Biological Research For Nursing http://brn.sagepub.com/

Potential Epigenetic Mechanism(s) Associated With the Persistence of Psychoneurological Symptoms in

Women Receiving Chemotherapy for Breast Cancer: A Hypothesis

Debra Lyon, Lynne Elmore, Noran Aboalela, Jacqueline Merrill-Schools, Nancy McCain, Angela Starkweather, R. K. Elswick, Jr. and Colleen Jackson-Cook

Biol Res Nurs published online 11 April 2013 DOI: 10.1177/1099800413483545

The online version of this article can be found at: http://brn.sagepub.com/content/early/2013/04/09/1099800413483545

Published by:

\$SAGE

http://www.sagepublications.com

Additional services and information for Biological Research For Nursing can be found at:

Email Alerts: http://brn.sagepub.com/cgi/alerts

Subscriptions: http://brn.sagepub.com/subscriptions

Reprints: http://www.sagepub.com/journalsReprints.nav

Permissions: http://www.sagepub.com/journalsPermissions.nav

>> OnlineFirst Version of Record - Apr 11, 2013

What is This?

Potential Epigenetic Mechanism(s) Associated With the Persistence of Psychoneurological Symptoms in Women Receiving Chemotherapy for Breast Cancer: A Hypothesis

Biological Research for Nursing 00(0) 1-15 © The Author(s) 2013 Reprints and permission: sagepub.com/journalsPermissions.nav DOI: 10.1177/1099800413483545 brn.sagepub.com



Debra Lyon, RN, PhD, FAAN^{1,5}, Lynne Elmore, PhD^{2,5}, Noran Aboalela, MS³, Jacqueline Merrill-Schools, PhD^{2,5}, Nancy McCain, RN, DSN, FAAN⁴, Angela Starkweather, RN, PhD⁴, R. K. Elswick, Jr., PhD^{1,6}, and Colleen Jackson-Cook, PhD, FACMG^{2,3,5}

Abstract

Due to recent treatment advances, there have been improvements in the proportion of women surviving a diagnosis of breast cancer (BC). However, many of these survivors report persistent adverse side effects following treatment, such as cognitive dysfunction, depressive symptoms, anxiety, fatigue, sleep disturbances, and pain. Investigators have examined circulating levels of inflammatory markers, particularly serum cytokines, for a potential causal relationship to the development/persistence of these psychoneurological symptoms (PNS). While inflammatory activation, resulting from perceived stress or other factors, may directly contribute to the development of PNS, we offer an alternative hypothesis, suggesting that these symptoms are an early step in a cascade of biological changes leading to epigenetic alterations at the level of deoxyribonucleic acid (DNA) methylation, histone modifications, and/or chromatin structure/chromosomal instability. Given that epigenetic patterns have plasticity, if this conjectured relationship between epigenomic/acquired genomic alterations and the development/persistence of PNS is confirmed, it could provide foundational knowledge for future research leading to the recognition of predictive markers and/or treatments to alleviate PNS in women with BC. In this article, we discuss an evolving theory of the biological basis of PNS, integrating knowledge related to inflammation and DNA repair in the context of genetic and epigenetic science to expand the paradigm for understanding symptom acquisition/persistence following chemotherapy.

Keywords

epigenetic, cognitive, methylation

In 2012, a total of 226,870 women were expected to receive a diagnosis of breast cancer (BC) in the United States (Siegel, Naishadham, & Jemal, 2012). Most of these women will have been diagnosed in the early stages of the disease (Stages I and II) and 90% can expect to survive at least 5 years due to improvements in adjuvant chemotherapy and targeted hormonal therapies. However, these treatments and, perhaps, the cancer, itself, contribute to a number of distressing short-term and long-term life-altering and debilitating side effects that may persist in some women long after active treatment has ended. In particular, the administration of adjuvant chemotherapy is frequently associated with multiple co-occurring distressing symptoms (Dodd, Cho, Cooper, & Miaskowski, 2010; Goedendorp, Gielissen, Verhagen, Peters, & Bleijenberg, 2008). Research has linked depressive symptoms, anxiety (Badger, Segrin, Dorros, Meek, & Lopez, 2007), fatigue (Berger, Wielgus, Hertzog, Fischer, & Farr, 2010), sleep disturbances (Lee,

Corresponding Author:

Debra Lyon, RN, PhD, FAAN, Virginia Commonwealth University, 1100 Leigh Street, Richmond, VA 23220, USA.

Email: delyon@vcu.edu

¹ Department of Family and Community Health Nursing, Virginia Commonwealth University School of Nursing, Richmond, VA, USA

²Department of Pathology, Virginia Commonwealth University School of Medicine, Richmond, VA, USA

³ Department of Human and Molecular Genetics, Virginia Commonwealth University School of Medicine, Richmond, VA, USA

⁴Department of Adult Health and Nursing Systems, Virginia Commonwealth University School of Nursing, Richmond, VA, USA

⁵ Massey Cancer Center, Virginia Commonwealth University, Richmond, VA,

⁶ Department of Biostatistics, School of Medicine, Virginia Commonwealth University School of Medicine, Richmond, VA, USA

Cho, Miaskowski, & Dodd, 2004), and pain (Utne, Miaskowski, Bjordal, Paul, & Rustoen, 2010; Valeberg et al., 2008) as a symptom cluster prominent across cancer types and stages. BC patients and survivors have also frequently noted the symptom of cognitive dysfunction, commonly referred to as chemobrain, during active treatment and, for some, after the completion of treatment as a seemingly long-lasting phenomenon (reviewed in Wefel, Vardy, Ahles, & Shagan, 2011). Cognitive dysfunction in cancer patients has, in fact, been a topic of intense research (Vardy, Wefel, Ahles, Tannock, & Schagen, 2008). Collectively, these symptoms can be described as psychoneurological symptoms (PNS) and often result in a significant decline in quality of life by contributing to adverse health outcomes over the active treatment period and into survivorship. PNS are associated with decreased functional status and work limitations (Hansen, Feuerstein, Calvio, & Olsen, 2008) as well as unemployment and early retirement (Mehnert, 2011).

With this growing awareness that cancer survivors may develop both short-term and long-term effects from cancer and its treatments, it is important that we increase our understanding of the molecular mechanisms leading to these side effects. In this article, we present an evolving theory of the biological basis of PNS, suggesting that their development and persistence may reflect a cascade of cellular and molecular events. We offer evidence supporting this hypothesis along with a discussion of how the knowledge of acquired epigenetic perturbation(s), if present, could be exploited to develop future, practical biomarkers for identifying women who are at an elevated risk of developing these potentially debilitating treatment-associated symptoms, with the ultimate goal of designing therapies to alleviate these symptoms.

PNS

The interrelationship of PNS in patients with cancer has led researchers to question whether or not at least some of these symptoms might share a common biological etiology and whether they could be collectively treated (Cleeland et al., 2003). The theoretical guidance from the symptom cluster literature presupposes that commonly occurring symptoms in patients with cancer share a common biological mediator. Inflammation, along with its systemic effects, is a likely candidate for a shared mechanism of not only the development, initiation, and progression of cancer (Mantovani, Allavena, Sica, & Balkwill, 2008) but also the commonly experienced symptoms in individuals with cancer. Epidemiological evidence points to a connection between inflammation and a predisposition for the development of cancer (Coussens & Werb, 2002; Mantovani et al., 2008). In addition, inflammatory processes are central to the pathogenesis of depression (Dantzer, O'Connor, Freund, Johnson, & Kelley, 2008) and other symptoms in the cluster including pain, fatigue, sleep disturbances (Raison, Capuron, & Miller, 2006), and cognitive dysfunction (Roberts et al., 2010).

Support for the premise that inflammatory activation is an early or initiating step in the cascade of events leading to PNS comes from investigations showing cytokine-induced sickness behaviors in animal models and select human studies (Raison

et al., 2006). The phrase sickness behaviors describes the subjective complaints of patients that may accompany a variety of conditions associated with inflammatory response, such as infectious diseases and cancers. Investigators have suggested that these behaviors (such as lethargy, depression, and fatigue) are actually caused by perturbations in the levels of cytokines acting in the brain (Bower, Ganz, Aziz, & Fahey, 2002; Dantzer et al., 2008; Kelley et al., 2003). The sickness behavior model is relevant to cancer in that chemotherapy has been associated with higher levels of interleukins in both humans and animal models, and researchers have noted cytokine elevations in cancer survivors with severe and persistent fatigue after treatment (Bower et al., 2002). Collectively, these findings indicate that the dysregulation of cytokines may be related to PNS in cancer patients and survivors before, during, and after active treatment. However, these studies, which have generally limited their foci to single symptoms and single markers of inflammation, have not resulted in a clear agreement regarding biological mechanisms of PNS, nor have they led to the development of effective therapeutic interventions.

Although a biological model that includes inflammation as an initiating step leading to PNS has partial empirical support, the evidence, to date, is not sufficient to conclude that peripheral inflammatory mechanisms are a complete or sufficient mechanism for treatment-related symptoms or persistent symptoms in survivors. Since most inflammatory molecules, such as cytokines, are relatively short lived, the mechanism(s) for how these changes could lead to long-term symptoms that persist and are embedded in an individual's biological memory beyond the time of chemotherapy treatment has been enigmatic (Esteller, 2008). One possible means for "biologically remembering" the effects of BC and/or its treatments would be deoxyribonucleic acid (DNA)-based changes in the individual's somatic (nonreproductive) cells. These acquired changes could result from either epigenomic modifications (which encompass alterations in DNA methylation patterns, modifications of histone proteins, and/or changes in chromatin structure; Feinberg et al., 2010) or genomic changes (which include, but are not limited to, telomere attrition, and acquired chromosomal instability).

Evidence for Epigenetic Alterations in BC and/or Its Treatment

Researchers have conjectured that epigenetic changes are highly relevant to the development of chronic health problems because they account for interactive relationships among environment, genetic background, and disease (Ptak & Petronis, 2008). An epigenetic alteration is defined as one "resulting from changes in a chromosome without alterations in the DNA sequence" (Collado-Hidalgo, Bower, Ganz, Irwin, & Cole, 2008). Thus, epigenetic modifications cause a change in gene activity without altering the underlying DNA sequence. Simplistically, one can liken the activity of the human genome to the wiring of a house (Figure 1). Appliances may be turned on or off in response to an environmental need (e.g., darkness

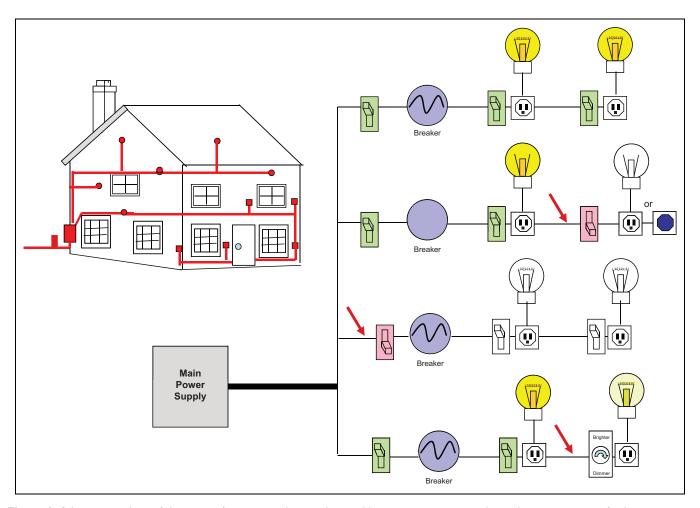


Figure 1. Schematic analogy of the types of epigenetic changes that could occur in response to chemotherapy treatment for breast cancer. Simplistically, one can liken the activity of the genome to the wiring of a house (top left diagram). Although wiring is available to all outlets for appliance/lights usage in a home, one typically turns them on or off in response to need, with the exception of some "housekeeping" appliances (i.e., refrigerator, etc.) that are continuously active. In the schematic representation shown on the right, the typical scenario may be to have activity for multiple regions or genes (Row 1; illustrated by both lights being on). Changes in functionality could impact a single gene (Row 2) or multiple genes in a network (e.g., if a breaker flips as shown in Row 3). These alterations can be complete (on or off) or they can be partial (i.e., dimmer switch as shown in the bottom row).

triggers the need for lighting). Similarly, genes may be turned on and off, via epigenetic modifications, in response to an environmental or exposure-related trigger, with this stimulus being termed an *epigenator*. Continuing with our analogy, to correct the need for light, one must turn on a lamp in a particular location (e.g., a lamp in a downstairs kitchen will not meet the need for light in an upstairs bedroom). Although wiring is available to all outlets for appliance/lights usage in a home, one typically turns them on or off in response to need, with the exception of some "housekeeping" appliances (e.g., refrigerator, etc.) that are continuously active. In the schematic representation (on the right of Figure 1), the typical scenario may be to have activity for multiple regions or genes (Row 1; illustrated by both lights being on). Changes in functionality could impact a single gene (Row 2), or multiple genes in a network (e.g., if a breaker flips as shown in Row 3). These alterations can be complete (on or off) or they can be partial (i.e., dimmer switch as shown in the schematic representation, bottom row). The biological change

involved in identifying the specific location for implementing epigenetic changes, termed the *initiator* event, is based on the underlying DNA sequences of the chromatin to be altered. Finally, the resulting response to the environmental stimulus (in our example, keeping the light turned on) is accomplished through the action of the *maintainer*, which for epigenetics is a persistent biological change that sustains an altered chromatin state (Collado-Hidalgo et al., 2008). Given that the maintainers are the persistent, or long-term, biological marks that sustain epigenetic alterations, most biological assays rely on the assessment of these markers to detect the presence of epigenetic changes (Gupta, Nagarajan, & Wajapeyee, 2010).

In Table 1, we summarize the results of studies in which investigators have identified perturbations in a variety of epigenetic markers related to BC and PNS for the following types of epigenetic/genetic alterations: (1) DNA methylation and hydroxymethylation, (2) histone modifications, and (3) chromatin structural alterations as well as chromosomal instability.

Table I. Summary of Genomic and/or Epigenomic Changes Associated with Breast Cancer, Chemotherapy, and/or Psychoneurological

Symptoms.	
Finding	References ^a
1. BC has been associated with the acquisition of DNA methylation	changes
DNA methylation	D
Altered epigenetic patterns have been observed in BC tumors and allow for recognition of distinct prognostic groups	Dedeurwaerder et al. (2011 ^b), Fang et al. (2011), Huang, Nayak, Jankowitz, Davidson, and Oesterreich (2011 ^b), Jovanovic et al. (2010 ^b), Parrella (2010 ^b), Veeck and Esteller (2010 ^b)
DNA methylation profiles have allowed for the recognition of an immune component in a subset of breast tumors	Dedeurwaerder et al. (2011)
BRCA1 methylation alterations have been seen in a portion of sporadic cases of BC tumors but have not been consistently observed	Bosveil et al. (2012), Butcher and Rodenhiser (2007)
Focused BRCA1 promoter methylation status in blood has been used to attempt to predict a woman's risk of developing BC, with varying results	Al-Moghrabi et al. (2011), Iwamoto et al. (2011)
Genome-wide DNA methylation profiles in blood have been associated with risk of developing BC	Terry et al. (2011)
Epigenetic changes have been implicated as a factor in disparities for cancer risk associated with socioeconomic status	Mohammed et al. (2012)
Stress	
Environmentally induced oxidative stress may induce changes in the 5-hydroxymethylation status of DNA	Chia et al. (2011)
Epigenetic alterations have been observed in response to a variety of stress situations and have been associated with psychiatric conditions as well as the function of neural and other tissues	Mathews et al. (2011), Mehler (2008 ^b), Schmidt et al. (2011), Toyokawa et al. (2012)
2 PC and/an the effects of showerth arrays are consisted with the co	anisisian of history and difference
2. BC and/or the effects of chemotherapy are associated with the ac EZH2 levels are elevated in BC tumors and it is thought that this histone methyltransferase may be an important early step in initiating a cascade of biological changes in cancer cells, including epigenetic alterations	Bachmann et al. (2006), Collett et al. (2006), Ding et al. (2006), Gonzalez et al. (2009), Grzenda, Ordog, and Urrutia (2011 ^b), Kleer et al. (2003), Raaphorst et al. (2003), Rush et al. (2009), Schlesinger et al. (2006), Simon and Lange (2008 ^b), Viré et al. (2006), Tsang and Cheng (2011 ^b)
EZH2 is frequently elevated in inflammatory BC and is predictive of a worse clinical outcome	Gong et al. (2011)
EZH2 is thought to mediate repression of DNA repair Reductions in histone (H3 and H4) acetylation lead to significant decreases in brain-derived neurotrophic factor gene expression and perturbations in hippocampus signaling/presence of dendritic spine	Chang et al. (2011), Stefansson and Esteller (2011) Zeng et al. (2011)
Chemotherapy agents adversely affect neural progenitor cell proliferation and are associated with epigenetic (histone) modifications in the hippocampus and prefrontal cortex of the brains of rats	Briones and Woods (2011)
3. BC, stress, or direct effects from chemotherapy lead to the acquirent Telomeres BC	sition of telomere alterations and/or chromosomal instability
Chromosomes from epithelial breast cells having shorter telomeres had higher frequencies of chromosomal instability	Diehl et al. (2011), Pampalona, Soler, Genescà, and Tusell (2010)
The value of lymphocyte telomere length as a biomarker for predicting BC risk is controversial Stress and inflammation	De Vivo et al. (2009), Gramatges et al. (2010), Kim et al. (2011), Pooley et al. (2010), Shen et al. (2007), Zeng et al. (2011)
Higher anticipatory threat appraisals have been associated with shortened telomere length	O'Donovan et al. (2012)
Cumulative inflammatory response and/or chronic pain/stress have been associated with telomere attrition Progressive telomere shortening has been observed in	Epel (2009 ^b), Epel et al. (2004), O'Donovan et al. (2011, 2012), Sibille et al. (2012), Strub et al. (2008) Flanary and Streit (2004)

(continued)

microglial rat cells, which are mitotically active

Table I. (continued)

Finding	References ^a
Interventions to reduce stress have been associated with increased telomere lengths and/or less attrition Chemotherapy and other treatments	Biegler, Anderson, Wenzel, Osann, and Nelson (2012), Peres (2011), Puterman et al. (2010)
Several chemotherapeutic agents lead to shortening of telomeres through a variety of mechanisms	Hewitt et al. (2012), McCormick and Silva (2006), Walker et al. (2012 ^b)
Changes in leukocyte telomere lengths have been observed in posttreatment specimens compared to pretreatment specimens in women with or without stem cell transplants	Schroeder et al. (2001)
Chromosomes from mesenchymal stem cells showed shortened telomeres and other attributes of "aging"	Buttiglieri et al. (2011)
following exposure to chemotherapeutic agents Related conditions	
Memory loss and compromised cognitive function have been associated with leukocyte telomere attrition for several conditions	Martin-Ruiz et al. (2006), Valdes et al. (2010), Yaffe et al. (2011)
Telomere attrition has been observed in people having depression, mood disorders, cardiovascular disease, atherosclerosis, diabetes, dementia, Alzheimer's disease, primary caregiver status to chronically ill family members, social stress, and numerous other health conditions Chromosomal instability (micronuclei)	Aviv and Aviv (1998), Cherkas et al. (2006), Damjanovic et al. (2007), Effros (2011), Epel (2009 ^b), Fitzpatrick et al. (2007), Humphreys et al. (2012), Kume et al. (2012), Parks et al. (2009), Serrano and Andres (2004 ^b), Simon et al. (2006), Wikgren et al. (2012)
BC	
Infiltrating ductal carcinoma tumor specimens have an increased frequency of micronuclei, which is correlated with tumor grade	Goel, Bhatia, and Dey (2011), Samanta, Dey, and Nijhawan (2011)
Increased frequency of micronuclei and other measures of chromosomal instability have been seen in lymphocytes of women with BC compared to controls, but results have varied between studies	Aristei et al. (2009), Cardinale, Bruzzi, and Bolognesi (2012 ^b), Djordjevic et al. (2010), Elsendoorn et al. (2001), Miloševic-Santos et al. (2010), Varga et al. (2006), Wang et al. (2006)
Folate levels appear to influence lymphocyte micronuclei values in women with and without BC	Wang et al. (2006)
Chemotherapy Increased frequency of micronuclei and other measures of chromosomal instability in lymphocytes have been found following therapy compared to baseline values in women	Aristei et al. (2009), Elsendoorn et al. (2001), Miloševic-Djordjevic et al. (2010)
with BC, with individual differences being noted Effect(s) of different therapy regimens on chromosomal instability or breakage varies between studies	Aristei et al. (2009), Miloševic-Djordjevic et al. (2010)
Chemotherapy-induced increases in chromosomal abnorm alities in lymphocytes persist for months after treatment	Bilban-Jakopin and Bilban (2001), Elsendoorn et al. (2001)
Age-related changes and complex diseases	
Acquired chromosomal aberrations have been observed to occur in an increased frequency in patients having Alzheimer's disease, Parkinson's disease, schizophrenia, heart disease, autoimmune conditions, atherosclerosis, and numerous other conditions	Aviv and Aviv (1998 ^b), lourov et al. (2008b ^b), Jackson-Cook (2011 ^b) Wojda and Witt (2003 ^b), Yurov et al. (2008)
Somatic mosaicism for DNA copy number alterations and loss of heterozygosity has been associated with an increased risk of developing cancer	Jacobs et al. (2013)

Note. BC = breast cancer; BRCAI = breast cancer susceptibility gene one; DNA = deoxyribonucleic acid; EZH2 = enhancer of zeste 2.

DNA Methylation and Hydroxymethylation Alterations

DNA methylation occurs when a methyl group $(-CH_3)$ is added to one of the four DNA bases (cytosines) in a DNA molecule

(Jones & Liang, 2009; Miranda & Jones, 2007). This process, which is catalyzed by enzymes called DNA methyltransferases (Carey, Marques, & Reik, 2011), usually results in the repression of gene transcription (turning genes "off"; Mazzio & Soliman, 2012). Hydroxymethylation is an alternative

^aThe references cited are representative of published studies in the field, but the list is not exhaustive.

^bReview article.

modification that can be found on cytosine bases (Kriaucionis & Heintz, 2009; Tahiliani et al., 2009). Research has not yet fully elucidated the biological role of hydroxymethylation, but investigators have hypothesized that it is associated with gene activity (rather than inactivity, as with methylation) and have observed the process in several different tissues, including neural (brain) tissue (Jin, Wu, Li, & Pfeifer, 2011). Methylation changes have been consistently observed in cells from BC tumors as well as in BC cell lines (Table 1). Overall, BC tumor cells have shown significantly increased levels of global DNA hypomethylation (leading to gene activation) as well as hypermethylation in targeted regions of DNA (leading to gene inactivation/repression) when compared to normal tissues (Wilson, Power, & Molloy, 2007). These genome-wide methylation patterns, or "signatures," have allowed for improvements in pathologists' ability to stratify patients by disease progression risk and have led to the recognition of an immune component to the BC tumors in a subset of patients (Dedeurwaerder et al., 2011). However, researchers have noted heterogeneity (between-patient differences) in the utility of single-gene methylation patterns, such as those seen for BC susceptibility gene one (BRCA1), in evaluating BC tumors (Bosveil et al., 2012; Butcher & Rodenhiser, 2007).

In addition to better characterizing BC tumors, epigeneticists have explored the potential for using lymphocyte epigenetic signatures to identify a woman's risk of developing BC a priori. The results of these projections have yielded varied outcomes, with some groups reporting significant odds ratios, while others (primarily based on assessments of just one gene) have found them to have no clear predictive value (Al-Moghrabi, Al-Qasem, & Aboussekhra, 2011; Iwamoto, Yamamoto, Taguchi, Tamaki, & Noguchi, 2011; Terry, Delgado-Cruzata, Vin-Raviv, Wu, & Santella, 2011). Excitingly, recent reports of associations between genome-wide lymphocyte epigenetic patterns and socioeconomic status and/or environmental exposures have shown promise for explaining the well-recognized, but poorly understood, disparity in adult cancer risk related to low socioeconomic status in childhood, even when that status is reversed in adulthood (Mohammed, Springfield, & Das, 2012).

In women with BC, stress associated with the diagnosis of cancer and the ensuing lifestyle disruptions and existential threats may further compound the biological disruptions related to cancer by triggering endocrine stress-response mechanisms through the hypothalamic-pituitary-adrenal axis and the autonomic nervous system (Sephton & Spiegel, 2003). Stress-related inflammation also elicits perturbations in the regulation/expression of several genes, including (but not limited to) those regulating nuclear factor-κB, interleukin-1β, interleukin-6, and tumor necrosis factor-α (Bartsch & Nair, 2006; Miller, Chen, & Parker, 2011). Further, stress may also lead to the disruption of neuroendocrine and immune circadian rhythms (Sephton & Spiegel, 2003), causing disruptions in sleep and metabolic and endocrine functions (Dedert et al., 2012). Researchers have also consistently observed that individuals experiencing stress acquire alterations in the methylation patterns of their lymphocytes (Mathews et al., 2011; Mehler, 2008; Toyokawa, Uddin, Koenen, & Galea, 2012). In summary, the pioneering studies investigating methylation patterns in BC have shown that changes are clearly present in BC tumors, with the results of recent genome-wide studies suggesting that methylation alterations are also acquired in lymphocytes in response to environmental exposures. Furthermore, studies have shown that these perturbations in methylation patterns persist for years (Toyokawa et al., 2012).

Histone Modifications

In vertebrates, the packaging of DNA into chromatin is dependent on four core histone proteins (called Histone 2A, Histone 2B, Histone 3, and Histone 4). These histones and DNA form a complex that provides the basic organizational structure for chromatin (Kouzarides, 2007). The "tail" portion of these histone proteins contains numerous amino acid residues that can be modified, with the location and types of modifications influencing how tightly the DNA is compacted, as well as serving to recruit other proteins that are essential for mediating the methylation of DNA (Arrowsmith, Bountra, Fish, Lee, & Schapira, 2012; Greer & Shi, 2012). Methylation is also one type of modification that can occur in histones. Researchers have hypothesized that the protein enhancer of zeste 2 (EZH2) is involved in the early "decision" steps leading to histone methylation, chromatin condensation, and gene silencing (Schlesinger et al., 2006; Viré et al., 2006). Although research has not fully resolved (Rush et al., 2009), the specific role EZH2 plays in initiating histone methylation, and perhaps DNA methylation as well (Viré et al., 2006), findings have shown that the expression of EZH2 is significantly altered (often overexpressed) in BC tumors, allowing the protein to serve as a prognostic indicator for BC tumor cell progression (Bachmann et al., 2006; Collett et al., 2006; Ding, Erdmann, Chinnaiyan, Merajver, & Kleer, 2006; Gong et al., 2011; Kleer et al., 2003; Raaphorst et al., 2003; Viré et al., 2006). Researchers have also noted that EZH2 silences the expression of several DNA repair genes. Its amplification/overexpression in tumor cells, then, may impair these cells' ability to repair DNA, thus predisposing them to metastasis (Chang et al., 2011; Stefansson & Esteller, 2011). As we noted above for DNA methylation, studies of histone modifications associated with BC in humans have primarily focused on tumor tissue. However, investigators have also completed studies of somatic cells, with Briones and Woods (2011) showing that histone alterations were induced in the brain cells of rats following exposure to chemotherapeutic agents. Moreover, brain cells in rats have consistently shown perturbations of gene expression, along with changes in brain cell function and structure, following the acquisition of histone modifications.

Chromatin Structural Alterations and Chromosomal Instability

A third type of epigenetic modification involves alterations in chromatin structure. Active genes tend to be localized to loosely compacted DNA (termed euchromatin), whereas

inactive genes tend to be found in tightly condensed DNA (called heterochromatin; Quivy et al., 2004). One chromatin structure that has been implicated in BC is the telomere or the end of the chromosome. Telomeres are normally heterochromatic and are interesting ribonuclear structures in that they are epigenetically regulated (through histone modifications). They can also influence gene expression via epigenetic mechanisms. In humans, telomeres consist of tandem copies of a hexameric sequence (TTAGGG) bound by numerous associated proteins, which form a protective cap-like structure important for ensuring chromosomal stability and regulating how many times a cell can divide (Jovanovic, Rønneberg, Tost, & Kristensen, 2010; O'Sullivan & Karlseder, 2010). Telomeres can shorten, which is a genetic change rather than an epigenetic alteration since the underlying DNA is altered. Alternatively, the protective telomeric structure or cap may become compromised, which is an epigenetic change since the underlying DNA is not altered. If either of these telomeric alterations occurs, the neighboring chromatin structure becomes more open or "relaxed," thereby allowing for transcription of genes adjacent to subtelomeric regions (an epigenetic change). Multiple investigators have observed telomere shortening in BC tumors (Diehl et al., 2011) as well during the early stages of mammary carcinogenesis (Meeker & Argani, 2004). Furthermore, data are accumulating to indicate that several exposure factors can result in telomere shortening in normal body cells. These exposures include (but are not limited to) stress, mainstay chemotherapeutic agents used in the treatment of BC, and inflammation (Epel, 2009; Epel et al., 2004; Hewitt et al., 2012; McCormick & Silva, 2006; O'Donovan et al., 2011, 2012; Sibille et al., 2012; Strub, Depcrynski, Elmore, & Holt, 2008; Walker et al., 2012). Research has also revealed associations between telomere shortening and many age-related health conditions (Aviv & Aviv, 1998; Humphreys et al., 2012; Kume et al., 2012; Martin-Ruiz et al., 2006; Simon et al., 2006; Valdes et al., 2010; Wikgren et al., 2012). However, the potential utility of the telomere as a biomarker for recognizing women at risk of developing BC is controversial, with some investigators identifying a clear association between telomere attrition and risk of BC, and others finding no relationship (De Vivo et al., 2009; Gramatges, Telli, Balise, & Ford, 2010; Kim et al., 2011; Pooley et al., 2010; Shen et al., 2007; Zheng et al., 2010).

One consequence of telomere shortening is chromosomal instability, which is a genetic change. Chromosomal aberrations, which can be identified using both conventional metaphase chromosome analysis and the micronucleus assay (which is a high throughput means for accurately quantifying chromosomal instability) are hallmark findings in many tumors and are not limited to BC (Janssen & Medema, 2012). In addition to tumor-specific chromosomal abnormalities, investigators have shown a significantly increased frequency of acquired chromosomal anomalies present in a variety of body cells from individuals with cancer. Researchers completed most of these assessments using noninvasively obtained tissues such as lymphocytes and buccal mucosa (Aristei et al., 2009; Cardinale, Bruzzi, & Bolognesi, 2012; Elsendoorn et al., 2001; Miloševic-Djordjevic, Grujicic,

Vaskovic, & Marinkovic, 2010; Santos et al., 2010; Varga et al., 2006; Wang et al., 2006). The frequency of these acquired chromosomal aberrations increases in response to chemotherapy (Aviv & Aviv, 1998; Iourov, Vorsanova, & Yurov, 2008b; Jackson-Cook, 2011; Wojda & Witt, 2003; Yurov et al., 2008). Though research has not yet fully determined the causes for these acquired somatic cell chromosomal abnormalities, age, telomere attrition, genetic makeup, diet, and environmental exposures can all be contributing factors (Jackson-Cook, 2011; Kirsch-Volders et al., 2011). Investigators have identified an increased frequency of acquired chromosomal abnormalities in a wide range of complex health problems in addition to cancer, leading to speculation that the abnormalities play a contributory/causal role in the development of these conditions (Table 1).

Hypothesized Model for a Biological Cascade Involving Inflammation, Epigenetic, and/or Genetic Changes in the Development and/or Persistence of PNS

We have integrated the existing knowledge about cancer and epigenetics to develop a hypothesis to explain the development and persistence of PNS following cancer treatment (Figure 2). We posit that the chemotherapy treatment (and possibly the cancer itself) initiates a cascade of biological changes that include inflammation and epigenetic and genetic alterations. Specifically, we propose that epigenetic alterations arise either in response to inflammatory activation or as a direct consequence of chemotherapy. The induced inflammatory activation/epigenetic alterations may lead directly to cellular changes but can also result in telomere attrition, with potential gene-byenvironment interactions influencing an individual's response. The proposed epigenetic changes may involve DNA methylation or histone modifications, possibly through perturbations in EZH2. Since EZH2 is important for DNA repair, alterations in its expression could lead to chromosomal instability. Similarly, epigenetic alterations and telomere shortening could lead to chromosomal instability. Each of these factors, acting either singly or in concert, could lead to cellular changes or may act synergistically.

The types of cellular responses arising as a consequence of these epigenetic and/or genetic changes could include cell death/cellular senescence, as well as true (due to aneuploidy) or functional (via gene expression changes) genetic imbalances. Each of these alterations would be expected to interfere with the cell's metabolic activities and could contribute to the development of PNS. For example, if true or functional genetic imbalances occur in neural tissue, the cells may no longer be capable of completing normal responses, thereby blocking the transmission of intercellular signals necessary for normal neural function. Support for this conjecture comes from the observation that enzymes involved in histone methylation (gene expression) are important components in learning and memory processes (Day & Sweatt, 2011; Lubin, Gupta, Parrish, Grissom, & Davis, 2011). Furthermore, researchers have shown that individuals

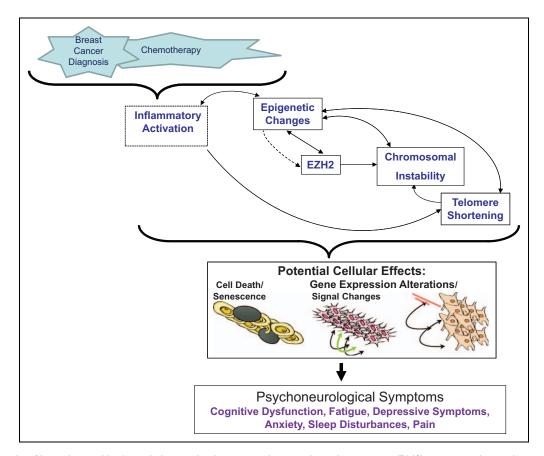


Figure 2. Cascade of hypothesized biological changes leading to psychoneurological symptoms (PNS) in women having breast cancer treated with chemotherapy. Breast cancer and chemotherapy lead to inflammatory activation and may also directly induce epigenetic, telomeric, and/or chromosomal changes. Briefly, the induced inflammatory activation may cause epigenetic changes as well as telomere attrition. Alternatively, epigenetic alterations may occur first, leading to inflammatory activation. These epigenetic changes may involve DNA methylation or histone modifications, possibly through perturbations in the protein enhancer of zeste 2 (EZH2). Changes in EZH2 expression could lead to chromosomal instability as well as other DNA methylation alterations. Telomere shortening, as well as epigenetic alterations, could also directly lead to chromosomal instability. Each of these factors, acting either singly or in concert, could lead to cellular changes. These induced cellular changes could then lead to the presentation and/or persistence of PNS. CRP = C-reactive protein; DNA = deoxyribonucleic acid; E × G interactions = environment × genetic interactions.

having a variety of health and/or psychiatric conditions (including but not limited to Alzheimer's disease, Parkinson disease, and schizophrenia) have higher levels of acquired chromosomal abnormalities (genetic imbalance) than age-matched controls in both organ-specific tissues (e.g., brain cells in people having psychopathology) and peripheral tissues (most frequently peripheral blood; Faggioli, Vijg, & Montagna, 2011; Iourov, Vorsanova, & Yurov, 2008a, 2008b; Kingsbury et al., 2005). If the cells experience senescence or death in response to the acquired epigenetic/genetic changes, the proliferative potential of the tissue would be compromised, with the effects potentially leading to physiological alterations that could culminate in phenotypic consequence. Briones and Woods (2011), for example, reported cognitive impairment in response to chemotherapy-induced decreases in cellular proliferation in rats.

In addition to contributing to the development of PNS through their response to environmental stimuli, epigenetic/genetic alterations could explain the persistence of PNS since epigenetic "marks" and chromosomal alterations can be stably retained for years and through multiple cell divisions in mitotically active cells (Feinberg et al., 2010). Similarly, this phenomenon could explain how epigenetic changes in postmitotic cells, such as neurons, could potentially contribute to long-term, treatment-related symptoms. The plasticity and sensitivity of epigenetic responses to genetic background, different environment influences, and the dynamic interplay between genetics and environment ($G \times E$ interaction) could explain why PNS only develop in a subset of women with BC who have had chemotherapy. Thus, epigenetic and chromosomal alterations fully meet the criteria necessary for providing a means to "biologically remember" the effects of chemotherapy (or the cancer itself).

Examples From Pilot Data

To date, there are no published reports directly testing our hypothesized association between the development and/or persistence of chemotherapy-induced PNS and epigenetic alterations, telomere attrition, or chromosomal instability. However, preliminary data from our pilot studies have shown that at least a subset of telomeres have alterations in length following

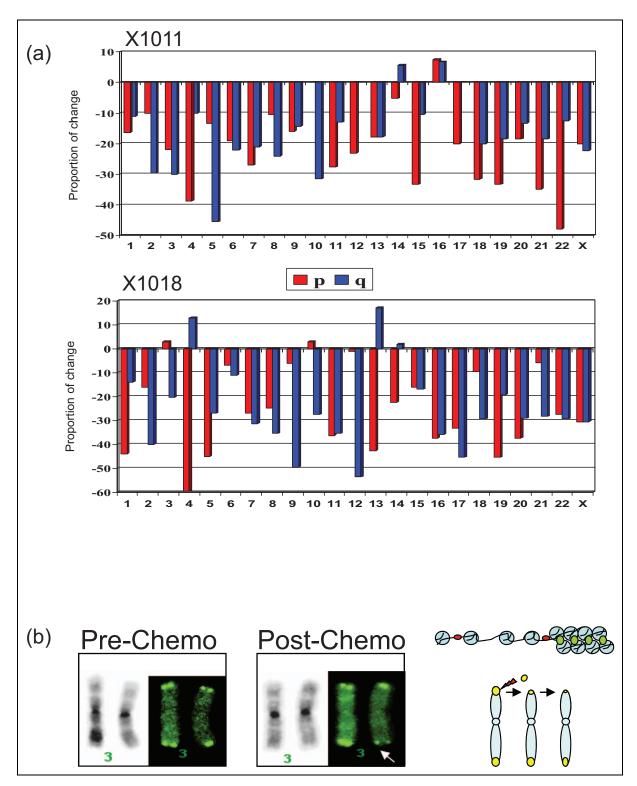


Figure 3. Differential chromosomal and epigenetic alterations acquired following chemotherapy for breast cancer in women evaluated in pilot study. (a) A chromosome-specific fluorescent in situ hybridization methodology was used to assess telomere length variation in lymphocyte chromosomes of two women before and after chemotherapy treatment for breast cancer. Genome-wide chromosome-specific telomere shortening (short arm [p] and long arm [q]) was seen for both women studied (X1011 [top panel] and X1018 [middle panel]), visualized by difference values less than 0. (b) To illustrate the method used for the telomere length determinations, a partial karyotype of the chromosomes 3 from case X1018 is shown (bottom left). Similar telomere intensities are present for the short arm (top of chromosomes), but the telomere was less bright on one of the long-arm telomeres, indicating that shortening occurred (arrow). A schematic illustrating how acquired telomeric shortening could lead to "opening" of nearby chromatin is also shown (bottom right).

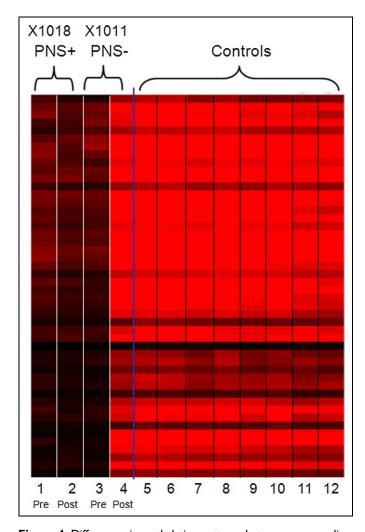


Figure 4. Differences in methylation patterns between women diagnosed with breast cancer and age-matched controls without cancer. This heatmap shows the methylation status of a subset of loci from 12 peripheral blood specimens. Columns 1 (pretreatment) and 2 (15 weeks posttreatment) show values from woman X1018, who had persistent psychoneurological symptoms (PNS). Columns 3 (pretreatment) and 4 (posttreatment) show the values from woman XI011, who did not have persistent PNS. The remaining columns (columns 5-12) are specimens collected from eight different healthy control subjects. Each row (Y axis) represents a different genetic locus, with a total of 48 loci being shown in this subset. The methylation status for each gene/gene region is quantified as a continuous number ranging from 0 to 1, with the value indicated by a shade of red to black, respectively. Both of the pretreatment specimens from the women with a diagnosis of breast cancer (Columns I and 3) had a different pattern (black-toned) from those seen in healthy, age-matched controls (Columns 5-12; red-toned patterns). The posttreatment methylation pattern (Column 4) for woman X1011 (no PNS) became more similar to that of the normal controls. In contrast, the posttreatment methylation pattern (Column 2) for woman X018 (who had persistent PNS) remained similar to the pretreatment pattern (dark-toned pattern).

chemotherapy, that epigenetic alterations are present in the leukocytes of women with BC compared to age-matched controls without BC, and that these epigenetic patterns can be altered in some women after chemotherapy (Figures 3 and 4). We used a chromosome-specific fluorescent in situ hybridization (FISH) methodology to assess telomere length variation in lymphocyte chromosomes of two women diagnosed with BC, completing evaluations before treatment and 15 weeks after chemotherapy (Figure 3). We found genome-wide chromosome-specific telomere shortening for both women and noted no clear pattern of chromosome-specific loss.

We also observed differences in methylation patterns (Figure 4) between women having a diagnosis of BC and age-matched controls without BC. Excitingly, our anecdotal assessment of two women receiving chemotherapy for their BC showed that the methylation signature for a subset of genes evaluated in a genome-wide analysis reverted back to a pattern similar to that of the "normal" controls 15 weeks after chemotherapy initiation for a woman who did not develop debilitating side effects. In contrast, a woman who experienced persistent adverse significant side effects maintained an aberrant methylation pattern 15 weeks following the initiation of chemotherapy. We currently have an ongoing longitudinal study of a larger cohort of women receiving treatment for BC to formally test the hypothesis presented in this article.

Implications for the Development of Future Biomarkers for Biobehavioral Research

We predict that our and other researchers' future investigations to test this hypothesized relationship of epigenetic alternations to the development and persistence of PNS will reveal biomarkers that will allow for the identification of individuals having an elevated risk of developing PNS when treated for BC. Given that there are numerous epigenetic mechanisms controlling gene expression within cells, it seems likely that multiple predictive biomarkers will be identified and that these biomarkers may ultimately provide a basis for a personalized medicine approach for empowering women to make informed decisions regarding treatment options. However, in concert with research to examine epigenetic mechanisms, further study of the incidence, prevalence, and co-occurrence of symptoms is needed to reach a consensus on the nature of symptom clusters.

Additional research may also culminate in an understanding of the molecular/cellular basis for PNS development and/or persistence. For example, discovering hypermethylation of DNA in promoter regions of genes implicated in DNA repair as well as neural cell function, memory, and other cognitive functions could guide future studies leading to the recognition of genes underpinning the development of PNS and may also lead to the discovery of biomarkers most amenable to diagnostic testing (i.e., those with a more rapid turnaround time and/or lower cost than a genome-wide platform). The facts that EZH2, a key epigenetic mediator, has been implicated in DNA repair (Stefansson & Esteller, 2011) and that basal levels of acquired chromosomal instability/DNA damage in peripheral blood lymphocytes are higher in a subset of untreated patients having BC compared to healthy women (Parrella, 2010) lend additional credence to our pursuit of these end points as potential predictive biomarkers related to the development of PNS symptoms.

Finally, since epigenetic changes can be reversed (Esteller, 2005; Issa, Shen, & Toyota, 2005; Mehler, 2008; Muotri & Gage, 2006), the recognition of clinically relevant epigenetic alterations could lead to the development of effective therapeutic interventions. By exploiting the knowledge gained about acquired epigenetic changes, researchers could develop targeted interventions to reduce stress, ameliorate symptoms, enhance quality of life, and possibly prevent or reverse the adverse health outcomes associated with BC and its treatment during the acute treatment phase as well as the survivorship period.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: R01 NR012667, Lyon/Jackson-Cook (MPI), 10/01/10–8/30/15, "Epigenetics and Psychoneurologic Symptoms in Women with Breast Cancer." P30 NR011403, Grap (PI), 8/10/2009–05/1/2014) Biobehavioral Science Core, CBCR Center of Excellence.

References

- Al-Moghrabi, N., Al-Qasem, A. J., & Aboussekhra, A. (2011). Methylation-related mutations in the BRCA1 promoter in peripheral blood cells from cancer-free women. *International Journal of Oncology*, 39, 129–135. doi:10.3892/ijo.2011.1021
- Aristei, C., Stracci, F., Guerrieri, P., Anselmo, P., Armellini, R., Rulli, A., & Menghini, A. R. (2009). Frequency of sister chromatid exchanges and micronuclei monitored over time in patients with early-stage breast cancer: Results of an observational study. Cancer Genetics and Cytogenetics, 192, 24–29. doi:10.1016/j. cancergencyto.2009.02.019
- Arrowsmith, C. H., Bountra, C., Fish, P. V., Lee, K., & Schapira, M. (2012). Epigenetic protein families: A new frontier for drug discovery. *Nature Reviews. Drug Discovery*, 11, 384–400. doi:10.1038/nrd3674
- Aviv, A., & Aviv, H. (1998). Telomeres, hidden mosaicism, loss of heterozygosity, and complex genetic traits. *Human Genetics*, 103, 2–4.
- Bachmann, I. M., Halvorsen, O. J., Collett, K., Stefansson, I. M., Straume, O., Haukaas, S. A., & Akslen, L. A. (2006). EZH2 expression is associated with high proliferation rate and aggressive tumor subgroups in cutaneous melanoma and cancers of the endometrium, prostate, and breast. *Journal of Clinical Oncology*, 24, 268–273. doi:10.1200/JCO.2005.01.5180
- Badger, T., Segrin, C., Dorros, S. M., Meek, P., & Lopez, A. M. (2007). Depression and anxiety in women with breast cancer and their partners. *Nursing Research*, 56, 44–53.
- Bartsch, H., & Nair, J. (2006). Chronic inflammation and oxidative stress in the genesis and perpetuation of cancer: Role of lipid peroxidation, DNA damage, and repair. *Langenbeck's Archives of Surgery*, 391, 499–510. doi:10.1007/s00423-006-0073-1
- Berger, A. M., Wielgus, K., Hertzog, M., Fischer, P., & Farr, L. (2010). Patterns of circadian activity rhythms and their

- relationships with fatigue and anxiety/depression in women treated with breast cancer adjuvant chemotherapy. *Support Care in Cancer*, 18, 105–114. doi:10.1007/s00520-009-0636-0
- Biegler, K. A., Anderson, A. K. L., Wenzel, L. B., Osann, K., & Nelson, E. L. (2012). Longitudinal change in telomere length and the chronic stress response in a randomized pilot biobehavioral clinical study: Implications for cancer prevention. *Cancer Preven*tion Research, 10, 1173–1182. doi:10.1158/1940-6207.CAPR-12-0008
- Bilban-Jakopin, C., & Bilban, M. (2001). Genotoxic effects of radiotherapy and chemotherapy on circulating lymphocytes in patients with Hodgkin's disease. *Mutation Research*, 497, 81–88.
- Bosveil, R., Durif, J., Guo, J., Mebrek, M., Kwiatkowski, F., Bignon, Y. J., & Bernard-Gallon, D. J. (2012). BRCA2 promoter hypermethylation in sporadic breast cancer. *OMICS: A Journal of Integrative Biology*, 16, 707–710.
- Bower, J. E., Ganz, P. A., Aziz, N., & Fahey, J. L. (2002). Fatigue and proinflammatory cytokine activity in breast cancer survivors. *Psychosomatic Medicine*, 64, 604–611.
- Briones, T. L., & Woods, J. (2011). Chemotherapy-induced cognitive impairment is associated with decreases in cell proliferation and histone modifications. *BMC Neuroscience*, 12, 124. doi:10.1186/ 1471-2202-12-124
- Butcher, D. T., & Rodenhiser, D. I. (2007). Epigenetic inactivation of BRCA1 is associated with aberrant expression of CTCF and DNA methyltransferase (DNMT3B) in some sporadic breast tumours. *European Journal of Cancer*, *43*, 210–219.
- Buttiglieri, S., Ruella, M., Risso, A., Spatola, T., Silengo, L., Avvedimento, E. V., & Tarella, C. (2011). The aging effect of chemotherapy on cultured human mesenchymal stem cells. *Experimental Hematology*, 39, 1171–1181. doi:10.1016/j.exphem.2011.08.009
- Cardinale, F., Bruzzi, P., & Bolognesi, C. (2012). Role of micronucleus test in predicting breast cancer susceptibility: A systematic review and meta-analysis. *British Journal of Cancer*, 106, 780–790. doi:10.1038/bjc.2011.567
- Carey, N., Marques, C. J., & Reik, W. (2011). DNA demethylases: A new epigenetic frontier in drug discovery. *Drug Discovery Today*, 16, 683–690. doi:10.1016/j.drudis.2011.05.004
- Chang, C. J., Yang, J. Y., Xia, W., Chen, C. T., Xie, X., & Chao, C. H., ... Hung M. C. (2011). EZH2 promotes expansion of breast tumor initiating cells through activation of RAF1-beta-catenin signaling. *Cancer Cell*, 19, 86–100. doi:10. 1016/j.ccr.2010.10.035
- Cherkas, L. F., Aviv, A., Valdes, A. M., Hunkin, J. L., Gardner, J. P., & Surdulescu, G. L., ... Spector T. D. (2006). The effects of social status on biological aging as measured by white-blood-cell telomere length. *Aging Cell*, 5, 361–365. doi:10.1111/j.1474-9726. 2006.00222.x
- Chia, N., Wang, L., Lu, X., Senut, M. C., Brenner, C., & Ruden, D. M.
 (2011). Hypothesis: Environmental regulation of 5-hydroxymethylcytosine by oxidative stress. *Epigenetics*, 6, 853–856.
- Cleeland, C. S., Bennett, G. J., Dantzer, R., Dougherty, P. M., Dunn, A. J., Meyers, C. A., & Lee, B. N. (2003). Are the symptoms of cancer and cancer treatment due to a shared biologic mechanism? A cytokine-immunologic model of cancer symptoms. *Cancer*, 97, 2919–2925. doi:10.1002/cncr.11382

- Collado-Hidalgo, A., Bower, J. E., Ganz, P. A., Irwin, M. R., & Cole, S. W. (2008). Cytokine gene polymorphisms and fatigue in breast cancer survivors: Early findings. *Brain, Behavior, and Immunity*, 22, 1197–1200. doi:10.1016/j.bbi.2008.05.009
- Collett, K., Eide, G. E., Arnes, J., Stefansson, I. M., Eide, J., & Braaten, A., ... Akslen L. A. (2006). Expression of enhancer of zeste homologue 2 is significantly associated with increased tumor cell proliferation and is a marker of aggressive breast cancer. *Clinical Cancer Research*, 12, 1168–1174. doi:10.1158/1078-0432.ccr-05-1533
- Coussens, L. M., & Werb, Z. (2002). Inflammation and cancer. *Nature*, *420*, 860–867. doi:10.1038/nature01322
- Damjanovic, A. K., Yang, Y., Glaser, R., Kiecolt-Glaser, J. K., Nguyen, H., & Laskowski, B., ... Weng N. P. (2007). Accelerated telomere erosion is associated with a declining immune function of caregivers of Alzheimer's disease patients. *Journal of Immunology*, 179, 4249–4254.
- Dantzer, R., O'Connor, J. C., Freund, G. G., Johnson, R. W., & Kelley, K. W. (2008). From inflammation to sickness and depression: When the immune system subjugates the brain. *Nature Reviews*. *Neuroscience*, 9, 46–56. doi:10.1038/nrn2297
- Day, J. J., & Sweatt, J. D. (2011). Cognitive neuroepigenetics: A role for epigenetic mechanisms in learning and memory. *Neurobiology* of *Learning and Memory*, 96, 2–12. doi:10.1016/j.nlm.2010.12. 008
- De Vivo, I., Prescott, J., Wong, J. Y., Kraft, P., Hankinson, S. E., & Hunter, D. J. (2009). A prospective study of relative telomere length and postmenopausal breast cancer risk. *Cancer Epidemiol*ogy *Biomarkers & Prevention*, 18, 1152–1156.
- Dedert, E., Lush, E., Chagpar, A., Dhabhar, F. S., Segerstrom, S. C., & Spiegel, D., ... Sephton S. E. (2012). Stress, coping, and circadian disruption among women awaiting breast cancer surgery. *Annals of Behavioral Medicine*, 44, 10–20. doi:10.1007/s12160-012-9352-y
- Dedeurwaerder, S., Desmedt, C., Calonne, E., Singhal, S., Haibe-Kains, B., & Defrance, M., ... Fuks F. (2011). DNA methylation profiling reveals a predominant immune component in breast cancers. *EMBO Molecular Medicine*, *3*, 726–741. doi:10.1002/emmm. 201100801
- Diehl, M. C., Idowu, M. O., Kimmelshue, K. N., York, T. P., Jackson-Cook, C. K., & Turner, K. C., ... Elmore L. W. (2011). Elevated TRF2 in advanced breast cancers with short telomeres. *Breast Cancer Research and Treatment*, 127, 623–630. doi:10.1007/s10549-010-0988-7
- Ding, L., Erdmann, C., Chinnaiyan, A. M., Merajver, S. D., & Kleer, C. G. (2006). Identification of EZH2 as a molecular marker for a precancerous state in morphologically normal breast tissues. *Cancer Research*, 66, 4095–4099. doi:10.1158/0008-5472.can-05-4300
- Dodd, M. J., Cho, M. H., Cooper, B. A., & Miaskowski, C. (2010). The effect of symptom clusters on functional status and quality of life in women with breast cancer. *European Journal of Oncology Nursing*, *14*, 101–110. doi:10.1016/j.ejon.2009.09.005
- Effros, R. B. (2011). Telomere/telomerase dynamics within the human immune system: Effect of chronic infection and stress. *Experimental Gerontology*, 46, 135–140.
- Elsendoorn, T. J., Weijl, N. I., Mithoe, S., Zwinderman, A. H., van Dam, F., & De Zwart, F. A., ... Osanto S. (2001). Chemotherapy-induced chromosomal damage in peripheral blood lymphocytes of cancer

- patients supplemented with antioxidants or placebo. *Mutation Research*, 498, 145–158.
- Epel, E. S. (2009). Psychological and metabolic stress: A recipe for accelerated cellular aging? *Hormones*, 8, 7–22.
- Epel, E. S., Blackburn, E., Lin, J., Dhabhar, F. S., Adler, N. E., Morrow, J. D., & Cawthon, R. M. (2004). Accelerated telomere shortening in response to exposure to life stress. *Proceedings on the National Academy of Sciences*, 101, 7312–17315.
- Esteller, M. (2005). Aberrant DNA methylation as a cancer-inducing mechanism. *Annual Review of Pharmacology and Toxicology*, 45, 629–656.
- Esteller, M. (2008). Epigenetics in cancer. New England Journal of Medicine, 358, 1148–1159. doi:10.1056/NEJMra072067
- Faggioli, F., Vijg, J., & Montagna, C. (2011). Chromosomal aneuploidy in the aging brain. *Mechanisms of Ageing and Development*, 132, 429–436. doi:10.1016/j.mad.2011.04.008
- Fang, F., Turcan, S., Rimner, A., Kaufman, A., Giri, D., & Morris, L. G. T., ... Chan T. A. (2011). Breast cancer methylomes establish an epigenomic foundation for metastasis. *Science Translational Medicine*, 3, 75ra25. doi:10.1126/scitranslmed.3001875
- Feinberg, A. P., Irizarry, R. A., Fradin, D., Aryee, M. J., Murakami, P., & Aspelund, T., ... Fallin, M. D. (2010). Personalized epigenomic signatures that are stable over time and covary with body mass index. *Science Translational Medicine*, 2, 49–67.
- Fitzpatrick, D. A., Kronmal, R. A., Gardner, J. P., Psaty, B. M., Jenny, N. S., & Tracy, R. P., ... Aviv A. (2007). Leukocyte telomere length and cardiovascular disease in the cardiovascular health study. *American Journal of Epidemiology*, 165, 14–21.
- Flanary, B. E., & Streit, W. J. (2004). Progressive telomere shortening occurs in cultured rat microglia, but not astrocytes. *Glia*, 45, 75–88. doi:10.1002/glia.10301
- Goedendorp, M. M., Gielissen, M. F., Verhagen, C. A., Peters, M. E., & Bleijenberg, G. (2008). Severe fatigue and related factors in cancer patients before the initiation of treatment. *British Journal of Cancer*, 99, 1408–1414. doi:10.1038/sj.bjc.6604739
- Goel, S., Bhatia, A., & Dey, P. (2011). Spontaneously occurring micronuclei in infiltrating ductal carcinoma of breast: A potential biomarker for aggressive phenotype detection? *Diagnostic Cyto*pathology, 1–7. doi:10.1002/dc.21836
- Gong, Y., Huo, L., Liu, P., Sneige, N., Sun, X., & Ueno, N. T., ... Cristofanilli M. (2011). Polycomb group protein EZH2 is frequently expressed in inflammatory breast cancer and is predictive of worse clinical outcome. *Cancer*, 117, 5476–5484. doi:10.1002/cncr.26179
- Gonzalez, M. E., Li, X., Toy, K., DuPrie, M., Ventura, A. C., & Banerjee, M., ... Kleer C. G. (2009). Downregulation of EZH2 decreases growth of estrogen receptor-negative invasive breast carcinoma and requires BRCA1. *Oncogene*, 28, 843–853. doi:10.1038/onc.2008.433
- Gramatges, M. M., Telli, M. L., Balise, R., & Ford, J. M. (2010). Longer relative telomere length in blood from women with sporadic and familial breast cancer compared with healthy controls. *Cancer Epidemiology Biomarkers & Prevention*, 19, 605–613. doi:10. 1158/1055-9965.epi-09-0896
- Greer, E. L., & Shi, Y. (2012). Histone methylation: A dynamic mark in health, disease and inheritance. *Nature Reviews Genetics*, *13*, 343–357.

- Grzenda, A., Ordog, T., & Urrutia, R. (2011). Polycomb and the emerging epigenetics of pancreatic cancer. *Journal of Gastrointestinal Cancer*, 42, 100–111.
- Gupta, R., Nagarajan, A., & Wajapeyee, N. (2010). Advances in genome-wide DNA methylation analysis. *BioTechniques*, 49, iii–xi. doi:10.2144/000113493
- Hansen, J. A., Feuerstein, M., Calvio, L. C., & Olsen, C. H. (2008). Breast cancer survivors at work. *Journal of Occupational and Environmen*tal Medicine, 50, 777–784. doi:10.1097/JOM.0b013e318165159e
- Hewitt, G., Jurk, D., Marques, F. D. M., Correia-Melo, C., Hardy, T., & Gackowska, A., ... Passos J. F. (2012). Telomeres are favoured targets of a persistent DNA damage response in ageing and stressinduced senescence. *Nature Communications*, 3, 708.
- Huang, Y., Nayak, S., Jankowitz, R., Davidson, N., & Oesterreich, S. (2011). Epigenetics in breast cancer: What's new? *Breast Cancer Research*, 13, 225.
- Humphreys, K., Epel, E. S., Cooper, B. A., Lin, J., Blackburn, E. H., & Lee, K. A. (2012). Telomere shortening in formerly abused and never abused women. *Biological Research for Nursing*, 14, 115–123. doi:10.1177/1099800411398479
- Iourov, I. Y., Vorsanova, S. G., & Yurov, Y. B. (2008a). Chromosomal mosaicism goes global. *Molecular Cytogenetics*, 1, 26. doi: 10.1186/1755-8166-1-26
- Iourov, I. Y., Vorsanova, S. G., & Yurov, Y. B. (2008b). Molecular cytogenetics and cytogenomics of brain diseases. *Current Genomics*, 9, 452–465. doi:10.2174/138920208786241216
- Issa, J. P. J., Shen, L., & Toyota, M. (2005). CIMP, at last. *Gastroenterology*, 129, 1121–1124. doi:10.1053/j.gastro.2005.07.040
- Iwamoto, T., Yamamoto, N., Taguchi, T., Tamaki, Y., & Noguchi, S. (2011). BRCA1 promoter methylation in peripheral blood cells is associated with increased risk of breast cancer with BRCA1 promoter methylation. *Breast Cancer Research and Treatment*, 129, 69–77. doi:10.1007/s10549-010-1188-1
- Jackson-Cook, C. (2011). Constitutional and acquired autosomal aneuploidy. *Clinics in Laboratory Medicine*, 31, 481–511. doi: 10.1016/j.cll.2011.08.002
- Jacobs, D. I., Hansen, J., Fu, A., Stevens, R. G., Tjonneland, A., & Vogel, U. B., ... Zhu Y. (2013). Methylation alterations at imprinted genes detected among long-term shiftworkers. *Environmental and Molecular Mutagenesis*, 54, 141–146.
- Janssen, A., & Medema, R. H. (2012). Genetic instability: Tipping the balance. *Oncogene*, 1–12. doi:10.1038/onc.2012.576
- Jin, S. G., Wu, X., Li, A. X., & Pfeifer, G. P. (2011). Genomic mapping of 5-hydroxymethylcytosine in the human brain. *Nucleic Acids Research*, 39, 5015–5024. doi:10.1093/nar/gkr120
- Jones, P. A., & Liang, G. (2009). Rethinking how DNA methylation patterns are maintained. *Nature Reviews Genetics*, 10, 805–811. doi:10.1038/nrg2651
- Jovanovic, J., Rønneberg, J. A., Tost, J., & Kristensen, V. (2010). The epigenetics of breast cancer. *Molecular Oncology*, 4, 242–254. doi: 10.1016/j.molonc.2010.04.002
- Kelley, K. W., Bluthe, R. M., Dantzer, R., Zhou, J. H., Shen, W. H., Johnson, R. W., & Broussard, S. R. (2003). Cytokine-induced sickness behavior. *Brain, Behavior, and Immunity*, 17, S112–118.
- Kim, S., Bi, X., Czarny-Ratajczak, M., Dai, J., Welsh, D. A., & Myers, L., ... Jazwinski S. M. (2011). Telomere maintenance genes

- SIRT1 and XRCC6 impact age-related decline in telomere length but only SIRT1 is associated with human longevity. *Biogerontology*, *13*, 119–131. doi:10.1007/s10522-011-9360-5
- Kingsbury, M. A., Friedman, B., McConnell, M. J., Rehen, S. K., Yang, A. H., Kaushal, D., & Chun, J. (2005). Aneuploid neurons are functionally active and integrated into brain circuitry. *Proceedings of the National Academy of Sciences of the United States of America*, 102, 6143–6147. doi:10.1073/pnas.0408171102
- Kirsch-Volders, M., Plas, G., Elhajouji, A., Lukamowicz, M., Gonzalez, L., Vande Loock, K., & Decordier, I. (2011). The in vitro MN assay in 2011: Origin and fate, biological significance, protocols, high throughput methodologies and toxicological relevance. *Archives of Toxicology*, 85, 873–899. doi:10.1007/s00204-011-0691-4
- Kleer, C. G., Cao, Q., Varambally, S., Shen, R., Ota, I., & Tomlins, S. A.,... Chinnaiyan A. M. (2003). EZH2 is a marker of aggressive breast cancer and promotes neoplastic transformation of breast epithelial cells. *Proceedings of the National Academy of Sciences*, 100, 11606–11611. doi:10.1073/pnas.1933744100
- Kouzarides, T. (2007). Chromatin modifications and their function. *Cell*, 128, 693–705. doi:10.1016/j.cell.2007.02.005
- Kriaucionis, S., & Heintz, N. (2009). The nuclear DNA base 5hydroxymethylcytosine is present in Purkinje neurons and the brain. Science, 324, 929–930. doi:10.1126/science.1169786
- Kume, K., Kikukawa, M., Hanyu, H., Takata, Y., Umahara, T., & Sakurai, H.,... Iwamoto T. (2012). Telomere length shortening in patients with dementia with Lewy bodies. *European Journal of Neurology*, 19, 905–910. doi:10.1111/j.1468-1331.2011.03655
- Lee, K., Cho, M., Miaskowski, C., & Dodd, M. (2004). Impaired sleep and rhythms in persons with cancer. *Sleep Medicine Reviews*, 8, 199–212. doi:10.1016/j.smrv.2003.10.001
- Lubin, F. D., Gupta, S., Parrish, R. R., Grissom, N. M., & Davis, R. L. (2011). Epigenetic mechanisms: Critical contributors to long-term memory formation. *Neuroscientist*, 17, 616–632. doi:10.1177/ 1073858410386967
- Mantovani, A., Allavena, P., Sica, A., & Balkwill, F. (2008). Cancerrelated inflammation. *Nature*, 454, 436–444. doi:10.1038/nature07205
- Martin-Ruiz, C., Dickinson, H. O., Keys, B., Rowan, E., Kenny, R. A., & Von Zglinicki, T. (2006). Telomere length predicts poststroke mortality, dementia, and cognitive decline. *Annals of Neurology*, 60, 174–180. doi:10.1002/ana.20869
- Mathews, H. L., Konley, T., Kosik, K. L., Krukowski, K., Eddy, J., Albuquerque, K., & Janusek, L. W. (2011). Epigenetic patterns associated with the immune dysregulation that accompanies psychosocial distress. *Brain, Behavior, and Immunity*, 25, 830–839. doi:10.1016/j.bbi.2010.12.002
- Mazzio, E. A., & Soliman, K. F. (2012). Basic concepts of epigenetics: Impact of environmental signals on gene expression. *Epigenetics*, 7, 119–130. doi:10.4161/epi.7.2.18764
- McCormick, V. A., & Silva, I. D. S. (2006). Breast density and parenchymal patterns as markers of breast cancer risk: A meta-analysis. *Cancer Epidemiology Biomarkers & Prevention*, 15, 1159–1169. doi:10.1158/1055-9965.epi-06-0034
- Meeker, A. K., & Argani, P. (2004). Telomere shortening occurs early during breast tumorigenesis: A cause of chromosome destabilization underlying malignant transformation? *Journal of Mammary Gland Biological Neoplasia*, 9, 285–296.

- Mehler, M. F. (2008). Epigenetic principles and mechanisms underlying nervous system functions in health and disease. *Progress in Neurobiology*, 86, 305–341.
- Mehnert, A. (2011). Employment and work-related issues in cancer survivors. Critical Reviews in Oncology/Hematology, 77, 109–130. doi:10.1016/j.critrevonc.2010.01.004
- Miller, G. E., Chen, E., & Parker, K. J. (2011). Psychological stress in childhood and susceptibility to the chronic diseases of aging: Moving toward a model of behavioral and biological mechanisms. *Psychological Bulletin*, 137, 959–997. doi:10.1037/a0024768
- Miloševic-Djordjevic, O., Grujicic, D., Vaskovic, Z., & Marinkovic, D. (2010). High micronucleus frequency in peripheral blood lymphocytes of untreated cancer patients irrespective of gender, smoking and cancer sites. *Tohoku Journal of Experimental Medicine*, 220, 115–120.
- Miranda, T. B., & Jones, P. A. (2007). DNA methylation: The nuts and bolts of repression. *Journal of Cellular Physiology*, 213, 384–390. doi:10.1002/jcp.21224
- Mohammed, S. I., Springfield, S., & Das, R. (2012). Role of epigenetics in cancer health disparities. *Methods in Molecular Biology*, 863, 395–410. doi:10.1007/978-1-61779-612-8-25
- Muotri, A., & Gage, F. (2006). Generation of neuronal variability and complexity. *Nature*, 441, 1087–1093.
- O'Donovan, A., Pantell, M. S., Puterman, E., Dhabhar, F. S., Blackburn, E. H., & Yaffe, K., . . . Epel E. S. (2011). Cumulative inflammatory load is associated with short leukocyte telomere length in the Health, Aging and Body Composition Study. *PloS One*, 6, e19687.
- O'Donovan, A., Tomiyama, A. J., Lin, J., Puterman, E., Adler, N. E., & Kemeny, M., . . . Epel E. S. (2012). Stress appraisals and cellular aging: A key role for anticipatory threat in the relationship between psychological stress and telomere length. *Brain, Behavior, and Immunity*, 26, 573–579. doi:10.1016/j.bbi.2012.01.007
- O'Sullivan, R. J., & Karlseder, J. (2010). Telomeres: Protecting chromosomes against genome instability. *Nature Reviews Molecular Cell Biology*, 11, 171–181. doi:10.1038/nrm2848
- Pampalona, J., Soler, D., Genescà, A., & Tusell, L. (2010). Whole chromosome loss is promoted by telomere dysfunction in primary cells. *Genes, Chromosomes and Cancer*, 49, 368–378. doi:10. 1002/gcc.20749
- Parks, C. G., Miller, D. B., McCanlies, E. C., Cawthon, R. M., Andrew, M. E., DeRoo, L. A., & Sandler, D. P. (2009). Telomere length, current perceived stress, and urinary stress hormones in women. *Cancer Epidemiology Biomarkers & Prevention*, 18, 551–560. doi:10.1158/1055-9965.epi-08-0614
- Parrella, P. (2010). Epigenetic signatures in breast cancer: Clinical perspective. *Breast Care*, 5, 66–73. doi:10.1159/000309138
- Peres, J. (2011). Telomere research offers insight on stress-disease link. *Journal of The National Cancer Institute*, 103, 848–850. doi:10.1093/jnci/djr214
- Pooley, K. A., Sandhu, M. S., Tyrer, J., Shah, M., Driver, K. E., & Luben, R. N., ... Dunning A. M. (2010). Telomere length in prospective and retrospective cancer case-control studies. *Cancer Research*, 70, 3170–3176. doi:10.1158/0008-5472.can-09-4595
- Ptak, C., & Petronis, A. (2008). Epigenetics and complex disease: From etiology to new therapeutics. *Annual Review of Pharmacology and Toxicology*, 48, 257–276.

- Puterman, E., Lin, J., Blackburn, E., O'Donovan, A., Adler, N., & Epel, E. (2010). The power of exercise: Buffering the effect of chronic stress on telomere length. *PLoS One*, 5, e10837.
- Quivy, V., Calomme, C., Dekoninck, A., Demonte, D., Bex, F., & Lamsoul, I., ... Van Lint C. (2004). Gene activation and gene silencing: A subtle equilibrium. *Cloning and Stem Cells*, 6, 140–149. doi:10.1089/1536230041372454
- Raaphorst, F. M., Meijer, C. J., Fieret, E., Blokzijl, T., Mommers, E., & Buerger, H., ... van Diest P. J. (2003). Poorly differentiated breast carcinoma is associated with increased expression of the human polycomb group EZH2 gene. *Neoplasia*, 5, 481–488.
- Raison, C. L., Capuron, L., & Miller, A. H. (2006). Cytokines sing the blues: Inflammation and the pathogenesis of depression. *Trends in Immunology*, 27, 24–31. doi:10.1016/j.it.2005.11.006
- Roberts, R. O., Geda, Y. E., Knopman, D. S., Cha, R. H., Boeve, B. F., & Ivnik, R. J., ... Petersen R. C. (2010). Metabolic syndrome, inflammation, and nonamnestic mild cognitive impairment in older persons: A population-based study. *Alzheimer Disease and Associated Disorders*, 24, 11–18. doi:10.1097/WAD.0b013e3181a4485c
- Rush, M., Appanah, R., Lee, S., Lam, L. L., Goyal, P., & Lorincz, M. C. (2009). Targeting of EZH2 to a defined genomic site is sufficient for recruitment of Dnmt3a but not de novo DNA methylation. *Epigenetics*, 4, 404–414.
- Samanta, S., Dey, P., & Nijhawan, R. (2011). The role of micronucleus scoring in fine needle aspirates of ductal carcinoma of the breast. *Cytopathology*, 22, 111–114. doi:10.1111/j.1365-2303. 2010.00773
- Santos, F., Hyslop, L., Stojkovic, P., Leary, C., Murdoch, A., & Reik, W., ... Dean W. (2010). Evaluation of epigenetic marks in human embryos derived from IVF and ICSI. *Human Reproduction*, *25*, 2387–2395.
- Schlesinger, Y., Straussman, R., Keshet, I., Farkash, S., Hecht, M., & Zimmerman, J., ... Cedar H. (2006). Polycomb-mediated methylation on Lys27 of histone H3 pre-marks genes for de novo methylation in cancer. *Nature Genetics*, 39, 232–236. doi:10.1038/ng1950
- Schmidt, R. J., Hansen, R. L., Hartiala, J., Allayee, H., Schmidt, L. C., & Tancredi, D. J., ... Hertz-Picciotto I. (2011). Prenatal vitamins, one-carbon metabolism gene variants, and risk for autism. *Epide-miology*, 22, 476–485. doi:10.1097/EDE.0b013e31821d0e30
- Schroeder, C. P., Wisman, G. B., de Jong, S., van der Graaf, W. T., Ruiters, M. H., Mulder, N. H., & de Vries, E. G. (2001). Telomere length in breast cancer patients before and after chemotherapy with or without stem cell transplantation. *British Journal of Cancer*, 18, 1348–1353.
- Sephton, S., & Spiegel, D. (2003). Circadian disruption in cancer: A neuroendocrine-immune pathway from stress to disease? *Brain, Behavior, and Immunity*, 17, 321–328.
- Serrano, A. L., & Andrés, V. (2004). Telomeres and cardiovascular disease: Does size matter? *Circulation Research*, 94, 575–584. doi:10.1161/01.RES.0000122141.18795.9C
- Shen, J., Terry, M. B., Gurvich, I., Liao, Y., Senie, R. T., & Santella, R. M. (2007). Short telomere length and breast cancer risk: A study in sister sets. *Cancer Research*, 67, 5538–5544. doi:10.1158/0008-5472.can-06-3490

- Sibille, K. T., Langaee, T., Burkley, B., Gong, Y., Glover, T. L., & King, C., ... Fillingim R. B. (2012). Chronic pain, perceived stress, and cellular aging: An exploratory study. *Molecular Pain*, 8, 12. doi:10.1186/1744-8069-8-12
- Siegel, R., Naishadham, D., & Jemal, A. (2012). Cancer statistics, 2012. CA: A Cancer Journal for Clinicians, 62, 10–29. doi:10. 3322/caac.20138
- Simon, J. A., & Lange, C. A. (2008). Roles of the EZH2 histone methyltransferase in cancer epigenetics. *Mutation Research*, 647, 21–29. doi:10.1016/j.mrfmmm.2008.07.010
- Simon, N. M., Smoller, J. W., McNamara, K. L., Maser, R. S., Zalta, A. K., & Pollack, M. H., ... Wong K. K. (2006). Telomere shortening and mood disorders: Preliminary support for a chronic stress model of accelerated aging. *Biological Psychiatry*, 60, 432–435. doi:10.1016/j.biopsych.2006.02.004
- Stefansson, O. A., & Esteller, M. (2011). EZH2-mediated epigenetic repression of DNA repair in promoting breast tumor initiating cells. *Breast Cancer Research*, 13, 309. doi:10.1186/bcr2871
- Strub, G. M., Depcrynski, A., Elmore, L. W., & Holt, S. E. (2008). Recovery from stress is a function of age and telomere length. *Cell Stress and Chaperones*, 13, 475–482.
- Tahiliani, M., Koh, K. P., Shen, Y., Pastor, W. A., Bandukwala, H., & Brudno, Y., ... Rao A. (2009). Conversion of 5-methylcytosine to 5-hydroxymethylcytosine in mammalian DNA by MLL partner TET1. *Science*, *324*, 930–935. doi:10.1126/science.1170116
- Terry, M. B., Delgado-Cruzata, L., Vin-Raviv, N., Wu, H. C., & Santella, R. M. (2011). DNA methylation in white blood cells: Association with risk factors in epidemiologic studies. *Epigenetics*, 6, 828–837.
- Toyokawa, S., Uddin, M., Koenen, K. C., & Galea, S. (2012). How does the social environment 'get into the mind'? Epigenetics at the intersection of social and psychiatric epidemiology. *Social Science* and Medicine, 74, 67–74. doi:10.1016/j.socscimed.2011.09.036
- Tsang, D. P., & Cheng, A. S. (2011). Epigenetic regulation of signaling pathways in cancer: Role of the histone methyltransferase EZH2. *Journal of Gastroenterology and Hepatology*, 26, 19–27. doi:10.1111/j.1440-1746.2010.06447.x
- Utne, I., Miaskowski, C., Bjordal, K., Paul, S. M., & Rustoen, T. (2010). The relationships between mood disturbances and pain, hope, and quality of life in hospitalized cancer patients with pain on regularly scheduled opioid analgesic. *Journal of Palliative Medicine*, 13, 311–318. doi:10.1089/jpm.2009.0294
- Valdes, A. M., Deary, I. J., Gardner, J., Kimura, M., Lu, X., & Spector, T. D., ... Cherkas L. F. (2010). Leukocyte telomere length is associated with cognitive performance in healthy women. *Neurobiological Aging*, 31, 986–992. doi:10.1016/j.neurobiolaging.2008.07. 012
- Valeberg, B. T., Rustoen, T., Bjordal, K., Hanestad, B. R., Paul, S., & Miaskowski, C. (2008). Self-reported prevalence, etiology, and characteristics of pain in oncology outpatients. *European Journal* of Pain, 12, 582–590. doi:10.1016/j.ejpain.2007.09.004
- Vardy, J., Wefel, J. S., Ahles, T., Tannock, I. F., & Schagen, S. B. (2008). Cancer and cancer-therapy related cognitive dysfunction: An international perspective from the Venice cognitive workshop. *Annals of Oncology*, 19, 623–629. doi:10.1093/annonc/mdm500

- Varga, D., Hoegel, J., Maier, C., Jainta, S., Hoehne, M., & Patino-Garcia, B., ... Vogel W. (2006). On the difference of micronucleus frequencies in peripheral blood lymphocytes between breast cancer patients and controls. *Mutagenesis*, 21, 313–320. doi:10.1093/mutage/gel035
- Veeck, J., & Esteller, M. (2010). Breast cancer epigenetics: From DNA methylation to microRNAs. *Journal of Mammary Gland Biology and Neoplasia*, 15, 5–17. doi:10.1007/s10911-010-9165-1
- Viré, E., Brenner, C., Deplus, R., Blanchon, L., Fraga, M., & Didelot, C., ... Fuks F. (2006). The Polycomb group protein EZH2 directly controls DNA methylation. *Nature*, 439, 871–874. doi:10.1038/ nature04431
- Walker, S. L., Ariga, J., Mathias, J. R., Coothankandaswamy, V., Xie, X., & Distel, M., ... Mumm J. S. (2012). Automated reporter quantification in vivo: High-throughput screening method for reporter-based assays in zebrafish. *PLoS One*, 7, e29916. doi:10. 1371/journal.pone.0029916
- Wang, X., Wu, X., Liang, Z., Huang, Y., Fenech, M., & Xue, J. (2006). A comparison of folic acid deficiency-induced genomic instability in lymphocytes of breast cancer patients and normal non-cancer controls from a Chinese population in Yunnan. *Muta-genesis*, 21, 41–47. doi:10.1093/mutage/gei069
- Wefel, J. S., Vardy, J., Ahles, T., & Shagan, S. (2011). International cognition and cancer task force recommendations to harmonise studies of cognitive function in patients with cancer. *The Lancet Oncology*, 12, 703–708.
- Wikgren, M., Maripuu, M., Karlsson, T., Nordfjäll, K., Bergdahl, J., & Hultdin, J., ... Norrback K. F. (2012). Short telomeres in depression and the general population are associated with a hypocortisolemic state. *Biological Psychiatry*, 71, 294–300. doi:10.1016/j.biopsych.2011.09.015
- Wilson, A. S., Power, B. E., & Molloy, P. L. (2007). DNA hypomethylation and human diseases. *Biochimica et Biophysica Acta—Reviews* on Cancer, 1775, 138–162. doi:10.1016/j.bbcan.2006.08.007
- Wojda, A., & Witt, M. (2003). Manifestations of ageing at the cytogenetic level. *Journal of Applied Genetics*, 44, 383–400.
- Yaffe, K., Lindquist, K., Kluse, M., Cawthon, R., Harris, T., & Hsueh, W.-C., ... Cummings S. R. (2011). Telomere length and cognitive function in community-dwelling elders: Findings from the Health ABC Study. *Neurobiology of Aging*, 32, 2055–2060. doi:10.1016/j. neurobiolaging.2009.12.006
- Yurov, Y. B., Iourov, I. Y., Vorsanova, S. G., Demidova, I. A., Kravetz, V. S., & Beresheva, A. K., ... Liehr T. (2008). The schizophrenia brain exhibits low-level aneuploidy involving chromosome 1. *Schizophrenia Research*, 98, 139–147. doi:10.1016/j.schres.2007.07.035
- Zeng, Y., Tan, M., Kohyama, J., Sneddon, M., Watson, J. B., Sun, Y. E., & Xie, C. W. (2011). Epigenetic enhancement of BDNF signaling rescues synaptic plasticity in aging. *Journal of Neuroscience*, 31, 17800–17810.
- Zheng, Y.-L., Ambrosone, C., Byrne, C., Davis, W., Nesline, M., & McCann, S. (2010). Telomere length in blood cells and breast cancer risk: Investigations in two case-control studies. *Breast Cancer Research and Treatment*, 120, 769–775. doi:10.1007/s10549-009-0440-z