightgrey",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 1,
  show_legend = TRUE
)
```

```
sdimx
                  x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimv
cell_locations_metadata_selected
                  annotated location from selected cells
cell_locations_metadata_other
                  annotated location from non-selected cells
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
{\tt cell\_color\_gradient}
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
```

```
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
show_other_cells
                  display not selected cells
other_cell_color
                  color for not selected cells
other_point_size
                  point size for not selected cells
show_legend
                  show legend
                  giotto object
gobject
```

Details

Description of parameters.

Value

ggplot

Examples

```
plot_spat_point_layer_ggplot(gobject)
```

```
plot\_spat\_point\_layer\_ggplot\_noFILL \\ plot\_spat\_point\_layer\_ggplot\_noFILL
```

Description

creat ggplot point layer for spatial coordinates without borders

Usage

```
plot_spat_point_layer_ggplot_noFILL(
 ggobject,
  sdimx = NULL,
  sdimy = NULL,
  cell_locations_metadata_selected,
  cell_locations_metadata_other,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 2,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 4,
  label_fontface = "bold",
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
  show_legend = TRUE
)
```

```
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
{\tt cell\_locations\_metadata\_selected}
                  annotated location from selected cells
cell_locations_metadata_other
                  annotated location from non-selected cells
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
                  size of point (cell)
point_size
```

print.giotto 185

```
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
label_fontface font of labels
show_other_cells
                  display not selected cells
other_cell_color
                  color for not selected cells
other_point_size
                  point size for not selected cells
show_legend
                  show legend
gobject
                  giotto object
```

Details

Description of parameters.

Value

ggplot

Examples

```
plot_spat_point_layer_ggplot_noFILL(gobject)
```

print.giotto

print method for giotto class

Description

print method for giotto class. Prints the chosen number of genes (rows) and cells (columns) from the raw count matrix. Also print the spatial locations for the chosen number of cells.

Usage

```
print.giotto(object, ...)
```

Arguments

nr_genes number of genes (rows) to print nr_cells number of cells (columns) to print 186 rankEnrich

rankEnrich

rankEnrich

Description

Function to calculate gene signature enrichment scores per spatial position using a rank based approach.

Usage

```
rankEnrich(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

Arguments

Details

sign_matrix: a rank-fold matrix with genes as row names and cell-types as column names. Alternatively a scRNA-seq matrix and vector with clusters can be provided to makeSignMatrixRank, which will create the matrix for you.

First a new rank is calculated as $R = (R1*R2)^{n}(1/2)$, where R1 is the rank of fold-change for each gene in each spot and R2 is the rank of each marker in each cell type. The Rank-Biased Precision is then calculated as: $RBP = (1 - 0.99) * (0.99)^{n}(R - 1)$ and the final enrichment score is then calculated as the sum of top 100 RBPs.

Value

data.table with enrichment results

See Also

```
make Sign Matrix Rank
```

rankEnrichDT 187

Examples

```
rankEnrich(gobject)
```

rankEnrichDT

rankEnrichDT

Description

Function to calculate gene signature enrichment scores per spatial position using a rank based approach.

Usage

```
rankEnrichDT(
  gobject,
  sign_matrix,
  expression_values = c("normalized", "scaled", "custom"),
  reverse_log_scale = TRUE,
  logbase = 2,
  output_enrichment = c("original", "zscore")
)
```

Arguments

```
gobject Giotto object

sign_matrix Matrix of signature genes for each cell type / process

expression_values

expression values to use

reverse_log_scale

reverse expression values from log scale

logbase log base to use if reverse_log_scale = TRUE

output_enrichment

how to return enrichment output
```

Details

sign_matrix: a rank-fold matrix with genes as row names and cell-types as column names. Alternatively a scRNA-seq matrix and vector with clusters can be provided to makeSignMatrixRank, which will create the matrix for you.

First a new rank is calculated as $R = (R1*R2)^{(1/2)}$, where R1 is the rank of fold-change for each gene in each spot and R2 is the rank of each marker in each cell type. The Rank-Biased Precision is then calculated as: $RBP = (1 - 0.99) * (0.99)^{(R - 1)}$ and the final enrichment score is then calculated as the sum of top 100 RBPs.

Value

data.table with enrichment results

See Also

```
make Sign Matrix Rank
```

Examples

```
rankEnrichDT(gobject)
```

```
rank Spatial Cor Groups \\ \hspace{0.5in} rank Spatial Cor Groups \\
```

Description

Rank spatial correlated clusters according to correlation structure

Usage

```
rankSpatialCorGroups(
  gobject,
  spatCorObject,
  use_clus_name = NULL,
  show_plot = NA,
  return_plot = FALSE,
  save_plot = NA,
  save_param = list(),
  default_save_name = "rankSpatialCorGroups")
```

Arguments

```
gobject
                  giotto object
spatCorObject
                  spatial correlation object
                  name of clusters to visualize (from clusterSpatialCorGenes())
use_clus_name
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

Value

data.table with positive (within group) and negative (outside group) scores

Examples

```
rankSpatialCorGroups(gobject)
```

rank_binarize 189

rank_binarize

rank_binarize

Description

create binarized scores using arbitrary rank of top genes

Usage

```
rank_binarize(x, max_rank = 200)
```

readGiottoInstructions

read Giot to Instrunctions

Description

Retrieves the instruction associated with the provided parameter

Usage

```
readGiottoInstructions(giotto_instructions, param = NULL)
```

Arguments

giotto_instructions

 $giot to\ object\ or\ result\ from\ create Giot to Instructions()$

param

parameter to retrieve

Value

specific parameter

Examples

readGiottoInstrunctions()

190 removeGeneAnnotation

remove Cell Annotation remove Cell Annotation

Description

removes cell annotation of giotto object

Usage

```
removeCellAnnotation(gobject, columns = NULL, return_gobject = TRUE)
```

Arguments

gobject giotto object

columns names of columns to remove

return_gobject boolean: return giotto object (default = TRUE)

Details

if return_gobject = FALSE, it will return the cell metadata

Value

giotto object

Examples

removeCellAnnotation(gobject)

removeGeneAnnotation removeGeneAnnotation

Description

removes gene annotation of giotto object

Usage

```
removeGeneAnnotation(gobject, columns = NULL, return_gobject = TRUE)
```

Arguments

gobject giotto object

columns names of columns to remove

return_gobject boolean: return giotto object (default = TRUE)

Details

if return_gobject = FALSE, it will return the gene metadata

Value

giotto object

Examples

```
removeGeneAnnotation(gobject)
```

```
{\tt replaceGiottoInstructions}
```

replace Giot to Instructions

Description

Function to replace all instructions from giotto object

Usage

```
replaceGiottoInstructions(gobject, instructions = NULL)
```

Arguments

```
gobject giotto object
instructions new instructions (e.g. result from createGiottoInstructions)
```

Value

named vector with giotto instructions

Examples

```
replaceGiottoInstructions()
```

runPCA

runPCA

Description

runs a Principal Component Analysis

```
runPCA(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  name = "pca",
  genes_to_use = NULL,
  return_gobject = TRUE,
  scale_unit = F,
  ncp = 200,
  ...
)
```

192 runtSNE

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
                  cells or genes
reduction
name
                  arbitrary name for PCA run
                  subset of genes to use for PCA
genes_to_use
return_gobject boolean: return giotto object (default = TRUE)
scale_unit
                  scale features before PCA
                  number of principal components to calculate
ncp
                  additional parameters for PCA (see details)
. . .
```

Details

See PCA for more information about other parameters.

Value

giotto object with updated PCA dimension recuction

Examples

```
runPCA(gobject)
```

runtSNE

runtSNE

Description

run tSNE

```
runtSNE(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "tsne",
  genes_to_use = NULL,
  return_gobject = TRUE,
  dims = 2,
  perplexity = 30,
  theta = 0.5,
  do_PCA_first = F,
  set\_seed = T,
  seed_number = 1234,
)
```

runtSNE 193

Arguments

gobject giotto object

expression_values

expression values to use

reduction cells or genes

dim_reduction_to_use

use another dimension reduction set as input

dim_reduction_name

name of dimension reduction set to use

dimensions_to_use

number of dimensions to use as input

name arbitrary name for tSNE run

genes_to_use if dim_reduction_to_use = NULL, which genes to use

return_gobject boolean: return giotto object (default = TRUE)

dims tSNE param: number of dimensions to return

perplexity tSNE param: perplexity

theta tSNE param: theta

do_PCA_first tSNE param: do PCA before tSNE (default = FALSE)

set_seed use of seed

seed_number seed number to use

... additional tSNE parameters

Details

See Rtsne for more information about these and other parameters.

- Input for tSNE dimension reduction can be another dimension reduction (default = 'pca')
- To use gene expression as input set dim_reduction_to_use = NULL
- multiple tSNE results can be stored by changing the *name* of the analysis

Value

giotto object with updated tSNE dimension recuction

Examples

runtSNE(gobject)

194 runUMAP

runUMAP

runUMAP

Description

run UMAP

Usage

```
runUMAP(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  dim_reduction_to_use = "pca",
  dim_reduction_name = "pca",
  dimensions_to_use = 1:10,
  name = "umap",
  genes_to_use = NULL,
  return_gobject = TRUE,
  n_neighbors = 40,
  n_{components} = 2,
  n_{epochs} = 400,
  min_dist = 0.01,
  n_{threads} = 1,
  spread = 5,
  set\_seed = T,
  seed_number = 1234,
)
```

```
gobject
                 giotto object
expression_values
                 expression values to use
                 cells or genes
reduction
dim_reduction_to_use
                 use another dimension reduction set as input
dim_reduction_name
                 name of dimension reduction set to use
dimensions_to_use
                 number of dimensions to use as input
                 arbitrary name for UMAP run
name
                 if dim_reduction_to_use = NULL, which genes to use
genes_to_use
return_gobject boolean: return giotto object (default = TRUE)
n\_neighbors
                 UMAP param: number of neighbors
                 UMAP param: number of components
n_components
n_epochs
                 UMAP param: number of epochs
```

selectPatternGenes 195

```
min_dist UMAP param: minimum distance
n_threads UMAP param: threads to use
spread UMAP param: spread
set_seed use of seed
seed_number seed number to use
additional UMAP parameters
```

Details

See umap for more information about these and other parameters.

- Input for UMAP dimension reduction can be another dimension reduction (default = 'pca')
- To use gene expression as input set dim_reduction_to_use = NULL
- multiple UMAP results can be stored by changing the *name* of the analysis

Value

giotto object with updated UMAP dimension recuction

Examples

```
runUMAP(gobject)
```

selectPatternGenes selectPatternGenes

Description

Select genes correlated with spatial patterns

Usage

```
selectPatternGenes(
   spatPatObj,
   dimensions = 1:5,
   top_pos_genes = 10,
   top_neg_genes = 10,
   min_pos_cor = 0.5,
   min_neg_cor = -0.5,
   return_top_selection = FALSE
)
```

Arguments

spatPatObj

```
dimensions dimensions to identify correlated genes for.

top_pos_genes Top positively correlated genes.

top_neg_genes Top negatively correlated genes.

min_pos_cor Minimum positive correlation score to include a gene.

min_neg_cor Minimum negative correlation score to include a gene.
```

Output from detectSpatialPatterns

show,giotto-method

Details

Description.

Value

Data.table with genes associated with selected dimension (PC).

Examples

```
selectPatternGenes(gobject)
```

```
select_expression_values
```

select_expression_values

Description

helper function to select expression values

Usage

```
select_expression_values(gobject, values)
```

Arguments

gobject giotto object

values expression values to extract

Value

expression matrix

show,giotto-method

show method for giotto class

Description

show method for giotto class

```
## S4 method for signature 'giotto'
show(object)
```

 $show Cluster Dendrogram \quad show Cluster Dendrogram \quad$

Description

Creates dendrogram for selected clusters.

Usage

```
showClusterDendrogram(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  h = NULL,
  h_color = "red",
  rotate = FALSE,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showClusterDendrogram",
  ...
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
distance
                  distance method to use for hierarchical clustering
h
                  height of horizontal lines to plot
h_color
                  color of horizontal lines
                  rotate dendrogram 90 degrees
rotate
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  additional parameters for ggdendrogram()
```

Details

Expression correlation dendrogram for selected clusters.

198 showClusterHeatmap

Value

ggplot

Examples

showClusterDendrogram(gobject)

showClusterHeatmap

showClusterHeatmap

Description

Creates heatmap based on identified clusters

Usage

```
showClusterHeatmap(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = "all",
  cluster_column,
  cor = c("pearson", "spearman"),
  distance = "ward.D",
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showClusterHeatmap",
  ...
)
```

```
giotto object
gobject
expression_values
                  expression values to use
                  vector of genes to use, default to 'all'
genes
cluster_column name of column to use for clusters
                  correlation score to calculate distance
cor
distance
                  distance method to use for hierarchical clustering
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
                  additional parameters for the Heatmap function from ComplexHeatmap
. . .
```

showCPGscores 199

Details

Correlation heatmap of selected clusters.

Value

ggplot

Examples

showClusterHeatmap(gobject)

showCPGscores

showCPGscores

Description

visualize Cell Proximity Gene enrichment scores

Usage

```
showCPGscores(
 gobject,
 CPGscore,
 method = c("volcano", "cell_barplot", "cell-cell", "cell_sankey"),
 min_cells = 5,
 min_fdr = 0.05,
 min_spat_diff = 0.2,
 min_log2_fc = 0.5,
 keep_int_duplicates = TRUE,
 direction = c("both", "up", "down"),
 cell_color_code = NULL,
  show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "showCPGscores"
)
```

```
CPGscore
                 CPGscore, output from getCellProximityGeneScores()
method
                 visualization method
                 min number of cells threshold
min_cells
min_fdr
                 fdr threshold
min_spat_diff
                 spatial difference threshold
                 min log2 fold-change
min_log2_fc
keep_int_duplicates
                 keep both cell_A-cell_B and cell_B-cell_A
direction
                 up or downregulation or both
```

```
cell_color_code
```

color code for cell types

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function

default_save_name

default save name for saving, don't change, change save_name in save_param

Details

Give more details ...

Value

Gene to gene scores in data.table format

Examples

```
showCPGscores(CPGscore)
```

showGeneExpressionProximityScore

show Gene Expression Proximity Score

Description

Create heatmap from cell-cell proximity scores

Usage

```
showGeneExpressionProximityScore(
  scores,
  selected_gene,
  sort_column = "diff_spat"
)
```

Arguments

scores CPscore, output from getAverageCellProximityGeneScores()

selected_gene gene to show

sort_column column name to use for sorting

Details

Give more details ...

Value

ggplot barplot

showGiottoInstructions 201

Examples

```
showGeneExpressionProximityScore(scores)
```

```
{\tt showGiottoInstructions}
```

showGiottoInstructions

Description

Function to display all instructions from giotto object

Usage

```
showGiottoInstructions(gobject)
```

Arguments

```
gobject giotto object
```

Value

named vector with giotto instructions

Examples

```
showGiottoInstructions()
```

showGTGscores

showGTGscores

Description

visualize Cell Proximity Gene enrichment scores

```
showGTGscores(
  GTGscore,
  method = c("cell_barplot", "cell-cell", "cell_sankey"),
  min_cells = 5,
  min_pval = 0.05,
  min_spat_diff = 0.2,
  min_log2_fc = 0.5,
  direction = c("both", "up", "down"),
  cell_color_code = NULL,
  show_plot = T,
  specific_genes_1 = NULL,
  specific_genes_2 = NULL,
  first_cell_name = "ligand cell",
  second_cell_name = "receptor cell",
  return_DT = F
)
```

Arguments

```
visualization method
method
min_cells
                  min number of cells threshold
                  p-value threshold
min_pval
min_spat_diff
                  spatial difference threshold
                  log2 fold-change threshold
min_log2_fc
direction
                  up or downregulation or both
cell_color_code
                  color code for cell types
show_plot
                  print plot
specific_genes_1
                  subset of genes, matched with specific_genes_2
specific_genes_2
                  subset of genes, matched with specific_genes_1
first_cell_name
                  name for first cells
second_cell_name
                  name for second cells
                  CPGscore, output from getCellProximityGeneScores()
CPGscore
```

Details

Give more details ...

Value

ggplot

Examples

```
showGTGscores(CPGscore)
```

```
show Int {\it Expression Proximity Score} \\ show Int {\it Expression Proximity Score}
```

Description

Create heatmap from cell-cell proximity scores

```
showIntExpressionProximityScore(
   scores,
   selected_interaction,
   sort_column = "diff_spat",
   show_enriched_n = 5,
   show_depleted_n = 5
```

showPattern 203

Arguments

scores scores, output from getAverageCellProximityGeneScores() selected_interaction interaction to show sort_column column name to use for sorting show_enriched_n show top enriched interactions show_depleted_n

show top depleted interactions

Details

Give more details ...

Value

ggplot barplot

Examples

showIntExpressionProximityScore(scores)

showPattern showPattern

Description

show patterns for 2D spatial data

Usage

```
showPattern(gobject, spatPatObj, ...)
```

Arguments

gobject giotto object

spatPatObj Output from detectSpatialPatterns

dimension dimension to plot

trim Trim ends of the PC values.

background_color

background color for plot

grid_border_color

color for grid

show_legend show legend of ggplot

show_plot show plot

return_plot return ggplot object

directly save the plot [boolean] save_plot

save_param list of saving parameters from all_plots_save_function

default_save_name

default save name for saving, don't change, change save_name in save_param

204 showPattern2D

Value

ggplot

See Also

showPattern2D

Examples

```
showPattern(gobject)
```

showPattern2D

showPattern2D

Description

show patterns for 2D spatial data

Usage

```
showPattern2D(
  gobject,
  spatPatObj,
  dimension = 1,
  trim = c(0.02, 0.98),
  background_color = "white",
  grid_border_color = "grey",
  show_legend = T,
  point_size = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showPattern2D"
)
```

Arguments

return_plot

giotto object gobject spatPatObj Output from detectSpatialPatterns dimension dimension to plot Trim ends of the PC values. trim background_color background color for plot grid_border_color color for grid show_legend show legend of ggplot show_plot show plot

return ggplot object

showPattern3D 205

Value

ggplot

Examples

```
showPattern2D(gobject)
```

showPattern3D

showPattern3D

Description

show patterns for 3D spatial data

Usage

```
showPattern3D(
 gobject,
 spatPatObj,
 dimension = 1,
  trim = c(0.02, 0.98),
 background_color = "white",
 grid_border_color = "grey",
 show_legend = T,
 point_size = 1,
 axis_scale = c("cube", "real", "custom"),
 custom_ratio = NULL,
 x_{ticks} = NULL,
 y_ticks = NULL,
 z_ticks = NULL,
 show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "showPattern3D"
)
```

```
gobject giotto object
spatPatObj Output from detectSpatialPatterns
dimension dimension to plot
trim Trim ends of the PC values.
background_color
background color for plot
```

206 showPatternGenes

```
grid_border_color
                  color for grid
show_legend
                  show legend of plot
                  adjust the point size
point_size
axis_scale
                  scale the axis
                  cutomize the scale of the axis
custom_ratio
                  the tick number of x_axis
x_ticks
y_ticks
                  the tick number of y_axis
                  the tick number of z_axis
z_ticks
show_plot
                  show plot
return_plot
                  return plot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

Value

plotly

Examples

showPattern3D(gobject)

showPatternGenes

showPatternGenes

Description

show genes correlated with spatial patterns

```
showPatternGenes(
  gobject,
  spatPatObj,
  dimension = 1,
  top_pos_genes = 5,
  top_neg_genes = 5,
  point_size = 1,
  return_DT = FALSE,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "showPatternGenes"
)
```

showProcessingSteps 207

Arguments

gobject giotto object

 ${\tt spatPatObj} \qquad {\tt Output} \ from \ detectSpatial Patterns$

dimension dimension to plot genes for.top_pos_genes Top positively correlated genes.top_neg_genes Top negatively correlated genes.

point_size size of points

return_DT if TRUE, it will return the data.table used to generate the plots

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function()

default_save_name

default save name for saving, don't change, change save_name in save_param

Value

ggplot

Examples

showPatternGenes(gobject)

showProcessingSteps showProcessingSteps

Description

shows the sequential processing steps that were performed in a summarized format

Usage

showProcessingSteps(gobject)

Arguments

gobject giotto object

Value

list of processing steps and names

Examples

showProcessingSteps(gobject)

208 showSpatialCorGenes

showSpatialCorGenes showSpatialCorGenes

Description

Shows and filters spatially correlated genes

Usage

```
showSpatialCorGenes(
  spatCorObject,
  use_clus_name = NULL,
  selected_clusters = NULL,
  genes = NULL,
  min_spat_cor = 0.5,
  min_expr_cor = NULL,
  min_cor_diff = NULL,
  min_rank_diff = NULL,
  show_top_genes = NULL
)
```

Arguments

```
spatCorObject
                  spatial correlation object
use_clus_name
                  cluster information to show
selected_clusters
                  subset of clusters to show
                  subset of genes to show
genes
                  filter on minimum spatial correlation
min_spat_cor
                  filter on minimum single-cell expression correlation
min_expr_cor
                  filter on minimum correlation difference (spatial vs expression)
min_cor_diff
                  filter on minimum correlation rank difference (spatial vs expression)
min_rank_diff
show_top_genes show top genes per gene
```

Value

data.table with filtered information

Examples

```
\verb|showSpatialCorGenes(gobject)|
```

showTopGeneToGene 209

showTopGeneToGene

show Top Gene To Gene

Description

Show enriched/depleted gene-gene enrichments

Usage

```
showTopGeneToGene(
  GTGscore,
  top_interactions = 10,
  direction = c("increased", "decreased"),
  complement_data = T,
  subset_cell_ints = NULL,
  subset_genes = NULL
)
```

Arguments

Details

Give more details ...

Value

ggplot barplot

Examples

```
showTopGeneToGene(scores)
```

210 signPCA

signPCA signPCA

Description

identify significant prinicipal components (PCs)

Usage

```
signPCA(
  gobject,
  method = c("screeplot", "jackstraw"),
  expression_values = c("normalized", "scaled", "custom"),
  reduction = c("cells", "genes"),
  genes_to_use = NULL,
  scale_unit = T,
  ncp = 50,
  scree_labels = T,
  scree_ylim = c(0, 10),
  jack_iter = 10,
  jack_threshold = 0.01,
  jack_verbose = T,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "signPCA",
)
```

```
gobject
                  giotto object
method
                  method to use to identify significant PCs
expression_values
                  expression values to use
                  cells or genes
reduction
genes_to_use
                  subset of genes to use for PCA
                  scale features before PCA
scale_unit
                  number of principal components to calculate
ncp
scree_labels
                  show labels on scree plot
                  y-axis limits on scree plot
scree_ylim
jack_iter
                  number of interations for jackstraw
jack_threshold p-value threshold to call a PC significant
                  show progress of jackstraw method
jack_verbose
show_plot
                  show plot
return_plot
                  return ggplot object
```

spatCellPlot 211

Details

Two different methods can be used to assess the number of relevant or significant prinicipal components (PC's).

- 1. Screeplot works by plotting the explained variance of each individual PC in a barplot allowing you to identify which PC does not show a significant contribution anymore (= 'elbow method').
- 2. The Jackstraw method uses the permutationPA function. By systematically permuting genes it identifies robust, and thus significant, PCs.

multiple PCA results can be stored by changing the name parameter

Value

ggplot object for scree method and maxtrix of p-values for jackstraw

Examples

```
signPCA(gobject)
```

spatCellPlot

spatCellPlot

Description

Visualize cells according to spatial coordinates

```
spatCellPlot(
  gobject,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_annotation_values = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 point_size = 3,
 point_border_col = "black",
  point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
  center_point_size = 4,
  center_point_border_col = "black",
```

212 spatCellPlot

```
center_point_border_stroke = 0.1,
 label_size = 4,
  label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
  show\_grid = F,
  spatial_grid_name = "spatial_grid",
 grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
 other_cells_alpha = 0.1,
  coord_fix_ratio = NULL,
  show_legend = T,
  legend_text = 8,
 legend_symbol_size = 1,
 background_color = "white",
 axis_text = 8,
 axis_title = 8,
 cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
 save_param = list(),
 default_save_name = "spatCellPlot"
)
```

```
giotto object
gobject
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
spat_enr_names names of spatial enrichment results to include
cell annotation values
                  numeric cell annotation columns
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
                  size of point (cell)
point_size
```

point_border_col color of border around points point_border_stroke stroke size of border around points show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points label_size size of labels label_fontface font of labels show_network show underlying spatial network spatial_network_name name of spatial network to use color of spatial network network_color network_alpha alpha of spatial network show_grid show spatial grid spatial_grid_name name of spatial grid to use grid_color color of spatial grid show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size point size of not selected cells other_cells_alpha alpha of not selected cells coord_fix_ratio fix ratio between x and y-axis show_legend show legend legend_text size of legend text legend_symbol_size size of legend symbols background_color color of plot background size of axis text axis_text size of axis title axis_title show_plot show plot return_plot return ggplot object directly save the plot [boolean] save_plot list of saving parameters from all_plots_save_function save_param default_save_name default save name for saving, don't change, change save_name in save_param 214 spatCellPlot2D

Details

Description of parameters.

Value

ggplot

Examples

```
spatCellPlot(gobject)
```

spatCellPlot2D

spatCellPlot2D

Description

Visualize cells according to spatial coordinates

```
spatCellPlot2D(
 gobject,
 sdimx = "sdimx",
 sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_annotation_values = NULL,
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
 select_cell_groups = NULL,
  select_cells = NULL,
  point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
 label_fontface = "bold",
 show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
  show\_grid = F,
  spatial_grid_name = "spatial_grid",
  grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
```

spatCellPlot2D 215

```
other_cells_alpha = 0.1,
      coord_fix_ratio = NULL,
      show_legend = T,
      legend_text = 8,
      legend_symbol_size = 1,
      background_color = "white",
      axis_text = 8,
      axis_title = 8,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatCellPlot2D"
    )
Arguments
   gobject
                     giotto object
                     x-axis dimension name (default = 'sdimx')
    sdimx
                     y-axis dimension name (default = 'sdimy')
    sdimy
    spat_enr_names names of spatial enrichment results to include
    cell_annotation_values
                     numeric cell annotation columns
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
    point_size
                     size of point (cell)
   point_border_col
                     color of border around points
   point_border_stroke
                     stroke size of border around points
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
                     size of center points
    label_size
                     size of labels
```

label_fontface font of labels

216 spatCellPlot2D

```
show underlying spatial network
show_network
spatial_network_name
                  name of spatial network to use
                  color of spatial network
network_color
                  alpha of spatial network
network_alpha
show_grid
                  show spatial grid
spatial_grid_name
                  name of spatial grid to use
grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
other_point_size
                  point size of not selected cells
other_cells_alpha
                  alpha of not selected cells
coord_fix_ratio
                  fix ratio between x and y-axis
show_legend
                  show legend
                  size of legend text
legend_text
legend_symbol_size
                  size of legend symbols
background_color
                  color of plot background
axis_text
                  size of axis text
axis_title
                  size of axis title
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function
save_param
```

default save name for saving, don't change, change save_name in save_param

Details

Description of parameters.

default_save_name

Value

ggplot

Examples

spatCellPlot2D(gobject)

spatDimCellPlot 217

spatDimCellPlot

spatDimCellPlot

Description

Visualize numerical features of cells according to spatial AND dimension reduction coordinates in 2D

```
spatDimCellPlot(
 gobject,
 plot_alignment = c("vertical", "horizontal"),
  spat_enr_names = NULL,
  cell_annotation_values = NULL,
  dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 dim_point_size = 1,
  dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 dim_show_cluster_center = F,
 dim_show_center_label = T,
 dim_center_point_size = 4,
  dim_center_point_border_col = "black",
 dim_center_point_border_stroke = 0.1,
 dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_center_point_border_col = "black",
  spat_center_point_border_stroke = 0.1,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
 nn_network_to_use = "sNN",
 nn_network_name = "sNN.pca",
 dim_edge_alpha = 0.5,
  spat_show_network = F,
```

218 spatDimCellPlot

```
spatial_network_name = "spatial_network",
      spat_network_color = "red",
      spat_network_alpha = 0.5,
      spat_show_grid = F,
      spatial_grid_name = "spatial_grid",
      spat_grid_color = "green",
      show_other_cells = TRUE,
      other_cell_color = "grey"
      dim_other_point_size = 0.5,
      spat_other_point_size = 0.5,
      spat_other_cells_alpha = 0.5,
      coord_fix_ratio = NULL,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h"
      show_legend = T,
      legend_text = 8,
      legend_symbol_size = 1,
      dim_background_color = "white",
      spat_background_color = "white",
      axis_text = 8,
      axis_title = 8,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimCellPlot"
   )
Arguments
                    giotto object
   gobject
    plot_alignment direction to align plot
    spat_enr_names names of spatial enrichment results to include
    cell_annotation_values
                    numeric cell annotation columns
    dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
    dim2_to_use
                    dimension to use on y-axis
    sdimx
                    = spatial dimension to use on x-axis
                    = spatial dimension to use on y-axis
    sdimy
    cell_color_gradient
                    vector with 3 colors for numeric data
    gradient_midpoint
                    midpoint for color gradient
    gradient_limits
```

vector with lower and upper limits

select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells dim_point_size size of points in dim. reduction space dim_point_border_col border color of points in dim. reduction space dim_point_border_stroke border stroke of points in dim. reduction space spat_point_size size of spatial points spat_point_border_col border color of spatial points spat_point_border_stroke border stroke of spatial points dim_show_cluster_center show the center of each cluster dim_show_center_label provide a label for each cluster dim_center_point_size size of the center point dim_center_point_border_col border color of center point dim_center_point_border_stroke stroke size of center point dim_label_size size of the center label dim_label_fontface font of the center label spat_show_cluster_center show the center of each cluster spat_show_center_label provide a label for each cluster spat_center_point_size size of the center point spat_label_size size of the center label spat_label_fontface font of the center label show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) nn_network_name name of NN network to use, if show_NN_network = TRUE dim_edge_alpha column to use for alpha of the edges spat_show_network show spatial network spatial_network_name name of spatial network to use

220 spatDimCellPlot

```
spat_network_color
                  color of spatial network
spat_show_grid show spatial grid
spatial_grid_name
                  name of spatial grid to use
spat_grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
dim_other_point_size
                  size of not selected dim cells
spat_other_point_size
                  size of not selected spat cells
spat_other_cells_alpha
                  alpha of not selected spat cells
coord_fix_ratio
                  ratio for coordinates
cow_n_col
                  cowplot param: how many columns
cow_rel_h
                  cowplot param: relative height
cow_rel_w
                  cowplot param: relative width
cow_align
                  cowplot param: how to align
show_legend
                  show legend
legend_text
                  size of legend text
legend_symbol_size
                  size of legend symbols
dim_background_color
                  background color of points in dim. reduction space
spat_background_color
                  background color of spatial points
                  size of axis text
axis_text
                  size of axis title
axis_title
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_param
                  list of saving parameters from all_plots_save_function
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

Details

Description of parameters.

Value

ggplot

spatDimCellPlot2D 221

Examples

```
spatDimCellPlot(gobject)
```

spatDimCellPlot2D

spatDimCellPlot2D

Description

Visualize numerical features of cells according to spatial AND dimension reduction coordinates in 2D

```
spatDimCellPlot2D(
  gobject,
 plot_alignment = c("vertical", "horizontal"),
  spat_enr_names = NULL,
 cell_annotation_values = NULL,
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 dim_point_size = 1,
  dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 dim_show_cluster_center = F,
 dim_show_center_label = T,
 dim_center_point_size = 4,
 dim_center_point_border_col = "black",
 dim_center_point_border_stroke = 0.1,
 dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_center_point_border_col = "black",
  spat_center_point_border_stroke = 0.1,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
```

222 spatDimCellPlot2D

```
nn_network_to_use = "sNN",
nn_network_name = "sNN.pca",
dim_edge_alpha = 0.5,
spat_show_network = F,
spatial_network_name = "spatial_network",
spat_network_color = "red",
spat_network_alpha = 0.5,
spat_show_grid = F,
spatial_grid_name = "spatial_grid",
spat_grid_color = "green",
show_other_cells = TRUE,
other_cell_color = "grey",
dim_other_point_size = 0.5,
spat_other_point_size = 0.5,
spat_other_cells_alpha = 0.5,
coord_fix_ratio = NULL,
cow_n_col = 2,
cow_rel_h = 1,
cow_rel_w = 1,
cow_align = "h",
show_legend = T,
legend_text = 8,
legend_symbol_size = 1,
dim_background_color = "white",
spat_background_color = "white",
axis_text = 8,
axis_title = 8,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spatDimCellPlot2D"
```

Arguments

```
giotto object
gobject
plot_alignment direction to align plot
spat_enr_names names of spatial enrichment results to include
cell_annotation_values
                  numeric cell annotation columns
dim_reduction_to_use
                  dimension reduction to use
dim_reduction_name
                  dimension reduction name
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
sdimx
                  = spatial dimension to use on x-axis
                  = spatial dimension to use on y-axis
sdimy
cell_color_gradient
                  vector with 3 colors for numeric data
```

gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells dim_point_size size of points in dim. reduction space dim_point_border_col border color of points in dim. reduction space dim_point_border_stroke border stroke of points in dim. reduction space spat_point_size size of spatial points spat_point_border_col border color of spatial points spat_point_border_stroke border stroke of spatial points dim_show_cluster_center show the center of each cluster dim_show_center_label provide a label for each cluster dim_center_point_size size of the center point dim_center_point_border_col border color of center point dim_center_point_border_stroke stroke size of center point dim_label_size size of the center label dim_label_fontface font of the center label spat_show_cluster_center show the center of each cluster spat_show_center_label provide a label for each cluster spat_center_point_size size of the center point spat_label_size size of the center label spat_label_fontface font of the center label show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) nn_network_name name of NN network to use, if show_NN_network = TRUE dim_edge_alpha column to use for alpha of the edges

224 spatDimCelIPlot2D

```
spat_show_network
                  show spatial network
spatial_network_name
                  name of spatial network to use
spat_network_color
                  color of spatial network
spat_show_grid show spatial grid
spatial_grid_name
                  name of spatial grid to use
spat_grid_color
                  color of spatial grid
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
dim_other_point_size
                  size of not selected dim cells
spat_other_point_size
                  size of not selected spat cells
spat_other_cells_alpha
                  alpha of not selected spat cells
coord_fix_ratio
                  ratio for coordinates
                  cowplot param: how many columns
cow_n_col
                  cowplot param: relative height
cow_rel_h
                  cowplot param: relative width
cow_rel_w
                  cowplot param: how to align
cow_align
show_legend
                  show legend
legend_text
                  size of legend text
legend_symbol_size
                  size of legend symbols
dim_background_color
                  background color of points in dim. reduction space
spat_background_color
                  background color of spatial points
axis_text
                  size of axis text
axis_title
                  size of axis title
{\sf show\_plot}
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, don't change, change save_name in save_param
```

Details

Description of parameters.

spatDimGenePlot 225

Value

ggplot

Examples

```
spatDimCellPlot2D(gobject)
```

spatDimGenePlot

spatDimGenePlot

Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

```
spatDimGenePlot(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("vertical", "horizontal"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  show_NN_network = F,
  show\_spatial\_network = F,
  show_spatial_grid = F,
  nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
  spatial_network_name = "spatial_network",
  spatial_grid_name = "spatial_grid",
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 midpoint = 0,
 genes_high_color = "red",
 genes_mid_color = "white",
 genes_low_color = "blue",
 cow_n_col = 2,
 cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_legend = T,
  show_plot = NA,
  return_plot = NA,
```

226 spatDimGenePlot

 $save_plot = NA$,

```
save_param = list(),
      default_save_name = "spatDimGenePlot"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
    plot_alignment direction to align plot
    genes
                     genes to show
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    point_size
                     size of point (cell)
    dim_point_border_col
                     color of border around points
    dim_point_border_stroke
                     stroke size of border around points
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    edge_alpha_dim dim reduction plot: column to use for alpha of the edges
    scale_alpha_with_expression
                     scale expression with ggplot alpha parameter
    spatial_network_name
                     name of spatial network to use
    spatial_grid_name
                     name of spatial grid to use
    spat_point_size
                     spatial plot: point size
    spat_point_border_col
                     color of border around points
    spat_point_border_stroke
                     stroke size of border around points
    midpoint
                     size of point (cell)
    cow_n_col
                     cowplot param: how many columns
    cow_rel_h
                     cowplot param: relative height
    cow_rel_w
                     cowplot param: relative width
                     cowplot param: how to align
    cow_align
```

spatDimGenePlot2D 227

Details

Description of parameters.

Value

ggplot

See Also

```
spatDimGenePlot3D
```

Examples

```
spatDimGenePlot(gobject)
```

spatDimGenePlot2D

spatDimGenePlot2D

Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

```
spatDimGenePlot2D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("vertical", "horizontal"),
  genes,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  point_size = 1,
  dim_point_border_col = "black",
  dim_point_border_stroke = 0.1,
  show_NN_network = F,
  show_spatial_network = F,
  show_spatial_grid = F,
  nn_network_to_use = "sNN",
```

228 spatDimGenePlot2D

```
network_name = "sNN.pca",
     edge_alpha_dim = NULL,
      scale_alpha_with_expression = FALSE,
      spatial_network_name = "spatial_network",
      spatial_grid_name = "spatial_grid",
      spat_point_size = 1,
      spat_point_border_col = "black",
      spat_point_border_stroke = 0.1,
     midpoint = 0,
     genes_high_color = "red",
      genes_mid_color = "white",
      genes_low_color = "blue",
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h"
      show_legend = T,
      show_plot = NA,
     return_plot = NA,
     save_plot = NA,
     save_param = list(),
     default_save_name = "spatDimGenePlot2D"
   )
Arguments
   gobject
                    giotto object
   expression_values
                    gene expression values to use
   plot_alignment direction to align plot
   genes
                    genes to show
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   point_size
                    size of point (cell)
   dim_point_border_col
                    color of border around points
   dim_point_border_stroke
                    stroke size of border around points
   show_NN_network
                    show underlying NN network
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
   network_name
                    name of NN network to use, if show_NN_network = TRUE
   edge_alpha_dim dim reduction plot: column to use for alpha of the edges
   scale_alpha_with_expression
```

scale expression with ggplot alpha parameter

spatDimGenePlot2D 229

```
spatial_network_name
                 name of spatial network to use
spatial\_grid\_name
                 name of spatial grid to use
spat_point_size
                 spatial plot: point size
spat_point_border_col
                 color of border around points
spat_point_border_stroke
                 stroke size of border around points
midpoint
                 size of point (cell)
                 cowplot param: how many columns
cow_n_col
                 cowplot param: relative height
cow_rel_h
                 cowplot param: relative width
cow_rel_w
                 cowplot param: how to align
cow_align
show_legend
                 show legend
show_plot
                 show plots
return_plot
                 return ggplot object
save_plot
                 directly save the plot [boolean]
save_param
                 list of saving parameters from all_plots_save_function
default_save_name
                 default save name for saving, don't change, change save_name in save_param
dim_point_size dim reduction plot: point size
```

Details

Description of parameters.

Value

ggplot

See Also

 ${\tt spatDimGenePlot3D}$

Examples

spatDimGenePlot2D(gobject)

230 spatDimGenePlot3D

spatDimGenePlot3D

spatDimGenePlot3D

Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

```
spatDimGenePlot3D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("horizontal", "vertical"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
 dim3_to_use = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy",
  sdimz = "sdimz",
 genes,
 cluster_column = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1.5,
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
  label_size = 16,
  genes_low_color = "blue",
 genes_mid_color = "white",
  genes_high_color = "red",
 dim_point_size = 3,
 nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
 network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  legend_text_size = 12,
  axis_scale = c("cube", "real", "custom"),
 custom_ratio = NULL,
 x_ticks = NULL,
 y_ticks = NULL,
```

spatDimGenePlot3D 231

```
z_ticks = NULL,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatDimGenePlot3D"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
    plot_alignment direction to align plot
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    genes
                     genes to show
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    dim_point_size dim reduction plot: point size
    spatial_network_name
                     name of spatial network to use
    spatial_grid_name
                     name of spatial grid to use
    spatial_point_size
                     spatial plot: point size
    show_plot
                     show plots
    return_plot
                     return plotly object
    save_plot
                     directly save the plot [boolean]
    save_param
                     list of saving parameters from all_plots_save_function
    default_save_name
                     default save name for saving, don't change, change save_name in save_param
    edge_alpha_dim dim reduction plot: column to use for alpha of the edges
    scale_alpha_with_expression
                     scale expression with ggplot alpha parameter
                     size of point (cell)
    point_size
```

show_legend

show legend

232 spatDimPlot

Details

Description of parameters.

Value

plotly

Examples

```
spatDimGenePlot3D(gobject)
```

spatDimPlot

spatDimPlot

Description

Visualize cells according to spatial AND dimension reduction coordinates 2D

```
spatDimPlot(
 gobject,
 plot_alignment = c("vertical", "horizontal"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 dim_point_shape = c("border", "no_border"),
 dim_point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_shape = c("border", "no_border"),
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
  dim_show_cluster_center = F,
 dim_show_center_label = T,
 dim_center_point_size = 4,
 dim_center_point_border_col = "black",
  dim_center_point_border_stroke = 0.1,
```

spatDimPlot 233

```
dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 nn_network_alpha = 0.05,
  show_spatial_network = F,
  spat_network_name = "spatial_network",
  spat_network_color = "blue",
  spat_network_alpha = 0.5,
  show_spatial_grid = F,
  spat_grid_name = "spatial_grid",
  spat_grid_color = "blue",
  show_other_cells = T,
  other_cell_color = "lightgrey",
 dim_other_point_size = 1,
  spat_other_point_size = 1,
  spat_other_cells_alpha = 0.5,
  dim\_show\_legend = F,
  spat_show_legend = F,
  legend_text = 8,
  legend_symbol_size = 1,
  dim_background_color = "white",
  spat_background_color = "white",
 axis_text = 8,
 axis_title = 8,
  show_plot = NA,
 return_plot = NA,
 save_plot = NA,
 save_param = list(),
 default_save_name = "spatDimPlot"
)
```

Arguments

```
gobject
                  giotto object
plot_alignment direction to align plot
{\tt dim\_reduction\_to\_use}
                  dimension reduction to use
dim_reduction_name
                  dimension reduction name
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
                  = spatial dimension to use on x-axis
sdimx
sdimy
                  = spatial dimension to use on y-axis
spat_enr_names names of spatial enrichment results to include
```

234 spatDimPlot

cell_color color for cells (see details) color_as_factor convert color column to factor cell_color_code named vector with colors cell_color_gradient vector with 3 colors for numeric data gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells dim_point_shape point with border or not (border or no_border) dim_point_size size of points in dim. reduction space dim_point_border_col border color of points in dim. reduction space ${\tt dim_point_border_stroke}$ border stroke of points in dim. reduction space spat_point_shape point with border or not (border or no_border) spat_point_size size of spatial points spat_point_border_col border color of spatial points spat_point_border_stroke border stroke of spatial points dim_show_cluster_center show the center of each cluster dim_show_center_label provide a label for each cluster ${\tt dim_center_point_size}$ size of the center point dim_center_point_border_col border color of center point dim_center_point_border_stroke stroke size of center point dim_label_size size of the center label dim_label_fontface font of the center label spat_show_cluster_center show the center of each cluster spat_show_center_label provide a label for each cluster ${\tt spat_center_point_size}$

size of the center point

spat_label_size size of the center label spat_label_fontface font of the center label show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) name of NN network to use, if show_NN_network = TRUE network_name nn_network_alpha column to use for alpha of the edges show_spatial_network show spatial network spat_network_name name of spatial network to use spat_network_color color of spatial network show_spatial_grid show spatial grid spat_grid_name name of spatial grid to use spat_grid_color color of spatial grid show_other_cells display not selected cells other_cell_color color of not selected cells dim_other_point_size size of not selected dim cells spat_other_point_size size of not selected spat cells spat_other_cells_alpha alpha of not selected spat cells dim_show_legend show legend of dimension reduction plot spat_show_legend show legend of spatial plot legend_text size of legend text legend_symbol_size size of legend symbols dim_background_color background color of points in dim. reduction space spat_background_color background color of spatial points size of axis text axis_text axis_title size of axis title show_plot show plot return_plot return ggplot object save_plot directly save the plot [boolean] list of saving parameters from all_plots_save_function save_param default_save_name default save name for saving, don't change, change save_name in save_param 236 spatDimPlot2D

Details

Description of parameters.

Value

ggplot

See Also

spatDimPlot2D and spatDimPlot3D for 3D visualization.

Examples

```
spatDimPlot(gobject)
```

spatDimPlot2D

spatDimPlot2D

Description

Visualize cells according to spatial AND dimension reduction coordinates 2D

```
spatDimPlot2D(
 gobject,
 plot_alignment = c("vertical", "horizontal"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2\_to\_use = 2,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_color = NULL,
  color_as_factor = T,
 cell_color_code = NULL,
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
  gradient_limits = NULL,
  select_cell_groups = NULL,
 select_cells = NULL,
 dim_point_shape = c("border", "no_border"),
 dim_point_size = 1,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  spat_point_shape = c("border", "no_border"),
  spat_point_size = 1,
  spat_point_border_col = "black",
  spat_point_border_stroke = 0.1,
 dim_show_cluster_center = F,
```

spatDimPlot2D 237

```
dim_show_center_label = T,
 dim_center_point_size = 4,
 dim_center_point_border_col = "black",
 dim_center_point_border_stroke = 0.1,
  dim_label_size = 4,
 dim_label_fontface = "bold",
  spat_show_cluster_center = F,
  spat_show_center_label = F,
  spat_center_point_size = 4,
  spat_label_size = 4,
  spat_label_fontface = "bold",
  show_NN_network = F,
  nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 nn_network_alpha = 0.05,
  show_spatial_network = F,
  spat_network_name = "spatial_network",
  spat_network_color = "blue",
  spat_network_alpha = 0.5,
  show_spatial_grid = F,
  spat_grid_name = "spatial_grid",
  spat_grid_color = "blue",
  show_other_cells = T,
 other_cell_color = "lightgrey",
 dim_other_point_size = 1,
  spat_other_point_size = 1,
  spat_other_cells_alpha = 0.5,
  dim\_show\_legend = F,
  spat_show_legend = F,
  legend_text = 8,
  legend_symbol_size = 1,
 dim_background_color = "white",
  spat_background_color = "white",
 axis_text = 8,
 axis_title = 8,
  show_plot = NA,
 return_plot = NA,
  save_plot = NA,
 save_param = list(),
 default_save_name = "spatDimPlot2D"
)
```

Arguments

238 spatDimPlot2D

sdimx = spatial dimension to use on x-axis = spatial dimension to use on y-axis sdimy spat_enr_names names of spatial enrichment results to include cell_color color for cells (see details) color_as_factor convert color column to factor cell_color_code named vector with colors cell_color_gradient vector with 3 colors for numeric data gradient_midpoint midpoint for color gradient gradient_limits vector with lower and upper limits select_cell_groups select subset of cells/clusters based on cell_color parameter select subset of cells based on cell IDs select_cells dim_point_shape point with border or not (border or no border) dim_point_size size of points in dim. reduction space dim_point_border_col border color of points in dim. reduction space dim_point_border_stroke border stroke of points in dim. reduction space spat_point_shape point with border or not (border or no_border) spat_point_size size of spatial points spat_point_border_col border color of spatial points spat_point_border_stroke border stroke of spatial points dim_show_cluster_center show the center of each cluster dim_show_center_label provide a label for each cluster dim_center_point_size size of the center point dim_center_point_border_col border color of center point dim_center_point_border_stroke stroke size of center point dim_label_size size of the center label dim_label_fontface font of the center label spat_show_cluster_center show the center of each cluster

spat_show_center_label provide a label for each cluster spat_center_point_size size of the center point spat_label_size size of the center label spat_label_fontface font of the center label show_NN_network show underlying NN network nn_network_to_use type of NN network to use (kNN vs sNN) name of NN network to use, if show_NN_network = TRUE network_name nn_network_alpha column to use for alpha of the edges show_spatial_network show spatial network spat_network_name name of spatial network to use spat_network_color color of spatial network show_spatial_grid show spatial grid spat_grid_name name of spatial grid to use spat_grid_color color of spatial grid show_other_cells display not selected cells other_cell_color color of not selected cells dim_other_point_size size of not selected dim cells spat_other_point_size size of not selected spat cells spat_other_cells_alpha alpha of not selected spat cells dim_show_legend show legend of dimension reduction plot spat_show_legend show legend of spatial plot legend_text size of legend text legend_symbol_size size of legend symbols dim_background_color background color of points in dim. reduction space spat_background_color background color of spatial points

size of axis text

axis_text

240 spatDimPlot3D

```
axis_title size of axis title

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_param list of saving parameters from all_plots_save_function

default_save_name

default save name for saving, don't change, change save_name in save_param
```

Details

Description of parameters.

Value

ggplot

See Also

```
spatDimPlot3D
```

Examples

```
spatDimPlot2D(gobject)
```

spatDimPlot3D

spatDimPlot3D

Description

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

```
spatDimPlot3D(
 gobject,
 plot_alignment = c("horizontal", "vertical"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 dim3_to_use = 3,
 sdimx = "sdimx",
  sdimy = "sdimy",
 sdimz = "sdimz",
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
```

spatDimPlot3D 241

```
label_size = 16,
     select_cell_groups = NULL,
     select_cells = NULL,
     show_other_cells = T,
     other_cell_color = "lightgrey",
     other_point_size = 1.5,
     cell_color = NULL,
     color_as_factor = T,
     cell_color_code = NULL,
     dim_point_size = 3,
     nn_network_alpha = 0.5,
     show_spatial_network = F,
     spatial_network_name = "spatial_network",
     network_color = "lightgray",
     spatial_network_alpha = 0.5,
     show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
     spatial_grid_color = NULL,
     spatial_grid_alpha = 0.5,
     spatial_point_size = 3,
     axis_scale = c("cube", "real", "custom"),
     custom_ratio = NULL,
     x_ticks = NULL,
     y_ticks = NULL,
     z_ticks = NULL,
     legend_text_size = 12,
     show_plot = NA,
     return_plot = NA,
     save_plot = NA,
     save_param = list(),
     default_save_name = "spatDimPlot3D"
Arguments
   gobject
                   giotto object
   plot_alignment direction to align plot
   dim_reduction_to_use
                   dimension reduction to use
   dim_reduction_name
                   dimension reduction name
   dim1_to_use
                   dimension to use on x-axis
   dim2_to_use
                   dimension to use on y-axis
```

```
dim3_to_use dimension to use on y-axis

sdimx = spatial dimension to use on x-axis

sdimy = spatial dimension to use on y-axis

sdimz = spatial dimension to use on y-axis

sdimz = spatial dimension to use on z-axis

show_NN_network

show underlying NN network

nn_network_to_use

type of NN network to use (kNN vs sNN)
```

242 spatDimPlot3D

network_name name of NN network to use, if show NN network = TRUE show_cluster_center show the center of each cluster show_center_label provide a label for each cluster center_point_size size of the center point size of the center label label_size select_cell_groups select subset of cells/clusters based on cell_color parameter select_cells select subset of cells based on cell IDs show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size size of not selected cells cell_color color for cells (see details) color_as_factor convert color column to factor cell_color_code named vector with colors dim_point_size size of points in dim. reduction space nn_network_alpha column to use for alpha of the edges show_spatial_network show spatial network spatial_network_name name of spatial network to use spatial_network_alpha alpha of spatial network show_spatial_grid show spatial grid spatial_grid_name name of spatial grid to use spatial_grid_color color of spatial grid spatial_point_size size of spatial points show_plot show plot return ggplot object return_plot save_plot directly save the plot [boolean] list of saving parameters from all_plots_save_function save_param default_save_name default save name for saving, don't change, change save_name in save_param dim_point_border_col border color of points in dim. reduction space

spatGenePlot 243

Details

Description of parameters.

Value

plotly

Examples

```
spatDimPlot3D(gobject)
```

spatGenePlot

spatGenePlot

Description

Visualize cells and gene expression according to spatial coordinates

```
spatGenePlot(
 gobject,
  expression_values = c("normalized", "scaled", "custom"),
 genes,
 genes_high_color = "darkred",
 genes_mid_color = "white",
 genes_low_color = "darkblue",
  show_network = F,
 network_color = NULL,
 spatial_network_name = "spatial_network",
 edge_alpha = NULL,
 show\_grid = F,
 grid_color = NULL,
 spatial_grid_name = "spatial_grid",
 midpoint = 0,
 scale_alpha_with_expression = FALSE,
 point_size = 1,
 point_border_col = "black",
```

244 spatGenePlot

```
point_border_stroke = 0.1,
      show_legend = T,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatGenePlot"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
    genes
                     genes to show
    genes_high_color
                     color represents high gene expression
    genes_mid_color
                     color represents middle gene expression
    genes_low_color
                     color represents low gene expression
                     show underlying spatial network
    show_network
    network_color
                     color of spatial network
    spatial_network_name
                     name of spatial network to use
                     show spatial grid
    show_grid
    grid_color
                     color of spatial grid
    spatial_grid_name
                     name of spatial grid to use
                     expression midpoint
    midpoint
    scale_alpha_with_expression
                     scale expression with ggplot alpha parameter
                     size of point (cell)
    point_size
    point_border_col
                     color of border around points
    point_border_stroke
                     stroke size of border around points
    show_legend
                     show legend
    cow_n_col
                     cowplot param: how many columns
                     cowplot param: relative height
    cow_rel_h
                     cowplot param: relative width
    cow_rel_w
    cow_align
                     cowplot param: how to align
    show_plot
                     show plots
```

spatGenePlot2D 245

Details

Description of parameters.

Value

ggplot

See Also

```
spatGenePlot3D and spatGenePlot2D
```

Examples

```
spatGenePlot(gobject)
```

spatGenePlot2D

spatGenePlot2D

Description

Visualize cells and gene expression according to spatial coordinates

```
spatGenePlot2D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 genes_high_color = "darkred",
 genes_mid_color = "white",
 genes_low_color = "darkblue",
  show_network = F,
 network_color = NULL,
 spatial_network_name = "spatial_network",
 edge_alpha = NULL,
  show_grid = F,
 grid_color = NULL,
  spatial_grid_name = "spatial_grid",
 midpoint = 0,
  scale_alpha_with_expression = FALSE,
 point_size = 1,
 point_border_col = "black",
 point_border_stroke = 0.1,
```

246 spatGenePlot2D

```
show_legend = T,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatGenePlot2D"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
                     genes to show
    genes
    genes_high_color
                     color represents high gene expression
    genes_mid_color
                     color represents middle gene expression
    genes_low_color
                     color represents low gene expression
                     show underlying spatial network
    show_network
                     color of spatial network
    network_color
    spatial_network_name
                     name of spatial network to use
    show_grid
                     show spatial grid
    grid_color
                     color of spatial grid
    spatial_grid_name
                     name of spatial grid to use
                     expression midpoint
    midpoint
    scale_alpha_with_expression
                     scale expression with ggplot alpha parameter
    point_size
                     size of point (cell)
    point_border_col
                     color of border around points
    point_border_stroke
                     stroke size of border around points
    show_legend
                     show legend
    cow_n_col
                     cowplot param: how many columns
    cow_rel_h
                     cowplot param: relative height
    cow_rel_w
                     cowplot param: relative width
                     cowplot param: how to align
    cow_align
    show_plot
                     show plots
```

return ggplot object

return_plot

spatGenePlot3D 247

Details

Description of parameters.

Value

ggplot

See Also

```
spatGenePlot3D
```

Examples

```
spatGenePlot2D(gobject)
```

spatGenePlot3D

spatGenePlot3D

Description

Visualize cells and gene expression according to spatial coordinates

```
spatGenePlot3D(
 gobject,
 expression_values = c("normalized", "scaled", "custom"),
 genes,
  show_network = F,
 network_color = NULL,
  spatial_network_name = "spatial_network",
 edge_alpha = NULL,
  show\_grid = F,
 cluster_column = NULL,
 select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
 genes_low_color = "blue",
  spatial_grid_name = "spatial_grid",
 point_size = 2,
```

248 spatGenePlot3D

```
show_legend = T,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatGenePlot3D"
    )
Arguments
    gobject
                     giotto object
    expression_values
                     gene expression values to use
    genes
                     genes to show
                     show underlying spatial network
    show_network
                     color of spatial network
    network_color
    spatial_network_name
                     name of spatial network to use
                     show spatial grid
    show_grid
    genes_high_color
                     color represents high gene expression
    genes_mid_color
                     color represents middle gene expression
    genes_low_color
                     color represents low gene expression
    spatial_grid_name
                     name of spatial grid to use
    point_size
                     size of point (cell)
    show_legend
                     show legend
    show_plot
                     show plots
    return_plot
                     return ggplot object
    save_plot
                     directly save the plot [boolean]
                     list of saving parameters from all_plots_save_function
    save_param
    default_save_name
                     default save name for saving, don't change, change save_name in save_param
    grid_color
                     color of spatial grid
    midpoint
                     expression midpoint
    scale_alpha_with_expression
```

scale expression with ggplot alpha parameter

parameters for cowplot::save_plot()

Spatial_AEH 249

Details

Description of parameters.

Value

ggplot

Examples

```
spatGenePlot3D(gobject)
```

Spatial_AEH

Spatial_AEH

Description

calculate automatic expression histology with spatialDE method

Usage

```
Spatial_AEH(
  gobject = NULL,
  results = NULL,
  pattern_num = 5,
  1 = 1.05,
  show_AEH = T,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_alpha = 1,
  low_color = "blue"
  mid_color = "white",
  high_color = "red",
  midpoint = 0,
  python_path = NULL
)
```

Arguments

gobject Giotto object

results output from spatial_DE

pattern_num the number of gene expression patterns

show_AEH show AEH plot

python_path specify specific path to python if required

Details

Description.

250 Spatial_DE

Value

a list or a dataframe of SVs

Examples

```
{\tt Spatial\_AEH(gobject)}
```

Spatial_DE

Spatial_DE

Description

calculate spatial varible genes with spatialDE method

Usage

```
Spatial_DE(
  gobject = NULL,
  show_plot = T,
  size = c(4, 2, 1),
  color = c("blue", "green", "red"),
  sig_alpha = 0.5,
  unsig_alpha = 0.5,
  python_path = NULL
)
```

Arguments

```
gobject Giotto object
show_plot show FSV plot
python_path specific path to python if required
```

Details

Description.

Value

```
a list or a dataframe of SVs
```

Examples

```
Spatial_DE(gobject)
```

spatNetwDistributions 251

```
{\tt spatNetwDistributions} \ \textit{spatNetwDistributionsDistance}
```

Description

This function return histograms displaying the distance distribution for each spatial k-neighbor

Usage

```
spatNetwDistributions(
  gobject,
  spatial_network_name = "spatial_network",
  distribution = c("distance", "k_neighbors"),
  hist_bins = 30,
  test_distance_limit = NULL,
  ncol = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "spatNetwDistributions"
)
```

Arguments

```
gobject
                  Giotto object
spatial_network_name
                  name of spatial network
distribution
                  show the distribution of cell-to-cell distance or number of k neighbors
hist_bins
                  number of binds to use for the histogram
test_distance_limit
                  effect of different distance threshold on k-neighbors
                  number of columns to visualize the histograms in
ncol
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_param
                  list of saving parameters from all_plots_save_function
default_save_name
                  default save name for saving, alternatively change save_name in save_param
```

Details

The **distance** option shows the spatial distance distribution for each nearest neighbor rank (1st, 2nd, 3th, ... neigbor). With this option the user can also test the effect of a distance limit on the spatial network. This distance limit can be used to remove neigbor cells that are considered to far away. The **k_neighbors** option shows the number of k neighbors distribution over all cells.

Value

```
ggplot plot
```

Examples

```
spatNetwDistributionsDistance(gobject)
```

```
spat {\tt NetwDistributionsDistance} \\ spat {\tt NetwDistributionsDistance}
```

Description

This function return histograms displaying the distance distribution for each spatial k-neighbor

Usage

```
spatNetwDistributionsDistance(
  gobject,
  spatial_network_name = "spatial_network",
  hist_bins = 30,
  test_distance_limit = NULL,
  ncol = 1,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "spatNetwDistributionsDistance")
```

Arguments

```
gobject
                  Giotto object
spatial_network_name
                  name of spatial network
                  number of binds to use for the histogram
hist_bins
test_distance_limit
                  effect of different distance threshold on k-neighbors
ncol
                  number of columns to visualize the histograms in
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, alternatively change save_name in save_param
```

Value

ggplot plot

Examples

```
spatNetwDistributionsDistance(gobject)
```

```
spat Netw Distributions Kneighbors \\ spat Netw Distributions Kneighbors
```

Description

This function returns a histogram displaying the number of k-neighbors distribution for each cell

Usage

```
spatNetwDistributionsKneighbors(
  gobject,
  spatial_network_name = "spatial_network",
  hist_bins = 30,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "spatNetwDistributionsKneighbors")
```

Arguments

```
Giotto object
gobject
spatial_network_name
                  name of spatial network
hist_bins
                  number of binds to use for the histogram
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
                  default save name for saving, alternatively change save_name in save_param
```

Value

ggplot plot

```
spatNetwDistributionsKneighbors(gobject)
```

254 spatPlot

spatPlot

spatPlot

Description

Visualize cells according to spatial coordinates

```
spatPlot(
 gobject,
 group_by = NULL,
 group_by_subset = NULL,
  sdimx = "sdimx",
  sdimy = "sdimy"
  spat_enr_names = NULL,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
 select_cell_groups = NULL,
  select_cells = NULL,
 point_shape = c("border", "no_border"),
 point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_cluster_center = F,
  show\_center\_label = F,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
  show_grid = F,
  spatial_grid_name = "spatial_grid",
 grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
 other_cells_alpha = 0.1,
  coord_fix_ratio = NULL,
  title = NULL,
  show_legend = T,
  legend_text = 8,
  legend_symbol_size = 1,
```

spatPlot 255

```
background_color = "white",
      axis_text = 8,
      axis_title = 8,
      cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
      cow_align = "h",
      show_plot = NA,
      return_plot = NA,
      save_plot = NA,
      save_param = list(),
      default_save_name = "spatPlot"
    )
Arguments
    gobject
                     giotto object
    group_by_subset
                     subset the group_by factor column
                     x-axis dimension name (default = 'sdimx')
    sdimx
                     y-axis dimension name (default = 'sdimy')
    sdimy
    spat_enr_names names of spatial enrichment results to include
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    cell_color_gradient
                     vector with 3 colors for numeric data
    gradient_midpoint
                     midpoint for color gradient
    gradient_limits
                     vector with lower and upper limits
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
   point_shape
                     point with border or not (border or no_border)
    point_size
                     size of point (cell)
    point_border_col
                     color of border around points
   point_border_stroke
                     stroke size of border around points
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
   center_point_size
```

size of center points

256 spatPlot

label_size size of labels label_fontface font of labels show_network show underlying spatial network spatial_network_name name of spatial network to use network_color color of spatial network network_alpha alpha of spatial network show_grid show spatial grid spatial_grid_name name of spatial grid to use grid_color color of spatial grid show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size point size of not selected cells other_cells_alpha alpha of not selected cells coord_fix_ratio fix ratio between x and y-axis title title of plot show_legend show legend legend_text size of legend text legend_symbol_size size of legend symbols background_color color of plot background axis_text size of axis text axis_title size of axis title cow_n_col cowplot param: how many columns cow_rel_h cowplot param: relative height cowplot param: relative width cow_rel_w cow_align cowplot param: how to align show_plot show plot return_plot return ggplot object save_plot directly save the plot [boolean] list of saving parameters from all_plots_save_function save_param default_save_name default save name for saving, don't change, change save_name in save_param create multiple plots based on cell annotation column groub_by

Details

Description of parameters.

spatPlot2D 257

Value

ggplot

See Also

```
spatPlot3D
```

Examples

```
spatPlot(gobject)
```

spatPlot2D

spatPlot2D

Description

Visualize cells according to spatial coordinates

```
spatPlot2D(
 gobject,
 group_by = NULL,
 group_by_subset = NULL,
  sdimx = "sdimx",
 sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_color = NULL,
 color_as_factor = T,
 cell_color_code = NULL,
 cell_color_gradient = c("blue", "white", "red"),
  gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
 point_shape = c("border", "no_border"),
 point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
  show_cluster_center = F,
  show_center_label = F,
 center_point_size = 4,
 center_point_border_col = "black",
  center_point_border_stroke = 0.1,
 label_size = 4,
 label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
  show\_grid = F,
```

258 spatPlot2D

```
spatial_grid_name = "spatial_grid",
      grid_color = NULL,
      show_other_cells = T,
     other_cell_color = "lightgrey",
     other_point_size = 1,
     other_cells_alpha = 0.1,
      coord_fix_ratio = NULL,
      title = NULL,
      show_legend = T,
     legend_text = 8,
     legend_symbol_size = 1,
     background_color = "white",
      axis_text = 8,
     axis_title = 8,
     cow_n_col = 2,
      cow_rel_h = 1,
      cow_rel_w = 1,
     cow_align = "h",
      show_plot = NA,
     return_plot = NA,
     save_plot = NA,
     save_param = list(),
     default_save_name = "spatPlot2D"
Arguments
   gobject
                    giotto object
   group_by_subset
                    subset the group_by factor column
   sdimx
                    x-axis dimension name (default = 'sdimx')
                    y-axis dimension name (default = 'sdimy')
   sdimy
   spat_enr_names names of spatial enrichment results to include
   cell_color
                    color for cells (see details)
```

convert color column to factor

vector with 3 colors for numeric data

vector with lower and upper limits

select subset of cells based on cell IDs

point with border or not (border or no_border)

select subset of cells/clusters based on cell_color parameter

named vector with colors

midpoint for color gradient

size of point (cell)

color_as_factor

cell_color_code

cell_color_gradient

gradient_midpoint

gradient_limits

select_cells

point_shape
point_size

select_cell_groups

spatPlot2D 259

point_border_col color of border around points point_border_stroke stroke size of border around points show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points label_size size of labels label_fontface font of labels show underlying spatial network show_network spatial_network_name name of spatial network to use network_color color of spatial network network_alpha alpha of spatial network show_grid show spatial grid spatial_grid_name name of spatial grid to use grid_color color of spatial grid show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size point size of not selected cells other_cells_alpha alpha of not selected cells coord_fix_ratio fix ratio between x and y-axis title title of plot show_legend show legend legend_text size of legend text legend_symbol_size size of legend symbols background_color color of plot background axis_text size of axis text axis_title size of axis title cowplot param: how many columns cow_n_col cowplot param: relative height cow_rel_h cowplot param: relative width cow_rel_w cow_align cowplot param: how to align

show_plot

show plot

260 spatPlot2D_single

Details

Description of parameters.

Value

ggplot

See Also

```
spatPlot3D
```

Examples

```
spatPlot2D(gobject)
```

Description

Visualize cells according to spatial coordinates

```
spatPlot2D_single(
 gobject,
  sdimx = "sdimx",
  sdimy = "sdimy",
  spat_enr_names = NULL,
 cell_color = NULL,
 color_as_factor = T,
  cell_color_code = NULL,
 cell_color_gradient = c("blue", "white", "red"),
 gradient_midpoint = NULL,
 gradient_limits = NULL,
  select_cell_groups = NULL,
 select_cells = NULL,
 point_shape = c("border", "no_border"),
 point_size = 3,
 point_border_col = "black",
 point_border_stroke = 0.1,
 show_cluster_center = F,
  show_center_label = F,
```

spatPlot2D_single 261

```
center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  show_network = F,
  spatial_network_name = "spatial_network",
 network_color = NULL,
 network_alpha = 1,
  show_grid = F,
  spatial_grid_name = "spatial_grid",
  grid_color = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 1,
  other_cells_alpha = 0.1,
  coord_fix_ratio = NULL,
  title = NULL,
  show_legend = T,
  legend_text = 8,
 legend_symbol_size = 1,
 background_color = "white",
 axis_text = 8,
  axis_title = 8,
  show_plot = NA,
 return_plot = NA,
  save_plot = NA,
 save_param = list(),
 default_save_name = "spatPlot2D_single"
)
```

Arguments

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
                  y-axis dimension name (default = 'sdimy')
sdimy
spat_enr_names names of spatial enrichment results to include
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
cell_color_gradient
                  vector with 3 colors for numeric data
gradient_midpoint
                  midpoint for color gradient
gradient_limits
                  vector with lower and upper limits
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
```

262 spatPlot2D_single

point_shape point with border or not (border or no border) point_size size of point (cell) point_border_col color of border around points point_border_stroke stroke size of border around points show_cluster_center plot center of selected clusters show_center_label plot label of selected clusters center_point_size size of center points size of labels label_size label_fontface font of labels show_network show underlying spatial network spatial_network_name name of spatial network to use network_color color of spatial network network_alpha alpha of spatial network show_grid show spatial grid spatial_grid_name name of spatial grid to use color of spatial grid grid_color show_other_cells display not selected cells other_cell_color color of not selected cells other_point_size point size of not selected cells other_cells_alpha alpha of not selected cells coord_fix_ratio fix ratio between x and y-axis title title of plot show legend show_legend legend_text size of legend text legend_symbol_size size of legend symbols background_color color of plot background axis_text size of axis text size of axis title axis_title show_plot show plot return_plot return ggplot object save_plot directly save the plot [boolean] list of saving parameters from all_plots_save_function save_param default_save_name

default save name for saving, don't change, change save_name in save_param

spatPlot3D 263

Details

Description of parameters.

Value

ggplot

See Also

spatPlot3D

Examples

```
spatPlot2D_single(gobject)
```

spatPlot3D

spatPlot3D

Description

Visualize cells according to spatial coordinates

```
spatPlot3D(
 gobject,
  sdimx = "sdimx",
 sdimy = "sdimy",
  sdimz = "sdimz",
 point_size = 3,
 cell_color = NULL,
 cell_color_code = NULL,
  select_cell_groups = NULL,
 select_cells = NULL,
  show_other_cells = T,
 other_cell_color = "lightgrey",
 other_point_size = 0.5,
 show_network = F,
 network_color = NULL,
 network_alpha = 1,
 other_cell_alpha = 0.5,
 spatial_network_name = "spatial_network",
 show_grid = F,
 grid_color = NULL,
 spatial_grid_name = "spatial_grid",
  title = "",
  show_legend = T,
 axis_scale = c("cube", "real", "custom"),
 custom_ratio = NULL,
 x_{ticks} = NULL,
 y_{ticks} = NULL,
```

264 spatPlot3D

```
z_ticks = NULL,
show_plot = NA,
return_plot = NA,
save_plot = NA,
save_param = list(),
default_save_name = "spat3D"
)
```

Arguments

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
                  z-axis dimension name (default = 'sdimy')
sdimz
point_size
                  size of point (cell)
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
select_cells
                  select subset of cells based on cell IDs
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
                  title of plot
title
show_legend
                  show legend
axis_scale
                  the way to scale the axis
custom_ratio
                  customize the scale of the plot
x_ticks
                  set the number of ticks on the x-axis
                  set the number of ticks on the y-axis
y_ticks
z_ticks
                  set the number of ticks on the z-axis
                  show plot
show_plot
                  return ggplot object
return_plot
                  directly save the plot [boolean]
save_plot
                  list of saving parameters from all_plots_save_function
save_param
default_save_name
```

default save name for saving, don't change, change save_name in save_param

Details

Description of parameters.

Value

ggplot

Examples

```
spatPlot3D(gobject)
```

```
specific Cell Cell communication Scores\\ specific Cell Cell communication Scores
```

Description

Specific Cell-Cell communication scores based on spatial expression of interacting cells

Usage

```
specificCellCellcommunicationScores(
  gobject,
  spatial_network_name = "spatial_network",
  cluster_column = "cell_types",
  random_iter = 100,
  cell_type_1 = "astrocyte",
  cell_type_2 = "endothelial",
  gene_set_1,
  gene_set_2,
  log2FC_addendum = 0.1,
  min_observations = 2,
  verbose = T
)
```

Arguments

```
gobject
                  giotto object to use
spatial_network_name
                  spatial network to use for identifying interacting cells
cluster_column cluster column with cell type information
random_iter
                  number of iterations
cell_type_1
                  first cell type
cell_type_2
                  second cell type
                  first specific gene set from gene pairs
gene_set_1
                  second specific gene set from gene pairs
gene_set_2
log2FC\_addendum
                  addendum to add when calculating log2FC
```

min_observations

minimum number of interactions needed to be considered

verbose verbose

Details

Statistical framework to identify if pairs of genes (such as ligand-receptor combinations) are expressed at higher levels than expected based on a reshuffled null distribution of gene expression values in cells that are spatially in proximity to eachother.. More details will follow soon.

Value

Cell-Cell communication scores for gene pairs based on spatial interaction

Examples

```
{\tt specificCellCellcommunicationScores(gobject)}
```

Description

Merge selected clusters based on pairwise correlation scores and size of cluster.

Usage

```
split_dendrogram_in_two(dend)
```

Arguments

dend

dendrogram object

Value

list of two dendrograms and height of node

```
split_dendrogram_in_two(dend)
```

stitchFieldCoordinates 267

```
stitchFieldCoordinates
```

stitchFieldCoordinates

Description

Helper function to stitch field coordinates together to form one complete picture

Usage

```
stitchFieldCoordinates(
  location_file,
  offset_file,
  cumulate_offset_x = F,
  cumulate_offset_y = F,
  field_col = "Field of View",
  X_coord_col = "X",
  Y_coord_col = "Y",
  reverse_final_x = F,
  reverse_final_y = T
)
```

Arguments

```
location_file location dataframe with X and Y coordinates
offset_file
                  dataframe that describes the offset for each field (see details)
cumulate_offset_x
                  (boolean) Do the x-axis offset values need to be cumulated?
cumulate_offset_y
                  (boolean) Do the y-axis offset values need to be cumulated?
field_col
                  column that indicates the field within the location_file
X_coord_col
                  column that indicates the x coordinates
Y_coord_col
                  column that indicates the x coordinates
reverse_final_x
                  (boolean) Do the final x coordinates need to be reversed?
reverse_final_y
                  (boolean) Do the final y coordinates need to be reversed?
```

Details

Stitching of fields:

- 1. have cell locations: at least 3 columns: field, X, Y
- 2. create offset file: offset file has 3 columns: field, x_offset, y_offset
- 3. create new cell location file by stitching original cell locations with stitchFieldCoordinates
- 4. provide new cell location file to createGiottoObject

Value

Updated location dataframe with new X ['X_final'] and Y ['Y_final'] coordinates

268 subClusterCells

Examples

```
stitchFieldCoordinates(gobject)
```

subClusterCells

subClusterCells

Description

subcluster cells

Usage

```
subClusterCells(
  gobject,
 name = "sub_clus",
 cluster_method = c("leiden", "louvain_community", "louvain_multinet"),
 cluster_column = NULL,
 selected_clusters = NULL,
 hvg_param = list(reverse_log_scale = T, difference_in_variance = 1, expression_values
   = "normalized"),
 hvg_min_perc_cells = 5,
 hvg_mean_expr_det = 1,
 use_all_genes_as_hvg = FALSE,
 min_nr_of_hvg = 5,
 pca_param = list(expression_values = "normalized", scale_unit = T),
 nn_param = list(dimensions_to_use = 1:20),
 k_neighbors = 10,
 resolution = 1,
  gamma = 1,
 omega = 1,
 python_path = NULL,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 return_gobject = TRUE,
 verbose = T
)
```

Arguments

```
gobject giotto object

name name for new clustering result

cluster_method clustering method to use

cluster_column cluster column to subcluster

selected_clusters

only do subclustering on these clusters

hvg_param parameters for calculateHVG

hvg_min_perc_cells

threshold for detection in min percentage of cells
```

subClusterCells 269

hvg_mean_expr_det

threshold for mean expression level in cells with detection

use_all_genes_as_hvg

forces all genes to be HVG and to be used as input for PCA

min_nr_of_hvg minimum number of HVG, or all genes will be used as input for PCA

pca_param parameters for runPCA

nn_param parameters for parameters for createNearestNetwork

k_neighbors number of k for createNearestNetwork

resolution resolution gamma gamma omega omega

python_path specify specific path to python if required

nn_network_to_use

type of NN network to use (kNN vs sNN)

network_name name of NN network to use

return_gobject boolean: return giotto object (default = TRUE)

verbose verbose

Details

This function performs subclustering on selected clusters. The systematic steps are:

- 1. subset Giotto object
- 2. identify highly variable genes
- 3. run PCA
- 4. create nearest neighbouring network
- 5. do clustering

Value

giotto object with new subclusters appended to cell metadata

See Also

doLouvainCluster_multinet, doLouvainCluster_community and @seealso doLeidenCluster

Examples

subClusterCells(gobject)

270 subsetGiottoLocs

subsetGiotto

subsetGiot to

Description

subsets Giotto object including previous analyses.

Usage

```
subsetGiotto(gobject, cell_ids = NULL, gene_ids = NULL, verbose = FALSE)
```

Arguments

```
gobject giotto object
cell_ids cell IDs to keep
gene_ids gene IDs to keep
verbose be verbose
```

Value

giotto object

Examples

```
subsetGiotto(gobject)
```

 ${\tt subsetGiottoLocs}$

subsetGiottoLocs

Description

subsets Giotto object based on spatial locations

```
subsetGiottoLocs(
  gobject,
  x_max = NULL,
  x_min = NULL,
  y_max = NULL,
  y_min = NULL,
  z_max = NULL,
  z_min = NULL,
  return_gobject = T,
  verbose = FALSE
)
```

trendSceek 271

Arguments

gobject	giotto object
x_max	maximum x-coordinate
x_min	minimum x-coordinate
y_max	maximum y-coordinate
y_min	minimum y-coordinate
z_max	maximum z-coordinate
z_min	minimum z-coordinate
return_gobject	return Giotto object

Details

if return_gobject = FALSE, then a filtered combined metadata data.table will be returned

Value

giotto object

Examples

subsetGiottoLocs(gobject)

trendSceek trendSceek

Description

Compute spatial variable genes with trendsceek method

Usage

```
trendSceek(
  gobject,
  expression_values = c("normalized", "raw"),
  subset_genes = NULL,
  nrand = 100,
  ncores = 8,
  ...
)
```

Arguments

```
gobject Giotto object
expression_values
gene expression values to use
subset_genes subset of genes to run trendsceek on
nrand An integer specifying the number of random resamplings of the mark distribution as to create the null-distribution.

ncores An integer specifying the number of cores to be used by BiocParallel
... Additional parameters to the trendsceek_test function
```

272 viewHMRFresults

Details

This function is a wrapper for the trendsceek_test method implemented in the trendsceek package

Value

data.frame with trendsceek spatial genes results

Examples

```
trendSceek(gobject)
```

viewHMRFresults

viewHMRFresults

Description

View results from doHMRF.

Usage

```
viewHMRFresults(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

Arguments

```
gobject giotto object
```

HMRF output from doHMRF k number of HMRF domains

betas_to_view results from different betas that you want to view

... paramters to visPlot()

Details

Description ...

Value

spatial plots with HMRF domains

See Also

```
visPlot
```

```
viewHMRFresults(gobject)
```

viewHMRFresults2D 273

viewHMRFresults2D

viewHMRFresults2D

Description

View results from doHMRF.

Usage

```
viewHMRFresults2D(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

Arguments

```
gobject giotto object
```

HMRF output from doHMRF k number of HMRF domains

number of fivire domains

betas_to_view results from different betas that you want to view

... paramters to visPlot()

Details

Description ...

Value

spatial plots with HMRF domains

See Also

```
spatPlot2D
```

```
viewHMRFresults2D(gobject)
```

274 viewHMRFresults3D

viewHMRFresults3D

viewHMRFresults3D

Description

View results from doHMRF.

Usage

```
viewHMRFresults3D(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  third_dim = NULL,
  ...
)
```

Arguments

```
gobject giotto object
```

HMRF output from doHMRF k number of HMRF domains

betas_to_view results from different betas that you want to view

... paramters to visPlot()

Details

Description ...

Value

spatial plots with HMRF domains

See Also

```
spatPlot3D
```

```
viewHMRFresults3D(gobject)
```

violinPlot 275

violinPlot

violinPlot

Description

Creates violinplot for selected clusters

Usage

```
violinPlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  cluster_column,
  cluster_custom_order = NULL,
  color_violin = c("genes", "cluster"),
  cluster_color_code = NULL,
  strip_position = c("top", "right", "left", "bottom"),
  strip\_text = 7,
  axis_text_x_size = 10,
  axis_text_y_size = 6,
  show_plot = NA,
  return_plot = NA,
  save_plot = NA,
  save_param = list(),
  default_save_name = "violinPlot"
)
```

Arguments

```
gobject
                  giotto object
expression_values
                  expression values to use
                  genes to plot
genes
cluster_column name of column to use for clusters
cluster_custom_order
                  custom order of clusters
color_violin
                  color violin according to genes or clusters
cluster_color_code
                  color code for clusters
strip_position position of gene labels
strip_text
                  size of strip text
\verb"axis_text_x_size"
                  size of x-axis text
axis_text_y_size
                  size of y-axis text
show_plot
                  show plot
return_plot
                  return ggplot object
```

276 visDimGenePlot

Value

ggplot

Examples

```
violinPlot(gobject)
```

visDimGenePlot

visDimGenePlot

Description

Visualize cells and gene expression according to dimension reduction coordinates

```
visDimGenePlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h"
  show_legend = T,
  plot_method = c("ggplot", "plotly"),
  show_plots = F
)
```

visDimGenePlot 277

Arguments

gobject giotto object

expression_values

gene expression values to use

genes genes to show

dim_reduction_to_use

dimension reduction to use

dim_reduction_name

dimension reduction name

dim1_to_use dimension to use on x-axis dim2_to_use dimension to use on y-axis dim3_to_use dimension to use on z-axis

show_NN_network

show underlying NN network

nn_network_to_use

type of NN network to use (kNN vs sNN)

 $network_name$ name of NN network to use, if $show_NN_network = TRUE$

edge_alpha column to use for alpha of the edges

scale_alpha_with_expression

scale expression with ggplot alpha parameter

point_size size of point (cell)

point_border_col

color of border around points

point_border_stroke

stroke size of border around points

midpoint size of point (cell)

cow_n_col cowplot param: how many columns
cow_rel_h cowplot param: relative height
cow_rel_w cowplot param: relative width
cow_align cowplot param: how to align

show_legend show legend show_plots show plots

Details

Description of parameters.

Value

ggplot

Examples

visDimGenePlot(gobject)

Description

Visualize cells and gene expression according to dimension reduction coordinates

Usage

```
visDimGenePlot_2D_ggplot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  genes_high_color = "red",
  genes_mid_color = "white",
  genes_low_color = "blue",
  point_border_col = "black",
  point_border_stroke = 0.1,
  midpoint = 0,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_legend = T,
  show_plots = F
```

Arguments

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
dim_reduction_to_use
dimension reduction to use
dim_reduction_name
dimension reduction name
dim1_to_use
dimension to use on x-axis
```

```
dimension to use on y-axis
dim2_to_use
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
edge_alpha
                 column to use for alpha of the edges
scale_alpha_with_expression
                 scale expression with ggplot alpha parameter
                 size of point (cell)
point_size
point_border_col
                 color of border around points
point_border_stroke
                 stroke size of border around points
                 size of point (cell)
midpoint
cow_n_col
                 cowplot param: how many columns
                 cowplot param: relative height
cow_rel_h
                 cowplot param: relative width
cow_rel_w
cow_align
                 cowplot param: how to align
show_legend
                 show legend
show_plots
                 show plots
```

Details

Description of parameters.

Value

ggplot

Examples

```
visDimGenePlot_2D_ggplot(gobject)
```

Description

Visualize cells and gene expression according to dimension reduction coordinates

Usage

```
visDimGenePlot_3D_plotly(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = 3,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  network_color = "lightgray",
  edge_alpha = NULL,
  point_size = 1,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_legend = T,
  show_plots = F
)
```

Arguments

```
gobject
                 giotto object
expression_values
                 gene expression values to use
genes
                 genes to show
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
dim2_to_use
                 dimension to use on y-axis
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
edge_alpha
                 column to use for alpha of the edges
point_size
                 size of point (cell)
show_legend
                 show legend
                 show plots
show_plots
```

Details

Description of parameters.

visDimPlot 281

Value

ggplot

Examples

```
visDimGenePlot_3D_plotly(gobject)
```

visDimPlot

visDimPlot

Description

Visualize cells according to dimension reduction coordinates

```
visDimPlot(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  dim3_to_use = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
  edge_alpha = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  plot_method = c("ggplot", "plotly"),
  show_legend = T,
  show_plot = F,
  return_plot = TRUE,
  save_plot = F,
  save_dir = NULL,
```

282 visDimPlot

```
save_folder = NULL,
      save_name = NULL,
      save_format = NULL,
      show_saved_plot = F,
    )
Arguments
                     giotto object
    gobject
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
                     dimension to use on z-axis
    dim3_to_use
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
                     size of center points
    label_size
                     size of labels
    label_fontface font of labels
    edge_alpha
                     column to use for alpha of the edges
    point_size
                     size of point (cell)
    point_border_col
                     color of border around points
    point_border_stroke
                     stroke size of border around points
    show_legend
                     show legend
    show_plot
                     show plot
    return_plot
                     return ggplot object
                     directly save the plot [boolean]
    save_plot
```

directory to save the plot

save_dir

visDimPlot_2D_ggplot 283

Details

Description of parameters.

Value

ggplot or plotly

Examples

```
visDimPlot(gobject)
```

```
visDimPlot_2D_ggplot visDimPlot_2D_ggplot
```

Description

Visualize cells according to dimension reduction coordinates

```
visDimPlot_2D_ggplot(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  center_point_border_col = "black",
  center_point_border_stroke = 0.1,
  label_size = 4,
  label_fontface = "bold",
```

```
edge_alpha = NULL,
      point_size = 1,
      point_border_col = "black",
      point_border_stroke = 0.1,
      show_legend = T,
      show_plot = F,
      return_plot = TRUE,
      save_plot = F,
      save_dir = NULL,
      save_folder = NULL,
      save_name = NULL,
      save_format = NULL,
      show_saved_plot = F,
    )
Arguments
    gobject
                     giotto object
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
    network_name
                     name of NN network to use, if show_NN_network = TRUE
                     color for cells (see details)
    cell_color
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
    select_cells
                     select subset of cells based on cell IDs
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    other_point_size
                     size of not selected cells
    show_cluster_center
                     plot center of selected clusters
    show_center_label
                     plot label of selected clusters
    center_point_size
```

size of center points

visDimPlot_2D_plotly 285

Details

Description of parameters.

Value

ggplot

Examples

```
visDimPlot_2D_ggplot(gobject)
```

```
visDimPlot_2D_plotly visDimPlot_2D_plotly
```

Description

Visualize cells according to dimension reduction coordinates

```
visDimPlot_2D_plotly(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  color_as_factor = T,
  cell_color = NULL,
  cell_color_code = NULL,
  show_cluster_center = F,
  show_center_label = T,
```

```
center_point_size = 4,
label_size = 4,
edge_alpha = NULL,
point_size = 5
)
```

Arguments

```
gobject
                 giotto object
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
                 dimension to use on x-axis
dim1_to_use
dim2_to_use
                 dimension to use on y-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
network_name
                 name of NN network to use, if show_NN_network = TRUE
color_as_factor
                 convert color column to factor
cell_color
                 color for cells (see details)
cell_color_code
                 named vector with colors
show_cluster_center
                 plot center of selected clusters
show_center_label
                 plot label of selected clusters
center_point_size
                 size of center points
label_size
                 size of labels
edge_alpha
                 column to use for alpha of the edges
point_size
                 size of point (cell)
```

Details

Description of parameters.

Value

plotly

```
visDimPlot_2D_plotly(gobject)
```

```
visDimPlot_3D_plotly
```

Description

Visualize cells according to dimension reduction coordinates

Usage

```
visDimPlot_3D_plotly(
  gobject,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = 3,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  color_as_factor = T,
  cell_color = NULL,
  cell_color_code = NULL,
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 4,
  edge_alpha = NULL,
  point_size = 1
```

Arguments

```
giotto object
gobject
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
dim1_to_use
                 dimension to use on x-axis
                 dimension to use on y-axis
dim2_to_use
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
```

288 visForceLayoutPlot

```
name of NN network to use, if show_NN_network = TRUE
network_name
color_as_factor
                  convert color column to factor
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
show_cluster_center
                  plot center of selected clusters
show_center_label
                  plot label of selected clusters
center_point_size
                  size of center points
label_size
                  size of labels
                  column to use for alpha of the edges
edge_alpha
point_size
                  size of point (cell)
```

Details

Description of parameters.

Value

plotly

Examples

```
visDimPlot_3D_plotly(gobject)
```

visForceLayoutPlot visForceLayoutPlot

Description

Visualize cells according to forced layout algorithm coordinates

```
visForceLayoutPlot(
  gobject,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  layout_name = "layout",
  dim1_to_use = 1,
  dim2_to_use = 2,
  show_NN_network = T,
  cell_color = NULL,
  color_as_factor = TRUE,
  cell_color_code = NULL,
  edge_alpha = NULL,
  point_size = 1,
```

visForceLayoutPlot 289

```
point_border_col = "black",
point_border_stroke = 0.1,
show_legend = T,
show_plot = F,
return_plot = TRUE,
save_plot = F,
save_dir = NULL,
save_folder = NULL,
save_name = NULL,
save_format = NULL,
show_saved_plot = F,
...
)
```

```
giotto object
gobject
nn_network_to_use
                  type of NN network to use (kNN vs sNN)
network_name
                  NN network to use
                  name of layout to use
layout_name
dim1_to_use
                  dimension to use on x-axis
dim2_to_use
                  dimension to use on y-axis
show_NN_network
                  show underlying NN network
cell_color
                  color for cells (see details)
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
                  column to use for alpha of the edges
edge_alpha
                  size of point (cell)
point_size
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
                  show legend
show_legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
                  directory to save the plot
save_dir
save_folder
                  (optional) folder in directory to save the plot
                  name of plot
save_name
save_format
                  format of plot (e.g. tiff, png, pdf, ...)
show_saved_plot
                  load & display the saved plot
```

290 visGenePlot

Details

Description of parameters.

Value

ggplot

Examples

visForceLayoutPlot(gobject)

visGenePlot

visGenePlot

Description

Visualize cells and gene expression according to spatial coordinates

```
visGenePlot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  show_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  edge_alpha = NULL,
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  midpoint = 0,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  plot_method = c("ggplot", "plotly"),
  show_plots = F
```

visGenePlot 291

Arguments

gobject giotto object expression_values

gene expression values to use

genes to show genes

genes_high_color

color represents high gene expression

genes_mid_color

color represents middle gene expression

genes_low_color

color represents low gene expression

show underlying spatial network show_network

network_color color of spatial network

spatial_network_name

name of spatial network to use

show spatial grid show_grid color of spatial grid grid_color

spatial_grid_name

name of spatial grid to use

midpoint expression midpoint scale_alpha_with_expression

scale expression with ggplot alpha parameter

size of point (cell) point_size

point_border_col

color of border around points

point_border_stroke

stroke size of border around points

show legend show_legend

cow_n_col cowplot param: how many columns cow_rel_h cowplot param: relative height cow_rel_w cowplot param: relative width cow_align cowplot param: how to align three mode to adjust axis scale axis_scale x_ticks number of ticks on x axis number of ticks on y axis y_ticks number of ticks on z axis z_ticks plot_method two methods of plot show_plots

show plots

Details

Description of parameters.

Value

ggplot or plotly

Examples

```
visGenePlot(gobject)
```

```
{\tt visGenePlot\_2D\_ggplot} \ \ \textit{visGenePlot\_2D\_ggplot}
```

Description

Visualize cells and gene expression according to spatial coordinates

Usage

```
visGenePlot_2D_ggplot(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  genes_high_color = "darkred",
  genes_mid_color = "white",
  genes_low_color = "darkblue",
  show_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  edge_alpha = NULL,
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  midpoint = 0,
  scale_alpha_with_expression = FALSE,
  point_size = 1,
  point_border_col = "black",
  point_border_stroke = 0.1,
  show_legend = T,
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  show_plots = F
)
```

```
gobject giotto object
expression_values
gene expression values to use
genes genes to show
genes_high_color
color represents high gene expression
genes_mid_color
color represents middle gene expression
```

visGenePlot_3D_plotly

```
genes_low_color
```

color represents low gene expression

show_network show underlying spatial network

network_color color of spatial network

spatial_network_name

name of spatial network to use

show_grid show spatial grid grid_color color of spatial grid

spatial_grid_name

name of spatial grid to use

midpoint expression midpoint

scale_alpha_with_expression

scale expression with ggplot alpha parameter

point_size size of point (cell)

point_border_col

color of border around points

point_border_stroke

stroke size of border around points

show_legend show legend

cow_n_colcowplot param: how many columnscow_rel_hcowplot param: relative heightcow_rel_wcowplot param: relative widthcow_aligncowplot param: how to align

show_plots show plots

Details

Description of parameters.

Value

ggplot

Examples

visGenePlot_2D_ggplot(gobject)

 ${\tt visGenePlot_3D_plotly} \ \ {\it visGenePlot_3D_plotly}$

Description

Visualize cells and gene expression according to spatial coordinates

Usage

```
visGenePlot_3D_plotly(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  genes,
  show_network = F,
  network_color = NULL,
  spatial_network_name = "spatial_network",
  edge_alpha = NULL,
  show\_grid = F,
  genes_high_color = NULL,
  genes_mid_color = "white",
  genes_low_color = "blue",
  spatial_grid_name = "spatial_grid",
  point_size = 1,
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  show_plots = F
gobject
                giotto object
expression_values
```

```
gene expression values to use
genes
                  genes to show
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
show_grid
                  show spatial grid
genes_high_color
                  color represents high gene expression
genes_mid_color
                  color represents middle gene expression
genes_low_color
                  color represents low gene expression
spatial_grid_name
                  name of spatial grid to use
                  size of point (cell)
point_size
show_legend
                  show legend
axis_scale
                  three mode to adjust axis scale
x_ticks
                  number of ticks on x axis
                  number of ticks on y axis
y_ticks
```

visPlot 295

Details

Description of parameters.

Value

plotly

Examples

```
visGenePlot_3D_plotly(gobject)
```

visPlot visPlot

Description

Visualize cells according to spatial coordinates

```
visPlot(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  show_network = F,
  network_color = NULL,
  network_alpha = 1,
  other_cell_alpha = 0.1,
  spatial_network_name = "spatial_network",
  show\_grid = F,
```

296 visPlot

```
grid_color = NULL,
  grid_alpha = 1,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 0.6,
  title = "",
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  plot_method = c("ggplot", "plotly"),
  show_plot = F,
  return_plot = TRUE,
  save_plot = F,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  save_format = NULL,
  show_saved_plot = F,
)
gobject
                giotto object
```

```
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
sdimz
                  z-axis dimension name (default = 'sdimz')
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
show_network
                  show underlying spatial network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
```

visPlot_2D_ggplot 297

```
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
spatial_grid_name
                  name of spatial grid to use
coord_fix_ratio
                  fix ratio between x and y-axis
title
                  title of plot
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
save_plot
                  directly save the plot [boolean]
save_dir
                  directory to save the plot
                  (optional) folder in directory to save the plot
save_folder
                  name of plot
save_name
save_format
                  format of plot (e.g. tiff, png, pdf, ...)
show_saved_plot
                  load & display the saved plot
```

Details

Description of parameters.

Value

ggplot

Examples

visPlot(gobject)

Description

Visualize cells according to spatial coordinates

```
visPlot_2D_ggplot(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  point_border_col = "black",
  point_border_stroke = 0.1,
  cell_color = NULL,
  cell_color_code = NULL,
```

298 visPlot_2D_ggplot

```
color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  show_network = F,
  network_color = NULL,
  network_alpha = 1,
  other_cells_alpha = 0.1,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  spatial_grid_name = "spatial_grid",
  coord_fix_ratio = 0.6,
  title = "",
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  show_plot = F,
  return_plot = TRUE,
  save_plot = F,
  save_dir = NULL,
  save_folder = NULL,
  save_name = NULL,
  save_format = NULL,
  show_saved_plot = F,
)
```

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
point_size
                  size of point (cell)
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
color_as_factor
                  convert color column to factor
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
```

visPlot_2D_ggplot 299

show_other_cells

display not selected cells

other_cell_color

color of not selected cells

show_network show underlying spatial network

network_color color of spatial network

spatial_network_name

name of spatial network to use

show_grid show spatial grid

grid_color color of spatial grid

spatial_grid_name

name of spatial grid to use

coord_fix_ratio

fix ratio between x and y-axis

title title of plot

show_legend show legend

show_plot show plot

return_plot return ggplot object

save_plot directly save the plot [boolean]

save_dir directory to save the plot

save_folder (optional) folder in directory to save the plot

save_name name of plot

save_format format of plot (e.g. tiff, png, pdf, ...)

show_saved_plot

load & display the saved plot

Details

Description of parameters.

Value

ggplot

Examples

visPlot_2D_ggplot(gobject)

300 visPlot_2D_plotly

```
visPlot_2D_plotly
```

Description

Visualize cells according to spatial coordinates

Usage

```
visPlot_2D_plotly(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  point_size = 3,
  cell_color = NULL,
  cell_color_code = NULL,
  color_as_factor = T,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_network = F,
  network_color = "lightgray",
  network_alpha = 1,
  other_cell_alpha = 0.5,
  spatial_network_name = "spatial_network",
  show\_grid = F,
  grid_color = NULL,
  grid_alpha = 1,
  spatial_grid_name = "spatial_grid",
  show_legend = T,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  show_plot = F
```

```
gobject giotto object

sdimx x-axis dimension name (default = 'sdimx')

sdimy y-axis dimension name (default = 'sdimy')

point_size size of point (cell)

cell_color color for cells (see details)

cell_color_code

named vector with colors

color_as_factor

convert color column to factor
```

visPlot_3D_plotly 301

```
select_cell_groups
                  select a subset of the groups from cell_color
                  show underlying spatial network
show_network
                  color of spatial network
network_color
spatial_network_name
                  name of spatial network to use
                  show spatial grid
show_grid
grid_color
                  color of spatial grid
                  alpha of spatial grid
grid_alpha
spatial_grid_name
                  name of spatial grid to use
                  show legend
show_legend
show_plot
                  show plot
```

Details

Description of parameters.

Value

plotly

Examples

```
visPlot_2D_plotly(gobject)
```

```
visPlot_3D_plotly
```

Description

Visualize cells according to spatial coordinates

```
visPlot_3D_plotly(
  gobject,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  point_size = 3,
  cell_color = NULL,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  other_point_size = 0.5,
  show_network = F,
```

302 visPlot_3D_plotly

```
network_color = NULL,
network_alpha = 1,
other_cell_alpha = 0.5,
spatial_network_name = "spatial_network",
spatial_grid_name = "spatial_grid",
title = "",
show_legend = T,
axis_scale = c("cube", "real", "custom"),
custom_ratio = NULL,
x_ticks = NULL,
y_ticks = NULL,
z_ticks = NULL,
show_plot = F
```

```
gobject
                  giotto object
sdimx
                  x-axis dimension name (default = 'sdimx')
sdimy
                  y-axis dimension name (default = 'sdimy')
sdimz
                  z-axis dimension name (default = 'sdimz')
point_size
                  size of point (cell)
cell_color
                  color for cells (see details)
cell_color_code
                  named vector with colors
select_cell_groups
                  select a subset of the groups from cell_color
                  show underlying spatial network
show_network
network_color
                  color of spatial network
spatial_network_name
                  name of spatial network to use
spatial_grid_name
                  name of spatial grid to use
                  title of plot
title
show_legend
                  show legend
show_plot
                  show plot
point_border_col
                  color of border around points
point_border_stroke
                  stroke size of border around points
color_as_factor
                  convert color column to factor
show_grid
                  show spatial grid
grid_color
                  color of spatial grid
coord_fix_ratio
                  fix ratio between x and y-axis
```

visSpatDimGenePlot 303

Details

Description of parameters.

Value

ggplot

Examples

```
visPlot_3D_plotly(gobject)
```

visSpatDimGenePlot

visSpatDimGenePlot

Description

integration of visSpatDimGenePlot_2D(ggplot) and visSpatDimGenePlot_3D(plotly)

```
visSpatDimGenePlot(
 gobject,
 plot_method = c("ggplot", "plotly"),
 expression_values = c("normalized", "scaled", "custom"),
 plot_alignment = c("horizontal", "vertical"),
 dim_reduction_to_use = "umap",
 dim_reduction_name = "umap",
 dim1_to_use = 1,
 dim2_to_use = 2,
 dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
 genes,
 dim_point_border_col = "black",
 dim_point_border_stroke = 0.1,
  show_NN_network = F,
 nn_network_to_use = "sNN",
 network_name = "sNN.pca",
 edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
 label_size = 16,
 genes_low_color = "blue",
 genes_mid_color = "white",
 genes_high_color = "red",
 dim_point_size = 3,
 nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
 network_color = "lightgray",
  spatial_network_alpha = 0.5,
```

304 visSpatDimGenePlot

```
show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_grid_alpha = 0.5,
      spatial_point_size = 3,
      spatial_point_border_col = "black",
      spatial_point_border_stroke = 0.1,
      legend_text_size = 12,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
     x_ticks = NULL,
     y_ticks = NULL,
     z_ticks = NULL,
     midpoint = 0,
     point_size = 1,
      cow_n_col = 2,
      cow_rel_h = 1,
     cow_rel_w = 1,
     cow_align = "h",
     show_legend = T,
      show_plots = F
   )
Arguments
   gobject
                    giotto object
   expression_values
                    gene expression values to use
   plot_alignment direction to align plot
   dim_reduction_to_use
                    dimension reduction to use
   dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
   dim2_to_use
                    dimension to use on y-axis
   dim3_to_use
                    dimension to use on z-axis
   sdimx
                    x-axis dimension name (default = 'sdimx')
   sdimy
                    y-axis dimension name (default = 'sdimy')
    sdimz
                    z-axis dimension name (default = 'sdimz')
   genes
                    genes to show
   dim_point_border_col
                    color of border around points
   dim_point_border_stroke
                    stroke size of border around points
   show_NN_network
                    show underlying NN network
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
                    name of NN network to use, if show_NN_network = TRUE
   network_name
```

visSpatDimGenePlot 305

edge_alpha_dim dim reduction plot: column to use for alpha of the edges scale_alpha_with_expression

scale expression with ggplot alpha parameter

size for the label label_size

genes_low_color

color to represent low expression of gene

genes_high_color

color to represent high expression of gene

dim_point_size dim reduction plot: point size

spatial_network_name

name of spatial network to use

spatial_grid_name

name of spatial grid to use

spatial_point_size

spatial plot: point size

spatial_point_border_col

color of border around points

spatial_point_border_stroke

stroke size of border around points

legend_text_size

the size of the text in legend

three modes to adjust axis scale ratio axis_scale custom_ratio set the axis scale ratio on custom

x_ticks number of ticks on x axis y_ticks number of ticks on y axis z_ticks number of ticks on z axis

size of point (cell) midpoint point_size size of point (cell)

cow_n_col cowplot param: how many columns cow_rel_h cowplot param: relative height cowplot param: relative width cow_rel_w cowplot param: how to align cow_align

show_legend show legend show_plot show plot

Details

Description of parameters.

Value

ggplot or plotly

Examples

visSpatDimGenePlot(gobject)

visSpatDimGenePlot_2D visSpatDimGenePlot_2D

Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot mode

Usage

```
visSpatDimGenePlot_2D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("horizontal", "vertical"),
  genes,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  point_size = 1,
  dim_point_border_col = "black",
  dim_point_border_stroke = 0.1,
  show_NN_network = F,
  show_spatial_network = F,
  show_spatial_grid = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  edge_alpha_dim = NULL,
  scale_alpha_with_expression = FALSE,
  spatial_network_name = "spatial_network",
  spatial_grid_name = "spatial_grid",
  spatial_point_size = 1,
  spatial_point_border_col = "black",
  spatial_point_border_stroke = 0.1,
  midpoint = 0,
  genes_high_color = "red",
  genes_mid_color = "white";
  genes_low_color = "blue",
  cow_n_col = 2,
  cow_rel_h = 1,
  cow_rel_w = 1,
  cow_align = "h",
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  show_legend = T,
  show_plots = F
```

Arguments

gobject giotto object

expression_values

gene expression values to use

plot_alignment direction to align plot

genes genes to show

dim_reduction_to_use

dimension reduction to use

dim_reduction_name

dimension reduction name

dim1_to_use dimension to use on x-axis dim2_to_use dimension to use on y-axis

point_size size of point (cell)

dim_point_border_col

color of border around points

dim_point_border_stroke

stroke size of border around points

show_NN_network

show underlying NN network

nn_network_to_use

type of NN network to use (kNN vs sNN)

network_name name of NN network to use, if show_NN_network = TRUE

edge_alpha_dim dim reduction plot: column to use for alpha of the edges

scale_alpha_with_expression

scale expression with ggplot alpha parameter

spatial_network_name

name of spatial network to use

spatial_grid_name

name of spatial grid to use

spatial_point_size

spatial plot: point size

spatial_point_border_col

color of border around points

 ${\tt spatial_point_border_stroke}$

stroke size of border around points

midpoint size of point (cell)

cow_n_col cowplot param: how many columns

cow_rel_h cowplot param: relative height cow_rel_w cowplot param: relative width cow_align cowplot param: how to align

show_legend show legend

dim_point_size dim reduction plot: point size

show_plot show plot

Details

Description of parameters.

Value

ggplot

Examples

```
visSpatDimGenePlot_2D(gobject)
```

```
visSpatDimGenePlot_3D visSpatDimGenePlot_3D
```

Description

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

```
visSpatDimGenePlot_3D(
  gobject,
  expression_values = c("normalized", "scaled", "custom"),
  plot_alignment = c("horizontal", "vertical"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  genes,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  label_size = 16,
  genes_low_color = "blue",
  genes_mid_color = "white",
  genes_high_color = "red",
  dim_point_size = 3,
  nn_network_alpha = 0.5,
  show\_spatial\_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  legend_text_size = 12,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_{ticks} = NULL,
```

y_ticks = NULL, z_ticks = NULL

```
Arguments
   gobject
                     giotto object
    plot_alignment direction to align plot
   dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
   dim1_to_use
                     dimension to use on x-axis
   dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
   network_name
    genes_low_color
                     color represent high gene expression (see details)
    genes_high_color
                     color represent high gene expression (see details)
    nn_network_alpha
                     column to use for alpha of the edges
    show_spatial_network
                     show spatial network
    spatial_network_name
                     name of spatial network to use
```

legend_text_size

spatial_network_alpha

show_spatial_grid

spatial_grid_name

spatial_grid_color

spatial_grid_alpha

text size of legend

color of spatial grid

alpha of spatial grid

alpha of spatial network

name of spatial grid to use

show spatial grid

network_color color of spatial/nn network

show_legend show legend
show_plot show plot

Details

Description of parameters.

310 visSpatDimPlot

Value

plotly

Examples

```
visSpatDimPlot_3D(gobject)
```

visSpatDimPlot

visSpatDimPlot

Description

integration of visSpatDimPlot_2D and visSpatDimPlot_3D

```
visSpatDimPlot(
  gobject,
  plot_method = c("ggplot", "plotly"),
  plot_alignment = NULL,
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = NULL,
  label_fontface = "bold",
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  select_cell_groups = NULL,
  select_cells = NULL,
  show_other_cells = T,
  other_cell_color = "lightgrey",
  dim_point_size = 3,
  dim_point_border_col = "black",
  dim_point_border_stroke = 0.1,
  nn_network_alpha = NULL,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
```

visSpatDimPlot 311

```
show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_grid_alpha = 0.5,
      spatial_point_size = 3,
      legend_text_size = 12,
      spatial_point_border_col = "black",
      spatial_point_border_stroke = 0.1,
      show_legend = T,
      axis_scale = c("cube", "real", "custom"),
      custom_ratio = NULL,
      x_ticks = NULL,
      y_ticks = NULL,
      z_ticks = NULL,
      show_plot = F
Arguments
    gobject
                     giotto object
    plot_alignment direction to align plot
    dim_reduction_to_use
                     dimension reduction to use
    dim_reduction_name
                     dimension reduction name
    dim1_to_use
                     dimension to use on x-axis
    dim2_to_use
                     dimension to use on y-axis
    dim3_to_use
                     dimension to use on z-axis
    show_NN_network
                     show underlying NN network
    nn_network_to_use
                     type of NN network to use (kNN vs sNN)
                     name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color
                     color for cells (see details)
    color_as_factor
                     convert color column to factor
    cell_color_code
                     named vector with colors
    select_cell_groups
                     select subset of cells/clusters based on cell_color parameter
                     select subset of cells based on cell IDs
    select_cells
    show_other_cells
                     display not selected cells
    other_cell_color
                     color of not selected cells
    nn_network_alpha
                     column to use for alpha of the edges
    show\_spatial\_network
                     show spatial network
```

312 visSpatDimPlot_2D

```
spatial_network_name
                 name of spatial network to use
spatial_network_alpha
                 alpha of spatial network
show_spatial_grid
                 show spatial grid
spatial_grid_name
                 name of spatial grid to use
spatial_grid_color
                 color of spatial grid
spatial_grid_alpha
                 alpha of spatial grid
legend_text_size
                 text size of legend
show_legend
                 show legend
show_plot
                 show plot
plot_mode
                 choose the mode to draw plot: ggplot or plotly
spatial_network_color
                 color of spatial network
```

Details

Description of parameters.

Value

ggplot or plotly

Examples

```
visSpatDimPlot(gobject)
```

```
visSpatDimPlot_2D
visSpatDimPlot_2D
```

Description

Visualize cells according to spatial AND dimension reduction coordinates in ggplot2 mode

```
visSpatDimPlot_2D(
  gobject,
  plot_alignment = c("vertical", "horizontal"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2_to_use = 2,
  sdimx = NULL,
  sdimy = NULL,
```

visSpatDimPlot_2D 313

 $show_NN_network = F,$

```
nn_network_to_use = "sNN",
     network_name = "sNN.pca",
      show\_cluster\_center = F,
      show_center_label = T,
      center_point_size = 4,
      label_size = 4,
      label_fontface = "bold",
      cell_color = NULL,
      color_as_factor = T,
      cell_color_code = NULL,
      select_cell_groups = NULL,
      select_cells = NULL,
      show_other_cells = T,
     other_cell_color = "lightgrey",
      dim_plot_mode = NULL,
     dim_point_size = 1,
     dim_point_border_col = "black",
     dim_point_border_stroke = 0.1,
     nn_network_alpha = 0.05,
      show_spatial_network = F,
      spatial_network_name = "spatial_network",
      spatial_network_color = NULL,
      show_spatial_grid = F,
      spatial_grid_name = "spatial_grid",
      spatial_grid_color = NULL,
      spatial_point_size = 1,
      spatial_point_border_col = "black",
      spatial_point_border_stroke = 0.1,
      show_legend = T,
      show_plot = F,
     plot_method = "ggplot"
Arguments
   gobject
                    giotto object
    plot_alignment direction to align plot
   dim_reduction_to_use
                    dimension reduction to use
    dim_reduction_name
                    dimension reduction name
   dim1_to_use
                    dimension to use on x-axis
    dim2_to_use
                    dimension to use on y-axis
    show_NN_network
                    show underlying NN network
   nn_network_to_use
                    type of NN network to use (kNN vs sNN)
                    name of NN network to use, if show_NN_network = TRUE
    network_name
    cell_color
                    color for cells (see details)
```

314 visSpatDimPlot_2D

```
color_as_factor
                  convert color column to factor
cell_color_code
                  named vector with colors
select_cell_groups
                  select subset of cells/clusters based on cell_color parameter
                  select subset of cells based on cell IDs
select_cells
show_other_cells
                  display not selected cells
other_cell_color
                  color of not selected cells
nn_network_alpha
                  column to use for alpha of the edges
show_spatial_network
                  show spatial network
spatial_network_name
                  name of spatial network to use
spatial_network_color
                  color of spatial network
show_spatial_grid
                  show spatial grid
spatial_grid_name
                  name of spatial grid to use
spatial_grid_color
                  color of spatial grid
show_legend
                  show legend
show_plot
                  show plot
return_plot
                  return ggplot object
                  directly save the plot [boolean]
save_plot
save_dir
                  directory to save the plot
                  (optional) folder in directory to save the plot
save_folder
                  name of plot
save_name
save_format
                  format of plot (e.g. tiff, png, pdf, ...)
show_saved_plot
                  load & display the saved plot
```

Details

Description of parameters.

Value

ggplot

Examples

```
visSpatDimPlot_2D(gobject)
```

visSpatDimPlot_3D 315

visSpatDimPlot_3D

visSpatDimPlot_3D

Description

Visualize cells according to spatial AND dimension reduction coordinates in plotly mode

Usage

```
visSpatDimPlot_3D(
  gobject,
  plot_alignment = c("horizontal", "vertical"),
  dim_reduction_to_use = "umap",
  dim_reduction_name = "umap",
  dim1_to_use = 1,
  dim2\_to\_use = 2,
  dim3_to_use = NULL,
  sdimx = NULL,
  sdimy = NULL,
  sdimz = NULL,
  show_NN_network = F,
  nn_network_to_use = "sNN",
  network_name = "sNN.pca",
  show_cluster_center = F,
  show_center_label = T,
  center_point_size = 4,
  label_size = 16,
  cell_color = NULL,
  color_as_factor = T,
  cell_color_code = NULL,
  dim_point_size = 3,
  nn_network_alpha = 0.5,
  show_spatial_network = F,
  spatial_network_name = "spatial_network",
  network_color = "lightgray",
  spatial_network_alpha = 0.5,
  show_spatial_grid = F,
  spatial_grid_name = "spatial_grid",
  spatial_grid_color = NULL,
  spatial_grid_alpha = 0.5,
  spatial_point_size = 3,
  axis_scale = c("cube", "real", "custom"),
  custom_ratio = NULL,
  x_ticks = NULL,
  y_ticks = NULL,
  z_ticks = NULL,
  legend_text_size = 12
```

Arguments

gobject giotto object

316 visSpatDimPlot_3D

```
plot_alignment direction to align plot
dim_reduction_to_use
                 dimension reduction to use
dim_reduction_name
                 dimension reduction name
                 dimension to use on x-axis
dim1_to_use
                 dimension to use on y-axis
dim2_to_use
dim3_to_use
                 dimension to use on z-axis
show_NN_network
                 show underlying NN network
nn_network_to_use
                 type of NN network to use (kNN vs sNN)
                 name of NN network to use, if show_NN_network = TRUE
network_name
cell_color
                 color for cells (see details)
color_as_factor
                 convert color column to factor
cell_color_code
                 named vector with colors
nn_network_alpha
                 column to use for alpha of the edges
show_spatial_network
                 show spatial network
spatial_network_name
                 name of spatial network to use
spatial_network_alpha
                 alpha of spatial network
show_spatial_grid
                 show spatial grid
spatial_grid_name
                 name of spatial grid to use
spatial_grid_color
                 color of spatial grid
spatial_grid_alpha
                 alpha of spatial grid
legend_text_size
                 text size of legend
spatial_network_color
                 color of spatial network
                 show legend
show_legend
                 show plot
show_plot
```

Details

Description of parameters.

Value

plotly

writeHMRFresults 317

Examples

```
visSpatDimPlot_3D(gobject)
```

writeHMRFresults

writeHMRFresults

Description

write results from doHMRF to a data.table.

Usage

```
writeHMRFresults(
  gobject,
  HMRFoutput,
  k = NULL,
  betas_to_view = NULL,
  print_command = F
)
```

Arguments

gobject giotto object

HMRF output HMRF output from doHMRF

k k to write results for

betas_to_view results from different betas that you want to view

print_command see the python command

Value

data.table with HMRF results for each b and the selected k

Examples

```
writeHMRFresults(gobject)
```

Description

write out factor-like annotation data from a giotto object for the Viewer

```
write_giotto_viewer_annotation(
  annotation,
  annot_name = "test",
  output_directory = getwd()
)
```

Arguments

```
annotation annotation from the data.table from giotto object
annot_name name of the annotation
output_directory
directory where to save the files
```

Value

write a .txt and .annot file for the selection annotation

Description

write out dimensional reduction data from a giotto object for the Viewer

Usage

```
write_giotto_viewer_dim_reduction(
  dim_reduction_cell,
  dim_red = NULL,
  dim_red_name = NULL,
  dim_red_rounding = NULL,
  dim_red_rescale = c(-20, 20),
  output_directory = getwd()
)
```

Arguments

```
dim_reduction_cell

dimension reduction slot from giotto object

dim_red high level name of dimension reduction

dim_red_name specific name of dimension reduction to use

dim_red_rounding

numerical indicating how to round the coordinates

dim_red_rescale

numericals to rescale the coordinates

output_directory

directory where to save the files
```

Value

write a .txt and .annot file for the selection annotation

```
write\_giotto\_viewer\_numeric\_annotation \\ write\_giotto\_viewer\_numeric\_annotation
```

Description

write out numeric annotation data from a giotto object for the Viewer

Usage

```
write_giotto_viewer_numeric_annotation(
  annotation,
  annot_name = "test",
  output_directory = getwd()
)
```

Arguments

```
annotation annotation from the data.table from giotto object
annot_name name of the annotation
output_directory
directory where to save the files
```

Value

write a .txt and .annot file for the selection annotation

Index

*Topic giotto,	cellProximityEnrichment, 25
giotto-class, 138	cellProximityHeatmap, 26
print.giotto, 185	cellProximityNetwork, 27
show, giotto-method, 196	cellProximitySpatPlot, 29
*Topic giotto	cellProximitySpatrlot2D, 30, 30
createGiottoObject, 47	cellProximitySpatPlot3D, 30, 32
*Topic object	cellProximitySpatriot3b, 30, 32
- •	cellProximityVisPlot_2D_ggplot, 36
giotto-class, 138	cellProximityVisPlot_2D_ggplot, 38
print.giotto, 185	
show, giotto-method, 196	cellProximityVisPlot_3D_plotly, 40 changeGiottoInstructions, 41
addCellMetadata, 7, 49	_
addCellStatistics, 8, 12	cluster_walktrap, 104
addGeneMetadata, 9, 49	clusterCells, 42
	clusterSpatialCorGenes, 45
addGeneStatistics, 9, 12	combineMetadata, 45
addHMRF, 10	convertEnsemblToGeneSymbol, 46
addNetworkLayout, 11	cowplot::save_plot, 138
addStatistics, 12	create_average_detection_DT, 58
adjustGiottoMatrix, 12	create_average_DT, 58
aes_string2, 13	create_cell_type_random_cell_IDs, 59
all_plots_save_function, 15, 21, 25, 27,	create_cluster_matrix, 60
28, 30, 32, 34, 66, 69, 71, 73, 75, 78,	create_dimObject, 60
81, 84, 86, 140, 151, 153, 157, 159,	createGiottoInstructions, 46, 48
161, 163, 165, 167, 169, 171, 173,	createGiottoObject, 47, 267
175, 177, 188, 197, 198, 200, 203,	createHeatmap_DT, 49
205, 206, 213, 216, 220, 224, 227,	createMetagenes, 50
229, 231, 235, 240, 242, 245, 247,	createNearestNetwork, 51
248, 251–253, 256, 260, 262, 264,	createSpatialEnrich, 53, 108
276	createSpatialGrid, 54
allCellCellcommunicationsScores, 14	createSpatialGrid_2D, 55
annotate_spatlocs_with_spatgrid_2D, 17	createSpatialGrid_3D, 56
annotate_spatlocs_with_spatgrid_3D, 18	createSpatialNetwork, 57, 62
annotateGiotto, 16	death alouten ander (1
annotateSpatialNetwork, 17	decide_cluster_order, 61
<pre>average_gene_gene_expression_in_groups,</pre>	detectSpatialCorGenes, 62
18	detectSpatialPatterns, 63
	dimCellPlot, 64
binGetSpatialGenes, 19	dimCellPlot2D, 66, 67
1 1	dimGenePlot, 69
calculate_spatial_genes_python, 23	dimGenePlot2D, 72
calculateHVG, 20	dimGenePlot3D, 71, 73, 74
calculateMetaTable, 22	dimPlot, 75
calculateMetaTableCells, 23	dimPlot2D, 78, 78, 162, 164, 167, 169, 173,
cellProximityBarplot, 24	175

INDEX 321

dimPlot2D_single, 82	get10Xmatrix, 130
dimPlot3D, 69, 78, 81, 84, 84	<pre>get_cell_to_cell_sorted_name_conversion,</pre>
direction_test (direction_test_CPG), 87	135
direction_test_CPG, 87	<pre>get_interaction_gene_enrichment, 136</pre>
do_spatial_grid_averaging, 105	<pre>get_specific_interaction_gene_enrichment.</pre>
do_spatial_knn_smoothing, 106	136
doHclust, 44, 87	<pre>getCellProximityGeneScores, 130</pre>
doHMRF, 88	getClusterSimilarity, 132
doKmeans, 44, 90	getDendrogramSplits, 133
doLeidenCluster, 44, 91, 94, 269	getDistinctColors, 134
doLeidenSubCluster, 92	getGeneToGeneScores, 134
doLouvainCluster, 44, 94	ggplot_save_function, 137
doLouvainCluster_community, 44, 95, 95,	giotto (giotto-class), 138
99, 101, 269	giotto-class, 138
doLouvainCluster_multinet, 44, 95, 97, 99,	glouvain_ml, 97
103, 269	giouvuiii_mi, //
doLouvainSubCluster, 98	hclust, 88
doLouvainSubCluster_community, 100	Heatmap, <i>140</i>
doLouvainSubCluster_multinet, 101	heatmSpatialCorGenes, 139
doRandomWalkCluster, 44, 103	hyperGeometricEnrich, 53, 140
doSNNCluster, 44, 104	, ,
dt_to_matrix, 107	kmeans, <i>91</i>
ut_to_matrix, 107	kmeans_binarize, 141
exportGiottoViewer, 107	kNN, <i>52</i>
exprOnlyCellCellcommunicationScores,	
109	layout_with_drl, <i>ll</i>
extended_gini_fun, 110	loadHMRF, 141
extractNearestNetwork, 110	
extractivearestivetwork, 110	make_simulated_network, 143
fDataDT, 111	makeSignMatrixPAGE, 142, 148
filterCombinations, 111, 115	makeSignMatrixRank, 142, <i>186</i> , <i>188</i>
filterCPGscores, 112	mergeClusters, 144
filterDistributions, 113	mygini_fun, 145
filterGiotto, 114	
find_grid_2D, 125	nnDT_to_kNN, 145
find_grid_3D, 125	node_clusters, 146
find_grid_x, 125	normalizeGiotto, 146
find_grid_y, 125	OD Comption 2 147
	OR_function2, 147
find_grid_z, 126	PAGEEnrich, 53, 142, 148
findGiniMarkers, 115, 117, 119	PCA, 192
findGiniMarkers_one_vs_all, 116, 120	
findMarkers, 118, 123	pDataDT, 149 permutationPA, <i>211</i>
findMarkers_one_vs_all, 119	
findMastMarkers, 119, 121, 122	plot_network_layer_ggplot, 177
findMastMarkers_one_vs_all, 120, 122	plot_point_layer_ggplot, 178
findScranMarkers, 119, 123, 124	plot_point_layer_ggplot_noFILL, 180
findScranMarkers_one_vs_all, 120, 124	plot_spat_point_layer_ggplot, 181
fish_function, 126	plot_spat_point_layer_ggplot_noFILL,
fish_function2, 126	183
FSV_show, 127	plotCPGscores, 149
0. 0. 1. 100	plotGTGscores, 150
GenePattern_show, 128	plotHeatmap, 152
general_save_function, <i>16</i> , 129	plotly_axis_scale_2D, 154

322 INDEX

plotly_axis_scale_3D, 154	spatDimGenePlot, 225
plotly_grid, 155	spatDimGenePlot2D, 227
plotly_network, 156	spatDimGenePlot3D, 227, 229, 230
plotMetaDataCellsHeatmap, 156, 160	spatDimPlot, 232
plotMetaDataHeatmap, 158, 158	spatDimPlot2D, 236, 236
plotPCA, 160	spatDimPlot3D, 236, 240, 240
plotPCA_2D, 162	spatGenePlot, 243
plotPCA_3D, <i>162</i> , <i>164</i> , 164	spatGenePlot2D, 245, 245
plotTSNE, 165	spatGenePlot3D, 245, 247, 247
plotTSNE_2D, 167	Spatial_AEH, 249
plotTSNE_3D, <i>167</i> , <i>169</i> , 170	Spatial_DE, 250
plotUMAP, 171	spatNetwDistributions, 251
plotUMAP_2D, 173	spatNetwDistributionsDistance, 252
plotUMAP_3D, <i>173</i> , <i>175</i> , 175	spatNetwDistributionsKneighbors, 253
print.giotto, 185	spatPlot, 254
	spatPlot2D, 257, 273
rank_binarize, 189	spatPlot2D_single, 260
rankEnrich, 53, 143, 186	spatPlot3D, 257, 260, 263, 263, 274
rankEnrichDT, 53, 187	specificCellCellcommunicationScores
rankSpatialCorGroups, 188	265
readGiottoInstructions, 189	split_dendrogram_in_two, 266
removeCellAnnotation, 190	stitchFieldCoordinates, 49, 267
removeGeneAnnotation, 190	subClusterCells, 268
replaceGiottoInstructions, 191	subsetGiotto, 270
Rtsne, <i>193</i>	subsetGiottoLocs, 270
runPCA, 191	SubsettiottoLocs, 270
runtSNE, 192	trendSceek, 271
runUMAP, 194	trendsceek_test, 271
	## CHASCON_2000, 271
select_expression_values, 196	umap, <i>195</i>
selectPatternGenes, 195	
show, giotto-method, 196	viewHMRFresults, 272
showClusterDendrogram, 197	viewHMRFresults2D, 273
showClusterHeatmap, 198	viewHMRFresults3D, 274
showCPGscores, 199	violinPlot, 275
showGeneExpressionProximityScore, 200	visDimGenePlot, 276
showGiottoInstructions, 201	visDimGenePlot_2D_ggplot, 278
showGTGscores, 201	visDimGenePlot_3D_plotly, 279
showIntExpressionProximityScore, 202	visDimPlot, 281
showPattern, 203	visDimPlot_2D_ggplot, 283
showPattern2D, 204, 204	visDimPlot_2D_plotly, 285
showPattern3D, 205	visDimPlot_3D_plotly, 287
showPatternGenes, 206	visForceLayoutPlot, 288
showProcessingSteps, 207	visGenePlot, 290
showSpatialCorGenes, 63, 208	visGenePlot_2D_ggplot, 292
showTopGeneToGene, 209	visGenePlot_3D_plotly, 293
signPCA, 210	visPlot, 272, 295
SNN, 52	visPlot_2D_ggplot, 297
sNNclust, 105	visPlot_2D_plotly, 300
spatCellPlot, 211	visPlot_3D_plotly, 301
spatCellPlot2D, 214	visSpatDimGenePlot, 303
spatDimCellPlot, 217	visSpatDimGenePlot_2D, 306
spatDimCellPlot2D, 221	<pre>visSpatDimGenePlot_3D, 308</pre>

INDEX 323

```
visSpatDimPlot, 310
visSpatDimPlot_2D, 312
visSpatDimPlot_3D, 315

write_giotto_viewer_annotation, 317
write_giotto_viewer_dim_reduction, 318
write_giotto_viewer_numeric_annotation, 319
writeHMRFresults, 317
zlm, 121
```