

Bioinformatics: Tool for Genome Analysis

Final Portfolio

Brian Wiley
Johns Hopkins University
MS Bioinformatics

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1. Purpose

Purpose text good old purpose

2. Tools Used

This section briefly lists tools used, their purpose, and how the tool is used.

2.1. General Annotations Tools

tbl2asn

Genbank

Sequin

2.2. Prokaryotic Annotation Tools

Glimmer3

ORF Finder

FGENESB

BROM

Prodigal

2.3. Eukaryotic Annotation Tools

Augustus

Genscan

FGENESH

HMMGene

Splign

2.4. Other Core Annotation Tools

samtools

bcftools

IGV

2.5. Protein Bioinformatics Tools (Might remove this)

All the tools from Protein Bioinformatics class. These are just listed to show my experience utilizing Protein Bioinformatics resources.

2.5.1. Alignment Tools

ClustalO, MAFFT, T-Coffee, Muscle, Kalign, PROBCONS

2.5.2. Profiles and Motifs

PROSITE, ProtScale Regular Expressions

2.5.3. Visualization

Chimera, Pymol

2.5.4. Secondary Structure and SASA Prediction

PredictProtein including PHDsec, PHDacc, PHDhtm by Rost Lab.

2.5.5. PDB/Structure Validation

PROCHECK, Bio3D (R), WHATCHECK

2.5.6. Comparative Modeling and Fold Recognition

FFAS, Fugue, I-Tasser, Modeller, SWISS-MODEL, Phyre2

2.6. Databases

Nuccore/Nucleotide

Pubmed

Protein

UniProt

PDBsum

Interpro

Pfam

PATRIC

Ensembl

UCSC Table Browser

dbSNP

ClinVar

2.7. Genome Browsers

NCBI Variation Viewer

NCBI Genome Data Viewer

Ensembl

UCSC

2.8. File Types

Bed

GFF/GFF3

GTF

Wig/Wiggle

Sam/Bam

Genbank

Sequin

ASN.1

Fasta/Fastq

PDB file (.pdb)

VCF/BCF

3. Software

Python

Example Scripts and Descriptions

R

Example Scripts and Descriptions

Perl

Example Scripts and Descriptions

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4. Case Studies

4.1. Week 1 - Gene annotation and structure

Genbank, Accessing Genbank files with Bio::DB::EUtilities in perl in database class.

In the first week of class we reviewed general annotation. A major topic was reading Genbank files. Interesting I started very early on week 3 in my Biological databases courses which had a module on E-utilities and accessing information from NCBI programmatically. I discovered a package for accessing Genbank file in perl.

I wrote a perl script in my database class, [eUtils_genbank.pl](#) to access Genbank files searching nucleotide/nuccore database and return exons of search hits or protein database and return CDSs of search hits. Usage is below:

Usage:

```
eUtils_genbank.pl -q [query] -e [e-mail] [OPTIONS...]
```

Options:

--max-return=<number>

How many max database entries to return

-t, --taxid=<number>

Option filter by tax id

-r, --refseq

Option to filter by only RefSeq entries

-d, --db=<String>

Database option. Default is 'nucleotide'. Only allows for 'nucleotide', 'nuccore', or 'protein'. I figured a way to get NG and NC reference number from 'gene' database but it requires parsing the Entrez gene file for the genomic accessions. It's a pain.

4.2. Week 2 - Prokaryotic genes and gene prediction

Using tools in the Glimmer package, genome for Halanaerobium praevalens and contig for strain of Halanaerobium predict coding sequences for the contig. Compare this to FGENESB.

```
# obtain open reading frames from complete genome in hprev_genome.fasta  
# using entropy threshold of t=1.15  
long-orfs -n -t 1.15 hprev_genome.fasta Hpraevalens.longorfs
```

```
# extract sequences for training using -t for no stop codon
extract -t hprev_genome.fasta Hpraevalens.longorfs > Hpraevalens.train
# create interpolated context model from train set
build_icm -r Hpraevalens.icm < Hpraevalens.train
# run glimmer3 to predict CDS' s
# maxoverlap=50, min_gene_length=110, threshold_score >= 30
glimmer3 -o50 -g110 -t30 halan.fasta Hpraevalens.icm halan
# extract sequences from halan.predict using -t for no stop codon
extract -t halan.fasta halan.predict > halan.glimmer
```

Results:

>Halanaerobium sp. MDAL1, whole genome shotgun sequence

orf00001	171	350	+3	11.68
orf00003	343	1626	+1	8.96
orf00004	1629	4733	+3	6.58
orf00005	5786	4971	-3	8.13

From Softberry homepage run FGENESB with full genome and contig to predict CDS sequences

Selecting BACTERIAL generic as closest organism returns following results:

Output:

Number of transcription units - 2, operons - 1

N	Tu/Op	Conserved pairs(N/Pv)	S	Start	End	Score
1	1	Op 1	.	+	CDS 3	- 350 298
2	1	Op 2	.	+	CDS 343	- 1626 1063
3	1	Op 3	.	+	CDS 1629	- 4733 1901
4	2	Tu 1	.	-	CDS 4971	- 5786 654

Install and use Prodigal for bacterial gene prediction to compare with Glimmer and FGENESB

Steps:q

1) Download binary with:

```
 wget https://github.com/hyattpd/Prodigal/releases/download/v2.6.3/prodigal.Linux
```

2) Change name to prodigal and move to desired directory

3) Alternatively download source code and make with:

```
 wget https://github.com/hyattpd/Prodigal/archive/v2.6.3.tar.gz
```

```
 tar -xvzf v2.6.3.tar.gz
```

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```
cd Prodigal-2.6.3
```

```
make install INSTALLDIR=/home/coyote/tools
```

4) Run prodigal in training mode with:

```
prodigal -i hprev_genome.fasta -t hprev_genome.train
```

5) Use training to predict CDS sequences

```
prodigal -i halan.fasta -t hprev_genome.train -a halan_protein.faa
```

Results for Prodigal are same as with FGENESB:

FEATURES	Location/Qualifiers
CDS	<3..350
CDS	343..1626
CDS	1629..4733
CDS	complement(4971..5786)

Use PATRIC to find information on bacterial genomes as well as transcriptomic experiments

The screenshot shows the PATRIC 3.6.7 interface for the **Bacillus anthracis str. CDC 684** genome. The main content area displays a table of genomic features:

Feature Type	PATRIC	RefSeq
CDS	6016	5902
tRNA	96	96
pseudogene	36	0
rRNA	22	33
misc_RNA	11	0
misc feature	0	70

Below this, the "Organism Info" section provides details:

- Genome ID: 568206.3
- Genome Name: Bacillus anthracis str. CDC 684
- NCBI Taxon ID: 568206
- Genome Status: Complete
- MLST: MLST.Bacillus_cereus.1
- Culture Collection: NRRRL 3495

The screenshot shows the PATRIC interface for transcriptomic experiments. The main table lists experiments:

Title	Comparisons	Genes	PubMed	Organism	Strain	Gene Modification	Experimental Condition	Time Series	Release Date
The Chlamydia trachomatis plasmid-enco 6	10614	23319558	Chlamydia trachomatis	L2,L2R,L2R	ppg4,ppg5	mutant vs wild typ		2013-02-28T00:00	
Developmental stage specific metabolic a 6	10614	23129646	Chlamydia trachomatis	LGV434		growth media		2012-11-20T00:00	
Identification of enteroxigenic Escherichia 18	1389	23115039	Escherichia coli	E2437A		growth condition	Time series	2012-11-26T00:00	
Gene expression analysis of Yersinia Pes 12	28357	22479471	Yersinia pestis,Yersinia pseud	CO92,PB1+,Pest		temperature	Time series	2011-07-14T00:00	
<input checked="" type="checkbox"/> Gene expression analysis of Salmonella t 29	17648		Salmonella enterica	Typhimurium 1402	STM3096,fts,hilD,phoPQ,ssrA	mutant vs wild typ	Time series	2011-04-08T00:00	
In vivo expression of Salmonella enterica 1	12901	22180799	Salmonella enterica	clinical		in vivo		2011-07-12T00:00	
Expression analysis of Bacteroides fragilis 1	7812	18723678	Bacteroides fragilis	NCTC 9343	unoD1,unoD2,PSI	mutant vs wild typ		2008-09-02T00:00	

A sidebar on the right provides detailed information for the selected experiment (Salmonella enterica Typhimurium 14028S):

- Organism: Salmonella enterica
- Strain: Typhimurium 14028S
- Gene: STM3096,fts,hilD,phoPQ,ssrA
- Modification: Time Series
- Experimental Condition: mutant vs wild type
- Comparisons: 29
- Platforms: 1
- Genes: 17648
- Genome IDs: 209261,6,220341,7,588858,6,5

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ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSM623568

bcftools Welcome,BRI... National Cen... UCSC Genome... The Sheridan... PC

Hybridization protocol Aminosilane-coated slides were prehybridized in 5x SSC (1x SSC is 0.15 M NaCl plus 0.015 M sodium citrate) (Invitrogen), 0.1% sodium dodecyl sulfate, and 1% bovine serum albumin at 42 °C degree for 60 min. The slides then were washed at room temperature with distilled water, dipped in isopropanol, and allowed to dry. Equal volumes of the appropriate Cy3- and Cy5-labeled probes were combined, dried and then resuspended in a solution of 40% formamide, 5x SSC, and 0.1% sodium dodecyl sulfate. Resuspended probe was applied to a slide at 42 °C. After hybridization, 2%誓 polyvinyl pyrrolidone was added to the microarray slide and allowed to hybridize overnight at 42 °C degree. Hybridized slides were washed sequentially in solutions of 1x SSC-0.2% SDS, 0.1x SSC-0.2% SDS, and 0.1x SSC at room temperature, then dried in air.

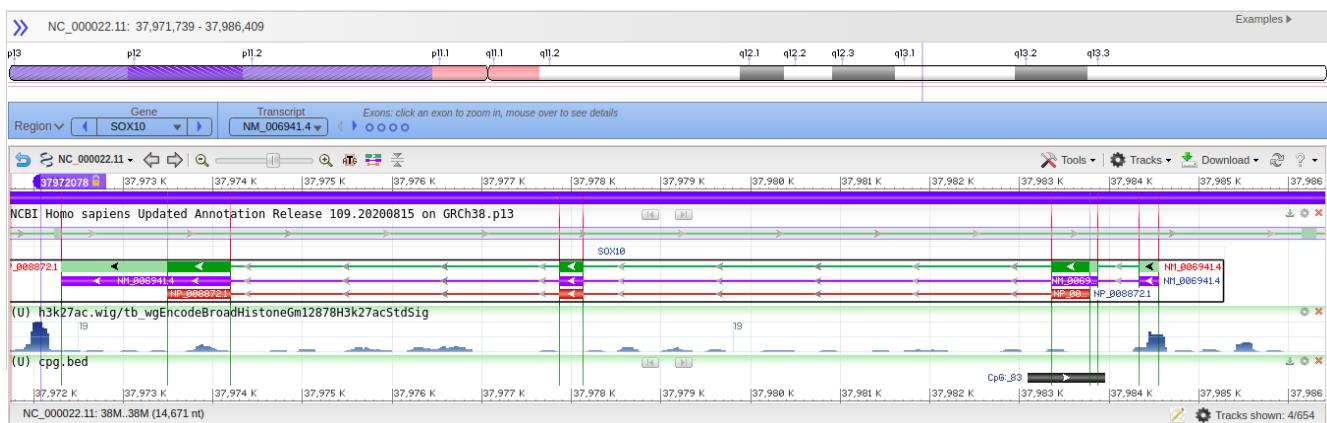
Scan protocol Hybridized slides were scanned with an Axon GenePix 4000 scanner

Data processing Individual TIFF images from each channel were analyzed with TIGR Spotfinder (available at (<http://pfgrc.jcvi.org/index.php/bioinformatics.html>)). Microarray data were normalized by LOWESS normalization and with in-slide replicate analysis using TM4 software MIDAS (available at (<http://pfgrc.jcvi.org/index.php/bioinformatics.html>)).

Submission date	Nov 15, 2010
Last update date	Apr 08, 2011
Contact name	John Braisted
E-mail(s)	jbraisted@jcvi.org
Organization name	J Craig Venter Institute

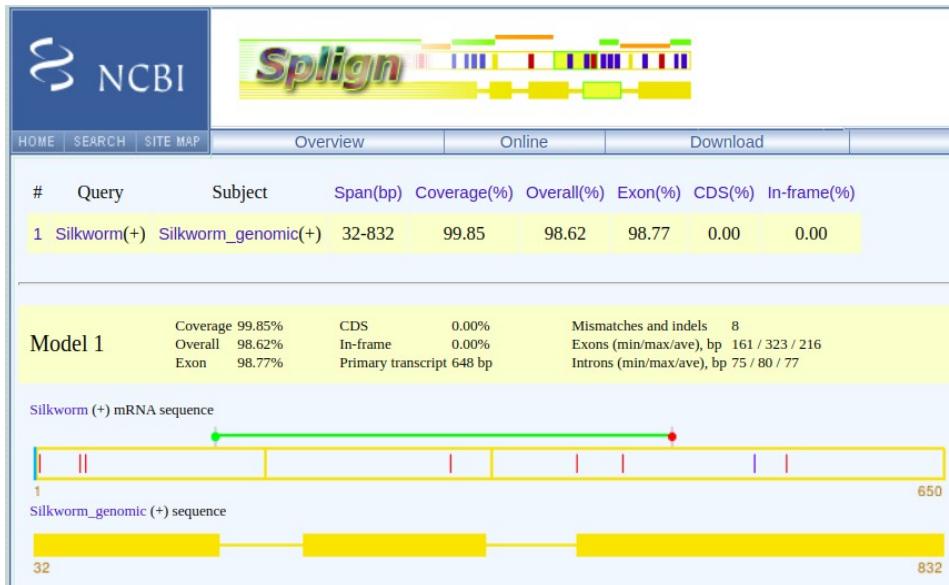
4.3. Week 3 - Eukaryotic genes and gene prediction

Add tracks to Variation Viewer: Added .bed file for CpG island and H3K27ac narrowPeaks wiggle file to NCBI Variation Viewer.



Align mRNA to genomic DNA in Splign. Then annotated with tbl2asn and Sequin to generate Genbank file. Steps below:

Step 1: Splign with cDNA input from silkworm mRNA fasta file and Genomic input from silkworm genomic input file for the gene FK506-binding protein (LOC733041)

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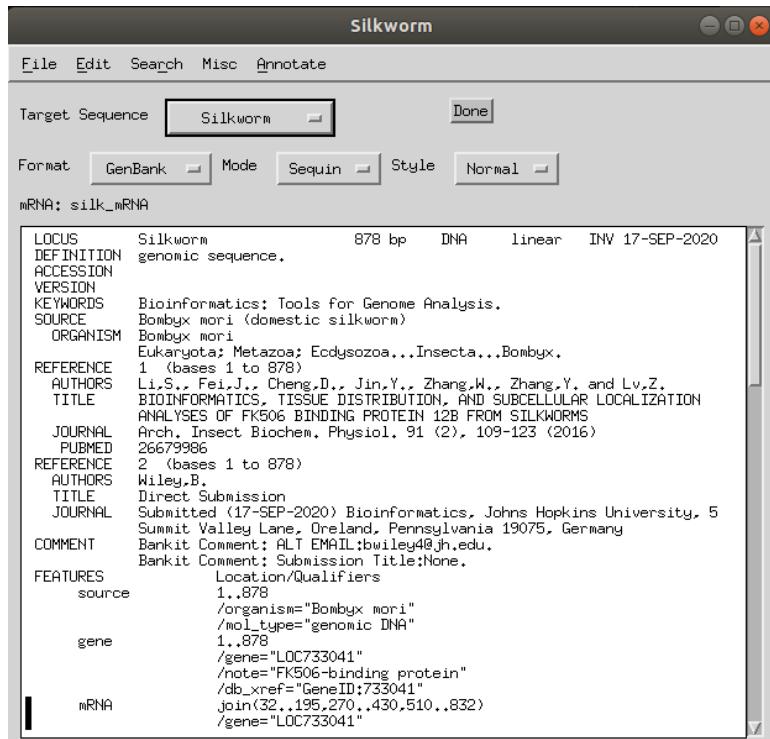
Step 2: install tbl2asn from ftp [here](#). Using the guide by NCBI for tbl2asn to create [Feature table](#).

```
1 >Feature Silkworm_genomic_sequence
2 1 878 REFERENCE
3     PubMed 26679986
4 1 878 gene
5     gene LOC733041
6     note FK506-binding protein
7     db_xref GeneID:733041
8 32 195 mRNA
9 270 430
10 510 832
11         product silk_mRNA
12 159 195 CDS
13 270 430
14 510 637
15         gene LOC733041
16         codon_start 1
17         product FK506-binding protein
18         protein_id NP_001040498.1
19 32 195 exon
20 270 430 exon
21 510 832 exon
22
```

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Step 3: Run tbl2asn with three required input files (silkworm genomic DNA, feature table, and submit file from NCBI) and update in Sequin before exporting genbank file (silkworm.gb)

```
tbl2asn -p silkworm_tbl2asn
```



Downloaded and install Augusts to run gene prediction analysis on genomic DNA from chicken from command line.

Steps:

1) Clone package from github with:

```
git clone https://github.com/Gaius-Augustus/Augustus.git
```

2) Install samtools, bcftools, and htslib to build Augustus with bam2wig

3) Run gene prediction with:

```
augustus --species=chicken chicken_gene.txt > chicken.out.gff
```

3) Obtain the proteins, coding exons, and coding sequence (mRNA) using getAnnoFasta.pl script:

```
getAnnoFasta.pl chicken.out.gff --seqfile=chicken_gene.txt
```

Exon sequences:

```

1 >g1.t1.cds1
2 atgaacttgcagaactgtcgacatccccaggcatccagaagctcaacagaggagtgcag
3 >g1.t1.cds2
4 gtcccttgcataatgacaccaggatggcacaagtggcttcaaggatcgaaag
5 >g1.t1.cds3
6 ctgtcagagcaagactgtcgccaggctgccccactgtggacaacatgaccgactgcaagaaggactacggagccctcatcaccagtctgaagagccgtcacggcatgacg
7 >g1.t1.cds4
8 aactgcccggcggacccgacaatggatctacctggaaacttcttgcggactgggaaactacccaggcgctgtaccggcggatctcgccacagctgcaaactg
9 >g2.t1.cds1
10 atgagctccactgtccacccctgtggcactgtggctgtggccggctgtgcccacgctgtgtttacagctctcagtgccgtatggagagcatccggatgtgaatgacatccaggag
11 >g2.t1.cds2
12 gtttcctgcgtcaagatggatctttcagacaataag
13 >g2.t1.cds3
14 acaataaaaaactgagctttatgaaagctccacaattttggagagccagactgccacaagaacctgtcagggctcttcatacatgcgtcagtcctgaatgccagcacccctcaag
15 >g2.t1.cds4
16 gcaccatgtccacccggcggcggcaactacttcaatggagaatgtccatgcggatgtttttcccaactagtagttaaaaataagtga
17

```

4.4. Week 4 - Ensembl and Biomart

Use BioMart interface on Ensembl website since “biomaRt” R package was working on updates.

This analysis was performed in Week 5 as when querying UCSC Table Browser when for the “Genes and Gene Predictions” group. Most tracks and tables, particularly GENCODE tracks, have a name column for “Name of gene” but list transcript IDs. The question asked how many Ensembl genes are listed in a particular region and the return from UCSC listed all the transcripts and so, after discussing with professors, to obtain the Ensembl genes we input the transcript IDs to BioMart interface and then we can export to R to count unique gene IDs in the gene ID column.

field	example	SQL type	info	description
name	ENST00000619216.1	varchar(255)	values	Name of gene
chrom	chr1	varchar(255)	values	Reference sequence chromosome or scaffold
strand	-	char(1)	values	+ or - for strand
txStart	17368	int(10) unsigned	range	Transcription start position (or end position for minus strand item)
txEnd	17436	int(10) unsigned	range	Transcription end position (or start position for minus strand item)
cdsStart	17368	int(10) unsigned	range	Coding region start (or end position if for minus strand item)
cdsEnd	17368	int(10) unsigned	range	Coding region end (or start position if for minus strand item)
exonCount	1	int(10) unsigned	range	Number of exons
exonStarts	17368,	longblob		Exon start positions (or end positions for minus strand item)
exonEnds	17436,	longblob		Exon end positions (or start positions for minus strand item)
proteinID		varchar(40)	values	UniProt display ID, UniProt accession, or RefSeq protein ID
alignID	uc031tla.1	varchar(255)	values	Unique identifier (GENCODE transcript ID for GENCODE Basic)

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The screenshot shows a search interface with the following components:

- Top Bar:** Buttons for New, Count, Results, URL, XML, Perl, and Help.
- Left Sidebar (Dataset):**
 - Human genes (GRCh38.p13)
 - Filters:** Transcript stable ID(s) [e.g. ENST00000000233] (with a note: [ID-list specified]).
 - Attributes:** Gene stable ID, Gene stable ID version, Transcript stable ID, Transcript stable ID version.
- Search Form:**
 - REGION:** A dropdown menu.
 - GENE:**
 - Limit to genes (external references)... Only Excluded
 - Input external references ID list [Max 500 advised] Transcript stable ID(s) [e.g. ENST00000000233]
- Buttons at the bottom:** Browse... transcripts.

Export all results to File TSV

Email notification to

View 10 rows as HTML Unique results only

Gene stable ID	Gene stable ID version	Transcript stable ID	Transcript stable ID version
ENSG00000076344	ENSG00000076344.16	ENST00000168869	ENST00000168869.12
ENSG00000086504	ENSG00000086504.17	ENST00000199706	ENST00000199706.13
ENSG00000086506	ENSG00000086506.3	ENST00000199708	ENST00000199708.3
ENSG00000185615	ENSG00000185615.16	ENST00000219406	ENST00000219406.11
ENSG00000242173	ENSG00000242173.10	ENST00000219409	ENST00000219409.8
ENSG00000103152	ENSG00000103152.12	ENST00000219431	ENST00000219431.4
ENSG00000103202	ENSG00000103202.13	ENST00000219479	ENST00000219479.7
ENSG00000242612	ENSG00000242612.7	ENST00000219481	ENST00000219481.10
ENSG00000129925	ENSG00000129925.11	ENST00000250930	ENST00000250930.7
ENSG00000188536	ENSG00000188536.13	ENST00000251595	ENST00000251595.11

```

31 # write transcript ids to table without version id
32 write.table(gsub("\\..*", "", bed$V4), quote = F, file="transcripts", col.names=F, row.names=F)
33
34 # read result from Ensembl BioMart
35 biomart <- read.table("/home/coyote/Downloads/mart_export(3).txt", sep="\t", header=T)
36 length(unique(biomart$Gene.stable.ID))

```

Use 'biomaRt' and 'GenomicRanges' bioconductor packages to obtain genes overlapping region

Part 3 - biomaRt

- What Ensembl genes are in a 100,000 base pair region of chromosome 20 from 5.0 to 5.3 Mb? What chromosome band are they on, what strand, and what type of genes are they?

Hint: Attributes are: hgnc_symbol, band, strand, gene_biotype

- Are there OMIM genes in this same region? Use "hgnc_symbol","band","strand","gene_biotype","mim_morbid_accession","mim_morbid_description" for the Attributes.

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I used ‘biomaRt’ package in R to send request to database to answer this question. The R script, titled [part3_2.R](#) is available on github. We discussed this problem on Slack and Discussion board as a class and there was some good conversation. We did come to agreement on interpreting the question to only include ranges that had with less than 100 kb which I performed with `BiocGenerics::width()` function and as long as any base of the region overlapped the 300 kb range we would include it. For this I also used the `GenomicRanges` Bioconductor package in R and use of the `findOverlaps()` function.

Use ‘biomaRt’ and ‘GenomicRanges’ Bioconductor packages to obtain mouse genes with RefSeq peptides and write GRanges to BED file using ‘rtracklayer’ package.

Use the web-based BioMart in Ensembl to create a dataset and save it as a TSV, CSV, or XLS file. Use the following parameters to make the dataset:

Dataset:

Ensembl Genes 93 (or the latest version)

Mouse genes (GRCm38.p6) (or the latest version)

Filters:

Chromosome 11

Band E2 only

Transcript count >=7

Limit to genes with RefSeq protein (peptide) IDs only

Attributes:

Default attributes

Add “RefSeq Protein (peptide) ID”

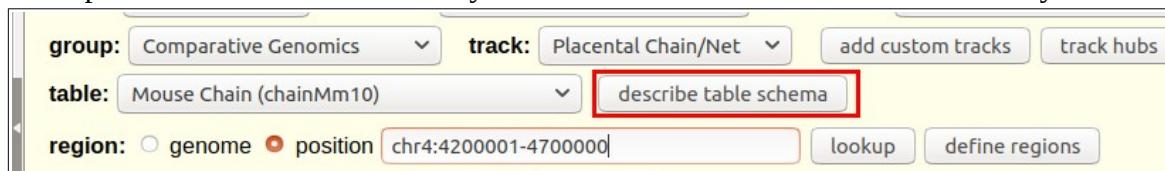
I performed the search with ‘biomaRt’ in R, put information into GRanges object, sorted by start position and wrote to BED6 file using ‘rtracklayer’. Script is named [week4_problem4_updated.R](#) and is available on github along with the bed file [week4_problem4.bed](#) with RefSeq accession ID for the peptide entry.

4.5. Week 5 - Genome browsers and analysis platforms

Create and document a workflow for students to follow for comparative genomics analysis using **Galaxy**, **UCSC Genome Browser**, and **IGV**. Posted to discussion forums for students to follow.

I decided to apply all that I learned in reading and in my own experience with all three tools, UCSC, Galaxy, & IGV that includes some extra nice stuff.

An interesting function in the UCSC Table browser is the “describe table” schema. For part 4 in peer collab clicking this for the Mouse Chain table shows the fields of the data in the table and gives a description. The name of the Primary Table: chainMm10 is what shows in Galaxy.



Format description: Summary info about a chain of alignments			
field	example	SQL type	description
bin	585 ???	smallint(5) unsigned	Indexing field to speed chromosome range queries.
score	170460	double	score of chain
tName	chr1	varchar(255)	Target sequence name
tSize	248956422	int(10) unsigned	Target sequence size
tStart	11677	int(10) unsigned	Alignment start position in target
tEnd	37616	int(10) unsigned	Alignment end position in target
qName	chr6	varchar(255)	Query sequence name
qSize	149736546	int(10) unsigned	Query sequence size
qStrand	-	char(1)	Query strand
qStart	28206106	int(10) unsigned	Alignment start position in query
qEnd	28238509	int(10) unsigned	Alignment end position in query
id	2478	int(10) unsigned	chain id

You may be asking yourself what is this “bin” field? It doesn’t seem important! Oh but wait...it is EXTREMELY important. The developers of the UCSC Table Browser, including a name you will see on about every single UCSC paper, James Kent, came up with a clever, “mystic” as they describe it, way to index the HUGE amount of data that can be queried and retrieved from extremely large tables and created a binning algorithm for faster data retrieval. You don’t need to worry about it from the user interface but if you are accessing information programatically, such as overnight batch retrieval, according to this “famous” [Biostars post](#), Heng Li (the creator along with R. Durbin of the bwa aligner program!) strongly urges learning how to implement "binning" coordinate intervals for 1) faster retrieval and 2) to not bog down the server. If you are interested in the algorithm you can find it [here](#) based on the advice of R. Durbin as well as Python implementation [here](#).

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The database used also shows up if you click the history title in Galaxy. Notice the wrong format of “tabular”.

A screenshot of a UCSC genome browser history item. The title is "3: UCSC Main on Human". Below it, a red box highlights "chainMm10 (chr4:4,200,001-4,700,000)". The text "144 lines, 1 comments" is followed by "format: tabular, database hg38". A red box highlights "hg38". Below this, a table is shown with columns 7, 8, 9, 10, and 11. The table data is:

qName	qSize	qStrand	qStart	qEnd
chr5	151834684	-	113526919	11641
chr5	151834684	-	113537000	11353
chr1	195471971	+	66032011	66032
chr1	195471971	-	129438536	12943

If you are trying to perform a function or use a tool on a data but that number is “unavailable” this means that you may have used an incorrect format. For instance for the Mouse Chain, I incorrectly did not set to “Bed Format” and so you would see:

Subtract

The Galaxy Subtract tool interface. The "Second dataset" dropdown contains "1: UCSC Main on Human: wgEncodeGencodeBasicV24 (chr4:4,200,001-4,700,...)". The "from" dropdown contains "3: (unavailable) UCSC Main on Human: chainMm10 (chr4:4,200,001-4,700,000)". The "First dataset" dropdown is empty.

Correcting this shows:

The Galaxy Subtract tool interface after correction. The "Second dataset" dropdown contains "1: UCSC Main on Human: wgEncodeGencodeBasicV24 (chr4:4,200,001-4,700,...)". The "from" dropdown contains "4: UCSC Main on Human: chainMm10 (chr4:4,200,001-4,700,000)".

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You can see the Galaxy tool parameters by clicking the circled info(i) button if you forgot which parameters you chose to run the tool.

Tool Parameters	
Input Parameter	Value
Subtract	• 1: UCSC Main on Human: wgEncodeGencodeBasicV24 (chr4:4,200,001-4,700,000)
from	• 4: UCSC Main on Human: chainMm10 (chr4:4,200,001-4,700,000)
Return	Intervals with no overlap
where minimal overlap is	1

You can actually visualize results directly from Galaxy in IGV and UCSC.

The screenshot shows a Galaxy tool interface for a subtraction operation. The tool name is "6: Subtract on data 4 and data 1". It indicates 186 regions and uses a bed format with hg38 database. Below the main information, there are three buttons: "display in IGB View", "display with IGV local Human hg38", and "display at UCSC main". The third option, "display at UCSC main", is highlighted with a red box. At the bottom, a table shows the columns: 1.Chrom, 2.Start, 3.End, 4.Name. An example row is shown: chr4 4183015 4188802 chr5.

I think this linking from Galaxy to IGV only works if you have IGV installed in Windows as the link asks you to open up an application and IGV is only installed as an “application” in windows. On my Linux machine I have to start IGV with their shell script that comes with the download. For me I can only save the XML file and source it into IGV.

For the “Non-overlapping pieces of intervals” return, if there is an overlapping interval then the pieces of the interval that remains after subtracting will still be returned. For the “Intervals with no overlap” return, if there is an overlapping interval then no interval will be returned after subtracting, i.e. you will have less regions returned from “Intervals with no overlap” than “Non-overlapping pieces of intervals”.

No overlap

vs. Overlapping pieces

The screenshot shows a Galaxy tool interface for a subtraction operation. The tool name is "5: Subtract on data 4 and data 1". It indicates 128 regions and uses a bed format with hg38 database. The interface is identical to the one above it, but the result count is lower due to the "Intervals with no overlap" setting.

The screenshot shows a Galaxy tool interface for a subtraction operation. The tool name is "6: Subtract on data 4 and data 1". It indicates 186 regions and uses a bed format with hg38 database. The interface is identical to the one above it, but the result count is higher due to the "Non-overlapping pieces of intervals" setting.

If you directly click the link from Galaxy to “display at UCSC main” you can see the User Track at the top by default and clicking “manage custom tracks” shows information about the track.

genome: Human assembly: Dec. 2013 (GRCh38/hg38) [hg38]

Name	Description	Type	Doc Items	Pos	delete	update
User Track	User Supplied Track	bed		186 chr4:		

view in: Genome Browser go add custom tracks

Clicking the name “User Track” link you would think you can change the name but no it’s greyed out. However it does show how to label your track like we did a few weeks back.

Please note a much more efficient way to load data is to use [Track Hubs](#), which are loaded from

Configuration:

```
track name='User Track' description='User Supplied Track'
```

Data:

```
Replace data at URL: https://usegalaxy.org/root/display_as?id=59988960&display_app=ucsc&authz_method=display_at
```

Submit

Adding a track is not that easy I found as uploading and writing in the configuration doesn’t work.

[Here are the steps for adding custom tracks.](#)

- 1) Format file type as bed, wig, gff3, etc.
- 2) Add browser line(s), e.g.
browser position chr4:4200001-4700000
- 3) Add track name and description, e.g.

```
track name='Interval No Overlap (I AM ON LEFT)' description='Interval No Overlap (I AM IN REGION)'
```

Note it is the Description that shows in the region of the browser and the Name is the track name to left.

Warning be aware of your quotes. It works like code.

- 4) Don’t worry about step 4. Go back to “add custom tracks” upload and Submit

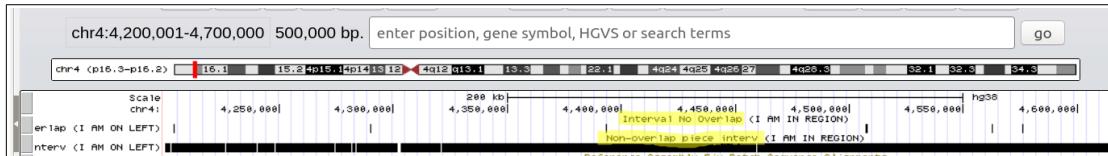
Paste URLs or data: Or upload: Intervals_with_no_overlap.bed

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Manage Custom Tracks

genome: Human assembly: Dec. 2013 (GRCh38/hg38) [hg38]

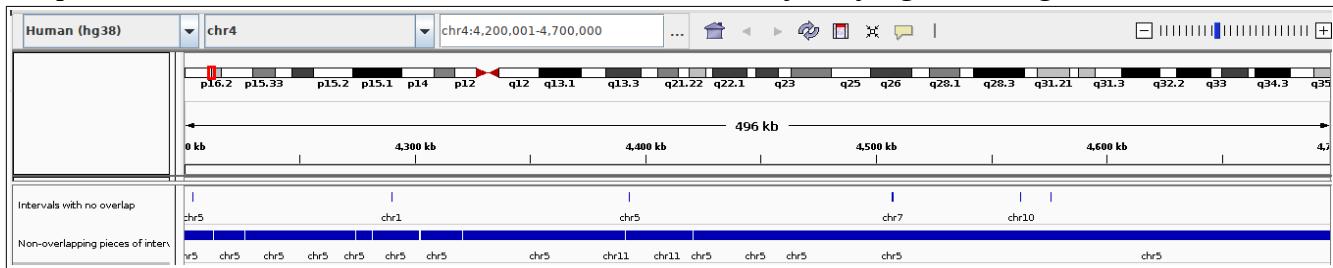
Name	Description	Type	Doc	Items	Pos	delete	view in
Non-overlap piece interv (I AM ON LEFT)	Non-overlap piece interv (I AM IN REGION)	bed		186	chr4:	<input type="checkbox"/>	go
Interval No Overlap (I AM ON LEFT)	Interval No Overlap (I AM IN REGION)	bed		128	chr4:	<input type="checkbox"/>	



As you can see, allowing overlapping pieces (bottom track) instead of getting rid of any pieces that overlap even just by 1 base (top track) removes a lot of region in width but not necessarily the # of regions from the database that conserved with this region in Mouse.

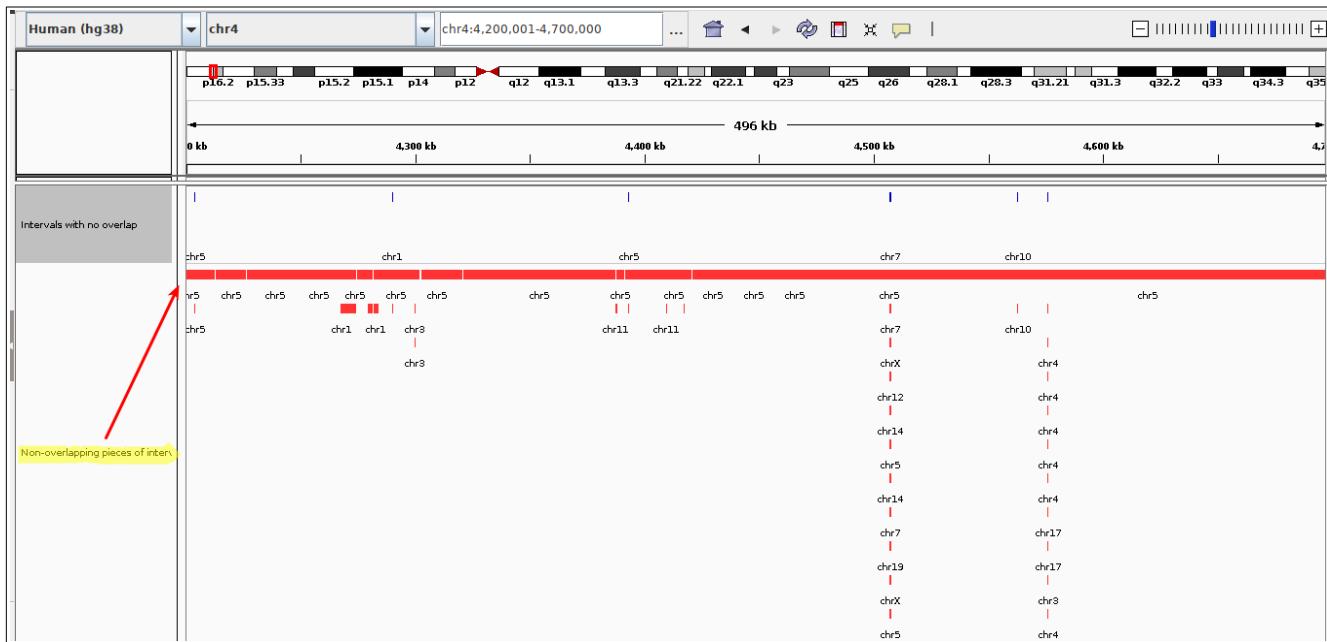


It's much easier to upload to IGV. Below you will see in IGV with “Collapsed View” as well as “Expanded View”. It is also much easier to rename the track just by right-clicking.



It's these really long pieces in red that get removed when you select the return type “Intervals with no overlap”.

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Also another interesting fact about IGV that Isabel elluded to with respect to Broad Institute developing the program for research related to The Cancer Genome Atlas (TCGA). One of the cool labs that is part of the Bioinformatics and Integrative Genomics (BIG) at Harvard University where [Heng Li](#) is associated with, the [Stegmaier Lab](#) works with Broad and one of her lab members works for the [Cancer Data Science](#) team at the Broad. I hope to write about my interest in this Data Science team in my BIG application.

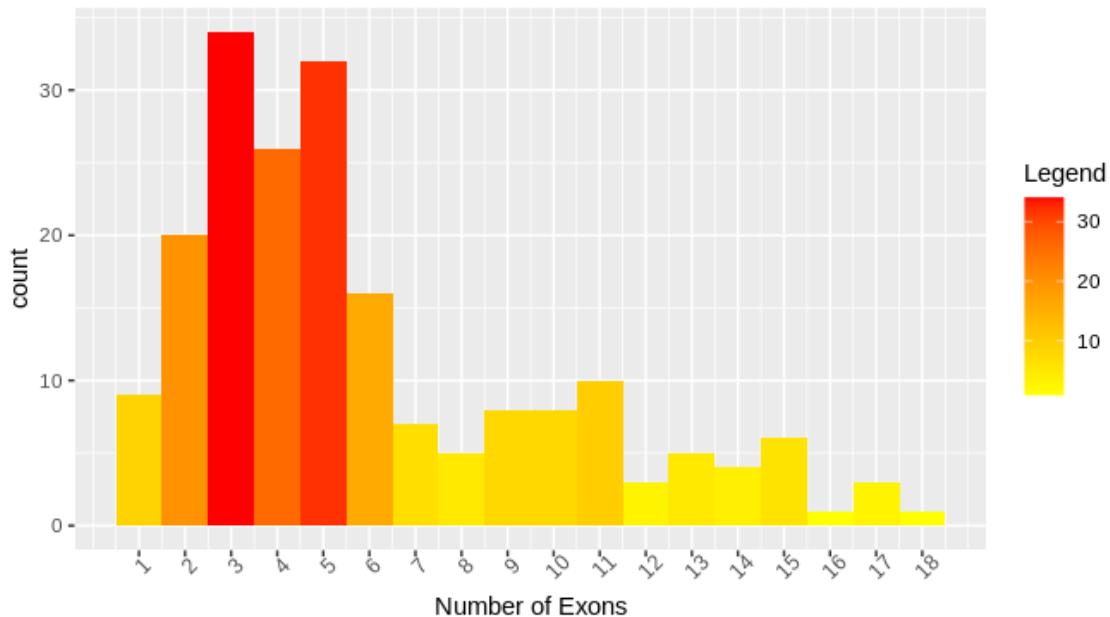
Resources:

1. Kent WJ, Sugnet CW, Furey TS, Roskin KM, Pringle TH, Zahler AM, Haussler D. The human genome browser at UCSC. *Genome Res.* 2002 Jun;12(6):996-1006. doi: 10.1101/gr.229102. PMID: 12045153; PMCID: PMC186604.

Reviewed and posted issue with the "Histogram of a numeric column" tool in Galaxy.

The breaks functionality that backends with the R hist() function incorrectly calculates the start break of range for bins. If we set the total range to be the number of breaks, i.e a bin width of one, then bins 1 and 2 were being combined. I open a github ticket under [galaxyproject/usegalaxy-playbook #309](#). As the ggplot() tool in Galaxy is not very customizable, I exported to R and plotted with ggplot2. The script is also available on github at file [galaxy_ggplot.R](#).

Histogram Gene (Transcript) count with X number of Exons



4.6. Week 6 - Single nucleotide and copy number variations

Compare information from Firehose IGV server dataset to the [Broad GDAC Firehose](#) website to learn more annotated information.

Noticing that there were different types of prefixes for Firehose data from servers datasets in IGV, I looked up the meaning of TP and TM on the Firehose website finding they stood for Tumor Primary and Tumor Metastatic respectively. I also looked up the samples for Copy Number datasets from Firehose from 2016 and found for the samples for SNP6 CopyNum, e.g. genome_wide_snp_6-segmented_scna_minus_germline_cnv_hg19 samples, where the SCNA stands for somatic copy-number alterations, was ran with the Affymetrix Genome-Wide Human SNP Array 6.0. Looking up information on this chip from Thermo Fisher website I found that of the 1.8 million genetic markers on this chip, 946,000 of the probes detect for CNV.

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Table 1. This table provides a breakdown of sample counts on a per sample type and, if applicable, per subtype basis. Each count is a link to a table containing a list of the samples that comprise that count and details pertaining to each individual sample (e.g. platform, sequencing center, etc.). Please note, there are usually multiple protocols per data type, so there are typically many more rows than the count implies.

Sample Type	BCR	Clinical	CN	LowP	Methylation	mRNA	mRNaseq	miR	miRSeq	RPPA	MAF	rawMAF
TP	1098	1097	1089	19	1097	526	1093	0	1078	887	977	0
TM	Z	Z	Z	0	Z	3	Z	0	Z	5	5	0
NB	1009	1008	979	19	0	0	0	0	0	0	0	0
NT	162	162	134	0	124	61	112	0	104	45	0	0
FFPE	0	0	Z	0	Z	0	0	0	12	0	0	0
Totals	1098	1097	1089	19	1097	526	1093	0	1078	887	977	0

The sample type short letter codes in the table above are defined in the following list.

- TP: Primary Solid Tumor
- TR: Recurrent Solid Tumor
- TB: Primary Blood Derived Cancer - Peripheral Blood
- TAP: Additional - New Primary
- TM: Metastatic
- TAM: Additional Metastatic
- NB: Blood Derived Normal

[UP](#) [<](#) [>](#) [EXPAND ALL](#) [COLLAPSE ALL](#) [SET AUTO WIDTH](#) [PRINT](#)

Table 1. This table provides a breakdown of sample counts on a per sample type and, if applicable, per subtype basis. Each count is a link to a table containing a list of the samples that comprise that count and details pertaining to each individual sample (e.g. platform, sequencing center, etc.). Please note, there are usually multiple protocols per data type, so there are typically many more rows than the count implies.

Sample Type	BCR	Clinical	CN	LowP	Methylation	mRNA	mRNaseq	miR	miRSeq	RPPA	MAF	rawMAF
TP	1098	1097	1089	19	1097	526	1093	0	1078	887	977	0
TM	Z	Z	Z	0	Z	3	Z	0	Z	5	5	0
NB	1009	1008	979	19	0	0	0	0	0	0	0	0
NT	162	162	134	0	124	61	112	0	104	45	0	0
FFPE	0	0	Z	0	Z	0	0	0	12	0	0	0

X [A12A-01](#) [Array 6.0](#) [Harvard](#)

TCGA-E2-A15E-06A-11D-A12A-01	Affymetrix Genome-Wide Human SNP Array 6.0	Broad Institute of MIT and Harvard	3	segmented_scna_minus_germline_cnv_hg18
TCGA-E2-A15E-06A-11D-A12A-01	Affymetrix Genome-Wide Human SNP Array 6.0	Broad Institute of MIT and Harvard	3	segmented_scna_minus_germline_cnv_hg19
TCGA-E2-A15K-06A-11D-A12N-01	Affymetrix Genome-Wide Human SNP Array 6.0	Broad Institute of MIT and Harvard	3	segmented_scna_hg18
TCGA-E2-A15K-06A-11D-A12N-01	Affymetrix Genome-Wide Human SNP Array 6.0	Broad Institute of MIT and Harvard	3	segmented_scna_hg19
TCGA-E2-A15K-06A-11D-A12N-01	Affymetrix Genome-Wide Human SNP Array 6.0	Broad Institute of MIT and Harvard	3	segmented_scna_minus_germline_cnv_hg18

Applied Biosystems™

Genome-Wide Human SNP Array 6.0



Catalog number:

Related applications: [Microarray](#)

	Catalog number
★	901153

Pure Power and Performance

The new Affymetrix Genome-Wide Human SNP Array 6.0 features 1.8 million genetic markers, including more than 906,600 single nucleotide polymorphisms (SNPs) and more than 946,000 probes for the detection of copy number variation. The SNP Array 6.0 is the only platform with analysis tools to truly bridge copy number and association, including a new, high-resolution reference map and a copy number polymorphism (CNP) calling algorithm developed by the Broad Institute. The SNP

This also shows that the software used by Thermo Fisher included with the chip is developed by Broad.

Use IGV to visualize whole-exome sequencing GBR (British in England and Scotland) sample data with the data from NCBI dbSNP 147 database.

Using IGV for hg19, load dbSNP 1.4.7 or newer (i.e. Available Datasets > Annotations > Variation and Repeats > dbSNP 1.4.7) and an exome sequencing track from the 1000 Genomes project (1000 Genomes > Alignments > GBR > exome > HG00096 exome). Go to the EPHX1 gene and zoom in on the exon #4.

- a. (0.25 pts) How many SNPs overlap this exon and what are the SNP IDs?
- b. (0.25 pts) At which SNP(s) in part a does this individual appear to be heterozygous? What is the sequence count for each nucleotide at this(these) position(s)

The sample HG00096 is a Bam (Binary alignment map) file with the results of runs from whole-exome sequencing (WES) on human B-lymphocytes from a donor from the United Kingdom as part of the 1000 Genomes project. I found that there were too many SNPs to count, even on 1 exon of EPHX1, and so I used the UCSC Table Browser, track All SNPs(147) primary table.snp147, with the coordinates of exon 4 of EPHX1 gene, obtained from NCBI Genome Data Viewer and confirmed with the GenBank record (see [7J](#) section under “Be a Teacher”) to obtain an output of all the SNPs listed on this track in IGV. Exporting from the table browser was then able to read into R to print out nicely in the script [galaxy_ggplot.R](#).

Table Browser

Use this program to retrieve the data associated with a track in text format, to calculate intersections between tracks, and this application see [Using the Table Browser](#) for a description of the controls in this form, and the [User's Guide](#) for gene may want to use [Galaxy](#) or our [public MySQL server](#). To examine the biological function of your set through annotation, the list of contributors and usage restrictions associated with these data. All tables can be downloaded in their entirety f

clade: Mammal **genome:** Human **assembly:** Feb. 2009 (GRCh37/hg19) **group:** Variation **track:** All SNPs(147) **table:**.snp147 **describe table schema** **add custom tracks** **tr**

Note: Most dbSNP tables are huge. Trying to download them through the Table Browser usually leads to a timeout. Please see our [Data Access FAQ](#) on how to download dbSNP data.

region: genome ENCODE Pilot regions position chr1:226026355-226026582 **lookup**

```
45 ## read in export from All SNPs(147) - Simple Nucleotide Polymorphisms (dbSNP 147)
46 test <- read.table("/home/coyote/Downloads/part3_out", header=T,
47                     sep="\t", comment.char = "")
48 paste(test$name, collapse = ", ")
```

As a class through discussion on Slack we discovered there were more than just one type of variation from the dbSNP database and that in addition to SNPs, there were other classes of variation list in red box below.

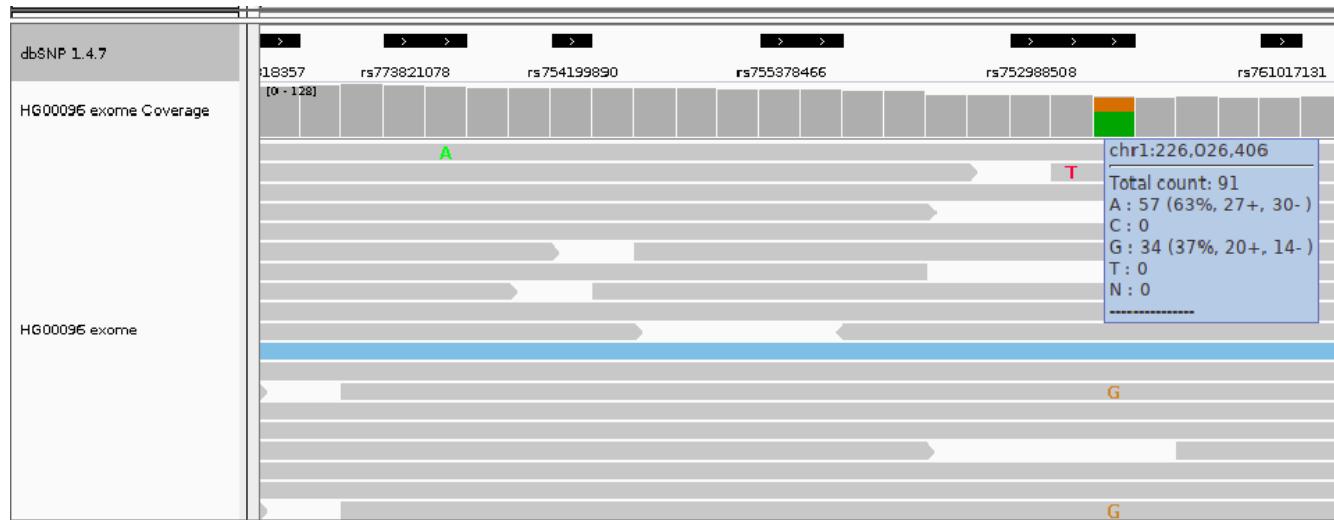
[Table of Contents](#)**Schema for All SNPs(147) - Simple Nucleotide Polymorphisms (dbSNP 147)**

Database: hg19 Primary Table:.snp147 Row Count: 154,822,082 Data last updated: 2016-07-29

Format description: Polymorphism data from dbSNP

field	example	SQL type
bin	585	smallint(5) unsigned
chrom	chr1	varchar(31)
chromStart	10019	int(10) unsigned
chromEnd	10020	int(10) unsigned
name	rs775809821	varchar(15)
score	0	smallint(5) unsigned
strand	+	enum('+', '-')
refNCBI	A	blob
refUCSC	A	blob
observed	-/A	varchar(255)
molType	genomic	enum('unknown', 'genomic', 'cDNA')
class	deletion	enum('single', 'in-del', 'microsatellite', 'named', 'mnp', 'insertion', 'deletion')

Next using the summary track for Bam file for sample HG00096, found which specific SNP the sample was heterozygous for. Another student and I found from help page for AlignmentData for IGV at Broad that “if a nucleotide differs from the reference sequence in greater than 20% of quality weighted reads, IGV colors the bar in proportion to the read count of each base (A, C, G, T).” (Broad Institute)



Use NCBI ClinVar and Ensembl to look up information on structural variants of a protein coding gene.

As a follow to my paper on the [A712T mutation of the high-voltage Cav2.1 Voltage-dependent P/Q-type calcium channel subunit alpha-1A](#) (Cav α 1) in Protein Bioinformatics regarding to the structural implications of the variant on epileptic pathologies, the goal was to complete a meta-analysis study to quantify the amount of variants for this large (>2500 residue) protein.

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First use ClinVar to determine the amount of deletion type variations and of those how many were classified as being pathogenic.

The screenshot shows the ClinVar website interface. At the top, there is a search bar with "CACNA1A[gene]" entered. Below the search bar are links for "Create alert" and "Advanced". A navigation bar at the bottom includes "Home", "About", "Access", "Help", "Submit", "Statistics", and "FTP". On the left, a sidebar titled "Variation type" lists categories: "✓ Deletion (102)", "Duplication (0)", "Indel (8)", "Insertion (8)", and "Single nucleotide (0)". On the right, under "Clinical significance", categories include "Conflicting interpretations (0)", "Benign (0)", "Likely benign (0)", "Uncertain significance (0)", "Likely pathogenic (1)", and "✓ Pathogenic (53)".

As a follow to my original paper, the p.A712T variant was also listed in ClinVar and in dbSNP. These NCBI databases usually will have links to each other, if in ClinVar, the dbSNP rs id entry will indicate this next to “Clinical Significance”, and if in dbSNP, the ClinVar entry will have the dbSNP link to the rs id next to “Links”. However neither of these entries have the valid link to eachother despite the variant being in both databases.

dbSNP:

The screenshot shows the dbSNP entry for rs886037945. The header indicates "Current Build 154" and "Released April 21, 2020". The main table contains the following information:

Organism	Homo sapiens
Position	chr19:13303584 (GRCh38.p12) ?
Alleles	C>T
Variation Type	SNV Single Nucleotide Variation
Frequency	None
Clinical Significance Not Reported in ClinVar	
Gene : Consequence CACNA1A : Missense Variant	
Publications	2 citations
LitVar 3	
Genomic View	See rs on genome

ClinVar:

Interpretation:	Pathogenic/Likely pathogenic
Review status:	criteria provided, multiple submitters, no conflicts
Submissions:	6 (Most recent: Jul 16, 2020)
Last evaluated:	Dec 4, 2019
Accession:	VCV000254268.4
Variation ID:	254268
Description:	single nucleotide variant

Variant details

Conditions	NM_001127222.2(CACNA1A):c.2134G>A (p.Ala712Thr)
Gene(s)	Allele ID: 248801 Variant type: single nucleotide variant Variant length: 1 bp Cytogenetic location: 19p13.13 Genomic location: 19: 13303584 (GRCh38) GRCh38 UCSC 19: 13414398 (GRCh37) GRCh37 UCSC

Links:

[ClinGen: CA10586394](#)
[OMIM: 601011.0036](#)

Finally, use Ensembl to count the structural variants as well as how many of the structural variants are listed in the dbVar at NCBI. We can then export and/or read into R to filter for columns based on database study, e.g. dbVar, Database of Genomic Variants Archive (DGV), etc. and the variant class, e.g. CNV, insertion, etc.



Structural variants					
Name	Chr:bp	Genomic size (bp)	Class	Source Study	Study description
nsv995068	19:10319474-13777860	3,458,387	CNV	DGVA:nstd37	Database of Genomic Variants Archive: Miller 2010 "Consensus statement: chromosomal microarray is a first-tier clinical diagnostic test for individuals with developmental disabilities or congenital anomalies."
nsv992659	19:13381400-13381401	-	mobile element insertion	DGVA:nstd94	Database of Genomic Variants Archive: Helman 2014 "Somatic retrotransposition in human cancer revealed by whole-genome and exome sequencing." PMID:24823667
nsv984831	19:90910-58586487	-	CNV	DGVA:nstd11	Database of Genomic Variants Archive: Walter 2009 "Acquired copy number alterations in adult acute myeloid leukemia genomes." PMID:19651600
nsv911096	19:13319510-13354187	34,678	CNV	DGVA:nstd71	Database of Genomic Variants Archive: Xu 2011 "SgD-CNV, a database for common and rare copy number variants in three Asian populations." PMID:21882294
nsv911095	19:13291258-13359502	68,245	CNV	DGVA:nstd71	Database of Genomic Variants Archive: Xu 2011 "SgD-CNV, a database for common and rare copy number variants in three Asian populations." PMID:21882294

4.7. Week 7 - Genomic data file formats and manipulation tools

Use bedtools sort, subtract, intersect, and closest tools on genomic annotated bed files.

First I installed the most recent version of bedtools from github:

1. Download bedtools-2.29.2.tar.gz from github
2. Run commands to compile

```
tar -xvzf bedtools-2.29.2.tar.gz  
cd bedtools2  
make
```

3. Move bedtools2 folder and add bedtools2/bin to PATH

A)

Returning to the exercise in [Week 5](#) for analyzing intervals of conserved regions in mouse and human on 0.5 mb interval of chromosome 4, we can perform similar operations using bedtools that we performed in galaxy.

Using the subtract tools in bedtools we can subtract overlapping intervals of human exons from the conserved regions with the command:

```
bedtools subtract -a mouse.bed -b human.bed > conserved_not_in_human_exons.bed
```

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And this gives us the same 186 regions in Galaxy using the parameter “Non-overlapping pieces of intervals”. If we wanted the opposite, i.e. only intervals in human exons that overlap conserved regions, we can use the intersect tool with the command:

```
bedtools intersect -a mouse.bed -b human.bed > conserved_only_in_human_exons.bed
```

If we wanted to include all parts of the interval of conserved mouse regions with this intersection, i.e the (-a) parameter mouse.bed, then we can pass the -wa parameter. This will NOT include regions of -a that have no overlap at all with -b.

```
bedtools intersect -a mouse.bed -b human.bed -wa
```

In order to mirror the subtract functionality of Galaxy with parameter “Intervals with no overlap” we use bedtools subtract and the parameter “-A Remove features with any overlap” with the command:

```
bedtools subtract -a mouse.bed -b human.bed -A > conserved_not_in_human_exons_no_overlap.bed
```

Alternatively we can obtain this same function with bedtools intersect passing the “-v Reporting the absence of any overlapping features” parameter with the command:

```
bedtools intersect -a mouse.bed -b human.bed -v > conserved_not_in_human_exons_no_overlap.bed
```

This last two calls returns the same 128 lines from my Galaxy tutorial in Week 5.

B)

In another example we can use bedtools intersect to look at regions of H3K4me3 methylation and intersection with RefSeq genes.

```
bedtools intersect -a hs_chr20_H3K4me3_for_linux.bed -b hs_chr20_refseq.bed > h3k4me3_intersects_RefSeq.bed
```

As well as passing the ‘-v’ parameter for A does not intersect with B and get regions of H3K3me3 that do not intersect exons.

C) A third example we use bedtools sort and bedtools closest to find regions of one annotation that are closest to regions in another. To summarize the closest tool we just run:

```
bedtools closest
```

Tool: bedtools closest (aka closestBed)

Version: v2.29.2

Summary: For each feature in A, finds the closest

feature (upstream or downstream) in B.

To find the closest exons to genome assembly gaps, we set the gap annotation file to be A and B to be the exons file. If the files are not sorted we should sort them first with bedtools sort:

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```
bedtools sort -i hs_chr20_refseq.bed > hs_chr20_refseq_sorted.bed
```

And then we can run:

```
bedtools closest -a hs_chr20_gaps.bed -b hs_chr20_refseq_sorted.bed > hs_chr20_nearest_exon_gap.bed
```

We can also create a new column to report the distance with the `-d` parameter which measures the distance from B in the exons file to the gaps in the gaps file. If there are exons for different transcripts but for the same region then we may get more lines than in file A since these exons would be the same distance. If we just want 1 of these exons we can set the ties `-t` parameter to “first” and it will report the first if there are ties such as:

```
bedtools closest -t first -a hs_chr20_gaps.bed -b hs_chr20_refseq_sorted.bed >  
hs_chr20_nearest_unique_exon_gap.bed
```

View histone modification BAM alignment files for multiple histone modification marks in IGV

If, because we have many and/or large alignment files, we want to just view the alignment(s) for a certain chromosome of the genome or genomic interval of a particular chromosome we can use samtools package to index and subset our SAM/BAM alignment files. For instance, if we were just interested in looking at the alignment near the PARD6B gene for the hg19 assembly and 0.5 Mb upstream and 0.5 Mb downstream, as the location for GRCh37 is NC_000020.10 (49347923..49373333) on chromosome 20, we first index the SAM/BAM file(s) with:

```
samtools index HUES64_rep1.H3K27me3.chr20.sorted.bam
```

```
samtools index HUES64_rep1.H3K4me3.chr20.sorted.bam
```

First to see the format of the ‘SN’ Reference sequence name, as we some plant genomes use a capital ‘C’ such as Chr4, while the more common type use a lowercase ‘c’ to be UCSC compatible, we check the header of the alignment file with:

```
samtools view -H HUES64_rep1.H3K27me3.chr20.sorted.bam
```

```
@HD VN:1.0 SO:coordinate  
@SQ SN:chr20 LN:63025520  
@PG ID:Bowtie VN:1.0.0
```

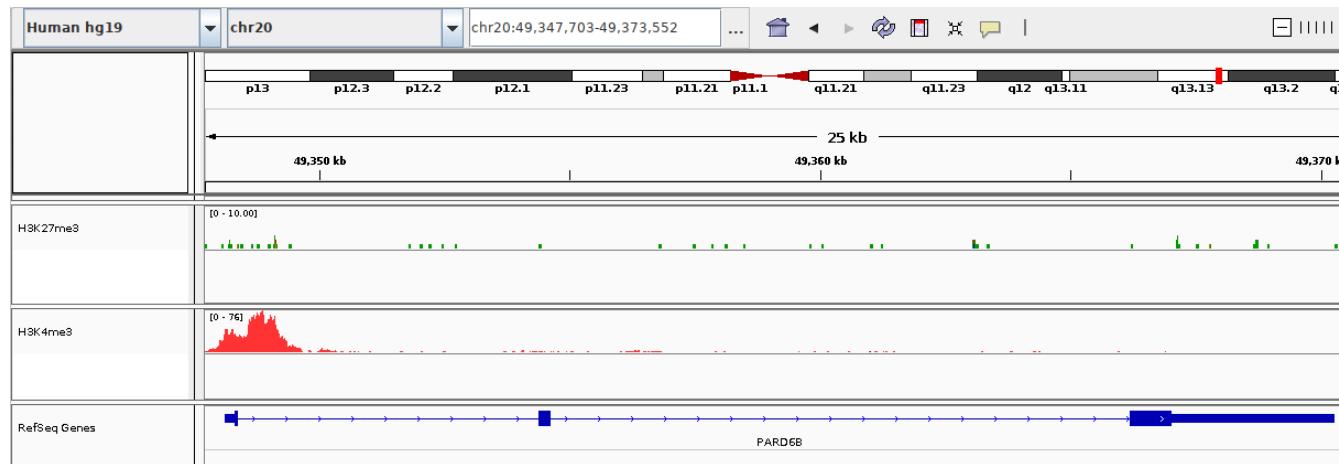
And then subset each SAM/BAM file using samtools view and keeping header `-h` and in BAM format `-b` with:

```
samtools view -h -b HUES64_rep1.H3K27me3.chr20.sorted.bam chr20:48847923-49873333 >  
PARD6B_HUES64_rep1.H3K27me3.chr20.sorted.bam
```

I have listed more about samtools is listed in [7L](#) section under “Be a Teacher”.

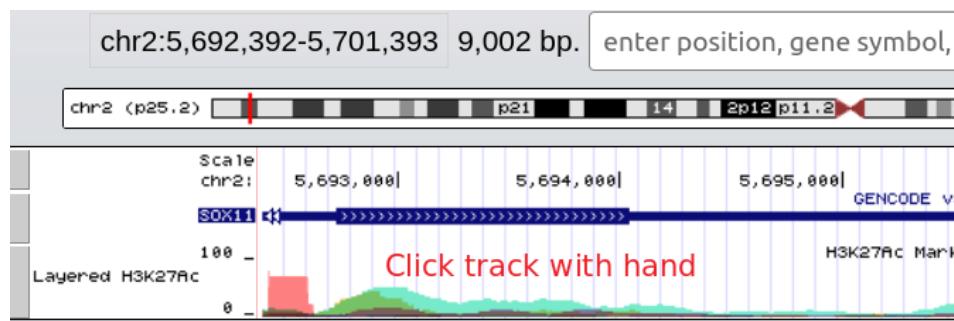
We can then view both alignment files within 0.5 MB of the PARD6B gene in IGV but only after we index the new subsetted alignments, as IGV required .bai index files, with:

```
samtools index PARD6B_HUES64_rep1.H3K27me3.chr20.sorted.bam
```



4.8. Week 8 - The ENCODE Project and model organism genomes

View SOX11 gene for the hg38 assembly in UCSC looking at the H3K27ac data for two cells lines HSSM and K562. Then add DNase hypersensitivity

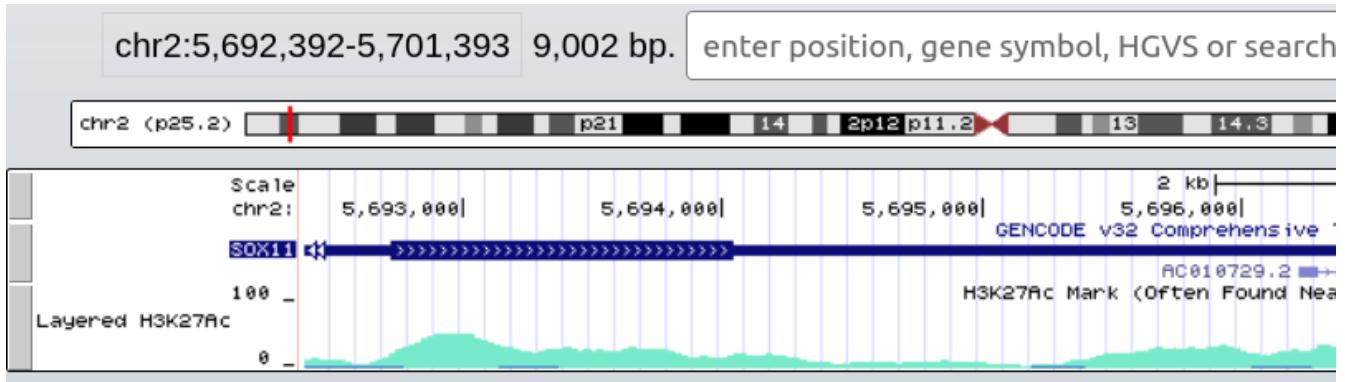


Select only these two cells lines

List subtracks: only selected/visible all (2 of 7 selected)

- GM12878 H3K27Ac Mark (Often Found Near Regulatory Elements) on GM12878 Cells from ENCODE [Schema](#)
- H1-hESC H3K27Ac Mark (Often Found Near Regulatory Elements) on H1-hESC Cells from ENCODE [Schema](#)
- HSMM H3K27Ac Mark (Often Found Near Regulatory Elements) on HSMM Cells from ENCODE [Schema](#)
- HUVEC H3K27Ac Mark (Often Found Near Regulatory Elements) on HUVEC Cells from ENCODE [Schema](#)
- K562 H3K27Ac Mark (Often Found Near Regulatory Elements) on K562 Cells from ENCODE [Schema](#)
- NHEK H3K27Ac Mark (Often Found Near Regulatory Elements) on NHEK Cells from ENCODE [Schema](#)
- NHLF H3K27Ac Mark (Often Found Near Regulatory Elements) on NHLF Cells from ENCODE [Schema](#)

2 of 7 selected



H3K27ac is high in the HSMM cell line at promoter and through the gene body while it is low for K562 cell line. The K562 cell line is a leukemia cell line from the blood of a 53 year old patient why HSMM is a skeleton muscle myoblast cell line. SOX11 is a HMG-box gene involved in embryonic development and determination of cell fate in normal cells. It would make sense that normal cell differentiation is disrupted and SOX11 is not activated with H3K27ac in a leukemia cell line why the gene would be activated on the normal differentiation from myoblast to myocyte.

Adding DNase HS sites for only these two cell lines

-

ENCODE
cCREs

P12 ENCODE
Regulation...

hide

show

full [P12 Layered H3K27Ac](#) H3K27Ac Mark (Often Found Near Regulatory Elements) on 7 cell lines from ENCODE

hide [P12 DNase Clusters](#) DNase I Hypersensitivity Peak Clusters from ENCODE (95 cell types)

hide [P12 DNase Signal](#) DNase I Hypersensitivity Signal Colored by Similarity from ENCODE

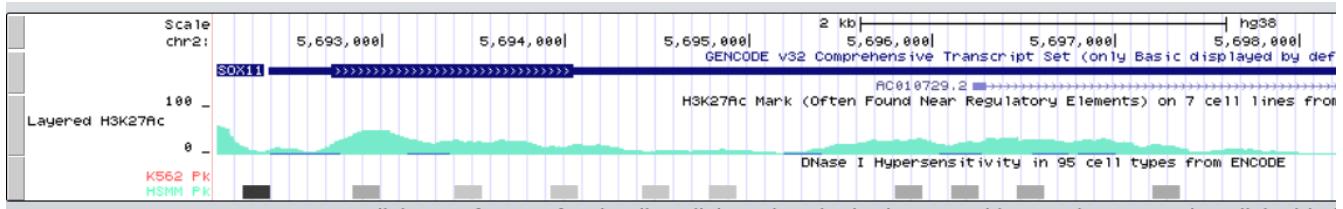
full [P12 DNase HS](#) DNase I Hypersensitivity in 95 cell types from ENCODE

List subtracks: only selected/visible all (2 of 285 selected)

views [Similarity](#) [Cell Type](#) [Tissue](#)

full [Peaks](#) K562 bone marrow
 full [Peaks](#) HSMM muscle

2 of 285 selected



The HSMM cell line has a high scoring DNase HS system just upstream of the promoter.

Use UCSC BLAT and Mouse Genome Informatics (MGI) to perform comparative genomics of human lysozyme with mouse genome.

1. Take the human lysozyme sequence (attached) and run BLAT against the mouse genome at UCSC. Click the browser link for the hit with the longest span. What are the chromosomal coordinates?

BLAT Search Genome

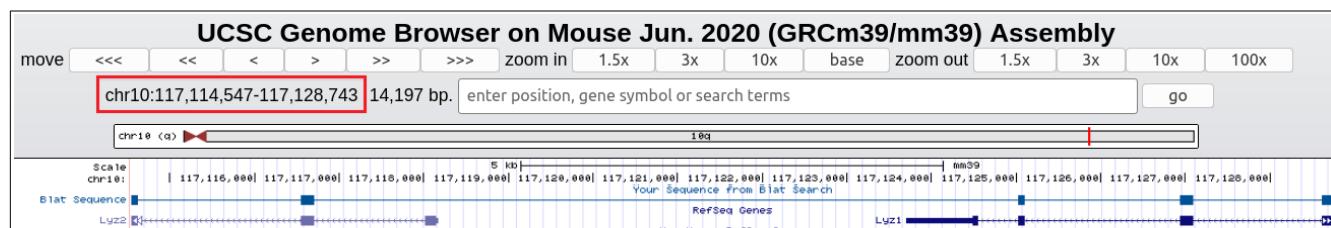
Genome: Search all
Assembly: Jun. 2020 (GRCm39/mm39)

Mouse

```
>human lysozyme precursor
MKALIVLGLVLLSVTVQGKVFERCELARTLKRLGMDGYRGISLANWMCLAKWESGYNTRATNYNAGDRST
DYGIFQINSRYWCNDGKTPGAVNACHLSCSALLQDNIADAVACAKRVVRDPQGIRAWVAWRNRCQNRDVR
QYVQGCGV
```

ACTIONS	QUERY	SCORE	START	END	QSIZE	IDENTITY	CHROM	STRAND	START	END	SPAN
browser details	human	219	1	127	148	79.6%	chr10	+-	117114547	117128743	14197
browser details	human	149	48	127	148	81.3%	chr10	+-	117125023	117127093	2071
browser details	human	60	48	99	148	69.3%	chr10	+-	117073526	117073681	156
browser details	human	45	67	85	148	89.5%	chr11	+-	103525864	103525920	57

chr10:117,114,547-117,128,743



2. Go to MGI with the human sequence and run Mouse BLAST. Be sure to choose BLASTP and the UniProt Mouse database. Your best hit should be Lyz1. What is its score and P value?

P value = 2e-77 or basically 0

Score is 575

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RecName: Full=Lysozyme C-1; AltName: Full=1,4-beta-N-acetylmuramidase C; AltName: Full=Lmusculus]

Sequence ID: [P17897.1](#) Length: 148 Number of Matches: 1

Range 1: 1 to 148 [GenPept](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Method	Identities	Positives	Gaps
226 bits(575)	2e-77	Compositional matrix adjust.	113/148(76%)	129/148(87%)	0/148(0%)

Query 1 MKALIVLGLVLLSVTVQGVFERCELARTLKRLGMDGYRGISLANWMCLAKWESGYNTRA 60
MKAL+ LGL+LLSVT Q KV+ RCELA LKR GMDGYRG+ LA+W+CLA+ ES YNTRA
Sbjct 1 MKALLTLGLLLSVTAQAKVYNRCELARILKRNMGMDGYRGVKLADWVCLAQHESNYNTRA 60

Query 61 TNYNAGDRSTDYGFQINSRYWCNDGKTPGAVNACHLSCSALLQDNIADAVACAKRVVRD 120
TNYN GDRSTDYGFQINSRYWCNDGKTP + NAC ++CSALLQD+I A+ CAKRVVRD
Sbjct 61 TNYNRGDRSTDYGFQINSRYWCNDGKTPRSKNACGINCSALLQDDITAAIQCAKRVVRD 120

Query 121 PQGIRAWVAWRNRQCQRDVRQYVQGCGV 148
PQGIRAWVAWR +CQRD+ QY++ CGV
Sbjct 121 PQGIRAWVAWRTQCQRDLSQYIRNCGV 148

3. Click the Lyz1 link. What synonyms are listed?

Lzp-s is listed at NCBI GenPept entry

gene 1..148 /gene="Lyz1" /gene_synonym="Lzp-s"

As well as in UniProt

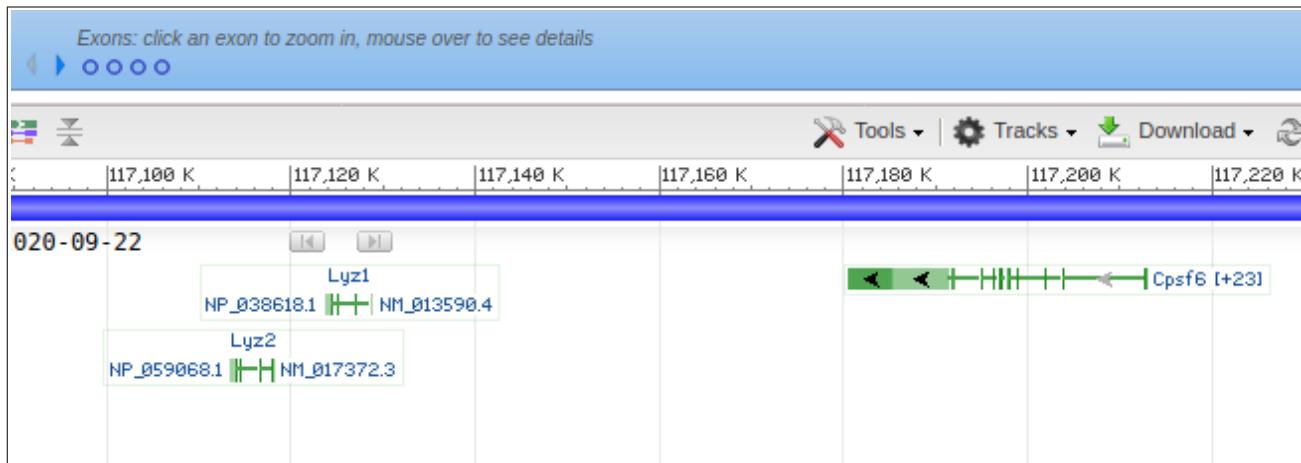
Protein names ⁱ Recommended name:
Lysozyme C-1 (EC:3.2.1.17)
Alternative name(s):

- 1,4-beta-N-acetylmuramidase C
- Lysozyme C type P

Gene names ⁱ Name:**Lyz1**
Synonyms:Lzp-s

4. What are the flanking genes?

Viewing in NCBI Genome Data Viewer we see Lyz2 is just down stream on the negative strand and Cpsf6 just upstream on the negative strand

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5. Click the Ensembl Gene Model under Sequences. How many paralogues in mouse?

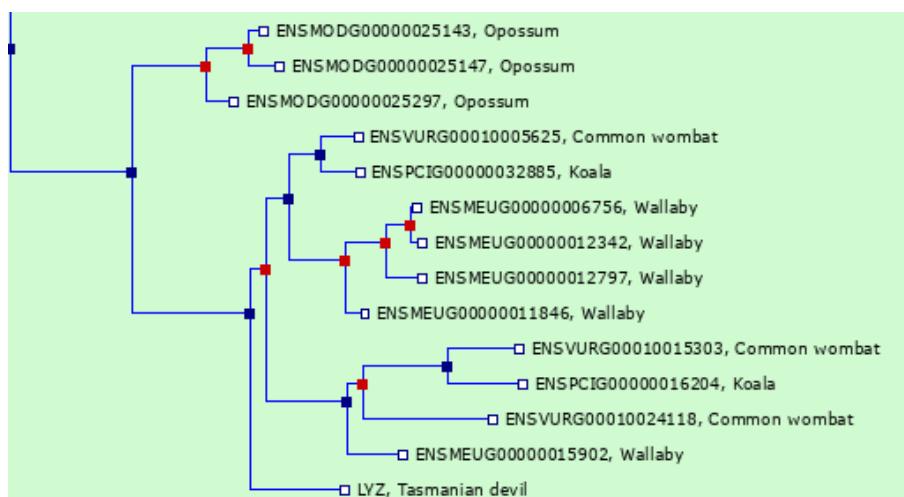
8 paralogues

Gene: Lyz1 ENSMUSG00000069515

Description	lysozyme 1 [Source:MGI Symbol;Acc: MGI:96902]
Gene Synonyms	Lyz, Lzp-s, renal amyloidosis
Location	Chromosome 10: 117,287,797-117,292,868 reverse strand. GRCh38:CM001003.2
About this gene	This gene has 1 transcript (splice variant), 340 orthologues , 8 paralogues & 147 variants .
Transcripts	Hide transcript table

6. Click the Gene Tree image. Expand Placental Mammals. List all primate species that come up.

There are no primate species that come up in Ensembl for Lyz1 despite BLAT having a hit for Human Lysozyme C with 79.6% identity.



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Summary of orthologues of this gene [Hide](#) ⊖

Click on 'Show details' to display the orthologues for one or more groups of species. Alternatively, click on 'Configure this page' to choose a custom list of species.

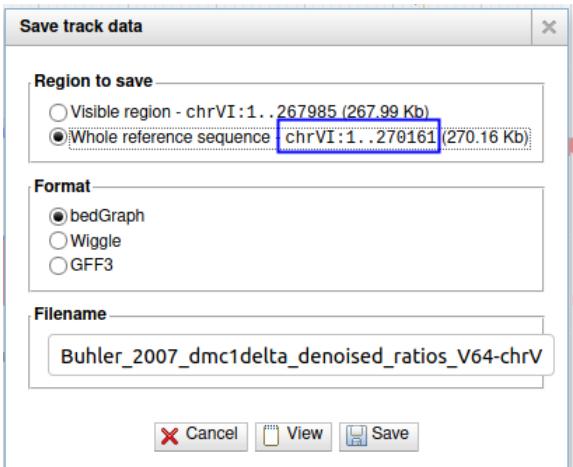
Species set	Show details	With 1:1 orthologues	With 1:many orthologues	With many:many orthologues	Without orthologues
Primates (27 species) Humans and other primates	<input checked="" type="checkbox"/>	0	0	0	27
Rodents and related species (31 species) Rodents, lagomorphs and tree shrews	<input type="checkbox"/>	2	5	4	20
Laurasiatheria (45 species) Carnivores, ungulates and insectivores	<input type="checkbox"/>	0	1	1	43
Placental Mammals (108 species) All placental mammals	<input type="checkbox"/>	2	6	5	95
Sauropsida (19 species) Birds and Reptiles	<input type="checkbox"/>	0	4	15	0
Fish (86 species) Ray-finned fishes	<input type="checkbox"/>	0	36	38	12
All (280 species) All species, including invertebrates	<input type="checkbox"/>	2	91	66	121

Create and document a workflow for students to follow for uploading custom yeast tracks from the Saccharomyces Genome Database (SGD) and NCBI GEO database to UCSC Genome Browser. Posted to discussion forums for students to follow.

I am going to post some screenshots since this is an all visual exercise. I was able to download a few tracks from the SGD database in wiggle format, as well as a BED and bigWig file from the two different not related studies from the NCBI GEO database.

You can download datasets while in the genome viewer of SGD but you can only download based on the chromosome you are viewing doing it this way.



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Alternatively you can download from the link in the (?) question mark "About this track" to get the full dataset for entire genome.

About track: Buhler_2007_dmc1delta_denoised_ratios_V64

Name	Buhler_2007_dmc1delta_denoised_ratios_V64
Accession	GSE8981
Assay Term Name	ChIP-chip assay
Assay term id	OBI:0001248
Biosample Term Name	DNA extract
Biosample id	OBI:0001051
Category	DNA replication, recombination and repair
Datafile name	http://downloads.yeastgenome.org/published_datasets/Buhler_2007_PMID_18076285/track_files/Buhler_2007_dmc1delta_denoised_ratios_V64.wig
Dataset description	Supplementary Table S1 from the supplementary materials accompanying the paper; represents the dmc1delta mutant (averaged between two independent experiments) microarray ratios.

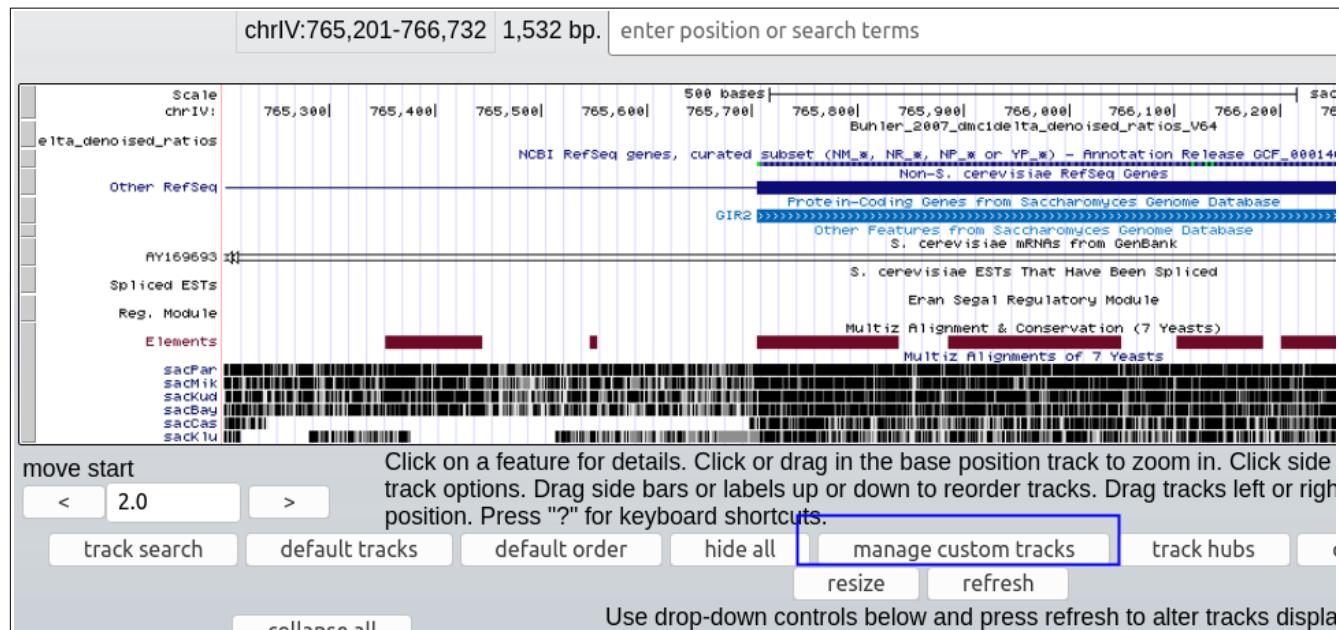
SGD Saccharomyces GENOME DATABASE

Analyze ▾ Sequence ▾ Function ▾

Date	Size	File
2019-10-25T18:45:58.000Z	0.3 kB	Buhler_2007_rad50S_DSB_hotspots_5x_threshold_V27toV64_unmapped.txt
2019-10-25T18:45:57.000Z	308.5 kB	Buhler_2007_rad50S_DSB_hotspots_2x_threshold_V64.gff3
2019-10-25T18:45:58.000Z	52.5 kB	Buhler_2007_rad50S_DSB_hotspots_2x_threshold_V64.bed
2019-10-25T18:45:57.000Z	0.5 kB	Buhler_2007_rad50S_DSB_hotspots_2x_threshold_V27toV64_unmapped.txt
2019-10-25T18:45:57.000Z	515.9 kB	Buhler_2007_dmc1delta_ratios_V64.wig
2019-10-25T18:45:58.000Z	1.0 MB	Buhler_2007_dmc1delta_ratios_V64.bedgraph
2019-10-25T18:45:57.000Z	515.6 kB	Buhler_2007_dmc1delta_denoised_ratios_V64.wig
2019-10-25T18:45:57.000Z	1.0 MB	Buhler_2007_dmc1delta_denoised_ratios_V64.bedgraph
2019-10-25T18:45:57.000Z	259.2 kB	Buhler_2007_dmc1delta_DSB_hotspots_5x_threshold_V64.gff3
2019-10-25T18:45:57.000Z	52.6 kB	Buhler_2007_dmc1delta_DSB_hotspots_5x_threshold_V64.bed
2019-10-25T18:45:57.000Z	469.8 kB	Buhler_2007_dmc1delta_DSB_hotspots_2x_threshold_V64.gff3

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To add to UCSC select "manage custom tracks"



You can upload files here. Depending on which version of UCSC yeast genome you could have issues because they change from Chr numbers (1, 5, 14) to Roman numerals (I, V, XIV).

view in [Genome Browser](#) go

[add custom tracks](#)

Manage Custom Tracks					
genome	S. cerevisiae	assembly	Apr. 2011 (SacCer_Apr2011/sacCer3)	[sacCer3]	v
Name	Description	Type	Doc	delete	
Doube_strand_break_hotspots	Pan_2011_Spo11_uniquely_mapped_read_density_V64	wiggle_0		<input type="checkbox"/>	
dmc1delta_denoised_ratios	Buhler_2007_dmc1delta_denoised_ratios_V64	wiggle_0		<input type="checkbox"/>	

Then you can get datasets from GEO as well.

BED:

[Investigation of impact of DNA synthesis during Break-induced replication on transcription](#)

2. (Submitter supplied) We discovered that during rPolII is enriched in the TES region due collision between transcription and BIR synthesis. This enrichment is specific to genes that are located in head-on orientation to BIR.

Organism: **Saccharomyces cerevisiae**

Type: Genome binding/occupancy profiling by high throughput sequencing

Platform: **GPL19756** 12 Samples

Download data: [BED](#), [BEDGRAPH](#)

Series Accession: GSE159384 ID: 200159384

[SRA Run Selector](#)

bigWig:

[Chromosome localization of cohesin oligomers in mid-M arrested yeast cells](#)

64. (Submitter supplied) We report the genomic localization of cohesin oligomers in nocodazole arrested **yeast** cells. Two alleles of SMC3 were expressed in **yeast** cells, one fused to BirA enzyme and the other tagged with AviTag. Cohesin oligomers were biotinylated and ChIP with streptavidin beads. As control experiments, cohesin localization on chromosome was determined in strains expresses freely diffusible BirA enzyme, where all Smc3 proteins were biotinylated; non-specific ChIP were determined in strains with no BirA.

Organism: **Saccharomyces cerevisiae**

Type: Genome binding/occupancy profiling by high throughput sequencing

Platform: **GPL27812** 3 Samples

Download data: [BW](#)

Series Accession: GSE157155 ID: 200157155

[SRA Run Selector](#)

However you cannot add binary bigWig files to UCSC this way.

See [here](#). If you do not have web accessible data storage available, please see the [Networking](#) section of the Track Hub Help documentation.

Please note a much more efficient way to load data is to use [Track Hubs](#), which are loaded from the [Track Hubs Portal](#) found in the menu under My Data.

Error File 'GSM4756782_O.bw' - It appears that you are directly uploading binary data of type bigWig. Custom tracks of this type require the files to be accessible by public http/https/ftp. Our [track hub documentation](#) lists third-party services where you can store custom track or track hub files. Once the files are available on the internet, file URLs can be entered as-is, one per line, or via the bigDataUrl setting on a "track" line. See [bigWig custom track documentation](#) for more information and examples.

Paste URLs or data:

Or upload:

[Browse...](#)

No file selected.

[Submit](#)

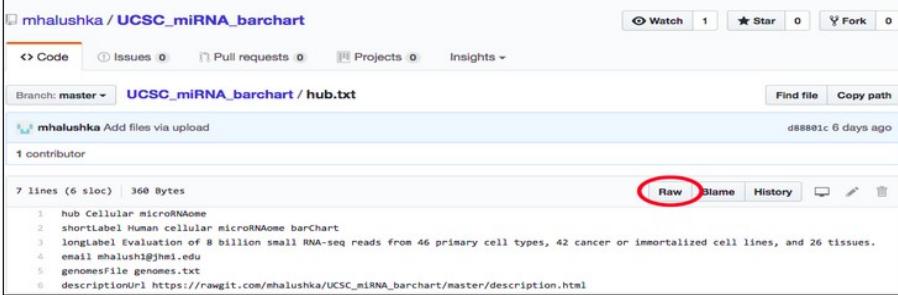
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However you can use github to upload files up to 100Mb in size and then copy the raw url to upload:

<https://genome.ucsc.edu/goldenPath/help/hgTrackHubHelp.html#Hosting>

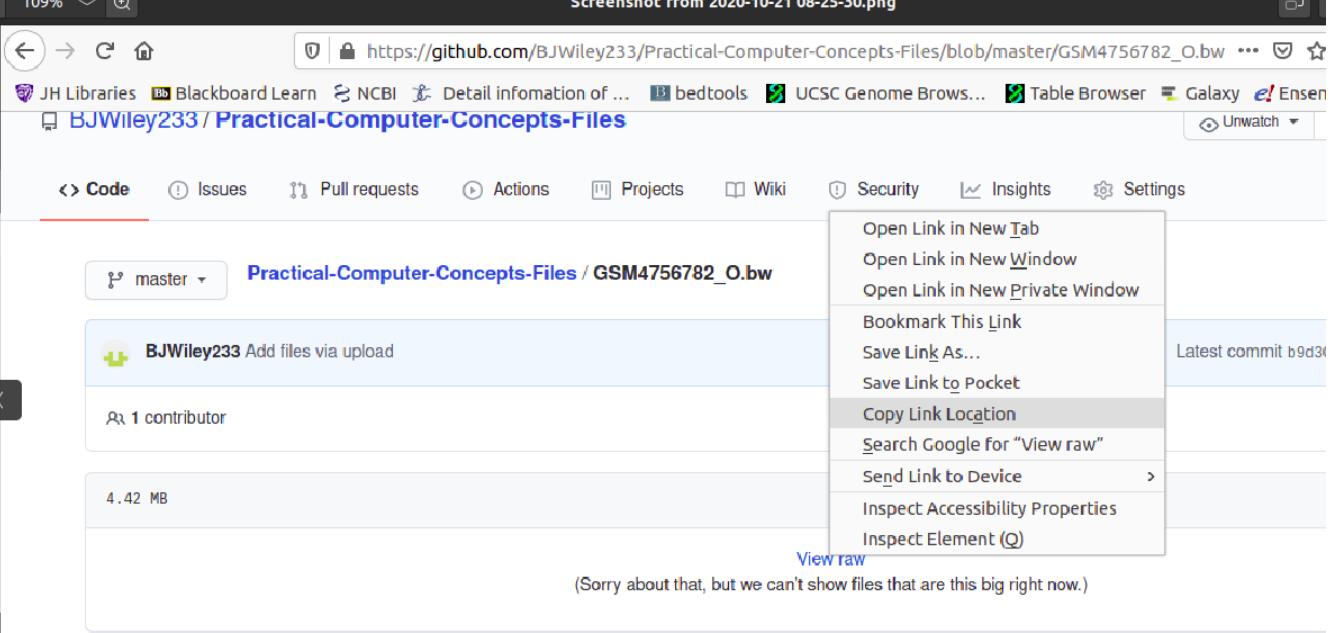
Hosting Hubs on Github

Github supports byte-range access to files when they are accessed via the `raw.githubusercontent.com` style URLs. To obtain a raw URL to a file already uploaded on Github, click the `Raw` button:



The screenshot shows a GitHub repository page for "mhalushka / UCSC_miRNA_barchart". A file named "hub.txt" is listed. At the bottom right of the file card, there is a "Raw" button, which is circled in red.

Screenshot from 2020-10-21 08:25:30.png



The screenshot shows a GitHub file page for "Practical-Computer-Concepts-Files / GSM4756782_O.bw". A context menu is open over the file, with the "Copy Link Location" option highlighted. The menu also includes options like "Open Link in New Tab", "Bookmark This Link", and "Save Link As...".

do not have web-accessible data storage available, please see the [Hosting](#) section of the Track Hub Help documentation.

Please note a much more efficient way to load data is to use [Track Hubs](#), which are loaded from the [Track Hubs Portal](#).

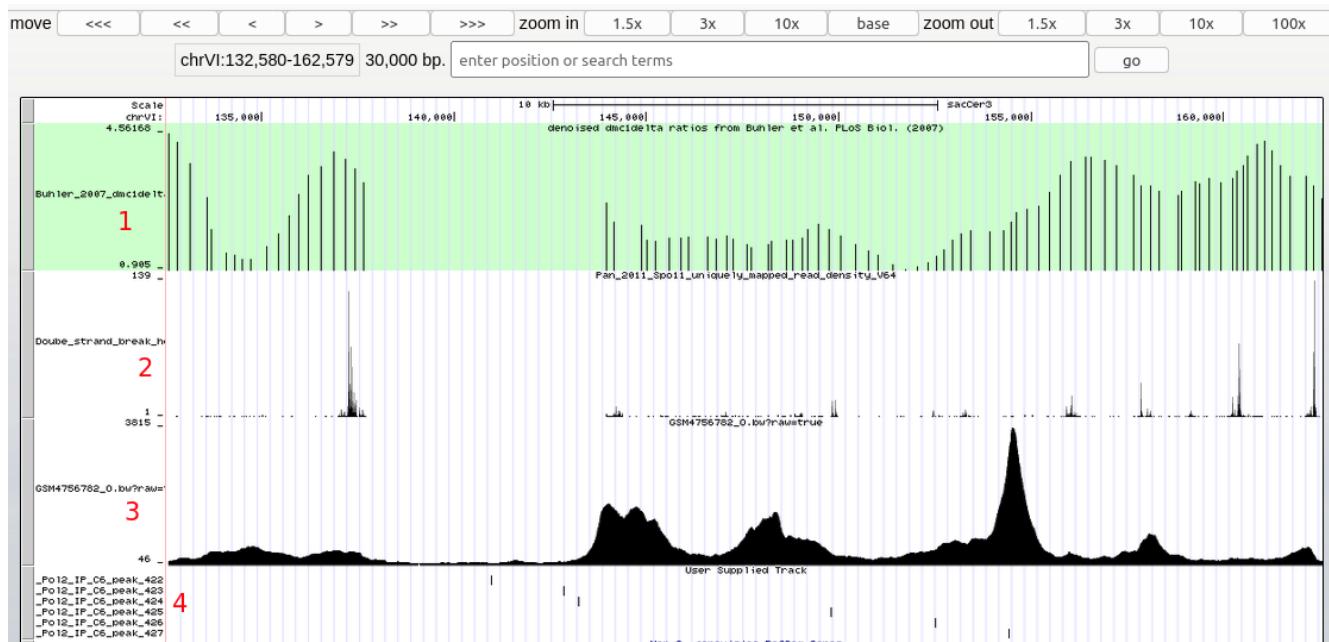
Error File 'GSM4756782_O.bw' - It appears that you are directly uploading binary data of type bigWig. Custom accessible by public http/https/ftp. Our [track hub documentation](#) lists third-party services where you can store are available on the internet, file URLs can be entered as-is, one per line, or via the bigDataUrl setting on a "track hub documentation" for more information and examples.

Paste URLs or data: Or upload: No file selected.

`https://github.com/BJWiley233/Practical-Computer-Concepts-Files/blob/master/GSM4756782_O.bw?raw=true`

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In the below I have 4 custom tracks. The top 2 from SGD are wiggle format (#1 and #2), then a BW (#3) from GEO and a BED from GEO (4):



4.9. Week 9 - Noncoding RNAs and ultraconserved regions

This week was an integration between the week we covered RNA databases and tools in my Biological Databases course. The project for this was to develop and create an RNA database schema for all types of non-coding RNAs including mRNA, lncRNA, tRNA, miRNA, miRNA, shRNA for the sake of the following 3 goals:

- First goal is to look for various RNA molecule types that are associated with the same or similar disease(s).
- The next is use sequence analysis algorithm (some sort of Neighborhoods analysis) to be able to search experimental sequences from Next gen sequencing to find existing similar RNA molecules or to determine which molecule your sequence is from.
- For the sequences attribute which is one of the extra attributes you can search with the lncRNA and interfering RNAs on the sequences of other RNA types including its same type (i.e. tsix and Xist) to predict additional targets then those that are listed from the source.

The interface would have a few search fields as well as filtering fields to update the search fields. Based on selected filters in first column of interface you can print reports or advance to second column to search based on those filters. Lastly you can search for alignments with high identity

to determine if your RNA-sequence is a type of RNA molecule or is an siRNA which have closer identity to targets or search for targets if lower threshold is selected such as that for miRNA.

The following RNA database sources were used:

1. [NCBI Gene](#) (mRNA)
 2. [NCBI GEO](#) (mRNA)
 3. [NONCODE](#) (lncRNA)
 4. [LncTarD](#) (lncRNA)
 5. [GtRNAdb](#) (tRNA)
 6. [miRBase](#) (miRNA)
 7. [DIANA miRGen](#) (miRNA)
 8. [DIANA TarBase](#) (miRNA)
 9. [MIT/ICBP siRNA Database](#) (siRNA & shRNA)

Interface:



Filters

RNA Type

Target type

Disease

Amino Acid/Codonon

Reports

} Filters Search/
Select on top
2 fields

Search>Select

Identifier (RNA or GEO)

Target/Target ID

Disease Name

Filters Search/
Select on top
2 fields

**Alignment/
Target Search**

Enter identifier...
or paste sequences in Fasta
(genomic or RNA)

>Fasta1
TTTTAATTG
>Fasta2
TTTTAATTA

Threshold

80%

Upload file

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Data Dictionary:

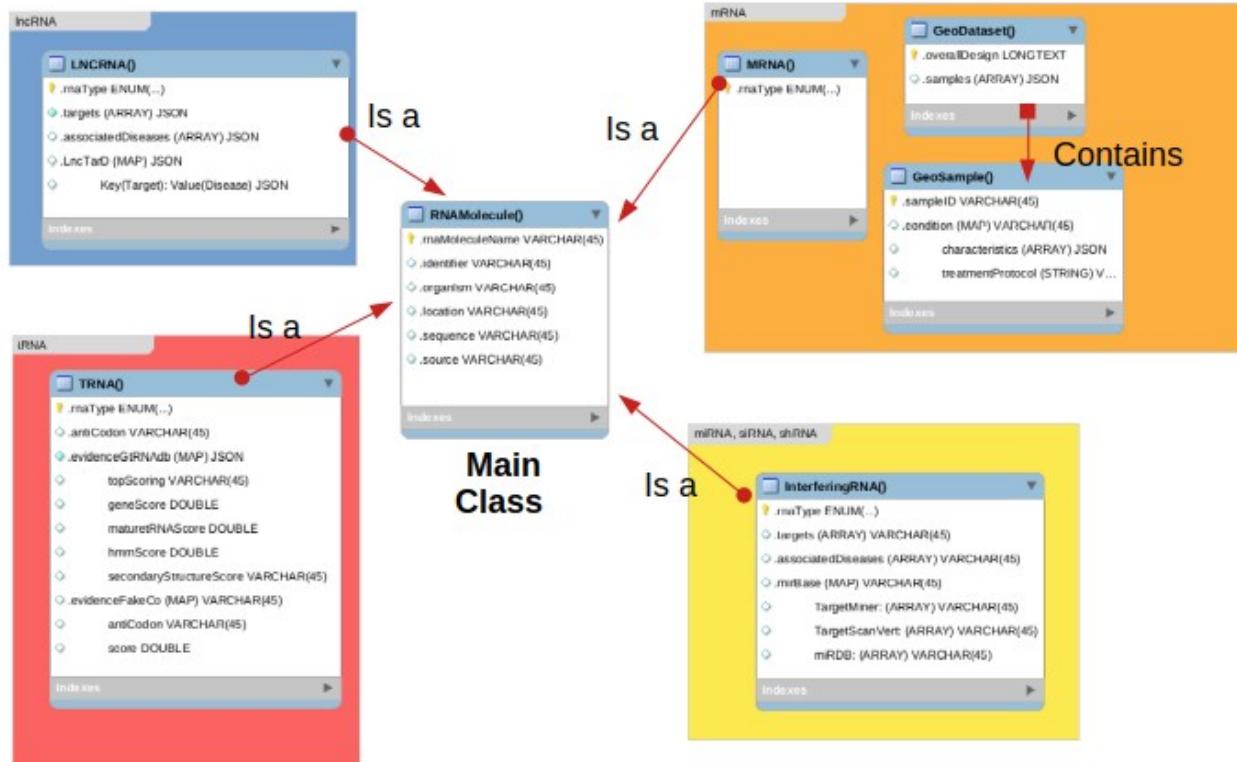
Data element name	Description	Source	Source field	Datatype	Example(s)
rnaMoleculeName	RNA molecule name from database	1. NCBI Gene GenBank file 2. NONCODE 3. GtRNAdb 4. miRBase 5. DIANA miRGen 6. MIT/ICBP siRNA Database	1. product 2. KeyName 3. GtRNAdb Gene Symbol 4. ID 5. miRNA name 6. N/A	String/ VARCHAR	1. BRCA1 DNA repair associated, transcript variant 1 2. lnc-BRCA1-1-5_dup1 3. tRNA-Arg-CCT-2-1 4. hsa-mir-4698 5. hsa-mir-192 6. NONE
identifier	The id for the lncRNA from database	1. NCBI Gene GenBank file 2. NONCODE 3. GtRNAdb 4. miRBase 5. DIANA miRGen 6. MIT/ICBP siRNA Database	1. transcript_id 2. NONCODE TRANSCRIPT ID / NONCODE ID 3. tRNAscan-SE ID 4. Accession 5. DIANA miRNAGen 6. siRNA ID#	String/ VARCHAR	1. NM_007294.4 2. NONHSAT053833.2 3. chr17.tRNA19 4. MI0017331 5. MI0000234 6. 1107
rnaType	1 of 5 types of RNA	1. NCBI Gene GenBank file 2. NONCODE 3. GtRNAdb 4. miRBase 5. DIANA miRGen 6. MIT/ICBP siRNA Database	1. FEATURES 2. NONCODE 3. GtRNAdb 4. N/A 5. N/A 6. Probe type	String enum(mRNA, lncRNA, tRNA, miRNA, siRNA, shRNA)	1. mRNA 2. lncRNA 3. tRNA 4. miRNA 5. miRNA 6. shRNA
targets	Target gene(s), protein (transcription factors), or RNA	1. LncTarD 2. miRBase 3. DIANA miRGen 4. MIT/ICBP siRNA Database	1. Target 2. Predicted targets 3. TF name 4. Target gene	MAP for LncTarD & miRBase or ARRAY	1. {miR-214-3p: Epithelial ovarian cancer, WNT1: Colon cancer} 2. {TargetMiner: [NM_001122679, NM_000024, NM_000024], TargetScanVert: [RP11-379H8.1, UBE2W, LAGE3], miRDB: [BRWD3, MED13L]} 3. [GATA3, Jun, Fos] 4. [AKT1]
antiCodon	The anticodon with the associated amino acid	GtRNAdb	Predicted tRNA Isotype / Anticodon	String/ VARCHAR	CCT (Arg)
evidenceGtRNAdb	The scores from the tRNAscan-SE for the prediction evidence in GtRNAdb	GtRNAdb		MAP of Top Scoring, Gene Score, Mature tRNA Score, HMM Score, Secondary Structure Score	Arg (99.8 bits) 71.1 71.1 52.80 18.30

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evidenceFakeCo	The evidence from Fake Company, Inc.	Evidence from user Community	Anticodon and score	MAP of Anticodon (Residue), Score	CCT (Arg) 85
geoID		NCBI GEO		String/ VARCHAR	GSE123068
geoCondition	This will be the condition being tested that is stated in the design of the dataset.	1. NCBI GEO Dataset 2. Internal GEO Sample within the dataset 3. Internal GEO Sample within the dataset	1. Overall design, 2. Characteristics 3. Treatment protocol	String/ LONGTEXT and/or Array of Sample Objects	1. 3' RNA-Seq (QuantSeq) profiling of sorted cDCs populations from WNT1 overexpressing and control (Empty) lung tumors. 2. cell type: Ovarian Clear Cell Carcinoma cell line 3. No treatment
genomicSequence	The genomic sequence. Some have both genomic and RNA with uracil	1. NCBI Gene GenBank file 2. NONCODE 3. GtRNAdb 4. miRBase 5. DIANA miRGen 6. MIT/ICBP siRNA Database	1. ORIGIN with join() 2. Sequence 3. Genomic Sequence 4. Sequence 5. DIANA miRNAGen 6. Sequence	String/ LONGTEXT	DNA sequence. Can be long for some and short for other short type RNAs.
location	Location in genome including chromosome and positions (concatenated if multiple fields, with hyphen if all 3 fields)	1. NCBI Gene GenBank file 2. NONCODE 3. GtRNAdb 4. miRBase 5. DIANA miRGen	1. chromosome, REGION 2. Chromosome, Start Site, End Site 3. Locus 4. Genome context 5. TSS Coordinates	String/ VARCHAR	1. 17:complement(43044295..43125364) 2. chr17:43221167-43303331 3. chr17:75034431-75034503 (-) 4. chr12: 47187812-47187891 [+] 5. chr11:64660708-64660709 [-]
associatedDiseases	Diseases associated with RNA	1. NONCODE 2. LncTarD 2. DIANA miRGen 3. DIANA TarBase	1. Disease name 2. DiseaseName 3. Cluster diseases 4. Related Diseases	ARRAY	1. AIDS, dermatomyositis 2. Hepatocellular carcinoma 3. Adenocarcinoma, Colorectal Neoplasms, Hematologic Neoplasms 4. Adenocarcinoma, Colorectal Neoplasms, Hematologic Neoplasms, Uterine Cervical Neoplasms
Source, sourceEvidence	Which database or source the database came from	The database that is queried		String/ VARCHAR, LONGTEXT	Database name from each database and all their evidence from that entry that was obtained.

Schema :

This is an Object Oriented database that can be implemented in Mongo DB or Hadoop Ozone.



4.10. Week 10 - Next Generation Sequencing and Analysis

Perform NGS analysis using command-line tools. Although these analysis were performed from the command-line, each one of them can also be performed in Galaxy. The example sample is one from the [SNP Project section 5](#) from the Sequence Read Archive (SRA). An entire workflow to run a list of SRR IDs is available on [github.com here](#).

NCBI SRA-toolkit to download SRR sample files with raw Illumina paired-end reads.

srr.ids file:

SRR10023023
SRR10022985
SRR10022871
SRR10022930

Command:

```
parallel -a srr.ids fastq-dump --split-files --origfmt --gzip;
```

fastqc

Command:

```
parallel -a srr.ids fastq-dump --split-files --origfmt --gzip;
```

Output:

Summary

- [Basic Statistics](#)
- [Per base sequence quality](#)
- [Per sequence quality scores](#)
- [Per base sequence content](#)
- [Per sequence GC content](#)
- [Per base N content](#)
- [Sequence Length Distribution](#)
- [Sequence Duplication Levels](#)
- [Overrepresented sequences](#)
- [Adapter Content](#)

Basic Statistics

Measure	Value
Filename	SRR10022871_1.fastq.gz
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	41517834
Sequences flagged as poor quality	0
Sequence length	35-76
%GC	50

Per base sequence quality

trimmomatic

Command:

```
java -jar /path/to/trimmomatic-0.39.jar PE SRR10022871_1.fastq.gz SRR10022871_2.fastq.gz \
    SRR10022871_1.trim.fastq.gz SRR10022871_1un.trim.fastq.gz \
    SRR10022871_2.trim.fastq.gz SRR10022871_2un.trim.fastq.gz \
    SLIDINGWINDOW:4:20 \
    MINLEN:20;
```

After running fastqc again:

Summary

- [Basic Statistics](#)
- [Per base sequence quality](#)
- [Per sequence quality scores](#)
- [Per base sequence content](#)
- [Per sequence GC content](#)
- [Per base N content](#)
- [Sequence Length Distribution](#)
- [Sequence Duplication Levels](#)
- [Overrepresented sequences](#)
- [Adapter Content](#)

Basic Statistics

Measure	Value
Filename	SRR10022871_1.trim.fastq.gz
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	38316105
Sequences flagged as poor quality	0
Sequence length	20-76
%GC	50

Per base sequence quality

bwa

bwa requires running alignment first for each sample in paired-end to output the forward and reverse sequence alignment index (sai) files before running alignment. It also requires creating the index for the reference.

Commands:

```
bwa index hg19.fa -p hg19
bwa aln hg19 SRR10022871_1.trim.fastq.gz > SRR10022871_1.trim.sai
bwa aln hg19 SRR10022871_2.trim.fastq.gz > SRR10022871_2.trim.sai
bwa sampe hg19 SRR10022871_1.trim.sai SRR10022871_2.trim.sai SRR10022871_1.trim.fastq.gz
SRR10022871_2.trim.fastq.gz > SRR10022871_paired.hg19.trim.sam
```

bowtie2

bowtie2 has indexes that can be downloaded but indexes can also be created before aligning.

Commands:

```
bowtie2-build hg19.fa hg19;
```

```
bowtie2 -x hg19 -1 SRR10022871_1.trim.fastq.gz -2 SRR10022871_2.trim.fastq.gz --threads 32 --fast -S SRR10022871_paired.hg19.trim.sam;
```

samtools sort

Sorting the alignment file as well as creating a bam index .bai file is required to run variant caller freebayes.

Commands:

```
samtools sort SRR10022871_paired.hg19.trim.sam -o SRR10022871_paired.hg19.trim.sorted.bam -O bam;  
samtools index SRR10022871_paired.hg19.trim.sorted.bam;
```

freebayes

Option to run freebayes on a specific region is used.

Command:

```
freebayes -f hg19.fa SRR10022871_paired.hg19.trim.sorted.bam -r chr17:0-81195210 >  
SRR10022871_paired.hg19.trim.vcf;
```

VCFlib

vcffilter

Filtering for allele frequency = 0.5 since we have two samples in the paired-end

Command:

```
vcffilter -f "DP > 10 & AF = 0.5" SRR10022871_paired.hg19.trim.vcf >  
SRR10022871_paired.hg19.trim.filtered.vcf
```

vcfannotate

Using the geneRef table for NCBI RefSeq as indicated by GATK, we can annotate our vcf file for variants that overlap certain regions.

Command:

```
vcfannotate -b refGene_BRCA1_exons_hg19_transcript1.bed \
-k REFGENE \
SRR10022871_paired.hg19.trim.filtered.vcf >
SRR10022871_paired.hg19.trim.filtered.annot.vcf
```

We can then get the lines with:

```
grep "^[^#]" SRR10022871_paired.hg19.trim.filtered.annot.vcf | grep "REFGENE"
```

4.11. Week 11 – ChIP-seq

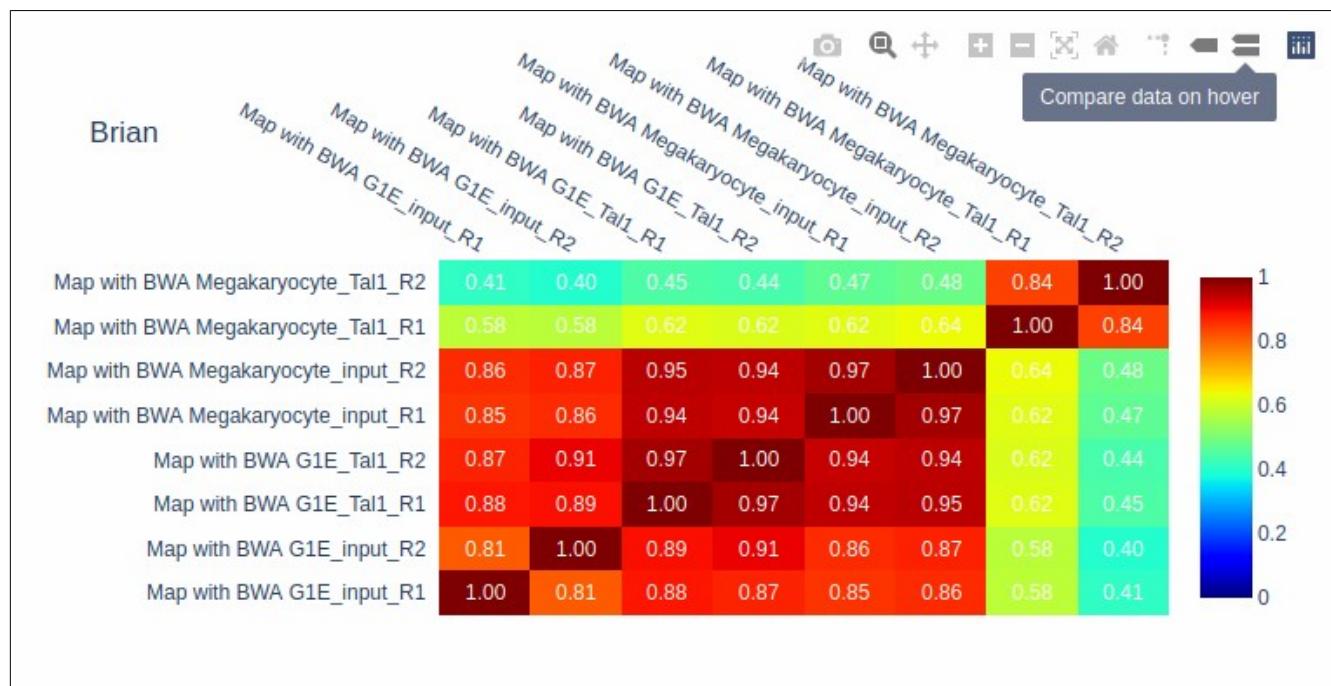
deeptools

```
plotCorrelation -in 'Galaxy57-[multiBamSummary_on_data_56,_data_55,_and_others__correlation_matrix].deeptools_coverage_matrix' --corMethod pearson --whatToPlot heatmap --plotTitle "Brian" --plotFileFormat plotly --plotNumbers --removeOutliers --skipZeros -o test.htm
```

<https://github.com/deeptools/deepTools/issues/1013> (enhancements)

@TODO

Add dendrogram to plotly code for deeptools



macs2

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4.12. Week 12 - RNA-seq: Galaxy

4.13. Week 13 - RNA-seq analysis using command-line and R

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4.14. Week 14 – Bioinformatics for gene editing (CRISPR-Cas9) & Precision medicine from genomics

5. SNP Project

SNP Choice

The HGVS for the SNP variant that we chose, rs1799966, for the hg38 assembly is NC_000017.11:g.43071077T>A[1]. The gene is the BRCA1 DNA repair associated gene and is located on the minus strand with a cytogenetic location of 17q21.31[2]. The primary isoform 1 transcript variant is 7,088 nt long and the most observed mutation, occurring at mRNA position 4950 in exon 15 of isoform 1,[3] is mutated from adenine to thymine responsible for the missense mutation from wild-type serine [AGT] to cysteine [TGT] at location 1613 of the 1863 residue length protein[4]. rs1799966 is associated with the age of diagnosis of hereditary breast cancer[5]. There have also been reports that this SNP affects chemotherapy response and survival of non-small cell lung cancer[6]. The inspiration for choosing this SNP comes from the paper “Next Generation Sequencing Reveals High Prevalence of BRCA1 and BRCA2 Variants of Unknown Significance in Early-Onset Breast Cancer in African American Women” (PMID 28439188)[7]. Another notable paper that has more extensive statistical analysis on rs1799966 is “Association between BRCA1 polymorphisms rs799917 and rs1799966 and breast cancer risk: a meta-analysis” (PMID 30832521)[8].

[Links to Resources](#) for above.

We utilized a variety of tools including but not limited to SIFT, Poly-Phen2, NCBI Genome Data Viewer, UCSC Genome Browser, Ensembl variation, dbSNP, ClinVar, UCSC Variant Annotation Integrator, Galaxy, VCFlib, bowtie2, freebayes, and samtools to perform SNP analysis on rs1799966 and within the BRCA1 gene.

Rough draft of our paper is available [here](#).

Final paper will be available in December, 2020.

6. Issues

This section describes unique issues that arise from using tools, communication and teaching between students, and opportunity for tool enhancements.

1. Issue regarding documentation @ NCBI’s dbSNP database for the frequency tab.

Sub issues:

- a. Entry in dbSNP and ClinVar for same variant but neither links to the other where this usually occurs. This issue is shown as [example under Week 5](#).

2. More issues available when portfolio is completed in December, 2020.

7. “Be a Teacher. Please, please, please be a teacher... Even if you’re not a teacher, be a teacher” -Tim Minchin

This section, maybe the longest one of them all, is inspired by the commencement speech by [Tim Minchin](#) to give a list of experiences where I could act as teacher to help my fellow students. This includes discussions, threads on slacks, and e-mails. I also include links to 3 of my PowerPoint presentations in which I felt a lot of passion during presenting those to my fellow classmates. Each entry or teaching experience is proceed by a letter starting with A) and going on to AA), AB)...BC) and so forth. Each entry briefly describes what the experiences entails.

A) During the course project, my teammate asked about why he/she was not able to find the join(..) statements for the CDS in a GenBank entry with the prefix NM. Here is my response:

Sometimes NM entries don't have the join. I saw this happen often or it will list the exons all as separate features so you can extract the exons however Biopython can do this with their parsing module with joins so separate features is not necessary. Mostly it's the "Gene" Genbank record that has the joins. So this mRNA entry has it here:

go to the

db_xref="GeneID:672"

in NM_007294.4. Then go to [Genomic regions, transcripts, and products](#) and select the [GenBank](#) link. It defaults to the region in genome:(edited)

GenBank ▾ Send to: ▾ Change region shown
Whole sequence (abbreviated view)
Selected region
from: 43044295 to: 43125364
Update View

Homo sapiens chromosome 17, GRCh38.p13 Primary Assembly
NCBI Reference Sequence: NC_000017.11
FASTA Graphics
LOCUS NC_000017 81070 bp DNA linear CON 17-AUG-2020

Then scroll down to CDSs (there are 5 of them) and confirm its the correct transcript variant in note:

```
CDS          join(1269..1348,9586..9639,18832..18909,20409..20497,  
21104..21243,25485..25590,28076..28121,29443..29519,  
30505..33930,34333..34421,42790..42961,48751..48877,  
50844..51034,54127..54437,57670..57757,61414..61491,  
61992..62032,68230..68313,74248..74302,76171..76244,  
77662..77722,79563..79687)  
/gene="BRCA1"  
/gene_synonym="BRCA1; BRCC1; BROVCA1; FANCS; IRIS; PNCA4;  
PPP1R53; PSCP; RNF53"  
/note="isoform 1 is encoded by transcript variant 1;  
Derived by automated computational analysis using gene  
prediction method: BestRefSeq."  
/codon_start=1  
/product="breast cancer type 1 susceptibility protein  
isoform 1"  
/protein_id="NP_009225.1"  
/db_xref="CDS:CDS11453.1"  
/db_xref="Ensembl:ENSP00000350283.3"  
/db_xref="GeneID:672"
```

B) In discussions for connecting to a remote server at the University, our professors asked what are ways to download information. After realizing BlackBoard would require UN and PW here is my response:

Hey All,

FYI for peer collab unit 2. Under Part Four if you want to upload files to the server from blackboard, you need to provide wget with username and password to access downloads from blackboard. So for example after you make directories for "unit2" and subdirectory for "fastas" (optional) you can get the fastas for S. malt by the code below with user name being your email without @jh.edu and your password:

```
wget --user bwiley4 --password <your password>
'https://blackboard.jhu.edu/bbcswebdav/pid-8336676-dt-content-rid-91961117_2/xid-91961117_2' -O smalprt.fasta
wget --user bwiley4 --password <your password>
'https://blackboard.jhu.edu/bbcswebdav/pid-8336676-dt-content-rid-91961111_2/xid-91961111_2' -O smalprt.fasta
```

Then you can check if it worked with this and you should see same results:

```
wc -l smalprt.fasta
# 93 smalprt.fasta
wc -l smalt.fasta
# 69320 smalt.fasta
```

C) I talk about here while reviewing different bacterial gene prediction programs my experience seeing different start codons being supplied.

So interestingly for part 1 of peer collab, I was seeing start codons in FGENESB of 'TTG' for 2247, 3653 start and stop (+) and also a start codon of 'GTG' 8859, 9728 start and stop (+) and I found [this paper](#) that indicates that FGENESB will use TTG as a start codon. The *brpA* gene was predicted to have different start codons using FGENESB depending on the settings used; the alternative start codon TTG (leucine) was predicted using "generic bacterial" And [this page](#) indicates that GTG along with ATG are the most common start promoters. So I did not know that programs and research has shown that start promoters can be other than ATG so it is important to check the docs for the program.

D) After hearing a lot about Augustus and downloading and trying out myself here I indicate my excitement about the tool in relation to functionality of that to another tool.

As I have read so much about Augustus I decided to download it and they have a cool script (getAnnoFasta.pl) to extract the sequences from the output which if you see on their website is a [gff](#) format output. This script is like that of glimmer "extract" protocol. Here is 1 of the ouput files with the exons extracted.

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```
1 >g1.t1.cds1
2 atgaacctgtccaaagctgaagctgtcgacatcacccaggcatccagaagctcaacagaggagtgcag
3 >g1.t1.cds2
4 gtccccttgcaatgacaccaggactggcacaaggcttcaaggatcgaa
5 >g1.t1.cds3
6 ctgtcagagaaggactgtgcaggctgccccacatggactgcagaaggactacgagccctcatcaccatcgtaagagccgcacggcatgacg
7 >g1.t1.cds4
8 aactgcccggccggacccgacaatgagatctaccctggacttggggactacacccaggcgctgtacggcgatctggccacagctgcaaaactga
9 >g2.t1.cds1
10 atgagctctcaactgcccacccctgtggactgtggctgtggccggctgggtgtgcccacgctgtgcttacagctctcagtgcgcgtatggagagcatccggatgtgaatgacatccaggagag
11 >g2.t1.cds2
12 gtttcctgcgtcaagatgaaacgtgacagatatcttgagacaataag
13 >g2.t1.cds3
14 acaaaataaaaaactgagctttatgcaaagccccaattttggagagccggactgcacaagaacctgcagggtctttcaacatgcgtcagctctgaatgccagcagcacccctcaag
15 >g2.t1.cds4
16 gcaccatgtccccccggcggcggcaactacttcaatggagaagttcttagcagacctacgtaccccttccaccaactagctaaaaataagtqa
17
```

E) At the start of the week on Biomart, I am excited to off my help as I had a lot of experience and self teaching after learning and researching Bioconductor “biomaRt” package. I received a lot of questions which was exciting.

Hi All! Welcome to week 4. I was super excited to see Ensembl and Biomart as the topic. I have been working with the "biomaRt" library in R for the past year and profusely with my database class last week. I would consider myself extremely experienced in the R package if you need help you have questions about your output.

Question 1:

Hi Brian, I have been trying the initial steps to install and get used to the biomaRt package by following the user guide provided. I selected a dataset of homosapiens and then started to build biomaRt query. So when I put in "listFilters()" it shows me error. Can you help me with the same? I will attach a picture here. Am I missing a step?

My response:

useMart() returns a "mart" object and listFilters() takes a "mart" object so you pass your mart to listFilters(). Such as listFilters(ensembl). you can see which type of parameters a function takes with ?listFilters

Question 2:

@Brian Wiley Thanks! I keep running into proxy problems even if I use Sys.setenv("http_proxy" = "http://my.proxy.org:9999"). It keeps saying Ensembl site unresponsive, trying useast mirror
Error in curl::curl_fetch_memory(url, handle = handle) :

Could not resolve proxy: my.proxy.org

My response:

you should be able to just load library and use a mart. What happens if you go to a new R script and run:

```
library(biomaRt)
human.mart <- useMart(biomart = "ENSEMBL_MART_ENSEMBL",
                      dataset = "hsapiens_gene_ensembl",
                      host = "useast.ensembl.org")
```

interestingly in 4 years of R I have never used Sys.setenv()

Question 3:

Here another question Brian.... some of the filter functions are obvious but for the less obvious is there a quick way to find out about their function? How do we find a filter for a specific task? Hopefully it makes sense....

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My response:

I use grep...ALOT when looking for filters (this uses regular expressions if you want to learn more you can google regex in R). There two columns in return from listFilters(mart). They are name and description. You can search both columns, for instance:

```
filters <- listFilters(mouse.mart)
filters[grep("[Rr]ef[Ss]eq", filters$name), ]
filters[grep("[Rr]ef[Ss]eq", filters$description), ]
```

This will search the description column and return both columns (i.e. the rows since subsetting is first index [something,]) where there is either "refseq" or "RefSeq" in the name or description. You can also do it for attributes.

```
attrs <- listAttributes(mouse.mart)
attrs[grep("[Bb]and", attrs$name), ]
```

Question 4:

Hi! I keep getting this error message when I try to run getBM

```
> getBM(attributes=c("ensembl_transcript_id", "hgnc_symbol"), filters="entrezgene_id", values="6928",
mart=ensembl)
```

Error: failed to load HTTP resource

Does anyone know how to fix this issue? @Brian Wiley? Thank you in advance!!

My response:

how did you call useMart()

I think there could be issues if you use the general ensembl host because of location which is why I do "host=useast.ensembl.org"

also looks like server might be down <https://www.ensembl.org/>

F) After email NCBI support to ask about GenBank submissions, I advised students about tbl2asn for input files to sequin. Below are the response exchanges.

Student 1:

One thing that confused me for a while was that the command doesn't include a flag for specifying the feature table name. It has to have the same name as the fasta file and be in the same folder (and end with ".tbl"). I used the simple command given in this description: <https://www.ncbi.nlm.nih.gov/genbank/tbl2asn2/> which was just "tbl2asn -t template.sbt -i x.fsa" (where "x" is whatever your .fsa and .tbl files are called)

My response:

yea so its this that needs to match (the second word after "Feature" in the feature table with the first word of sequence in fasta file

```
1 >Feature Silkworm genomic_sequence
2 1 878 REFERENCE
3          PubMed 26679986
4 1 878 gene
5          gene LOC733041
6          note FK506-binding protein
7          db_xref GeneID:733041
```

```
1 >Silkworm genomic sequence
2 TATTATTCCGATTTGCATGCCGTGATGCCATAATTGCCATAGTTGCTACTTACTACTAC
3 TACCTATTAAATTACGTTTCACATAAAGAGCTATTAACCTCGTAGTTGTAATTCCCTCAATT
4 TTTAACATTATAAAACCATGGGAGTCGACGTGGAAACTATTCACTGGAAATGGTTGTA
5 GACATTAATTTCATACAAAATCCAGTCGTAATACTATCCGATTAAAAATATAATTATAGGAT
```

example:

>Feature Sc_16

with

>Sc_16 [organism=Saccharomyces cerevisiae]

ttt

The Sc_16 matches

Student 2:

@Brian Wiley I guess I will try tbl2asn because I can't quite get Sequin to work right and nobody else seems to have input on it. If you wouldn't mind detailing the process for tbl2asn, that would be awesome. Did you tbl2asn for peer-collaborating practice problem #1 for Unit 4?

My response:

sorry just seeing this now while working on portfolio, I used the tutorials at these links below:

<https://www.ncbi.nlm.nih.gov/genbank/tbl2asn2/>

more on feature table format:

<https://www.ncbi.nlm.nih.gov/genbank/tbl2asn2/#tbl>

<https://www.ncbi.nlm.nih.gov/Sequin/table.html>

G) Here is a discussion I posted about prefixes for accession ids in NCBI

Thanks for the example. The graphics format shows both transcript variant 1 and 2 and their CDS respectively. It also shows RNA alignments. One is from mRNA in this GenBank entry for each variant NM_000162.5 and NM_033507.3. There are other alignments from other studies. The 'M' in NM is an mRNA entry. 'NG' prefix like in your example is incomplete genomic region, 'NC' complete genomic region, etc. See prefixes [here](#). The NM_001354800.1 alignment is from [this entry](#) and the NM_033508.3 is from [this entry](#). Many NG and NC entries are going to have other NM entries in the graphics under alignments I assume.

[EDIT] Just found another great [resource](#) for prefixes from Sequin.

H) This discussion talks about RNA-seq software I have used, a few annotation file formats, as well as an alignment software.

Thanks for the post. I have not come across seeing WIG file with "auxillary" probability scores or GC content. I guess you may see this with really large step sizes. The only experience I have with WIG files is in this tutorial for [RSEM](#) which is a tool for RNA-seq analysis. It comes with a script to convert BAM to WIG called 'rsem-bam2wig'. I would also like to add two more format files that we will see coming up that are closer to the representation of BED files. They are GTF and GFF3. Like BED which has many [optional columns](#) they have many columns which are required, each has 9 columns. I have posted links to the most formal and extensive descriptions I have seen for these files.

GTF: <https://mblab.wustl.edu/GTF22.html>

GFF3: <https://learn.genome.org/units/ngs-analysis-formats/gff3-format/>

The Hisat2 tutorial for building index can also use GTF files to create exons and splice sites to incorporate in building index. Here is link: <http://daehwankimlab.github.io/hisat2/howto/>. It gets the GTF for human Hg38 assembly here http://ftp.ensembl.org/pub/release-84/gtf/homo_sapiens/

I) Instead of writing a post on reading for one of the modules, I decided to post an applied summary of the material that combines the use of Galaxy, UCSC Genome Browser and IGV. This is posted under Week 5 which was the most appropriate location for it.

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J) For a particular exon of a gene we were to list all the SNPs listed in the dbSNP 147 database for the hg19 assembly. A fellow student was getting way more SNPs then myself and another student.

Student 1:

For part 3, writing all the SNPs that are in the exon, does anyone have a faster way than just writing them all down manually? There are a lot of SNPs and it's going slowly, I keep losing my place and it's very tedious. I tried exporting features to a BED file, but when I do that it just has the ranges, with no names. I also tried going to Ensembl and exporting the SNPs from dbSNP in the 4th exon of the gene to an excel file. It seems like this basically matches up but I wish there was some way to verify...

My initial response to student 1:

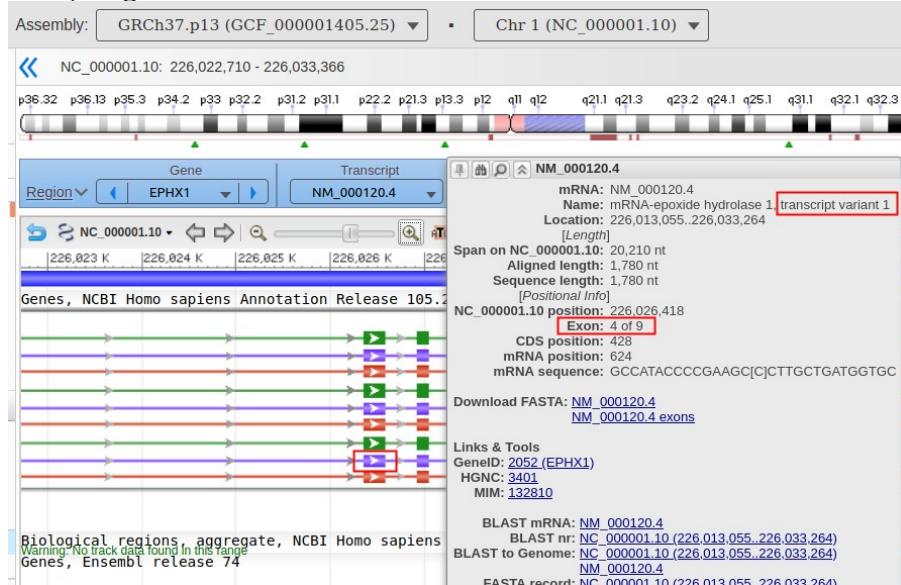
I used UCSC "All SNPs(147)" on the region for transcript variant 1 exon #4 coordinates for the hg19 assembly. Exported to table, read into R and printed with `paste(data$variant_ids, sep=",")`

Student 2:

Hey, how do you specify the exon? I just put in the location and got 204 results in the table browser

My response to student 2:

I looked at coordinates for exon #4 for the main transcript variant 1. If you zoom in close enough in NCBI Genome Viewer you can hover to see exons. Then you can either zoom in more (to the base level) to get the coordinates



Alternatively you can go to the GenBank record: NC_000001.10 (226,013,055..226,033,264) > Select full view > Look for transcript 1 > go to the 4th mRNA join and add those positions to first position of gene

mRNA

```
/un_ncbi_mim_132810
join(<1..191,3372..3559,6426..6606,13301..13528,
13864..13993,14476..14684,17013..17121,19145..19270,
19793..20210)
/gene="EPHX1"
/gene_synonym="EPHX; EPOX; HYL1; MEH"
/product="epoxide hydrolase 1, microsomal (xenobiotic),
transcript variant 1"
/note="Derived by automated computational analysis using
gene prediction method: BestRefSeq."
/transcript_id="NM\_000120.3"
/db_xref="GeneID:2052"
/db_xref="HGNC:HGNC:3401"
/db_xref="MIM:132810"
```

Student 2:

My bad, I misspoke. I got 225997776-226033264 from IGV but when I put that region into the table browser, I get way more than 44 SNPs

My response:

I see you hovered over the Gene in IGV. Try looking a little more closely and also see how many NM_** there are? I usually check the shortest NM first to confirm its transcript variant 1

Student 2:

oh wow, the collapsed view messed me up. I didn't realize the transcripts were all on top of each other and the first set of coordinates in IGV are for the whole gene. I got it now, Thank you!

K) Slack post about Variant Annotation Integrator tool at UCSC

Also check out this COOL tool from UCSC. It's under "Tools > Variant Annotation Integrator". You can search by variant ID such as rs770317327 for question 2b and you will see the predictions for SIFT, PolyPhen2 using HumVar training set and HumDiv training set. We used this for our Part 4 of project. Can you tell which training set Ensembl uses?

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Variant Annotation Integrator

Select Genome Assembly and Region

clade	genome	assembly
Mammal	Human	Dec. 2013 (GRCh38/hg38)

region to annotate

genome

Select Variants

Your session doesn't have any custom tracks or hub tracks in [pgSnp](#) or [VCF](#) format. [add pgSnp or VCF custom track](#) [add track hub](#)

variants: Variant Identifiers

Enter dbSNP rs# identifiers separated by whitespace or commas:
rs770317327

maximum number of variants to be processed: 10,000

[add custom tracks](#) [track hubs](#) To reset all user cart settings (including custom tracks), [click here](#).

```
## ENSEMBL VARIANT EFFECT PREDICTOR format (UCSC Variant Annotation Integrator)
## Output produced at 2020-10-13 03:21:33
## Connected to UCSC database hg38
## Variants: Variant Identifiers
## Transcripts: GENCODE v32 Comprehensive Transcript Set (only Basic displayed by default) (hg38.knownGene)
## dbSNP: Simple Nucleotide Polymorphisms (dbSNP 151) (/gddb/hg38/vai/snpl51.bed4.bb)
## Keys for Extra column items:
## SIFT: (http://sift.bii.a-star.edu.sg/) SIFT (D = damaging, T = tolerated)
## PP2HVAR: (http://genetics.bwh.harvard.edu/phh2/) PolyPhen-2 with HumVar training set (D = probably damaging, P = possibly damaging, B = benign)
## PP2HDIV: (http://genetics.bwh.harvard.edu/phh2/) PolyPhen-2 with HumDiv training set (D = probably damaging, P = possibly damaging, B = benign)
Uploaded Variation Location Allele Gene Feature Feature type Consequence Position in cDNA Position in CDS Position in protein Amino acid change
Codon change Co-located Variation Extra
rs770317327 chr1:225828763 A EPHX1 ENST00000445856.5 Transcript missense_variant 117 34 12 G/S Ggc/Agc rs770317327
SIFT=T(0.054,0.094,0.039,,0.094);PP2HVAR=P(0.642);PP2HDIV=P(0.951);EXON=2/4
rs770317327 chr1:225828763 A EPHX1 ENST00000272167.9 Transcript missense_variant 114 34 12 G/S Ggc/Agc rs770317327
SIFT=T(0.054,0.094,0.039,,0.094);PP2HVAR=P(0.642);PP2HDIV=P(0.951);EXON=2/9
rs770317327 chr1:225828763 A EPHX1 ENST00000448202.5 Transcript missense_variant 521 34 12 G/S Ggc/Agc rs770317327
SIFT=T(0.054,0.094,0.039,,0.094);PP2HVAR=P(0.642);PP2HDIV=P(0.951);EXON=2/4
rs770317327 chr1:225828763 A EPHX1 ENST00000614058.4 Transcript missense_variant 243 34 12 G/S Ggc/Agc rs770317327
SIFT=T(0.054,0.094,0.039,,0.094);PP2HVAR=P(0.642);PP2HDIV=P(0.951);EXON=2/9
rs770317327 chr1:225828763 A EPHX1 ENST00000366837.5 Transcript missense_variant 230 34 12 G/S Ggc/Agc rs770317327
SIFT=T(0.054,0.094,0.039,,0.094);PP2HVAR=P(0.642);PP2HDIV=P(0.951);EXON=2/9
rs770317327 chr1:225828763 A EPHX1 ENST00000467015.1 Transcript upstream_gene_variant - - - - - rs770317327 DISTANCE=80
```

L) Discussion posts about some tools in the samtools and bcftools packages of tools

Post 1:

I found 3 nice tidbits of ways to get information on a SAM/BAM file.

1. If you are interested in getting the unique RNAMEs, i.e. the unique chromosomes of the reference genome, you can use the 'awk' command in addition to samtool tools view. To get unique on a column using awk I found this post:

<https://www.commandlinefu.com/commands/view/10840/display-unique-values-of-a-column>

And so you would pipe the 'samtools view' command to only see lines without the header with something like:

```
samtools view plant.sam | awk '{ a[$3]++ } END { for (b in a) { print b } }'
```

2. How do I know my SAM/BAM file is sorted?

You may want to check if your file is sorted (this is required by most variant callers), before you sort it because this can use a lot of memory. After [samtools sort](#) is called to sort a SAM/BAM file, it will include a new line (maybe overwrite existing line) in the comments at the header. So you can call samtools view with the -H parameter to see the header.

```
samtools view -H plant.sam
```

If the file is unsorted you will see a line for example from this [post](#)

```
@HD VN:1.0 SO:unsorted
```

If it is sorted like this plant.sam file it will show

```
@HD VN:1.3 SO:coordinate
```

meaning it was sorted by coordinate.

3. Some times you have a SAM/BAM file with more reference chromosomes than you wish to analyze, i.e. you have a SAM/BAM alignment file for the entire human genome what you only want chromosome 22.

See this post: <https://carleshf87.wordpress.com/2013/10/28/extracting-reads-for-a-single-chromosome-from-bamsam-file-with-samtools/>

First you need an index file. If you don't have one you can create one with the command with [samtools index](#). However you need to convert bam to sam first and index only works on a binary compressed files.

```
samtools view -b plant.sam > plant.bam
```

```
samtools index plant.bam
```

gives you the file "plant.bam.bai". Now to extract just the 1 chromosome of *A. thal* you can call command (-h will include header so you keep this in the subset):

```
samtools view -h plant.bam Chr1 > plant.Chr1.sam
```

Then checking the new file to confirm we just call view again with the first 10 lines:

```
samtools view plant.Chr1.sam | head -n10
```

To really confirm your new sam file is a subset you can use the word count command with lines parameter to count lines:

```
samtools view plant.Chr1.sam | wc -l  
1185
```

```
samtools view plant.sam | wc -l  
104154
```

If you only want a region of the 1st chromosome, we can use a familiar region format we have seen with biomaRt, for instance just the first 1 mega base pair of chromosome 1 with:

```
samtools view -h plant.bam Chr1:1-1000000 > plant.Chr1_1mb.sam
```

Then we can see that we cut out only about 125 lines showing that about 90% of the reads from the first chromosome are in the the 1st mega base pairs:

```
samtools view plant.Chr1_1mb.sam | wc -l  
1062
```

Post 2:

FYI I just found out that you would have to pass the binary -b parameter in the subsetting view command so that you can create the index .bai file required for viewing in IGV.

Post 3:

Thanks for this post Anonomous. There is a sister package to samtools, called bcftools in which some of those tools are used in the variant calling process. These two packages in addition to htseqlib are all written by same people. The two most popular tools of bcftools for variant calling are:

bcftools mpileup

bcftools call

You usually use mpileup before call. You used to use samtools mpileup and now its under the bcftools package. See comment from bcftools website:

The **mpileup** command was transferred to bcftools in order to avoid errors resulting from use of incompatible versions of samtools and bcftools when using in the mpileup+bcftools call pipeline.

Below is example of protocol:

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Examples:

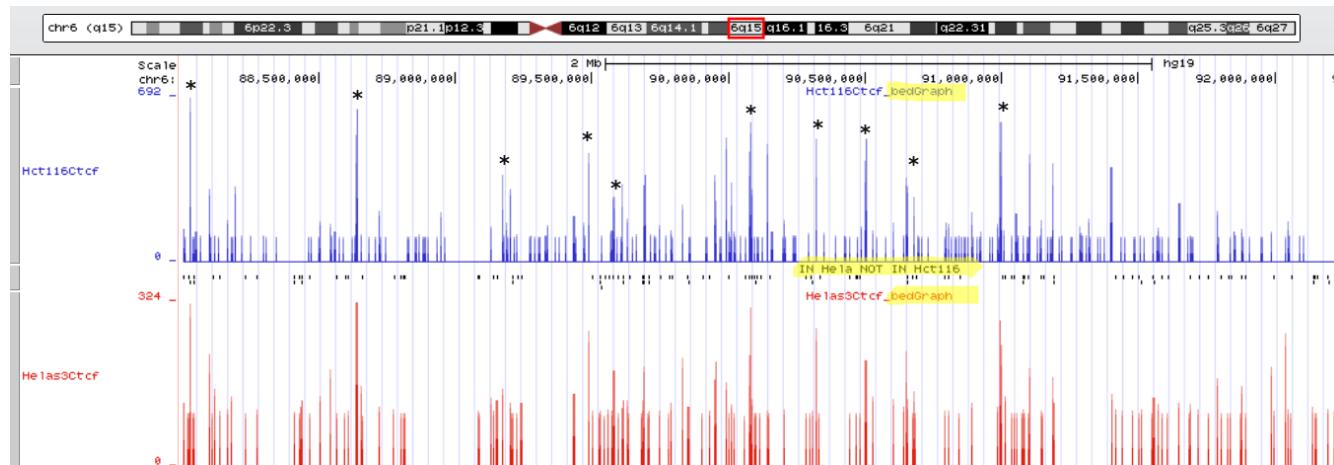
Call SNPs and short INDELS, then mark low quality sites and sites with the read depth exceeding a limit. (The read depth should be adjusted to about twice the average read depth as higher read depths usually indicate problematic regions which are often enriched for artefacts.) One may consider to add **-C50** to mpileup if mapping quality is overestimated for reads containing excessive mismatches. Applying this option usually helps for BWA-backtrack alignments, but may not other aligners.

```
bcftools mpileup -Ou -f ref.fa aln.bam | \
bcftools call -Ou -mv | \
bcftools filter -s LowQual -e '%QUAL<20 || DP>100' > var.flt.vcf
```

M) This is a Slack post I added after performing some analysis on intersecting regions between annotation files of 2 different cell lines. I wanted to let students know that sometimes you need to question the results you get and you may have to perform additional work to make more appropriate interpretations.

For part 2 of peer collab, I created a custom Python script to convert discontinuous bed files that have scores into a continuous bedGraph files so that you can view a bar graph in UCSC with the score column. Since although you may see from question 4 of part 2 there are many regions in Hela cell line not in HCT116 (there were 148 of the 211 regions in Hela cell not in HCT116 cell lines for 6q15) and think "yes CTCF binding is completely different in the cell lines because ~75% of regions of Hela are not in HCT116" but if you look at the spikes from the bedGraphs of each they really coincide. You can see all the regions in the middle track in Hela not in HCT116 and there are a lot but most of the peaks (noted with *) are very close on both tracks.

I have attached Python script if you are interested.



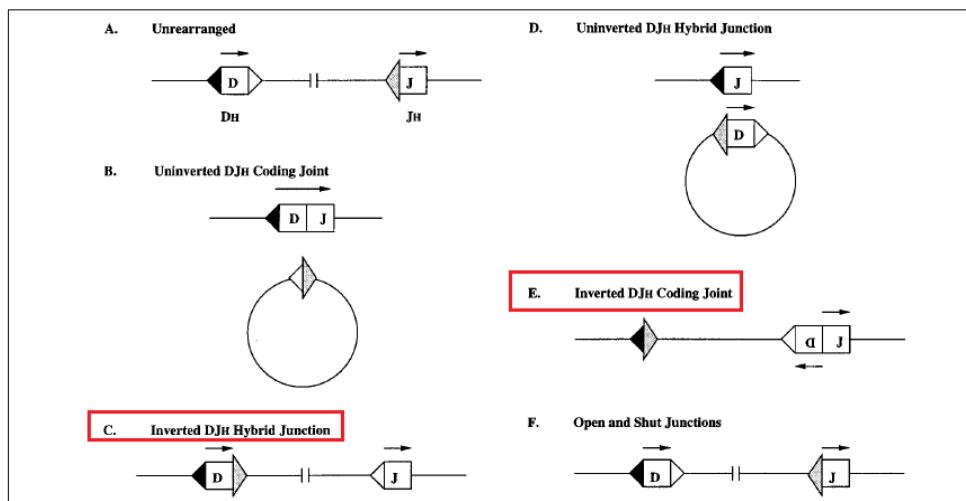
BedGraph Track Format

The bedGraph format allows display of continuous-valued data in track format. This display type is useful for probability scores and transcriptome data. This track type is similar to the wiggle (WIG) format, but unlike the wiggle format, data exported in the bedGraph format are preserved in their original state. This can be seen on export using the table browser. For more details on data compression in wiggle tracks see the notes section of the wiggle [track description page](#). If you have a very large data set and you would like to keep it on your own server, you should use the [bigWig](#) data format. In fact, an attempt to load a bedGraph custom track over 50,000,000 lines will result in an error message, but can be addressed by turning the bedGraph into a bigWig (see [Example 3](#)). Note that bedGraph files cannot easily be converted to wiggle files; converting bedGraph to bigWig and using [bigWigToWig](#) will return the original bedGraph file.

The simple script [bedToBedGraph.py](#) is only for sorted bed file with no overlapping regions where column 5 is a score column.

N) Here I add a post to discussion in which in many scenarios we associate chromosomal inversions with mutation and disease while in the immune this system this is occurring frequently and is necessary for diverse immunity.

Chromosomal inversions are not always considered to be associated with disease or mutation. In fact they provide a critical (crucial?) source of variation in the immune system during maturation of B and T cells in which BCR and TCR genes undergo V(D)J recombination. In molecular biology we learn of three types of recombination, more formally conservative site specific recombination (CSSR): 1) deletion, 2) duplication (insertion) and 3) inversion (Watson et al., 2013). A study (a LONG time ago) of VDJ recombination of the Heavy chain IgH of the BCR, probably the most studied gene in Cancer from a recombination perspective, in mouse indicated that "There are approximately 20,000 inverted DJH coding joints and inverted DJH hybrid junctions per day 16 fetal liver" (Sollbach et al., 1995). Below shows a summary described by authors research indicating two possible DJH recombination events 1) uninverted (coding joint) DJH coding points and 2) inverted DJH coding points.



- Watson, J. D., Baker, T. A., Bell, S. P., Gann, A., Levine, M., & Losick, R. M. (2013). Molecular biology of the gene. 7th Edition.
- Sollbach, A. E., & Wu, G. E. (1995). Inversions produced during V(D)J rearrangement at IgH, the immunoglobulin heavy-chain locus. *Molecular and cellular biology*, 15(2), 671–681.
<https://doi.org/10.1128/mcb.15.2.671>

O)

P)

Q)

R)

S) Here I discuss my project for epigenetics and Leukemic Stem Cells (aka Cancer Stem Cells CSCs) on a forum discussion for epigenetics.

In my Epigenetic course we gave a presentation for our final project on "Cancer Stem Cells" also known as "Leukemic Stem Cells" where we identified Epigenetic regulation in the onset of hematopoietic cancers. The influence for our project was Dr. Andrew Feinberg from Hopkins as well as Dr. Stephen Baylin from Hopkins (Papers below). The interesting finding from these researchers was titled the "Epigenetic progenitor model of Cancer" which is different from the typical viewed "clonal genetic model" where cancer is thought to derived from one ancestral cell with tumor suppressor mutation or oncogene progression. Instead in the Epigenetic progenitor model, erratic or abnormal epigenetic programming goes awry, either by means of tumor progenitor genes, chronic inflammation, aging, etc. leading to polyclonal expansion of undifferentiated stem cells.

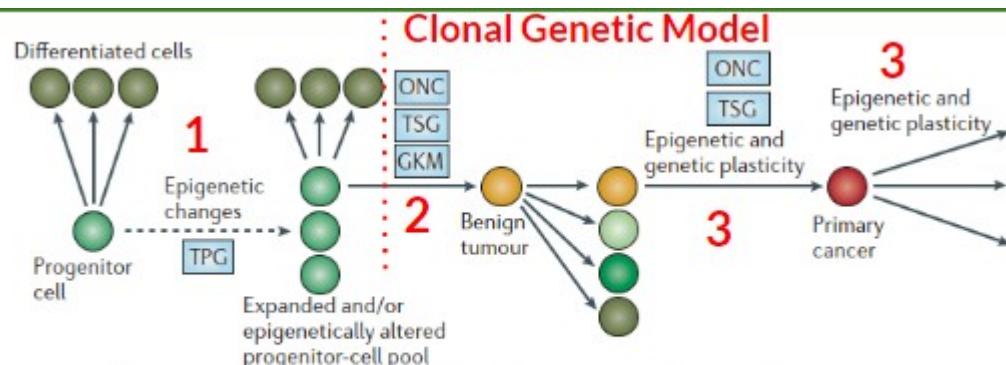


Figure 2 | The epigenetic progenitor model of cancer. According to this model, cancer arises in an epigenetic alteration of stem/progenitor cells within a given tissue, which is mediated by abe tumour-progenitor genes (TPG). This alteration can be due to events within the stem cells thems

Figure from Feinberg.

Attractive therapies, called differentiation therapies include targets in epigenetics. For example H3K79 methylation by DOT1L activates self-renewal homeobox genes HOXA9 and MEIS1. These genes are responsible for maintenance of the proper pool level of self-renewing HSCs in the bone marrow. When DOT1L is upregulated this leads to increased marks in H3K79me which positively regulates transcription of HOXA9 and MEIS1 leading to increased level of undifferentiated HSCs in multiple leukemias such as MLL-fusion type leukemias, AML and ALL. Pinonmetostat is a DOT1L Histone Lysine Methyltransferase inhibitor that is on trial. By inhibiting DOT1L methyltransferase activity on H3K79 this leads to downstream differentiation of these leukemic stem cells to eradicate the cancer.

An interesting lab and Dana Farber in Boston works with differentiation therapy specifically with HDACi (Histone deacetylase inhibitors) in neuroblastomas.

<http://stegmaierlab.dfcf.harvard.edu/research.html>

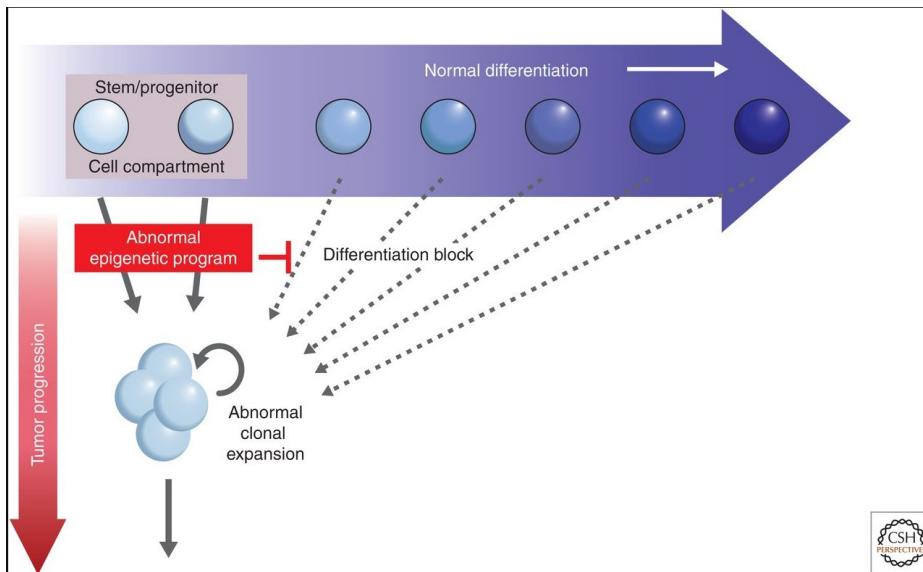


Figure from Baylin and Jones.

T) Here I look into a bug/issue on Galaxy for adding additional tracks to Trackster to an initially visualized track.

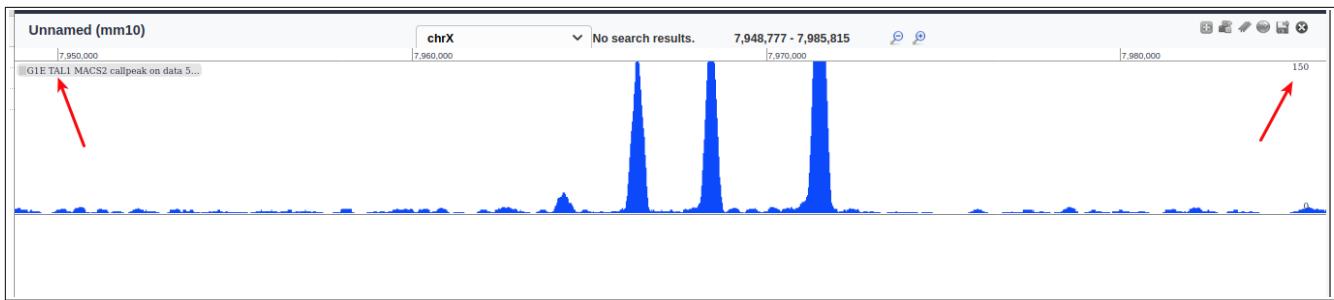
Hey All, as a reference there has been a bug in Galaxy for some time now when using the "Add tracks" function in Trackster to an initially added track. You can see the first track added in the new visualization but any track added there after will not show, basically making it unusable to visualize control vs treatment because they need to be on the same scale and not their individual scales in separate visualizations. Therefore you need to use either IGV or UCSC genome browsers or NCBI Genome Data Viewer, or other if you have found others.

See response from Jenna J, main Galaxy maintainer, in first post:

<https://help.galaxyproject.org/t/trackster-not-loading-dataset-with-server-indexed-database-assigned/2308/3>

<https://github.com/galaxyproject/usegalaxy-playbook/issues/276>

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8. Future Opportunity & Outlook

Examples job fields:

Bioinformatics

Genomics

Statistical Genetics

Computer Aided Drug Design (CADD)

Computational Protein Engineering

Biological Data Science

Examples jobs applied to:

Software Engineering Intern

Data Sciences & AI Graduate Programme

Bioinformatics Engineer

Bioinformatics Co-Op

R&D Bioinformatics Analyst

Computational & Structural Chemistry Co-Op

Intern, Computational Biology

Early Career Biological Data Liaison

Data Scientist Intern

Co-op, Computational Sciences Bioinformatics

Data Analyst Bioinformatics

Computational Biologist - Summer Intern - Single Cell Analysis

Protein Design Data Scientist Intern

Cheminformatics Intern

CADD & Molecular Analytics Intern

...

R&D Data Science Graduate Program

Short 5 year plan Option 1:

Apply for 2 year graduate programs and internships in the fields above. My goals during these 2 years are to use computational tools and algorithms to model small molecules and protein drugs as therapies for blood cancers. I hope to narrow down a specific type of hematological cancer throughout this time. I completed a project on Leukemia stems cells in my Epigenetics course on the role of differentiation in ALL and AML which was really exciting however I am also interested in immunotherapies targeting receptors in aggressive B-cell lymphomas and B-cell acute lymphoblastic leukemia.

After 2 years apply to roles in Level 2 and 3 Bioinformatician and Computed Aided-Drug Design to define molecular targets in pediatric leukemia.

Short 5 year plan Option 2:

Applying to 5 PhD programs and 1 Master's programs below in Bioinformatics and Medicinal Chemistry to continue my education in Cancer Bioinformatics and Computed Aided-Drug Design for targets in pediatric leukemia.

University of California San Francisco – [PhD Bioinformatics](#)

European Bioinformatics Institute (EMBL-EBI) – [PhD Bioinformatics](#), [Cortes-ciriano group](#)

Harvard University – [BIG PhD Bioinformatics](#)

University of Michigan – [Medicinal Chemistry](#) (Bioinformatics Tab)

University of Illinois at Chicago – [Pharmaceutical Science](#) (Chemistry in Drug Discovery Concentration)

Stevens Institute of Technology - [Computational and Medicinal Chemistry](#)